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Supplementary material

Designing of Coumarinyl-Picolinoyl hydrazide Schiff base for the fluorescence turn-on-off sequential sensing of Al³⁺ and nitroaromatics, and

electronic device fabrication

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Table S1

Sl	Probe		Sensing ion,	Electrical	Ref.
no		AIE	solvent, LOD	conductivity	
1		No	Mg ²⁺ , Ethanol, 105 nM	No	31
2		No	Zn^{2+} (72 nM) and	No	32
			AcO ⁻ (94 nM) in		
	N //		DMSO/H ₂ O		
	HN S		(v/v = 3:7)		
	HOOO				
3		No	Zn ²⁺ , THF, 100 nM	No	33
	HOLOO				
4		No	Zn ²⁺ ,	No	34
	s		MeOH/H ₂ O(v/v,		
			3:1), 78 nM		
	HOLOO				

5	H ₂ N O O N N	No	Zn ²⁺ (26 nM) and ClO ⁻ (2 μ M) in 10 mM HEPES	No	35
6		No	Al ³⁺ , Ethanol, 820 nM	No	36
7	OH N S O O O	No	Zn ²⁺ , CH ₃ CN/H ₂ O (95:5, v/v), 35 nM	No	37
8		No	Ca ²⁺ , DMF/HEPES buffer 1:1(v/v), 33.3 µM	No	38
9		No	Zn ²⁺ (19 nM) and Cu ²⁺ (1.87 nM) in CH ₃ CN : H ₂ O (1 : 1, v/v,)	No	39
10		No	Zn ²⁺ , CH ₃ CN:H ₂ O 9 : 1(v/v), 11 nM	No	40
11	HN N OH	Yes	Al ³⁺ , Water, 6.99 nM	Yes	This work

Materials and methods

Picolinic acid and hydrazine hydrate were purchased from High media and Merck respectively. All other organic chemicals and inorganic salts were obtained from commercial suppliers Merck and used without further purification. Aqueous solutions were prepared using Milli-Q water (Millipore). Elemental analyses were performed using a Perkin-Elmer USA elemental analyzer. UV-vis spectra were recorded on Perkin Elmer Lambda 25 spectrophotometer and fluorescence spectra were obtained using a Perkin Elmer spectrofluorimeter model LS55, Luminescence lifetime measurements were carried out by using time-correlated single photon counting set up from Horiba Jobin-Yvon; FT-IR spectra (KBr disk, 4000-400 cm⁻¹) from a Perkin Elmer LX-1 FTIR spectrophotometer. NMR spectra were obtained on a Bruker (AC) 300 MHz FT-NMR spectrometer using TMS as an internal standard. ESI mass spectra were recorded from a Water HRMS model XEVO-G2QTOF#YCA351 spectrometer. All of the measurements were conducted at room temperature. To calculate band gap we have use the tauc's equation,

$$(\alpha h_{\nu})^2 = A(h_{\nu} - E_g) \tag{1}$$

Where α , E_g , h, v is the absorption coefficient, optical band gap, Planck's constant and frequency of light. A is a constant which is considered as 1 for ideal case. By extrapolating the linear region of the plot $(\alpha hv)^2$ vs. hv (Figure 1) to $\alpha = 0$ absorption and using the above equation, the direct optical band gap of our synthesized compound has been evaluated as 2.78 eV. The dielectric study of our synthesized compound has been performed by the impedance spectrum which has been evaluated as a function of frequency and recorded at room temperature by the computer controlled Agilent make precision 4294A LCR meter at the frequency range 40 Hz to 11 MHz. In this regard we have calculated the capacitance (C), impedance (Z) and phase angle (θ) of the sample.

Preparation of probe, H-CPh

Picolinohydrazide was prepared from Picolinic acid as per report and also 7-hydroxy-4methyl-2-oxo-2H-chromene-8-carbaldehyde was synthesized from 7-hydroxy-4-methylcoumarin following reported method. The condensation of Picolinohydrazide (137 mg, 1.0 mmol) was dissolved in 7 ml MeOH and 7 ml MeOH solution of 7-hydroxy-4-methyl-2-oxo-2H-chromene-8-carbaldehyde (804 mg, 1.0 mmol) was added to amine solution drop by drop. Then the mixture was stirred for 5 hr to get a pale greenish yellow precipitate of (E)-N'-((7hydroxy-4-methyl-2-oxo-2H-chromen-8-yl)methylene)picolinohydrazide (H-CPh). The Collected precipitate of H-CPh was dried in open air. Yield: 88%. M.P. >200 °C (Scheme 1). Microanalytical data: $C_{20}H_{13}N_3O_3$ calcd (found): C, 63.16(63.05); H, 4.05(4.13); N, 13.00(12.85) %. ¹H NMR (300 MHz, DMSO-d₆): 12.95 (s, 1H, NH), 12.93 (s, 1H, -OH), 9.41 (s, 1H, imine-H), 8.74 (d, 1H, 4.5 Hz), 8.15-8.04 (m, 2H), 7.73-7.67 (m, 2H), 6.97 (d, 1H, 8.7 Hz), 6.25 (s, 1H) 2.40 (s, 3H, -CH₃) (Fig. S1); ESI-mass peak for CPh at 323.99 (calculated mass of **CPh**, 323.30) (**Fig. S2**); FTIR: v3461 cm⁻¹(OH), v3299 cm⁻¹(NH), v1685 $cm^{-1}(C=O)$, v1609 $cm^{-1}(C=N)$ (Fig S3)

General method for UV-Vis and fluorescence studies

CPh (1.73 mg, 0.001 mmol) was dissolved in DMSO and diluted to prepare 20 μ L using 2 ml required solvent. The metals solutions (40 μ l) were transferred to solution of **H-CPh**. After mixing both absorption and emission spectra were recorded at room temperature. Fixing excitation slit = 15.0 and emission slit = 5.0, fluorescence experiment had been carried out using excitation wavelength 340 nm for AIEE study and 400 nm for cation sensing study.



Fig S1 Mass spectra of H-CPh

1H_NMR_CPh



Fig S2 1H NMR of H-CPh in DMSO-d₆



Fig S3 FT-IR spectrum of H-CPh



Fig S4 UV-Vis spectra of H-CPh on increasing water percentage in DMSO-water mixture



Fig S5 DLS spectra of H-CPh in DMSO



Fig S6 DLS spectra of H-CPh in 90% water



Fig S7 UV-vis spectrum of H-CPh in water



Fig S8 Fluorescence spectra of H-CPh in presence of various cations in DMSO



Fig S9 Fluorescence spectra of H-CPh in presence of various cations 3:1 DMSO-water



Fig S10 Fluorescence spectra of H-CPh in presence of various cations in 1:1 DMSO-Water



Fig S11 Fluorescence spectra of H-CPh in presence of various cations in 1:3 DMSO-Water



Fig S12 UV-vis spectra of **H-CPh** on incremental addition of Al³⁺ in water (hepes buffer, pH 7.2) medium.

Fig S13 Job'S plot for binding of Al³⁺ with H-CPh by fluoremetric method

Fig S14 Benesi-Hildebrand plot for determining binding constant of Al³⁺ with H-CPh

Fig S15 1H NMR spectra of H-CPh on gradual addition of Al^{3+} in DMSO-d₆

Fig S16 Mass spectrum of Al³⁺ complex

Fig S17 FT-IR spectrum of Al³⁺ complex

Fig S18 Limit of detection determining plot for Al³⁺

Fig S19 Interfering study on Al³⁺ sensitivity

Fig S20 pH dependence fluorescence sensitivity of Al³⁺ by the probe H-CPh

Fig S21 Limit of detection determining plot for TNP

Fig S22 Limit of detection determining plot for DNP

Fig S23 Theoretical electronic transitions of H-CPh and its Al³⁺ complex