

Supporting Information

Side-chain deuterated cholesterol probe as a molecular probe to determine membrane order and cholesterol partitioning

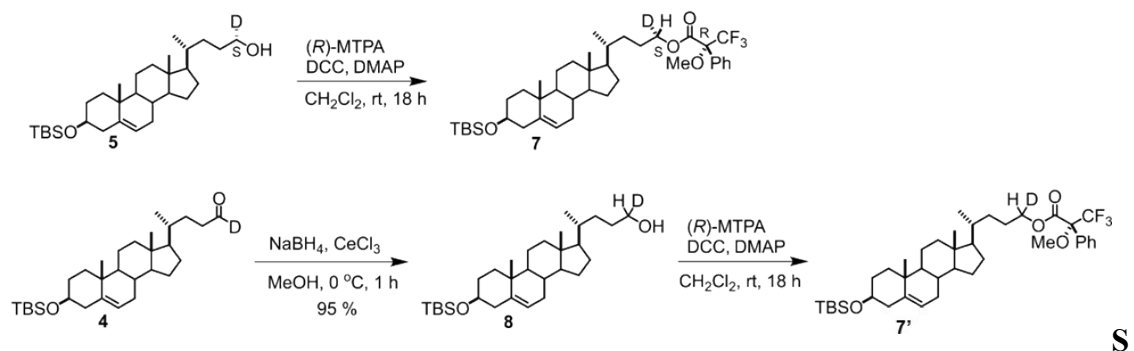
Shinya Hanashima, Yuki Ibata, Hirofumi Watanabe, Tomokazu Yasuda,
Hiroshi Tsuchikawa, Michio Murata

Determination of the stereochemistry and optical purity of the intermediate **5** and **24dCho**

(*R*)-MTPA ester **7** was prepared and the stereochemistry at C24 was determined by comparison of the ¹H NMR spectra with the racemic mixture **7'** (Fig.S1). **7'** was prepared from aldehyde **4** via a reduction with NaBH₄ and CeCl₃ to give alcohol **8** in 95% yield. Then, MTPA ester was introduced to give **7'** as a diastereomeric mixture (Scheme S1).

¹H NMR spectra of **7** and its diastereomeric mixture **7'**, focusing on the H24 signal, are shown in Figure S1. The (*R*)-MTPA group more shields the proton on the *S* isomer than the *R* isomer due to the fixed phenyl group of the MTPA ester. Similar difference of the chemical shifts had been reported in the (*R*)-MTPA ester of (*S*)-1-*d*-1-octanol.^[S1] Thus, the stereochemistry of C24 was determined in *S* configuration. Furthermore, the other stereoisomer was not observed in the ¹H NMR spectrum of **7**, thus the purity could be more than 95% d.e.

The stereochemistry at C24 of 24dCho was again confirmed by comparison of ¹H-¹³C HSQC spectra between 24dCho and native Cho (Fig.S2). In f2 dimension (¹H), the signal at pro-chiral C24-H(*proR*) of Cho was 1.09 ppm, while C24-H(*proS*) was 1.11 ppm (Fig.S2A).^[S2] On the other hand, the signal from the remaining proton at C24 of 24dCho was found at 1.07 ppm with a triplet in f1 dimension because of the geminal ¹³C-²H coupling (¹J_{C-D} = 17.1 Hz) (Fig.S2B). The disagreement of the chemical shifts can be accounted by deuterium induced isotope (DIS) shifts. DIS of ¹H affected from the geminal ²H can be estimated within the range between -0.01 and -0.02 ppm.^[S3] Thus, the stereochemistry at C24 was confirmed as *S* because the C-H correlation corresponding to the C24-H(*proR*) was appeared in the spectrum.



cheme S1. Synthesis of the (*R*)-MTPA ester **7** and the diastereomeric mixture **7'** from aldehyde **4**.

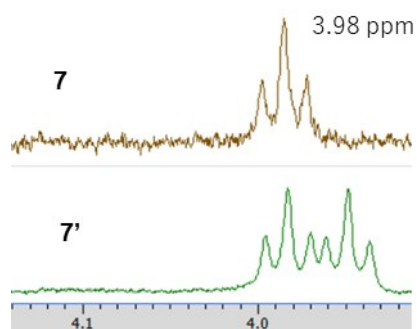


Figure S1. ^1H NMR spectra at C24-H of the synthetic (*R*)-MTPA esters of **7** and its diastereomeric mixture **7'**.

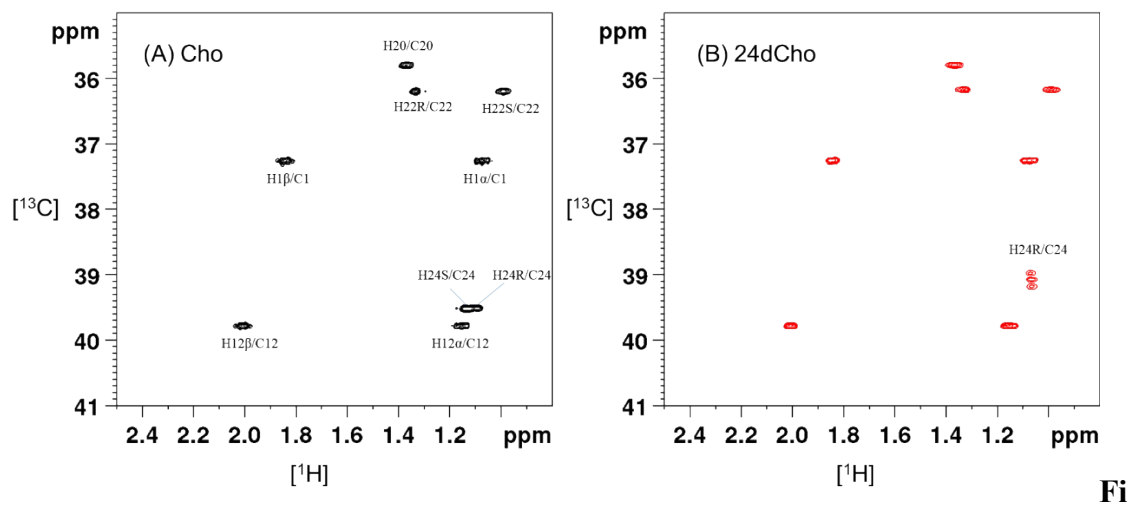


Figure S2. ^1H - ^{13}C HSQC spectra of Cho (A) and 24dCho (B) in CDCl_3 . Only one HSQC correlation was observed from the C24-H of 24dCho. The spectra were collected with 700 MHz spectrometer (Bruker AVANCE700).

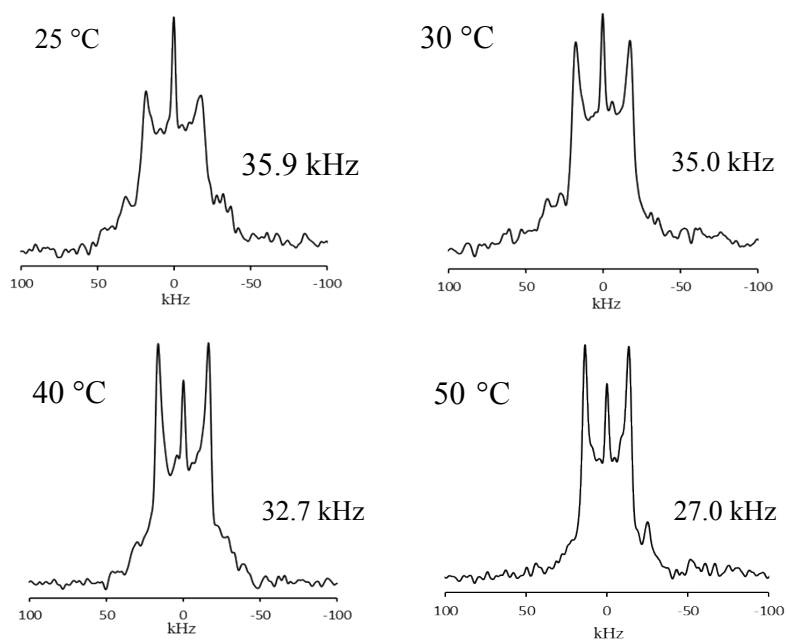


Figure S3. ^2H NMR spectra of 24dCho in PSM/24dCho 8:2 membrane.

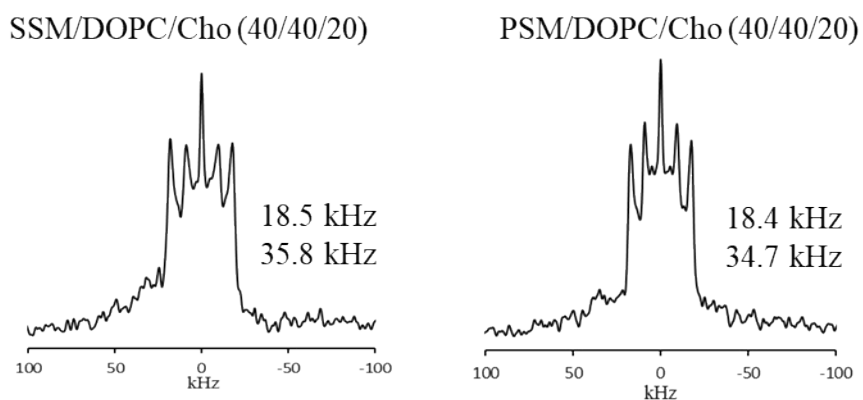


Figure S4. ^2H NMR spectra of 24dCho in SSM/DOPC/Cho 40:40:20 membrane and PSM/DOPC/Cho 40:40:20 membrane at 30 °C.

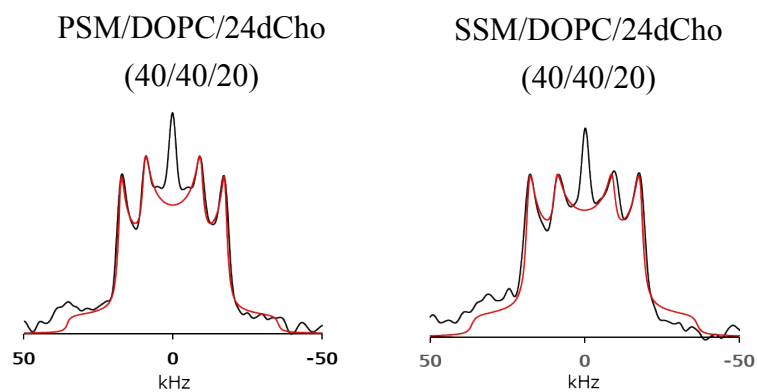


Figure S5. The ^2H NMR spectra and theoretical fitting curves of the SM/DOPC/24dCho 40:40:20 membranes at 30 °C. The experimental spectra were shown with the black traces and the fittings were shown with the red traces.

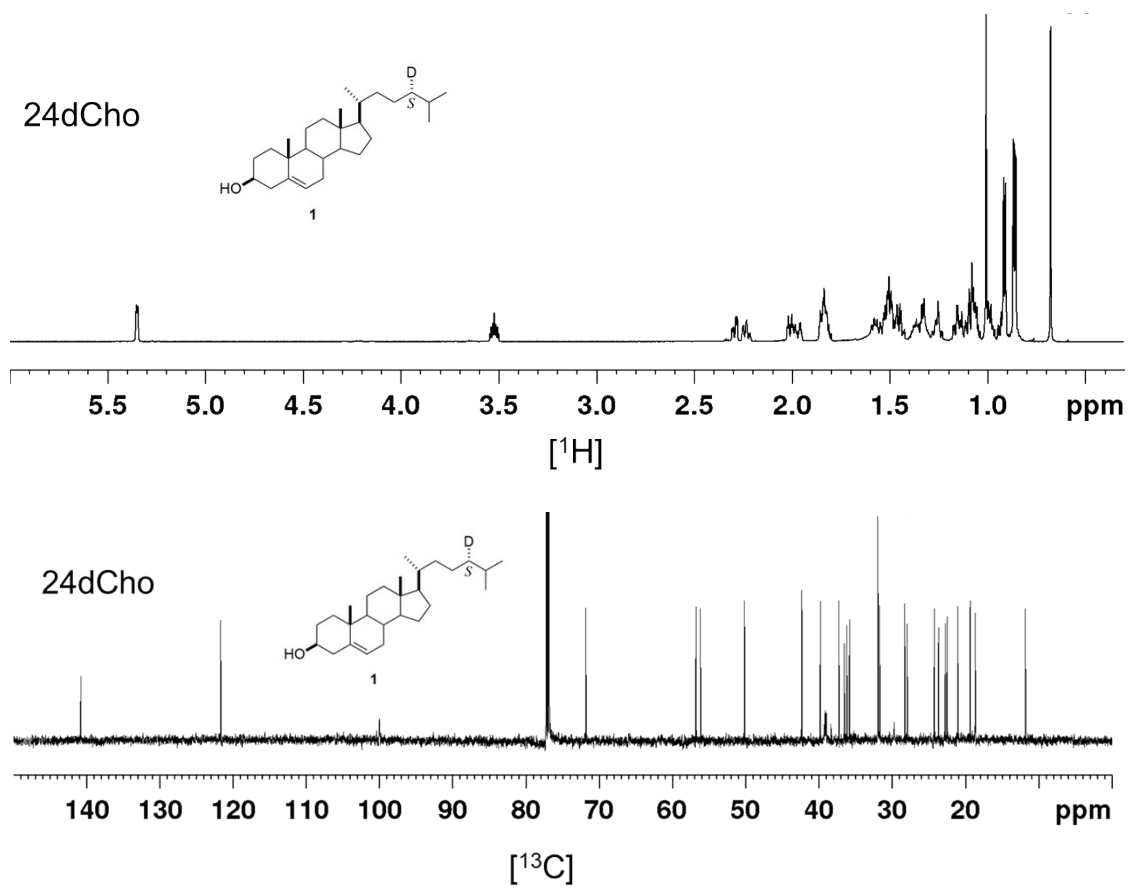


Figure S6. ¹H and ¹³C NMR spectra of 24dCho in CDCl₃. The spectra were collected using 700 MHz spectrometer.

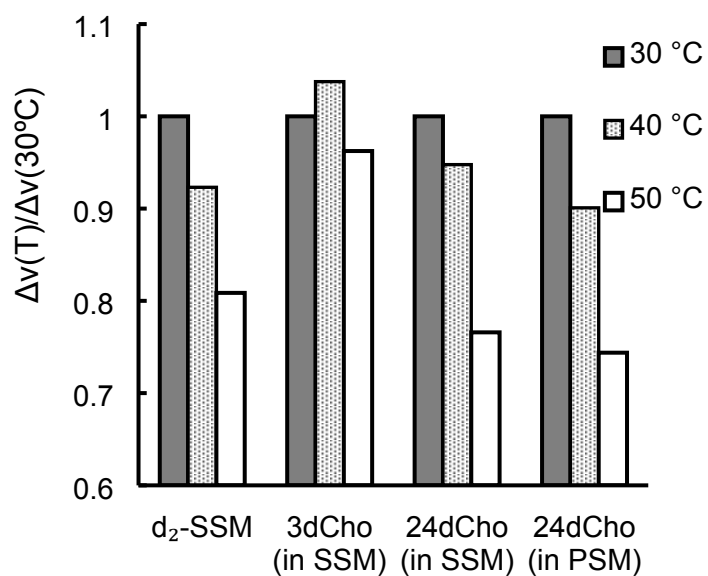


Figure S7. The relative $\Delta\nu$ values of d_2 -SSM, 3dCho, and 24dCho in SSM/Cho (8:2) or PSM/Cho (8:2) membranes by changing temperature (T) at 30, 40 and 50 °C. The y-axis values were normalized by that at 30 °C. The $\Delta\nu$ values are shown in Table 1.

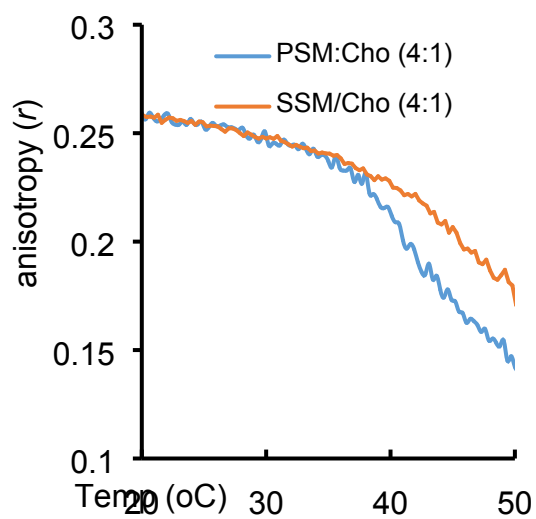
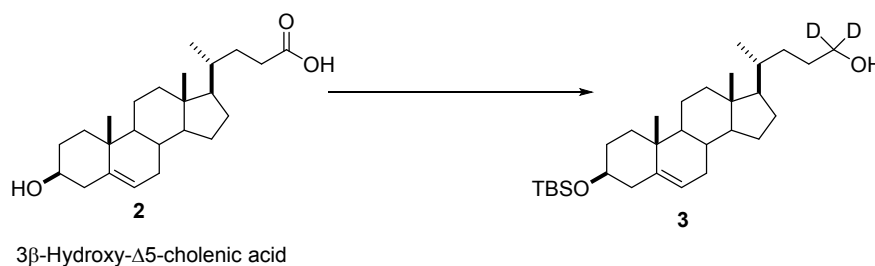


Figure S8. DPH anisotropy of the binary and ternary SM membranes whose lipid compositions were used for ^2H NMR measurements; PSM/Cho (8:2) and SSM/Cho (8:2). MLV was prepared with 200 nmol lipid plus 1 nmol DPH. The data was taken using JASCO FP-6500 spectrometer with 2 mL sample in a quartz cell. DPH; diphenylhexatriene

Synthetic Procedures

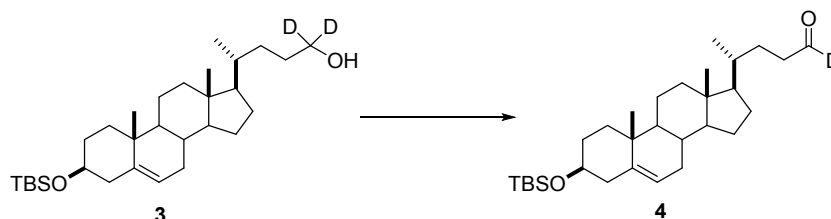
24-*d*₂-(3β)-3-*O*-*tert*-Butyldimethylsilyl-cholest-5-ene-3,24-diol (**3**)^[4]



To a solution of the commercial 3-β-hydroxy choleonic acid **2** (244 mg, 0.65 mmol) in DMF (5.5 mL) was added TBSCl (313 mg, 2.08 mmol), imidazole (359 mg, 5.27 mmol), and DMAP (171 mg, 1.40 mmol). The reaction mixture was stirred overnight at room temperature, and then quenched with water. The aqueous phase was extracted with EtOAc (×3), and the combined organic layers were washed with water (×2) and brine (×2), and dried over Na₂SO₄. After filtration, the organic layer was concentrated and dried under high vacuum to obtain the crude material (312.5 mg). A part of the crude material (150 mg) was dissolved in dry THF (4 mL) and cooled on ice-water bath for 5 min. LiAlD₄ (63 mg, 1.50 mmol) was added to the solution on ice-water bath. The reaction mixture was stirred overnight at 0 °C to room temperature, and quenched with the sodium potassium tartrate solution. After additional stirring for 5 h, the aqueous layer was extracted with EtOAc (×3). The combined organic phase was washed with water (×2) and brine (×2), and dried over Na₂SO₄. After filtration, the organic phase was evaporated and purified with SiO₂ chromatography (Hex/EtOAc = 5:1) to give **3**^[S4] (96.1 mg, 0.197 mmol, 66%, 2 steps). ¹H NMR (500 MHz, CDCl₃) δ 5.31 (1H, m, H6), 3.47 (1H, m, H3), 2.25 (2H, m, H4), 0.99 (3H, s, H19), 0.93 (3H, d, *J* = 6.5 Hz, H21), 0.88 (9H, s, TBS-*t*Bu),

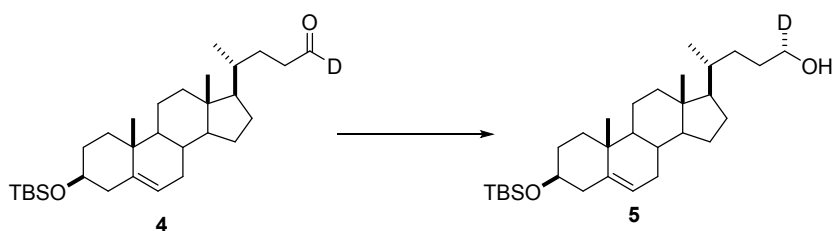
0.67 (3H, s, H18), 0.06 (6H, s, TBS-Me).

24-*d*₁-(3 β)-3-O-*tert*-Butyldimethylsilyl-cholest-5-ene-3-oxy-24-al (4)



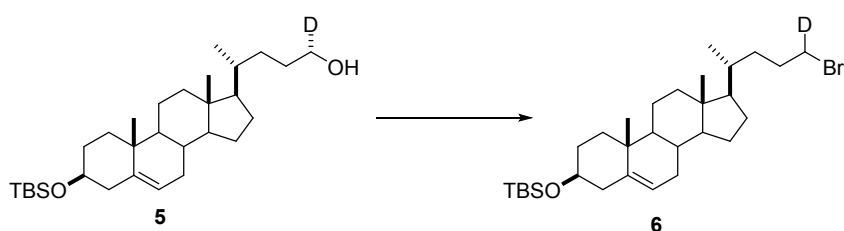
To a solution of the compound **4** (405 mg, 0.85 mmol) in CH₂Cl₂ (10 mL) was added NaOAc (25 mg, 0.31 mmol) and PCC (239 mg, 1.11 mmol) and stirred overnight at room temperature. Then, the solution was evaporated, and the residue was purified with SiO₂ chromatography (Hex/EtOAc = 2:1) to give **4** (215.6 mg, 0.455 mmol, 54%). The spectral data matches those reported in the literature for the non-deuterated compound.^[S5] ¹H NMR (500 MHz, CDCl₃) δ 5.31 (1H, m, H6), 3.47 (1H, m, H3), 2.39 (2H, m, H23), 2.21 (2H, m, H4), 0.99 (3H, s, H19), 0.92 (3H, d, J = 6.5 Hz, H21), 0.88 (9H, s, TBS-*t*Bu), 0.67 (3H, s, H18), 0.05 (6H, s, TBS-Me). ¹³C NMR (125 MHz, CDCl₃) δ 202.9 (C24), 141.5 (C5), 121.1 (C6), 72.6 (C3), 56.8 (C14), 55.8 (C17), 50.2 (C9), 42.8 (C4), 42.3 (C13), 40.8 (C23), 39.8 (C12), 37.3 (C1), 36.6 (C10), 35.3 (C20), 32.1 (C22), 31.9 (C7,C8), 28.1, 27.9 (C16), 25.9 (3C, TBS-*t*Bu), 24.2 (C15), 21.0 (C11), 19.4 (C19), 18.4 (C21), 18.3 (TBS), 11.9 (C18), -4.6 (2C, TBS-Me); HR-ESI-MS C₃₀H₅₂D₂O₂Si [M+H]⁺ calcd. 474.3878, found 474.3967.

24-(*S*)-*d*₁-(3 β)-3-O-*tert*-Butyldimethylsilyl-cholest-5-ene-3,24-diol (5)



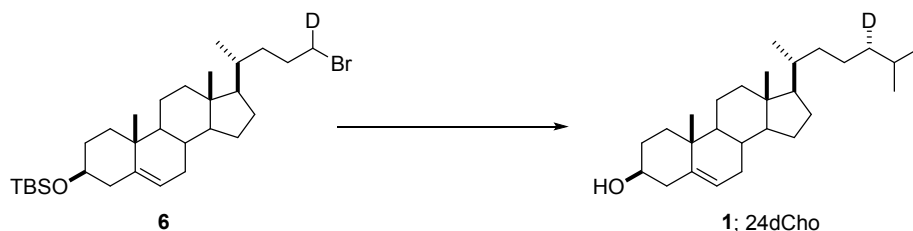
(*R*)-Alpine borane (0.5 M in THF, 1.82 mL, 0.91 mmol) was added to a solution of **4** (216 mg, 0.455 mmol) in dry THF (4 mL) and stirred 1 day at room temperature. Then, the mixture was quenched with acetaldehyde and the solvent was evaporated. The residue was dissolved in Et₂O (5 mL), cooled on ice-water bath, and then ethanolamine (0.1 mL) was added. After stirring for 1 h at room temperature, the mixture was diluted with water, and extracted with Et₂O (×3). The combined organic phase was washed with water (×2) and brine (×2), and dried over Na₂SO₄. After filtration, the residue was purified with SiO₂ chromatography (Hex/EtOAc = 10:1) to give **5** (207.9 mg, 0.437 mmol, 96%). The spectral data matches those reported for the di-deuterated **3**. ¹H NMR (500 MHz, CDCl₃) δ 5.31 (1H, m, H6), 3.65 (1H, m, H24), 3.47 (1H, m, H3), 2.21 (2H, m, H4), 0.99 (3H, s, H19), 0.93 (3H, d, *J* = 6.5 Hz, H21), 0.88 (9H, s, TBS-*t*Bu), 0.67 (3H, s, H18), 0.05 (6H, s, TBS-Me); ¹³C-NMR (125 MHz, CDCl₃) δ 141.6 (C5), 121.2 (C6), 72.6 (C3), 70.8 (C24), 56.8 (C14), 56.0 (C17), 50.2 (C9), 42.8 (C4), 42.3 (C13), 39.8 (C12), 37.4 (C1), 36.6 (C10), 35.6 (C20), 32.1 (C22), 31.9 (C2), 31.84 (C7), 31.81 (C8), 29.2 (C23), 28.2 (C16), 25.9 (3C, TBS-*t*Bu), 24.3 (C15), 21.1 (C11), 19.4 (C19), 18.7 (C21), 18.2 (TBS), 11.8 (C18), -4.6 (2C, TBS-Me).; HR-ESI-MS C₃₀H₅₃DO₂SiNa [M+Na]⁺ calcd. 498.3854, found 498.3882.

24-(*S*)-*d*₁-(3β)-3-O-*tert*-Butyldimethylsilyl-cholest-5-ene-3-oxy-24-bromide (6**)**



To a solution of **5** (214 mg, 0.45 mmol) in CH_2Cl_2 (4.9 mL) was added Ph_3P (205 mg, 0.78 mmol), imidazole (53 mg, 0.78 mmol), and CBr_4 (226 mg, 0.68 mmol), and stirred for 3 h at room temperature. Upon addition of MeOH for quenching, the mixture was evaporated and purified with SiO_2 chromatography (Hex/EtOAc = 10:1) to give **6** (170 mg, 0.316 mmol, 70%). The spectral data matches those reported for non-deuterated compound.^[S6] ^1H NMR (500 MHz, CDCl_3) δ 5.31 (1H, m, H6), 3.45 (1H, m, H3), 3.38 (1H, t, H24), 2.21 (2H, m, H4), 0.99 (3H, s, H19), 0.93 (3H, d, H21), 0.89 (9H, s, TBS-tBu), 0.68 (3H, s, H18), 0.05 (6H, s, TBS-Me); ^{13}C -NMR (125 MHz, CDCl_3) δ 141.6 (C5), 121.1 (C6), 72.6 (C3), 56.8 (C14), 55.8 (C17), 50.2 (C9), 42.8 (C4), 42.3 (C13), 39.8 (C12), 37.4 (C1), 36.6 (C10), 35.2 (C20), 34.5 (C24), 32.1 (C22), 31.90 (C7), 31.88 (C8), 29.5 (C23), 28.2 (C16), 25.9 (3C, TBS-tBu), 24.2 (C15), 21.0 (C11), 19.4 (C19), 18.7 (C21), 18.2 (TBS), 11.8 (C18), -4.6 (2C, TBS-Me); HR-ESI-MS $\text{C}_{30}\text{H}_{53}\text{DOBrSi}$ $[\text{M}+\text{H}]^+$ calcd.538.3190, found 538.3333.

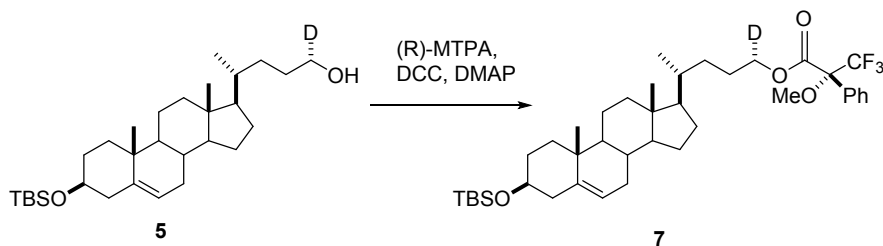
24-(*S*)- d_1 -cholesterol (**1**)



The bromide **6** (170 mg, 0.32 mmol) was dissolved in dry THF (3 mL) and cooled on ice-water bath. Then, the mixed solution of Li_2CuCl_4 (0.1 M in THF, 0.47 mL, 0.047 mmol)

and *i*-PrMgBr (1 M in THF, 1.9 mL, 1.9 mmol) was dropwise to the reaction mixture on ice-water bath. After completion of the addition, the mixture was stirred for 3 h at room temperature. Then, the reaction mixture was quenched with NH₄Cl/Et₂O=1/1 solution, and the aqueous phase was extracted with Et₂O (×2). The combined organic phase was washed with brine (×2) and dried over Na₂SO₄. After filtration, the solution was evaporated and purified with short SiO₂ column (Hex/EtOAc = 100:1) to give a crude material. The crude was then dissolved in THF (4.5 mL) and the TBAF solution (1 M in THF, 2.87 mL, 2.87 mmol) was added. The mixture was stirred overnight at room temperature and then diluted with EtOAc. The organic phase was washed with 1 M HCl (×2) and brine (×2), and dried over Na₂SO₄. After filtration, the mixture was purified with SiO₂ chromatography (Hex:EtOAc = 6:1) to give 24dCho **1** (102 mg, 0.26 mmol, 82%). The spectral data matches those commercial Cho. ¹H NMR (400 MHz, CDCl₃) δ 5.35 (1H, m, H6), 3.52 (1H, m, H3), 2.25 (2H, m, H4), 1.99 (2H, m, H7, H12), 1.83 (3H, m, H1, H2, H16), 1.61-1.41 (6H), 1.40-1.03 (10H), 1.01 (3H, s, H19), 0.99 (2H), 0.95 (1H), 0.91 (3H, d, H21), 0.86 (6H, dd, H26, H27), 0.68 (3H, s, H18). ¹³C NMR (175 MHz, CDCl₃) δ 140.7, 121.7, 71.8, 56.8, 56.1, 50.1, 42.3, 39.8, 39.1 (dd, ¹J_{C-D} = 18.9 Hz), 37.2, 36.5, 36.2, 35.8, 31.9, 31.7, 28.2, 27.9, 24.3, 23.7, 22.8, 22.5, 21.1, 19.4, 18.7, 11.9. HR-ESI-MS C₂₇H₄₇DO₂ [M+H₂O]⁺ calcd. 405.3712, found 405.3782.

24-(S)-d₁-(3β)-3-O-tert-Butyldimethylsilyl-cholest-5-ene-3,24-diol (R)-MTPA ester
(7)



To a solution of **5** (5.5 mg, 0.012 mmol) in CH₂Cl₂ (0.50 mL) was added DCC (26.4 mg, 0.13 mmol), DMAP (15.6 mg, 0.13 mmol), and (*R*)-MTPA (24.0 mg, 0.10 mmol), and stirred overnight at room temperature. Then, the mixture was diluted with EtOAc, and the organic phase was washed with 1 M HCl and brine, and dried over Na₂SO₄. After filtration, the solvent was evaporated and the residue was purified with SiO₂ chromatography (Hex:EtOAc = 20:1) to give ester **7** (6.4 mg, 0.094 mmol, 78%). ¹H NMR (500 MHz, CDCl₃) δ 7.67-6.94 (5H, m, Ph), 5.36 (1H, m, H₆), 3.99 (1H, t, H₂₄), 3.60 (1H, m, H₃), 0.99-0.92 (12H, m, H₁₉, TBS-tBu), 0.77 (3H, d, H₂₁), 0.54 (3H, s, H₁₈), 0.07 (6H, d TBS-Me).

References

- [S1] D. C. Kapeller, L. Brecker F. Hammerschmidt, *Chem. E. J.* **2007**, *13*, 9582-9588.
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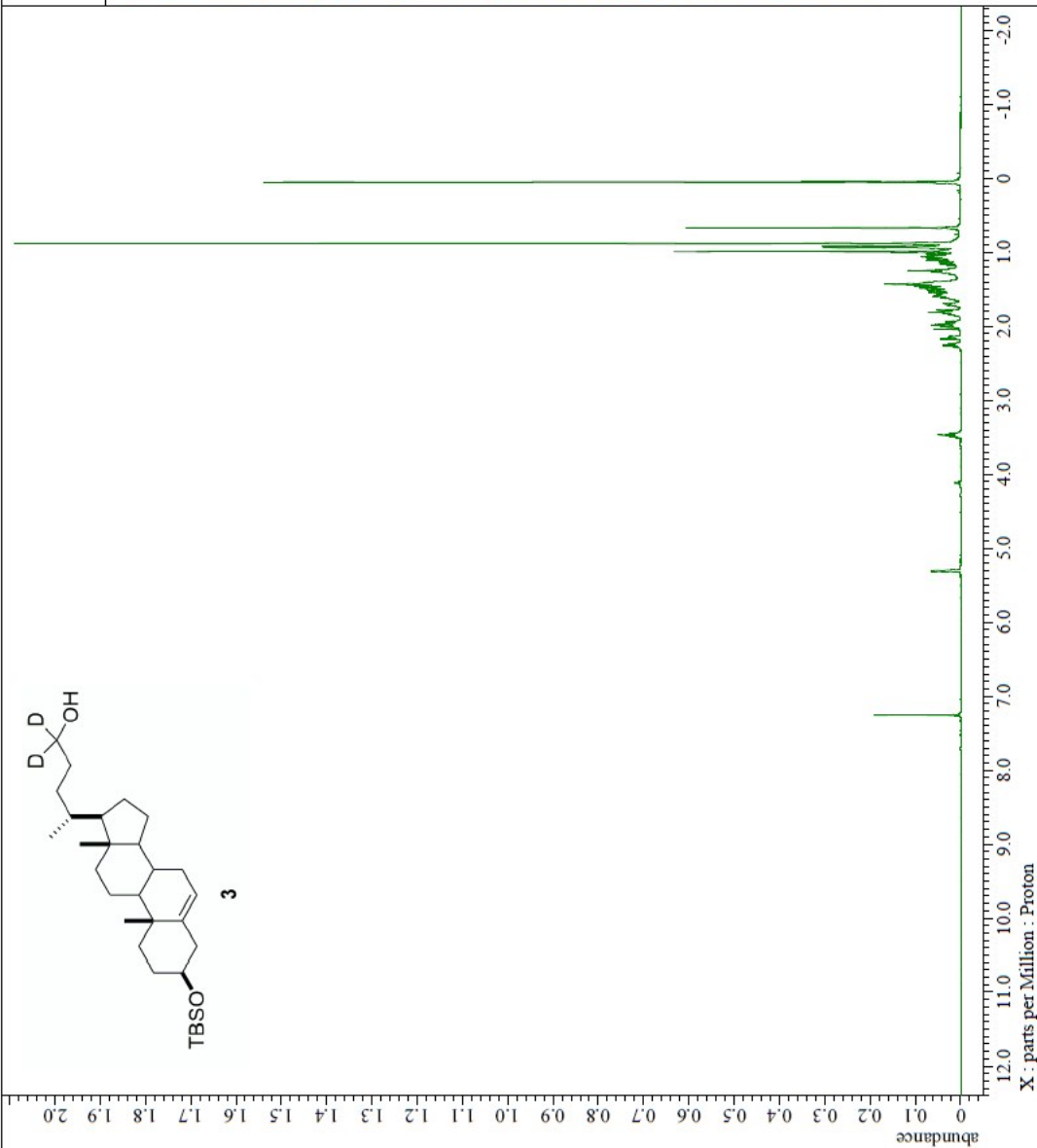
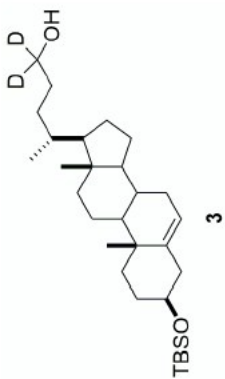
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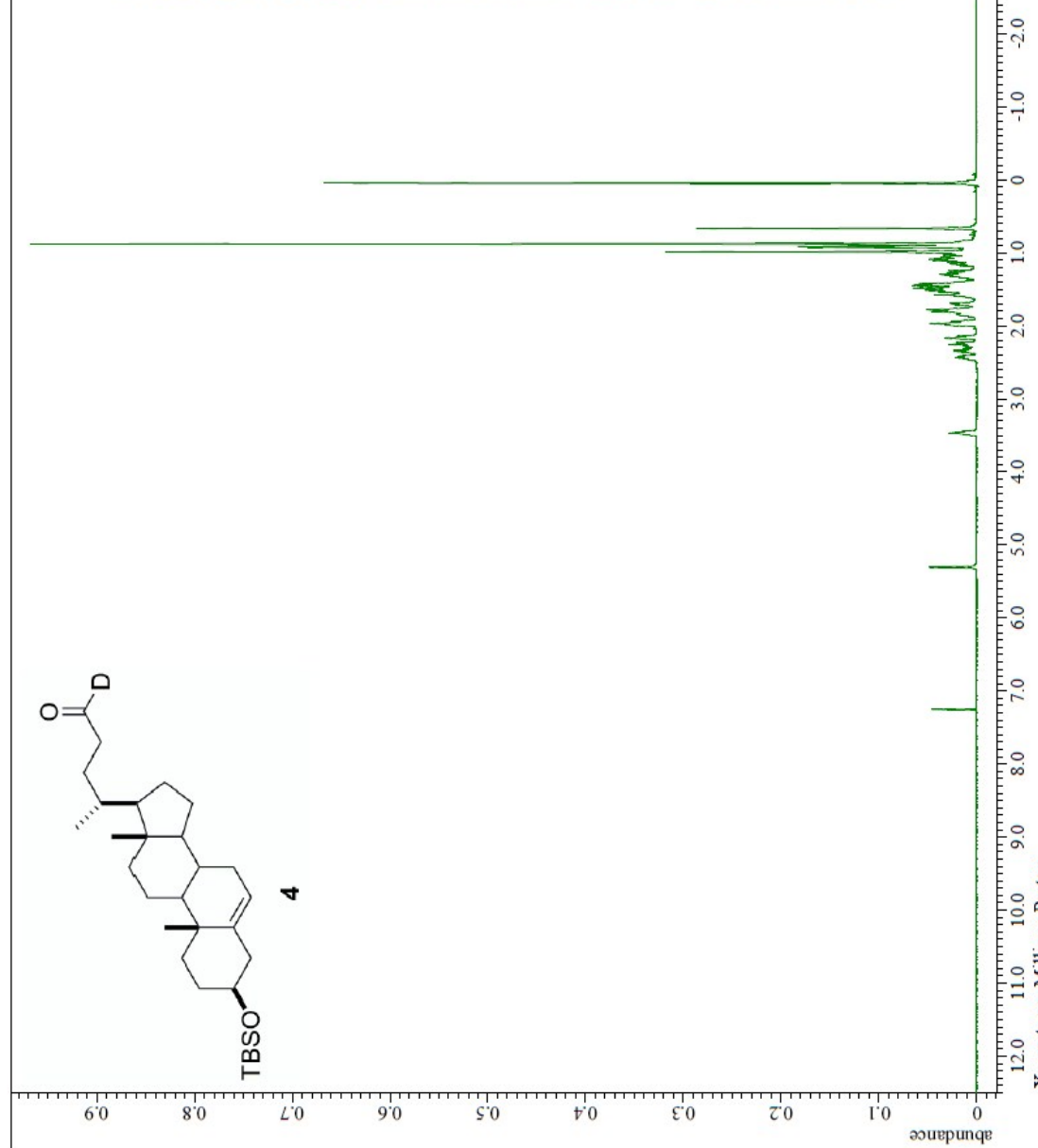
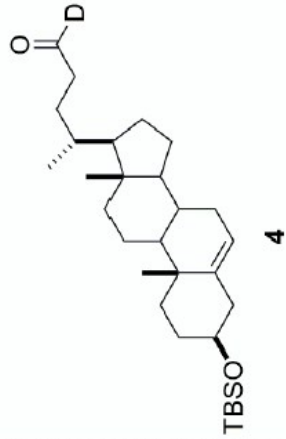


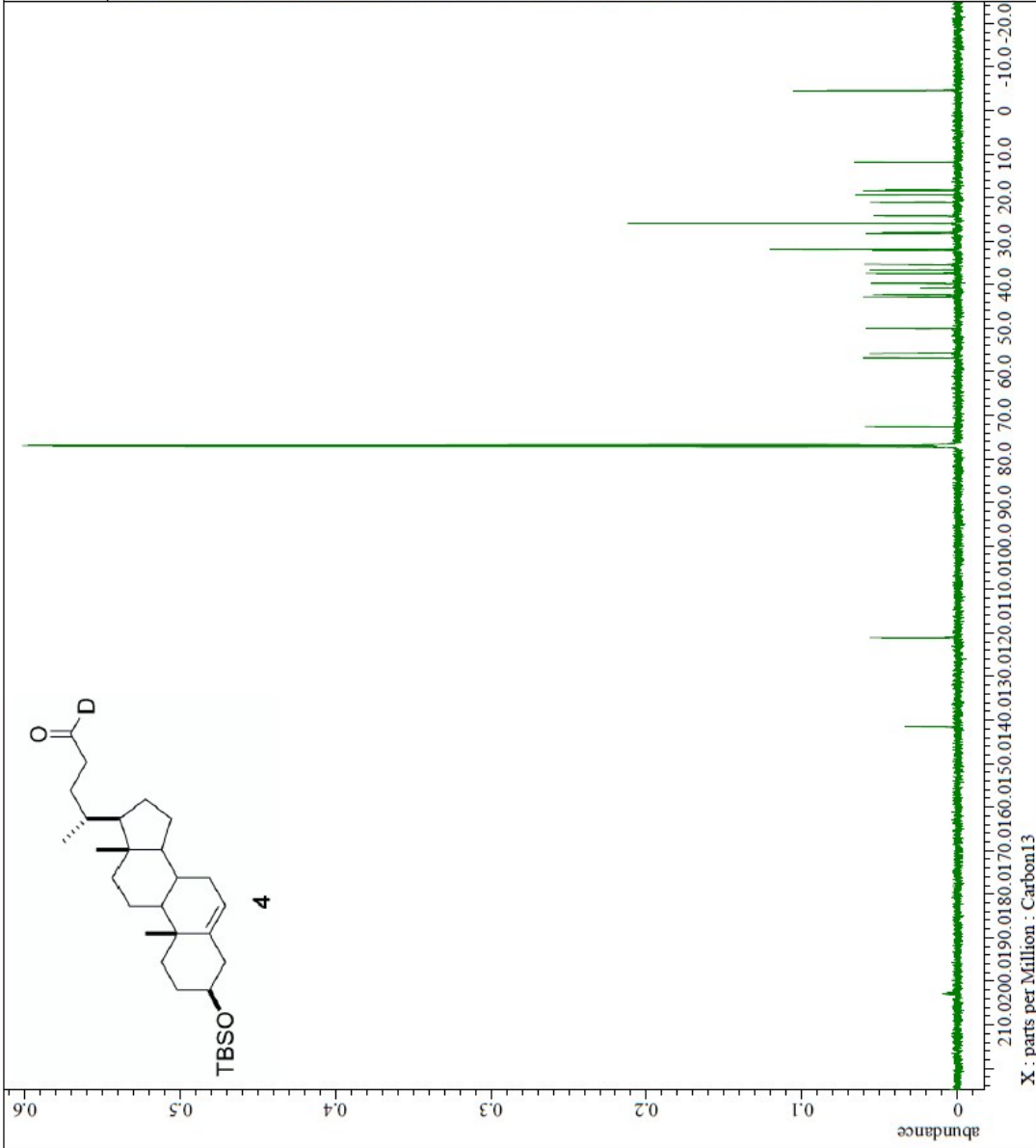
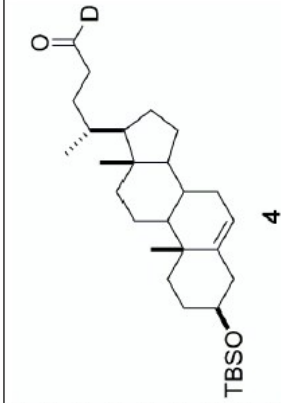
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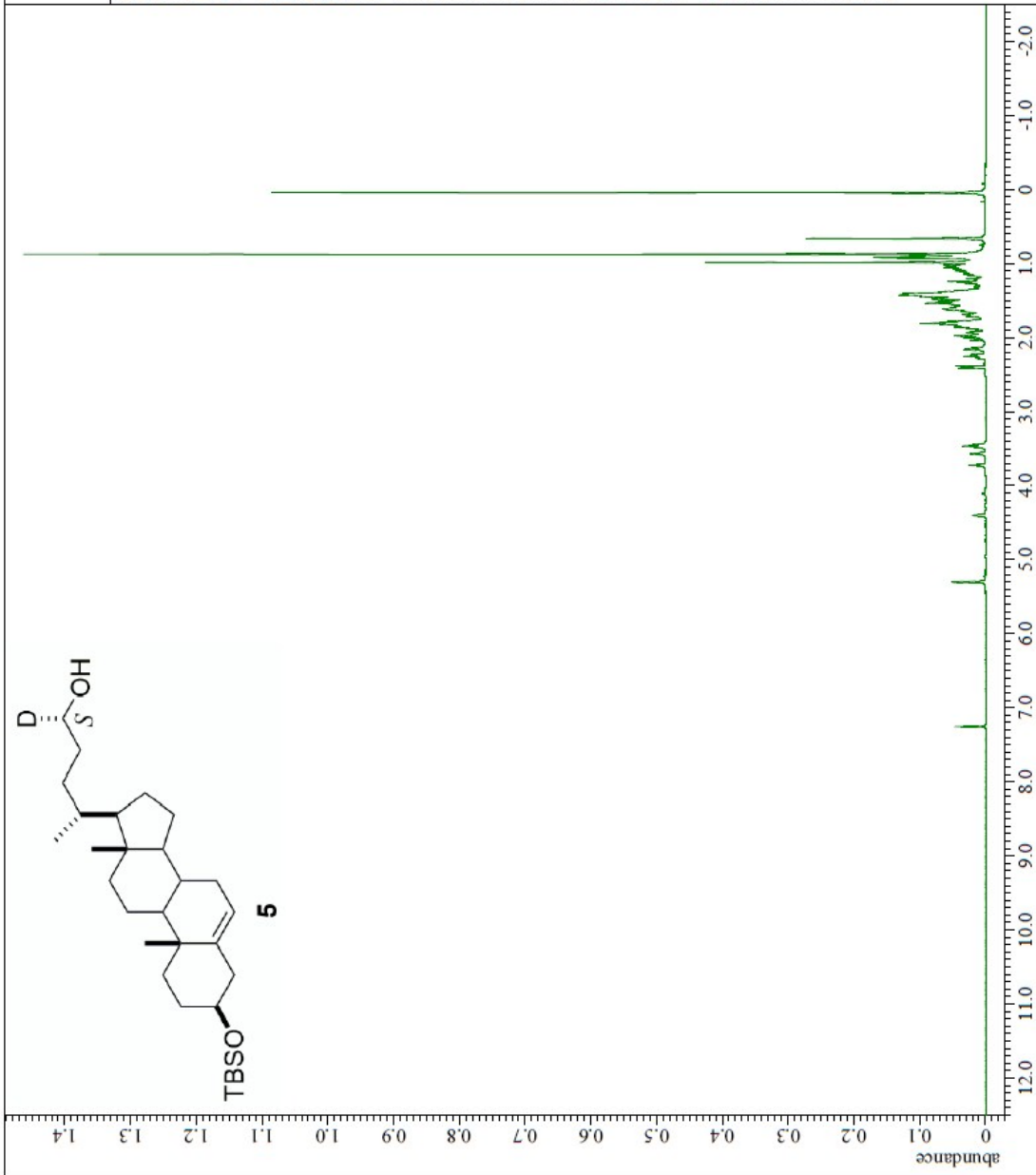
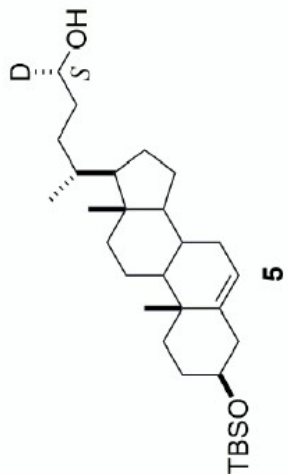


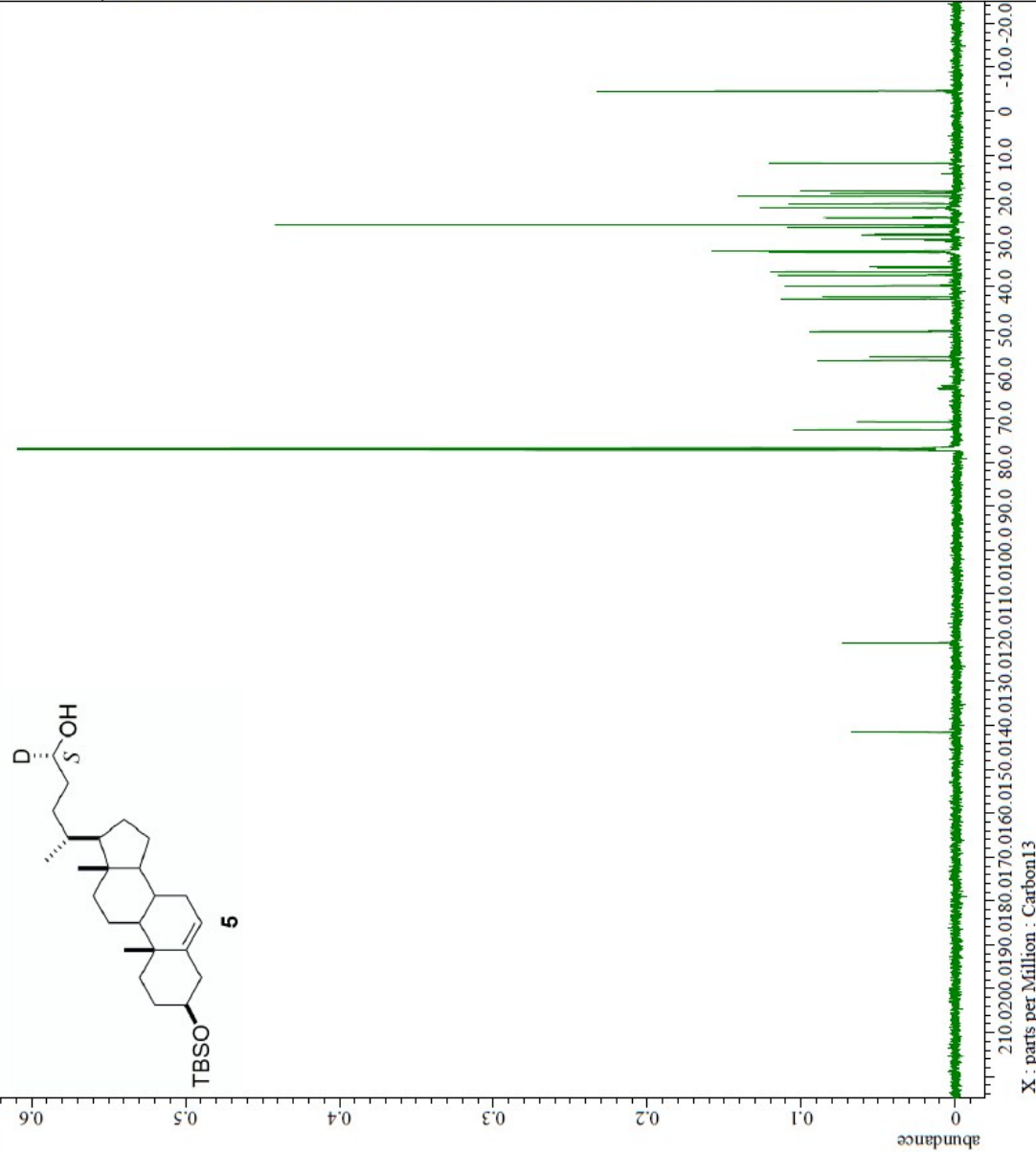
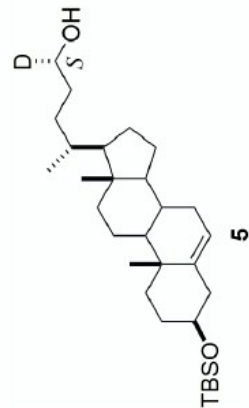
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 Current_Time = 12-MAR-2019 13:26:22
 Comment = single pulse decoupled gat
 Data_Format = 1D COMPMX
 Dia_Size = 52429
 Dia_Title = Carbon13
 Dia_Units = [ppm]
 Dimensions = X
 Site = MCA 500
 Spectrometer = DELTA2_NMR
 Field Strength = 11.7473579[T] (500 [MHz])
 X_Acq Duration = 1.66723584[s]
 X_Domain = 13C
 X_Freq = 125.76529768 [MHz]
 X_Offset = 100 [ppm]
 X_Points = 65536
 X_Prescans = 4
 X_Resolution = 0.59979517 [Hz]
 X_Sweep = 39.3081761 [kHz]
 X_Sweep_Clippped = 31.44654088 [kHz]
 Irr_Domain = Proton
 Irr_Freq = 500.15991521 [MHz]
 Irr_Offset = 5.0 [ppm]
 Clipped = FALSE
 Scans = 921
 Total_Scans = 921
 Relaxation_Delay = 2 [s]
 Recvr_Gain = 58
 Temp_Get = 25 [dC]
 X_90_Width = 11.6 [us]
 X_Acq_Time = 1.66723584 [s]
 X_Angle = 30 [deg]
 X_Atn = 10 [db]
 X_Pulse = 3.86666667 [us]
 Irr_Atn_Dec = 20.5 [db]
 Irr_Atn_Noise = 20.5 [db]
 Irr_Noise = WALTZ
 Irr_Width = 92 [us]
 Decoupling = PRNP
 Initiat_Wait = 1 [s]
 Noe_Time = 2 [s]
 Repetition_Time = 3.66723584 [s]

Y12-119-F28-62_proton-1-2.

Filename = Y12-119-F28-62_proton-1-2.
 Author = Murata
 Experiment = Proton_1xp
 Sample_Id = Y12-119-F28-62
 Solvent = CHLOROFORM-D
 Creation_Time = 14-FEB-2018 11:31:59
 Revision_Time = 30-JAN-2019 15:29:12
 Current_Time = 30-JAN-2019 15:29:30
 Comment = single_pulse
 Data_Format = 1D_COMPLEX
 Dim_Size = 52429
 Dim_Title = Proton
 Dim_Units = [ppm]
 Dimensions = X
 Site = BCA 500
 Spectrometer = JNM-ECX500
 Field_Strength = 11.747379[T] (500[MHz])
 X_Acq_Duration = 6.98351616[s]
 X_Domain = IR
 X_Freq = 500.15991621[MHz]
 X_Offset = 5.0[ppm]
 X_Points = 65536
 X_Prescans = 1
 X_Resolution = 0.14319434[Hz]
 X_Sweep = 9.38438438[Hz]
 X_Sweep_Clipped = 7.50750751[Hz]
 Ix1_Domain = Proton
 Ix1_Freq = 500.15991621[MHz]
 Ix1_Offset = 5.0[ppm]
 Tx1_Domain = Proton
 Tx1_Freq = 500.15991621[MHz]
 Tx1_Offset = 5.0[ppm]
 Clipped = FALSE
 Scans = 8
 Total_Scans = 8
 Relaxation_Delay = 5[s]
 Recvr_Gain = 22
 Temp_Get = 21.5[degC]
 X_90_Width = 6.4[us]
 X_Acq_Time = 6.98351616[s]
 X_Angle = 45[deg]
 X_Atn = 4.5[dB]
 X_Pulse = 3.2[us]
 Ix1_Mode = Off
 Tx1_Mode = Off
 Data_Preset = FALSE
 Initial_Weit = 1[s]
 Repetition_Time = 11.98351616[s]





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File Name = Y13-05-FZ6-S1_carbon-1-2.j
 Author = Hurata
 Experiment = carbon.jcf
 Sample_Id = Y13-05-FZ6-S1
 Solvent = CHLOROFORM-D
 Creation_Time = 28-FEB-2019 14:11:34
 Revision_Time = 12-MAR-2019 13:25:20
 Current_Time = 12-MAR-2019 13:25:24

Comment = single pulse decoupled gat
 Data_Format = 1D COMPLEX
 Dim_Size = 32429
 Dim_Title = Carbon13
 Dim_Units = [ppm]
 Dimensions = 1
 Site = NCA_500
 Spectrometer = DELTA2_NMR

Field_Strength = 11.7473579 [T] (500 [MHz])
 X_Acq_Duration = 1.66723584 [s]
 X_Det_Acquisition = 13C_76529768 [MHz]
 X_Freq = 125.76529768 [MHz]
 X_Offset = 100 [ppm]
 X_Points = 65536
 X_Precans = 4
 X_Resolution = 0.59979517 [Hz]
 X_Sweep_Duration = 39.3081761 [kHz]
 X_Sweep_Clippped = 31.44654088 [kHz]
 X_Domain = Proton
 Xr_Freq = 500.15991521 [MHz]
 Xr_Offset = 5.0 [ppm]
 Clipped = FALSE
 Scans = 691
 Total_Scans = 691

Relaxation_Delay = 2 [s]
 Recv_Gain = 58
 Temp [C] = 25 [dC]
 X_90_Width = 11.6 [us]
 X_Acq_Time = 1.66723584 [s]
 X_Angle = 30 [deg]
 X_Atn = 10 [dB]
 X_Pulse = 3.86666667 [us]
 Xr_Atn_Dec = 20.5 [dB]
 Xr_Atn_Noise = 20.5 [dB]
 Xr_Noise = WALTZ
 Xr_Width = 92 [us]
 Decoupling = TRUE
 Initial_Wait = 1 [s]
 Noe_Time = TRUE
 Noe_Time = 2 [s]
 Repetition_Time = 3.66723584 [s]

Filename = Y12-163-Fl-7_Proton-1-2.jd
 Author = Murata
 Experiment = Proton.jfif
 Sample_Id = Y12-163-Fl-7
 Solvent = CHLOROFORM-D
 Creation_Time = 21-JUN-2018 17:15:39
 Revision_Time = 30-JAN-2019 15:31:04
 Current_Time = 30-JAN-2019 15:31:16

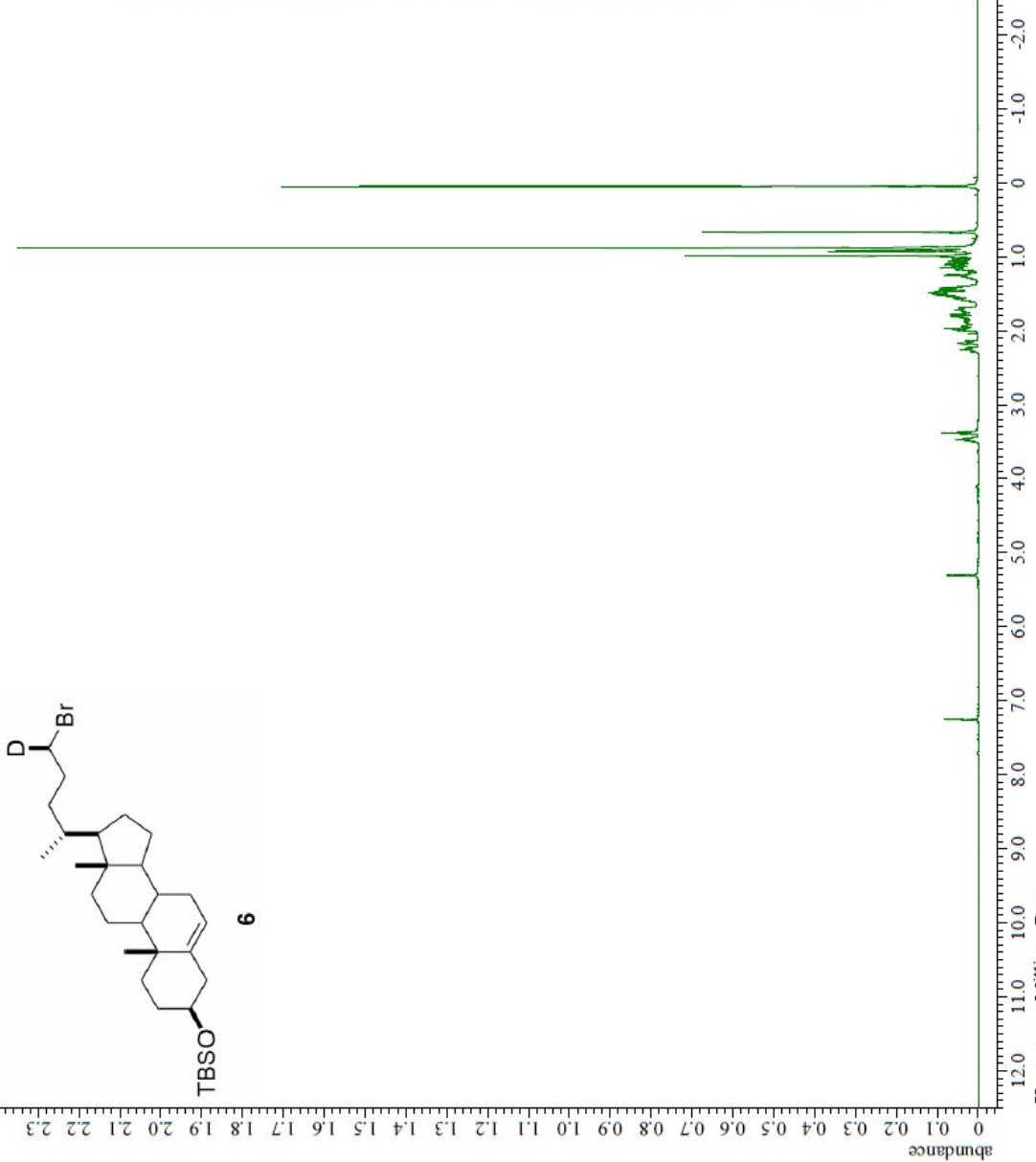
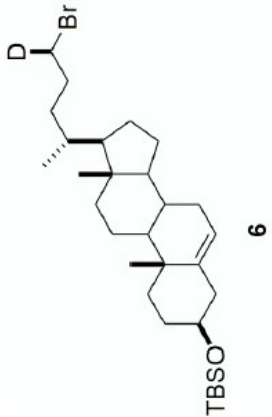
Comment = single_pulse
 Data_Format = ID COMPLEX
 Diam_Size = 52429
 Diam_Title = Proton
 Diam_Units = [ppm]
 Dimensions = X
 Site = MCA 500
 Spectrometer = DELTA2_NMR

Field_Strength = 11.7473579 [T] (500 [MHz])
 X_Acq_Duration = 6.98351616 [s]
 X_Domain = IR
 X_Freq = 500.15991521 [MHz]
 X_Offset = 5.0 [ppm]
 X_Points = 65536

X_Prescans = 1
 X_Resolution = 0.14319434 [Hz]
 X_Sweep = 9.38438438 [kHz]
 X_Sweep_Clippped = 7.50750751 [kHz]
 Irx_Domain = Proton
 Irx_Freq = 500.15991521 [MHz]
 Irx_Offset = 5.0 [ppm]

Tri_Domain = Proton
 Tri_Freq = 500.15991521 [MHz]
 Tri_Offset = 5.0 [ppm]
 Clipped = FALSE
 Scans = 8
 Total_Scans = 8

Relaxation_Delay = 5 [s]
 Recvr_Gain = 30
 Temp_Get = 20.8 [dC]
 X_90_Width = 12.7 [us]
 X_Acq_Time = 6.98351616 [s]
 X_Angle = 45 [deg]
 X_Atn = 3.3 [dB]
 X_Pulse = 6.35 [us]
 Irx_Mode = Off
 Tri_Mode = Off
 Dante_Preset = FALSE
 Initial_Wait = 1 [s]
 Repetition_Time = 11.98351616 [s]

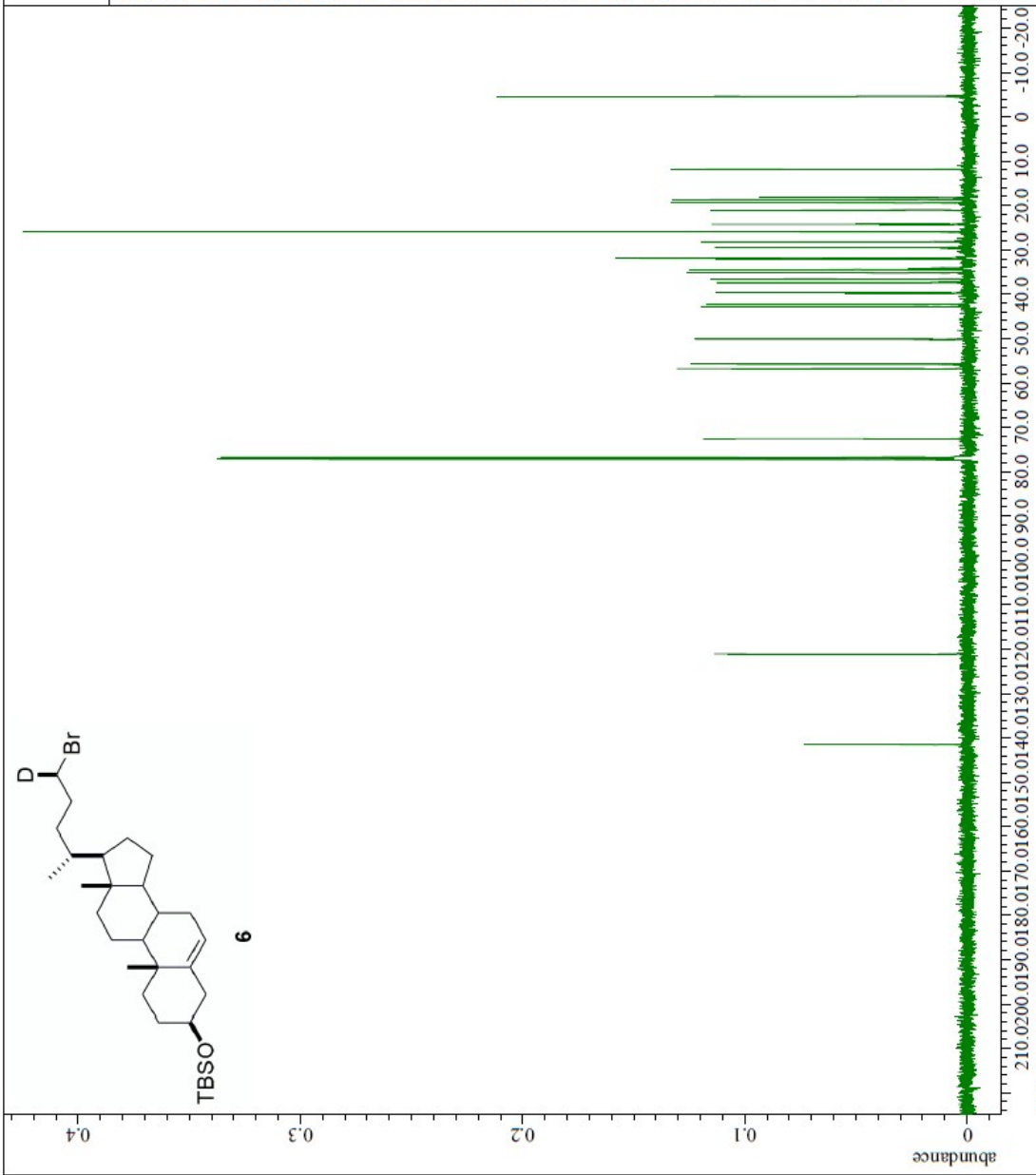
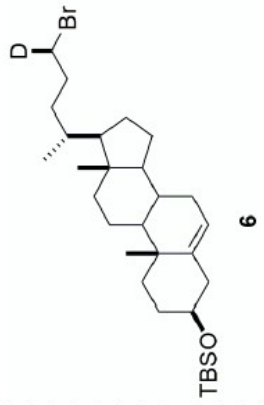


Filename = Y13-87-F3-6_carbon-1-2.jdf
 Author = Murata
 Experiment = carbon.jxp
 Sample_Id = Y13-87-F3-6
 Solvent = CHLOROFORM-D
 Creation_Time = 27-FEB-2019 09:55:39
 Revision_Time = 12-MAR-2019 13:23:44
 Current_Time = 12-MAR-2019 13:23:50

Comment = single pulse decoupled gat
 Data_Format = ID COMPLEX
 Data_Size = 52429
 Data_Title = Carbon13
 Data_Units = [ppm]
 Dimensions = X
 Site = BCCA 500
 Spectrometer = JNM-BCA500

Field_Strength = 11.7473579 [T] (500 [MHz])
 X_Acq_Duration = 1.66723884 [s]
 X_Domain = 13C
 X_Freq = 125.76529768 [MHz]
 X_Offset = 100 [ppm]
 X_Points = 65536
 X_Prescans = 4
 X_Resolution = 0.59979517 [Hz]
 X_Sweep = 39.3061761 [kHz]
 X_Sweep_Clippped = 31.44654088 [kHz]
 Irr_Domain = Proton
 Irr_Freq = 500.15991521 [MHz]
 Irr_Offset = 5.0 [ppm]
 Clipped = TRUE
 Scans = 551
 Total_Scans = 551

Relaxation_Delay = 2 [s]
 Recvrz_Gain = 54
 Temp_Get = 25 [dC]
 X_90_Width = 22 [us]
 X_Acq_Time = 1.66723884 [s]
 X_Angle = 30 [deg]
 X_Atn = 7 [dB]
 X_Pulse = 7.33333333 [us]
 Irr_Atn_Dec = 27.65 [dB]
 Irr_Atn_Noise = 27.65 [dB]
 Irr_Noise = WALTZ
 Irr_Width = 92 [us]
 Decoupling = TRUP
 Initial_Wait = 1 [s]
 Noe_Time = TRUP
 Repetition_Time = 3.66723884 [s]



```

Filename = YI-137-F7-13-benzen_proton
Author = Murata
Experiment = proton_jkp
Sample Id = YI-137-F7-13-benzen
Solvent = BENZENE-D6
Creation Time = 18-JAN-2017 11:03:39
Revision Time = 26-FEB-2019 15:16:01
Current Time = 26-FEB-2019 15:16:57

Comment = single pulse
Data Format = 1D COMPLEX
Dim Size = 52429
Dim Title = Proton
Dim Units = [ppm]
Dimensions = X
Site = ECA 500
Spectrometer = DELTA2_NMR

Field Strength = 11.7473579[T] (500 [MHz])
X_Acq_Duration = 6.98351616[s]
X_Domain = 1H
X_Freq = 500.15991521 [MHz]
X_Offset = 5.0 [ppm]
X_Points = 65536
X_Prescans = 1
X_Resolution = 0.14319434 [Hz]
X_Sweep = 9.38438438 [kHz]
X_Sweep_Clippped = 7.50750751 [kHz]
Irr_Domain = Proton
Irr_Freq = 500.15991521 [MHz]
Irr_Offset = 5.0 [ppm]
Tri_Domain = Proton
Tri_Freq = 500.15991521 [MHz]
Tri_Offset = 5.0 [ppm]
Clipped = FALSE
Scans = 8
Total_Scans = 8

Relaxation_Delay = 5 [s]
Recvr_Gain = 44
Temp_Get = 18.2 [dc]
X_90_Width = 11.4 [us]
X_Acq_Time = 6.98351616 [s]
X_Angle = 45 [deg]
X_Atn = 2.7 [dB]
X_Pulse = 5.7 [us]
Irr_Mode = Off
Tri_Mode = Off
Dante_Presat = FALSE
Initial_Wait = 1 [s]
Repetition_Time = 11.98351616 [s]
  
```

