

SUPPORTING INFORMATION

***P*-Chiral, *N*-Phosphoryl Sulfonamide Brønsted Acids with an Intramolecular Hydrogen Bond Interaction that Modulates Organocatalysis**

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General Experimental Methods

All reactions were carried out under anhydrous conditions and under an atmosphere of dry argon unless otherwise indicated. Compounds were purified by normal phase flash column chromatography on silica gel (SDS, 60 Å C. C. 40-63 mm) as the stationary phase. Thin Layer Chromatography (TLC) was performed on alumina plates pre-coated with silica gel (Merck silica gel, 60 F254), which were visualized by UV when applicable ($\lambda_{\text{max}} = 254 \text{ nm}$ and/or 366 nm) and/or by staining with vanillin or anisadehyde in acidic ethanol and/or KMnO_4 in basic water followed by heating. Key compounds were fully characterized by ^1H , $^{13}\text{C}\{^1\text{H}\}$ and $^{31}\text{P}\{^1\text{H}\}$ NMR and HRMS. Chemical shifts (δ) are reported in ppm relative to the internal deuterated solvent or external H_3PO_4 ($\delta 0.00$ ^{31}P), unless indicated otherwise. High-resolution MS spectra were recorded using electrospray ionization (ESI+/-) and Fourier transform ion cyclotron resonance mass analyzer (FTMS).

The reactions were monitored either by TLC or analytical HPLC/MS to confirm completion and homogeneity of the products. Analytical HPLC was performed using a reversed phase C18 5 μm column on a Waters Atlantis T3 instrument and the solvent system indicated below:

Solvent A: H_2O , 0.1% formic acid

Solvent B: CH_3CN , 0.1% formic acid

Mobile phase: linear gradient from 95%A and 5%B to 5%A and 95%B in 13 min, then 2 min at 100% B

Flow rate: 1 mL/min

Compounds 6-bromo-2-methylquinoline (**10a**), 2-methylquinoline (**10b**), 2-phenylquinoline (**10c**) and 4-methylquinoline (**10d**) were purchased from Sigma Aldrich. The 2-ethylquinoline (**10e**), ¹ 2-isopropylquinoline (**10f**), ² 6-nitro-2-methylquinoline (**10g**)³ and 6-methoxy-2-methylquinoline (**10h**)³ were synthesized according to the literature procedures indicated.

The enantiomeric purity of chiral compounds was determined by chiral HPLC using an Agilent 1100 or Agilent 1260 series instrument and the column and solvent system indicated for each compound. The absolute stereochemistry of all compounds was assigned based on several factors, including the single crystal X-ray of the previously reported key precursor compound **6**,⁴ the single crystal X-ray structures of intermediate phosphinic amide **8d** (refer to SI Table 3), the single crystal X-ray structures of catalysts **5a** and **5c**, the single crystal X-ray structure of compound (*S*)-2-bromo-6-methyl-3,4-dihydro-2*H*-1 λ^2 -quinoline (**11a**), and by analogy with previously reported compounds in the literature.

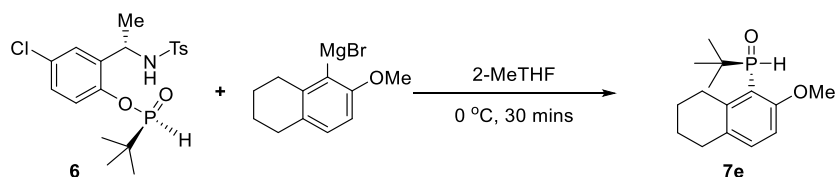
The names of all compounds were generated using ChemBioDraw Ultra 12.0.

General synthesis of secondary phosphine oxides (SPOs) **7**:

We recently reported the synthesis of SPO intermediates **6**, **7a**, **7c** and **7d**.⁴ The synthesis of analogs **7e** and **7f** was achieved using the same protocol.⁴ The synthesis of SPO analogs **7b** was achieved using the previously reported methodology.⁵

(S)-tert-Butyl(2-methoxy-5,6,7,8-tetrahydronaphthalen-1-yl)phosphine oxide (**7e**):

Precursor compound 5-bromo-6-methoxy-1,2,3,4-tetrahydronaphthalene (used to prepare the Grignard reagent) was synthesized according to the method reported by Smith and co-workers.⁶



A three neck flask under argon was charged with SPO **6** (1 mmol) in 2-MeTHF (3 mL) and cooled to 0 °C. 2-Methoxy-5,6,7,8-tetrahydronaphthalen-1-yl magnesium bromide (1 M in 2-MeTHF, 4 mmol, 4 mL) was added slowly while keeping the internal temperature <5 °C. The reaction mixture was stirred for 40 min to completion. Saturated and degassed aqueous NH₄Cl solution (5 mL) was added slowly to quench the reaction. The organic layer was collected and the aqueous residue was extracted with DCM (25 mL x3). The combined organic extracts were dried over anhydrous Na₂SO₄ and concentrated. The residue was purified by flash column chromatography on silica gel (deactivated with 10% water) using a solvent gradient of hexane/EtOAc (from 50:50 to 0:100, v/v) to obtain the desired product (141 mg) in 53% yield.

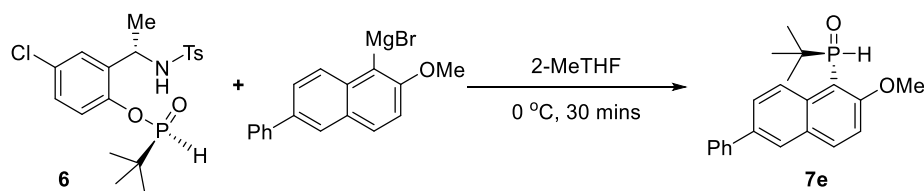
¹H NMR (400 MHz, CDCl₃): δ 8.28 (s, 0.5 H), 7.17 (d, *J* = 8.5 Hz, 1H), 7.06 (s, 0.5 H), 6.71 (dd, *J* = 8.5, 5.1 Hz, 1H), 3.77 (s, 3H), 3.54 – 3.39 (m, 1H), 2.93 (dt, *J* = 17.0, 5.6 Hz, 1H), 2.72 (t, *J* = 6.2 Hz, 2H), 1.83 – 1.66 (m, 4H), 1.20 (d, *J* = 16.6 Hz, 9H).

³¹P NMR (162 MHz, CDCl₃): δ 36.57.

More detailed characterization and estimation of the enantiomeric purity was performed at the subsequent step, when **7e** was converted to the corresponding *P*-chiral (*tert*-butyl)-*P*-arylphosphinic amide **8e**.

(S)-tert-Butyl(2-methoxy-6-phenylnaphthalen-1-yl)phosphine oxide (**7f**):

The precursor 1-bromo-2-methoxy-6-phenylnaphthalene (used to prepare the Grignard reagent) was synthesized according to the method reported by Smith and co-workers.⁶



A three-neck flask under argon was charged with SPO **6** (1 mmol) dissolved in 2-MeTHF (3 mL) and cooled to 0 °C. A solution of 2-methoxy-6-phenylnaphthalen-1-yl

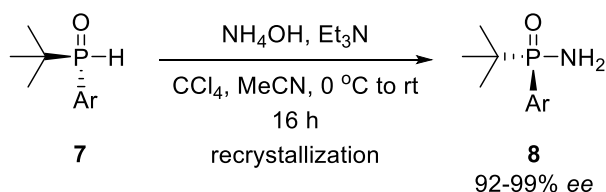
magnesium bromide (1 M in 2-MeTHF, 4 mmol, 4 ml; prepared as previously reported⁶) was added slowly, while keeping the internal temperature <5 °C. The reaction mixture was stirred for 40 min to complete the reaction. Saturated and degassed aqueous NH₄Cl solution (5 mL) was added slowly to quench the reaction. The organic layer was collected and the aqueous residue was extracted with DCM (25 mL x3). The combined organic extracts were dried over anhydrous Na₂SO₄ and concentrated. The residue was purified by silica gel chromatography (on deactivated silica with 10% water) eluted with a solvent gradient of hexane/EtOAc gradient (50:50 to 0:100, v/v) to obtain the desired product (189 mg) in 56% yield.

¹H NMR (400 MHz, CDCl₃): δ 9.04 (d, *J* = 9.0 Hz, 1H), 8.54 (s, 0.5 H), 8.07 (d, *J* = 9.1 Hz, 1H), 7.98 (s, 1H), 7.81 (dd, *J* = 9.0, 2.1 Hz, 1H), 7.70 (d, *J* = 7.1 Hz, 2H), 7.48 (t, *J* = 7.7 Hz, 2H), 7.37 (t, *J* = 7.4 Hz, 1H), 7.31 (s, 0.5 H), 7.30 – 7.26 (m, 1H), 3.98 (s, 3H), 1.26 (d, *J* = 16.8 Hz, 9H).

³¹P NMR (162 MHz, CDCl₃): δ 36.19.

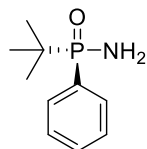
More detailed characterization and estimation of the enantiomeric purity was performed at the subsequent step, when **7f** was converted to the corresponding *P*-chiral (*tert*-butyl)-*P*-arylphosphinic amide **8f**.

General procedure for the conversion of SPOs **7** to the *P*-Chiral (*t*-butyl)-*P*-arylphosphinic amides **8**:



Chiral SPO **7** (1.0 mmol) was dissolved in 6 mL of degassed acetonitrile and cooled to 0 °C. CCl₄ (1.0 mL), Et₃N (2.0 mmol) and saturated aqueous solution of NH₄OH (28% in water, 0.5 mL) were sequentially added dropwise while stirring. The solution was stirred at 0 °C for 30 min and then warmed to RT and allowed to stir for 16 h. Water (5 mL) was added to the reaction mixture and then extracted with EtOAc, the organic layers were combined, dried over anhydrous Na₂SO₄ and concentrated to give the crude product. The pure product was obtained after first passing the crude through a short silica gel column and then doing a crystallization in DCM/Et₂O (1:5, v/v) at -20 °C to obtain the phosphoramidate products as highly enriched single enantiomers (92-99% ee).

(*R*)-*P*-(*tert*-butyl)-*P*-phenylphosphinic amide (**8a**); characterization data consistent with previously reported.⁷



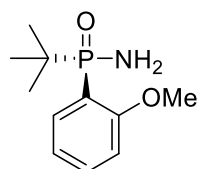
Isolated as a white solid in 82% yield (162 mg) and 96.7% ee.

¹H NMR (500 MHz, CDCl₃): δ 7.90–7.84 (m, 2H), 7.57–7.52 (m, 1H), 7.49–7.43 (m, 2H), 2.72 (brs, 2H), 1.16 (d, *J* = 15.3 Hz, 9H).

³¹P NMR (202 MHz, CDCl₃): δ 41.34.

Chiral HPLC method: Chiralcel OD, hexane/IPA = 80/20, 1.0 mL/min, λ = 220 nm; (*R*)-enantiomer *t*_R (major) = 5.88 min, (*S*)-enantiomer *t*_R (minor) = 7.60 min.

(*R*)-*P*-(*tert*-butyl)-*P*-(2-methoxyphenyl)phosphinic amide (**8b**):



Isolated as a white solid in 73% yield (166 mg) and 95% ee.

¹H NMR (500 MHz, CDCl₃): δ 7.90 (ddd, *J* = 11.9, 7.5, 1.8 Hz, 1H), 7.49 – 7.40 (m, 1H), 7.10 – 7.02 (m, 1H), 6.93 – 6.87 (m, 1H), 3.83 (s, 3H), 3.17 (s, 2H), 1.08 (d, *J* = 15.9 Hz, 9H).

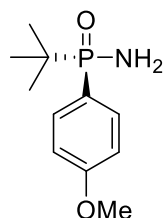
¹³C NMR (126 MHz, CDCl₃): δ 159.2 (d, *J* = 3.9 Hz), 135.5 (dd, *J* = 5.5, 3.0 Hz), 133.3, 121.0 (dd, *J* = 10.6, 2.5 Hz), 119.4 (d, *J* = 101.4 Hz), 110.6 (d, *J* = 7.0 Hz), 55.2, 34.3 (d, *J* = 93.7 Hz), 24.2.

³¹P NMR (203 MHz, CDCl₃): δ 46.01.

HRMS: calculated for C₁₁H₁₈NNaO₂P⁺ [M+H]⁺: 250.0967, found: 250.0967.

Chiral HPLC method: Chiralcel OD, hexane/IPA = 80/20, 1.0 mL/min, λ = 220 nm; (*S*)-enantiomer *t*_R (minor) = 7.38 min, (*R*)-enantiomer *t*_R (major) = 10.23 min.

(*R*)-*P*-(*tert*-butyl)-*P*-(4-methoxyphenyl)phosphinic amide (**8c**)



Isolated as a white solid in 48% yield (109 mg) and >99% ee.

¹H NMR (500 MHz, CDCl₃) δ 7.77 – 7.70 (m, 2H), 6.92 (dd, *J* = 8.9, 2.4 Hz, 2H), 3.83 (s, 3H), 2.84 (s, 2H), 1.11 (d, *J* = 15.2 Hz, 9H).

¹³C NMR (126 MHz, CDCl₃): δ 162.4 (d, *J* = 2.9 Hz), 135.0 (d, *J* = 9.6 Hz), 121.4 (d,

$J = 123.0$ Hz), 113.6 (d, $J = 12.5$ Hz), 55.2, 32.3 (d, $J = 93.5$ Hz), 24.8.

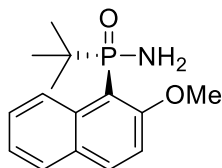
^{31}P NMR (203 MHz, CDCl_3): δ 41.44.

HRMS: calculated for $\text{C}_{11}\text{H}_{18}\text{NNaO}_2\text{P}^+$ $[\text{M}+\text{H}]^+$: 250.0967, found: 250.0968.

Chiral HPLC method: Chiralcel OD, hexane/IPA = 80/20, 1.0 mL/min, $\lambda = 220$ nm;

(*R*)-enantiomer t_{R} (major) = 7.08 min, (*S*)-enantiomer t_{R} (minor) = 12.49 min.

(*R*)-*P*-(*tert*-butyl)-*P*-(2-methoxynaphthalen-1-yl)phosphinic amide (**8d**)



Isolated as a white solid in 42% yield (116 mg) and >99% ee.

^1H NMR (400 MHz, CDCl_3): δ 9.59 (d, $J = 8.5$ Hz, 1H), 7.99 (d, $J = 9.1$ Hz, 1H), 7.76 (dt, $J = 8.2, 1.7$ Hz, 1H), 7.54 (ddd, $J = 8.6, 6.7, 1.5$ Hz, 1H), 7.38 (ddd, $J = 8.0, 6.8, 1.2$ Hz, 1H), 7.31 – 7.23 (m, 1H), 3.99 (s, 3H), 3.22 (s, 2H), 1.16 (d, $J = 16.0$ Hz, 9H).

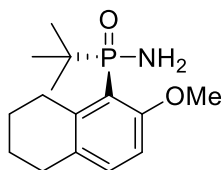
^{13}C NMR (101 MHz, CDCl_3): δ 158.7 (d, $J = 3.2$ Hz), 136.7 (d, $J = 6.7$ Hz), 134.7, 129.4 (d, $J = 9.1$ Hz), 128.0, 127.6 (d, $J = 2.2$ Hz), 127.5, 124.2, 112.8, 111.9, 111.8, 56.0, 35.9 (d, $J = 93.2$ Hz), 24.4.

^{31}P NMR (162 MHz, CDCl_3): δ 50.06.

HRMS: calculated for $\text{C}_{15}\text{H}_{20}\text{NNaO}_2\text{P}^+$ $[\text{M}+\text{H}]^+$: 300.1124, found: 300.1115.

Chiral HPLC method: Chiralcel OD, hexane/IPA = 80/20, 1.0 mL/min, $\lambda = 220$ nm; (*S*)-enantiomer $t_{\text{R}} = 8.46$ min, (*R*)-enantiomer t_{R} (single peak) = 28.96 min.

(*R*)-*P*-(*tert*-butyl)-*P*-(2-methoxy-5,6,7,8-tetrahydronaphthalen-1-yl)phosphinic amide (**8e**)



Isolated as a white solid in 45% yield (126 mg) and >99% ee.

^1H NMR (500 MHz, CDCl_3): δ 7.15 (d, $J = 8.5$ Hz, 1H), 6.72 (dd, $J = 8.5, 4.8$ Hz, 1H), 3.79 (s, 3H), 3.65 – 3.56 (m, 1H), 3.20 (dt, $J = 17.7, 5.7$ Hz, 1H), 3.12 (s, 2H), 2.72 (t, $J = 6.6$ Hz, 2H), 1.86 – 1.77 (m, 1H), 1.75 – 1.66 (m, 2H), 1.65 – 1.55 (m, 1H), 1.13 (d, $J = 15.8$ Hz, 9H).

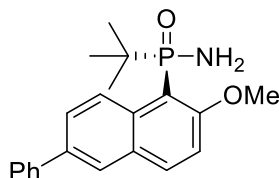
^{13}C NMR (126 MHz, CDCl_3): δ 158.09 (d, $J = 4.6$ Hz), 145.82 (d, $J = 7.0$ Hz), 133.25 (d, $J = 2.1$ Hz), 131.73 (d, $J = 10.3$ Hz), 117.61 (d, $J = 95.2$ Hz), 108.24 (d, $J = 7.4$ Hz), 55.16, 35.83 (d, $J = 91.8$ Hz), 29.87, 28.51 (d, $J = 2.2$ Hz), 24.41 (d, $J = 0.9$ Hz), 22.39 (d, $J = 84.5$ Hz).

^{31}P NMR (203 MHz, CDCl_3): δ 50.18.

HRMS: calculated for $C_{15}H_{24}NNaO_2P^+$ $[M+H]^+$: 304.1437, found: 304.1142.

Chiral HPLC method: Chiralcel OD, hexane/IPA = 80/20, 1.0 mL/min, $\lambda = 220$ nm; (*S*)-enantiomer $t_R = 4.96$ min, (*R*)-enantiomer t_R (single peak) = 6.20 min.

(*R*)-*P*-(*tert*-butyl)-*P*-(2-methoxy-6-phenylnaphthalen-1-yl)phosphinic amide (**8f**)



Isolated as a white solid in 60% yield (212 mg) and >99% ee.

1H NMR (500 MHz, $CDCl_3$): δ 9.64 (d, $J = 9.1$ Hz, 1H), 8.00 (d, $J = 9.0$ Hz, 1H), 7.94 (s, 1H), 7.80 (dd, $J = 9.1, 2.1$ Hz, 1H), 7.70 (d, $J = 7.0$ Hz, 2H), 7.47 (t, $J = 7.7$ Hz, 2H), 7.36 (t, $J = 7.4$ Hz, 1H), 7.24 (dd, $J = 9.0, 4.3$ Hz, 1H), 3.95 (s, 3H), 3.31 (brs, 2H), 1.17 (d, $J = 16.1$ Hz, 9H).

^{13}C NMR (126 MHz, $CDCl_3$): δ 158.7 (d, $J = 3.2$ Hz), 140.6, 136.5, 135.9 (d, $J = 6.7$ Hz), 134.9 (d, $J = 2.1$ Hz), 129.7 (d, $J = 9.0$ Hz), 128.9, 128.2 (d, $J = 2.1$ Hz), 127.3, 127.2, 127.0, 125.63, 112.4 (d, $J = 93.6$ Hz), 112.3 (d, $J = 7.7$ Hz), 56.0, 35.9 (d, $J = 93.2$ Hz), 24.4.

^{31}P NMR (203 MHz, $CDCl_3$): δ 50.11.

HRMS: calculated for $C_{21}H_{25}NO_2P^+$ $[M+H]^+$: 354.1617, found: 354.1618.

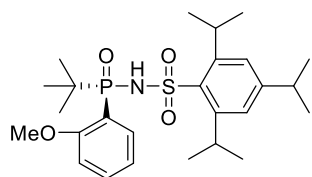
Chiral HPLC method: Chiralcel OD, hexane/IPA = 80/20, 1.0 mL/min, $\lambda = 220$ nm; (*S*)-enantiomer $t_R = 10.91$ min, (*R*)-enanatiomer (single peak) $t_R = 15.34$ min.

General procedure for the conversion of arylphosphinic amides **8** to Brønsted acids **3b** and **4**:

A slurry of NaH (3 equiv of 60% NaH in oil) in anhydrous THF (3.0 mL) at 0 °C was added to a solution of phosphinamide **8** (0.5 mmol, 1 equiv) and the mixture was stirred for 30 min. The arylsulfonyl chloride (1.5 equiv) was added slowly, and the mixture was warmed to RT and monitored by TLC. After complete conversion (~12-15 h), NH_4Cl (0.1 g) was added portion-wise, the mixture was diluted with THF and filtered. The filtrate was concentrated and the crude residue was purified by flash column chromatography on silica gel to give the desired product. The product was dissolved in DCM (15 mL) and thoroughly washed with 4 M HCl (2x) to remove any salt impurities and completely protonate the catalyst. The organic layer was separated and concentrated under reduced pressure. The residue was taken up in toluene (5 mL), evaporated to dryness again and dried under high vacuum for 24 h to give the catalyst.

Note: Upon completion of the coupling reaction between intermediate **8** and the sulfonyl chloride, some analogs **4** were used directly in the subsequent demethylation step (without isolation/purification) to get the final catalysts **5**.

(R)-*N*-(*tert*-butyl(2-methoxyphenyl)phosphoryl)-2,4,6-triisopropylbenzenesulfonamide (**3b**): This compound was recently reported by Han and coworkers.⁸



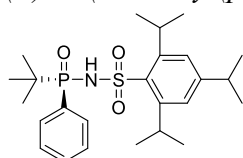
Isolated as a yellow solid in 97% yield (240 mg).

¹H NMR (500 MHz, CD₃OD): δ 7.74 (ddd, *J* = 7.0, 1.5 Hz, 1H), 7.37 (t, *J* = 7.3 Hz, 1H), 7.11 (s, 2H), 6.91 (dd, *J* = 7.8, 5.3 Hz, 1H), 6.77 (t, *J* = 7.0 Hz, 1H), 4.49 - 4.40 (m, 2H), 3.54 (s, 3H), 2.88 (hept, *J* = 6.9 Hz, 1H), 1.27 - 1.21 (m, 12H), 1.14 - 1.04 (m, 15H).

³¹P NMR (202 MHz, CD₃OD): δ 33.97;

HRMS: calculated for C₂₆H₄₀NO₄PSNa [M+Na]: 516.2308, found: 516.2305.

(R)-*N*-(*tert*-butyl(phenyl)phosphoryl)-2,4,6-triisopropylbenzenesulfonamide (**4a**):



Isolated as a yellow solid in 96% yield (223 mg).

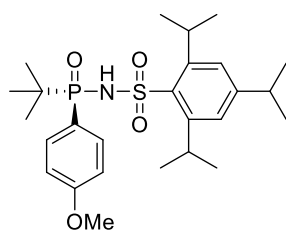
¹H NMR (500 MHz, CD₃OD): δ 7.66-7.62 (m, 2H), 7.35 (td, *J* = 7.5, 1.5 Hz, 1H), 7.22 (td, *J* = 8.0, 3.0 Hz, 2H), 7.04 (s, 2H), 4.51-4.45 (m, 2H), 2.88-2.82 (m, 1H), 1.23 (d, *J* = 7.0 Hz, 6H), 1.21 (d, *J* = 6.5 Hz, 6H), 1.06 (d, *J* = 7.0 Hz, 6H), 1.01 (d, *J* = 15.5 Hz, 9H);

¹³C NMR (125 MHz, CD₃OD): δ 150.8, 149.2, 142.7, 134.8 (d, *J* = 112.0 Hz), 134.5 (d, *J* = 8.1 Hz), 131.3 (d, *J* = 1.8 Hz), 128.1 (d, *J* = 10.9 Hz), 123.6, 35.3, 33.9 (d, *J* = 103.8 Hz), 30.2, 25.6, 25.2, 25.1, 24.33, 24.30;

³¹P NMR (202 MHz, CD₃OD): δ 31.74;

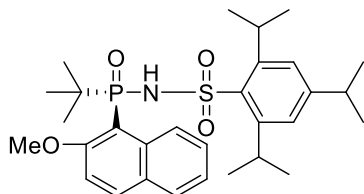
HRMS: calculated for C₂₅H₃₈NO₃PSNa [M+Na]⁺: 486.2202, found: 486.2194.

(R)-*N*-(*tert*-butyl(4-methoxyphenyl)phosphoryl)-2,4,6-triisopropylbenzenesulfonamide (**4b**):



Upon completion of the coupling reaction with the sulfonyl chloride, compound **4b** was used directly in the subsequent demethylation step to get the final catalyst **5b**.

(R)-*N*-(*tert*-butyl(2-methoxynaphthalen-1-yl)phosphoryl)-2,4,6-triisopropylbenzenesulfonamide (**4c**):



Compound was isolated as a white solid in 98% yield (173 mg).

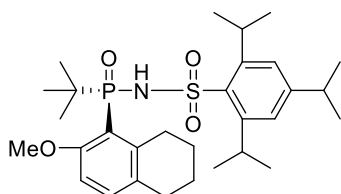
^1H NMR (500 MHz, CDCl_3): δ 9.43 (d, $J = 8.9$ Hz, 1H), 8.02 (d, $J = 9.1$ Hz, 1H), 7.75 (d, $J = 8.0$ Hz, 1H), 7.56 – 7.43 (m, 2H), 7.37 (t, $J = 8.0$ Hz, 1H), 7.32 – 7.27 (m, 1H), 7.05 (s, 2H), 4.16 (hept, $J = 6.5$ Hz, 2H), 4.10 (s, 3H), 2.81 (hept, $J = 7.0$ Hz, 1H), 1.31 – 1.11 (m, 21H), 1.10 (d, $J = 6.8$ Hz, 6H).

^{13}C NMR (126 MHz, CDCl_3): δ 158.42, 152.56, 150.27, 136.80 (d, $J = 7.6$ Hz), 135.92 (d, $J = 2.3$ Hz), 135.43, 129.60 (d, $J = 10.0$ Hz), 128.28, 127.99, 127.57 (d, $J = 2.3$ Hz), 124.64, 123.75, 111.89 (d, $J = 8.2$ Hz), 110.79, 110.03, 56.53, 37.40 (d, $J = 89.4$ Hz), 34.18, 29.91, 24.91 (d, $J = 3.4$ Hz), 24.60, 23.63 (d, $J = 5.8$ Hz).

^{31}P NMR (203 MHz, CDCl_3): δ 43.1.

HRMS: calculated for $\text{C}_{30}\text{H}_{41}\text{O}_4\text{NPS}$: 542.2499, found: 542.2491.

(R)-*N*-(*tert*-butyl(2-methoxy-5,6,7,8-tetrahydronaphthalen-1-yl)phosphoryl)-2,4,6-triisopropylbenzenesulfonamide (**4d**):

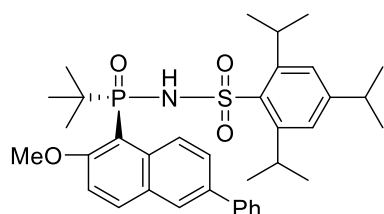


^1H NMR (500 MHz, CDCl_3): δ 7.47 (s, 1H), 7.17 (d, $J = 8.5$ Hz, 1H), 7.08 (s, 2H), 6.74 (dd, $J = 8.5, 5.2$ Hz, 1H), 4.18 (hept, $J = 6.7$ Hz, 2H), 3.89 (s, 3H), 3.40 (dt, $J = 18.0, 6.2$ Hz, 1H), 3.03 (dt, $J = 17.9, 5.5$ Hz, 1H), 2.85 (hept, $J = 6.9$ Hz, 1H), 2.74 – 2.64 (m, 2H), 1.73 – 1.56 (m, 4H), 1.26 (d, $J = 6.7$ Hz, 6H), 1.24 – 1.15 (m, 15H), 1.13 (d, $J = 6.7$ Hz, 6H).

^{31}P NMR (203 MHz, CDCl_3): δ 44.0.

Upon completion of the coupling reaction with the sulfonyl chloride, the crude compound **4d** was used directly in the subsequent demethylation step to get the final catalyst **5d**.

(*R*)-*N*-(*tert*-butyl(2-methoxy-6-phenylnaphthalen-1-yl)phosphoryl)-2,4,6-triisopropylbenzenesulfonamide (**4e**):

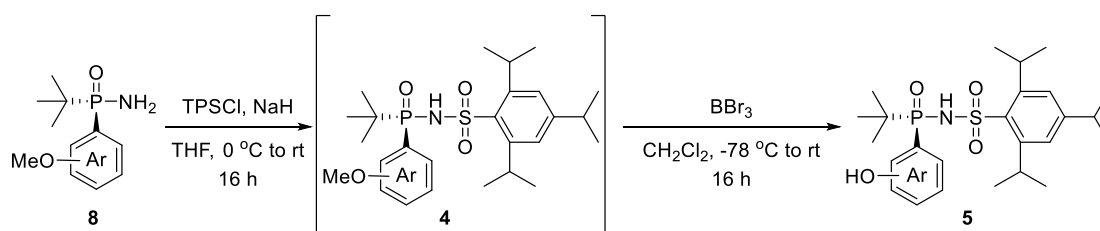


^1H NMR (500 MHz, CDCl_3) δ 9.51 (d, $J = 9.2$ Hz, 1H), 8.08 (d, $J = 9.1$ Hz, 1H), 7.95 (t, $J = 2.1$ Hz, 1H), 7.75 (dd, $J = 9.2, 2.1$ Hz, 1H), 7.68 (dd, $J = 8.3, 1.3$ Hz, 2H), 7.52 (d, $J = 6.4$ Hz, 1H), 7.47 (t, $J = 7.7$ Hz, 2H), 7.39 – 7.34 (m, 1H), 7.31 (dd, $J = 9.1, 4.7$ Hz, 1H), 7.06 (s, 2H), 4.18 (hept, $J = 6.7$ Hz, 2H), 4.11 (s, 3H), 2.81 (hept, $J = 6.8$ Hz, 1H), 1.26 – 1.20 (m, 15H), 1.18 (dd, $J = 6.9, 2.4$ Hz, 6H), 1.12 (d, $J = 6.7$ Hz, 6H).

^{31}P NMR (203 MHz, CDCl_3): δ 43.2.

Upon completing of the coupling reaction with the sulfonyl chloride, compound **4e** the crude product was used directly in the subsequent demethylation step to get the final catalyst **5e**.

General procedure for the synthesis of Brønsted acids **5**:

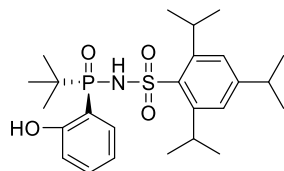


Note: Upon completion of the coupling reaction between intermediate **8** and the sulfonyl chloride, some analogs **4** were used directly in the subsequent demethylation step to get the final catalysts **5**.

Demethylation step: A solution of intermediates **3b** or **4** in dry DCM (5 mL) was cooled to -78 °C, then BBr_3 (1.2 equiv in hexane) was added slowly over a 5 min period. After the addition was finished, the reaction mixture was allowed to warm-up to RT and stirred overnight. The reaction was quenched with water and diluted with DCM. The organic fraction was washed with 1 N HCl, dried over anhydrous MgSO_4 , concentrated and purified by flash column chromatography on silica gel. The isolated product was re-dissolved in DCM (15 mL) and thoroughly washed with 4 M HCl (15 mL x 2) to remove any metal impurities and completely protonate the catalyst. The organic layer

was separated and concentrated under reduced pressure. The residue was taken up in toluene (5 mL), evaporated to dryness again and allowed to dry under high vacuum for a minimum of 24 h to give (*R*)-phenolic catalyst.

(R)-*N*-(*tert*-butyl(2-hydroxyphenyl)phosphoryl)-2,4,6-triisopropylbenzenesulfonamide (**5a**):



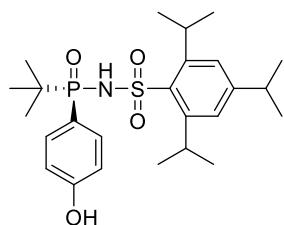
Compound was isolated as a yellow solid in 92% yield (221 mg).

^1H NMR (400 MHz, CDCl_3): δ 10.60 (s, 1H), 7.42 (ddt, $J = 8.4, 7.1, 1.4$ Hz, 1H), 7.31–7.23 (m, 1H), 7.14 (s, 2H), 6.97 – 6.89 (m, 1H), 6.84 – 6.75 (m, 1H), 6.41 (d, $J = 8.7$ Hz, 1H), 3.91 (hept, $J = 6.6$ Hz, 2H), 2.90 (hept, $J = 6.9$ Hz, 1H), 1.30 – 1.14 (m, 27H).
 ^{13}C NMR (126 MHz, CDCl_3): δ 163.6 (d, $J = 5.0$ Hz), 153.7, 150.3, 135.1 (d, $J = 2.1$ Hz), 134.2 (s), 133.0 (d, $J = 8.4$ Hz), 123.9, 118.8 (d, $J = 12.5$ Hz), 118.0 (d, $J = 9.3$ Hz), 107.5 (d, $J = 117.5$ Hz), 34.6 (d, $J = 86.2$ Hz), 34.2, 30.1, 24.7 (d, $J = 47.0$ Hz), 23.8, 23.5 (d, $J = 4.2$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 45.42.

HRMS: calculated for $\text{C}_{25}\text{H}_{39}\text{NO}_4\text{PS}^+$ $[\text{M}+\text{H}]^+$: 480.2332, found: 480.2336.

(R)-*N*-(*tert*-butyl(4-hydroxyphenyl)phosphoryl)-2,4,6-triisopropylbenzenesulfonamide: (**5b**)



Isolated as a yellow solid, in 80% yield (193 mg).

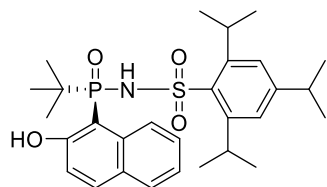
^1H NMR (500 MHz, MeOD): δ 7.51–7.44 (m, 2H), 7.17 (s, 2H), 6.77 (dd, $J = 8.5, 2.5$ Hz, 2H), 4.27–4.14 (m, 2H), 2.96–2.87 (m, 1H), 1.31–1.20 (m, 12H), 1.18–1.03 (m, 15H).

^{13}C NMR (126 MHz, MeOD): δ 161.4, 152.4, 149.6, 136.6, 134.8 (d, $J = 11.0$ Hz), 123.1, 116.5 (d, $J = 125.4$ Hz), 114.7 (d, $J = 13.8$ Hz), 34.0, 33.1 (d, $J = 93.4$ Hz), 28.9, 23.8 (d, $J = 38.7$ Hz), 23.0, 22.7 (d, $J = 6.2$ Hz).

^{31}P NMR (203 MHz, MeOD): δ 41.15.

HRMS: calculated for $\text{C}_{25}\text{H}_{39}\text{NO}_4\text{PS}^+$ $[\text{M}+\text{H}]^+$: 480.2332, found: 480.2335.

(R)-*N*-(*tert*-butyl(2-hydroxynaphthalen-1-yl)phosphoryl)-2,4,6-triisopropylbenzenesulfonamide (**5c**):



Isolated as a yellow solid in 95% yield (252 mg).

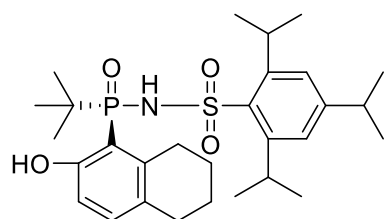
^1H NMR (400 MHz, CDCl_3): δ 12.55 (s, 1H), 8.15 (m, 1H), 7.88 (d, $J = 8.9$ Hz, 1H), 7.70 (m, 1H), 7.25 – 7.20 (m, 2H), 7.15 – 7.06 (m, 3H), 6.56 (s, 1H), 3.88 (hept, $J = 6.7$ Hz, 2H), 2.88 (hept, $J = 6.9$ Hz, 1H), 1.36 – 1.14 (m, 21H), 1.11 (d, $J = 6.7$ Hz, 6H).

^{13}C NMR (126 MHz, CDCl_3): δ 166.5 (d, $J = 4.8$ Hz), 153.4, 150.1, 136.5 (d, $J = 2.3$ Hz), 134.6, 133.6 (d, $J = 8.6$ Hz), 128.4 (d, $J = 10.2$ Hz), 128.0 (d, $J = 197.2$ Hz), 125.5 (d, $J = 4.2$ Hz), 123.9, 123.2, 120.4 (d, $J = 11.1$ Hz), 36.8 (d, $J = 85.8$ Hz), 34.2, 30.2, 24.8, 24.7 (d, $J = 39.8$ Hz), 23.5 (d, $J = 4.6$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 47.72.

HRMS: calculated for $\text{C}_{29}\text{H}_{41}\text{NO}_4\text{PS}^+$ $[\text{M}+\text{H}]^+$: 530.2488, found: 530.2496.

(R)-*N*-(*tert*-butyl(2-hydroxy-5,6,7,8-tetrahydronaphthalen-1-yl)phosphoryl)-2,4,6-triisopropylbenzenesulfonamide (**5d**):



Isolated as a yellow solid in 93% yield (248 mg).

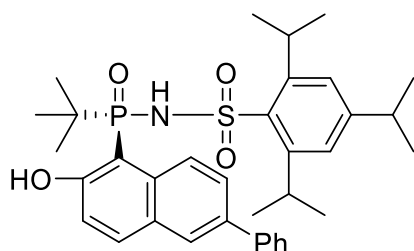
^1H NMR (400 MHz, CDCl_3): δ 11.50 (s, 1H), 7.17 (s, 2H), 7.14 (d, $J = 8.4$ Hz, 1H), 6.75 (dd, $J = 8.4, 5.2$ Hz, 1H), 6.19 (d, $J = 10.9$ Hz, 1H), 3.97 (hept, $J = 6.5$ Hz, 2H), 3.13 (ddd, $J = 15.9, 10.1, 5.4$ Hz, 1H), 2.95 – 2.78 (m, 2H), 2.75 – 2.67 (m, 2H), 1.98 – 1.68 (m, 4H), 1.31 (d, $J = 6.6$ Hz, 6H), 1.27 – 1.22 (m, 12H), 1.18 (d, $J = 16.6$ Hz, 9H).

^{13}C NMR (126 MHz, CDCl_3): δ 162.9 (d, $J = 5.6$ Hz), 153.3, 150.2, 140.3 (d, $J = 9.1$ Hz), 136.0, 135.0, 128.9 (d, $J = 10.4$ Hz), 124.0, 116.4 (d, $J = 11.0$ Hz), 106.3 (d, $J = 109.8$ Hz), 36.2 (d, $J = 83.9$ Hz), 34.1, 30.3, 29.5, 24.8 (d, $J = 84.6$ Hz), 24.8 (d, $J = 14.8$ Hz), 24.3, 23.6, 22.4, 22.1.

^{31}P NMR (162 MHz, CDCl_3): δ 49.00.

HRMS: calculated for $\text{C}_{29}\text{H}_{45}\text{NO}_4\text{PS}^+$ $[\text{M}+\text{H}]^+$: 534.2801, found: 534.2808.

(*R*)-*N*-(*tert*-butyl(2-hydroxy-6-phenylnaphthalen-1-yl)phosphoryl)-2,4,6-triisopropylbenzenesulfonamide (**5e**):



Isolated as a yellow solid in 88% yield (267 mg).

^1H NMR (500 MHz, CDCl_3) δ 12.49 (s, 1H), 8.30 (d, $J = 8.9$ Hz, 1H), 7.85 (d, $J = 8.9$ Hz, 1H), 7.64 (s, 1H), 7.44 – 7.30 (m, 6H), 7.17 (s, 2H), 7.13 (dd, $J = 8.9, 4.7$ Hz, 1H), 3.98 (hept, $J = 6.7$ Hz, 2H), 2.89 (hept, $J = 6.9$ Hz, 1H), 1.35 (d, $J = 6.7$ Hz, 6H), 1.28 – 1.12 (m, 21H).

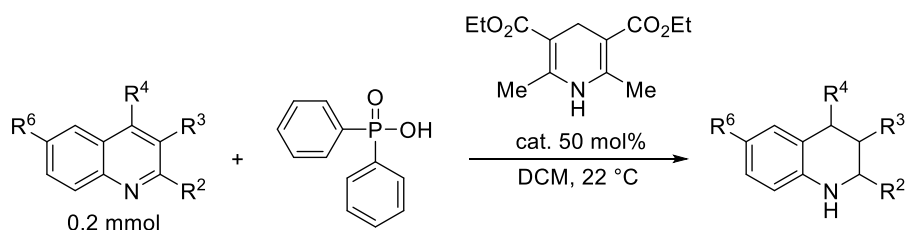
^{13}C NMR (126 MHz, CDCl_3): δ 166.2 (d, $J = 4.3$ Hz), 153.6, 150.6, 140.0, 136.8, 135.3, 134.7, 132.9 (d, $J = 8.7$ Hz), 128.6 (d, $J = 10.1$ Hz), 128.3, 126.9, 126.6 (d, $J = 126.9$ Hz), 126.3 (d, $J = 4.1$ Hz), 124.0, 120.3 (d, $J = 10.8$ Hz), 98.3 (d, $J = 111.8$ Hz), 36.9 (d, $J = 85.1$ Hz), 34.2, 30.3, 25.2, 24.6, 23.5 (d, $J = 6.9$ Hz).

^{31}P NMR (203 MHz, CDCl_3): δ 48.88.

HRMS: calculated for $\text{C}_{35}\text{H}_{45}\text{NO}_4\text{PS}^+$ $[\text{M}+\text{H}]^+$: 606.2801, found: 606.2803.

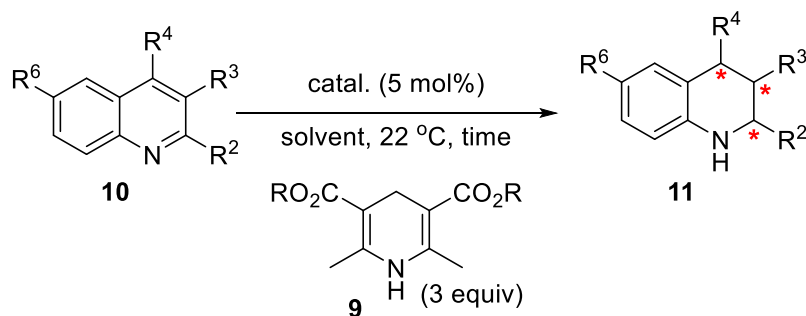
General procedure for the transfer hydrogenation of quinolines **10** to the tetrahydroquinolines **11**:

General procedure for the synthesis of the racemic tetrahydroquinolines:



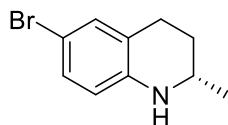
An oven dried 2 dram vial equipped with a stir bar was cooled to ambient temperature in a desiccator and subsequently charged with the requisite quinoline (0.200 mmol). 1 mL of DCM, Hantzsch ester (152 mg, 0.600 mmol) and diphenylphosphinic acid (21.8 mg, 0.500 mmol). The vial was capped under air, sealed with parafilm and the mixture was stirred at RT for 2-24 h. Progress of the reaction was monitored by TLC (20% EtOAc and 80% hexanes). The crude product was purified by flash chromatography on silica gel (using EtOAc/hexanes) to afford the desired tetrahydroquinoline.

General procedure for the asymmetric transfer hydrogenation of quinolines **10** to the tetrahydroquinolines **11**:



An oven-dried flask was fitted with magnetic stirring bar and charged with the quinoline (reactions were typically carried out at a 0.1-0.2 mmol scale), catalyst (5 mol%), Hantzsch ester (3.0 equiv) and solvent (0.5 mL). The resulting mixture was stirred at RT (~22 °C), unless otherwise indicated and monitored by TLC. When all starting material was consumed, the solvent was removed under reduced pressure and the residue was purified by flash column chromatography on silica gel using the solvent system indicated to isolate the corresponding product.

(*S*)-6-bromo-2-methyl-1,2,3,4-tetrahydroquinoline (**11a**):



Reaction time: 5 h using catalyst **5c**. Known compound⁹, purified using a 0-2% EtOAc/hexanes eluent gradient; isolated as white solid in 98% yield (44.2 mg) and 88% ee. The compound was crystallized from DCM/hexanes to afford the tetrahydroquinoline **11e** in 93% ee.

¹H NMR (500 MHz, CDCl₃): δ 7.09 – 7.05 (d, *J* = 2.4 Hz, 1H), 7.03 (dd, *J* = 8.4, 2.4 Hz, 1H), 6.34 (d, *J* = 8.4 Hz, 1H), 3.76 (s, 1H), 3.38 (dq, *J* = 9.3, 6.3, 2.9 Hz, 1H), 2.80 (ddd, *J* = 17.0, 11.5, 5.7 Hz, 1H), 2.75 – 2.64 (dt, 1H), 1.98 – 1.87 (m, 1H), 1.55 (m, 1H), 1.20 (d, *J* = 6.3 Hz, 3H).

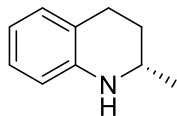
¹³C NMR (101 MHz, CDCl₃): δ 143.9, 131.8, 129.5, 123.3, 115.5, 108.4, 47.2, 29.8, 26.5, 22.6.

Chiral HPLC method: Chiralpak OD-H, hexane/IPA = 98/2, 1.0 mL/min, λ = 254 nm; (*R*)-enantiomer *t_R* (minor) = 8.6 min, (*S*)-enantiomer *t_R* (major) = 11.2 min.

For the purpose of comparison the product was also analyzed using the same chiral HPLC column and solvent system as previously reported:⁹ Chiralcel OJ-H, hexane/IPA = 95/5, 0.8 mL/min, λ = 254 nm; (*S*)-enantiomer *t_R* (major) = 19.04 min, (*R*)-enantiomer *t_R* (minor) = 23.19 min.

(S)-2-methyl-1,2,3,4-tetrahydroquinoline (**11b**):

NMR and chiral HPLC data consistent with those previously reported.¹⁰



Compound was purified using a 0-1% EtOAc/hexanes as the eluent; isolated as a pale-yellow oil in 73% yield (21.6 mg) and 88% ee.

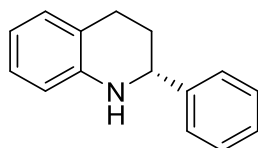
¹H NMR (500 MHz, CDCl₃): δ 7.02 – 6.90 (m, 2H), 6.60 (td, *J* = 7.3, 1.2 Hz, 1H), 6.47 (dd, *J* = 8.4, 1.2 Hz, 1H), 3.70 (broad s, 1H), 3.40 (dq, *J* = 10.0, 6.3, 2.8 Hz, 1H), 2.90 – 2.78 (m, 1H), 2.73 (ddd, *J* = 16.3, 5.2, 3.4 Hz, 1H), 1.93 (dddd, *J* = 12.8, 5.6, 3.4, 2.8 Hz, 1H), 1.63 – 1.55 (m, 1H), 1.21 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃): δ 144.9, 129.4, 126.8, 121.3, 117.1, 114.1, 47.3, 30.3, 26.7, 22.8.

Chiral HPLC method: Chiralcel OD, hexane/IPA = 98/2, 1.0 mL/min, λ = 254 nm; (*R*)-enantiomer *t*_R (minor) = 6.78 min, (*S*)-enantiomer (*major*) *t*_R = 7.79 min.

(R)-2-phenyl-1,2,3,4-tetrahydroquinoline (**11c**):

NMR and chiral HPLC data consistent with those previously reported.¹⁰



Compound was purified using a 0-3% EtOAc/hexanes eluent and isolated as a white solid 95% yield (39.9 mg) and 59.3% ee.

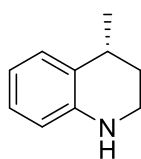
¹H NMR (500 MHz, CDCl₃): δ 7.45 – 7.33 (m, 4H), 7.32 – 7.27 (m, 1H), 7.07 – 6.97 (m, 2H), 6.66 (td, *J* = 7.3, 1.2 Hz, 1H), 6.55 (d, *J* = 7.5 Hz, 1H), 4.45 (dd, *J* = 9.4, 3.3 Hz, 1H), 4.05 (s, 1H), 2.93 (ddd, *J* = 16.2, 10.7, 5.5 Hz, 1H), 2.75 (dt, *J* = 16.3, 4.8 Hz, 1H), 2.13 (dddd, *J* = 13.1, 5.4, 4.5, 3.3 Hz, 1H), 2.00 (dddd, *J* = 13.0, 10.7, 9.3, 5.1 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃): δ 144.9, 144.9, 129.4, 128.7, 127.6, 127.0, 126.7, 121.0, 117.3, 114.1, 56.4, 31.1, 26.5.

Chiral HPLC method: Chiralcel OD, hexane/IPA = 98/2, 1.0 mL/min, λ = 254 nm; (*S*)-enantiomer *t*_R (minor) = 15.13 min, (*R*)-enantiomer (*major*) *t*_R = 21.39 min (*major*).

(R)-4-methyl-1,2,3,4-tetrahydroquinoline (**11d**):

NMR and chiral HPLC data consistent with those previously reported¹¹



Compound purified using a 0-20% Et₂O in pentane and isolated a pale-yellow oil in 70% yield and 30% ee.

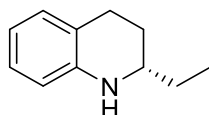
¹H NMR (500 MHz, CDCl₃): δ 7.06 (d, *J* = 7.6 Hz, 1H), 6.96 (tdd, *J* = 7.3, 1.7, 0.8 Hz, 1H), 6.63 (td, *J* = 7.4, 1.2 Hz, 1H), 6.48 (dd, *J* = 7.9, 1.2 Hz, 1H), 3.91 (s, 1H), 3.39 – 3.23 (m, 2H), 2.92 (h, *J* = 6.6 Hz, 1H), 2.04 – 1.94 (m, 1H), 1.68 (dddd, *J* = 13.0, 6.9, 6.1, 3.5 Hz, 1H), 1.29 (d, *J* = 7.0 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃): δ 144.3, 128.6, 126.9, 126.8, 117.1, 114.3, 39.2, 30.4, 30.0, 22.8.

Chiral HPLC method: Chiralcel OD, hexane/IPA = 98/2, 0.6 mL/min, λ = 254 nm; (*S*)-enantiomer *t*_R (minor) = 16.41 min, (*R*)-enantiomer *t*_R (major) = 17.67 min.

(S)-2-ethyl-1,2,3,4-tetrahydroquinoline (**11e**):

NMR and chiral HPLC data consistent with those previously reported.¹⁰



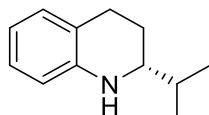
Compound was purified using a 0-2% EtOAc in hexanes and isolated as a pale-yellow oil in 72% yield (23.1 mg) and 75% ee.

¹H NMR (500 MHz, CDCl₃): δ 6.96 (ddt, *J* = 8.2, 7.4, 0.8 Hz, 2H), 6.60 (td, *J* = 7.4, 1.2 Hz, 1H), 6.48 (dt, *J* = 7.4, 1.3 Hz, 1H), 3.77 (s, 1H), 3.17 (dtd, *J* = 9.4, 6.4, 2.9 Hz, 1H), 2.88 – 2.77 (m, 1H), 2.73 (ddd, *J* = 16.3, 5.4, 4.0 Hz, 1H), 1.98 (dddd, *J* = 12.7, 5.6, 4.0, 2.9 Hz, 1H), 1.65 – 1.56 (m, 1H), 1.56 – 1.49 (m, 2H), 1.00 (t, *J* = 7.5 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃): δ 144.9, 129.4, 126.8, 121.5, 117.0, 114.1, 53.2, 29.6, 27.7, 26.6, 10.2.

Chiral HPLC method: Chiralcel OD, hexane/IPA = 98/2, 1.0 mL/min, λ = 254 nm; (*R*)-enantiomer *t*_R (minor) = 6.56 min, (*S*)-enantiomer *t*_R (major) = 7.91 min.

(R)-2-isopropyl-1,2,3,4-tetrahydroquinoline (**11f**):



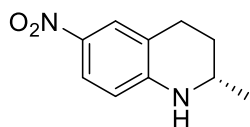
NMR and chiral HPLC data consistent with those previously reported,¹⁰ purified using a 0-2% EtOAc/hexanes eluent; isolated as a pale-yellow oil in 77% yield (27.0 mg) and 66% ee.

¹H NMR (500 MHz, CDCl₃): δ 7.00 – 6.91 (m, 2H), 6.59 (td, *J* = 7.3, 1.2 Hz, 1H), 6.52 – 6.45 (m, 1H), 3.76 (s, 1H), 3.04 (ddd, *J* = 10.0, 5.9, 2.9 Hz, 1H), 2.81 (ddd, *J* = 16.5, 11.3, 5.5 Hz, 1H), 2.77 – 2.70 (m, 1H), 1.92 (dddd, *J* = 12.5, 5.5, 3.9, 2.9 Hz, 1H), 1.77 – 1.60 (m, 2H), 1.00 (d, *J* = 6.8 Hz, 3H), 0.98 (d, *J* = 6.8 Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3): δ 145.2, 129.3, 126.8, 121.6, 116.9, 114.1, 57.4, 32.7, 26.8, 24.7, 18.7, 18.4.

Chiral HPLC method: Chiralcel OD, hexane/IPA = 98/2, 1.0 mL/min, λ = 254 nm; (*S*)-enantiomer t_{R} (minor) = 5.90 min, (*R*)-enantiomer t_{R} (major) = 8.53 min.

(S)-2-methyl-6-nitro-1,2,3,4-tetrahydroquinoline (**11g**):



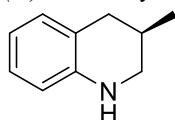
NMR and chiral HPLC data consistent with those previously reported;¹² compound was purified by flash column chromatography on silica gel (silica gel was deactivated with a 1% Et_3N in hexanes solution) using a 0-15% EtOAc/hexanes eluent gradient; isolated an orange solid in 83% yield (44.2 mg) and 86% ee. The compound was recrystallized from DCM/hexanes to afford the tetrahydroquinoline in 96% ee.

^1H NMR (500 MHz, CDCl_3): δ 7.95 – 7.86 (m, 2H), 6.42 – 6.32 (m, 1H), 4.53 (s, 1H), 3.55 (dq, J = 9.7, 6.4, 3.4 Hz, 1H), 2.90 – 2.74 (m, 2H), 2.00 (dtd, J = 12.9, 4.8, 3.4 Hz, 1H), 1.58 (dtd, J = 13.0, 9.8, 6.2 Hz, 1H), 1.28 (d, J = 6.4 Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3): δ 150.4, 137.5, 125.9, 124.4, 119.8, 112.2, 47.6, 29.0, 26.3, 22.4.

Chiral HPLC method: Chiralcel OD, hexane/IPA = 95/5, 1 mL/min, λ = 254 nm; (*R*)-enantiomer t_{R} (minor) = 19.35 min, (*S*)-enantiomer t_{R} (major) = 20.67 min (major).

(R)-3-methyl-1,2,3,4-tetrahydroquinoline (**11h**):



NMR and chiral HPLC data consistent with those previously reported.¹³

Compound was purified by flash column chromatography on silica gel using a 0-10% pentane/ Et_2O eluent; isolated as a pale-yellow oil in 71% yield (20.9 mg) and 14% ee.

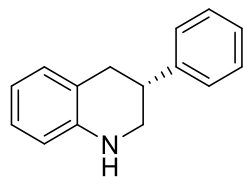
^1H NMR (400 MHz, CDCl_3): δ 7.01-6.91 (m, 2H), 6.61 (td, J = 7.4, 1.2 Hz, 1H), 6.49 (dd, J = 7.9, 1.2 Hz, 1H), 3.89 (s, 1H), 3.27 (ddd, J = 11.0, 3.7, 2.0 Hz, 1H), 2.90 (dd, J = 11.0, 9.6 Hz, 1H), 2.78 (ddd, J = 16.0, 5.0, 2.0 Hz, 1H), 2.43 (dd, J = 16.0, 10.2 Hz, 1H), 2.14 – 1.99 (m, 1H), 1.05 (d, J = 6.6 Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 144.4, 129.7, 126.8, 121.3, 117.1, 114.0, 49.0, 35.6, 27.3, 19.2.

Chiral HPLC method: Chiralcel OJ-H, hexane/IPA = 90/10, 0.5 mL/min, λ = 210 nm; (*R*)-enantiomer t_{R} = 30.77 min (major), (*S*)-enantiomer t_{R} = 37.77 min (minor).

(R)-3-phenyl-1,2,3,4-tetrahydroquinoline (**11i**):

The precursor 3-phenylquinoline was synthesized according to literature procedure.¹⁴



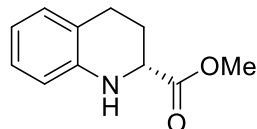
NMR and chiral HPLC data consistent with those previously reported.¹⁵ Compound was purified by flash column chromatography on silica gel using 0-6% EtOAc/hexanes as the eluent; isolated as a pale-yellow solid in 51% yield (10.5 mg; yield based on recovered starting material) and 4% ee.

¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.30 (m, 2H), 7.30 – 7.22 (m, 3H), 7.02 (d, *J* = 7.4 Hz, 2H), 6.66 (td, *J* = 7.4, 1.2 Hz, 1H), 6.57 (dd, *J* = 8.4, 1.3 Hz, 1H), 4.15 (s, 1H), 3.47 (ddd, *J* = 11.2, 3.7, 1.9 Hz, 1H), 3.35 (t, *J* = 10.7 Hz, 1H), 3.16 (tdd, *J* = 10.2, 5.8, 3.7 Hz, 1H), 3.09 – 2.93 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 144.1, 144.0, 129.7, 128.8, 127.4, 127.1, 126.8, 121.6, 117.3, 114.3, 48.5, 38.8, 34.8.

Chiral HPLC method: Chiralcel OD, hexane/IPA = 98/2, 1 mL/min, λ = 254 nm; (*R*)-enantiomer *t*_R = 18.81 min (major), (*S*)- 10.5 mg *t*_R = 24.04 min (minor).

Methyl (R)-1,2,3,4-tetrahydroquinoline-2-carboxylate (**11j**)



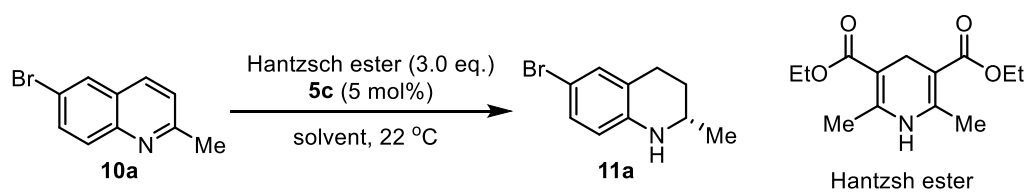
Reaction time: 4 h at 50 °C using catalyst **5c**. NMR and chiral HPLC data consistent with those previously reported.¹⁶

purified using a 0-8% Et₂O/hexanes eluent; isolated as a colourless oil in 71% yield (27.2 mg) and 30% ee.

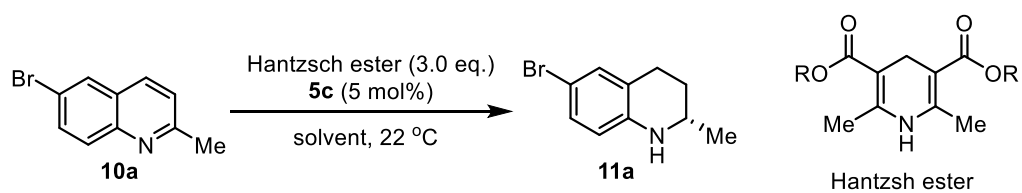
¹H NMR (400 MHz, CDCl₃) δ 7.05 – 6.96 (m, 1H), 6.96 (d, *J* = 7.5 Hz, 1H), 6.65 (td, *J* = 7.3, 1.2 Hz, 1H), 6.59 (dd, *J* = 8.0, 1.2 Hz, 1H), 4.40 (s, 1H), 4.05 (dd, *J* = 8.8, 3.8 Hz, 1H), 3.78 (s, 3H), 2.90 – 2.71 (m, 2H), 2.29 (dtd, *J* = 13.0, 5.6, 3.8 Hz, 1H), 2.01 (dtd, *J* = 13.0, 9.1, 5.3 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 173.8, 143.1, 129.3, 127.2, 120.7, 117.8, 114.7, 54.1, 52.5, 26.0, 24.8.

Chiral HPLC method: Chiralpak AD, hexane/IPA = 80/20, 1 mL/min, λ = 254 nm; (*R*)-*t*_R = 7.62 min (major), (*S*)-*t*_R = 9.04 min (minor).

Table 1: Solvent Screening and Optimization of Reaction Conditions

Entry	Solvent	Time (h)	Yield (%)	ee (%)
1	toluene	2	99	80
2	CH ₂ Cl ₂	1.5	99	75
3	CHCl ₃	0.5	99	80
4	CCl ₄	1.5	99	86
5	DCE	1.5	99	78
6	cyclohexane	5	99	89
7	<i>n</i> -hexane	5	99	86
8	Et ₂ O	3	99	85
9	<i>t</i> -BuOMe	3	99	85
10	EtOAc	2	99	84

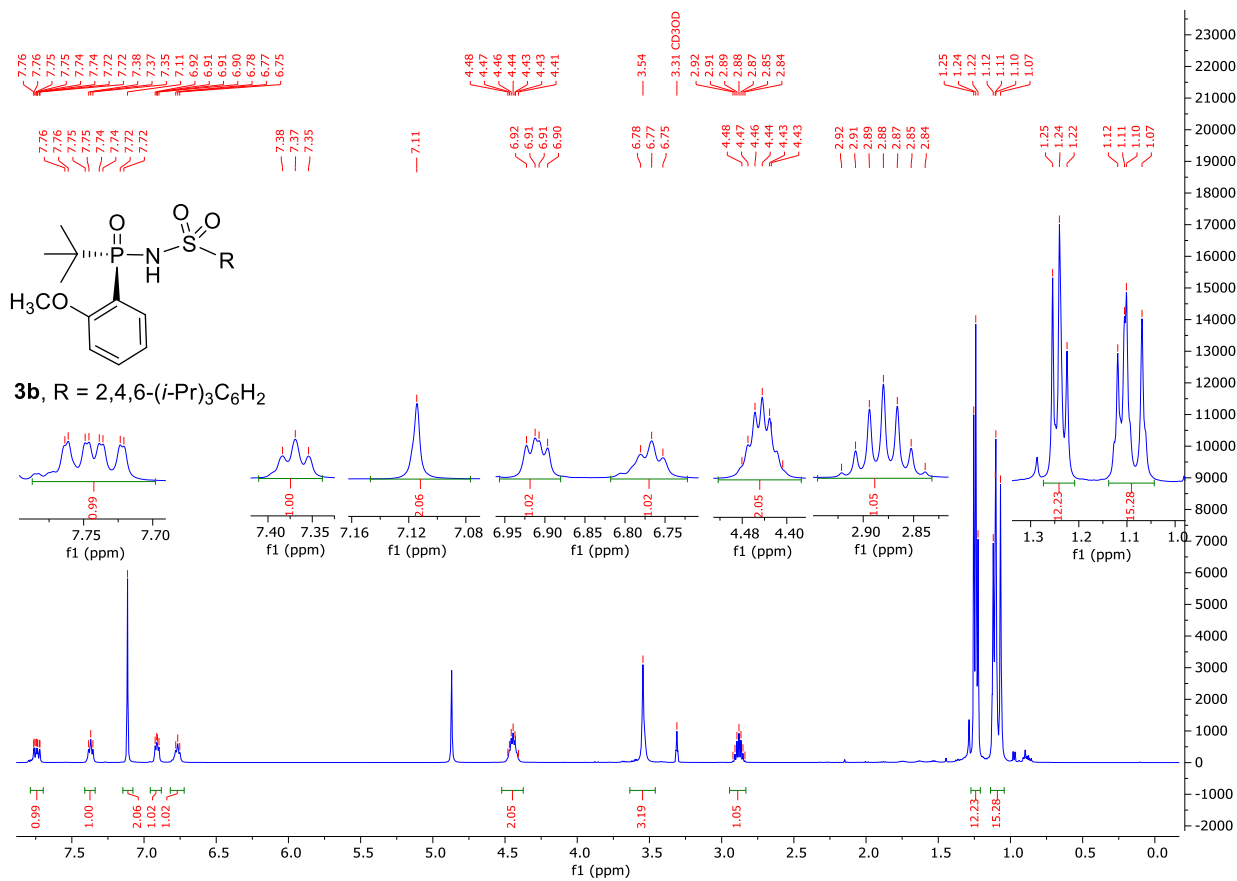
Table 2: Optimization of Hantzsh Ester

Entry	R	Time (h)	Yield (%)	ee (%)
1	Me	5	99	88
2	Et	5	99	89
3	<i>t</i> -Bu	3	99	85

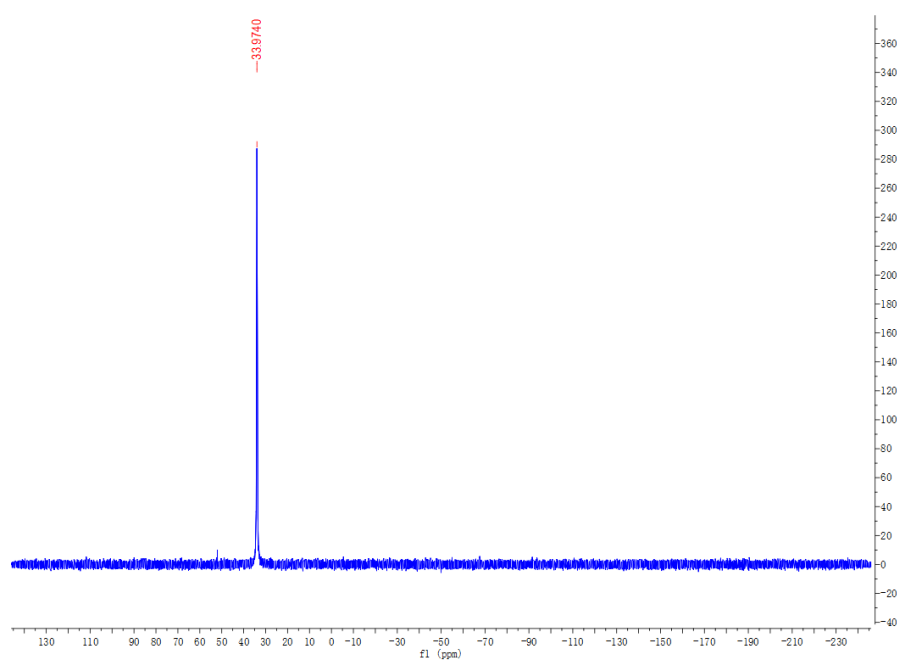
^1H , ^{13}C , ^{31}P NMR spectra and chiral HPLC chromatograms

Compounds Listed in Numerical Order

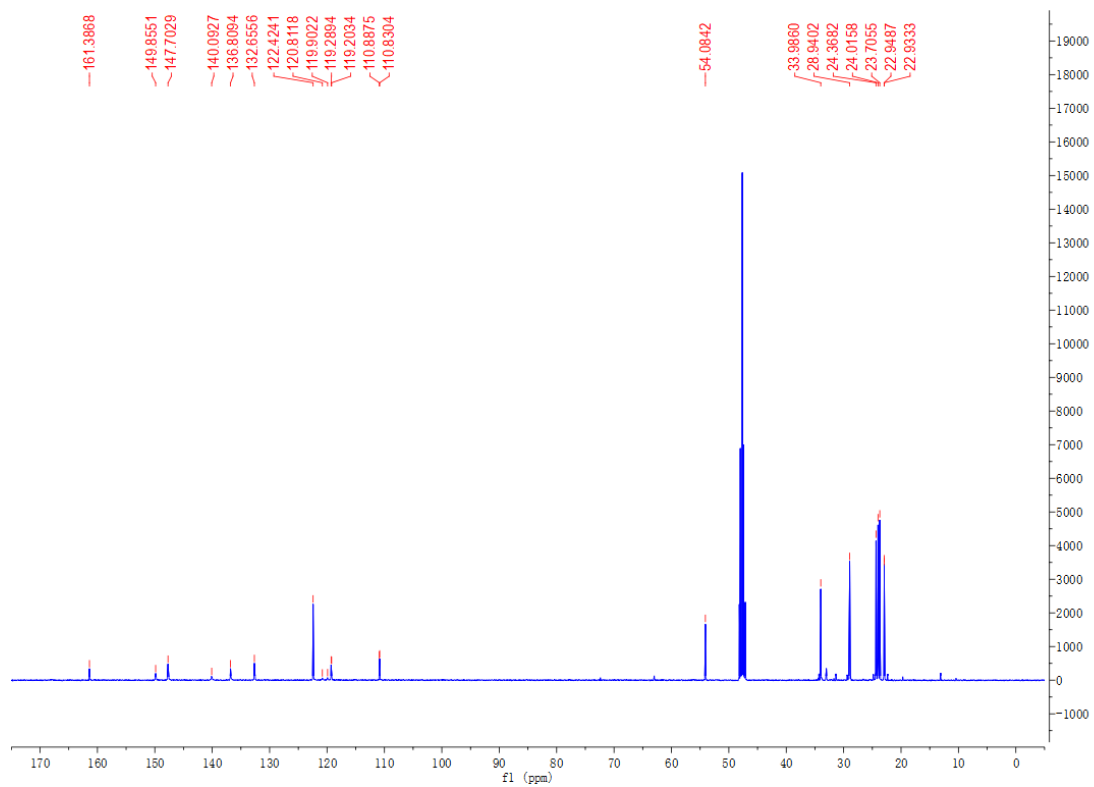
^1H NMR of **3b**



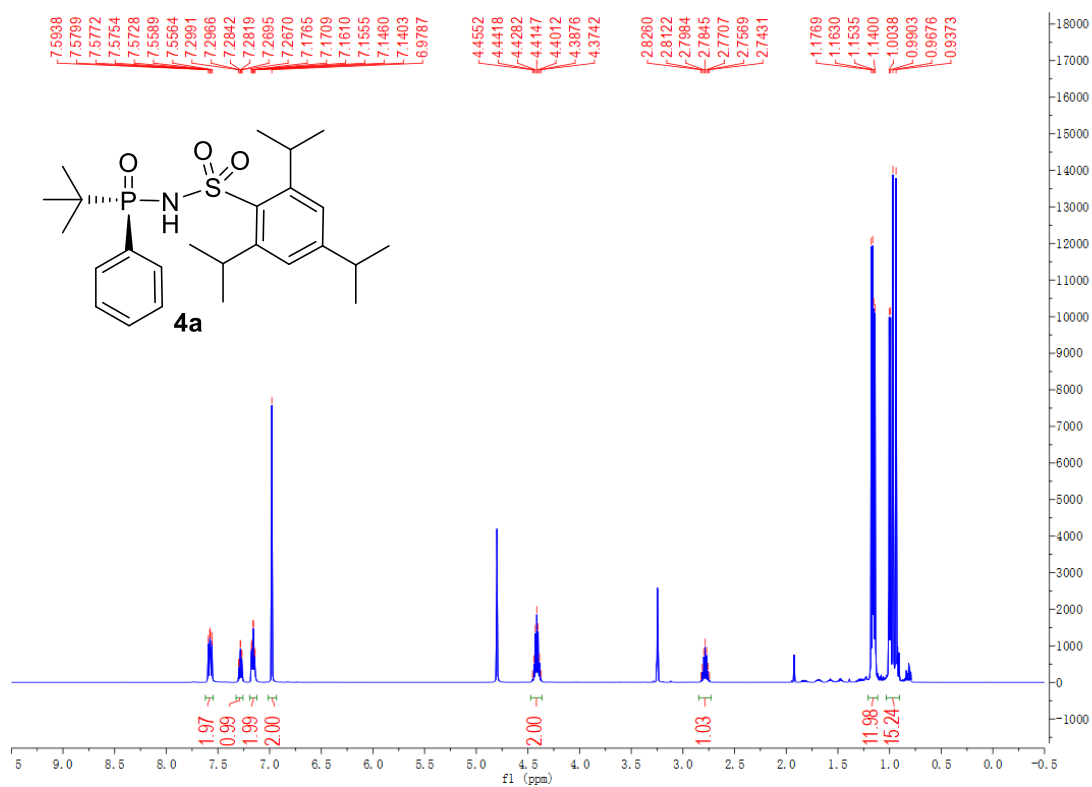
^{31}P NMR of **3b**



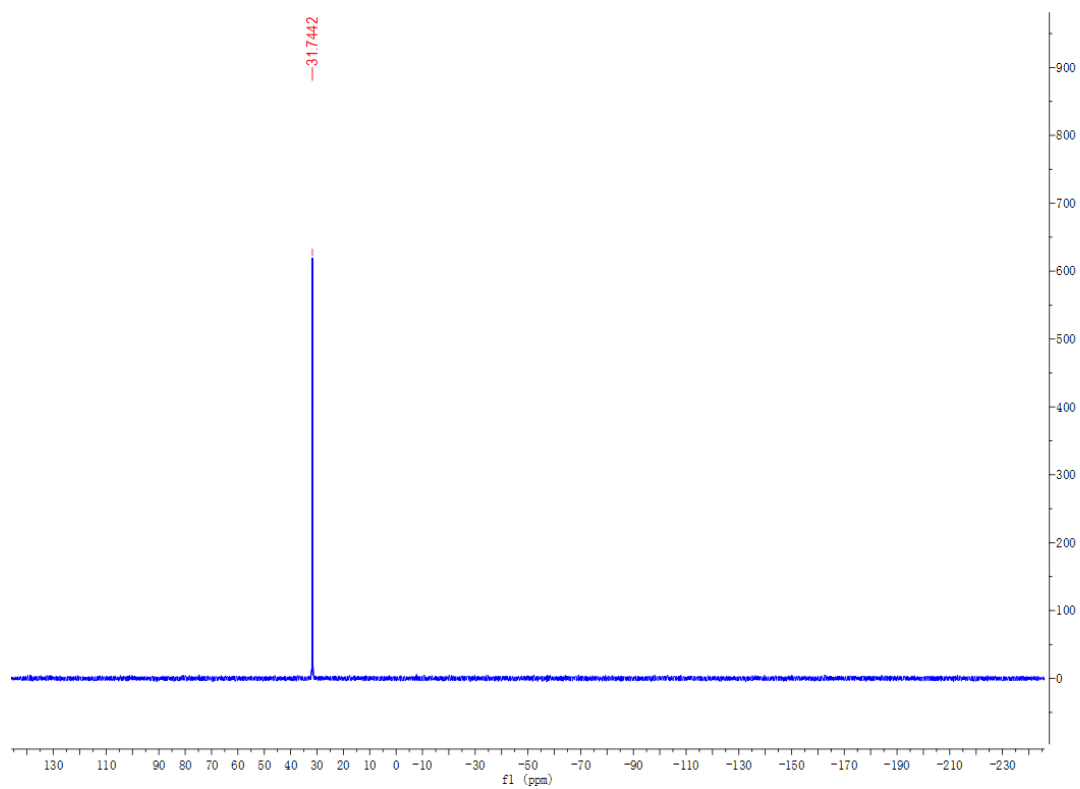
¹³C NMR of **3b**



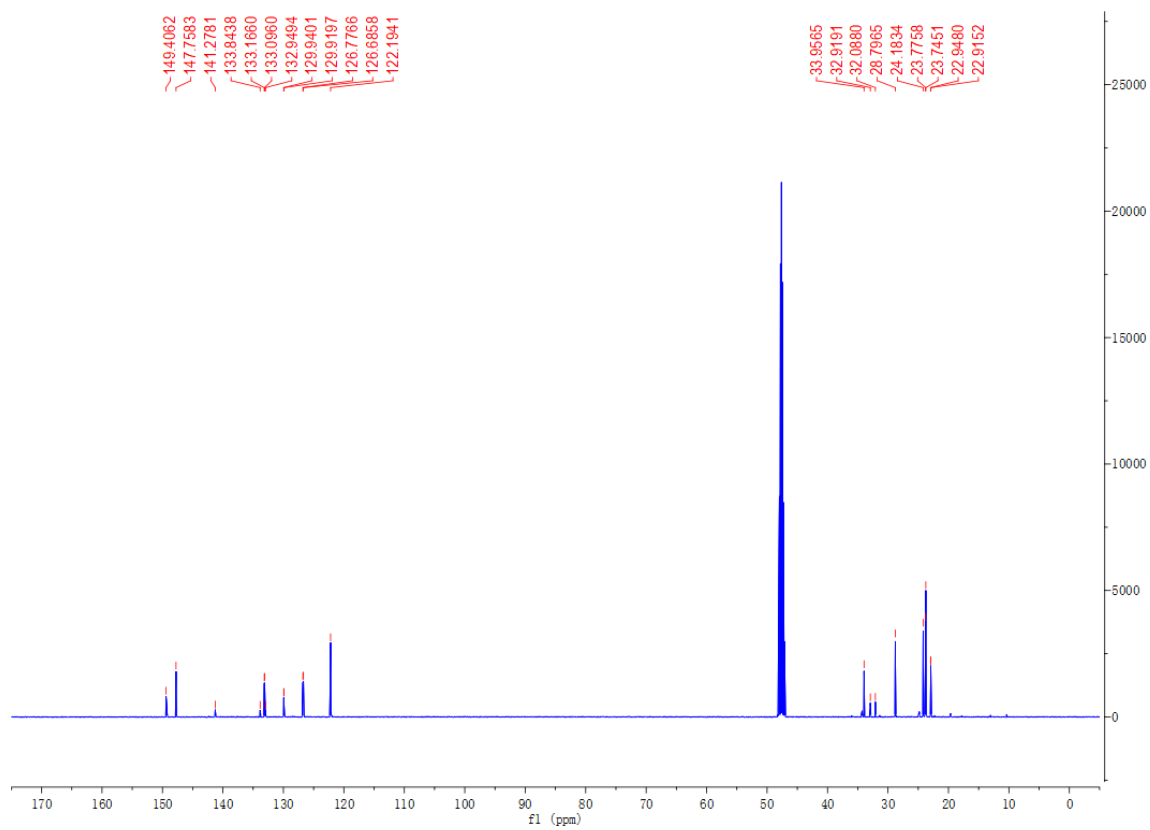
¹H NMR of 4a



³¹P NMR of 4a



¹³C NMR of 4a



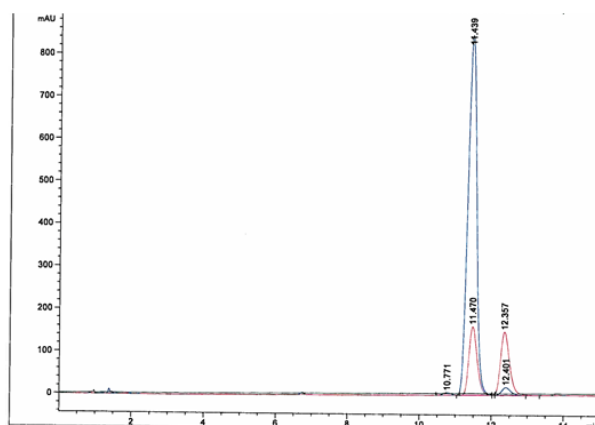
Chiral HPLC Chromatograms of **4a**

Method: HPLC instrument: Agilent 1260 HPLC; $\lambda = 220$ nm

Column: Phenomenex Lux Cellulose-2, 4.6x100 mm

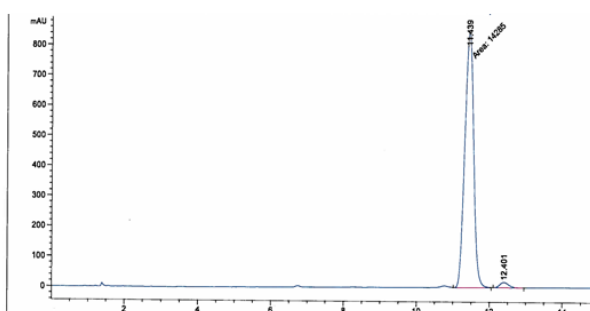
Solvent A: 0.1% (v/v) HClO₄ in water, Solvent B: CH₃CN; 50-90% solvent B in 15 min
at a flow rate of 1.2 mL/min

Top panel, racemic (in red) superimposed with (*R*)-enantiomer (in blue); bottom panel
(*R*)-**4a**; (*R*)-enantiomer $t_R = 11.44$ min (major), (*S*)- enantiomer $t_R = 12.36$ min (minor).

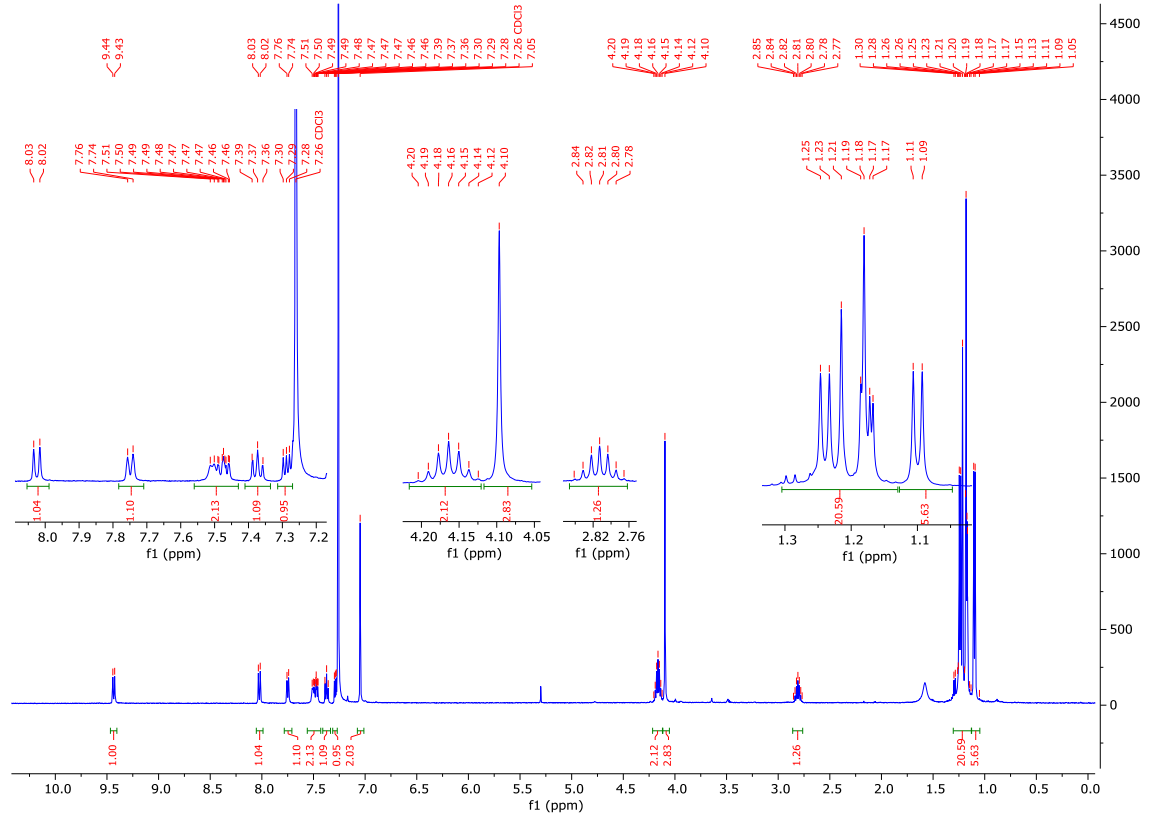
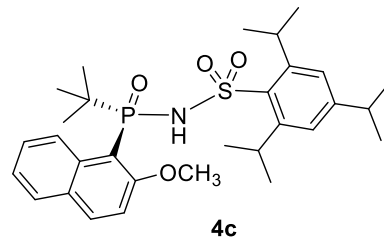


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.439	MM	0.2819	1.42850e4	844.44775	98.0089
2	12.401	BB	0.2276	290.20435	17.39890	1.9911
Totals :				1.45752e4	861.84665	

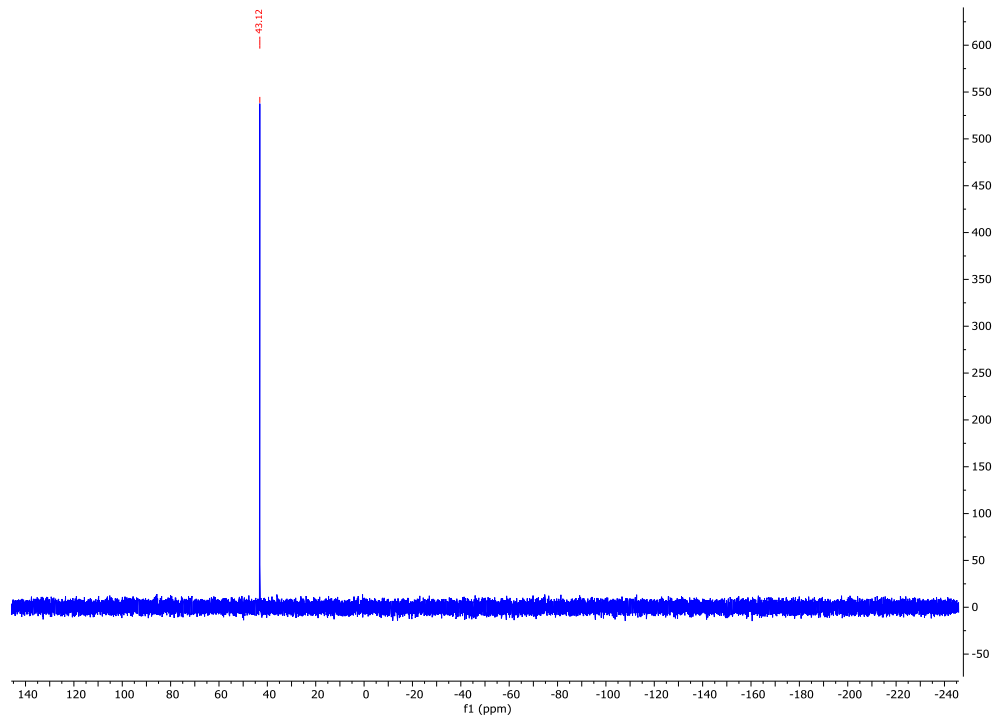
*** End of Report ***



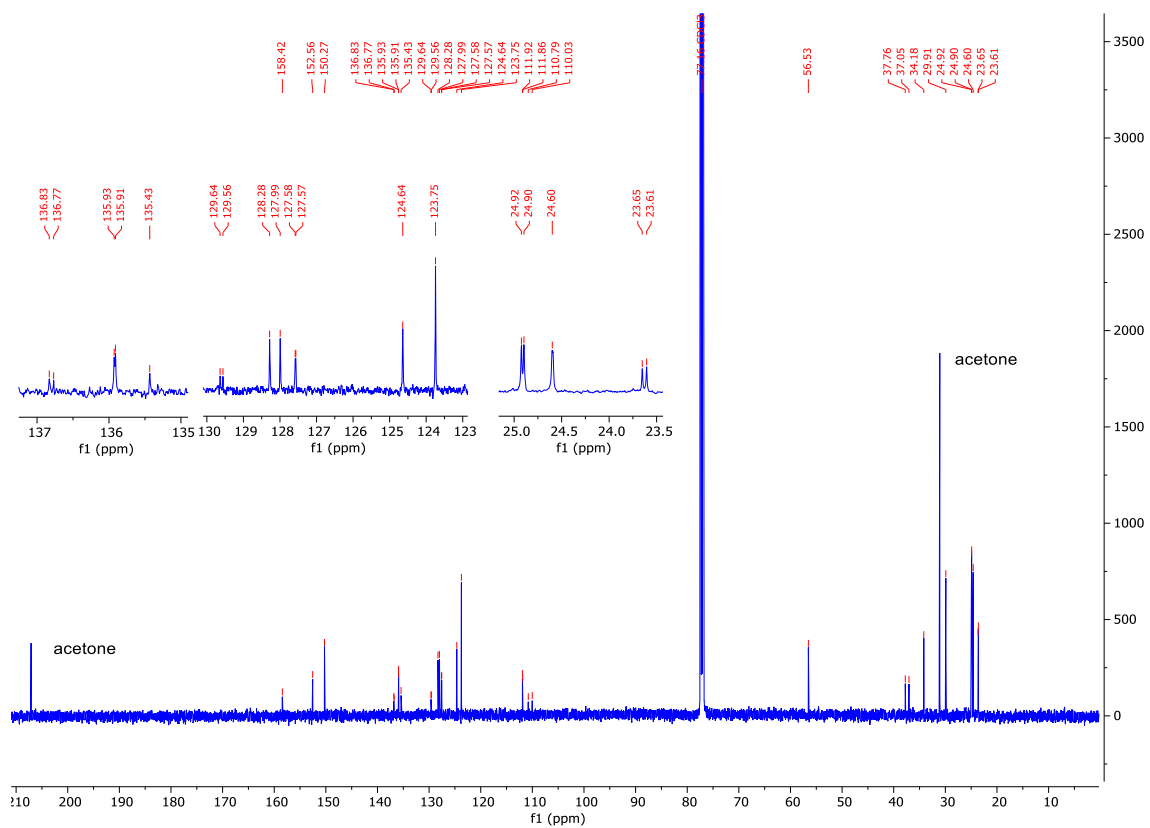
¹H NMR of 4c



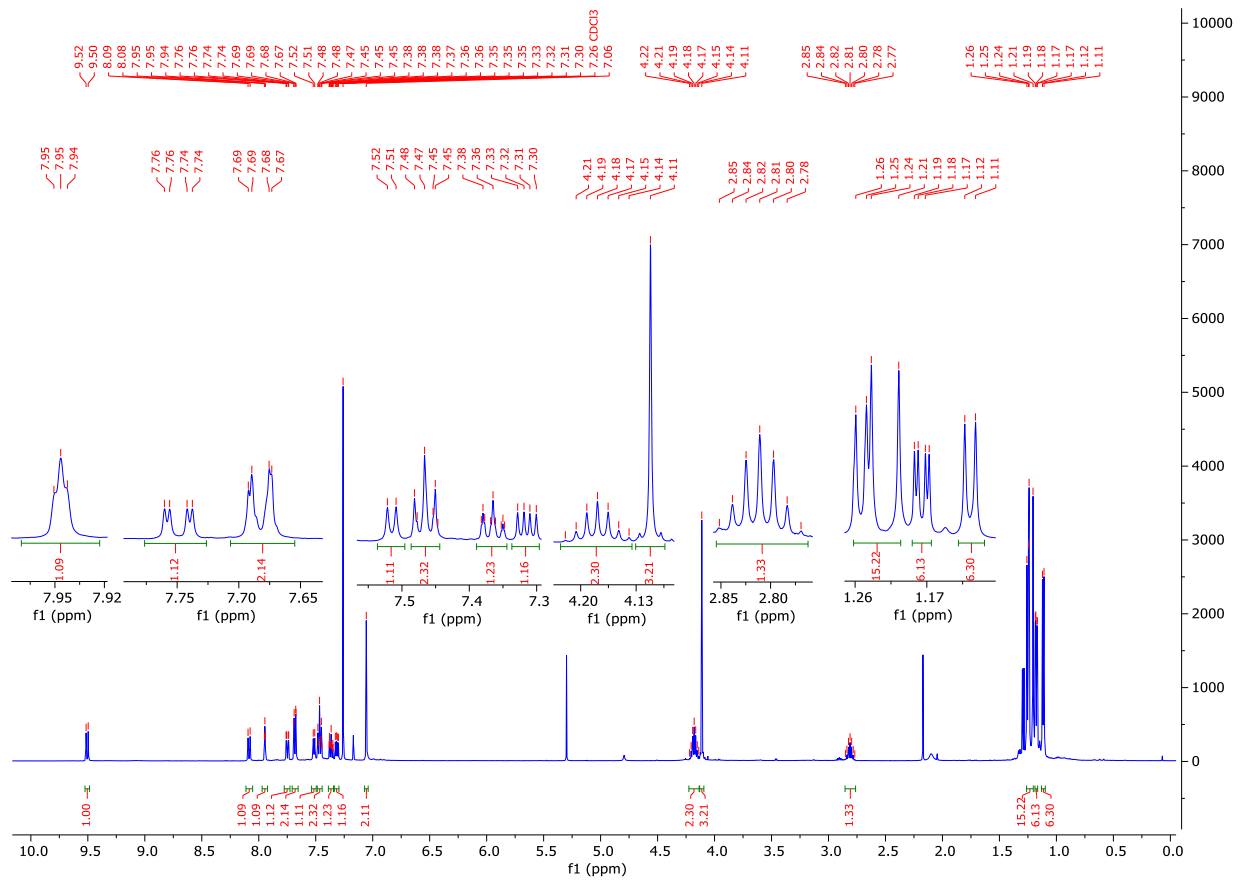
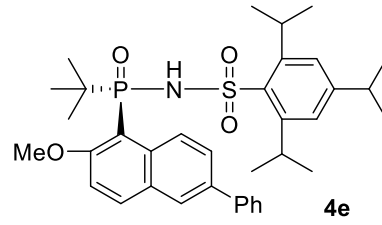
³¹P NMR of 4c



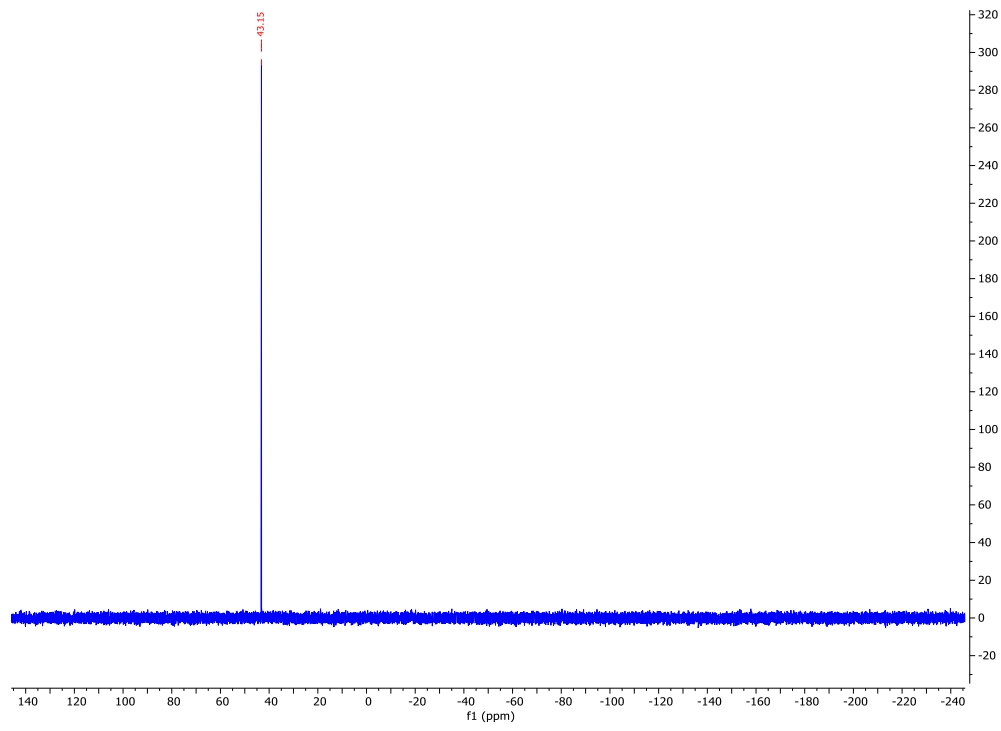
^{13}C NMR of 4c



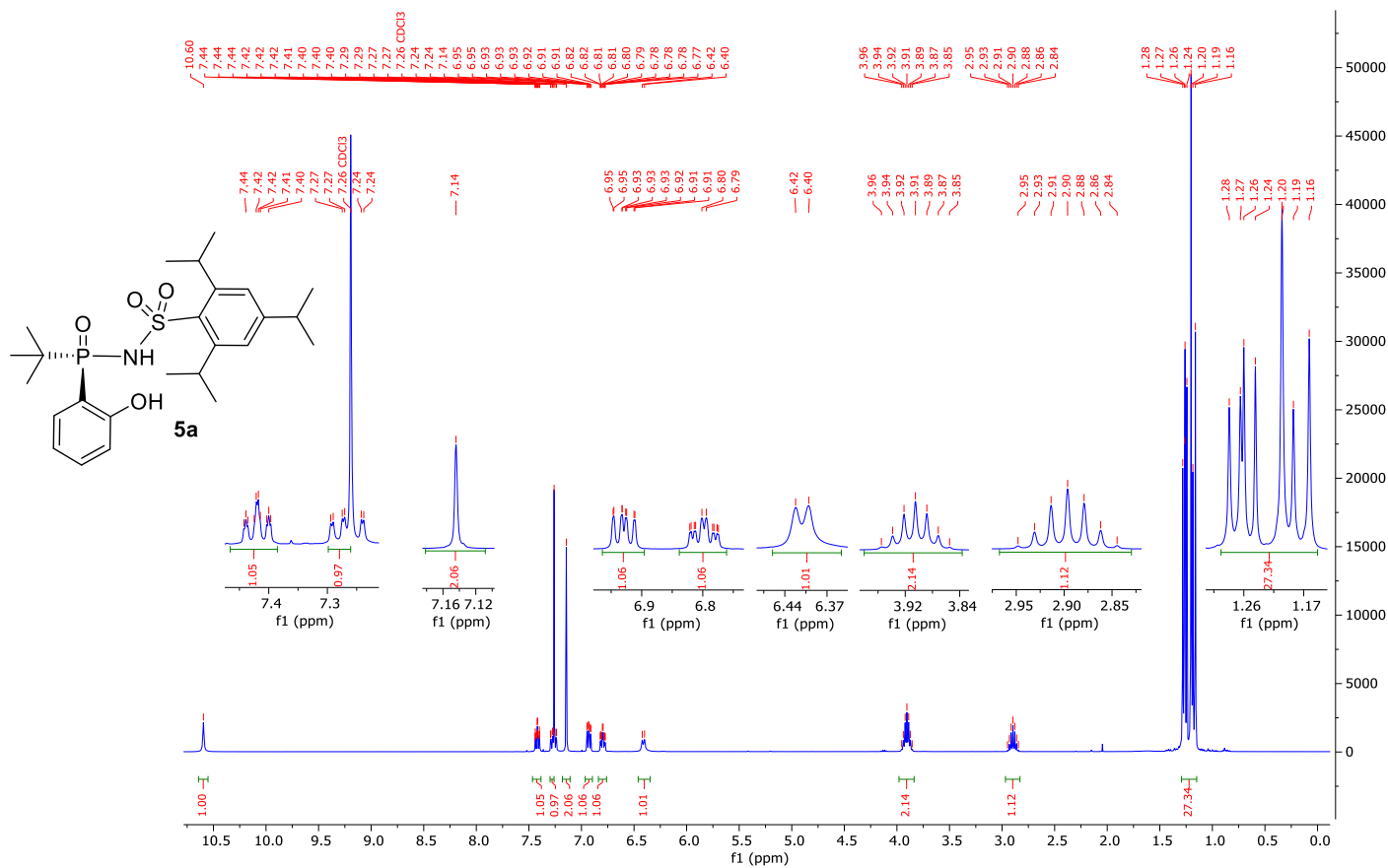
¹H NMR of 4e



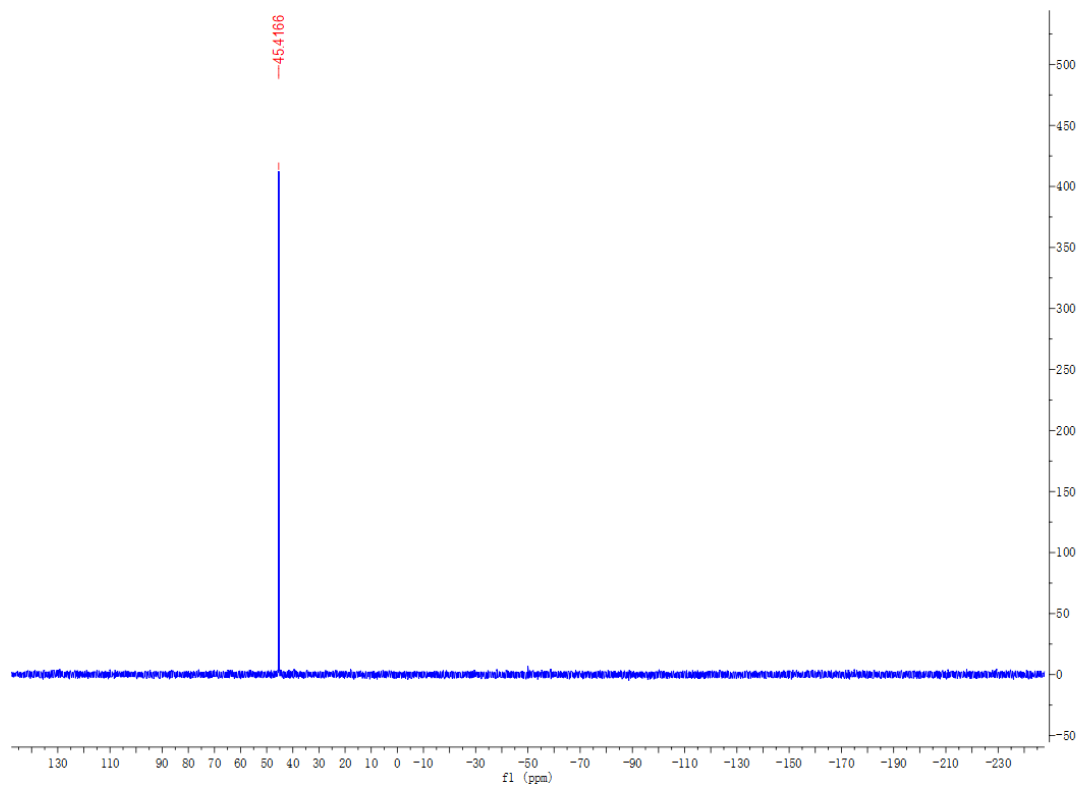
³¹P NMR of 4e



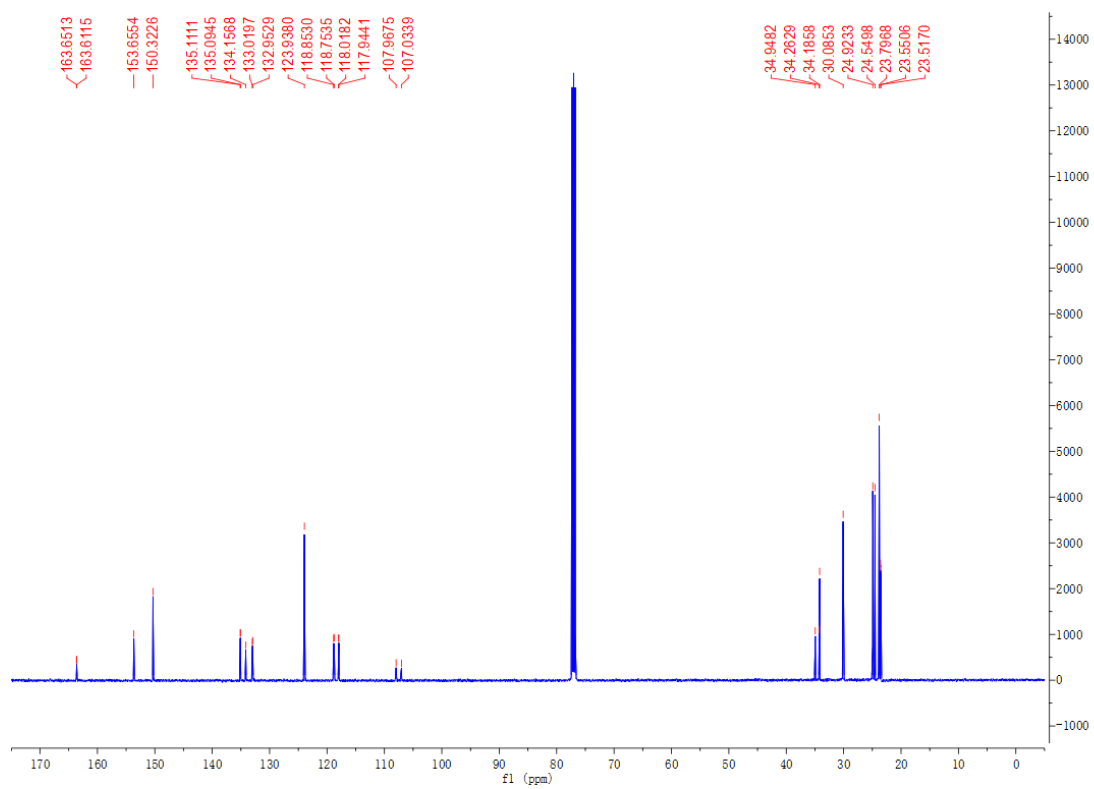
¹H NMR of **5a**



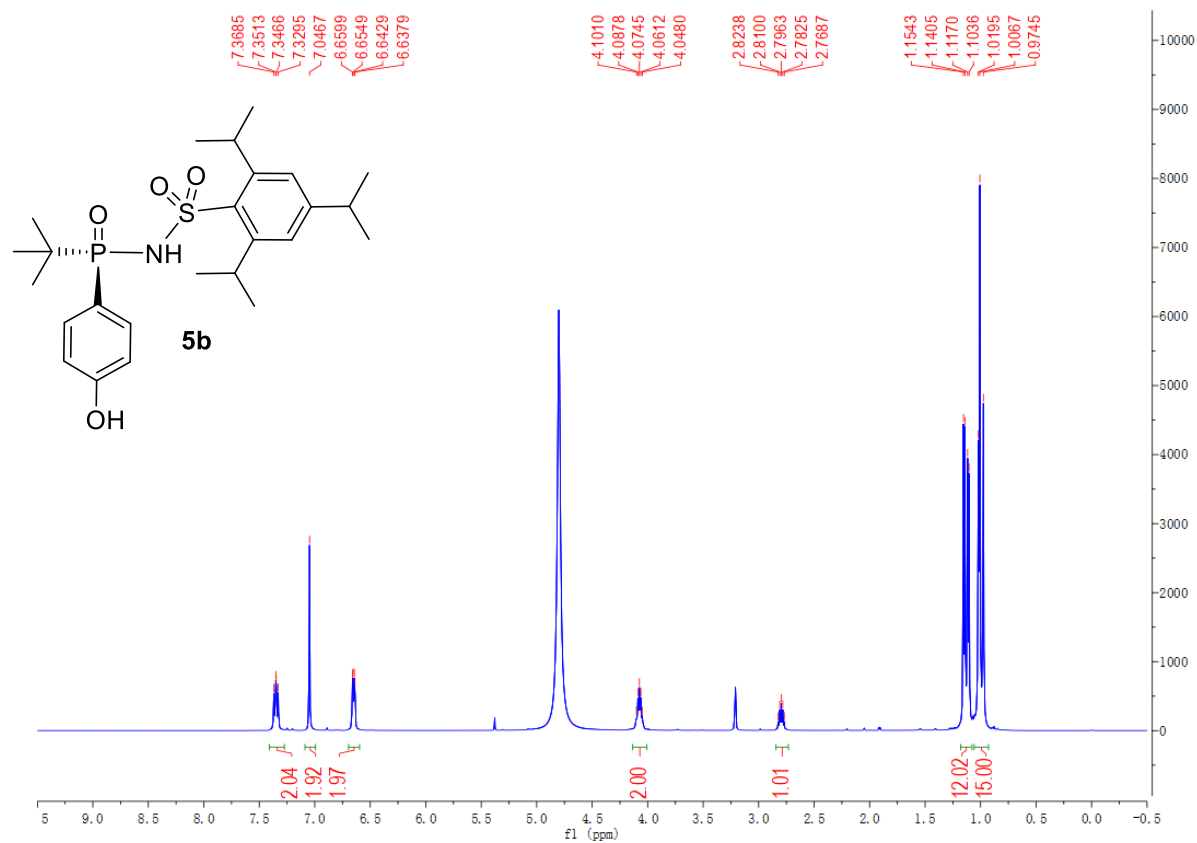
³¹P NMR of **5a**



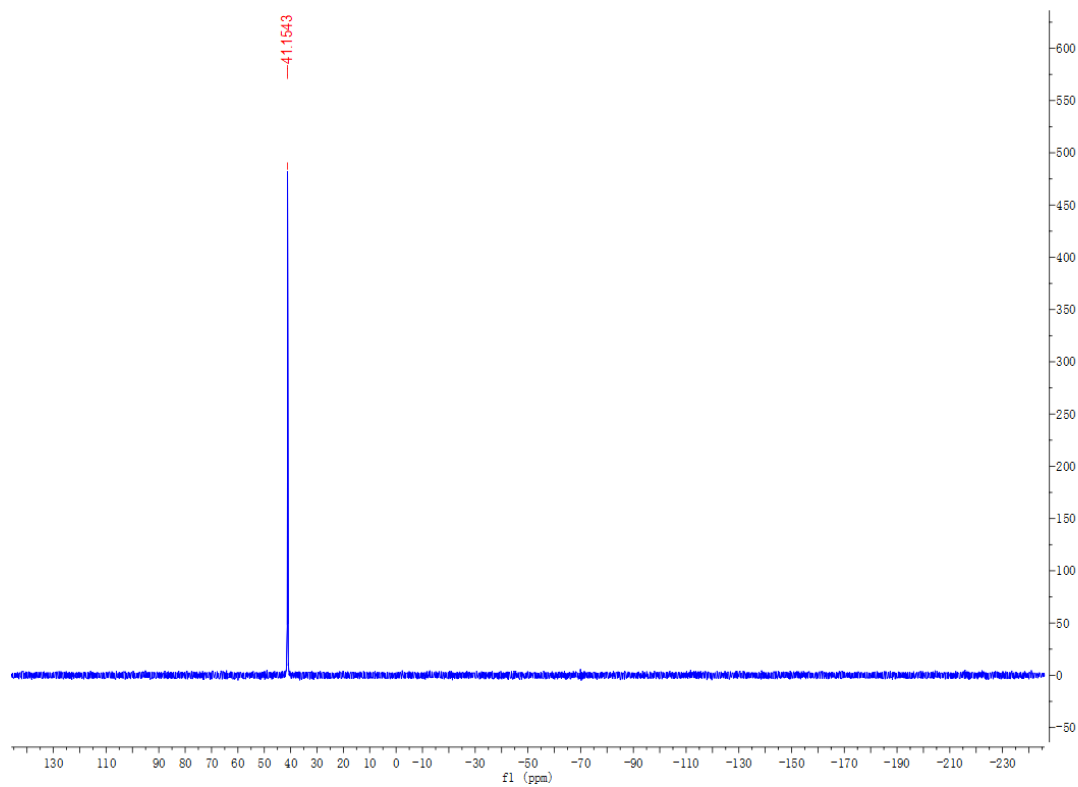
¹³C NMR of 5a



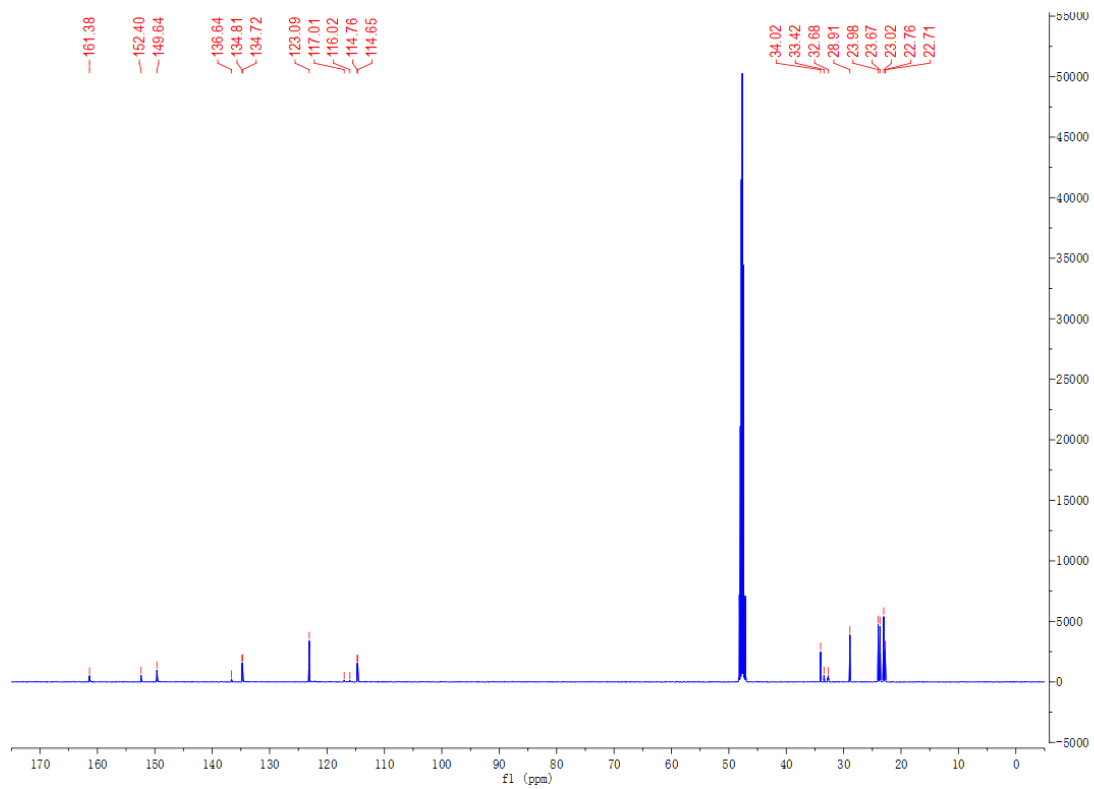
^1H NMR of **5b**



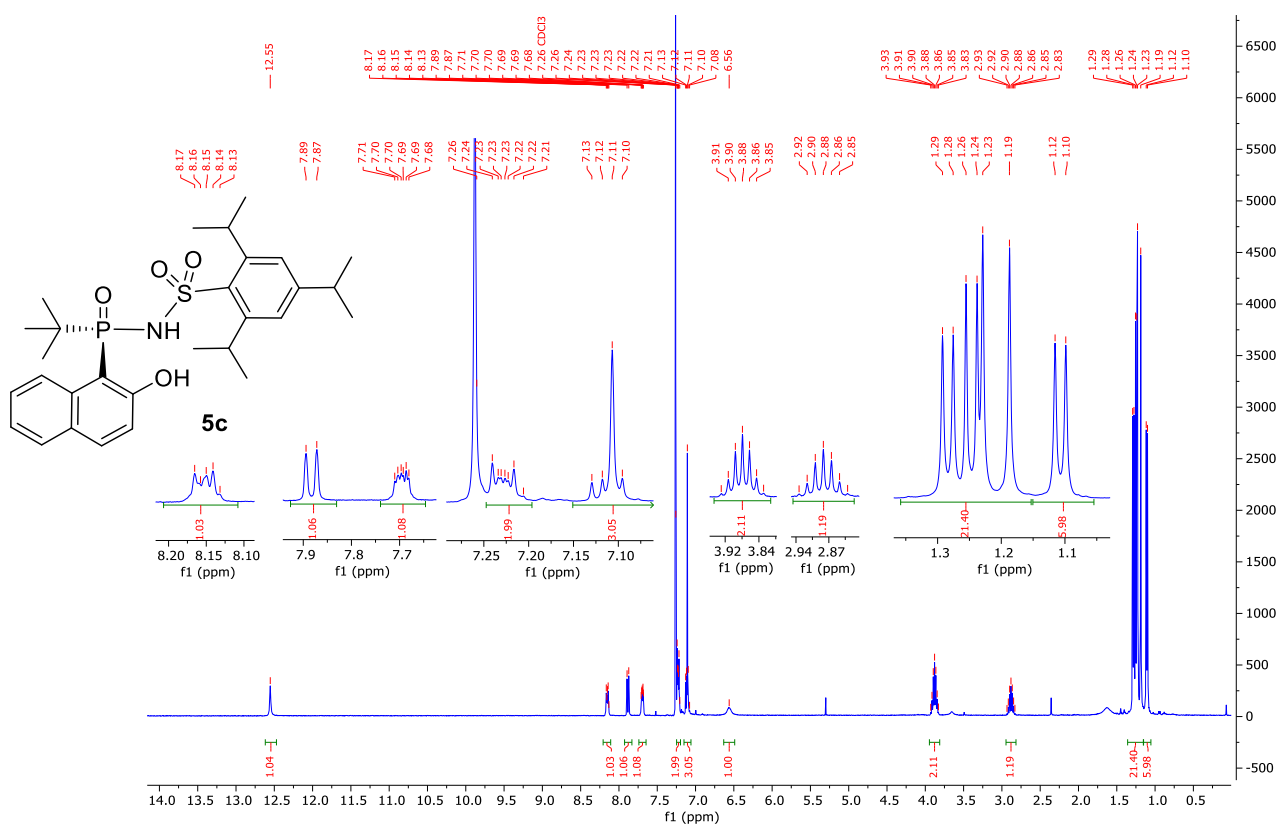
^{31}P NMR of **5b**



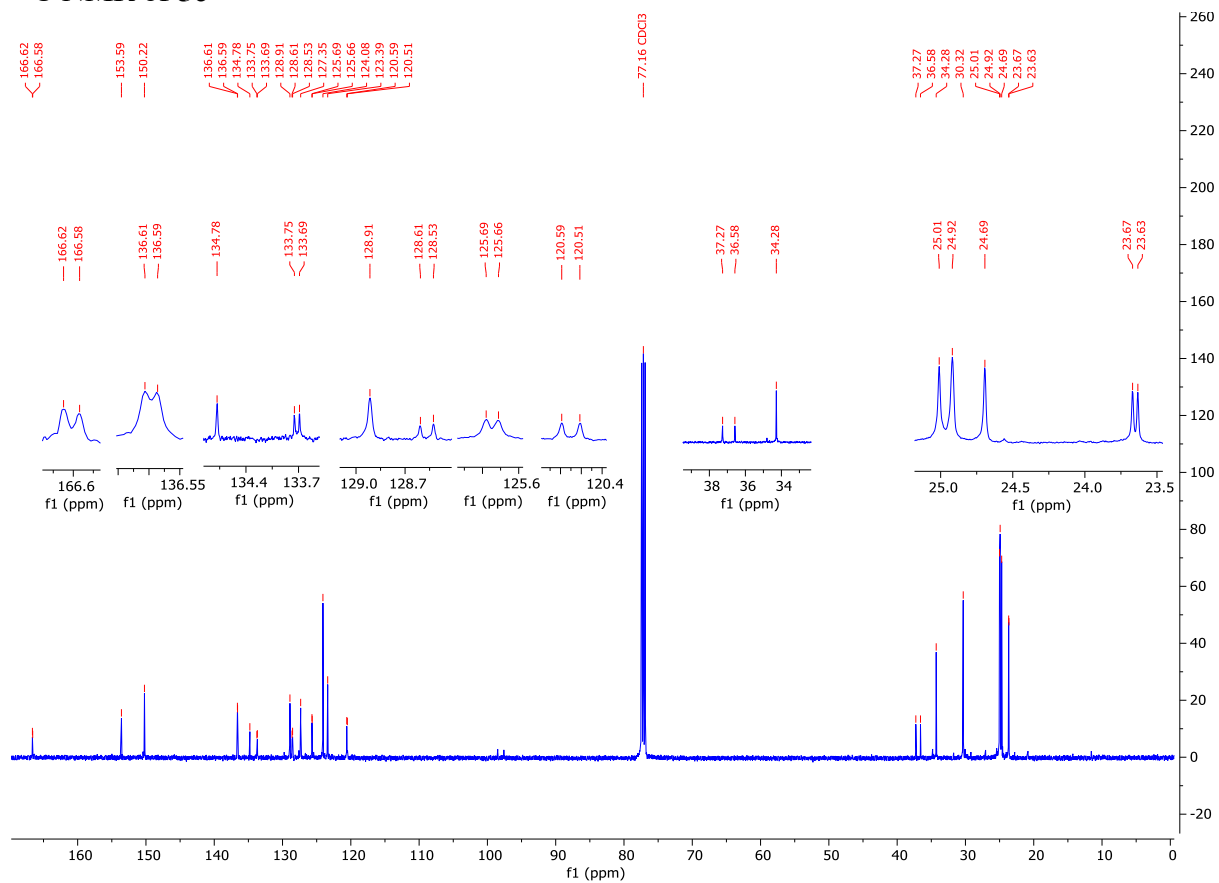
^{13}C NMR of **5b**



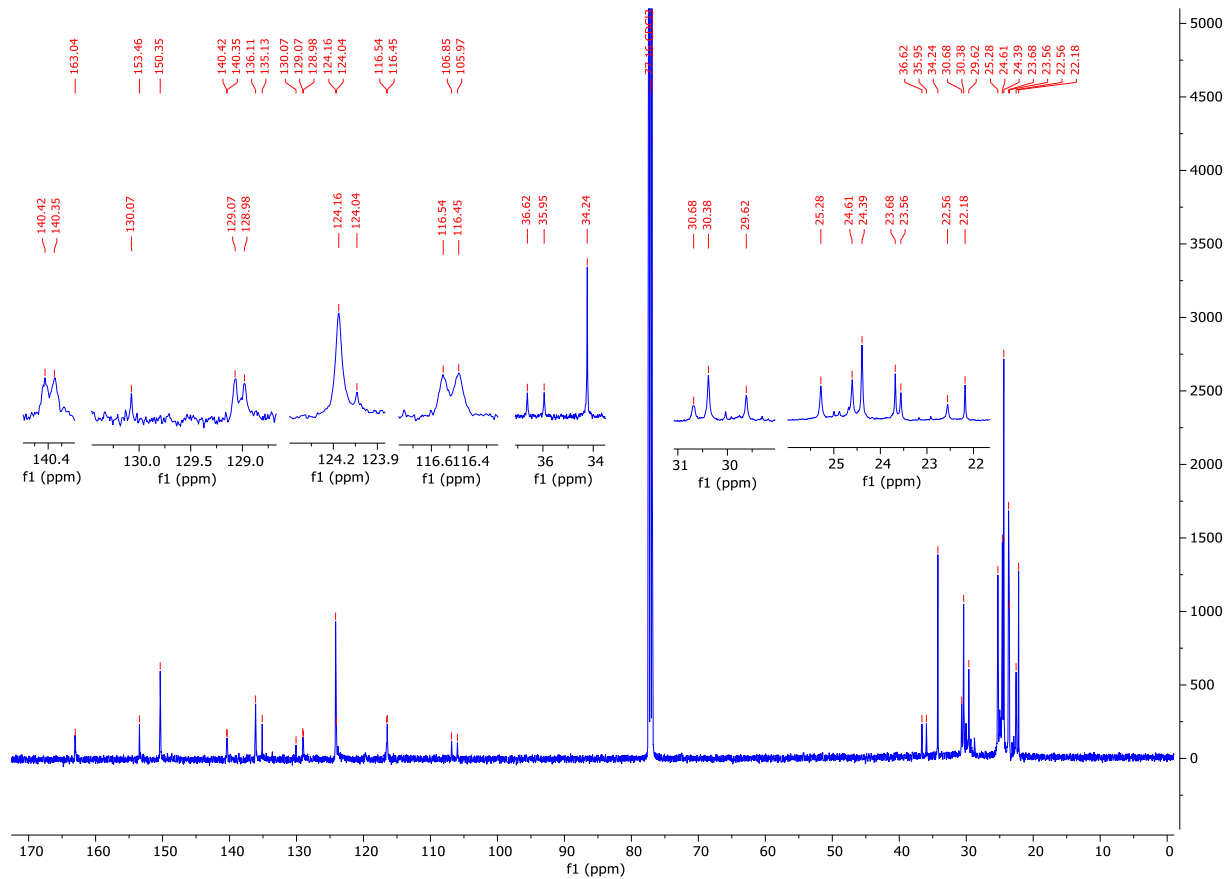
^1H NMR of **5c**



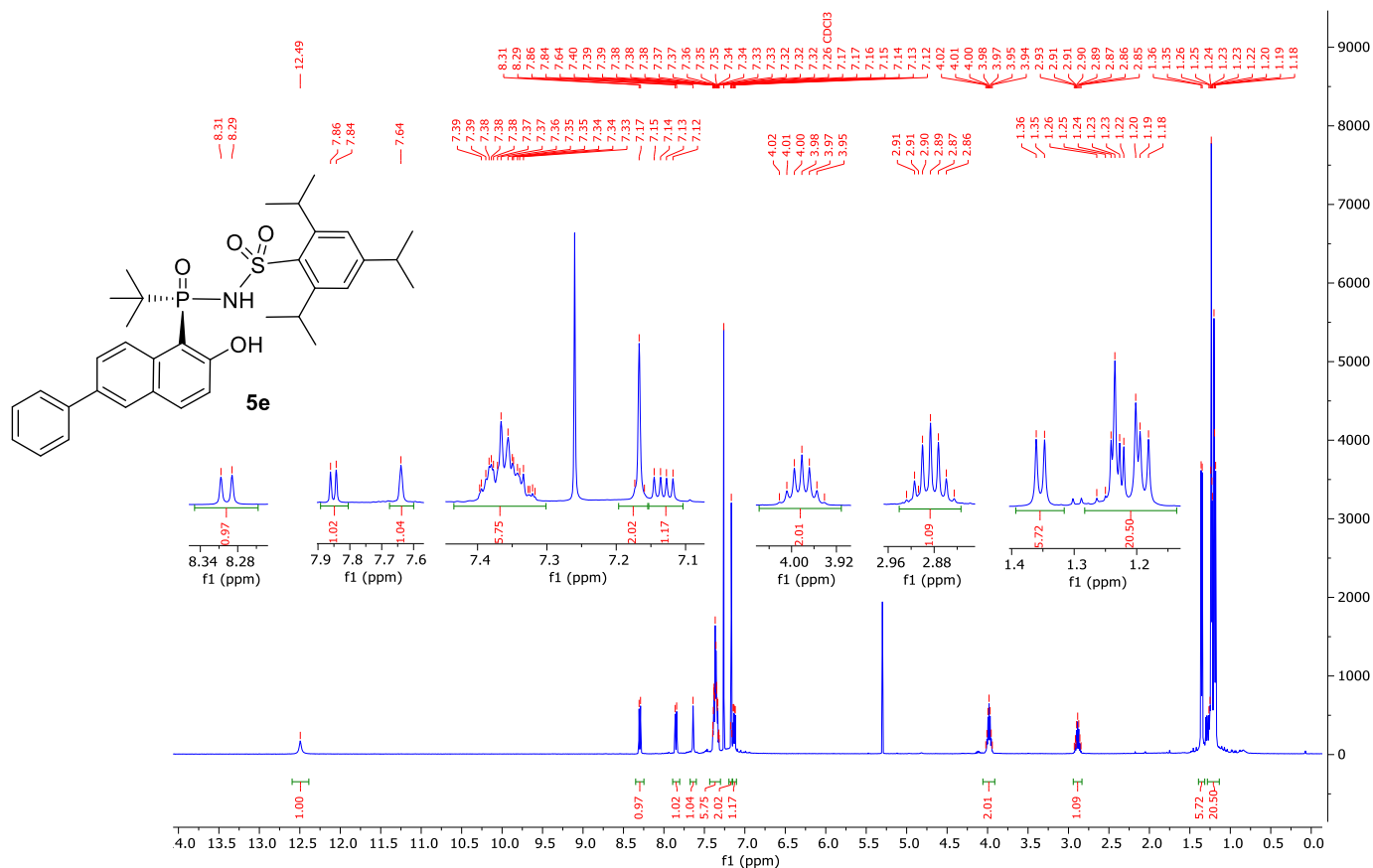
^{13}C NMR of 5c



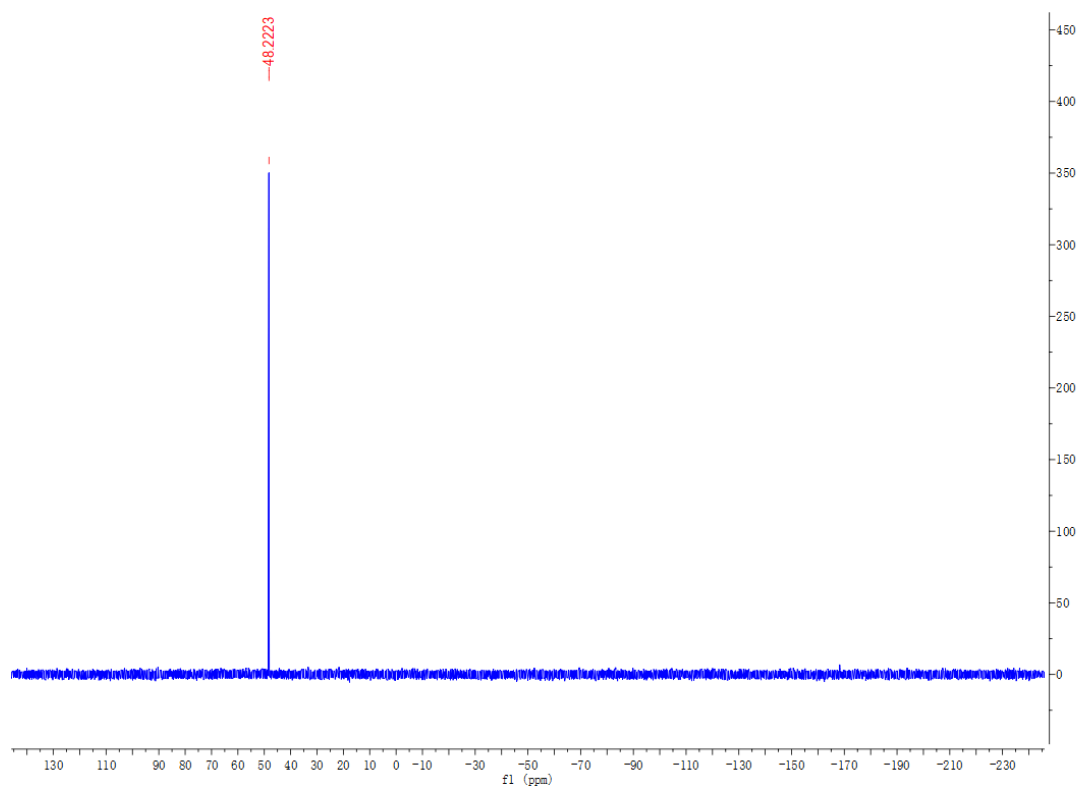
¹³C NMR of 5d



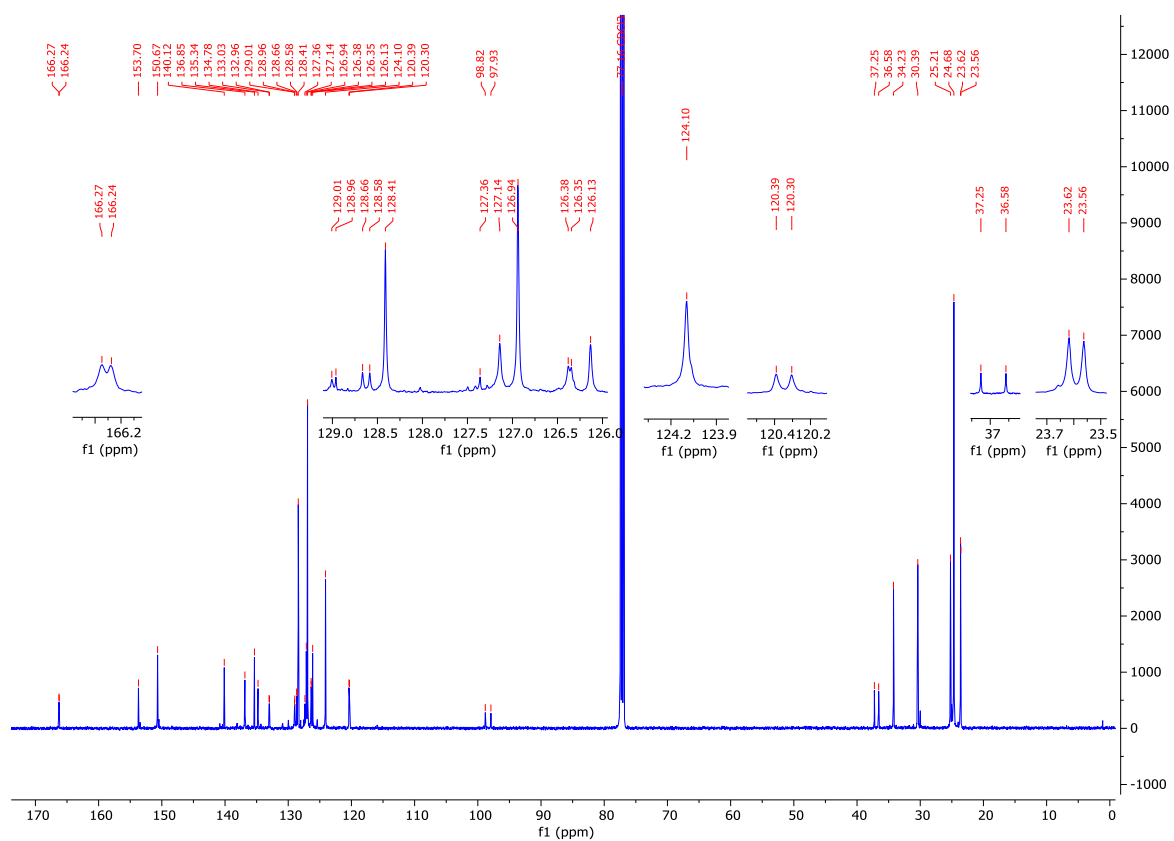
¹H NMR of 5e



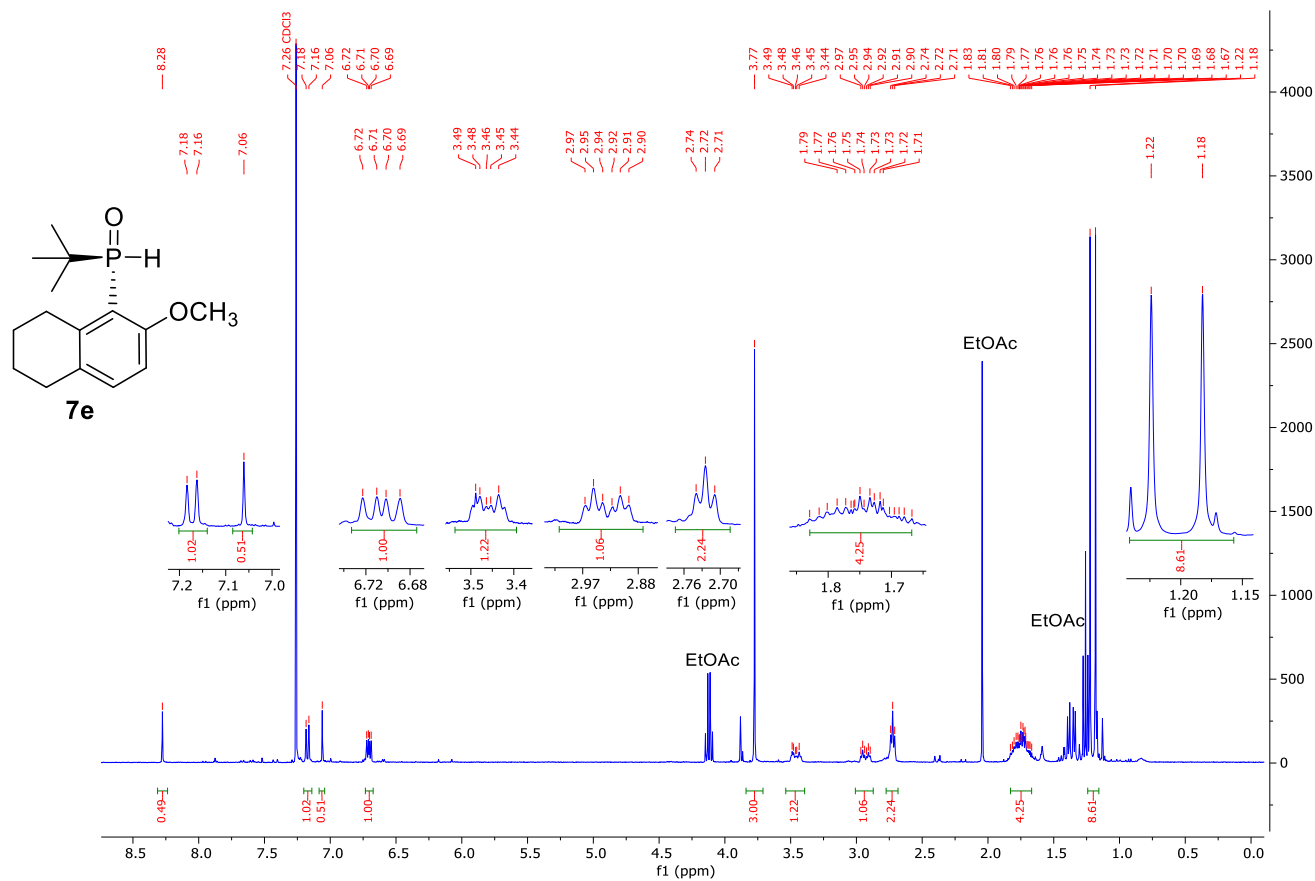
³¹P NMR of 5e



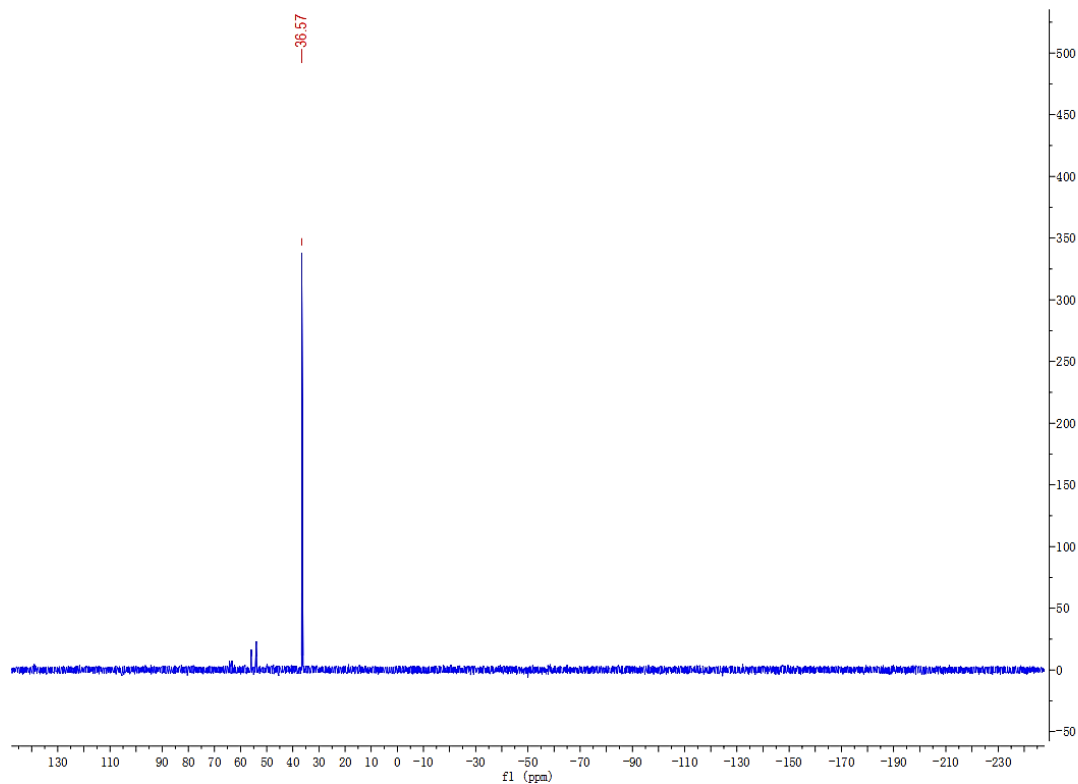
¹³C NMR of 5e



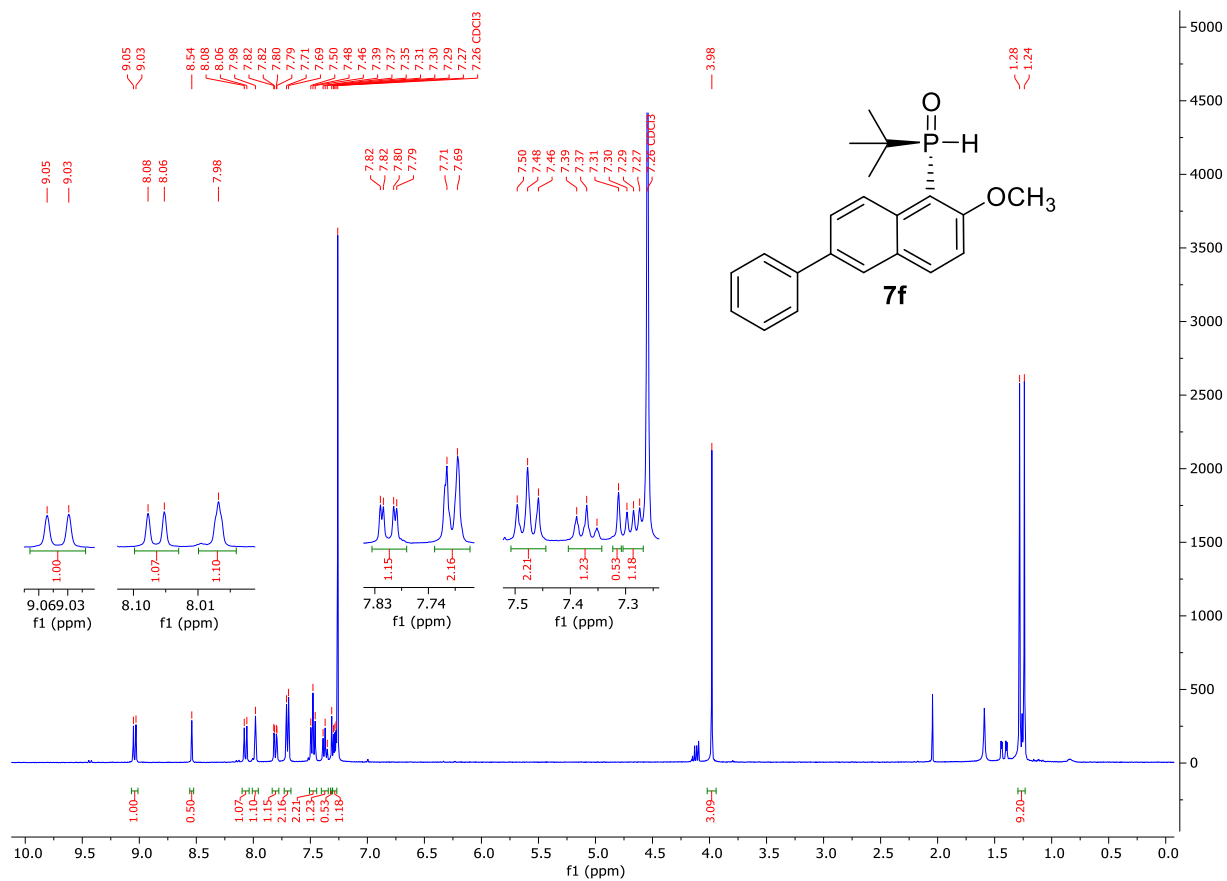
¹H-NMR of 7e



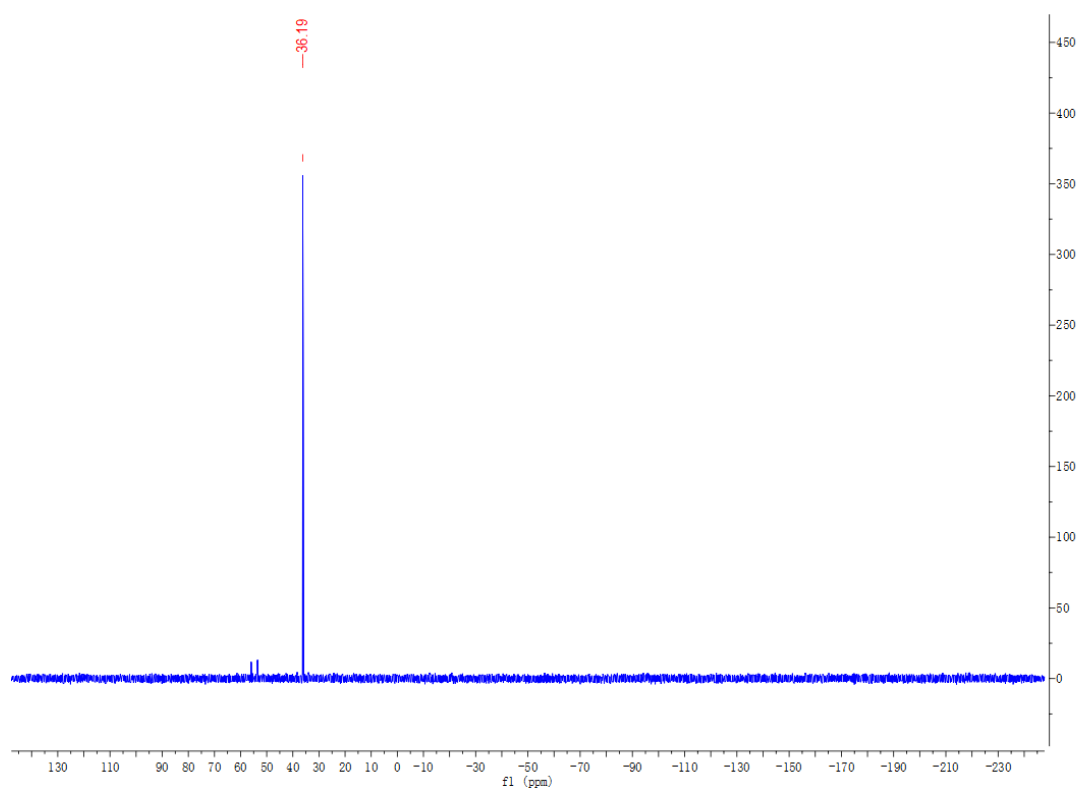
³¹P-NMR of 7e



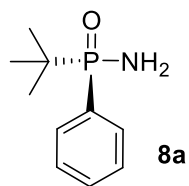
$^1\text{H-NMR}$ of **7f**



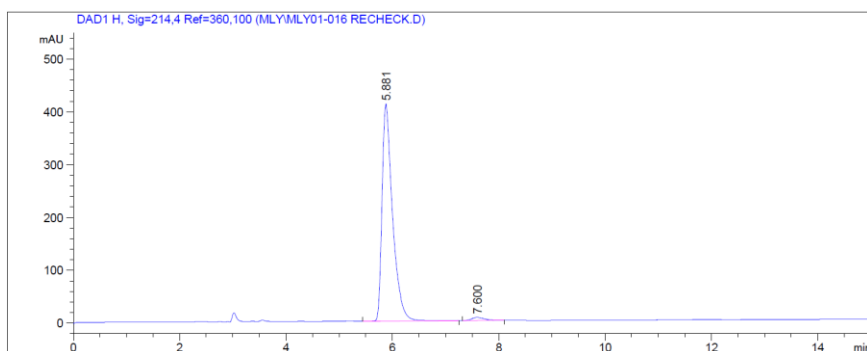
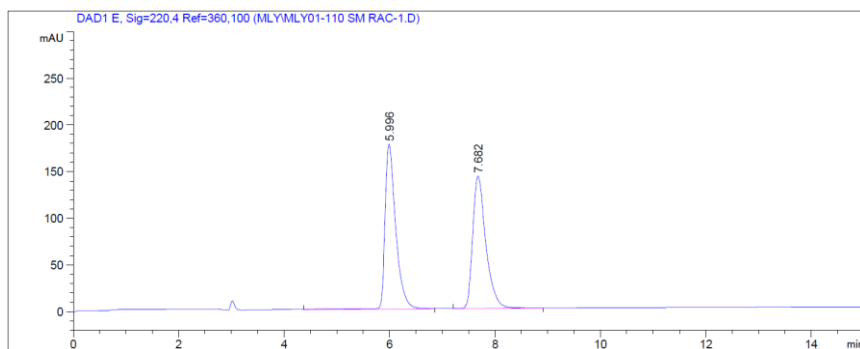
$^{31}\text{P-NMR}$ of **7f**



$^1\text{H-NMR}$, ^{13}C and ^{31}P NMR spectra of compound **8a** were consistent with those previously reported.⁷

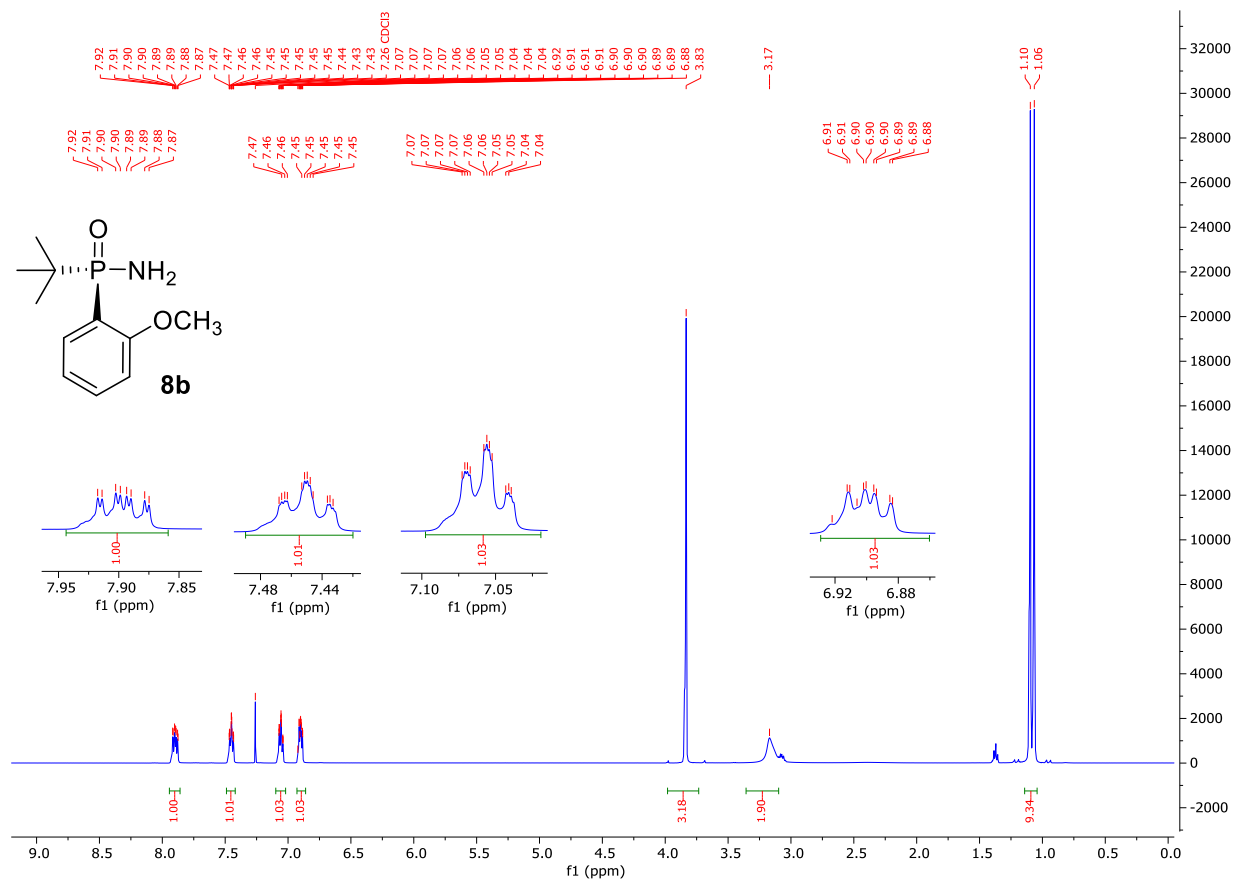


Chiral HPLC chromatograms of **8a**

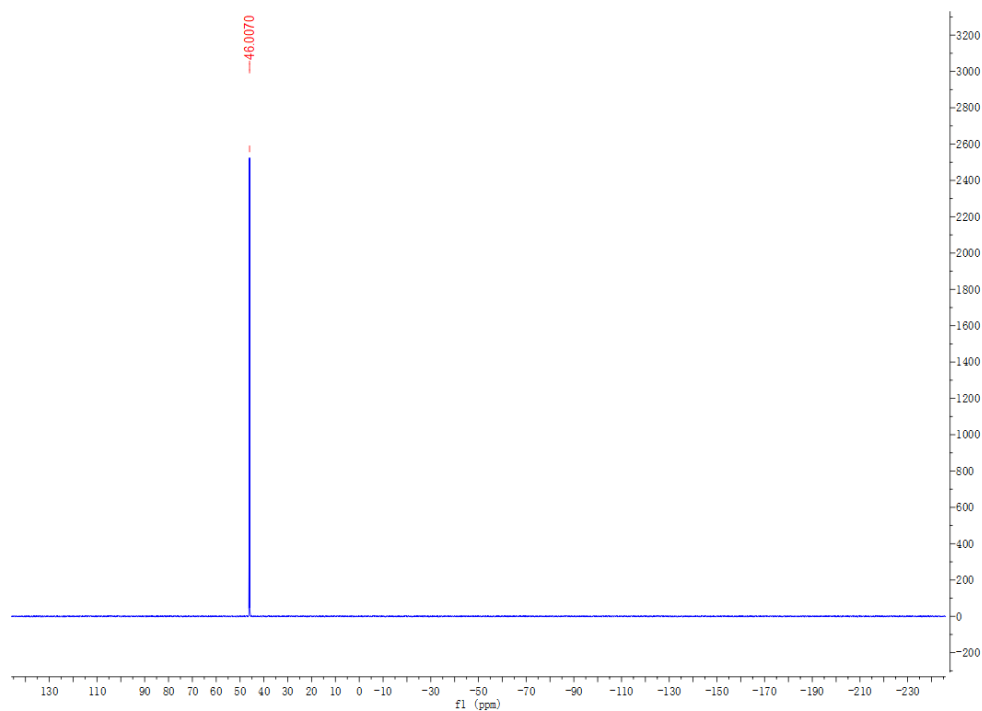


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.881	BB	0.2069	5673.06494	411.40399	98.3002
2	7.600	BB	0.2540	98.09958	5.90015	1.6998

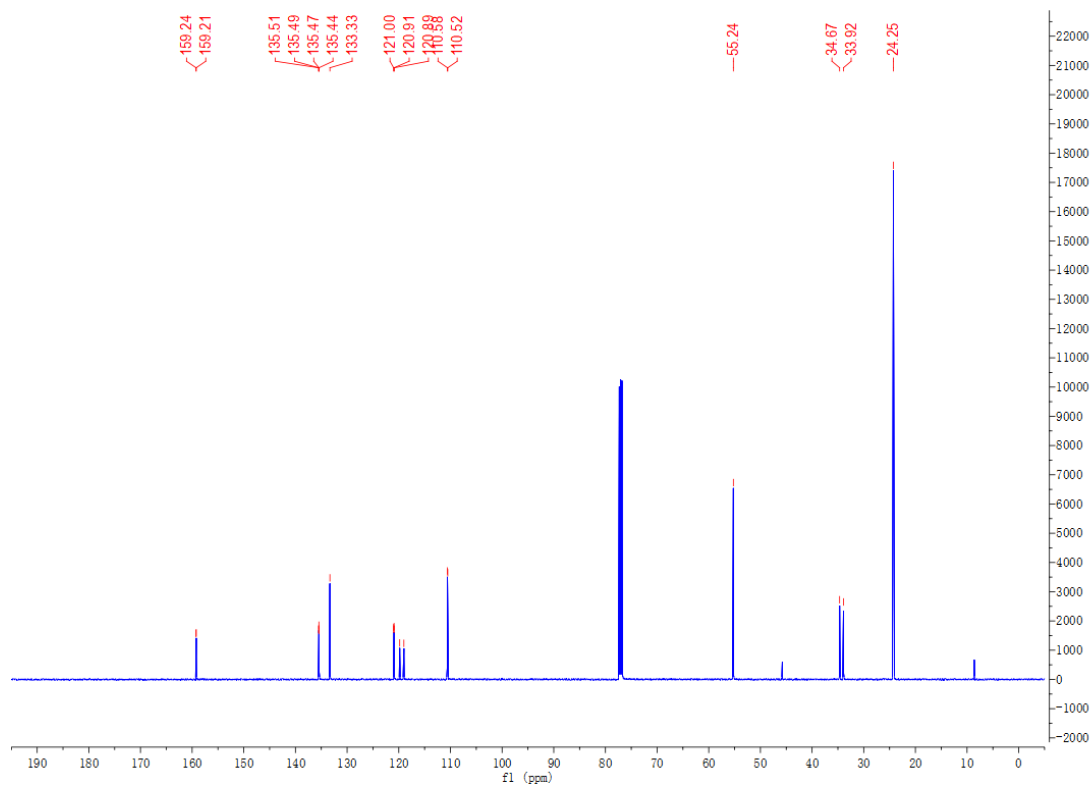
¹H NMR of **8b**



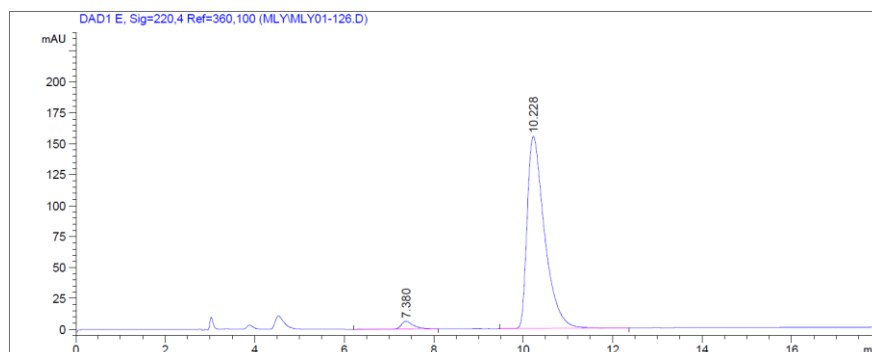
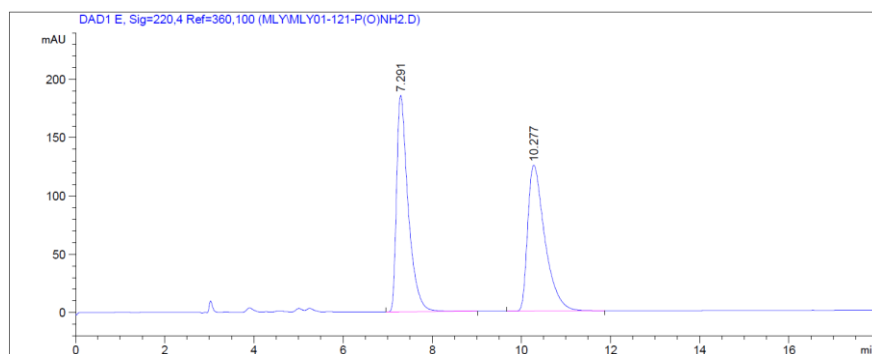
³¹P NMR of **8b**



¹³C NMR of **8b**

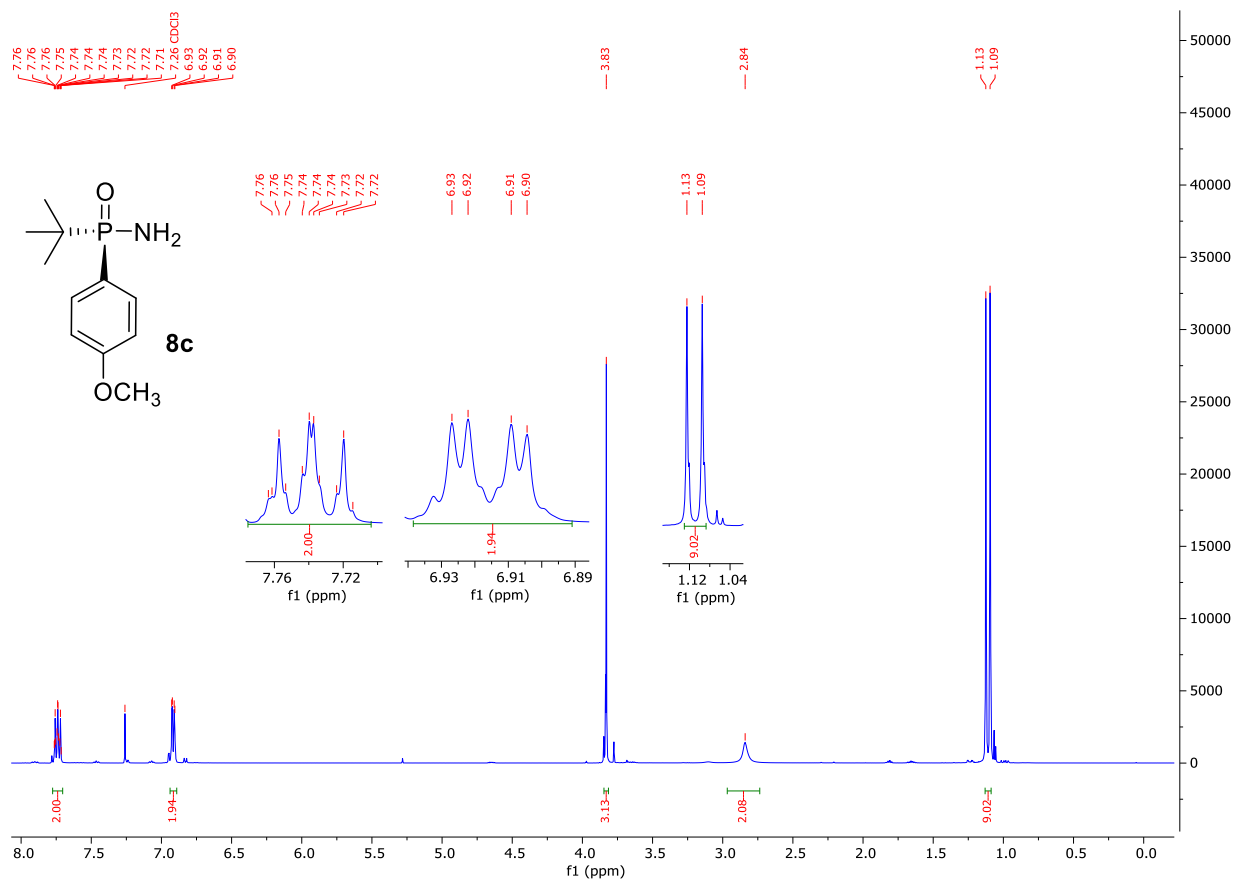


Chiral HPLC chromatograms of **8b**

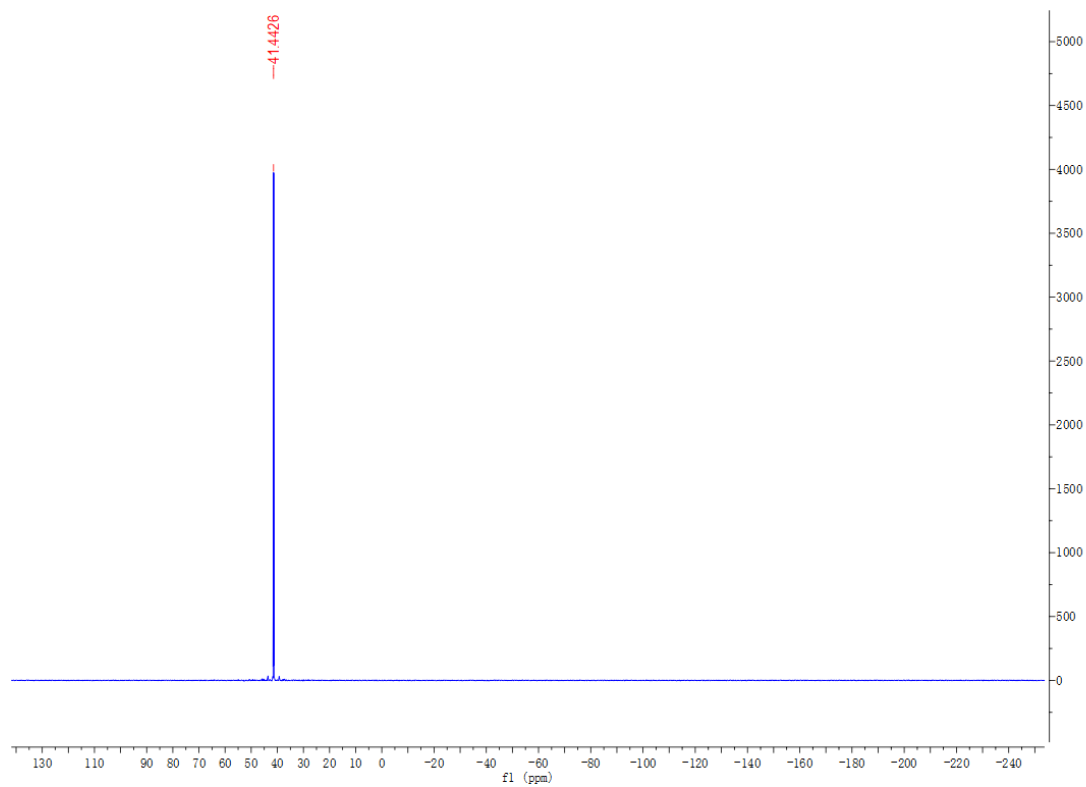


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
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2	10.228	BB	0.4025	4181.28223	155.10017	97.2392

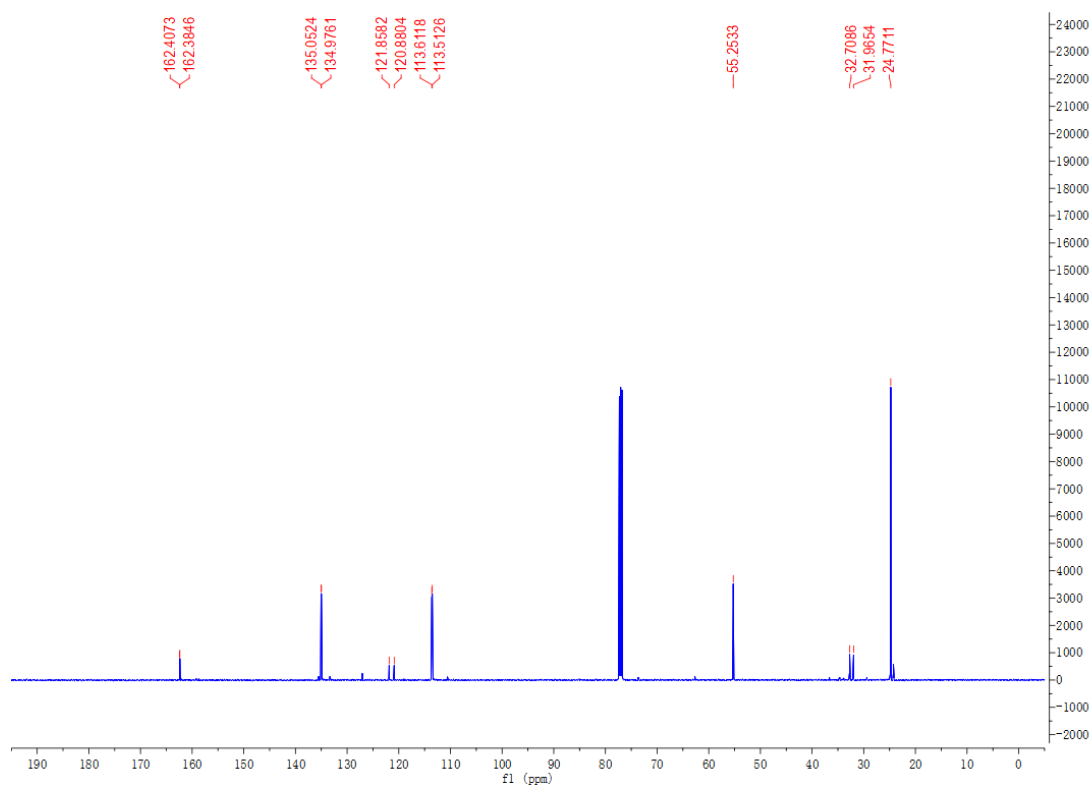
^1H NMR of **8c**



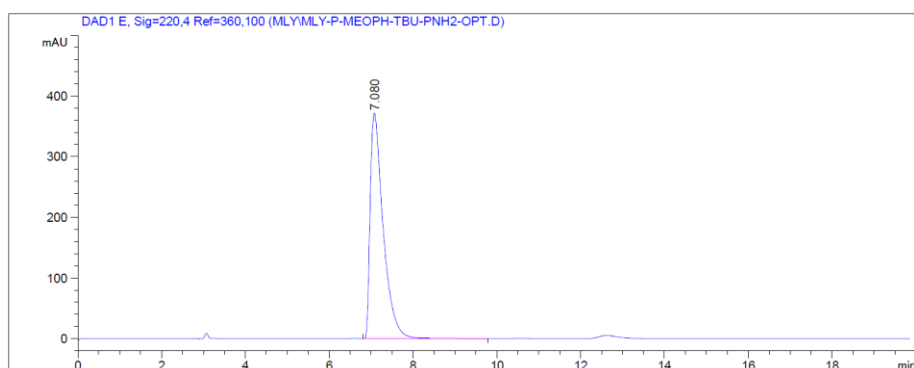
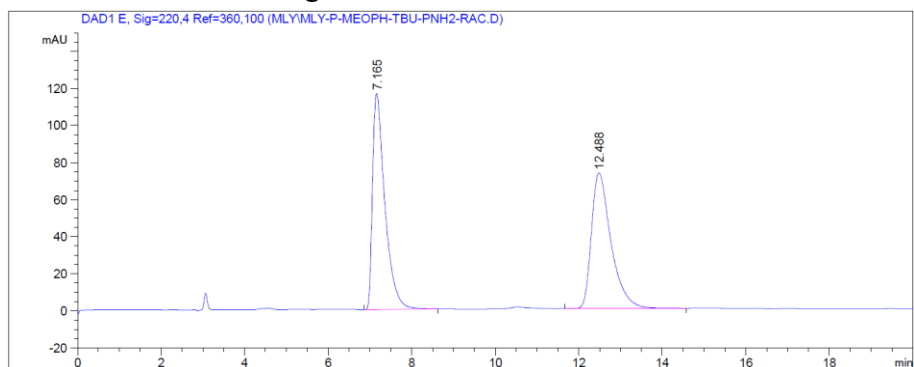
^{31}P NMR of **8c**



¹³C NMR of **8c**

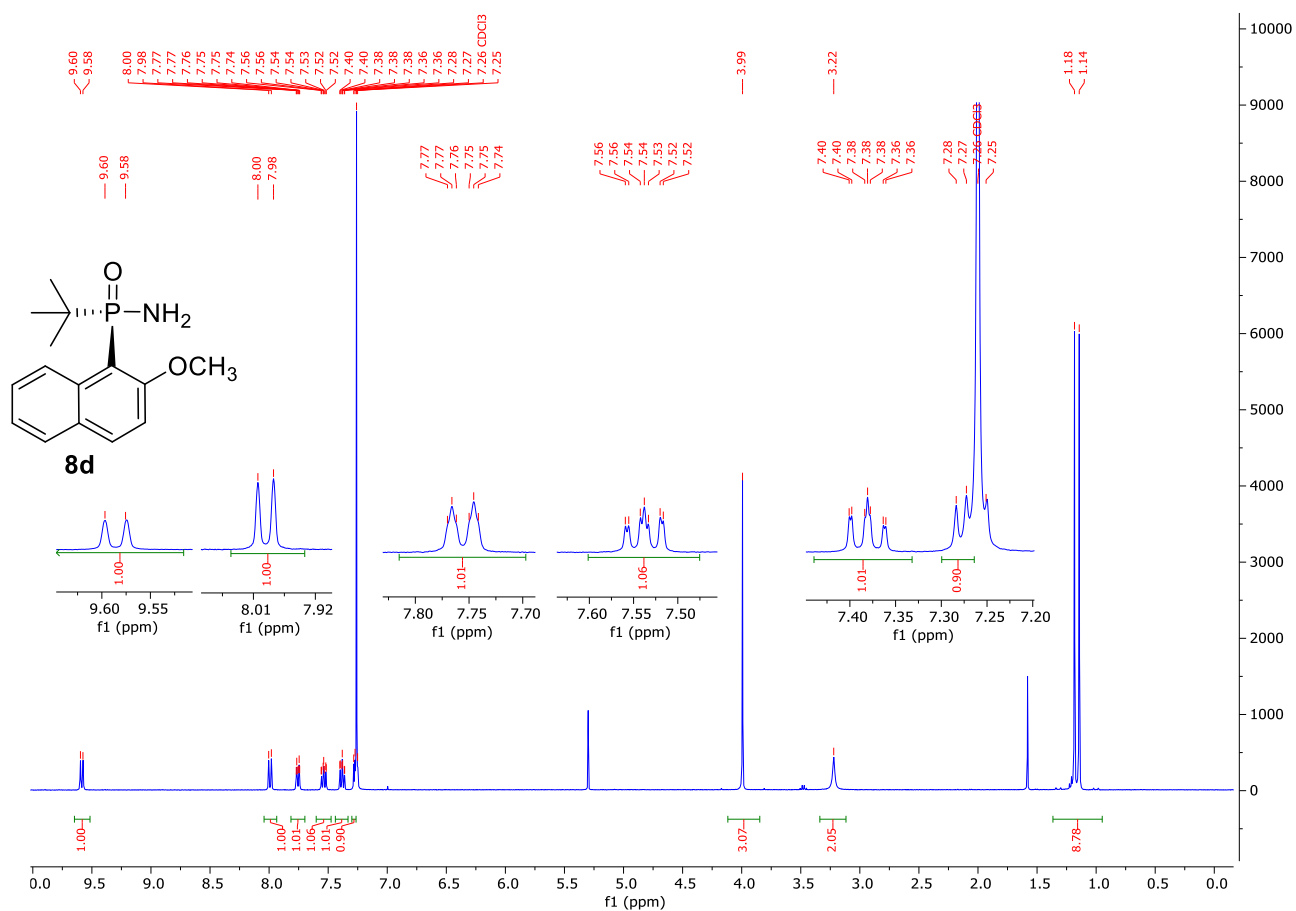


Chiral HPLC chromatograms of **8c**

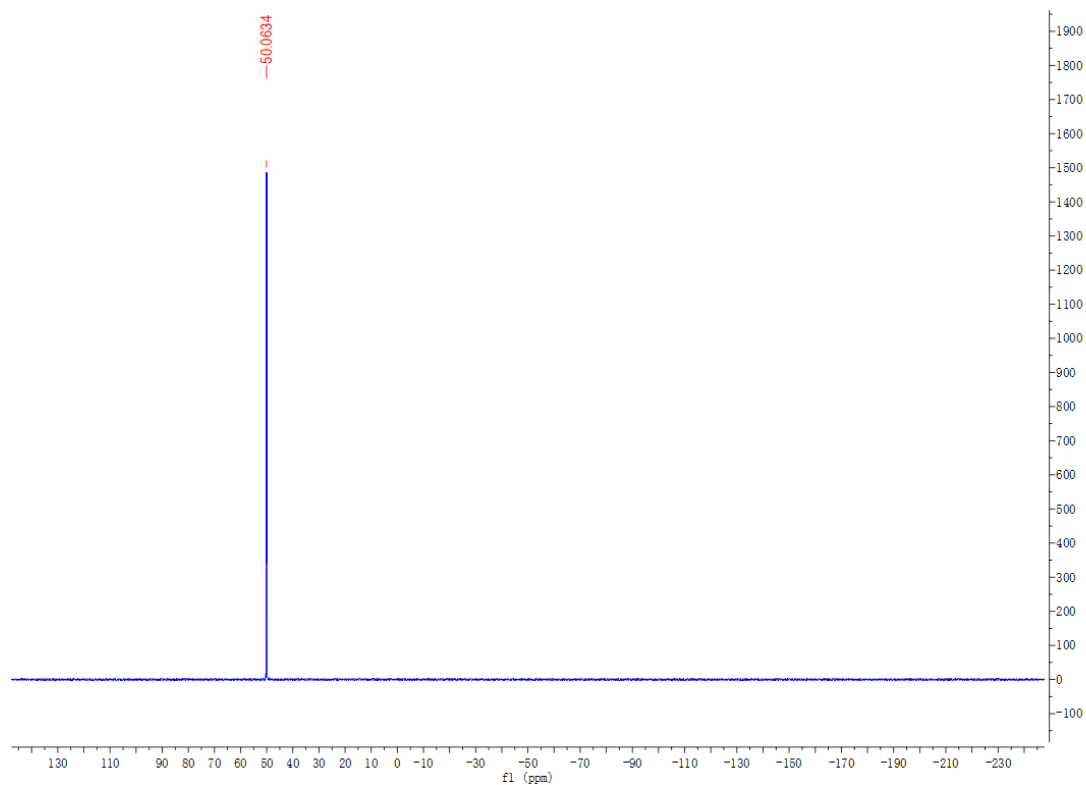


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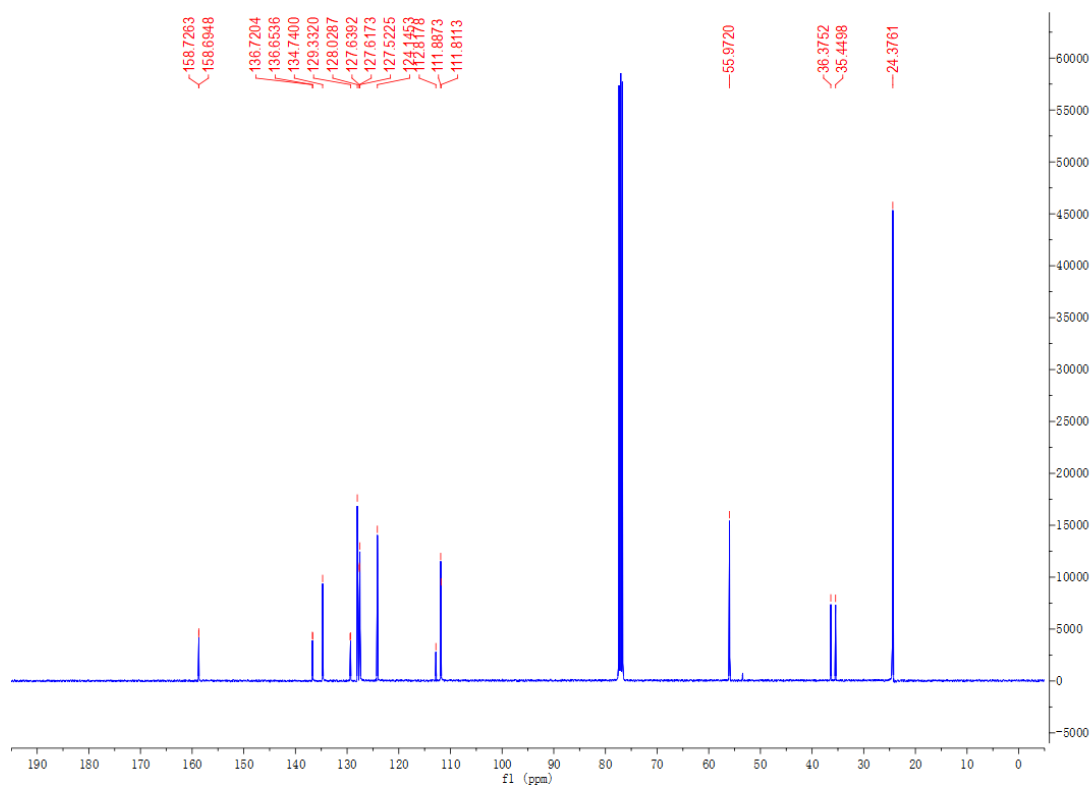
¹H NMR of 8d



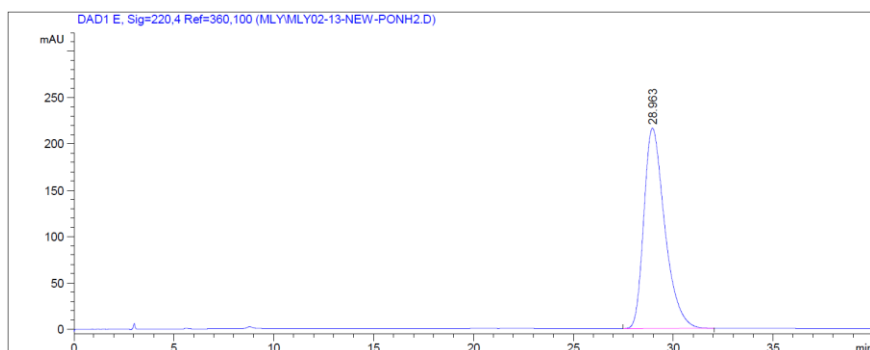
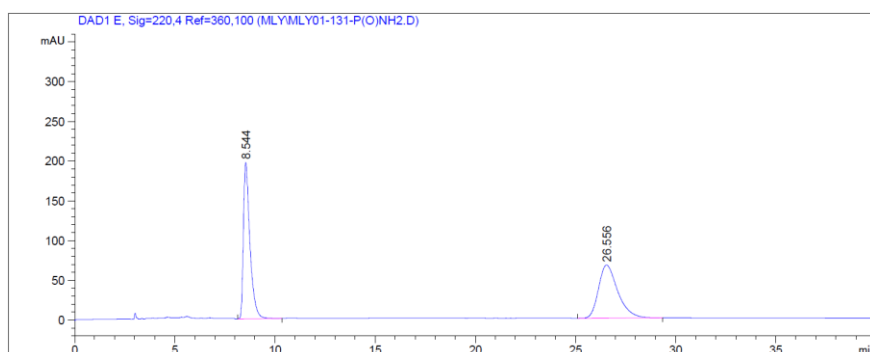
³¹P NMR of 8d



¹³C NMR of **8d**

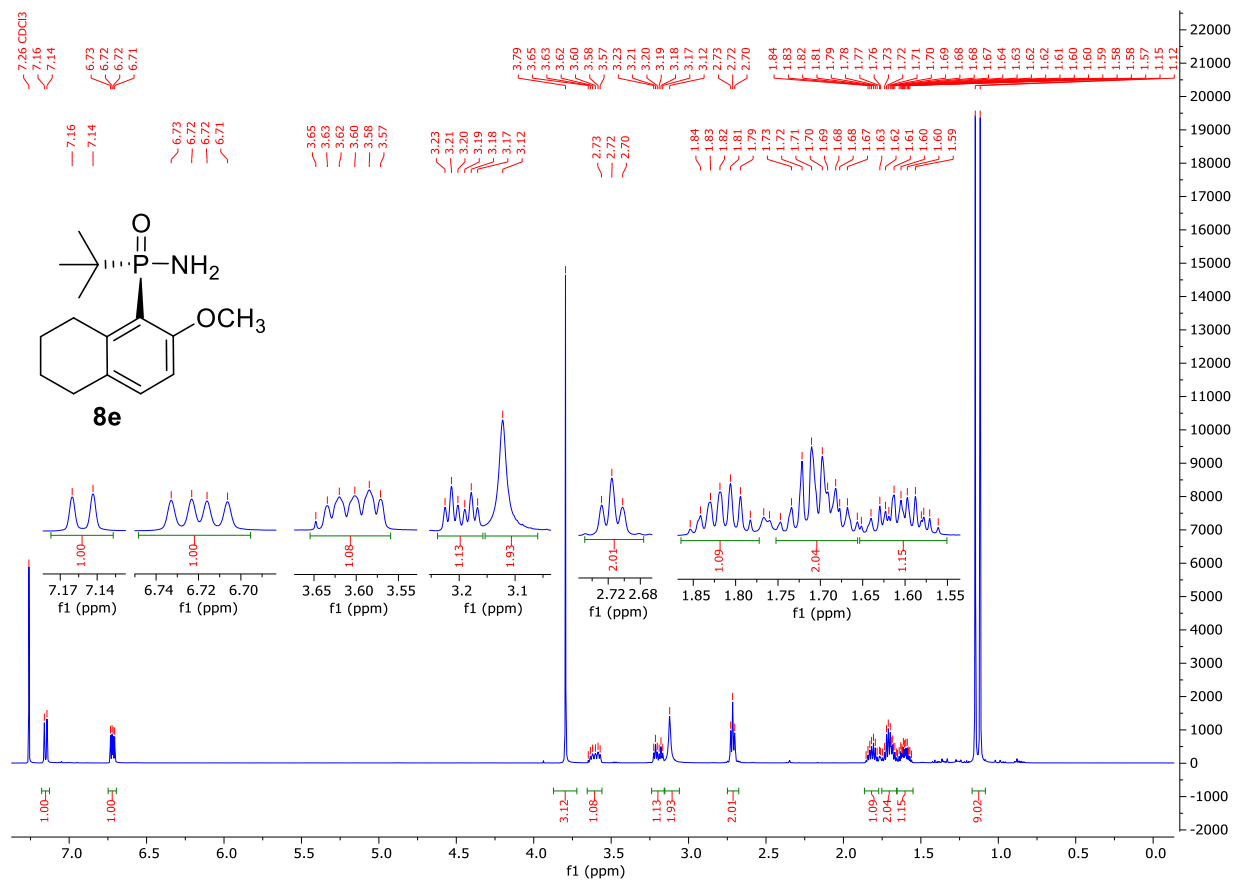


Chiral HPLC chromatograms of **8d**

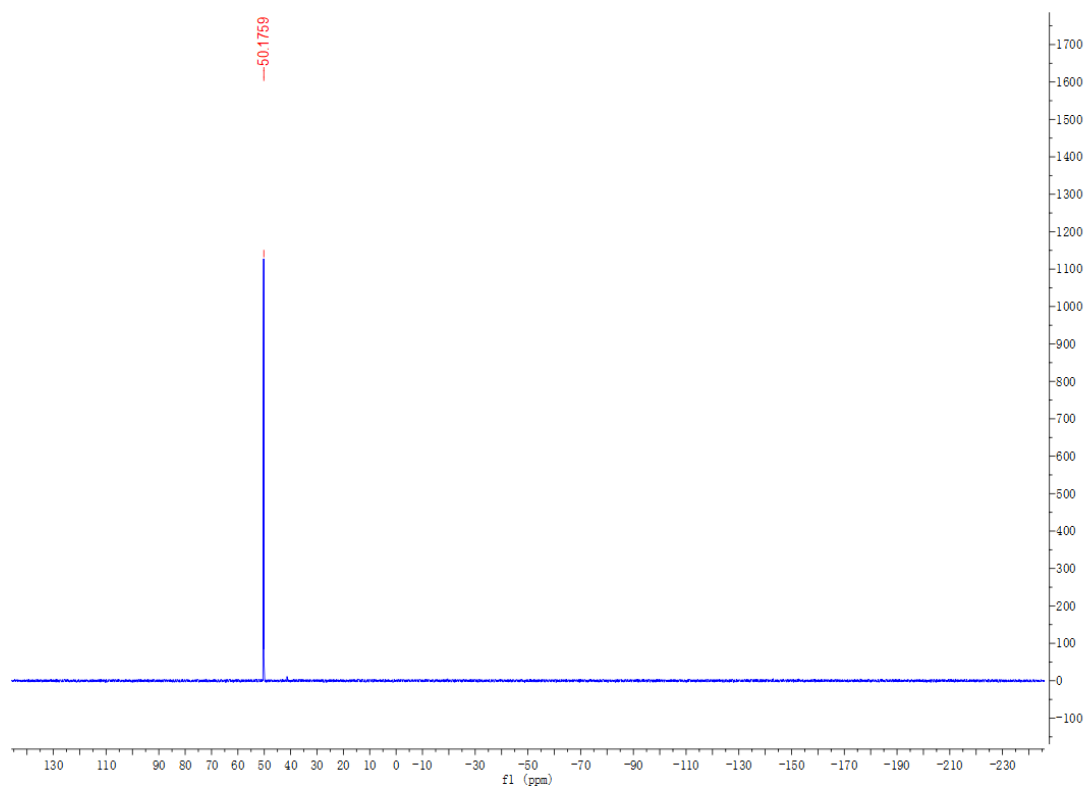


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
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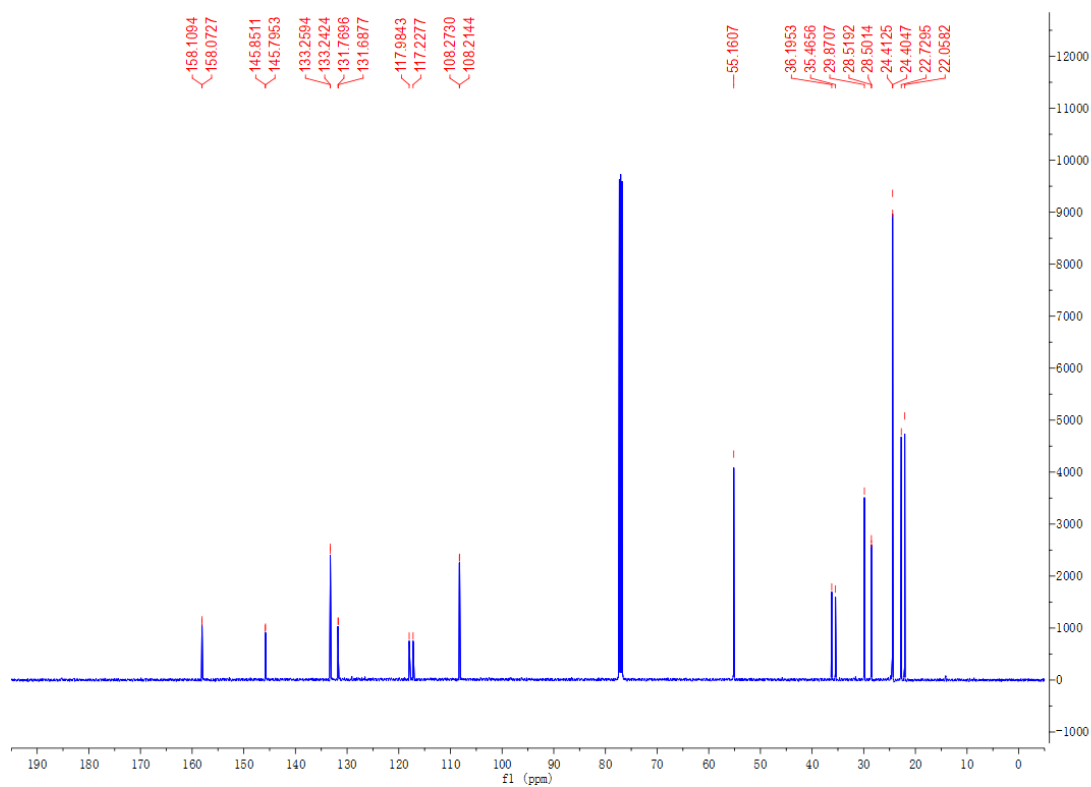
¹H NMR of 8e



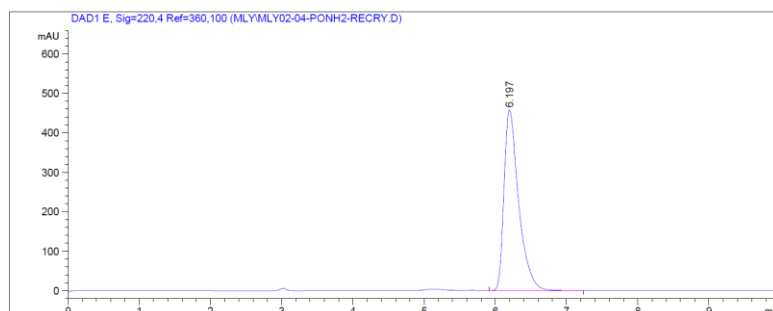
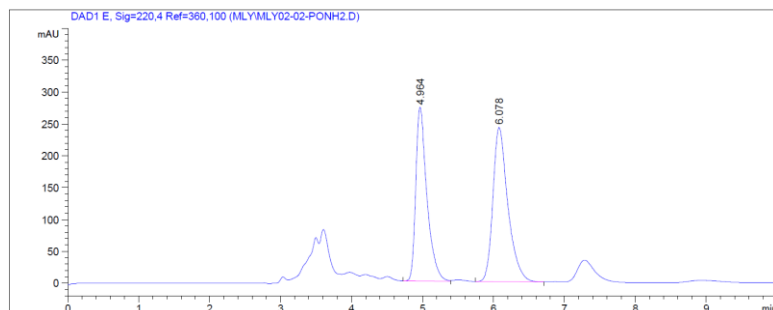
³¹P NMR of 8e



¹³C NMR of **8e**

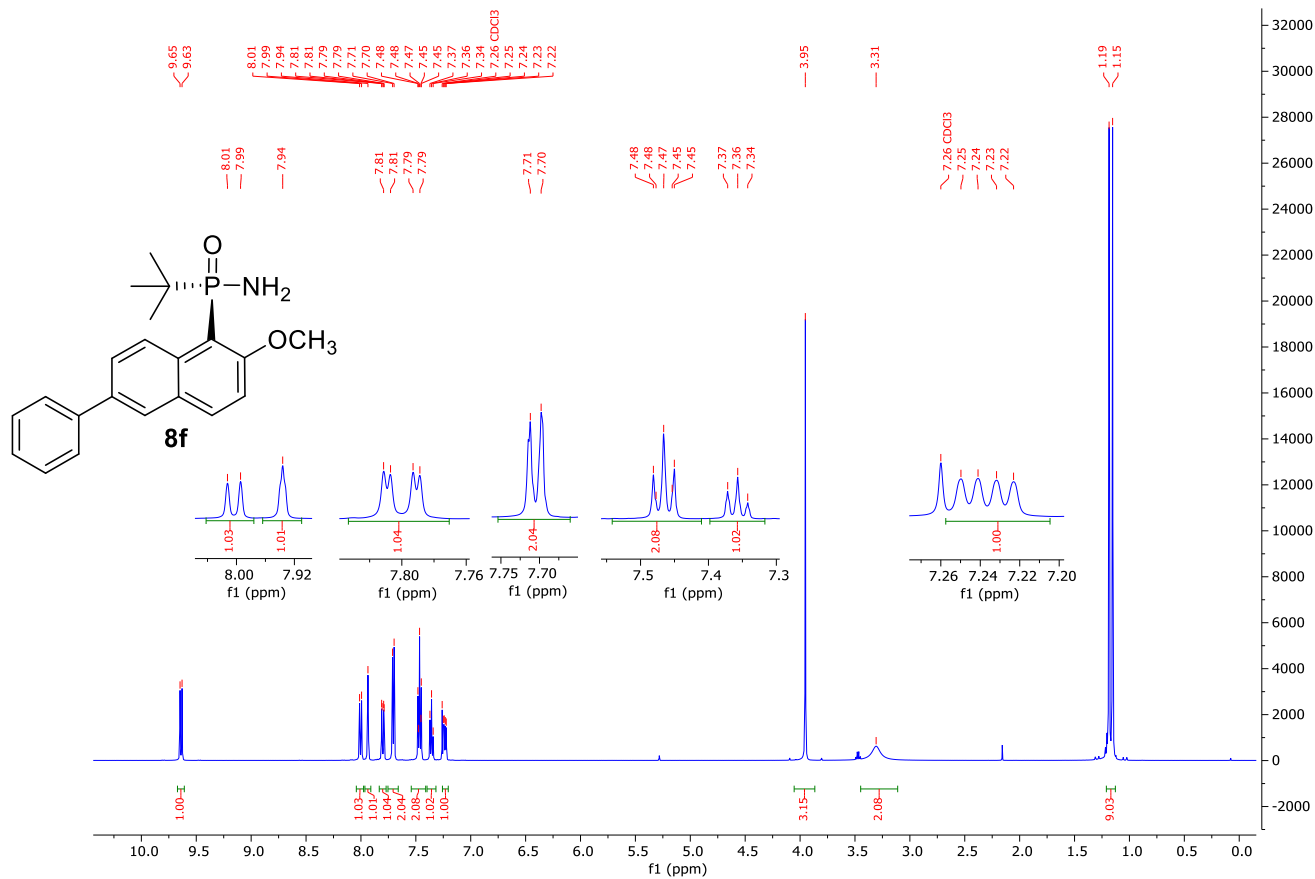


Chiral HPLC chromatograms of **8e**

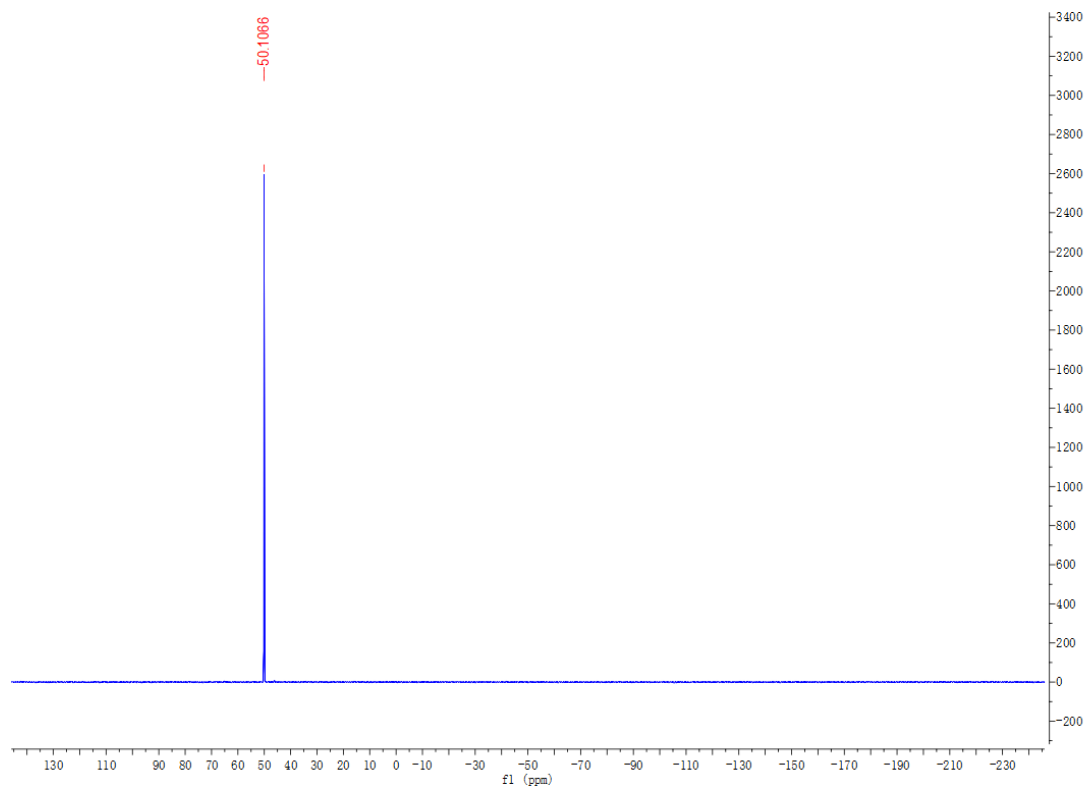


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.197	BV	0.2148	6538.37207	457.20520	100.0000

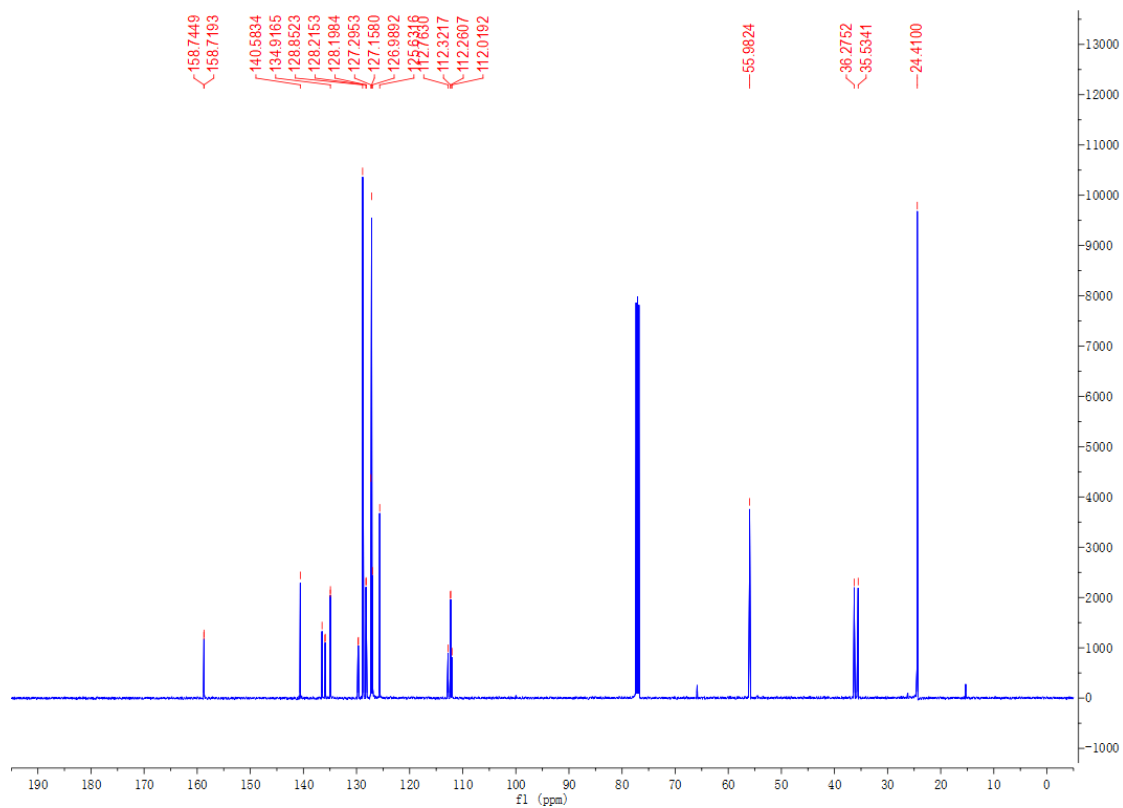
¹H NMR of **8f**



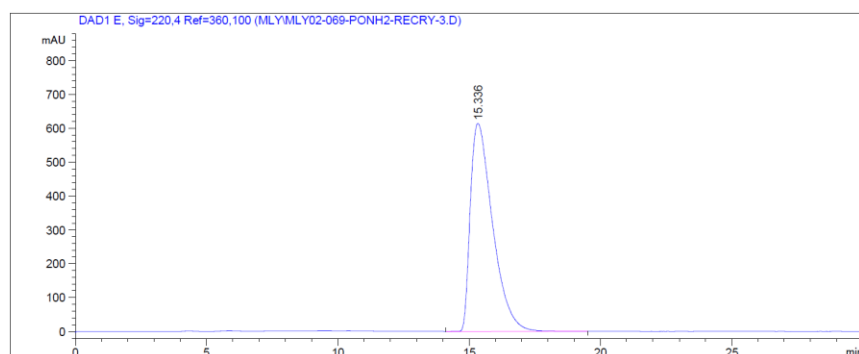
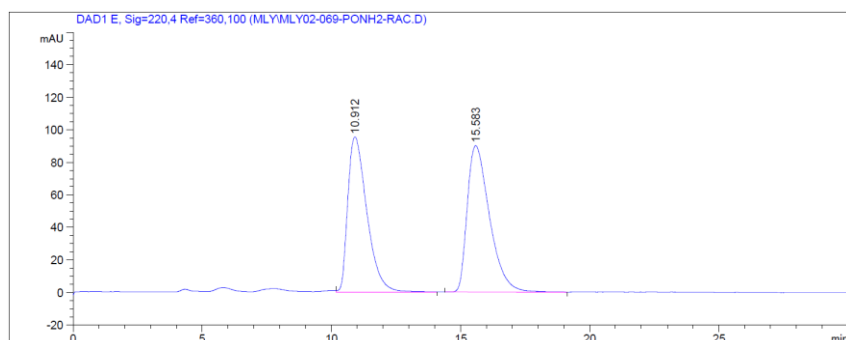
³¹P NMR of **8f**



¹³C NMR of **8f**

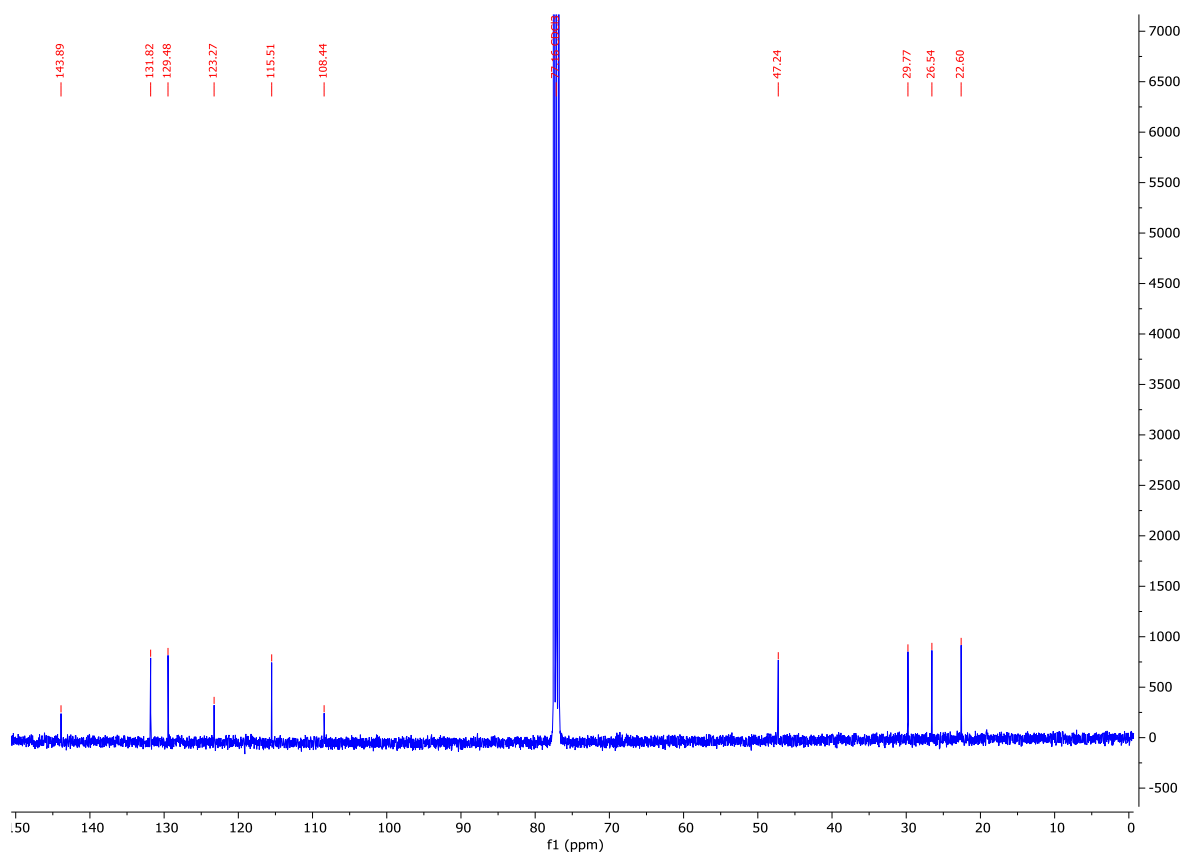


Chiral HPLC chromatograms of **8f**

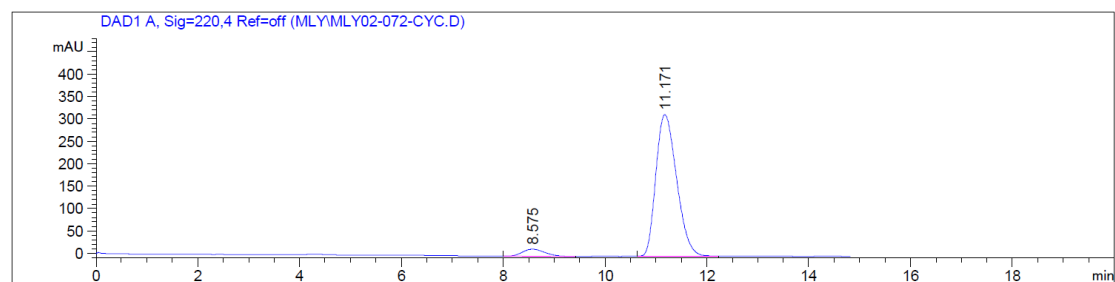
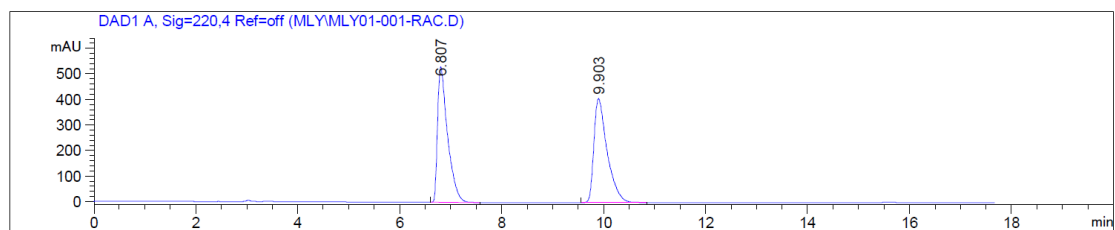


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.336	BV	0.9039	3.62257e4	614.41528	100.0000

¹³C NMR of 11a



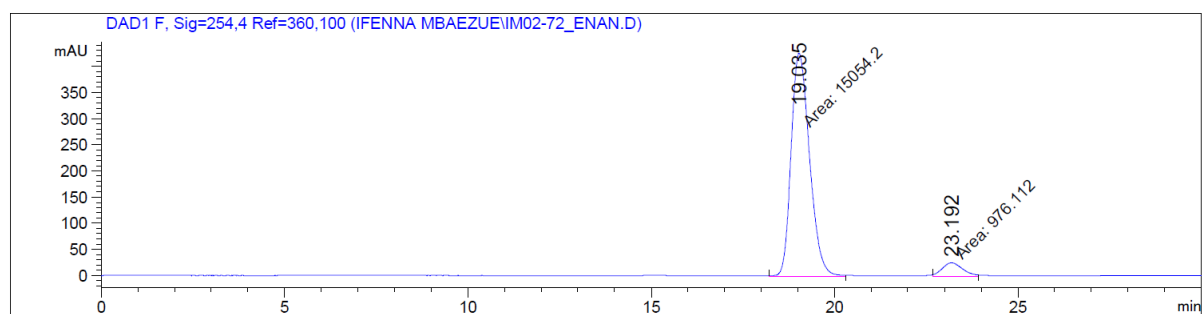
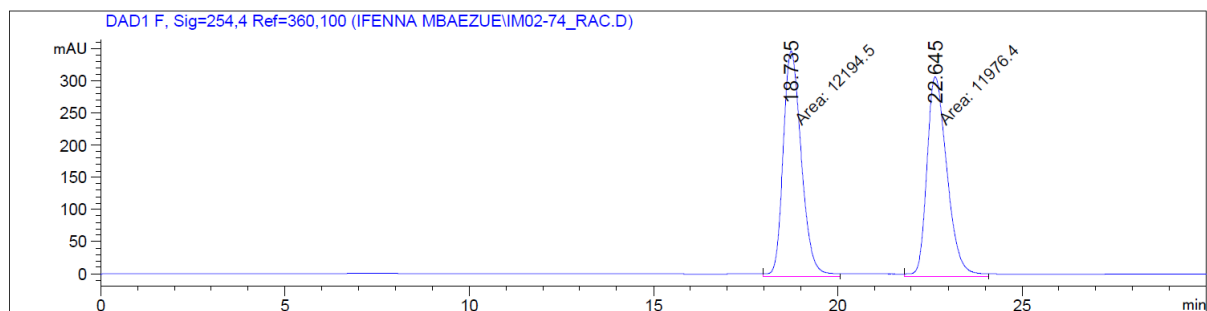
Chiral HPLC Chromatograms of 11a



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.575	BB	0.4773	492.90408	16.00656	5.2228
2	11.171	BB	0.4480	8944.63574	316.22061	94.7772

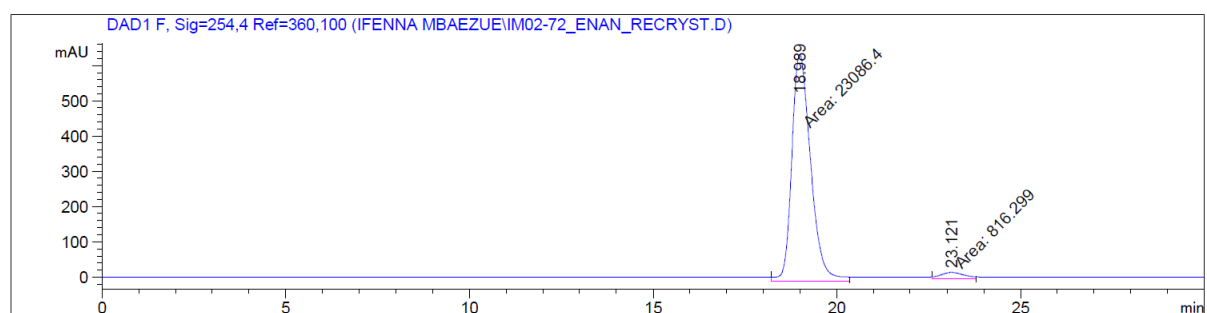
For comparison compound **11a** was also analyzed using a chiracel OJ-H comlun in comparison with the literature.⁹

Chiral HPLC Chromatograms of **11a**



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.035	MM	0.5867	1.50542e4	427.68332	93.9108
2	23.192	MM	0.6424	976.11206	25.32313	6.0892

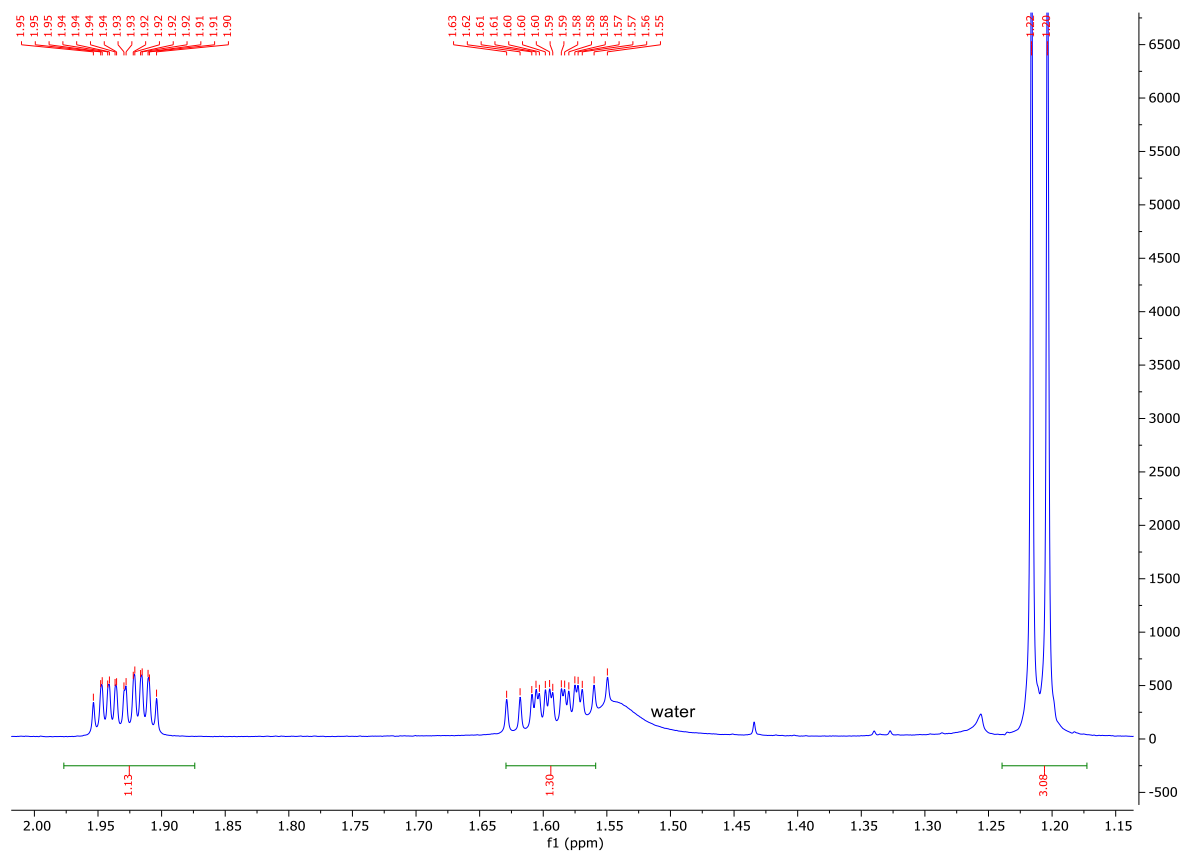
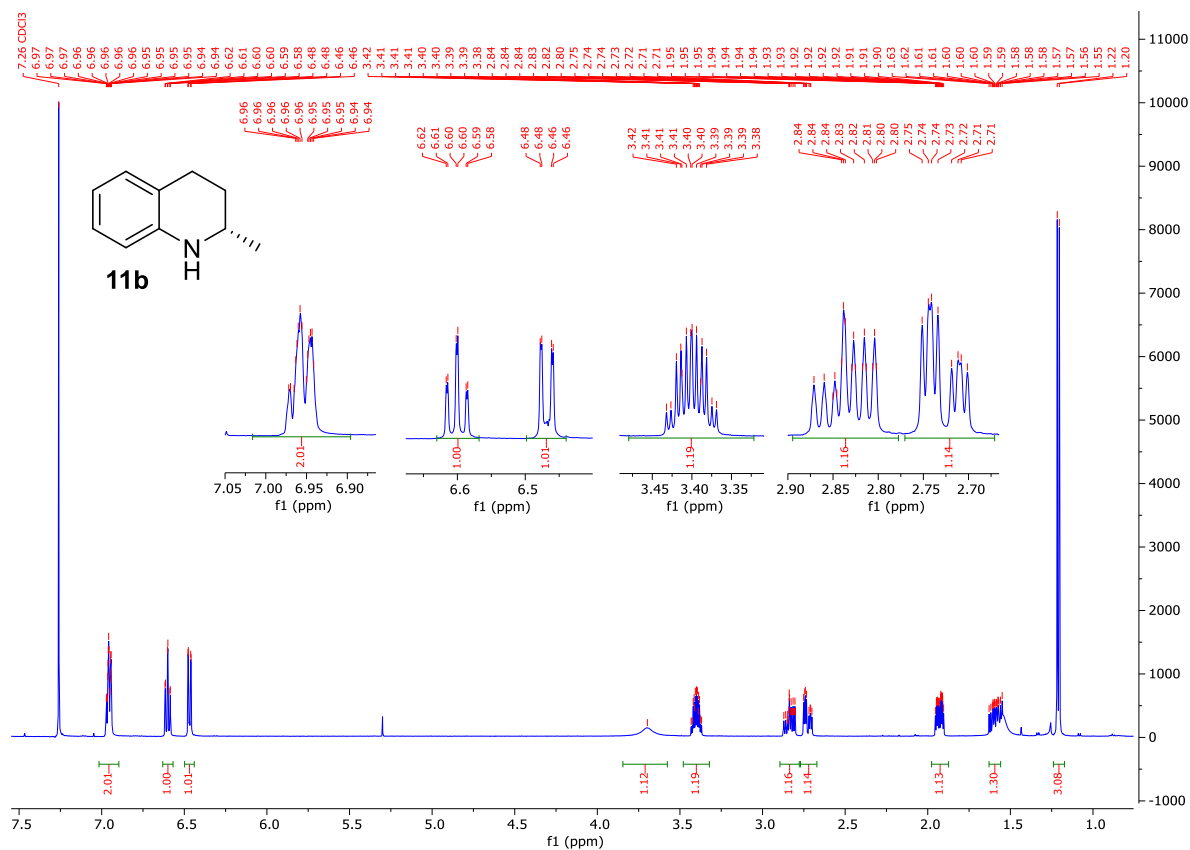
Chiral HPLC Chromatogram of crystallized product **11a**



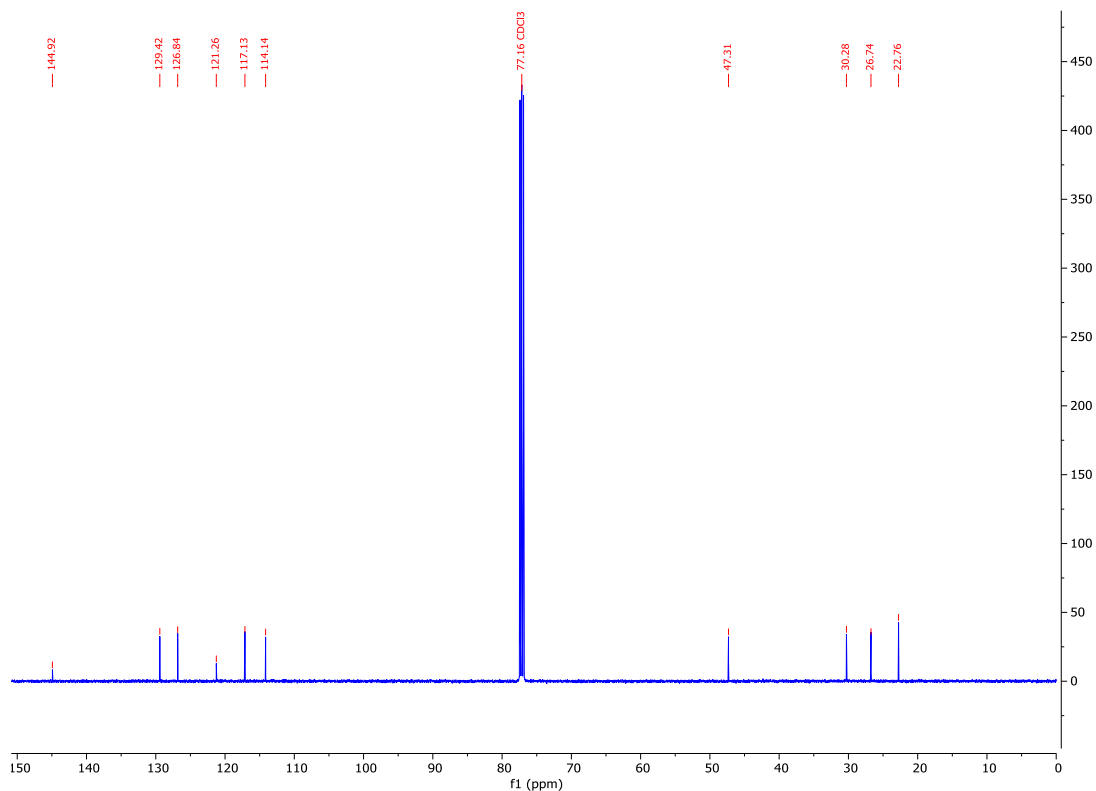
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.989	MM	0.5980	2.30864e4	643.44177	96.5849
2	23.121	MM	0.7581	816.29865	17.94708	3.4151

(S)-2-methyl-1,2,3,4-tetrahydroquinoline (**11b**)

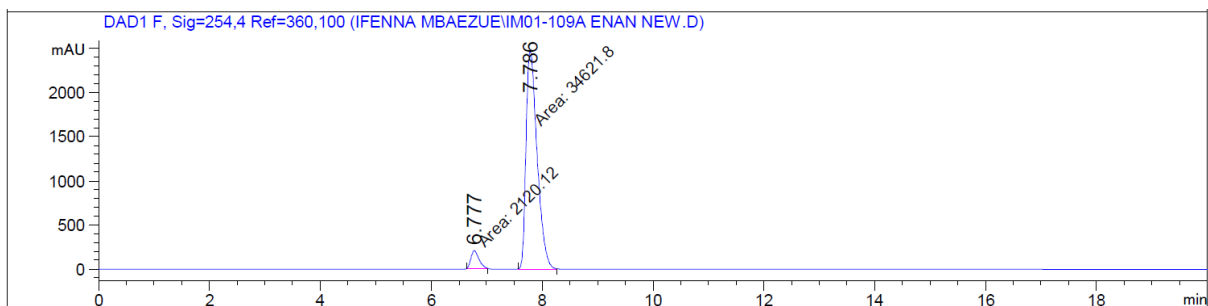
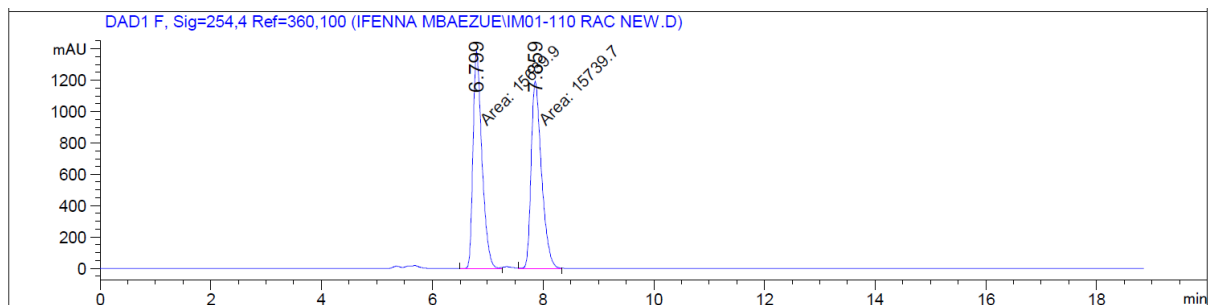
¹H NMR of **11b**



¹³C NMR of **11b**



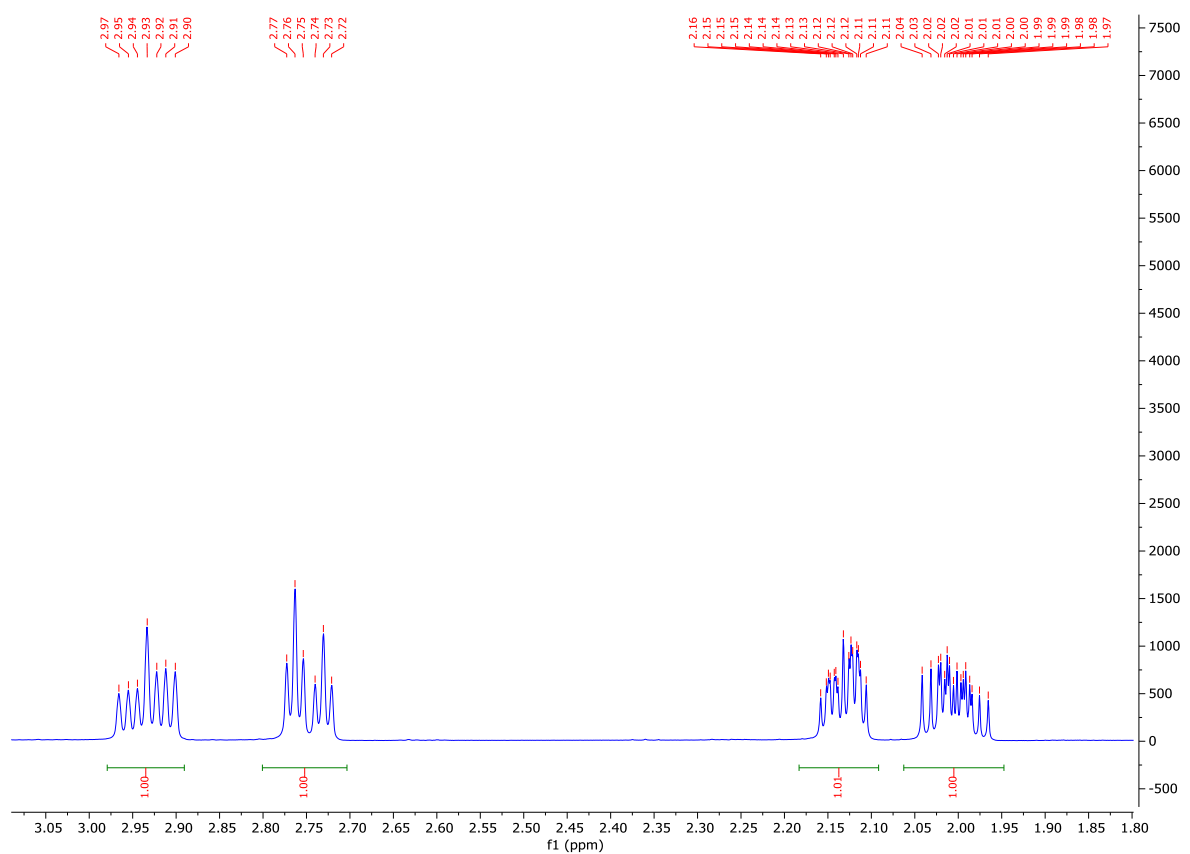
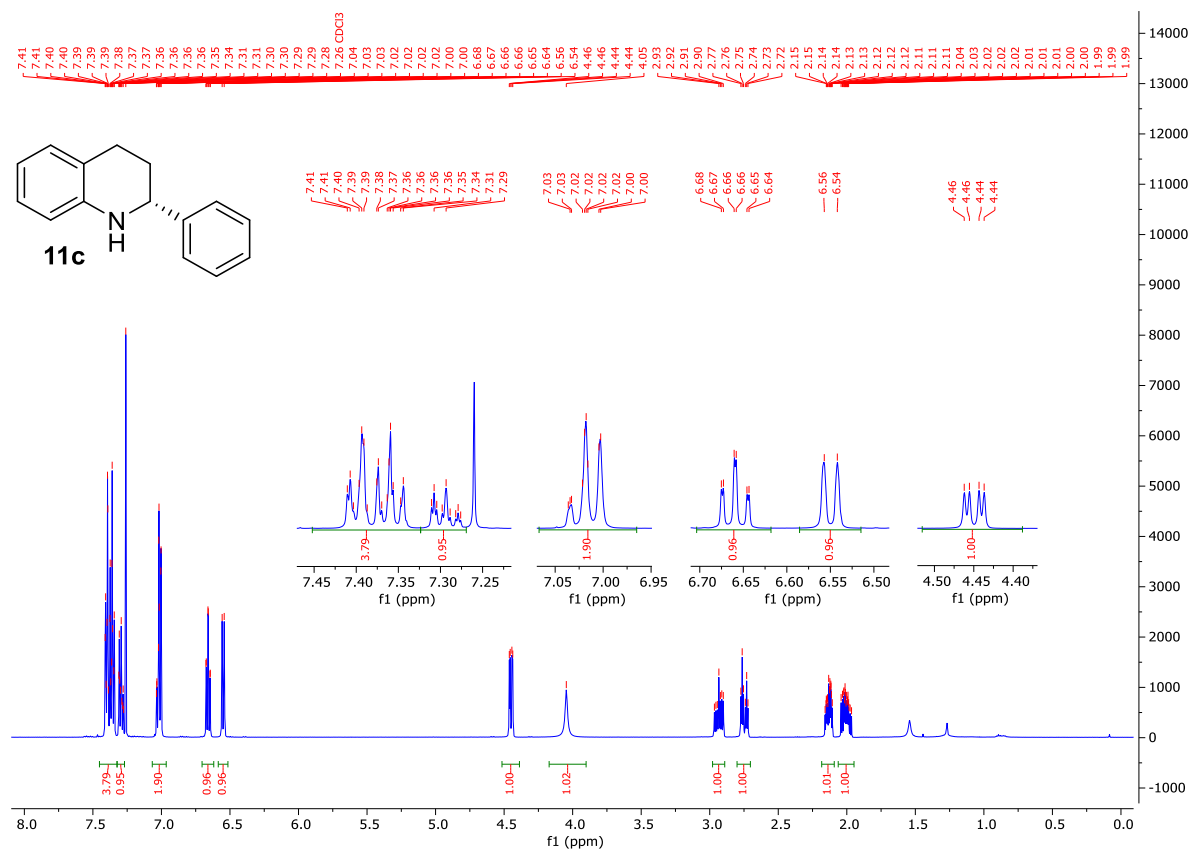
Chiral HPLC Chromatograms of **11b**



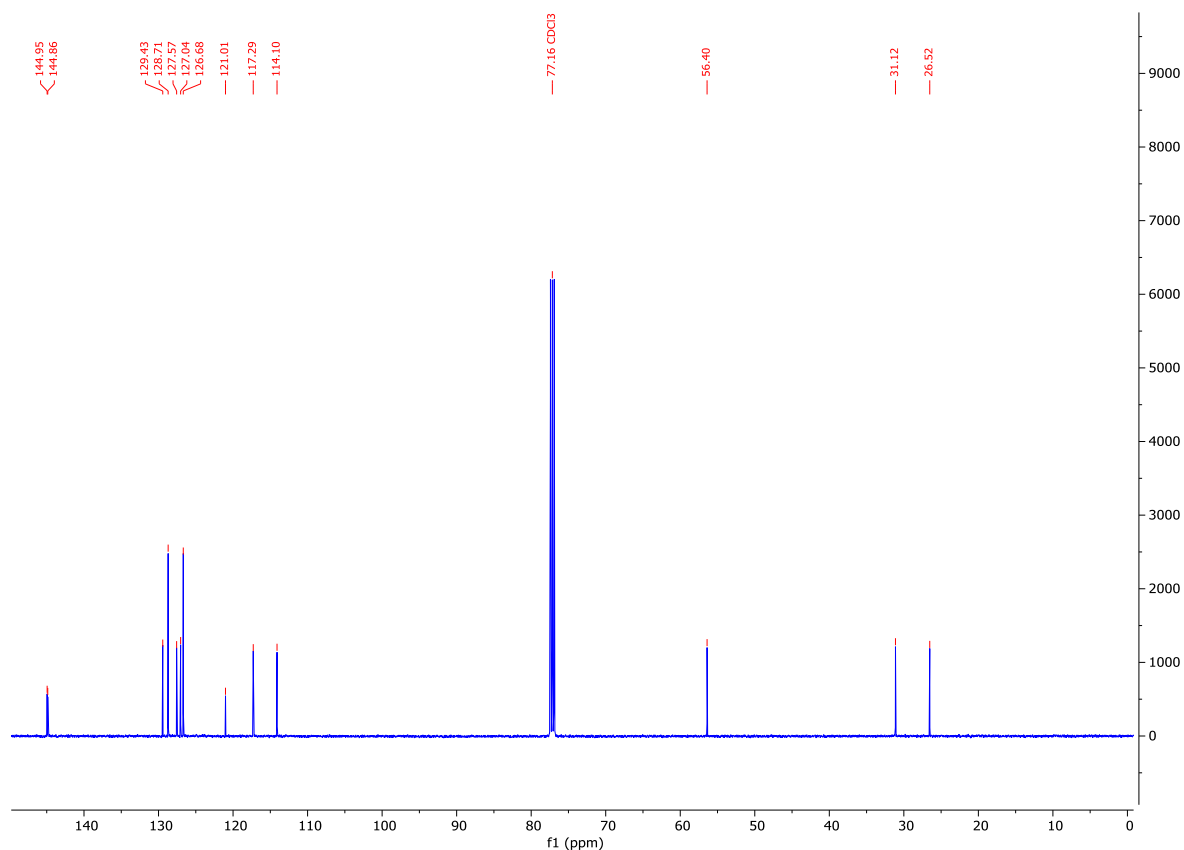
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.777	MM	0.1708	2120.12207	206.83456	5.7703
2	7.786	MM	0.2332	3.46218e4	2474.55640	94.2297

(R)-2-phenyl-1,2,3,4-tetrahydroquinoline (**11c**)

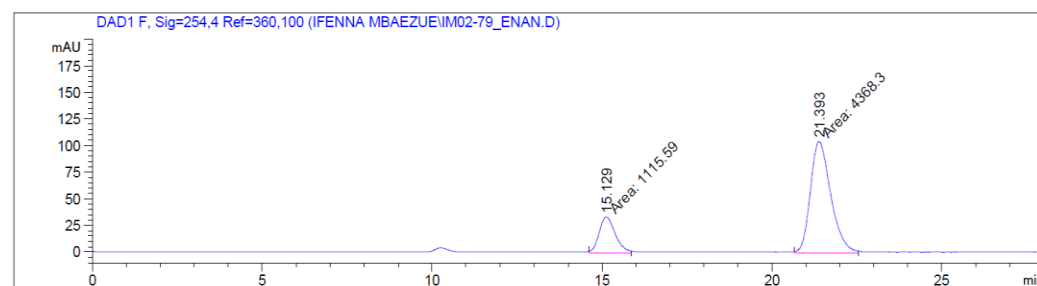
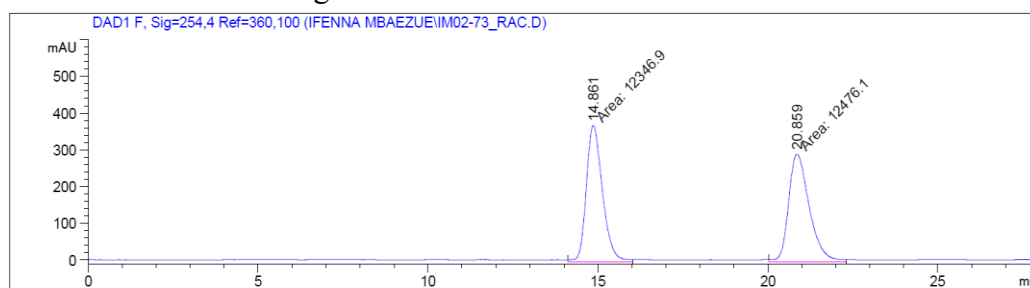
¹H NMR of **11c**



¹³C NMR of 11c



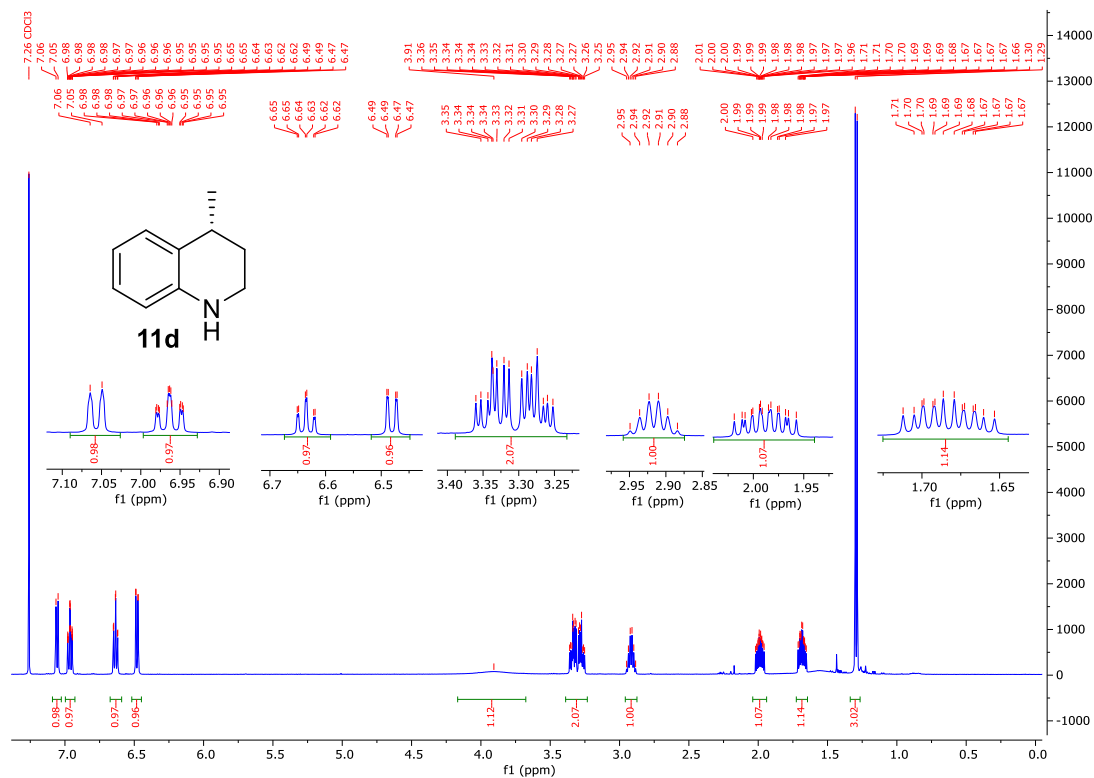
Chiral HPLC Chromatograms of 11c



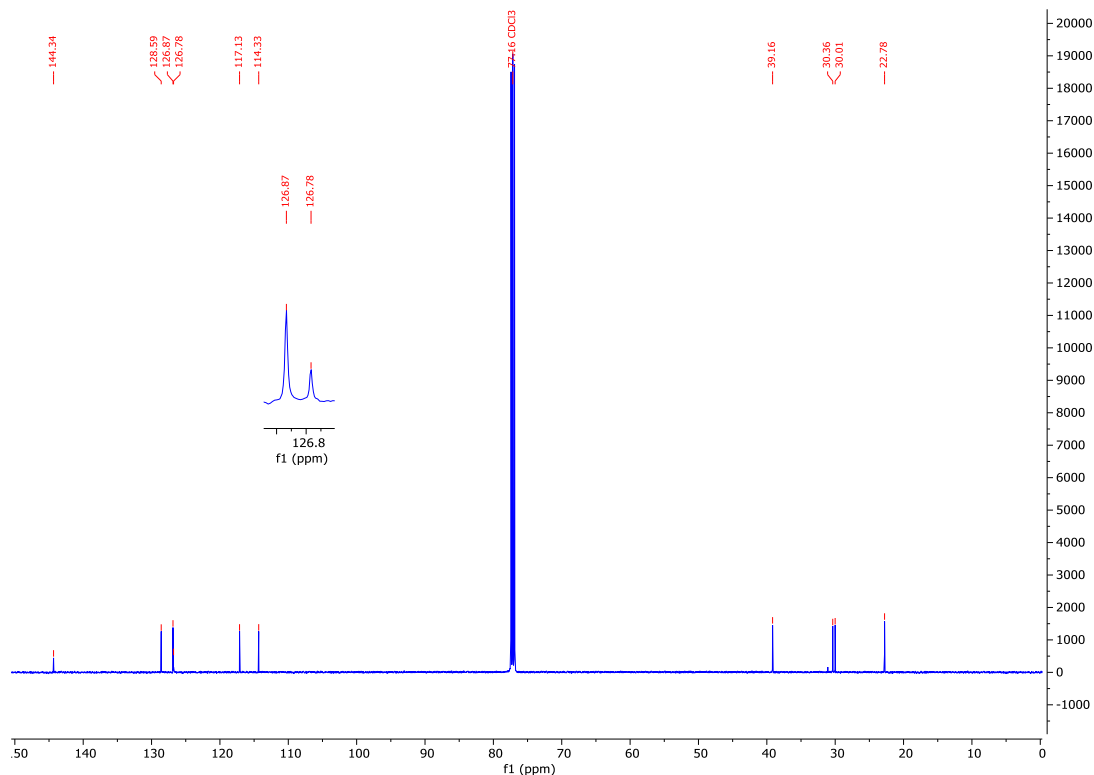
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.129	MM	0.5518	1115.59338	33.69407	20.3431
2	21.393	MM	0.6967	4368.30176	104.49825	79.6569

(R)-4-methyl-1,2,3,4-tetrahydroquinoline (11d)

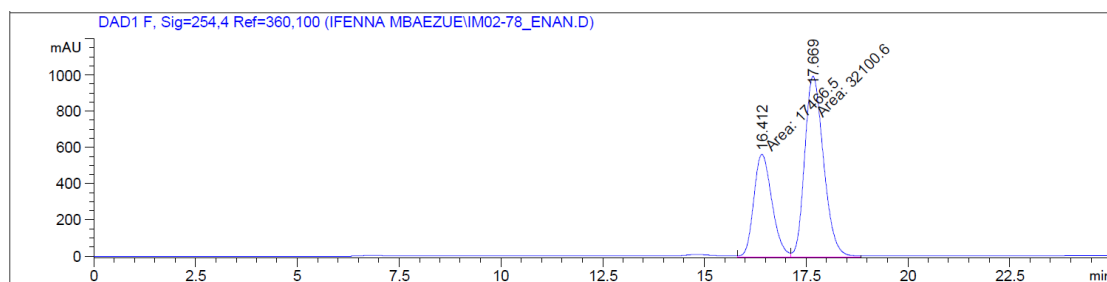
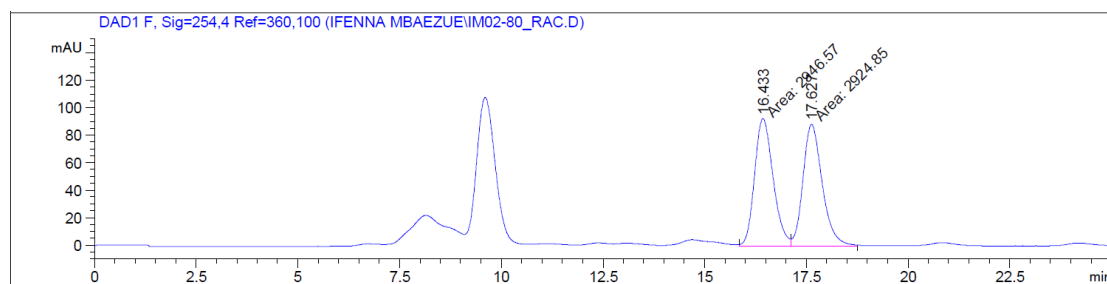
¹H NMR of 11d



¹³C NMR of 11d



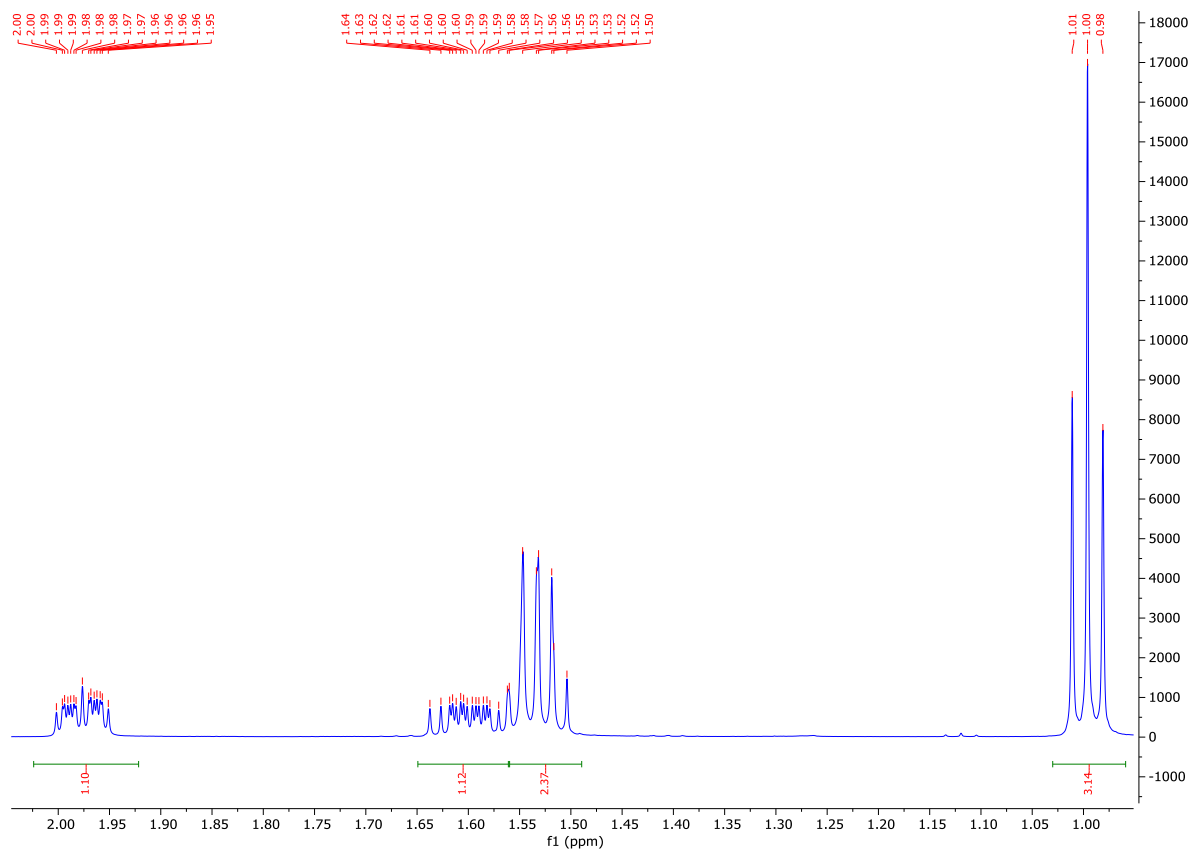
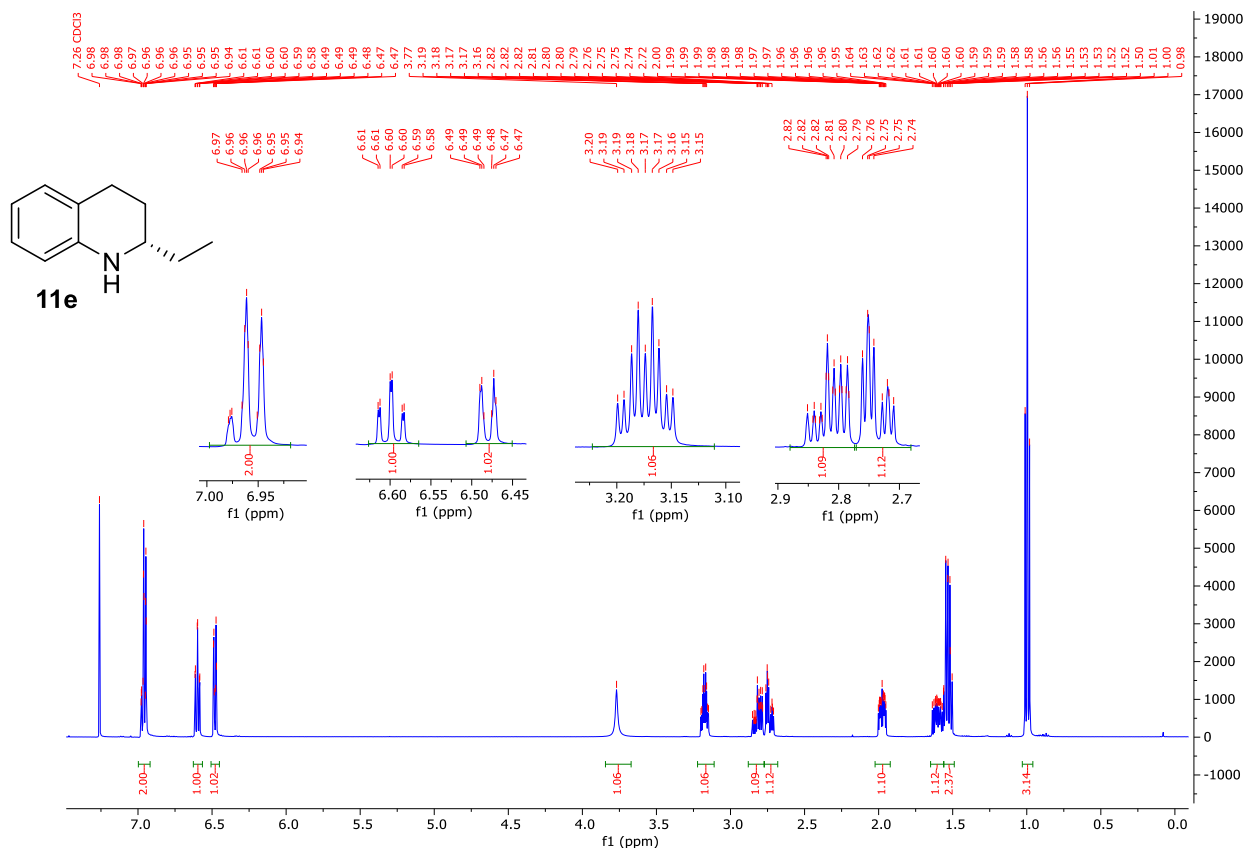
Chiral HPLC Chromatograms of **11d**



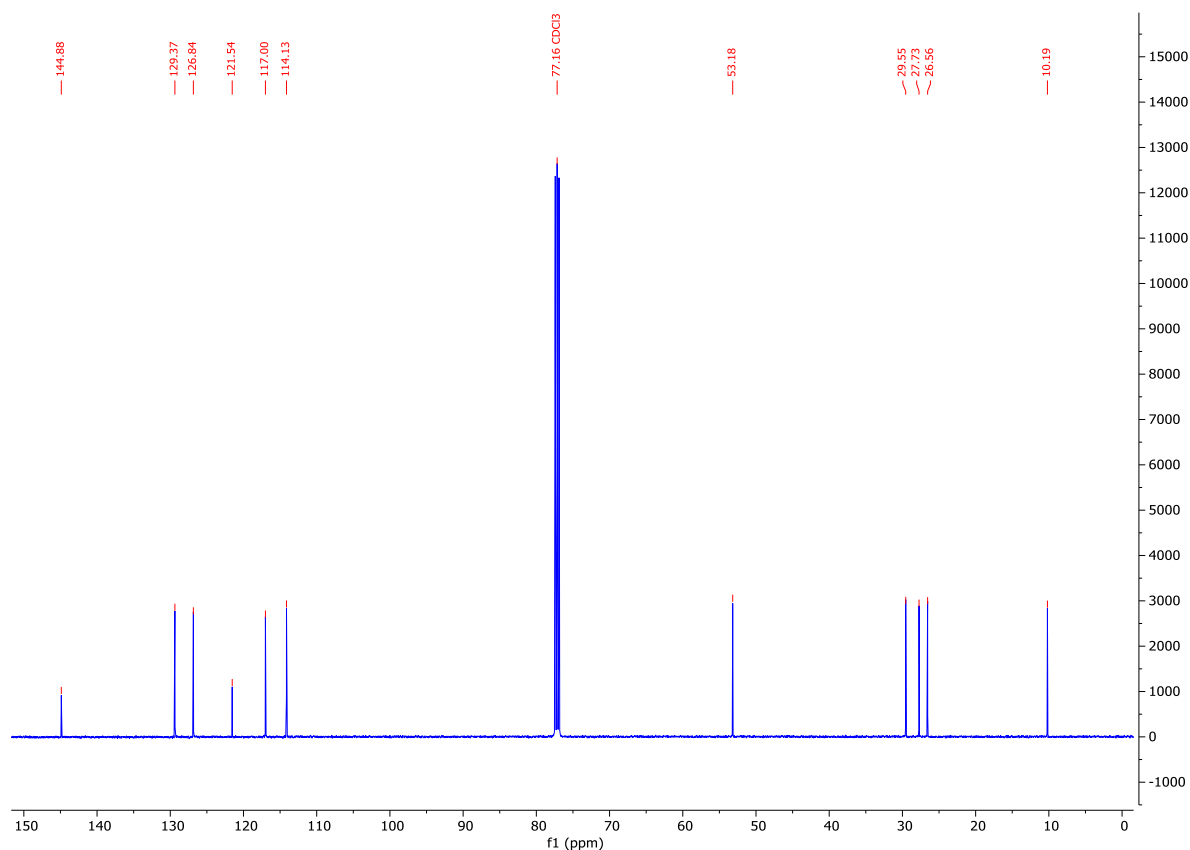
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.412	MM	0.5150	1.74665e4	565.20856	35.2382
2	17.669	MM	0.5373	3.21006e4	995.67267	64.7618

(S)-2-ethyl-1,2,3,4-tetrahydroquinoline (11e)

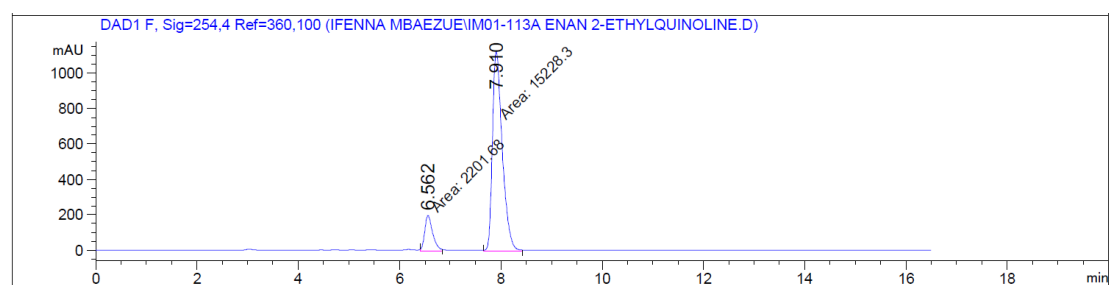
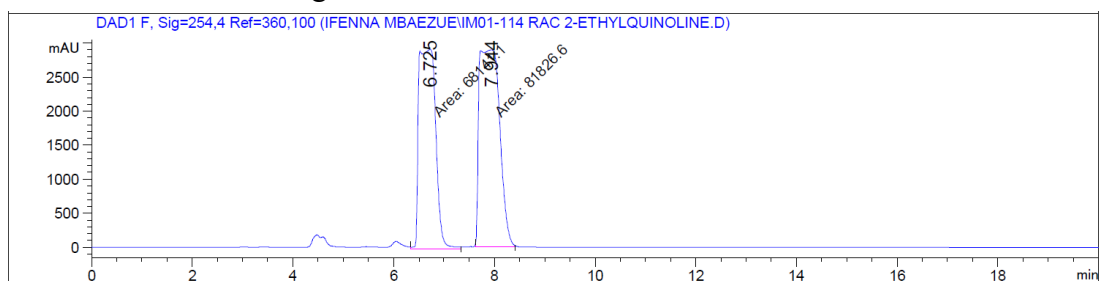
¹H NMR of 11e



¹³C NMR of 11e

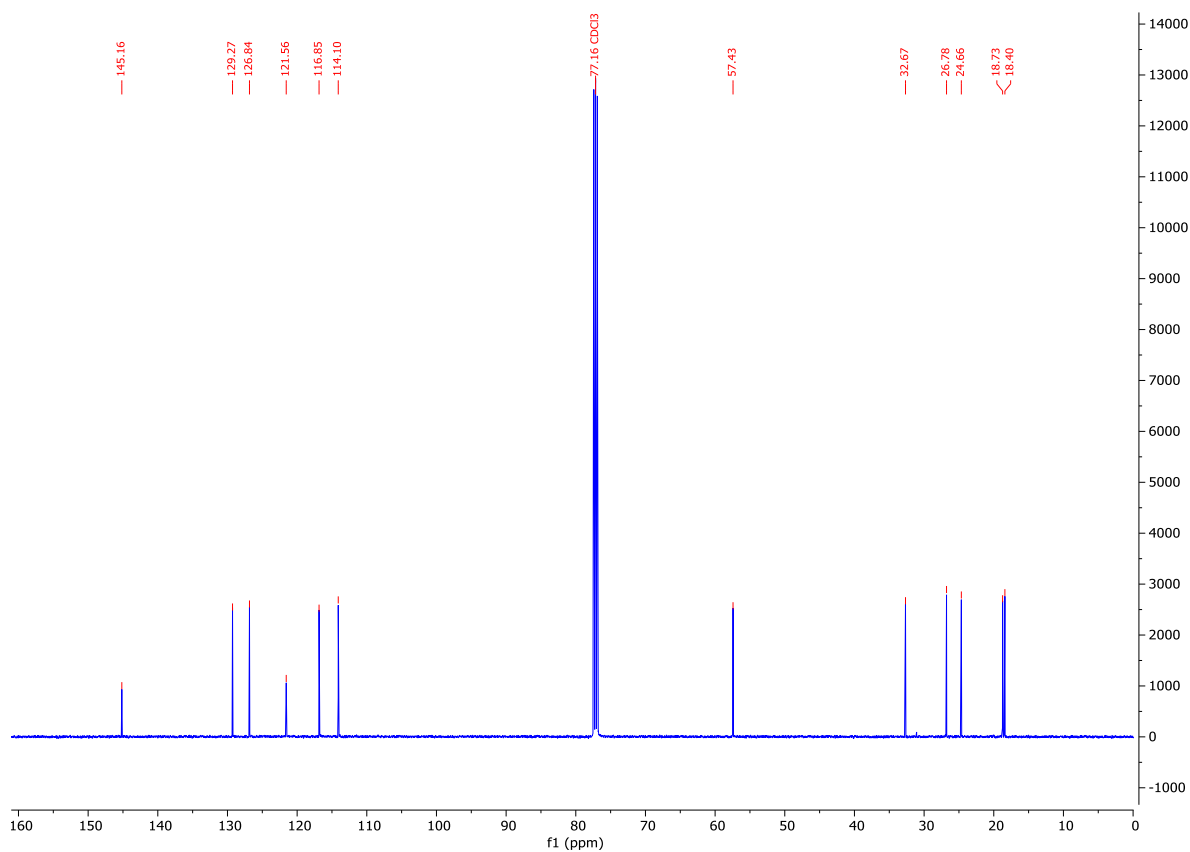


Chiral HPLC Chromatograms of 11e

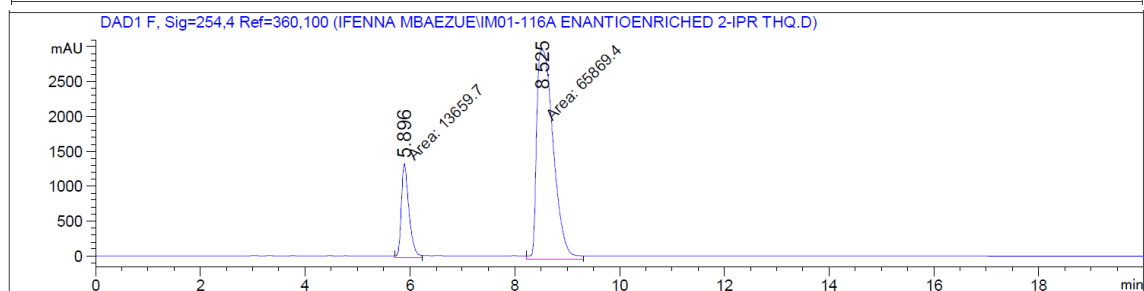
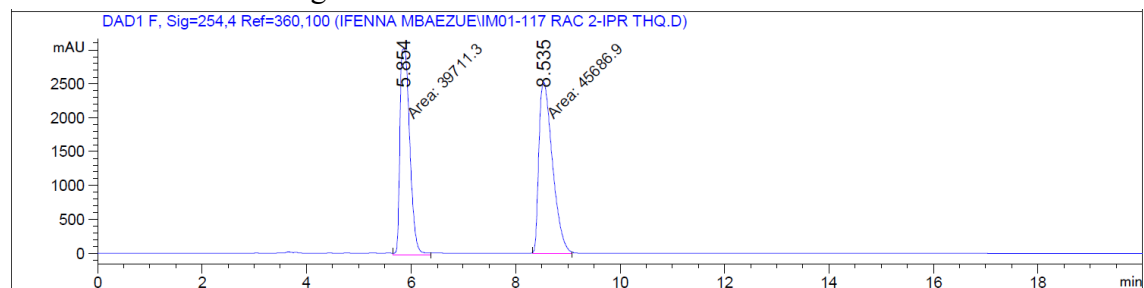


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.562	MM	0.1823	2201.67993	201.33543	12.6316
2	7.910	MM	0.2248	1.52283e4	1129.25708	87.3684

¹³C NMR of **11f**



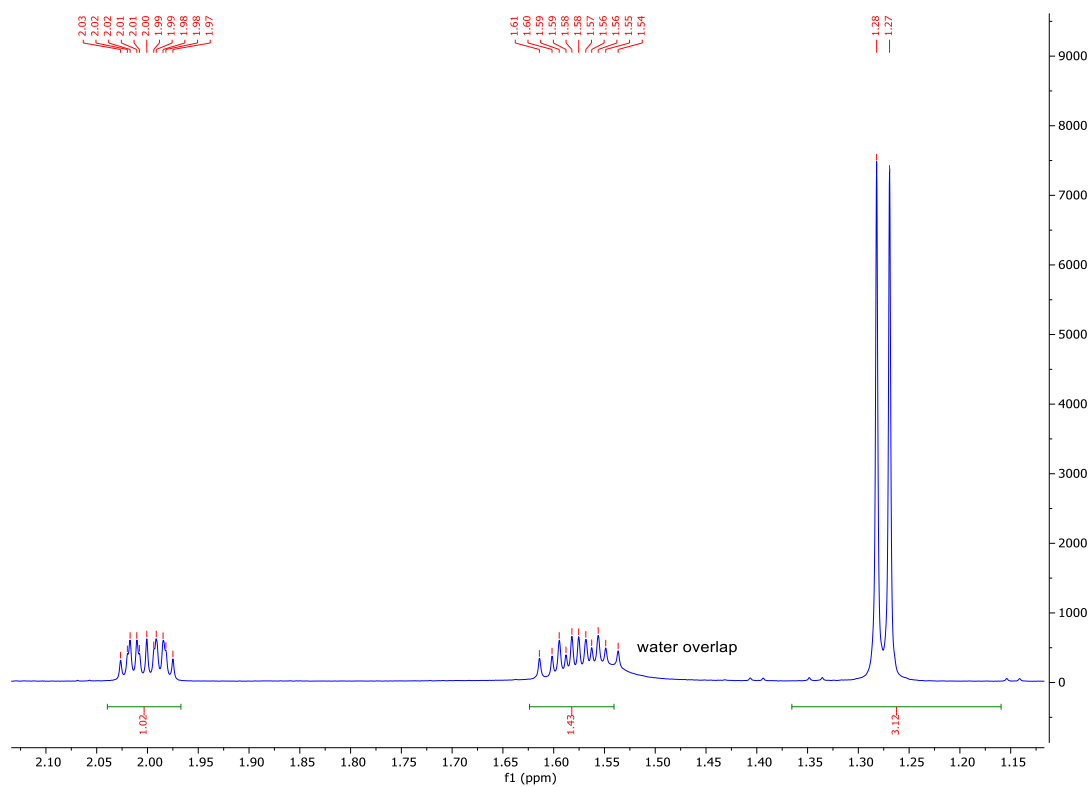
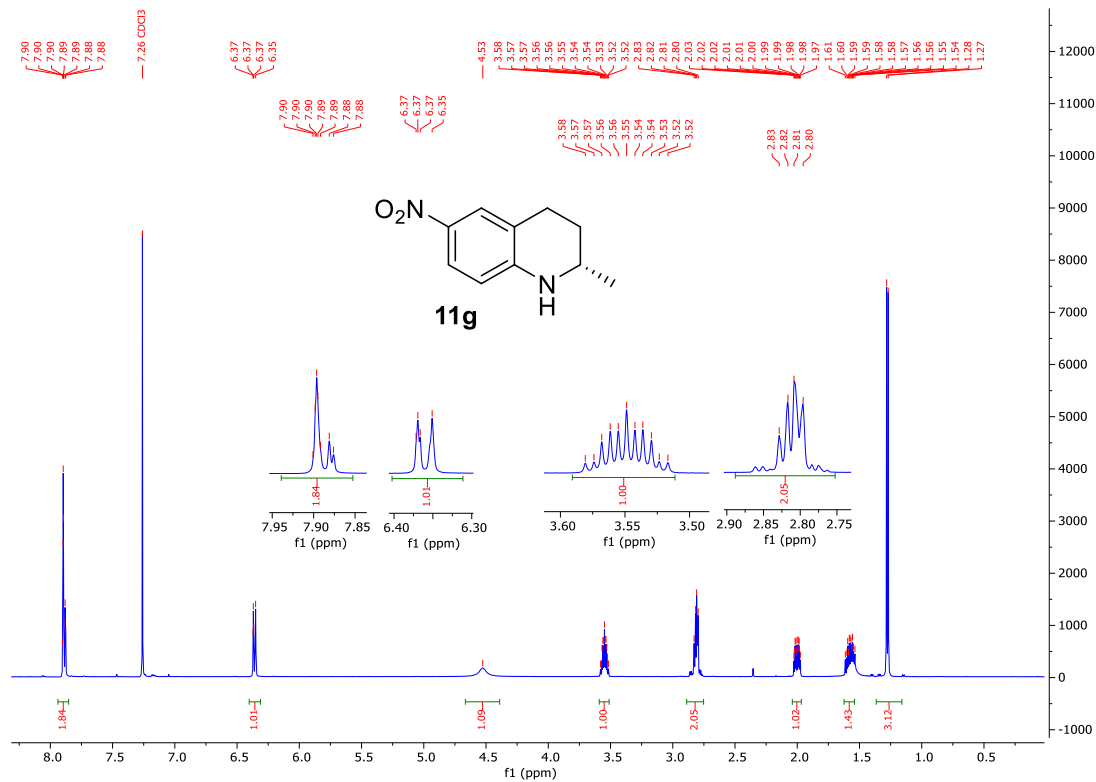
Chiral HPLC Chromatograms of **11f**



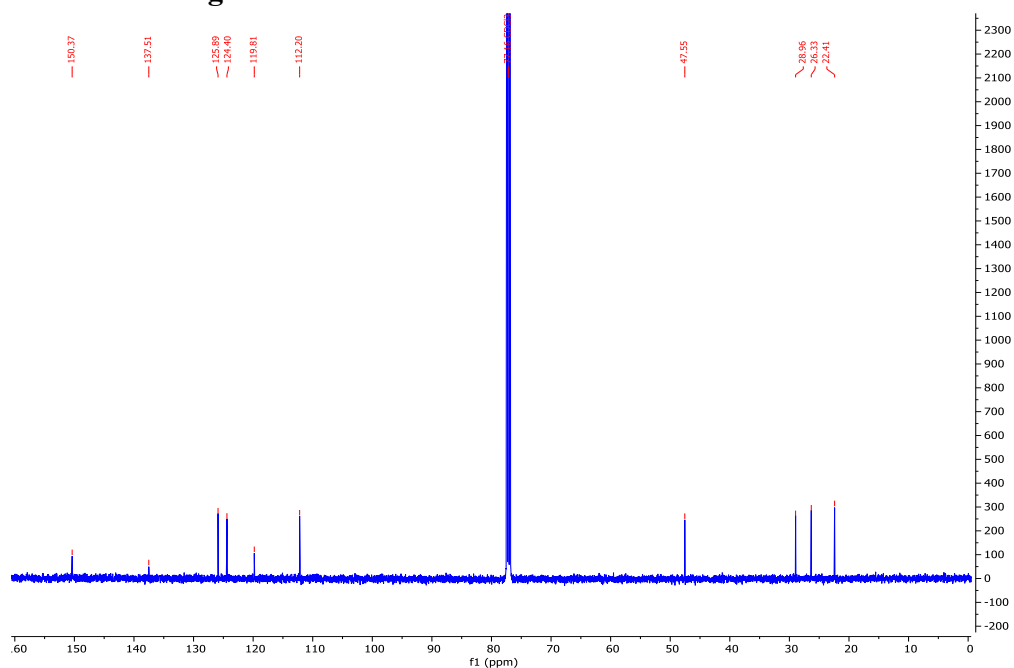
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.896	MM	0.1692	1.36597e4	1345.61450	17.1757
2	8.525	MM	0.3666	6.58694e4	2994.45923	82.8243

(S)-2-methyl-6-nitro-1,2,3,4-tetrahydroquinoline (**11g**)

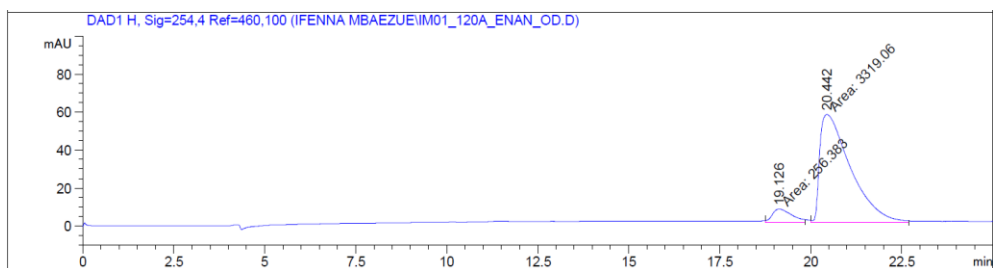
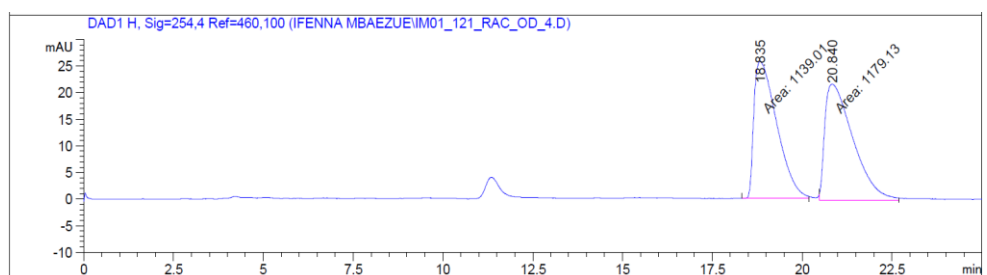
¹H NMR of **11g**



¹³C NMR of **11g**

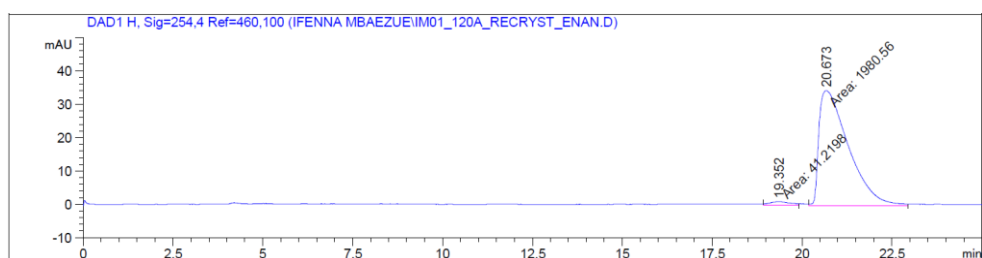
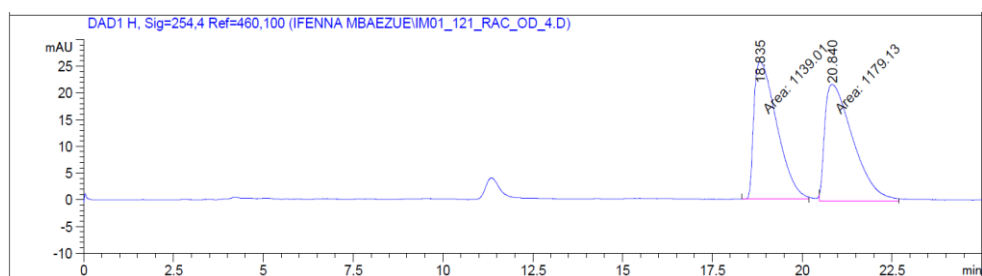


Chiral HPLC Chromatogram of **11g**



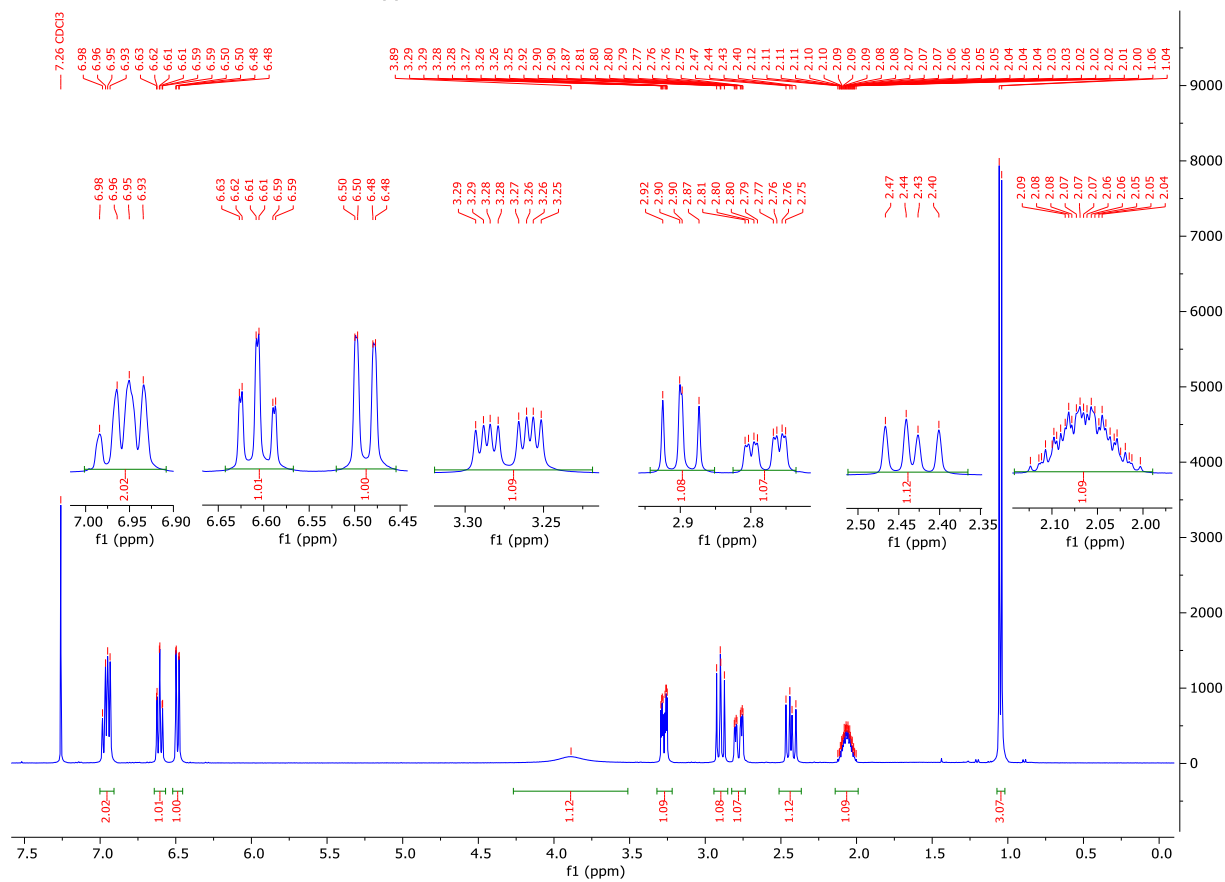
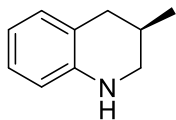
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.126	MM	0.6119	256.38251	6.98332	7.1706
2	20.442	MM	0.9745	3319.06494	56.76318	92.8294

Chiral HPLC Chromatogram of crystallized product **11g**

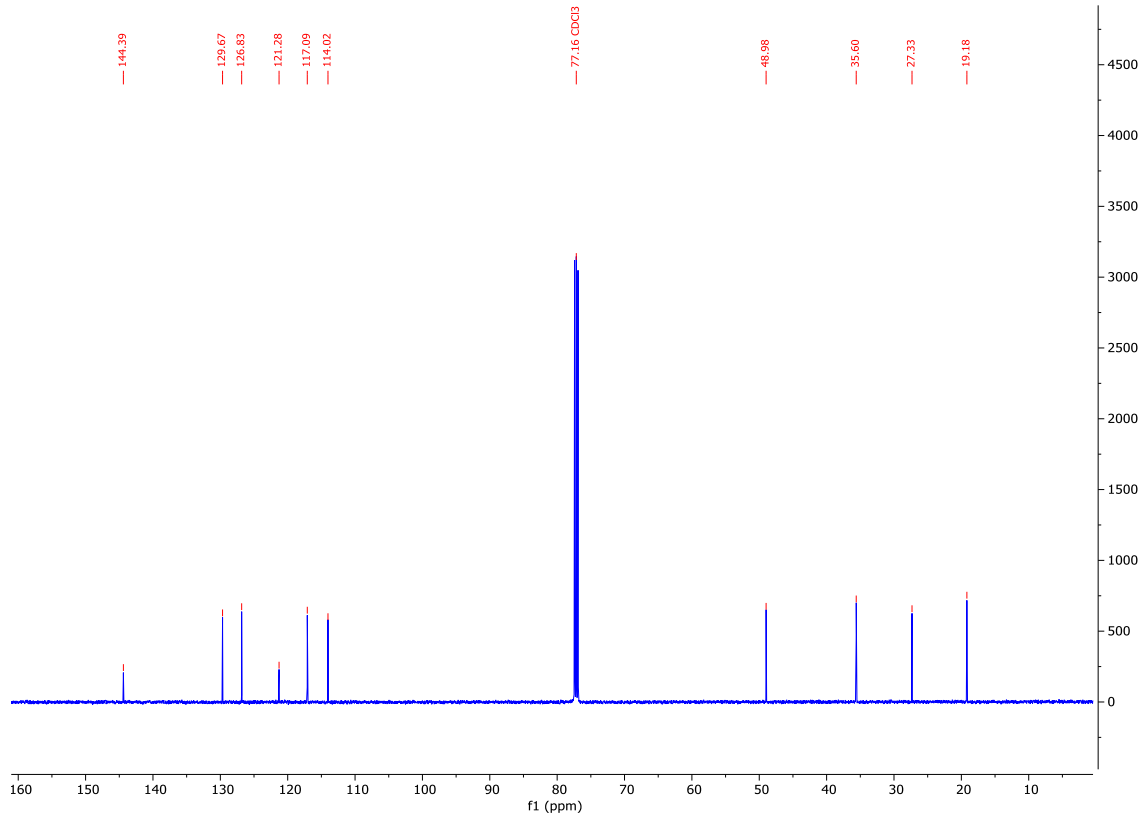


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.352	MM	0.6727	41.21978	1.02125	2.0388
2	20.673	MM	0.9551	1980.56396	34.56263	97.9612

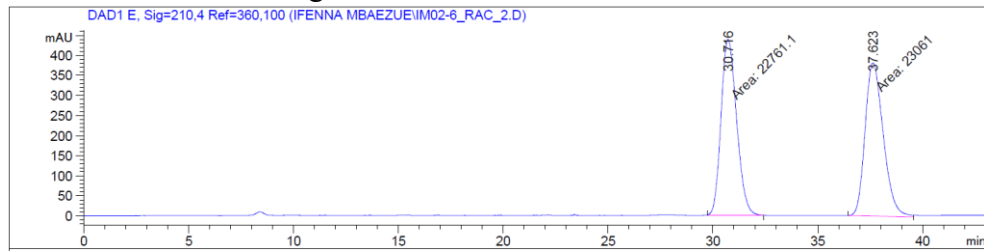
¹H NMR of 11h



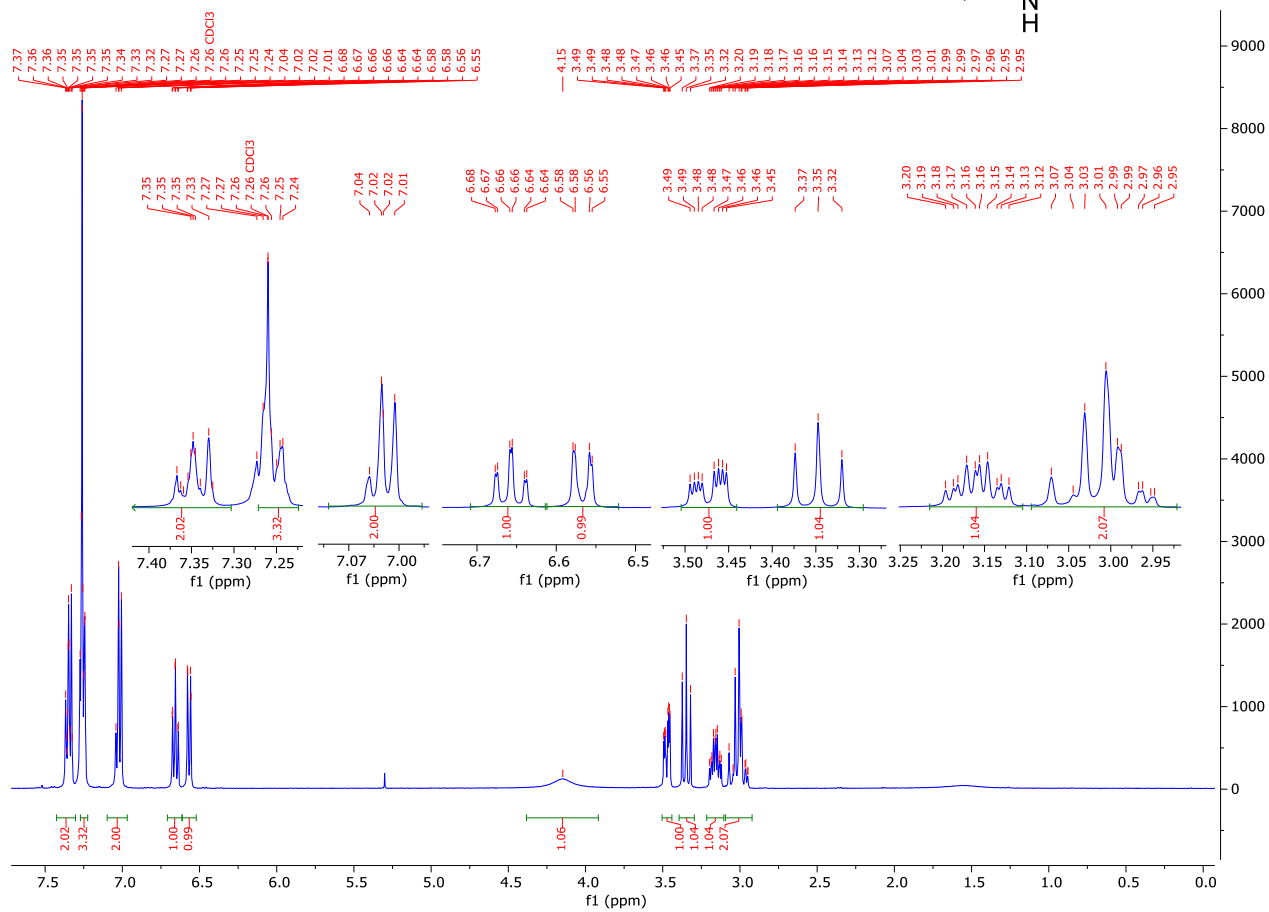
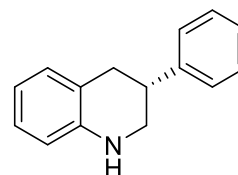
¹³C NMR of **11h**



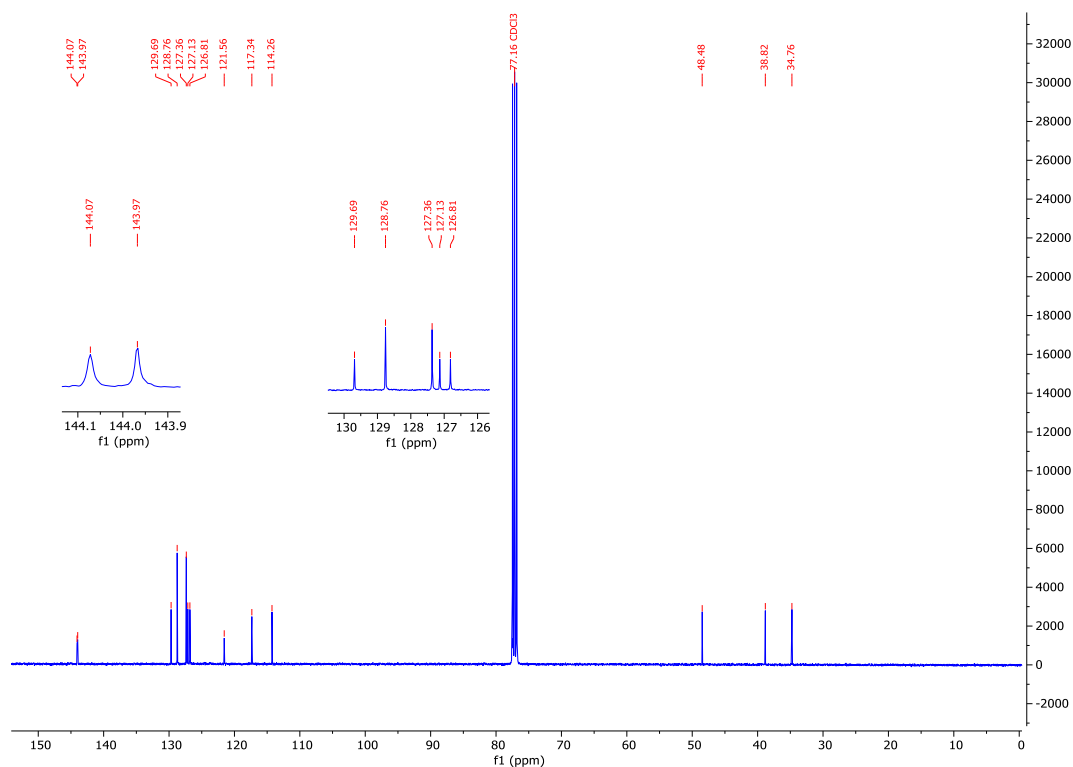
Chiral HPLC Chromatograms of **11h**



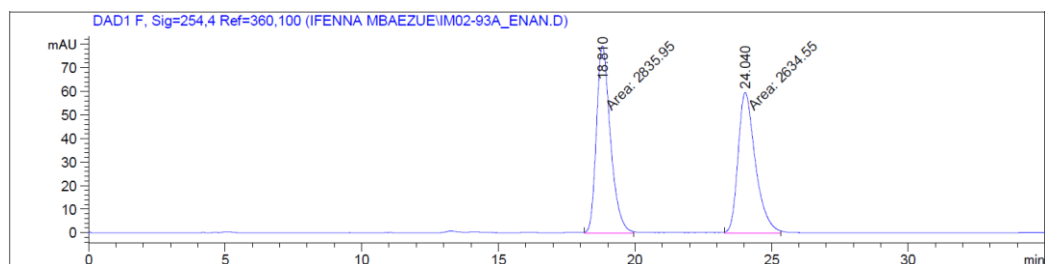
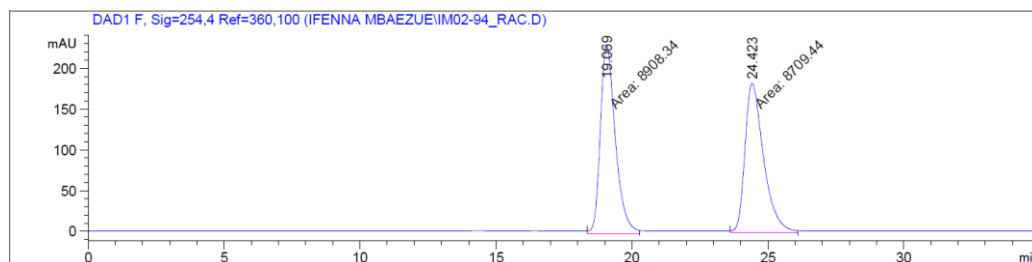
¹H NMR of 11i



¹³C NMR of **11i**



Chiral HPLC Chromatograms of **11i**

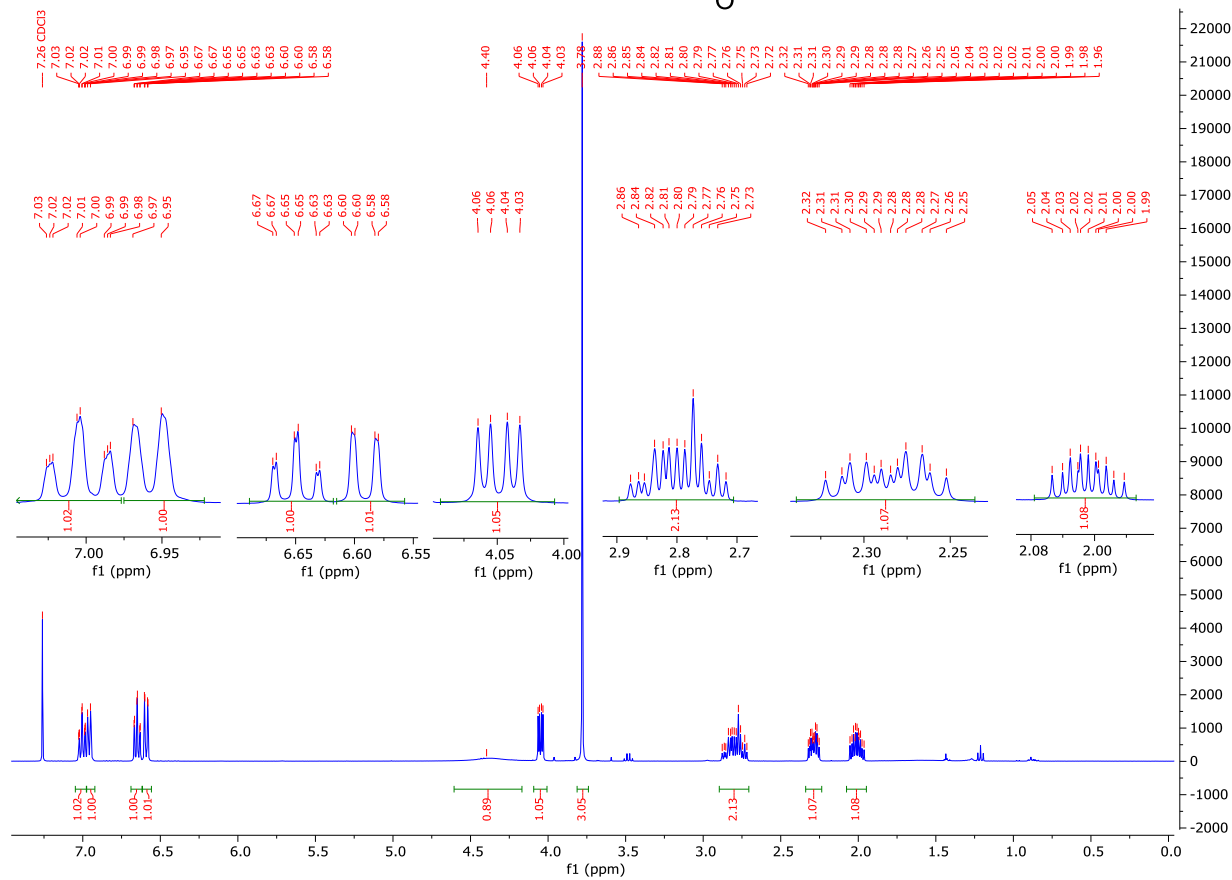
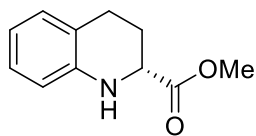


Signal 6: DAD1 F, Sig=254,4 Ref=360,100

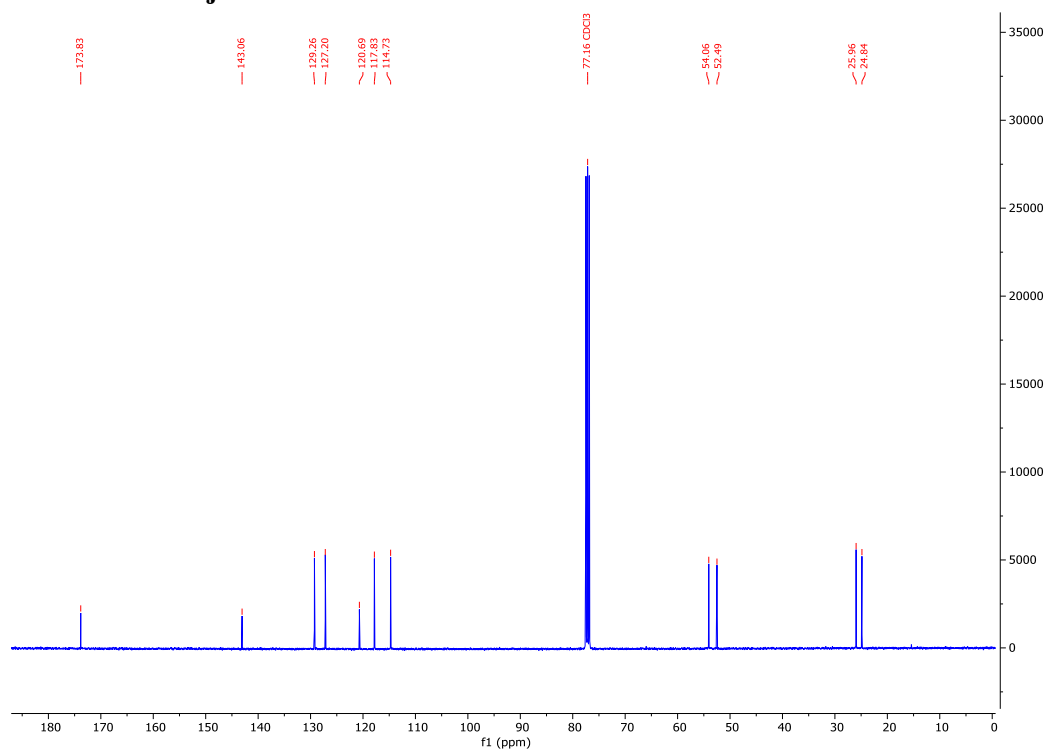
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.810	MM	0.5948	2835.94873	79.46480	51.8408
2	24.040	MM	0.7355	2634.54956	59.70087	48.1592

Totals : 5470.49829 139.16567

¹H NMR of 11j



¹³C NMR of 11j



X-Ray Data Collection and Structural Refinement Statistics

The X-ray data were collected on a Bruker D8 Venture dual-source diffractometer equipped with a PHOTON II detector and an Oxford Cryostream 800 cooling system, using mirror-monochromatized CuK α radiation ($\lambda = 1.54184 \text{ \AA}$) from a microfocus source, in a series of φ - and ω -scans. APEX3 software was used for data collection, integration and reduction.¹⁷ Semi-empirical absorption correction was applied using SADABS-2016/2.¹⁸

The structures were solved using SHELXT-2014/5 (**5a_123K**, **11a_253K**) or SHELXT-2018/2 (**5c_253K**, **8d_253K**)¹⁹ and refined by full-matrix least-squares using SHELXL-2018/3²⁰ within Olex2²¹ and WinGX²² packages. All non-hydrogen atoms were refined anisotropically. All carbon-bound hydrogen atoms were calculated to their optimal positions and treated as riding atoms using isotropic displacement parameters 1.2 (or 1.5 in case of methyl groups) times larger than the respective parent atoms. Nitrogen- and oxygen-bound hydrogen atoms were found in the difference electron density map and were modelled as constrained, with isotropic displacement parameters 1.2 (for nitrogen-bound) or 1.5 (for oxygen-bound) times larger than those of the respective parent atoms. In case of **11a_253K**, the amino group was instead allowed to refine as a rigid body to allow for the partial sp³ character of the nitrogen, i.e. the out-of-plane position of the attached hydrogen atom. For disordered moieties, 1,2- and 1,3-interatomic distances were restrained to be equal and the anisotropic displacement parameters of the atoms were restrained to be equal for bonded and spatially close atoms. In case of **5c_253K**, the minor disorder component of the 2,4,6-triisopropylbenzenesulfonyl group was partially refined as a rigid body including the benzene ring with the attached secondary carbon atoms of isopropyl groups and the sulfonyl ($-\text{SO}_2-$) group. The occupancies of the disordered moieties were either allowed to refine freely (**5c_253K**) or were fixed to 0.5 as required by the proximity of a two-fold rotation axis (**8d_253K**).

CCDC 1935843–1935846 contain the supplementary crystallographic data for this paper. The data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/structures.

Table 3. Crystallographic data

Complex	5a 123K	5c 253K	8d 253K	11a 253K
CCDC Number	1935843	1935844	1935845	1935846
Empirical formula	C ₂₅ H ₃₈ NO ₄ PS	C ₃₀ H ₄₄ NO ₅ PS	C ₃₁ H ₄₂ Cl ₂ N ₂ O ₄ P ₂	C ₁₀ H ₁₂ BrN
Formula weight	479.59	561.69	639.50	226.12
<i>T</i> /K	123.0(1)	253.0(1)	253.0(1)	253.0(1)
Crystal system	Monoclinic	Orthorhombic	Tetragonal	Orthorhombic
Space group	<i>P</i> 2 ₁	<i>P</i> 2 ₁ 2 ₁ 2 ₁	<i>P</i> 4 ₁ 2 ₁ 2	<i>P</i> 2 ₁ 2 ₁ 2 ₁
<i>a</i> /Å	10.6790(7)	10.5840(6)	10.7127(2)	16.0321(5)
<i>b</i> /Å	12.0178(8)	15.7917(8)	10.7127(2)	6.1532(2)
<i>c</i> /Å	11.2241(7)	18.4493(10)	27.9476(6)	9.7828(3)
α /°	90	90	90	90
β /°	112.3257(14)	90	90	90
γ /°	90	90	90	90
<i>V</i> /Å ³	1332.50(15)	3083.6(3)	3207.32(14)	965.06(5)
<i>Z</i>	2	4	4	4
ρ_{calc} /g cm ⁻³	1.195	1.210	1.324	1.556
μ /mm ⁻¹	1.878	1.721	3.070	5.338
Max. and min. transmission	0.7543 and 0.5780	0.7536 and 0.6107	0.7536 and 0.6332	0.7543 and 0.5764
<i>F</i> (000)	516.0	1208.0	1352.0	456.0
Crystal color and shape	colorless, prism	yellow, lath	colorless, prism	colorless, block
Crystal size/mm ³	0.655×0.634×0.502	0.644×0.628×0.384	0.314×0.283×0.221	0.337×0.312×0.294
2 θ range for data collection/°	8.952 to 161.056	7.368 to 145.612	8.84 to 144.914	10.594 to 161.068
Index ranges	-13 ≤ <i>h</i> ≤ 13, -15 ≤ <i>k</i> ≤ 14, -14 ≤ <i>l</i> ≤ 14	-13 ≤ <i>h</i> ≤ 13, -19 ≤ <i>k</i> ≤ 19, -22 ≤ <i>l</i> ≤ 22	-13 ≤ <i>h</i> ≤ 13, -13 ≤ <i>k</i> ≤ 13, -34 ≤ <i>l</i> ≤ 32	-20 ≤ <i>h</i> ≤ 20, -7 ≤ <i>k</i> ≤ 7, -12 ≤ <i>l</i> ≤ 12
Reflections collected	31499	60068	45989	25154
Reflections [<i>R</i> _{int}]	5643 [0.0313]	6006 [0.0411]	3167 [0.0835]	2073 [0.0457]
Data completeness (%)	99.5 to 2 θ = 135.500°	97.6 to 2 θ = 135.358°	99.2 to 2 θ = 135.500°	97.6 to 2 θ = 135.358°
Data/restraints/parameters	5643/1/300	6006/981/483	3167/20/208	2073/0/114
Goodness-of-fit on <i>F</i> ²	1.034	1.137	1.054	1.145
Final <i>R</i> indices [<i>I</i> > 2 σ (<i>I</i>)]	<i>R</i> _{<i>I</i>} = 0.0271, <i>wR</i> ₂ = 0.0709	<i>R</i> _{<i>I</i>} = 0.0437, <i>wR</i> ₂ = 0.1133	<i>R</i> _{<i>I</i>} = 0.0278, <i>wR</i> ₂ = 0.0741	<i>R</i> _{<i>I</i>} = 0.0656, <i>wR</i> ₂ = 0.1579
Final <i>R</i> indices [all data]	<i>R</i> _{<i>I</i>} = 0.0273, <i>wR</i> ₂ = 0.0711	<i>R</i> _{<i>I</i>} = 0.0440, <i>wR</i> ₂ = 0.1137	<i>R</i> _{<i>I</i>} = 0.0280, <i>wR</i> ₂ = 0.0744	<i>R</i> _{<i>I</i>} = 0.0662, <i>wR</i> ₂ = 0.1581
Largest diff. peak/hole (<i>e</i> Å ⁻³)	0.346/-0.258	0.477/-0.282	0.253/-0.190	0.475/-0.632
Flack parameter <i>x</i>	0.014(9)	0.029(4)	-0.007(5)	0.050(18)
Extinction coefficient	0.0090(10)	0.0130(15)	-	0.014(2)

References

1. Qian, B.; Guo, S.; Shao, J.; Zhu, Q.; Yang, L.; Xia, C.; Huang, H. Palladium-Catalyzed Benzylic Addition of 2-Methyl Azaarenes to N-Sulfonyl Aldimines via C–H Bond Activation. *J. Am. Chem. Soc.* **2010**, *132*, 3650–3651.
2. Wen, J.; Tan, R.; Liu, S.; Zhao, Q.; Zhang, X. Strong Brønsted acid promoted asymmetric hydrogenation of isoquinolines and quinolines catalyzed by a Rh–thiourea chiral phosphine complex via anion binding. *Chem. Sci.*, **2016**, *7*, 3047–3051.
3. (a) Matsugi, M.; Tabusa, F.; Minamikawa, J. Doebner-Miller synthesis in a two-phase system: practical preparation of quinolines. *Tetrahedron Lett.* **2000**, *41*, 8523–8525. (b) Tahtaoui, C.; Guillier, F.; Klotz, P.; Galzi, J-L.; Hibert, M.; Ilien, B. On the Use of Nonfluorescent Dye Labeled Ligands in FRET-Based Receptor Binding Studies. *J. Med. Chem.* **2005**, *48*, 7847-7859.
4. Li, S.G.; Yuan, M.; Topic, F.; Han, Z.S.; Senanayake, C.H.; Tsantrizos, Y.S. Asymmetric library synthesis of *P*-chiral *t*-butyl-substituted secondary and tertiary phosphine oxides. *J. Org. Chem.* **2019**, *84*, 7291-7302.
5. Han, Z.S.; Wu, H.; Xu, Y.; Zhang, Y.; Qu, B.; Li, Z.; Caldwell, D.R.; Fandrick, K.R.; Zhang, L.; Roschangar, F.; Song, J.J.; Senanayake, C.H. General and stereoselective method for the synthesis of sterically congested and structurally diverse *P*-stereogenic secondary phosphine oxides. *Org. Lett.* **2017**, *19*, 1796.
6. Jolliffe, J.D.; Armstrong, R.J.; Smith, M.D. Catalytic enantioselective synthesis of atropisomeric biaryl by a cation-directed *O*-alkylation. *Nature Chem.* **2017**, *9*, 558-562.
7. Han, Z. S.; Zhang, L.; Xu, Y.; Sieber, J. D.; Marsini, M. A.; Li, Z.; Reeves, J. T.; Fandrick, K. R.; Patel, N. D.; Desrosiers, J.; Qu, B.; Chen, A.; Rudzinski, D. M.; Samankumara, L. P.; Ma, S.; Grinberg, N.; Roschangar, F.; Yee, N. K.; Wang, G.; Song, J. J.; Senanayake, C. H. Efficient asymmetric synthesis of structurally diverse *P*-stereogenic phosphinamides for catalyst design. *Angew. Chem. Int. Ed.* **2015**, *54*, 5474-5477.
8. Han, Z.S.; Wu, H.; Qu, B.; Wang, Y.; Wu, L.; Zhang, L.; Xu, Y.; Wu, L.; Zhang, Y.; Lee, H.; Roschangar, F.; Song, J.J.; Senanayake, C.H. New class of *P*-stereogenic chiral Brønsted acids catalysts derived from chiral phosphinamides. *Tetrahedron Lett.* **2019**, *60*, 1834-1837.
9. Yang, T.; Yin, Q.; Gu, G.; Zhang, X. A one-pot process for the enantioselective synthesis of tetrahydroquinolines and tetrahydroisoquinolines via asymmetric reductive amination (ARA). *Chem. Commun.* **2018**, *54*, 7247-7250.
10. Wang, C.; Li, C.; Wu, X.; Pettman A.; Xiao, J. pH-Regulated Asymmetric Transfer Hydrogenation of Quinolines in Water. *Angew. Chem. Int. Ed.* **2009**, *48*, 6524–6528.

-
11. Rueping, M.; Theissmann, T.; Stoeckel, M.; Antonchick, A. P. Direct enantioselective access to 4-substituted tetrahydroquinolines by catalytic asymmetric transfer hydrogenation of quinolines *Org. Biomol. Chem.* **2011**, *9*, 6844–6850
12. Gou, F-R.; Zhang, X.; Liang, Y-M. Iridium-Catalyzed Asymmetric Hydrogenation of Quinoline Derivatives with C3*-TunePhos. *Adv. Synth. Catal.* **2010**, *352*, 2441-2444.
13. Zhao, X.; Xiao, J.; Tang, W. Enantioselective reduction of 3-substituted quinolones with a cyclopentadiene-based chiral Brønsted acid. *Synthesis*, **2017**, *49*, 3157–3164.
14. Batsyts, S.; Vedmid, R.; Namyslo, J. C.; Nieger, M.; Schmidt, A. 3-Aryl substituted 1-methylquinolinium salts as carbene precursors. *Eur. J. Org. Chem.*, **2019**, 1301–1310.
15. Rueping, M.; Theissmann, T.; Raja, S.; Bats, J. W. Asymmetric counterion pair catalysis: An enantioselective Brønsted acid-catalyzed protonation. *Adv. Synth. Catal.* **2008**, *350*, 1001-1006.
16. Takamura, M.; Funabashi, K.; Kanai, M.; Shibasaki, M. *J. Am. Chem. Soc.* **2000**, *122*, 6327-6328.
17. Bruker. APEX3. Bruker AXS Inc., Madison, Wisconsin, USA, 2012.
18. Krause, L.; Herbst-Irmer, R.; Sheldrick, G.M.; Stalke, D. Comparison of silver and molybdenum microfocus X-ray sources for single-crystal structure determination. *J. Appl. Cryst.* **2015**, *48*, 3.
19. Sheldrick, G. M. *SHELXT* – Integrated space-group and crystal-structure determination. *Acta Cryst.* **2015**, *A71*, 3.
20. Sheldrick, G. M. Crystal structure refinement with SHELXL. *Acta Cryst.* **2015**, *C71*, 3.
21. Dolomanov, O.V.; Bourhis, L.J.; Gildea, R.J.; Howard, J. A. K.; Puschmann, H. *OLEX2*: a complete structure solution, refinement and analysis program. *J. Appl. Cryst.* **2009**, *42*, 339.
22. Farrugia, L. J. *WinGX* and *ORTEP* for Windows: an update. *J. Appl. Cryst.* **2012**, *45*, 849.