

## Supporting Information

### **A Chiral-at-Metal Asymmetric Catalyzed Vinylogous Michael Addition of Ortho-Methyl Aromatic Nitro Compounds for Isoxazole Derivatives Synthesis**

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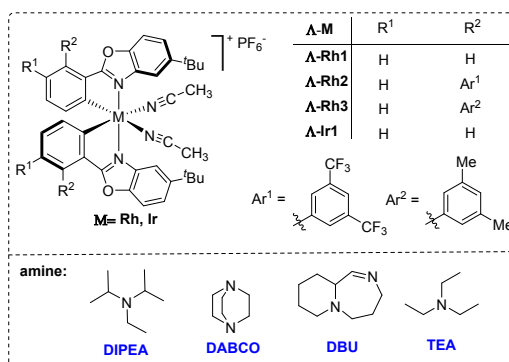
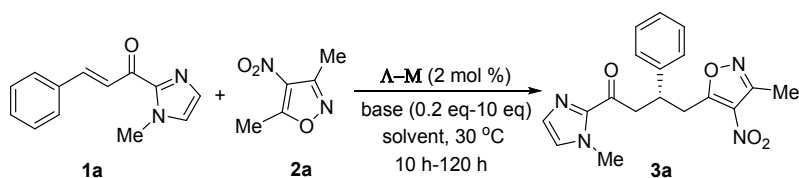
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## I General Information

All reactions were performed in Schlenk tubes at 30 °C using oven-dried glassware. Commercially obtained reagents were used without further purification, unless otherwise noted. Dry 1,2-dichloroethane (DCE) was obtained from solvent distillation machine (Vigor VSPS-5) and stored under argon over 4 Å type molecular sieves. THF and toluene were distilled freshly before use over sodium and benzophenone. Dichloromethane (DCM) was distilled from CaH<sub>2</sub>. Methanol was used without further purification. Reactions were checked by TLC analysis and plates were visualized with short-wave UV light (254 nm). The <sup>1</sup>H and <sup>13</sup>C NMR spectra were obtained in CDCl<sub>3</sub> using a Bruker-BioSpin AVANCE III HD NMR spectrometer at 400 and 100 MHz, respectively. Chemical shifts are reported in parts per million (δ value) calibrated against the residual solvent peak. Signal patterns are indicated as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet. Coupling constants (*J*) are given in hertz (Hz). HPLC analyses of the compounds were performed on chiralcel IA-IF columns and chiralcel AD-H, AS-H, OJ-H and OD-H columns using hexane and isopropanol as eluent. The infrared spectra were recorded on a Bruker VERTEX 70 IR spectrometer as KBr pellets, with absorption reported in cm<sup>-1</sup>. High-resolution mass spectra were recorded on a Bruker Impact II UHR TOF LC/MS Mass Spectrometry.

## II Optimization of Reaction Conditions

**Table 1. Optimization of the reaction conditions<sup>a</sup>**



entry	$\Delta$ -M (x mol %)	base (y equiv.)	solvent	time (h)	yield (%) <sup>b</sup>	ee (%) <sup>c</sup>
1	$\Delta$ -Rh1 (2)	DIPEA (0.2)	DCE	96	95	94
2	$\Delta$ -Rh2 (2)	DIPEA (0.2)	DCE	96	90	94
3	$\Delta$ -Rh3 (2)	DIPEA (0.2)	DCE	96	92	79
4	$\Delta$ -Ir1 (2)	DIPEA (0.2)	DCE	120	70	92
5	/	DIPEA (0.2)	DCE	24	NR	NR
6	$\Delta$ -Rh1 (2)	/	DCE	24	trace	trace
7	$\Delta$ -Rh1 (2)	DIPEA (0.2)	DCM	96	75	93
8	$\Delta$ -Rh1 (2)	DIPEA (0.2)	THF	96	86	90
9	$\Delta$ -Rh1(2)	DIPEA (0.2)	CH <sub>3</sub> OH	96	95	88
10	$\Delta$ -Rh1(2)	DIPEA (0.2)	toluene	96	85	78
11	$\Delta$ -Rh1(2)	DABCO (0.2)	DCE	96	68	93
12	$\Delta$ -Rh1(2)	DBU (0.2)	DCE	96	80	92
13	$\Delta$ -Rh1(2)	TEA (0.2)	DCE	96	90	94
15 <sup>d</sup>	$\Delta$ -Rh1(2)	DIPEA (0.2)	DCE	40	92	94
16 <sup>d</sup>	$\Delta$ -Rh1(2)	DIPEA (0.5)	DCE	18	96	95

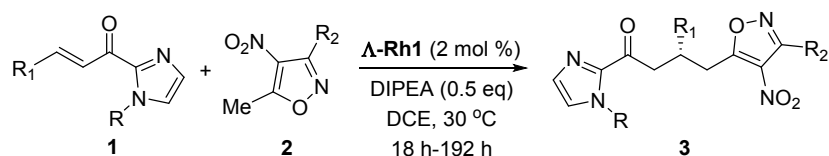
17 <sup>d</sup>	<b>Λ-Rh1</b> (2)	<b>DIPEA</b> (2.0)	DCE	16	95	94
18 <sup>d</sup>	<b>Λ-Rh1</b> (2)	<b>DIPEA</b> (5.0)	DCE	14	90	94
19 <sup>d</sup>	<b>Λ-Rh1</b> (2)	<b>DIPEA</b> (10.0)	DCE	10	93	95
20 <sup>d</sup>	<b>Λ-Rh1</b> (1)	<b>DIPEA</b> (0.5)	DCE	60	92	94

<sup>a</sup>Reaction conditions: **1a** (0.20 mmol), **2a** (0.40 mmol), **Λ-M** (2 mol %), base (0.2 - 10 eq), solvent (0.5-1 mL) at 30 °C under argon atmosphere. <sup>b</sup>Isolated yields. <sup>c</sup>Determined by chiral HPLC. <sup>d</sup>1,2-dichloroethane (0.5 mL). DIPEA = *N,N*-Diisopropylethylamine, DABCO = Triethylenediamine, DBU= 1,8-Diazabicyclo[5.4.0]undec-7-ene, TEA= Triethylamine.

### III Experimental Section

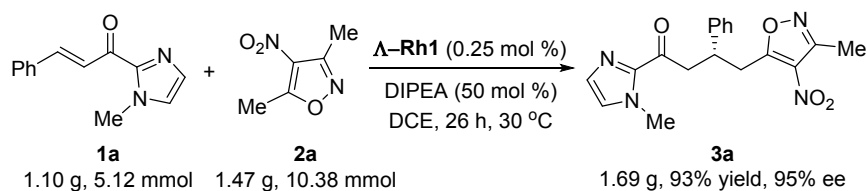
**$\Lambda$ -Rh1,  $\Lambda$ -Rh2,  $\Lambda$ -Rh3 and  $\Lambda$ -Ir1** were prepared according to reported procedure.<sup>1-2</sup>  $\alpha,\beta$ -unsaturated 2-acyl imidazoles<sup>3</sup> and 3,5-disubstituted-4-nitroisoxazole substrates **2a-2f** were synthesized according to reported procedures.<sup>4</sup>

#### General procedure for catalytic asymmetric synthesis of isoxazole analogues.



To an oven-dried 25 mL Schlenk tube equipped with a stir bar,  **$\Lambda$ -Rh1** (2.0 mol %) was added along with  $\alpha,\beta$ -unsaturated 2-acyl imidazole **1** (1 equiv., 0.2 mmol), DIPEA (50 mol %) and DCE (0.5 mL). After being stirred at room temperature for 5 min, 3,5-disubstituted-4-nitroisoxazole substrates **2** (2.0 equiv., 0.40 mmol) was added. The reaction was stirring at 30 °C until consumption of the 2-acyl imidazole (monitored by TLC). The reaction mixture was concentrated and directly purified by silica gel column chromatography (with ethyl acetate-petroleum ether as the eluent) to afford the desired products **3** or **4**.

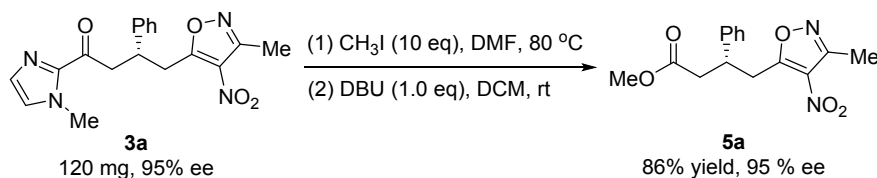
#### General procedure for gram-scale experiments with lower catalyst loading.



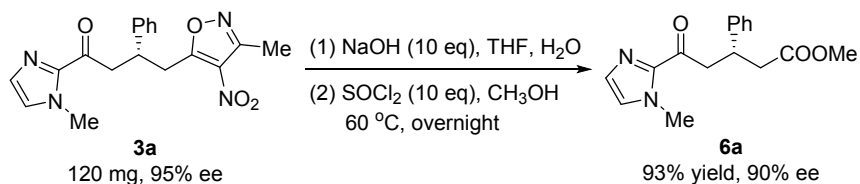
To an oven-dried 50 mL Schlenk tube equipped with a stir bar,  **$\Lambda$ -Rh1** (0.25 mol%) was added along with  $\alpha,\beta$ -unsaturated 2-acyl imidazole **1a** (1.10 g, 5.12 mmol, 1.0 equiv.), DIPEA (50 mol %) and DCE (6 mL). After being stirred at room temperature for 5 min, 3,5-dimethyl-substituted 4-nitroisoxazole **2a** (1.47 g, 10.38 mmol, 2.0 equiv.) was added. The reaction was stirring at room temperature until consumption of the 2-acyl imidazole as monitored by TLC for 26 h. The reaction mixture was

concentrated and directly purified by silica gel column chromatography (EtOAc/Petroleum ether = 1:4-1:3) to afford **3a** (white solid, 1.69 g, 93% yield, 95% ee).

### General procedure for synthetic transformation.



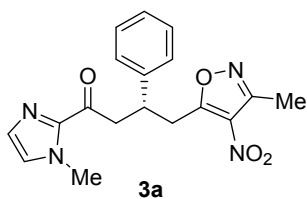
A solution of **3a** (120 mg, 0.34 mmol, 1 equiv.) and methyl iodide (10 equiv.) in DMF (2 mL) was stirred in a sealed tube at 80 °C overnight, after **3a** was fully converted to salt (monitored by TLC), the reaction was cooled down to room temperature. The reaction mixture was concentrated and CH<sub>3</sub>OH (2 mL) was added to the mixture, and then DBU (10 equiv.) was added to the solution. The resulting mixture was stirred for 12 hours at room temperature before it was quenched with NH<sub>4</sub>Cl (20 mL, sat. aq.), the aqueous layer was extracted with EtOAc (3 × 20 mL). The combined organic layers were washed with brine (20 mL), dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated in vacuo. Flash column chromatography (silica gel) afforded the desired product **5a** (89 mg, 86% yield, 95% ee).



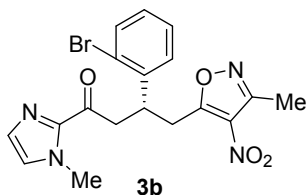
To the compound **3a** (120 mg, 0.34 mmol, 1 equiv.) in 5 mL solvent (5:1 THF/H<sub>2</sub>O) was added NaOH (10 equiv.) at 0 °C. The resulting mixture was refluxed for 3 h. The reaction mixture was quenched with 4 M aqueous HCl and the solvent was removed under reduced pressure. The remaining precipitation dissolves in methanol, and then 5 mL SOCl<sub>2</sub> was added to the above reaction solution. The reaction was stirred at 60 °C overnight. After evaporation of solvent, dissolved the crude product in 20 mL

saturated sodium carbonate solution and 30 mL ethyl acetate, then the reaction was stirred under the room temperature for 120 min. The aqueous layer was extracted three times with ethyl acetate (20 mL  $\times$  3). The combined organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>. The volatile solvent was removed under reduced pressure at last, the product **6a** was obtained after purified by flash column chromatography on silica gel (90 mg, 93% yield, 90% ee) as light yellow oil.

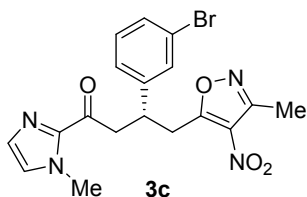




White solid, m.p. 119.1-119.5 °C, 68 mg, 96% yield, 95% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 9.50 min, tr (minor) = 10.88 min);  $[\alpha]_D^{25} = +16.8$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.26 (d, *J* = 4.3 Hz, 4H), 7.20-7.18 (m, 1H), 7.13 (s, 1H), 7.01 (s, 1H), 4.01-3.92 (m, 1H), 3.91 (s, 3H), 3.74-3.62 (m, 2H), 3.58-3.50 (m, 2H), 2.47 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 189.9, 173.2, 155.4, 142.8, 141.8, 130.4, 129.2, 128.7, 127.3, 127.2, 44.5, 38.9, 36.2, 34.2, 11.7. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2921, 2851, 2359, 1676, 1603, 1519, 1415, 1379, 1365, 1289, 1075, 985, 915, 829. HRMS (ESI, *m/z*) calcd for C<sub>18</sub>H<sub>18</sub>N<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 377.1219, found: 377.1220.

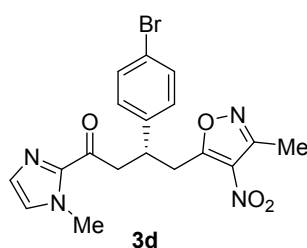


Light yellow oil, 79 mg, 92% yield, 97% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 9.74 min, tr (minor) = 10.93 min);  $[\alpha]_D^{25} = +8.4$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.51 (d, *J* = 8.0 Hz, 1H), 7.34 (d, *J* = 7.7 Hz, 1H), 7.26 (d, *J* = 7.8 Hz, 1H), 7.13 (s, 1H), 7.13-7.03 (m, 1H), 7.03 (s, 1H), 4.58-4.51 (m, 1H), 3.94 (s, 3H), 3.68-3.56 (m, 4H), 2.49 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 189.4, 172.5, 155.4, 142.7, 140.7, 133.3, 130.6, 129.2, 128.7, 127.9, 127.3, 124.5, 43.6, 37.4, 36.2, 32.8, 11.7. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2922, 2852, 2361, 1675, 1603, 1519, 1413, 1379, 1365, 1289, 1155, 1023, 985, 829, 759. HRMS (ESI, *m/z*) calcd for C<sub>18</sub>H<sub>17</sub>BrN<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 455.0324, found: 455.0325.

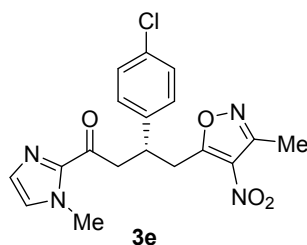


Yellow oil, 75 mg, 87% yield, 95% ee (HPLC: chiralpak IC column, 254 nm,

hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 8.21 min, tr (minor) = 8.93 min);  $[\alpha]_D^{25} = +12.4$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.42 (s, 1H), 7.33 (d, *J* = 7.9 Hz, 1H), 7.22 (d, *J* = 7.7 Hz, 1H), 7.20-7.10 (m, 1H), 7.14 (s, 1H), 7.04 (s, 1H), 4.00-3.95 (m, 1H), 3.93 (s, 3H), 3.71-3.50 (m, 4H), 2.49 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 189.3, 172.6, 155.5, 144.2, 142.5, 130.5, 130.4, 130.3, 129.2, 127.4, 126.0, 122.7, 44.3, 38.5, 36.2, 34.0, 11.6. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2923, 2360, 1676, 1615, 1519, 1413, 1383, 1289, 984. HRMS (ESI, *m/z*) calcd for C<sub>18</sub>H<sub>17</sub>BrN<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 455.0325, found: 455.0325.

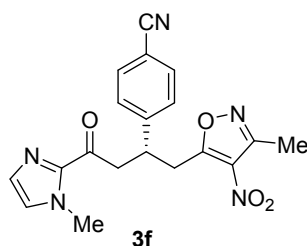


Light yellow oil, 77 mg, 89% yield, 96% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 8.58 min, tr (minor) = 9.40 min);  $[\alpha]_D^{25} = +15.9$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.38 (d, *J* = 8.4 Hz, 2H), 7.16 (d, *J* = 8.4 Hz, 2H), 7.13 (s, 1H), 7.03 (s, 1H), 4.01-3.96(m, 1H), 3.92 (s, 3H), 3.73-3.60 (m, 2H), 3.53-3.46 (m, 2H), 2.49 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 189.5, 172.7, 155.5, 142.7, 140.8, 131.8, 130.4, 129.3, 129.1, 127.4, 121.1, 44.3, 38.3, 36.2, 34.0, 11.7. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2922, 2360, 1674, 1603, 1519, 1414, 1382, 1289, 985, 828. HRMS (ESI, *m/z*) calcd for C<sub>18</sub>H<sub>17</sub>BrN<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 455.0322, found: 455.0325.

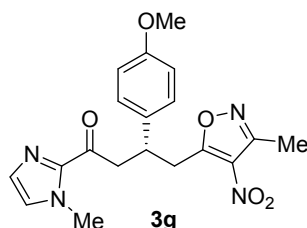


Light yellow oil, 74 mg, 95% yield, 96% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 8.24 min, tr (minor) = 9.05 min);  $[\alpha]_D^{25} = +12.1$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.22 (brs,

4H), 7.13 (s, 1H), 7.03 (s, 1H), 4.03-3.95(m, 1H), 3.92 (s, 3H), 3.73-3.60 (m, 2H), 3.54-3.47 (m, 2H), 2.49 (s, 3H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 100 MHz):  $\delta$  = 189.5, 172.8, 155.5, 142.7, 140.3, 132.9, 130.4, 129.3, 128.9, 128.7, 127.4, 44.4, 38.3, 36.2, 34.1, 11.7. IR (KBr):  $\nu$  ( $\text{cm}^{-1}$ ) 2922, 2360, 2340, 1674, 1603, 1519, 1492, 1415, 1380, 1092, 829. HRMS (ESI,  $m/z$ ) calcd for  $\text{C}_{18}\text{H}_{17}\text{ClN}_4\text{NaO}_4$  [ $\text{M}+\text{Na}$ ] $^+$ : 411.0841, found: 411.0831.

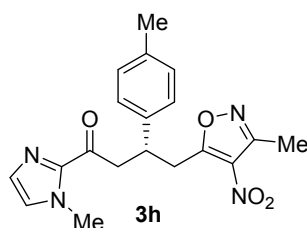


Light yellow oil, 68 mg, 90% yield, 92% ee (HPLC: chiralpak OD-H column, 254 nm, hexane/isopropanol = 70/30, flow rate 1.0 mL/min, tr (major) = 14.08 min, tr (minor) = 16.97 min);  $[\alpha]_{\text{D}}^{25}$  = +7.8 (c = 0.5,  $\text{CHCl}_3$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  = 7.57 (d,  $J$  = 8.1 Hz, 2H), 7.42 (d,  $J$  = 8.1 Hz, 2H), 7.15 (s, 1H), 7.06 (s, 1H), 4.10-4.00 (m, 1H), 3.93 (s, 3H), 3.79-3.73 (m, 1H), 3.69-3.63 (m, 1H), 3.58-3.52 (m, 2H), 2.49 (s, 3H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 100 MHz):  $\delta$  = 188.9, 172.2, 155.6, 147.3, 142.3, 132.6, 130.5, 129.2, 128.3, 127.6, 118.6, 111.3, 44.1, 38.8, 36.2, 33.7, 11.6. IR (KBr):  $\nu$  ( $\text{cm}^{-1}$ ) 2922, 2360, 1672, 1603, 1593, 1517, 1409, 1385, 1366, 702. HRMS (ESI,  $m/z$ ) calcd for  $\text{C}_{19}\text{H}_{17}\text{N}_5\text{NaO}_4$  [ $\text{M}+\text{Na}$ ] $^+$ : 402.1170, found: 402.1173.

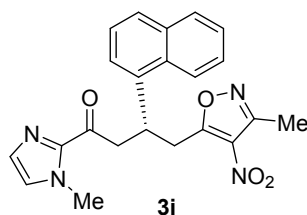


Light yellow oil, 65 mg, 85% yield, 94% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 13.38 min, tr (minor) = 15.38 min);  $[\alpha]_{\text{D}}^{25}$  = +18.0 (c = 0.5,  $\text{CHCl}_3$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  = 7.18 (d,  $J$  = 8.5 Hz, 2H), 7.12 (s, 1H), 7.01 (s, 1H), 6.78 (d,  $J$  = 8.5 Hz, 2H), 3.99-3.95 (m, 1H), 3.91 (s, 3H), 3.74 (s, 3H), 3.71-3.59 (m, 2H), 3.53-3.46 (m, 2H), 2.47 (s, 3H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 100 MHz):  $\delta$  = 190.0, 173.3, 158.5, 155.4, 142.8, 133.8, 130.4,

129.2, 128.3, 127.2, 114.0, 55.2, 44.7, 38.2, 36.1, 34.4, 11.7. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2923, 2360, 2341, 1672, 1604, 1514, 1410, 1382, 1249, 985, 829. HRMS (ESI,  $m/z$ ) calcd for C<sub>19</sub>H<sub>20</sub>N<sub>4</sub>NaO<sub>5</sub> [M+Na]<sup>+</sup>: 407.1326, found: 407.1326.

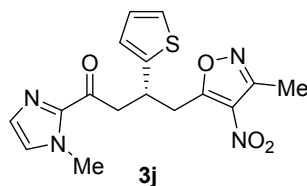


Light yellow oil, 66 mg, 90% yield, 94% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 9.90 min, tr (minor) = 10.99 min);  $[\alpha]_D^{25} = +14.3$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.15 (d,  $J$  = 7.9 Hz, 2H), 7.12 (s, 1H), 7.06 (d,  $J$  = 7.9 Hz, 2H), 7.01 (s, 1H), 3.98-3.94 (m, 1H), 3.91 (s, 3H), 3.71-3.59 (m, 2H), 3.56-3.47 (m, 2H), 2.47 (s, 3H), 2.27 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 190.0, 173.3, 155.4, 142.8, 138.8, 136.8, 130.4, 129.4, 129.2, 127.2, 127.1, 44.6, 38.5, 36.2, 34.3, 21.1, 11.7. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2922, 2360, 2341, 1673, 1603, 1518, 1492, 1410, 1379, 1365, 985, 829. HRMS (ESI,  $m/z$ ) calcd for C<sub>19</sub>H<sub>20</sub>N<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 391.1376, found: 391.1377.

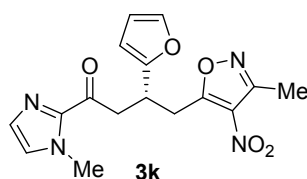


White solid, 75 mg, m.p. 130.1-130.5 °C, 93% yield, 96% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 9.98 min, tr (minor) = 10.92 min);  $[\alpha]_D^{25} = +13.2$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 8.24 (d,  $J$  = 8.5 Hz, 1H), 7.83 (d,  $J$  = 8.0 Hz, 1H), 7.72 (d,  $J$  = 8.1 Hz, 1H), 7.56-7.39 (m, 4H), 7.13 (s, 1H), 6.99 (s, 1H), 4.98 (brs, 1H), 3.85 (s, 3H), 3.83-3.60 (m, 4H), 2.41 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 190.0, 173.0, 155.4, 142.7, 138.1, 133.9, 131.1, 130.5, 129.3, 129.0, 127.8, 127.3, 126.5, 125.7, 125.5, 123.5, 122.6, 44.5, 36.1, 33.7, 32.8, 11.6. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2921, 2851, 2361, 2341,

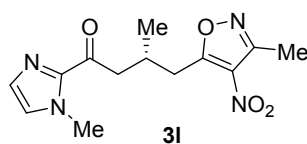
1672, 1601, 1518, 1409, 1155, 829, 778. HRMS (ESI,  $m/z$ ) calcd for  $C_{22}H_{20}N_4NaO_4$   $[M+Na]^+$ : 427.1377, found: 427.1377.



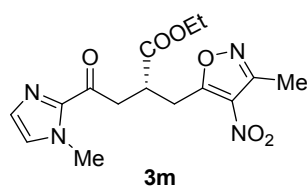
Light yellow oil, 66 mg, 91% yield, 95% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min,  $t_r$  (major) = 11.20 min,  $t_r$  (minor) = 12.22 min);  $[\alpha]_D^{25} = +16.7$  ( $c = 0.5$ ,  $CHCl_3$ ).  $^1H$  NMR (400 MHz,  $CDCl_3$ ):  $\delta = 7.18$ -7.11 (m, 1H), 7.13 (s, 1H), 7.04 (s, 1H), 6.86 (t,  $J = 3.5$  Hz, 2H), 4.37-4.30 (m, 1H), 3.95 (s, 3H), 3.77-3.65 (m, 2H), 3.60-3.54 (m, 2H), 2.50 (s, 3H).  $^{13}C$  NMR ( $CDCl_3$ , 100 MHz):  $\delta = 189.3$ , 172.6, 155.5, 145.2, 142.6, 130.6, 129.3, 127.4, 126.8, 124.5, 124.1, 45.4, 36.2, 35.1, 34.2, 11.7. IR (KBr):  $\nu$  ( $cm^{-1}$ ) 2361, 2341, 1675, 1603, 1519, 1414, 1378, 981, 915, 829, 703. HRMS (ESI,  $m/z$ ) calcd for  $C_{16}H_{16}N_4NaO_4S$   $[M+Na]^+$ : 383.0785, found: 383.0784.



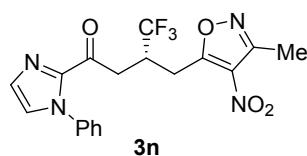
Light yellow oil, 60 mg, 87% yield, 93% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 87/13, flow rate 1.0 mL/min,  $t_r$  (major) = 16.37 min,  $t_r$  (minor) = 17.80 min);  $[\alpha]_D^{25} = +27.6$  ( $c = 0.5$ ,  $CHCl_3$ ).  $^1H$  NMR (400 MHz,  $CDCl_3$ ):  $\delta = 7.29$  (s, 1H), 7.14 (s, 1H), 7.05 (s, 1H), 6.22 (m, 1H), 6.06 (d,  $J = 3.2$  Hz, 1H), 4.14-4.06 (m, 1H), 3.97 (s, 3H), 3.73-3.64 (m, 2H), 3.58-3.47 (m, 2H), 2.52 (s, 3H).  $^{13}C$  NMR ( $CDCl_3$ , 100 MHz):  $\delta = 189.4$ , 172.7, 155.5, 154.6, 142.6, 141.9, 130.5, 129.3, 127.3, 110.2, 106.0, 42.1, 36.2, 32.5, 31.9, 11.7. IR (KBr):  $\nu$  ( $cm^{-1}$ ) 2922, 2361, 2341, 1675, 1604, 1519, 1415, 1380, 1151, 915, 830. HRMS (ESI,  $m/z$ ) calcd for  $C_{16}H_{16}N_4NaO_5$   $[M+Na]^+$ : 367.1013, found: 367.1013.



Light yellow oil, 47 mg, 80% yield, 94% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 94/6, flow rate 1.0 mL/min, tr (major) = 21.72 min, tr (minor) = 23.38 min);  $[\alpha]_D^{25} = +9.2$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.13 (s, 1H), 7.05 (s, 1H), 4.00 (s, 3H), 3.32-3.12 (m, 4H), 2.84-2.76 (m, 1H), 2.55 (s, 3H), 1.10 (d,  $J$  = 6.8 Hz, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 191.0, 174.0, 155.6, 142.9, 130.4, 129.0, 127.2, 45.2, 36.3, 33.9, 28.6, 20.3, 11.7. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2360, 2341, 1690, 1609, 1521, 1418, 1382, 1169, 829. HRMS (ESI,  $m/z$ ) calcd for C<sub>13</sub>H<sub>16</sub>N<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 315.1062, found: 315.1064.

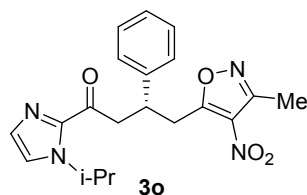


Light yellow oil, 62 mg, 89% yield, 93% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 75/25, flow rate 1.0 mL/min, tr (major) = 17.74 min, tr (minor) = 22.34 min);  $[\alpha]_D^{25} = +5.6$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.08 (s, 1H), 6.99 (s, 1H), 4.08 (q,  $J$  = 7.1 Hz, 2H), 3.92 (s, 3H), 3.68-3.58 (m, 2H), 3.53-3.35 (m, 3H), 2.49 (s, 3H), 1.13 (t,  $J$  = 7.1 Hz, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 188.0, 171.6, 171.5, 154.6, 141.2, 129.4, 128.2, 126.3, 60.4, 39.0, 36.8, 35.2, 28.1, 12.9, 10.6. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2921, 2360, 2341, 1734, 1679, 1607, 1520, 1417, 1383, 1155, 829. HRMS (ESI,  $m/z$ ) calcd for C<sub>15</sub>H<sub>18</sub>N<sub>4</sub>NaO<sub>6</sub> [M+Na]<sup>+</sup>: 373.1120, found: 373.1119.

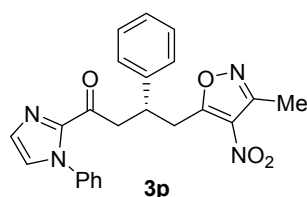


Light yellow oil, 68 mg, 83% yield, 97% ee (HPLC: chiralpak AD-H column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 11.74 min, tr (minor) = 9.50 min);  $[\alpha]_D^{25} = -15.0$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.49-7.43 (m, 3H), 7.29-7.18 (m, 4H), 3.68-3.58 (m, 2H), 3.54-3.45 (m, 2H), 3.28 (dd,  $J$  = 14.8, 7.2 Hz, 1H), 2.50 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 185.8, 170.8, 155.7, 141.9, 137.9, 130.7, 130.1, 129.1, 129.0, 127.9, 126.8 (q,  $J$  = 279.7 Hz), 125.9, 37.4, 36.6 (q,  $J$  = 27.8 Hz), 26.6 (q,  $J$  = 2.8 Hz), 11.6. <sup>19</sup>F NMR (376.4 MHz, CDCl<sub>3</sub>):  $\delta$  = -

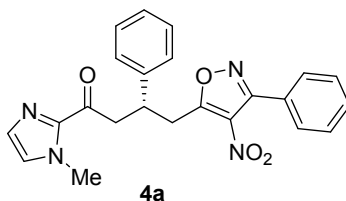
71.7. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2360, 2341, 1690, 1608, 1521, 1418, 1382, 1168, 829. HRMS (ESI,  $m/z$ ) calcd for C<sub>18</sub>H<sub>15</sub>F<sub>3</sub>N<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 431.0936, found: 431.0938.



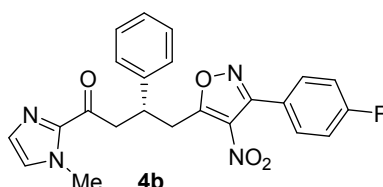
Light yellow oil, 73 mg, 95% yield, 92% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 9.19 min, tr (minor) = 8.30 min); [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -17.2 (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.27-7.23 (m, 5H), 7.20-7.14 (m, 2H), 5.46-5.36 (m, 1H), 4.04-3.96 (m, 1H), 3.74-3.50 (m, 4H), 2.47 (s, 3H), 1.38 (d,  $J$  = 6.6 Hz, 3H), 1.35 (d,  $J$  = 6.6 Hz, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 190.1, 173.2, 155.4, 142.0, 141.8, 130.4, 129.7, 128.7, 127.3, 127.2, 121.4, 49.2, 45.1, 39.0, 34.3, 23.6, 23.4, 11.6. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2361, 2341, 1674, 1603, 1519, 1396, 1381, 1365, 1255, 985, 829. HRMS (ESI,  $m/z$ ) calcd for C<sub>20</sub>H<sub>22</sub>N<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 405.1531, found: 405.1533.



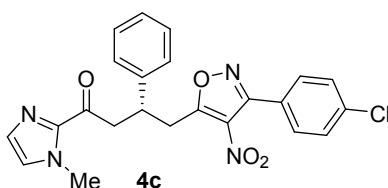
Light yellow oil, 75 mg, 90% yield, 95% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 11.63 min, tr (minor) = 14.78 min); [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -31.3 (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.43-7.38 (m, 3H), 7.26-7.11 (m, 9H), 3.98-3.91 (m, 1H), 3.74-3.54 (m, 3H), 3.50-3.44 (m, 1H), 2.45 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 188.5, 173.1, 155.4, 142.7, 141.8, 138.1, 130.4, 129.7, 129.0, 128.8, 128.7, 127.4, 127.3, 125.8, 44.7, 38.8, 34.2, 11.7. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2923, 2360, 2341, 1685, 1602, 1517, 1493, 1407, 1378, 1364, 829, 763, 700. HRMS (ESI,  $m/z$ ) calcd for C<sub>23</sub>H<sub>20</sub>N<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 439.1375, found: 439.1377.



Light yellow oil, 73 mg, 88% yield, 90% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 9.39 min, tr (minor) = 10.57 min);  $[\alpha]_D^{25} = +18.7$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.53$ -7.44 (m, 5H), 7.31-7.18 (m, 5H), 7.14 (s, 1H), 7.01 (s, 1H), 4.10-4.03 (m, 1H), 3.91 (s, 3H), 3.78-3.57 (m, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta = 189.9$ , 173.8, 157.6, 142.8, 141.8, 130.7, 129.8, 129.2, 128.8, 128.5, 127.3, 125.8, 44.4, 38.9, 36.2, 34.5. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2922, 2851, 2360, 2341, 1676, 1636, 1617, 1593, 1515, 1413, 1367, 829. HRMS (ESI, *m/z*) calcd for C<sub>23</sub>H<sub>20</sub>N<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 439.1375, found: 439.1377.



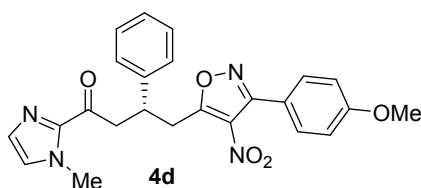
Light yellow oil, 78 mg, 90% yield, 95% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 8.45 min, tr (minor) = 9.30 min);  $[\alpha]_D^{25} = +7.9$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.55$ -7.51 (m, 2H), 7.31-7.26 (m, 4H), 7.21-7.12 (m, 4H), 7.01 (s, 1H), 4.09-4.02 (m, 1H), 3.90 (s, 3H), 3.78-3.57 (m, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta = 189.8$ , 174.1, 164.2 (d, *J* = 249.9 Hz), 156.7, 142.7, 141.8, 131.5 (d, *J* = 8.7 Hz), 129.7, 129.2, 128.8, 127.3, 127.3, 121.9 (d, *J* = 3.5 Hz), 115.7 (d, *J* = 21.9 Hz), 44.5, 38.9, 36.2, 34.5. <sup>19</sup>F NMR (376.4 MHz, CDCl<sub>3</sub>):  $\delta = -109.2$ . IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2922, 2360, 2341, 1674, 1606, 1617, 1593, 1515, 1413, 1367, 828, 699. HRMS (ESI, *m/z*) calcd for C<sub>23</sub>H<sub>19</sub>FN<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 457.1283, found: 457.1283.



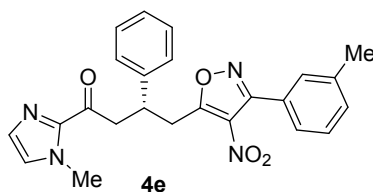
White solid, 83 mg, m.p. 128.4-128.8 °C, 92% yield, 95% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 8.83



min, tr (minor) = 9.69 min);  $[\alpha]_D^{25} = +8.0$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.48-7.41 (m, 4H), 7.31-7.25 (m, 4H), 7.21-7.18 (m, 1H), 7.13 (s, 1H), 7.01 (s, 1H), 4.10-4.02 (m, 1H), 3.90 (s, 3H), 3.77-3.57 (m, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 189.8, 174.2, 156.7, 142.7, 141.7, 137.0, 130.7, 129.7, 129.2, 128.8, 127.6, 127.4, 127.3, 124.3, 44.5, 38.9, 36.2, 34.5. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2918, 2849, 2360, 2341, 1671, 1604, 1521, 1507, 1409, 1384, 831, 701. HRMS (ESI, *m/z*) calcd for C<sub>23</sub>H<sub>19</sub>ClN<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 473.0983, found: 473.0987.

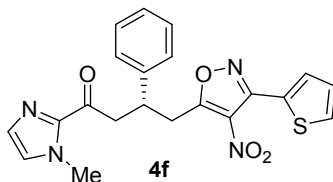


White solid, 85 mg, m.p. 126.5-126.9 °C, 95% yield, 95% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 13.60 min, tr (minor) = 15.61 min);  $[\alpha]_D^{25} = +9.1$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.49 (d, *J* = 8.7 Hz, 2H), 7.31-7.25 (m, 4H), 7.20 (t, *J* = 7.0 Hz, 1H), 7.13 (s, 1H), 7.00 (s, 1H), 6.96 (d, *J* = 8.7 Hz, 2H), 4.09-4.02 (m, 1H), 3.90 (s, 3H), 3.84 (s, 3H), 3.77-3.55 (m, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 189.9, 173.8, 161.5, 157.2, 142.7, 141.9, 130.8, 129.8, 129.2, 128.8, 127.4, 127.3, 117.9, 113.9, 55.4, 44.4, 38.9, 36.2, 34.5. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2924, 2360, 1674, 1598, 1520, 1411, 1365, 1155, 982, 827, 702. HRMS (ESI, *m/z*) calcd for C<sub>24</sub>H<sub>22</sub>N<sub>4</sub>NaO<sub>5</sub> [M+Na]<sup>+</sup>: 469.1479, found: 469.1482.

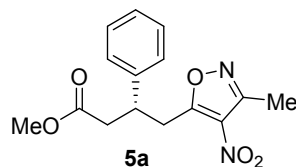


Yellow oil, 77 mg, 90% yield, 95% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 8.88 min, tr (minor) = 10.15 min);  $[\alpha]_D^{25} = +4.8$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.36-7.25 (m, 8H), 7.20 (t, *J* = 6.8 Hz, 1H), 7.13 (s, 1H), 7.00 (s, 1H), 4.10-4.02 (m, 1H), 3.90 (s, 3H), 3.78-3.55 (m, 4H), 2.40 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 189.9, 173.7, 157.7, 142.8, 141.8, 138.3, 131.4, 129.8, 129.7, 129.2, 128.8, 128.4, 127.3,

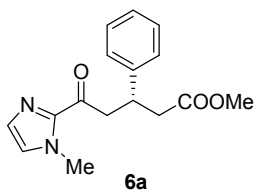
126.3, 125.7, 44.4, 38.9, 36.2, 34.5, 21.4. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2924, 2360, 1674, 1598, 1520, 1411, 1365, 1155, 982, 702. HRMS (ESI,  $m/z$ ) calcd for C<sub>24</sub>H<sub>22</sub>N<sub>4</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 453.1532, found: 453.1533.



White solid, 75 mg, m.p. 120.2-120.5 °C, 89% yield, 95% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 80/20, flow rate 1.0 mL/min, tr (major) = 10.75 min, tr (minor) = 11.80 min);  $[\alpha]_D^{25} = +13.0$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.80 (t,  $J$  = 3.1 Hz, 1H), 7.53-7.51 (m, 1H), 7.31-7.25 (m, 4H), 7.21-7.12 (m, 3H), 7.00 (s, 1H), 4.08-4.01 (m, 1H), 3.89 (s, 3H), 3.75-3.55 (m, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 189.9, 174.5, 151.7, 142.7, 141.8, 131.8, 129.9, 129.4, 129.2, 128.8, 127.8, 127.3, 127.2, 125.6, 44.5, 38.9, 36.2, 34.6. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2922, 2360, 1673, 1594, 1517, 1409, 1385, 1365, 702. HRMS (ESI,  $m/z$ ) calcd for C<sub>21</sub>H<sub>18</sub>N<sub>4</sub>NaO<sub>4</sub>S [M+Na]<sup>+</sup>: 445.0943, found: 445.0941.



Light yellow oil, 88.6 mg, 86% yield, 95% ee (HPLC: chiralpak OD-H column, 254 nm, hexane/isopropanol = 90/10, flow rate 1.0 mL/min, tr (major) = 9.88 min, tr (minor) = 15.62 min);  $[\alpha]_D^{25} = +5.6$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.28 (t,  $J$  = 6.9 Hz, 2H), 7.23-7.19 (m, 3H), 3.82-3.74 (m, 1H), 3.65-3.59 (m, 1H), 3.60 (s, 3H), 3.55-3.49 (m, 1H), 2.82-2.72 (m, 2H), 2.48 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 172.8, 171.6, 155.5, 141.1, 130.4, 128.9, 127.5, 127.0, 51.9, 40.3, 39.9, 33.8, 11.6. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2921, 2360, 1733, 1679, 1607, 1520, 1416, 1384, 1155, 829. HRMS (ESI,  $m/z$ ) calcd for C<sub>15</sub>H<sub>16</sub>N<sub>2</sub>NaO<sub>5</sub> [M+Na]<sup>+</sup>: 327.0949, found: 327.0951.



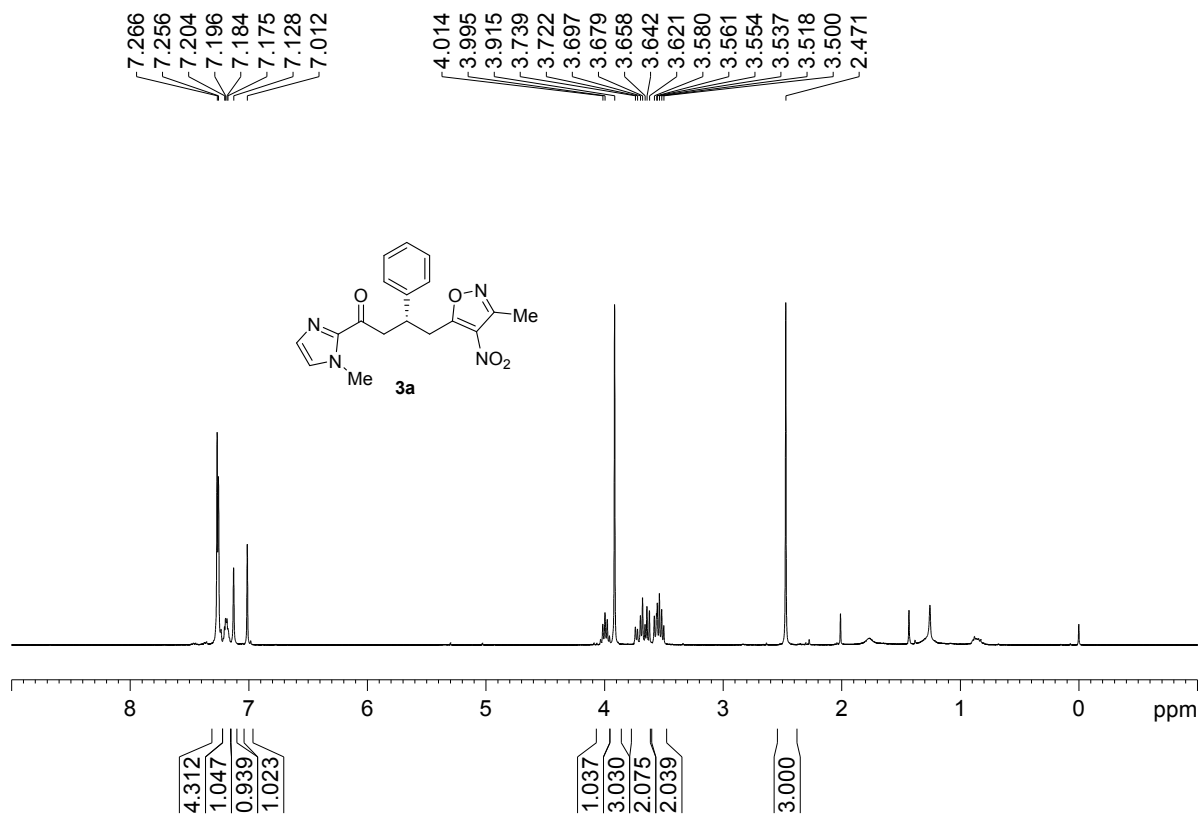
Light yellow oil, 90.0 mg, 93% yield, 90% ee (HPLC: chiralpak IC column, 254 nm, hexane/isopropanol = 85/15, flow rate 1.0 mL/min, tr (major) = 11.68 min, tr (minor) = 13.53 min);  $[\alpha]_D^{25} = +12.3$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.30-7.22 (m, 4H), 7.19-7.14 (m, 1H), 7.12 (s, 1H), 7.00 (s, 1H), 3.91 (s, 3H), 3.88-3.83 (m, 1H), 3.65 (dd,  $J$  = 17.0, 7.7 Hz, 1H), 3.57 (s, 3H), 3.47 (dd,  $J$  = 17.0, 6.7 Hz, 1H), 2.77 (dd,  $J$  = 15.6, 7.0 Hz, 1H), 2.67 (dd,  $J$  = 15.6, 8.0 Hz, 1H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 190.4, 172.3, 143.2, 128.7, 128.5, 127.5, 126.9, 126.7, 51.6, 44.8, 41.0, 37.2, 36.2. IR (KBr):  $\nu$  (cm<sup>-1</sup>) 2924, 2361 1733, 1679, 1607, 1520, 1416, 1384, 1155, 828. HRMS (ESI,  $m/z$ ) calcd for C<sub>16</sub>H<sub>18</sub>N<sub>2</sub>NaO<sub>3</sub> [M+Na]<sup>+</sup>: 309.1215, found: 309.1216.

## IV References

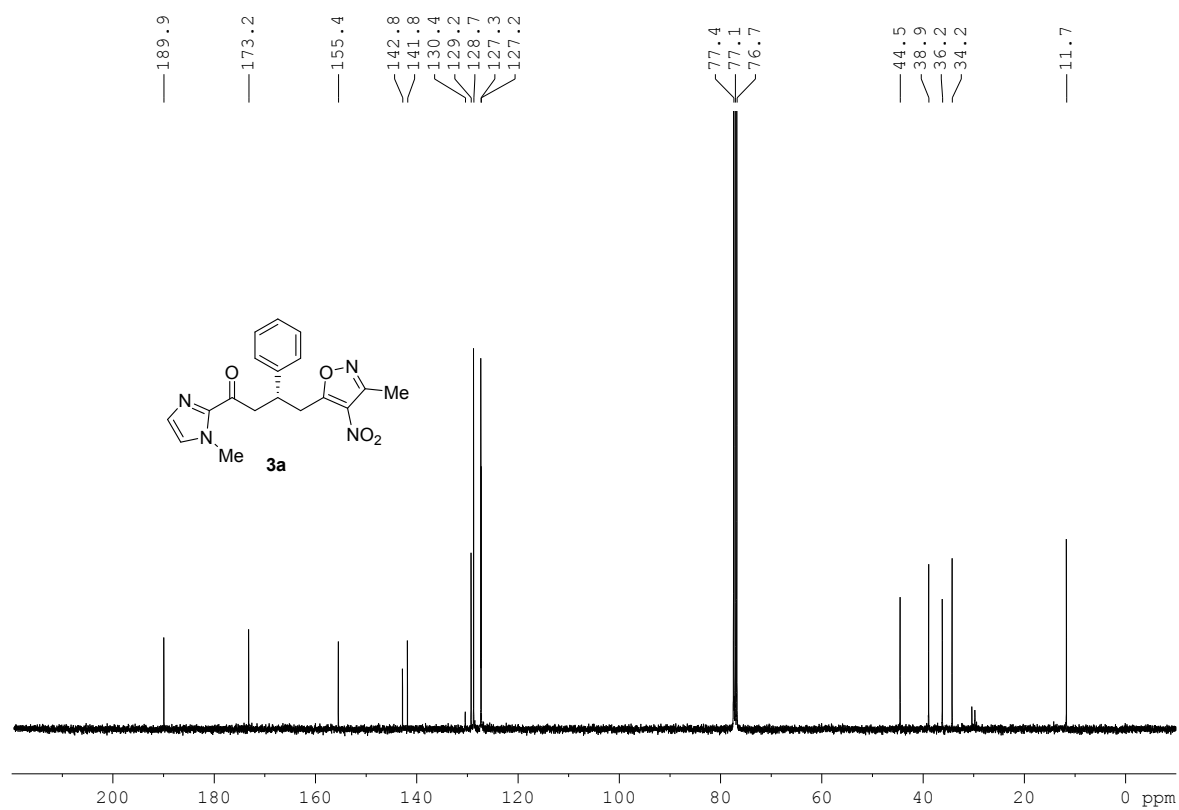
- (1) (a) C. Wang, L.-A. Chen, H. Huo, X. Shen, K. Harms, L. Gong and E. Meggers, *Chem. Sci.*, 2015, **6**, 1094; (b) G.-J. Sun, J. Gong and O. Kang, *J. Org. Chem.*, 2017, **82**, 796; (c) S.-W. Li, J. Gong and Q. Kang, *Org. Lett.*, 2017, **19**, 1350; (d) K. Li, Q. Wan and Q. Kang, *Org. Lett.*, 2017, **19**, 3299; (e) S.-X. Lin, G.-J. Sun and Q. Kang, *Chem. Commun.*, 2017, **53**, 7665.
- (2) H. Huo, X. Shen, C. Wang, L. Zhang, P. Rose, L.-A. Chen, K. Harms, M. Marsch, G. Hilt and E. Meggers, *Nature*, 2014, **515**, 100.
- (3) H. Huo, C. Fu, K. Harms and E. Meggers, *J. Am. Chem. Soc.*, 2014, **136**, 2990.
- (4) (a) J.-L. Zhang, X.-H. Liu, X.-J. Ma and R. Wang, *Chem. Commun.*, 2013, **49**, 9329; (b) G. Fuller, B. Kelly and M. F. A. Adamo, *Org. Process Res. Dev.*, 2015, **19**, 1274; (c) C. Balsamini, G. Spadoni, L. Staccioli and E. Duranti, *J. Org. Chem.*, 1988, **53**, 2870; (d) S. Chimichi, P. Sarti-Fantoni, A. Buzzi, D. Giomi and R. Nesi, *Heterocycles*, 1985, **23**, 1465.

## V NMR Spectrum

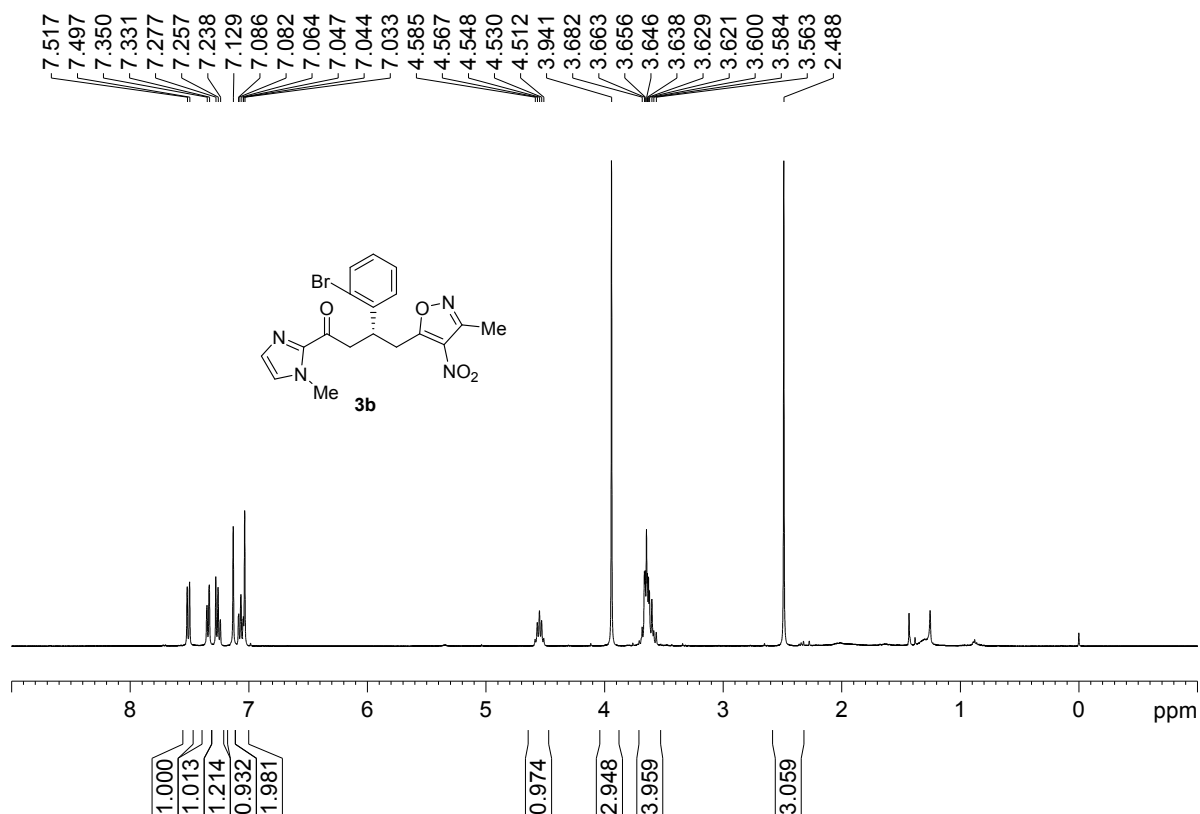
$^1\text{H}$  NMR (400 MHz) spectra of **3a**



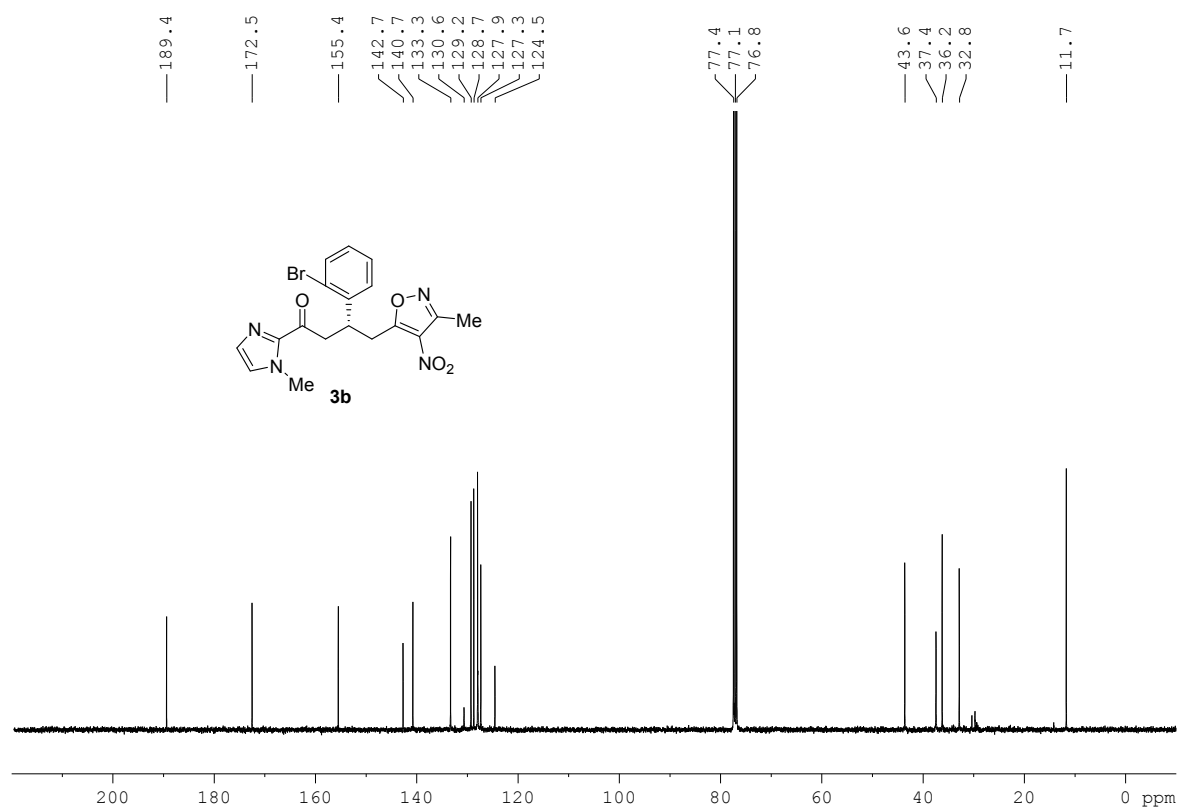
$^{13}\text{C}$  NMR (100 MHz) spectra of **3a**



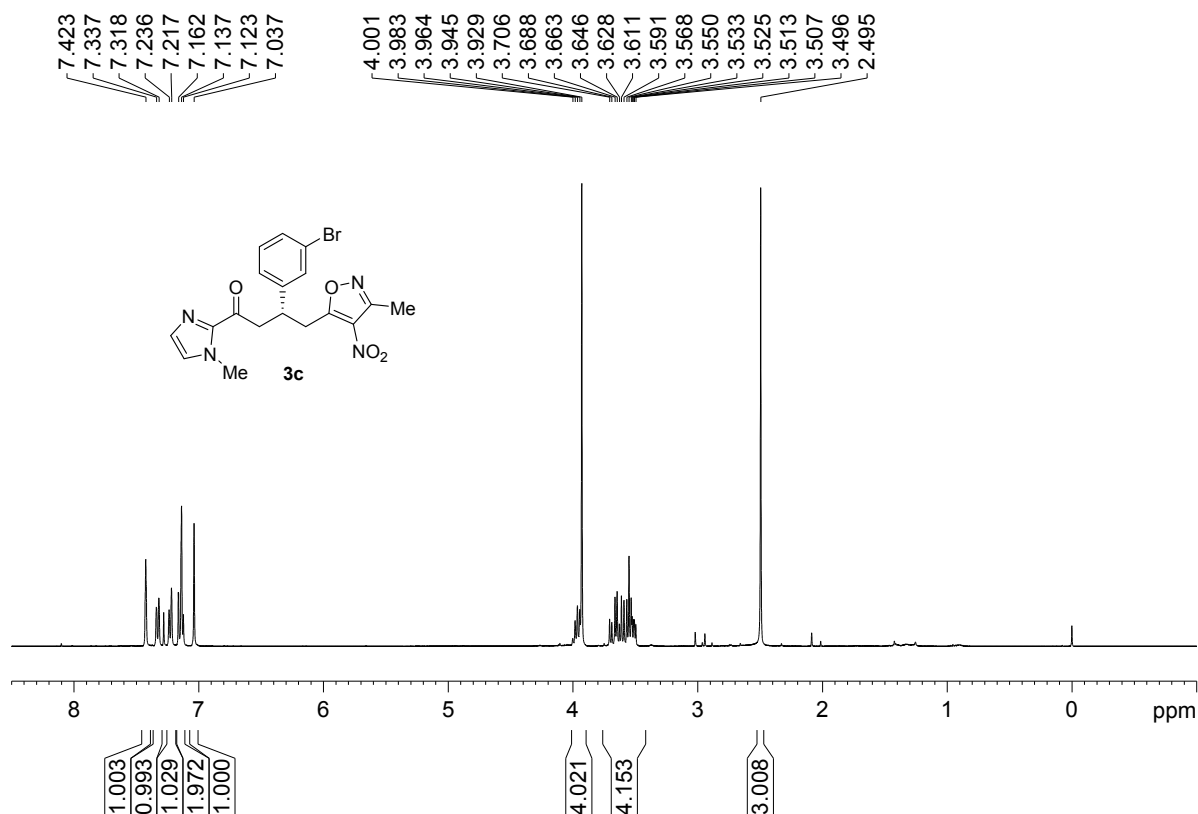
<sup>1</sup>H NMR (400 MHz) spectra of **3b**



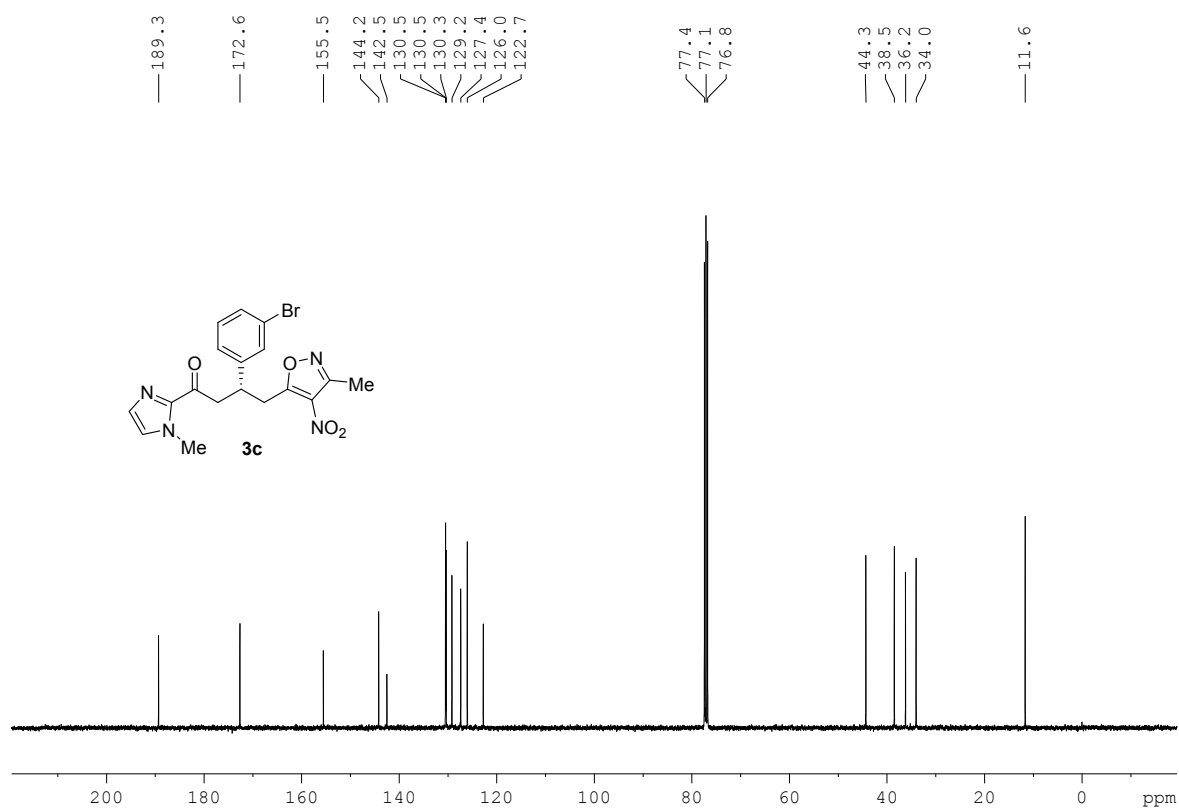
<sup>13</sup>C NMR (100 MHz) spectra of **3b**



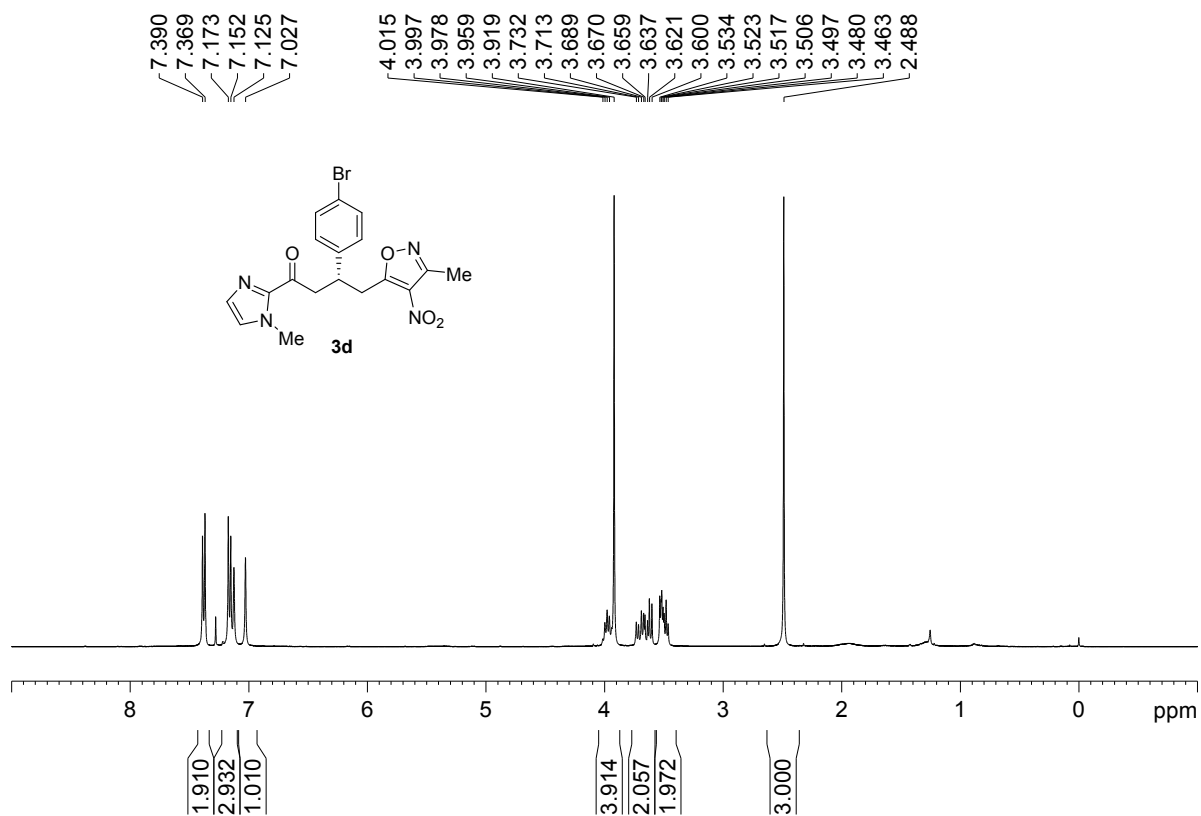
<sup>1</sup>H NMR (400 MHz) spectra of **3c**



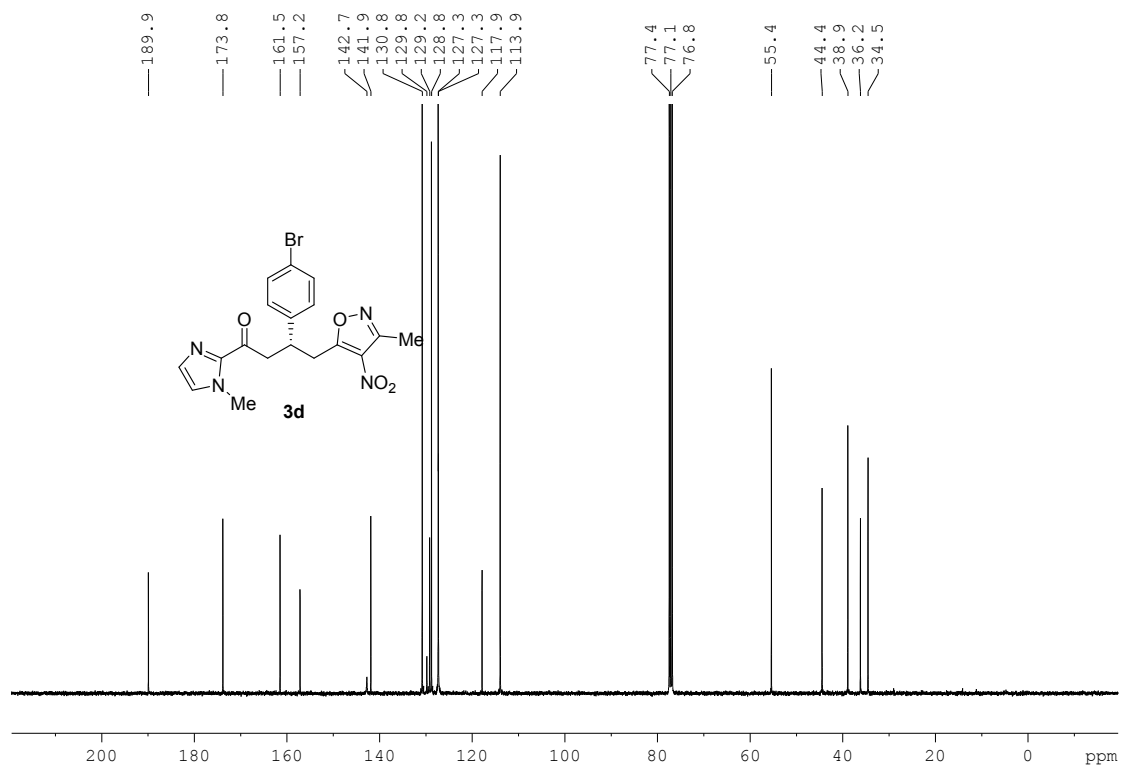
<sup>13</sup>C NMR (100 MHz) spectra of **3c**



<sup>1</sup>H NMR (400 MHz) spectra of **3d**

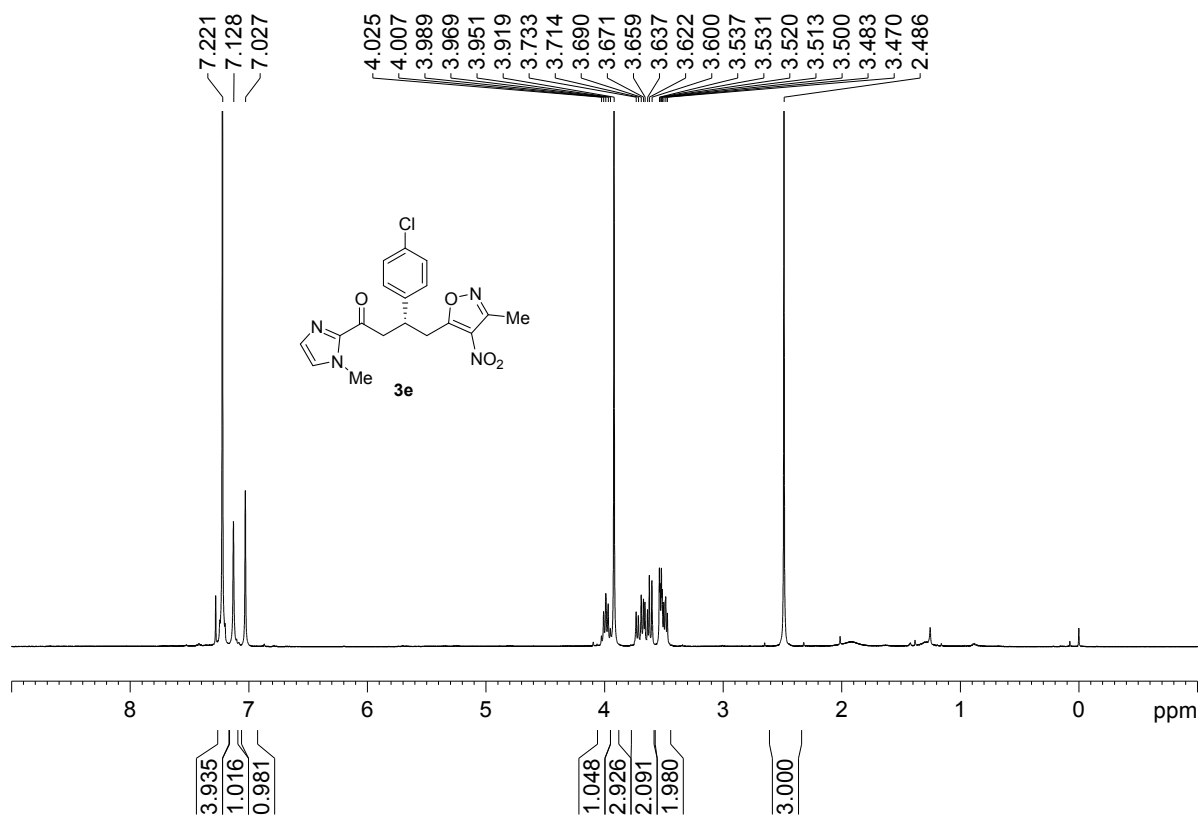


<sup>13</sup>C NMR (100 MHz) spectra of **3d**

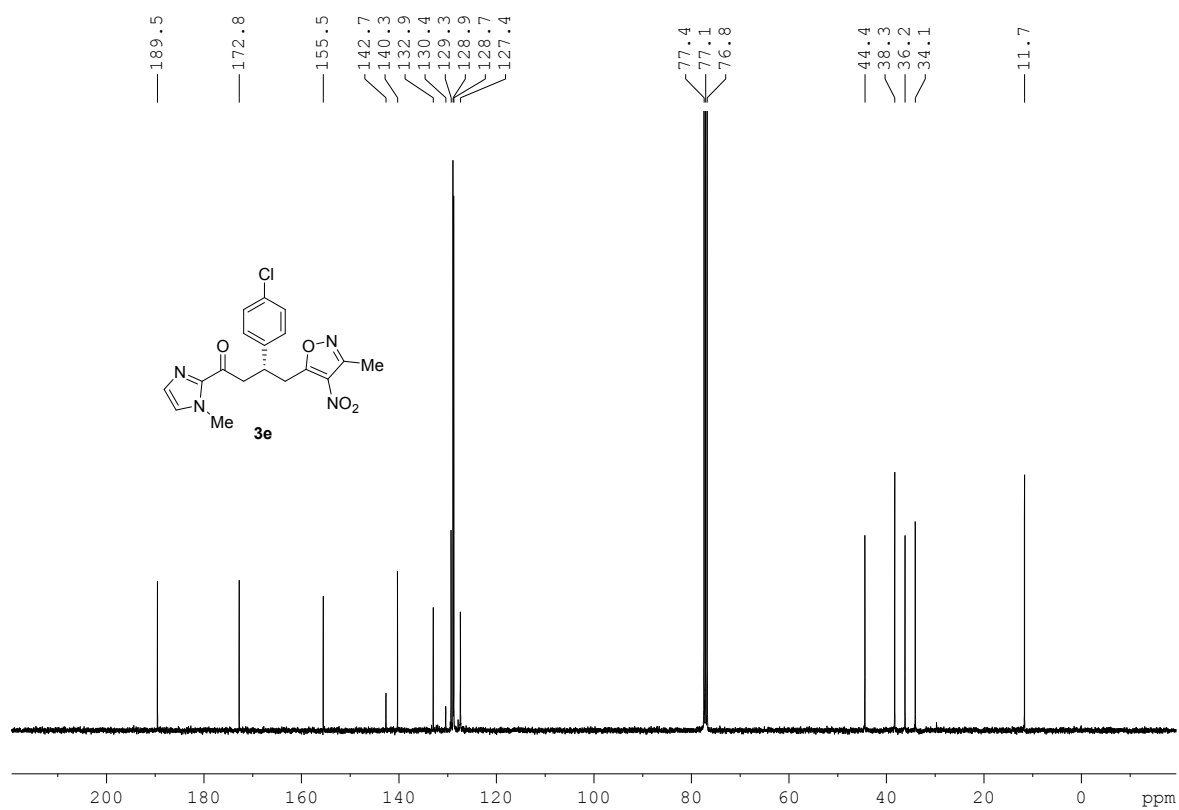




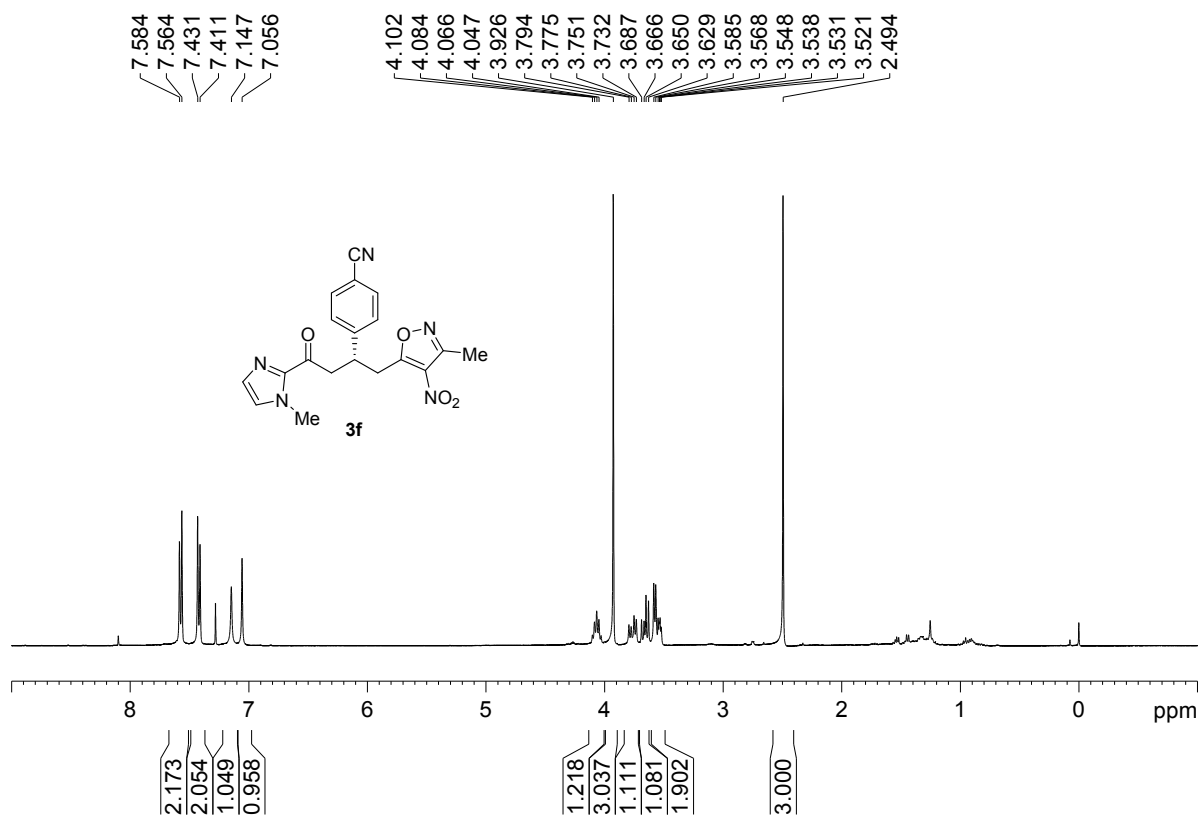
<sup>1</sup>H NMR (400 MHz) spectra of **3e**



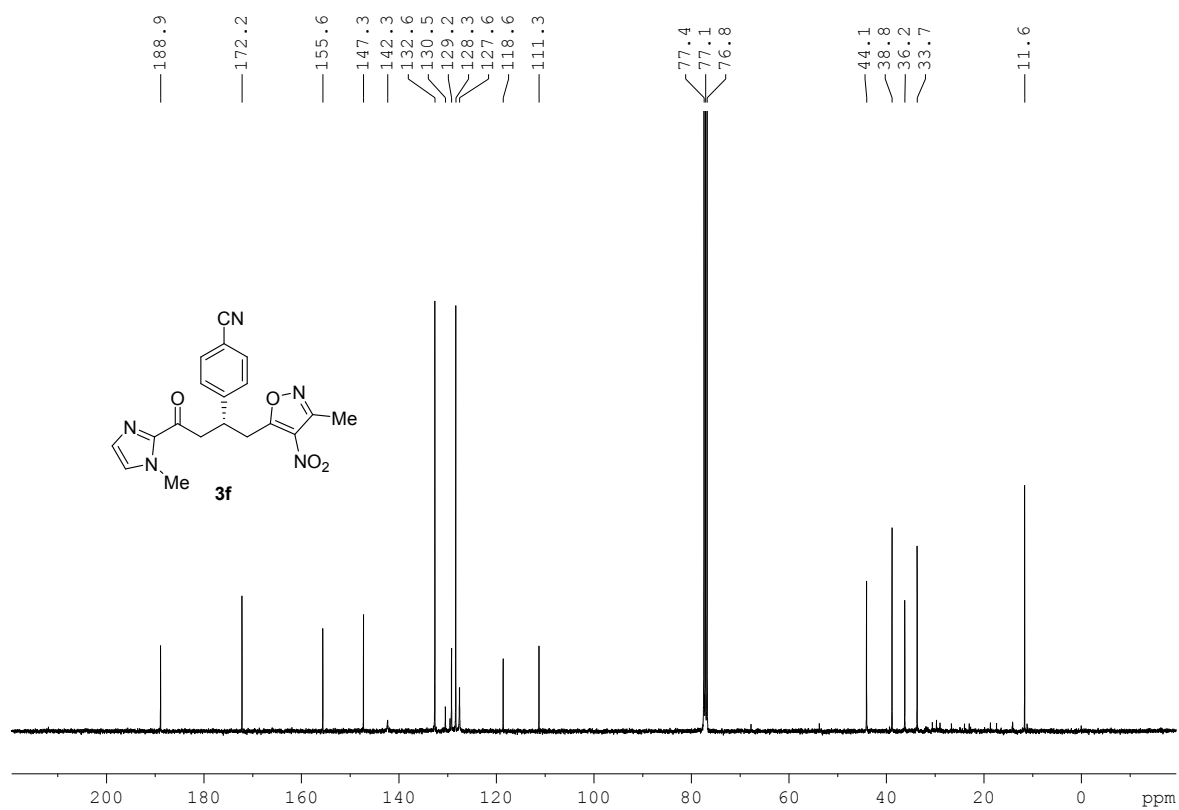
<sup>13</sup>C NMR (100 MHz) spectra of **3e**



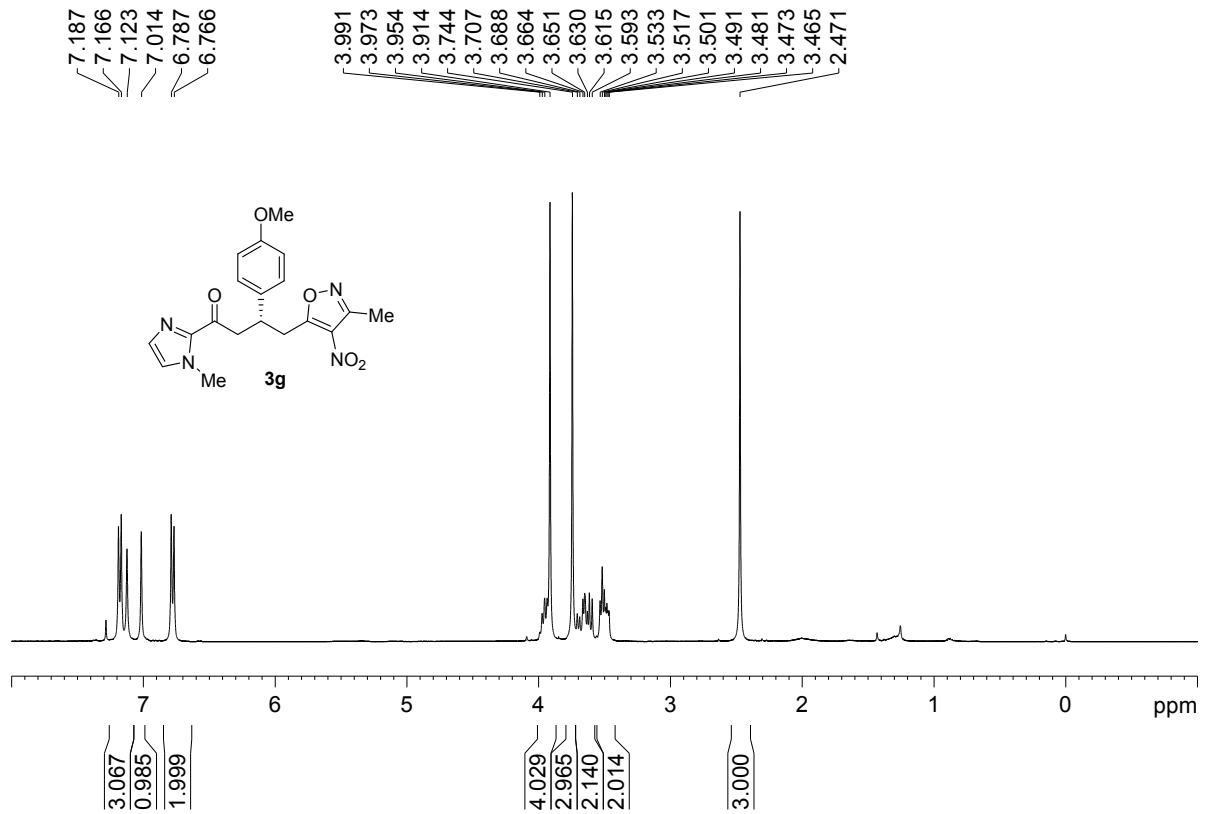
<sup>1</sup>H NMR (400 MHz) spectra of **3f**



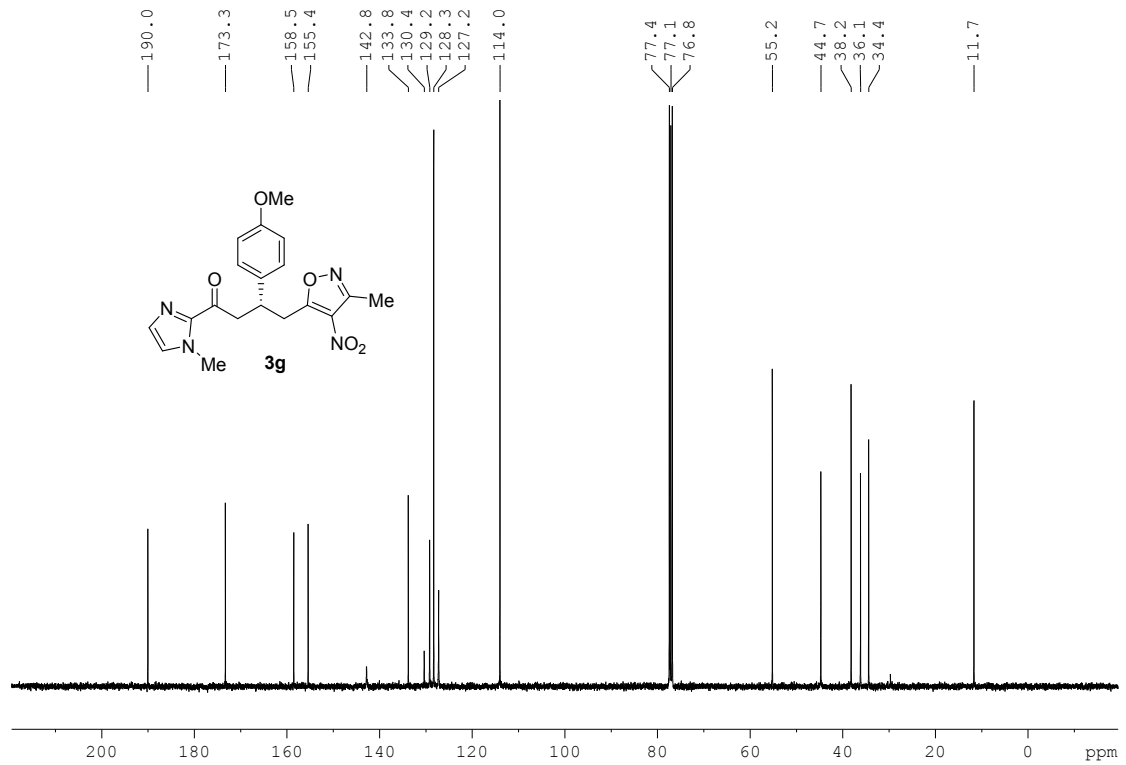
<sup>13</sup>C NMR (100 MHz) spectra of **3f**



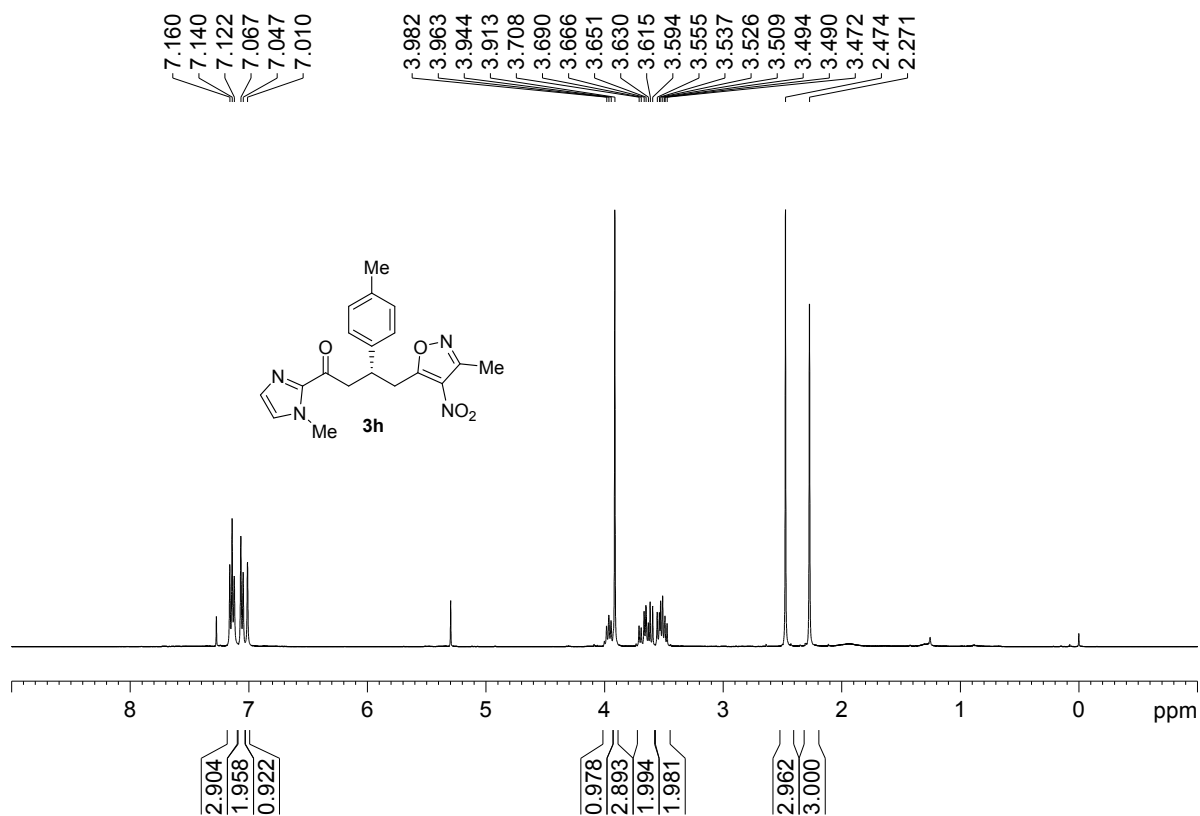
<sup>1</sup>H NMR (400 MHz) spectra of **3g**



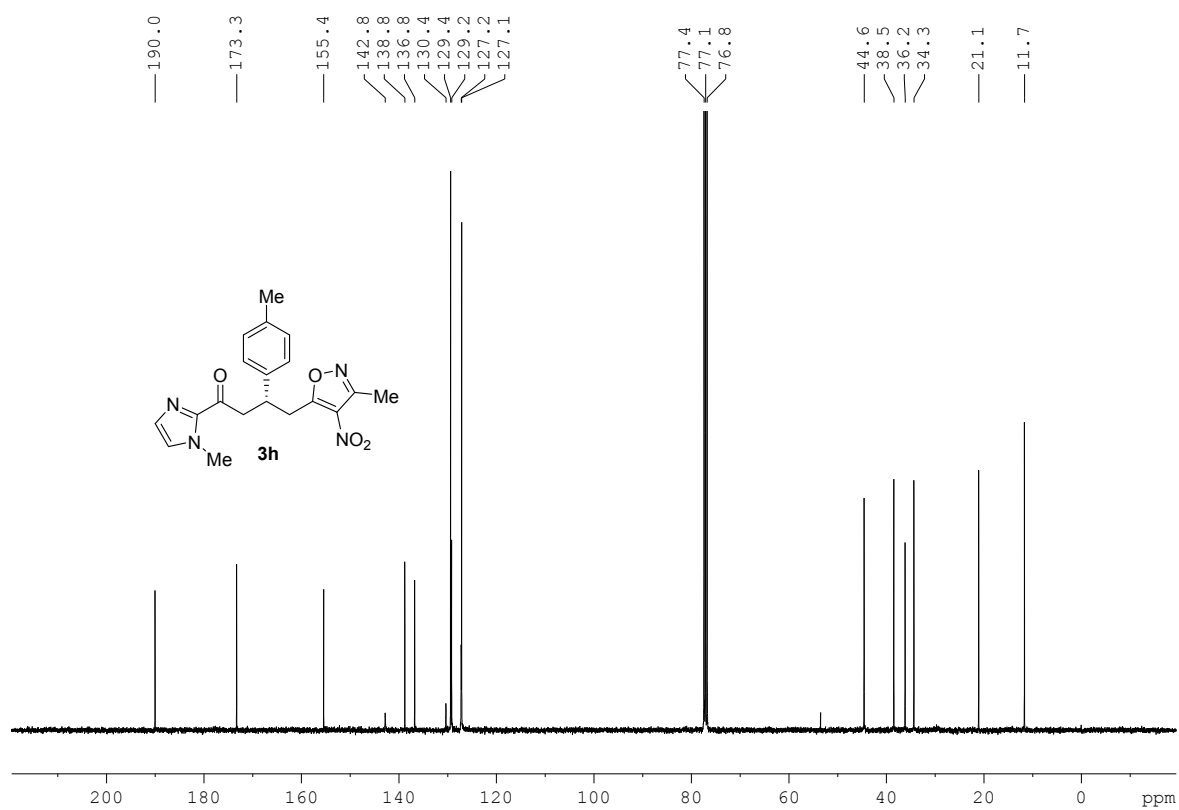
<sup>13</sup>C NMR (100 MHz) spectra of **3g**



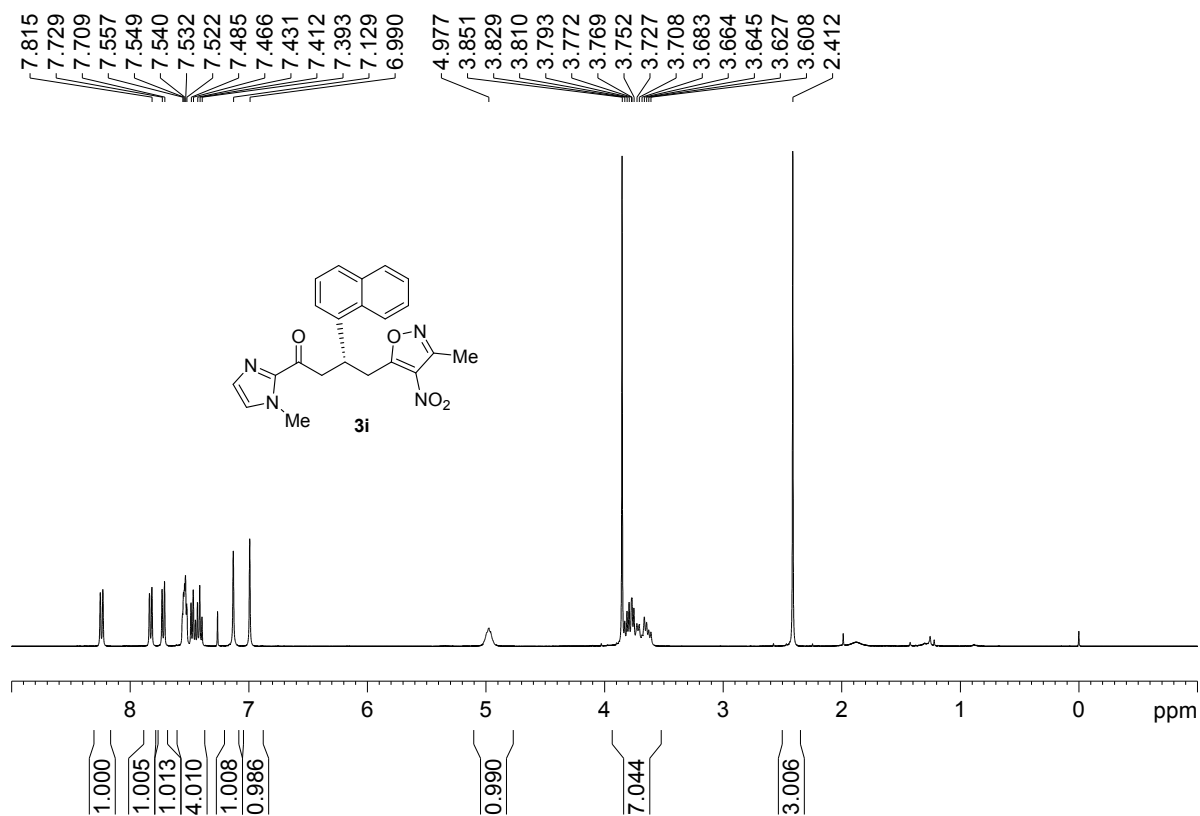
<sup>1</sup>H NMR (400 MHz) spectra of **3h**



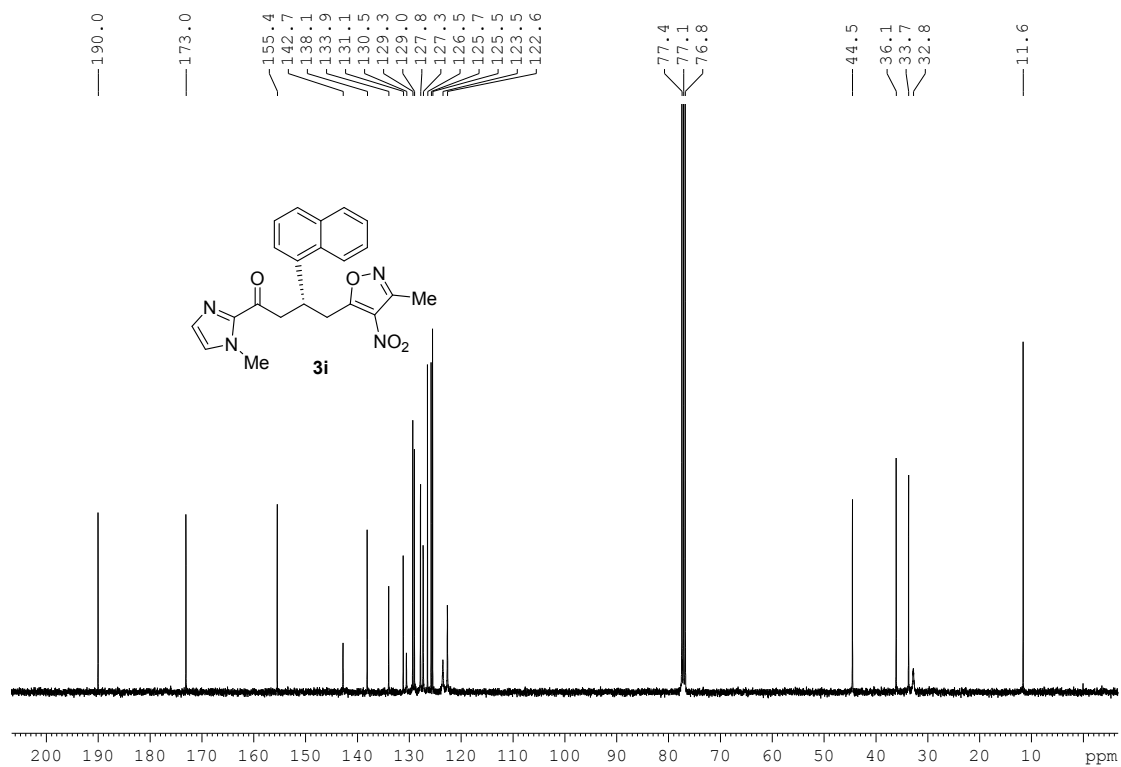
<sup>13</sup>C NMR (100 MHz) spectra of **3h**



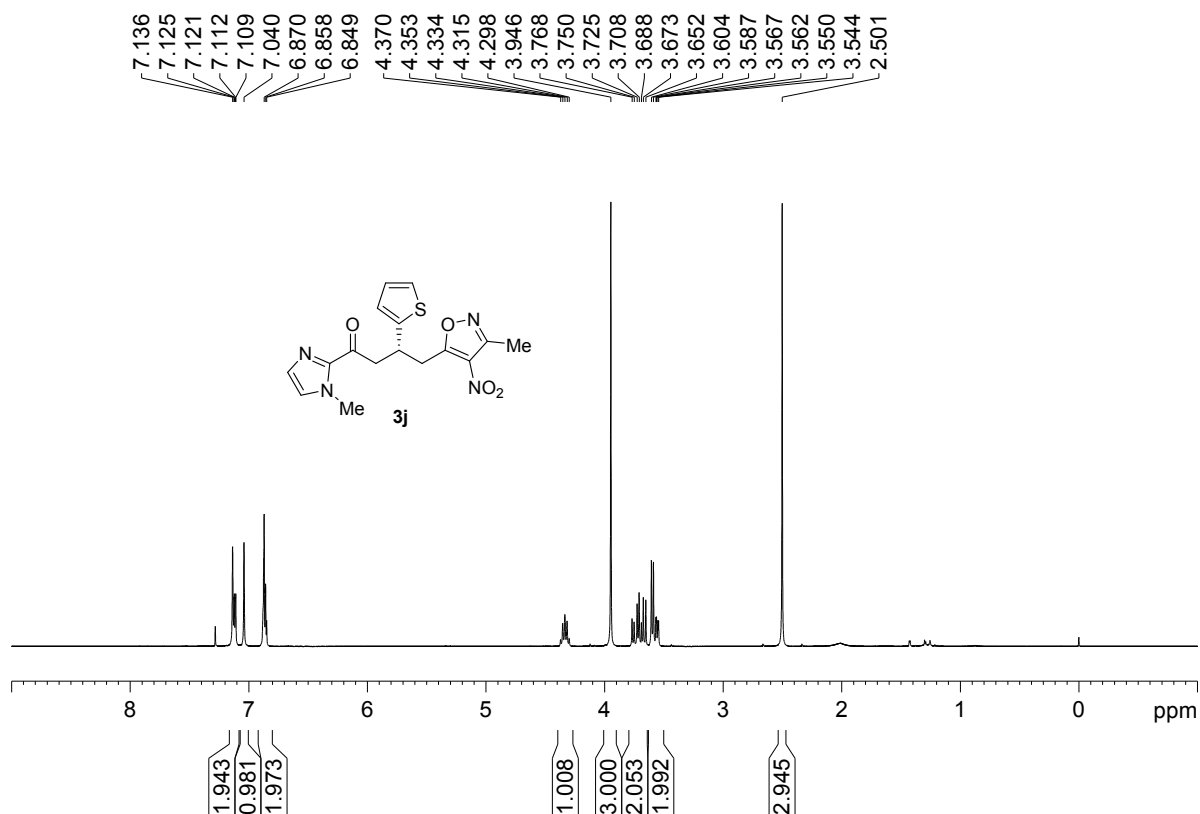
<sup>1</sup>H NMR (400 MHz) spectra of **3i**



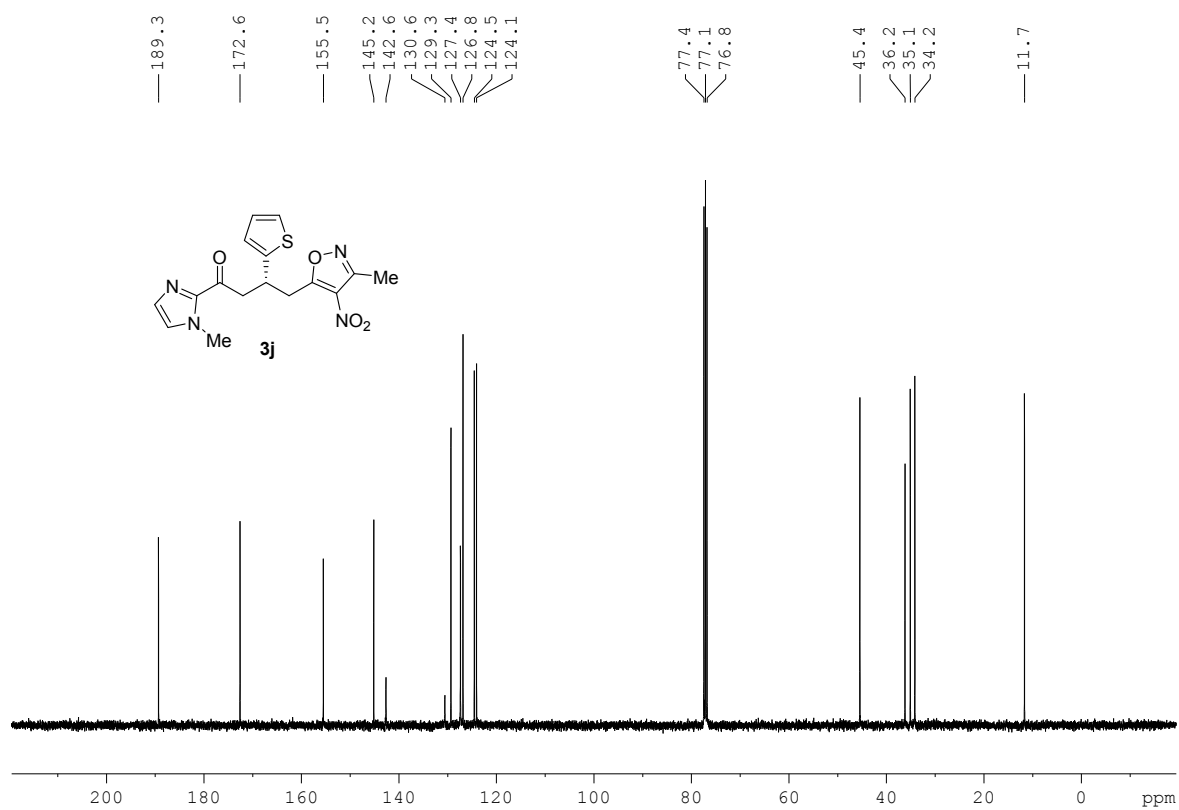
<sup>13</sup>C NMR (100 MHz) spectra of **3i**



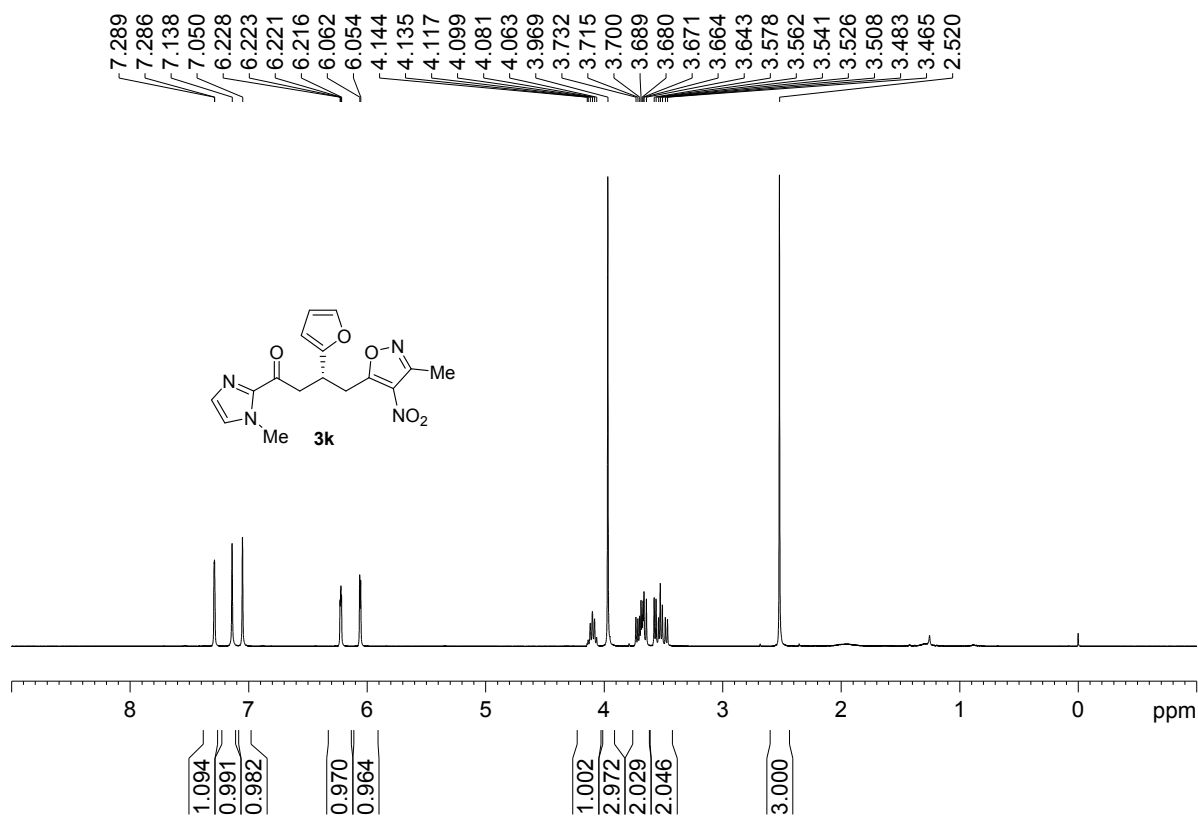
<sup>1</sup>H NMR (400 MHz) spectra of **3j**



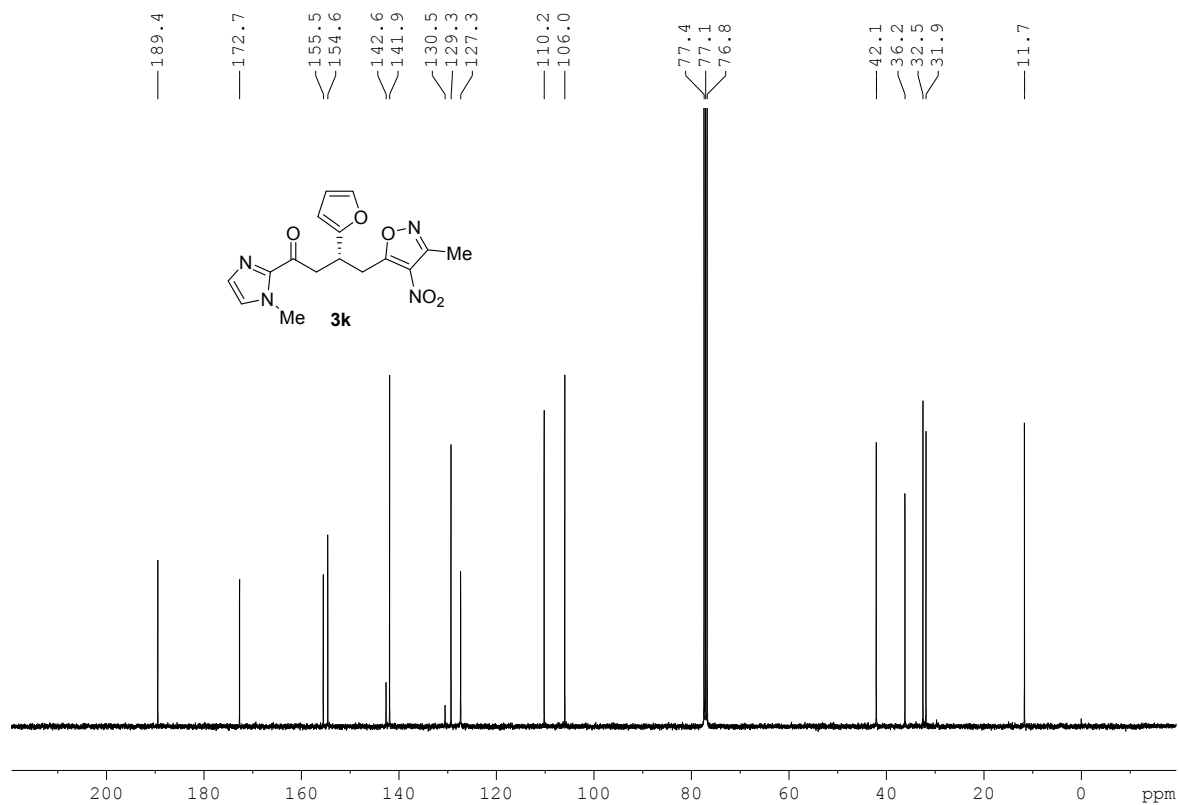
<sup>13</sup>C NMR (100 MHz) spectra of **3j**



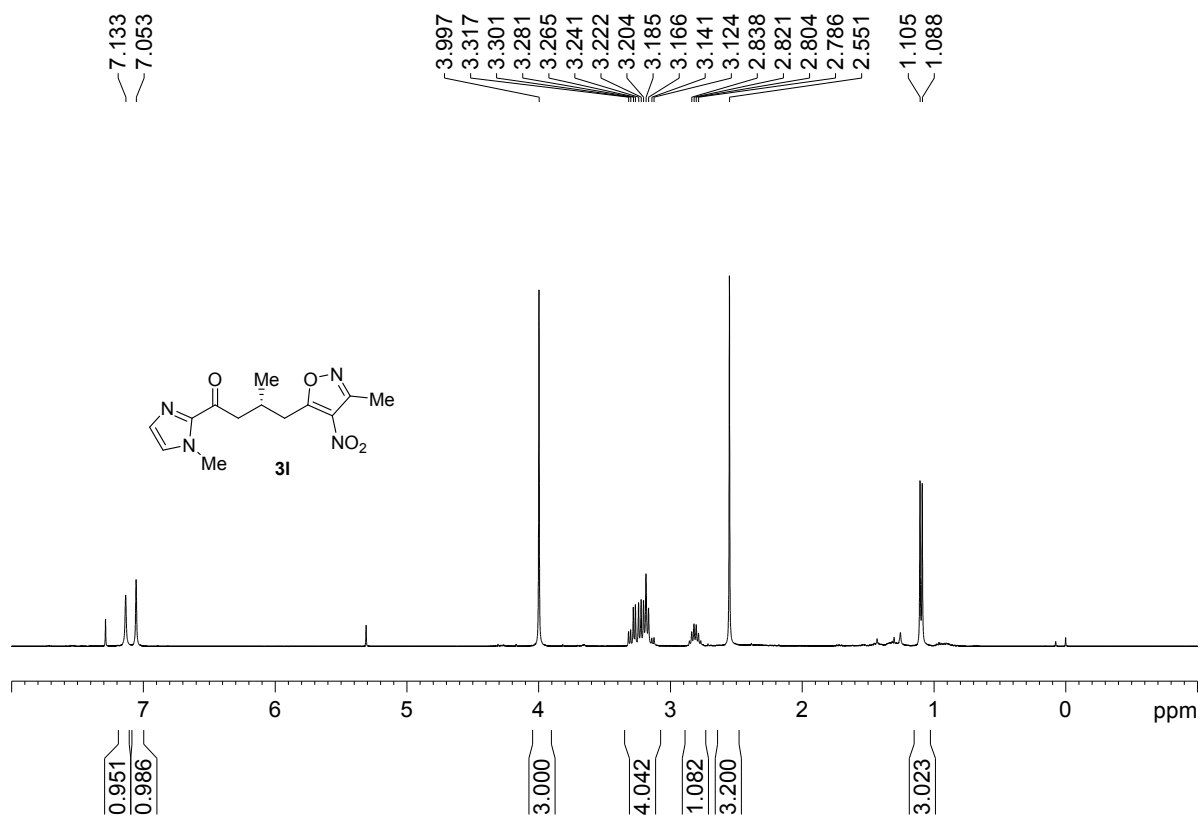
<sup>1</sup>H NMR (400 MHz) spectra of **3k**



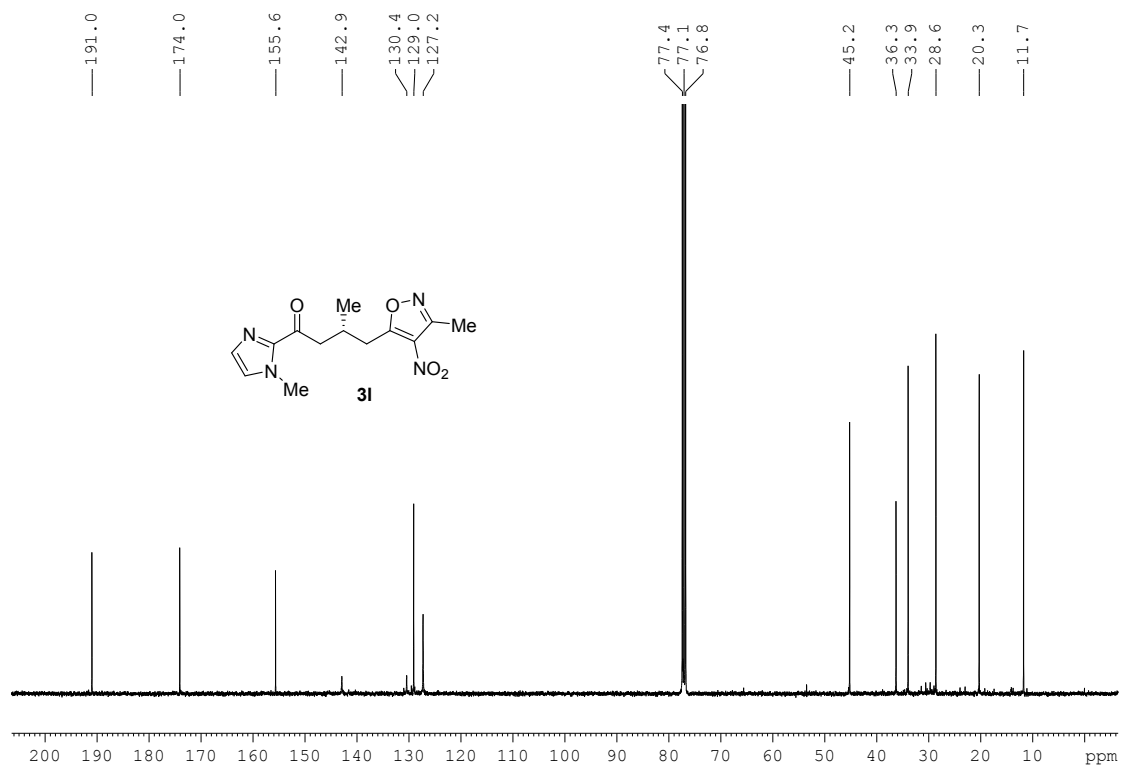
<sup>13</sup>C NMR (100 MHz) spectra of **3k**



<sup>1</sup>H NMR (400 MHz) spectra of **31**

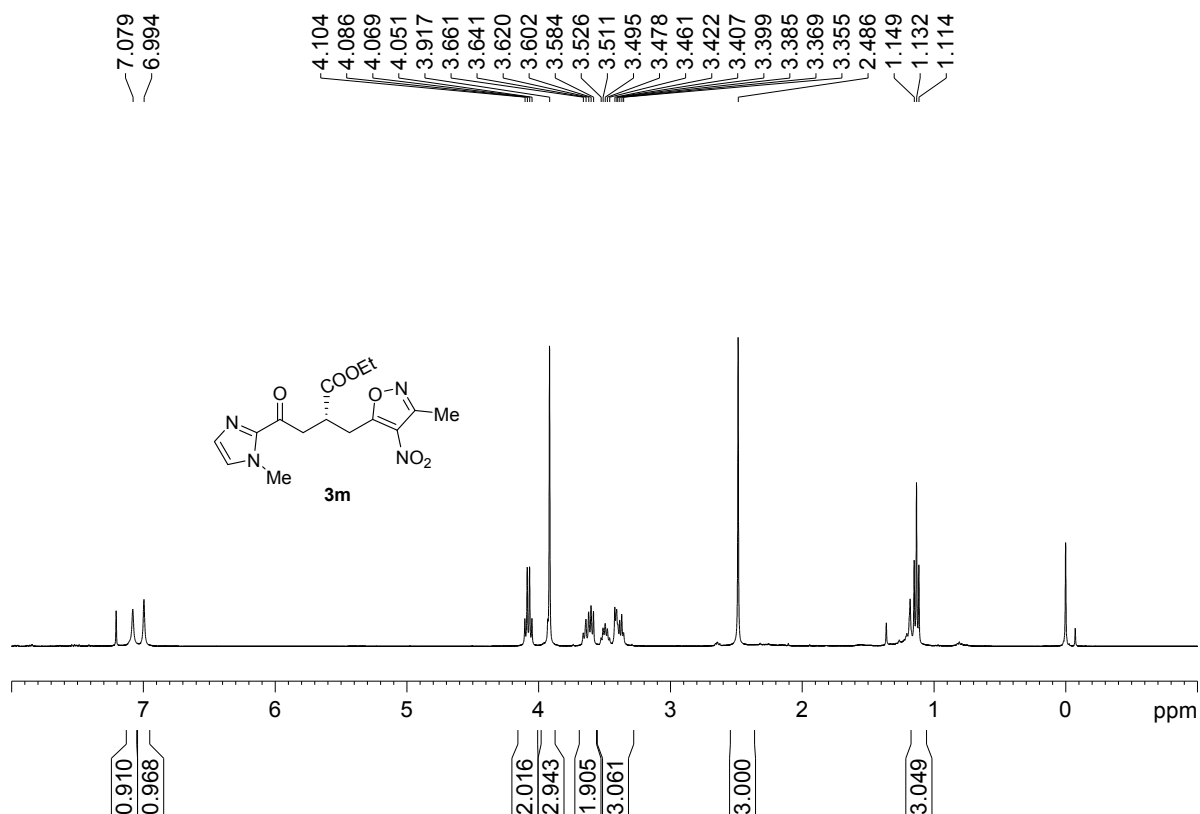


<sup>13</sup>C NMR (100 MHz) spectra of **31**

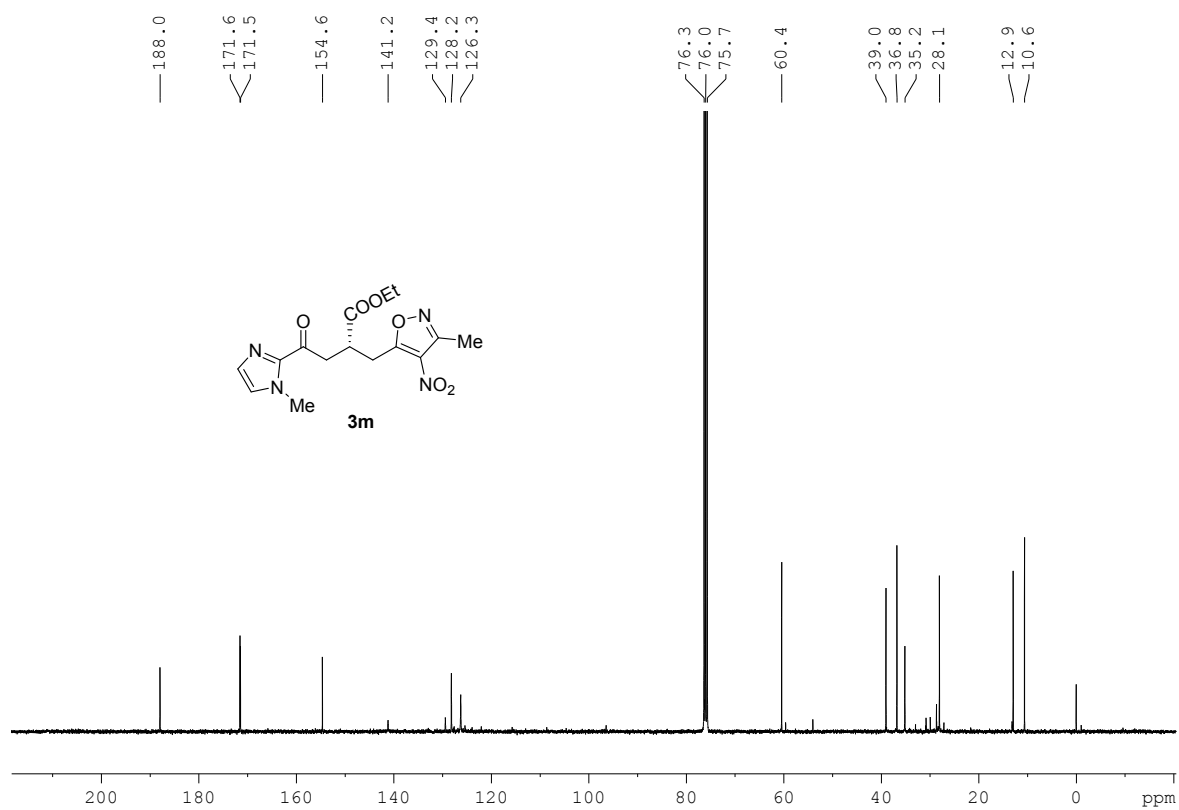




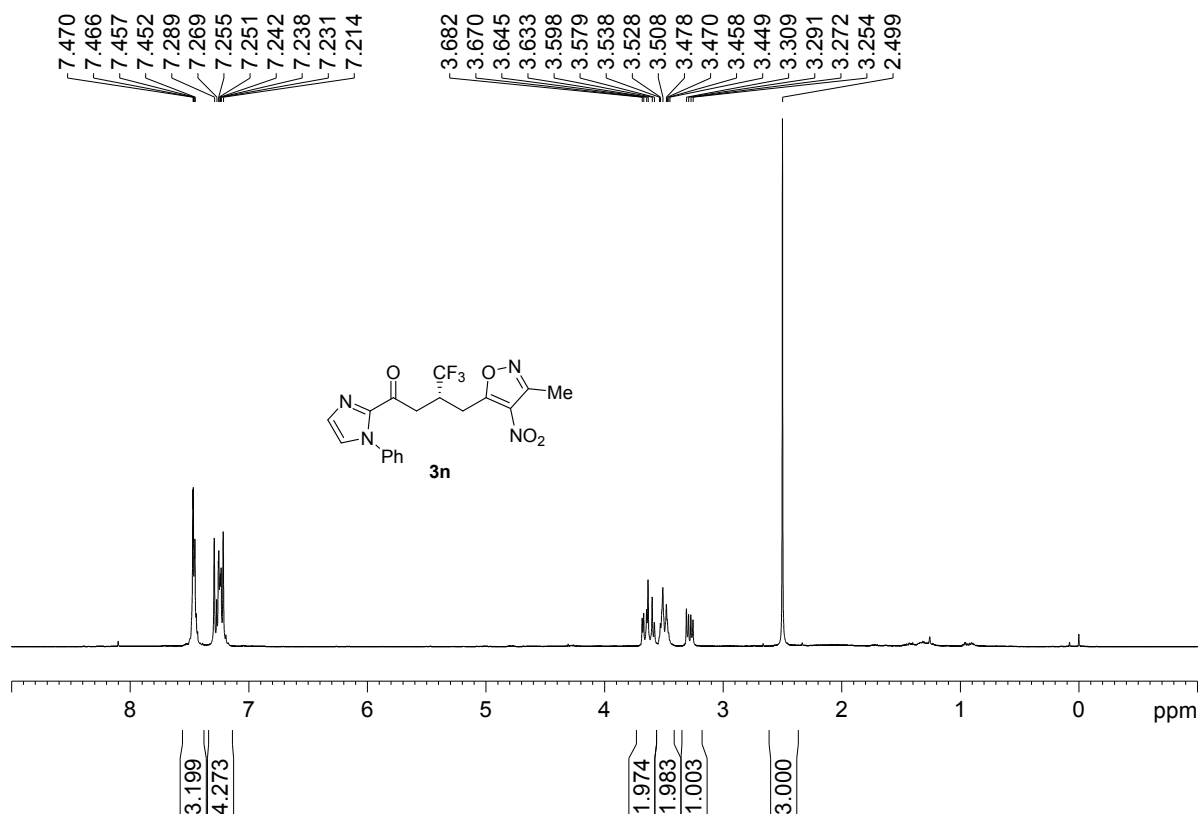
<sup>1</sup>H NMR (400 MHz) spectra of **3m**



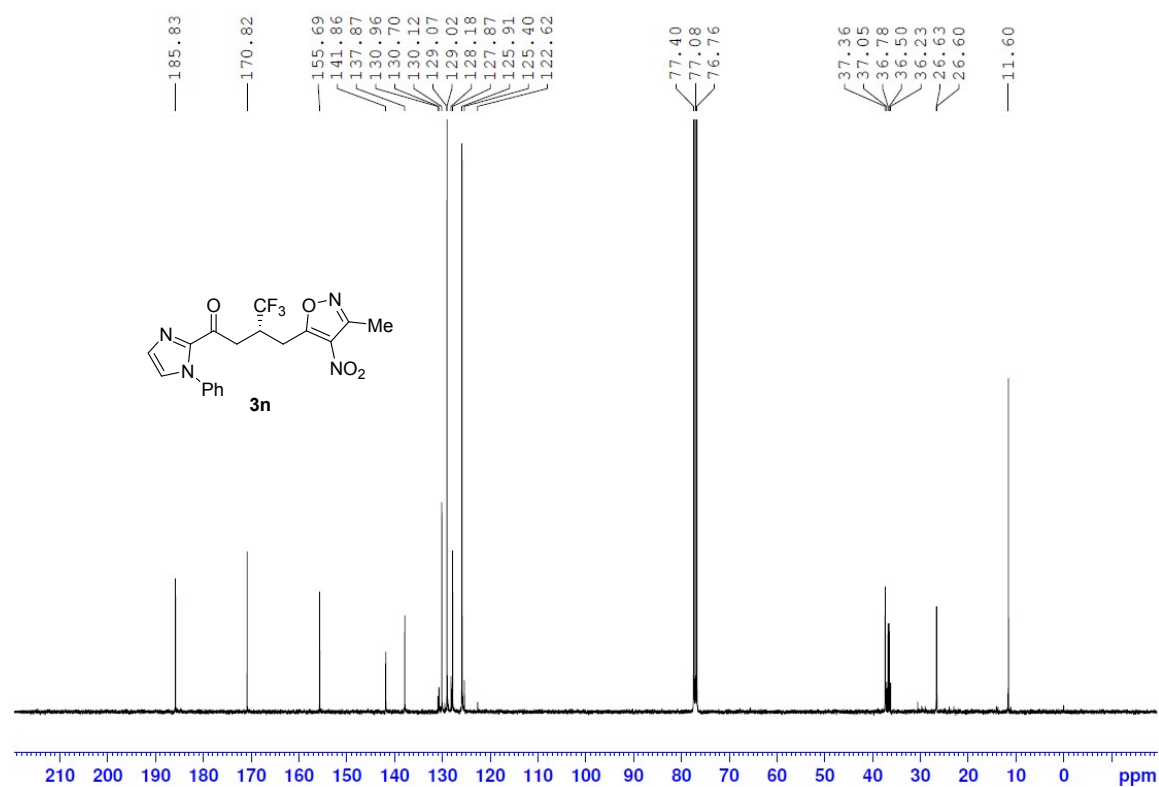
<sup>13</sup>C NMR (100 MHz) spectra of **3m**



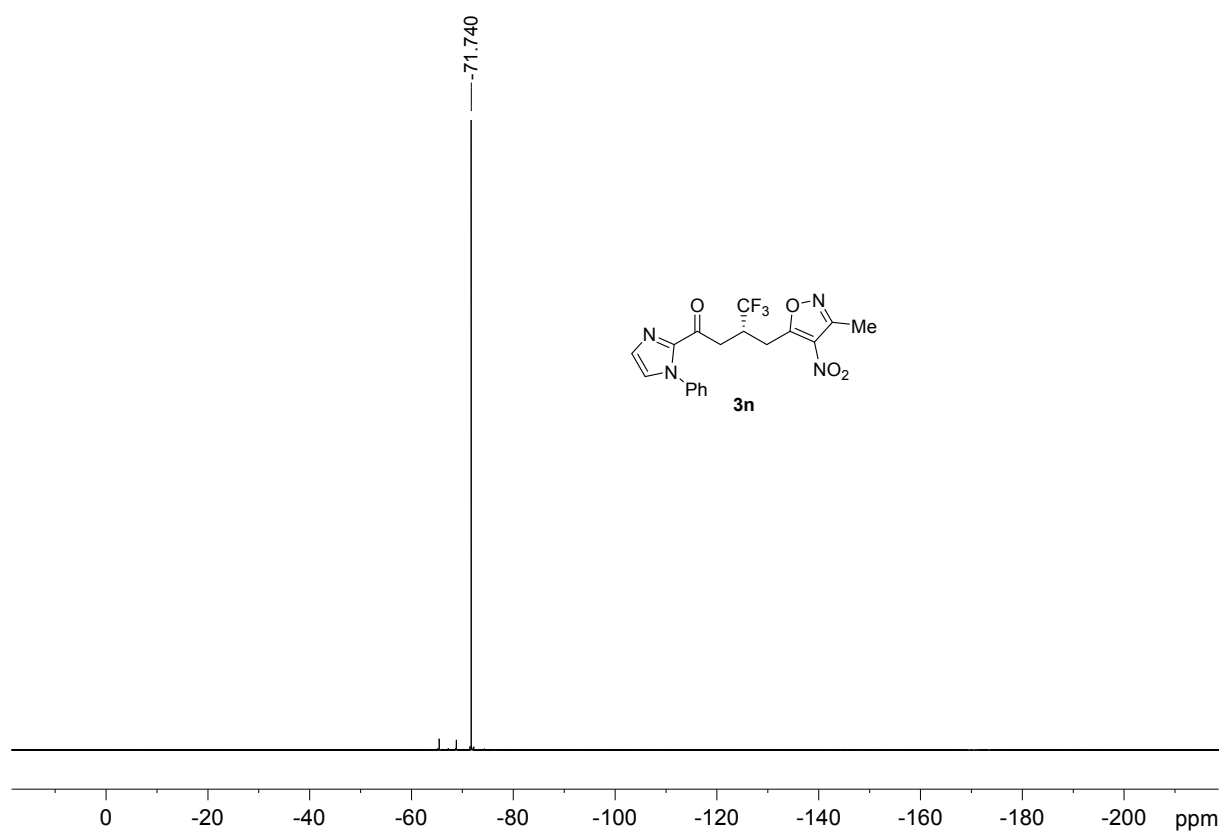
<sup>1</sup>H NMR (400 MHz) spectra of **3n**



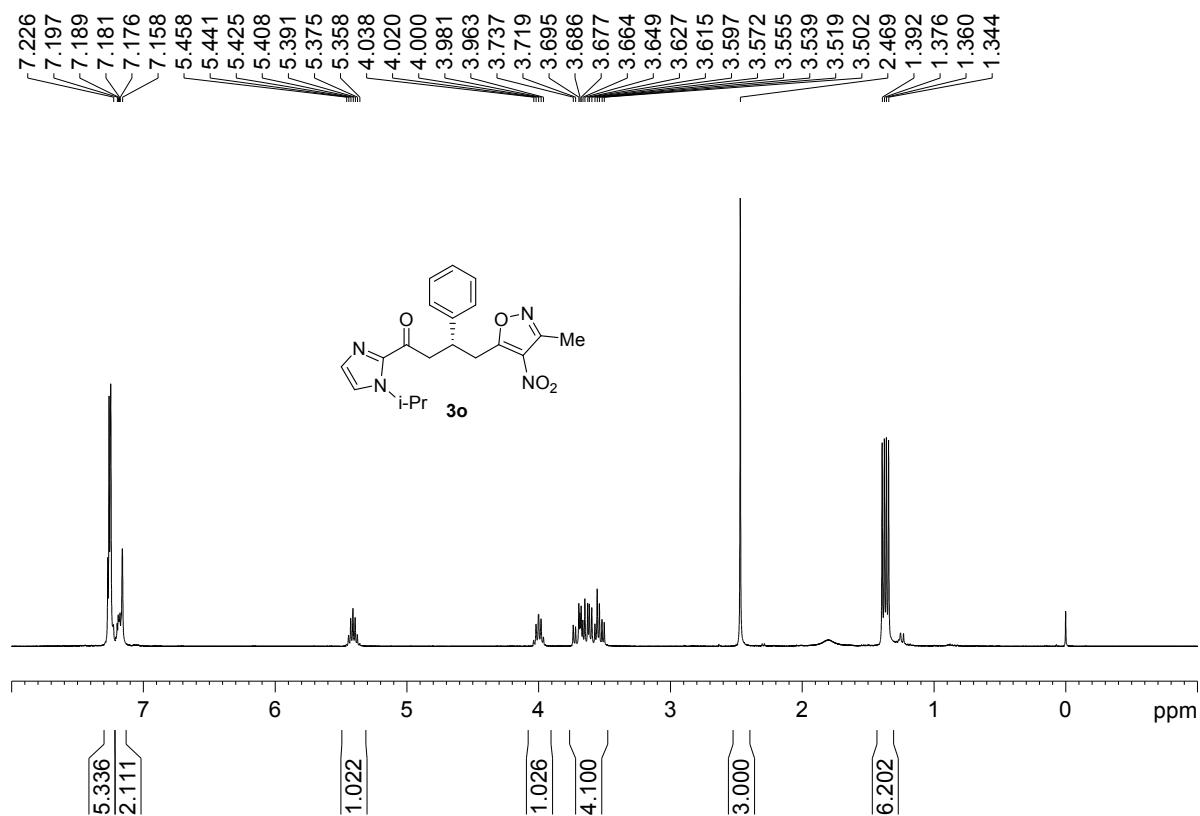
<sup>13</sup>C NMR (100 MHz) spectra of **3n**



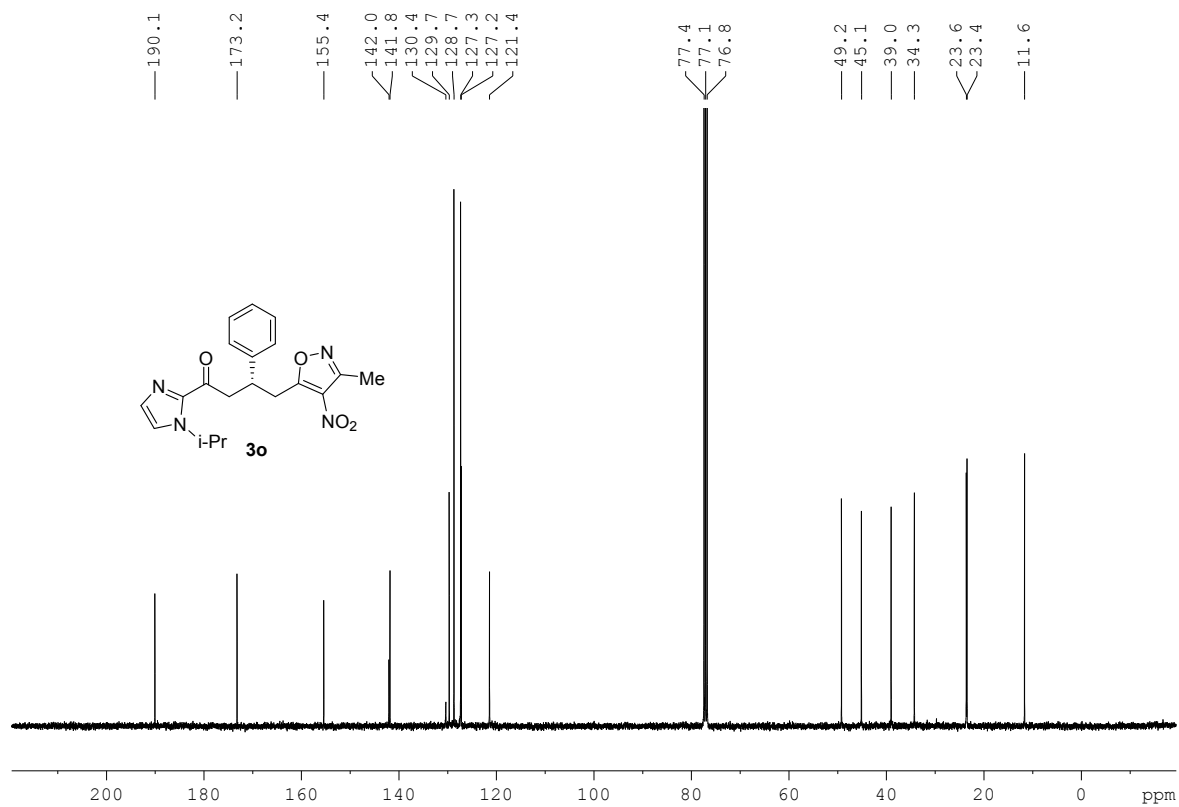
$^{19}\text{F}$  NMR (376.4 MHz) spectra of **3n**



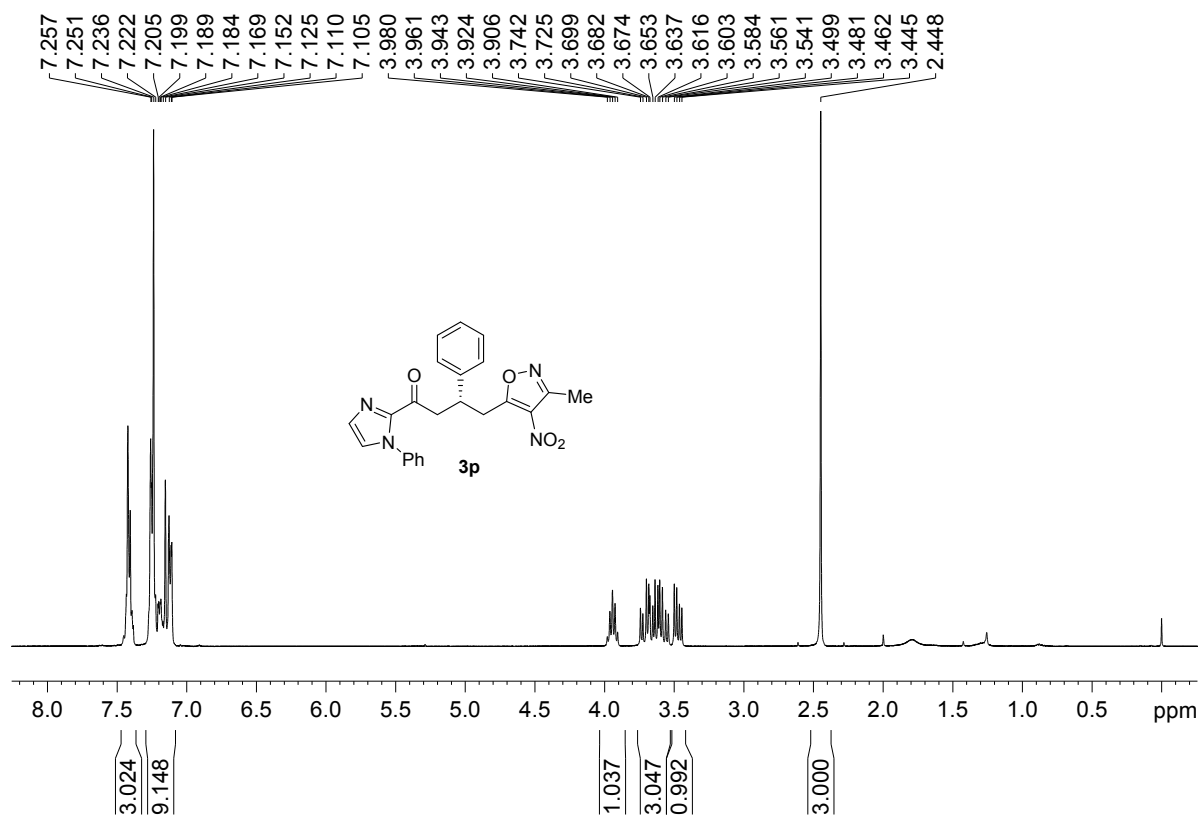
<sup>1</sup>H NMR (400 MHz) spectra of **3o**



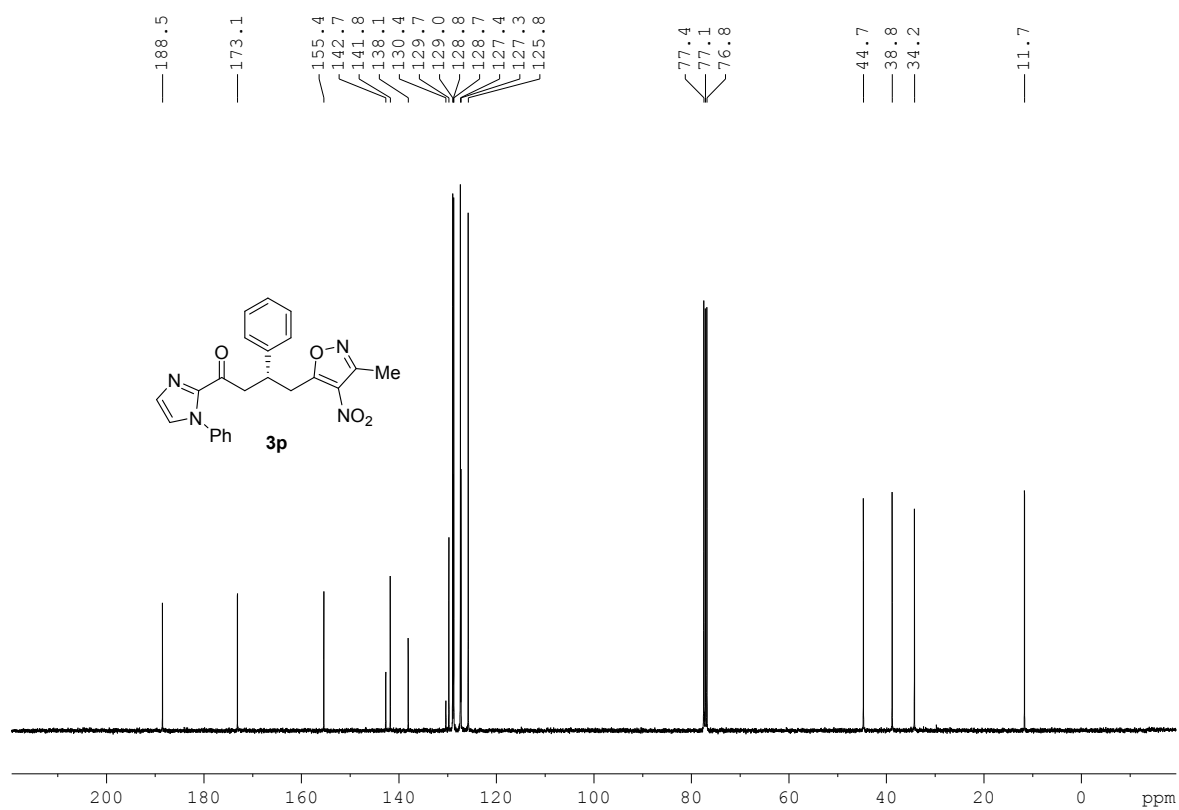
<sup>13</sup>C NMR (100 MHz) spectra of **3o**



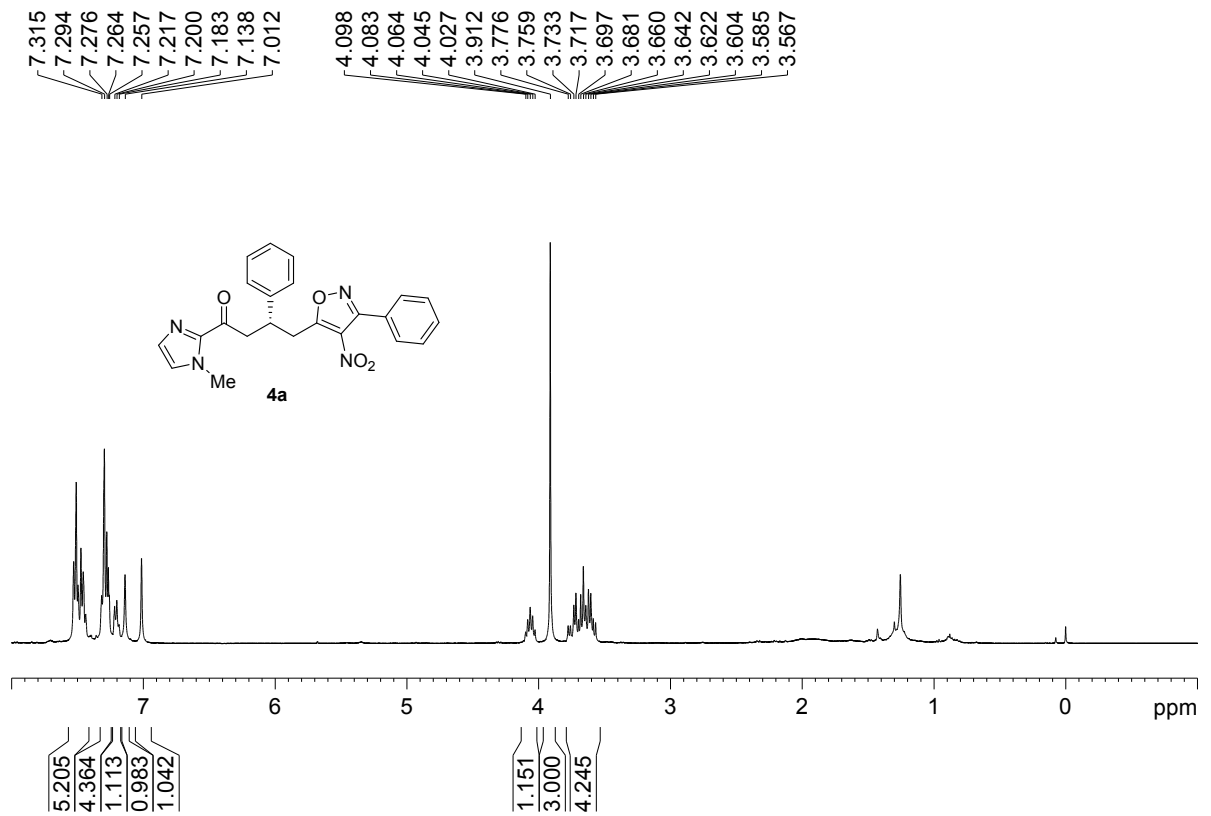
<sup>1</sup>H NMR (400 MHz) spectra of **3p**



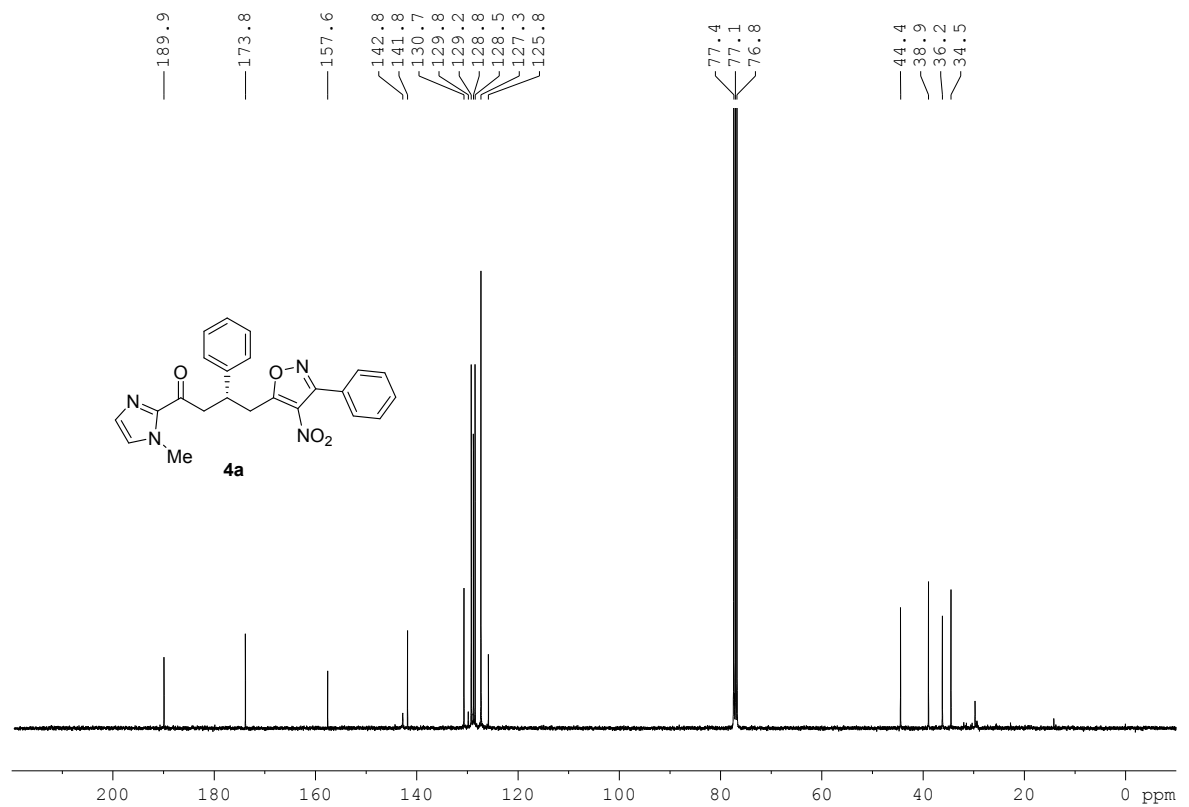
<sup>13</sup>C NMR (100 MHz) spectra of **3p**



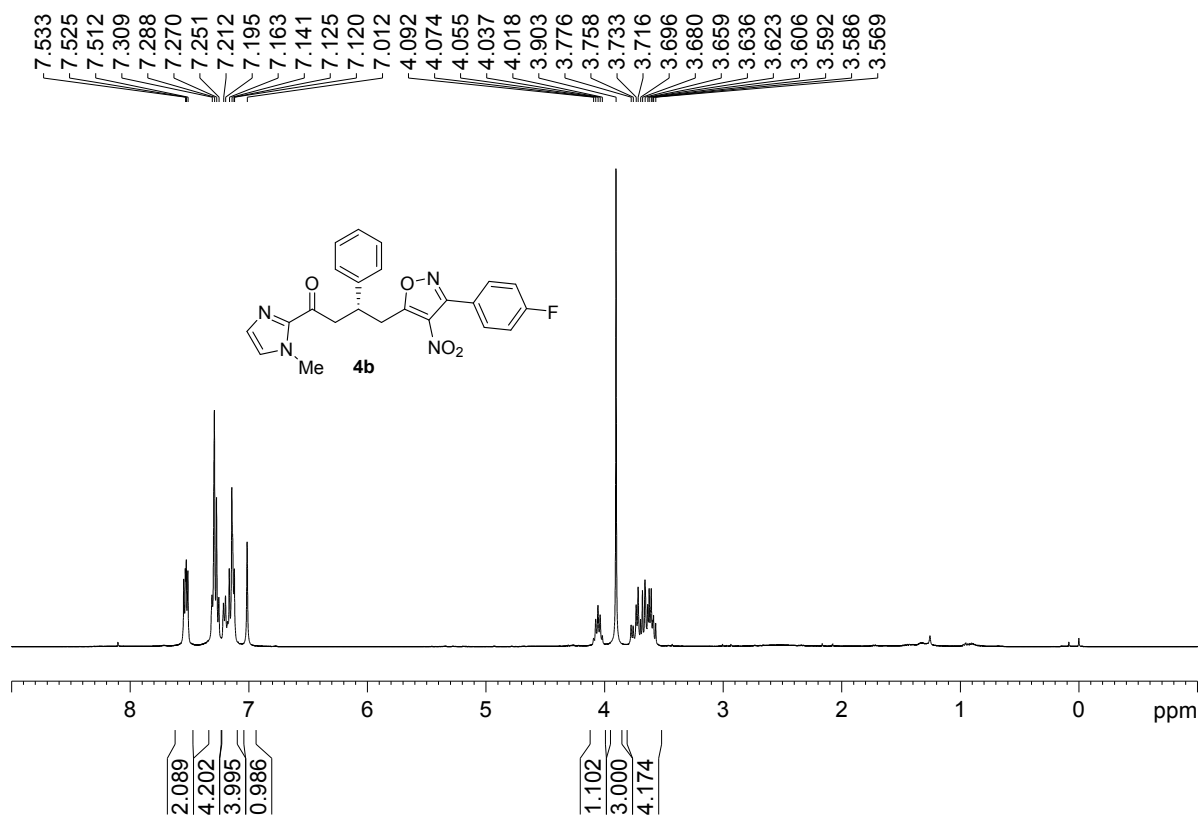
<sup>1</sup>H NMR (400 MHz) spectra of **4a**



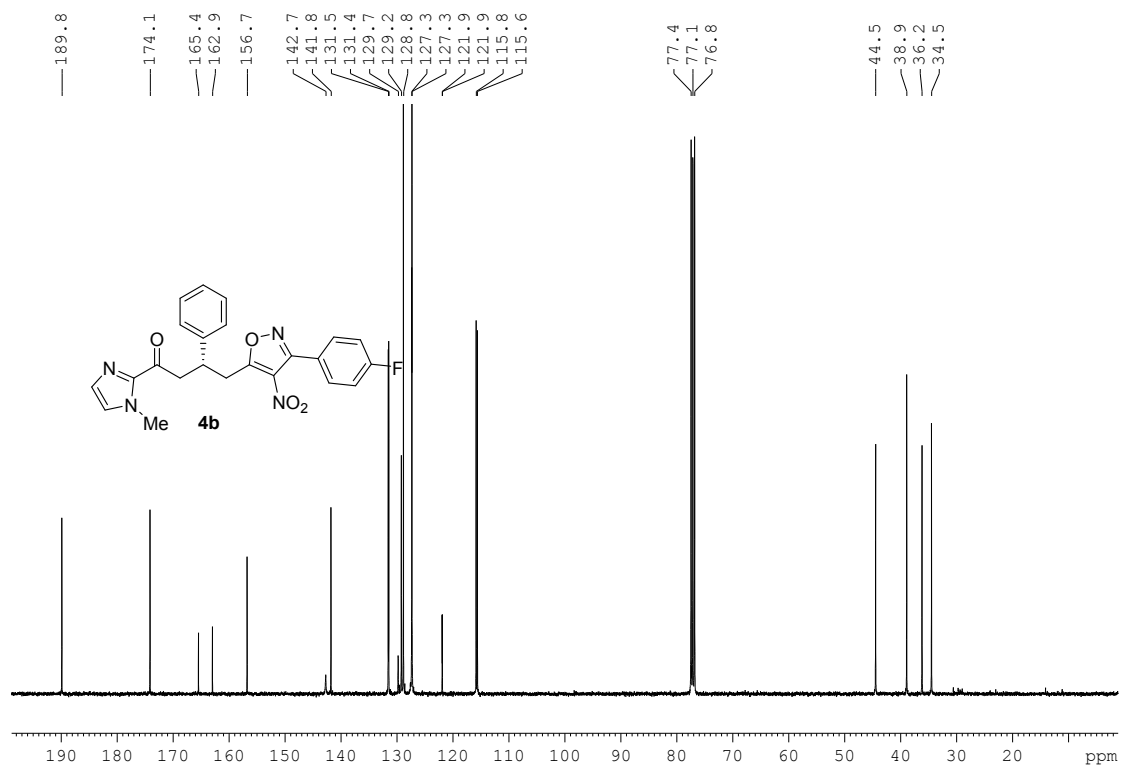
<sup>13</sup>C NMR (100 MHz) spectra of **4a**



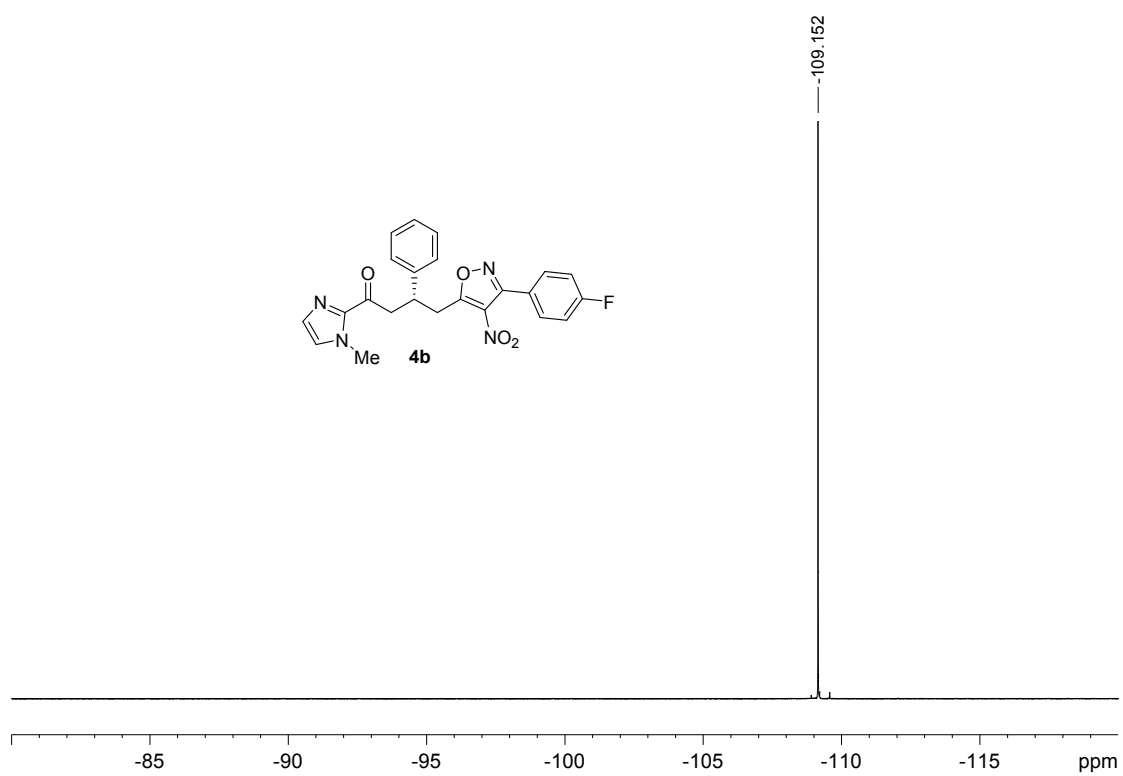
<sup>1</sup>H NMR (400 MHz) spectra of **4b**



<sup>13</sup>C NMR (100 MHz) spectra of **4b**

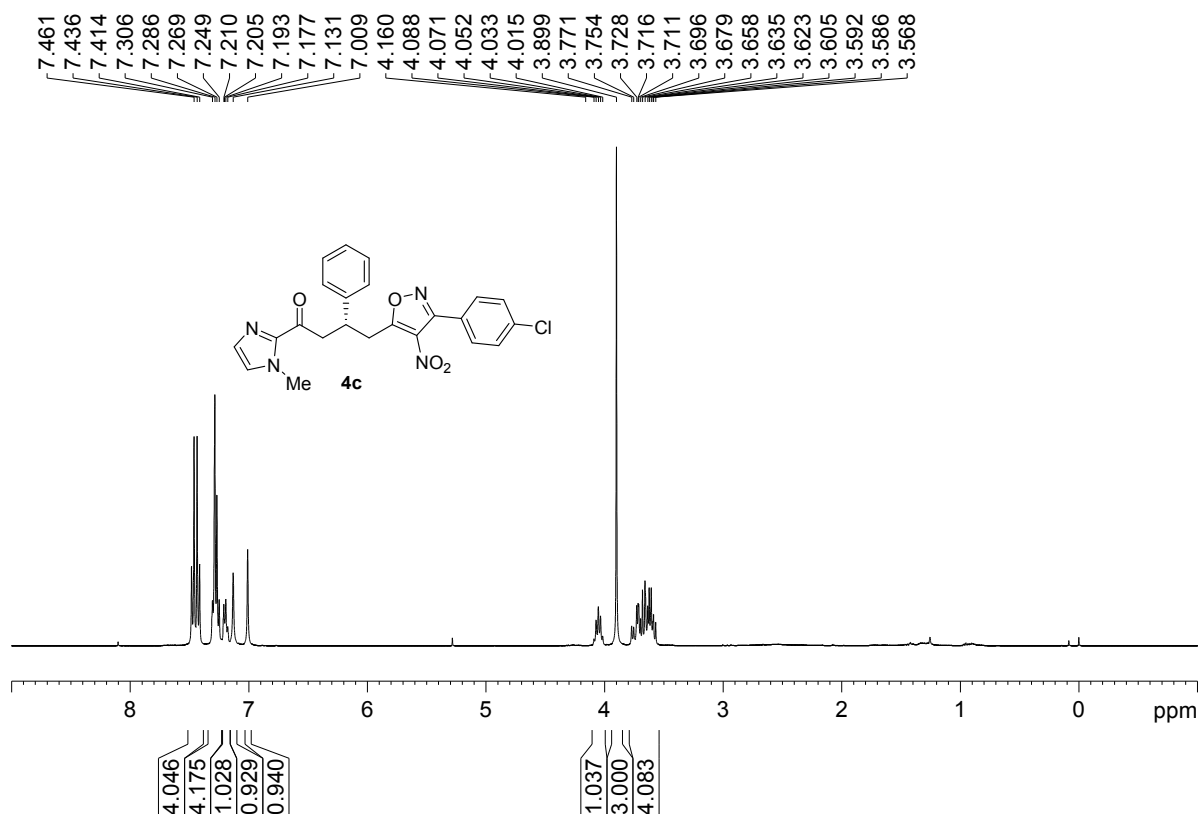


$^{19}\text{F}$  NMR (376.4 MHz) spectra of **4b**

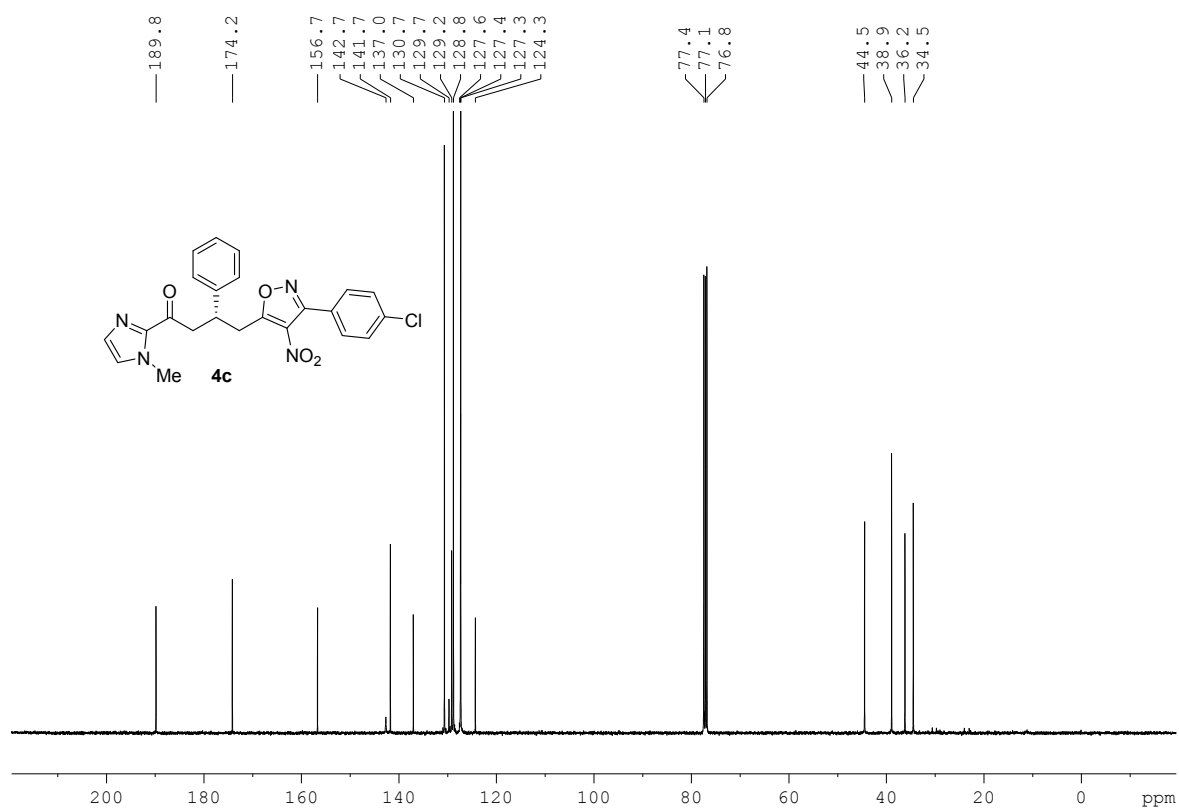




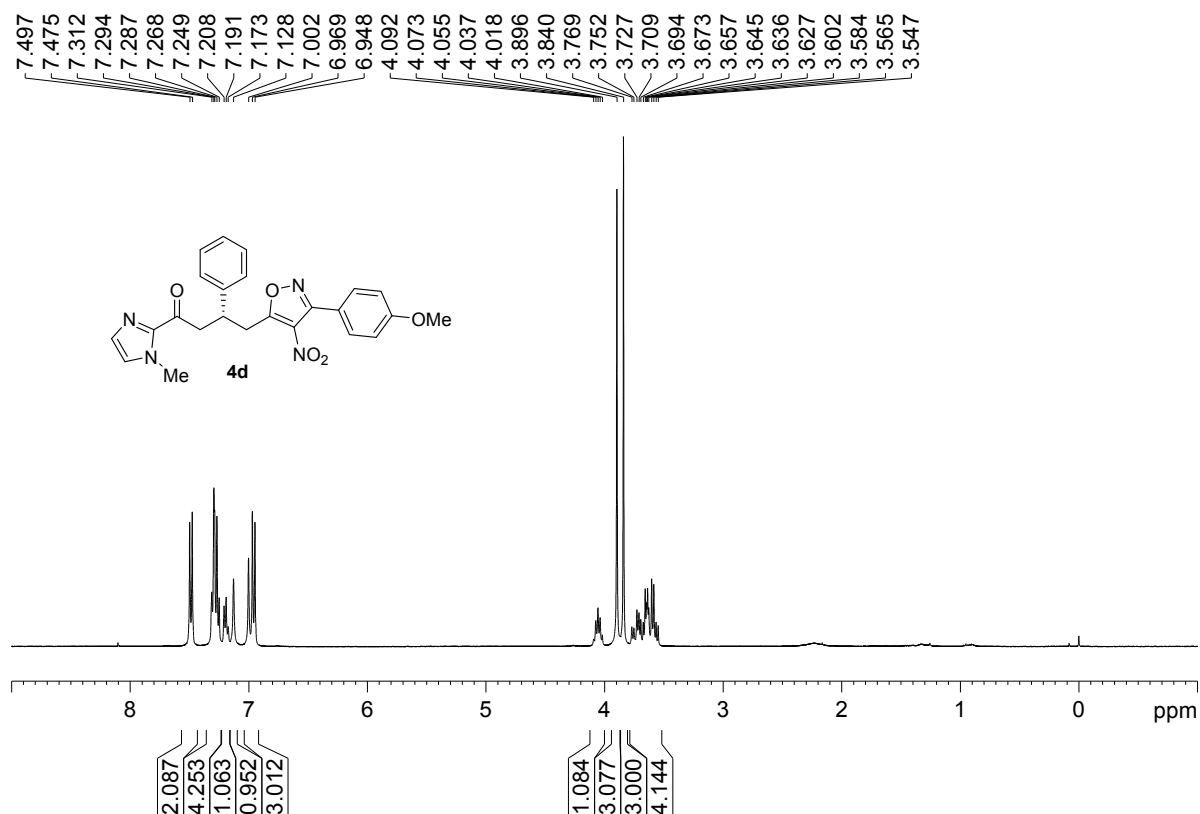
<sup>1</sup>H NMR (400 MHz) spectra of **4c**



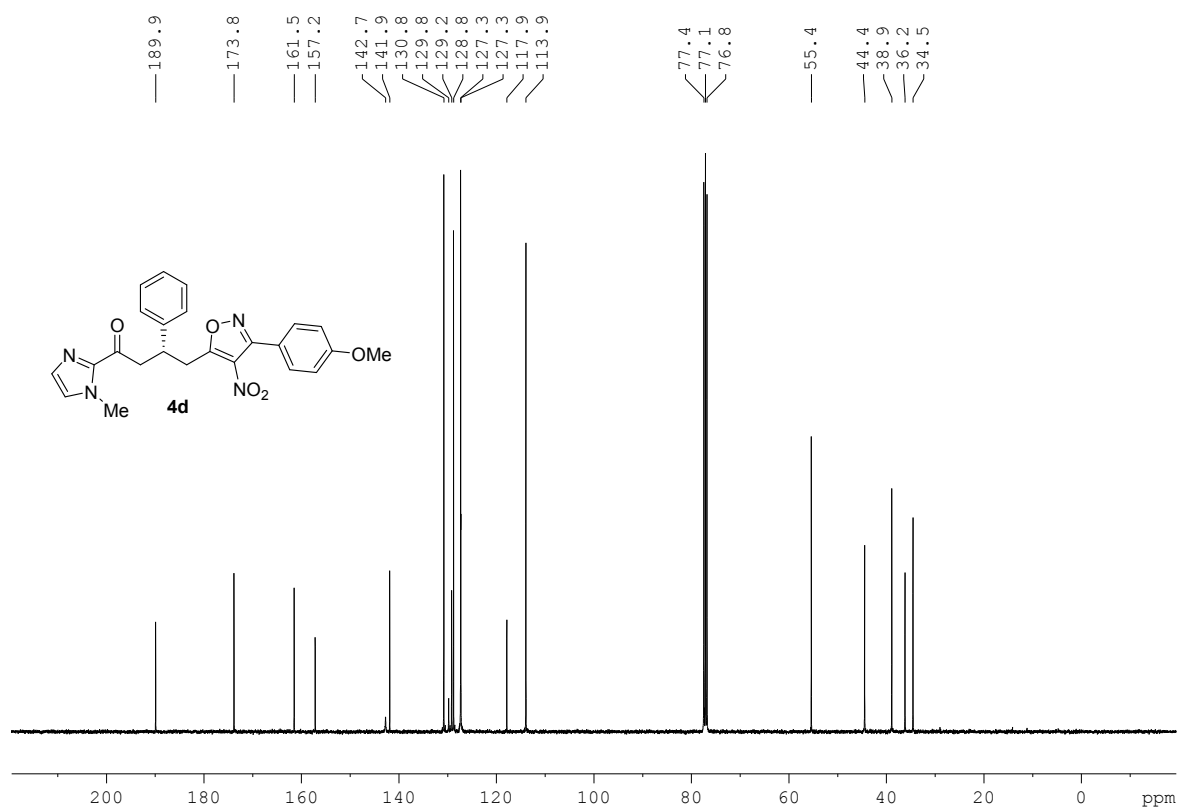
<sup>13</sup>C NMR (100 MHz) spectra of **4c**



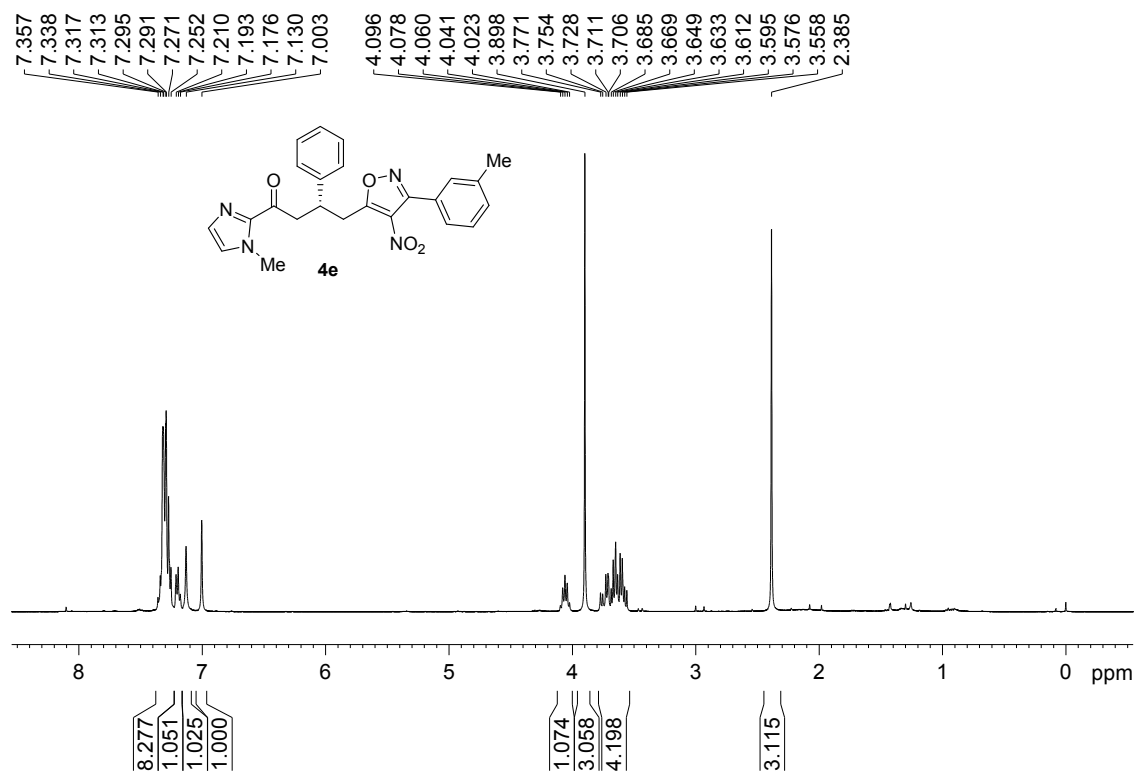
<sup>1</sup>H NMR (400 MHz) spectra of **4d**



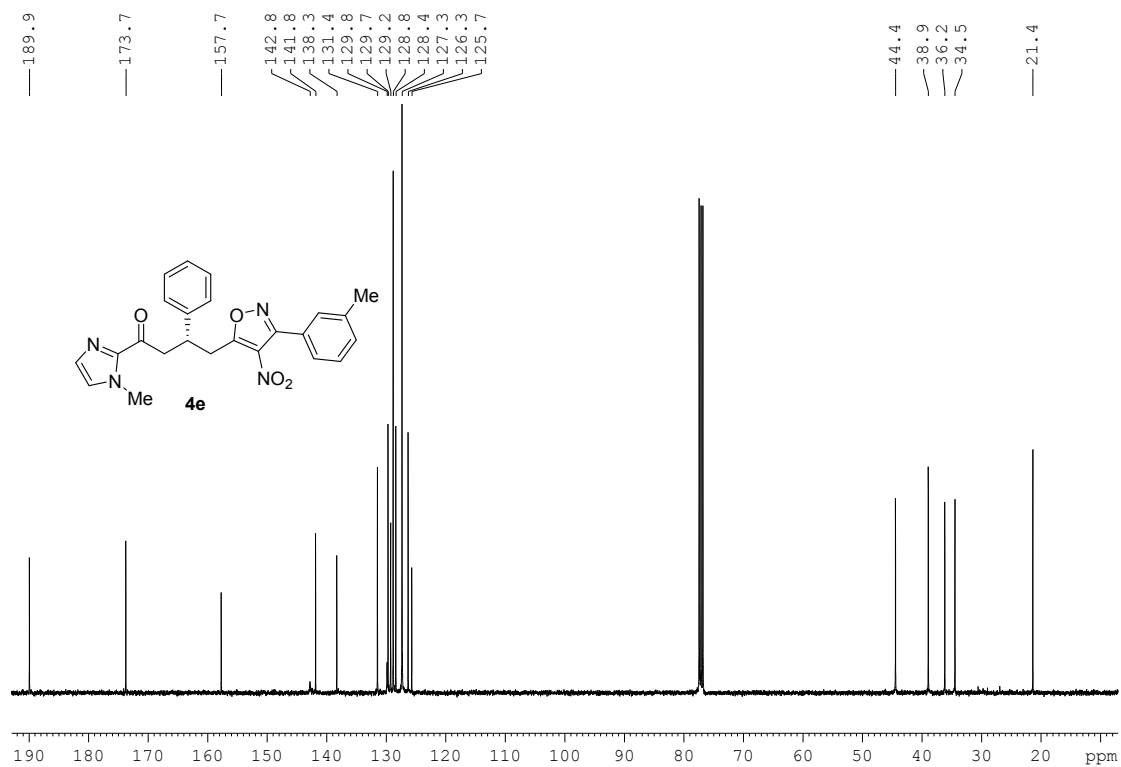
<sup>13</sup>C NMR (100 MHz) spectra of **4d**



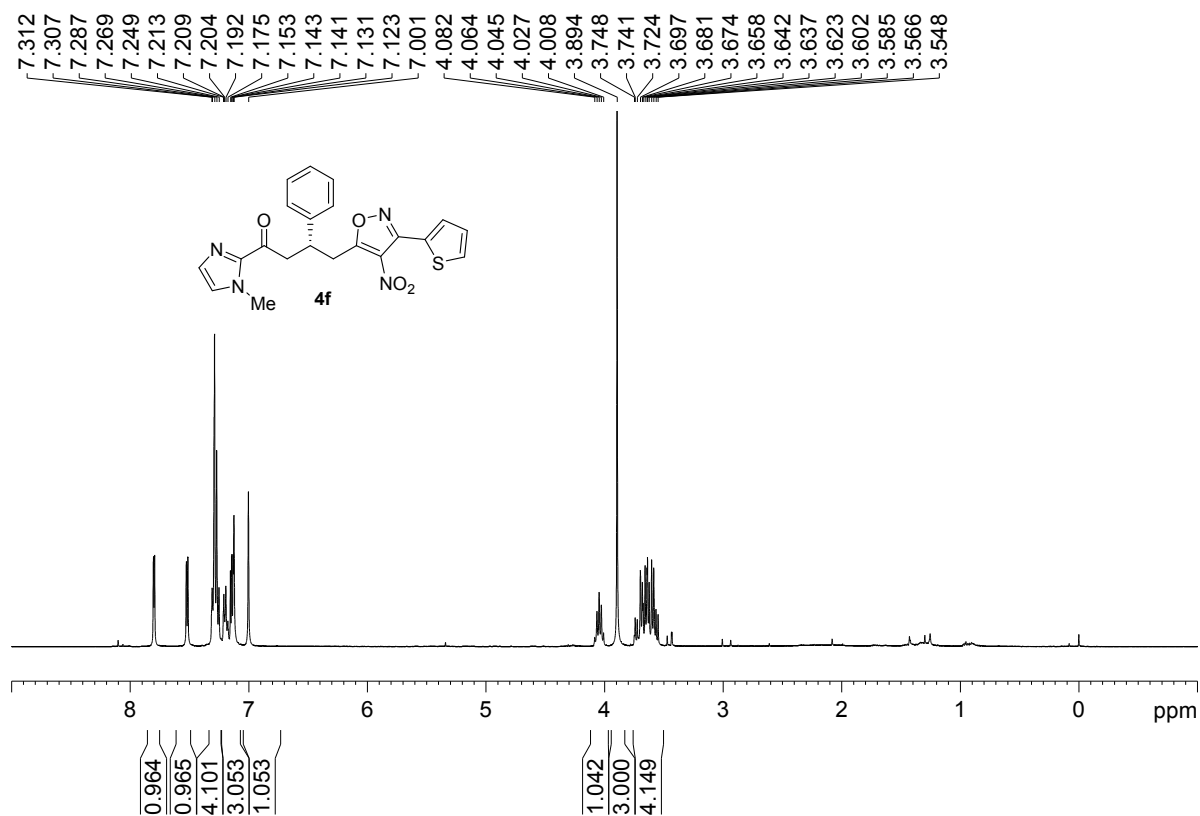
<sup>1</sup>H NMR (400 MHz) spectra of **4e**



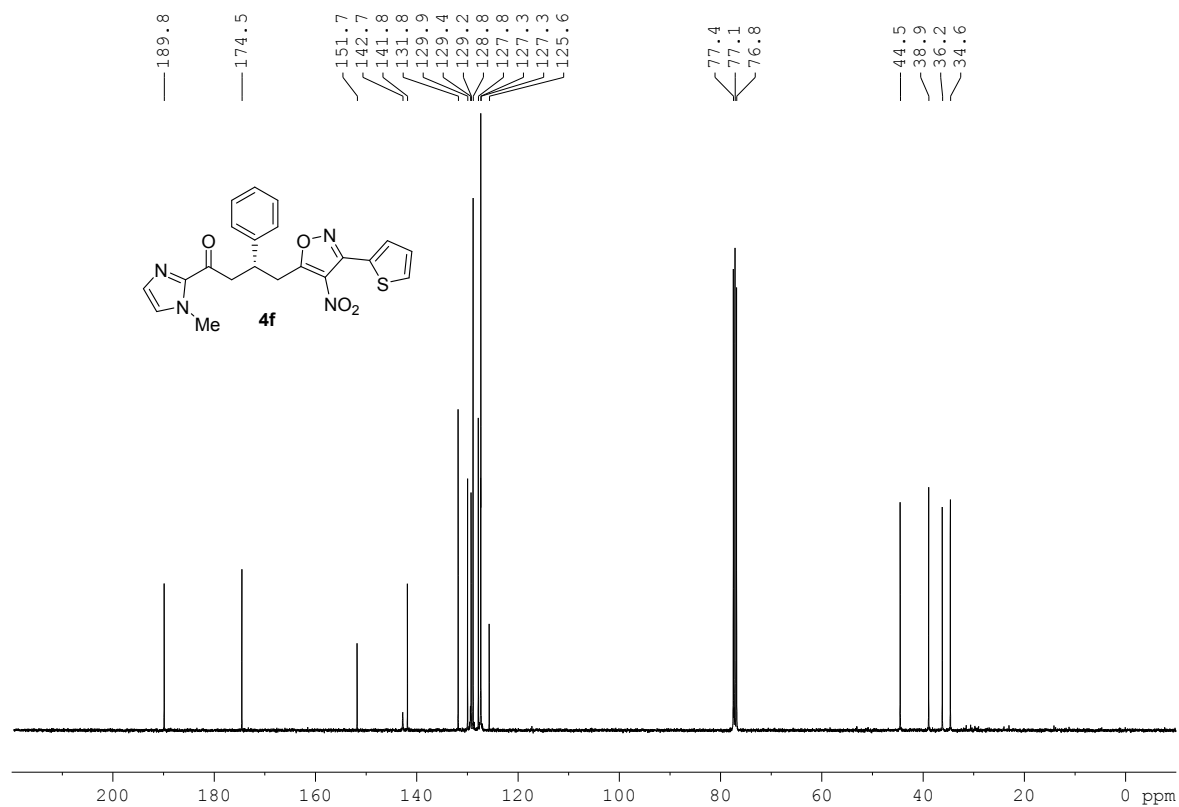
<sup>13</sup>C NMR (100 MHz) spectra of **4e**



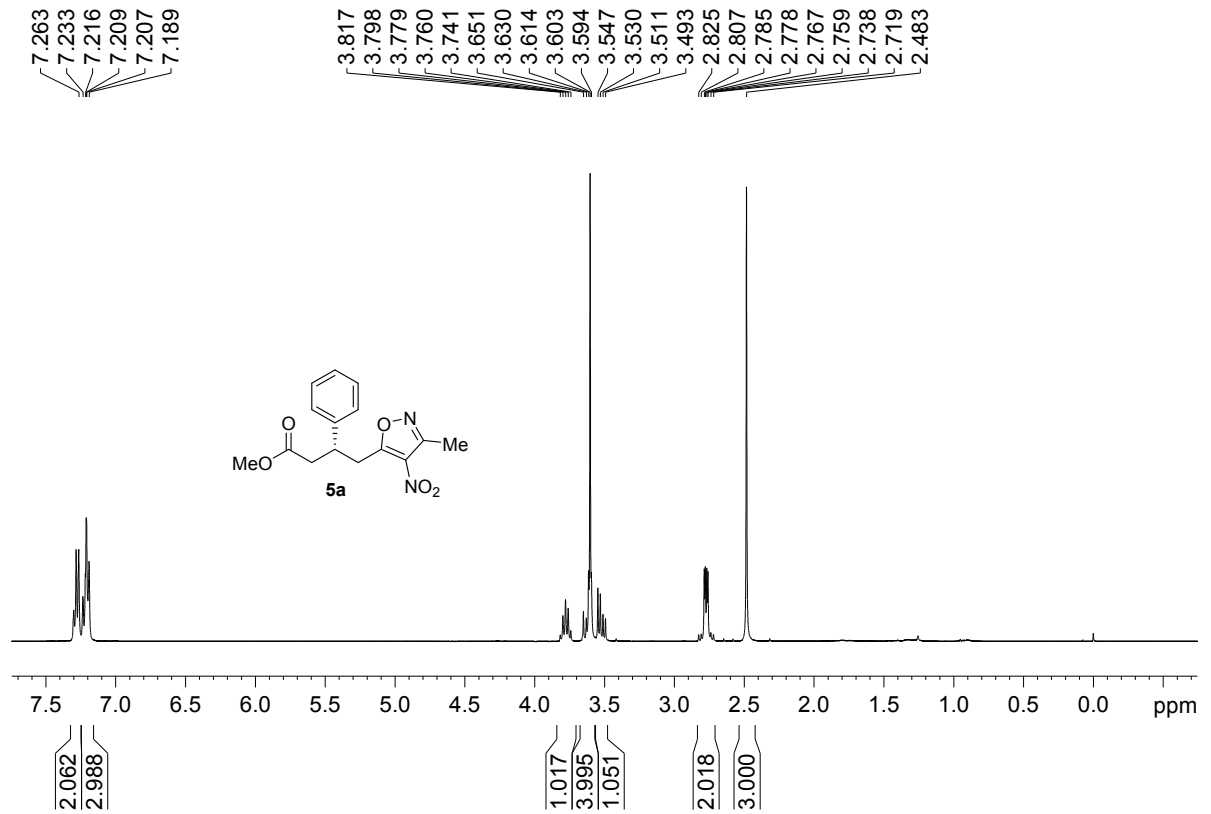
<sup>1</sup>H NMR (400 MHz) spectra of **4f**



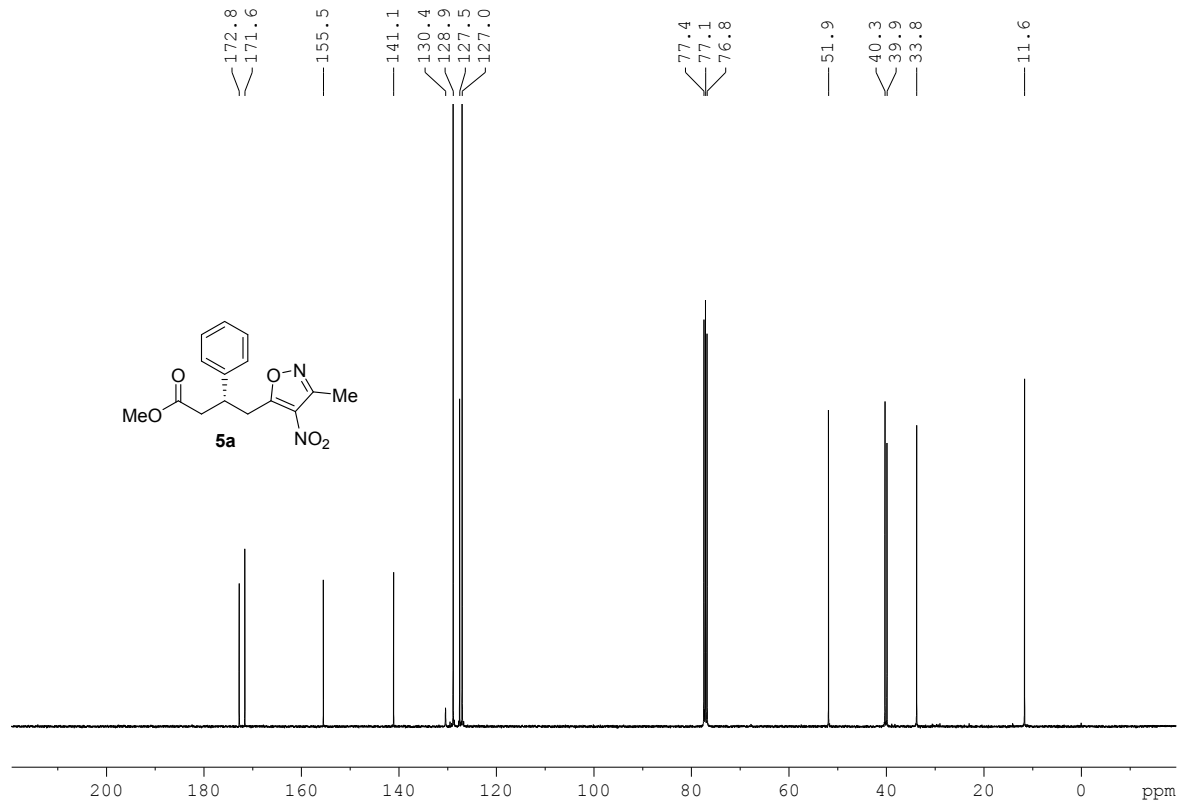
<sup>13</sup>C NMR (100 MHz) spectra of **4f**



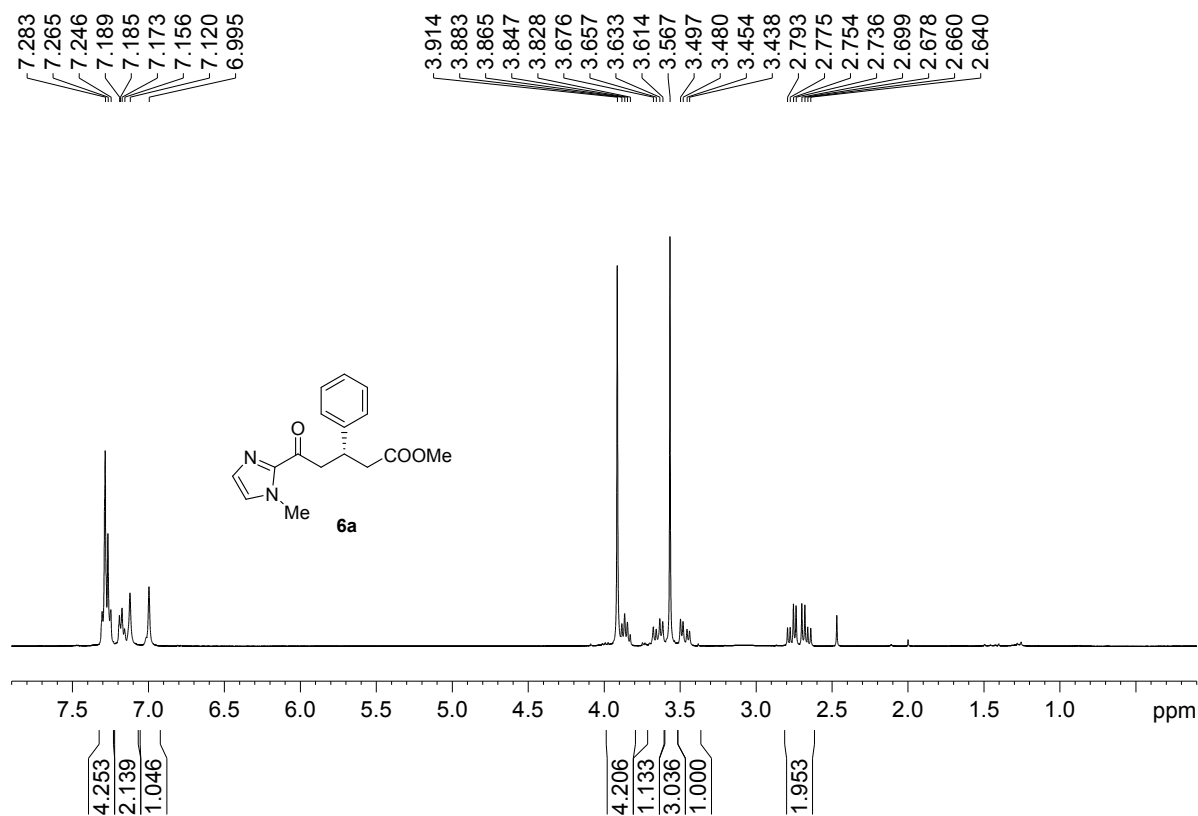
<sup>1</sup>H NMR (400 MHz) spectra of **5a**



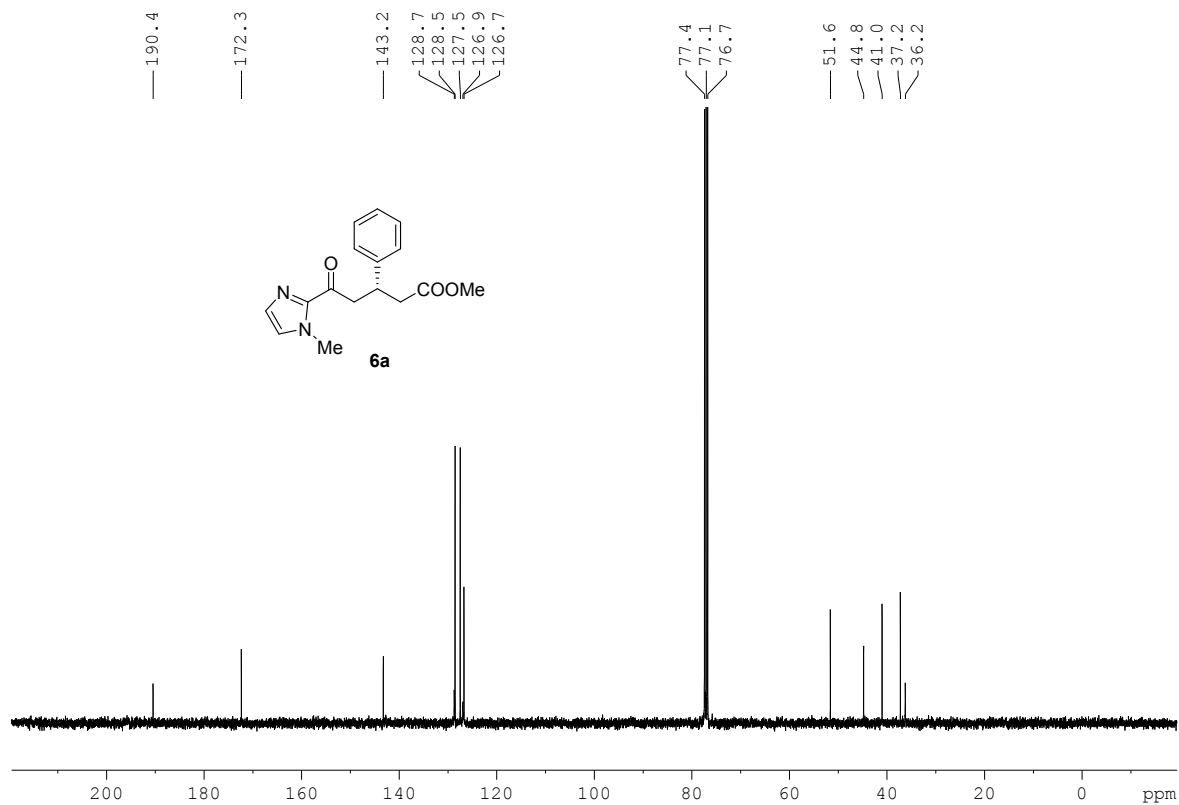
<sup>13</sup>C NMR (100 MHz) spectra of **5a**



<sup>1</sup>H NMR (400 MHz) spectra of **6a**



<sup>13</sup>C NMR (100 MHz) spectra of **6a**

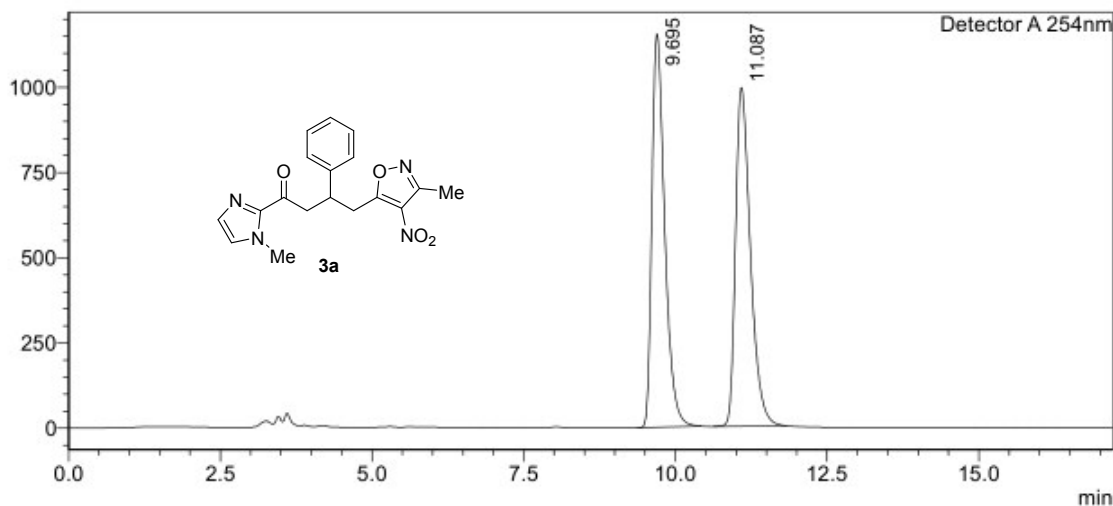


## VI Chiral HPLC trace analysis

### Racemic **3a**

#### <Chromatogram>

mV



#### <Peak Table>

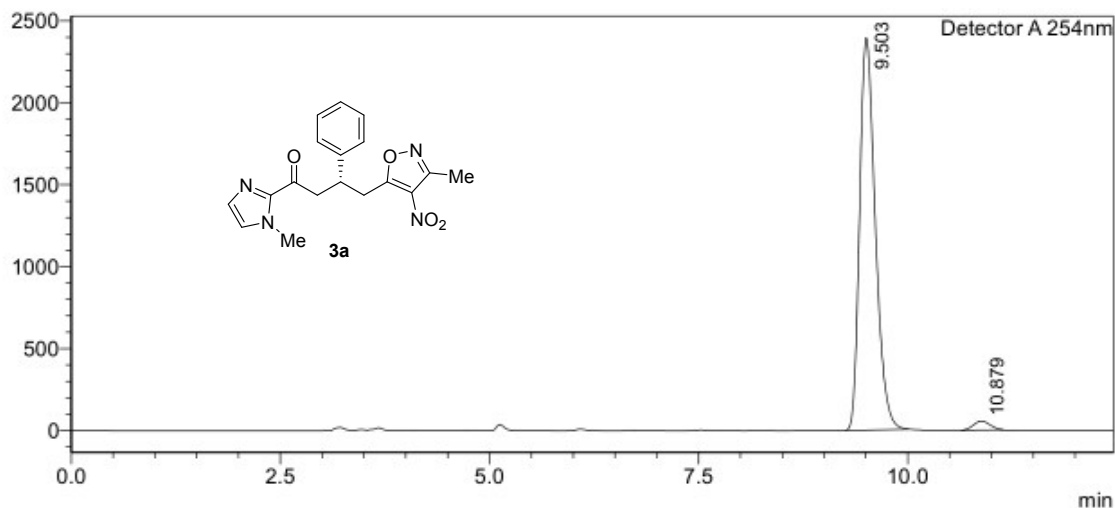
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.695	16955282	1153931	49.995		M	
2	11.087	16958842	993837	50.005		M	
Total		33914124	2147769				

### Chiral **3a**:

#### <Chromatogram>

mV



#### <Peak Table>

Detector A 254nm

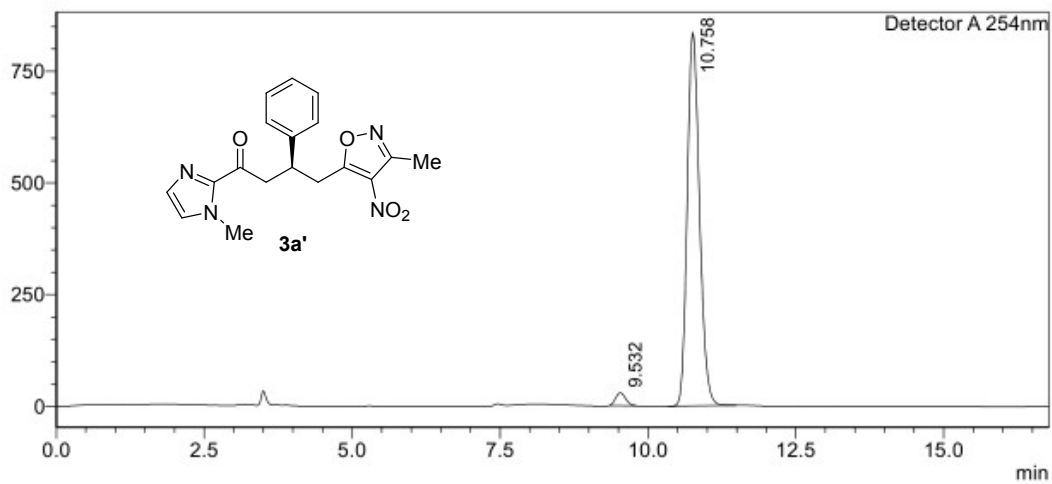
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.503	31145211	2391785	97.624		M	
2	10.879	757906	54922	2.376		M	
Total		31903117	2446707				

HPLC traces of racemic **3a** and chiral **3a**. Area integration = 95% ee.

## Chiral **3a'**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
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2	10.758	12040221	835239	97.271		M	
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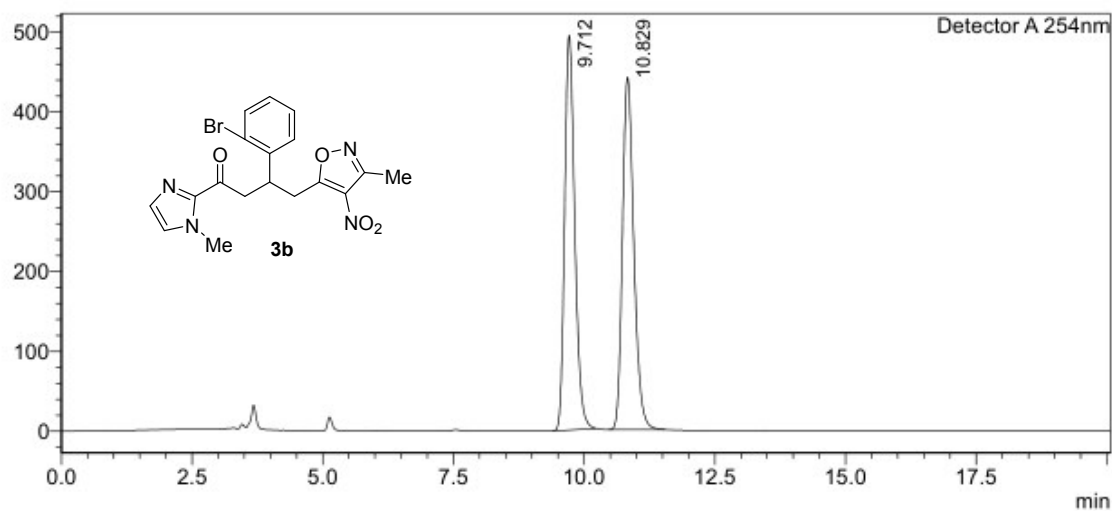
HPLC traces of racemic **3a** and chiral **3a'**. Area integration = 95% ee.



## Racemic **3b**

### <Chromatogram>

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### <Peak Table>

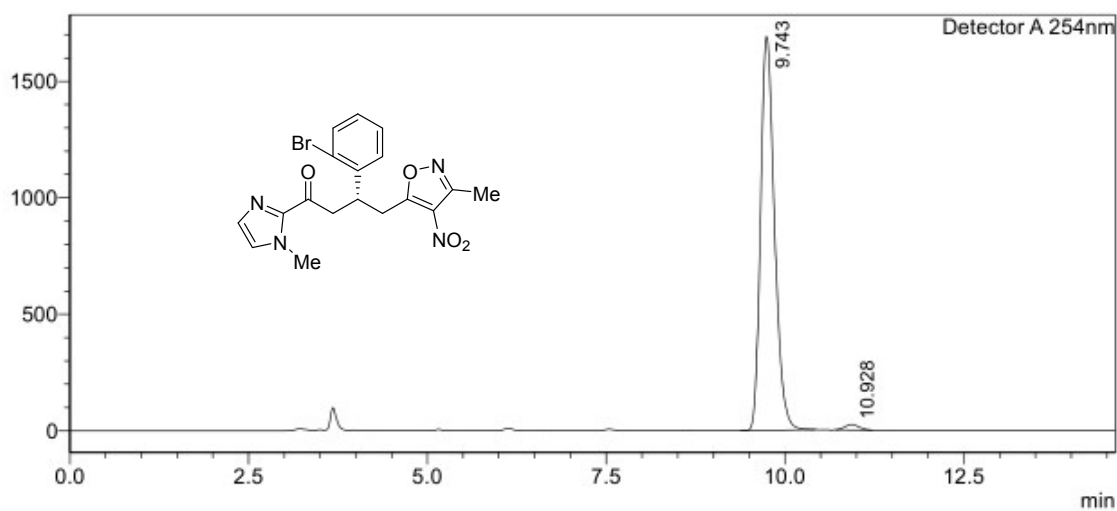
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.712	6653930	494288	49.835		M	
2	10.829	6698012	441259	50.165		M	
Total		13351942	935547				

## Chiral **3b**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

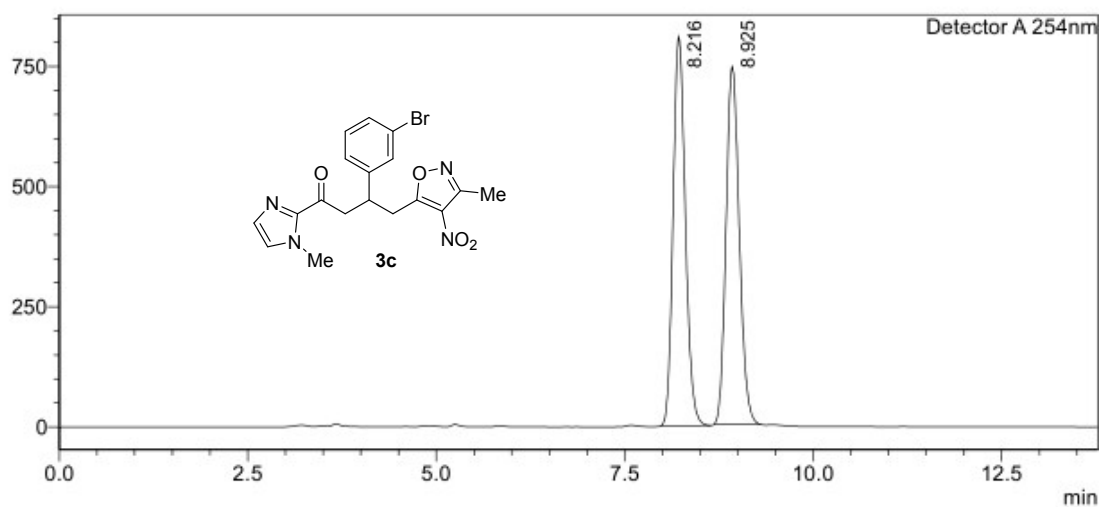
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.743	22915072	1689189	98.624		M	
2	10.928	319774	22953	1.376		M	
Total		23234846	1712142				

HPLC traces of racemic **3b** and chiral **3b**. Area integration = 97% ee.

## Racemic **3c**

### <Chromatogram>

mV



### <Peak Table>

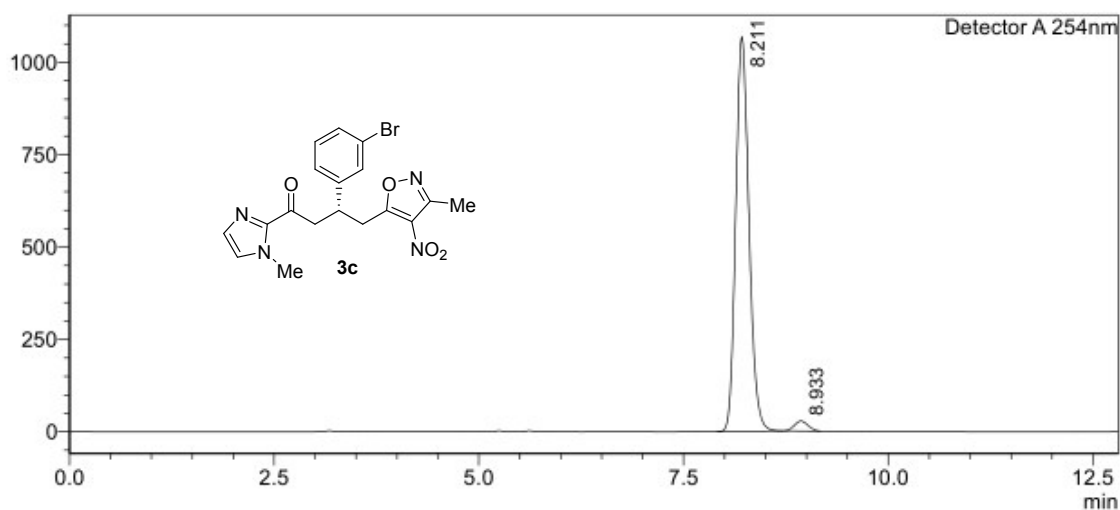
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.216	9177025	809348	50.162		M	
2	8.925	9117879	743878	49.838		M	
Total		18294904	1553226				

## Chiral **3c**:

### <Chromatogram>

mV



### <Peak Table>

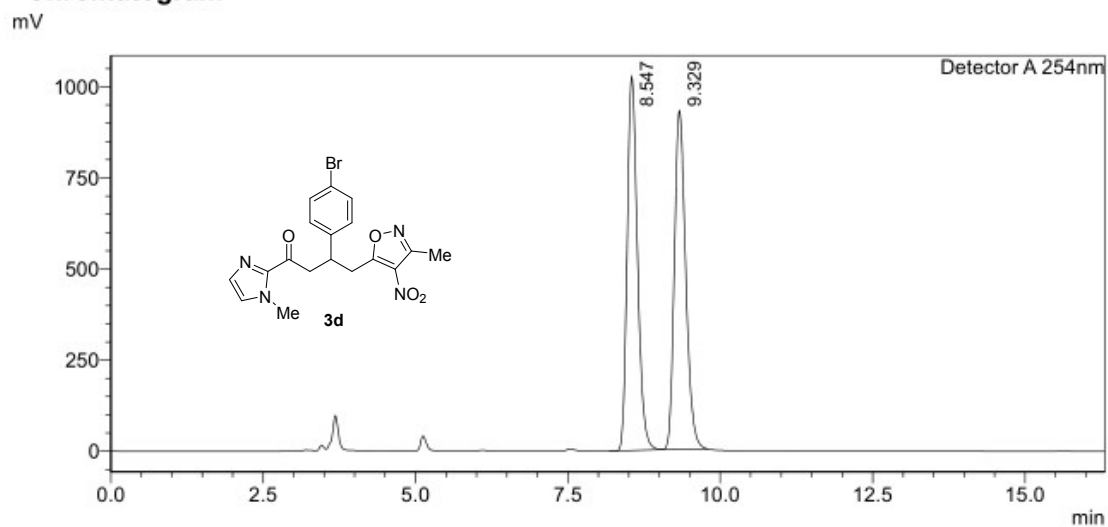
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.211	12042807	1067304	97.495		M	
2	8.933	309397	26922	2.505		M	
Total		12352204	1094225				

HPLC traces of racemic **3c** and chiral **3c**. Area integration = 95% ee.

## Racemic **3d**

### <Chromatogram>

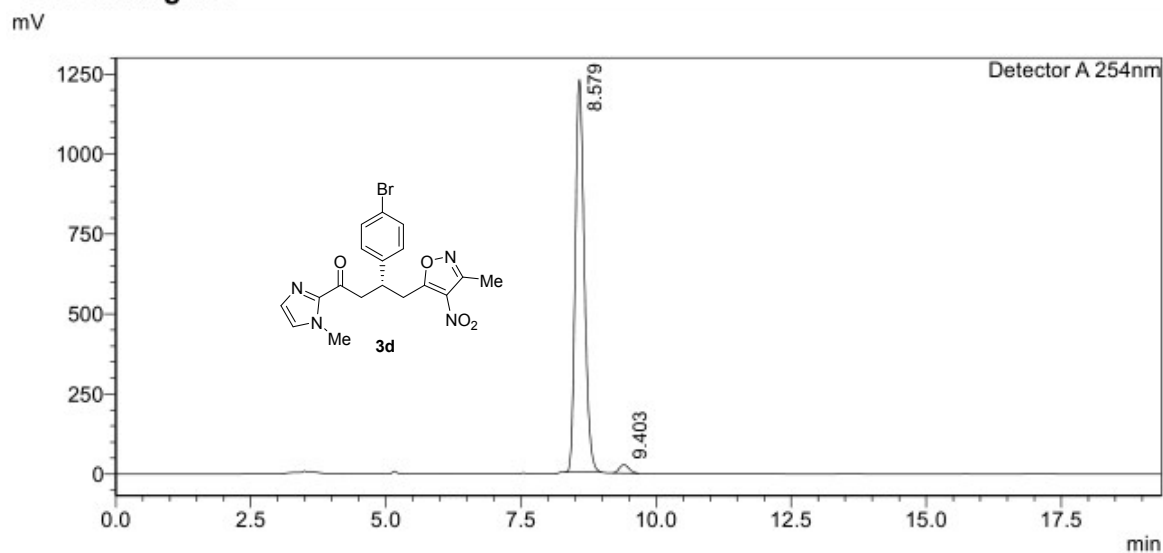


### <Peak Table>

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.547	12041027	1026400	50.112		M	
2	9.329	11987310	929522	49.888		M	
Total		24028337	1955923				

## Chiral **3d**:

### <Chromatogram>



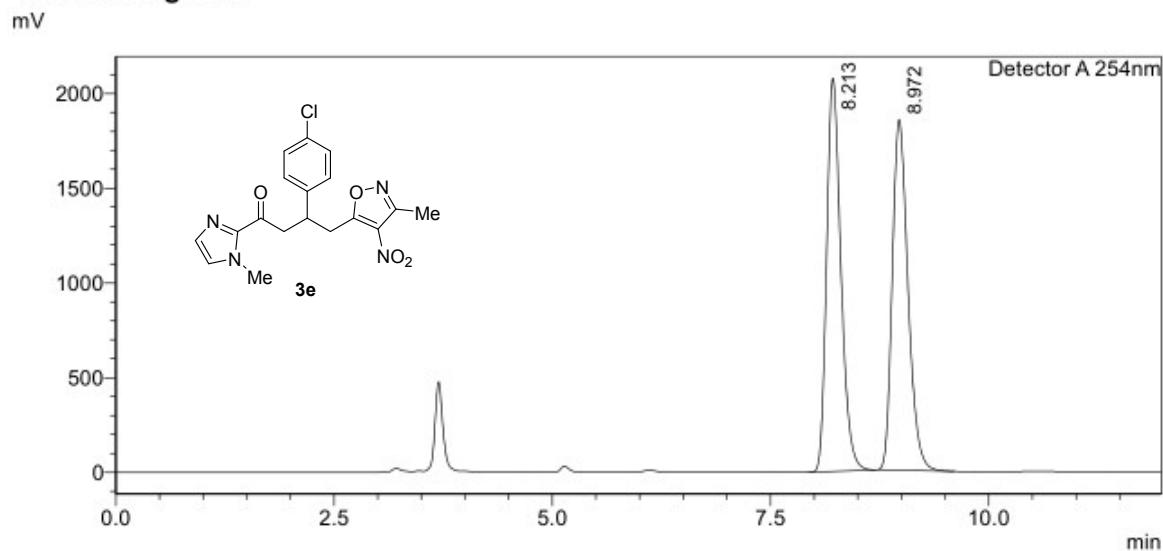
### <Peak Table>

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.579	14272337	1225708	97.779		M	
2	9.403	324251	26514	2.221		M	
Total		14596588	1252221				

HPLC traces of racemic **3d** and chiral **3d**. Area integration = 96% ee.

## Racemic **3e**

### <Chromatogram>



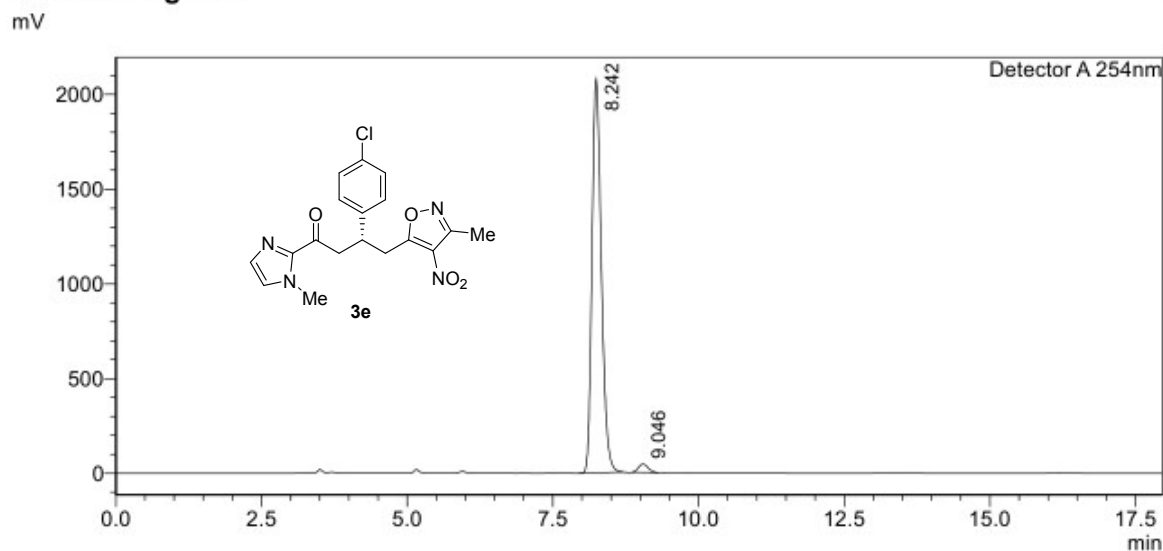
### <Peak Table>

Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.213	23522410	2074775	50.446		M	
2	8.972	23106479	1850310	49.554		M	
Total		46628889	3925085				

## Chiral **3e**:

### <Chromatogram>



### <Peak Table>

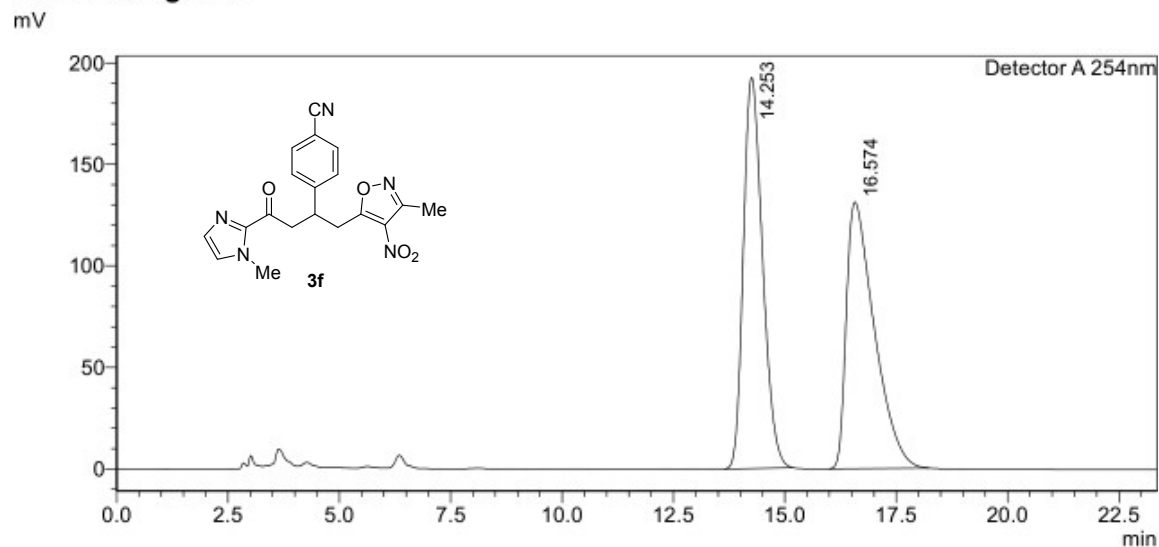
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.242	23252587	2077797	97.879		M	
2	9.046	503940	43597	2.121		M	
Total		23756527	2121394				

HPLC traces of racemic **3e** and chiral **3e**. Area integration = 96% ee.

## Racemic **3f**

### <Chromatogram>

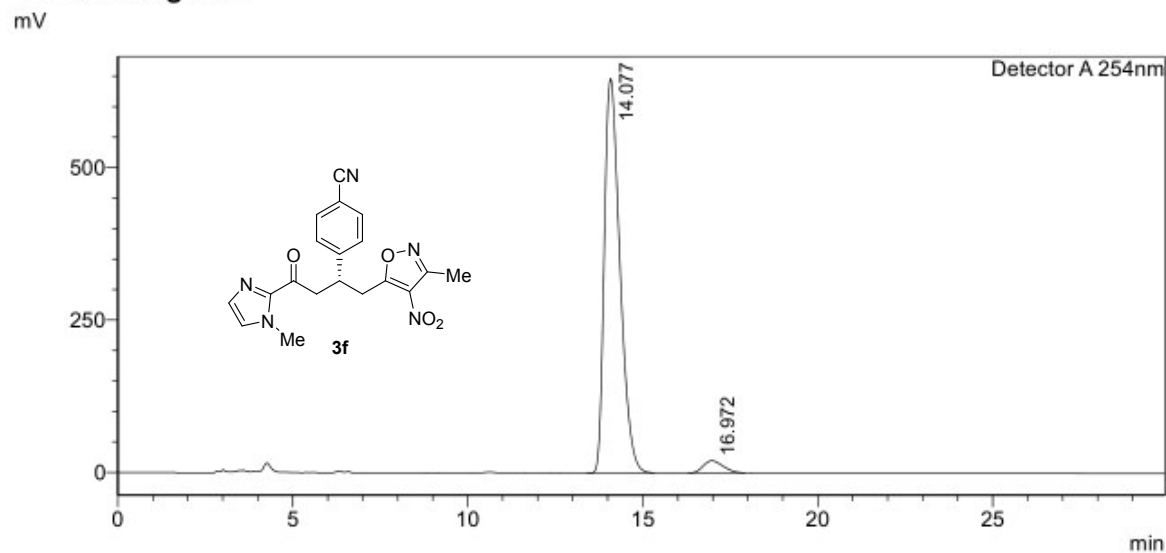


### <Peak Table>

Detector A 254nm							
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	14.253	5754970	192435	50.040		M	
2	16.574	5745712	131208	49.960		M	
Total		11500682	323643				

## Chiral **3f**:

### <Chromatogram>



### <Peak Table>

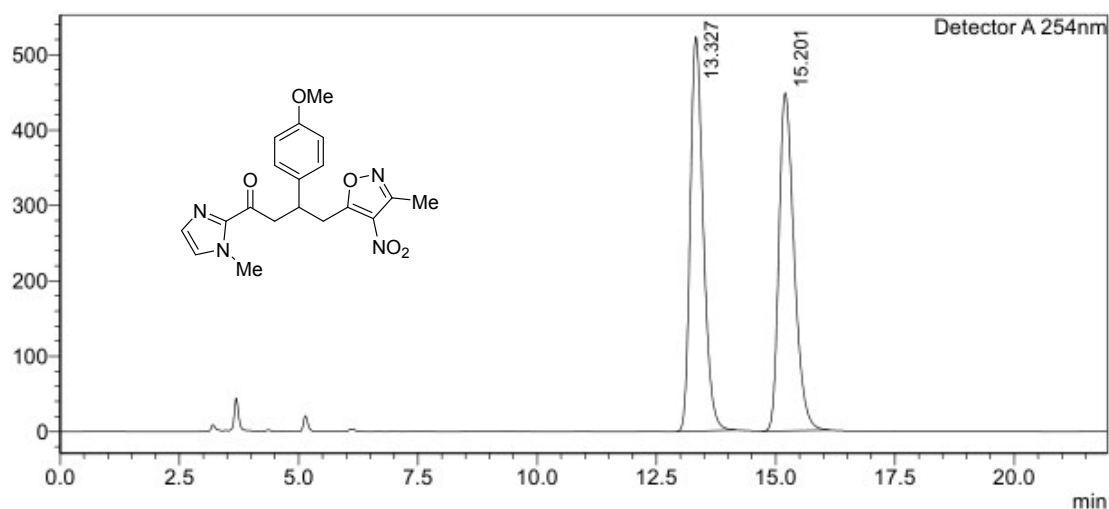
Detector A 254nm							
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	14.077	19971644	647547	96.146		M	
2	16.972	800481	20071	3.854		M	
Total		20772126	667618				

HPLC traces of racemic **3f** and chiral **3f**. Area integration = 92% ee.

## Racemic **3g**

### <Chromatogram>

mV



### <Peak Table>

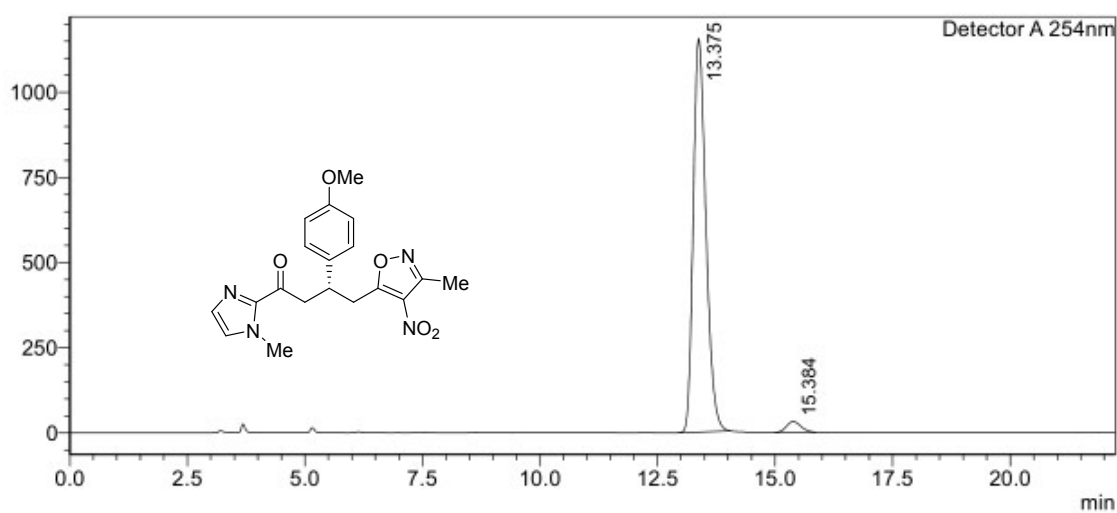
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	13.327	9922123	522787	49.955		M	
2	15.201	9940127	447939	50.045		M	
Total		19862249	970726				

## Chiral **3g**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

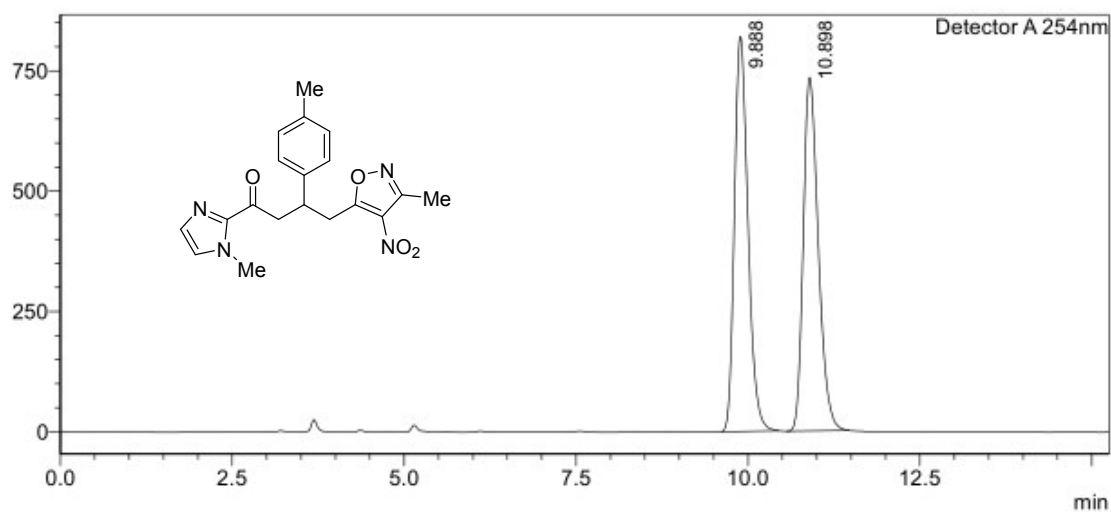
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	13.375	21695310	1155453	96.959		M	
2	15.384	680473	31750	3.041		M	
Total		22375782	1187202				

HPLC traces of racemic **3g** and chiral **3g**. Area integration = 94% ee.

### Racemic **3h**

#### <Chromatogram>

mV



#### <Peak Table>

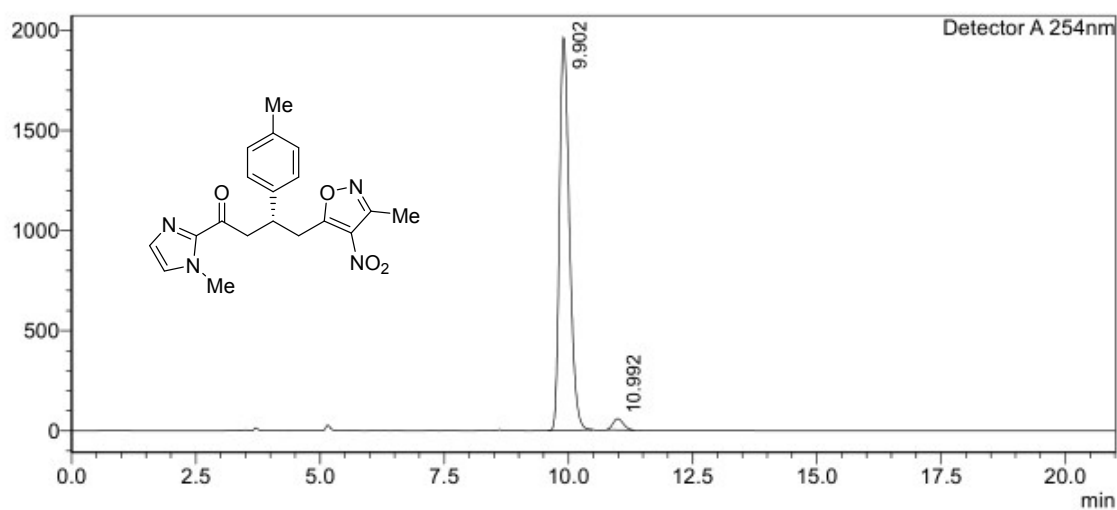
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.888	11186002	819737	49.981		M	
2	10.898	11194686	734463	50.019		M	
Total		22380688	1554200				

### Chiral **3h**:

#### <Chromatogram>

mV



#### <Peak Table>

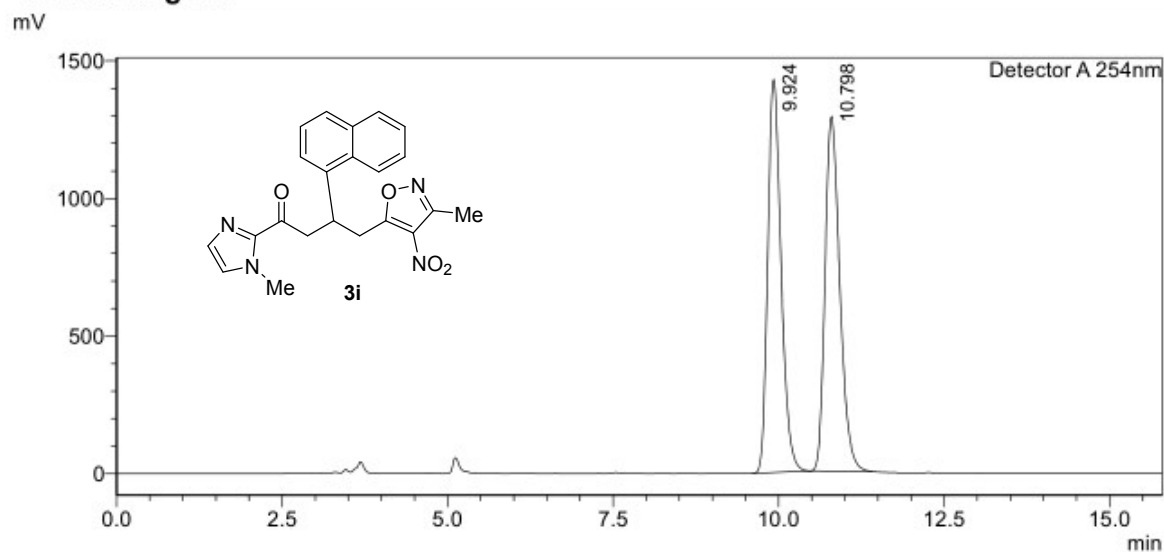
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.902	27122716	1960924	97.116		M	
2	10.992	805399	55903	2.884		M	
Total		27928115	2016827				

HPLC traces of racemic **3h** and chiral **3h**. Area integration = 94% ee.

## Racemic **3i**

### <Chromatogram>



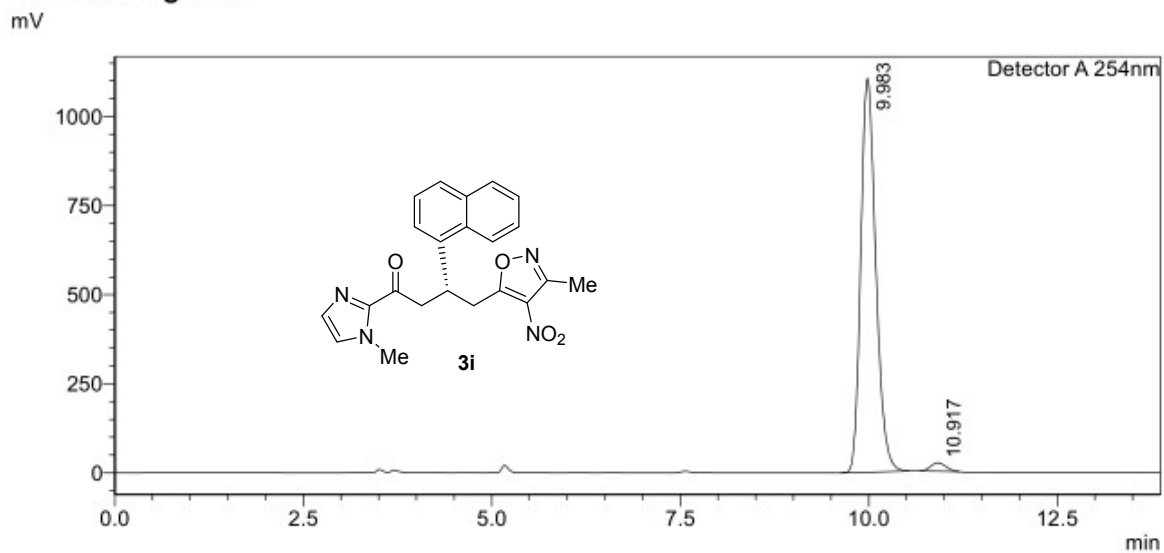
### <Peak Table>

Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.924	20026174	1427749	50.043		M	
2	10.798	19991830	1288658	49.957		M	
Total		40018004	2716407				

## Chiral **3i**:

### <Chromatogram>



### <Peak Table>

Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.983	15347054	1104425	97.845		M	
2	10.917	338001	23830	2.155		M	
Total		15685055	1128255				

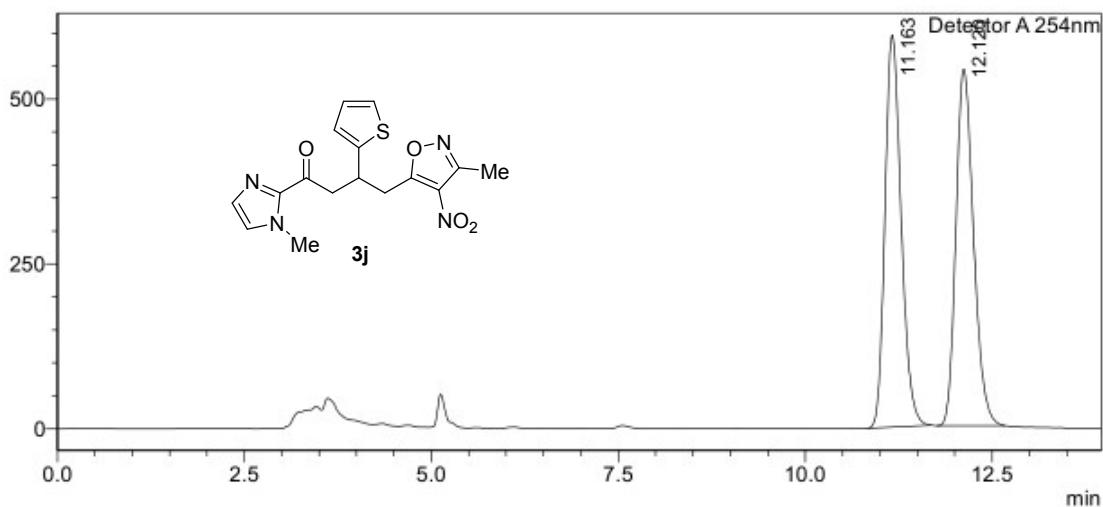
HPLC traces of racemic **3i** and chiral **3i**. Area integration = 96% ee.



## Racemic **3j**

### <Chromatogram>

mV



### <Peak Table>

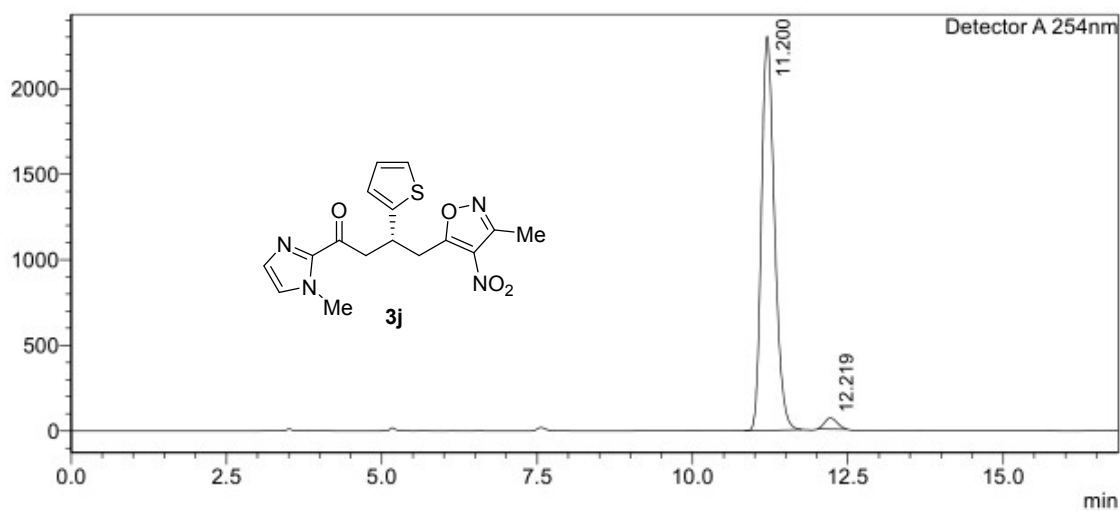
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	11.163	8937292	594584	50.129		M	
2	12.120	8891338	541169	49.871		M	
Total		17828630	1135753				

## Chiral **3j**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

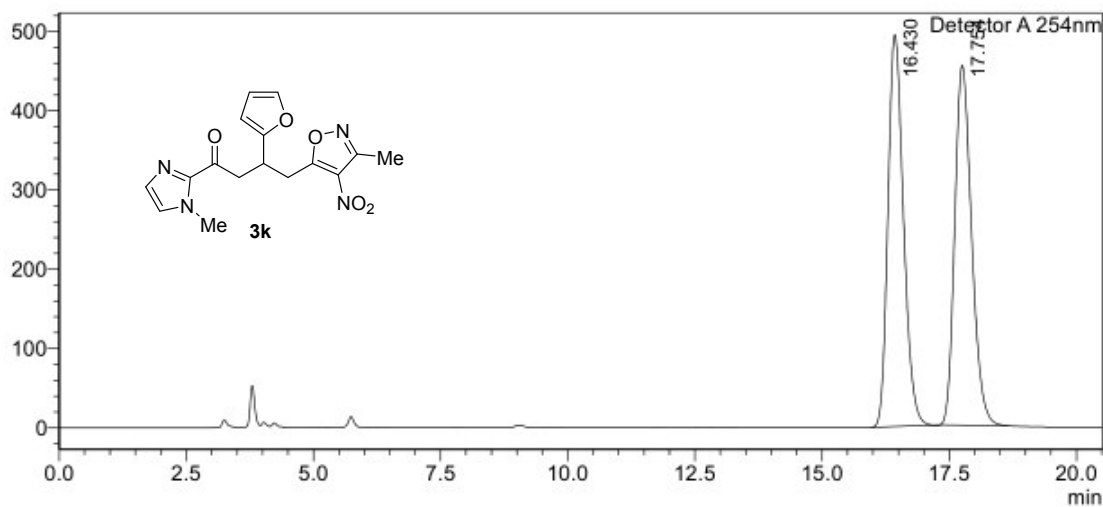
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	11.200	33729975	2300888	97.302		M	
2	12.219	935358	65509	2.698		M	
Total		34665333	2366396				

HPLC traces of racemic **3j** and chiral **3j**. Area integration = 95% ee.

## Racemic **3k**

### <Chromatogram>

mV



### <Peak Table>

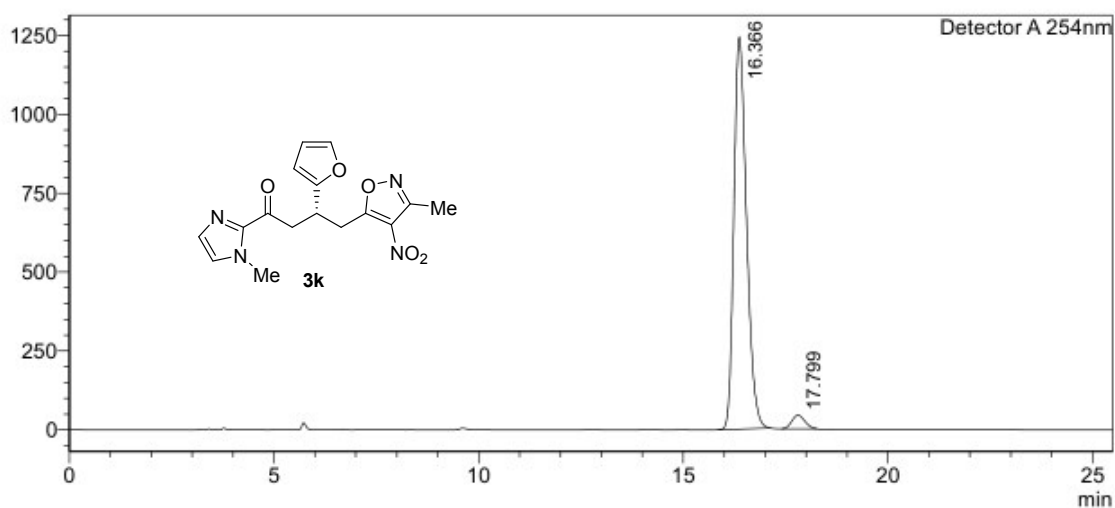
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	16.430	10483783	494062	50.059		M	
2	17.754	10459034	453586	49.941		M	
Total		20942816	947648				

## Chiral **3k**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

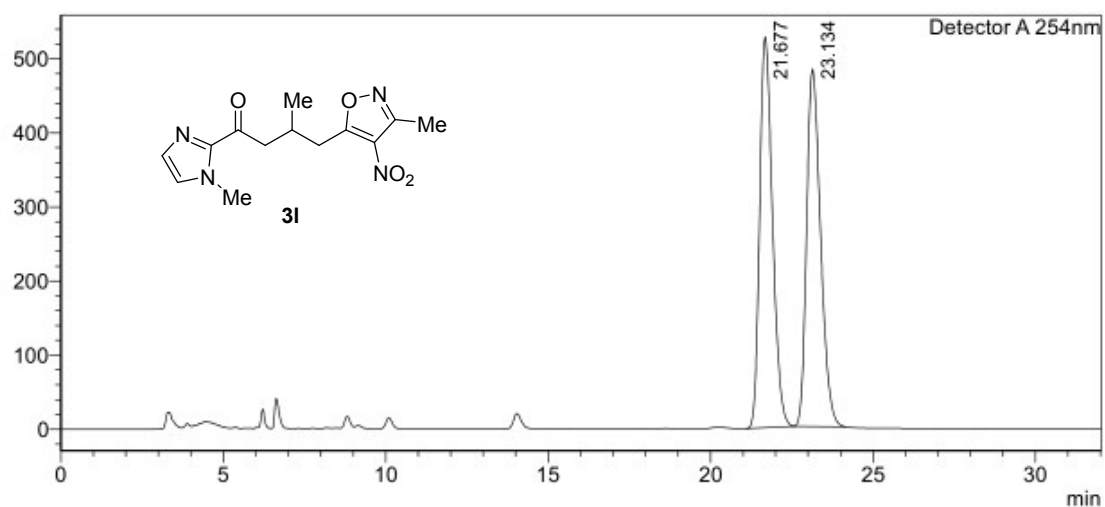
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	16.366	26696099	1242572	96.521		M	
2	17.799	962190	43305	3.479		M	
Total		27658289	1285877				

HPLC traces of racemic **3k** and chiral **3k**. Area integration = 93% ee.

## Racemic **3I**

### <Chromatogram>

mV



### <Peak Table>

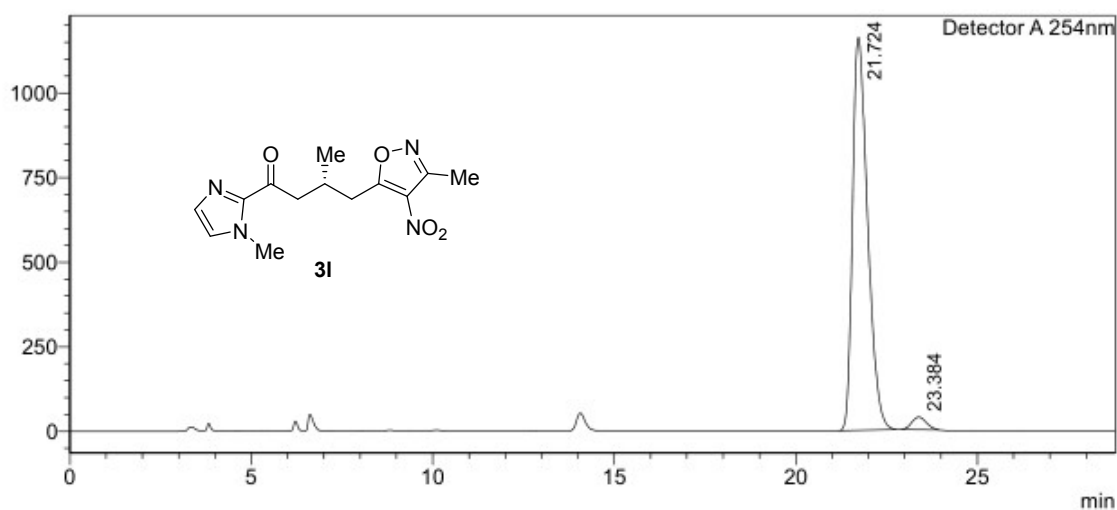
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	21.677	14421247	527230	50.086		M	
2	23.134	14371569	481769	49.914		M	
Total		28792816	1008999				

## Chiral **3I**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

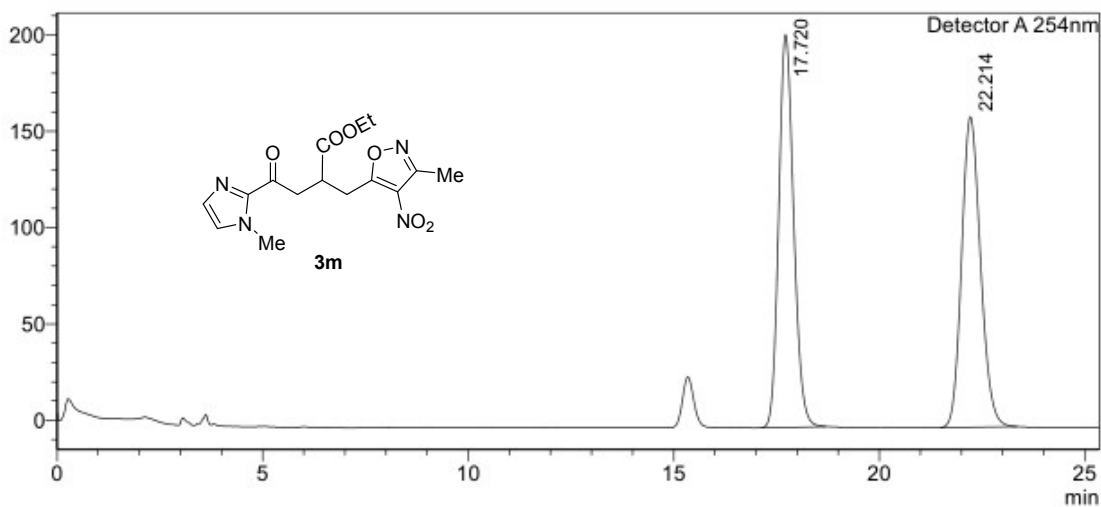
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	21.724	33507590	1160794	96.989		M	
2	23.384	1040204	37068	3.011		M	
Total		34547794	1197862				

HPLC traces of racemic **3I** and chiral **3I**. Area integration = 94% ee.

## Racemic **3m**

### <Chromatogram>

mV



### <Peak Table>

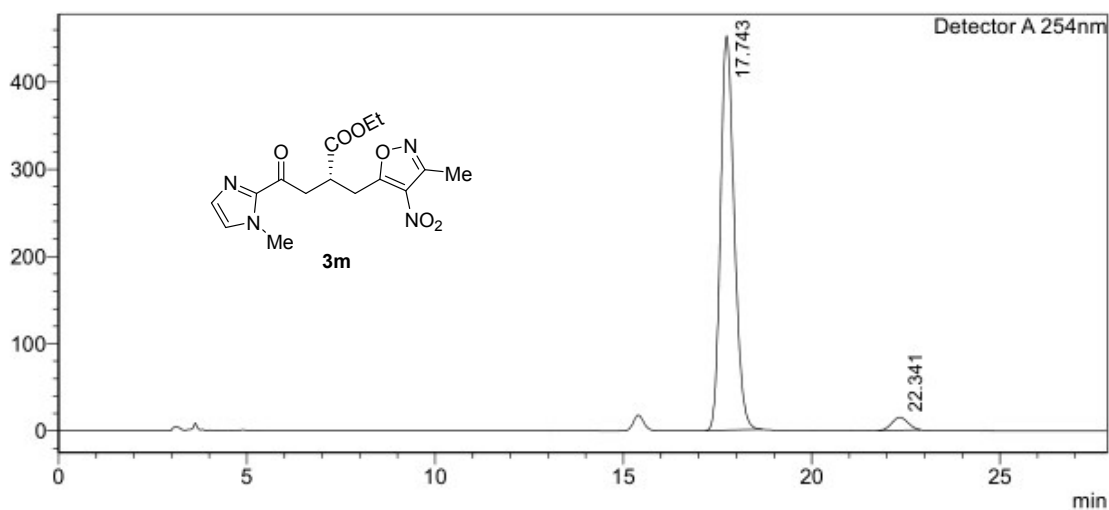
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	17.720	5138844	203634	50.007		M	
2	22.214	5137369	161020	49.993		M	
Total		10276212	364654				

## Chiral **3m**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

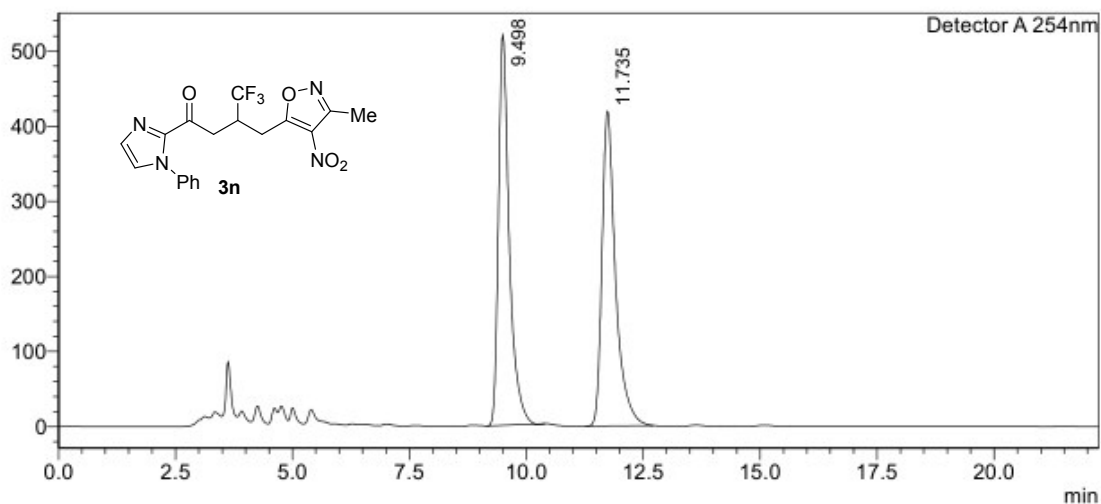
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	17.743	11458044	452001	96.304		M	
2	22.341	439773	14684	3.696		M	
Total		11897817	466685				

HPLC traces of racemic **3m** and chiral **3m**. Area integration = 93% ee.

## Racemic **3n**

### <Chromatogram>

mV



### <Peak Table>

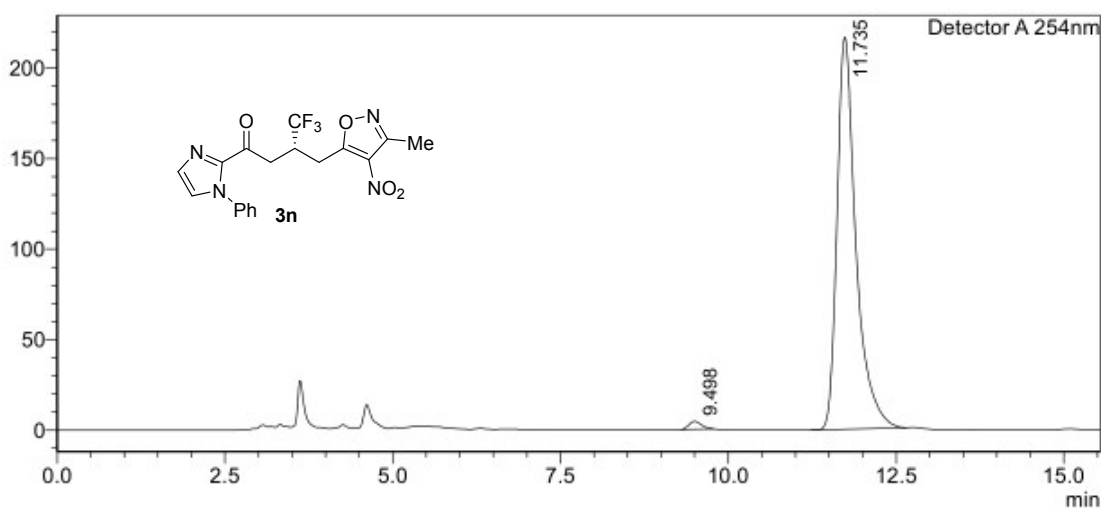
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.498	8356380	519836	49.780		M	
2	11.735	8430089	419047	50.220		M	
Total		16786469	938884				

## Chiral **3n**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

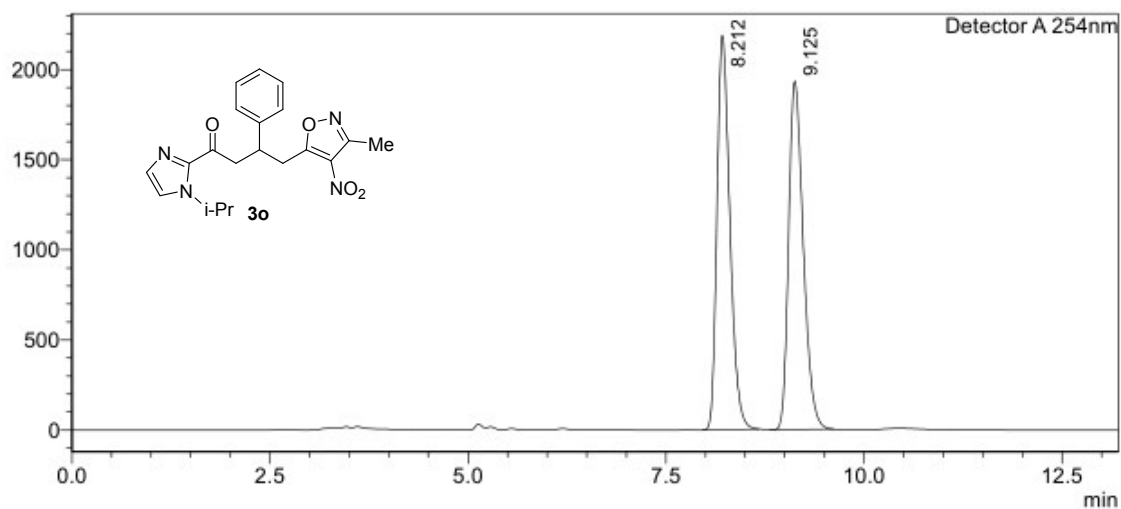
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.498	56546	4334	1.315		M	
2	11.735	4241944	216661	98.685		M	
Total		4298490	220995				

HPLC traces of racemic **3n** and chiral **3n**. Area integration = 97% ee.

## Racemic **3o**

### <Chromatogram>

mV



### <Peak Table>

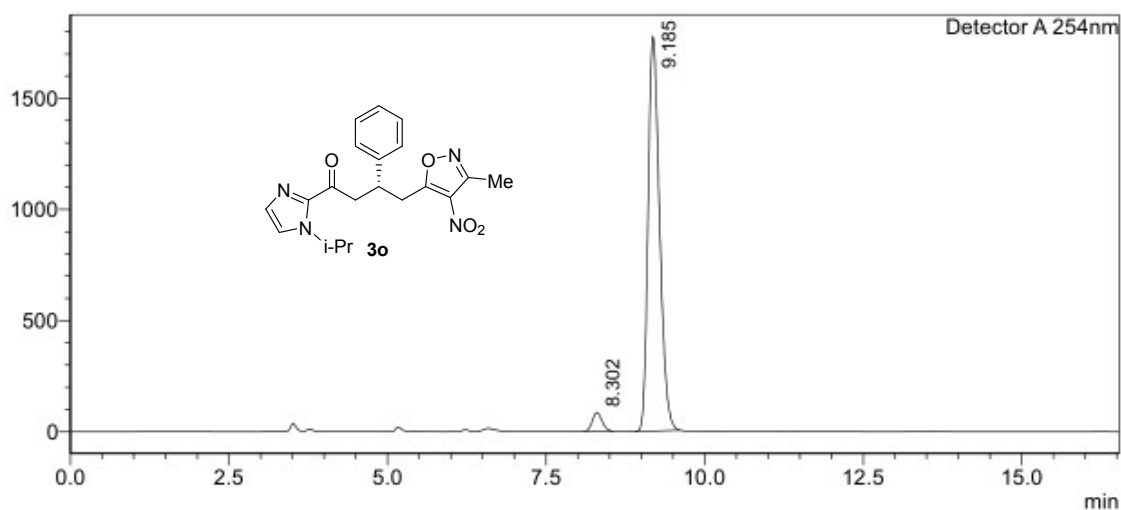
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.212	24757215	2186058	49.977		M	
2	9.125	24780101	1932033	50.023		M	
Total		49537316	4118091				

## Chiral **3o**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

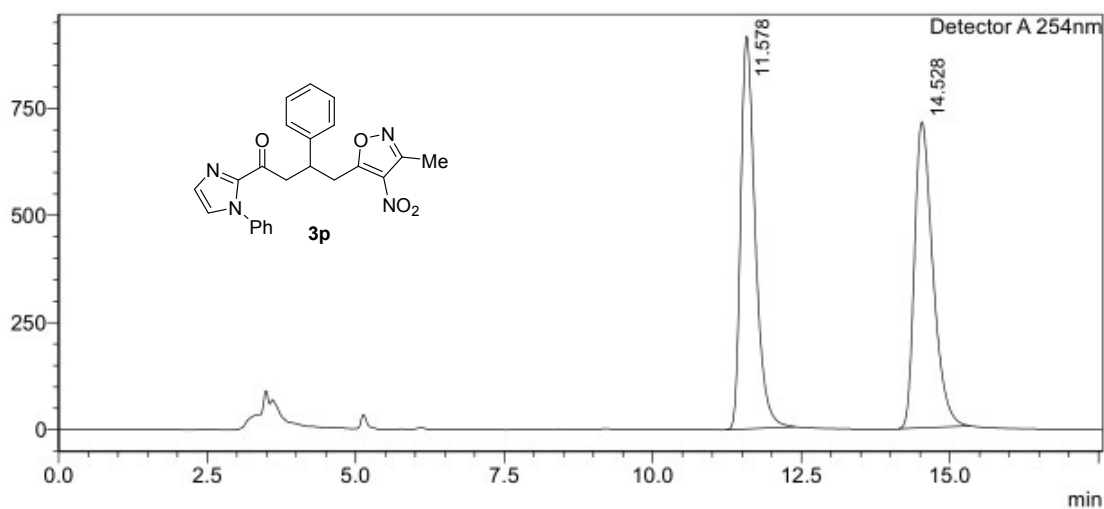
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.302	908655	84546	3.907		M	
2	9.185	22346885	1774020	96.093		M	
Total		23255540	1858566				

HPLC traces of racemic **3o** and chiral **3o**. Area integration = 92% ee.

## Racemic **3p**

### <Chromatogram>

mV



### <Peak Table>

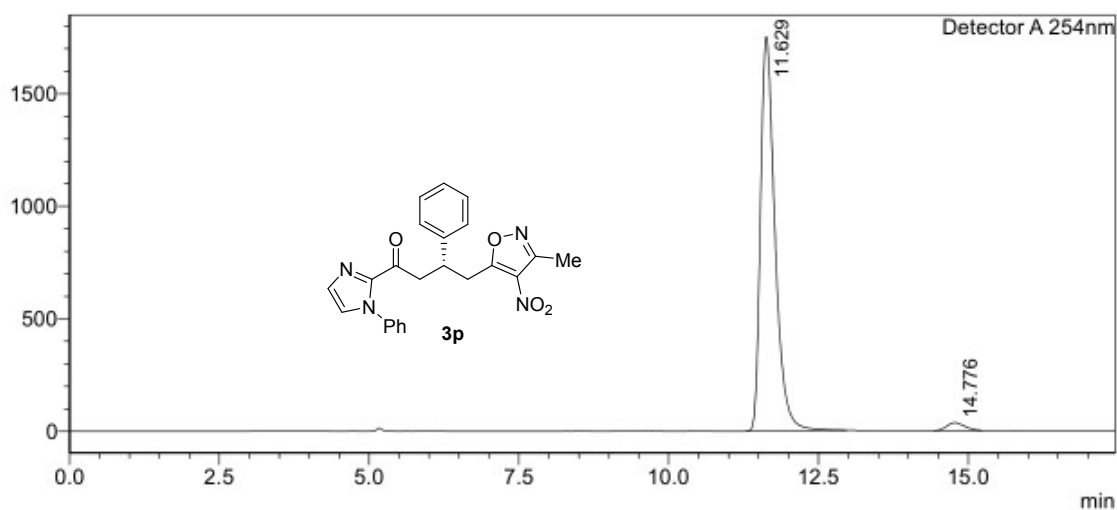
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	11.578	15534464	915869	50.150		M	
2	14.528	15441811	713729	49.850		M	
Total		30976275	1629598				

## Chiral **3p**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

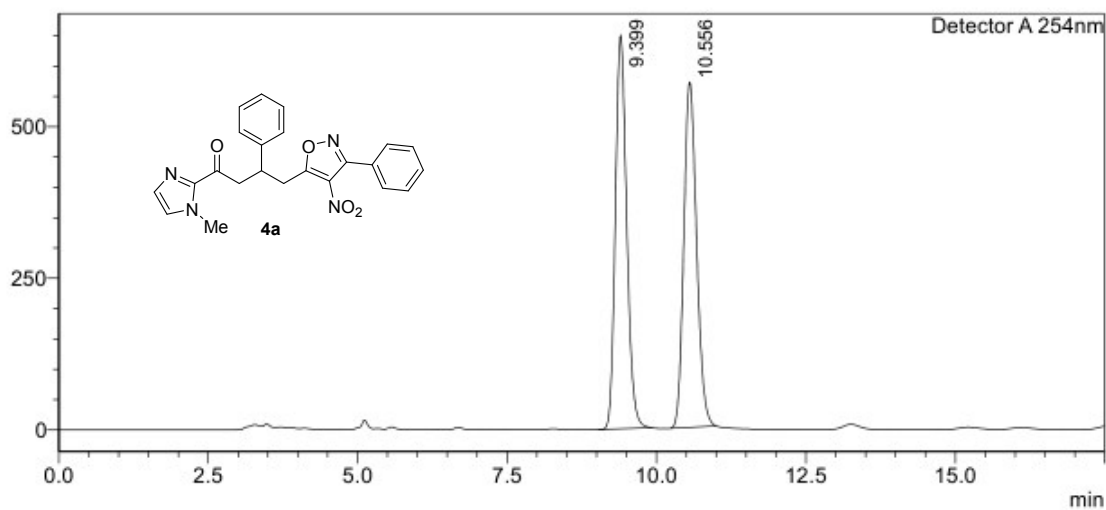
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	11.629	29400503	1751578	97.575		M	
2	14.776	730556	34795	2.425		M	
Total		30131059	1786373				

HPLC traces of racemic **3p** and chiral **3p**. Area integration = 95% ee.

## Racemic **4a**

### <Chromatogram>

mV



### <Peak Table>

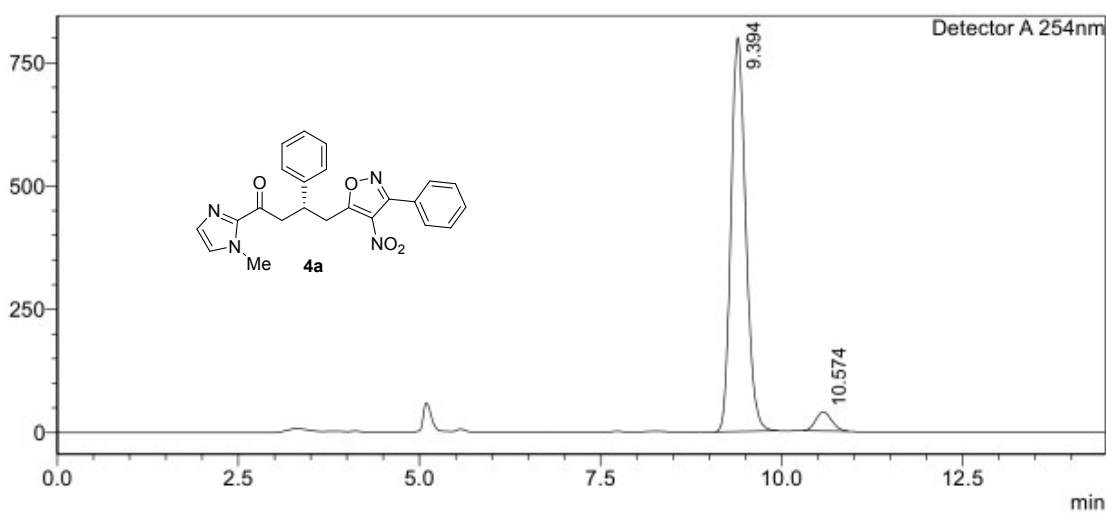
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.399	8544594	648449	50.291		M	
2	10.556	8445820	569130	49.709		M	
Total		16990414	1217579				

## Chiral **4a**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.394	11305455	798357	95.014		M	
2	10.574	593249	38521	4.986		M	
Total		11898704	836878				

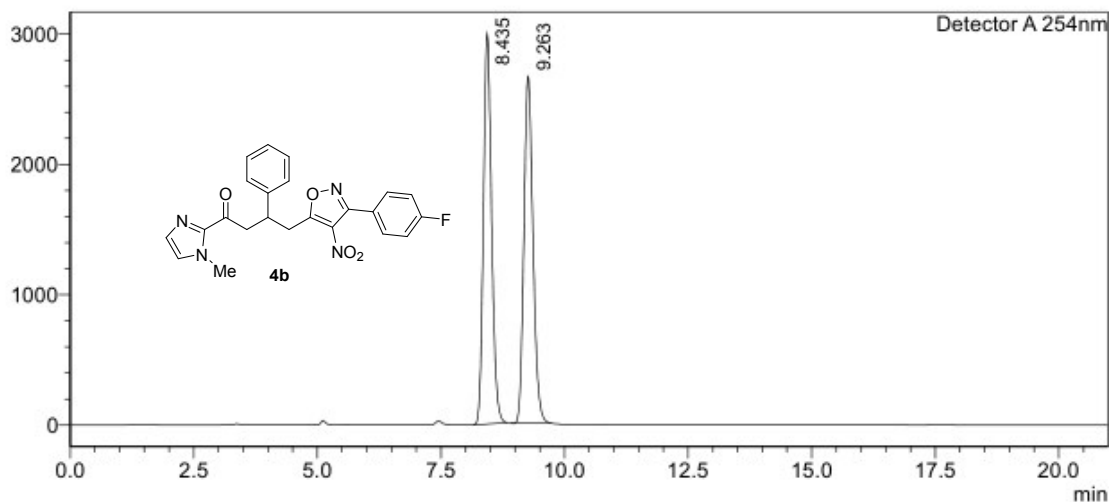
HPLC traces of racemic **4a** and chiral **4a**. Area integration = 90% ee.



## Racemic **4b**

### <Chromatogram>

mV



### <Peak Table>

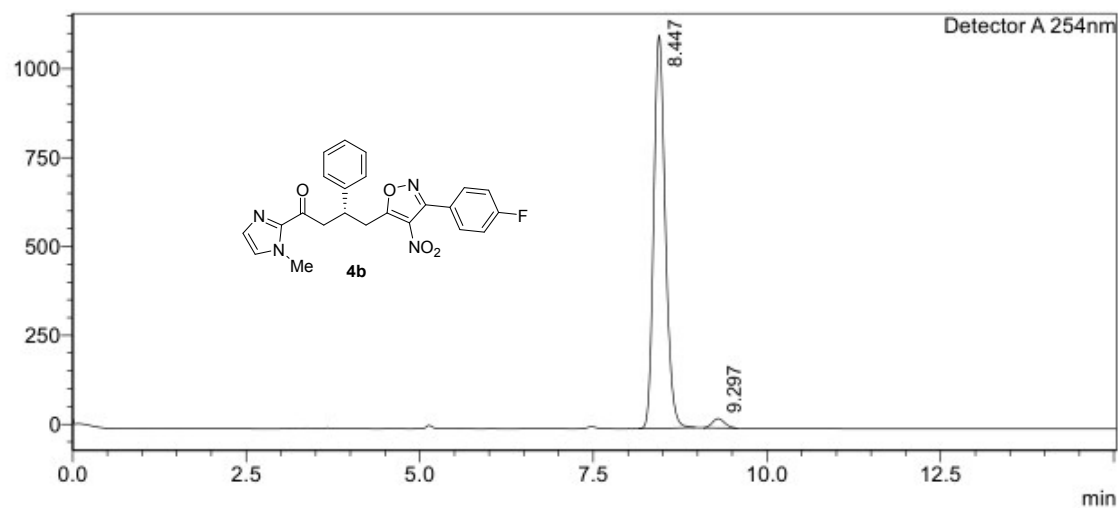
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.435	34328716	2994168	50.170		M	
2	9.263	34095892	2664272	49.830		M	
Total		68424607	5658440				

## Chiral **4b**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

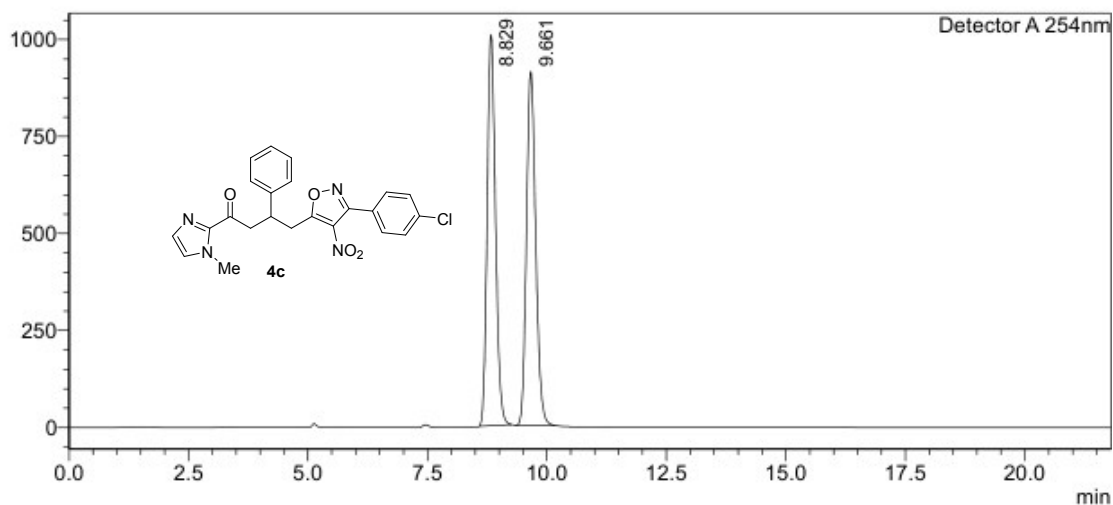
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.447	12822034	1105791	97.536		M	
2	9.297	323950	26181	2.464		M	
Total		13145984	1131973				

HPLC traces of racemic **4b** and chiral **4b**. Area integration = 95% ee.

## Racemic 4c

### <Chromatogram>

mV



### <Peak Table>

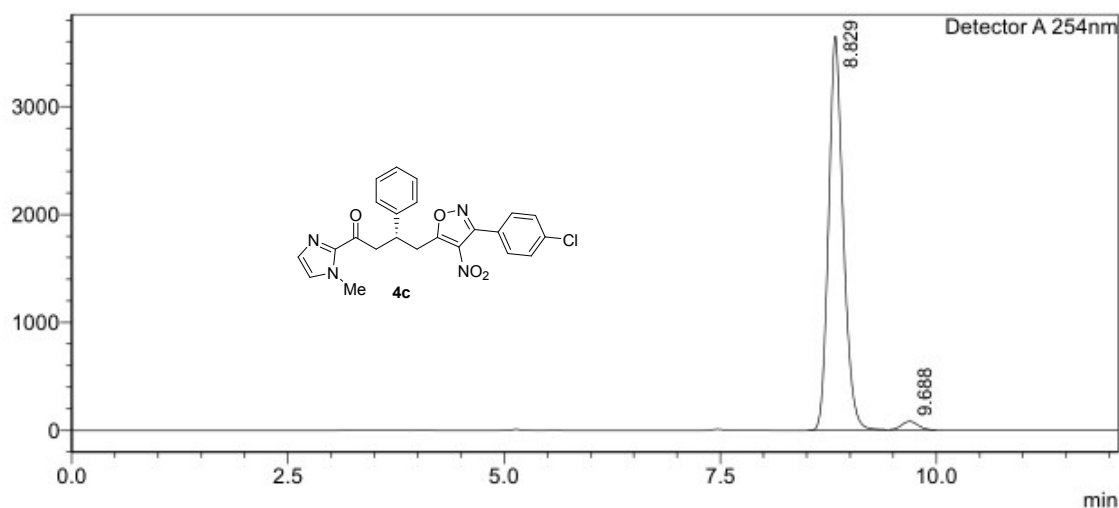
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.829	12209382	1006916	49.896		M	
2	9.661	12260227	911600	50.104		M	
Total		24469609	1918516				

## Chiral 4c:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

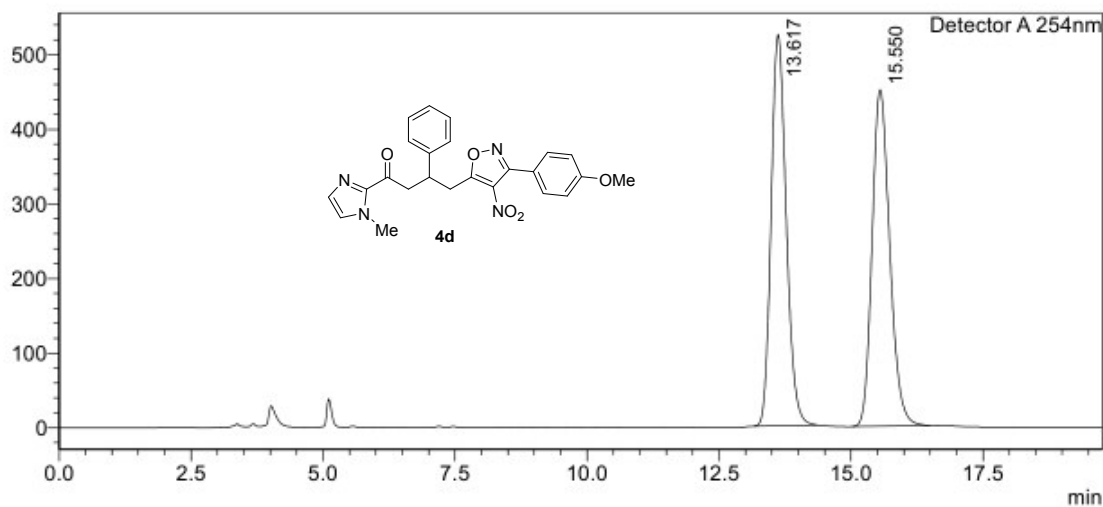
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.829	42962531	3644639	97.641			
2	9.688	1038158	78082	2.359		M	
Total		44000689	3722721				

HPLC traces of racemic 4c and chiral 4c. Area integration = 95% ee.

## Racemic **4d**

### <Chromatogram>

mV



### <Peak Table>

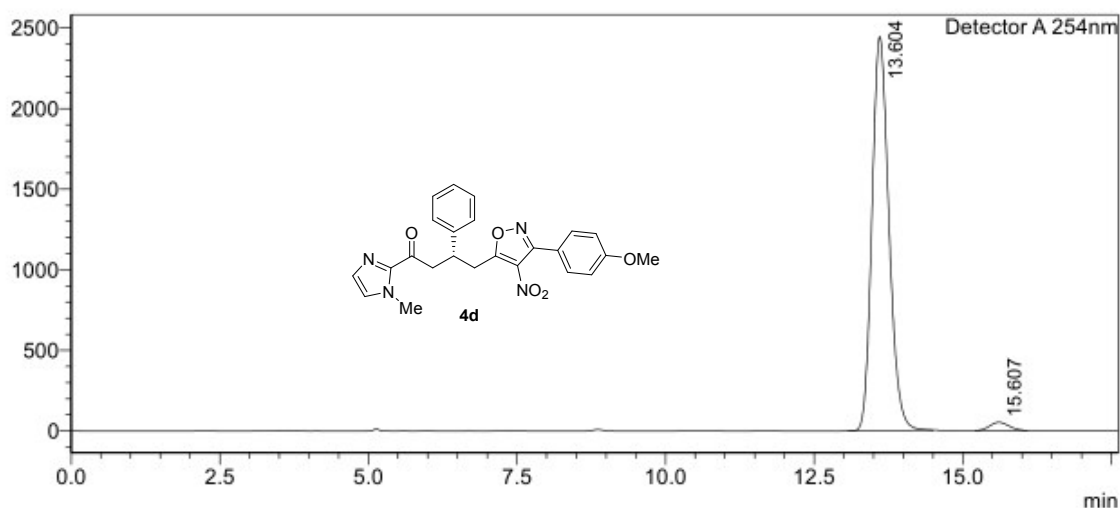
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	13.617	10410507	523983	49.814		M	
2	15.550	10488264	450753	50.186		M	
Total		20898772	974736				

## Chiral **4d**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

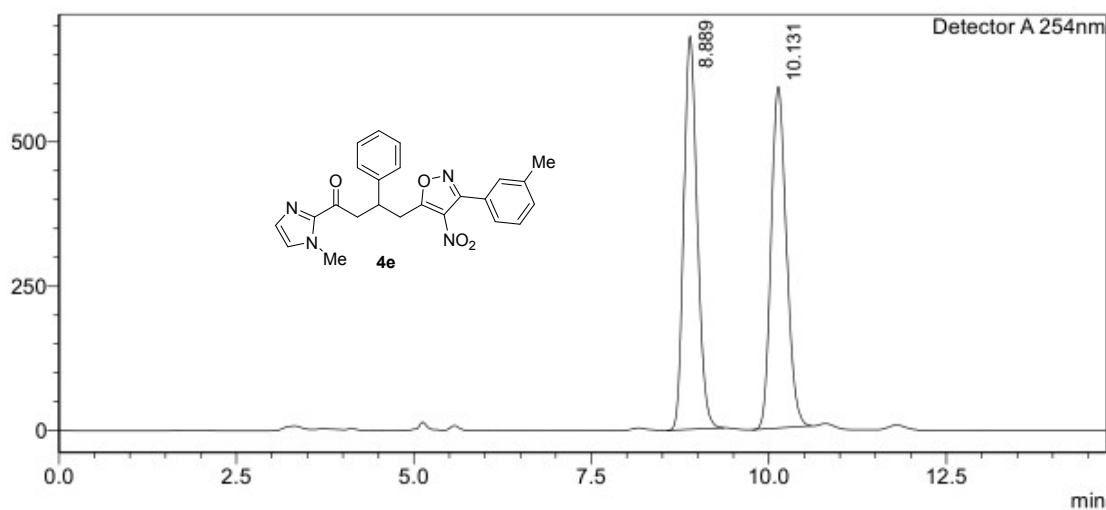
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	13.604	48249351	2444097	97.658		M	
2	15.607	1157216	51417	2.342		M	
Total		49406567	2495514				

HPLC traces of racemic **4d** and chiral **4d**. Area integration = 95% ee.

## Racemic 4e

### <Chromatogram>

mV



### <Peak Table>

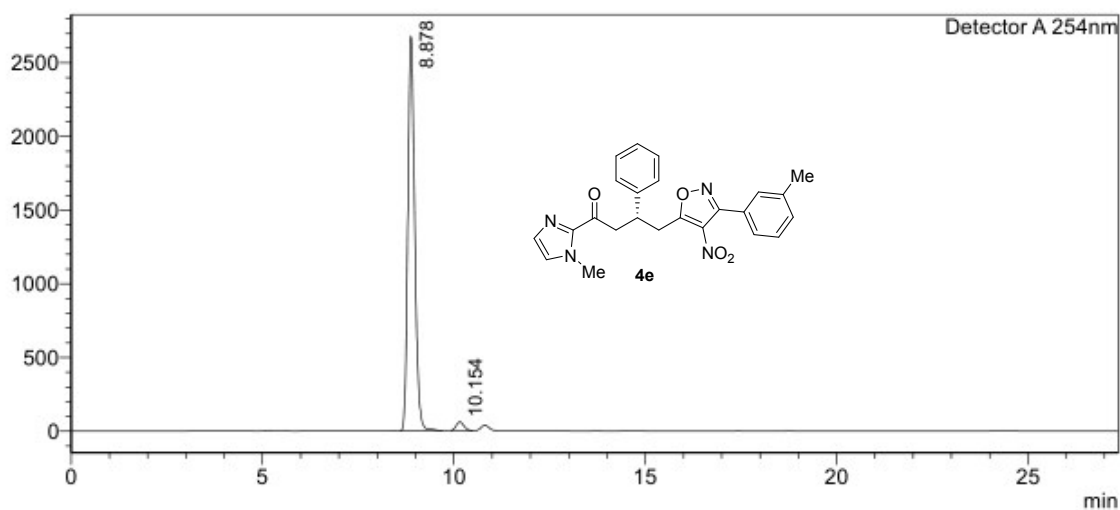
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.889	8982493	679657	50.273		M	
2	10.131	8884810	590707	49.727		M	
Total		17867303	1270364				

## Chiral 4e:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

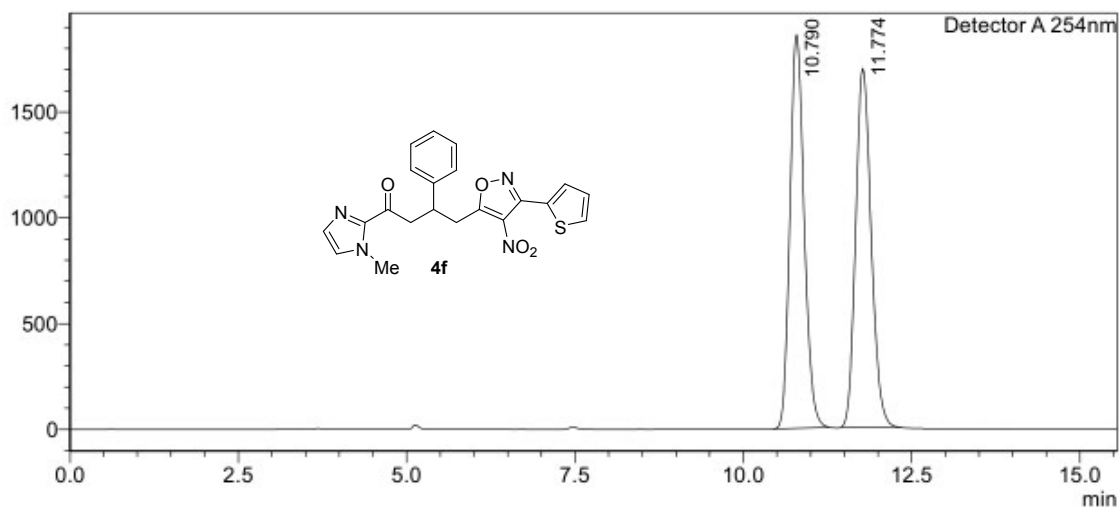
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	8.878	33472472	2676927	97.592		M	
2	10.154	825896	58752	2.408		M	
Total		34298368	2735678				

HPLC traces of racemic 4e and chiral 4e. Area integration = 95% ee.

## Racemic **4f**

### <Chromatogram>

mV



### <Peak Table>

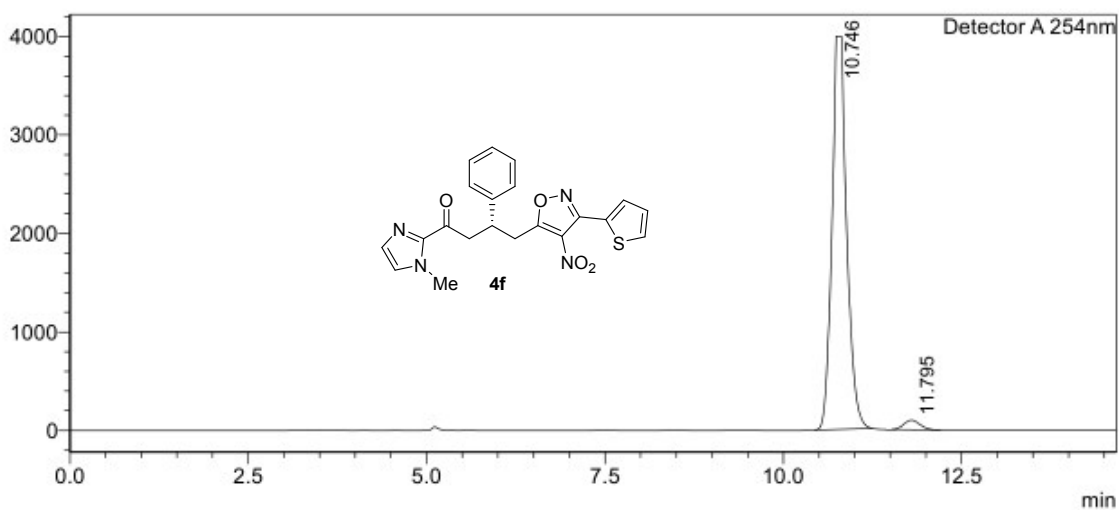
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	10.790	27332141	1858620	50.074		M	
2	11.774	27251763	1693892	49.926		M	
Total		54583904	3552513				

## Chiral **4f**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

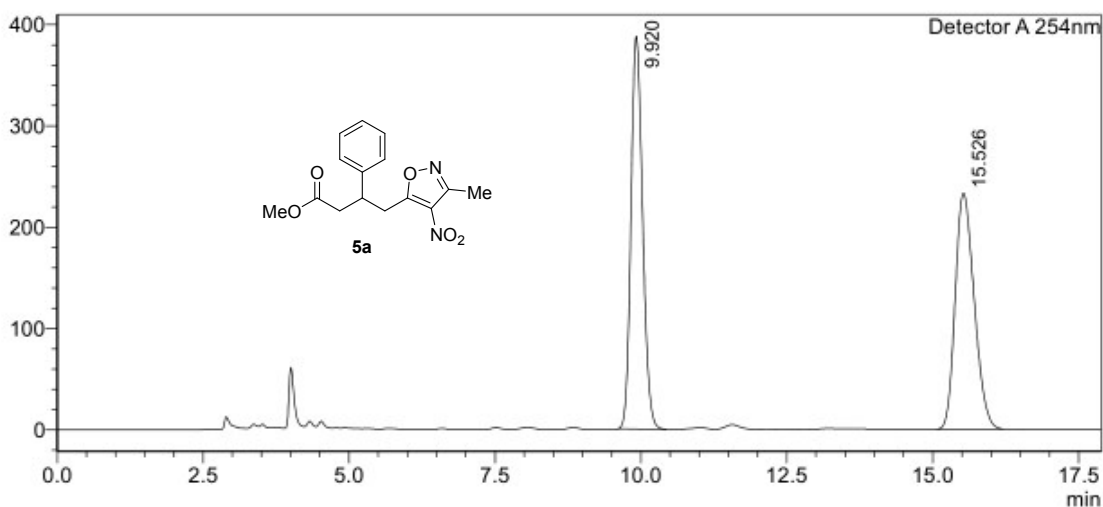
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	10.746	57174480	3988903	97.354		M	
2	11.795	1553883	97673	2.646		M	
Total		58728364	4086576				

HPLC traces of racemic **4f** and chiral **4f**. Area integration = 95% ee.

## Racemic **5a**

### <Chromatogram>

mV



### <Peak Table>

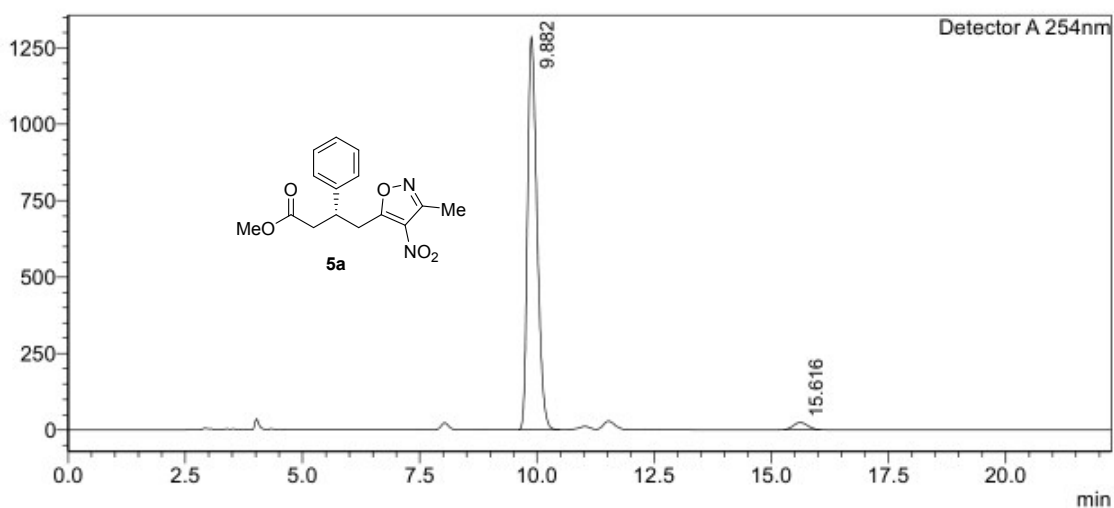
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.920	5369427	387537	50.483		M	
2	15.526	5266774	232719	49.517		M	
Total		10636202	620256				

## Chiral **5a**:

### <Chromatogram>

mV



### <Peak Table>

Detector A 254nm

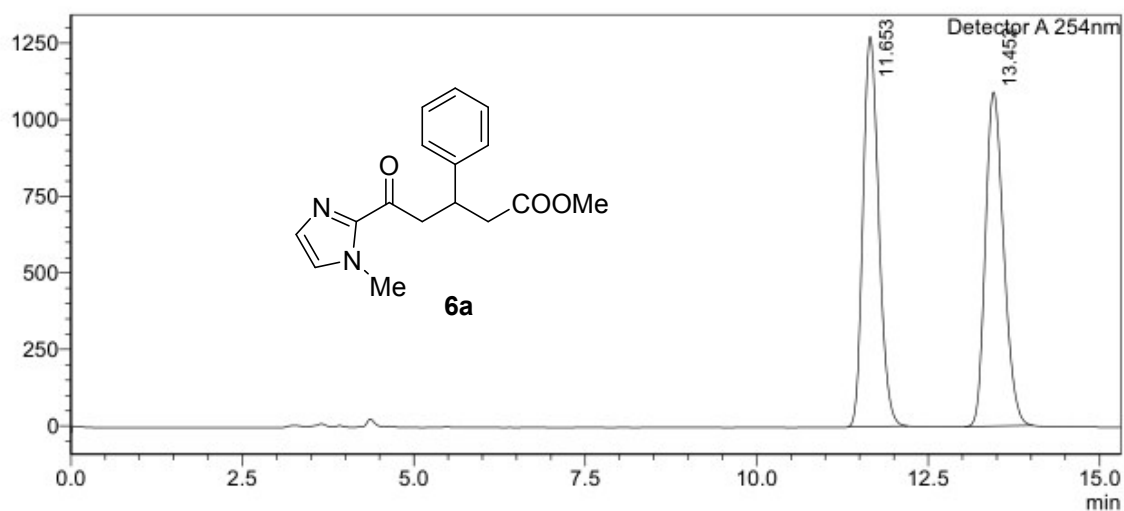
Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	9.882	18122350	1284357	97.397		M	
2	15.616	484276	23255	2.603		M	
Total		18606626	1307612				

HPLC traces of racemic **5a** and chiral **5a**. Area integration = 95% ee.

## Racemic **6a**

### <Chromatogram>

mV



### <Peak Table>

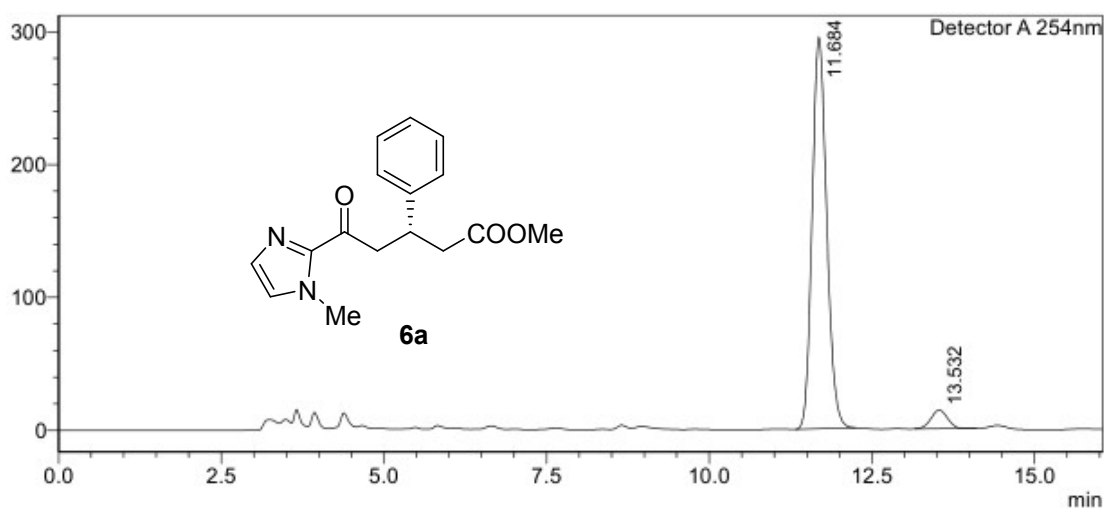
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	11.653	20262375	1272560	50.081		M	
2	13.452	20196793	1090887	49.919		M	
Total		40459169	2363447				

## Chiral **6a**:

### <Chromatogram>

mV



### <Peak Table>

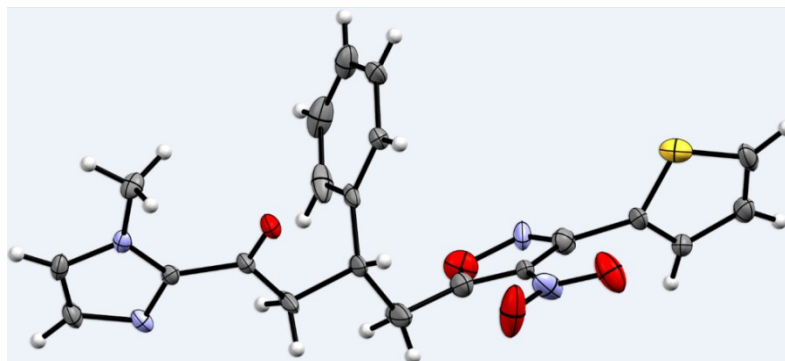
Detector A 254nm

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	11.684	4586840	294511	94.928		M	
2	13.532	245064	13960	5.072		M	
Total		4831904	308471				

HPLC traces of racemic **6a** and chiral **6a**. Area integration = 90% ee.

## VII. Single Crystal X-Ray Diffraction of 4f

**Table 1. Crystal data and structure refinement for 4f.**



CCDC 1903727.

**Table 1.** Crystal data and structure refinement for **4f**.

Identification code	compound- <b>4f</b>	
Empirical formula	C <sub>21</sub> H <sub>18</sub> N <sub>4</sub> O <sub>4</sub> S	
Formula weight	422.45	
Temperature (K)	127(2)	
Wavelength (Å)	1.54184	
Crystal system	orthorhombic	
Space group	<i>P</i> 2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>	
Unit cell dimensions (Å, °)	<i>a</i> = 6.0211(2)	$\alpha$ = 90
	<i>b</i> = 16.1241(5)	$\beta$ = 90
	<i>c</i> = 20.6340(5)	$\gamma$ = 90
Volume (Å <sup>3</sup> )	2003.25(10)	
<i>Z</i>	4	
Calculated density (g cm <sup>-3</sup> )	1.401	
Absorption coefficient (mm <sup>-1</sup> )	1.753	
<i>F</i> <sub>000</sub>	880	
Crystal size (mm <sup>3</sup> )	0.6 × 0.4 × 0.3	
$\theta$ range for data collection (°)	3.479 to 67.079	
Miller index ranges	-7 ≤ <i>h</i> ≤ 7, -16 ≤ <i>k</i> ≤ 19, -23 ≤ <i>l</i> ≤ 24	
Reflections collected	16910	
Independent reflections	3543 [ <i>R</i> <sub>int</sub> = 0.0657]	
Completeness to $\theta_{\max}$ (%)	0.987	
Max. and min. transmission	0.3203 and 0.7521	
Refinement method	Full-matrix least-squares on <i>F</i> <sup>2</sup>	
Data / restraints / parameters	3543 / 15 / 272	



Goodness-of-fit on $F^2$	1.064
Final $R$ indices [ $I > 2\sigma(I)$ ]	$R1 = 0.0916$ , $wR2 = 0.2275$
$R$ indices (all data)	$R1 = 0.1040$ , $wR2 = 0.2394$
Largest diff. peak and hole ( $e \text{ \AA}^{-3}$ )	1.320 and -0.460
Absolute structure parameter	.018(13)