Supporting Information

Construction of highly fluorescent N-O sevenmembered heterocycles via thermo-oxidation of oxazolidines

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1. General experimental information

Materials

Phenylhydrazine (98 %), 4-nitrophenylhydrazine (98 %), 4-methoxyphenylhydrazine hydrochloride (98 %), 4-dimethylaminobenzaldehyde (99 %) and diethyl malonate (99 %) were purchased from Sinopharm Chemical Reagent Co., Ltd. (Beijing, China). 3-Methyl-2-butanone (98 %), 2-bromoethanol (98 %), anisic aldehyde (99 %), 4- (Diethylamino)salicylaldehyde (98 %), Iodomethane (99.5 %), Iodine (98 %) and *N*-iodosuccinimide (98 %) were purchased from Energy Chemical (Shanghai, China). Sodium chloride (NaCl), sodium carbonate (Na₂CO₃) and anhydrous sodium sulfate (Na₂SO₄) were purchased from Beijing Chemical Factory (Beijing, China). Lyso Tracker Green and CellLightTM Mitochondria-GFP were obtained from Thermo Fisher (Eugene, OR). 4,6-diamidino-2-phenylindole (DAPI) were obtained from Tianjin Guangfu Fine Chemical Research Institute (Tianjin, China); dimethylformamide (DMF) from Beijing Chemical Factory (Beijing, China). Unless otherwise stated, all reagents and solvents were used without further purification.

Instruments

The UV-Vis absorption spectra were measured using a 0.1 cm quartz cuvette on a Shimadzu UV-2550 PC double-beam spectrophotometer. The fluorescent emission spectra were measured using a 0.1 cm quartz cuvette on a Shimadzu RF-5301 PC spectrofluorophotometer with a xenon lamp as a light source. The fluorescence quantum yields (Φ_f) and fluorescence lifetime (under the excitation at 400 nm) were measured on Edinburgh FLS 920 steady state spectrometer. The LC-HRMS (ESI) analysis was performed on an Agilent 1290-micro TOF-Q II mass spectrometer. ¹H NMR and ¹³C NMR spectra were recorded on a Bruker AVANCE500 (500M) and Wuhan Zhongke Niujin As 400 (400M) at room temperature and were reported in ppm and determined with tetramethylsilane (TMS) or respect to residual signals of the deuterated solvents as internal standards (TMS, 0.00; CDCl₃, 7.26; DMSO-*d*₆, 2.50 for ¹H NMR and CDCl₃, 77.0; DMSO-*d*₆, 39.5 for ¹³C NMR). Melting point was determined using a SGW X-4B microscopy melting point apparatus. Flow cytometry (cyto FLEX, Beckman COULTER). Confocal laser scanning microscope (Carl Zeiss Microscopy LLC, Jena, Germany).

Methods

Cytotoxicity assays. The viabilities of cells treated with **2e** were measured by a wellestablished 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide (MTT) assay. Generally, HeLa cells, A549 cells and SW480 cells were seeded in 96-well plates harboring 2 mL of 10% FBS-containing DMEM at a density of 8000 cells per well. The cells were incubated with drugs at concentrations from 0 to 100 μ g/mL, and cells without treatment were used as a control. After the incubation at 37 °C for 20 h, the standard MTT assay was used to determine the cell viability. Five repeats were conducted for each sample.

Co-localization experiments of 2e with commercial organelle-specific probes. HeLa cells were seeded into 6-well plates at a density of 2.5×10^5 cells/well and cultured with a sterilized coverslip for 12 h. Then the cells were treated with **2e** at the concentration of 1µg/mL for 4 h, respectively. Afterwards, the cells were stained with CellLightTM Mitochondria-GFP and Lyso Tracker Green DND-26 according to the manufacturer's protocol, respectively. Then the cells were washed with PBS for three times, followed by fixing with cold 75% ethanol at 4 °C for 20 min and then stained with DAPI solution. Finally, the cover slips were taken from the wells and observed with confocal laser scanning microscope (CLSM).

Visualizing extracellular fluctuations under oxidative stress and with drug treatment. Rosup, 5-fluorouracil (5-FU) and methotrexate (MTX) were used to construct oxidative and drug environments and then the endocytosis of 2e was detected under these conditions. HeLa cells were seed into 6-well plates at a density of 2.5×10^5 cells/well, and first, the cells were treated with Rosup, 5-FU and MTX, respectively. Then the medium was discarded after 1 hour treatment, and the cells were washed with phosphate buffer saline (PBS) three times. After that, 2e was incubated with cells at concentration of 1 µg/mL for 4 h. The endocytosis efficiency of 2e was determined by flow cytometry and CLSM.

2. Synthesis of 2a-2f, I and 3

2.1 Synthesis of thermo-oxidation products oxazolidines (2a-2f)



General synthesis method 1 of **2a-2f**. Starting materials **1a-OF-1f-OF** were prepared as reported.^{S1} **1a-OF-1f-OF** (100 mg) was stirred in dimethyl sulphoxide (2 mL) at 120 °C for 2 ~ 11 h (for **1f-OF**, 10 mg I₂ was added to prevent ring-closing of it). The reaction mixture was added with saturated solution of NaCl (20 mL). Dichloromethane (CH₂Cl₂) was added to extract product for 2 times and the CH₂Cl₂ layer was washed with saturated solution of NaCl for 1 time. Then the solvent was dried with anhydrous sodium sulfate and removed under reduced pressure. The crude mixture was precipitated as black solid. Purification of the crude mixture by chromatography (CH₂Cl₂ / MeOH = 50 / 1 to 20 / 1) afforded the product **2a-2f**. The products for characterizations below were obtained by method 1.



General synthesis method 2 of **2a-2f**. Starting materials **1a-1f** were prepared as reported.^{S1} A mixture of **1**s (0.37 mmol) and iodomethane (8 mmol) was stirred in dimethyl sulphoxide (2 mL) at 120 °C for 11 h. The reaction mixture was precipitated by addition of saturated solution of NaCl (20 mL) and filtered as brown powder. Purification of the crude reaction mixture by chromatography (CH₂Cl₂ / MeOH = 50 / 1 to 20 / 1) afforded the product **2a-2f**.



Yield: 31 %. m.p. 239.5 – 240.5 °C; ¹H NMR (500 MHz, DMSO- d_6): δ 8.03 (d, J = 9.2 Hz, 2H), 7.69 (d, J = 7.4 Hz, 1H), 7.58 (d, J = 7.4 Hz, 1H), 7.51 (t, J = 7.4 Hz, 1H), 7.41 (t, J = 7.4 Hz, 1H), 6.85 (d, J = 9.2 Hz, 2H), 6.59 (s, 1H), 5.02 (s, 2H), 4.65

(s, 2H), 3.12 (s, 6H), 1.62 (s, 6H); ¹³C NMR (126 MHz, DMSO-*d*₆): δ 176.9, 176.5, 154.1, 142.3, 139.9, 131.1, 128.5, 126.4, 123.1, 117.9, 112.2, 111.5, 85.8, 70.2, 51.2, 49.0, 24.7; LC-HRMS (ESI): m/z calculated for [*M* + H]⁺ 333.1961, found 333.1967.



Yield: 28 %. m.p. 246.9 – 247.9 °C; ¹H NMR (400 MHz, DMSO- d_6): δ 8.62 (s, 1H), 8.43 (d, J = 8.7 Hz, 1H), 8.08 (d, J = 8.7 Hz, 2H), 7.73 (d, J = 8.7 Hz, 1H), 6.89 (d, J = 8.7 Hz, 2H), 6.68 (s, 1H), 5.07 (s, 2H), 4.66 (s, 2H), 3.16 (s,

6H), 1.68 (s, 6H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 178.3, 178.0, 154.9, 147.4, 145.2, 140.9, 132.1, 125.3, 118.9, 117.4, 112.4, 111.8, 86.9, 70.6, 50.9, 49.3, 39.77, 24.8; LC-



HRMS (ESI): m/z calculated for $[M + H]^+$ 378.1812, found 378.1815.

2c Yield: 30 %. m.p. 229.8 – 230.7 °C; ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.15 (d, *J* = 8.9 Hz, 2H), 7.74 (d, *J* = 8.0 Hz, 1H), 7.69 (d, *J* = 8.0 Hz, 1H), 7.56 (t, *J* = 7.5 Hz, 1H), 7.48 (t, *J* = 7.5 Hz, 1H), 7.17 (d, *J* = 8.9 Hz, 2H), 6.78 (s, 1H), 5.10 (s, 2H), 4.77 (s, 2H), 3.90 (s, 3H), 1.65 (s, 6H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 178.9, 176.0, 164.0, 142.1, 140.4, 131.1, 128.6, 127.3, 124.7, 123.2, 114.6, 113.1, 88.0, 70.7, 55.9, 51.9, 49.5, 24.0. LC-HRMS (ESI): m/z calculated for [*M* + H]⁺ 320.1645, found 320.1644.



Yield: 30 %. m.p. 225.1 – 226.0 °C; ¹H NMR (400 MHz, DMSO- d_6): δ 7.99 (d, J = 9.1 Hz, 2H), 7.51 (d, J = 8.7 Hz, 1H), 7.35 (d, J = 2.3 Hz, 1H), 7.06 (dd, J = 8.7, 2.3 Hz, 1H), 6.84 (d, J = 9.1 Hz, 2H), 6.53 (s, 1H), 4.98 (s, 2H),

4.63 (s, 2H), 3.83 (s, 3H), 3.11 (s, 6H), 1.61 (s, 6H); ¹³C NMR (126 MHz, DMSO-*d*₆): δ 175.9, 175.4, 158.8, 153.9, 141.9, 135.8, 130.8, 118.1, 113.5, 113.3, 111.5, 109.5, 85.7, 70.0, 55.9, 51.3, 49.3, 24.6. LC-HRMS (ESI): m/z calculated for $[M + H]^+$ 363.2067, found 363.2073.



Yield: 29 %. m.p. 214.3 – 215.2 °C; ¹H NMR (400 MHz, DMSO- d_6): δ 8.70 (s, 1H), 7.75 (d, J = 7.2 Hz, 1H), 7.72 (d, J = 9.1 Hz, 1H), 7.66 (d, J = 8.0 Hz, 1H), 7.56 (t, J = 7.4 Hz, 1H), 7.47 (t, J = 7.4 Hz, 1H), 7.38 (s, 1H), 6.92 (d, J = 9.1

Hz, 1H), 6.69 (s, 1H), 5.03 (s, 2H), 4.73 (s, 2H), 3.56 (q, J = 7.0 Hz, 4H), 1.57 (s, 6H), 1.17 (t, J = 7.0 Hz, 6H); ¹³C NMR (100 MHz, DMSO- d_6): δ 177.9, 171.3, 158.7, 158.0, 154.1, 147.3, 142.4, 140.0, 132.8, 128.7, 127.2, 123.3, 113.0, 111.2, 108.7, 108.0, 96.0, 90.5, 70.1, 51.3, 49.7, 44.8, 24.7, 12.4. LC-HRMS (ESI): m/z calculated for $[M + H]^+$ 429.2173, found 429.2174.



Yield: 16 %. m.p. 174.5 – 175.2 °C; ¹H NMR (400 MHz, DMSO- d_6): δ 8.74 (s, 1H), 8.71 (d, J = 2.3 Hz, 1H), 8.45 (dd, J = 8.8, 2.3 Hz, 1H), 7.84 (d, J = 8.8 Hz, 1H), 7.75 (d, J = 9.5 Hz, 1H), 7.43 (s, 1H), 6.95 (dd, J = 9.1, 2.1 Hz,

1H), 6.71 (d, J = 2.1 Hz, 1H), 5.07 (s, 2H), 4.74 (s, 2H), 3.59 (q, J = 6.9 Hz, 4H), 1.64 (s, 6H), 1.18 (t, J = 6.9 Hz, 6H); ¹³C NMR (100 MHz, DMSO- d_6): δ 179.8, 173.8, 158.6, 158.3, 154.7, 148.0, 147.2, 145.8, 140.9, 133.3, 125.4, 119.2, 113.4, 111.6, 109.2, 107.5, 96.2, 91.1, 70.6, 51.2, 49.9, 44.9, 24.7, 12.5. LC-HRMS (ESI): m/z calculated for [M + H]⁺ 474.2023, found 474.2015.

2.2 Synthesis of I



A mixture of **1b** (380 mg, 1 mmol) and iodomethane (1.2 mL, 19.2 mmol) was stirred in acetone (6 mL) at 56 °C for 11 h. Light yellow solid was produced and precipitated as reaction proceeded. After removal of the solvent, the crude product was washed with acetone for 5 times to afford the product as a light yellow solid **I** (470 mg, Yield: 90.2 %). m.p. 138.6 – 139.5 °C; ¹H NMR (500 MHz, DMSO- d_6): δ 8.13 (dd, J = 8.7, 2.1 Hz, 1H), 8.06 (d, J = 2.1 Hz, 2H), 7.93 (d, J = 8.9 Hz, 2H), 7.84 (d, J = 8.9 Hz, 2H), 7.12 (d, J = 8.7 Hz, 1H), 6.90 (d, J = 16.1 Hz, 1H), 6.56 (d, J = 16.1 Hz, 1H), 3.45 – 3.89 (m, 4H), 3.60 (s, 9H), 1.46 (s, 3H), 1.15 (s, 3H); ¹³C NMR (126 MHz, DMSO- d_6): δ 157.1, 146.5, 141.9, 140.4, 137.5, 130.1, 128.2, 127.6, 125.1, 120.7, 118.5, 111.9, 109.2, 63.3, 56.4, 48.9, 46.9, 30.7, 27.5, 20.1; LC-HRMS (ESI): m/z calculated for $[M + H]^+$ 394.2125, found 394.2125.

2.3 Synthesis of 3



A mixture of **1b** (400 mg, 1.05 mmol) and iodomethane (0.62 mL, 10 mmol) was stirred in dimethylformamide (2 mL) at 110 °C for 36 h. Then the solvent was removed under reduced pressure and the crude mixture was precipitated as black solid. Purification of the crude mixture by chromatography (Petroleum ether / EtOAc = 6 / 1) afforded the product as a yellow solid **3** (20 mg, Yield: 5.4 %). m.p. 221.9 – 222.8 °C; ¹H NMR (500 MHz, DMSO-*d*₆): δ 11.87 (s, 1H), 8.18 (dd, *J* = 8.6, 2.1 Hz, 1H), 8.10 (d, *J* = 2.1 Hz, 1H), 7.92 (d, *J* = 8.9 Hz, 2H), 6.93 (d, *J* = 8.6 Hz, 1H), 6.73 (d, *J* = 8.9 Hz, 2H), 6.18 (s, 1H), 3.07 (s, 6H), 1.51 (s, 6H); ¹³C NMR (126 MHz, DMSO-*d*₆): δ 189.2, 170.6, 148.7, 142.7, 138.7, 129.7, 126.0, 118.6, 111.3, 109.0, 90.2, 46.9, 40.4, 28.0; LC-HRMS (ESI): m/z calculated for [*M* + H]⁺ 352.1656, found 352.1659.

3. ¹H NMR spectra monitoring heating 1a-1f in DMSO- d_6



Figure S1. (a) Reaction of **1a-1f** in DMSO- d_6 . (b) ¹H NMR spectra of **1a-1f** in DMSO- d_6 (5mg / 0.5 mL) before and after heating for 3 h at 120 °C.

4. Structural characterizations of intermediate product I of 1b reacting with CH₃I



Figure S2. ¹H NMR spectra comparison between **I** (blue line) and intermediate products of **1b** and CH₃I reacting in DMF at 10 minutes (red line).

a). 2b b) 0h I b 1b ↑↓ 1h 2h £Î. 3h ↑↓II 2b M. 4h Hels A 6h M 8h 10h 15h 20h 32h 1b 11 П 2b CH₃I 9.0 8.5 8.0 7.5 7.0 6.5 f1 (ppm) 4.5 6.0 5.5 5.0 C) Qh I 1þ 1h

5. ¹H NMR spectra monitoring heating I in DMSO- d_6



Figure S3. (a) Probable reaction process of **I** in DMSO- d_6 at 120 °C. (b) ¹H NMR spectra variation of **I** in DMSO- d_6 (5mg / 0.5 mL) in (b) low and (c) high magnetic fields at 120 °C with reaction time.

6. Photographs of heating CH₃I in DMSO



Figure S4. Photographs of CH₃I heated in DMSO over time in the dark at 120 °C.

The DMSO solution of CH_3I gradually changed from colorless to brown with heating time, which indicated the generation of I_2 from thermolysis of CH_3I .

7. HRMS and ¹H NMR spectra monitoring reaction of 1g and CH₃I in DMF at 120 °C



Figure S5. (a) Probable reaction process of 1g and CH_3I in DMF at 120 ^oC. (b) HRMS spectra of 1g and CH_3I in DMF at 120 ^oC for 40 min (above) and HRMS spectra of 1g-Me (below).



Figure S6. (a) Probable reaction process of **1g** and CH_3I in DMSO- d_6 at 120 °C. (b) ¹H NMR spectra comparison of **1g** (purple), **1g** after heating 3 h (blue), **1g** and CH_3I after heating 3 h (green), **1g-OF** (yellow) and **1g-OF** after heating 3 h (dark red) (5 mg / 0.5 mL in DMSO- d_6).

8.

¹H NMR spectra monitoring heating 1b-OF in DMSO-*d*₆



Figure S7. (a) Probable reaction process of **1b-OF** in DMSO- d_6 at 120 °C. (b) ¹H NMR spectra variation of **1b-OF** in DMSO- d_6 (5 mg / 0.5 mL) in (b) low and (c) high magnetic fields at 120 °C with reaction time.



Figure S8. (a) HRMS spectrum of 1b-Br and (b) ¹H NMR spectrum of 1b-OF in DMSO- d_6 (5 mg / 0.5 mL) after reacting 3h at 120 °C and the speculated structure of 1b-Br.

9. Comparison of 1b-OF reaction in DMSO under aerobic and anaerobic conditions



Figure S9. The UV-Vis spectra of the diluted reaction solution of **1b-OF** after heating for 4 hours in DMSO under aerobic (blcak line) and anaerobic (red line) conditions.

1b-OF could be oxidized to **2b** both under aerobic and anaerobic condition, but it exhibits a slower reaction rate under anaerobic condition within the same reaction time of 4 h. It proved that except O_2 , DMSO also works as oxidant in this reaction.

10. Reaction of 1a-OF with H_2O_2 in DMF



Figure S10. (a) Reaction process of 1a-OF and H_2O_2 in DMF solution at 100 ^oC. (b) HRMS spectra for reaction solutions of 1a-OF (1.0 eq) and H_2O_2 (2.4 eq) in DMF solution at 100 ^oC for 5 h.

11. Reaction of 1b-OF in DMSO/DMF solutions with addition of radical inhibitor BHT



Figure S11. UV-Vis spectra for the diluted reaction solutions of **1b-OF** with (blue line) and without (black line) addition of butylated hydroxytoluene (BHT) in (a) DMSO and (b) DMF solutions, respectively.

12. HRMS spectra for reaction solutions of 1b with NIS, NBS and NCS



Figure S12. (a) Reaction process of 1b (1.0 eq) with N-iodosuccinimide (NIS), N-bromosuccinimide (NBS), and N-chlorosuccinimide (NCS) (1.2 eq) in DMF solution. (b) - (d) HRMS spectra for reaction solutions of 1b with NBS at 100 °C for 6 h, NCS at 100 °C for 6 h and NIS at 80 °C for 80 min, respectively.

13. Crystal data and structure refinement for 2a, 2b, 2c and 3

Single crystals of **2a**, **2b**, **2c** and **3** were all obtained by vapor diffusion of n-hexane into their acetonitrile solutions of **2a**, **2b** and **2c**. Single crystal of **3** was obtained by slow evaporation in mixed solution of n-hexane and EtOAc.



Figure S13. Single-crystal X-ray structures of 2a, 2b, 2c and 3 (50 % probability ellipsoids). The anion in 2a, 2b and 2c is I⁻.

Compound	2b	2a	2c	3
Formula	C ₂₂ H ₂₄ IN ₃ O ₃	C ₂₂ H ₂₅ IN ₂ O	$C_{21}H_{22}INO_2$	$C_{20}H_{21}N_3O_3$
Formula mass	505.34	460.34	447.29	351.40
Space group	monoclinic	monoclinic	triclinic	monoclinic
	P 21/n	P 21/c	P-1	P 21/c
<i>a</i> / Å	6.4834 (13)	20.1317(10)	9.4276(5)	10.118(2)
b/ Å	23.560 (5)	7.4780(4)	10.1879(5)	9.1993(18)
c/ Å	14.458(3)	13.4306(6)	11.1539(6)	20.098(4)
lpha/ °	90	90	111.6410(10)	90
eta/ °	100.21(3)	94.394(2)	97.790(2)	100.49(3)
γ/ °	90	90	99.874(2)	90
V / Å3	2173.4(8)	2015.96(17)	957.65(9)	1839.4(7)
Ζ	4	4	2	4
$\rho/\text{ g.cm}^{-3}$	1.544	1.517	1.551	1.269
μ / mm ⁻¹	1.501	1.601	1.685	0.087
F000	1016	928	448	744.0
Temp, (K)	293(2) K	273(2) K	273(2) K	293 K
No. of reflns. collected	19917	12417	6501	17394
No. of unique reflns.	. 4975	3542	3553	4194
R _{int}	0.0276	0.0437	0.0215	0.0325
Final <i>R1</i> values ($I >$	2 <i>σ</i> (<i>I</i>)) 0.0320	0.0378	0.0318	0.0466
Final $wR(F^2)$ values (I	$> 2\sigma(I))0.0724$	0.0777	0.0732	0.1199
Final R1 values (all	data) 0.0405	0.0576	0.0398	0.0690
Final $wR(F^2)$ values (al	ll data) 0.0774	0.0869	0.0789	0.1289
Goodness of fit on F	1.124	1.023	1.015	1.057
CCDC numbers	1562292	1562290	1562291	1571735

Table S1. Summary of crystal data and intensity collection parameters for 2a, 2b, 2c and3.

14. Spectral data of 1a-OF-1f-OF and 2a-2f in DCM solutions and PMMA films

14.1 Maximum absorption wavelength, maximum emission wavelength and molar absorption coefficient of 1a-OF-1f-OF and 2a-2f.

Table S2. Spectral data including maximum absorption wavelength, maximum emission wavelength and molar absorption coefficient of **1a-OF** to **1f-OF** and **2a** to **2f** (a) in DCM solutions $(3.0 \times 10^{-5} \text{ M})$ and (b) in PMMA films (**2e** is 0.6% and others are 1.0% weight percent).

a)	Solution (in DCM)	λ _{1,abs} (nm)	λ _{1,em} (nm)	ε1 (L/mol·cm)	Solution (in DCM)	λ _{2,abs} (nm)	λ _{2,em} (nm)	ε2 (L/mol	·cm)	$\Delta \lambda_{abs} = \\ \lambda_{2,abs} - \lambda_{1,abs}$	$\Delta \lambda_{em} = \lambda_{2,em} - \lambda_{1,em}$	$\Delta \varepsilon = \varepsilon 2 - \varepsilon 1$ (L/mol·cm)
	1a-OF	559	592	86530	2a	517	557	8419	90	-42	-35	-2340
	1b-OF	593	624	137380	2b	535	578	904	50	-58	-47	-46930
	1c-OF	445	482	36760	2c	427	480	4242	20	-18	-2	5660
	1d-OF	561	603	73550	2d	522	565	6613	30	-39	-38	-7420
	1e-OF	622	666	87100	2e	572	609	1057	10	-50	-57	18610
	1f-OF	666	701	104680	2f	598	642	826	70	-68	-59	-22010
b)	Film (in PMI	MA)	λ _{1,abs} (nm)	λ _{1,em} (nm)	(in	Film PMMA)	λ (2,abs nm)	λ _{2,er} (nm	n Δλ _{at} 1) λ _{2,abs} –	_{os} = ·λ _{1,abs}	$\Delta \lambda_{em} = \lambda_{2,em} - \lambda_{1,em}$
	1a-C	F	547	601		2a	4	95	563	3 -5	2	-38
	1b-C	F	586	631		2b	5	17	589	9 -6	9	-42
	1c-0	F	433	-		2c	4	15	503	3 -1	8	-
	1d-C	F	539	609		2d	4	93	573	3 -4	6	-36
	1e-C	F	603	677		2e	5	649	614	4 -5	4	-63
	1f-0	F	655	700		2f	1.5	68	640	8	7	-60



14.2 Measurement of molar absorption coefficient of 1a-OF-1f-OF and 2a-2f.

Figure S14. Plots of molar absorption coefficient for (a) 1a-OF-1f-OF and (b) 2a-2f in DCM.

14.3 Fluorescent lifetime measurements of 1a-OF-1f-OF and 2a-2f.

		$\tau_1(ns)$	$\tau_2(ns)$	$\tau_{avg}(ns)$	χ^2
1a-OF	solution ^{a)}	0.34	2.54	0.58	1.363
	film ^{a)}	0.94	2.17	1.38	1.119
1b-OF	solution ^{a)}	0.61	3.25	1.36	1.653
	film ^{a)}	0.72	2.59	1.03	1.402
1c-OF	solution ^{a)}	0.26	2.59	0.49	1.309
	film ^{a)}	0.71	2.25	1.60	1.253
1d-OF	solution ^{a)}	0.30	2.73	0.56	1.381
	film ^{a)}	0.70	1.84	1.10	1.253
1e-OF	solution ^{b)}	1.84	-	1.84	1.643
	film ^{a)}	2.08	14.1	2.27	1.395
1f-OF	solution ^{b)}	3.67	-	3.67	1.419
	film ^{a)}	2.29	15.73	2.56	1.182

Table S3. Fluorescence lifetime of **1a-OF** to **1f-OF** in DCM solutions (5.0 X 10⁻⁵ M) and PMMA films (1.0 % weight percent). ^{a)} $\lambda_{ex} = 400$ nm. ^{b)} $\lambda_{ex} = 375$ nm.

		$\tau_1(ns)$	$\tau_2(ns)$	$\tau_{avg}(ns)$	χ^2
2a	solution ^{b)}	1.70	-	1.70	1.991
	film ^{a)}	1.99	4.23	2.71	1.378
2b	solution ^{b)}	1.53	-	1.53	1.577
	film ^{a)}	1.14	2.53	1.74	1.360
2c	solution ^{b)}	1.53	-	1.53	1.790
	film ^{a)}	0.97	2.22	1.62	1.147
2d	solution ^{b)}	1.38	-	1.38	2.381
	film ^{a)}	1.82	3.59	2.31	1.108
2e	solution ^{b)}	3.27	-	3.27	1.722
	film ^{a)}	1.93	4.06	2.29	1.234
2f	solution ^{b)}	2.56	-	2.56	1.445
	film ^{a)}	1.08	2.22	1.81	1.704

Table S4. Fluorescence lifetime of **2a** to **2f** in DCM solutions (5.0 X 10⁻⁵ M) and PMMA films (1.0 % weight percent). ^{a)} $\lambda_{ex} = 400$ nm. ^{b)} $\lambda_{ex} = 375$ nm.

Fluorescent photographs of solid powders of 1a-OF-1f-OF and 2a 2f



Figure S15. Fluorescent photographs of solid powder of 1a-OF-1f-OF (above) and 2a-2f (below) under 365 nm handhold UV lamp, and the absolute fluorescence quantum yields of 2a, 2c, 2d and 2e.



16. Solvent polarity effects on optical properties of 2a-2f

Figure S16. Structures and photographs of **2a-2f** in different solvents $(1.0 \times 10^{-5} \text{ M})$ under visible and UV light by 365 nm handhold UV lamp.



Figure S17. UV-Vis spectra of 2a-2f in different solvents (1.0×10^{-5} M).



Figure S18. Fluorescence emission spectra of 2a-2f in different solvents (1.0×10^{-5} M).

17. Photographs, UV-Vis spectra and fluorescent spectra of 2a-2f in PMMA with different weight percent



Figure S19. Photographs of spin-coated **2a-2f** PMMA films with weight percent of 0.2%, 0.6%, 1%, 1.5%, 5% and 10% respectively (a) under visible light and (b) 365 nm handhold UV lamp.



Figure S20. (a) - (f) UV-Vis spectra of spin-coated 2a-2f PMMA films with weight percent of 0.2%, 0.6%, 1%, 1.5%, 5% and 10% respectively.

2b is not fully soluble when the weight percent reaches 10%, so the baseline increased.



Figure S21. (a) - (f) Fluorescence spectra of **2a-2f** in PMMA films by spinning coating of different weight percent (0.2%, 0.6%, 1%, 1.5%, 5% and 10%).

18. Comparison of fluorescence quantum efficiency of 2a and 2c with I⁻, Br⁻ and PF₆⁻

Synthesis of 2a and 2c with different anions (I⁻ or PF₆-): 2a (I⁻)/2c (I⁻) were obtained by the reaction of 1a/1c with CH₃I in DMSO at 120 °C. 2a (PF₆⁻)/2c (PF₆⁻) were obtained by counterion exchange with the bromide anions. The synthetic method is that we synthesized the crude product 2a (Br⁻) and 2c (Br⁻) in DMSO, respectively and added 15 equivalents of ammonium hexafluorophosphate into the DMSO solution. After stirring 20 minutes and adding saturated saline, the crude product 2a and 2c with hexafluorophosphate anion precipitated out. The product were purified by chromatography (dichloromethane / methanol = 60 / 1).

Structures confirmation of 2a (PF₆⁻) and 2c (PF₆⁻): The ¹H NMR spectra in Fig. S19 indicated that 2a (PF₆⁻) and 2c (PF₆⁻) may be obtained, but the anions can't be determined. Moreover, compared to 2a (Br⁻) and 2c (Br⁻), the significantly reduced mobile phase polarity when doing chromatography, as well as 1~4 nm red shift of absorption spectra confirmed that we indeed obtained product 2a and 2c with PF₆⁻ and I⁻ (Fig. S20).



Figure S22. ¹H NMR spectra of 2a and 2c with hexafluorophosphate anion.



Figure S23. Normalized UV-Vis spectra of 2a and 2c with I⁻, Br⁻ and PF₆⁻ in dichloromethane, respectively.



Figure S24. Comparison of photos and absolute fluorescence quantum yields of 2a and 2c with I⁻, Br⁻ and PF₆⁻ in dichloromethane and in solid states, respectively.

19. Theoretical calculations of energy levels of frontier molecular orbitals for ground states of 2a-2f

All the Density functional theory (DFT) calculations were carried out using the GAUSSIAN 09 series of programs.^{S2} DFT and B3LYP with a standard 6-31g (d) basis set were used for geometry optimizations of ground states of **2a-2f**. And the vibrational spectrum of each molecule was calculated at the same level of theory to ensure that all of the structures correspond to the true minima of the potential energy surface.

	2a	2b	2c	2d	2e	2f
LUMO (eV)	-5.30	-5.68	-5.62	-5.16	-5.42	-5.74
HOMO (eV)	-8.17	-8.48	-8.83	-7.95	-8.03	-8.29
Band gap (eV)	2.87	2.80	3.21	2.79	2.61	2.55

Table S5. Calculated energy levels of frontier molecular orbitals for ground states of 2a-2f.

20. Photostability and thermal stability of 2a-2f



20.1 Measuring the photostability of 2a-2f in dichloromethane.

Figure S25. Measuring the photostability of **2a-2f**. (a) ~ (g) UV-Vis absorption spectra of **2a-2f** and the Rhodamine B (a commercial dye) in dichloromethane (2.0 X 10^{-5} M) with different UV irradiation time by 365 nm hand hold UV lamp (power: 12 W, distance: 5 cm). (h) Remaining percentage of the dyes **2a-2f** and Rhodamine B after photodegradation.



20.2 Measuring the thermal stability for solids of 2a-2f.

Figure S26. Measuring thermal stability of 2a-2f's solids. (a) ~ (f) Thermo gravimetric analysis (TGA) of 2a-2f.

For thermal stability, the starting decomposition temperatures for 2a, 2b, 2c, 2d, 2e and 2f were 206 °C, 162 °C, 206 °C, 206 °C, 186 °C and 157 °C, respectively. It indicates that these heterocycles have moderate thermal stability to resist high temperature. The results suggest that in these molecules, the coumarin group of the molecule has a weak thermal stability than the phenyl groups of the molecules (i.e., 2e and 2f vs. 2a, 2b and 2d), and the nitro group on the indole subunit will relatively decrease the thermal stability of molecules (i.e., 2b vs. 2a, 2f vs. 2e).

21. HRMS spectra of 2b-OH and UV-Vis spectra measurement for switch property of 2b



Figure S27. HRMS spectrum of **2b-OH** from the reaction of **2b** (4.0×10^{-4} M in DMSO) and aqueous solution of Na₂CO₃.



Figure S28. UV-Vis spectra of **2b** in DMSO (4.0×10^{-4} M) after repeated addition of aqueous solution of Na₂CO₃ and HCl.

22. Measurement of the pKa' of 2a-2f

The pKa'(s) of these heterocycles were tested by measuring the UV-Vis spectra of these heteroclcles in buffers containing 1% DMSO with pH between 6 and 13. Although **2a-2f** are responsive to OH⁻, they are not really weak acid and with no dissociation equilibrium. Considering that they could react with OH⁻ and are Lewis acid, therefore we use pKa'(s) to evaluate their acidity and the ability to react with base.



Figure S29. (a) ~ (f) UV-Vis absorption spectra of 2a-2f in buffers with pH between 6.35 and 12.96 (C = 1.0×10^{-5} M) (above) and the intensity changes at its λ_{max} (2a-2f) with pH of buffers (below).



23. HRMS spectra monitoring reaction of 2b with amines

Figure S30. 2b in MeCN reacting with multiple nucleophilic reagents. (a) Photos and structural illustration of **2b** reacting with nucleophilic reagents. HRMS spectra of **2b** reacting with (b) methylamine, (c) dimethylamine, (d) diethylamine and (e) pyrrolidine.

24. The selectivity to pH of 2b over reactive nitrogen and oxygen species



Figure S31. Effect of pH on the reaction of **2b** with nucleophilic reagents and hydrogen peroxide. The maximum absorption variation at 520 nm of **2b**'s buffer solutions (1.0×10^{-5} M) of different pH values before (grey bar) and after (light yellow or light cyan bar) adding (a) dimethylamine (1.45×10^{-4} M), (b) pyrrolidine (1.81×10^{-4} M), (c) sodium methoxide (2.44×10^{-4} M) and (d) hydrogen peroxide (1.50×10^{-4} M).

25. The cytotoxicity of 2e



Figure S32. Effects of **2e** at varied concentrations on the viability of (a) SW480, (b) A549 and (c) HeLa cells.

26. Effect of extracellular fluctuations under oxidative stress and with drug treatment on endocytosis efficiency and fluorescence of 2e



Figure S33. Measurement for effect of extracellular fluctuations of oxidative stress (Rosup) and drug treatment (5-fluorouracil (5-FU) and methotrexate (MTX)) on endocytosis efficiency of **2e** in HeLa cells by (a) flow cytometry and (b) CLSM.

27. ¹H NMR and ¹³C NMR spectra



Figure S34. ¹H NMR spectrum of 2a (400 MHz, DMSO-*d*₆).



Figure S35. ¹³C NMR spectrum of **2a** (125 MHz, DMSO-*d*₆).



Figure S36. ¹H NMR spectrum of 2b (400 MHz, DMSO-*d*₆).



Figure S37. ¹³C NMR spectrum of **2b** (100 MHz, DMSO-*d*₆).



Figure S38. ¹H NMR spectrum of 2c (500 MHz, DMSO- d_6).





Figure S40. ¹H NMR spectrum of **2d** (400 MHz, DMSO-*d*₆).



Figure S41. ¹³C NMR spectrum of 2d (126 MHz, DMSO- d_6).



Figure S42. ¹H NMR spectrum of 2e (400 MHz, DMSO-*d*₆).



Figure S43. ¹³C NMR spectrum of **2e** (100 MHz, DMSO-*d*₆).



Figure S44. ¹H NMR spectrum of **2f** (400 MHz, DMSO-*d*₆).



Figure S45. ¹³C NMR spectrum of 2f (100 MHz, DMSO- d_6).



Figure S46. ¹H NMR spectrum of I (500 MHz, DMSO-*d*₆).



Figure S47. ¹³C NMR spectrum of I (126 MHz, DMSO-*d*₆).



Figure S48. ¹H NMR spectrum of 3 (500 MHz, CDCl₃).



Figure S49. ¹³C NMR spectrum of 3 (126 MHz, CDCl₃).

28. The coordination of structures



2a

С	3.96644271	-0.84461912	0.02769471
С	5.22420769	-1.43131355	0.00684497
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Н	2.03181132	3.04334669	-0.07095273
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Н	-4.82401245	-2.26400391	-0.59884677
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Н	-8. 30759327	-3. 52106248	-1. 29297055
Н	-8.66293055	-2.01392978	-2.15316402

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