## Organocatalytic Asymmetric 5-Hydroxyisoxazolidinone Synthesis: A Highly Enantioselective Route to β-Amino acids

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## **Supporting Information**

General. Chemicals and solvents were either purchased *puriss p.A.* from commercial suppliers or purified by standard techniques. Catalyst 9 was synthesized according to litterature procedures.<sup>1</sup> For thin-layer chromatography (TLC), silica gel plates Merck 60 F254 were used and compounds were visualized by irradiation with UV light and/or by treatment with a solution of phosphomolybdic acid (25 g), Ce(SO<sub>4</sub>)<sub>2</sub>·H<sub>2</sub>O (10 g), conc. H<sub>2</sub>SO<sub>4</sub> (60 mL), and H<sub>2</sub>O (940 mL) followed by heating or by treatment with a solution of *p*-anisaldehyde (23 mL), conc. H<sub>2</sub>SO<sub>4</sub> (35 mL), acetic acid (10 mL), and ethanol (900 mL) followed by heating. Flash chromatography was performed using silica gel Merck 60 (particle size 0.040-0.063 mm), <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded on Varian AS 400. Chemical shifts are given in  $\delta$  relative to tetramethylsilane (TMS), the coupling constants J are given in Hz. The spectra were recorded in CDCl<sub>3</sub> as solvent at room temperature, TMS served as internal standard  $(\delta = 0 \text{ ppm})$  for <sup>1</sup>H NMR, and CDCl<sub>3</sub> was used as internal standard ( $\delta = 77.0 \text{ ppm}$ ) for <sup>13</sup>C NMR. HPLC was carried out using a Waters 2690 Millennium with photodiode array detector. Optical rotations were recorded on a Perkin Elemer 241 Polarimeter  $(\lambda = 589 \text{ nm}, 1 \text{ dm cell})$ . High-resolution mass spectra were recorded on a Bruker MicrOTOF spectrometer.

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**Typical experimental procedure:** To a stirred solution of catalyst **9** (20 mol %) in chloroform (0.5 mL) at 4 °C was added  $\alpha$ , $\beta$ -unsaturated aldehyde **2** (1.0 equiv. 0.25 mmol) and hydroxycarbamate **1** (1.2 equiv. 0.3 mmol). The reaction was vigorously stirred for 3 hours or 16 hours. Next, the reaction mixture was directly loaded upon a silica-gel column and immediate chromatography (pentane:EtOAc-mixtures) furnished the pure 5-hydroxyisoxazolidines **3**.



**3a**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.35-7.28$  (m, 5H), 5.89 (d, J = 4.9Hz, 1H), 5.29 (t, J = 8.4Hz, 1H), 2.76 (dd, J = 8.3Hz, J' = 12.5Hz, 1H), 2.31-2.25 (m, 1H), 1.43 (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 160.0$ , 142.3, 128.8, 127.5, 126.4, 98.8, 82.8, 61.7, 45.5, 28.3;  $[\alpha]_D^{25} = -8.0$  (c = 1.0, CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC on Daicel Chiralpak OD-H with *iso*-hexane/*i*-PrOH (97:3) as the eluent. Flow: 0.5 mL/min; minor isomer:  $t_R = 14.9$  min; major isomer:  $t_R = 13.4$  min.; HRMS (ESI): calcd. for [M+Na]<sup>+</sup>(C<sub>14</sub>H<sub>19</sub>NO<sub>4</sub>) requires m/z 288.1206, found 288.1197.



**3b**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.40-7.20$  (m, 10H), 5.84 (d, *J*=4.4Hz, 1H), 5.39 (t, *J*=8.4Hz, 1H), 5.18 (s, 2H), 2.78 (dd, *J*=8.4Hz, *J'*=12.8Hz, 1H), 2.32-2.28 (m, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 159.3$ , 141.4, 135.6, 128.6, 128.4, 128.1, 127.7, 127.4, 126.0, 98.7, 68.1, 61.3, 45.2.  $[\alpha]_D^{25} = -22.2$  (c = 1.0, CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC on Daicel Chiralpak OD-H with *iso*-hexane/*i*-PrOH (98:2) as the eluent; Flow: 1.0 mL/min; minor isomer: t<sub>R</sub> = 35.89 min; major isomer: t<sub>R</sub> = 30.78 min. HRMS (ESI): calcd. for [M+H]<sup>+</sup>(C<sub>17</sub>H<sub>17</sub>NO<sub>4</sub>) requires m/z 300.1230, found 300.1233.



**3c**: <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta = 7.30$  (d, *J*=8.8Hz, 2H), 7.27 (d, *J*=8.8Hz, 2H), 5.85 (d, *J*=4.5Hz, 1H), 5.26 (t, *J*=8.4Hz, 1H), 2.76 (dd, *J*=8.3Hz, *J'*=12.6Hz, 1H), 2.25-2.19 (m, 1H), 1.41 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 159.0$ , 141.0, 133.3, 129.0, 127.7, 98.8, 83.0, 61.2, 45.5, 28.3. [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -10.7 (c = 1.0, CHCl<sub>3</sub>).The enantiomeric excess was determined by HPLC on Daicel Chiralpak OD-H with *iso*-hexane/*i*-PrOH (98:2) as the eluent; Flow: 0.5 mL/min; minor isomer: t<sub>R</sub> = 16.5 min; major isomer: t<sub>R</sub> = 18.3 min. HRMS (ESI): calcd. for [M+Na]<sup>+</sup>(C<sub>14</sub>H<sub>18</sub>CINO<sub>4</sub>) requires m/z 322.0817, found 322.0820.



**3d**:<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta = 7.45$  (d, *J*=8.3Hz, 2H), 7.20 (d. *J*=8.3Hz, 2H), 5.83 (d, *J*=4.5Hz, 1H), 5.20 (t, *J*=8.2Hz, 1H), 2.74 (dd, *J*=8.3Hz, *J'*=12.6Hz, 1H), 2.25-2.19 (m, 1H), 1.41 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 159.0$ , 141.4, 132.0, 128.1, 121.4, 99.0, 83.1, 61.2, 45.5, 28.3.  $[\alpha]_D^{25} = -11.2$  (c = 1.0, CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC on Daicel Chiralpak OD-H with *iso*-hexane/*i*-PrOH (98:2) as the eluent; Flow: 0.5 mL/min; minor isomer: t<sub>R</sub> = 18.8 min; major isomer: t<sub>R</sub> = 20.9 min. HRMS (ESI): calcd. for [M+Na]<sup>+</sup>(C<sub>14</sub>H<sub>18</sub>Br<sup>79</sup>NO<sub>4</sub>) requires m/z 366.0311, found 366.0326.



**3e**:<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta = 7.63$  (d, *J*=8.2Hz, 2H), 7.44 (d. *J*=8.2Hz, 2H), 5.85 (d, *J*=4.5Hz, 1H), 5.31 (t, *J*=8.3Hz, 1H), 2.79 (dd, *J*=8.3Hz, *J'*=12.6Hz, 1H), 2.22-2.18 (m, 1H), 1.41 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 159.0$ , 147.7, 132.8, 127.1, 119.0, 111.5, 98.7, 83.4, 61.4, 45.4, 28.3.  $[\alpha]_D^{25} = -12.1$  (c = 1.0, CHCl<sub>3</sub>).The enantiomeric excess was determined by HPLC on Daicel Chiralpak AD with *iso*-hexane/*i*-PrOH (96:4) as the eluent; Flow: 0.5 mL/min; minor isomer: t<sub>R</sub> =

52.9 min; major isomer:  $t_R = 58.8$  min. HRMS (ESI): calcd. for  $[M+Na]^+(C_{15}H_{18}N_2O_4)$  requires m/z 313.1159, found 313.1147.



**3f**:<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 8.20$  (d, *J*=11.7Hz, 2H), 7.51 (d, *J*=11.7Hz, 2H), 5.87 (d, *J*=4.2Hz, 1H), 5.38 (t, *J*=8.1Hz, 1H), 2.83 (dd, *J*=8.1Hz, *J'*=12.3Hz, 1H), 2.23-2.19 (m, 1H), 1.43 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 158.5$ , 149.3, 147.3, 126.9, 124.0, 98.5, 83.2, 60.9, 45.2, 28.1.  $[\alpha]_D^{25} = -15.5$  (c = 1.0, CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC on Daicel Chiralpak OD-H with *iso*-hexane/*i*-PrOH (90:10) as the eluent; Flow: 1.0 mL/min; minor isomer: t<sub>R</sub> = 13.16 min; major isomer: t<sub>R</sub> = 10.75 min. HRMS (ESI): calcd. for  $[M+Na]^+(C_{14}H_{18}N_2O_6)$  requires m/z 333.1057, found 333.1041.



**3g**:<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.84-7.78$  (m, 4H), 7.48-7.42 (m, 3H), 5.90 (d, *J*=4.2Hz, 1H), 5.46 (t, *J*=8.4Hz, 1H), 2.84 (dd, *J*=8.4Hz, *J'*=12.6Hz, 1H), 2.38-2.34 (m, 1H), 1.43 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 158.8$ , 139.3, 133.3, 132.8, 128,6, 127.8, 127.6, 126.2, 125.8, 124.8, 124.1, 98.6, 82.5, 61.5, 45.4, 28.1.  $[\alpha]_D^{25} = -6.6$  (c = 1.0, CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC on Daicel Chiralpak OD-H with *iso*-hexane/*i*-PrOH (95:5) as the eluent; Flow: 1.0 mL/min; minor isomer: t<sub>R</sub> = 6.77 min; major isomer: t<sub>R</sub> = 8.24 min. HRMS (ESI): calcd. for  $[M+H]^+(C_{18}H_{22}NO_4)$  requires m/z 316.1543, found 316.1531.



**3h**:<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta = 7.39-7.31$  (m, 5H), 5.73 (d, *J*=4.4Hz, 1H), 5.23 (d, *J*=12.4Hz, 1H), 5.17 (d, *J*=12.4Hz, 1H), 4.90 (t, *J*=8.4Hz, 1H), 4.20 (q, *J*=7.2Hz, 2H), 2.59 (dd, *J*=8.4Hz, *J*'=12.8Hz, 1H), 2.49-2.45 (m, 1H) 1.25 (t, *J*=7.2Hz, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 170.5$ , 159.1, 135.5, 128.7, 128.7, 128.5, 128.4, 128.1, 98.4, 68.7, 62.0, 58.9, 39.7, 14.3.  $[\alpha]_D^{25} = -34.2$  (c = 1.0, CHCl<sub>3</sub>).The enantiomeric excess was determined by HPLC on Daicel Chiralpak OD-H with *iso*-hexane/*i*-PrOH (95:5) as the eluent; Flow: 1.0 mL/min; minor isomer: t<sub>R</sub> = 27.2 min; major isomer: t<sub>R</sub> = 23.8 min. HRMS (ESI): calcd. for [M+Na]<sup>+</sup>(C<sub>14</sub>H<sub>18</sub>N<sub>2</sub>O<sub>6</sub>) requires m/z 296.1129, found 296.1138.



**3i**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  5.69 (d, J = 4.4 Hz, 1H), 4.21 (t, J = 7.2 Hz, 1H), 2.36 (dd, J = 12.4, 8.0 Hz, 1H), 1.83-1.88 (m, 1H), 1.61-1.62 (m, 1H), 1.49-1.51 (m, 2H), 1.47 (s, 9H), 1.30-1.37 (m, 3H), 0.89 (t, J = 6.0 Hz, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  159.4, 98.8, 82.2, 58.2, 41.8, 36.1, 28.6, 28.3, 22.5, 14.1;  $[\alpha]_D^{25} = -3.4$  (c = 1.0, CHCl<sub>3</sub>); The enantiomeric excess was determined on a Chromasil CP-Chirasil-DexCB column, temperature program: 70-170 °C, rate: 10 °C/min, hold 10 min, 170-200 °C, rate: 80 °C/min, hold 5 min. Major isomer: t<sub>R</sub> = 13.958 min; minor isomer: t<sub>R</sub> = 14.211 min; HRMS (ESI): calcd. for [M+Na]<sup>+</sup>(C<sub>12</sub>H<sub>23</sub>NO<sub>4</sub>) requires m/z 268.1519, found 268.1524.



**3j**: <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) major rotamer  $\delta$  5.70 (d, J = 3.9 Hz, 1H), 5.45 (br s, 1H), 4.23 (t, J = 6.6 Hz, 1H), 2.37 (dd, J = 12.4, 8.4 Hz, 1H), 1.85-1.94 (m, 1H), 1.55-1.66 (m, 1H), 1.48-1.49 (m, 2H), 1.46 (s, 9H), 1.32-1.39 (m, 1H), 0.90-0.96 (m, 3H); minor rotamer  $\delta$  5.73 (d, J = 3.6 Hz, 1H), 4.38 (t, J = 5.7 Hz, 1H), 3.82 (br s, 1H), 2.63 (dd, J = 12.9, 6.6 Hz, 1H), 2.03-2.05 (m, 1H), 1.55-1.66 (m, 1H), 1.48-1.49 (m, 2H), 1.46 (s, 9H), 1.32-1.39 (m, 1H), 0.90-0.96 (m, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) mixture of two rotamer:  $\delta$  159.4, 155.4, 98.7, 82.4, 82.1, 57.9, 42.7, 41.8,

38.6, 28.4, 28.3, 19.3, 19.8, 14.0, 13.9;  $[\alpha]_D^{25} = -1.2$  (c = 1.0, CHCl<sub>3</sub>); The enantiomeric excess was determined on a Chromasil CP-Chirasil-DexCB column, temperature program: 70-160 °C, rate: 10 °C/min, hold 1 min, 160-200 °C, rate: 80 °C/min, hold 5 min. Major isomer:  $t_R = 11.164$  min; minor isomer:  $t_R = 11.248$  min; HRMS (ESI): calcd. for  $[M+Na]^+(C_{11}H_{21}NO_4)$  requires m/z 254.1363, found 254.1373.



**3k**: Mixture of rotamers: <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta = 7.43-7.31$  (m, 5H), 5.80 (dd, *J*=3.6Hz, *J*'=6.4Hz, 1H minor rotamer), 5.69 (dd, *J*=4.8Hz, *J*'=32.4Hz, 1H major rotamer), 5.25-5.20 (m, 2H), 4.45-4.30 (m, 1H, major rotamer), 4.05-4.00 (m, 1H, minor rotamer), 2.7-2.6 (m, 1H, minor rotamer), 2.43-2.28 (m, 1H, major rotamer), 2.10-1.35 (m, 5H), 0.96-0.93 (m, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 155.6$ , 135.9, 135.6, 128.6, 128.5, 128.4, 128.2, 128.1, 128.1 98.7, 79.6, 67.9, 67.8, 58.0, 42.3, 41.5, 41.4, 38.2, 34.9, 34.6, 19.4, 19.0, 19.0, 13.9, 13.7.  $[\alpha]_D^{25} = -5.4$  (c = 1.0, CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC on Daicel Chiralpak AD with *iso*-hexane/*i*-PrOH (98.5:1.5) as the eluent; Flow: 1.0 mL/min; minor isomer: t<sub>R</sub> = 44.1 min; major isomer: t<sub>R</sub> = 37.4 min. HRMS (ESI): calcd. for  $[M+Na]^+(C_{14}H_{19}NO_4)$  requires m/z 288.1206, found 288.1198.

Typical experimental procedure for the direct enantioselective catalytic Synthesis of 5-isoxazolidinones 10: To a stirred solution of catalyst 9 (20 mol %) in chloroform (0.5 mL) at 4 °C was added  $\alpha$ , $\beta$ -unsaturated aldehyde 2 (1.0 equiv. 0.25 mmol) and hydroxycarbamate 1 (1.2 equiv. 0.3 mmol). (In the case of the synthesis of 10a, the reaction temperature was 25 °C). The reaction was vigorously stirred for 3 hours. Upon completion (a small aliquot was removed for ee determination) The reaction temperature was increased to room temperature, isobutene (0.1 mL), *tert*butanol (0.4 mL), H<sub>2</sub>O (0.2mL) KH<sub>2</sub>PO<sub>4</sub> (54.4 mg, 4 mmol), and NaClO<sub>2</sub> (36 mg, 4mmol) were added sequentially. After 16h, the crude product 10 was purified by column chromatography (pentane/EtOAC mixtures) to afford the desired 5isoxazolidinones 10.



**10a**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.42-7.29$  (m, 5H), 5.50 (d, *J*=8.1Hz, 1H), 3.25 (dd, *J*=8.4Hz, *J*'=15.0Hz, 1H), 2.85 (dd, *J*=4.8Hz, *J*'=15.4Hz, 1H), 1.43 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 175.7$ , 157.4, 138.9, 128.8, 128.2, 127.3, 83.2, 59.6, 37.4, 28.5.  $[\alpha]_D^{25} = -13.0$  (c = 1.0, CHCl<sub>3</sub>). HRMS (ESI): calcd. for [M+Na]<sup>+</sup>(C<sub>14</sub>H<sub>17</sub>NO<sub>4</sub>) requires m/z 286.1050, found 286.1047.



**10b**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.42-7.20$  (m, 10H), 5.60 (dd, *J*=5.1Hz, *J*'=10.5Hz, 1H), 5.20-5.10 (m, 2H), 3.26 (dd, *J*=10.5Hz, *J*'=15.9Hz, 1H), 2.85 (dd, *J*=5.1Hz, *J*'=15.9Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 175.3$ , 157.4, 138.1, 135.5, 128.6, 128.5, 128.2, 128.1, 127.9, 127.2, 68.3, 59.1, 36.6.  $[\alpha]_D^{25} = -33.2$  (c = 1.0, CHCl<sub>3</sub>). HRMS (ESI): calcd. for  $[M+H_2O+Na]^+(C_{17}H_{15}NO_4)$  requires m/z 338.0999, found 338.1002.



**10c**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.38-7.32$  (m, 5H), 5.19 (s, 2H), 4.20-4.10 (m, 1H), 2.61 (t, *J*=5.2Hz, 2H), 1.72-1.35 (m, 4H), 0.91 (t, J=6.4Hz, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 175.6$ , 158.9, 135.1, 128.6, 128.5, 128.4, 82.6, 68.2, 38.5, 34.5, 18.7, 13.8.  $[\alpha]_D^{25} = -24.2$  (c = 0.7, CHCl<sub>3</sub>). HRMS (ESI): calcd. for  $[M+H_2O+Na]^+(C_{14}H_{17}NO_4)$  requires m/z 304.1155, found 304.1159.

**β**-amino acid synthesis: To a stirred solution of Cbz-protected isoxazolidinones 10 in MeOH (0.1 M), was added 10% (in weight) of Pd/C (10%). The reaction was stirred under 90 atm of Hydrogen overnight. Then the cude reaction was filtered

through a plug of Celite $\mathbb{R}$ . The solvent was removed under reduced pressure to afford the pure  $\beta$ -aminoacid **11**.



**11b**:<sup>2 1</sup>H NMR (400 MHz, D<sub>2</sub>O/K<sub>2</sub>CO<sub>3</sub>):  $\delta$  = 7.40-7.30 (m, 5H), 4.27 (t, *J*=7.2Hz, 1H), 2.60-2.45 (m, 2H); <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O/K<sub>2</sub>CO<sub>3</sub>):  $\delta$  = 179.4, 128.9, 128.8, 128.7, 127.6, 53.1, 46.7.; [ $\alpha$ ]<sub>D</sub><sup>25</sup> = - 6.9 (c = 1, H<sub>2</sub>O).



**11c**:<sup>3 1</sup>H NMR (300 MHz, MeOD)  $\delta$  3.96-3.90 (m, 1H), 2.40-2.25 (m, 2H), 1.50-1.35 (m, 4H), 0.93 (t, *J*=7.2Hz, 3H);  $[\alpha]_D^{25} = +30.8$  (c = 1, H<sub>2</sub>O).

**Experimental procedure for the one-pot synthesis of amino alcohols 12:** To a stirred solution of the catalyst **9** (16 mg, 20 mol %) in CHCl<sub>3</sub> (1 mL) was added *trans*cinnamaldehyde **2a** (33 mg, 0.25 mmol) and **1a** (40 mg, 0.3 mmol). The reaction was vigorously stirred at room temperature for 4 hours. Then the reaction mixture was diluted with MeOH (1 mL) and cooled to  $0^{\circ}$ C followed by addition of NaBH<sub>4</sub> (19 mg, 0.5 mmol). The mixture was then stirred for 10 min., quenched with HCl (1 N), and extracted with EtOAc. The organic layer was separated, dried over Na<sub>2</sub>SO<sub>4</sub> and the solvent was removed. The residue was purified by silica gel (pentane: ethyl acetate = 4:1) to give the product 58 mg (yield 87%).

**12a**: <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.27-7.42 (m, 5H), 6.91 (br s, 1H), 5.21 (dd, J = 10.8, 2.1 Hz, 1H), 3.76-3.81 (m, 2H), 2.36-2.47 (m, 1H), 2.02-2.11 (m, 1H), 1.43 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  157.2, 140.3, 128.5, 127.6, 127.4, 82.0, 60.3, 60.1, 34.2, 28.4;  $[\alpha]_D^{25} = -52.0$  (c = 0.5, CHCl<sub>3</sub>). HRMS (ESI): calcd. for [M+Na]<sup>+</sup>(C<sub>14</sub>H<sub>21</sub>NO<sub>4</sub>) requires m/z 290.1363, found 290.1355.

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