Supramolecular Gel Formation and Self-correction Induced by Aggregation-driven Conformational Changes

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Synthetic procedures:

Preparation of diamino amides shown below has been previously reported¹:



General procedure for the preparation of N-Boc-protected compounds 1a-c:

A solution of diamino amide (5.51 mmol) in 100 mL of dry DME was added dropwise over a solution of Boc-L-Pro-OSuc (11.50 mmol) in 100 mL of dry DME. The mixture was stirred at room temperature for 24 h and then at 40 °C for 5 h. Solvent was evaporated under vacuum and the resulting white solid was dissolved in 50 mL of dichloromethane and washed with NaHCO₃ (3 x 15 mL). Afterwards the organic layers were dried (Na₂SO₄) and the solvent was evaporated under vacuum to yield N-Boc-protected **1a-c** as a white solids.

N-Boc-protected 1a



Yield 89 %; m.p. 163 °C; IR (KBr) v = 3303, 2970, 2876, 1702, 1649, 1545 cm⁻¹; ¹H NMR (500 MHz, DMSO-d6): $\delta = 0.89$ (12 H, br, m), 1.34-1.54 (20H, br, m), 1.76-2.12 (10H, br, m), 3.03-3.23 (4H, br, m), 3.30-3.48 (4H, br, m), 4.11 (2H, dd, J = 8.7, 7.9 Hz), 4.20-4.31 (2H, br, m), 7.72 (2H, br, d, J = 7.8 Hz), 7.90-8.08 (2H, br, m) ppm; ¹³C NMR (300 MHz, DMSO-d6): $\delta = 19.157$, 19.904, 23.657, 28.670, 29.830, 31.295, 31.790, 36.901, 47.224, 58.599, 60.125, 79.099, 154.055, 171.411, 172.868 ppm. ESI-MS (*m/z*) = 335.3 [M + 2H]²⁺, 667.6 [M + H]⁺, 689.6 [M + Na]⁺, 705.5 [M + K]⁺.

N-Boc-protected 1b:



Yield = 83 %; m.p. 173 °C; IR (KBr) v = 3299, 2968, 2874, 1702, 1647, 1545 cm⁻¹; ¹H NMR (500 MHz, DMSO-d6): δ = 0.89 (12 H, br, m), 1.26-1.52 (26H, br, m), 1.76-2.21 (10H, br, m), 3.02-3.18 (4H, br, m), 3.30-3.47 (4H, br, m), 4.13 (2H, dd, J = 8.1, 7.9 Hz),

¹ Becerril, J; Bolte, M.; Burguete, M.I.; Galindo, F.; García-España, E.; Luis, S.V.; Miravet, J.F. *J. Am. Chem. Soc.* **2003**, *125*, 6677-6686.

4.20-4.31 (2H, br, m), 7.68 (2H, br, d, J = 8.5 Hz), 7.90-8.03 (2H, br, m) ppm; ¹³C NMR (500 MHz, DMSO-d6): δ = 19.133, 19.912, 23.643, 26.701, 28.655, 29.650, 31.386, 31.813, 38.961, 47.223, 58.484, 60.177, 79.105, 154.067, 171.255, 172.773 ppm; ESI-MS (*m*/*z*) = 709.7 [M + H]⁺, 731.6 [M + Na]⁺, 747.6 [M + K]⁺.

N-Boc-protected 1c:



Yield = 85 %; m.p. = 109 °C. IR (KBr) v = 3307, 2968, 2872, 1703, 1644, 1543 cm⁻¹; ¹H NMR (500 MHz, DMSO-d6): δ = 0.90 (12 H, br, m), 1.25-1.50 (30H, br, m), 1.75-2.20 (10H, br, m), 2.98-3.16 (4H, br, m), 3.29-3.47 (4H, br, m), 4.13 (2H, dd, J = 8.0, 7.8 Hz), 4.22-4.29 (2H, br, m), 7.67 (2H, br, d, J = 7.9 Hz), 7.87-8.05 (2H, br, m) ppm; ¹³C NMR (300 MHz, DMSO-d6): δ = 19.111, 19.897, 23.650, 26.915, 28.663, 29.303, 29.607, 31.379, 31.798, 38.984, 47.232, 58.469, 60.163, 79.106, 154.062, 171.228, 172.761 ppm. ESI-MS (*m*/*z*) = 737.6 [M + H]⁺, 759.6 [M + Na]⁺, 775.7 [M + K]⁺.

Synthesis of N-Boc-protected 2:



A solution of amino amide (6.32 mmol) in 100 mL of dry DME was added dropwise over a solution of Boc-L-Pro-OSuc (6.57 mmol) in 100 mL of dry DME. The mixture was stirred at room temperature for 24 h and then at 40 °C for 5 h. Solvent was evaporated under vacuum and the resulting white solid was dissolved in 50 mL of dichloromethane and washed with NaHCO₃ (3 x 15 mL). Afterwards the organic layers were dried (Na₂SO₄) and the solvent was evaporated under vacuum to yield N-Boc-protected **2c** as a white solid.

Yield 84 %; m.p. = 104-105 °C; IR (KBr) v = 3298, 2966, 2876, 1703, 1647, 1545 cm⁻¹; ¹H NMR (500 MHz, DMSO-d6): δ = 0.86 (9 H, m), 1.39 (9H, s), 1.41-1.51 (2H, m), 1.77-2.23 (5H, m), 3.02-3.16 (2H, m), 3.32-3.52 (2H, m), 4.11-4.19 (1H, dd, J = 8.1, 7.9 Hz), 4.21-4.31 (1H, m), 7.69 (1H, br, d, J = 8.6 Hz), 7.91-8.06 (1H, br, m) ppm; ¹³C NMR (500 MHz, DMSO-d6): δ = 12.016, 19.134, 19.927, 22.949, 23.658, 28.647, 31.394, 31.813, 40.877, 47.232, 58.507, 60.201, 79.113, 154.075, 171.309, 172.782 ppm; ESI-MS (*m*/*z*) = 356.4 [M + H]⁺, 378.4 [M + Na]⁺, [M + K]⁺.

Synthesis of compounds 1a-c and 2.

N-protected compounds (3.00 mmol) were dissolved in dichloromethane and after addition of 15 mL of TFA the mixture was stirred at room temperature for 2 h. The solvent was evaporated under vacuum and then the resulting crude oil was dissolved in water (50 mL). The solution was treated with NaOH (pH =12), and extracted with

chloroform (3 x 15 mL). The organic layers were washed with water and dried (Na_2SO_4) . Solvent was evaporated under vacuum to yield compounds **1a-c** and **2** as white solids.

Compound 1a:



Yield 84 %, m.p. 170 °C; IR (KBr) v = 3289, 2959, 2870,1638, 1545 cm⁻¹;¹H NMR (500 MHz, CD₃CN): δ = 0.89 (12 H, dd, J =19.9, 6.8 Hz), 1.53-1.60 (4H, m), 1.65-1.72 (4H, m), 1.76-1.84 (2 H, m), 2.02-2.14 (4 H, m), 2.31 (2H, br, s), 2.84-2.88 (2H, m), 3.12-3.20 (4 H, m), 3.69 (2H, dd, J = 8.8, 4.8 Hz), 4.06 (2H, dd, J = 8.8, 5.5 Hz), 6.97 (2H, br, s), 8.08 (2H, br, d, 8 Hz) ppm; ¹³C NMR (500 MHz, CD₃CN): δ = 17.171, 19.071, 26.258, 29.264, 30.752, 30.927, 35.986, 47.141, 58.002, 60.916, 171.720, 175.321 ppm; ESI-MS (*m*/*z*) = 234 [M+2]²⁺ C₂₃H₄₂N₆O₄.

Compound 1b:



Yield 87 %; m.p. = 190 °C; IR (KBr) v =3285, 2956, 2861, 1641, 1541 cm⁻¹; ¹H NMR (500 MHz, CD₃CN): δ = 0.90 (12 H, dd, J =19.8, 7 Hz), 1.28-1.34 (4H, m), 1.42-1.49 (4H, m), 1.64-1.71 (4 H, m), 1.77-1.84 (2H, m), 2.02-2.10 (4 H, m), 2.22 (2H, br, s), 2.82-2.87 (2H, m), 3.00-3.04 (2H, m), 3.12-3.18 (4H, m) 3.65 (2H, dd, J = 4.5, 9.4 Hz), 4.09 (2H, dd, J = 8.9, 6.0 Hz), 6.77 (2H, br, s), 8.08 (2H, br, d, J = 6.70 Hz) ppm; ¹³C NMR (500 MHz, CD₃CN): δ = 17.333, 19.047, 22.706, 26.061, 26.298, 29.219, 30.875, 31.348, 38.672, 47.209, 57.867, 60.903, 171.289, 175.149 ppm; ESI-MS (*m/z*) = 256 [M+2]²⁺.

Compound 1c:



Yield 79 %; m.p. 177 °C; IR (KBr) v = 3289, 2957, 2854, 1640, 1540 cm⁻¹; ¹H NMR (500 MHz, CD₃CN): δ = 0.90 (12 H, dd, J =19.4, 6.9 Hz), 1.25-1.36 (8H, m), 1.41-1.50 (4H, m), 1.65-1.72 (4 H, m), 1.78-1.85 (2H, m), 2.03-2.10 (4 H, m), 2.29 (2H, br, s), 2.83-2.88 (2H, m), 3.00-3.04 (2H, m), 3.12-3.19 (4H, m) 3.67 (2H, dd, J = 4.6, 9.3 Hz), 4.01 (2H, dd, J = 9.0, 6.2 Hz), 6.77 (2H,br, s), 8.08 (2H, br, d, J = 9.04 Hz) ppm; ¹³C NMR (500 MHz, CD₃CN): δ = 17.289, 19.087, 22.706, 26.291, 26.467, 28.839, 29.289, 30.836, 31.410, 38.955, 47.217, 57.791, 60.896, 171.188, 175.102 ppm; ESI-MS (*m/z*)= 269 [M+2]²⁺.

Compound 2:



Yield = 79 %; m.p. = 127 °C; IR (KBr) v = 3303, 2963, 2873, 1467, 1518 cm⁻¹; ¹H NMR (500 MHz, CD₃CN): δ = 0.86-0.92 (9 H, m), 1.45-1.52 (2 H, m), 1.66-1.71 (2 H, m), 1.78-1.84 (1 H, m), 2.02-2.10 (2H, m), 2.39 (1H, br, s), 2.83-2.87 (1H, m), 3.00-3.05 (1H, m), 3.07-3.19 (2H, m), 3.67 (1H, dd, J = 9.2, 5Hz), 4.11 (1 H, dd, J = 8.9, 6.1 Hz), 6.70 (1H, br, s), 8.07 (1H, br, d, J = 6.70 Hz) ppm; ¹³C NMR (500 MHz, CD₃CN): δ = 10.971, 173.323, 19.032, 22.706, 26.288, 30.878, 31.484, 40.838, 47.205, 57.757, 60.878, 171.231, 175.089 ppm; ESI-MS (*m/z*) = 256 [M+H]⁺, 278 [M+Na]⁺.

Gelation studies.

Typically, the desired amount of the gelator in the corresponding solvent was heated in a screw-capped vial until it was completely dissolved and, afterwards, it was left to cool by two different methods:

Spontaneous cooling:

The sample was cooled at room temperature during 24 h and afterwards it was stabilized at r.t. during 12 h.

Slow cooling:

The sample was cooled in a thermoregulated bath (0.5 $^{\circ}\mathrm{C}$ / min) and stabilized at r.t. during 12 h.

To determine the minimum gel concentration values reported, 5-20 mmol of the studied compounds were dissolved in 1 mL of hot solvent in a screw-capped cylindrical glass vial (diameter = 2 cm). The formation of a gel was checked by turning the vial upside down. Thus, when, upon vial inversion, all the solvent remains entrapped within the gel and does not fall-down we consider it as a gel (G).

Table S1. Gelation of compounds **1a-c** in different organic solvents after spontaneous cooling at 25 $^{\circ}$ C.^a

	1a	1b	1c
Tetrahydrofuran	G (30)	S	S
Dimethoxyethane	G (24)	S	S
Toluene	G (50)	G (7)	G (11)
Ethyl acetate	G (6)	G (7)	G (9)
Acetonitrile	G (34)	G (16)	G (14)

a. G: gel; S: soluble. Minimum gel concentration (mM) in parentheses.

Scanning Electron Microscopy. Scanning electron micrographs were taken in a LEO 440I microscope equipped with a digital camera. Samples of the xerogels were prepared by placing the gel on top of a tin plate and, after vacuum drying of the solvent, sputtering with Au/Pd in a Polaron SC7610 Sputter Coater from Fisons Instruments.

NMR. ¹H and ¹³C NMR spectra were recorded in a Varian Mercury 300 MHz spectrometer at 30 °C. NOE (1D-NOESY and NOESY) measurements were carried out in a Varian Inova 500 MHz spectrometer. Gel samples were prepared by transferring a hot solution of the gelator in CD₃CN to the NMR tube. Upon cooling to room temperature a gel was formed and the sample obtained was analyzed using the same parameters as for solution NMR (except in the case of NOE experiments where the mixing time for gel samples was 20 ms -to avoid spin diffusion- and for solutions 500 ms).

Molecular modeling. The models reported were obtained by molecular mechanics calculations performed with MACROMODEL 8.0 using AMBER* as force-field and a GB/SA simulation of chloroform as solvent. The structures of the isolated molecules correspond to minimum energy conformers found after exhaustive Monte Carlo conformational search. The models for the aggregates were energetically minimized.

X-ray powder diffraction. Data collection was performed at room temperature on a Siemens D5000 diffractometer using Cu-K_{α} radiation. Samples of the powdered solids were placed on a quartz sample holder and data were collected for 2 θ values between 3° and 35° with a step size of 0.05° and a time step of 20 s.

CD spectroscopy. CD spectra were recorded in a Jasco J-810 spectropolarimeter equipped with a Peltier heating device. The samples were prepared in quartz cuvettes of 1 mm and 0.5 mm of path length.



N-Boc-protected 1a.





N-Boc-protected 1b.





N-Boc-protected 1c.





N-Boc-protected 2.







Compound 1a:





Compound 1b:









Compound 1c:

Compound 1d:





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¹H-RMN CD₃CN



	Compound 1a 0.002 M	Compound 1 b 0.001 M	Compound 1 c 0.002 M	Compound 2 0.004 M
Signal	δ (ppm)	δ (ppm)	δ (ppm)	δ (ppm)
а	6.98	6.71	6.72	6.61
b	8.09	8.07	8.07	8.04
с	3.67	3.67	3.69	3.68
d	3.17	3.15	3.16	3.13
e	1.54	1.45	1.46	1.49
f	2.08	2.05	2.07	2.05
g	0.89	0.90	0.87	0.90
h	4.05	4.08	4.10	4.10
i	2.08	2.05	2.07	2.05
j	1.78	1.80	1.82	1.80
k	1.67	1.69	1.70	1.69
1	1.67	1.69	1.70	1.69
m	3.00	3.02	3.04	3.02
n	2.87	2.85	2.87	2.84
0	2.15	2.15	2.18	2.15
р		1.32	1.30	0.87
q			1.30	

¹³C-RMN CD₃CN



	Compound 1a 0.004 M	Compound 1 b 0.004 M	Compound 1 c 0.004 M	Compound 2 0.004 M
Signal	δ (ppm)	δ (ppm)	δ (ppm)	δ (ppm)
0			26.467	
1		26.061	28.839	10,927
2	29.278	29.219	29,289	22,694
3	35.973	38.672	38,955	40,820
4	171.715	171.289	171,188	171,193
5	58.003	57.867	57,791	57,830
6	30.929	31.348	31,410	31,382
7	19.073	19.047	19,089	19,015
8	17.167	17.333	17,289	17,249
9	175.278	175.149	175,102	174,854
10	60.923	60.903	60,896	60,850
11	30.753	30.875	30,836	30,835
12	26.261	26.298	26,291	26,237
13	47.149	47.209	47,217	47,201



Figure S1. SEM images of xerogels of compound **1b**: A) CH₃CN, spontaneous cooling; B) *idem*, slow cooling; C) toluene, spontaneous cooling.



Figure S2. SEM images of xerogels and crystalline precipitates of compound 1c: A) CH₃CN, spontaneous cooling; B,C) *idem*, slow cooling; D) EtOAc, spontaneous cooling.



Figure S3. $\Delta\delta$ vs concentration for the amide signals **a** and **b** of compounds **1a-c** and **2** (CD₃CN, 30 °C)

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Compound 1a



Compound 1b













Compound 2



Figure S4. $\Delta\delta$ vs temperature for the amide signals **a** and **b** of compounds **1a-c** and **2** (CD₃CN, 30 °C)



Figure S5. Informative NOE contacts for non-aggregated solutions of compounds **1a** (CD₃CN, 30 °C)



Figure S6. Transfer-NOE contacts for gels of compounds 1a (CD₃CN, 30 °C). Arrows indicate variations from diluted solution.



Figure S7. NOE spectra for a non-aggregated solution (top) and a gel (bottom) of compound **1a** (CD_3CN , 30 °C).



Figure S8. informative NOE contacts for non-aggregated solutions of compounds **1b** - **c** (CD₃CN, 30 $^{\circ}$ C)



Figure S9. Changes in-NOE contacts for gels of compounds **1b-c** $(CD_3CN, 30 \ ^{\circ}C)$

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Figure S10. WAXD patters of xerogels of compound 1a.



Figure S11. WAXD patters of compound **1b**.



Figure S12. WAXD patters of compound **1c**.





В



1b

1c

Figure S13. Molecular models for compounds **1b** and **1c** in solution (A) and in aggregates (B). (Macromodel 9, AMBER*, $CHCl_3$)