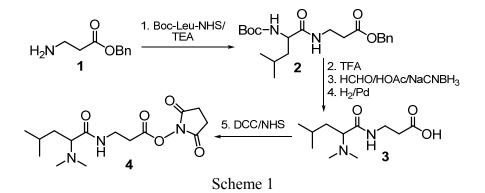
## **Supporting Information**

#### 1. Synthesis of non-isotope labeled DiART reagent



#### **Compound 2**

The synthesis of compound  $\mathbf{2}$  was previously reported<sup>1</sup>. It was synthesized from compound  $\mathbf{1}$  here by using a different method.

Compound **1** (528 mg, 1.5 mmol)<sup>2</sup>, Boc-Leu-NHS (493 mg, 1.5 mmol)<sup>3</sup>, and TEA (700  $\mu$ L, 5.0 mmol) were added into 20 mL DMF. After the mixture was stirred for 5 h at room temperature, the solvent was removed *in vacuo*. The residue was dissolved in 60 mL DCM, washed with 10% K<sub>2</sub>CO<sub>3</sub> (2×60 mL) and 1M KHSO<sub>4</sub> (2×60 mL), then dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. After the solvent was removed, the residue was purified by silica gel chromatography to give compound **2** (447 mg, 76% yield) as colorless oil. <sup>1</sup>**H-NMR (CDCl<sub>3</sub>, 500 MHz)**  $\delta$  = 7.35-7.23 (m, 5H), 5.11 (s, 2H), 4.20 (m, 1H), 3.50 (t, J = 6.1 Hz, 2H), 2.56 (t, J = 6.0 Hz, 2H), 1.70-1.64 (m, 2H), 1.53-1.46 (m, 1H), 1.41 (S, 9H), 0.92 (d, J = 5.5 Hz, 3H), 0.89 (d, J = 5.6 Hz, 3H). **MS** (M+H)<sup>+</sup> = 393.31, Cal. (M+H)<sup>+</sup> = 393.23.

#### **Compound 3**

Compound **2** (392 mg, 1.0 mmol) was stirred in 10 mL deprotection cocktail (47.5% TFA, 2.5% H<sub>2</sub>O, 50% DCM) for 1 hr, then the solvent was removed *in vacuo* and the residue was dried in a lyophilizer. To the reaction flask, 30 mL CH<sub>3</sub>CN and 1.0 mL HOAc were added, followed by addition of aqueous HCHO (40%, 400  $\mu$ L). After the solution was stirred for 5 min, NaCNBH<sub>3</sub> (124mg, 2.0 mmol) was added. The reaction mixture was stirred for another 4 h at room temperature. After the solvent was removed, the residue was dissolved in 80 mL ethyl acetate, washed with 10% K<sub>2</sub>CO<sub>3</sub> (2×60 mL), and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. After the organic solvent was removed, the crude product was dissolved in 20 mL methanol, and then 30 mg Pd/C catalyst was added to the solution. The hydrogenation was carried out with a 200 mL hydrogen balloon. After the reaction mixture was stirred overnight, the solid catalyst was filtered out and the filtrate was evaporated. The residue was purified by Dowex 50 (H<sup>+</sup> form) chromatography to give compound **3** (163 mg, 71% yield) as white solid.

<sup>1</sup>**H-NMR (D<sub>2</sub>O, 500 MHz)**  $\delta$  =3.56 (m, 1H), 3.28 (t, J = 6.5 Hz, 2H), 2.67 (S, 6H), 2.22 (t, J = 6.3 Hz, 2H), 1.64-1.51 (m, 2H), 1.35-1.24 (m, 1H), 0.72 (d, J = 6.7 Hz, 3H), 0.70

(d, J = 6.7 Hz, 3H); <sup>13</sup>C-NMR (D<sub>2</sub>O, 500 MHz)  $\delta$  = 177.0, 167.9, 67.7, 41.4, 36.9, 36.0, 33.1, 25.1, 22.8, 20.4 ; MS (M+H)<sup>+</sup> = 231.10, Cal. (M+H)<sup>+</sup> = 231.16

# **Compound 4**

Compound **3** (115 mg, 0.50 mmol) was added to the mixture of NHS (59 mg, 0.51mmol) and DCC (113 mg, 0.55mmol) in 10 mL DCM. The reaction mixture was stirred for 2 days at room temperature and insoluble solid was filtered out. After the solvent was evaporated, the residue was recrystallized from diisopropyl ether to afford compound **4** (106 mg, 65% yield) as white solid.

<sup>1</sup>**H-NMR (CDCl<sub>3</sub>, 500 MHz)**  $\delta$  =3.61 (m, 1H), 3.27 (t, J = 6.5 Hz, 2H), 2.82 (t, J = 6.3 Hz, 2H), 2.66 (S, 6H), 2.61 (S, 4H), 1.74-1.61(m, 2H), 1.43-1.37 (m, 1H), 0.94 (d, J = 6.7 Hz, 3H), 0.92 (d, J = 6.7 Hz, 3H). <sup>13</sup>**C-NMR (CDCl<sub>3</sub>, 500 MHz)**  $\delta$  = 174.0, 169.1, 168.0, 66.8, 41.8, 36.4, 34.8, 31.4, 25.6, 25.0, 23.5, 21.7. **MS** (M+H)<sup>+</sup> = 328.11, Cal. (M+H)<sup>+</sup> = 328.18.

	DiART-114	DiART-115	DiART-116	DiART-117	DiART-118	DiART-119
Leu-OH	1- <sup>13</sup> C	<sup>15</sup> N	1- <sup>13</sup> C	<sup>15</sup> N	1- <sup>13</sup> C	<sup>15</sup> N
НСНО	unlabeled	unlabeled	unlabeled	unlabeled	DCDO	DCDO
NaCNBH <sub>3</sub>	unlabeled	unlabeled	NaCNBD <sub>3</sub>	NaCNBD <sub>3</sub>	unlabeled	unlabeled
β-Ala-OH	$H_2N \xrightarrow{D} D \xrightarrow{O} OH$	$\begin{array}{c} D & D \\ D & D \\ H_2 N \\ D \\ D \\ D \end{array} OH$	D O H₂N D OH		unlabeled	unlabeled

Table 1. Isotope-labeled starting materials for DiART reagents

 $1^{-13}$ C and  $^{15}$ N labeled leucine were purchased from Cambridge Isotope Laboratory, Inc. DCDO and NaCNBD<sub>3</sub> were purchased from Sigma. D<sub>4</sub>- $\beta$ -alanine was

purchased from C/D/N Isotopes Inc.  $D_2$ - $\beta$ -alanine was synthesized in house<sup>4</sup>.

# 2. HPLC of DiART-labeled phenylalanine

Six different amount of phenylalanine were reacted with each excess DiART reagent in a mixture (NaHCO<sub>3</sub>, 50 mM, and 1,4-dioxane, 50%), respectively, mixed, and then purified with  $C_{18}$ -HPLC (Buffer A: water, 0.1% TFA; Buffer B: acetonitrile, 0.1% TFA; gradient: 0-6 mins 10% B, 6-8 mins 10% to 20% B, 8-48 mins 20% B to 60% B; flow rate: 1 mL/min) (see Figure 1 in this supporting information). The purified product was re-injected into HPLC using the same program. A single broad peak of the product was observed. Three time fractions of this peak were collected and analyzed with MALDI-MS/MS (see Figure 2a in paper).

# 3. Protein quantitation with DiART reagents and Mascot software

# 3.1 Detailed procedure

BSA (20  $\mu$ g), bovine catalase (20  $\mu$ g), and chicken ovalbumin (10  $\mu$ g) were dissolved in 100  $\mu$ L denaturing/reducing solution (8 M urea, 50 mM Sodium borate buffer, pH = 8.3, 5 mM TCEP) and incubated at 37 °C for 30 min. Then, 20 mM 2-bromoacetamide was added to alkylate free cysteine residues. The proteins were

precipitated with acetone, dissolved again in 100  $\mu$ L buffer (200 mM Sodium borate buffer, pH = 8.3, 0.8 M urea), and then digested with trypsin (10  $\mu$ g) at 37 °C overnight. Six fractions of samples (10  $\mu$ L) were mixed with 20  $\mu$ L DiART reagents (2 mg/mL in ethanol), respectively and the reaction was incubated at room temperature for 4 h. All of six samples were then mixed together, dried in a SpeedVac, then dissolved again in a SCX (strong ion exchange) loading buffer (10 mM KH<sub>2</sub>PO<sub>4</sub>, pH = 3.0, 25% acetonitrile). This sample was loaded onto a SCX column, washed, and eluted with 500  $\mu$ L elution buffer (10 mM KH<sub>2</sub>PO<sub>4</sub>, pH = 3.0, 400 mM KCl, 25% acetonitrile). The elution was dried in SpeedVac and dissolved in 100  $\mu$ L 5% acetonitrile. 4  $\mu$ L of labeled peptide sample was injected into a capillary reverse phase HPLC (gradient, 5% acetonitrile, 0.1% TFA to 50% acetonitrile, 0.1% TFA in 60 min; column, Agilent Zorbax C18, 5  $\mu$ m, 150×0.5 mm; flow rate, 15  $\mu$ L/min) and each fraction was analyzed with ABI-4700 MALDI-MS/MS.

# 3.2. Isotope purity of DiART reagents

The isotope purity of DiART reagents was determined as described previously<sup>5</sup>. These numbers were included in a Mascot server configuration file in order to get correct quantitation.

# Table-2. Isotope purity

			· · · · · · · ·		
$\Delta(M/Z)$	-2	-1	0	+1	+2
DiART-114	0.000	0.000	0.925	0.075	0.000
DiART-115	0.000	0.065	0.842	0.093	0.000
DiART-116	0.000	0.076	0.840	0.084	0.000
DiART-117	0.000	0.116	0.815	0.069	0.000
DiART-118	0.000	0.000	0.914	0.086	0.000
DiART-119	0.000	0.092	0.821	0.087	0.000

3.3. Compatibility of DiART reagents with custom-configured Mascot sever

Only two configuration files in Mascot sever (version 2.2 only) need to be updated to make the server compatible with DiART-based protein identification and quantitation. These two files (unimod.xml and quantitation.xml) will be distributed freely upon request. A result of Mascot searching is attached as an example.

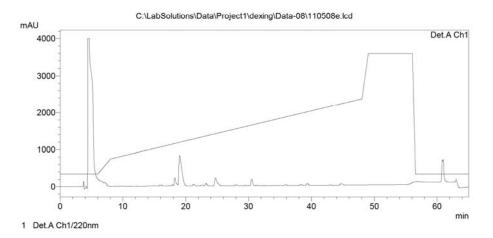
# 4. References

- 1) Murphy, R.F.; Douglas, A.J.; Walker, B. US Patent 4997950, 1991.
- 2) Erickson, B.W. US Patent 4515920, 1985.
- Nakamura, M.; Yamaguchi, M.; Sakai, O.; Inoue, J. Bioorg. Med. Chem. 2003, 11, 1371–1379.
- 4) Hanai, K.; Kuwae, A. J. Labelled Comp. Rad., 1988, 25, 217-224.
- 5) Zeng, D.; Li, S.W. Bioorg. Med. Chem. Lett., 2009, 19, 2059-2061.

## ==== Shimadzu LCsolution Analysis Report ====

	C:\LabSolutions\Data\Project1\dexing\Data-08\110508e.lcd
Acquired by	: Admin
Sample Name	:1
Sample ID	
Tray#	:1
Vail #	:2
Injection Volume	: 100 uL
Data File Name	: 110508e.lcd
Method File Name	: 40min-philic.lcm
Batch File Name	: 050908.lcb
Report File Name	: Default.lcr
Data Acquired	: 2008-11-5 22:10:13
Data Processed	: 2008-11-5 23:15:15

#### <Chromatogram>



C:\LabSolutions\Data\Project1\dexing\Data-08\110508e.lcd

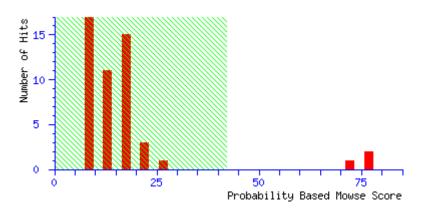
Figure 1

# SCIENCE Superascotris Searchic Results ns This journal is (c) The Royal Society of Chemistry 2009

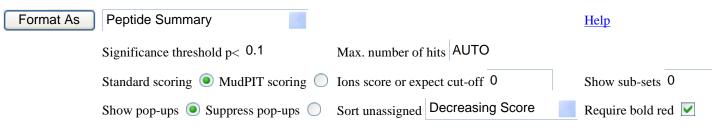
User Email Search title MS data file Database Quantitation Timestamp	: E:\Laboratory\Projects\Proteomics\DiART\NewMultiFile2.mgf : MSDB 20060831 (3239079 sequences; 1079594700 residues)									
Protein hits	: 115/114	116/114	117/114	118/114	119/114					
	0.996	1.022	1.056	1.095	0.910	CATA_BOVIN	Catalase (EC 1.11.1.6) Bos taurus (Bovine).			
						OACH	ovalbumin [validated] - chicken			
	1.017	1.053	1.086	1.142	1.044	ABBOS	serum albumin precursor [validated] - bovine			

# **Probability Based Mowse Score**

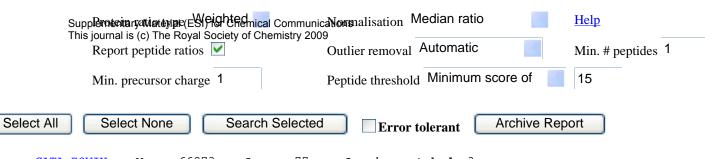
Ions score is -10\*Log(P), where P is the probability that the observed match is a random event. Individual ions scores > 42 indicate identity or extensive homology (p<0.1). Protein scores are derived from ions scores as a non-probabilistic basis for ranking protein hits.



# **Peptide Summary Report**



Peptide Summary Report (Submitted from DiART-New by Mascot Daemon on LIW-XP)



1. <u>CATA\_BOVIN</u> Mass: 66273 Score: 77 Queries matched: 3 Catalase (EC 1.11.1.6).- Bos taurus (Bovine).

Check to include this hit in error tolerant search or archive report

Quantitation:	Ratio	Weighted	N	SD(geo)
	115/114	0.996	3	1.073
	116/114	1.022	3	1.039
	117/114	1.056	3	NN
	118/114	1.095	3	1.061
	119/114	0.910	3	1.060

Quer	У	Observed	Mr(expt)	Mr(calc)	Delta	Miss	Score	Expect	Rank	115/114	116/114	117/114	118/114	119/114	Peptide
~	<u>1</u> 1	173.7300	1172.7227	1172.7260	-0.0033	0	31	1.8	1	0.901	0.950	1.170	1.024	0.818	K.LNSLTVGPR.G
~	<u>2</u> 1	263.6600	1262.6527	1262.6751	-0.0223	0	18	48	1	1.076	1.066	1.000	1.162	0.973	R.THFSGDVQR.F
~	<u>4</u> 1	502.8000	1501.7927	1501.7908	0.0019	0	30	1.7	1	0.889	1.015	1.000	0.977	0.875	R.LAHEDPDYGLR.D

Proteins matching the same set of peptides:
CSBO Mass: 63859 Score: 77 Queries matched: 3
catalase (EC 1.11.1.6) [validated] - bovine
<u>4BLCA</u> Mass: 62642 Score: 77 Queries matched: 3
catalase (EC 1.11.1.6), chain A - bovine
8CATA Mass: 62513 Score: 77 Queries matched: 3
catalase (EC 1.11.1.6), chain A - bovine
AAI03067 Mass: 66404 Score: 77 Queries matched: 3
BC103066 NID: - Bos taurus

2. <u>OACH</u> Mass: 47756 Score: 75 Queries matched: 1 ovalbumin [validated] - chicken

Check to include this hit in error tolerant search or archive report

Quantitation:	Ratio	Weighted	N	SD(geo)
	115/114		1	
	116/114		1	

117/114 --- 1 ---Supplementary Material (ESI) for Chemical Gommunications This journal is (ch19/1R0yal Society of Chemistry 2009

Query Observed Mr(expt) Mr(calc) Delta Miss Score Expect Rank 115/114 116/114 117/114 118/114 119/114 Peptide 6 1904.9800 1903.9727 1904.0135 -0.0408 0 75 4e-005 1 1.028 0.985 0.964 0.877 1.201 R.GGLEPINFQTAADQAR.E

#### Proteins matching the same set of peptides:

OVAL CHICK Mass: 47625 **Score:** 75 Oueries matched: 1 Ovalbumin (Plakalbumin) (Allergen Gal d 2) (Gal d II).- Gallus gallus (Chicken). 10VAB2 Mass: 37887 **Score:** 75 Queries matched: 1 ovalbumin, chain B, fragment 2 - chicken 10VAC1 Mass: 42648 **Score:** 75 Queries matched: 1 ovalbumin, chain C, fragment 1 - chicken CAA23681 Mass: 19424 **Score:** 75 Queries matched: 1 GGALB1 NID: - Gallus gallus CAA23682 Mass: 47786 **Score:** 75 Queries matched: 1 GGALB2 NID: - Gallus gallus CAC28424 Mass: 116079 Score: 75 Oueries matched: 1 Sequence 20 from Patent WO0104344.- Cloning vector pINT1. CAC28434 Mass: 50198 Queries matched: 1 **Score:** 75 Sequence 54 from Patent WO0104344. - Cloning vector pINT1. AA043266 Mass: 47780 **Score:** 75 Queries matched: 1 AY223553 NID: - Gallus gallus

3. <u>ABBOS</u> Mass: 84469 Score: 73 Queries matched: 2 serum albumin precursor [validated] - bovine

Check to include this hit in error tolerant search or archive report

Quantitation:	Ratio	Weighted	N	SD(geo)
	115/114	1.017	2	1.036
	116/114	1.053	2	1.169
	117/114	1.086	2	1.257
	118/114	1.142	2	1.194
	119/114	1.044	2	1.026

Q	uery	Observed	Mr(expt)	Mr(calc)	Delta	Miss	Score	Expect	Rank	115/114	116/114	117/114	118/114	119/114	Peptide
~	<u>3</u>	1356.8200	1355.8127	1355.8426	-0.0299	0	39	0.25	1	0.973	0.860	0.802	0.905	1.078	K.AEFVEVTK.L
~	<u>5</u>	1502.8200	1501.8127	1501.7961	0.0166	0	34	0.71	1	1.038	1.145	1.220	1.254	1.028	K.QNCDQFEK.L

Serum albumin (Fragment).- Bos indicus (Zebu). <u>AAI0 Supplementary Material (ESI) for Chemical Communications</u> BC102742 NID: - Bos taurus <u>AAN17824</u> Mass: 84522 Score: 73 Queries matched: 2 AF542068 NID: - Bos taurus <u>AAA51411</u> Mass: 84492 Score: 73 Queries matched: 2 BOVALBUMIN NID: - Bos taurus

## **Search Parameters**

Type of search : MS/MS Ion Search Enzyme : Trypsin Fixed modifications : Carbamidomethyl (C), DiART6plex (K), DiART6plex (N-term), DiART6plex (K), DiART6plex (N-term) Mass values : Monoisotopic Protein Mass : Unrestricted Peptide Mass Tolerance : ± 0.8 Da Fragment Mass Tolerance: ± 0.5 Da Max Missed Cleavages : 0 Instrument type : Default Number of queries : 6

Mascot: http://www.matrixscience.com/