

An Efficient and Recyclable Hybrid Nanocatalyst to Promote Enantioselective Radical Cascade Rearrangements of Eneidyne

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1. Preparation of hybrid materials

1.1. MCM41

The MCM-41 mesoporous silica was synthesized by adopting the procedure reported by Grün et al.¹ In a standard method of synthesis, 0.80 g of cetyltrimethylammonium bromide (CTAB), was dissolved in 17.7 mL of water under vigorous stirring at 70 °C for 10 min. After cooling (30 °C), a mixture of 10.3 mL of absolute ethanol and 8.1 mL of ammonium hydroxide 32% was added and the solution was stirred for an additional 10 min. Then, 1.5 g (7.2 mmol) of tetraethyl orthosilicate (TEOS (98%), commercially available from Aldrich) was added dropwise to the solution with stirring for an additional 24 h. The molar composition of the resultant mixture was 1 TEOS: 0.3 CTAB: 11 NH₃: 144 H₂O: 58 EtOH. After hydrothermal treatment at 90 °C for 24 hours, the precipitate was filtered, washed with deionized water, ethanol and diethyl ether. For acidic extraction, the as-obtained material (1 g) was treated with a mixture of ethanol (100 ml) and concentrated HCl (1 mL, 38% in weight) at 70 °C for 8 h. The resulting (surfactant removed) solid product was filtered and washed with ethanol, and then dried at 60 °C.

1.2. SBA15

In a typical synthesis, 4 g of Pluronic P123 were dissolved in 30 mL of deionized water and 120 mL of 2M HCl under constant stirring for three hours, prior to the addition of 8.5 g of TEOS. This mixture was further stirred for 24 hours before hydrothermal treatment at 100 °C for 48 hours. The resulting white precipitate was filtered, which was followed by soxhlet extraction of the polymeric template with ethanol for 24 hours. The molar composition of the synthesis mixture was as follows: 1 TEOS: 0.017M P123 Polymer: 188M H₂O: 5.8M HCl. The resulting (surfactant removed) solid product was filtered and washed with ethanol, acetone and diethyl ether and then dried at 60 °C.

1.3. Passivated SBA15

In a typical synthesis, 4 g of Pluronic P123 were dissolved in 30 mL of deionized water and 120 mL of 2M HCl under constant stirring for three hours, prior to the addition of 8.5 g of TEOS. This mixture was further stirred for 24 hours before hydrothermal treatment at 100 °C for 48 hours. The resulting white precipitate was filtered, and washed with ethanol, acetone and diethyl ether and then dried at 60 °C. The molar composition of the synthesis mixture was as follows: 1 TEOS: 0.017M P123 Polymer: 188M H₂O: 5.8M HCl. SBA15 still containing the surfactant (1 g) was loaded in dry toluene (75 ml) and triethylamine (4.04 g, 40 mmol), then chlorotrimethylsilane (3.56 g, 33.3 mmol) was added, and the mixture was heated at 70 °C overnight. The resulting white precipitate was filtered, which was followed by soxhlet extraction of the polymeric template with ethanol for 24 hours. The resulting (surfactant removed) solid product was filtered and washed with ethanol, acetone and diethyl ether and then dried at 60°C. The grafting procedure is described in § 1.4.4.

¹ M. Grün, K.K. Unger, A. Matsumoto and K. Tsutsumi, *Micropor. Mesopor. Mater.*, 1999, **27**, 207.

1.4. Grafting procedure

- 1.4.1.** Diethylaminopropyltrimethoxysilane (DEAP) was anchored onto the silica surface via post-grafting procedure. After SBA15 (1 g, 16.7 mmol) was loaded in dry toluene (16 ml), DEAP (1.35 g, 5.7 mmol) was added, and mixture was heated at 70 °C overnight. The precipitate was filtered, washed (ethanol and ether) and dried which was referred as **GA-SBA15**. The quantity of organic group was determined by TGA analysis; it is 1.45 mmol/g.
- 1.4.2.** Diethylaminopropyltrimethoxysilane (DEAP) was anchored onto the silica surface via post-grafting procedure. After MCM41 (1 g, 16.7 mmol) was loaded in dry toluene (16 ml), DEAP (1.35 g, 5.7 mmol) was added, and mixture was heated at 70 °C overnight. The precipitate was filtered, washed (ethanol and ether) and dried which was referred as **GA-MCM41**. The quantity of organic group was determined by TGA analysis; it is 1.57 mmol/g.
- 1.4.3.** Diethylaminopropyltrimethoxysilane (DEAP) was anchored onto the silica surface via post-grafting procedure. After Si2000 (1 g, 16.7 mmol) was loaded in dry toluene (16 ml), DEAP (1.35 g, 5.7 mmol) was added, and mixture was heated at 70 °C overnight. The precipitate was filtered, washed (ethanol and ether) and dried which was referred as **GA-Si2000**. The quantity of organic group was determined by TGA analysis; it is 0.21 mmol/g.
- 1.4.4.** Diethylaminopropyltrimethoxysilane (DEAP) was anchored onto the silica surface via post-grafting procedure. After SBA15 (0.6 g) was loaded in dry toluene (11 ml), DEAP (0.78 g, 3.3 mmol) was added, and mixture was heated at 70 °C overnight. The precipitate was filtered, washed (ethanol and ether) and dried which was referred as passivated **GA-SBA15**.

1.5. Characterization of Mesoporous Silicas

Thermogravimetric (TGA) measurements were carried out with a TGA Q500 apparatus (TA Instruments) under dynamic air atmosphere (sample flow rate 60 ml/min). XRD patterns were measured on a Siemens D5000; XRD diffractometer using Cu (40 Kv, 40 mA) Ka radiation and detected by Scintillator (Thêta-2 Thêta). The N₂ adsorption/desorption isotherms were obtained at 77 K on a Micrometrics ASAP2010. The specific surface area was determined with the Brunauer, Emmett, and Teller (BET) method and the pore size distribution was calculated from the desorption isotherms using the Barrett Joyner Halenda (BJH) method^{1,2}. Prior to adsorption, the samples were out gassed at 393 K overnight under a vacuum pressure of 2×10^{-3} mbar. All solid-state Cross Polarization Magic Angle Spinning (CPMAS) NMR spectra were obtained on a Bruker Avance-400 MHz NMR spectrometer operating at a ¹³C and ²⁹Si resonance frequency of 101.6 MHz and 79.5MHz, respectively. ¹³C and ²⁹Si CPMAS experiments were performed with a commercial Bruker Double-bearing probe. About 100 mg of samples were placed in zirconium dioxide rotors of 4-mm outer diameter and spun at a Magic Angle Spinning rate of 10 kHz. The CP technique³ was applied with a ramped 1H-pulse starting at 100 % power and decreasing until 50% during the contact time in order to

² F. Rouquerol, J. Rouquerol, K. S. W. Sing, Adsorption by Powders and Porous Solids: Principles, Methodology and Applications, Academic Press: London, 1999.

³ J. Schaefer and E. O. R. Stejskal, *J. Am. Chem. Soc.*, 1976, **98**, 1031.

circumvent Hartmann-Hahn mismatches (Peersen *et al.*, 1993; Cook *et al.*, 1996)⁴. The contact times were 2 ms for ¹³C CPMAS and 5ms for ²⁹Si CPMAS. To improve the resolution, a dipolar decoupling GT8 pulse sequence (Gerbaud *et al.* 2003)⁵ was applied during the acquisition time. To obtain a good signal-to-noise ratio, 6144 scans were accumulated using a delay of 2 s in ¹³C CPMAS experiment, and 4096 scans with a delay of 5 s in ²⁹Si CPMAS experiment. The ¹³C and ²⁹Si chemical shifts were referenced to tetramethylsilane.

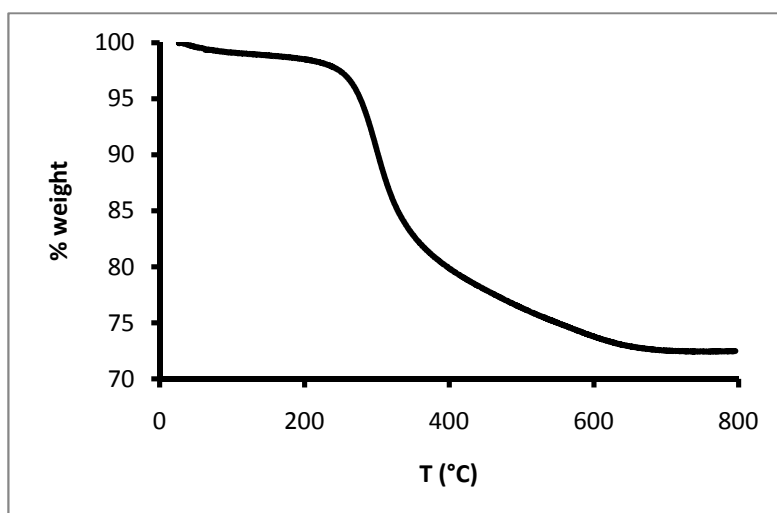


Figure 1: TGA analysis of GA-SBA15 (5.8 nm)

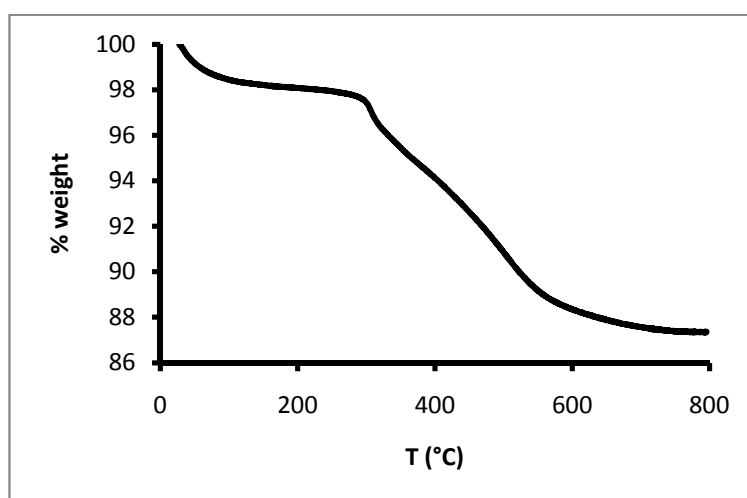


Figure 2: TGA analysis of passivated SBA15 before grafting (5.8 nm)

⁴ a) O. B. Peersen, X. Wu, I. Kustanovich and S.O. Smith, *J. Magn. Reson.*, 1993, **104**, 334. b) R. L. Cook, C. H. Langford, R. Yamdagni and C. M. Preston, *Anal. Chem.*, 1996, **68**, 3979.

⁵ G. Gerbaud, F. Ziarelli and S. Caldarelli, *Chem. Phys. Lett.*, 2003, **377**, 1.

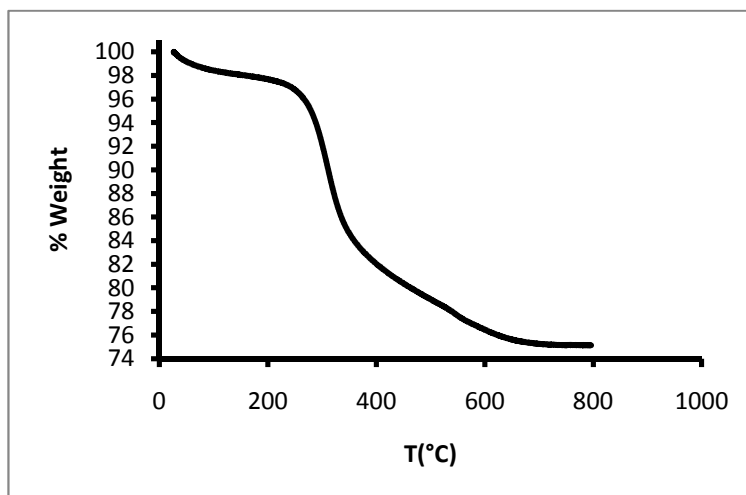


Figure 3: TGA analysis of GA-MCM41 (2 nm)

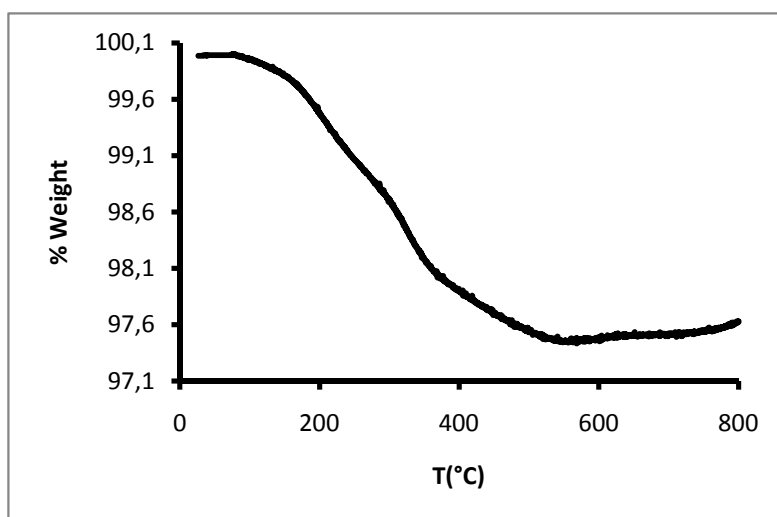


Figure 4: TGA analysis of GA-Si2000 (200 nm)

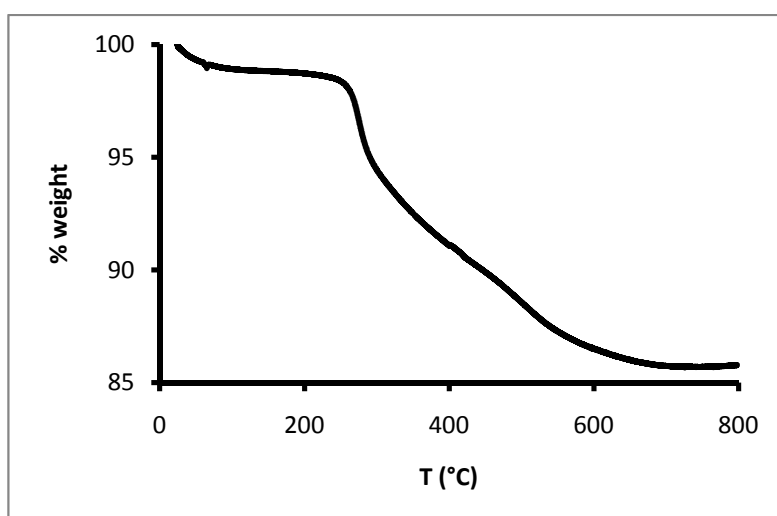


Figure 5: TGA analysis of passivated GA-SBA15 (5.8 nm) after grafting

TGA experiments were confirmed by elemental analyses.

GA-SBA15: found %N=3.23, C=20.13, H=4.28 (the experimental ratio between C/N is 7.29, close to the theoretical ratio of 7).

GA-MCM41: found % N=2.71, C=17.29, H=3.77 (the experimental ratio between C/N is 7.46, close to the theoretical ratio of 7).

GA-Si2000: found % N=0.09, C=0.75, H=0.38 (owing the experimental errors the values are not significant, but they are in agreement with TGA data).

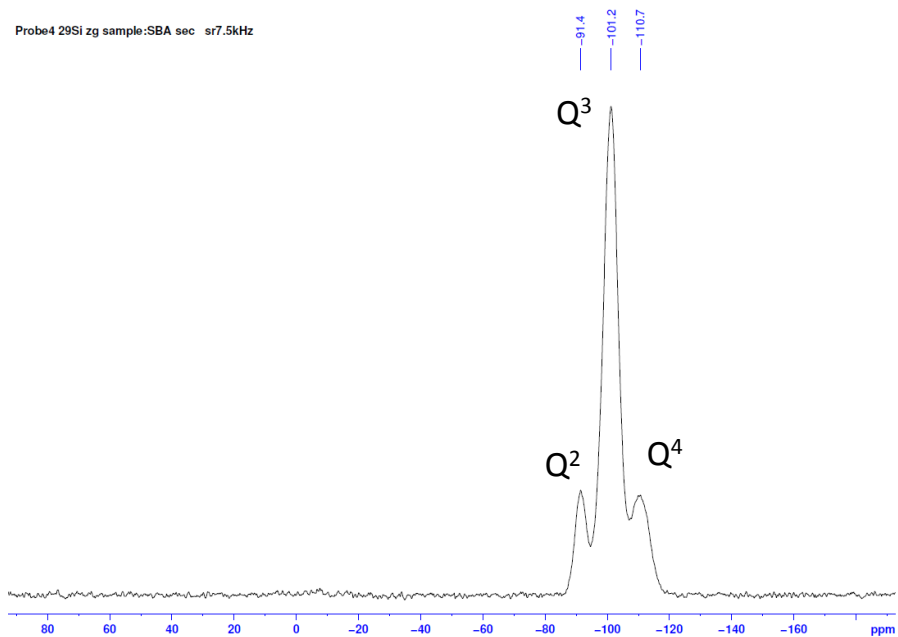


Figure 6: ²⁹Si CP-MAS solid state NMR of SBA15 (7.9 nm)

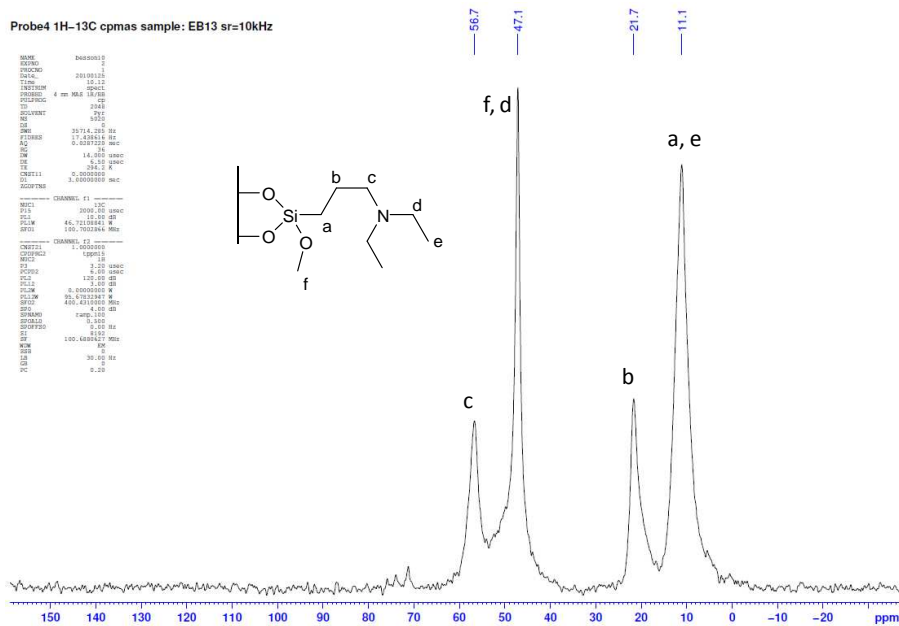


Figure 7: ^{13}C CP-MAS solid state NMR of **GA-SBA15** (5.8 nm)

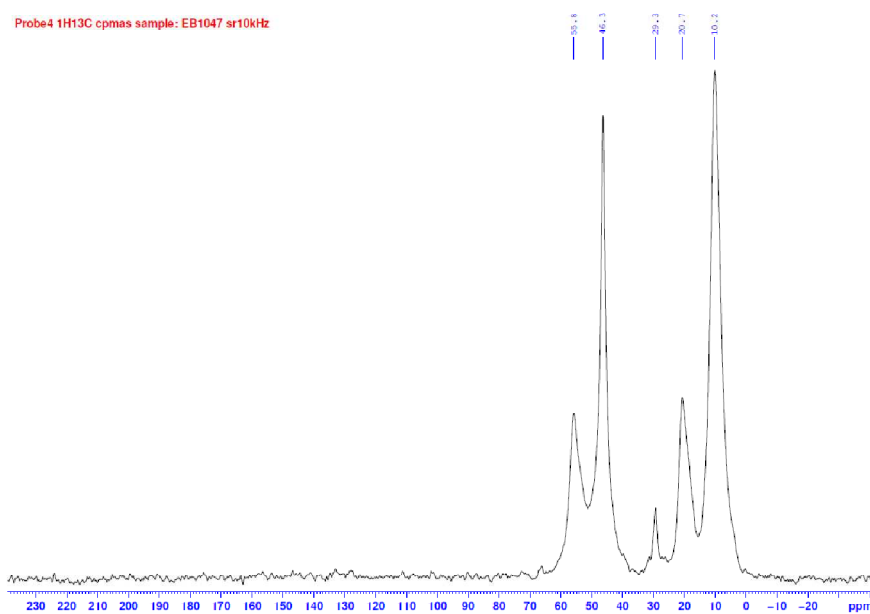


Figure 8: ^{13}C CP-MAS solid state NMR of **GA-MCM41** (2.0 nm)

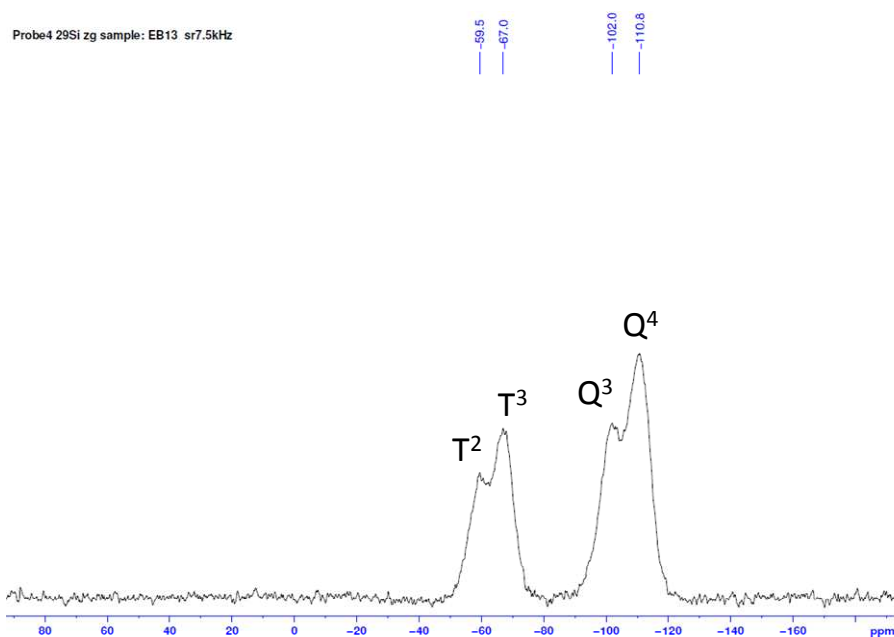


Figure 9: ^{29}Si CP-MAS solid state NMR of **GA-SBA15** (5.8 nm)

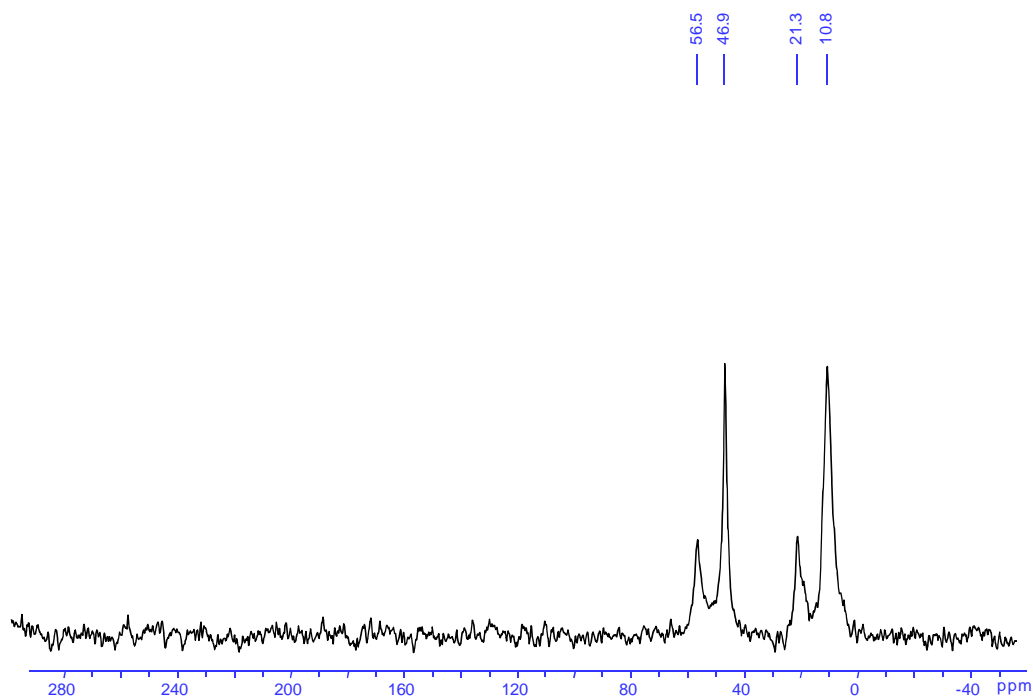


Figure 10: ^{13}C CP-MAS solid state NMR of recycled GA-SBA15 (5.8 nm)

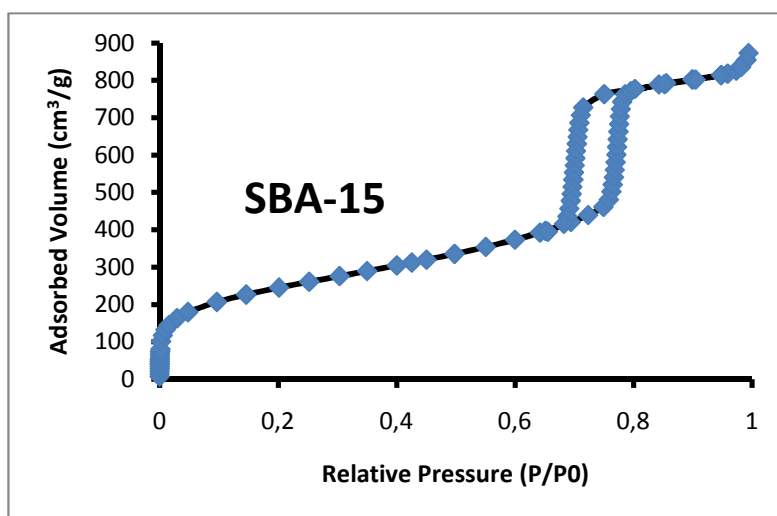


Figure 11: Nitrogen adsorption-desorption isotherm of SBA15. Isotherm of type IV, characteristic of mesoporous materials with a narrow pore size distribution S_{BET} : 874 ± 3 m^2/g , V_p : $1.1 \text{ cm}^3/\text{g}$ and $D_{p_{\text{BJHdes.}}}$: 7.9 nm .

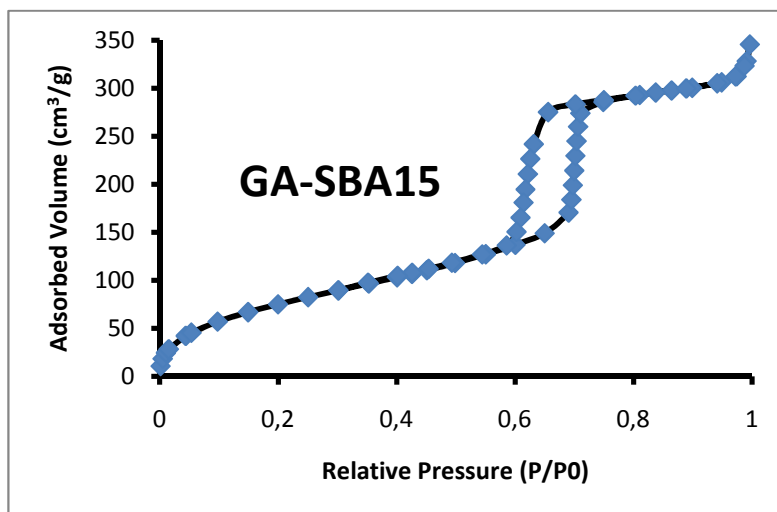


Figure 12: Nitrogen adsorption-desorption isotherm of GA-SBA15. Isotherm of type IV, characteristic of mesoporous materials with a narrow pore size distribution S_{BET} : 296 ± 3 m^2/g , V_p : $0.51 \text{ cm}^3/\text{g}$ and D_{pBJHdes} : 5.8 nm.

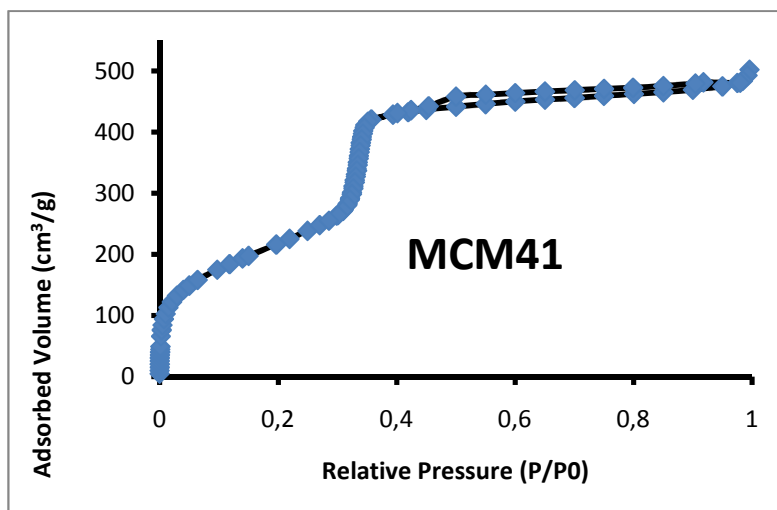


Figure 13: Nitrogen adsorption-desorption isotherm of GA-MCM41. Isotherm of type IV, characteristic of mesoporous materials with a narrow pore size distribution S_{BET} : 720 ± 3 m^2/g , V_p : $0.78 \text{ cm}^3/\text{g}$ and D_{pBJHdes} : 2.4 nm.

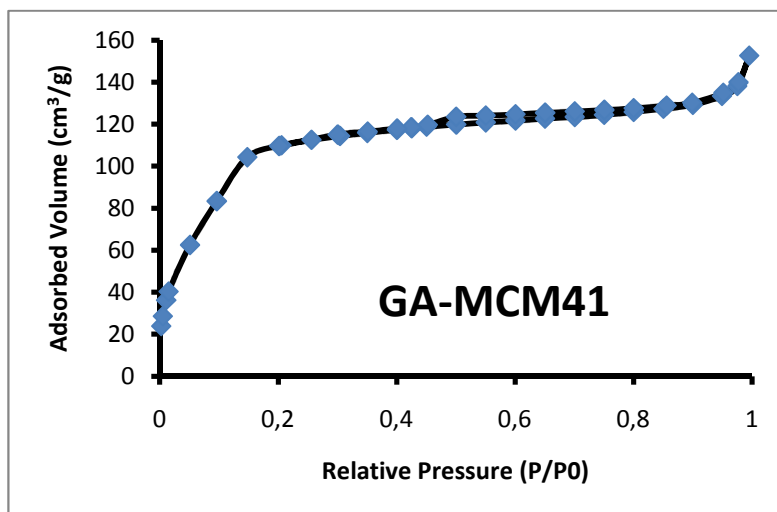


Figure 14: Nitrogen adsorption-desorption isotherm of GA-MCM41. Isotherm of type IV, characteristic of mesoporous materials with a narrow pore size distribution S_{BET} : $200 \pm 3 \text{ m}^2/\text{g}$ and $D_{p_{est}}$: 2 nm.

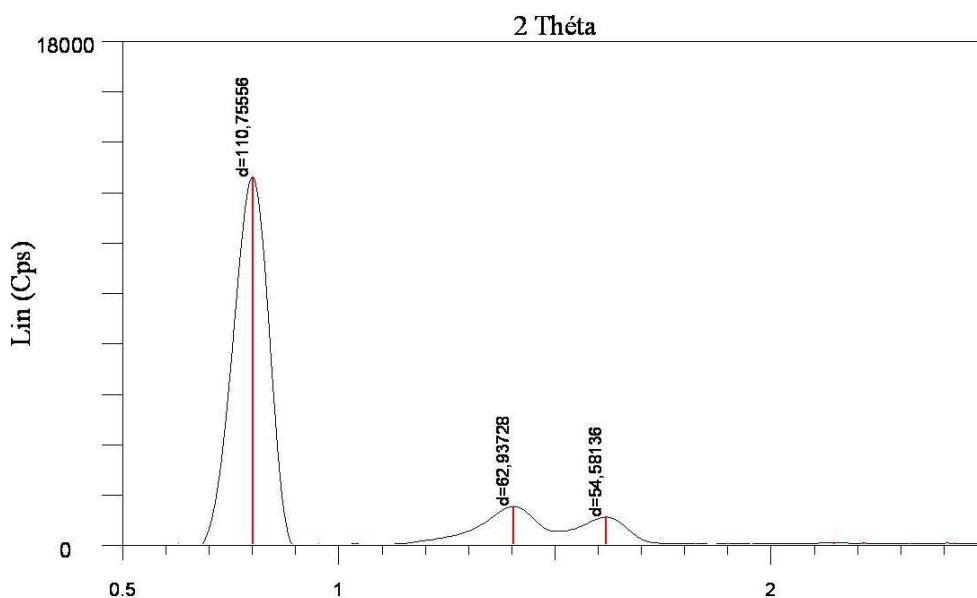


Figure 15: Powder Small Angle X-Ray Diffraction pattern of SBA15

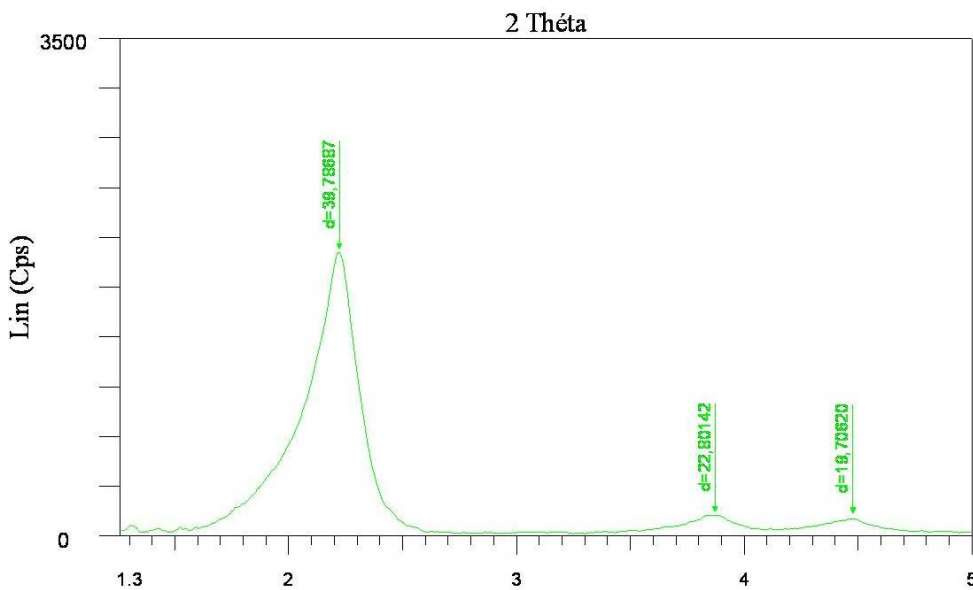


Figure 16: Powder Small Angle X-Ray Diffraction pattern of MCM41

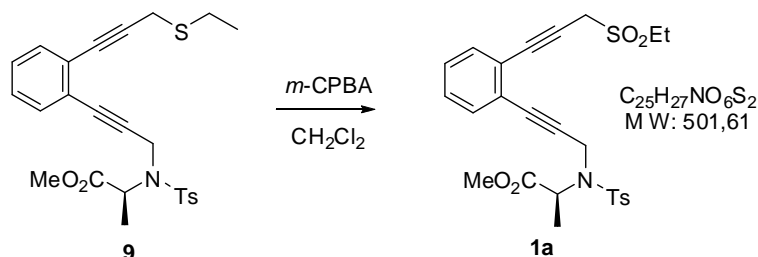
2. Synthesis and rearrangement of substrates 1a, 5a, and 5c

General information

All reactions were carried out in dry glassware using magnetic stirring and a positive pressure of argon. Solvents are commercially available, most of them were used as purchased (analytic grade), without further purification. THF was distilled over sodium benzophenone ketyl prior to use. Dry state adsorption conditions and purification were performed on silica gel 60 Å (70-230 mesh). Analytical thin layer chromatography was performed on pre-coated silica gel plates. Visualization was accomplished by UV (254 nm) and with phosphomolybdic acid in ethanol. ¹H NMR spectra were recorded on 400 MHz-spectrometers. ¹³C spectra were recorded at 100 MHz. Chemical shifts (δ) are reported in ppm. Signals due residual protonated solvent (¹H NMR) or to the solvent (¹³C NMR) served as the internal standard: CDCl₃ (7.26 ppm and 77.16 ppm). Multiplicity is indicated by one or more of the following: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), br (broad). The lists of coupling constants (*J*) correspond to the order of multiplicity assignment and are reported in Hertz (Hz). APT was used for ¹³C spectra assignment. All melting points were uncorrected and were recorded in open capillary tubes using a Buchi melting point apparatus.

High resolution mass spectra were obtained on QStar Elite (Applied Biosystems SCIEX).

Crude mixture and purified compounds were analysed by chiral HPLC with double detection, with UV and circular dichroism detectors. The solvents for chiral chromatography (*n*-hexane, 2-PrOH, ethanol) were HPLC grade, they were degassed and filtered on a 0.45 μm membrane before use. The columns used are Chiralpak IC (250*4.6 mm, cellulose tris(3,5-dichlorophenylcarbamate)), Chiralpak AS-3 (250*4.6 mm, amylose tris[(*S*)alpha-phenethyl]carbamate), and Chiralpak IA (250*4.6 mm, amylose tris-(3,5-dimethylphenylcarbamate)). Enantiomeric excesses were determined by integration of the peaks on the chromatograms obtained by UV detection at 254, 240, 230 or 220 nm, and confirmed by circular dichroism detection at 254 nm. The sign given by the on-line circular dichroism detector for one enantiomer is the sign in the solvent used for the chromatographic separation.



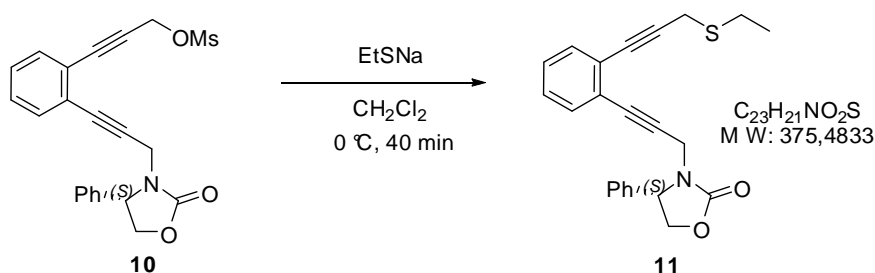
m-Chloroperbenzoic acid (75%) (346 mg, 1.5 mmol) was dissolved in dichloromethane (15 mL), and the solution was cooled to 0 °C. To this mixture a solution of **9** (323 mg, 0.688 mmol) in dichloromethane (5 mL) was added *via* syringe, and the reaction mixture was stirred for 40 min at 0 °C. The crude product was purified by flash chromatography on silica gel (pentane/CH₂Cl₂, 50/50 to 15/85) then the product was washed with a saturated solution of NaHCO₃, dried over MgSO₄ and concentrated to afford **1a** as a yellow oil (334 mg, 97%). ee=88% (Chiralpak AS-3, hexane/EtOH 50/50, R_t(S) = 13.22 min, R_t(R) = 10.54 min). [α]_D²⁰ = -11.1 (c=0.9, CHCl₃). HRMS (ESI): *m/z*: calcd for [M+NH₄]⁺ C₂₅H₃₁N₂O₆S₂: 519.1618, found: 519.1619.

¹H NMR (400 MHz, CDCl₃) δ: 7.79 (2H, d, *J*=8.3, CH_{ar}), 7.47-7.43 (1H, m, CH_{ar}), 7.30-7.24 (5H, m, CH_{ar}), 4.67 (1H, q, *J*=7.3, CHCH₃), 4.52 (1H, d, *J*=18.8, CH₂N, A part of an AB pattern), 4.37 (1H, d, *J*=18.8, CH₂N, B part of an AB pattern), 4.15 (2H, AB pattern, *J*=17.1, Δ*v*=10.6, CH₂SO₂Et), 3.58 (3H, s, CH₃O), 3.32 (2H, q, *J*=7.5, SO₂CH₂CH₃), 2.38 (3H, s, CH₃C), 1.53 (3H, d, *J*=7.3, CH₃CH), 1.47 (3H, t, *J*=7.5, SO₂CH₂CH₃).

¹³C NMR (100 MHz, CDCl₃) δ: 171.8 (CO₂Me), 143.8 (C_{ar}), 137.1 (C_{ar}), 132.4 (CH_{ar}), 132.1 (CH_{ar}), 129.6 (2xCH_{ar}), 128.9 (CH_{ar}), 128.4 (CH_{ar}), 127.6 (2xCH_{ar}), 125.7 (C_{ar}), 124.4 (C_{ar}), 89.4 (C_{C≡C}), 85.9 (C_{C≡C}), 82.6 (C_{C≡C}), 80.8 (C_{C≡C}), 55.0 (CHN), 52.4 (CH₃O), 46.2 (CH₂SO₂), 45.5 (CH₂SO₂), 35.1 (CH₂N), 21.6 (CH₃), 16.5 (CH₃), 6.8 (CH₃).

1.7. Synthesis of 5a

(S)-3-{3-[2-(3-Ethylsulfanyl-prop-1-ynyl)-phenyl]-prop-2-ynyl}-4-phenyl-oxazolidin-2-one (**11**)



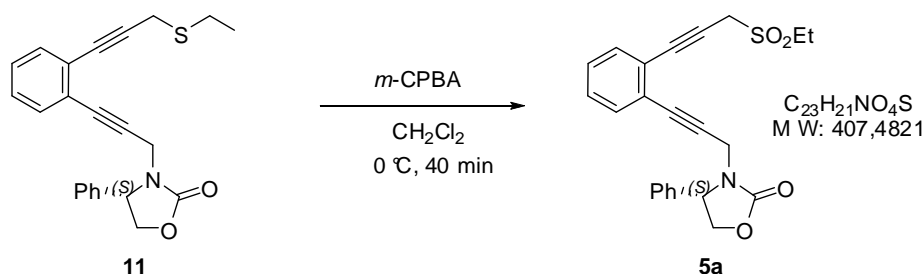
Sulfide **11** was prepared according to the procedure already described for the synthesis of **9**, using ethanethiol (130 μL, 1.72 mmol) in THF (10 mL), 210 μL of a 16 M solution of NaOH and **10**⁶ (600 mg, 1.46 mmol). After work-up, purification by liquid chromatography on silica gel (pentane/CH₂Cl₂, 100/0 to 40/60) led to **11** (407 mg, 74%) as a pale yellow oil. [α]_D²⁰ = +227 (c=0.8, CH₂Cl₂). HRMS (ESI): *m/z*: calcd for [M+H]⁺ C₂₃H₂₂NO₂S: 376.1366, found: 376.1365.

¹H NMR (400 MHz, CDCl₃) δ: 7.45-7.36 (7H, m, CH_{ar}), 7.29-7.22 (2H, m, CH_{ar}), 5.13 (1H, pseudo t, X part of an ABX pattern, *J*=8.3), 4.71 (1H, t, *J*=8.8), 4.67 (1H, superimposed d, *J*=17.8, CH₂N, A part of an AB pattern), 4.18 (1H, pseudo t, *J*=8.3), 3.67 (1H, d, *J*=17.6,

CH₂N, B part of an AB pattern), 3.52 (2H, s, CH₂SEt), 2.73 (2H, q, *J*=7.5, CH₃CH₂S), 1.28 (3H, t, *J*=7.5, CH₃CH₂SO₂).

¹³C NMR (100 MHz, CDCl₃) δ: 157.9 (CO), 137.1 (C_{ar}), 132.4 (CH_{ar}), 132.3 (CH_{ar}), 129.5 (2xCH_{ar}), 129.3 (CH_{ar}), 128.4 (CH_{ar}), 128.0 (CH_{ar}), 127.4 (2xCH_{ar}), 125.9 (C_{ar}), 124.8 (C_{ar}), 89.8 (C≡C), 85.8 (C≡C), 83.9 (C≡C), 81.5 (C≡C), 70.1 (CH₂oxa), 59.1 (CH_{oxa}), 33.1 (CH₂N), 25.7 (CH₂S), 20.0 (CH₂S), 14.4 (CH₃).

(S)-3-{3-[2-(3-Ethanesulfonyl-prop-1-ynyl)-phenyl]-prop-2-ynyl}-4-phenyl-oxazolidin-2-one (5a)



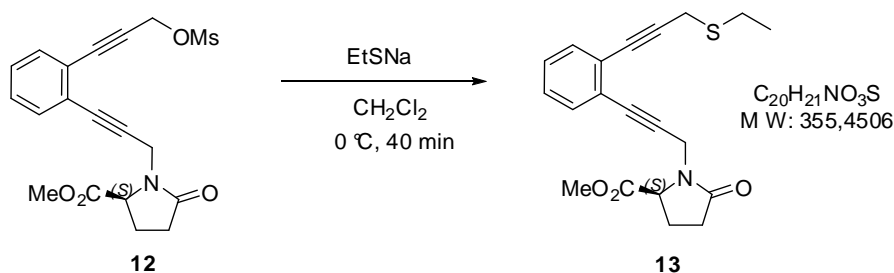
The procedure already described for the synthesis of **1a** was followed, starting from **11** (375 mg, 1 mmol) and 75% *m*-CPBA (540 mg, 2.2 mmol) in dichloromethane (10 mL). Substrate **5a** was isolated as a pale yellow oil (355 mg, 87%) after purification by flash chromatography on silica gel (pentane/CH₂Cl₂, 50/50 to 0/100). ee > 99% (Chiralpak IA, hexane/EtOH 50/50, R_t(S) = 9.83 min, R_t(R) = 7.61 min). [α]_D²⁰ = +143.8 (c=0.675, CHCl₃). HRMS (ESI): *m/z*: calcd for [M+H]⁺ C₂₃H₂₂NO₄S: 408.1264, found: 408.1266.

¹H NMR (400 MHz, CDCl₃) δ: 7.47-7.36 (7H, m, CH_{ar}), 7.33-7.27 (2H, m, CH_{ar}), 5.06 (1H, t, *J*=8.5), 4.71 (1H, t, *J*=8.8), 4.62 (1H, d, *J*=17.8, CH₂N), 4.14 (1H, t, *J*=8.5), 4.11 (2H, superimposed AB pattern, *J*=17.1, Δ*v*=11, CH₂SO₂Et), 3.69 (1H, d, *J*=17.8, CH₂N), 3.25 (2H, q, *J*=7.5, CH₃CH₂SO₂), 1.43 (3H, t, *J*=7.5, CH₃CH₂SO₂).

¹³C NMR (100 MHz, CDCl₃) δ: 158.1 (CO), 136.9 (C_{ar}), 132.5 (CH_{ar}), 132.3 (CH_{ar}), 129.5 (2xCH_{ar}), 129.4 (CH_{ar}), 128.9 (CH_{ar}), 128.5 (CH_{ar}), 127.4 (2xCH_{ar}), 125.3 (C_{ar}), 124.5 (C_{ar}), 86.5 (C≡C), 85.9 (C≡C), 83.1 (C≡C), 80.6 (C≡C), 70.2 (CH₂oxa), 59.4 (CH_{oxa}), 46.1 (CH₂S), 45.2 (CH₂S), 33.1 (CH₂N), 6.8 (CH₃).

1.8. Synthesis of 5c

(S)-1-{3-[2-(3-Ethylsulfanyl-prop-1-ynyl)-phenyl]-prop-2-ynyl}-5-oxo-pyrrolidine-2-carboxylic acid methyl ester (13)



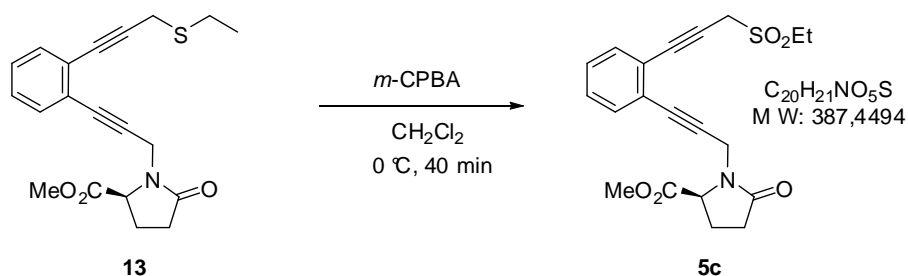
Sulfide **13** was prepared according to the procedure already described for the synthesis of **9**, using ethanethiol (30 μL, 0.39 mmol) in THF (2 mL), 57 μL of a 7 M solution of NaOH and

12⁶ (150 mg, 0.385 mmol). After work-up, purification by liquid chromatography on silica gel (pentane /CH₂Cl₂, 100/0 to 40/60) led to **13** (91 mg, 67%) as a pale yellow oil. [α]_D²⁰ = -11.7 (c=0.366, CH₂Cl₂). HRMS (ESI): *m/z*: calcd for [M+H]⁺ C₂₀H₂₂NO₃S: 356.1315, found: 356.1314.

¹H NMR (400 MHz, CDCl₃) δ : 7.43-7.37 (2H, m, CH_{ar}), 7.27-7.21 (2H, m, CH_{ar}), 4.89 (1H, d, *J*=17.8, CH₂N), 4.62-4.59 (1H, m, CH), 4.04 (1H, br d, *J*=17.8, CH₂N), 3.77 (3H, s, CH₃O), 3.55 (2H, s, CH₂SEt), 2.78 (2H, q, *J*=7.5, SCH₂CH₃), 2.57-2.38 (3H, m), 2.15-2.09 (1H, m), 1.33 (3H, t, *J*=7.5, SCH₂CH₃).

¹³C NMR (100 MHz, CDCl₃) δ : 174.5 (CO), 172.4 (CO), 132.3 (CH_{ar}), 132.3 (CH_{ar}), 128.3 (CH_{ar}), 128.0 (CH_{ar}), 125.8 (C_{ar}), 125.0 (C_{ar}), 89.8 (C_{C≡C}), 86.3 (C_{C≡C}), 83.6 (C_{C≡C}), 81.5 (C_{C≡C}), 58.6 (CH), 52.6 (CH₃O), 32.4 (CH₂N), 29.6 (CH₂), 25.7 (CH₂SEt), 23.0 (CH₂), 19.9 (CH₂), 14.4 (CH₃).

(S)-1-{3-[2-(3-Ethanesulfonyl-prop-1-ynyl)-phenyl]-prop-2-ynyl}-5-oxo-pyrrolidine-2-carboxylic acid methyl ester (**5c**)



The procedure already described for the synthesis of **1a** was followed, when reacting **13** (80 mg, 0.225 mmol) with 75% *m*-CPBA (113 mg, 0.49 mmol) in dichloromethane (3 mL). Substrate **5c** was isolated as a yellow solid (68 mg, 78%) after purification by flash chromatography on silica gel (CH₂Cl₂/Et₂O, 100/0 to 80/20). ee=97% (Chiralpak IA, hexane/EtOH 50/50, 1mL/min, R_t(S) = 7.63 min, R_t(R) = 8.99 min). [α]_D³⁰ = -2.1 (c=0.6, CH₂Cl₂). HRMS (ESI): *m/z*: calcd for [M+H]⁺ C₂₀H₂₂NO₅S: 388.1213, found: 388.1210.

¹H NMR (400 MHz, CDCl₃) δ : 7.46-7.40 (2H, m, CH_{ar}), 7.32-7.27 (2H, m, CH_{ar}), 4.82 (1H, d, *J*=17.8, CH₂N), 4.54-4.52 (1H, m, CH), 4.15 (2H, s, CH₂SO₂Et), 4.07 (1H, br d, *J*=17.8, CH₂N), 3.76 (3H, s, CH₃O), 3.30 (2H, q, *J*=7.5, SO₂CH₂CH₃), 2.56-2.36 (3H, m), 2.15-2.03 (1H, m), 1.47 (3H, t, *J*=7.5, SO₂CH₂CH₃).

¹³C NMR (100 MHz, CDCl₃) δ : 174.7 (CO), 172.4 (CO), 132.5 (CH_{ar}), 132.4 (CH_{ar}), 129.0 (CH_{ar}), 128.5 (CH_{ar}), 125.5 (C_{ar}), 124.3 (C_{ar}), 87.1 (C_{C≡C}), 86.0 (C_{C≡C}), 82.7 (C_{C≡C}), 80.6 (C_{C≡C}), 58.8 (CH), 52.7 (CH₃O), 46.0 (CH₂SO₂), 45.2 (CH₂SO₂), 32.2 (CH₂N), 29.5 (CH₂), 23.0 (CH₂), 6.9 (CH₃).

1.9. General procedure for the stoichiometric cyclisation

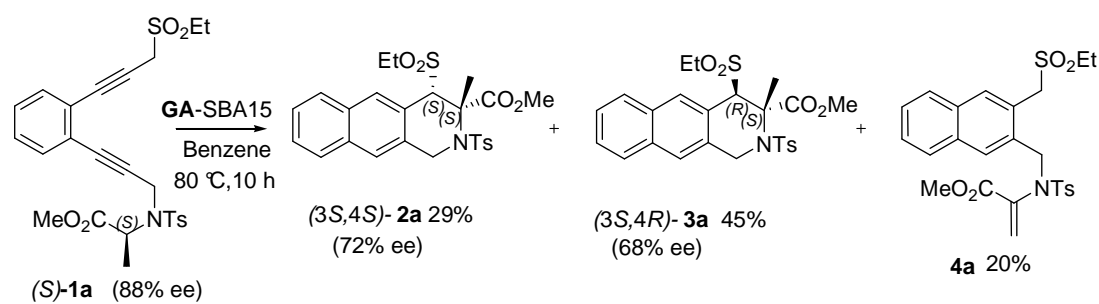
In a typical experiment, 400 mg (0.566 mmol of amine) of mesoporous silica grafted with the tertiary amine group (**GA-SBA15**) were added to a solution of enediyne (**S-1b**) (130 mg, 0.259 mmol) in benzene (20 mL). The mixture was stirred at 80 °C for 10h. The reaction was monitored by CCM (CH₂Cl₂/Et₂O, 98/2). After filtration and recovery of silica (rinsed with ether), purification by liquid chromatography on silica gel (pentane/CH₂Cl₂, from 70/30 to

0/100) led to the isolation of (3*S*,4*R*)-**3a** (58 mg, 45%, ee=68%) as a yellowish oil, **4a** (26 mg, 20%), and (3*S*,4*S*)-**2a** (38 mg, 29%, ee=72%) as yellowish oil.

1.10. General procedure for the catalytic cyclisation

In a typical experiment, 26 mg (0.037 mmol, 20 mol %) of mesoporous silica grafted with the tertiary amine group (GA-SBA15) were added to a solution of enediyne (*S*)-**5c** (100 mg, 0.184 mmol) in benzene (4 mL). The mixture stirred at 80 °C for 13h. The reaction was monitored by CCM (CH₂Cl₂/Et₂O, 95/5). The crude was then purified by liquid chromatography on silica gel (pentane/ CH₂Cl₂, from 70/30 to 0/100). This led to the isolation of (*R,S*)-**6c** (45 mg, 60%, ee=93.5%) as colourless oil and (*S,S*)-**7c** (22 mg, 30%, ee=62%) as a colourless oil.

1.11. Cycloaromatisation of (*S*)-**1a**



(3*S*,4*S*)-4-Ethanesulfonyl-3-methyl-2-(toluene-4-sulfonyl)-1,2,3,4-tetrahydrobenzo[*g*]isoquinoline-3-carboxylic acid methyl ester (**2a**)

ee=72% (Chiralpak IC, hexane/EtOH 50/50, 1mL/min $R_t(\text{major}) = 16.93$ min, $R_t(\text{minor}) = 22.23$ min). $[\alpha]_D^{30} = +9.6$ (c=0.3, CH₂Cl₂). HRMS (ESI): m/z : calcd for $[M+NH_4]^+$ C₂₅H₃₁N₂O₆S₂: 519.1618, found: 519.1614.

¹H NMR (400 MHz, CDCl₃) δ: 8.00 (2H, d, $J=8.3$, CH_{ar}), 7.86-7.82 (2H, m, CH_{ar}), 7.82 (1H, superimposed s, CH_{ar}), 7.69 (1H, s, CH_{ar}), 7.59-7.53 (2H, m, CH_{ar}), 7.32 (2H, d, $J=8.0$, CH_{ar}), 4.96 (1H, br d, $J=14.3$, CH₂N, A part of an AB pattern), 4.65 (1H, d, $J=15.1$, CH₂N, B part of an AB pattern), 4.59 (1H, s, CHSO₂Et), 3.93 (3H, s, CH₃O), 2.96 (1H, dq, $J=13.3$ and 7.5, SO₂CH₂CH₃, A part of ABX₃ pattern), 2.85 (1H, dq, $J=13.3$ and 7.5, SO₂CH₂CH₃, B part of an ABX₃ pattern), 2.42 (3H, s), 1.79 (3H, s), 1.26 (3H, t, $J=7.5$, SO₂CH₂CH₃).

¹³C NMR (100 MHz, CDCl₃) δ: 171.2 (CO₂Me), 143.7 (C_{ar}), 137.8 (C_{ar}), 134.0 (C_{ar}), 132.6 (C_{ar}), 131.8 (C_{ar}), 130.6 (CH_{ar}), 129.8 (2xCH_{ar}), 128.0 (CH_{ar}), 127.99 (2xCH_{ar}), 127.95 (CH_{ar}), 127.8 (CH_{ar}), 127.2 (CH_{ar}), 126.5 (CH_{ar}), 124.8 (C_{ar}), 74.4 (CHSO₂), 64.7 (C), 53.4 (CH₃O), 48.0 (CH₂), 46.7 (CH₂), 29.3 (CH₃), 21.7 (CH₃), 5.7 (CH₃).

(3*S*,4*R*)-4-Ethanesulfonyl-3-methyl-2-(toluene-4-sulfonyl)-1,2,3,4-tetrahydrobenzo[*g*]isoquinoline-3-carboxylic acid methyl ester (**3a**)

ee=68% (Chiralpak IC, hexane/EtOH 50/50, 1mL/min, $R_t(\text{major}) = 13.44$ min, $R_t(\text{minor}) = 18.32$ min). $[\alpha]_D^{30} = +63.7$ (c=0.85, CHCl_3). **HRMS** (ESI): m/z : calcd for $[\text{M}+\text{NH}_4]^+$ $\text{C}_{25}\text{H}_{31}\text{N}_2\text{O}_6\text{S}_2$: 519.1618, found: 519.1609.

^1H NMR (400 MHz, CDCl_3) δ : 7.91 (1H, s, CH_{ar}), 7.86 (2H, d, $J=8.5$, CH_{ar}), 7.84 (1H, d, $J=7.8$, CH_{ar}), 7.78 (1H, d, $J=7.8$, CH_{ar}), 7.67 (1H, s, CH_{ar}), 7.54-7.46 (2H, m, CH_{ar}), 7.35 (2H, d, $J=8.0$), 5.29 (1H, d, $J=15.9$, CH_2N , A part of an AB pattern), 4.98 (1H, d, $J=15.9$, CH_2N , B part of an AB pattern), 4.72 (1H, s, CHSO_2Et), 3.24 (3H, s, CH_3O), 3.09 (1H, dq, $J=13.3$ and 7.5, $\text{SO}_2\text{CH}_2\text{CH}_3$, A part of ABX_3 pattern), 2.69 (1H, dq, $J=13.3$ and 7.5, $\text{SO}_2\text{CH}_2\text{CH}_3$, B part of an ABX_3 pattern), 2.44 (3H, s), 2.32 (3H, s), 1.21 (3H, t, $J=7.5$, $\text{SO}_2\text{CH}_2\text{CH}_3$).

^{13}C NMR (100 MHz, CDCl_3) δ : 171.5 (CO_2Me), 143.8 (C_{ar}), 138.0 (C_{ar}), 133.6 (C_{ar}), 132.1 (C_{ar}), 131.0 (CH_{ar}), 130.2 (C_{ar}), 129.6 (2x CH_{ar}), 128.3 (CH_{ar}), 127.8 (2x CH_{ar}), 127.6 (CH_{ar}), 127.5 (CH_{ar}), 126.7 (CH_{ar}), 125.4 (CH_{ar}), 125.1 (C_{ar}), 72.1 (CHSO_2), 66.5 (C), 53.1 (CH_3O), 49.3 (CH_2), 46.1 (CH_2), 23.2 (CH_3), 21.7 (CH_3), 5.1 (CH_3).

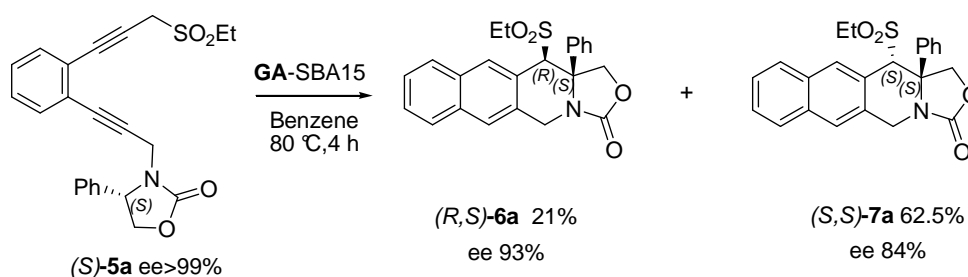
2-[(3-Ethanesulfonylmethyl-naphthalen-2-ylmethyl)-(toluene-4-sulfonyl)-amino]-acrylic acid methyl ester (4a)

HRMS (ESI): m/z : calcd for $[\text{M}+\text{NH}_4]^+$ $\text{C}_{25}\text{H}_{31}\text{N}_2\text{O}_6\text{S}_2$: 519.1618, found: 519.1615.

^1H NMR (400 MHz, CDCl_3) δ : 7.90 (1H, br s, CH_{ar}), 7.83-7.81 (1H, m, CH_{ar}), 7.75 (2H, d, $J=8.5$, CH_{ar}), 7.69-7.67 (1H, m, CH_{ar}), 7.51-7.44 (2H, m, CH_{ar}), 7.41 (1H, s, CH_{ar}), 7.35 (2H, d, $J=8.0$, CH_{ar}), 6.18 (1H, s, =CH), 5.45 (1H, s, =CH), 4.89 (2H, s, CH_2), 4.85 (2H, s, CH_2), 3.57 (3H, s, CH_3O), 3.29 (2H, q, $J=7.5$, $\text{SO}_2\text{CH}_2\text{CH}_3$), 2.47 (3H, s), 1.56 (3H, t, $J=7.5$, $\text{SO}_2\text{CH}_2\text{CH}_3$).

^{13}C NMR (100 MHz, CDCl_3) δ : 164.3 (CO_2Me), 144.4 (C_{ar}), 135.9 (C_{ar}), 134.4 (C_{ar}), 133.2 (CH_{ar}), 133.1 (C=), 133.0 (C_{ar}), 131.7 (CH_{ar}), 131.4 (C_{ar}), 129.8 (2x CH_{ar}), 128.2 (2x CH_{ar}), 127.9 (CH_{ar}), 127.6 (CH_{ar}), 127.4 (CH_2), 127.1 (CH_{ar}), 127.0 (CH_{ar}), 124.5 (C_{ar}), 54.4 (CH_2), 52.5 (CH_3O), 52.3 (CH_2), 48.2 (CH_2), 21.8 (CH_3), 6.9 (CH_3).

1.12. Cycloaromatisation of (S)-5a



(11R,11aS)-11-Ethanesulfonyl-11a-phenyl-1,4,11,11a-tetrahydro-2-oxa-3a-azacyclopenta[*b*]anthracen-3-one (6a)

ee=93% (Chiralpak IC, hexane/EtOH 50/50, 1mL/min $R_t(\text{minor}) = 12.77$ min, $R_t(\text{major}) = 14.31$ min). $[\alpha]_D^{30} = +106.6$ (c=0.9, CHCl_3). **HRMS** (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{23}\text{H}_{22}\text{NO}_4\text{S}$: 408.1264, found: 408.1265.

^1H NMR (400 MHz, CDCl_3) δ : 8.26 (1H, s, CH_{ar}), 7.88 (1H, d, $J=7.5$, CH_{ar}), 7.82 (1H, d, $J=7.5$, CH_{ar}), 7.72 (1H, s, CH_{ar}), 7.59-7.49 (4H, m, CH_{ar}), 7.32-7.29 (3H, m, CH_{ar}), 5.12 (1H,

d, $J=9.5$, CH₂O, A part of an AB pattern), 5.09 (1H, s, CHSO₂Et), 4.99 (1H, d, $J=16.5$, CH₂N, A part of an AB pattern), 4.74 (1H, d, $J=9.8$, CH₂O, B part of an AB pattern), 4.65 (1H, br d, $J=16.5$, CH₂N, B part of an AB pattern), 2.99 (1H, dq, $J=13.8$ and 7.5, CH₃CH₂SO₂, A part of an ABX₃ pattern), 2.57 (1H, dq, $J=13.8$ and 7.5, CH₃CH₂SO₂, B part of an ABX₃ pattern), 1.31 (3H, t, $J=7.5$, CH₃CH₂SO₂).

¹³C NMR (100 MHz, CDCl₃) δ: 157.2 (CO), 137.1 (C_{ar}), 132.8 (C_{ar}), 132.4 (C_{ar}), 130.8 (C_{ar}), 129.3 (CH_{ar}), 129.0 (CH_{ar}), 128.8 (2xCH_{ar}), 128.3 (CH_{ar}), 127.9 (CH_{ar}), 127.5 (CH_{ar}), 127.4 (2xCH_{ar}), 126.9 (CH_{ar}), 126.3 (CH_{ar}), 124.9 (C_{ar}), 76.9 (CH₂O), 71.1 (CHSO₂Et), 64.6 (C), 49.8 (CH₂), 43.6 (CH₂), 6.1 (CH₃).

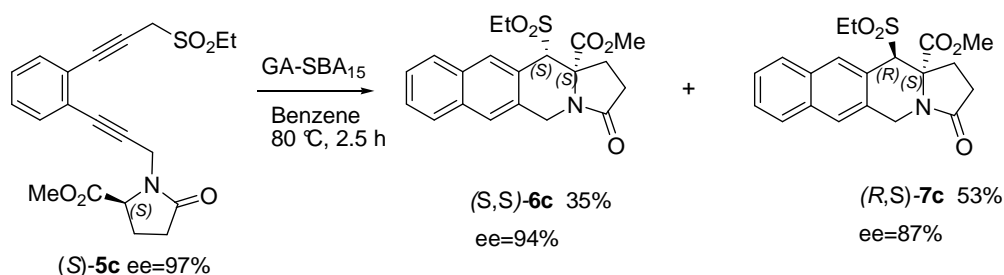
(11S,11aS)-11-Ethanesulfonyl-11a-phenyl-1,4,11,11a-tetrahydro-2-oxa-3a-azacyclopenta[*b*]anthracen-3-one (7a)

ee=84% (Chiralpak IC, hexane/EtOH 50/50, 1mL/min R_t (major) = 32.96 min, R_t (minor)=53.62 min). [α]_D³⁰ = +33 (c=1.5, CHCl₃). HRMS (ESI): *m/z*: calcd for [M+H]⁺ C₂₃H₂₂NO₄S: 408.1264, found: 408.1270.

¹H NMR (400 MHz, CDCl₃) δ: 7.75-7.72 (2H, m, CH_{ar}), 7.70 (2H, br s, CH_{ar}), 7.50-7.42 (2H, m, CH_{ar}), 7.21 (4H, m, CH_{ar}), 7.14-7.09 (1H, m, CH_{ar}), 5.90 (1H, d, $J=9.3$, CH₂O), 5.31 (1H, d, $J=16.8$, CH₂N, A part of an AB pattern), 5.07 (1H, s, CHSO₂Et), 4.86 (1H, d, $J=17.0$, CH₂N, B part of an AB pattern), 4.31 (1H, d, $J=9.0$, CH₂O), 2.98 (1H, dq, $J=13.5$ and 7.5, CH₃CH₂SO₂, A part of an ABX₃ pattern), 2.82 (1H, dq, $J=13.5$ and 7.5, CH₃CH₂SO₂, B part of an ABX₃ pattern), 1.27 (3H, t, $J=7.5$, CH₃CH₂SO₂).

¹³C NMR (100 MHz, CDCl₃) δ: 158.4 (CO), 142.4 (C_{ar}), 133.5 (C_{ar}), 131.8 (C_{ar}), 131.3 (CH_{ar}), 129.5 (2xCH_{ar}), 129.3 (C_{ar}), 128.3 (CH_{ar}), 127.8 (CH_{ar}), 127.7 (CH_{ar}), 127.5 (CH_{ar}), 126.7 (CH_{ar}), 126.6 (CH_{ar}), 124.6 (C_{ar}), 124.0 (2xCH_{ar}), 72.5 (CH₂O), 67.5 (CHSO₂Et), 65.2 (C), 46.5 (CH₂), 44.9 (CH₂), 5.3 (CH₃).

1.13. Cycloaromatisation of (*S*)-5c



(11S,11aS)-11-Ethanesulfonyl-3-oxo-2,3,4,11-tetrahydro-1*H*-3a-azacyclopenta[*b*]anthracene-11a-carboxylic acid methyl ester (6c)

ee=94% (Chiralpak IC, hexane/EtOH 50/50, 1mL/min R_t (major) = 13.43 min, R_t (minor)=18.46 min). [α]_D³⁰ = -172 (c=0.186, CH₂Cl₂). HRMS (ESI): *m/z*: calcd for [M+H]⁺ C₂₀H₂₂NO₅S: 388.1213, found: 388.1211.

¹H NMR (400 MHz, CDCl₃) δ: 8.15 (1H, s, CH_{ar}), 7.85-7.80 (2H, m, CH_{ar}), 7.72 (1H, s, CH_{ar}), 7.56-7.49 (2H, m, CH_{ar}), 5.18 (1H, d, $J=16.3$, CH₂N, A part of an AB pattern), 4.93

(1H, s, CHSO₂Et), 4.38 (1H, d, $J=16.3$, CH₂N, B part of an AB pattern), 3.83 (3H, s, OCH₃), 3.19-3.13 (1H, m), 3.17 (2H, superimposed q, $J=7.5$, SO₂CH₂CH₃), 2.69-2.59 (1H, m), 2.44-2.37 (1H, m), 2.25-2.17 (1H, m), 1.34 (3H, t, $J=7.5$, SO₂CH₂CH₃).

¹³C NMR (100 MHz, CDCl₃) δ : 174.3 (CO), 170.6 (CO), 133.0 (C_{ar}), 132.3 (C_{ar}), 131.4 (C_{ar}), 129.0 (CH_{ar}), 128.1 (CH_{ar}), 127.7 (CH_{ar}), 127.6 (CH_{ar}), 126.8 (CH_{ar}), 126.1 (CH_{ar}), 124.5 (C_{ar}), 74.0 (CHSO₂Et), 67.6 (C), 53.5 (OCH₃), 47.5 (CH₂), 41.3 (CH₂), 33.7 (CH₂), 30.0 (CH₂), 5.9 (CH₃).

(11R,11aS)-11-Ethanesulfonyl-3-oxo-2,3,4,11-tetrahydro-1H-3a-aza-cyclopenta[*b*]anthracene-11a-carboxylic acid methyl ester (7c)

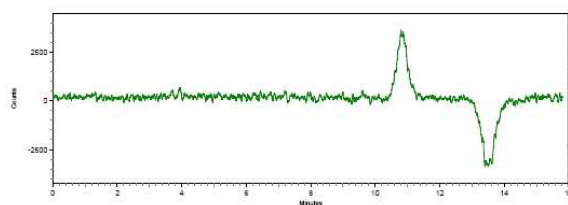
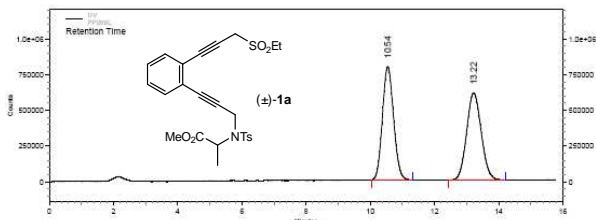
ee=87% (Chiralpak IC, hexane/EtOH 50/50, 1mL/min $R_t(\text{minor})=17.46$ min, $R_t(\text{major})=24.17$ min). $[\alpha]_D^{30} = -11.1$ (c=0.18, CH₂Cl₂). HRMS (ESI): m/z : calcd for [M+H]⁺ C₂₀H₂₂NO₅S: 388.1213, found: 388.1212.

¹H NMR (400 MHz, CDCl₃) δ : 7.87 (1H, s, CH_{ar}), 7.78-7.74 (2H, m, CH_{ar}), 7.69 (1H, br s, CH_{ar}), 7.57-7.49 (2H, m, CH_{ar}), 5.12 (1H, d, $J=17.6$, CH₂N, A part of an AB pattern), 5.02 (1H, s, CHSO₂Et), 4.74 (1H, d, $J=17.6$, CH₂N, B part of an AB pattern), 3.66-3.56 (1H, ddd, $J=14.0, 10.5$ and 6.0), 3.51 (3H, s, OCH₃), 2.84-2.54 (4H, m superimposed to the ABX₃ pattern of SO₂CH₂CH₃), 2.34-2.26 (1H, ddd, $J=14.0, 10.5$ and 6.0), 1.24 (3H, t, $J=7.5$, SO₂CH₂CH₃).

¹³C NMR (100 MHz, CDCl₃) δ : 175.1 (CO), 171.8 (CO), 133.9 (C_{ar}), 132.0 (C_{ar}), 131.4 (CH_{ar}), 129.7 (C_{ar}), 128.0 (CH_{ar}), 127.9 (CH_{ar}), 127.6 (CH_{ar}), 126.9 (CH_{ar}), 126.9 (CH_{ar}), 124.6 (C_{ar}), 67.9 (C), 66.7 (CHSO₂Et), 53.5 (OCH₃), 47.9 (CH₂), 43.5 (CH₂), 29.6 (CH₂), 26.3 (CH₂), 5.2 (CH₃).

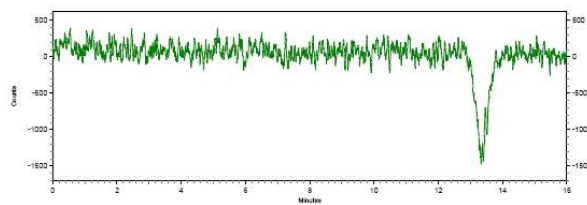
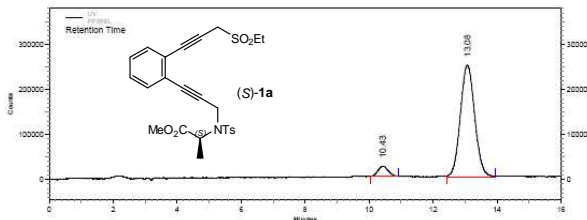
3. Chiral HPLC Spectra

Method description : Chiralpak AS-3, Hexane/ethanol 50/50, 1 ml/min, UV 254 nm et CD254



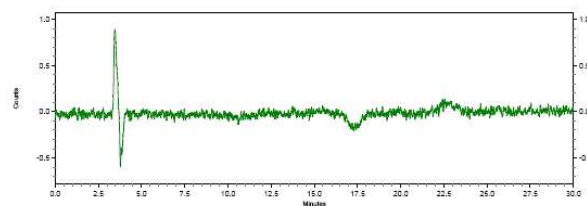
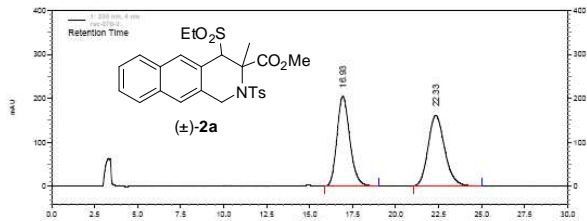
UV Results					
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
10.54	19197391	49.71	2.51	1.00	0.00
13.22	19420628	50.29	3.41	1.35	3.56
Totals		38618019	100.00		

Method description : Chiralpak AS-3, Hexane/ethanol 50/50, 1 ml/min, UV 254 nm et CD254



UV Results					
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
10.43	499974	6.09	2.48	0.00	0.00
13.08	7715680	93.91	3.36	0.00	3.71
Totals		8215654	100.00		

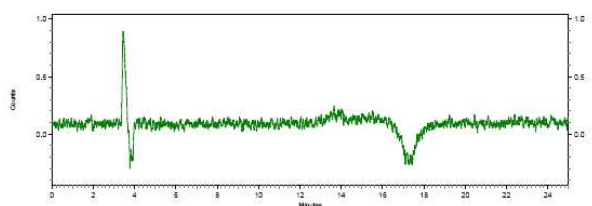
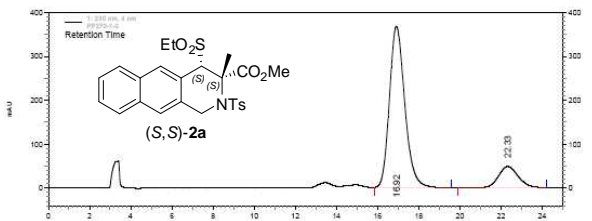
Method description : Chiralpak IC, Hexane/Ethanol 50/50, 1 ml/min, DAD and CD 254nm



1: 230 nm, 4 nm

Results					
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
16.93	42435389	49.44	4.64	0.00	0.00
22.33	43390341	50.56	6.44	0.00	3.43
Totals		85825730	100.00		

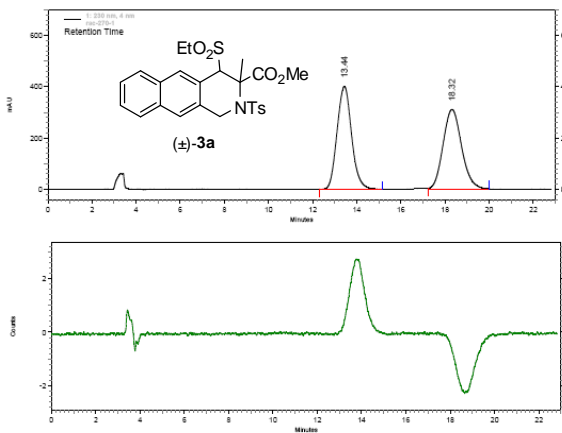
Method description : Chiralpak IC, Hexane/Ethanol 50/50, 1 ml/min, DAD and CD 254nm



1: 230 nm, 4 nm

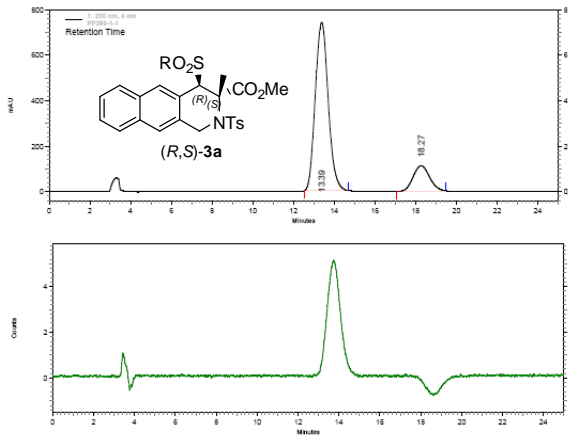
Results					
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
16.92	76175172	83.76	4.64	0.00	0.00
22.33	12650006	14.24	6.44	0.00	3.46
Totals		88825178	100.00		

Method description : Chiralpak IC, Hexane/Ethanol 50/50, 1 ml/min, DAD and CD 254nm



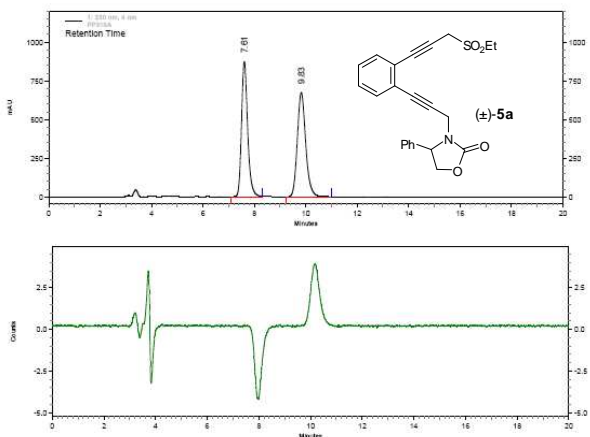
1: 230 nm, 4 nm					
Results					
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
13.44	73231572	50.38	3.48	0.00	0.00
18.32	72129016	49.62	5.11	0.00	3.54
Totals		145360588	100.00		

Method description : Chiralpak IC, Hexane/Ethanol 50/50, 1 ml/min, DAD and CD 254nm



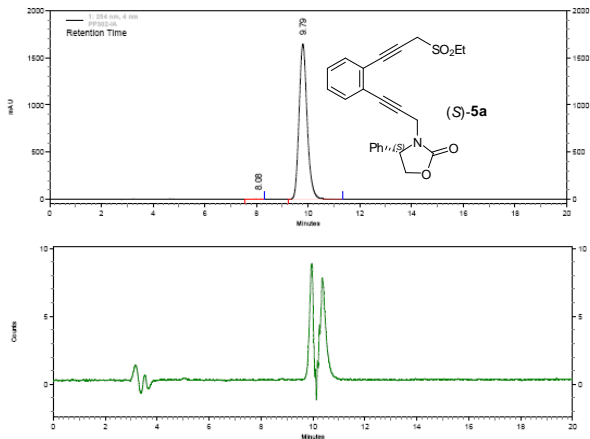
1: 230 nm, 4 nm					
Results					
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
13.39	131052673	83.92	3.46	1.00	0.00
18.27	25113774	16.08	5.09	1.47	3.64
Totals		156166447	100.00		

Method description : Chiralpak IA, Hexane/Ethanol 50/50, 1 ml/min, DAD and CD 254nm

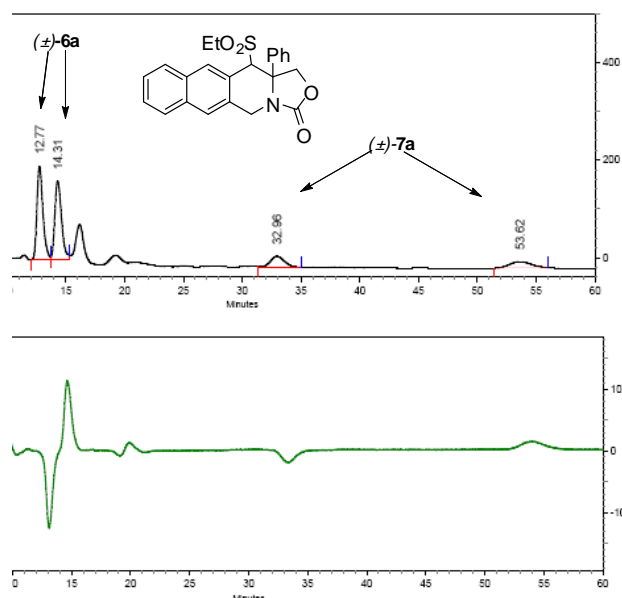


1: 230 nm, 4 nm					
Results					
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
7.61	57976728	48.65	1.54	0.00	0.00
9.83	61182287	51.35	2.28	0.00	4.35
Totals		119159015	100.00		

Method description : Chiralpak IA, Hexane/Ethanol 50/50, 1 ml/min, DAD and CD 254nm

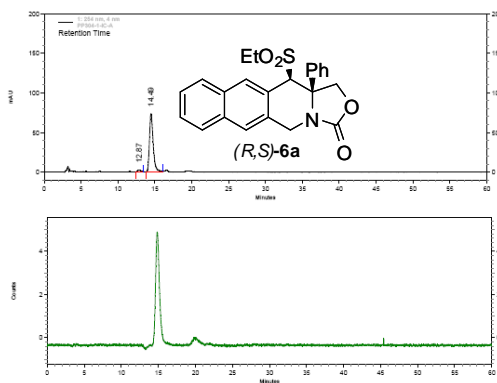


1: 254 nm, 4 nm					
Results					
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
8.08	341994	0.24	1.69	1.00	0.00
9.79	142436364	99.76	2.26	1.34	0.00
Totals		142778358	100.00		



Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
12.77	27408866	40.66	3.26	0.00	0.00
14.31	26047444	38.64	3.77	0.00	1.55
32.96	7426692	11.02	9.99	0.00	11.46
53.62	6522680	9.68	16.87	0.00	7.27
Totals	67405682	100.00			

Method description : Chiralpak IC, Hexane/Ethanol 50/50, 1 ml/min, DAD and CD 254nm

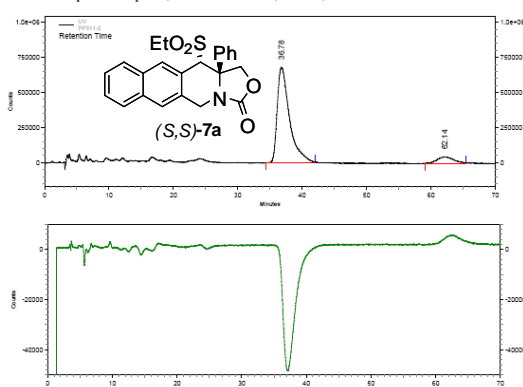


1: 254 nm, 4 nm

Results

Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
12.87	329364	3.07	3.29	1.00	0.00
14.49	10407176	96.93	3.83	1.16	1.82
Totals	10736540	100.00			

Method description : Chiralpak IC, Hexane/ethanol 50/50, 1 ml/min, UV 254 nm et CD254

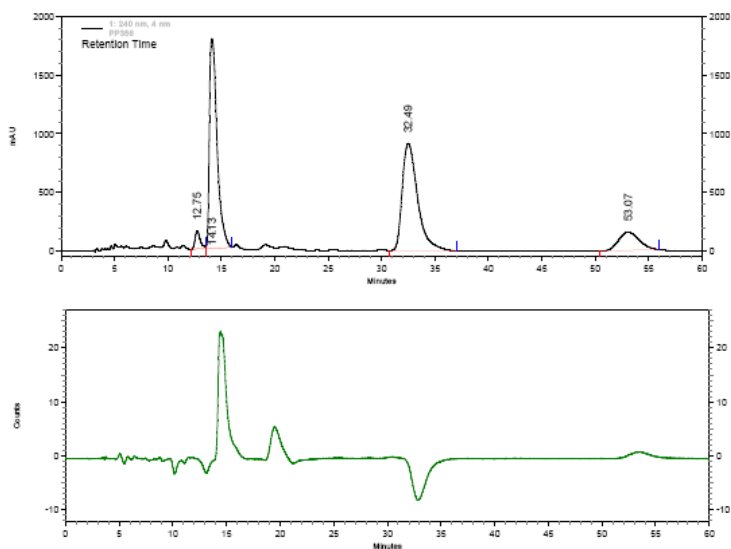


UV Results

Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
36.78	90997649	91.78	11.26	1.00	0.00
62.14	8154687	8.22	19.71	1.75	6.14
Totals	99152336	100.00			

Crude of the cyclization of **5a** (catalytic version)

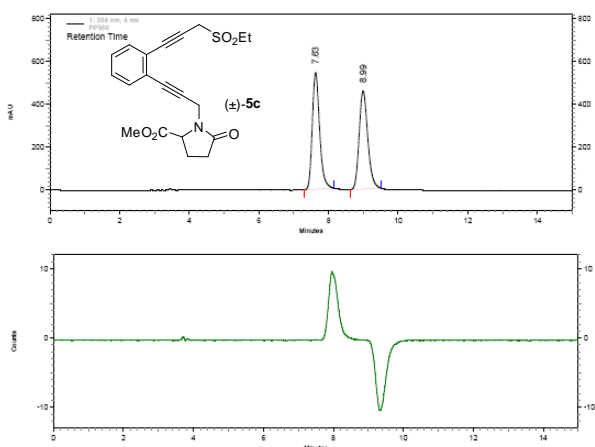
Method description : Chiralpak IC, Hexane/Ethanol 50/50, 1 ml/min, DAD and CD 254nm



1: 240 nm, 4 nm

Results						
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)	
12.75	20766024	2.52	3.25	0.00	0.00	
14.13	356360957	43.23	3.71	0.00	1.34	
32.49	363249939	44.06	9.83	0.00	9.83	
53.07	84018930	10.19	16.69	0.00	6.69	
Totals		824395850	100.00			

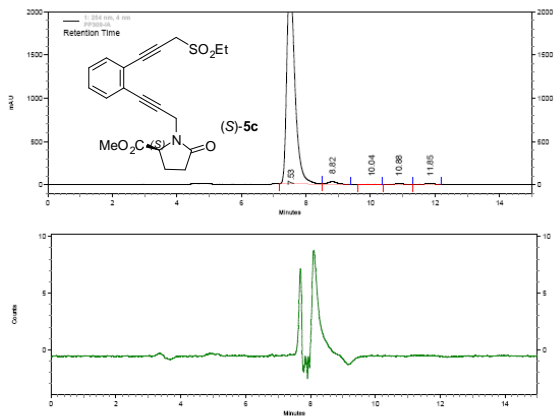
Method description : Chiralpak IA, Hexane/Ethanol 50/50, 1 ml/min, DAD and CD 254nm



1: 254 nm, 4 nm

Results						
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)	
7.83	32003960	50.36	1.54	1.00	0.00	
8.99	31547370	49.64	2.00	1.30	3.27	
Totals		63551330	100.00			

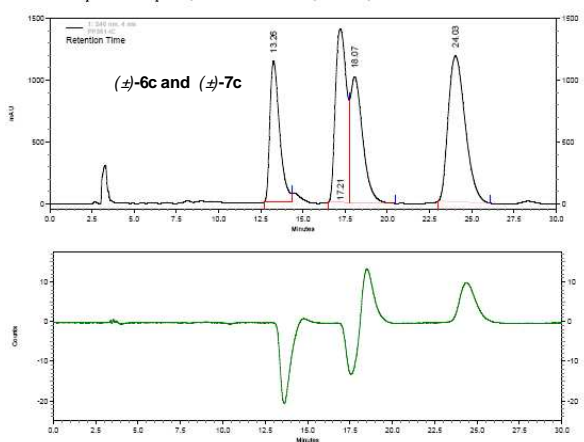
Method description : Chiralpak IA, Hexane/ethanol 50/50, 1 ml/min, UV 254 nm et CD 254nm



1: 254 nm, 4 nm

Results						
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)	
7.53	159147398	98.02	0.00	0.00	0.00	
8.82	2311169	1.42	0.00	0.00	0.00	
10.04	82410	0.05	0.00	0.00	0.00	
10.88	414360	0.26	0.00	0.00	0.00	
11.85	407759	0.25	0.00	0.00	0.00	
Totals		162363096	100.00			

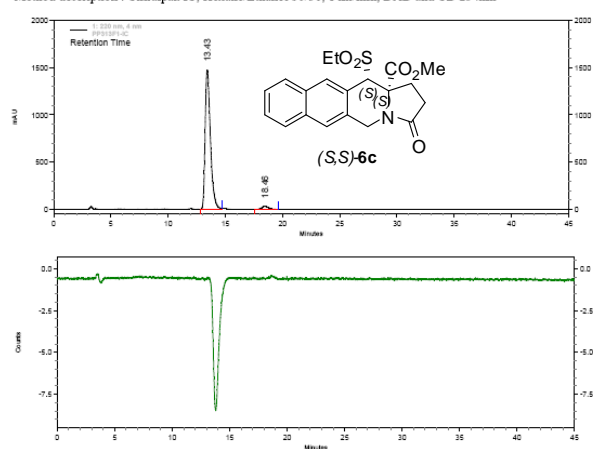
Method description : Chiralpak IC, Hexane/Ethanol 50/50, 1 ml/min, DAD and CD 254nm



1: 240 nm, 4 nm

Results					
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
13.26	180674909	18.59	3.42	0.00	0.00
17.21	254014493	26.14	4.74	0.00	3.17
18.07	208118076	21.42	5.02	0.00	0.49
24.03	328896300	33.85	7.01	0.00	3.04
Totals		971703778	100.00		

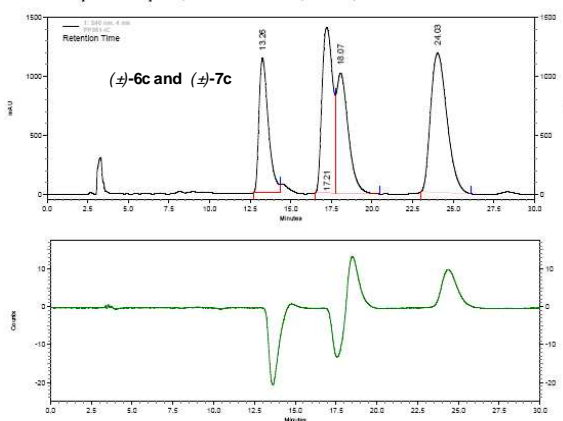
Method description : Chiralpak IC, Hexane/Ethanol 50/50, 1 ml/min, DAD and CD 254nm



1: 220 nm, 4 nm

Results					
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
13.43	190666299	97.21	3.48	1.00	0.00
18.46	5469928	2.79	5.15	1.48	5.06
Totals		196136227	100.00		

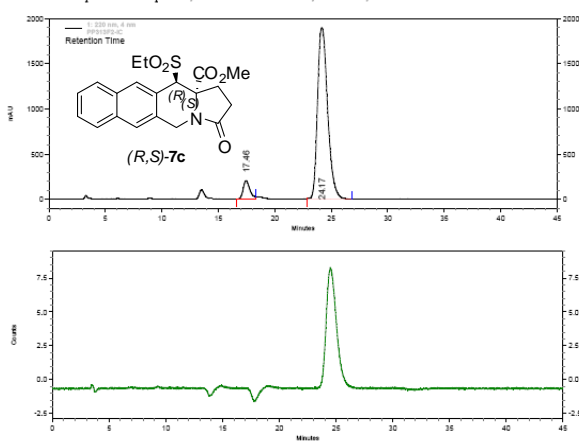
Method description : Chiralpak IC, Hexane/Ethanol 50/50, 1 ml/min, DAD and CD 254nm



1: 240 nm, 4 nm

Results					
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
13.26	180674909	18.59	3.42	0.00	0.00
17.21	254014493	26.14	4.74	0.00	3.17
18.07	208118076	21.42	5.02	0.00	0.49
24.03	328896300	33.85	7.01	0.00	3.04
Totals		971703778	100.00		

Method description : Chiralpak IC, Hexane/ethanol 50/50, 1 ml/min, UV 254 nm et CD 254nm



1: 220 nm, 4 nm

Results					
Retention Time	Area	Area %	Capacity factor	Relative RT	Resolution (USP)
17.46	33877384	6.71	0.00	0.00	0.00
24.17	470906845	93.29	0.00	0.00	0.00
Totals		504784229	100.00		

4. NMR Spectra

