

## Synthesis of a four-armed cage molecule and its pH-controlled complexation with paraquat

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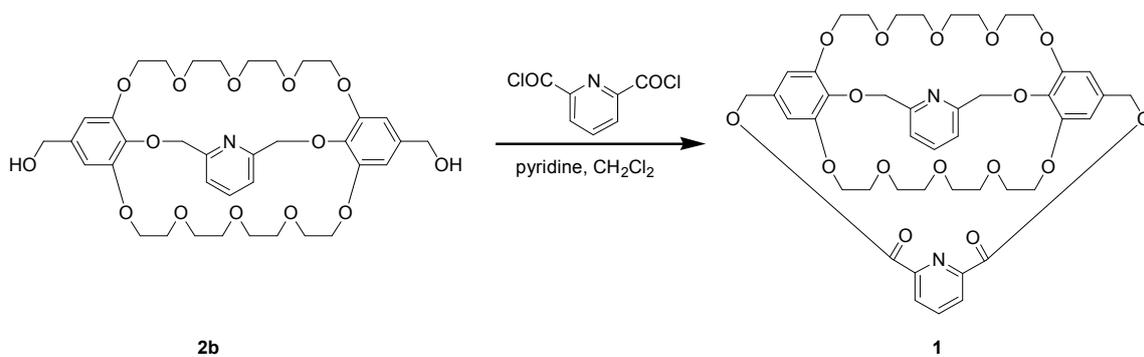
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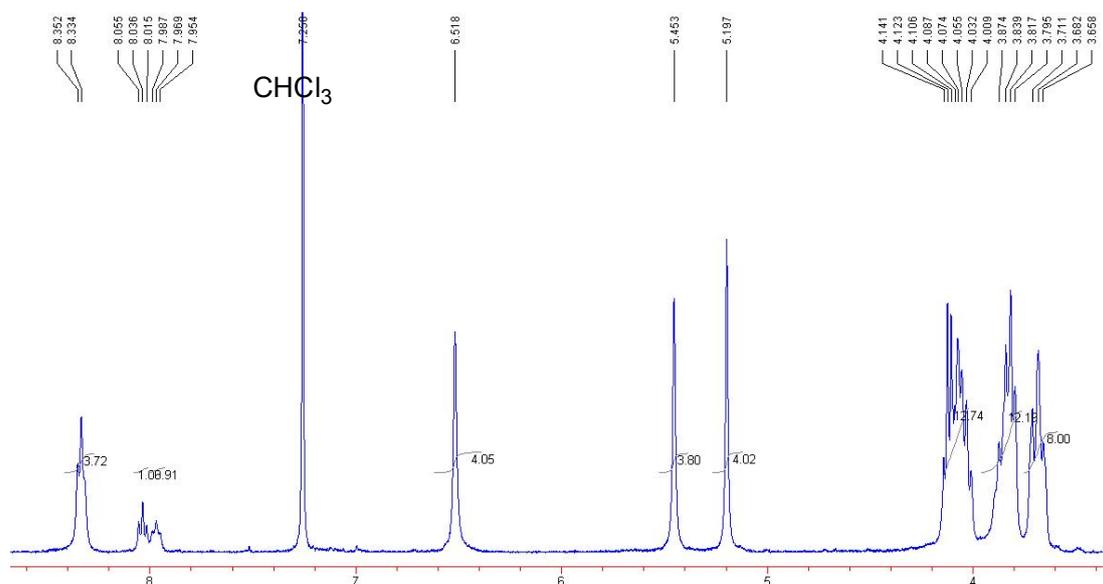
### 1. Materials and methods

All reagents were commercially available and used as supplied without further purification. BMP32C10 (**1**), BMP32C10 diol (**5**),<sup>S1</sup> 2,2'-dihydroxy-BMP32C10,<sup>S2</sup> bis(1,2,3-phenylene) cryptand **2b**<sup>S3</sup> and 2,6-bis(*p*-toluenesulfonyloxymethyl)pyridine<sup>S4</sup> were synthesized by published literature procedures. <sup>1</sup>H NMR spectra were collected on a temperature-controlled 400 MHz or 500 MHz spectrometer. <sup>13</sup>C NMR spectra were recorded on a Bruker AVANCE DMX-400 or DMX-500 spectrometer. Low-resolution electrospray ionization mass spectra (LRESI-MS) were obtained on a Bruker Esquire 3000 plus mass spectrometer (Bruker-Franzen Analytik GmbH Bremen, Germany) equipped with ESI interface and ion trap analyzer. High-resolution electrospray ionization mass spectra (HRESI-MS) were obtained on a Bruker 7-Tesla FT-ICR mass spectrometer equipped with an electrospray source (Billerica, MA, USA). The melting points were collected on a SHPSIC WRS-2 automatic melting point apparatus.

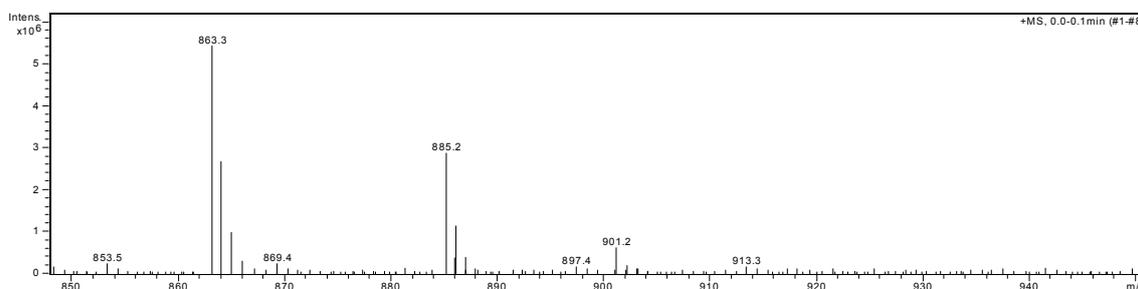
## 2. Synthesis of compound **1**



Pyridine (0.01 mL, 0.100 mmol) and dichloromethane (800 mL) were added into a 1000 mL round-bottom flask. Pyridine-2,6-dicarbonyl (91.8 mg, 0.450 mmol) in dichloromethane (50.0 mL) and bis(1,2,3-phenylene) cryptand **2b** (220 mg, 0.30 mmol) in dichloromethane (50.0 mL) were added at a speed of 2.5 mL/h, separately. Then the mixture was left reacting at room temperature for 4 days. The solvent was removed and the residue was purified by flash column chromatography ( $\text{SiO}_2$ : methanol:ethyl acetate = 1:100) to give **1** as a white solid (30 mg, 12%); m.p. 135.8–136.6 °C. The product has been identified by  $^1\text{H}$  NMR. The  $^1\text{H}$  NMR spectrum is shown in Fig. S1.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ , room temperature)  $\delta$  (ppm): 8.34 (4H, m), 8.05 (1H, m), 8.00 (1H, m), 6.52 (4H, s), 5.45 (4H, s), 5.20 (4H, s), 4.01–4.14 (12H, m), 3.80–3.88 (12H, m), 3.68 (8H, m). LRESIMS is shown in Fig. S2:  $m/z$  863.3 [ $\text{M} + \text{H}$ ] $^+$  (100%), 885.2 [ $\text{M} + \text{Na}$ ] $^+$  (52.9%), 901.2 [ $\text{M} + \text{K}$ ] $^+$  (11.6%). HRESIMS:  $m/z$  calcd for [ $\text{M} + \text{Na}$ ] $^+$   $\text{C}_{44}\text{H}_{50}\text{N}_2\text{NaO}_{16}$ , 885.3053; found 885.3029, error  $-2.7$  ppm.

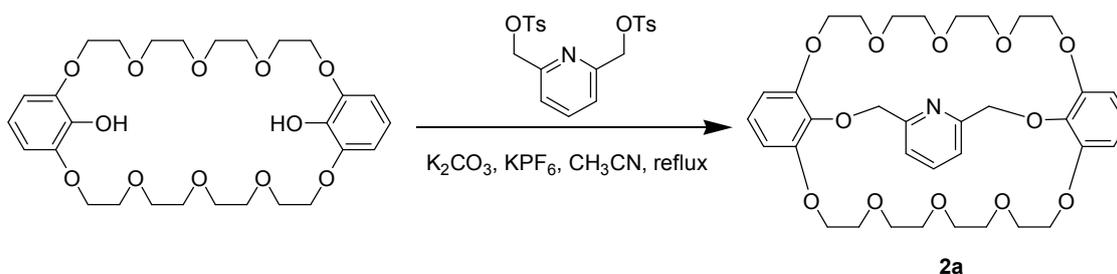


**Fig.S2**  $^1\text{H}$  NMR spectrum (400 MHz,  $\text{CDCl}_3$ , room temperature) of **1**.

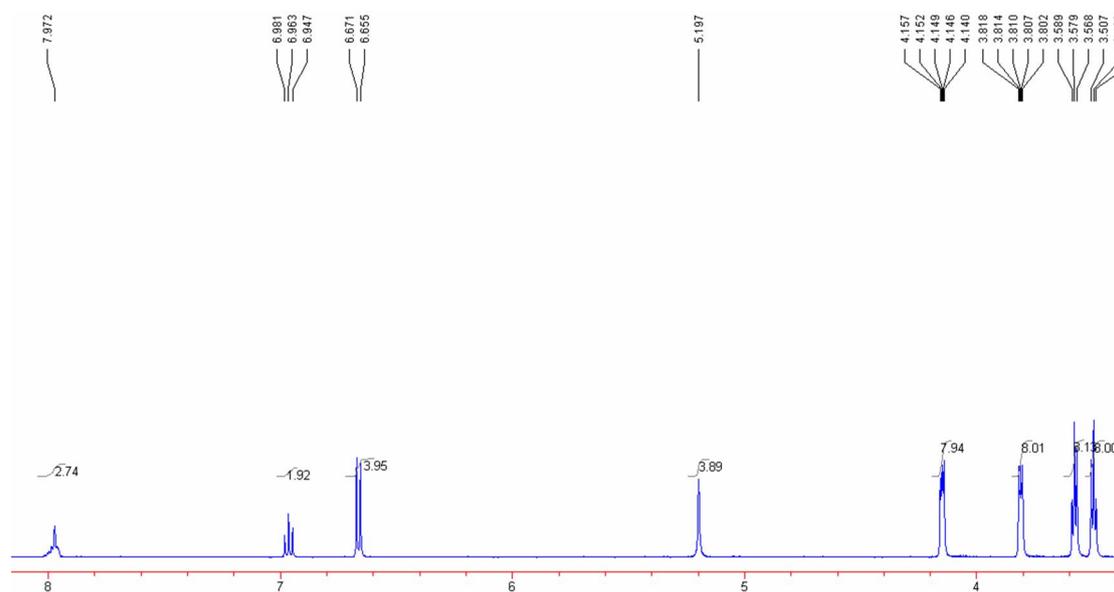


**Fig. S2** LRESI mass spectrum of **1**.

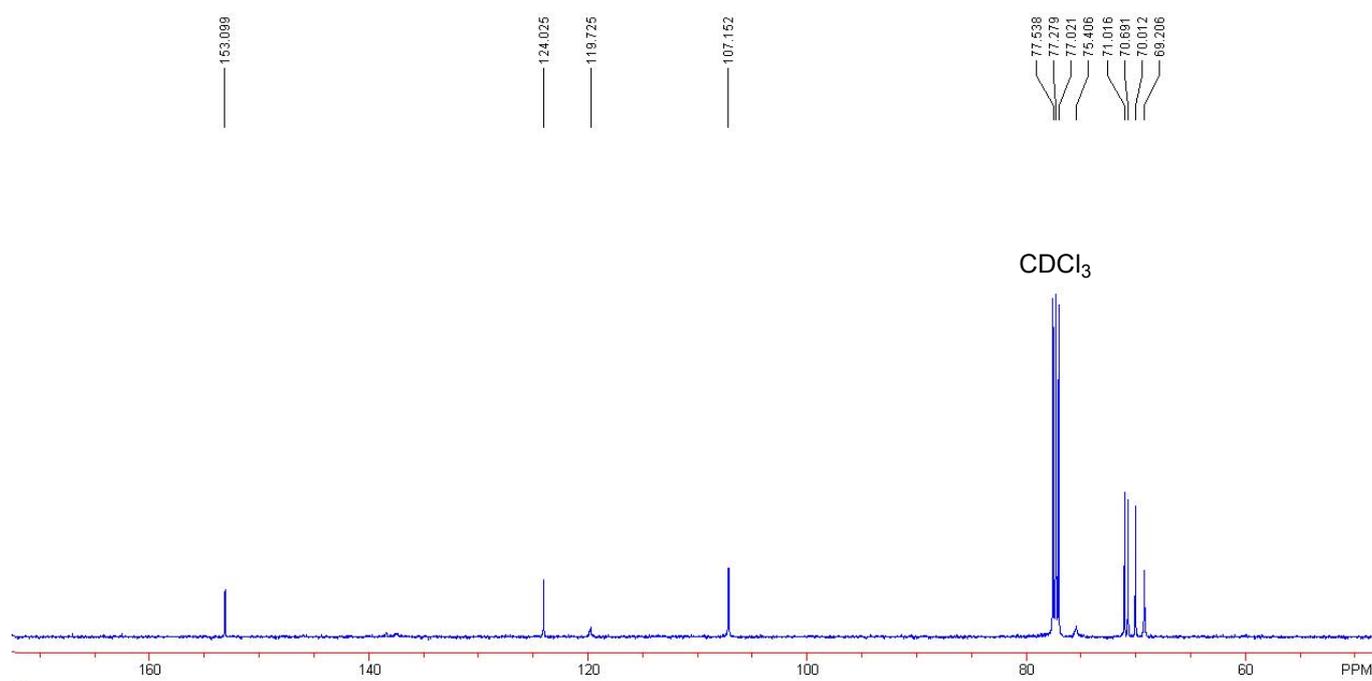
### 3. Synthesis of compound **2a**



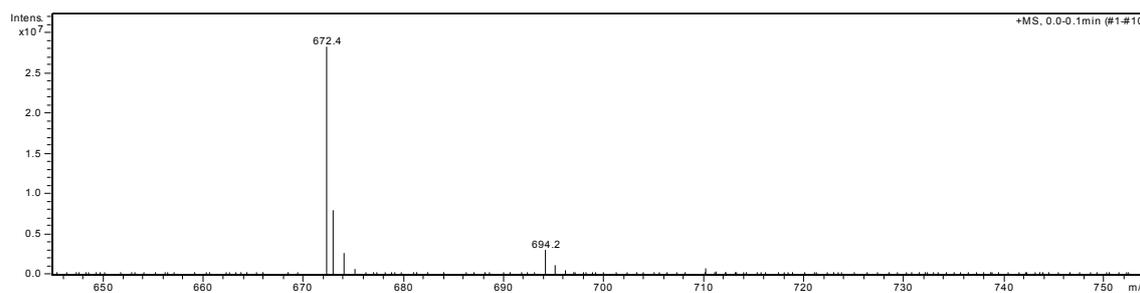
$\text{K}_2\text{CO}_3$  (276 mg, 2.00 mmol),  $\text{KPF}_6$  (92.0 mg, 0.500 mmol) and  $\text{CH}_3\text{CN}$  (80.0 mL) were added into a 150 mL round-bottom flask under nitrogen atmosphere. A  $\text{CH}_3\text{CN}$  (20.0 mL) solution of 2,2'-dihydroxy BMP32C10 (114 mg, 0.200 mmol) and 2,6-bis(*p*-toluenesulfonyloxymethyl)pyridine (89.5 mg, 0.200 mmol) were added at a speed of 1.00 mL/h at reflux. The mixture was then stirred at reflux for 5 days. The solution was filtrated and concentrated to give a crude product, which was purified by flash column chromatography (ethyl acetate) to give **2a** (75 mg, 56%) as a white solid; m.p. 103.3–105.4 °C. The  $^1\text{H}$  NMR spectrum of **2a** was shown in Fig. S3.  $^1\text{H}$  NMR (500 MHz,  $\text{CD}_3\text{COCD}_3$ , room temperature)  $\delta$  (ppm): 7.97 (3H, m), 6.96 (2H,  $J = 8.5$  Hz, t), 6.66 (4H,  $J = 8.5$  Hz, d), 5.20 (4H, s), 4.14–4.16 (8H, m), 3.80–3.82 (8H, m), 3.57–3.59 (8H, m), 3.48–3.51 (8H, m). The  $^{13}\text{C}$  NMR spectrum of **2a** is shown in Fig. S4.  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ , room temperature)  $\delta$  (ppm): 153.1, 124.0, 119.7, 107.2, 75.4, 71.0, 70.7, 70.0, 69.2. LRESIMS is shown in Figure S5:  $m/z$  672.4  $[\text{M} + \text{H}]^+$  (100%), 694.2  $[\text{M} + \text{Na}]^+$  (16.7%). HRESIMS:  $m/z$  calcd for  $[\text{M} + \text{H}]^+ \text{C}_{35}\text{H}_{45}\text{NNaO}_{12}^+$ , 672.3015; found 672.2988, error  $-4.0$  ppm.



**Fig. S3**  $^1\text{H}$  NMR spectrum (500 MHz, acetone- $d_6$ , room temperature) of **2a**.

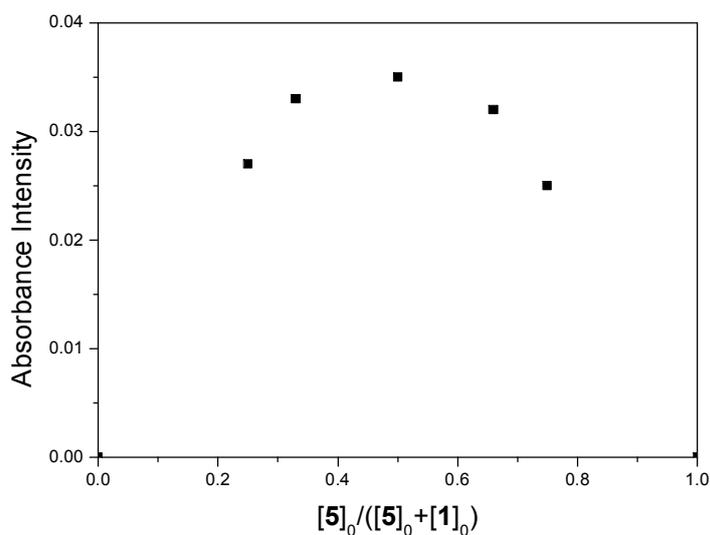


**Fig. S4**  $^{13}\text{C}$  NMR spectrum (100 MHz, chloroform- $d$ , room temperature) of **2a**.



**Fig. S5** LRESI mass spectrum of **2a**.

#### 4. Job plot of **1**↷**5** based on UV-Vis data in 1:1 acetonitrile/chloroform



**Fig. S6** Job plot showing the 1:1 stoichiometry of the complex **1**↷**5** in 1:1 acetonitrile/chloroform.  $[1]_0 + [5]_0 = 0.50$  mM.  $[1]_0$  and  $[5]_0$  are the initial concentrations of **1** and **5**, respectively.

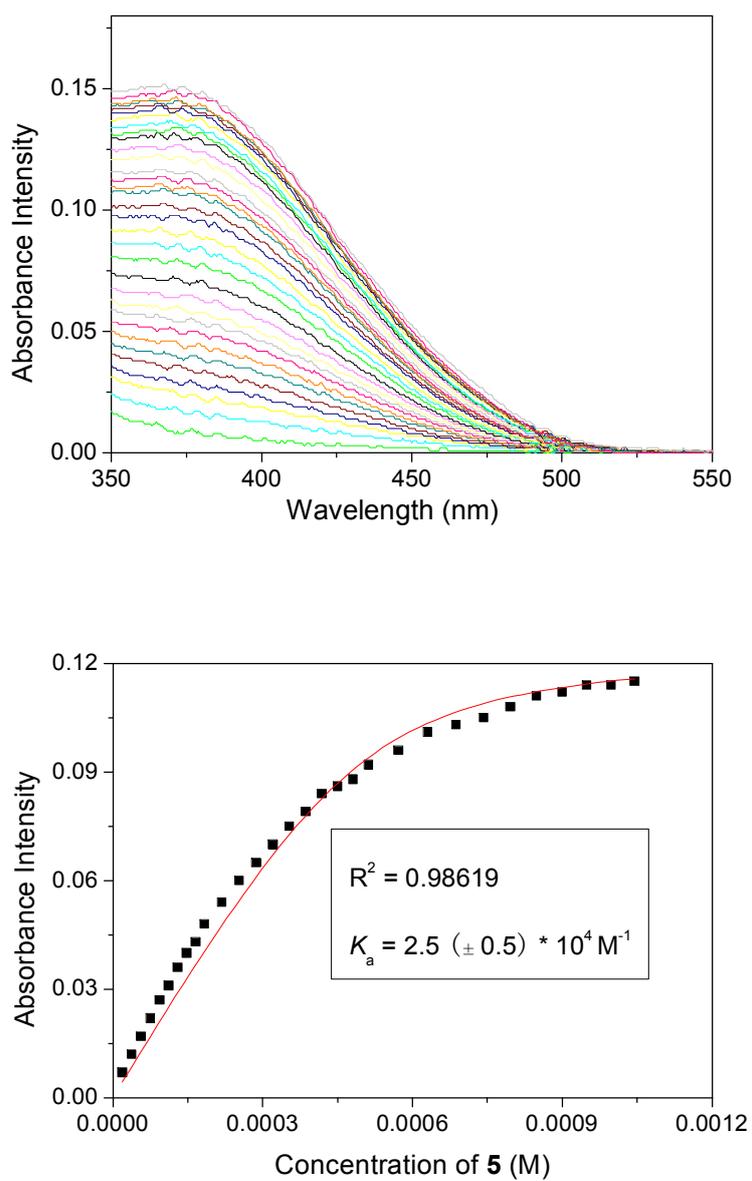
#### 5. Determination of association constants of **1**↷**5** and **4**↷**5**<sup>S5</sup>

The association constants of complexes **1**↷**5** and **4**↷**5** were determined by probing the charge-transfer band of the complexes by UV-vis spectroscopy and employing a titration method. Progressive addition of a 1:1 acetonitrile/chloroform solution with high guest **5** concentration and low host **1** or **4** concentration to a 1:1 acetonitrile/chloroform solution with the same concentration of host **1** or **4** resulted in an increase of the intensity of the charge-transfer band of the complex. Treatment of the collected absorbance data at  $\lambda = 403$  nm with a non-linear curve-fitting program afforded the corresponding association constants ( $K_a$ ):  $2.5 (\pm 0.5) \times 10^4 \text{ M}^{-1}$  for **1**↷**5**, and  $2.7 (\pm 1.4) \times 10^6 \text{ M}^{-1}$  for **4**↷**5**.

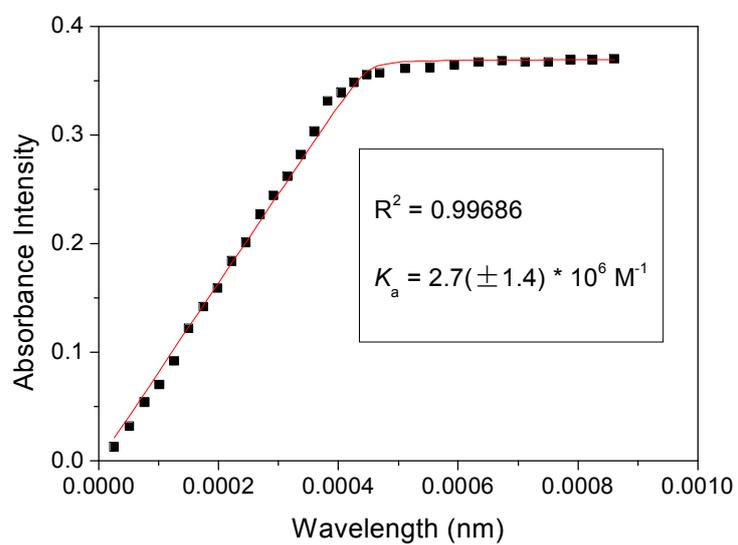
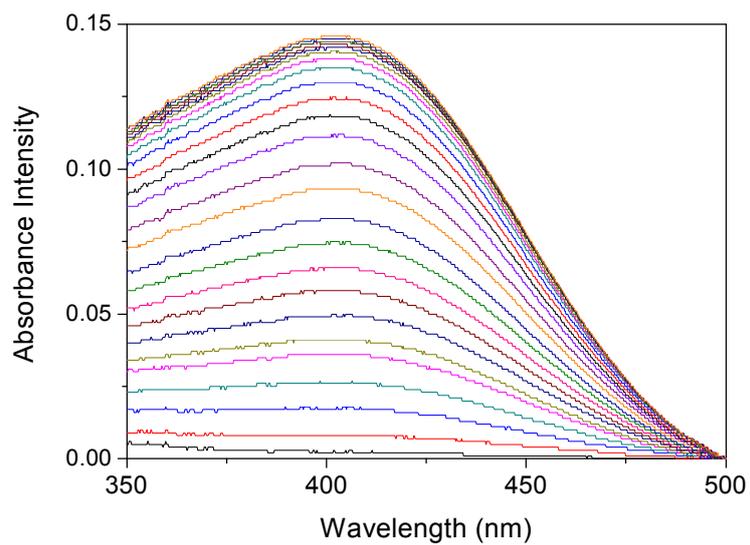
The non-linear curve-fitting was based on the equation:

$$A = (A_\infty/[H]_0) (0.5[G]_0 + 0.5([H]_0 + 1/K_a) - (0.5([G]_0^2 + (2[G]_0(1/K_a - [H]_0) + (1/K_a + [H]_0)^2)^{0.5})) \quad (\text{Eq. S1})$$

Where  $A$  is the absorption intensity of the charge-transfer band ( $\lambda = 403$  nm) at  $[G]_0$ ,  $A_\infty$  is the absorption intensity of the charge-transfer band ( $\lambda = 403$  nm) when the host is completely complexed,  $[H]_0$  is the fixed initial concentration of the host, and  $[G]_0$  is the initial concentration of the guest.



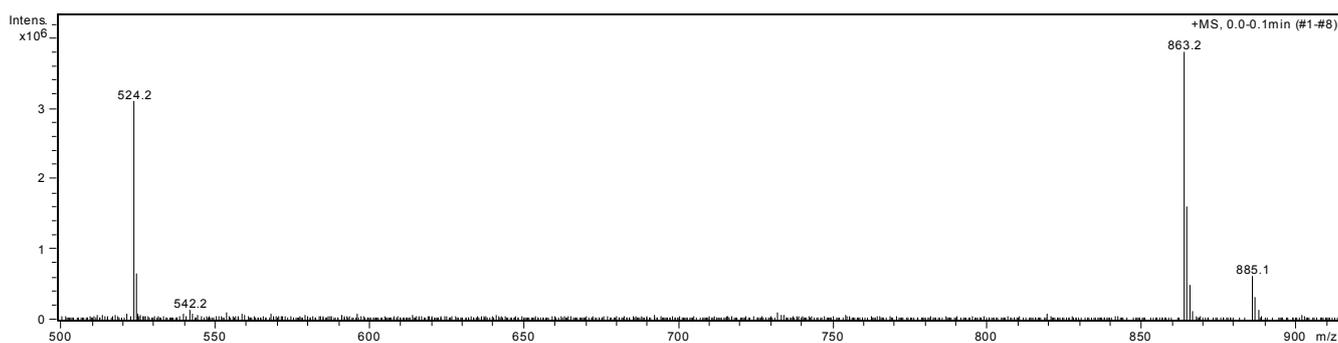
**Fig. S7** Titration curve (top) and non-linear fitting curve (bottom) of host **1** and guest **5**.



**Fig. S8** Titration curve (top) and non-linear fitting curve (bottom) of host **4** and guest **5**.

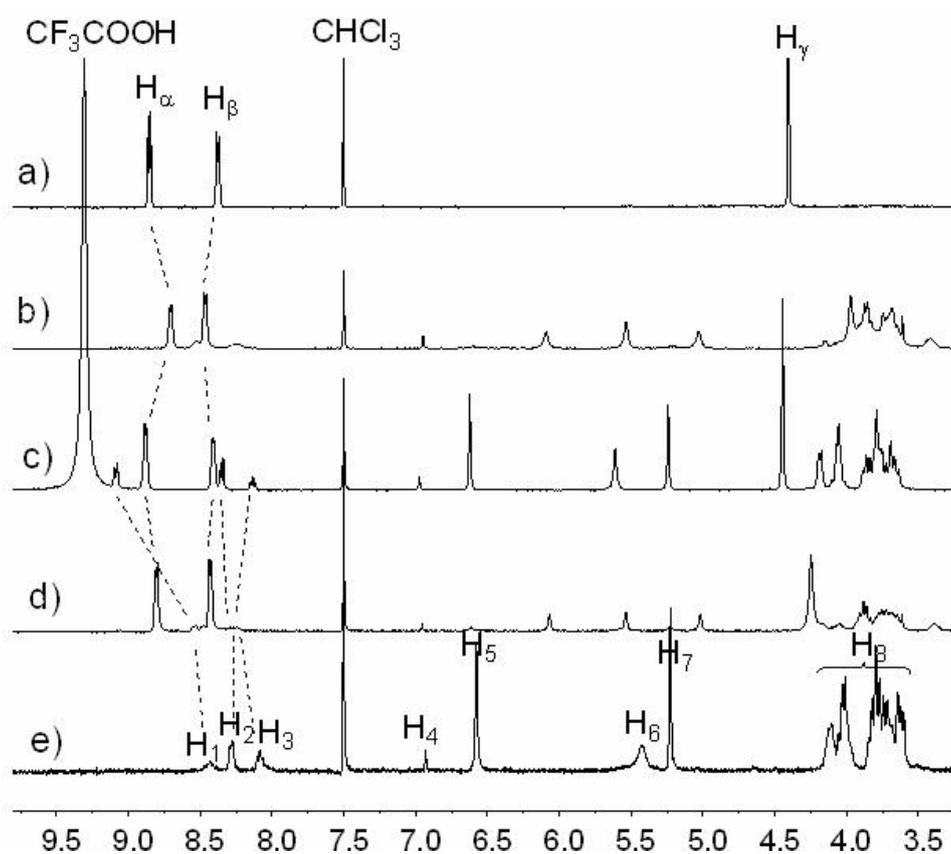
6. Electrospray ionization mass spectrum of an equimolar 1:1 acetonitrile/chloroform solution of **1** and **5**

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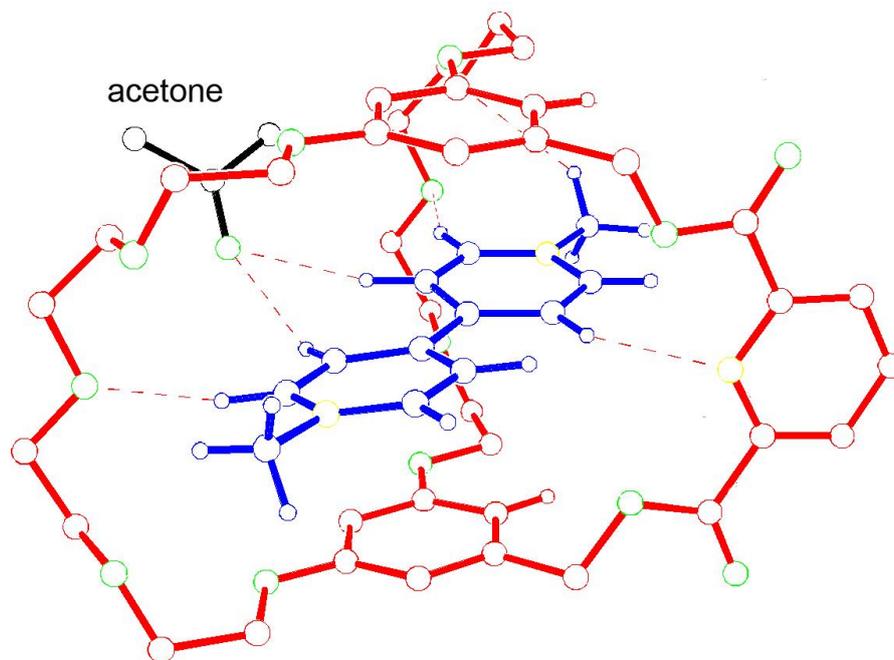
**Fig. S9** Positive electrospray ionization mass spectrum of an equimolar 1:1 acetonitrile/chloroform solution of **1** and **5** gave strong mass fragments at  $m/z$  524.2 (81.3%), 863.2 (100%) and 885.1 (20.5%), corresponding to  $[\mathbf{1}\rightarrow\mathbf{5} - 2\text{PF}_6]^{2+}$ ,  $[\mathbf{1} + \text{H}]^+$  and  $[\mathbf{1} + \text{Na}]^+$ , respectively.

7. Partial  $^1\text{H}$  NMR spectra on the pH-controlled complexation between **1** and **5**



**Fig. S10** Partial  $^1\text{H}$  NMR spectra (400 MHz,  $\text{CDCl}_3/\text{CD}_3\text{CN}$  1:1, 295 K) of: a) **5**, b) 4.00 mM **1** and **5**, c) a solution of 4.0  $\mu\text{L}$  of trifluoroacetic acid-*d* and 0.5 mL of 4.00 mM **1** and **5**, d) a solution of 4.0  $\mu\text{L}$  of trifluoroacetic acid-*d*, 8.0  $\mu\text{L}$  of triethylamine, and 0.5 mL of 4.00 mM **1** and **5**, and e) **1**.

### 8. Crystal structure of complex 4 $\rightarrow$ 5<sup>S6</sup>



**Fig. S11** Crystal structure of complex 4 $\rightarrow$ 5: 4 is red, 5 is blue, and the acetone molecule is black.

#### References:

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