- Electronic Supplementary Information -

Pd²⁺...O₃SR⁻ interaction encourages anion encapsulation of a quadruply-stranded Pd complex to achieve chirality or high solubility

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References

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Experimental Section

Materials and Instrumentations

All chemicals and solvents were purchased from Kanto Chemical Co., Ltd., Wako Pure Chemical Co., Ltd., and Tokyo Kasei Kogyo Co., Ltd., and were used as received without further purification. Melting points were measured on Yanaco Micro Melting Point Apparatus 3120. ¹H NMR (500MHz), ¹³C NMR (125 MHz), and ¹⁹F NMR (470 MHz) spectra were recorded on a JEOL α -500 spectrometer. Unless stated otherwise, the chemical shift values reported here are with respect to DMSO ($\delta = 2.48$ ppm (¹H), 39.5 ppm (¹³C)) or monofluorobenzene ($\delta = -113.15$ ppm, (¹⁹F)). ESI MS spectra were acquired on MarinerTM ESI-TOF Biospectrometry Workstation. Solution-state CD spectra were measured on a JASCO J-720 spectrometer.

Solid-state CD Measurements

Solid-state CD spectra were recorded using Universal Chiroptical Spectrophotometer (UCS-1).¹ A crystal of compounds (1(d-camS)₄ (100 µg) or 1(l-camS)₄ (108 µg)) and dry KBr matrix were finely ground, and the powder was pressed at 4.8 MP/ cm² for 10 min to prepare a clear disk of 1 cm diameter. The KBr disk, held by a specially devised disk holder, was placed normal to a light beam. True CD spectra in the solid state were obtained by a specially devised set of procedures based on the Storkes-Mueller matrix method.¹

X-ray Crystallography

Single crystals of $1(ONs)_4$ were obtained from a DMF solution of $1(ONs)_4$ at ca. 70 °C. Single crystals of 1(d-camS)₄ were obtained from a DMSO solution of 1(d-camS)₄ at rt. X-ray crystallographic data were collected on a Bruker SMART APEX CCD diffractometer using graphite-monochromatized Mo K α radiation ($\lambda = 0.71073$ Å) at 103 K. Crystallographic parameters are listed in Table S1. The crystal structures were solved by direct method using the *SHELXS-97* program and refined by successive differential Fourier syntheses and full-matrix least-squares procedures using the *SHELXL-97* program.² Anisotropic thermal factors were applied to all non-hydrogen atoms. In the case of $1(ONs)_4$, the diffuse electron density due to the disordered and unidentified moieties was treated with the SQUEEZE routine within the PLATON software package,³ and the final d_{calc} , and Formula weight reflect known contents only. Computer graphics of the X-ray crystal structures were portrayed with Mercury 2.3 program.⁴

NMR Titrations.

All titrations were performed with the starting concentration of $1(\text{OTf})_4$ at 1.0 mM (DMSO- d_6) and the addition of appropriate aliquots of titrant (0.5, 1.0, 1.5, 2.0, 3.0, 4.0, 5.0, 6.0, 7.0, and 8.0 equiv) to the NMR sample with a 40-200 µl Finnpipette[®] (ThermoLabsystems) as appropriate. α -pyridyl protons H_a and H_b of **1** were followed during the course of the titration, and the data were fitted and analyzed to give association constants (log₁₀(K_{a1}) and log₁₀(K_{a})) using WinEQNMR2 program.⁵

Table S1. Crystallographic parameters

Crystal	1 (ONs) ₄	1 (<i>d</i> -camS) ₄
Formula	$[Pd_2(C_{25}H_{20}N_2O_3)_4]$ ·	$[Pd_2(C_{25}H_{20}N_2O_3)_4]$ ·
	(ONs) ₄ ·(DMF) ₄	$(d-\text{camS})_4$ ·(DMSO) ₆
Formula weight	2919.81	3192.4
Crystal system	Triclinic	Triclinic
Space group	P 1 (#2)	P1 (#1)
<i>a</i> / Å	14.075(6)	14.6994(11)
b/ Å	16.502(7)	16.5936(12)
<i>c</i> / Å	19.601(8)	17.1117(12)
α⁄/°	92.546(7)	83.298(1)
<i>β</i> / °	108.332(7)	85.992(1)
γ°	109.447(7)	63.960(1)
$V/ \text{\AA}^3$	4018(3)	3723.7(5)
$D_{\text{calc.}}/\text{g cm}^{-3}$	1.21	1.42
Ζ	1	1
$2\theta_{\rm max}/^{\rm o}$	52.8	55.8
μ (MoK α) /mm ⁻¹	0.344	0.461
Temperature /K	103	103
Crystal form	block	block
Crystal size /mm ³	0.2 imes 0.1 imes 0.05	0.2 imes 0.2 imes 0.2
Crystal colour	colourless	colourless
<i>h</i> range	$-17 \rightarrow 17$	$-19 \rightarrow 8$
<i>k</i> range	$-20 \rightarrow 20$	$-21 \rightarrow 19$
<i>l</i> range	$-24 \rightarrow 23$	$-22 \rightarrow 21$
# of total reflections	32666	23115
# of unique reflections	16175	19148
# of observed reflections	10148	17459
R _{int}	0.0722	0.0168
Criterion for observed		
reflections	$I > 2\sigma(F_o)$	$I > 2\sigma(F_o)$
R1 (observed)	0.0737	0.0361
wR2 (observed)	0.1735	0.0953
<i>G. O. F.</i>	0.983	1.057
# of parameters used	892	1875
$\Delta \rho_{\rm max} ({\rm e}{\rm \AA}^{-3})$	+1.152	+1.515
$\Delta \rho_{\min} (e \text{\AA}^{-3})$	-0.766	-0.637
Flack's parameter	-	-0.018(13)
CCDC number	742382	827758

Experimental Detail

Synthesis of 1(OTf)₄



PdCl₂ (44.4 mg, 2.50×10^{-1} mmol) was dissolved in DMF (4 mL) and stirred for 1 h at room temperature with AgOTf (129 mg, 5.00×10^{-1} mmol). After removal of AgCl by filtration addition of compound L⁶ (208 mg, 5.25×10^{-1} mmol) to the filtrate, the mixture was stirred for 1 h at 70 °C. Et₂O (20 mL) was added to the resultant pale yellow solution. White precipitate was isolated by filtration and washed with CH₂Cl₂ and dried in vacuo to give 1(OTf)₄ in 91 % yield (272 mg, 1.14×10^{-1} mmol). ¹H NMR (500 MHz, DMSO-*d*₆, rt): δ 9.40 (s, 8H), 9.33 (d, *J* = 5.8 Hz, 8H), 8.16 (d, *J* = 7.6 Hz, 8H), 7.82 (dd, *J* = 5.8, 7.6 Hz, 8H), 7.54 (d, *J* = 8.5 Hz, 16H), 7.10 (d, *J* = 8.5 Hz, 16H), 5.29 (s, 16H); ¹³C NMR (125 MHz, DMSO-*d*₆, rt): δ 193.01, 160.92, 150.61, 149.09, 139.79, 136.22, 131.65, 130.74, 127.12, 114.35, 66.57; ¹⁹F NMR (470 MHz, DMSO-*d*₆, rt): δ -77.8 (s); MS (ESI, *m/z*) calcd for C₁₀₀H₈₀N₈O₁₂Pd₂(OTf)₄: 2394.87; Found: 1047.8 [1+(OTf)₂]²⁺, 649.5 [1+(OTf)]³⁺, 449.4 1. (1 = C₁₀₀H₈₀N₈O₁₂Pd₂ = [Pd₂L₄]⁴⁺).

Synthesis of 1(d-camS)₄



Sodium *d*-camphorsulfonate (13.7 mg, 5.40×10^{-2} mmol) was added to $1(\text{OTf})_4$ (32.3 mg, 1.35×10^{-2} mmol) at room temperature in DMF (2 mL). Upon addition, the mixture was stirred at 60 °C for 1.5 h, after which white precipitate was isolated by filtration and washed several times with small quantities of distilled water and dried in vacuo to give $1(d\text{-camS})_4$ in 75 % yield (27.4 mg, 1.01×10^{-2} mmol).

Synthesis of 1(*l*-camS)₄



Ammonium *l*-camphorsulfonate (13.1 mg, 5.27×10^{-2} mmol) was added to $1(\text{OTf})_4$ (31.5 mg, 1.31×10^{-2} mmol) at room temperature in DMF (2 mL). Upon addition, the mixture was stirred at 60 °C for 1.5 h, after which white precipitate was isolated by filtration and washed several times with small quantities of distilled water and dried in vacuo to give $1(l\text{-camS})_4$ in 68 % yield (24.1 mg, 8.9×10^{-3} mmol).

Synthesis of Na(DenS)



Sodium *p*-hydroxybenzenesulphonate (0.18 g, 7.7×10^{-1} mmol) in dry DMF (5 mL) was added slowly to 3,5-bis(benzyloxy)benzoic acid (0.26 g, 7.8×10^{-1} mmol)⁷ and *N*,*N*^c-dicyclohexylcarbodiimide (0.16 g, 7.7×10^{-1} mmol) at 0 °C in dry CH₂Cl₂ (10 mL). The solution was stirred at rt for 22 h and concentrated under reduced pressure to give the crude product. The crude product was purified by column chromatography using silica gel (CH₂Cl₂:MeOH = 9:1, v/v) to give a white powder. Repeated crystallization from methanol-CH₂Cl₂ mixed solvent solutions gave colourless crystals of Na(DenS)(H₂O)_{1.5} in 30 % yield (0.12 g, 2.26×10^{-1} mmol). ¹H NMR (500 MHz, DMSO-*d*₆, rt): δ 7.66 (d, *J* = 5.8 Hz, 2H), 7.46 (d, *J* = 8.5 Hz, 4H), 7.39 (t, *J* = 7.3 Hz, 4H), 7.33 (m, 4H), 7.20 (d, *J* = 8.5 Hz, 2H), 7.04 (t, *J* = 2.1 Hz, 1H), 5.19 (s, 4H); ¹³C NMR (125 MHz, DMSO-*d*₆): δ 164.01, 159.44, 150.37, 145.78, 136.45, 130.65, 128.05, 127.84, 127.63, 127.27, 126.84, 121.05, 108.40, 69.64; Elemental Analysis calcd. for C₂₇H₂₁O₇S Na(H₂O)_{1.5}: C, 60.10; H, 4.48; Found: C, 60.17; H, 4.66; Melting Point: > 300 °C (decomp.).

Supporting Figures



Fig. **S1** (a) An energy minimized structure (universal force field method) of $1^{.6,8}$ Colour scheme: gray (carbon), white (hydrogen), blue (nitrogen), red (oxygen), green (palladium). (b) Cavity volume of 1.



Fig. S2 (a) Chemical shift change of H_a upon dilution ([1]_{start} = 1.0 mM, [OTf⁻]_{start} = 4.0 mM). (b) and (c) NMR titrations (OTf⁻) monitoring H_a and H_b , respectively. [1] = 1.0 mM.



Fig. S3 ¹H NMR titration (OTs⁻) monitoring (a) H_a and (b) H_b . [1] = 1.0 mM. (c) H_a (obs.) – H_a (calc.). H_a (calc.) were calculated using WinEQNMR2 program⁵ (see also p. S20).



Fig. S4 ¹H NMR titration (ONs⁻) monitoring (a) H_a and (b) H_b . [1] = 1.0 mM. (c) H_a (obs.) – H_a (calc.). H_a (calc.) were calculated using WinEQNMR2 program⁵ (see also p. S21).



Fig. **S5** Observed and calculated ion peaks of (a) $[1+(OTs)_2]^{2+}$ and (b) $[1+(ONs)_2]^{2+}$.



Fig. **S6** Partial views of the 2D NOESY spectrum (500 MHz, DMSO- d_6 , rt) of **1** with 4 equiv OTs⁻. Mixing time = 1000 ms. The arrows denote NOEs.







Fig. S8 Coordination environment around the Pd^{2+} centre of $1 \supset (ONs)_2$ with atomic numbering scheme. Red dotted lines denote C–H···O hydrogen bonding. Colour scheme: gray (carbon), white (hydrogen), light blue (nitrogen), red (oxygen), orange (sulphur), purple (palladium).



Fig. S9 Chemical shift change of d-camS⁻ upon host-guest complexation (500 MHz, DMSO- d_6 , rt). (a) d-camS⁻. (b) 1 + d-camS⁻ ([1] = 1.25 mM, [d-camS⁻] = 5.0 mM).



Fig. **S10** A partial view of the 2D NOESY NMR spectrum (500 MHz, DMSO- d_6 , rt) of **1** with 4 equiv d-camS⁻. ([**1**] = 2.5 mM and [d-camS⁻] = 10.0 mM). Mixing time = 1000 ms.



Fig. S11 (a) ESI MS spectrum of the DMSO- d_6 solution containing $1_{\text{major}} \supset (d\text{-cam})_2$ and $1_{\text{minor}} \supset (d\text{-cam})_2$ (See also Fig. 1d). (b) and (c) Observed and calculated ion peaks of $[1+(d\text{-camS})]^{3+}$ and $[1+(d\text{-camS})_2]^{2+}$, respectively.



Fig. **S12** ¹H NMR titration (*d*-camS⁻) monitoring (a) H_{*a*} and (b) H_{*b*}. [1] = 2.5 mM.



Fig. S13 ¹H NMR spectrum (500 MHz, DMSO- d_6 , rt) of **1** with 1 equiv *d*-camS⁻. ([**1**] = 2.5 mM and [*d*-camS⁻] = 2.5 mM).

Pd…O distance

Pd1…O102: 3.038 Å Pd1…O401 :3.122 Å

Pd–N distance

Pd1-N1: 2.023 Å Pd1-N3 :2.003 Å Pd1-N5: 2.023 Å Pd1-N7 :2.027 Å

C–H···O hydrogen bonding

 O102····H1 : 2.868 Å
 O102····C1 : 3.614 Å

 O102····H26 : 2.242 Å
 O102···C26 : 3.075 Å

 O102····H51 : 2.786 Å
 O102···C51 : 3.469 Å

 O102····H51 : 2.786 Å
 O102···C76 : 3.364 Å

 O102····H51 : 2.552 Å
 O102···C76 : 3.374 Å

 O401····H5 : 2.514 Å
 O401···C30 : 3.264 Å

 O401···H30 : 2.488 Å
 O401···C30 : 3.174 Å

 O401···H80 : 2.626 Å
 O401···C80 : 3.463 Å



∠ O···Pd···O angle

∠ O102…Pd1…O401 : 171.36 °

∠ O…Pd–N angle

- ∠ O102…Pd1–N1 : 95.60 °
- ∠ O102…Pd1–N3 : 85.92 °
- ∠ O102…Pd1–N5 : 90.31 °
- \angle O102···Pd1–N7 : 91.05 ° \angle O401···Pd1–N1 : 92.25 °
- ∠ O401…Pd1–N3 : 86.35 °
- ∠ O401…Pd1–N5 : 85.67 °
- ∠ O401…Pd1–N7 : 93.20 °

\angle C–H···O angle

- ∠ C1-H1…O102 : 136.18°
 ∠ C26-H26…O102 : 145.78°
 ∠ C51-H51…O102 : 129.55°
 ∠ C76-H76…O102 : 143.79°
 ∠ C5-H5…O401 : 150.64°
 ∠ C30-H30…O401 : 138.79°
 ∠ C55-H55…O401 : 139.06°
- ∠ C80–H80…O401 : 147.17 °

Pd…O distance

Pd2…O302 : 3.039 Å Pd2…O202 : 3.214 Å

Pd–N distance

Pd1-N2: 2.034 Å Pd1-N4 :2.020 Å Pd1-N6: 2.030 Å Pd1-N8 :2.027 Å

C-H···O hydrogen bonding

 O302…H23: 2.229 Å
 O302…C23: 3.100 Å

 O302…H50: 2.716 Å
 O302…C50: 3.483 Å

 O302…H75: 2.485 Å
 O302…C75: 3.301 Å

 O302…H75: 2.485 Å
 O302…C75: 3.301 Å

 O302…H100: 2.596 Å
 O302…C100: 3.328 Å

 O202…H22: 2.433 Å
 O202…C22: 3.274Å

 O202…H47: 2.633 Å
 O202…C47: 3.500 Å

 O202…H71: 2.616 Å
 O202…C71: 3.4351 Å

 O202…H97: 2.637 Å
 O202…C97: 3.438 Å



∠ O…Pd…O angle

∠ O202…Pd2…O120 : 173.08 °

∠ O…Pd–N angle



Fig. S14 Coordination environments around the Pd^{2+} centres of $1 \supset (d\text{-camS})_2$ with atomic numbering scheme. Red dotted lines denote C–H···O hydrogen bonding. Colour scheme: gray (carbon), white (hydrogen), light blue (nitrogen), red (oxygen), orange (sulphur), purple (palladium).



Fig. **S15** Solid-state transmittance spectra (KBr disk) of $1(d-\text{camS})_4$ (blue line) and $1(l-\text{camS})_4$ (red line). Positive and negative CD originate from $d-\text{camS}^-$ and $l-\text{camS}^-$, respectively.



Fig. **S16** ORTEP drawing (50 % probability ellipsoids) of the X-ray crystal structures of receptors **1** with *P*-form (both receptors are the components of an interlocked metallohelicate with *P*-form).⁶ The benzophenone cores possess *P*-helicity. Colour scheme: gray (carbon), white (hydrogen), blue (nitrogen), red (oxygen), purple (palladium).



Fig. S17 Partial views of the 2D NOESY spectrum (500 MHz, DMSO- d_6 , rt) of **1** with 4 equiv DenS⁻. ([**1**] = 2.5 mM and [DenS⁻] = 10.0 mM). Mixing time = 1000 ms.



Fig. S18 Chemical shift change of the signals of d-camS⁻ upon host-guest complexation. (a) DenS⁻. (b) $1 + \text{DenS}^-$ ([1] = 2.5 mM, [DenS⁻] = 10.0 mM).



Fig. S19 (a) Observed and (b) calculated ion peaks of $[1+(DenS)_2]^{2+}$.



Fig. S20 (a), (b) ¹H NMR spectra (500 MHz, DMSO- d_6 +CD₂Cl₂, rt) of **1** with 4 equiv DenS⁻. The chemical shifts are with respect to DMSO ($\delta = 2.48$ ppm). (c) ¹H NMR spectrum (500 MHz, DMSO- d_6 +CD₂Cl₂, rt) of the suspended solution of **1**(OTf)₄. Noise reduction was applied to the spectrum. (•) and (•) denote partially duterated dichloromethane and free L, respectively.⁹



Fig. S21 (a), (b) ¹H NMR spectra (500 MHz, DMSO- d_6 +CDCl₃, rt) of **1** with 4 equiv DenS⁻. The chemical shifts are with respect to DMSO ($\delta = 2.48$ ppm). (c) ¹H NMR spectrum (500 MHz, DMSO- d_6 +CDCl₃, rt) of the suspended solution of **1**(OTf)₄. Noise reduction was applied to the spectrum. (•) and (•) denote partially duterated chloroform and free L, respectively.⁹



Fig. S22 (a), (b) ¹H NMR spectra (500 MHz, DMSO- d_6 +THF- d_8 , rt) of **1** with 4 equiv DenS⁻. The chemical shifts are with respect to DMSO ($\delta = 2.48$ ppm). (c) ¹H NMR spectrum (500 MHz, DMSO- d_6 +THF- d_8 , rt) of the suspended solution of **1**(OTf)₄. Noise reduction was applied to the spectrum. (\circ) denotes free L.⁹

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Fig. **S23** ¹H NMR titration (DenS⁻) monitoring (a) H_a and (b) H_b . [1] = 1.0 mM. (c) H_a (obs.) – H_a (calc.). H_a (calc.) were calculated using WinEQNMR2 program⁵ (see also p. S22).

1000000000000000000000000000000000000				
Anion	$\log_{10}(K_{a1})$	$\log_{10}(K_a)$		
OTs ⁻	2.9(3)	5.1(5)		
ONs	3.01(10)	5.57(14)		
DenS	3.7(2)	5.9(4)		
$K_{a1} = [1 \supset (X)]/([1] \cdot [X]) \text{ and } K_a = [1 \supset (X)_2]/([1] \cdot [X]^2)$				

Table S2 Association constants $log_{10}(K_{a1})$ and $log_{10}(K_a)$ in DMSO- d_6 .

Data fitting of the NMR titrations $(1 + OTs^{-})^{5}$

Calculations by WinEQNMR2 Version 2.00 by Michael J. Hynes Program run at 19:25:21 on 12/03/2010

Reactions: M + L = ML (beta1 = K1); M + 2L = (beta2 = K1K2) Data 2010 05 14 === Anion Recognition ===

Equilibrium constants are log₁₀ values

NO. A PARAMETER DELTA ERROR CONDITION DESCRIPTION

- 1 1 2.93323E+00 1.000E-01 2.580E-01 2.496E+03 Stability constant(K1)
- 2 1 5.12105E+00 1.000E-01 5.104E-01 1.401E+04 Stability constant(K1K2)
- 3 1 9.44643E+00 1.000E-06 2.168E-03 5.890E+00 Shift of 1
- 4 1 9.89682E+00 1.000E-04 1.533E-01 1.823E+04 Shift of 1:1_complex
- 5 1 1.01493E+01 1.000E-04 1.872E-02 9.554E+01 Shift of 1:2_complex

0RMS ERROR = 1.65E-03 MAX ERROR = 1.95E-03 AT OBS.NO. 3 RESIDUALS SQUARED = 1.36E-05 RFACTOR = 0.0119 PERCENT

NO. AEXPT. DELCALC. DELRESIDUAL% DEVWEIGHTGuest(OTs)Host(1)pH19.5390E+009.5400E+00-9.8324E-04-1.0308E-021.0000E+005.0000E+041.0000E-030.0000E+00219.6170E+009.6170E+001.9073E-061.9833E-051.0000E+001.0000E-031.0000E-030.0000E+00319.6830E+009.6810E+001.9512E-032.0151E-021.0000E+001.5000E-031.0000E-030.0000E+00419.7330E+009.7322E+007.8106E-048.0249E-031.0000E+002.0000E-031.0000E-030.0000E+00519.8080E+009.8096E+00-1.6432E-03-1.6753E-021.0000E+003.0000E-031.0000E-030.0000E+00619.8650E+009.8644E+006.4468E-046.5351E-031.0000E+004.0000E-031.0000E+030.0000E+00719.9030E+009.9047E+00-1.6518E-03-1.6679E-021.0000E+005.0000E-031.0000E+030.0000E+00819.9350E+009.9354E+00-3.5954E-04-3.6189E-031.0000E+006.0000E-031.0000E+030.0000E+00919.9610E+009.9595E+001.4944E-031.5003E-021.0000E+007.0000E-031.0000E+030.0000E+00109.9790E+009.9789E+001.2302E-041.2328E-031.0000E+008.0000E-031.0000E-030.0000E+00

TOLERANCE ON SUM OF SQUARES0.0100TOLERANCE ON EIGEN VALUES0.0001CONVERGANCE AFTER7 ITERATIONS

Data fitting of the NMR titrations $(1 + ONs^{-})^{5}$

Calculations by WinEQNMR2 Version 2.00 by Michael J. Hynes Program run at 19:54:43 on 12/03/2010

Reactions: M + L = ML (beta1 = K1); M + 2L = (beta2 = K1K2) Data 2010 05 14 === Anion Recognition ===

Equilibrium constants are log₁₀ values

NO. A PARAMETER DELTA ERROR CONDITION DESCRIPTION

- 1 1 3.00538E+00 1.000E-01 9.944E-02 7.556E+02 Stability constant(K1)
- 2 1 5.56744E+00 1.000E-01 1.441E-01 2.460E+03 Stability constant(K1K2)
- 3 1 9.43506E+00 1.000E-06 2.356E-03 8.241E+00 Shift of 1
- 4 1 1.00402E+01 1.000E-04 7.674E-02 4.693E+03 Shift of 1:1_complex
- 5 1 1.02835E+01 1.000E-04 1.367E-02 1.958E+02 Shift of 1:2_complex

0RMS ERROR = 1.30E-03 MAX ERROR = 1.40E-03 AT OBS.NO. 8 RESIDUALS SQUARED = 8.42E-06 RFACTOR = 0.0092 PERCENT

NO. AEXPT. DELCALC. DELRESIDUAL% DEVWEIGHTGuest(ONs)Host(1)pH19.5730E+009.5736E+00-6.4945E-04-6.7842E-031.0000E+005.0000E-041.0000E-030.0000E+00219.6890E+009.6892E+00-1.9264E-04-1.9883E-031.0000E+001.0000E-031.0000E-030.0000E+00319.7870E+009.7863E+007.3624E-047.5226E-031.0000E+001.5000E-031.0000E-030.0000E+00419.8630E+009.8638E+00-7.5817E-04-7.6870E-031.0000E+002.0000E-031.0000E-030.0000E+00519.9780E+009.9772E+008.3923E-048.4108E-031.0000E+003.0000E+031.0000E-030.0000E+00611.0052E+011.0051E+018.6975E-048.6525E-031.0000E+004.0000E-031.0000E+030.0000E+00711.0099E+011.0100E+01-1.3247E-03-1.3117E-021.0000E+005.0000E-031.0000E+030.0000E+00811.0133E+011.0134E+01-1.3952E-03-1.3769E-021.0000E+007.0000E-031.0000E-030.0000E+00911.0160E+011.0177E+011.2846E-031.2644E-021.0000E+008.0000E-031.0000E-030.0000E+001011.0177E+011.8406E-041.8086E-031.0000E+008.0000E-031.0000E-030.0000E+00

TOLERANCE ON SUM OF SQUARES0.0100TOLERANCE ON EIGEN VALUES0.0001CONVERGANCE AFTER8 ITERATIONS

Data fitting of the NMR titrations $(1 + \text{DenS}^{-})^{5}$

Calculations by WinEQNMR2 Version 2.00 by Michael J. Hynes Program run at 19:33:52 on 12/07/2010

Reactions: M + L = ML (beta1 = K1); M + 2L = (beta2 = K1K2) Data 2010 05 14 === Anion Recognition ===

Equilibrium constants are log₁₀ values

NO. A PARAMETER DELTA ERROR CONDITION DESCRIPTION

- 1 1 3.68282E+00 1.000E-01 1.785E-01 1.444E+03 Stability constant(K1)
- 2 1 5.86840E+00 1.000E-01 3.552E-01 7.728E+03 Stability constant(K1K2)
- 3 1 9.44991E+00 1.000E-06 2.259E-03 1.498E+00 Shift of 1
- 4 1 9.79416E+00 1.000E-02 3.263E-02 1.703E+03 Shift of 1:1_complex
- 5 1 1.00910E+01 1.000E-02 3.728E-02 5.218E+02 Shift of 1:2_complex

0RMS ERROR = 1.52E-03 MAX ERROR = 2.45E-03 AT OBS.NO. 2 RESIDUALS SQUARED = 1.16E-05 RFACTOR = 0.0110 PERCENT

NO. AEXPT. DELCALC. DELRESIDUAL% DEVWEIGHTGuest(DenS)Host(1)pH19.5790E+009.5794E+00-4.2248E-04-4.4105E-031.0000E+005.0000E+041.0000E+030.0000E+00219.6780E+009.6756E+002.4452E+032.5266E+021.0000E+001.0000E+031.0000E+030.0000E+00319.7370E+009.7389E+00-1.9217E+03-1.9736E+021.0000E+001.5000E+031.0000E+030.0000E+00419.7780E+009.7786E+00-6.2180E+04-6.3591E+031.0000E+002.0000E+031.0000E+030.0000E+00519.8290E+009.8288E+002.2316E+042.2704E+031.0000E+003.0000E+031.0000E+030.0000E+00619.8620E+009.8618E+001.6499E+041.6729E+031.0000E+004.0000E+031.0000E+030.0000E+00719.8870E+009.8865E+004.7398E+044.7939E+031.0000E+005.0000E+031.0000E+030.0000E+00819.9060E+009.9062E+00-1.6022E+04-1.6174E+031.0000E+005.0000E+031.0000E+030.0000E+00919.9230E+009.9233E+006.9237E+046.9774E+031.0000E+007.0000E+031.0000E+030.0000E+00109.9358E+007.6008E+04-7.6505E+031.0000E+008.0000E+031.0000E+030.0000E+00

TOLERANCE ON SUM OF SQUARES0.0100TOLERANCE ON EIGEN VALUES0.0001CONVERGANCE AFTER7 ITERATIONS

References

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- (2) G. M. Sheldrick, Acta Cryst., 2008, A64, 112.
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- (6) M. Fukuda, R. Sekiya and R. Kuroda, Angew. Chem. Int. Ed., 2008, 47, 706.
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- (8) A. K. Rappe', K. S. Colwell and C. J. Casewit, *Inorg. Chem.*, 1993, 32, 3438.
- (9) Since bridging ligand L is slightly soluble in CH₂Cl₂, CHCl₃, and THF, small amount of **1** was decomposed and free L dissolved in the mixed solvent solutions.