Supplementary Information

Convergent heteroditopic cyclo[6]aramides as macrocyclic ion-pair receptors for constructing [2]pseudorotaxanes

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1. Materials and methods

The ¹H NMR and ¹³C NMR spectra were recorded on Bruker AVANCE AV II-400 MHz (¹H: 400 MHz; ¹³C: 100 MHz). CDCl₃, CD₃COCD₃, CD₃CN and DMSO-*d*₆ were purchased from Cambridge Isotope Laboratories, used for the titration experiments without further drying. Chemical shifts are reported in δ values in ppm using tetramethylsilane (TMS) and coupling constants (J) are denoted in Hz. Multiplicities are denoted as follows: s = singlet, d = doublet, t = triplet, dd = double doublet and m = multiplet. High resolution mass (HRMS) data were collected by WATERS Q-TOF Premier. All chemicals were obtained from commercial suppliers and were used as received unless other-wise noted. CH₂Cl₂ was dried over CaH₂. Column chromatography was carried out using silica gel (300-400 mesh). Solvents for extraction and chromatography were reagent grade. Crystallographic studies were performed on compounds **2b** and [2] pseudorotaxane **2b G1**. Data were collected on a Xcalibur E diffractometer with graphite monochromated Mo-Ka radiation (λ = 0.7107 Å).

2. Synthesis



Scheme S1. Synthetic routes for macrocycles 2.

Compounds 3a, 3b, 5a and 5b were converted into 3a', 3b', 5a' and 5b' by catalytic hydrogenation, respectively. Compounds 3a', 3b', 5a' and 5b' were used directly in the subsequent reaction without further purification.

Compounds 3a, $^{1} 3b$, $^{1} 4a$, $^{2} 4b$, $^{2} 6a^{3}$ and $6b^{3}$ were synthesized according to analogous literature procedures.

Compound **3a**: A yellow solid; ¹H NMR (400 MHz, CDCl₃, 298 K): δ = 9.73 (s, 2H), 9.20 (s, 2H), 8.86 (s, 1H), 6.41 (s, 3H), 4.12 (d, *J*=6.0 Hz, 4H), 4.05 (s, 6H), 3.83 (s, 6H), 2.01 (m, 2H), 1.45-1.24 (m, 48H), 0.85 (m, 12H); ¹³C NMR (100 MHz, CDCl₃, 298 K): δ 161.86, 160.26, 153.59, 150.95, 137.26, 131.27, 121.05, 118.41, 114.51, 96.40, 95.43, 73.23, 56.58, 56.45, 37.59, 31.88, 31.79, 31.03, 30.09, 29.72, 29.57, 29.33, 26.60, 22.64, 14.06; ESI-HRMS (m/z) calcd for C₅₆H₈₆N₄O₁₂ [M+H]⁺ 1007.6320, [M+Na]⁺ 1029.6140, found [M+H]⁺ 1007.6320, [M+Na]⁺ 1029.6033.

Compound **5a**: Trimer **3a** (500 mg, 0.50 mmol) was hydrogenated in the presence of 20% Pd/C (100 mg) in CHCl₃/CH₃OH (60 mL, v/v=5:1) for 10 h at 40 °C. The

solution was filtered in darkness as fast as possible followed by immediate removal of the solvent. The reduced diamine was used for the immediate coupling reaction. DMF (5 uL) was added to a suspension of compound 4a (416 mg, 1.02 mmol) and oxalyl chloride (388 mg, 3.06 mmol) in CH₂Cl₂. The mixture was stirred for 40 min at room temperature. The solvent was evaporated and the resulting acid chloride was dried in vacuum at room temperature for 30 min to get compound 4a'. Compound 4a' was dissolved in CH₂Cl₂ (60 mL) and added dropwise to a mixture of the above 3a' and Et₃N (200 mg, 1.98 mmol) in CH₂Cl₂ (20 mL) at room temperature. The solution was stirred under N₂ for 7 h. The organic layer was washed with water (20 mL \times 3) and dried over anhydrous Na₂SO₄. Concentration under reduced pressure afforded a vellow solid which was triturated with ethyl acetate and filtered to give pentamer 5a (626 mg, 83.4%) as a yellow solid. ¹H NMR (400 MHz, CDCl₃, 298 K): $\delta = 9.63$ (s, 2H), 9.57 (s, 2H), 9.25 (s, 2H), 9.21 (s, 1H), 9.18 (d, J = 2.8 Hz, 2H), 8.21 (q, $J_1 = 2.8$ Hz, $J_2 = 9.2$ Hz, 2H), 7.09 (d, J = 9.2 Hz, 2H), 6.54 (s, 1H), 6.48 (s, 2H), 4.21 (d, J =5.6 Hz, 4H), 4.12 (d, J=6.2 Hz, 4H), 3.87 (s, 12H), 1.98 (m, 4H), 1.53-1.22 (m, 64H), 0.94-0.84 (m, 24H); ¹³C NMR (100 MHz, CDCl₃, 298 K): δ 162.09, 161.28, 160.51, 160.10, 146.93, 146.51, 141.58, 128.74, 127.73, 123.22, 120.67, 119.59, 117.46, 115.86, 112.70, 96.59, 74.81, 73.30, 72.83, 55.85, 38.61, 37.67, 31.79, 31.08, 30.00, 29.67, 29.59, 29.32, 28.76, 26.71, 23.45, 22.95, 22.64, 14.08, 13.99, 10.62; ESI-HRMS (m/z) calcd for $C_{86}H_{128}N_6O_{16}$ [M+H]⁺ 1501.9465, [M+Na]⁺ 1523.9285, found [M+H]⁺ 1501.9465, [M+Na]⁺ 1523.9282.

Compound 2a: Pentamer 5a (400 mg, 0.27 mmol) was hydrogenated in the presence of 20% Pd/C (80 mg) in CHCl₃/CH₃OH (100 mL, v/v = 5:1) for 10 h at 40 °C. The solution was filtered in darkness as fast as possible followed by immediate removal of the solvent. The reduced diamine was used for the immediate coupling reaction. DMF (5 uL) was added to a suspension of compound **6a** (82 mg, 0.28 mmol) and oxalyl chloride (105 mg, 0.84 mmol) in CH₂Cl₂. The mixture was stirred for 40 min at room temperature. The solvent was evaporated and the resulting acid chloride was dried in vacuum at room temperature for 30 min to get compound 6a'. Compound 6a' was dissolved in CH₂Cl₂ (60 mL) and added dropwise to a mixture of the above 5a' and Et₃N (162 mg, 1.60 mmol) in CH₂Cl₂ (20 mL) at 0 °C. The solution was stirred under N_2 for 7 h. The organic layer was washed with water (20 mL \times 3) and dried over anhydrous Na₂SO₄ and filtered. The crude product was purified by chromatography on silica gel (CH₂Cl₂/MeOH = 20: 1) to provide the product 2a as a white solid (339) mg, 74.6%). ¹H NMR (400 MHz, CDCl₃, 298 K): $\delta = 10.20$ (s, 2H), 9.42 (s, 2H), 9.39 (s, 2H), 9.16 (s, 2H), 9.15 (s, 1H), 8.51 (d, J = 6.2 Hz, 2H), 8.23 (s, 1H), 8.19 (s, 2H), 7.75 (s, 2H), 7.01 (d, J = 6.2 Hz, 2H), 6.50 (s, 2H), 6.49 (s, 1H), 4.06 (m, 10H), 3.89 (s, 6H), 3.88 (s, 6H), 2.00 (m, 4H), 1.77 (m, 1H), 1.53-1.26 (m, 72H), 0.92-0.85 (m, 30H); ¹³C NMR (100 MHz, CDCl₃, 298 K): δ 164.86, 163.45, 162.54, 159.98, 159.73, 153.26, 145.94, 135.55, 132.77, 125.17, 123.74, 121.86, 120.80, 119.48, 118.02, 117.61, 112.68, 94.73, 72.51, 55.92, 55.51, 39.37, 38.66, 37.81, 31.89, 30.91, 30.45, 30.03, 29.90, 29.72, 29.59, 29.34, 29.08, 28.74, 26.63, 23.80, 23.29, 23.07, 22.64, 14.10, 11.13, 10.41. ESI-HRMS (m/z) calcd for $C_{102}H_{150}N_6O_{15}$ [M+Na]⁺ 1723.1090,

found [M+Na]⁺ 1723.1082.

Compound **2b**: Prepared according to the same method as for **2a**. ¹H NMR (400 MHz, CDCl₃, 298 K, containing 1.5 equiv Et₂NH₂Cl): δ =10.80 (s, 2H), 10.50 (s, 2H), 9.76 (s, 2H), 9.35 (s, 2H), 9.26 (s, 1H), 8.99 (s, 1H), 8.72 (d, *J*=9.2 Hz, 2H), 8.63 (s, 2H), 7.80 (s, 2H), 7.05 (d, *J*=9.2 Hz, 2H), 6.57 (s, 2H), 6.53 (s, 1H), 4.26 (m, 8H), 4.11 (t, *J*=6.3 Hz, 2H), 3.94 (s, 12H), 1.97 (m, 8H), 1.89 (m, 2H), 1.71 (m, 5H), 1.07 (d, *J*=5.1 Hz, 12H), 1.01 (d, *J*=5.6 Hz, 12H), 0.96 (d, *J*=6.4 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃ + 40% CD₃OD, 298 K): δ 166.69, 166.08, 163.96, 163.15, 156.55, 148.75, 142.95, 142.35, 138.30, 136.18, 135.06, 132.62, 127.74, 123.60, 122.57, 117.09, 115.08, 96.93, 72.31, 58.84, 42.08, 41.69, 41.19, 35.78, 33.45, 33.15, 29.15, 26.60, 26.38, 17.64, 14.57; ESI-HRMS (m/z) calcd for C₇₁H₈₈N₆O₁₅ [M+H]⁺ 1265.6386, found [M+H]⁺ 1265.6390.

3. ¹H NMR and ¹³C NMR spectra of new compounds







Figure S2. ¹³C NMR spectrum of **3a** (100 MHz, CDCl₃, 298 K).



Figure S3. ¹H NMR spectrum of **5a** (400 MHz, CDCl₃, 298 K).



Figure S4. ¹³C NMR spectrum of **5a** (100 MHz, CDCl₃, 298 K).



Figure S5. ¹H NMR spectrum of **2a** (400 MHz, CDCl₃, 298 K).



Figure S6. ¹³C NMR spectrum of **2a** (100 MHz, CDCl₃, 298 K).



Figure S7. ¹H NMR spectrum of **2b** (400 MHz, CDCl₃, 298 K, containing 1.5 equiv Et₂NH₂Cl).



Figure S8. ¹³C NMR spectrum of **2b** (100 MHz, CDCl₃+40% CD₃OD, 298 K).

4. Concentration-dependent ¹H NMR spectra of **2a**



Figure S9. Stacked partial ¹H NMR spectra (400 MHz, acetone- d_6 , 298 K) of **2a** at 1.5 mM (a), 1.0 mM (b), 0.7 mM (c), 0.5 mM (d) and 0.3 mM (e).



Figure S10. Stacked partial ¹H NMR spectra (400 MHz, $CDCl_3$, 298 K) of **2a** at 2.0 mM (a), 1.5 mM (b), 1.0 mM (c), 0.7 mM (d), 0.5 mM (e) and 0.3 mM (f).

5. HR ESI-MS for complex



Figure S11. The HRESI MS spectrum of an equimolar solution of **2a** and **G1** in methanol in the positive ion mode.



Figure S12. The HRESI MS spectrum of an equimolar solution of **2a** and **G1** in methanol in the negative ion mode.



Figure S13. The HRESI MS spectrum of an equimolar solution of **2a** and TBACl in methanol in the negative ion mode.



Figure S14. The HRESI MS spectrum of an equimolar solution of **2b** and **G1** in methanol in the positive ion mode.



Figure S15. The HRESI MS spectrum of an equimolar solution of **2b** and **G1** in methanol in the negative ion mode.



Figure S16. The HRESI MS spectrum of an equimolar solution of **2b** and TBACl in methanol in the negative ion mode.

6. 2D ROESY spectra





Figure S17. Expanded 2D ROESY spectra of 2a (10 mM, 2a:G1=1:1) (CDCl₃, 400 MHz, 298 K).



Figure S18. Expanded 2D ROESY spectra of **2a** and **G1** (10 mM, **2a**:**G1**=1:1) (CDCl₃, 400 MHz, 298 K).



Figure S19. Stacked plots of ¹H NMR spectra of compound **G1** (1.0 mM) with cyclo[6]aramide **2a** at different concentration in CDCl₃ (400MHz, 298K).



Figure S20. Mole ratio plot for the complexation between 2a and G1, indicating a 1:1 stoichiometry.



Figure S21. Determination of the binding constant of **2a** G1 in $CDCl_3$ at 298 K. Fitting result based on H_1 of compound G1.

Protons	2a	2a+G1	
PIOLOIIS	δ_{f}	δ_b	$\Delta \delta = \delta_b - \delta_f$
H _b	9.18	9.32	0.14
H _c	9.18	9.23	0.05
H _d	9.45	9.63	0.18
\mathbf{H}_{f}	10.25	10.42	0.17
\mathbf{H}_{g}	8.17	8.58	0.41
H_{i}	8.53	8.70	0.17
\mathbf{H}_{j}	9.18	10.82	1.64
H_k	8.26	8.96	0.70

Table S1. The chemical shifts δ (ppm) for the 1:1 solution of **2a** and **G1** in CDCl₃.



Figure S22. Stacked plots of ¹H NMR spectra of compound **G1** (1.0 mM) with cyclo[6]aramide **2a** at different concentration in CDCl₃/DMSO- d_6 (v/v = 17:3)(400MHz, 298K).



Figure S23. Determination of the binding constant of **2a** G1 in CDCl₃/DMSO- d_6 (v/v = 17:3) at 298 K. Fitting result based on H₁ of compound G1.



Figure S24. Determination of the binding constant of **2a** G2 in $CDCl_3$ at 298 K. Fitting result based on H_2 of compound G2.





Figure S25. Determination of the binding constant of **2a** G**3** in CDCl₃ at 298 K. Fitting result based on H_3 of compound G**3**.





Figure S26. Determination of the binding constant of **2a** G4 in $CDCl_3/CD_3CN$ (v/v = 3:2) at 298 K. Fitting result based on H₄ of compound G4.



Figure S27. Determination of the binding constant of **2a** G**4** in CDCl₃ at 298 K. Fitting result based on H_5 of compound G**4**.

Table S2. Association constants $(K_a/M^{-1})^a$ for complexation of host **2a** and guest (**G1**, **G2**, **G3** and **G4**) at 298 K.

- /				
Guest	Solvent	2a		
G1	CDCl ₃	$(1.8 \pm 0.8) \times 10^5$		
G1	CDCl ₃ /DMSO- <i>d</i> ₆ (17:3, v/v)	$(2.9 \pm 0.4) \times 10^2$		
G2	CDCl ₃	$(1.2 \pm 0.8) \times 10^4$		
G3	CDCl ₃	$(5.5 \pm 1.7) \times 10^3$		
G4	CDCl ₃	$(2.2 \pm 1.1) \times 10^5$		
G4	CD ₃ CN/CDCl ₃ (2:3, v/v)	$(2.3 \pm 0.9) \times 10^4$		

^{*a*} The association constant K_a values were obtained by ¹H NMR titration.



Figure S28. Stacked plots of ¹H NMR spectra of compound **G1** (1.0 mM) with cyclo[6]aramide **1** at different concentration in CDCl₃ (400MHz, 298K).



Figure S29. Mole ratio plot for the complexation between 1 and G1, indicating a 1:1 stoichiometry.



Figure S30. Determination of the binding constant of **1** G**1** in $CDCl_3$ at 298 K. Fitting result based on H₁ of compound G**1**.

12. X-ray single crystal structures of **2b** and

[2]pseudorotaxane 2b G1



Figure S31. Section of the crystal packing of **2b**: (a) top and (b) side views.



Figure S32. Space-filling packing of the solid-state structure for [2]pseudorotaxane **2b** G1: (a) top and (b) side views.

Identification code	2b	2b G1
CCDC	976842	976751
Empirical formula	$C_{71}H_{88}N_6O_{15}$	$C_{81}H_{114}Cl_5N_7O_{16}\\$
Formula weight	1265.47	1619.04
Temperature/K	143.00(10)	143.00(10)
Crystal system	triclinic	monoclinic
Space group	P-1	$P2_1/n$
a/Å	15.0617(6)	15.3283(4)
b/Å	15.8532(5)	24.6571(4)
c/Å	20.3366(6)	23.0091(5)
α/°	86.099(3)	90
β/°	78.810(3)	107.044(2)
$\gamma/^{\circ}$	67.459(4)	90
Volume/Å ³	4399.6(3)	8314.4(3)
Z	2	4
$\rho_{calc}mg/mm^3$	0.955	1.293
μ (Mo K α) /mm ⁻¹	0.067	0.243
F(000)	1352.0	3448.0
Crystal size/mm ³	$0.40\times 0.35\times 0.20$	$0.3 \times 0.3 \times 0.2$
2Θ range for data collection	5.8 to 52.74 $^\circ$	5.796 to 52.744 °
Index ranges	$-18 \le h \le 16, -19 \le k \le 19,$	-16 \leq h \leq 19, -18 \leq k \leq 30, -24 \leq
	$-25 \le l \le 25$	$l \le 28$
Reflections collected	36605	35769
Independent reflections	17942[R(int) = 0.0290]	16968[R(int) = 0.0212]
Data/restraints/parameters	17942/2/852	16968/0/1001
Goodness-of-fit on F ²	0.946	1.067
Final R indexes [I>=2 σ (I)]	$R^1 = 0.0848, wR^2 = 0.2359$	$R^1 = 0.0672, wR^2 = 0.1722$
Final R indexes [all data]	$R^1 = 0.1306, wR^2 = 0.2638$	$R^1 = 0.0905, wR^2 = 0.1869$
Largest diff. peak/hole / e Å $^{\text{-}3}$	1.43/-0.40	1.30/-1.35

Table S3. Crystallographic data for macrocycle 2b and [2]pseudorotaxane 2b G1

13. General description of *ab initio* molecular modeling

All the calculations were carried out by utilizing the Gaussian 09 program package.⁴ The geometry optimizations were performed at the density functional theory (DFT) level, and the Becke's three parameter hybrid functional with the Lee-Yang-Parr correlation functional $(B3LYP)^5$ method was employed to do the calculations. The 6-31G(d,p)⁶ basic from the Gaussian basis set library has been used in all the structural optimizations. The harmonic vibrational frequencies and

zero-point energy corrections were calculated at the same level of theory. Single point energy were obtained at the B3LYP level in conjunction with the 6-311+G (2d, p) basis set with the use of the above optimized geometries, i.e., B3LYP/6-311+G(2d,p)//B3LYP/6-31G(d,p). Number of imaginary frequencies is zero.

14. References

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