Supplementary Information-1

Stereoselective Synthesis of 4-Substituted-Cyclic Sulfamidate-5-Carboxylates By Asymmetric Transfer Hydrogenation Accompanying Dynamic Kinetic Resolution and Its Use in Concise Stereoselective Synthesis of (-)-*epi*-Cytoxazone and Taxotere Side-Chain.

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General

All commercial reagents were used as obtained commercially unless otherwise noted. Reactions were performed using oven dried glassware under an atmosphere of nitrogen. Dichloromethane (DCM), ether, THF were dried and purified using a solvent purification system. Flash column chromatography was carried out on Fuji Chromatorex silica gel (38-75 μ m). Analytical thin layer chromatography (TLC) was performed on Merck silica gel 60 F₂₅₄ plates. Preparative thin layer chromatography (PLC) was performed on Merck silica gel 60 F₂₅₄ 2mm plates. Visualization of the developed chromatogram was accomplished with UV light and by staining with ethanolic phosphomolybdic acid (PMA) solution or ninhydrin solution followed by heating.

Nuclear magnetic resonance (NMR) spectra were recorded using Bruker 500 MHz NMR instrument (¹H NMR at 500 MHz and ¹³C NMR at 125 MHz) or Bruker 300 MHz NMR instrument (¹H NMR at 300 MHz and ¹³C NMR at 75 MHz). ¹H NMR data are reported as follows: chemical shift (δ , ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad), integration, coupling constants (Hz). Data for ¹³C NMR are reported in terms of chemical shift (δ , ppm). High performance liquid chromatography (HPLC) was carried out on a Young Lin HPLC system (7725i Injector, SDV 30 Plus Solvent Degassor & Valve Module (Helium Sparging), SP930D Solvent Delivery Pump, UV 730D Absorbance Detector) equipped with a Chiralpak IB or Chiralpak AD-H column or an Aglient 1100 Series HPLC equipped with Chiralpak IB or Chiralpak IC column. Specific rotations were measured on a Rudolph Autopol IV (Automatic polarimeter). High-resolution mass spectra and elemental analysis were obtained from the Korea Research Institute of Chemical Technology (EI) or Korea Basic Science Institute (ESI). HR-MS were measured with electron impact (EI) ionization via double focusing mass analyzer (magnetic and electric fields) or electrospray ionization (ESI) via time of flight (TOF) analyzer.

The formic acid/triethylamine mixtures (molar ratio = 5/2 or 1:1) are commercially available. (*R*,*R*)-**1a**¹ and (*R*,*R*)-**1b**² were prepared according to the literature procedures. Chiral catalysts, (*R*,*R*)-**1c**, (*R*,*R*)-**1d**, and (*R*,*R*)-**1e** are commercially available.

¹ Mao, J. M.; Baker, D. C. Org. Lett. 1999, 1, 841.

² K. Mashima; T. Abe; K. Tani, *Chem. Lett.* 1998, 1199.

1. Optimization of the ATH-DKR reaction of 6

1-1. Optimization of ATH-DKR reaction of 6a with various catalysts

	0, 0 N S Ph CO ₂ Me 6a	cat -1 (0.5 HCO ₂ H/E EtOAc(0.1M),	mol%) t ₃ N(5:2) 25°C, 0.5 h	$0 0 0$ $HN_{5}^{S} 0$ 4^{-5}^{5} Ph $CO_{2}N$ 7a	1e
Entry	Cat.1	Convn (%) ^b	dr (syn:anti)	ee(%) ^d	config ^e
1	(<i>R</i> , <i>R</i>)- 1a	>99	>25:1 [°]	98	<i>S</i> , <i>S</i>
2	(<i>R</i> , <i>R</i>)- 1b	>99	>25:1 ^c	30	<i>S</i> , <i>S</i>
3	(<i>R</i> , <i>R</i>)-1c	13	-	95	S, S
4	(<i>R</i> , <i>R</i>)-1d	6	-	-	-
5	(<i>R</i> , <i>R</i>)- 1e	17	-	83	<i>S</i> , <i>S</i>

Table S1. Optimization of chiral catalysts 1a-e in ATH-DKR of 6a^a



^aReaction conditions: **6a** (0.5 mmol), cat-**1** (0.5 mol%), HCO₂H/Et₃N (5:2, 0.5 ml), EtOAc (5 mL), rt. ^bDetermined by ¹H NMR analysis of the crude products. ^cOnly 4,5-*cis* products were detected in ¹H NMR of crude products. ^dDetermined by chiral HPLC. ^eSee, Scheme S-1 below

1-2. Optimization of ATH-DKR reaction of 6a in various solvents

Table S2. Optimization of solvent effect in ATH-DKR of 3^{a}

O N 4 Ph 6a	O O 5 CO ₂ Me So (<i>R,R</i> HCO ₂ H/E	olvent (0.1M))-1a (0.5 mol%) t ₃ N(5:2), 25ºC, 0.5 h	$0, 0$ $HN^{5} 0$ $4^{4} 5^{5}$ Ph CO ₂ Me 7a
Entry	Solvent	Convn (%) ^b	ee (%) ^c
1	EtOAc	>99	98.1
2	CH_2Cl_2	>99	98.9
3	Cl(CH ₂) ₂ Cl	>99	98.1
4	CHCl ₃	>99	84.1
5	Toluene	>99	85.1
6	DMF	>99	88.1
7	MeOH	>99	97.1
8	THF	96	90.9
9	2-Propanol	>99	96.8

^aReaction conditions: **6a** (0.5 mmol), (*R*,*R*)-**1a** (0.5 mol%), HCO₂H/Et₃N (5:2, 0.5 ml), in 5.0 mL of solvent at rt. ^bDetermined by ¹H NMR analysis of the crude products. ^cDetermined by chiral HPLC of the crude products.

2. General procedure for the synthesis of α -hydroxy- β -keto esters from β -keto-ester

[Method A]

$$R_{1} \xrightarrow{O} OR_{2} + TsN_{3} \xrightarrow{K_{2}CO_{3}} R_{1} \xrightarrow{O} OR_{2} \xrightarrow{Rh_{2}(OAc)_{4}} R_{1} \xrightarrow{O} OR_{2} \xrightarrow{Rh_{2}(OAc)_{4}} R_{1} \xrightarrow{O} OR_{2}$$

4-4-1.³

To a mixture of β -keto-ester (1.0 eq.) and potassium carbonate (1.25 eq.) in acetonitrile cooled in a water ice-bath, was added dropwise with stirring a solution of tosyl azide (1.25 eq) in acetonitrile. The reaction mixture was stirred at room temperature. The disappearance of

³ Leost, F.; Chantegrel B.; Deshayes C., *Tetrahedron*, 1997, **5**, 7557.

ester was monitored by TLC. Potassium carbonate was filtered off and the filterate was evaporated in *vacuo* to afford a residue which was purified by silica-gel chromatography to give the diazo compund.

Step-2⁴

A solution of the diazo compound (1.0 eq) and $Rh_2(OAc)_4$ (0.03 eq) in THF- H_2O (2:1 ratio) was refluxed overnight and allowed to cool to room temperature. The mixture was concentrated in *vacuo* and the aqueous residue was extracted with EtOAc (x3). The combined organic layers were washed with water and brine, dried over anhydrous MgSO₄, filtered, and evaporated to dryness. The residue was purified by silica-gel chromatography to give the α -hydroxy β -ketocarbonates.

[Method B]



Step-1

To a solution of **1** (4 g, 21.7 mmol) and PhI(OAc)₂ (7 g, 21.7 mmol) in dichloromethane (50 mL) was added BF₃·OEt₂ (1.4 mL, 10.85 mmol). The reaction mixture was stirred at room temperature for 20 min and then quenched by NaHCO₃. The layers were separated and the aqueous layer was extracted with dichloromethane(x3). The combined organic layers were washed with brine, dried over anhydrous MgSO₄, and evaporated. The residue was purified by silica-gel chromatography (Hexane/EtOAc=5:1) to give **2** (4.96 g, 94.4% yield).

Step-2

⁴ Sun, C.-Q.; Cheng, P. T. W.; Stevenson, J.; Dejneka, T.; Brown, B.; Wang, T. C.; Robal, J. A.; Poss, M. A., *Tetrahedron Lett.*, 2002, **43**, 1161.

A solution of **2** (1.3 g, 5.4 mmol) in anhydrous MeOH (10 mL) was added dropwise to a solution of KCN (175 mg, 2.7 mmol) in anhydrous MeOH (40 mL) at 0 °C. The resulting mixture was stirred at 0 °C for 2 h. After removal of the solvent, the residue was dissolved in CH_2Cl_2 and then washed with brine. The combined organic layers were dried over anhydrous MgSO₄, filtered, and concentrated. The residue was purified by silica-gel chromatography (Hexane/EtOAc, 5:1) to give **3** (0.815 g, 75.4% yield).

[Method C]⁵

$$R_1 \longrightarrow OR_2^+ Phl(OCOCF_3)_2 \longrightarrow R_1 \longrightarrow OR_2^+ OR_2^+$$

To a suspension of β -keto-ester in H₂O was added PIFA (2.0 eq) portionwise for 10 minutes. The reaction mixture was stirred at room temperature until TLC indicated the total consumption of the β -keto-ester. Then the reaction mixture was treated with saturated NaHCO₃ (aq) and extracted with EtOAc (x3). The combined organic layers were washed with water and brine, dried over anhydrous MgSO₄, filtered, and concentrated. The residue was purified by silica-gel chromatography to give the α -hydroxy- β -keto ester.

3-Phenyl-2-hydroxy-3-oxo-propionic acid methyl ester⁶ [Method A]

yield: 89.3% (1.9 g as a yellow oil); ¹H NMR (500 MHz, CDCl₃) δ 8.04-8.05 (m, 2H), 7.59-7.62 (m, 1H), 7.45-7.48 (m, 2H), 5.60 (s, 1H), 4.42-4.49 (brs, 1H), 3.68 (s, 3H).; ¹³C NMR (75 MHz, CDCl₃) δ 193.8, 169.1, 134.7, 133.0, 129.5, 128.9, 74.3, 53.1.; HRMS (EI): m/z calcd for C₁₀H₁₀O₄ 194.0579, found 194.0542.

3-(2-Tolyl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method C]



The ptoduct was unstable in silica-gel chloromatography and used next step without further purification. HRMS (EI): m/z calcd for $C_{11}H_{12}O_4$

⁵ Wang, J.; Yuan, Y.; Xiong, R.; Zhang-Negrerie, D.; Du, Y.; Zhano, K. Org. Lett. 2012, **14**, 2210.

⁶ (a) Scholte, Andrew A.; An, M. H.; Snapper, Marc L. Org. Lett. 2006, 8, 4759. (b) Plietker, B. J. Org. Chem. 2004, 69, 8287.

208.0736, found 208.0737.

3-(3-Tolyl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method B]

129.8, 128.7, 126.7, 74.3, 53.3, 21.3.; HRMS (EI): m/z calcd for C₁₁H₁₂O₄ 208.0736, found 208.0732.

3-(4-Tolyl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method C]

yield: 84.8% (84.8 mg as a yellow oil); ¹H NMR (500 MHz, CDCl₃) δ 7.98 (d, 2H, J = 8.3 Hz), 7.30 (d, 2H, J = 8.2 Hz), 5.59 (s, 1H), 4.33-4.37 (brs, 1H), 3.71 (s, 3H), 2.43 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 193.1, 169.2, 146.1, 130.4, 129.6, 129.6, 74.2, 53.0, 21.9.; HRMS (EI): m/z calcd for C₁₁H₁₂O₄ 208.0736, found 208.0733.

3-(3-Chloro-phenyl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method B]



yield: 41.7% (0.11 g as a yellow oil); ¹H NMR (500 MHz, CDCl₃) δ 8.06 (s, 1H), 7.97 (d, 1H, J = 7.8 Hz), 7.62 (m, 1H), 7.47 (m, 1H),
5.57 (s, 1H), 3.76 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 192.7, 168.8, 135.3, 134.6, 134.6, 130.2, 129.4, 127.6, 74.5, 53.3.; HRMS

(EI): m/z calcd for $C_{10}H_9ClO_4$ 228.0189, found 228.0177.

3-(4-Chloro-phenyl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method A]



yield: 35.7% (0.24 g as a yellow oil); ¹H NMR (500 MHz, CDCl₃) δ 8.02 (d, 2H, J = 8.3 Hz), 7.48 (d, 2H, J = 8.3 Hz), 5.56 (s, 1H), 4.21 (brs, 1H), 3.73 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 192.6, 168.9, 141.4, 131.3, 130.9, 129.3, 74.4, 53.2.; HRMS (EI): m/z

calcd for C₁₀H₉ClO₄ 228.0189, found 228.0183.

3-(4-Methoxy-phenyl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method A]



yield: 78% (5.6 g as a yellow oil); ¹H NMR (500 MHz, CDCl₃) δ 8.07 (d, 2H, J = 8.5 Hz), 6.97 (d, 2H, J = 8.5 Hz), 5.55 (d, 1H, J = 5.6 Hz), 4.35 (d, 1H, J = 7.1 Hz), 3.88 (s, 3H), 3.71 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 191.7, 169.3, 164.9, 132.0, 125.7, 114.2,

74.1, 55.6, 53.0.; HRMS (EI): m/z calcd for $C_{11}H_{12}O_5$ 224.0685, found 224.0673.

3-(4-Fluoro-phenyl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method C]

ÔН

yield: 66% (0.66 g as a pale yellow oil); ¹H NMR (500 MHz, CDCl₃) δ 8.13 (m, 2H), 7.13 (m, 2H), 5.38 (s, 1H), 3.88 (s, 3H).; ¹³C NMR $(75 \text{ MHz}, \text{CDCl}_3) \delta 196.5, 167.3, 166.3 (q, J_{CF}= 248.7 \text{ Hz}), 133.1 (d, J_{CF}= 248.7$ J_{CF} = 10.0 Hz), 130.2 (d, J_{CF} = 3.4 Hz), 115.8 (d, J_{CF} = 21.7 Hz), 85.3, 53.9.; HRMS (EI): m/z

calcd for C₁₀H₉FO₄ 212.0485, found 212.0456.

3-(4-Trifluoromethyl-phenyl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method A]



yield: 61% (0.77 g as a pale yellow oil); ¹H NMR (500 MHz, CDCl₃) δ 8.20 (d, 2H, J = 7.9 Hz), 7.78 (d, 2H, J = 8.1 Hz), 5.65 (s, 1H), 3.76 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 193.2, 168.9, 135.4 (q, *J_{CF}*= 32.6 Hz), 130.5, 129.8, 125.9, 125.5 (q, *J_{CF}* = 271.2

Hz), 74.7, 53.3.; HRMS (EI): m/z calcd for C₁₁H₉F₃O₄ 262.0453, found 262.0441.

3-(4-Cyano-phenyl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method A]



yield: 49.6% (0.93 g as a yellow solid), mp = 100.4-102.6 °C, 1 H NMR (500 MHz, CDCl₃) δ 8.17 (d, 2H, J = 8.0 Hz), 7.82 (d, 2H, J= 8.0 Hz), 5.59 (s, 1H), 4.09-4.17 (brs, 1H), 3.76 (s, 3H).; ^{13}C NMR (125 MHz, CDCl₃) δ 192.9, 168.7, 136.2, 132.6, 129.9,

117.7, 117.6, 74.7, 53.4.; HRMS (EI): m/z calcd for C₁₁H₉NO₄ 219.0532, found 219.0528.

3-(4-Methoxycarbonyl-phenyl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method A]

yield: 42% (0.71 g as a pale yellow oil); ¹H NMR (500 MHz, CDCl₃) δ 8.06-8.11 (m, 4H), 5.60 (s, 1H), 3.91 (s, 3H), 3.68 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 193.6, 168.9, 165.9, MeO₂C

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136.4, 135.0, 129.9, 129.3, 74.6, 53.2, 52.6.; HRMS (EI): m/z calcd for $C_{12}H_{12}O_6$ 252.0634, found 252.0632.

3-(Naphthalen-2-yl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method A]

yield: 59% (1.64 g as a yellow oil).; ¹H NMR (500 MHz, CDCl₃) δ 8.68 (s, 1H), 8.11 (d, 1H, J = 8.6 Hz), 8.03 (d, 1H, J = 8.2 Hz), 7.90-7.95 (m, 2H), 7.67 (t, 1H, J = 7.1 Hz), 7.61 (t, 1H, J = 7.4 Hz), 5.81 (s, 1H), 4.46 (brs, 1H), 3.75 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 193.6, 169.2, 136.3, 132.3, 132.2, 130.3, 130.0, 129.5, 128.8, 127.9, 127.2, 124.2, 74.4, 53.1.; HRMS (EI): m/z calcd for C₁₄H₁₂O₄ 244.0736, found 244.0729.

3-(Furan-2-yl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method B]

yield: 69.3% (0.95 g as a yellow oil); ¹H NMR (500 MHz, CDCl₃) δ 7.71-7.72 (m, 1H), 7.49 (d, 1H, J = 3.5 Hz), 6.62-6.63 (m, 1H), 5.35 (d, 1H, J = 8.3 Hz), 4.01 (d, 1H, J = 8.3 Hz), 3.78 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 181.8, 169.1, 149.8, 148.5, 121.4, 113.1, 74.3, 53.3.; HRMS (EI): m/z calcd for C₈H₈O₅ 184.0372, found 184.0353.

3-(Thiophen-2-yl)-2-hydroxy-3-oxo-propionic acid methyl ester [Method B]



yield: 75.4% (0.81 g as a brown oil); ¹H NMR (500 MHz, CDCl₃) δ
8.01 (d, 1H, , J = 3.6 Hz), 7.79 (d, 1H, J = 4.9 Hz), 7.19 (t, 1H, J = 4.4
e Hz), 5.42 (s, 1H), 4.22 (brs, 1H), 3.76 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 186.1, 169.2, 139.4, 136.4, 135.3, 128.7, 75.1. 53.3.; HRMS

(EI): m/z calcd for $C_8H_8O_4S$ 200.0143, found 200.0139.

2-Hydroxy-3-oxo-hexanoic acid methyl ester [Method A]

yield: 80.1% (1.36 g as a coloress oil); ¹H NMR (300 MHz, CDCl₃) δ 4.77 (s, 1H), 4.32 (brs, 1H), 3.80 (s, 3H), 2.05-2.78 (m, 2H), 1.57-1.69 (m, 2H), 0.90 (t, 3H, J = 7.4 Hz).; ¹³C NMR (75 MHz, CDCl₃) δ 204.4,

169.0, 76.7, 53.2, 40.6, 17.0, 13.6.; HRMS (EI): m/z calcd for $C_7H_{12}O_4$ 160.0736, found 160.0728.

2-Hydroxy-3-oxo-5-phenyl-pentanoic acid methyl ester [Method A]



126.4, 77.9, 53.2, 40.4, 29.3.; HRMS (EI): m/z calcd for C₁₂H₁₄O₄ 222.0892, found 222.0888.

3-Cyclohexyl-2-hydroxy-3-oxo-propionic acid methyl ester⁷ [Method A]

yield: 78.2% (2.18 g as a pale yellow oil); ¹H NMR (500 MHz, CDCl₃) δ 5.24 (s, 1H), 3.71 (s, 3H), 3.05-3.10 (m, 1H), 1.63-1.87 (m, 5H), 1.17-1.36 (m, 6H).; ¹³C NMR (125 MHz, CDCl₃) δ 212.1, 168.7, 84.0, 53.3, 46.5, 29.0, 28.6, 25.7.; HRMS (EI): m/z calcd for C₁₀H₁₆O₄ 200.1049, found 200.1040.

3. General procedure for the synthesis of cyclic imine 6 from α-hydroxy-β-keto-ester.^{8,9}



To the solution of 3-phenyl-2-hydroxy-3-oxo-propionic acid methyl ester (1.85 g, 9.53 mmol) in DMA (*N*,*N*-dimethyl acetamide, 18 mL) was added sulfamoyl chloride (2.2 g, 19 mmol) at 0 \degree . The reaction mixture was stirred at room temperature for 1.5 h and diluted with EtOAc (30 mL). The reaction mixture washed with brine and the organic layer was dried over anhydrous MgSO₄ and the solvent was evaporated under reduced pressure. The residue was re-dissolved in toluene (15 mL) and catalytic amount of PTSA (*p*-toluenesulfonic acid) was added. The reaction mixture was heated for 1 h at 110 \degree and cooled to room temperature. The solvent was removed and the reaction mixture was diluted with EtOAc (30 mL) and

⁷ Plietker, B. J. Org. Chem. 2004, **69**, 8287.

⁸ Lee, H.-K.; Kang, S.; Choi, E. B., J. Org. Chem., 2012, 77, 5454.

⁹ Han, J. A.; Kang, S. Y.; Lee, H-K. Chem. Commun. 2011, **47**, 4004.

washed brine. The organic layers were dried over anhydrous $MgSO_4$ and the solvent was evaporated under reduced pressure. The residue was recrystallized from EtOAc/Hexane, to give the desired imine, as white crystals **6a**.

Methyl 2,2-dioxo-4-phenyl-5H-1,2,3-oxathiazole-5-carboxylate, 6a



yield: 74.8% (1.81g as a white solid), mp = 149.5-151.5 °C, ¹H NMR (300 MHz, CDCl₃) δ 8.09 (d, 2H, *J* = 7.6 Hz), 7.77 (t, 1H, *J* = 7.5 Hz), 7.58 (t, 2H, *J* = 7.8 Hz), 6.19 (s, 1H), 3.82 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 171.7, 163.5, 136.3, 130.3, 129.6, 126.3, 83.7, 54.4.; HRMS

(EI): m/z calcd for $C_{10}H_9NO_5S$ 255.0201, found 255.0233.

Isopropyl 2,2-dioxo-4-phenyl-5H-1,2,3-oxathiazole-5-carboxylate, 6b



Yield: 53%; ¹H NMR (300 MHz, CDCl₃) δ 8.08 (d, 2H, *J* = 7.8), 7.77-7.72 (t, 1H, *J* = 7.4), 7.62-7.55 (t, 2H, *J* = 7.8), 6.13 (s, 1H), 5.05 (m, 1H), 1.18 (d, 6H, *J* = 6.21); ¹³C NMR (500 MHz, CDCl₃) δ 172.0, 162.3, 136.1, 130.3, 129.4, 126.5, 84.1, 72.6, 21.3, 21. 2; HRMS (EI): m/z calcd for C₁₂H₁₃NO₅S 283.0514, found 283.0503.

Benzyl 2,2-dioxo-4-phenyl-5H-1,2,3-oxathiazole-5-carboxylate, 6c



Yield: 66%; ¹H NMR (300 MHz, CDCl₃) δ 7.97 (d, 2H, *J* = 7.4), 7.70 (t, 1H, *J* = 7.5), 7.49 (t, 2H, *J* = 7.8), 7.31-7.18 (m, 6H), 6.17 (s, 1H), 5.23 (q, 2H, *J* = 12.1).; ¹³C NMR (500 MHz, CDCl₃) δ 171.7, 162.7, 136.1, 133.6, 130.3, 129.5, 128.9, 128.7, 128.4, 126.3, 83.8, 69.3.; HRMS (EI): m/z calcd for C₁₆H₁₃NO₅S 331.0514, found 331.0519.

t-Butyl 2,2-dioxo-4-phenyl-5H-1,2,3-oxathiazole-5-carboxylate, 6d



Yield: 48%; ¹H NMR (300 MHz, CDCl₃) δ 8.07 (d, 2H, *J* = 7.5), 7.73 (t, 1H, *J* = 7.5), 7.57 (t, 2H, *J* = 7.7), 6.06 (s, 1H), 1.36 (s, 9H); ¹³C NMR (500 MHz, CDCl₃) δ 172.3, 161.6, 136.0, 130.3, 129.4, 126.6, 86.2, 84.9, 27.5.; HRMS (EI): m/z calcd for C₁₃H₁₅NO₅S 297.0671, found 297.0680.

Methyl 4-(2-tolyl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6e



yield: 53.3% (0.93 g as a white solid), mp = 106.1-107.4 °C, ¹H NMR (300 MHz, CDCl₃) δ 7.68 (d, 1H, *J* = 7.9 Hz), 7.57 (t, 1H, *J* = 7.5 Hz), 7.37-7.43 (m, 2H), 6.24 (s, 1H), 3.75 (s, 3H), 2.69 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 171.7, 163.4, 142.8, 134.8, 133.1, 130.9, 126.5,

125.2, 84.6, 54.2, 22.9.; HRMS (EI): m/z calcd for $C_{11}H_{11}NO_5S$ 269.0358, found 269.0354.

Methyl 4-(3-tolyl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6f



yield: 71.4% (498.2 mg as a white solid), mp = 128.3-129.1 $^{\circ}$ C,¹H NMR (300 MHz, CDCl₃) δ 7.90 (s, 1H), 7.82-7.84 (m, 1H), 7.45-7.53 (m, 2H), 6.17 (s, 1H), 3.81 (s, 3H), 2.45 (s, 3H).; ¹³C NMR (75 MHz, CDCl₃) δ 171.8, 163.7, 139.9, 137.3, 130.8, 129.5, 127.6, 126.4, 83.8, 54.5, 21.4.; HRMS (EI): m/z calcd for C₁₁H₁₁NO₅S

269.0358, found 269.0364.

Methyl 4-(4-tolyl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6g



yield: 53% (670 mg as a white solid), mp = 147.7-148.3 °C, ¹H NMR (500 MHz, CDCl₃) δ 7.96 (d, 2H, *J* = 8.3 Hz), 7.37 (d, 2H, *J* = 8.1 Hz), 6.16 (s, 1H), 3.81 (s, 3H), 2.48 (s, 3H).; ¹³C NMR (75 MHz, CDCl₃) δ 171.5, 163.8, 148.2, 130.5, 130.5, 123.8, 83.7, 54.4, 22.2.; HRMS (EI): m/z calcd for C₁₁H₁₁NO₅S 269.0358, found 269.0355.

Methyl 4-(3-chloro-phenyl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6h



yield: 50.7% (176 mg as a white solid), mp = 149.0-150.5 °C, ¹H
NMR (500 MHz, CDCl₃) δ 8.10 (s, 1H), 7.93-7.95 (m, 1H), 7.707.72 (m, 1H), 7.52-7.55 (m, 1H), 6.15 (s, 1H), 3.84 (s, 3H).; ¹³C
NMR (75 MHz, CDCl₃) δ 170.7, 163.3, 136.2, 136.1, 130.9, 130.2,

128.5, 128.1, 83.6, 54.7.; HRMS (EI): m/z calcd for C₁₀H₈ClNO₅S 288.9812, found 288.9818.

Methyl 4-(4-chloro-phenyl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6i



yield: 68% (650 mg as a white solid), mp = 140.4-142.8 °C, ¹H NMR (500 MHz, CDCl₃) δ 8.03 (d, 2H, *J* = 8.7 Hz), 7.57 (d, 2H, *J* = 8.7 Hz), 6.15 (s, 1H), 3.83 (s, 3H).; ¹³C NMR (75 MHz, CDCl₃) δ 170.7, 163.5, 143.3, 131.7, 130.2, 124.8, 83.6, 54.6.; HRMS (EI): m/z calcd for C₁₀H₈ClNO₅S 288.9812, found 288.9817.

Methyl 4-(4-methoxy-phenyl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6j



285.0308.

yield: 63% (0.5 g as a white solid), mp = 160.8-163.1 °C, ¹H NMR (500 MHz, CDCl₃) δ 8.11 (d, 2H, J = 8.4 Hz), 7.17 (d, 2H, J = 8.5 Hz), 6.83 (s, 1H), 3.95 (s, 3H), 3.78 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 172.2, 172.2, 166.2, 164.0, 133.0, 119.0, 115.0, 83.9, 55.6, 53.5.; HRMS (EI): m/z calcd for C₁₁H₁₁NO₆S 285.0307, found

Methyl 4-(4-fluoro-phenyl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6k



yield: 69% (750 mg as a white solid), mp = 130.6-133.5 °C, ¹H NMR (500 MHz, CDCl₃) δ 8.15-8.18 (m, 2H), 7.29-7.32 (m, 2H), 6.18 (s, OMe 1H), 3.86 (s, 3H).; ¹³C NMR (75 MHz, CDCl₃) δ 170.5, 167.6 (d, J_{CF} = 259.8 Hz), 163.5, 133.4 (d, J_{CF} = 9.9 Hz), 122.8 (d, J_{CF} = 3.1 Hz), 117.3 (d, J_{CF} = 22.3 Hz), 83.6, 54.6.; HRMS (EI): m/z calcd for C₁₀H₈FNO₅S

273.0107, found 273.0107.

Methyl 4-(4- trifluoromethyl-phenyl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6l



yield: 44% (0.37 g as a white solid), mp = 191.1-196.6 °C, ¹H NMR (500 MHz, CDCl₃) δ 8.23 (d, 2H, *J* = 8.1 Hz), 7.86 (d, 2H, *J* = 8.1 Hz), 6.24 (s, 1H), 3.85 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 170.5, 163.0, 137.1 (q, *J*_{CF} = 33.2 Hz), 130.7, 129.4, 126.5, 123.0 (q, *J*_{CF} = 271.4 Hz), 83.6, 54.6.; HRMS (EI): m/z calcd for C₁₁H₈F₃NO₅S

323.0075, found 323.0077.

Methyl 4-(4-cyano-phenyl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6m



Methyl 4-(4-methoxycarbonyl-phenyl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6n



yield: 60.4% (1.13 g as a brown solid), mp = 118.2-123 °C, ¹H NMR (500 MHz, CDCl₃) δ 8.21 (d, 2H, *J* = 8.1 Hz), 8.14 (d, 2H, *J* Me = 8.2 Hz), 6.23 (s, 1H), 3.97 (s, 3H), 3.81 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 171.0, 165.4, 163.1, 136.5, 130.4, 130.3, 129.8, 83.7, 54.5, 52.9.; HRMS (EI): m/z calcd for C₁₂H₁₁NO₇S 313.0256,

found 313.0246.

Methyl 4-(naphthalen-2-yl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 60



yield: 59% (1.1 g as a white solid), mp = 151.5-155.7 °C, ¹H NMR (500 MHz, CDCl₃) δ 8.60 (s, 1H), 8.11 (d, 1H, *J* = 8.7 Hz), 8.01(t, 2H, *J* = 8.6 Hz), 7.95 (d, 1H, *J* = 8.2 Hz), 7.65-7.75 (m, 2H), 6.35 (s, 1H), 3.83 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 171.5, 163.9, 136.9, 133.5, 132.4, 130.6, 130.1, 129.9, 128.3, 128.0, 124.6, 123.9,

83.9, 54.5.; HRMS (EI): m/z calcd for C₁₄H₁₁NO₅S 305.0358, found 305.0356.

Methyl 4-(furan-2-yl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6p



yield: 18% (0.14 g as a ivory solid), mp = 130.6-133.5 °C, ¹H NMR (300 MHz, CDCl₃) δ 7.86-7.87 (m, 1H), 7.67-7.68 (m, 1H), 6.75-6.77 (m, 1H), 5.97 (s, 1H), 3.87 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 163.4, 159.8, 151.1, 143.3, 124.9, 114.6, 82.6, 54.5.; HRMS (EI): m/z calcd for C₈H₇NO₆S 244.9994, found 244.9997.

Methyl 4-(thiophen-2-yl)-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6q

yield: 88.7% (0.51g as a ivory solid), mp = 106.2-110.2 °C, ¹H NMR (300 MHz, CDCl₃) δ 8.07-8.08 (m, 1H), 7.98-7.99 (m, 1H), 7.29-7.31 (m, 1H), 6.09 (s, 1H), 3.90 (s, 3H).; ¹³C NMR (75 MHz, CDCl₃) δ 164.7, 163.9, 139.1, 138.0, 130.2, 129.8, 83.3, 54.6.; HRMS (EI): m/z calcd for

 $C_8H_7NO_5S_2$ 260.9766, found 260.9765.

Methyl 4-propyl-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6r



yield: 63.8% (2.79 g as a yellow oil); ¹H NMR (500 MHz, CDCl₃) δ 5.56 (s, 1H), 3.91 (s, 3H), 2.65-2.75 (m, 2H), 1.79-1.87 (m, 2H), 1.04 (t, 3H, J -OMe = 7.4 Hz).; ¹³C NMR (75 MHz, CDCl₃) δ 179.9, 162.8, 85.1, 54.4, 33.2, 18.9, 13.9.; HRMS (EI): m/z calcd for C₇H₁₁NO₅S 221.0358, found

221.0365.

Methyl 4-phenethyl-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6s



yield: 45% (0.63 g as a ivory solid), mp = 83-87.1 °C, ¹H NMR (500 MHz, CDCl₃) δ 7.20-7.37 (m, 5H), 5.54 (s, 1H), 3.86 (s, 3H), 3.05Me 3.13 (m, 4H).; ¹³C NMR (125 MHz, CDCl₃) δ 179.6, 162.4, 138.7, 129.0, 128.4, 127.0, 85.2, 54.3, 33.1, 31.3.; HRMS (EI): m/z calcd for C₁₂H₁₃NO₅S 283.0514, found 283.0516.

Methyl 4-cyclohexyl-5H-1,2,3-oxathiazole-5-carboxylate 2,2-dioxide, 6t



yield: 79.4% (2.26g as a pale yellow oil); ¹H NMR (500 MHz, CDCl₃) δ 5.64 (s, 1H), 3.86 (s, 3H), 2.69-2.70 (m, 1H), 1.37-2.06 (m, 10H).; ¹³C NMR (125 MHz, CDCl₃) δ 183.0, 162.9, 83.9, 54.2, 40.0, 30.8, 29.1, 25.7, 25.3, 24.9.; HRMS (EI): m/z calcd for C₁₀H₁₅NO₅S 261.0671,

found 261.0683.

4. General procedure for the ATH-DKR reaction of 4,5-disubstituted cyclic imine 6 to 7



To the solution of **6a** (255 mg, 1.0 mmol) in EtOA (10 mL) was added added (*R*,*R*)-Cp*RhCl(TsDPEN) (**1a**) catalyst (3.2 mg, 0.5 mol%), and then added slowly an azeotroic mixtrure of HCO₂H/Et₃N (molar ratio = 5:2, 1.0 mL) *via* a syringe. The reaction mixture was stirred for 0.5 h at room temperature and diluted with EtOAc. The reaction mixture was washed with water and brine. The organic layer was dried over anhydrous MgSO₄ and evaporated under reduced pressure. The residue was purified by column chromatography on silica gel (Hexane/EtOAc, 2:1) to give a white solid (234 mg, 91.6%).

Methyl (4S,5S)-4-phenyl-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (S,S)-7a



yield: 91.6% (234 mg as a white solid), mp = 101.9-104.7 °C, 97% ee: Chiralpak IB, 20% ethanol/n-hexane, 1.0 ml/min, 215 nm $t_{\rm R}$ (major) = 10.7 min, $t_{\rm R}$ (minor) = 12.5 min; $[\alpha]_{\rm D}^{30}$ = +102.9 (c 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.41-7.42 (m, 3H), 7.29-7.32 (m, 2H), 5.39 (d, 1H, J = 6.8 Hz), 5.28 (m, 1H), 5.07 (brs, 1H), 3.40 (s, 3H).; ¹³C

NMR (75 MHz, CDCl₃) δ 165.7, 132.0, 129.8, 129.1, 126.6, 81.8, 61.4, 52.7.; HRMS (EI): m/z calcd for C₁₀H₁₁NO₅S 257.0358, found 257.0340.

Methyl (4R,5R)-4-phenyl-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (R,R)-7a



yield: 94% (24 mg as a white solid), mp = 103.2-105.4 °C, 98.1% ee: Chiralpak IB, 20% ethanol/n-hexane, 1.0 ml/min, 215 nm $t_{\rm R}$ (major) = 9.9 min, $t_{\rm R}$ (minor) = 13.7 min; $[\alpha]_{\rm D}^{30}$ = -101.7 (c 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.41-7.42 (m, 3H), 7.30-7.31 (m, 2H), 5.38 (d, 1H, J = 6.8 Hz), 5.28 (d, 1H, J = 6.8 Hz), 5.02 (brs, 1H), 3.40 (s, 3H).; ¹³C

NMR (125 MHz, CDCl₃) δ 165.6, 131.9, 129.9, 129.2, 126.5, 81.6, 61.4, 52.7.; HRMS (EI): m/z calcd for C₁₀H₁₁NO₅S 257.0358, found 257.0343.

Isopropyl (45,55)-4-phenyl-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (5,5)-7b



yield: 85%; $[\alpha]_D^{23} = +76.8$ (c 0.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.39-7.34 (m, 5H), 5.33 (m, 3H), 4.69 (s, 1H), 1.02 (s, 3H), 0.72 (s, 3H); ¹³C NMR (500 MHz, CDCl₃) δ 164.8, 132.2, 129.7, 129.1, 126.9, 81.6, 70.8, 61.5, 21.4, 20.8; HRMS (EI): m/z calcd for C₁₂H₁₅NO₅S

285.0671, found 285.0664.

Benzyl (45,55)-4-phenyl-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (5,5)-7c



yield: 87%; $[\alpha]_D^{21} = +71.5$ (c 0.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.39-7.29 (m, 8H), 7.04 (d, 2H. J = 6.8), 5.44 (d, 1H, J = 6.7), 5.33 (d, 1H, J = 6.9), 4.93 (d, 1H, J = 12.0), 4.68 (d, 1H, J = 12.0); ¹³C NMR (500 MHz, CDCl₃) δ 165.2, 133.7, 131.9, 129.8, 129.2, 128.8, 128.7, 128.6, 126.7, 81.8, 68.1, 61.5; HRMS (EI): m/z calcd for C₁₆H₁₅NO₅S

333.0671, found 333.0662.

t-Butvl (45,55)-4-phenvl-1,2,3-oxathiazolidine-5-carboxvlate 2,2-dioxide, (5,5)-7d



yield: 87%; 99.9% ee (Chiralpak AD-H, 5% isopropanol/hexanes, 1.0 mL/min, 215 nm, $t_r(minor) = 32.27 \text{ min}, t_r(major) = 36.25 \text{ min}); [\alpha]_D^{20}$ = +89.3 (c 0.3, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.41-7.33 (m, 5H), 5.27 (m, 3H), 1.08 (s, 9H); ¹³C NMR (500 MHz, CDCl₃) δ 164.0, 132.6, 129.6, 129.1, 127.0, 84.5, 81.7, 61.5, 27.4; HRMS (EI): m/z

calcd for C₁₃H₁₇NO₅S 299.0827, found 299.0838.

Methyl (45,55)-4-(o-tolyl)-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (5,5)-7e



yield: 95.1% (25.7 mg as a white solid), mp = 119.8-120.4 °C, 92.1% ee: Chiralpak AD-H, 20% isopropanol/n-hexane, 1.0 ml/min, 215 nm $t_{\rm R}({\rm major}) = 8.4 {\rm min}, t_{\rm R}({\rm minor}) = 10.1 {\rm min}; \left[\alpha\right]_{\rm D}^{30} = +91.3 {\rm (c \ 0.15,}$ MeOH); ¹H NMR (500 MHz, CDCl₃) δ 7.22-7.29 (m, 3H), 7.18 (d, 1H, J = 7.6 Hz), 5.52 (d, 1H, J = 7.0 Hz), 5.36 (d, 1H, J = 7.0 Hz), 3.31 (s,

3H), 2.41 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 165.7, 136.1, 131.2, 129.7, 129.6, 126.8, 124.2, 80.6, 57.8, 52.5, 19.4.; HRMS (EI): m/z calcd for C₁₁H₁₃NO₅S 271.0514, found 271.0510.

Methyl (45,55)-4-(m-tolyl)-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (5,5)-7f



yield: 99.3% (31.4 mg as a white solid), mp = 97.1-99.4 °C, 98.7% ee: Chiralpak AD-H, 10% isopropanol/n-hexane, 1.3 ml/min, 215 nm $t_{\rm R}$ (major) = 15.9 min, $t_{\rm R}$ (minor) = 17.0 min; $[\alpha]_{\rm D}^{29}$ = +54.2 (c 0.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.29-7.33 (m, 1H), 7.23 (d, 1H, *J* = 7.6 Hz), 7.10-7.13 (m, 2H), 5.38 (d, 1H, *J* = 6.7

Hz), 5.28 (d, 1H, J = 6.5 Hz), 5.19 (brs, 1H), 3.44 (s, 3H), 2.39 (s, 3H).; ¹³C NMR (75 MHz, CDCl₃) δ 165.9, 139.3, 131.8, 130.7, 129.2, 127.2, 123.6, 81.9, 61.5, 52.7, 21.5.; HRMS (EI): m/z calcd for C₁₁H₁₃NO₅S 271.0514, found 271.0507.

Methyl (45,55)-4-(p-tolyl)-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (5,5)-7g



yield: 98.6% (50.8 mg as a white solid), mp = 103.8-105.8 °C ,99.2% ee: Chiralpak AD-H, 20% isopropanol/n-hexane, 1.0 ml/min, 215 nm $t_{\rm R}$ (major) = 10.1 min, $t_{\rm R}$ (minor) = 12.4 min; $[\alpha]_{\rm D}^{30}$ = +93.0 (c 0.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.18-7.22 (m, 4H), 5.35 (d, 1H, J = 6.8 Hz), 5.25 (d, 1H, J = 6.9 Hz), 5.20 (brs, 1H), 3.42 (s, 3H), 2.36

(s, 3H).; ¹³C NMR (75 MHz, CDCl₃) δ 165.8, 140.1, 130.0, 128.9, 126.5, 81.9, 61.4, 52.8, 21.3.; HRMS (EI): m/z calcd for C₁₁H₁₃NO₅S 271.0514, found 271.0515.

Methyl (4*S*,5*S*)-4-(3-chloro-phenyl)-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (*S*,*S*)-7h



yield: 99.6% (36.1 mg as a white solid), mp = 79.6-80.8 °C, 96.7% ee: Chiralpak AD-H, 10% isopropanol/n-hexane, 1.2 ml/min, 215 nm $t_{\rm R}$ (minor) = 13.8 min, $t_{\rm R}$ (major) = 15.0 min; $[\alpha]_{\rm D}^{29}$ = +69.4 (c 0.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.33-7.40 (m, 3H), 7.24-7.25 (m, 1H), 5.39 (d, 1H, *J* = 6.8 Hz), 5.25-5.26 (m, 2H), 3.47

(s, 3H).; ¹³C NMR (75 MHz, CDCl₃) δ 165.4, 135.3, 134.4, 130.6, 130.2, 127.2, 124.9, 81.4, 60.9, 53.0.; HRMS (EI): m/z calcd for C₁₀H₁₀ClNO₅S 290.9968, found 290.9957.

Methyl (4*S*,5*S*)-4-(4-chloro-phenyl)-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (*S*,*S*)-7i



yield: 92.1% (28.3 mg as a white solid), mp = 123.2-125.8 °C, 97.3% ee: Chiralpak AD-H, 20% isopropanol/n-hexane, 1.5 ml/min, 215 nm $t_{\rm R}$ (minor) = 9.3 min, $t_{\rm R}$ (major) = 14.1 min; $[\alpha]_{\rm D}^{29}$ = +69.9 (c 0.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.40 (d, 2H, *J* = 8.5 Hz), 7.28 (d, 2H, *J* = 8.5 Hz), 5.39 (d, 1H, *J* = 6.8 Hz), 5.25 (t, 1H, *J* = 8.0 Hz),

5.14 (d, 1H, J = 7.5 Hz), 3.46 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 165.4, 136.2, 130.9, 129.6, 128.2, 81.3, 61.0, 53.0.; HRMS (EI): m/z calcd for C₁₀H₁₀ClNO₅S 290.9968, found 290.9943.

Methyl (4*S*,5*S*)-4-(4-methoxy-phenyl)-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (*S*,*S*)-7j



yield: 99.2% (28.5 mg as a white solid), mp = 120.8-123.7 °C, 100% ee: Chiralpak AD-H, 20% isopropanol/n-hexane, 1.0 ml/min, 215 nm $t_{\rm R}$ (major) = 14.0 min, racemic: $t_{\rm R}$ (minor) = 18.7 min; $[\alpha]_{\rm D}^{30}$ = +91.6 (c 0.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.23 (d, 2H, J = 8.1 Hz), 6.91 (d, 2H, J = 8.1 Hz), 5.33 (d, 1H, J = 6.8 Hz), 5.22 (d,

MeÓ

1H, J = 6.8 Hz), 3.81 (s, 3H), 3.45 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 165.8, 160.8, 128.1, 123.9, 114.7, 81.8, 61.2, 55.5, 52.9.; HRMS (EI): m/z calcd for C₁₁H₁₃NO₆S 287.0464, found 287.0466.

Methyl (4*S*,5*S*)-4-(4-fluoro-phenyl)-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (*S*,*S*)-7k



yield: 96.1% (45 mg as a white solid), mp = 142.5-146.4 °C, 97.3% ee: Chiralpak IA, 20% ethanol/n-hexane, 1.0 ml/min, 215 nm $t_{\rm R}$ (minor) = 12.5 min, $t_{\rm R}$ (major) = 16.2 min; $[\alpha]_{\rm D}^{29}$ = +78.3 (c 0.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.32-7.35 (m, 2H), 7.10-7.13 (m, 2H), 5.38 (d, 1H, J = 6.7 Hz), 5.22-5.28 (m, 2H), 3.45 (s, 3H).; ¹³C NMR (125

MHz, CDCl₃) δ 165,6, 163.5 (d, J_{CF} = 248.8 Hz), 128.9 (d, J_{CF} = 8.5 Hz), 128.4, 116.5 (d, J_{CF} = 22.1 Hz), 81.6, 61.0, 53.0.; HRMS (EI): m/z calcd for C₁₀H₁₀FNO₅S 275.0264, found 275.0234.

Methyl (4*S*,5*S*)-4-(4-trifluoromethyl-phenyl)-1,2,3-oxathiazolidine-5-carboxylate 2,2dioxide, (*S*,*S*)-7l



Methyl (4*S*,5*S*)-4-(4-cyano-phenyl)-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (*S*,*S*)-7m



3H), 2.92 (brs, 1H).; ¹³C NMR (125 MHz, acetone- d_6) δ 165.8, 141.0, 133.3, 129.5, 119.0, 113.8, 81.9, 61.5, 52.8.; HRMS (EI): m/z calcd for C₁₁H₁₀N₂O₅S 282.0310, found 282.0304.

Methyl (4*S*,5*S*)-4-(4-methoxycarbonyl-phenyl)-1,2,3-oxathiazolidine-5-carboxylate 2,2dioxide, (*S*,*S*)-7n



yield: 92% (29 mg as a white solid), mp = 145.6-147.1 °C, 96.7% ee: Chiralpak IA, 20% ethanol/n-hexane, 1.5 ml/min, 215 nm $t_{\rm R}({\rm minor}) = 12.0 \text{ min}, t_{\rm R}({\rm major}) = 14.0 \text{ min}; [\alpha]_{\rm D}^{20} = +57.4 (c 0.6, CHCl_3);$ ¹H NMR (500 MHz, CDCl₃) δ 8.08 (d, 2H, *J* = 7.9 Hz), 7.44 (d, 2H, *J* = 7.9 Hz), 5.51-5.61 (brs, 1H), 5.45 (d, 1H, *J* = 6.8

Hz), 5.37 (d, 1H, J = 6.9 Hz), 3.94 (s, 3H), 3.41 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 166.2, 165.3, 137.1, 131.5, 130.3, 126.8, 81.2, 61.0, 52.8, 52.4.; HRMS (EI): m/z calcd for C₁₂H₁₃NO₇S 315.0413, found 315.0411.

Methyl (4*S*,5*S*)-4-(naphthalen-2-yl)-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (*S*,*S*)-70



yield: 90.8% (27 mg as a white solid), mp = 134.9-137.4 °C, 96.7% ee: Chiralpak AD-H, 20% isopropanol /n-hexane, 1.0 ml/min, 215 nm $t_{\rm R}$ (major) = 11.5 min, $t_{\rm R}$ (minor) = 13.7 min; $[\alpha]_{\rm D}^{20}$ = +94.1 (c 0.4, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.80-7.90 (m, 4H), 7.53-7.56 (m, 2H), 7.37 (d, 1H, *J* = 8.6 Hz), 5.46 (s, 2H), 3.29 (s,

3H).; ¹³C NMR (125 MHz, CDCl₃) δ 165.7, 133.5, 132.9, 129.2, 128.2, 127.8, 127.3, 127.1, 126.4, 123.3, 81.7, 61.6, 52.7.; HRMS (EI): m/z calcd for C₁₄H₁₃NO₅S 307.0514, found 307.0513.

Methyl (45,55)-4-(furan-2-yl)-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (5,5)-7p



yield: 91.6% (22 mg as a white solid), mp = 97.9-110.2 °C, 94.9% ee: Chiralpak IA, 30% ethanol /n-hexane, 1.3 ml/min, 215 nm $t_{\rm R}$ (major) = 8.7 min, $t_{\rm R}$ (minor) = 14.3 min; $[\alpha]_{\rm D}^{28}$ = +95.8 (c 0.3, CDCl₃); ¹H NMR (500 MHz, CHCl₃) δ 7.44 (s, 1H), 6.43-6.53 (m, 2H), 5.29-5.32 (m, 1H), 5.22-5.23 (m, 1H), 5.16-5.18 (m, 1H), 3.65 (s, 3H).; ¹³C NMR (75 MHz,

CDCl₃) δ 166.1, 144.3, 143.8, 111.5, 111.4, 80.8, 55.8, 53.5.; HRMS (EI): m/z calcd for C₈H₉NO₆S 247.0151, found 247.0155.

Methyl (4*S*,5*S*)-4-(thiophen-2-yl)-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (*S*,*S*)-7q



yield: 93.8% (38.3 mg as a white solid), mp = 96-97 °C, 98.7% ee: Chiralpak IA, 20% ethanol /n-hexane, 1.5 ml/min, 215 nm $t_{\rm R}$ (minor) = 12.5 min, $t_{\rm R}$ (major) = 14.2 min; $[\alpha]_{\rm D}^{29}$ = +70.6 (c 0.3, CDCl₃); ¹H NMR (500 MHz, CHCl₃) δ 7.37-7.38 (m, 1H), 7.14 (s, 1H), 7.04-7.05 (m, 1H), 5.49 (d, 1H, J = 5.9 Hz), 5.35 (d, 1H, J = 6.5 Hz), 5.13 (s, 1H), 3.59 (s,

3H).; ¹³C NMR (125 MHz, CDCl₃) δ 165.5, 133.5, 127.6, 127.4, 127.2, 81.7, 57.7, 53.0.; HRMS (EI): m/z calcd for C₈H₉NO₅S₂ 262.9922, found 262.9918.

Methyl (45,55)-4-propyl-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (5,5)-7r

yield: 69% (98.5 mg as a coloress oil); $[\alpha]_D^{29} = +52.5$ (c 0.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 5.06 (d, 1H, J = 5.3 Hz), 4.48 (brs, 1H), 4.13 (brs, 1H), 3.86 (s, 3H), 1.48-1.61 (m, 4H), 0.97 (t, 3H, J = 6.9 Hz).; ¹³C NMR (75 MHz, CDCl₃) δ 167.1, 82.3, 58.5, 53.2, 30.8, 19.8, 13.7.;

HRMS (EI): m/z calcd for C₇H₁₃NO₅S 223.0514, found 223.0537.

Methyl (45,55)-4-phenethyl-1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (5,5)-7s



yield: 53.5% (14 mg as a ivory solid), mp = 90.6-92.3 °C, 76.1% ee: Chiralpak IA, 20% ethanol /n-hexane, 1.5 ml/min, 215 nm $t_{\rm R}$ (minor) = 6.0 min, $t_{\rm R}$ (major) = 8.0 min; $[\alpha]_{\rm D}^{20}$ = +41.3 (c 0.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.33-7.36 (m, 2H), 7.26-7.29 (m, 1H), 7.20 (d, 2H, *J* = 7.4 Hz), 5.08 (d, 1H, *J* = 6.1 Hz), 4.66 (d, 1H, *J* =

2.2 Hz), 4.12-4.14 (m, 1H), 3.88 (s, 3H), 2.86-2.90 (m, 1H), 2.75-2.81 (m, 1H), 1.89-1.97 (m, 2H).; 13 C NMR (125 MHz, CDCl₃) δ 166.9, 139.2, 128.9, 128.4, 126.8, 82.0, 57.7, 53.2, 32.3, 30.5.; HRMS (EI): m/z calcd for C₁₂H₁₅NO₅S 285.0671, found 285.0666.

Methyl (45,55)-4-cyclohexyl -1,2,3-oxathiazolidine-5-carboxylate 2,2-dioxide, (5,5)-7t

yield: 66.7% (179 mg as a coloress oil); $[\alpha]_D^{30} = +41.4$ (c 0.68, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 5.09(d, 1H, J = 5.8 Hz), 4.60(bs, 1H), ^{3.87(s, 3H), 3.84 (m, 1H), 1.02-1.88 (m, 11H).; ¹³C NMR (75 MHz, CDCl₃) δ 167.4, 82.5, 63.9, 53.1, 37.8, 30.7, 29.3, 25.7, 25.2, 25.1.;}

HRMS (EI): m/z calcd for $C_{10}H_{17}NO_5S$ 263.0827, found 263.0848.

5. Assignment of absolute stereochemistry of 7a via converting to the known 8a

In order to determine the absolute stereochemistry of the ATH-DKR product, **7a** was converted to the known 2-azido-3-(*Boc*-amino)-3-phenyl propionic acid methyl ester **8a** and it was compared with the stereochemically defined **8a** which was derived from Sharpless asymmetric amino hydroxylation reaction.¹⁰ The spectroscopic data and specific rotation data of synthetic **8a** were full agreement with those of the known ((2R,3S)-**8a**) in the

¹⁰ S.-H. Lee, J. Yoon, S.-H. Chung and Y.-S. Lee, *Tetrahedron*, 2001, **57**, 2139.

literature. Additionally, the absolute stereochemistry of **7j** was unambiguously assigned by using single-crystal X-ray crystallographic analysis (deposited, CCDC-1007235).

Scheme S1.



Reaction conditions: (a) (Boc)₂O, Et₃N, cat. DMAP, CH₂Cl₂, 100%; (b) i. NaN₃, DMF, 60 °C, 6 h; ii. 1N HCl, Et₂O, 12 h, rt, 92%.

5-1. Synthesis of (*S*,*S*)-*N*-Boc-7a

To a stirred mixture of (S,S)-**7a** (0.23 g, 0.92 mmol) and triethylamine (0.15 mL, 1.1 mmol) in dichloromethane (2.5 mL) was added successively di-*tert*-butyl dicarbonate (0.4 g, 1.83 mmol) and DMAP (catalytic amount). The reaction mixture was stirred at room temperature for 40 min. The reaction mixture was diluted with diethyl ether (20 mL) and washed successively with 1 N HCl, saturated aqueous NaHCO₃ solution and brine. The organic layer was dried over anhydrous MgSO₄, filtered and evaporated under reduced pressure. The residue was purified by column chromatography (Hexane/EtOAc=3:1) to give 0.33 g (99.7% yield) of (S,S)-*N*-**Boc-7a**.

yield: 99.7% (0.33 g as a white solid), mp=126.3-130.2 °C, 97.8% ee: Chiralpak AD-H, 10% isopropanol/n-hexane, 1.0 ml/min, 215 nm $t_{\rm R}({\rm minor}) = 9.2 {\rm min}, t_{\rm R}({\rm major}) = 13.4 {\rm min}; [\alpha]_{\rm D}^{20} = -14.6 {\rm (c} 1.0, {\rm CHCl}_3);$ ¹H NMR (500 MHz, CDCl₃) δ 7.35-7.39 (m, 5H), 5.50 (d, 1H, *J* = 6.35 Hz), 5.45 (d, 1H, *J* = 6.3 Hz), 3.42 (s, 3H), 1.44 (s, 9H).; ¹³C NMR (75 MHz, CDCl₃) δ 163.1, 147.8, 133.6, 129.8, 129.0, 127.5, 86.4, 62.9, 52.9, 28.0,; HRMS (EI): m/z calcd for C₁₅H₁₉NO₇S 357.0882, found 357.0887.

5-2. 2-Azido-3-tert-butoxycarbonylamino-3-phenyl-propionic acid methyl ester, (2*R*,3*S*)-8a

NaN₃ (22.3 mg, 0.343 mol, 5.0 equiv) was added in a single portion to a solution of (*S*,*S*)-*N*-**Boc-7a** (24.5 mg, 68.6 mmol, 1.0 equiv) in DMF (2 ml) at 25 °C. The resulting mixture was warmed to 60 °C and stirred for 6 h. Upon completion, the reaction mixture was cooled to rt and the contents were diluted with Et_2O (3 mL), treated with 1*N* aqueous HCl (3 mL), and allowed to stir for an additional 12 h at 25 °C. Once this operation was complete, the reaction mixture was poured into saturated NaHCO₃ solution and extracted with Et_2O . The combined organic layers were then washed with water, dried (MgSO₄), and concentrated. The resultant light yellow solid was purified by flash column chromatography (silica gel, EtOAc/hexanes, 5:1).

NHBocWhite solid, yield: 92%; mp 133-134°C; $[\alpha]_D^{20} = +17.06$ (c 0.8, CHCl₃);Ph $\stackrel{:}{S}$ R CO2Me 1 H NMR (300 MHz, CDCl₃) δ 7.37-7.30 (m, 5H), 5.33 (brs, 2H), 4.38 (brs, 1H), 3.82 (s, 3H), 1.42 (s, 9H); 13 C NMR (500 MHz, CDCl₃) δ 168.8, 155.0, 138.4, 128.9, 128.3, 126.7, 80.4, 66.9, 55.3, 53.2, 28.4.

The spectroscopic and specific rotation data of synthetic **8a** were full agreement with those of the known ((2R, 3S)-**8a**) in the literature.¹¹

Lit.¹¹ mp 133–134°C; $[\alpha]_D^{20} = +16.5^\circ$ (*c* 1, CHCl₃); ¹H NMR (CDCl₃, 300 MHz) δ 7.25–7.50 (m, 5H), 5.34 (br s, 2H), 4.38 (br s, 1H), 3.81 (s, 3H), 1.42 (s, 9H); ¹³C NMR (CDCl₃, 75 MHz) δ 168.62, 154.78, 138.27, 128.71, 128.09, 126.51, 80.21, 66.71, 55.18, 52.95, 28.19.

6. Evaluation of ee's of 7r and 7t by convertion to ring-opened derivatives

Because of difficults in chiral separation of **7r** and **7t** themselves in various chiral columns and conditions, **7r** and **7t** were converted to the corresponding ring opened derivatives **8r** and **8t** and ee values were indirectly determined with these compounds.

Scheme S2.

¹¹ S.-H. Lee, J. Yoon, S.-H. Chung and Y.-S. Lee, *Tetrahedron*, 2001, **57**, 2139.



Reaction conditions: (a) $(Boc)_2O$, Et_3N , cat. DMAP, CH_2Cl_2 ; (b) i. PhCO₂NH₄, DMF, 55 °C, 12 h; ii. 1N HCl, CH_2Cl_2 , 6 h, rt.

6-1. Synthesis of N-Boc-7r and N-Boc-7t

6-1-1. N-Boc-7t

To a stirred mixture of (S,S)-**7t** (16.4 mg, 0.062 mmol) and triethylamine (0.01mL, 0.072 mmol) in dichloromethane (1 mL) at 0 °C was added successively di-*tert*-butyl dicarbonate (27.2 mg, 0.125 mmol) and DMAP (catalytic amount). The reaction mixture was stirred at room temperature for 2 h, diluted with CH₂Cl₂ (5 mL) and washed successively with 1 N HCl, saturated NaHCO₃ solution and brine. The organic layer was dried over anhydrous MgSO₄, and evaporated. The residue was purified by column chromatography (Hexane/EtOAc=10:1) to give 20 mg (87% yield) of (*S*,*S*)-*N*-**Boc-7t**.



Yield: 87% (20 mg); $[\alpha]_D{}^{30} = -15.01$ (c 0.92, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 5.30 (d, J = 4.7 Hz, 1H), 4.56 (m, 1H), 3.89 (s, 3H), 1.66-1.83 (m, 6H), 1.56 (s, 9H), 1.08-1.26 (m, 5H).; ¹³C NMR (125 MHz, CDCl₃) δ 164.1, 149.1, 85.9, 77.2, 63.8, 53.2, 40.1, 30.7, 27.8, 27.0, 26.3, 25.9, 25.6.; HRMS (EI): m/z calcd for C₁₅H₂₅NO₇S 363.1352, found 363.1364.

6-1-2. N-Boc-7r



Yield: 97% (270 mg); $[\alpha]_D^{29} = -26.77$ (c 1.48, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 5.28 (d, J = 5.4 Hz, 1H), 4.61-4.67 (m, 1H), 3.89 (s, 3H), 1.62-1.77 (m, 2H), 1.55 (s, 9H), 1.35-1.43 (m, 2H), 0.90-0.95 (t, J = 7.3 Hz, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 163.9, 148.4, 86.0, 77.0, 59.2,

53.3, 31.9, 27.9, 18.1, 13.9.; HRMS (EI): m/z calcd for $C_{12}H_{21}NO_7S$ 323.1039, found 323.1014.

6-2. Synthesis of 8r and 8t

6-2-1. (2R,3S)-8t

Ammonium benzoate (65.24 mg, 0.47 mmol) was added to a solution of (4*S*,5*S*)-*N*-*Boc*-7t (85.2 mg, 0.23 mmol) in dry DMF (10 mL). The solution was heated to 60 °C under inert atmosphere (N_2) and stirred for 14 h at that temperature. The solvent was evaporated under reduced pressure and the residue was re-dissolved in dichloromethane (10 mL) and 1N HCl 10 mL) was added. The reaction mixture was stirred at room temperature for 6 h before the pH was adjusted to 8 with saturated aquous NaHCO₃ solution. The layers were separated and the aqueous layer was extracted three times with dichloromethane. The combined organic layers were washed with water and brine, dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by column chromatography on silica-gel (Hexane/EtOAc, 4:1) to give 66 mg of **8t**.



yield: 70.8% (66 mg); 47% ee, dr = 31:1.; Chiralpak IC, 10% isopropanol/n-hexane, 0.7 ml/min, 254 nm $t_{\rm R}$ (minor) = 8.35 min, $t_{\rm R}$ (major) = 9.55 min; $[\alpha]_{\rm D}^{30}$ = -35.44 (c 0.35, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.07 (d, J = 7.3 Hz, 2H), 7.61 (m, 1H), 7.48 (m, 2H), 5.44 (d, J = 2.25 Hz, 1H), 4.78 (d, J = 10.5 Hz, 1H), 4.09 (m, 1H), 3.75

(s, 3H), 1.61-1.92 (m, 6H), 1.43 (s, 9H), 1.06-1.15 (m, 5H).; ¹³C NMR (125 MHz, CDCl₃) δ 169.2, 165.7, 155.5, 133.6, 129.9, 129.2, 128.5, 79.6, 72.4, 56.0, 52.5, 39.8, 29.7, 29.6, 28.3, 26.0, 25.9.; HRMS (EI): m/z calcd for C₂₂H₃₁NO₆ 405.2151, found 405.2158.

6-2-2. (2R,3S)-8r

 $\begin{array}{rcl} & \text{Boc} \\ & \text{HN} & \text{OBz} \\ & \text{Sr} & \text{OBz} \\ & \text{Br} & \text{OBz} \end{array} \begin{array}{r} \text{yield: } 72.3\% & (220 \text{ mg}); \ 91\% \text{ ee, } dr \ = \ 100:0.; \ \text{Chiralpak IC, } 10\% \\ & \text{isopropanol/n-hexane, } 0.7 \text{ ml/min, } 254 \text{ nm } t_{\text{R}}(\text{minor}) \ = \ 9.27 \text{ min, } t_{\text{R}}(\text{major}) \\ & = \ 11.15 \text{ min; } [\alpha]_{\text{D}}^{29} \ = \ -70.5 \ (c \ 0.99, \ \text{CHCl}_3); \ ^1\text{H NMR} \ (300 \text{ MHz, } \text{CDCl}_3) \\ & \delta \ 8.07 \ (d, \ J \ = \ 8.4 \text{ Hz, } 2\text{H}), \ 7.61 \ (m, \ 1\text{H}), \ 7.47 \ (m, \ 2\text{H}), \ 5.27 \ (d, \ J \ = \ 2.3 \text{ Hz, } \\ & 1\text{H}), \ 4.77(d, \ J \ = \ 9.9 \text{ Hz, } 1\text{H}), \ 4.32 \ (m, \ 1\text{H}), \ 3.77 \ (s, \ 3\text{H}), \ 1.53-1.62 \ (m, \ 2\text{H}), \ 1.46 \ (s, \ 9\text{H}), \end{array}$

1.36-1.48 (m, 2H), 0.93-0.97 (t, J = 7.3 Hz, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 168.8, 165.7, 155.3, 133.6, 129.9, 129.1, 128.5, 79.7, 74.2, 52.5, 51.1, 34.4, 28.3, 19.2, 13.7.; HRMS (EI): m/z calcd for C₁₉H₂₇NO₆ 365.1838, found 365.1856.

7. Synthesis of Taxotere side chain, (*2R*,*3S*)-10 Scheme S3.



7-1. Synthesis of (4S,5S)-N-Boc-7a

To a stirred mixture of (S,S)-**7a** (0.23 g, 0.92 mmol) and triethylamine (0.15 mL, 1.1 mmol) in dichloromethane (2.5 mL) was added successively di-*tert*-butyl dicarbonate (0.4 g, 1.83 mmol) and DMAP (cat) and the mixture was stirred at room temperature for 40 min. It was then diluted with diethyl ether (20 mL) and washed successively with 1 N HCl, NaHCO₃ and brine. The organic layer was dried over anhydrous MgSO₄, and evaporated under reduced pressure. The residue was purified by column chromatography on silicagel (Hexane/EtOAc = 3:1) to give 0.33 g (99.7% yield) of (4*S*,5*S*)-*N*-*Boc*-**7a**.



yield: 99.7% (0.33 g as a white solid), mp=126.3-130.2 °C, 97.8% ee: Chiralpak AD-H, 10% isopropanol/n-hexane, 1.0 ml/min, 215 nm $t_{\rm R}$ (minor) = 9.2 min, $t_{\rm R}$ (major) = 13.4 min; $[\alpha]_{\rm D}^{20}$ = -14.6 (c 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.35-7.39 (m, 5H), 5.50 (d, 1H, *J* = 6.35 Hz), 5.45 (d, 1H, *J* = 6.3 Hz), 3.42 (s, 3H), 1.44 (s, 9H).; ¹³C NMR (75

MHz, CDCl₃) δ 163.1, 147.8, 133.6, 129.8, 129.0, 127.5, 86.4, 62.9, 52.9, 28.0,; HRMS (EI): m/z calcd for C₁₅H₁₉NO₇S 357.0882, found 357.0887.

7-2. Synthesis of (4R,5R)-N-Boc-7a



yield: 94.2% (109 mg as a white solid), mp=127.2-129.4 °C, 98.1% ee: Chiralpak AD-H, 10% isopropanol/n-hexane, 1.0 ml/min, 215 nm $t_{\rm R}$ (major) = 9.3 min, $t_{\rm R}$ (minor) = 13.8 min; $[\alpha]_{\rm D}^{20}$ = +12.3 (c 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.35-7.38 (m, 5H), 5.49 (d, 1H, *J* = 6.35 Hz), 5.44 (d, 1H, *J* = 6.3 Hz), 3.42 (s, 3H), 1.43 (s, 9H).; ¹³C NMR (125

MHz, CDCl₃) & 163.0, 147.7, 133.4, 129.6, 128.8, 127.3, 86.2, 62.8, 52.8, 27.8.

7-3. Synthesis of (2R,3S)-8a

Ammonium benzoate (0.16 g, 1.17 mmol) was added to a solution of (4*S*,5*S*)-*N*-*Boc*-7a (0.21 g, 0.58 mmol) in dry DMF (2 mL). The solution was heated to 60 °C and stirred for 12 h at that temperature. The solvent was evaporated under reduced pressure and the residue was redissolved in dichloromethane (3 mL) and 1N HCl (3 mL) was added. The reaction mixture was stirred at room temperature for 6 h before the pH was adjusted to 8 with saturated aquous NaHCO₃ solution. The layers were separated and the aqueous layer was extracted three times with dichloromethane. The combined organic layers were washed with water and brine, dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by column chromatography on silica-gel (Hexane/EtOAc, 3:1) to give (2*R*,3*S*)-**8a** (0.23 g, 82.1% yield).



yield: 82.1% (0.23 g as a colorless oil), ; $[\alpha]_D{}^{22} = +11.49$ (c 1.0, CHCl₃), (lit.¹² $[\alpha]_D{}^{21} = -9.1$ (c 0.86, CHCl₃)); ¹H NMR (500 MHz, CDCl₃) δ ² 7.96-7.98 (m, 2H), 7.55-7.58 (m, 1H), 7.41-7.45 (m, 2H), 7.31-7.37 (m, 4H), 7.25-7.27 (m, 1H), 5.49 (s, 2H), 3.76 (s, 3H), 1.42 (s, 9H), 1.23-

1.29 (brs, 1H).; ¹³C NMR (125 MHz, CDCl₃) δ 168.3, 165.5, 155.0, 138.0, 133.6, 130.0, 129.9, 129.8, 128.9, 128.7, 128.5, 128.0, 127.2, 126.5, 80.3, 75.3, 54.9, 52.7, 31.6, 28.3, 22.7, 14.1.; HRMS (EI): m/z calcd for C₂₂H₂₅NO₆ 399.1682, found 399.1690.

7-4. Synthesis of (2R,3S)-9a

KCN (14 mg, 0.22 mmol) was added to a stirred solution of (2R,3S)-**8a** (0.17 g, 0.44 mmol) in MeOH (2 mL). The resulting mixture was stirred at 65 °C for 2 h. After removal of the solvent, the residue was dissolved in CH₂Cl₂ and then washed with brine. The combined organic layers were dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by column chromatography on silica-gel (Hexane/EtOAc, 3:1) to give (2*R*,3*S*)-**9a** (0.11 g, 84.5% yield).



yield: 84.5% (0.11 g as a white solid), mp=113.2-116.2 °C, $[\alpha]_D^{23} = -5.49$ (c 1.0, CHCl₃), (lit.¹³ $[\alpha]_D^{23} = -6.7$ (c 0.85, CHCl₃)); ¹H NMR (500 MHz, CDCl₃) δ 7.37-7.39 (m, 4H), 7.30-7.33 (m, 1H), 5.41 (d, 1H, *J* = 8.6 Hz),

¹² For enantiomeric (2S,3R)-8. Bunnage, M. E.; Davies, S. G.; Goodwi, C. J., J. Chem. Soc., Perkin Trans. 1, 1994, 2385.

¹³ Harris, L.; Mee, S. P. H.; Furneaux, R. H.; Gainsford, G. J.; Luxenburger, A., *J. Org. Chem.*, 2011, **76**, 358.

5.24 (d, 1H, J = 9.2 Hz), 4.50 (s, 1H), 3.87 (s, 3H), 3.15 (d, 1H, J = 3.7 Hz), 1.45 (s, 9H).; ¹³C NMR (125 MHz, CDCl₃) δ 173.4, 155.1, 139.1, 128.6, 127.8, 126.7, 80.0, 73.5, 56.1, 28.3.; HRMS (ESI): m/z calcd for C₁₅H₂₁NO₅Na 318.1317, found 318.1313.

7-5. Synthesis of (2R,3S)-10, Toxotere side chain

To a solution of compound (2R,3S)-**9a** (100 mg, 0.34 mmol) in methanol (2 mL) and THF (2 mL) was added 1N NaOH (1 mL). After being stirred at room temperature for 0.5 h, the solution was concentrated and H₂O (10 mL) was added. The aqueous solution was extracted with dichloromethane (3x10 mL). The remaining aqueous layer was acidified to pH 3~4 with 1N HCl and extracted with ethyl acetate (6x10 mL). The combined ethyl acetate fractions were dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The resulting residue was washed with H₂O and collected via filtration affording (2*R*,3*S*)-**10** (83.5 mg, 88%).



yield: 88% (83.5 mg as a white solid), mp = 122.5-123.9 °C; $[\alpha]_D^{23}$ = +26.58 (c 1.0, MeOH), (lit.¹⁴ mp = 123.7-124.9 °C, $[\alpha]_D^{25}$ = +24.9 (c 1.0, MeOH)) ; ¹H NMR (500 MHz, DMSO-*d*₆) δ 7.27-7.31 (m, 4H), 7.20-7.23 (m, 1H), 7.06 (d, 1H, *J* = 9.4 Hz), 4.91-4.94 (dd, 1H, *J* = 3.5, 9.4 Hz),

4.17 (d, 1H, J = 3.5 Hz), 1.34 (s, 9H), 1.21-1.22 (brs, 1H); ¹³C NMR (125 MHz, DMSO d_6) δ 173.9, 155.4, 141.3, 128.4, 127.3, 127.3, 78.6, 74.2, 57.3, 28.6.; HRMS (ESI): m/z calcd for C₁₄H₁₉NO₅Na 304.1161, found 304.1159.

8. Synthesis of (-)-epi-Cytoxazone, (2R,3S)-12

Scheme S4.



¹⁴ Shen, X.; Yang, J.; Zhan, H.; Wang, H.; Wu, S.; Chen, Z. Chin. J. Chem. 2013, **31**, 31.

8-1. Synthesis of (4S,5S)-N-Boc-7j

The (4*S*,5*S*)-*N*-*Boc*-**7j** was prepared by using the procedure for the synthesis of (4*S*,5*S*)-*N*-*Boc*-**7a** as depicted in **7-1**.



(EI): m/z calcd for $C_{16}H_{21}NO_8S$ 387.0988, found 387.0991.

8-2. Synthesis of (2R,3S)-8j

The (2R,3S)-8j was prepared by using the procedure for the synthesis of (2R,3S)-8a as depicted in 7-3.



114.1, 55.3, 52.7, 28.3.; HRMS (EI): m/z calcd for C₂₃H₂₇NO₇ 429.1788, found 429.1781.

8-3. Synthesis of (2R,3S)-9j

The (2R,3S)-9j was prepared by using the procedure for the synthesis of (2R,3S)-9a as depicted in 7-4.



yield: 86.3% (63 mg as a white solid), mp = 127.1-129.2 °C, $[\alpha]_D^{20}$ = +3.9 (c 0.5, CHCl₃), (lit.¹⁵ mp = 110-112 °C, $[\alpha]_D^{23}$ = -3.8 (c 0.5, CHCl₃)); ¹H NMR (500 MHz, CDCl₃) δ 7.29 (d, 2H, *J* = 7.9 Hz), 6.88 (t, 1H, *J* = 8.3 Hz), 5.35 (d, 1H, *J* = 9.3 Hz), 5.15 (d, 1H, *J* = 8.8 Hz), 4.43 (brs, 1H), 3.83 (s, 3H), 3.79 (s, 3H), 3.20 (brs, 1H), 1.41 (s, 9H).;

¹⁵ For enantiomeric (2*S*,3*R*)-9j. Mishra, R. K.; Coates, C. M.; Revell, K. D.; Turos, E., Org. Lett., 2007, 9, 575.

¹³C NMR (125 MHz, CDCl₃) δ 173.4, 159.1, 155.1, 131.3, 127.9, 114.0, 79.9, 73.6, 55.6, 55.3, 53.0, 28.3.; HRMS (ESI); m/z calcd for $C_{16}H_{23}NO_6Na$ 348.1423, found 348.1417.

8-4. Synthesis of (2*R*,3*S*)-11

The (2R,3S)-**9j** (63mg, 0.19 mmol) was dissolved in ethanol (2 mL) and cooled to 0 °C. To this was added dropwise NaBH₄ (24 mg, 0.63 mmol) as a solution in ethanol (1 mL). After the addition was complete the ice bath was removed and the reaction mixture was stirred at room temperature for 4 h before the addition of a saturated aqueous NH₄Cl solution (5 mL). The ethanol was evaporated under reduced pressure. The remaining aqueous solution was extracted with ethyl acetate (3 x 5 mL) and the combined organic layers were washed with brine and dried over MgSO₄. Solvents were removed under reduced pressure and the residue was purified by silicagel chromatography eluting with CH₂Cl₂/MeOH (20/1), affording (2R,3S)-**11** as a white solid (51.7 mg, 91.7%).



yield: 91.7% (51.7 mg as a white solid), mp = 125.7-127.9 °C, $[\alpha]_D^{20}$ = +36.9 (c 0.4, CHCl₃), (lit.¹⁶ $[\alpha]_D^{23}$ = -36.1 (c 1.0, CHCl₃)); ¹H NMR (500 MHz, CD₃OD) δ 7.23 (d, 2H, *J* = 7.8 Hz), 6.86 (d, 2H, *J* = 7.6 Hz), 6.72 (brs, 1H), 4.62 (brs, 1H), 3.76 (s, 3H), 3.73 (brs, 1H), 3.43-3.46 (m, 1H), 3.35-3.39(m, 1H), 3.30 (s, 1H), 1.41 (s, 9H).; ¹³C NMR

(125 MHz, CDCl₃) δ 159.1, 156.9, 128.6, 127.9, 114.3, 114.2, 80.4, 75.3, 63.7, 55.3, 28.3.; HRMS (EI): m/z calcd for C₁₅H₂₃NO₅ 297.1576, found 297.1556.

8-5. Synthesis of (2R,3S)-12, (-)-epi-Cytoxazone.

To a stirred solution of (2R,3S)-11 (41 mg, 0.14 mmol) in dry THF (2 mL) at 0 °C was added sodium hydride (20 mg, 60% w/w in mineral oil, 0.45 mmol) at room temperature, and the reaction mixture was refluxed for 1 h. The reaction mixture was cooled to room temperature and quenched with a saturated aqueous NH₄Cl (10 mL), then extracted with ethyl acetate (2x10 mL). The organic layer was washed with water and brine, dried over anhydrous MgSO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica-gel (Hexane/EtOAc = 2:1) to give (2*R*,3*S*)-12 as a white solid (29 mg, 95.1%).

¹⁶ For enantiomeric (2*S*,3*R*)-**11**. Kim, S.-G.; Park, T.-H., *Tetrahedron Asymmetry*, 2008, **19**, 1626.

yield: 95.1% (29 mg as a white solid), mp = 141-142 °C, $[\alpha]_D^{19} = -28.2$ (c 0.5, MeOH), lit.¹⁷ $[\alpha]_D^{25} = -27.2$ (c 1.1, MeOH), lit.¹⁸ $[\alpha]_D^{28} = -22.8$ (c 0.5, MeOH); ¹H NMR (500 MHz, CD₃OD-*d*₄) δ 7.36 (d, 2H, *J* = 8.2 Hz), 7.02 (d, 2H, *J* = 8.1 Hz), 4.81 (d, 1H, *J* = 6.5 Hz), 4.38-4.39 (m, 1H), 3.88 (m, 1H), 3.86 (s, 3H), 3.73-3.76 (dd, 1H, *J* = 4.2, 12.5 Hz).; ¹³C NMR (125 MHz, CD₃OD-*d*₄) δ 160.1, 160.0, 132.2, 127.2, 114.0, 85.4, 61.1, 57.2, 54.4.; HRMS (EI): m/z calcd for C₁₁H₁₃NO₄ 223.0845, found 223.0829.

9. X-ray crystallography analysis data of (S,S)-7j

CCDC-1007235 contains the supplementary crystallographic data for (*S*,*S*)-**7j**. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre *via* www.ccdc.cam.ac.uk/data_request/cif.



 ¹⁷ Matsunaga, S.; Yoshida, T.; Morimoto, H.; Kumagai, N.; Shibasaki, M., J. Am. Chem. Soc., 2004, **126**, 8777.

¹⁸ Kim, I. S.; Kim, J. D.; Ryu, C. B.; Zee, O. P.; Jung, Y. H., *Tetrahedron*, 2006, **62**, 9349.

Crystal data and structure refinement for (S,S)-7j

Identification code	20131022lt_0m			
Empirical formula	C11 H13 N O6 S			
Formula weight	287.28			
Temperature	100(1) K			
Wavelength	0.71073 Å			
Crystal system	Orthorhombic			
Space group	P2(1)2(1)2(1)			
Unit cell dimensions	a = 5.3793(2) Å	<i>α</i> = 90°.		
	b = 13.1499(5) Å	β= 90°.		
	c = 17.4076(6) Å	$\gamma = 90^{\circ}.$		
Volume	1231.37(8) Å ³			
Z	4			
Density (calculated)	1.550 Mg/m^3			
Absorption coefficient	0.286 mm ⁻¹	0.286 mm ⁻¹		
F(000)	600	600		
Crystal size	0.25 x 0.20 x 0.08 mm ³	0.25 x 0.20 x 0.08 mm ³		
Theta range for data collection	1.94 to 28.47 $^\circ$	1.94 to 28.47°		
Index ranges	-7<=h<=7, -17<=k<=17	-7<=h<=7, -17<=k<=17, -22<=l<=23		
Reflections collected	34700			
Independent reflections	3107 [R(int) = 0.0235]			
Completeness to theta = 28.47°	99.8 %	99.8 %		
Absorption correction	Multi-scan	Multi-scan		
Max. and min. transmission	0.9775 and 0.9319	0.9775 and 0.9319		
Refinement method	Full-matrix least-squares	Full-matrix least-squares on F ²		
Data / restraints / parameters3107 / 0 / 172				
Goodness-of-fit on F ²	1.051			
nal R indices [I>2sigma(I)] R1 = 0.0253, wR2 = 0.0722				
R indices (all data)	dices (all data) $R1 = 0.0261, wR2 = 0.0732$			
Absolute structure parameter	-0.01(6)	-0.01(6)		
Largest diff. peak and hole	0.404 and -0.530 e.Å $^{-3}$	0.404 and -0.530 e.Å ⁻³		