SUPPORTING INFORMATION

High surface and magnetically recoverable mPANI/pFe₃O₄ nanocomposites for C-S bond formation in water

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General Information:

All reagents were commercial grade materials and were used without further purification. All solvents were dried and distilled by standard methods. Purification of products was carried out by column chromatography using commercial column chromatography grade silica gel (60-120 mesh) purchased from s. d. fine-chemicals Ltd. using mixture of ethyl acetate and hexane as eluting agent. All known compounds were characterized and compared with the literature reports. Visualization was accomplished with UV lamp.

The ¹H NMR and ¹³C NMR spectra were recorded on a Bruker-Avance (300 MHz), Varian-Gemini (200 MHz) and Varian-Inova (500 MHz) spectrophotometer, in CDCl₃ using TMS as the internal standard, Chemical shifts were reported in parts per million (ppm, δ) downfield from tetramethylsilane. All the chemicals were purchased from Sigma Aldrich, USA. and used without further purification. ¹H NMR and ¹³C NMR of the compounds were proved either by comparison to the known compounds or the synthesized compounds according to the literature.^[1-5] General Information: ACME SILICA GEL (100-200 mesh) was used for column chromatography and thin layer chromatography was performed on Merck precoated silica gel 60-F254 plates.

Procedure for the synthesis of mPANI/pFe₃O₄

[A] Synthesis and chemical modification of porous Fe₃O₄ nanoparticles

The magnetic Fe₃O₄ particles were prepared through a solvothermal reaction. FeCl₃·6H₂O (1.35 g) and sodium acetate (3.6 g) were dissolved in ethylene glycol (40 mL) under stirring condition. The homogeneous yellow solution was obtained and then transferred to a teflon-lined stainless-steel autoclave and sealed to heat at 200 °C. After 8 h, the autoclave was cooled to room temperature. The obtained black magnetite particles were washed with ethanol several times and then dried in vacuum at 60 °C for 12 h. Subsequently, the Fe₃O₄ particles were chemically modified by using APTMS. Typically, porous Fe₃O₄ particles (0.1 g) and APTMS (2 mL) were dissolved in anhydrous ethanol (50 mL). The mixture was refluxed for 12 h under dry nitrogen. The resulting modified porous Fe₃O₄ particles were separated with the help of an external magnet and then washed with ethanol. Finally, the product was dried in vacuum at 60 °C for 24 h to obtain the amine functionalized Fe_3O_4 particles (NH₂-Fe₃O₄).

[B] Synthesis of porous mPANI/pFe₃O₄ nanocomposites

The bifunctional mPANI/pFe₃O₄ nanocomposite was prepared by an in situ surface polymerization method in the presence of PVP and SDBS. In a typical procedure, PVP (0.05 g) and SDBS (0.01 g) were dissolved in 130 mL of deionized water, and then the NH₂-Fe₃O₄ particles (0.015 g) were added. The mixture was then ultrasonically dispersed, and a solution of aniline (20 μ L) in HCl (0.1 mL) was added into the mixture under vigorous stirring. Afterwards, the mixture was mechanically stirred for 30 min at 20 °C. Then an aqueous solution (20 mL) of APS (0.2 g) was added into the above mixture instantly to start the oxidative polymerization. The polymerization was performed under mechanical stirring for 5 h (stirring speed 2000 rotation per minute (rpm)). The resulting precipitates were washed with deionized water and ethanol several times. Finally, the solid was dried in vacuum at 60 °C for 24 h to obtain of the desired mPANI/pFe₃O₄ composite as a dark powder.



Figure 1. Step-wise preparation of the porous Fe₃O₄ nanoparticles@mesoporous PANI core-shell nanocomposites

XRD analysis:

X-ray diffraction (XRD) data were collected on a Simens/D-5000 diffractometer using Cu K α radiation. XPS spectra were recorded on a Kratos AXIS 165 with a dual anode (Mg and Al) apparatus using the Mg K α anode. The pressure in the spectrometer was about 10⁻⁹ torr. The XRD patterns of magnetic Fe₃O₄ nanoparticles reveal that the crystal structure is indexed as face centered cubic (fcc) (JCPDS card No. 19-629).



Figure 2. XRD pictures of [A] Fe₃O₄ nanoparticles, and [B] Fe₃O₄ @ mesoporous PANI.

No other characteristic peaks due to the impurities of hematite or hydroxides were detected (Fig 1). It is found that the strong diffraction peaks situated at 2Θ of 18.9° , 30.22° , 32.53° , 43.24° , 53.97° , 57.44° , and 67.94° , corresponding to the diffractions of [111], [220], [311], [400], [422], [511], and [440] crystal faces of porous Fe₃O₄ structures.

FT-IR analysis:

FT-IR spectra of these samples were recorded on KBr pellets by using a Nicolet NEXUS-FT IR 670 spectrometer (Nicolet Corporation Ltd. USA).







Figure 3. FTIR spectrum of Fe₃O₄ nanoparticles



Figure 3. FTIR spectrum of mPANI/pFe₃O₄

XPS analysis:

The XPS of the fresh pFe₃O₄@*m*PANI nano composites are shown in Figure 4. [A]. The XPS of C_{1s}, O_{2p} , N_{1s} and Fe_{2p} level give a proof for approximate chemical structure of mPANI coated pFe₃O₄ nanocomposites. Peaks at 284, 398, 538 eV were ascribed to the carbon, nitrogen and oxygen in polyaniline. The binding energies at 712 and 725 eV the characteristic peaks from Fe_{2p3/2} and Fe_{2p1/2} core level electron.





Figure 4. [A] XPS spectrum of the as-synthesized magnetite mPANI/ pFe₃O₄ nano composite. [B] XPS spectrum of the reused mPANI/ pFe₃O₄ nano composite after 5th cycle.

The binding energies for Fe_3O_4 of fresh mPANI/pFe₃O₄ nanocomposites (Figure 4. [A]) and reused mPANI/pFe₃O₄ nanocomposites (Figure 4. [B]) are in good agreement with the values reported for Fe_3O_4 in the literature.^[7] indicates that there is no chemical change occurred in $Fe_{2p3/2}$ and $Fe_{2p1/2}$ core level.

TEM analysis:

The TEM image of a Fe₃O₄ [A-C], Fe₃O₄@PVP+SDBS [D-F] and Fe₃O₄@mPANI[G-I] sample synthesized are shown in Figure 5, respectively. The image confirms the formation of low electron density spherical white spots of ~2.0 and ~6.0 nm sizes, corresponding to small to medium size mesopores formation in the bare Fe₃O₄ nanoparticles and mPANI/pFe₃O₄ microspheres. The presence of the pores for both the samples were further confirmed through N₂ adsorption-desorption experiments. Thus from the XRD patterns and the TEM image analysis, we could conclude porous Fe₃O₄ /mPANI nano composite microspheres.



Figure 5. TEM images of bare porous Fe₃O₄ nanoparticles [A-C], Fe₃O₄@PVP+SDBS [D-F] and mPANI/p Fe₃O₄ [G-I].

SEM:



Figure 6. [a] SEM images of the as- prepared magnetite mPANI/pFe₃O₄ nanocomposite. [b] and the mPANI/pFe₃O₄ nanocomposite after 5th cycle.

From the images of Scanning Electron Microscopy of freshly synthesized mPANI/pFe₃O₄ and recycled nano composite after its 5^{th} cycle, we observed that there is no change in morphology of the catalyst before the reaction and after its successive cycles.

[A] General procedure for the synthesis of unsymmetrical diaryl sulfides:

To develop new synthetic methods with inexpensive and efficient heterogeneous catalysts we have recently developed high surface mesoporous polyaniline/porous magnetic Fe₃O₄, mPANI/pFe₃O₄ nanocomposite for the *O*- arylation reaction of aryl chlorides with phenols.^[6] In continuation of our work, herein we report C-S cross coupling using mPANI/pFe₃O₄ as a catalyst. The catalyst was used for the S- arylation of thiolphenol with deactivated aryl chlorides also examined for the synthesis of

symmetrical diaryl sulfides from the reaction of aryl iodides with thiourea under mild reaction condition using water as green solvent.

The reaction vessel was charged with aryl chloride (1.0 mmol), thiophenol (1.0 mmol), KOH (1.5 mmol) and the catalyst (mPANI/pFe₃O₄) (25 mg, 5 mol %) in water (3 mL). The reaction mixture was then stirred for 8h at reflux temperature. The progress of the reaction was monitored by TLC. At the end of the reaction, the reaction mixture was allowed to cool to room temperature. Then reaction mixture was treated with excess of cold water, organic phase was extracted by adding ethyl acetate and dried over anhydrous Na_2SO_4 . The crude mixture was purified by chromatography on silica gel to afford the coupled product.

[B] General procedure for the synthesis of symmetrical diaryl sulfides:

A mixture of aryl iodide (1.0 mmol), thiourea (0.75 mmol), mPANI/pFe₃O₄ (25 mg, 6.6 mol %), and KOH (1.5 equiv.) was stirred at reflux temperature for 24h in water (3.0 mL) solvent. The progress of the reaction was monitored by TLC. When the reaction was complete, the reaction mixture was allowed to cool, a mixture of ethyl acetate and water (20 mL) were added and the catalyst was separated with the use of an external magnet. The organic solution was washed with brine water and dried with Na₂SO₄. The solvent and volatiles were completely removed under vacuum to give the crude product, which was purified by column chromatography on silica gel to obtain the desired symmetrical diaryl sulfide.

Reusability Study:

For any heterogeneous system, it is important to know its ease of separation and possible reuse. The mPANI/pFe₃O₄ catalyst can easily be separated by an external magnet. The recovered catalyst, after washing with ethyl acetate and acetone followed by drying at 65° C, was used in the next run and consistent activity was observed (for e.g Table 1, entry 1, 1st cycle 88%, 5th cycle 86%). Next to see whether the reaction was occurring mainly due to leached metal or the supported catalyst, a reaction between 4-nitro chlorobenzene and thiophenol was terminated after 20% conversion (80 min) and the

catalyst was separated using an external magnet and the reaction was continued with the filtrate for 12 hours. No change in the conversion of 4-nitro chlorobenzene was observed. These studies clearly prove that the *S*- arylation reaction occurs only heterogeneously.

Table 1.Reusability study of mPANI/pFe₃O₄ catalyst in the *S*-arylation of 4-nitro chlorobenzene with thiophenol $[A]^a$ and in synthesis symmetric diaryl sulfide from thiourea and 4-methoxy iodobenzene $[B]^b$.

S-arylation	No. of cycles	Fresh	Run 1	Run 2	Run 3	Run 4	Run 5
[A]	Yield [%] ^[c]	88	88	86	87	86	86
[B]	Yield [%] ^[c]	92	90	90	88	85	87

^a Reaction conditions: 4-nitro chlorobenzene (1.0 mmol), thiophenol (1.0 mmol), catalyst (25 mg, 5 mol%), KOH (1.5 equiv.), water (3.0 mL), reflux temperature for 8 h.

^b Reaction conditions: 4-methoxy iodobenzene (1.0 mmol), thiourea (0.75 mmol), catalyst (25 mg, 6.6 mol%), KOH (1.5 equiv.) water (3.0 mL), reflux temperature for 24 h.

^c Isolated yields of product.

Spectroscopic data of representative compounds:



Diphenylsulfane: (Table 2 and 3, entries 1 and 3)

¹H NMR (300 MHz, CDCl₃, ppm): δ = 7.47 – 7.16 (m, 10H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 137.06, 128.99, 127.50, 127.05. EI-MS (m/z) = 186.



Phenyl(*p*-tolyl)sulfane: (Table 2, entry 2)

¹HNMR (300 MHz, CDCl₃, ppm): δ = 7.33-7.23 (m, 4H), 7.20 – 7.15 (m, 3H), 7.12 (d, *J* = 7.9 Hz, 2H), 3.78 (s, 3H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 137.58, 137.03, 132.30, 131.30, 129.74, 129.01,126.41, 21.07. EI-MS (m/z) = 200.



(4-Methoxyphenyl)(phenyl)sulfane: (Table 2, entry 3)

¹H NMR (300 MHz, CDCl₃, ppm): $\delta = 7.37$ (d, J = 8.0 Hz, 2H), 7.24 – 7.15 (m, 2H), 7.10-7.05 (m, 3H), 6.85 (d, J = 8.7 Hz, 2H), 3.80 (s, 3H); ¹³C NMR (75 MHz, CDCl₃, ppm): $\delta = 159.8,138.53$, 129.04, 128.04, 128.25, 125.80, 124.16, 114.9, 55.2. EI-MS (m/z) = 216.



(4-Nitrophenyl)(phenyl)sulfane: (Table 2, entry 4)

¹H NMR (300 MHz, CDCl₃, ppm): $\delta = 8.07$ (d, J = 9.0 Hz, 2H), 7.60–7.36 (m, 5H), 7.18 – 7.14 (d, J = 9.0 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃, ppm): $\delta = 149.2$, 146.1, 136.3, 131.5, 130.1, 130.2, 128.2, 125.4. EI-MS (m/z) = 231.



2- nitro diphenyl sulfane (Table 2, entry 5)

¹HNMR (300 MHz, CDCl₃, ppm): δ = 7.42 (d, J = 7.5, 1H), 7.24-7.01 (m, 6H), 6.73 (d, J = 9.0, 2H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 148.73, 137.49, 131.13, 129.12, 126.5, 125.5, 118.75, 115.38. EI-MS (m/z) = 231.27.



3-methyl diphenyl sulfane (Table 2, entry 6)

¹HNMR (300 MHz, CDCl₃, ppm): δ = 7.34-7.23 (m, 4H), 7.22-7.10 (m, 4H), 7.02 (d, J = 7.25, 1H) 2.32 (S, 3H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 138.95, 136.06, 135.16, 131.75, 129.03, 128.28, 127.94, 126.75, 21.23. EI-MS (m/z) = 200.



(3,5-dimethylphenyl)(phenyl)sulfane (Table 2, entry 7)

¹HNMR (300 MHz, CDCl₃, ppm): δ = 7.33-7.16 (m, 5H), 6.97 (s, 2H), 6.86 (s, 1H) 2.29 (s, 6H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 138.8, 136.3, 134.7, 130.4, 129.1, 126.6, 21.4. EI-MS (m/z) = 214.



(4-Flurophenyl)(phenyl)sulfane: (Table 2, entry 8)

¹HNMR (300 MHz, CDCl₃, ppm): δ = 7.36 – 7.30 (m, 2H), 7.26 – 7.13 (m, 5H), 6.99 (t, *J* = 8.3 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ =163.9, 160.6, 136.6, 34.0, 133.9, 130.1, 129.8, 129.1, 126.7, 116.5, 116.2. EI-MS (m/z) = 204.



4-(Phenylthio)benzenamine: (Table 2, entry 9)

¹H NMR (300 MHz, CDCl₃, ppm): δ = 7.30 – 7.23(m, 2H), 7.19 – 7.12 (m, 2H), 7.09 – 7.01 (m, 3H), 6.61 (d, *J* = 8.05 Hz, 2H) 3.67 (bs, 2H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 146.6, 139.69, 136.02, 128.80, 127.41, 125.19, 121.02, 116.07. EI-MS (m/z) = 201.



(4-(Trifluoromethyl)phenyl)(phenyl)sulfane: (Table 2, entry 10)

¹HNMR (300 MHz, CDCl₃, ppm): δ = 7.50 – 7.42 (m, 4H), 7.39 – 7.31 (m, 3H), 7.24 (d, *J* = 7.6 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 142.89, 133.50, 132.71, 129.61, 128.71, 128.28, 127.79, 125.7, 122.28. EI-MS (m/z) = 254.



(4-*Tert*-butylphenyl)(phenyl)sulfane: (Table 2, entry 11)

¹HNMR (300 MHz, CDCl₃, ppm): $\delta = 7.28 - 7.16$ (m, 9H), 1.27 (s, 9H); ¹³C NMR (75 MHz, CDCl₃, ppm): $\delta = 150.43$, 136.54, 131.37, 130.14, 128.96, 128.36, 126.54, 126.12, 124.25, 34.55, 31.24. EI-MS (m/z) = 242.



Naphthalen-2-yl(phenyl)sulfane: (Table 2, entry 12)

¹H NMR (300 MHz, CDCl₃, ppm): δ = 7.85 – 7.71 (m, 4H), 7.46 – 7.18 (m, 8H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 138.6, 135.4, 134.5, 133.3, 131.7, 130.8, 129.9, 129.4, 128.2, 127.8, 127.5, 127.2, 125.8, 127.3. EI-MS (m/z) = 236.



3-(Phenylthio)pyridine: (Table 2, entry 13)

¹H NMR (300 MHz, CDCl₃, ppm): $\delta = 8.77 - 8.18$ (m, 2H), 7.55 (d, J = 7.3 Hz, 1H), 7.40 - 7.05 (m, 5H), 7.19 (dd, J = 5.12, 2.93 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃, ppm): $\delta = 149.6$, 137.1, 135.3, 130.2, 129.9, 129.5, 129.2, 125.8, 122.0. EI-MS (m/z) = 187.



Butyl(phenyl)sulfane: (Table 2, entry 14)

¹H NMR (300 MHz, CDCl₃, ppm): δ = 7.33–7.18 (m, 4H), 7.16–7.04 (m, 1H), 2.90 (t, *J* = 7.5, 6.7 Hz, 2H), 1.59 (m, 2H), 1.50 (m, 2H), 0.95 (t, *J* = 7.5, 6.7 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 138.2, 131.3, 130.0, 129.3, 128.1, 125.4, 34.3, 32.1, 22.2, 13.9. EI-MS *m/z* = 166.



Pentyl(phenyl)sulfane: (Table 2, entry 15)

¹H NMR (300 MHz, CDCl₃, ppm): δ = 7.33–7.06 (m, 5H), 2.87 (t, *J* = 7.5 Hz, 2H), 1.63 (p, J = 7.5 H

2H), 1.47-1.23 (m, 4H), 0.90 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 136.97, 128.81, 125.63, 33.45, 30.95, 28.80, 22.23, 14.06. EI-MS *m*/*z* = 180.



hexyl(phenyl)sulfane (Table 2, entry 16)

¹HNMR (300 MHz, CDCl₃, ppm): δ = 7.36-7.02 (m, 5H), 2.87 (t, *J* = 7.54, 2H), 1.69-1.56 (m, 2H) 1.47-1.22 (m, 6H), 0.90 (t, *J* = 7.54, 3H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 137.1, 128.9, 128.8, 125.6, 33.6, 31.3, 29.1, 28.5, 22.5, 14.0. EI-MS (m/z) = 214.



Bis(4-Methoxyphenyl)sulfane: (Table 3, entry 1)

¹HNMR (300 MHz, CDCl₃, ppm): δ = 7.60-7.43 (m, 4H), 6.97 (d, *J* = 7.9 Hz, 4H), 3.82 (s, 6H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 160.01, 134.26, 128.64, 115.19, 55.33. EI-MS (m/z) = 246.



Di-*p*-tolylsulfane: (Table 3, entry 2)

¹H NMR (300 MHz, CDCl₃, ppm): δ = 7.19 (d, *J* = 8.1 Hz, 4H), 7.08 (d, *J* = 7.9 Hz, 4H), 2.34 (s, 6H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 138.8, 131.84, 130.07, 129.20, 21.37. EI-MS (m/z) = 214.

NO₂ O_2N

Bis(3-Nitrophenyl)sulfane: (Table 3, entry 4):

¹HNMR (300 MHz, CDCl₃, ppm): δ = 8.76-8.23 (m, 4H), 7.82 (d, *J* = 7.85 Hz, 2H), 7.31 (t, *J* = 8.3 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ =149.10, 136.82, 131.26, 126.07, 122.47. EI-MS (m/z) = 276.



Bis(4-(trifluoromethyl)phenyl)sulfane: (Table 3, entry 5)

¹HNMR (300 MHz, CDCl₃, ppm): δ = 7.78-7.35 (m, 8H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 140.9, 136.4, 134.2, 129.8, 127.8, 124.4, 121.8. EI-MS (m/z) = 322.



Dithiophen-3-ylsulfane (Table 3, entry6)

¹HNMR (300 MHz, CDCl₃, ppm): δ = 7.33-7.27 (m, 2H), 7.15-7.10 (m, 2H), 7.0-6.92 (m, 2H); ¹³C NMR (75 MHz, CDCl₃, ppm): δ = 130.17, 126.76, 125.00. EI-MS (m/z) = 197.



Dithiophen-2-ylsulfane (Table 3, entry 7)

¹HNMR (300 MHz, CDCl₃, ppm): δ = 7.74-7.58 (m, 1H), 7.53-7.41 (m, 2H), 7.29-7.15 (m, 3H). EI-MS (m/z) = 197.



Dipyrazin-2-ylsulfane (Table 3, entry 8)

¹HNMR (300 MHz, CDCl₃, ppm): δ =8.75-8.68 (m, 2H), 8.50-8.38 (m, 4H); ¹³C NMR (75 MHz, CDCl₃,

ppm): δ = 147.12, 145.07, 143.04. EI-MS (m/z) = 190.

¹H NMR spectra of the products:























9.0 8.5 7.5 7.0 6.5 2.5 2.0 1.5 1.0 0.5 0.0 8.0 5.0 4.5 3.5 4.0 3.0 6.0 5.5 0.0









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¹³C NMR spectra of the products:





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150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

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