

Supplementary Information

Diastereo- and Enantioselective Reductive Amination of Cycloaliphatic Ketones by Preformed Chiral Palladium Complexes

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Table of contents:

General and materials.....	S2
Crystal structure of Pd[(<i>R</i>)-Tol-BINAP]Br ₂ (1e).....	S3
Preparation of complex and crystal structure of 1h	S3
Preparation of complex and crystal structure of 1i	S4
Preparation of complex 1j	S6
General procedure for reductive amination of cyclic ketones.....	S6
General procedure for asymmetric reductive amination of 2-methylcyclopentanone.....	S10
General procedure for asymmetric reductive amination of 2-methylcyclohexanone.....	S16
General procedure for asymmetric reductive amination of (<i>R</i>)-3-methylcyclohexanone.....	S19
General procedure for asymmetric reductive amination of other substituted rings.....	S21
References.....	S23
Scanned spectra of NMR and GC-MS (EI) or HPLC for all compounds.....	S24

General:

All reactions and manipulations were carried out by using Schlenk-type techniques. Flash column chromatography was performed on silica gel (70-230 mesh). ^1H NMR, ^{13}C NMR and ^{31}P NMR spectra were obtained on a JEOL GX-300, Bruker-Avance 300, Varian Unity 300 (300, 75 and 121 MHz respectively), and Varian Inova Plus 500 (500 for ^1H and 125 MHz for ^{13}C) spectrometers using TMS as the internal reference in CDCl_3 as solvent. All chemical shifts (δ) are reported in ppm and coupling constants (J) are reported in Hz to apparent peak multiplications. 2D NOESY, DEPT and $^1\text{H}/^{13}\text{C}$ HSQC sequences were used for help the assignments of the ^1H and ^{13}C spectra. IR spectra were recorded on a Nicolet FTIR Magna 750 spectrophotometer and the characteristic absorption frequencies are reported in cm^{-1} . Optical rotations were performed on a Perkin-Elmer 343 spectropolarimeter (589 nm). Mass spectra were obtained using a JEOL JMS-SX102A instrument with *m*-nitrobenzyl alcohol as the matrix (FAB^+ mode), and a JEOL JMS-AX505-A (EI mode at 70 eV). Elemental compositions were calculated within an uncertainty of 5 ppm by using the program installed in the computer system. The enantiomeric excess were determined by GC-MS analyses employing a Hewlett Packard 5890 (series II) instrument coupled with a JEOL JMS-AX505-A GC/MS-EI and Agilent Technologies 6890N coupled with JMS-GC/MS at 70 eV instruments employing a chiral capillary column Cyclodex- β (0.32 mm x 0.32 mm x 50 m) and He as a carrier gas. HPLC analyses were performed on a Hewlett Packard 1100 system with UV-DAD. Separations were achieved on a Daicel Chiracel OD-H (25 x 4.6mm) column. X-ray determination was collected on a Bruker SMART APEX CCD area diffractometer by the ω -scan method.

Materials:

All reagents were obtained from commercial suppliers and used without further purification. Diethyl ether and benzene were distilled from sodium-benzophenone and chloroform was distilled from P_2O_5 under nitrogen. Other solvents were HPLC grade. The $\text{Pd}(\text{MeCN})_2\text{Br}_2$, $\text{Pd}[(R)\text{-BINAP}]\text{Cl}_2$ (**1b**), $\text{Pd}[(R)\text{-BINAP}]\text{Br}_2$ (**1c**), $\text{Pd}[(S)\text{-BINAP}]\text{Br}_2$ (**1d**), $\text{Pd}[(R)\text{-Tol-BINAP}]\text{Br}_2$ (**1e**), $\text{Pd}[(S)\text{-Tol-BINAP}]\text{Br}_2$ (**1f**) and $\text{Pd}[(S,S)\text{-CHIRAPHOS}]\text{Br}_2$ (**1g**) complexes were prepared according to the literature procedures.¹⁻³

Crystal structure of Pd[(*R*)-Tol-BINAP]Br₂ (**1e**):

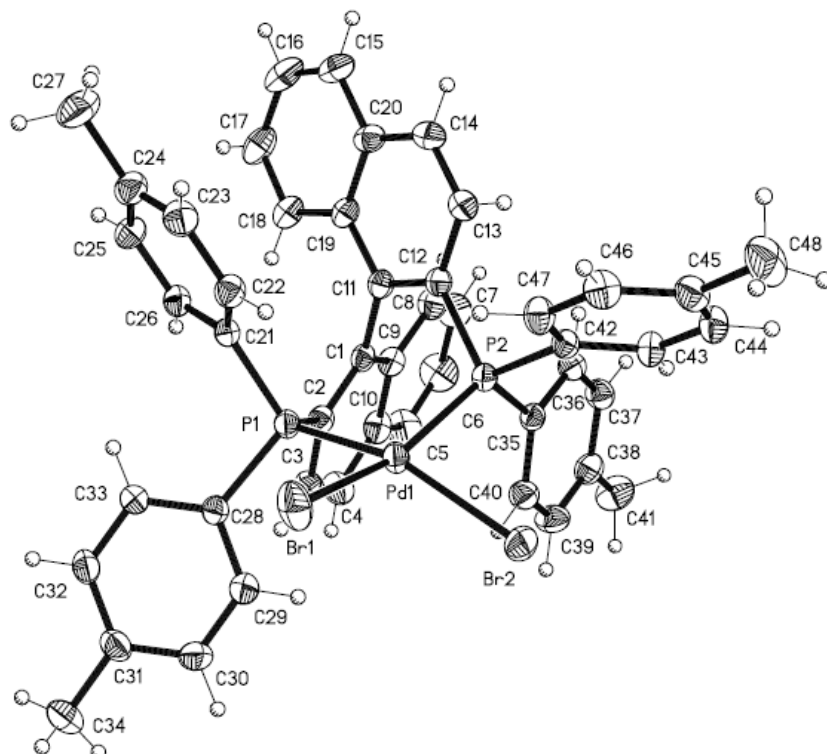
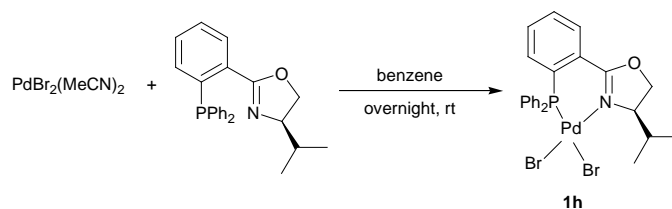


Figure S1. X-Ray diffraction structure of catalyst Pd[(*R*)-Tol-BINAP]Br₂ (**1e**). Selected bond lengths (Å): Pd(1)–Br(1) 2.4918(5); Pd(1)–Br(2) 2.4898(5); Pd(1)–P(1) 2.2473(1); Pd(1)–P(2) 2.2613(9); selected angles (°): P(1)–Pd(1)–Br(2) 158.08(3); P(2)–Pd(1)–Br(1) 159.00(3); P(1)–Pd(1)–P(2) 94.14(3); P(1)–Pd(1)–Br(1) 90.27(3); P(2)–Pd(1)–Br(2) 88.80(3); Br(2)–Pd(1)–Br(1) 94.733(1).

Preparation of complex **1h**:



This complex was synthesized by modified method described for the synthesis of PdCl₂[(*S*)-P[^]N] reported in the literature.⁴ In a Schlenk tube, PdBr₂(MeCN)₂ (174 mg, 0.5 mmol) was suspended in 10 mL of benzene. (*R*)-2-(2-(diphenylphosphino)phenyl)-4-isopropyl-1,3-oxazoline (186.5 mg, 0.5 mmol) was added. The suspension was stirred at room temperature for overnight. The yellow precipitate was collected by filtration, washed several times with diethyl ether and dried in vacuum. The complex was pure enough for further purposes, but it can be crystallized by the slow diffusion of diethyl ether into a concentrated solution of the solid in a mixture of dichloromethane:acetone (1:1) to obtain orange

crystals (yield: 82%). The presence of crystallization solvent in the product was ascertained by ^1H NMR spectroscopy. ^1H NMR (300 MHz, CDCl_3) δ 8.16-8.12 (m, 1H, ArH), 7.75-7.29 (m, 12H, ArH), 6.98-6.92 (m, 1H, ArH), 5.71-5.68 (m, 1H, -CHN), 4.51 (t, 1H, $J = 9.3$ Hz, $-\text{CH}_2\text{O}$), 4.36 (q, 1H, $J = 4.4$ Hz, $-\text{CH}_2\text{O}$), 2.60-2.54 (m, 1H, $\text{CH}(\text{CH}_3)_2$), 0.78 (d, 3H, $J = 7.1$ Hz, $-\text{CH}(\text{CH}_3)_2$), -0.03 (d, 3H, $J = 7.1$ Hz, $-\text{CH}(\text{CH}_3)_2$); ^{31}P NMR (121 MHz, CDCl_3) δ 26.7 (s, 1P, $\{(R)\text{-P}^*\text{N}\}$); IR (dissolution in CHCl_3 , cm^{-1}) 1624, 1570, 1247, 1100; MS (FAB $^+$) m/z : 638 $[\text{M} + 1]^+$, 560 (100), 478 (13), 402 (18); HRMS (FAB) m/z calcd for $\text{C}_{24}\text{H}_{25}\text{Br}_2\text{NOP}_2\text{Pd}$ $[\text{M} + 1]^+$ 637.9075, found 637.9076; $[\alpha]_{\text{D}}^{20}$ -258.5 (c 0.4, acetone).

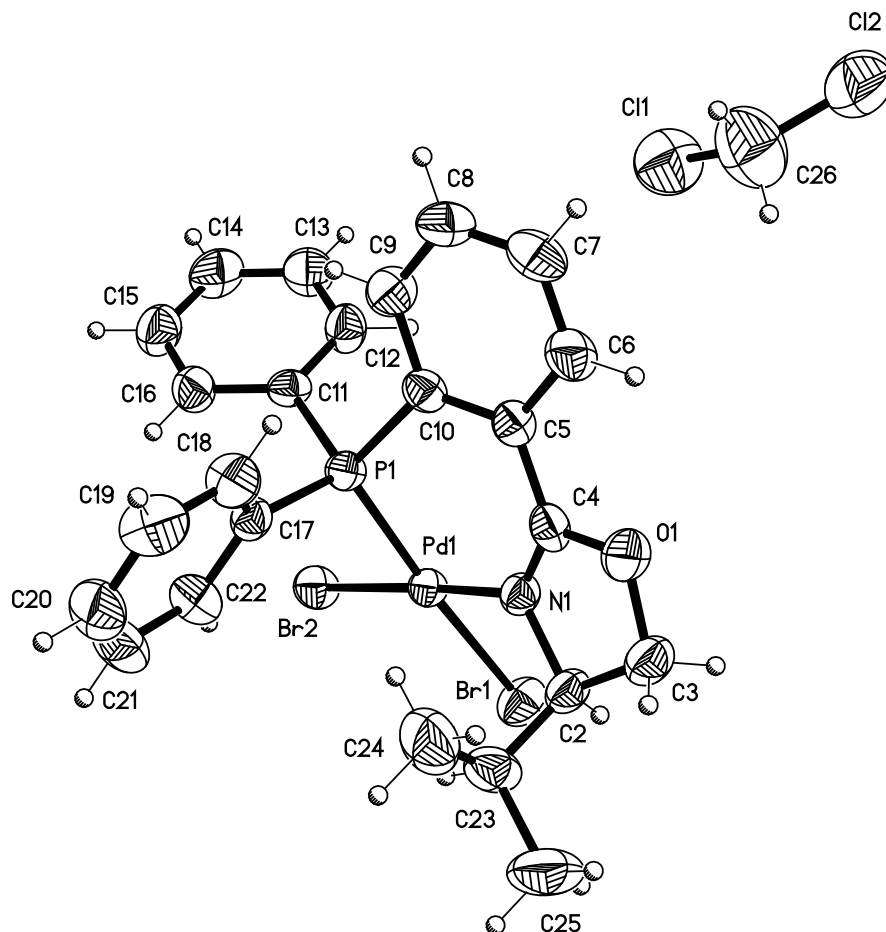
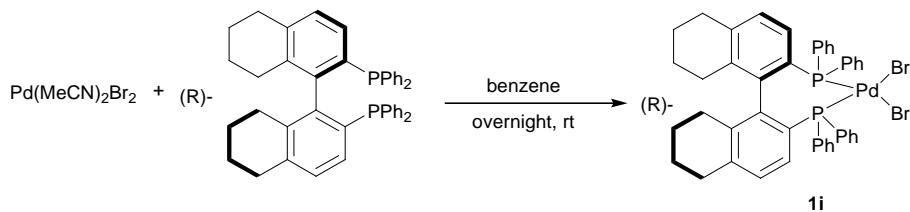


Figure S2. X-Ray diffraction structure of catalyst $\text{Pd}[(R)\text{-PHOX}]\text{Br}_2\cdot\text{CH}_2\text{Cl}_2$ (**1h**); selected bond lengths (\AA): Pd(1)–Br(1) 2.511(1); Pd(1)–Br(2) 2.414(1); Pd(1)–P(1) 2.223(1); Pd(1)–N(1) 2.048(3); selected angles ($^\circ$): N(1)–Pd(1)–P(1) 88.40(9); N(1)–Pd(1)–Br(2) 174.42(9); P(1)–Pd(1)–Br(2) 89.79(3); N(1)–Pd(1)–Br(1) 92.61(9); P(1)–Pd(1)–Br(1) 175.06(3); Br(2)–Pd(1)–Br(1) 89.645(1).

Preparation of complex 1i:



Complex **1i** was prepared by modified method described for the synthesis of Pd[(*R*)-BINAP]Cl₂ reported in the literature.² In a Schlenk tube, [(MeCN)₂]PdBr₂ (174 mg, 0.5 mmol) and (*R*)-(H₈)-BINAP (315 mg, 0.5 mmol) were suspended in 10 mL of benzene. The suspension was stirred at room temperature for overnight. The yellow-orange precipitate was collected by filtration, washed several times with diethyl ether and dried in vacuum. Yield: 75%. ³¹P NMR (121 MHz, CDCl₃) δ 24.40 (s, 2P, BINAP); ¹H NMR (300 MHz, CDCl₃) δ 7.87-7.74 (m, 8H, ArH), 7.36-7.18 (m, 12H, ArH), 6.99-6.93 (m, 2H, ArH), 6.65 (d, 2H, *J* = 8.1 Hz, ArH), 2.48-2.40 (m, 2H, -CH₂), 2.23-2.18 (m, 2H, -CH₂), 1.85-1.77 (m, 2H, -CH₂), 1.51-1.20 (m, 10H, -CH₂); FAB MS (positive ion mode): *m/z*: 815 [M⁺ - Br]; HRMS-FAB (*m/z*): calcd for C₄₄H₄₀BrP₂Pd [M - Br]⁺ 815.0823, found: 815.0827; [α]_D²⁰ +364 (c 0.2, CHCl₃).

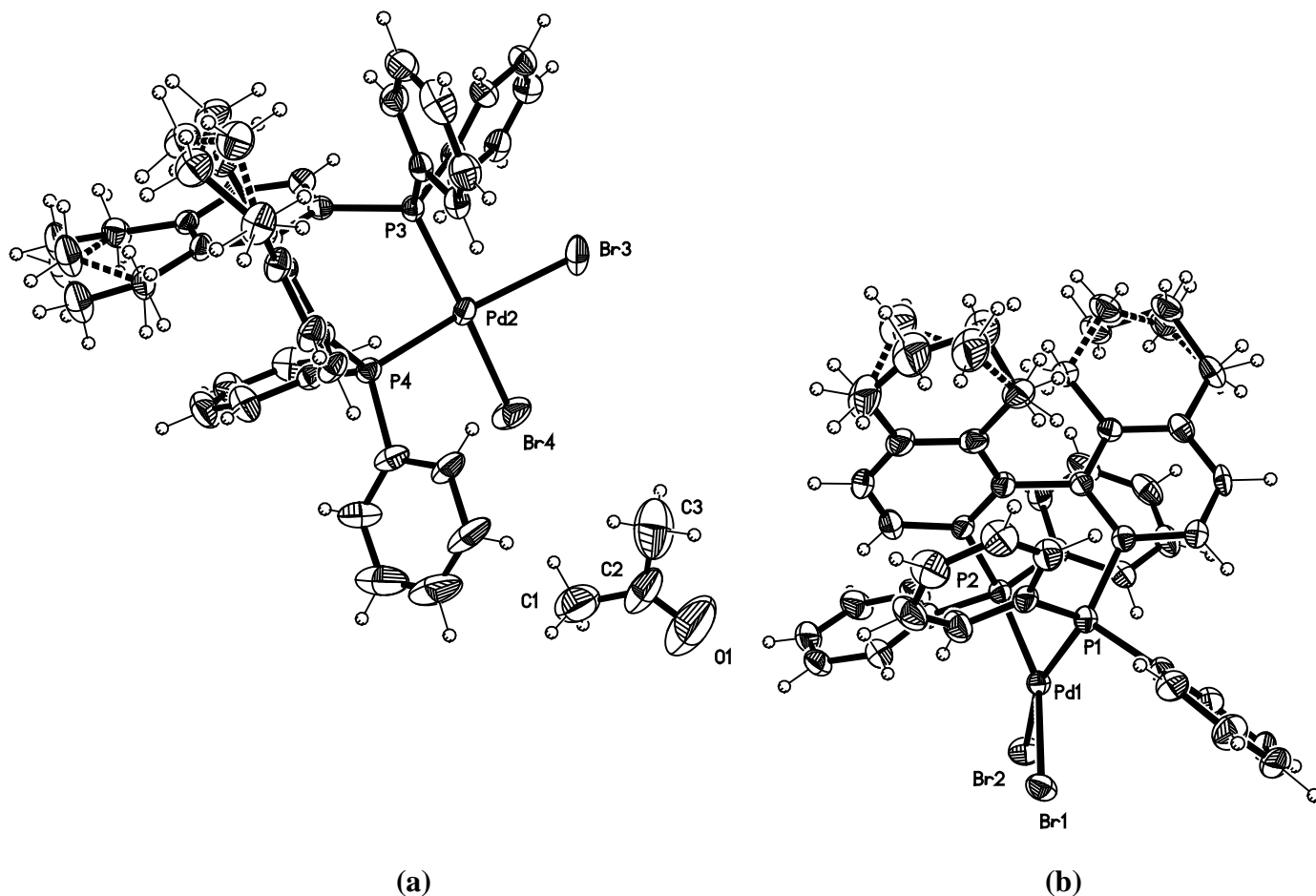
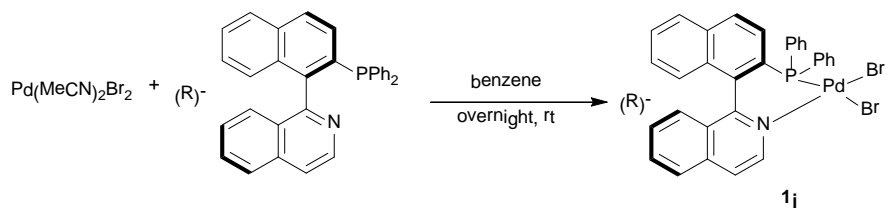


Figure S3. X-Ray diffraction structure of catalyst Pd[(*R*)-H₈-BINAP]Br₂.COMe₂ (**1h**); selected bond lengths (Å): (a) Pd(2)–Br(3) 2.4778(9); Pd(2)–Br(4) 2.4784(9); Pd(2)–P(3) 2.2731(1); Pd(2)–P(4) 2.2586(1); selected angles (°): P(4)–Pd(1)–Br(3) 169.15(5); P(3)–Pd(1)–Br(4) 163.97(5); P(4)–Pd(1)–P(3) 92.94(6); P(3)–Pd(2)–Br(3) 90.46(5); P(4)–Pd(2)–Br(4) 89.76(5); Br(3)–Pd(2)–Br(4) 89.81(4).

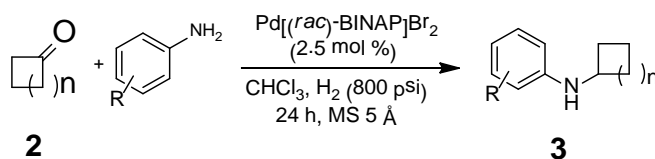
(b) Pd(1)–Br(1) 2.4842(8); Pd(1)–Br(2) 2.4948(8); Pd(1)–P(1) 2.2536(1); Pd(1)–P(2) 2.2632(1); selected angles (°): P(1)–Pd(1)–Br(2) 163.04(4); P(2)–Pd(1)–Br(1) 160.57(5); P(1)–Pd(1)–P(2) 93.00(6); P(1)–Pd(1)–Br(1) 88.95(5); P(2)–Pd(1)–Br(2) 90.66(5); Br(2)–Pd(1)–Br(1) 93.08(3).

Preparation of complex 1j:



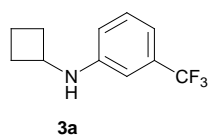
In a Schlenk tube, [(MeCN)₂]PdBr₂ (174 mg, 0.5 mmol) and (*R*)-QUINAP (219.5 mg, 0.5 mmol) were suspended in 10 mL of benzene. The suspension was stirred at room temperature for overnight. The yellow precipitate was collected by filtration, washed several times with diethyl ether and dried in vacuum. Yield: 68%. ¹H NMR (300 MHz, CDCl₃) δ 9.57 (d, 1H, *J* = 8.0 Hz, ArH), 8.05 (d, 2H, *J* = 8.0 Hz, ArH), 7.95 (d, 1H, *J* = 8.4 Hz, ArH), 7.61 (d, 1H, *J* = 8.5 Hz, ArH), 7.59-7.52 (m, 5H, ArH), 7.41-7.37 (m, 4H, ArH), 7.31-7.19 (m, 4H, ArH), 6.99-6.91 (m, 3H, ArH), 6.81 (d, 1H, *J* = 8.4 Hz, ArH); ³¹P NMR (121 MHz, CDCl₃) δ 35.43 (s, 1P, BINAP); FAB MS (positive ion mode): *m/z*: 706 [M⁺]; HRMS-FAB (*m/z*): calcd for C₄₄H₄₀BrP₂Pd [M⁺] 706.9204, found: 706.9209; ; [α]_D²⁰ +468 (c 0.21, CHCl₃).

General procedure for reductive amination of cyclic ketones.



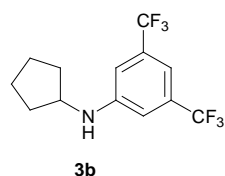
1.0 mmol of cycloaliphatic ketone and 1.3 mmol of aniline derivative were added to a stirred solution of 0.025 mmol of Pd[(*rac*)-BINAP]Br₂, **1k**, in 10 mL of dry chloroform (in a Schlenk tube) and stirred for 10 minutes. The solution was transferred to a 45 ml stainless steel autoclave (PARR) previously purged with vacuum-N₂ and containing 50 mg of MS 5Å. Subsequently, the reaction was taken to the desired pressure (800 psi H₂), stirred in an oil bath at 80°C for 24 h. At the end of this period, the gas was liberated. The solution was analyzed by GC-MS to quantify the remaining substrate, and was later

concentrated under reduced pressure, affording a crude residue, which was purified by column chromatography over silica gel (70-230 mesh), and eluted with hexane-ethyl acetate (99/1) to isolate the corresponding product.



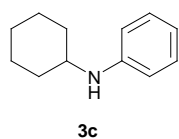
***N*-cyclobutyl-3-(trifluoromethyl)benzenamine (3a)**. Prepared according to the general procedure from 2-cyclobutanone (75 μ L, 1.0 mmol), *m*-trifluoromethyl aniline (162 μ L, 1.3 mmol) and Pd[(*rac*)-BINAP]Br₂, **1k**, (22 mg, 0.025 mmol) at 80

$^{\circ}$ C for 24 h, to provide the title compound as colorless oil (95%). ¹H NMR (300 MHz, CDCl₃) δ 7.14 (t, 2H, *J* = 7.8 Hz, ArH), 6.82 (d, 1H, *J* = 7.5 Hz, ArH), 6.64 (s, 1H, ArH), 6.57 (d, 1H, *J* = 8.1 Hz, ArH), 3.89-3.78 (m, 2H, -NHCH + -NHCH), 2.39-2.30 (m, 2H, -CH₂), 1.80-1.67 (m, 4H, -CH₂); ¹³C NMR (75 MHz, CDCl₃) δ 147.3, 131.5 (q, *J* = 31.6 Hz), 129.6, 124.5 (d, *J* = 272.0 Hz), 115.8, 113.5 (q, *J* = 3.9 Hz), 108.9 (q, *J* = 3.9 Hz), 48.7 (-NHCH), 31.0 (-2CH₂), 15.2 (-CH₂); IR (film, cm⁻¹) 3418, 2926, 2855, 1615, 1517; MS (EI) *m/z*: 215 (M⁺), 187 (100), 166 (29), 145 (21); HRMS (EI) *m/z* calcd for C₁₁H₁₂NF₃ (M⁺) 215.0922, found 215.0922.



***N*-cyclopentyl-3,5-bis(trifluoromethyl)benzenamine (3b)**. Prepared according to the general procedure from cyclopentanone (88 μ L, 1.0 mmol), 3,5-bis(trifluoromethyl)aniline (203 μ L, 1.3 mmol) and Pd[(*rac*)-BINAP]Br₂, **1k**, (22 mg, 0.025 mmol) at 80 $^{\circ}$ C for 24 h, to provide the title compound as colorless oil

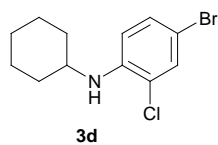
(93%). ¹H NMR (300 MHz, CDCl₃) δ 7.14 (s, 1H, ArH), 6.95 (s, 2H, ArH), 4.14 (bs, 1H, -NHCH), 3.87 – 3.83 (m, 1H, -NHCH), 2.14-2.05 (m, 2H, -CH₂), 1.82-1.67 (m, 4H, -CH₂), 1.57-1.48 (m, 2H, -CH₂); ¹³C NMR (75 MHz, CDCl₃) δ 148.4, 132.3 (q = 32.4 Hz), 123.6 (q = 272.5 Hz), 112.0 (q = 3.0 Hz), 109.5 (q = 3.9 Hz), 54.5 (-NHCH), 33.2 (-CH₂), 23.9 (CH₂); IR (film, cm⁻¹) 3414, 2925, 2848, 1615, 1517; MS (EI) *m/z*: 297 (M⁺), 78 (13), 268 (100), 255 (16), 242 (11), 213 (14); HRMS (EI) *m/z* calcd for C₁₃H₁₃NF₆ (M⁺) 297.0957, found 297.0952.



***N*-cyclohexylbenzenamine (3c)**. Prepared according to the general procedure from cyclohexanone (103 μ L, 1.0 mmol), 3,5-bis(trifluoromethyl)aniline (203 μ L, 1.3 mmol) and Pd[(*rac*)-BINAP]Br₂, **1k**, (22 mg, 0.025 mmol) at 80 $^{\circ}$ C for 24 h, to provide the

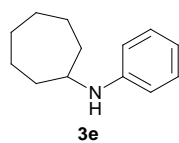
title compound as colorless oil (89%). ¹H NMR (300 MHz, CDCl₃) δ 7.17-7.10 (m, 2H, ArH), 6.67-6.55 (m, 3H, ArH), 3.48 (bs, 1H, -NHCH), 3.28-3.19 (m, 1H, -NHCH), 2.08-2.01 (m, 2H, -CH₂), 1.79-1.72 (m, 2H, -CH₂), 1.67-1.61 (m, 1H, -CH₂), 1.43-1.07 (m, 5H, -CH₂); ¹³C NMR (75 MHz, CDCl₃) δ 147.3, 116.7, 113.1, 51.6 (-NHCH), 33.4 (-CH₂), 25.9 (-CH₂), 25.0 (-CH₂); EM-IE (70 eV)

m/z : 175 (M^+), 146 (8), 132 (100), 119 (22); HRMS (EI) m/z calcd for $C_{12}H_{17}N_1$ (M^+) 175.1361, found 175.1356.



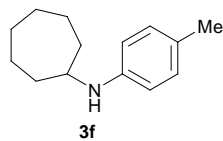
4-bromo-2-chloro-N-cyclohexylbenzenamine (3d). Prepared according to the general procedure from cyclohexanone (103 μ L, 1.0 mmol), 4-bromo-2-chloroaniline (266.5 mg, 1.3 mmol) and Pd[(*rac*)-BINAP]Br₂, **1k**, (22 mg, 0.025 mmol) at 80 °C

for 24 h, to provide the title compound as colorless oil (74%). ¹H NMR (300 MHz, CDCl₃) δ 7.38 (s, 1H, ArH), 7.22 (dd, 1H, J = 8.7 Hz, ArH), 6.55 (s, 1H, ArH), 4.23 (d, 1H, J = 6.60 Hz, -NHCH), 3.33-3.24 (m, 2H, -NHCH), 2.07-1.84 (m, 2H, -CH₂), 1.82-1.78 (m, 2H, -CH₂), 1.71-1.64 (m, 1H, -CH₂), 1.48-1.17 (m, 5H, -CH₂); ¹³C NMR (75 MHz, CDCl₃) δ 142.3, 131.4, 130.4, 119.5, 112.6, 106.7, 51.5 (-NHCH), 33.0 (-CH₂), 25.8 (CH₂), 24.8 (CH₂); IR (film, cm⁻¹) 3415, 2927, 2850, 1615, 1517; MS (EI) m/z : 287 (M^+), 187 (100), 166 (29), 145 (21); HRMS (EI) m/z calcd for $C_{12}H_{15}NBrCl$ (M^+) 287.0076, found 287.0070.



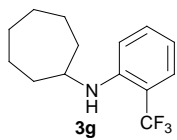
N-phenylcycloheptanamine (3e). Prepared according to the general procedure from cycloheptanone (126 μ L, 1.0 mmol), aniline (120 μ L, 1.3 mmol) and Pd[(*rac*)-BINAP]Br₂, **1k**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to provide the title compound

as colorless oil (92%). ¹H NMR (300 MHz, CDCl₃) δ 7.24-7.18 (m, 2H, ArH), 6.73-6.69 (m, 1H, ArH), 6.65-6.58 (m, 2H, ArH), 3.63 (bs, 1H, -NHCH), 3.54-3.47 (m, 1H, -NHCH), 2.09-2.01 (m, 2H, -CH₂), 1.76-1.45 (m, 10H, -CH₂); ¹³C NMR (75 MHz, CDCl₃) δ 147.4, 29.4, 116.9, 113.3, 53.7 (-NHCH), 39.9 (-CH₂), 28.5 (-CH₂), 24.5 (-CH₂); IR (film, cm⁻¹) 3404, 2925, 2854, 1602, 1503; MS (EI) m/z : 189 (M^+), 146 (14), 132 (100), 120 (38); HRMS (EI) m/z calcd for $C_{13}H_{19}N_1$ (M^+) 189.1517, found 189.1520.

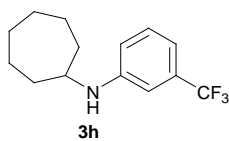


N-p-tolylcycloheptanamine (3f). Prepared according to the general procedure from cycloheptanone (126 μ L, 1.0 mmol), *p*-toluidine (139 mg, 1.3 mmol) and Pd[(*rac*)-BINAP], Br₂**1k**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to provide the title

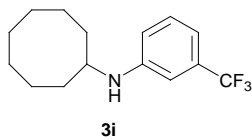
compound as colorless oil (91%). ¹H NMR (300 MHz, CDCl₃) δ 6.97 (d, 2H, J = 7.9 Hz, ArH), 6.51 (d, 2H, J = 8.2 Hz, ArH), 3.76 (bs, 1H, -NHCH), 3.45-3.38 (m, 1H, -NHCH), 2.23 (s, 3H, -Me), 2.03-1.95 (m, 2H, -CH₂), 1.70-1.39 (m, 10H, -CH₂); ¹³C NMR (75 MHz, CDCl₃) δ 144.6, 129.8, 126.5, 113.9, 54.4 (-NHCH), 34.7 (-CH₂), 28.4 (-CH₂), 24.4 (-CH₂), 20.4 (-Me); IR (film, cm⁻¹) 3400, 2924, 2855, 1617, 1517; MS (EI) m/z : 203 (M^+), 160 (5), 146 (25), 68 (100); HRMS (EI) m/z calcd for $C_{14}H_{21}N_1$ (M^+) 203.1674, found 203.1678.



***N*-(2-(trifluoromethyl)phenyl)cycloheptanamine (3g)**. Prepared according to the general procedure from cycloheptanone (118 μL , 1.0 mmol), *o*-trifluoromethyl aniline (163 μL , 1.3 mmol) and Pd[(*rac*)-BINAP]Br₂, **1k**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to provide the title compound as colorless oil (86%). ¹H NMR (300 MHz, CDCl₃) δ 7.45 (dd, 1H, *J* = 7.96, 1.20 Hz, ArH), 7.37 (t, 1H, *J* = 7.81 Hz, ArH), 6.71-6.66 (m, 2H, ArH), 4.32 (bs, 1H, -NHCH), 3.59-3.56 (m, 1H, -NHCH), 2.07-2.00 (m, 2H, -CH₂), 1.76-1.55 (m, 10H, -CH₂); ¹³C NMR (75 MHz, CDCl₃) δ 144.8, 132.9, 126.7 (*q* = 5.5 Hz), 125.3 (*d* = 272.1 Hz), 115.1, 113.1 (*q* = 29.0 Hz), 53.3 (-NHCH), 34.5 (-CH₂), 28.2 (CH₂), 24.2 (CH₂); IR (film, cm⁻¹) 3414, 2925, 2849, 1615, 1517; MS (EI) *m/z*: 257 (M⁺, 83), 238 (13), 228 (14), 214 (24), 200 (100); HRMS (EI) *m/z* calcd for C₁₄H₁₈NF₃ (M⁺) 257.1391, found 257.1389.

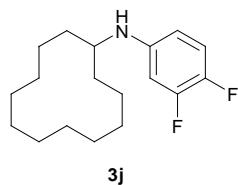


***N*-(3-(trifluoromethyl)phenyl)cycloheptanamine (3h)**. Prepared according to the general procedure from cycloheptanone (118 μL , 1.0 mmol), *m*-trifluoromethylaniline (162 μL , 1.3 mmol) and Pd[(*rac*)-BINAP]Br₂, **1k**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to provide the title compound as colorless oil (88%). ¹H NMR (300 MHz, CDCl₃) δ 7.25 (t, 1H, *J* = 7.9 Hz, ArH), 6.91 (d, 1H, *J* = 7.7 Hz, ArH), 6.76 (s, 1H, ArH), 6.68 (dd, 1H, *J* = 8.3, 2.2 Hz, ArH), 3.84 (bs, 1H, -NHCH), 3.53-3.46 (m, 1H, -NHCH), 2.05-1.97 (m, 2H, -CH₂), 1.74-1.50 (m, 10H, -CH₂); ¹³C NMR (75 MHz, CDCl₃) δ 147.5, 131.6 (*q*, *J* = 32.4 Hz), 129.7, 124.5 (*d*, *J* = 272.2 Hz), 116.0, 113.1 (*q*, *J* = 3.4 Hz), 109.3 (*q*, *J* = 3.4 Hz), 53.6 (-NHCH), 34.7 (-CH₂), 28.4 (-CH₂), 24.3 (-CH₂); IR (film, cm⁻¹) 3425, 2929, 2857, 1614, 1515; MS (EI) *m/z*: 257 (M⁺), 228 (3), 214 (24), 200 (100), 187 (19). HRMS (EI) *m/z* calcd for C₁₄H₁₈NF₃ (M⁺) 257.1391, found 257.1388.



***N*-(3-(trifluoromethyl)phenyl)cyclooctanamine (3i)**. Prepared according to the general procedure from cyclooctanone (132 μL , 1.0 mmol), *m*-trifluoromethyl aniline (162 μL , 1.3 mmol) and Pd[(*rac*)-BINAP]Br₂, **1k**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to provide the title compound as colorless oil (88%). ¹H NMR (300 MHz, CDCl₃) δ 7.13 (dd, 1H, *J* = 7.8 Hz, ArH), 6.78 (d, 1H, *J* = 7.8 Hz, ArH), 6.64 (s, 1H, ArH), 6.57 (dd, 1H, *J* = 8.1, 2.0 Hz, ArH), 3.70 (bs, 1H, -NHCH), 3.45-3.38 (m, 1H, -NHCH), 1.83-1.76 (m, 2H, -CH₂), 1.66-1.44 (m, 12H, -CH₂); ¹³C NMR (75 MHz, CDCl₃) δ 147.4, 131.5 (*q*, *J* = 31.5 Hz), 129.6, 124.0 (*d*, *J* = 272.0 Hz), 115.9, 112.9 (*q*, *J* = 3.8 Hz), 109.2 (*q*, *J* = 3.8 Hz), 52.4 (-NHCH), 32.3 (-CH₂), 27.1 (-CH₂), 25.8 (-CH₂), 23.9 (-CH₂); IR (film, cm⁻¹) 3416, 2926, 2850, 1614, 1515; MS (EI) *m/z*: 271 (M⁺), 252 (5),

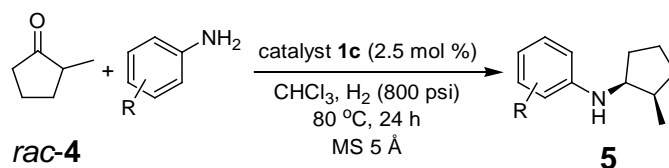
228 (8), 214 (7), 200 (100), 187 (31); HRMS (EI) m/z calcd for $C_{15}H_{20}F_3N$ (M^+) 271.1548, found 271.1550.



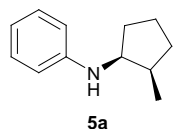
***N*-(3,4-difluorophenyl)cyclododecanamine (3j)**. Prepared according to the general procedure from cyclododecanone (182 mg, 1.0 mmol), 3,4-difluoromethyl aniline (130 μ L, 1.3 mmol) and Pd[(*rac*)-BINAP]Br₂, **1k**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to provide the title compound as colorless oil (81%).

¹H NMR (500 MHz, CDCl₃) δ 6.92 (ddd, 1H, $J_{H-H} = 9.0$, $J_{H-F} = 9.0$, 1.5 Hz, ArH), 6.35 (ddd, 1H, $J_{H-H} = 9.5$, $J_{H-F} = 6.5$, 3.0 Hz, ArH), 6.23-6.19 (m, 1H, ArH), 3.39-3.35 (m, 2H, -NHCH + -NHCH), 1.636-1.57 (m, 1H, -CH₂), 1.49-1.24 (m, 21H, -CH₂). ¹³C NMR (125 MHz, CDCl₃) δ 150.9 (dd = 243.7, 13.7 Hz), 144.8 (d = 8.7Hz), 142.5 (dd = 233.7, 12.5 Hz), 117.3 (d = 12.5 Hz), 108.0 (d = 12.5 Hz), 101.3 (d = 25.0 Hz), 50.2 (-NHCH), 40.3 (-CH₂), 29.5 (-CH₂), 24.3 (-CH₂), 24.0 (-CH₂), 23.2 (-CH₂), 23.1(-CH₂), 21.1 (-CH₂); IR (film, cm⁻¹) 3475, 2955, 2873, 1614, 1518; MS (EI) m/z : 295 (M^+ , 100), 266 (3), 252 (8), 238 (4), 224 (8), 168 (71); HRMS (EI) m/z calcd for $C_{18}H_{27}F_2N$ (M^+) 295.2112, found 295.2115.

General procedure for asymmetric reductive amination of 2-methylcyclopentanone.

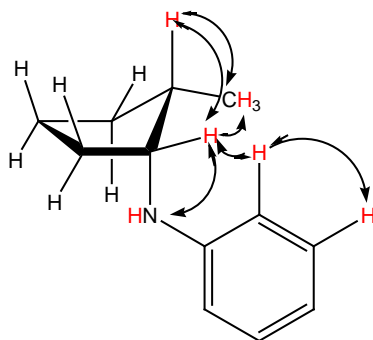


1.0 mmol of *rac*-**4** and 1.3 mmol of aniline derivative were added to a stirred solution of 0.025 mmol of preformed chiral palladium complex in 10 mL of dry chloroform (in a Schlenk tube) and stirred under nitrogen atmosphere for 10 minutes. The solution was transferred to a 45 ml stainless steel autoclave (PARR) previously purged with vacuum-N₂. Subsequently, the reaction was taken to the desired pressure (900 psi H₂), stirred in an oil bath at 80°C for 24 h. At the end of this period, the gas was liberated. The solution was analyzed by NMR to quantify the remaining substrate and d.r. respectively, and was later concentrated under reduced pressure, affording a crude residue, which was purified by column chromatography over silica gel (70-230 mesh), and eluted with hexane-ethyl acetate (99/1) to isolate the corresponding product.



***cis*-N-(phenyl)-2-methylcyclopentanamine (5a)**. Prepared according to the general procedure from 2-methylcyclopentanone (106 μ L, 1.0 mmol), aniline (120 μ L, 1.3

mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to provide the title compound as light yellow oil (90%). ¹H NMR (500 MHz, CDCl₃) δ 7.16-7.12 (m, 2H, ArH), 6.64 (t, 1H, *J* = 7.8 Hz, ArH), 6.60 (dd, 2H, *J* = 1.0, 7.8 Hz, ArH), 3.72 (m, 1H, -NHCH), 3.56 (bs, 1H, -NHCH), 2.31-2.22 (m, 1H, -CHCH₃), 2.02-1.36 (m, 6H, -CH₂), 0.88 (d, 3H, *J* = 7.0 Hz, -CHCH₃); ¹³C NMR (125 MHz, CDCl₃) δ 148.2, 129.1, 116.6, 112.9, 57.2 (-NHCH), 35.8 (-CHCH₃), 32.0 (-CH₂), 31.5 (-CH₂), 21.2 (-CH₂), 14.3 (-CHCH₃); IR (film, cm⁻¹) 3412, 2957, 2871, 1603, 1507; MS (EI) *m/z*: 175 (M⁺), 146 (19), 132 (100), 119 (21); HRMS (EI) *m/z* calcd for C₁₂H₁₇N(M⁺) 175.1361, found 175.1364; [α]_D²⁰ +9.2 (c 0.5, CHCl₃); ee = 98% by GC-MS (EI) [chiral column: Cyclodex-β; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 31.3 min (minor), t_R = 32.2 min (major)]. The relative stereochemistry was assigned by NOE analysis.



Interactions observed by NOE.

With Pd/C the selectivity *cis/trans* was 62/38.

With catalyst Pd[(*R*)-BINAP]Cl₂, **1b**: [α]_D²⁰ +7.8 (c 0.48, CHCl₃); ee = 92% by GC-MS (EI) [chiral column: Cyclodex-β; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 33.1 min (minor), t_R = 34.3 min (major)].

With catalyst Pd[(*S*)-BINAP]Br₂, **1d**: [α]_D²⁰ -11.2 (c 0.5, CHCl₃); ee = 95% by GC-MS (EI) [chiral column: Cyclodex-β; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 31.4 min (major), t_R = 32.3 min (minor)].

With catalyst Pd[(*R*)-Tol-BINAP]Br₂, **1e**: [α]_D²⁰ +8.1 (c 0.5, CHCl₃); ee = 89% by GC-MS (EI) [chiral column: Cyclodex-β; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 32.2 min (minor), t_R = 33.3 min (major)].

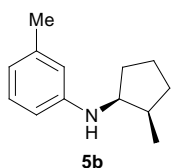
With catalyst Pd[(*S*)-Tol-BINAP]Br₂, **1f**: [α]_D²⁰ -10.9 (c 0.51, CHCl₃); ee = 93% by GC-MS (EI) [chiral column: Cyclodex-β; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 33.2 min (major), t_R = 34.3 min (minor)].

With catalyst Pd[(*S,S*)-CHIRAPHOS]Br₂, **1g**: [α]_D²⁰ +8.6 (c 0.49, CHCl₃); ee = 96% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 33.4 min (minor), t_R = 34.5 min (major)].

With catalyst Pd[(*R*)-PHOX]Br₂, **1h**: [α]_D²⁰ -7.2 (c 0.53, CHCl₃); ee = 87% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 33.1 min (minor), t_R = 34.2 min (major)].

With catalyst Pd[(*R*)-H₈BINAP]Br₂, **1i**: [α]_D²⁰ +9.0 (c 0.5, CHCl₃); ee = 98% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 31.3 min (minor), t_R = 32.2 min (major)].

With catalyst Pd[(*R*)-QUINAP]Br₂, **1b**: [α]_D²⁰ -8.5 (c 0.51, CHCl₃); ee = 89% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 32.4 min (minor), t_R = 33.8 min (major)].

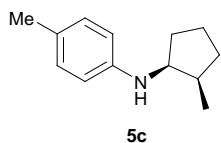


***cis*-N-(3-methylphenyl)-2-methylcyclopentaneamine (5b)**. Prepared according to the

general procedure from 2-methylcyclopentanone (106 μ L, 1.0 mmol), *m*-toluidine (140 μ L, 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to

provide the title compound as light yellow oil (90%). ¹H NMR (300 MHz, CDCl₃) δ

7.10 (t, 1H, *J* = 7.9 Hz, ArH), 6.55-6.47 (m, 3H, ArH), 3.77 (m, 1H, -NHCH), 3.63 (bs, 1H, -NHCH), 2.37-2.27 (m, 4H, -CHCH₃ + -Me), 2.10-1.39 (m, 6H, -CH₂), 0.93 (d, 3H, *J* = 6.8 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 148.3, 139.0, 129.2, 117.8, 113.8, 110.2, 57.2 (-NHCH), 35.8 (-CHMe), 32.1 (-CH₂), 31.6 (-CH₂), 21.8 (-CH₃), 21.3 (-CH₂), 14.5 (-CHCH₃); IR (film, cm⁻¹) 3410, 2955, 2868, 1604, 1511; MS (EI) *m/z*: 189 (M⁺), 160 (12), 146 (100), 133 (22); HRMS (EI) *m/z* calcd for C₁₃H₁₉N(M⁺) 189.1517, found 189.1515; [α]_D²⁰ +8.2 (c 0.5, CHCl₃); ee = 93% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 30.2 min (minor), t_R = 31.2 min (major)].

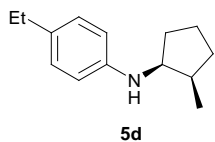


***cis*-N-(4-methylphenyl)-2-methylcyclopentaneamine (5c)**. Prepared according to

the general procedure from 2-methylcyclopentanone (106 μ L, 1.0 mmol), *p*-toluidine (139 mg, 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at

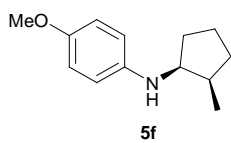
80 °C for 24 h, to provide the title compound as yellow oil (91%). ¹H NMR (500 MHz, CDCl₃) δ 6.95 (d, 2H, *J* = 8.0 Hz, ArH), 6.53 (d, 2H, *J* = 8.5 Hz, ArH), 3.69 (m, 1H, -NHCH), 3.35 (bs, 1H, -NHCH), 2.29-2.21 (m, 4H, -CHCH₃ + -Me), 2.00-1.35 (m, 6H, -CH₂), 0.87 (d, 3H, *J* = 7.0 Hz, -CHCH₃); ¹³C

NMR (125 MHz, CDCl₃) δ 145.9, 129.6, 125.8, 113.1, 57.5 (-NHCH), 35.7 (-CHCH₃), 31.9 (-CH₂), 31.5 (-CH₂), 21.1 (-CH₂), 20.3 (-CH₃), 14.3 (-CHCH₃); IR (film, cm⁻¹) 3409, 2955, 2868, 1618, 1518; MS (EI) m/z : 189 (M⁺), 160 (17), 146 (100), 133 (37); HRMS (EI) m/z calcd for C₁₃H₁₉N(M⁺) 189.1517, found 189.1521; [α]_D²⁰ +5.2 (c 0.5, CHCl₃); ee = 95% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1 °/min; t_R = 39.0 min (minor), t_R = 40.2 min (major)].



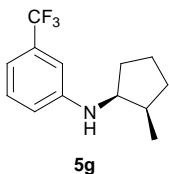
***cis*-N-(4-ethylphenyl)-2-methylcyclopentaneamine (5d)**. Prepared according to the general procedure from 2-methylcyclopentanone (106 μ L, 1.0 mmol), *p*-ethylaniline (160 μ L, 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at

80 °C for 24 h, to provide the title compound as yellow oil (86%). ¹H NMR (300 MHz, CDCl₃) δ 7.00 (d, 2H, J = 7.9 Hz, ArH), 6.57 (d, 2H, J = 7.9 Hz, ArH), 3.73-3.56 (m, 2H, -NHCH + -NHCH), 2.53 (q, 2H, J = 7.5 Hz, -CH₂CH₃), 2.33-2.22 (m, 1H, -CHCH₃), 1.99-1.39 (m, 6H, -CH₂), 1.19 (t, 3H, J = 7.5 Hz, -CH₂CH₃), 0.88 (d, 3H, J = 7.1 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 146.1, 132.6, 128.5, 113.1, 57.6 (-NHCH), 35.8 (-CHCH₃), 32.0 (-CH₂), 31.5 (-CH₂), 27.9 (-CH₂CH₃), 21.2 (-CH₂), 16.9 (-CH₂CH₃), 14.4 (-CHCH₃); IR (film, cm⁻¹) 3413, 2958, 2869, 1616, 1517; MS (EI) m/z : 203 (M⁺), 174 (11), 160 (65), 147 (21), 106 (100); HRMS (EI) m/z calcd for C₁₄H₂₁N(M⁺) 203.1674, found 203.1674; [α]_D²⁰ +5.9 (c 0.17, CHCl₃); ee = 83% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1 °/min; t_R = 51.8 min (minor), t_R = 53.1 min (major)].



***cis*-N-(4-methoxyphenyl)-2-methylcyclopentaneamine (5f)**. Prepared according to the general procedure from 2-methylcyclopentanone (106 μ L, 1.0 mmol), *p*-anisidine (160 mg, 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at

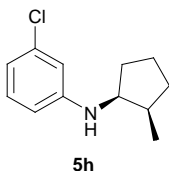
80 °C for 24 h, to provide the title compound as yellow oil (78%). ¹H NMR (300 MHz, CDCl₃) δ 6.76 (d, 2H, J = 8.8 Hz, ArH), 6.60 (d, 2H, J = 8.8 Hz, ArH), 3.74 (s, 4H, -OMe), 3.66 (m, 1H, -NHCH), 3.21 (bs, 1H, -NHCH), 2.30-2.21 (m, 1H, -CHCH₃), 1.99-1.34 (m, 6H, -CH₂), 0.88 (d, 3H, J = 6.9 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 142.3, 114.9, 114.4, 111.7, 58.3 (-NHCH), 55.9 (-OCH₃), 35.7 (-CHCH₃), 32.0 (-CH₂), 31.3 (-CH₂), 21.2 (-CH₂), 14.3 (-CHCH₃); IR (film, cm⁻¹) 3398, 2954, 2869, 1615, 1512; MS (EI) m/z : 205 (M⁺), 190 (10), 176 (18), 162 (100), 149 (38); HRMS (EI) m/z calcd for C₁₃H₁₉NO(M⁺) 205.1473, found 205.1467; [α]_D²⁰ +8.4 (c 0.5, CHCl₃); ee = 96% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1 °/min; t_R = 54.6 min (minor), t_R = 56.2 min (major)].



cis-N-(3-trifluoromethylphenyl)-2-methylcyclopentaneamine (5g). Prepared according to the general procedure from 2-methylcyclopentanone (106 μL , 1.0 mmol), *m*-trifluoromethylaniline (162 μL , 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to provide the title compound as colorless oil (80%).

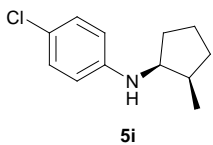
¹H NMR (300 MHz, CDCl₃) δ 7.22 (t, 1H, *J* = 7.7 Hz, ArH), 6.88 (d, 1H, *J* = 7.7 Hz, ArH), 6.81 (s, 1H, ArH), 6.74 (dd, 1H, *J* = 2.0, 7.7 Hz, ArH), 3.90 (bs, 1H, -NHCH), 3.78-3.71 (m, 1H, -NHCH), 2.36-2.22 (m, 1H, -CHCH₃), 2.05-1.38 (m, 6H, -CH₂), 0.89 (d, 3H, *J* = 7.1 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 148.3, 131.5 (q, *J* = 31.3 Hz), 129.6, 124.5 (d, *J* = 272.2 Hz), 115.9, 113.1 (q, *J* = 3.6 Hz), 109.0 (q, *J* = 3.6 Hz), 57.2 (-NHCH), 35.8 (-CHCH₃), 32.0 (-CH₂), 31.5 (-CH₂), 21.2 (-CH₂), 14.4 (-CHCH₃); IR (film, cm⁻¹) 3436, 2960, 2873, 1615, 1517; MS (EI) *m/z*: 243 (M⁺), 214 (16), 200 (100), 187 (15); HRMS (EI) *m/z* calcd for C₁₃H₁₆NF₃(M⁺) 243.1235, found 243.1239; [α]_D²⁰ +7.2 (c 0.58, CHCl₃); ee = 98% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 17.2 min (minor), t_R = 17.6 min (major)].

With Pd[(*S*)-BINAP]Br₂, **1d**, [α]_D²⁰ -10.9 (c 0.55, CHCl₃).



cis-N-(3-chlorophenyl)-2-methylcyclopentaneamine (5h). Prepared according to the general procedure from 2-methylcyclopentanone (106 μL , 1.0 mmol), *m*-chloroaniline (136 μL , 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to provide the title compound as colorless oil (86%).

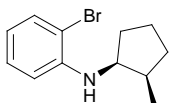
¹H NMR (300 MHz, CDCl₃) δ 7.05 (t, 1H, *J* = 7.8 Hz, ArH), 6.63 (s, 1H, ArH), 6.60-6.58 (m, 1H, ArH), 6.47 (dd, 1H, *J* = 1.8, 7.8 Hz, ArH), 3.83 (bs, 1H, -NHCH), 3.73-3.66 (m, 1H, -NHCH), 2.35-2.21 (m, 1H, -CHCH₃), 2.05-1.34 (m, 6H, -CH₂), 0.88 (d, 3H, *J* = 7.2 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 149.1, 134.9, 130.0, 116.5, 112.4, 111.3, 57.1 (-NHCH), 35.6 (-CHCH₃), 31.8 (-CH₂), 31.4 (-CH₂), 21.1 (-CH₂), 14.3 (-CHCH₃); IR (film, cm⁻¹) 3423, 2957, 2870, 1598, 1501; MS (EI) *m/z*: 209 (M⁺), 180 (21), 166 (100), 153 (22); HRMS (EI) *m/z* calcd for C₁₂H₁₆NCl(M⁺) 209.0971, found 209.0969; [α]_D²⁰ +7.9 (c 0.58, CHCl₃); ee = 91% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 39.8 min (minor), t_R = 41.0 min (major)].



5i

cis-N-(4-chlorophenyl)-2-methylcyclopentaneamine (5i). Prepared according to the general procedure from 2-methylcyclopentanone (106 μL , 1.0 mmol), *p*-chloroaniline (165 mg, 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol)

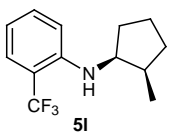
at 80 °C for 24 h, to provide the title compound as light yellow oil (88%). ¹H NMR (300 MHz, CDCl₃) δ 7.09 (d, 2H, *J* = 8.7 Hz, ArH), 6.53 (d, 2H, *J* = 8.7 Hz, ArH), 3.90 (bs, 1H, -NHCH), 3.71-3.65 (m, 1H, -NHCH), 2.37-2.22 (m, 1H, -CHCH₃), 2.03-1.34 (m, 6H, -CH₂), 0.87 (d, 3H, *J* = 7.2 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 146.6, 128.9, 121.2, 114.0, 57.4 (-NHCH), 35.6 (-CHCH₃), 31.9 (-CH₂), 31.4 (-CH₂), 21.1 (-CH₂), 14.3 (-CHCH₃); IR (film, cm⁻¹) 3421, 2957, 2870, 1599, 1499; MS (EI) *m/z*: 209 (M⁺), 180 (16), 166 (100), 153 (27); HRMS (EI) *m/z* calcd for C₁₂H₁₆NCl(M⁺) 209.0971, found 209.0968; [α]_D²⁰ +10.4 (c 0.5, CHCl₃); ee = 84% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 32.3 min (minor), t_R = 33.7 min (major)].



5j

cis-N-(2-bromophenyl)-2-methylcyclopentaneamine (5j). Prepared according to the general procedure from 2-methylcyclopentanone (106 μL , 1.0 mmol), *o*-bromoaniline (147 μL , 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at 80 °C for 24 h,

to provide the title compound as colorless oil (75%). ¹H NMR (300 MHz, CDCl₃) δ 7.40 (d, 1H, *J* = 8.1 Hz, ArH), 7.15 (td, 1H, *J* = 1.5, 8.1 Hz, ArH), 6.70 (dd, 1H, *J* = 1.3, 8.2 Hz, ArH), 6.53 (td, 1H, *J* = 1.5, 8.0 Hz, ArH), 4.13 (bs, 1H, -NHCH), 3.80-3.73 (m, 1H, -NHCH), 2.37-2.21 (m, 1H, -CHCH₃), 2.07-1.41 (m, 6H, -CH₂), 0.93 (d, 3H, *J* = 6.9 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 144.6, 132.3, 128.3, 117.2, 112.0, 109.8, 57.4 (-NHCH), 35.9 (-CHCH₃), 31.9 (-CH₂), 31.5 (-CH₂), 21.3 (-CH₂), 14.4 (-CHCH₃); IR (film, cm⁻¹) 3423, 2957, 2870, 1598, 1501; MS (EI) *m/z*: 253 (M⁺), 224 (19), 210 (100), 197 (21); HRMS (EI) *m/z* calcd for C₁₂H₁₆NBr(M⁺) 253.0466, found 253.0467; [α]_D²⁰ -2.2 (c 0.23, CHCl₃); ee = 18% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 32.3 min (minor), t_R = 32.5 min (major)].



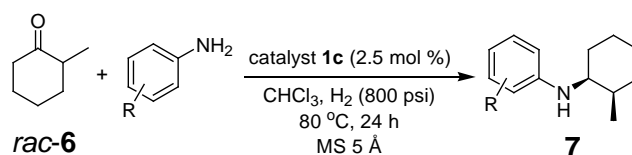
5l

cis-N-(2-trifluoromethylphenyl)-2-methylcyclopentaneamine (5l). Prepared according to the general procedure from 2-methylcyclopentanone (106 μL , 1.0 mmol), *o*-trifluoromethylaniline (163 μL , 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025

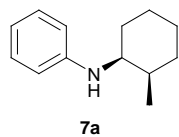
mmol) at 80 °C for 24 h, to provide the title compound as light yellow oil (69%). ¹H NMR (300 MHz, CDCl₃) δ 7.37 (dd, 1H, *J* = 7.96, 1.20 Hz, ArH), 7.28 (t, 1H, *J* = 7.81 Hz, ArH), 6.84-6.76 (m, 2H, ArH), 3.90 (bs, 1H, -NHCH), 3.79-3.72 (m, 1H, -NHCH), 2.36-2.22 (m, 1H, -CHCH₃), 2.05-1.38 (m,

6H, -CH₂), 0.90 (d, 3H, *J* = 7.8 Hz, -CHCH₃); IR (film, cm⁻¹) 3440, 2960, 2877, 1615, 1516; MS (EI) *m/z*: 243 (M⁺), 214 (16), 200 (100); HRMS (EI) *m/z* calcd for C₁₃H₁₆NF₃(M⁺) 243.1235, found 243.1239; ee = 95% by GC-MS (EI) [chiral column: Cyclodex-β; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 37.7 min (minor), t_R = 38.2 min (major)].

General procedure for asymmetric reductive amination of 2-methylcyclohexanone.

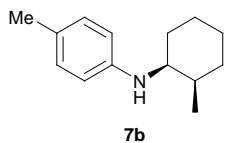


The procedure was the same as 2-methylcyclopentanone.



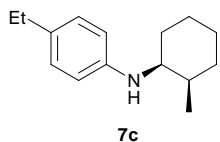
***cis*-N-(phenyl)-2-methylcyclohexaneamine (7a)**. Prepared according to the general procedure from 2-methylcyclohexanone (121 μL, 1.0 mmol), aniline (120 μL, 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to provide the title compound as colorless oil (91%). ¹H NMR (300 MHz, CDCl₃) δ 7.18-7.13 (m, 2H, ArH), 6.67-6.56 (m, 3H, ArH), 3.64 (bs, 1H, -NHCH), 3.52-3.47 (m, 1H, -NHCH), 2.06-1.98 (m, 1H, -CHCH₃), 1.80-1.35 (m, 8H, -CH₂), 0.92 (d, 3H, *J* = 6.9 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 147.6, 129.2, 116.5, 113.1, 53.1 (-NHCH), 33.0 (-CHCH₃), 30.3 (-CH₂), 28.5 (-CH₂), 22.9 (-CH₂), 22.7 (-CH₂), 15.4 (-CHCH₃); IR (film, cm⁻¹) 3415, 2926, 2854, 1601, 1504; MS (EI) *m/z*: 189 (M⁺), 160 (4), 146 (27), 132 (100); HRMS (EI) *m/z* calcd for C₁₃H₁₉N(M⁺) 189.1517, found 189.1520; [α]_D²⁰ -13.3 (c 0.18, CHCl₃); ee = 79% by GC-MS (EI) [chiral column: Cyclodex-β; conditions: t_{inj} = 280°; t_{initial} = 30° for 3 min; rate = 1.2°/min; t_R = 39.3 min (minor), t_R = 40.3 min (major)]. The relative stereochemistry of the product was assigned by comparison of ¹H NMR spectral data reported in the literatur for non chiral compound.⁵

With Pd[(*R*)-Tol-BINAP]Br₂, **1e**, [α]_D²⁰ -11.5 (c 0.53, CHCl₃); ee = 73% by GC-MS (EI) [chiral column: Cyclodex-β; conditions T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1.2°/min; t_R = 39.4 min (minor), t_R = 40.3 min (major)].



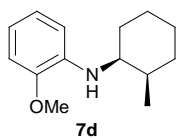
cis-N-(4-methylphenyl)-2-methylcyclohexaneamine (7b). Prepared according to the general procedure from 2-methylcyclohexanone (121 μL , 1.0 mmol), *p*-toluidine (139 mg, 1.3 mmol) and Pd[(*S*)-BINAP]Br₂, **1d**, (22 mg, 0.025 mmol) at

80 °C for 24 h, to provide the title compound as light yellow oil (91%). ¹H NMR (300 MHz, CDCl₃) δ 6.98 (d, 2H, *J* = 8.2 Hz, ArH), 6.55 (d, 1H, *J* = 8.5 Hz, ArH), 3.49-3.38 (m, 2H, -NHCH + -NHCH), 2.24 (s, 3H, -CH₃), 2.06-2.00 (m, 1H, -CHCH₃), 1.80-1.35 (m, 8H, -CH₂), 0.92 (d, 3H, *J* = 7.1 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 145.5, 129.8, 125.8, 113.4, 53.5 (-NHCH), 33.1 (-CHCH₃), 30.5 (-CH₂), 28.7 (-CH₂), 23.0 (-2CH₂), 20.4 (-CH₃), 15.5 (-CHCH₃); IR (film, cm⁻¹) 3412, 2924, 2855, 1617, 1518; MS (EI) *m/z*: 203 (M⁺), 174 (9), 160 (22), 146 (100); HRMS (EI) *m/z* calcd for C₁₄H₂₁N(M⁺) 203.1670, found 203.1674; [α]_D²⁰ +7.6 (c 0.5, CHCl₃); ee = 75% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1.0°/min; t_R = 138.6 min (minor), t_R = 139.9 min (major)].



cis-N-(4-ethylphenyl)-2-methylcyclohexaneamine (7c). Prepared according to the general procedure from 2-methylcyclohexanone (121 μL , 1.0 mmol), *p*-ethylaniline (160 μL , 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at 80 °C for

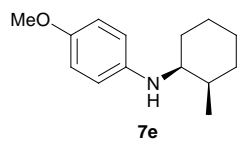
24 h, to provide the title compound as yellow oil (74%). ¹H NMR (300 MHz, CDCl₃) δ 7.02 (d, 2H, *J* = 8.4 Hz, ArH), 6.58 (d, 1H, *J* = 8.7 Hz, ArH), 3.51-3.47 (m, 2H, -NHCH + -NHCH), 2.56 (q, 2H, *J* = 7.6 Hz, -CH₂CH₃), 2.08-1.99 (m, 1H, -CHCH₃), 1.80-1.31 (m, 8H, -CH₂), 1.2 (t, 3H, *J* = 7.5 Hz, -CH₂CH₃), 0.94 (d, 3H, *J* = 6.9 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 145.6, 132.4, 128.5, 113.2, 53.4 (-NHCH), 33.1 (-CHCH₃), 30.3 (-CH₂), 28.6 (-CH₂), 27.8 (-CH₂CH₃), 22.9 (-CH₂), 22.7 (-CH₂), 15.8 (-CH₂CH₃), 15.5 (-CHCH₃); IR (film, cm⁻¹) 3418, 2926, 2855, 1615, 1517; MS (EI) *m/z*: 217 (M⁺), 188 (5), 174 (29), 160 (100); HRMS (EI) *m/z* calcd for C₁₅H₂₃N(M⁺) 217.1830, found 217.1832; [α]_D²⁰ -8.8 (c 0.42, CHCl₃); ee = 62% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: t_{inj} = 280°; t_{initial} = 30° for 3 min; rate = 1.2°/min; t_R = 49.8 min (minor), t_R = 51.4 min (major)].



cis-N-(2-methoxyphenyl)-2-methylcyclohexaneamine (7d). Prepared according to the general procedure from 2-methylcyclohexanone (121 μL , 1.0 mmol), *o*-anisidine (150 μL , 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at 80 °C for 24

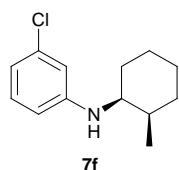
h, to provide the title compound as colorless oil (83%). ¹H NMR (300 MHz, CDCl₃) δ 6.87 (td, 1H, *J* = 1.3, 7.15 Hz, ArH), 6.79 (d, 1H, *J* = 7.7 Hz, ArH), 6.66-6.61 (m, 2H, ArH), 4.34 (bs, 1H, -NHCH), 3.87 (s, 3H, -OMe), 3.55-3.51 (m, 1H, -NHCH), 2.10-2.00 (m, 1H, -CHCH₃), 1.76-1.38 (m, 8H, -CH₂), 0.95 (d, 3H, *J* = 6.8 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 146.9, 137.8, 121.3, 115.5, 110.2,

109.6, 55.6 (-OCH₃), 53.0 (-NHCH), 33.3 (-CHCH₃), 30.5 (-CH₂), 28.8 (-CH₂), 23.2 (-CH₂), 22.9 (-CH₂), 15.7 (-CHCH₃); IR (film, cm⁻¹) 3421, 2926, 2855, 1594, 1498; MS (EI) *m/z*: 219 (M⁺), 190 (4), 176 (28), 162 (100); HRMS (EI) *m/z* calcd for C₁₄H₂₁NO(M⁺) 219.1623, found 219.1619; [α]²⁰_D -7.8 (c 0.54, CHCl₃); ee = 98% by GC-MS (EI) [chiral column: Cyclodex-β; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1.0°/min; t_R = 156.1 min (minor), t_R = 157.5 min (major)].



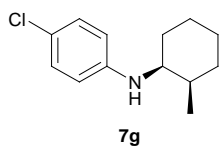
***cis*-N-(4-methoxyphenyl)-2-methylcyclohexaneamine (7e)**. Prepared according to the general procedure from 2-methylcyclohexanone (121 μL, 1.0 mmol), *p*-anisidine (160 mg, 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at

80 °C for 24 h, to provide the title compound as light yellow oil (85%). ¹H NMR (300 MHz, CDCl₃) δ 6.76 (d, 2H, *J* = 8.7 Hz, ArH), 6.58 (d, 2H, *J* = 8.7 Hz, ArH), 4.14 (bs, 1H, -NHCH), 3.74 (s, 3H, -OMe), 3.43-3.38 (m, 1H, -NHCH), 2.06-1.97 (m, 1H, -CHCH₃), 1.65-1.35 (m, 8H, -CH₂), 0.91 (d, 3H, *J* = 7.2 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 151.6, 141.9, 114.9, 114.6, 55.8 (-OCH₃), 54.2 (-NHCH), 33.0 (-CHCH₃), 30.4 (-CH₂), 28.5 (-CH₂), 22.9 (-CH₂), 22.8 (-CH₂), 15.2 (-CHCH₃); IR (film, cm⁻¹) 3393, 2925, 2853, 1617, 1512; MS (EI) *m/z*: 219 (M⁺), 190 (4), 176 (22), 162 (100); HRMS (EI) *m/z* calcd for C₁₄H₂₁NO(M⁺) 219.1623, found 219.1625; [α]²⁰_D -9.3 (c 0.4, CHCl₃); ee = >99% by GC-MS (EI) [chiral column: Cyclodex-β; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1.0°/min; t_R = 143.8 min (minor), t_R = 144.9 min (major)].



***cis*-N-(3-chlorophenyl)-2-methylcyclohexaneamine (7f)**. Prepared according to the general procedure from 2-methylcyclohexanone (121 μL, 1.0 mmol), *m*-chloroaniline (136 μL, 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at 80 °C for 24

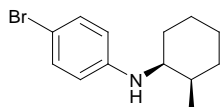
h, to provide the title compound as colorless oil (90%). ¹H NMR (300 MHz, CDCl₃) δ 7.07-7.00 (m, 1H, ArH), 6.62-6.57 (m, 2H, ArH), 6.46 (dd, 1H, *J* = 1.0, 8.3 Hz ArH), 3.70 (bs, 1H, -NHCH), 3.48-3.44 (m, 1H, -NHCH), 2.04-1.96 (m, 1H, -CHCH₃), 1.83-1.35 (m, 8H, -CH₂), 0.91 (d, 3H, *J* = 6.6 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 148.8, 134.9, 130.0, 116.2, 112.4, 111.3, 53.0 (-NHCH), 32.9 (-CHCH₃), 30.2 (-CH₂), 28.5 (-CH₂), 22.9 (-CH₂), 22.6 (-CH₂), 15.6 (-CHCH₃); IR (film, cm⁻¹) 3425, 2927, 2855, 1597, 1502; MS (EI) *m/z*: 223 (M⁺), 194 (5), 180 (32), 166 (100); HRMS (EI) *m/z* calcd for C₁₃H₁₈NCl(M⁺) 223.1128, found 223.1123; [α]²⁰_D -9.5 (c 0.46, CHCl₃); ee = 93% by GC-MS (EI) [chiral column: Cyclodex-β; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1.2°/min; t_R = 28.4 min (minor), t_R = 29.7 min (major)].



7g

cis-N-(4-chlorophenyl)-2-methylcyclohexaneamine (7g). Prepared according to the general procedure from 2-methylcyclohexanone (121 μ L, 1.0 mmol), *p*-chloroaniline (165 mg, 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol)

at 80 °C for 24 h, to provide the title compound as colorless oil (82%). ¹H NMR (300 MHz, CDCl₃) δ 7.09 (d, 2H, *J* = 8.8 Hz, ArH), 6.51 (d, 2H, *J* = 8.8 Hz, ArH), 3.59 (bs, 1H, -NHCH), 3.46-3.41 (m, 1H, -NHCH), 2.04-1.97 (m, 1H, -CHCH₃), 1.75-1.36 (m, 8H, -CH₂), 0.90 (d, 3H, *J* = 7.2 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 146.4, 129.1, 121.0, 114.2, 53.4 (-NHCH), 33.1 (-CHCH₃), 30.3 (-CH₂), 28.6 (-CH₂), 23.0 (-CH₂), 22.8 (-CH₂), 15.5 (-CHCH₃); IR (film, cm⁻¹) 3423, 2927, 2856, 1599, 1500; MS (EI) *m/z*: 223 (M⁺), 194 (6), 180 (37), 166 (100); HRMS (EI) *m/z* calcd for C₁₃H₁₈NCl(M⁺) 223.1128, found 223.1135; [α]_D²⁰ -6.4 (c 0.5, CHCl₃); ee = 89% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1.0°/min; t_R = 157.1 min (minor), t_R = 157.8 min (major)].

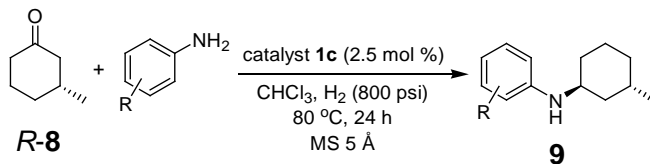


7h

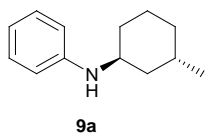
cis-N-(4-bromophenyl)-2-methylcyclohexaneamine (7h). Prepared according to the general procedure from 2-methylcyclohexanone (121 μ L, 1.0 mmol), *p*-bromoaniline (222 mg, 1.3 mmol) and Pd[(*S*)-BINAP]Br₂, **1d**, (22 mg, 0.025 mmol)

at 80 °C for 24 h, to provide the title compound as colorless oil (71%). ¹H NMR (300 MHz, CDCl₃) δ 7.21 (d, 2H, *J* = 8.8 Hz, ArH), 6.48 (d, 2H, *J* = 8.8 Hz, ArH), 3.67 (bs, 1H, -NHCH), 3.46-3.41 (m, 1H, -NHCH), 2.02-1.97 (m, 1H, -CHCH₃), 1.77-1.22 (m, 8H, -CH₂), 0.90 (d, 3H, *J* = 7.2 Hz, -CHCH₃); ¹³C NMR (75 MHz, CDCl₃) δ 146.8, 132.0, 114.7, 108.0, 53.4 (-NHCH), 33.0 (-CHCH₃), 30.3 (-CH₂), 28.6 (-CH₂), 23.0 (-CH₂), 22.8 (-CH₂), 15.5 (-CHCH₃); IR (film, cm⁻¹) 3434, 2926, 2854, 1600, 1515; MS (EI) *m/z*: 267 (M⁺), 240 (2), 226 (19), 212 (100); HRMS (EI) *m/z* calcd for C₁₃H₁₈NBr(M⁺) 267.0623, found 267.0631; [α]_D²⁰ +6.5 (c 0.52, CHCl₃); ee = 83% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1.0°/min; t_R = 167.3 min (minor), t_R = 168.0 min (major)].

General procedure for asymmetric reductive amination of (*R*)-3-methylcyclohexanone.



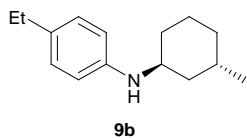
The procedure was the same as 2-methylcyclopentanone.



***trans*-N-(phenyl)-3-methylcyclohexaneamine (9a)**. Prepared according to the general procedure from (*R*)-(+)-3-methylcyclohexanone (122 μ L, 1.0 mmol), aniline (120 μ L, 1.3 mmol) and Pd[(*S*)-BINAP]Br₂, **1d**, (22 mg, 0.025 mmol) at 80 °C for 24

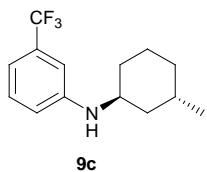
h, to provide the title compound as colorless oil (87%). ¹H NMR (500 MHz, CDCl₃) δ 7.17-7.14 (m, 2H, ArH), 6.68 (t, 1H, *J* = 7.2 Hz, ArH), 6.63 (d, 2H, *J* = 8.0 Hz, ArH), 4.21 (bs, 1H, -NHCH), 3.68-3.64 (m, 1H, -NHCH), 1.78-1.50 (m, 7H, -CHCH₃ + -CH₂), 1.38-1.33 (m, 1H, -CH₂), 1.09-1.01 (m, 1H, -CH₂), 0.91 (d, 3H, *J* = 6.5 Hz, -CHCH₃); ¹³C NMR (125 MHz, CDCl₃) δ 146.8, 129.2, 117.2, 113.6, 48.1 (-NHCH), 38.6 (-CH₂), 33.8 (-CH₂), 30.3 (-CH₂), 27.1 (-CHCH₃), 21.5 (-CHCH₃), 20.5 (-CH₂); IR (film, cm⁻¹) 3415, 2923, 2850, 1602, 1504; MS (EI) *m/z*: 189 (M⁺), 174 (8), 160 (5), 146 (100), 132 (87); HRMS (EI) *m/z* calcd for C₁₃H₁₉N(M⁺) 189.1517, found 189.1514; [α]_D²⁰ -13.3 (c 0.48, CHCl₃); ee = 91% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1^o/min; t_R = 132.7 min (major), t_R = 133.2 min (minor)].

The relative configuration was confirmed by NOE experiment and compared with a similar reported compound.⁶



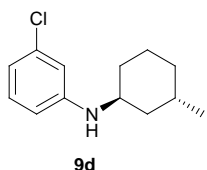
***trans*-N-(*p*-ethylphenyl)-3-methylcyclohexaneamine (9b)**. Prepared according to the general procedure from (*R*)-(+)-3-methylcyclohexanone (122 μ L, 1.0 mmol), *p*-ethylaniline (160 μ L, 1.3 mmol) and Pd[(*S*)-BINAP]Br₂, **1e**, (22 mg,

0.025 mmol) at 80 °C for 24 h, to provide the title compound as light yellow oil (90%). ¹H NMR (500 MHz, CDCl₃) δ 6.99 (d, 2H, *J* = 8.5 Hz, ArH), 6.54 (d, 2H, *J* = 8.5 Hz, ArH), 4.02 (bs, 1H, -NHCH), 3.65-3.62 (m, 1H, -NHCH), 2.52 (q, 2H, *J* = 7.5 Hz, -CH₂CH₃), 1.76-1.49 (m, 7H, -CHCH₃ + -CH₂), 1.36-1.30 (m, 1H, -CH₂), 1.18 (t, 3H, *J* = 7.5 Hz, -CH₂CH₃), 1.07-1.00 (m, 1H, -CH₂), 0.90 (d, 3H, *J* = 6.5 Hz, -CHCH₃); ¹³C NMR (125 MHz, CDCl₃) δ 145.3, 132.6, 128.5, 113.2, 47.8 (-NHCH), 38.9 (-CH₂), 33.9 (-CH₂), 30.5 (-CH₂), 27.8 (-CH₂CH₃), 27.1 (-CHCH₃), 21.6 (-CHCH₃), 20.5 (-CH₂), 15.8 (-CH₂CH₃); IR (film, cm⁻¹) 3411, 2924, 2864, 1615, 1517; MS (EI) *m/z*: 217 (M⁺), 202 (9), 173 (3), 160 (100), 146 (82); HRMS (EI) *m/z* calcd for C₁₅H₂₃N(M⁺) 217.1830, found 217.1829; [α]_D²⁰ -8.4 (c 0.5, CHCl₃); ee = 95% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1^o/min; t_R = 149.8 min (major), t_R = 150.5 min (minor)].



***trans*-N-(3-trifluoromethylphenyl)-3-methylcyclohexaneamine (9c).** Prepared according to the general procedure from (*R*)-(+)-3-methylcyclohexanone (122 μ L, 1.0 mmol), *m*-trifluoromethyl aniline (162 μ L, 1.3 mmol) and Pd[(*S*)-BINAP]Br₂, **1d**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to provide the title compound as colorless

oil (84%). ¹H NMR (500 MHz, CDCl₃) δ 7.21 (t, 1H, *J* = 8.0 Hz, ArH), 6.87 (d, 1H, *J* = 7.5 Hz, ArH), 6.77 (s, 1H, ArH), 6.70 (d, 1H, *J* = 8.0 Hz, ArH), 4.06 (bs, 1H, -NHCH), 3.70-3.66 (m, 1H, -NHCH), 1.76-1.45 (m, 7H, -CHCH₃ + -CH₂), 1.38-1.32 (m, 1H, -CH₂), 1.09-1.01 (m, 1H, -CH₂), 0.92 (d, 3H, *J* = 7.0 Hz, -CHCH₃); ¹³C NMR (125 MHz, CDCl₃) δ 147.4, 131.6 (q, *J* = 31.4 Hz), 129.6, 124.6 (d, *J* = 271.0 Hz), 115.9, 113.1 (q, *J* = 3.7 Hz), 109.1 (q, *J* = 3.8 Hz), 47.6 (-NHCH), 38.6 (-CH₂), 33.8 (-CH₂), 30.2 (-CH₂), 27.1 (-CHCH₃), 21.6 (-CHCH₃), 20.4 (-CH₂); IR (film, cm⁻¹) 3443, 2926, 2857, 1614, 1515; MS (EI) *m/z*: 257 (M⁺), 242 (7), 214 (100), 228 (2), 200 (74); HRMS (EI) *m/z* calcd for C₁₄H₁₈NF₃(M⁺) 257.1391, found 257.1389; [α]_D²⁰ -13.1 (c 0.48, CHCl₃); ee = >99% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 135.8 min (major)].

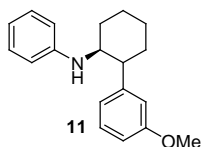


***trans*-N-(3-chlorophenyl)-3-methylcyclohexaneamine (9d).** Prepared according to the general procedure from (*R*)-(+)-3-methylcyclohexanone (122 μ L, 1.0 mmol), *m*-chloroaniline (136 μ L, 1.3 mmol) and Pd[(*S*)-BINAP]Br₂, **1d**, (22 mg, 0.025 mmol) at 80 °C for 24 h, to provide the title compound as colorless oil (89%). ¹H NMR (500

MHz, CDCl₃) δ 7.02 (t, 1H, *J* = 8.2 Hz, ArH), 6.62 (d, 1H, *J* = 8.0 Hz, ArH), 6.54 (s, 1H, ArH), 6.43 (d, 1H, *J* = 8.5 Hz, ArH), 3.90 (bs, 1H, -NHCH), 3.63-3.60 (m, 1H, -NHCH), 1.72-1.43 (m, 7H, -CHCH₃ + -CH₂), 1.35-1.29 (m, 1H, -CH₂), 1.07-0.99 (m, 1H, -CH₂), 0.91 (d, 3H, *J* = 6.5 Hz, -CHCH₃); ¹³C NMR (125 MHz, CDCl₃) δ 148.4, 135.0, 130.1, 116.5, 112.5, 111.3, 47.6 (-NHCH), 38.6 (-CH₂), 33.8 (-CH₂), 30.2 (-CH₂), 27.1 (-CHCH₃), 21.5 (-CHCH₃), 20.4 (-CH₂); IR (film, cm⁻¹) 3425, 2924, 2861, 1598, 1500; MS (EI) *m/z*: 223 (M⁺), 208 (7), 180 (100), 166 (73), 153 (17); HRMS (EI) *m/z* calcd for C₁₃H₁₈NCl(M⁺) 223.1128, found 223.1125; [α]_D²⁰ -12.7 (c 0.47, CHCl₃); [α]_D²⁰ -20.3 (c 0.52, benzene); ee = >99% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 °C; T_{det} = 280 °C; T_{initial} = 30 °C for 3 min; T_{final} = 200 °C; rate = 1°/min; t_R = 189.4 min (major)].

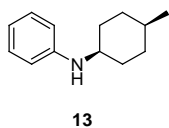
General procedure for asymmetric reductive amination of other substituted rings.

The procedure was the same as 2-methylcyclopentanone.



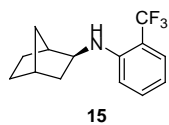
cis-N-[2-(3-methoxyphenyl)cyclohexyl]benzenamine (11). Prepared according to the general procedure from 2-(3-methoxyphenyl)cyclohexanone (185 μL , 1.0 mmol), aniline (120 μL , 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at 80

$^{\circ}\text{C}$ for 24 h, to provide the title compound as light yellow oil (77%). ^1H NMR (500 MHz, CDCl₃) δ 7.16 (t, 1H, $J = 7.5$ Hz, ArH), 7.03 (dd, 2H, $J = 8.5$ Hz, ArH), 6.84 (dt, 1H, $J = 8.5, 1.0$ Hz, ArH), 6.80 (t, 1H, $J = 8.5$ Hz, ArH), 6.68 (dd, 1H, $J = 8.0, 2.5$ Hz, ArH), 6.56 (tt, 1H, $J = 7.5, 1.0$ Hz, ArH), 6.41 (d, 2H, $J = 8.0$ Hz, ArH), 3.82-3.79 (m, 1H, -NHCH), 3.70 (s, 3H, -OMe), 3.66 (bs, 1H, -NHCH), 2.98-2.92 (ddd, 1H, $J = 11.5, 4.0, 4.0$ Hz, -CHArH), 2.12-2.08 (m, 1H, -CH₂), 1.92-1.82 (m, 3H, -CH₂), 1.61-1.42 (m, 4H, -CH₂); ^{13}C NMR (125 MHz, CDCl₃) δ 159.5, 147.8, 145.4, 129.1, 128.9, 119.9, 116.6, 113.7, 113.2, 111.3, 55.0 (-OMe), 53.2 (-NHCH), 46.1 (-CHArH), 30.1 (-CH₂), 25.87 (-CH₂), 25.83 (-CH₂), 20.3 (-CH₂); IR (film, cm⁻¹) 3424, 2926, 2861, 1598, 1502; MS (EI) m/z : 281 (M⁺), 252 (3), 238 (18), 224 (3), 159 (19), 132 (100); HRMS (EI) m/z calcd for C₁₉H₂₃ON (M⁺) 281.1780, found 281.1784; $[\alpha]_{\text{D}}^{20}$ -36.6 (c 0.21, CHCl₃); ee = 80% by GC-MS (EI) [chiral column: Cyclodex- β ; conditions: T_{inj} = 280 $^{\circ}\text{C}$; T_{det} = 280 $^{\circ}\text{C}$; T_{initial} = 30 $^{\circ}\text{C}$ for 3 min; T_{final} = 200 $^{\circ}\text{C}$; rate = 1 $^{\circ}$ /min; t_R = 147.0 min (minor), t_R = 148.4 min (major)].



cis-N-(4-methylcyclohexyl)aniline (13). Prepared according to the general procedure from 4-methylcyclohexanone (123 μL , 1.0 mmol), aniline (120 μL , 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at 80 $^{\circ}\text{C}$ for 24 h, to provide the title

compound as light yellow oil (74%). ^1H NMR (500 MHz, CDCl₃) δ 7.15 (dd, 2H, $J = 8.5$ Hz, ArH), 6.65 (t, 1H, $J = 8.0$ Hz, ArH), 6.61-6.58 (m, 2H, ArH), 3.73 (bs, 1H, -NHCH), 3.56-3.52 (m, 1H, -NHCH), 1.77-1.72 (m, 2H, -CH₂), 1.66-1.60 (m, 2H, -CH₂), 1.57-1.51 (m, 3H, -CHCH₃+ -CH₂), 0.92 (d, 3H, $J = 6.5$ Hz, -CHCH₃); ^{13}C NMR (125 MHz, CDCl₃) δ 147.3, 129.2, 116.7, 113.1, 48.1 (-NHCH), 30.8 (-CHCH₃), 29.7 (-2CH₂), 29.2 (-2CH₂), 21.3 (-CHCH₃); IR (film, cm⁻¹) 3425, 2924, 2860, 1601, 1514; MS (EI) m/z : 189 (M⁺), 157 (3), 144 (6), 132 (100), 118 (14), 106 (11); HRMS (EI) m/z calcd for C₁₃H₁₉N (M⁺) 189.1517, found 189.1515.



N-(2-(trifluoromethyl)phenyl)bicyclo[2.2.1]heptan-2-amine (15). Prepared according to the general procedure from norcamphor (110 mg, 1.0 mmol), *o*-trifluoromethyl aniline (163 μL , 1.3 mmol) and Pd[(*R*)-BINAP]Br₂, **1c**, (22 mg, 0.025 mmol) at 80 $^{\circ}\text{C}$

for 24 h, to provide the title compound as colorless oil (74%). ^1H NMR (500 MHz, CDCl₃) δ 7.41 (d, 1H, $J = 8.0$ Hz, ArH), 7.32 (t, 1H, $J = 7.5$ Hz, ArH), 6.71 (d, 1H, $J = 8.5$ Hz, ArH), 6.67 (t, 1H, $J = 7.5$ Hz, ArH), 4.51 (bs, 1H, -NHCH), 3.76-3.73 (m, 1H, -NHCH), 2.55-2.53 (m, 1H, -CH), 2.32-2.26 (m,

1H, -CH), 2.20-2.14 (m, 1H, -CH₂), 1.68-1.55 (m, 2H, -CH₂), 1.50-1.47 (m, 1H, -CH₂), 1.42-1.35 (m, 2H, -CH₂), 1.30-1.16 (m, 1H, -CH₂), 0.84-0.80 (m, 1H, -CH₂) ; ¹³C NMR (125 MHz, CDCl₃) δ 145.6, 132.9, 126.8 (q, *J* = 5.3 Hz), 119.8 (d = 272.1 Hz), 115.3, 113.2 (q, *J* = 29.4 Hz), 112.6, 54.2 (-NHCH), 39.7 (-CH), 38.9 (-CH₂), 38.1 (-CH₂), 36.8 (-CH), 29.7 (-CH₂), 21.2 (-CH₂); IR (film, cm⁻¹) 3476, 2956, 2873, 1615, 1519; MS (EI) *m/z*: 255 (M⁺, 100), 226 (23), 214 (12), 200 (29), 187 (57), 174 (21); HRMS (EI) *m/z* calcd for C₁₄H₁₆NF₃(M⁺) 255.1235, found 235.1239; [α]_D²⁰ -15.7 (c 0.50, CHCl₃); ee = 87% by HPLC (Daicel Chiracel OD-H, eluent hexane-iPrOH = 99 1, flow rate = 1mL min, t_R = 6.58 min (major), t_R = 7.58 min (minor)].

References:

1. M. A. Andrews, T. C. T. Chang, C. W. F. Cheng, T. J. Emge, K. P. Kelly, T. F. Koetzle, *J. Am. Chem. Soc.*, 1984, **106**, 5913-5920.
2. F. Ozawa, A. Kubo, Y. Matsumoto, T. Hayashi, *Organometallics*, 1993, **12**, 4188-4196.
3. L. Rubio-Pérez, F. J. Pérez-Flores, P. Sharma, A. Cabrera, *Org. Lett.*, 2009, **11**, 265-268.
4. A. J. Blacker, M. L. Clarke, M. S. Loft, M. F. Mahon, M. E. Humphries, J. M. J. Williams, *Chem. A. Eur. J.*, 2000, **6**, 353-360.
5. T. Suwa, E. Sugiyama, I. Shibata, A. Baba, *Synthesis*, 2000, 789-800.
6. J. Zhou, B. List, *J. Am. Chem. Soc.*, 2007, **129**, 7498-7499.

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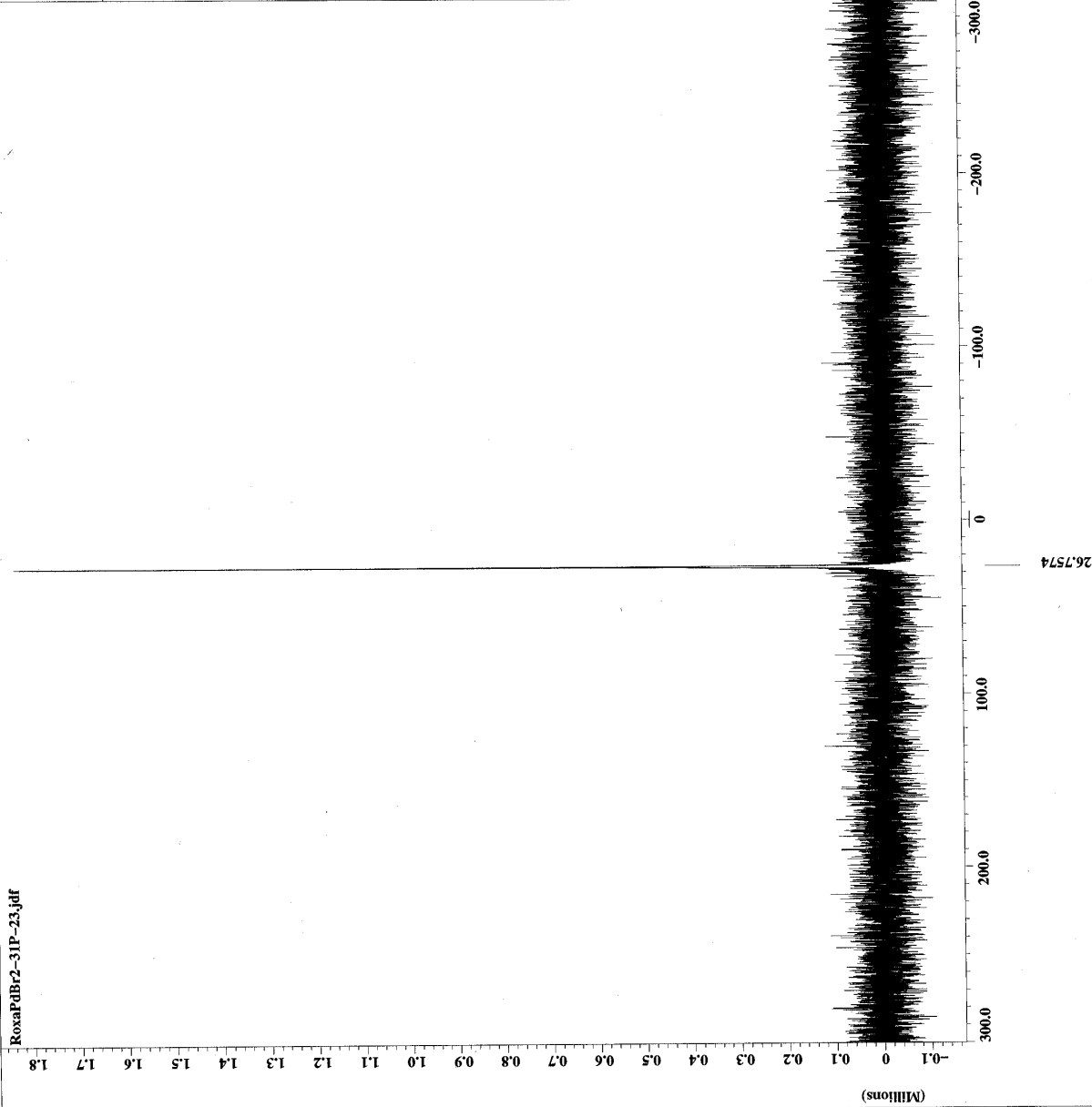
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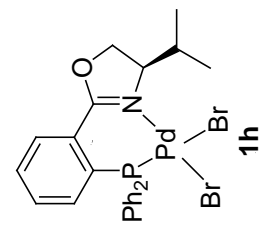
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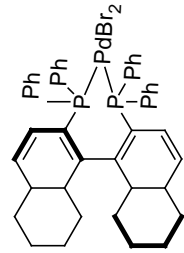


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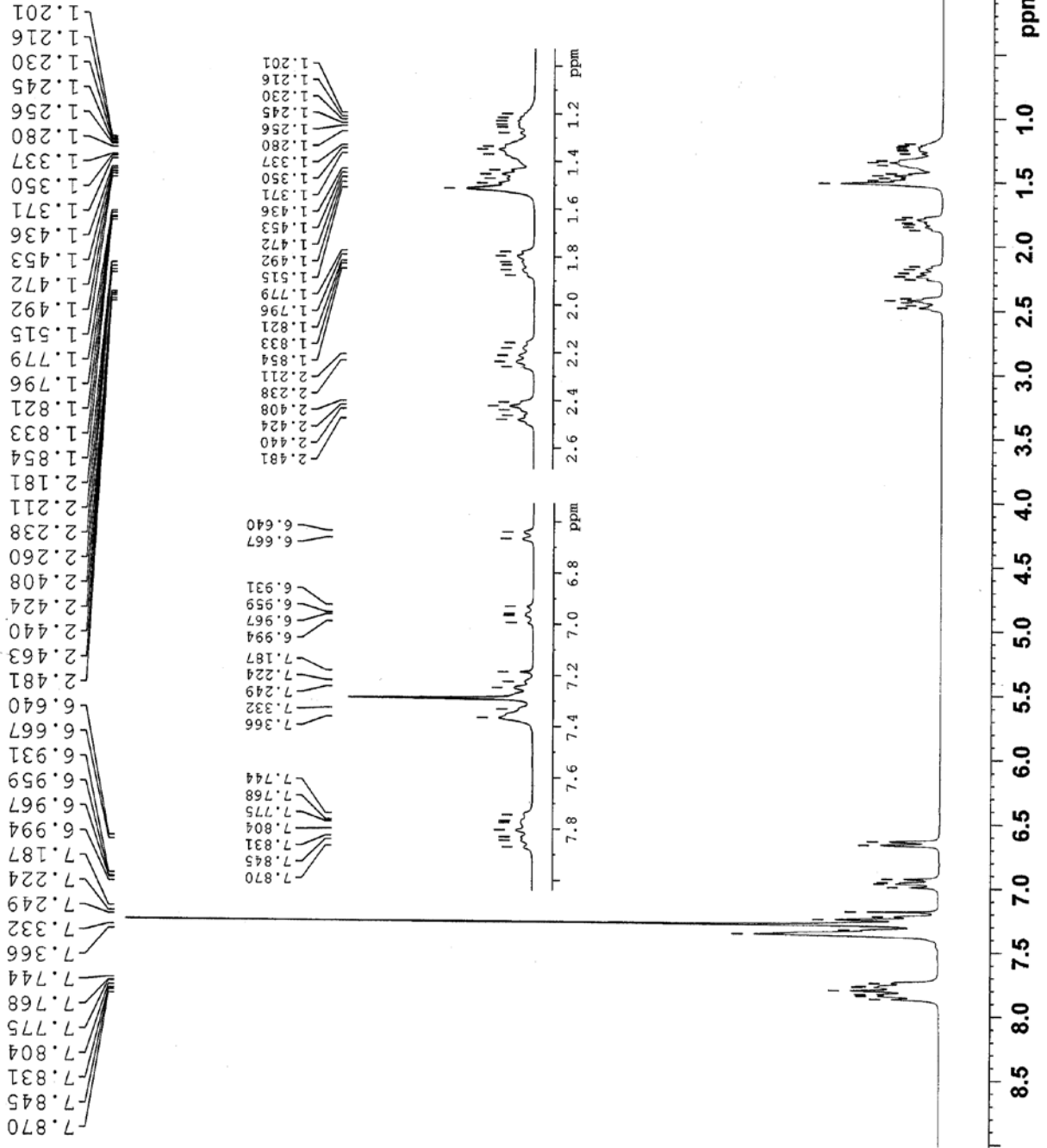
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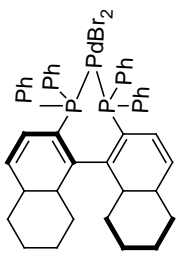
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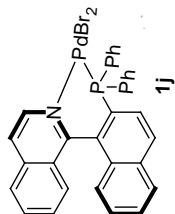
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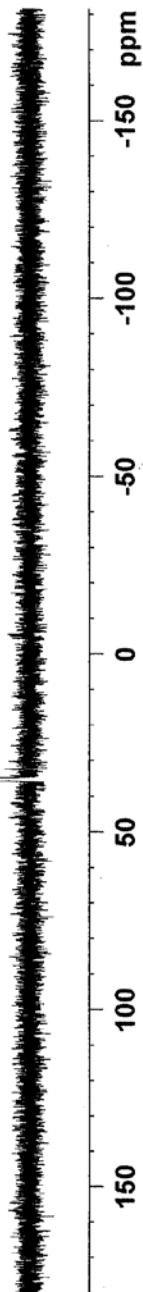
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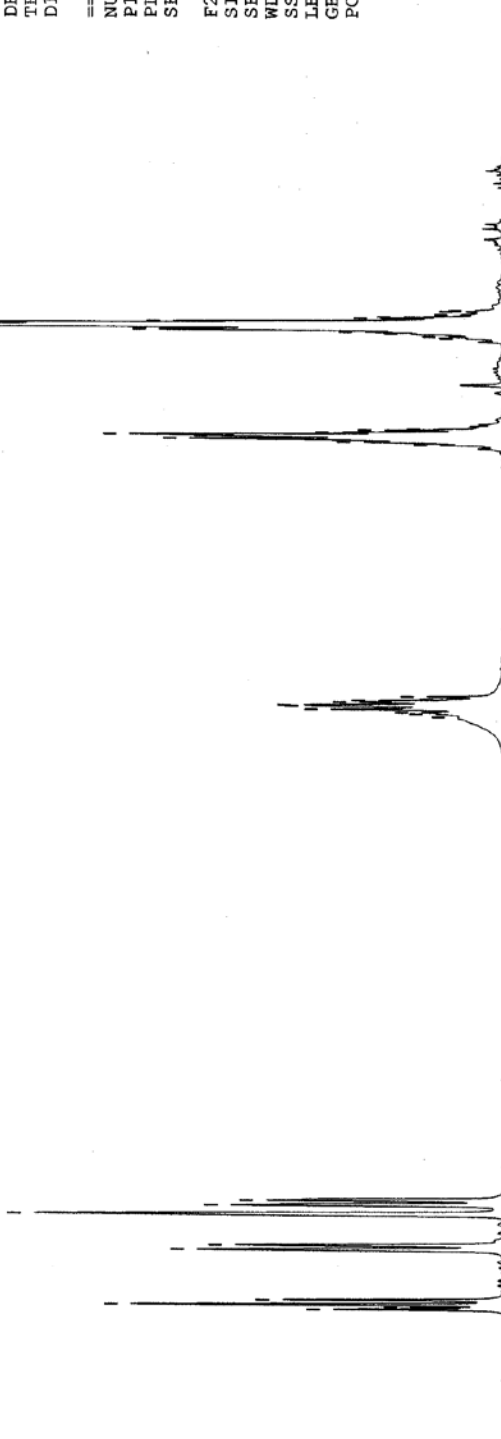
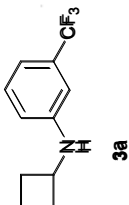
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Time 12.52
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDC13
NS 32
DS 1
SWH 6203.474 Hz
FIDRES 0.094658 Hz
AQ 5.2822518 sec
RG 71.8
LW 80.600 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec

==== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PLW1 9.24709988 W
SF01 300.5218558 MHz

F2 - Processing parameters
SI 65536
SF 300.5200414 MHz
WDW EM
SSB 0 0.30 Hz
LB 0
GB 0
PC 1.00

7.167
7.157
7.141
7.119
6.838
6.813
6.640
6.592
6.565
3.897
3.880
3.872
3.855
3.846
3.837
3.832
3.815
3.806
3.800
3.784
2.395
2.380
2.361
2.346
2.339
2.330
2.326
2.313
2.308
1.808
1.804
1.797
1.784
1.779
1.772
1.761
1.756
1.745
1.725
1.723
1.714
1.696



7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 ppm



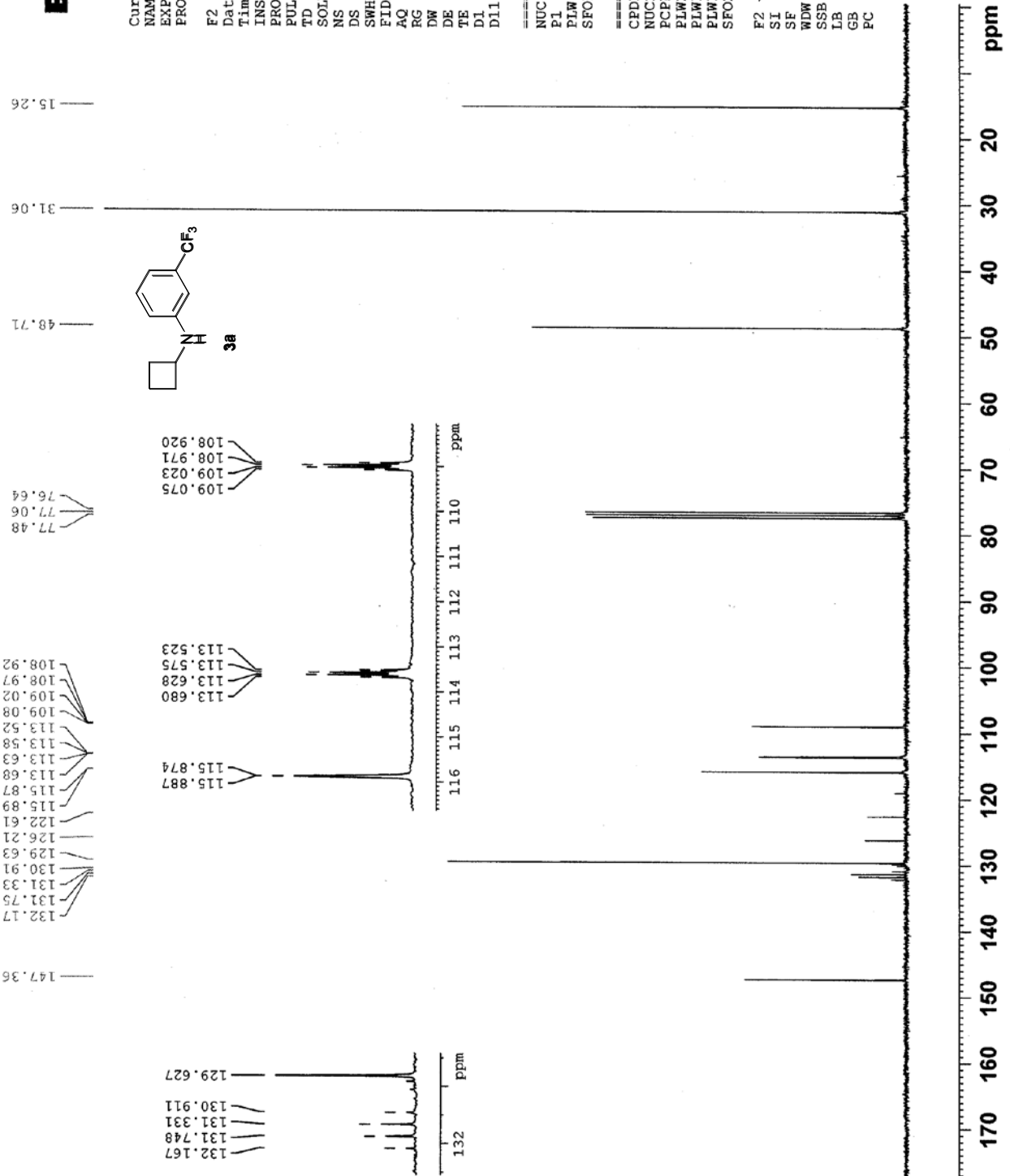
Current Data Parameters
 NAME Reacc707F19
 EXPNO 21
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120222
 Time 13.45
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 800
 DS 1
 SWH 18028.846 Hz
 FIDRES 0.275098 Hz
 AQ 1.8175818 sec
 RG 203
 DW 27.733 usec
 DE 6.50 usec
 TE 300.0 K
 D1 2.0000000 sec
 D11 0.0300000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 7.90 usec
 PLW1 48.9669905 W
 SFO1 75.5733703 MHz

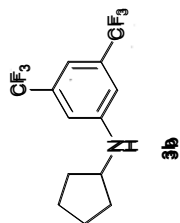
==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PLW2 9.24709988 W
 PLWI2 0.21741000 W
 PLWI3 0.17610000 W
 SFO2 300.5212021 MHz

F2 - Processing parameters
 SI 32768
 SF 75.5658140 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 FC 1.40





1.481
1.488
1.492
1.496
1.512
1.520
1.530
1.535
1.551
1.553
1.568
1.572
1.670
1.680
1.686
1.706
1.713
1.719
1.724
1.731
1.734
1.740
1.749
1.757
1.764
1.782
1.791
1.801
1.817
1.826
2.048
2.052
2.058
2.071
2.083
2.091
2.106
2.111
2.114
2.123
2.132
2.145
3.836
3.855
3.874
4.143
6.952
7.149
7.295

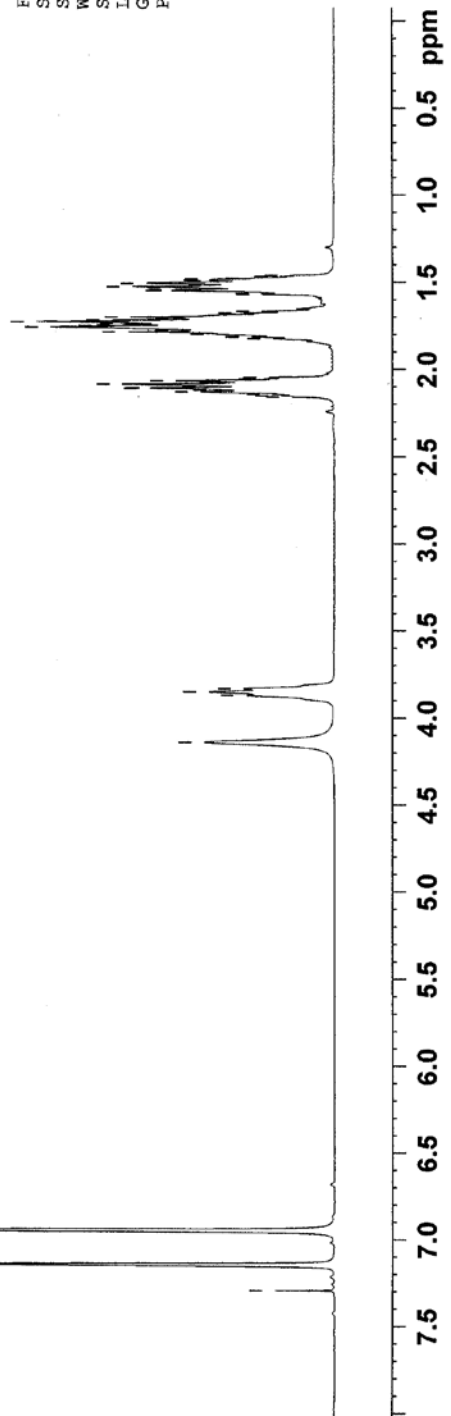


Current Data Parameters
NAME Reacc711f1314
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120227
Time_ 9.48
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 64
DS 2
SWH 6203.474 Hz
FIDRES 0.094658 Hz
AQ 5.2822518 sec
RG 90.5
DW 80.600 usec
DE 6.50 usec
TE 295.3 K
D1 1.00000000 sec

==== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PLW1 9.24709988 W
SF01 300.5218558 MHz

F2 - Processing parameters
SI 65536
SF 300.5200000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00





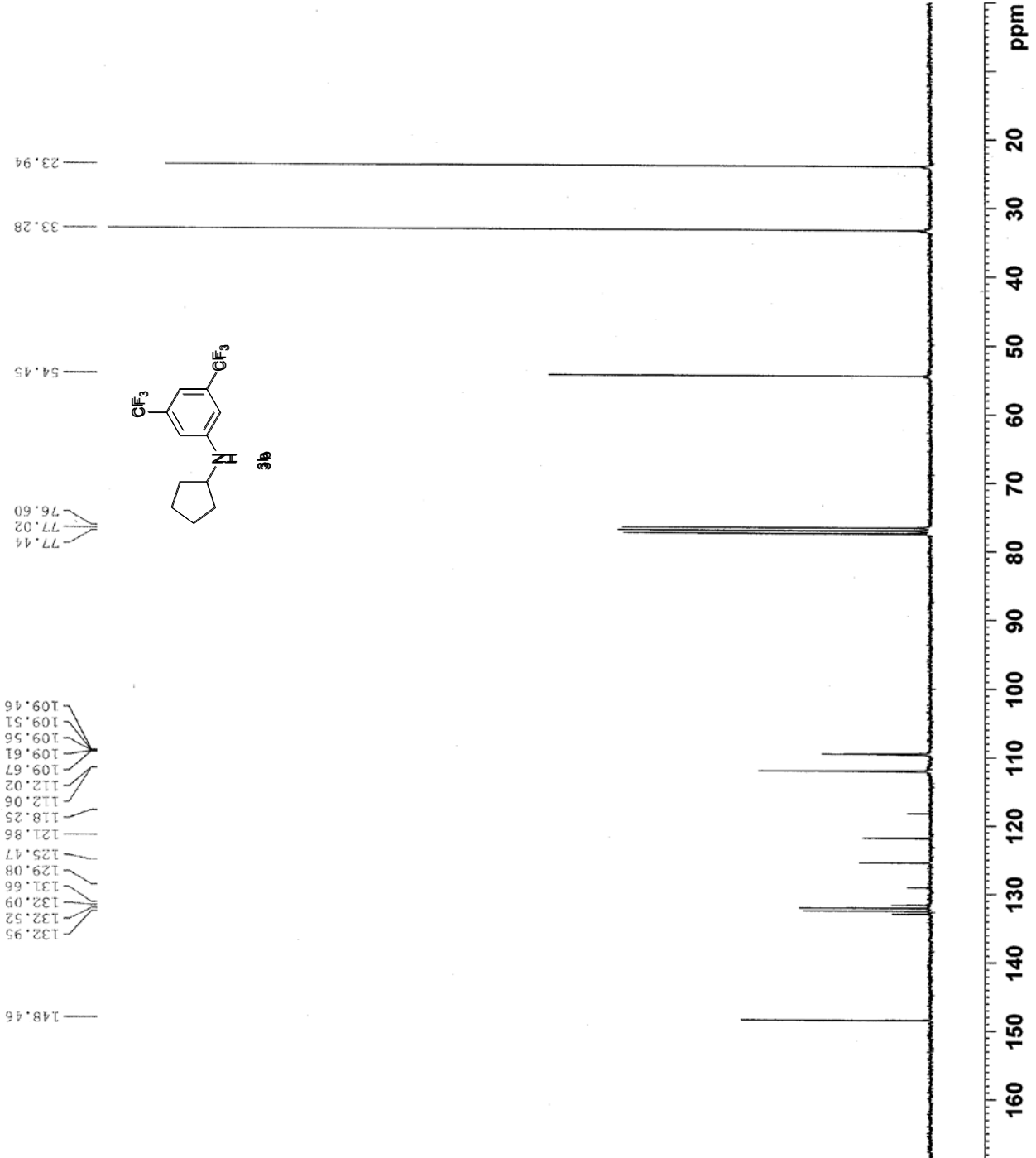
Current Data Parameters
NAME Reacc711F1314
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120227
Time 10.35
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 700
DS 2
SWH 18028.846 Hz
FIDRES 0.275098 Hz
AQ 1.8175818 sec
RG 203
DW 27.733 usec
DE 6.50 usec
TE 295.9 K
D1 2.0000000 sec
D11 0.0300000 sec

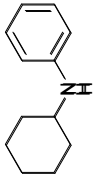
==== CHANNEL f1 =====
NUC1 13C
P1 7.90 usec
PLW1 48.9669905 W
SF01 75.5733703 MHz

==== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PLW2 9.24709988 W
PLW12 0.21741000 W
PLW13 0.17610000 W
SFO2 300.5212021 MHz

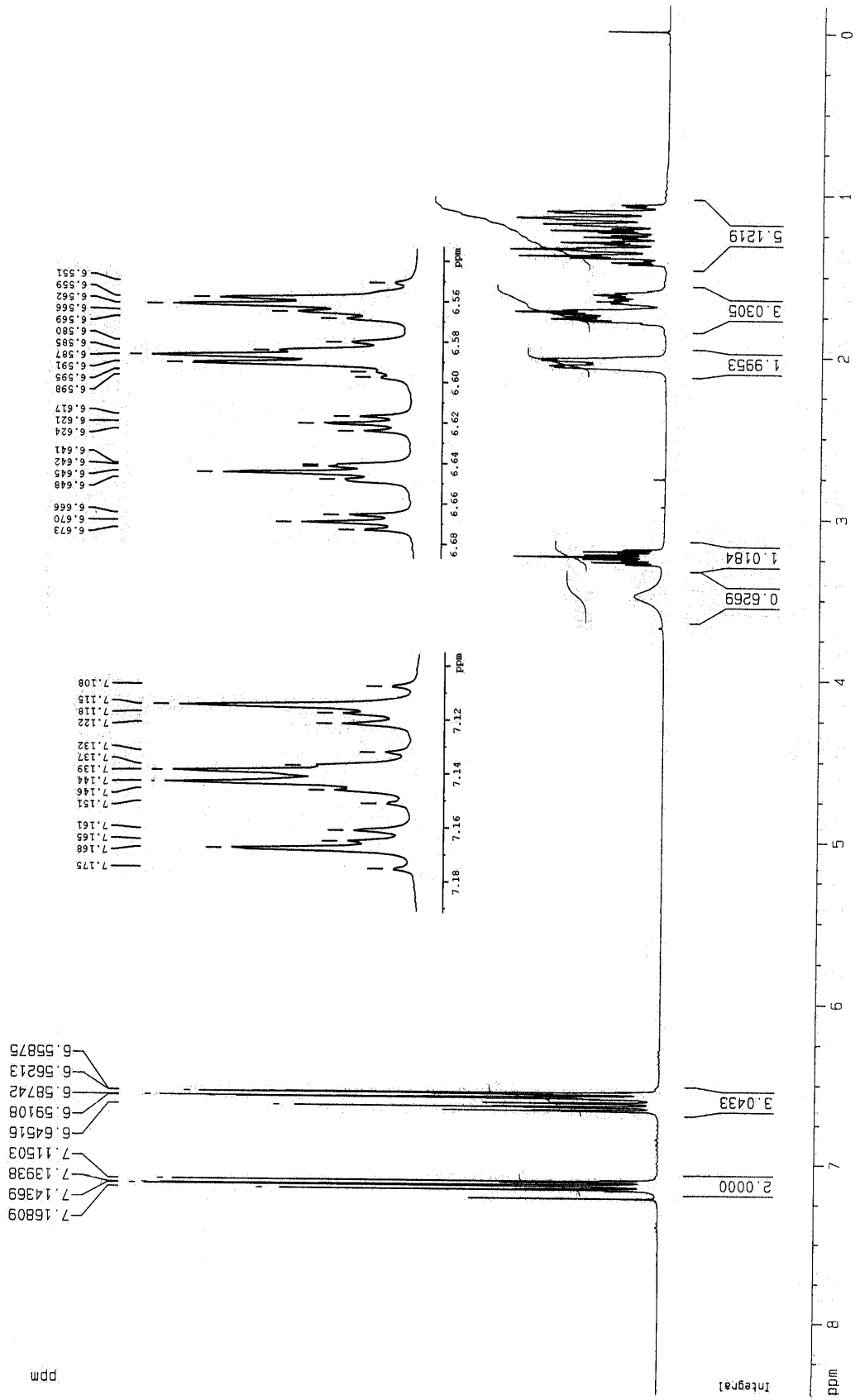
F2 - Processing parameters
SI 32768
SF 75.5658140 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



Instituto de Química UNAM NZ
 Dr. A. Cabrera/Laura R.
 Clave: Reacc 520
 CDC13
 Bruker-Avance 300MHz-F
 1H
 No. Registro: 442
 9-02-09



36



Instituto de Química UNAM NZ

Dr. A. Cabrera/Laura R.

Clave: Reacc 520

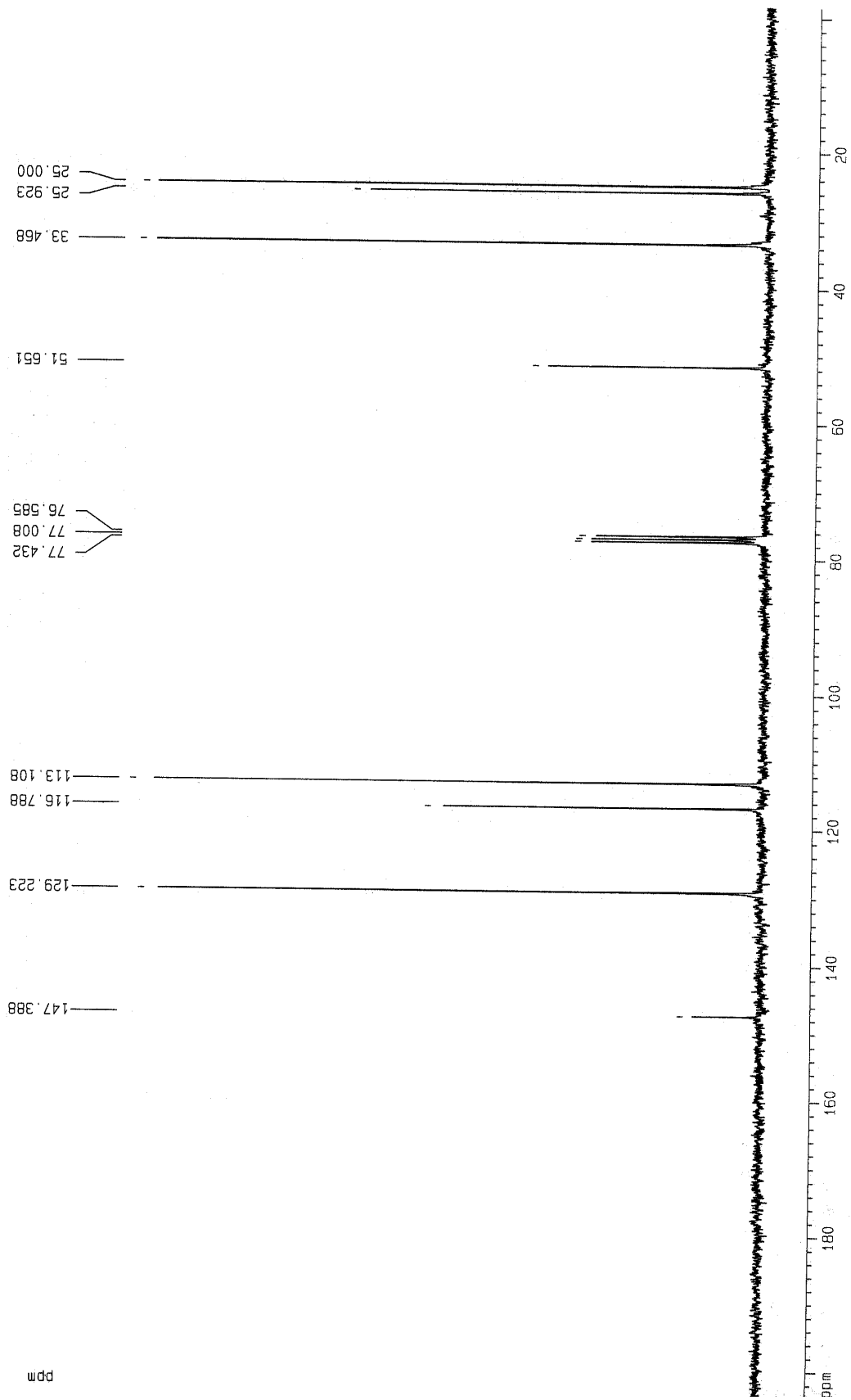
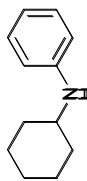
CDCl₃

Bruker-Avance 300MHz-F

¹³C

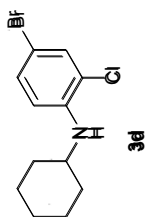
No. Registro: 442

9-02-09





4.250
4.228
3.337
3.304
3.292
3.279
3.246
2.078
2.066
2.061
2.045
2.037
1.841
1.829
1.815
1.798
1.785
1.723
1.714
1.711
1.698
1.685
1.679
1.671
1.659
1.644
1.486
1.476
1.465
1.460
1.448
1.438



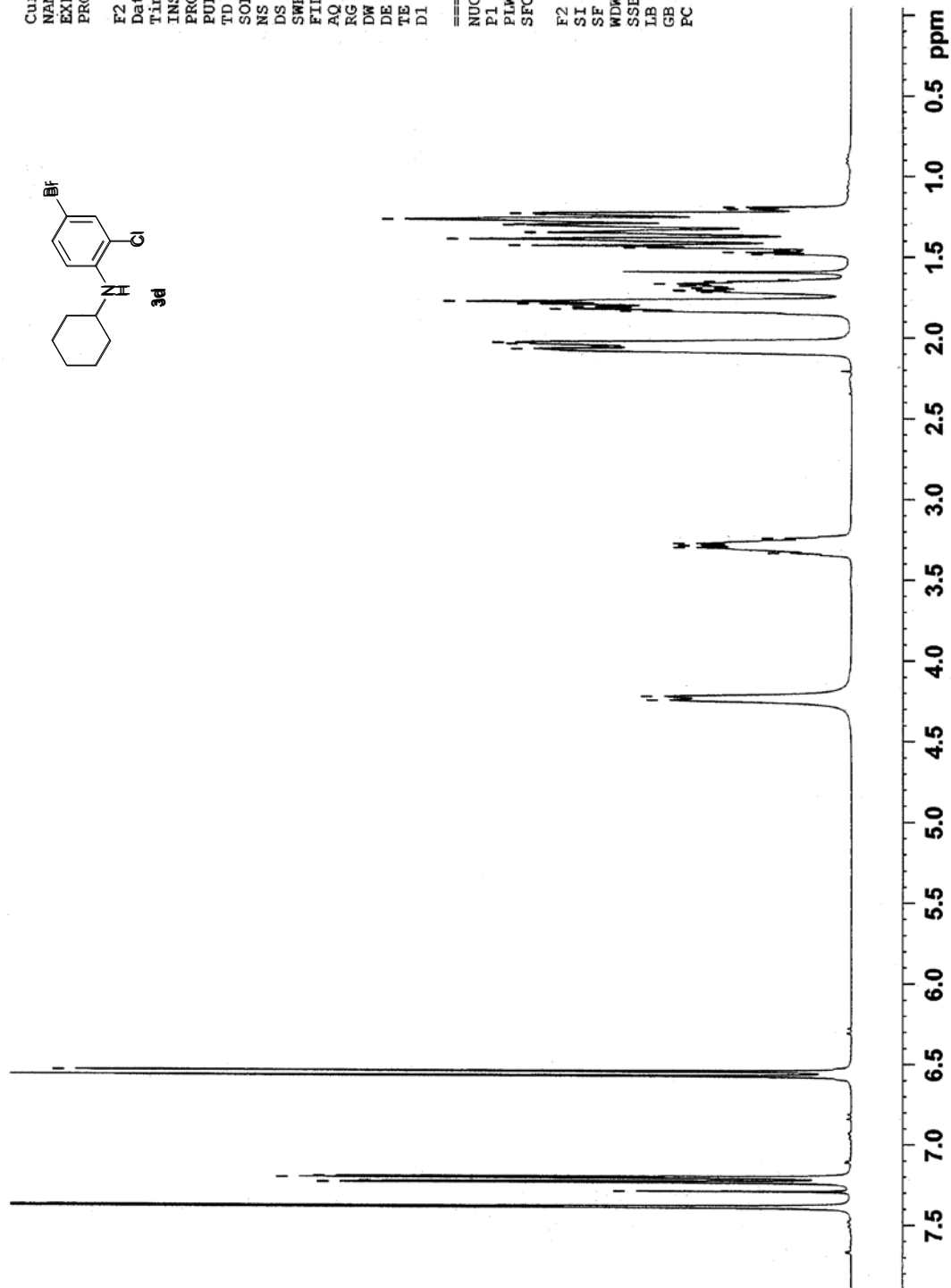
Current Data Parameters
NAME Reacc710F14
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120223
Time_ 9.31
INSTRUM spect
PROBHD 5 mm FAPBO BB-
PULPROG zg30
TD 65536
SOLVENT CDC13
NS 64
DS 1
SWH 6203.474 Hz
FIDRES 0.094658 Hz
AQ 5.2822518 sec
RG 114
DW 80.600 usec
DE 6.50 usec
TE 295.4 K
D1 1.0000000 sec

==== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PLW1 9.24709988 W
SF01 300.5218558 MHz

F2 - Processing parameters
SI 65536
SF 300.5200000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

7.395
7.387
7.294
7.241
7.233
7.212
7.204
6.578
6.548





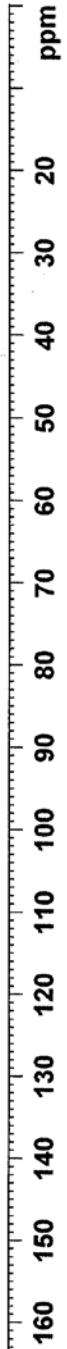
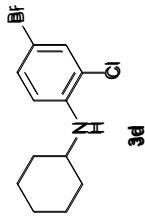
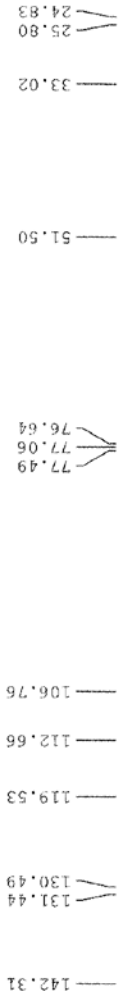
Current Data Parameters
NAME Reacc710F14
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120223
Time_ 10.38
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 1024
DS 1
SWH 18028.846 Hz
FIDRES 0.275098 Hz
AQ 1.8175818 sec
RG 203
DW 27.733 usec
DE 6.50 usec
TE 296.0 K
D1 2.00000000 sec
D11 0.03000000 sec

==== CHANNEL f1 =====
NUC1 13C
P1 7.90 usec
PLW1 48.96699905 W
SFO1 75.5733703 MHz

==== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PLW2 9.24709988 W
PLW12 0.21741000 W
PLW13 0.17610000 W
SFO2 300.5212021 MHz

F2 - Processing parameters
SI 32768
SF 75.5658140 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





```

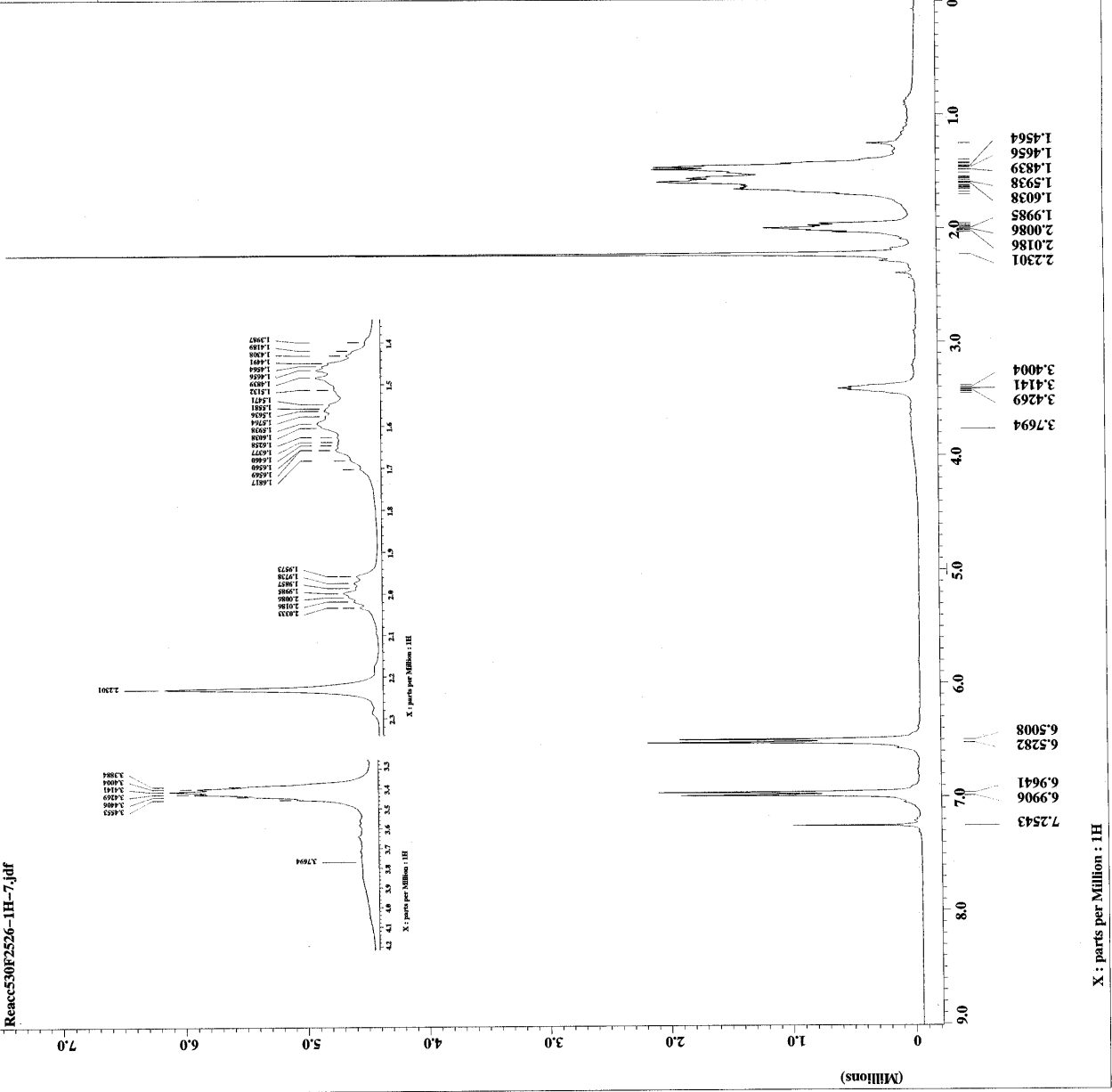
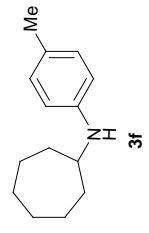
Filename = Reacc530F2526-1H-7.jd
Author = Cabrera
Experiment = single_pulse.exp
Sample_id = Laura
Solvent = CHLOROFORM-D
Creation time = 22-DEC-2008 08:03:03
Revision time = 5-MAR-2009 16:16:43
Current_time = 5-MAR-2009 16:16:42

Comment = Single Pulse Experiment
Data_format = 1D COMPLEX
Dim_size = 16384
Dim_title = 1H
Dim_units = [ppm]
Dimensions = X
Site = Eclipse+ 300
Spectrometer = DELTA_RMR

Field_strength = 7.0586013[T] (300[MEZ]
X_acq_duration = 3.6339712[fs]
X_domain = 1H
X_freq = 300.52965592[MEHz]
X_offset = 5[ppm]
X_points = 0
X_prescan = 0
X_resolution = 0.27518105[Hz]
X_sweep = 4.50856628[MEHz]
Clipped = FALSE
Mod_return = 1
Scans = 64
Total_scans = 64

X_90_width = 9.5[us]
X_acq_time = 3.6339712[fs]
X_angle = 45[deg]
X_pulse = 4.75[us]
Initial_wait = 1[fs]
Phase_preset = 3[us]
Recvr_gain = 20
Relaxation_delay = 1[fs]
Temp_get = 19.3[dc]
Unblank_time = 2[us]

```





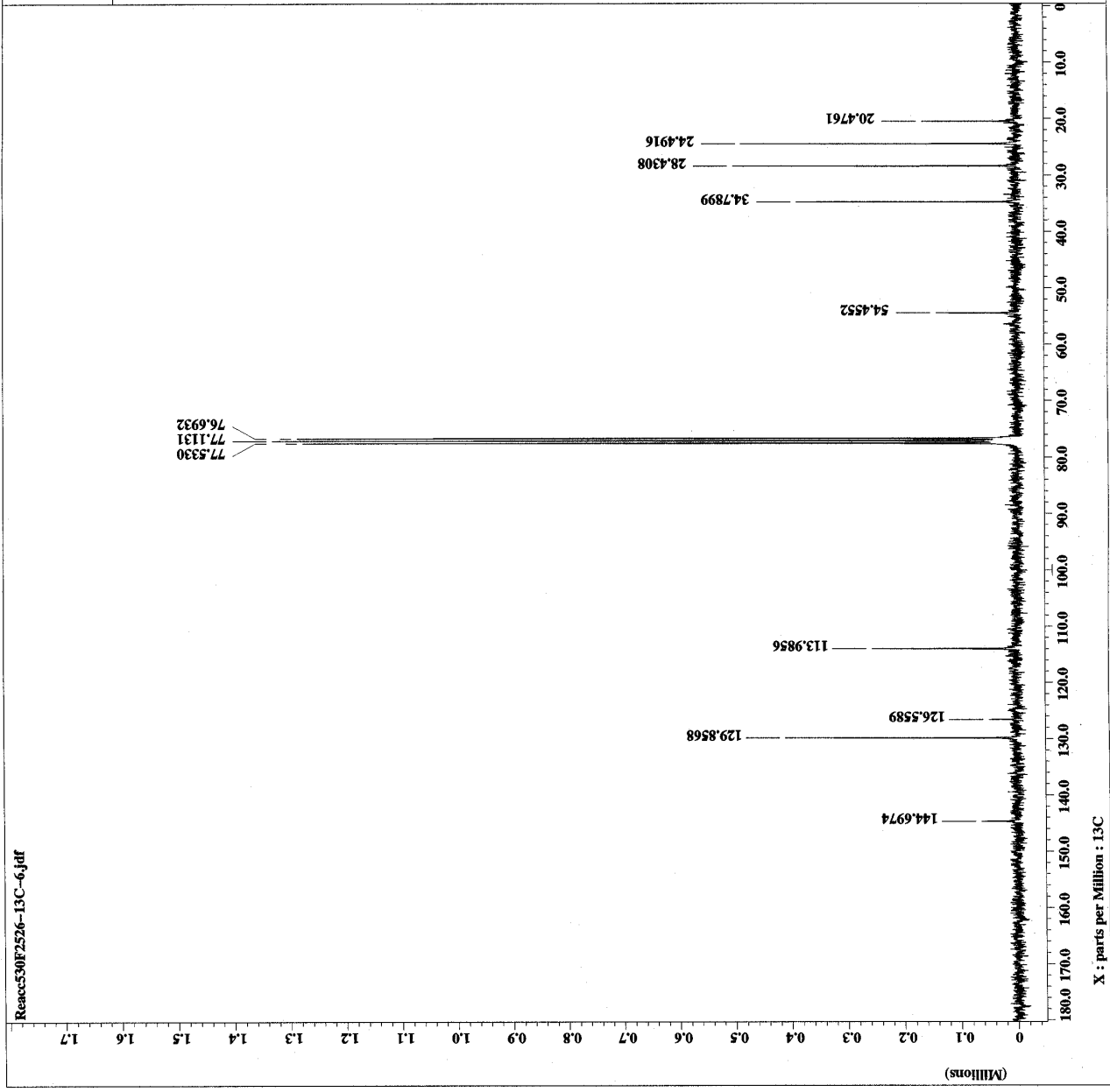
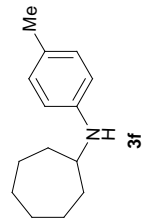
```

Filename = Reacc530F2526-13C-6.j
Author = Cabrera
Experiment = single_pulse_dec
Sample_id = Laura
Solvent = CHLOROFORM-D
Creation_time = 22-DEC-2008 08:50:02
Revision_time = 5-MAR-2009 16:56:22
Current_time = 5-MAR-2009 16:56:48

Comment = Single Pulse with Bro
Data_format = 1D COMPLEX
Dim_size = 32768
Dim_title = 13C
Dim_units = [ppm]
Dimensions = 2
Site = Scipase+ 300
Spectrometer = DELTA_NMR

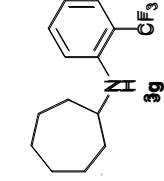
Field_strength = 7.0586013[F] (300[MHz]
X_acq_duration = 1.7334272[s]
X_domain = 13C
X_freq = 75.56823426[MHz]
X_offset = 100[ppm]
X_points = 32768
X_prescans = 4
X_resolution = 0.57689184[Hz]
X_sweep = 18.90359168[kHz]
Irr_domain = 1H
Irr_freq = 300.52965592[MHz]
Irr_offset = 5[ppm]
Clipped = FALSE
Mod_return = 1
Scans = 943
Total_scans = 943

X_90_width = 11.3[us]
X_acq_time = 1.7334272[s]
X_angle = 30[deg]
X_pulse = 3.76666667[us]
Initial_wait = 1[s]
Phase_preset = 3[us]
Recvr_gain = 30
Relaxation_delay = 1[s]
Temp_get = 21.1[dc]
Unblank_time = 2[us]
    
```





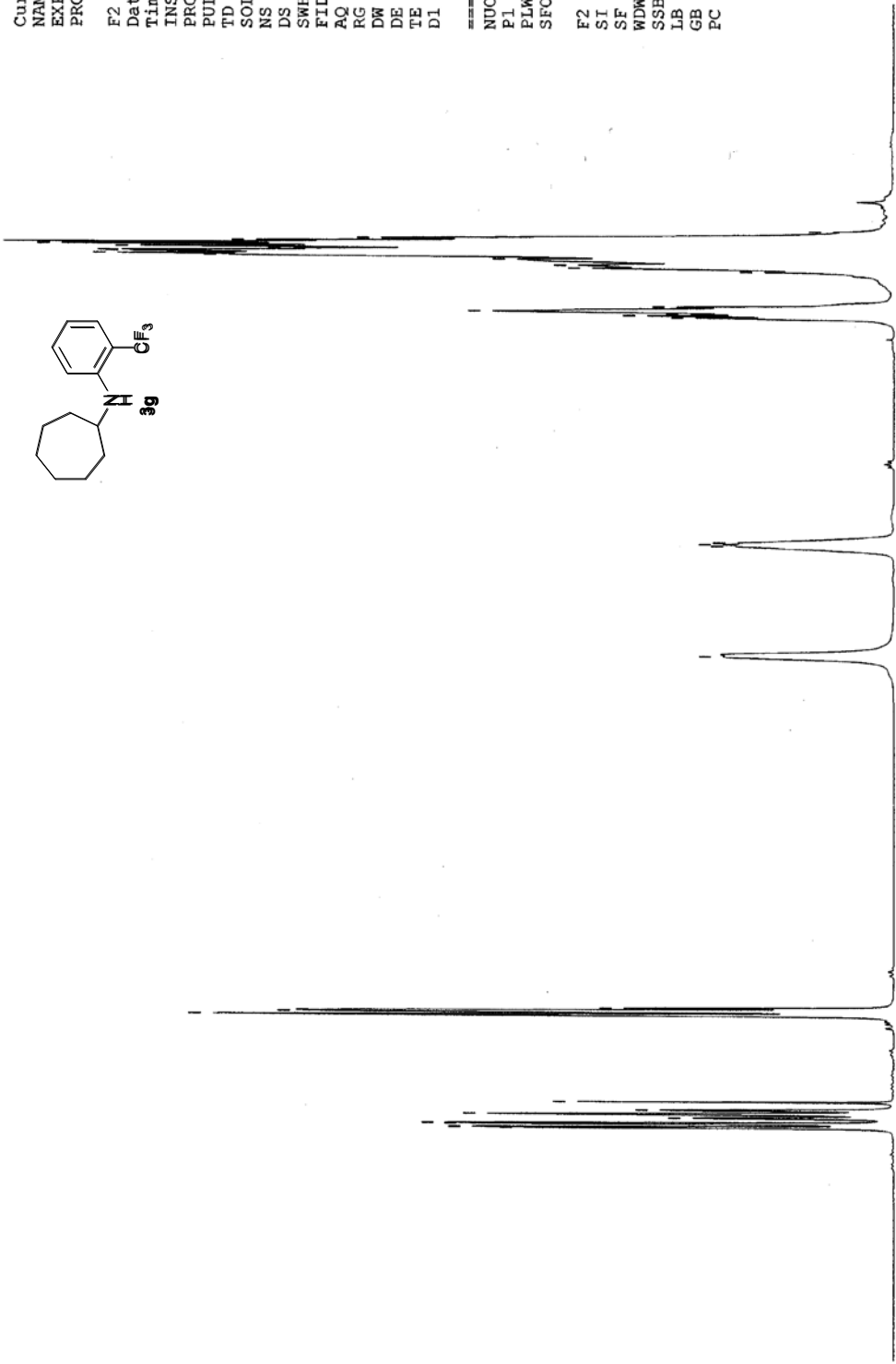
4.329
3.591
3.580
3.569
2.074
2.060
2.049
2.032
2.002
1.767
1.744
1.725
1.686
1.662
1.655
1.637
1.633
1.621
1.608
1.601
1.592
1.579
1.565
1.556



7.467
7.463
7.440
7.437
7.402
7.375
7.350
7.295
6.714
6.692
6.689
6.686
6.671
6.669

Current Data Parameters
NAME Reacc546F7-8
EXPNO 10
PROCNO 1
F2 - Acquisition Parameters
Date_ 20120302
Time_ 7.53
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 64
DS 2
SWH 6203.474 Hz
FIDRES 0.094658 Hz
AQ 5.2822518 sec
RG 128
DW 80.600 usec
DE 6.50 usec
TE 295.4 K
D1 1.00000000 sec

==== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PLW1 9.24709988 W
SF01 300.5218558 MHz
F2 - Processing parameters
SI 65536
SF 300.5200000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 ppm



Current Data Parameters
NAME Reacc546F7-8
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters

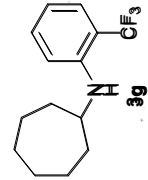
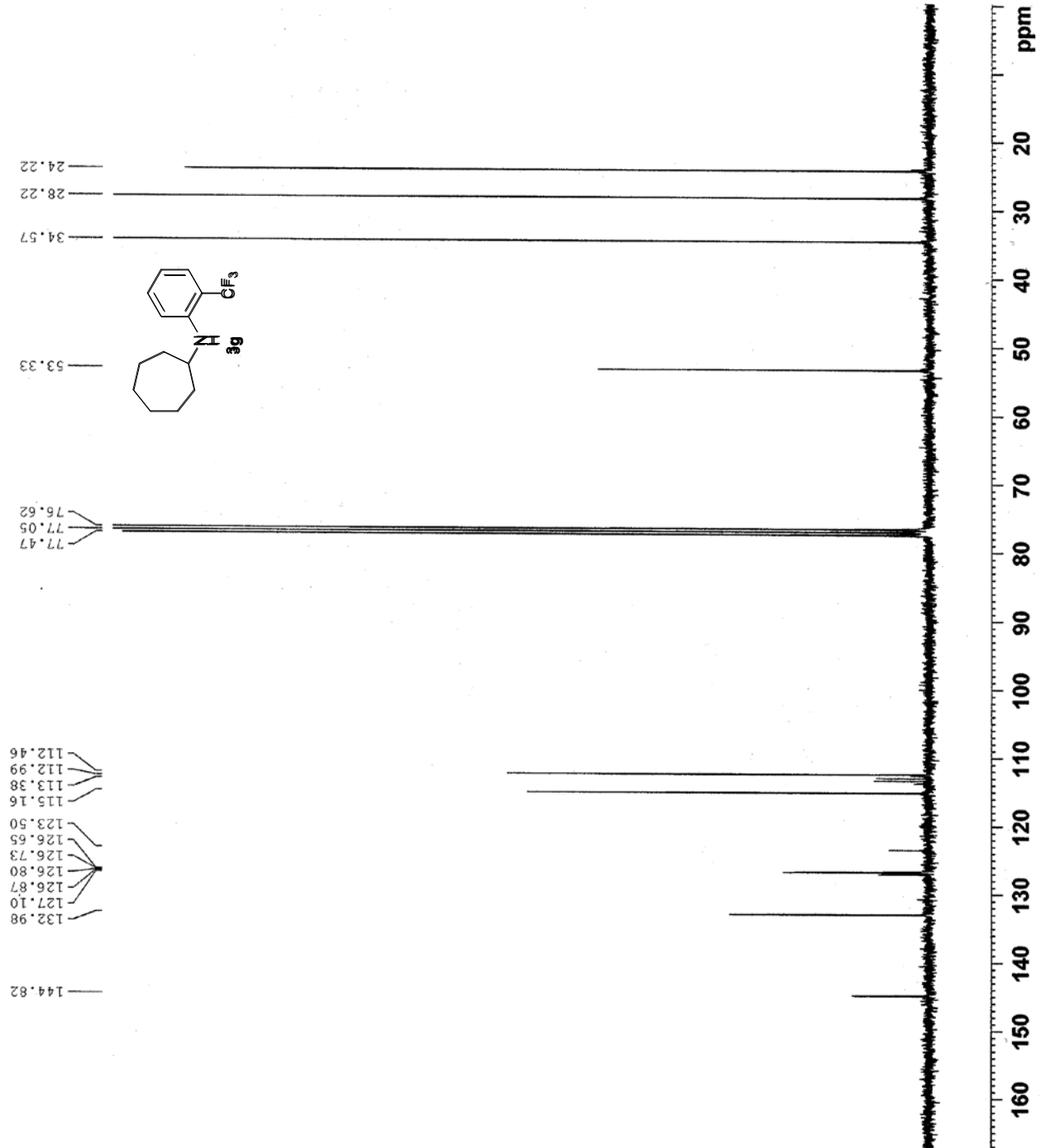
Date_ 20120302
Time_ 8.47
INSTRUM spect
PROBHD 5 mm FAPBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 800
DS 1
SWH 18028.846 Hz
FIDRES 0.275098 Hz
AQ 1.8175818 sec
RG 203
DM 27.733 usec
DE 6.50 usec
TE 296.0 K
D1 2.00000000 sec
D11 0.03000000 sec

==== CHANNEL f1 =====
NUC1 13C
P1 7.90 usec
PLW1 48.9669905 W
SFO1 75.5733703 MHz

==== CHANNEL f2 =====
CPDPRG2 waitz16
NUC2 1H
PCPD2 90.00 usec
PLW2 9.24709988 W
PLW12 0.21741000 W
PLW13 0.17610000 W
SFO2 300.5212021 MHz

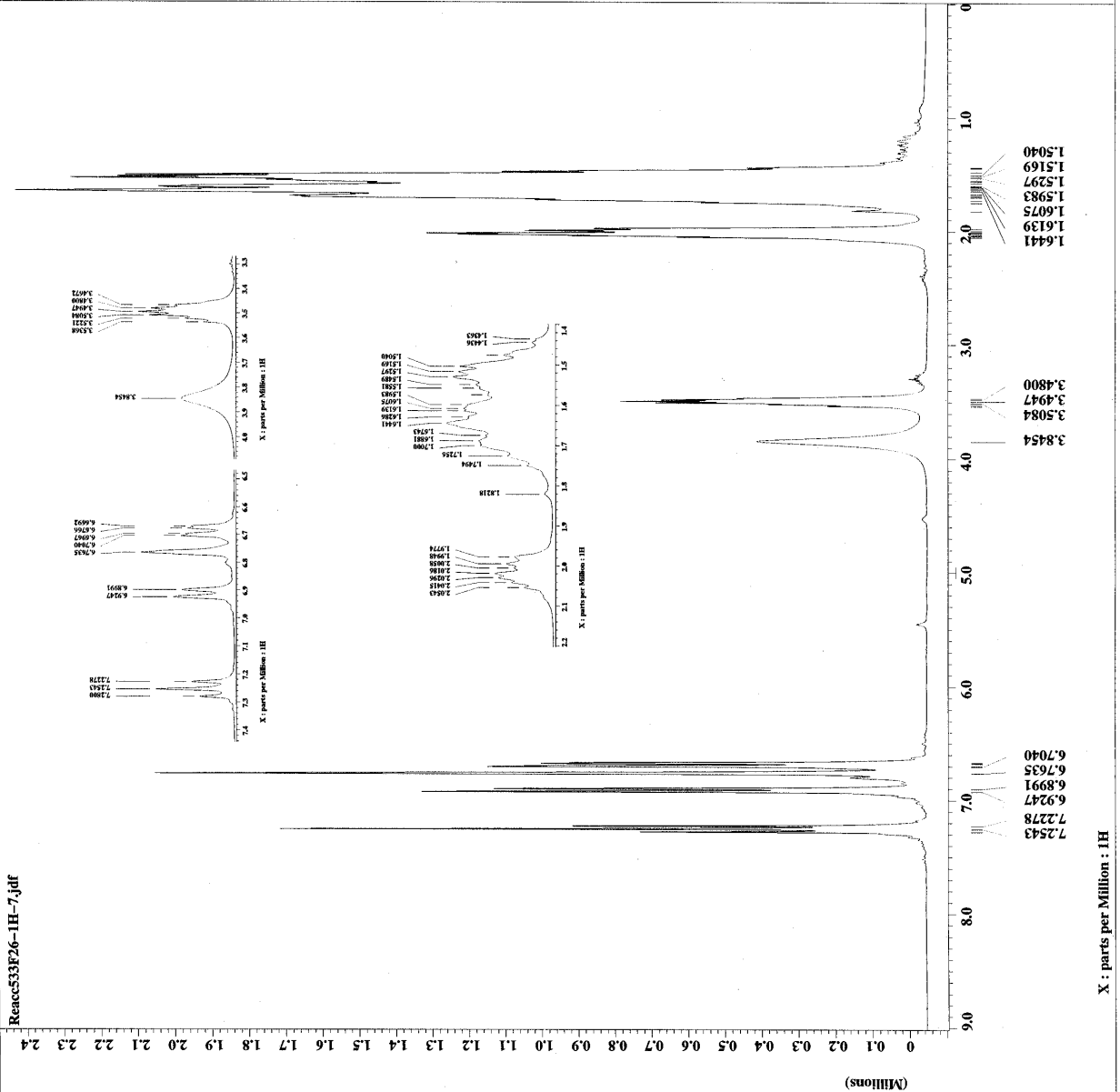
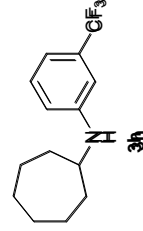
F2 - Processing parameters

SI 32768
SF 75.5658140 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





Reacc533F26-1H-7.jdf
= Cabrera
= single_pulse.exp
= laura.ppmk.D
= 28-DEC-2008 08:01:30
= 17-MAR-2009 13:07:04
= 17-MAR-2009 13:07:53
= Single Pulse Experiment
= 1D COMPLEX
= 16384
= 1H
= [ppm]
= X
= Eclipse+ 300
= DELTA_NMR
Spectrometer
Field_strength = 7.0586013[T] (300[Mhz])
X_acq_duration = 3.6339712[s]
X_domain = 1H
X_freq = 300.52965592[Mhz]
X_offset = 5.199ppm
X_points = 65384
X_time = 1.000000000
X_resolution = 0.27518105[Hz]
X_sweep = 4.50856628[Mhz]
Clipped = FALSE
Mod_return = 1
Scans = 32
Total_scans = 32
X_90_width = 9.5[us]
X_acq_time = 3.6339712[s]
X_angle = 45[deg]
X_pulse = 4.75[us]
Initial_wait = 1[s]
Phase_preset = 3[us]
Recvr_gain = 13
Relaxation_delay = 1[s]
Temp_get = 30.3[degC]
Onbftank_time = 2[us]





Current Data Parameters
NAME Reacc545F12
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120228
Time_ 8.21
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 64
DS 1
SWH 6203.474 Hz
FIDRES 0.094658 Hz
AQ 5.2822518 sec
RG 57
DW 80.600 usec
DE 6.50 usec
TE 295.2 K
D1 1.00000000 sec

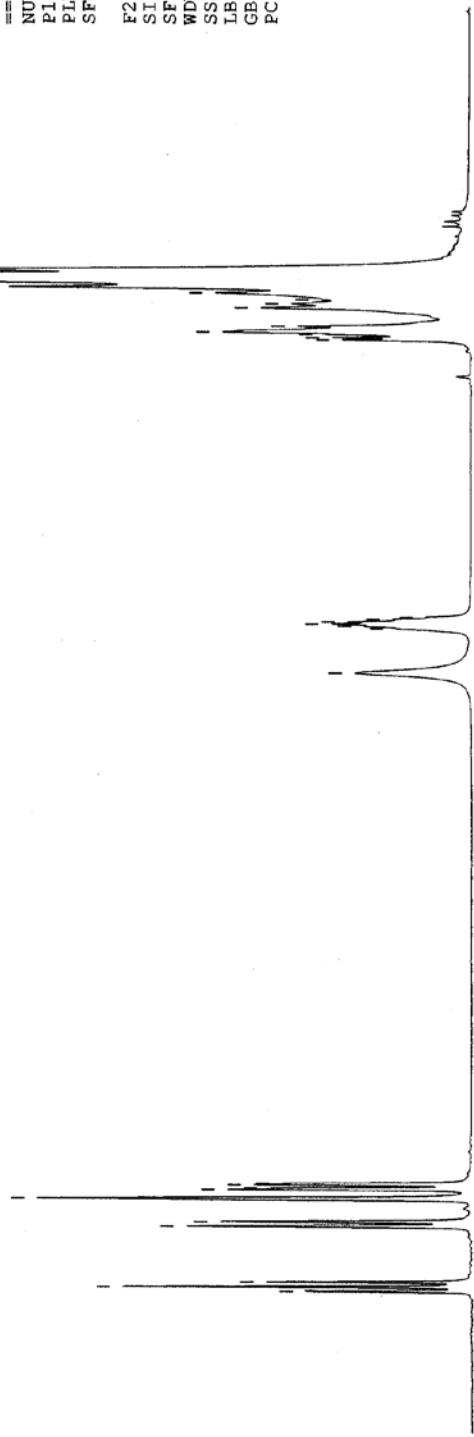
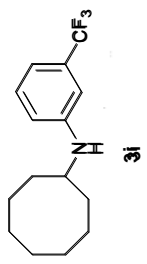
==== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PLW1 9.24709988 W
SFO1 300.5218558 MHz

F2 - Processing parameters
SI 65536
SF 300.5200407 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

7.164
7.159
7.138
7.112
6.799
6.773
6.644
6.593
6.586
6.566
6.559

3.701
3.451
3.438
3.426
3.414
3.400
3.387

1.839
1.827
1.809
1.796
1.763
1.661
1.638
1.615
1.578
1.547
1.518
1.513
1.478
1.449



7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 ppm



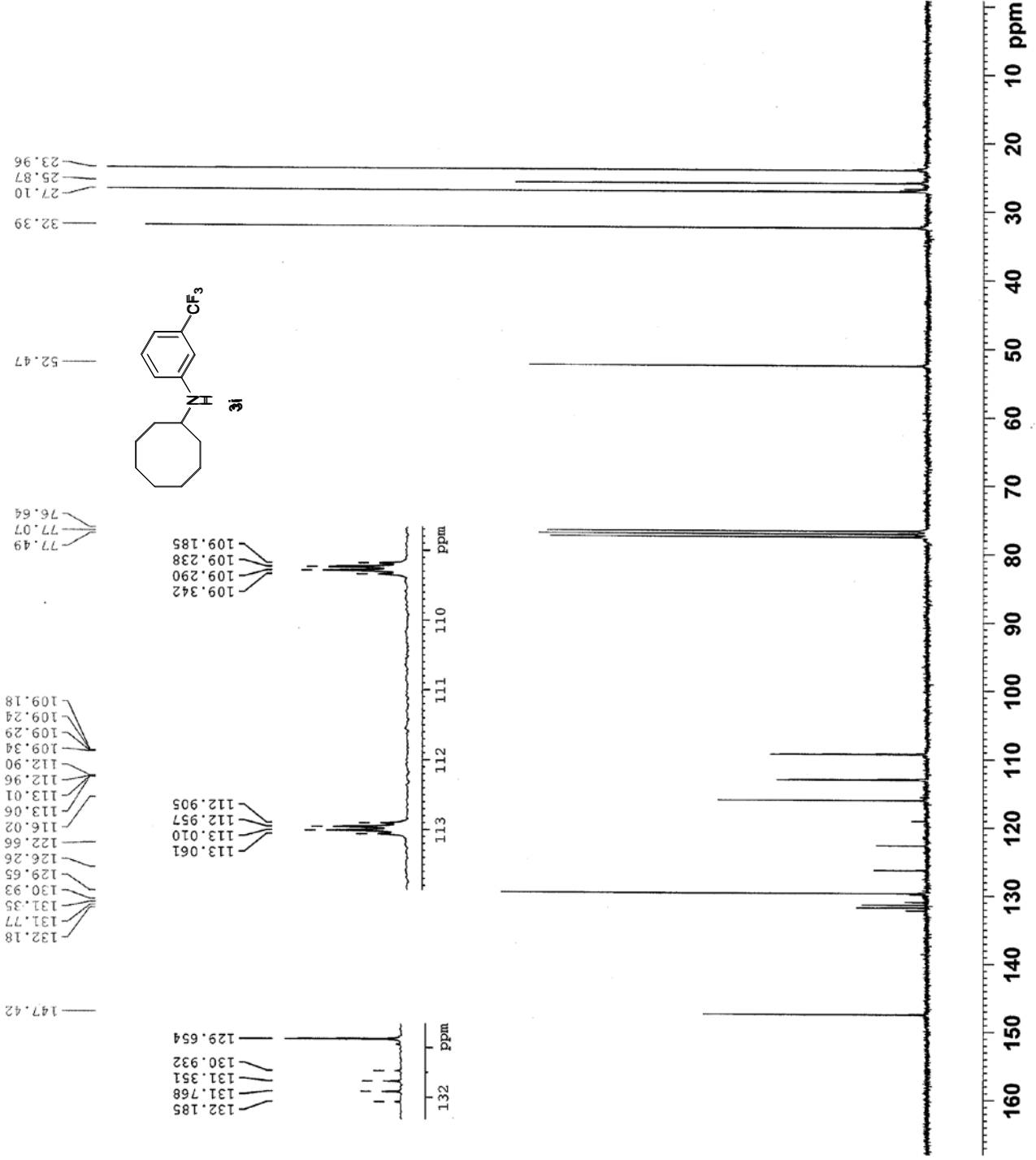
Current Data Parameters
NAME Reacc545F12
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120228
Time 9.08
INSTRUM spect
PROBHD 5 mm PABDO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 700
DS 1
SWH 18028.846 Hz
FIDRES 0.275098 Hz
AQ 1.8175818 sec
RG 203
DW 27.733 usec
DE 6.50 usec
TE 295.8 K
D1 2.0000000 sec
D11 0.03000000 sec

==== CHANNEL f1 =====
NUC1 13C
P1 7.90 usec
PLW1 48.9669905 W
SFO1 75.5733703 MHz

==== CHANNEL f2 =====
CFPRG2 waltz16
NUC2 1H
ECPD2 90.00 usec
PLW2 9.24709988 W
PLW12 0.21741000 W
PLW13 0.17610000 W
SFO2 300.5212021 MHz

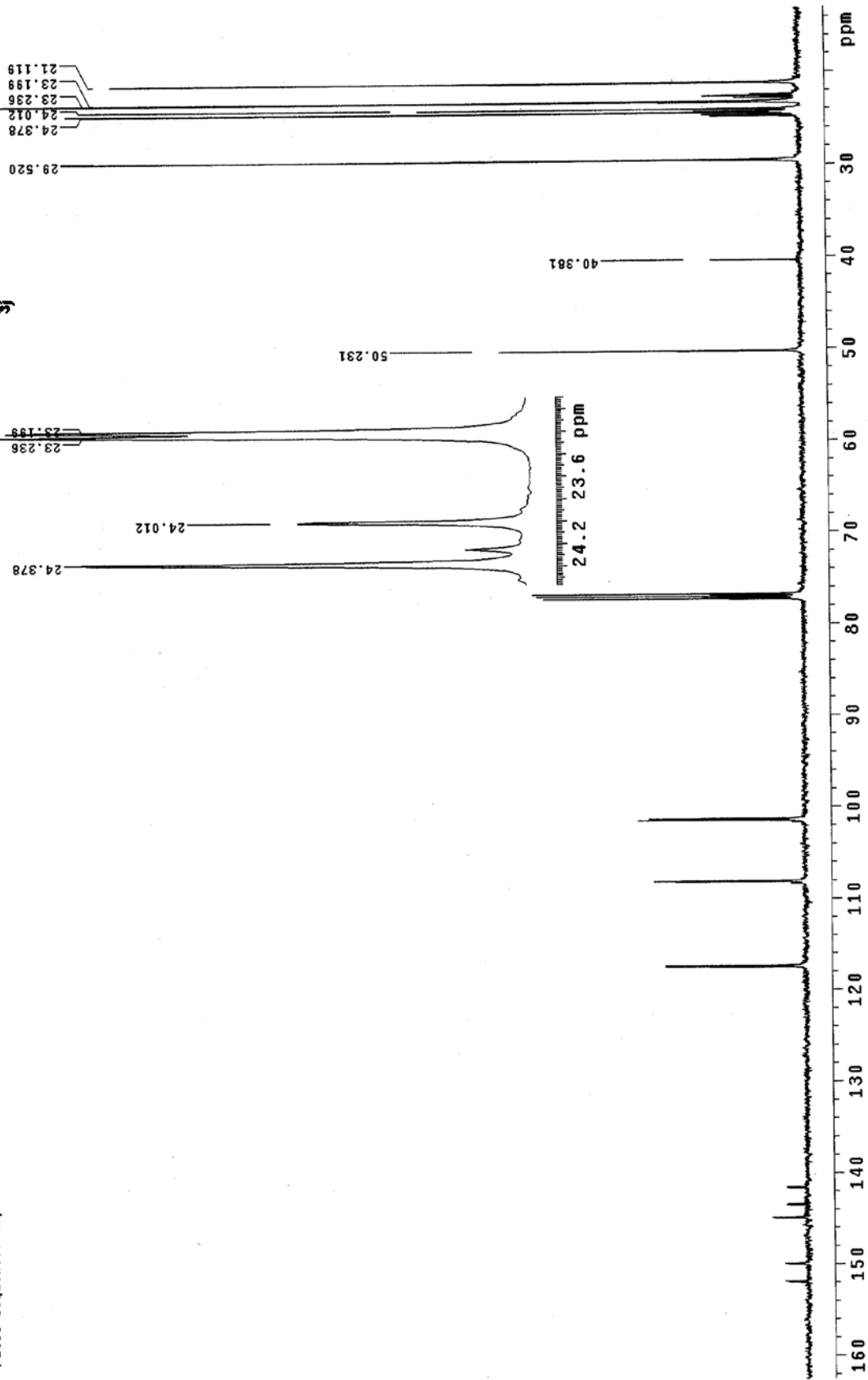
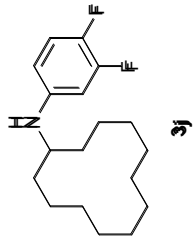
F2 - Processing parameters
SI 32768
SF 75.5658140 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



UNAM - Instituto de Química (H. Ríos)
Dr.-Acabrera
Clave: Reacc724F30
No. registro: 1114
Experimento: RMN-C13
Disolvente: CDCl3
Unity/Inova-125.71MHz (G)
18-Mayo-2012

File: Carbon

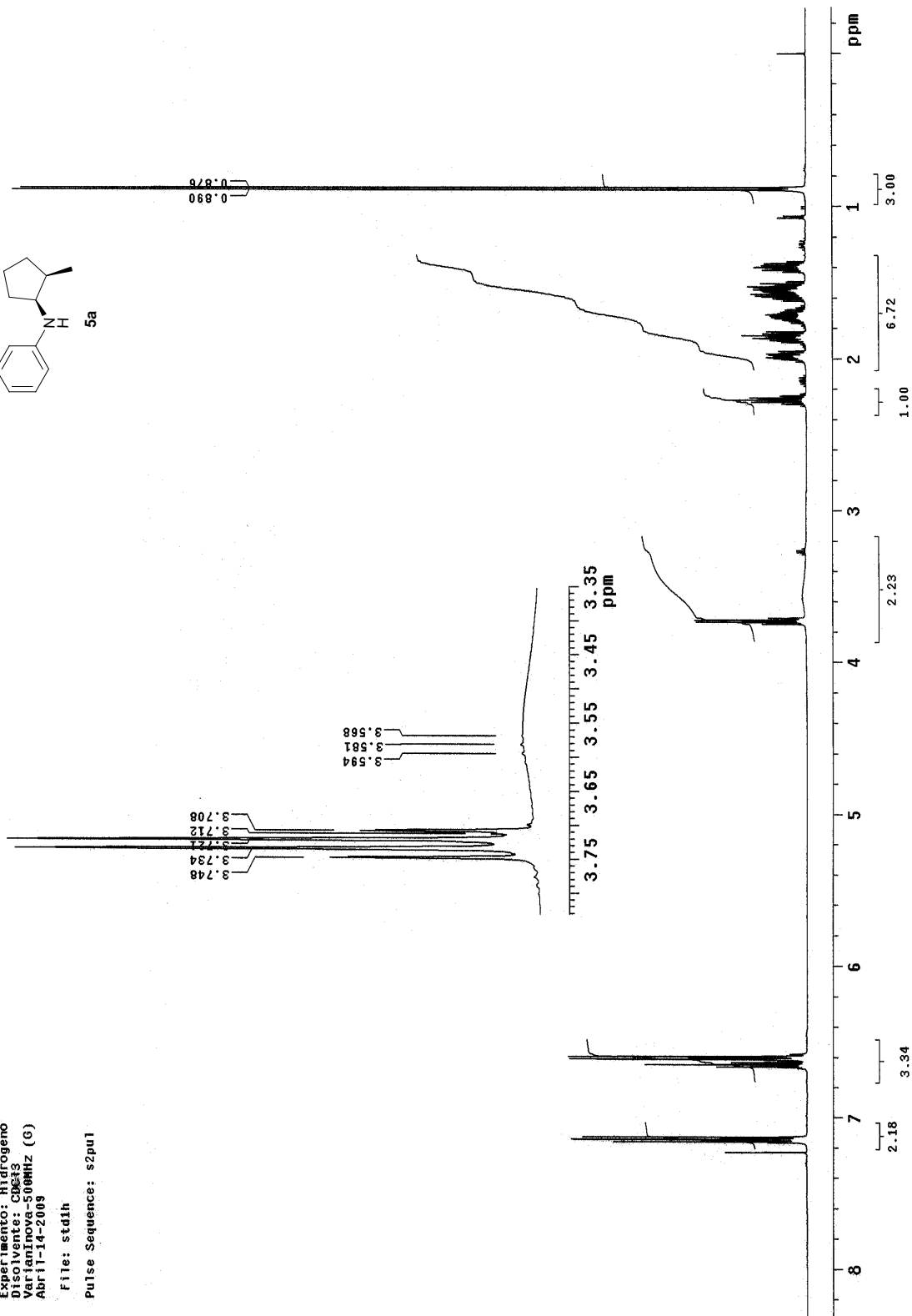
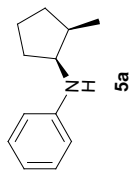
Pulse Sequence: s2pul



UNAM, Instituto de Química. (H. Ríos)
Dr-A-Cabrera/Laura-R.P.
Clave: Reacc952
No. Registro: 996
Experimento: Hidrogeno
Disolvente: CDCl3
VarianInova-500MHz (S)
Abril-14-2009

File: std1h

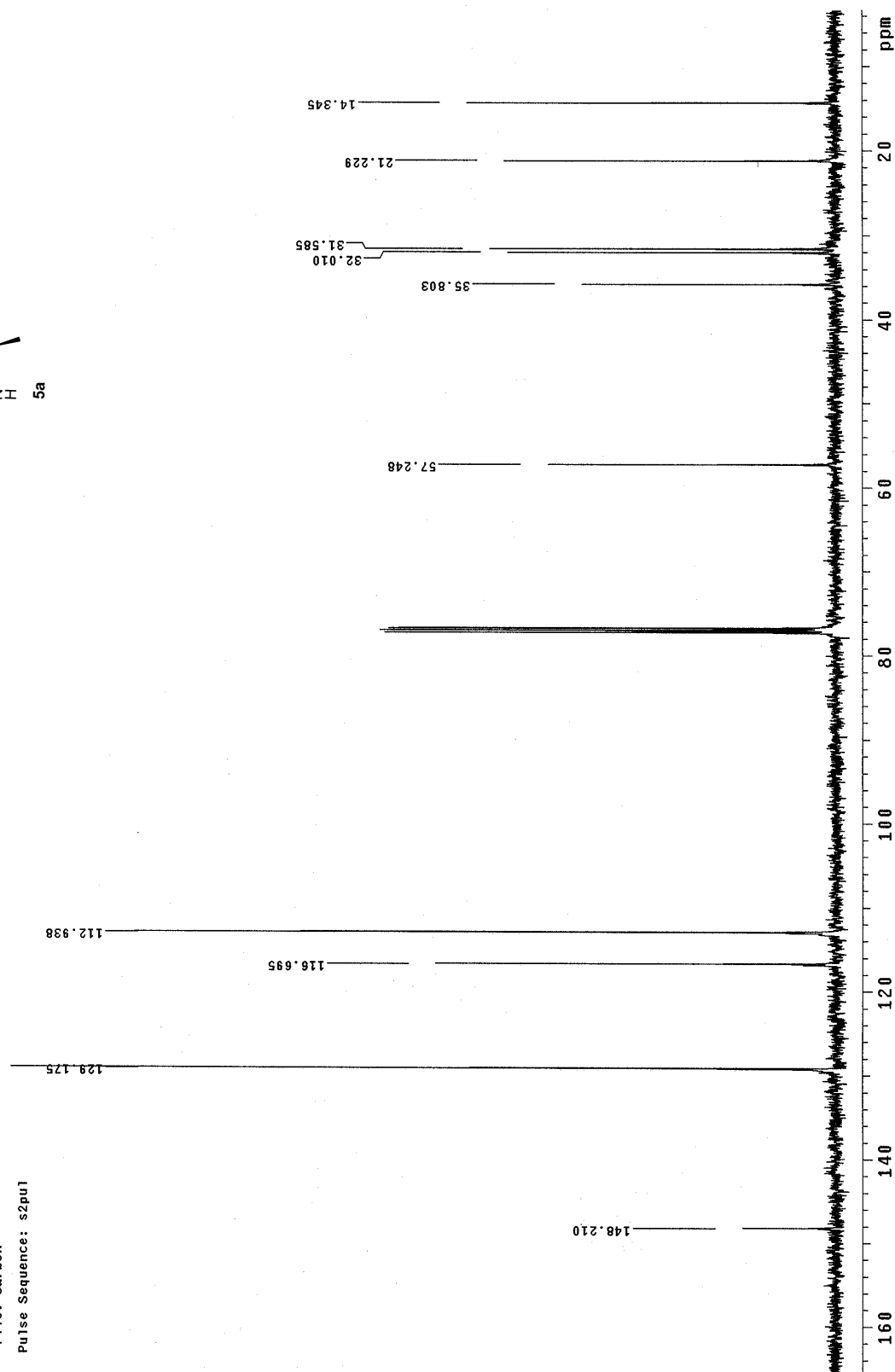
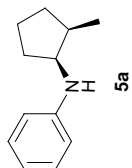
Pulse Sequence: s2pul



UNAM. Instituto de Química (H. Ríos)
Dr-A-cabrera/laura-R.P
Clave: Reacc352
No. Registro: 996
Experimento: C13
Disolvente: CDCl3
UnityInova-125.71MHz (G)
Abr11-14-2009

File: Carbon

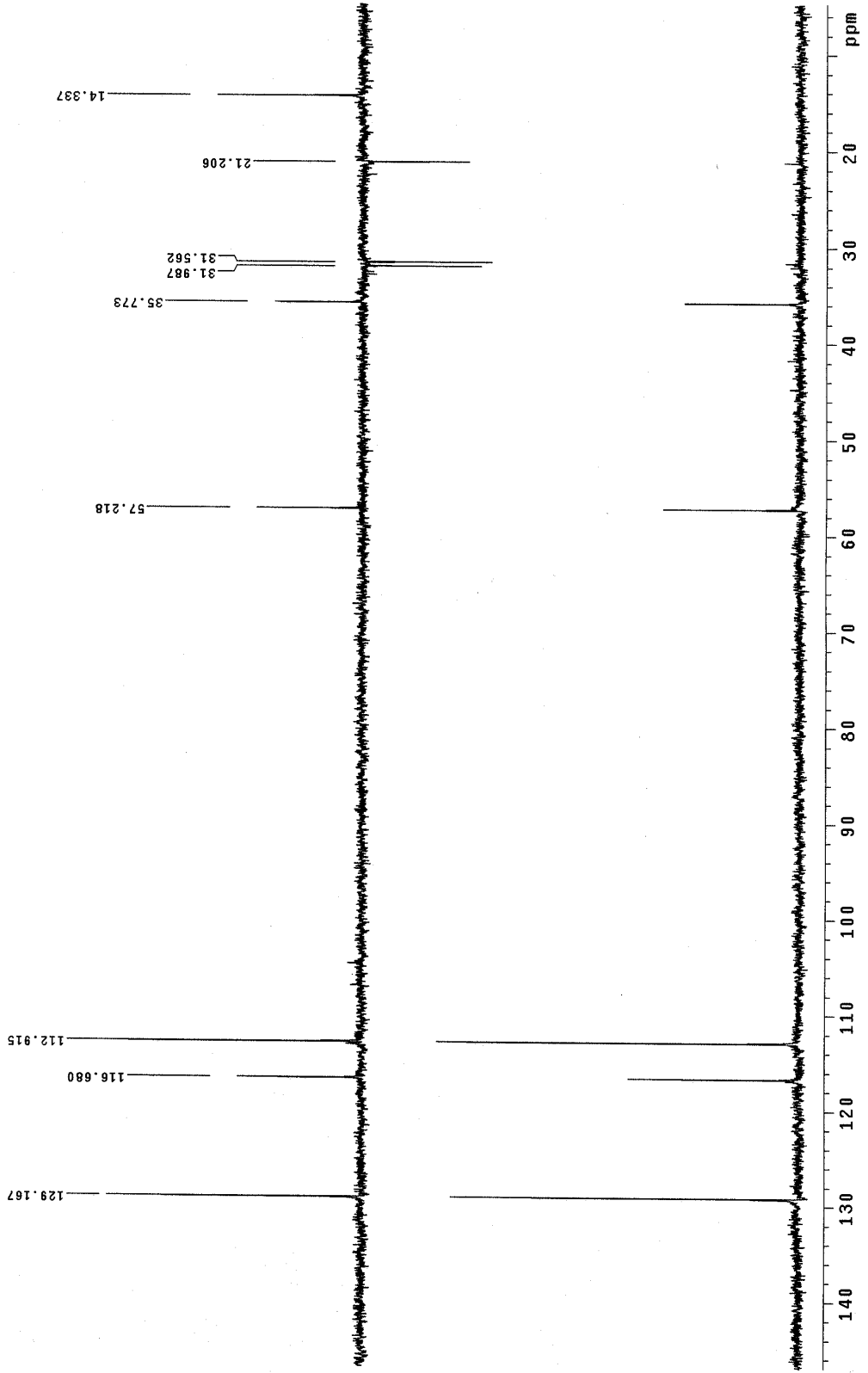
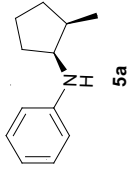
Pulse Sequence: s2pu1



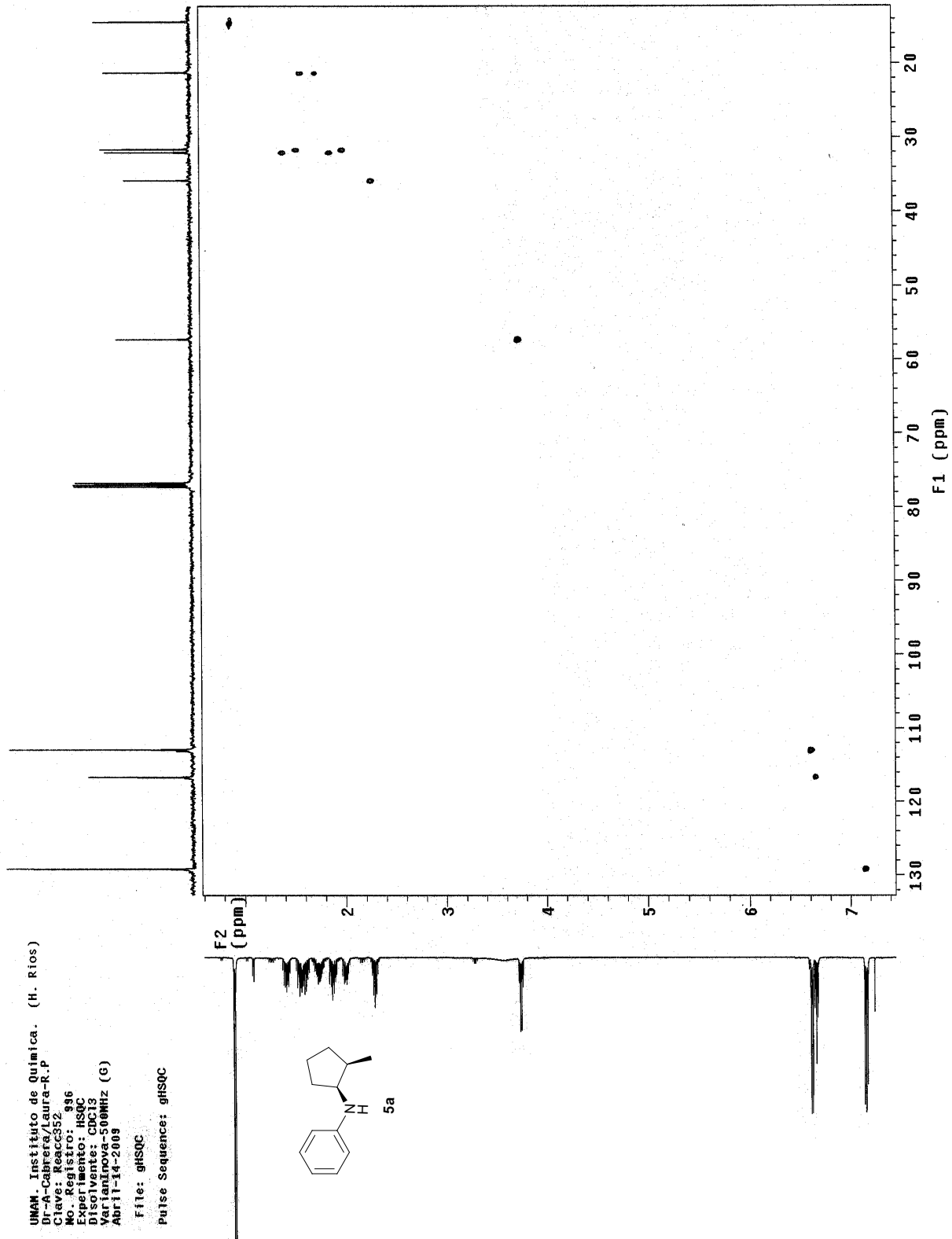
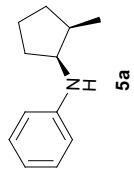
UNAM, Instituto de Química. (H. Ríos)
Dr-A-Cabrera/Laura-K.P
Clave: Reacc352
No. Registro: 396
Experimento: Dept
Carilini/125.7 MHz (G)
Abr11-14-2008

File: Dept

Pulse Sequence: DEPT

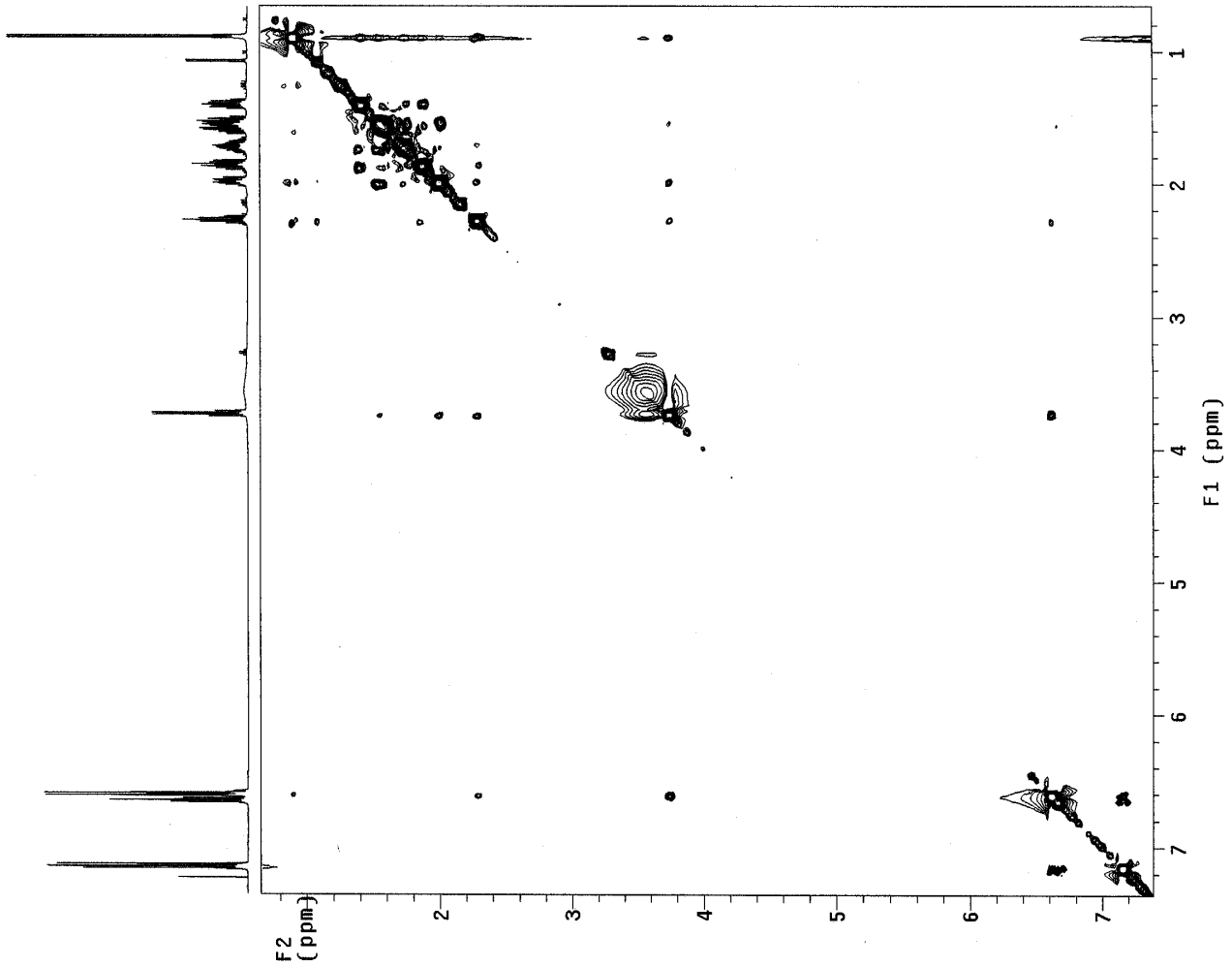
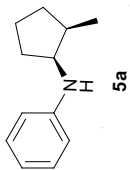


UNAM. Instituto de Quimica. (H. Rios)
Dr-A-Cabrera/Laura-R.P
Clave: Reacc52
Mo. Registro: 986
Experimento: HSQC
Disolvente: CDCl3
Varian/ova-500MHz (G)
Abril-14-2009
File: gHSQC
Pulse Sequence: gHSQC

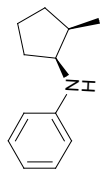


UNAM. Instituto de Quimica.
Dr-A-Cabrera/Laura-R.P
Clave: Reacc352
No. Registro: 996
Experimento: Noesy
Disolvente: CDCl3
Variante: 500MHZ (G)
Abril-14-2009

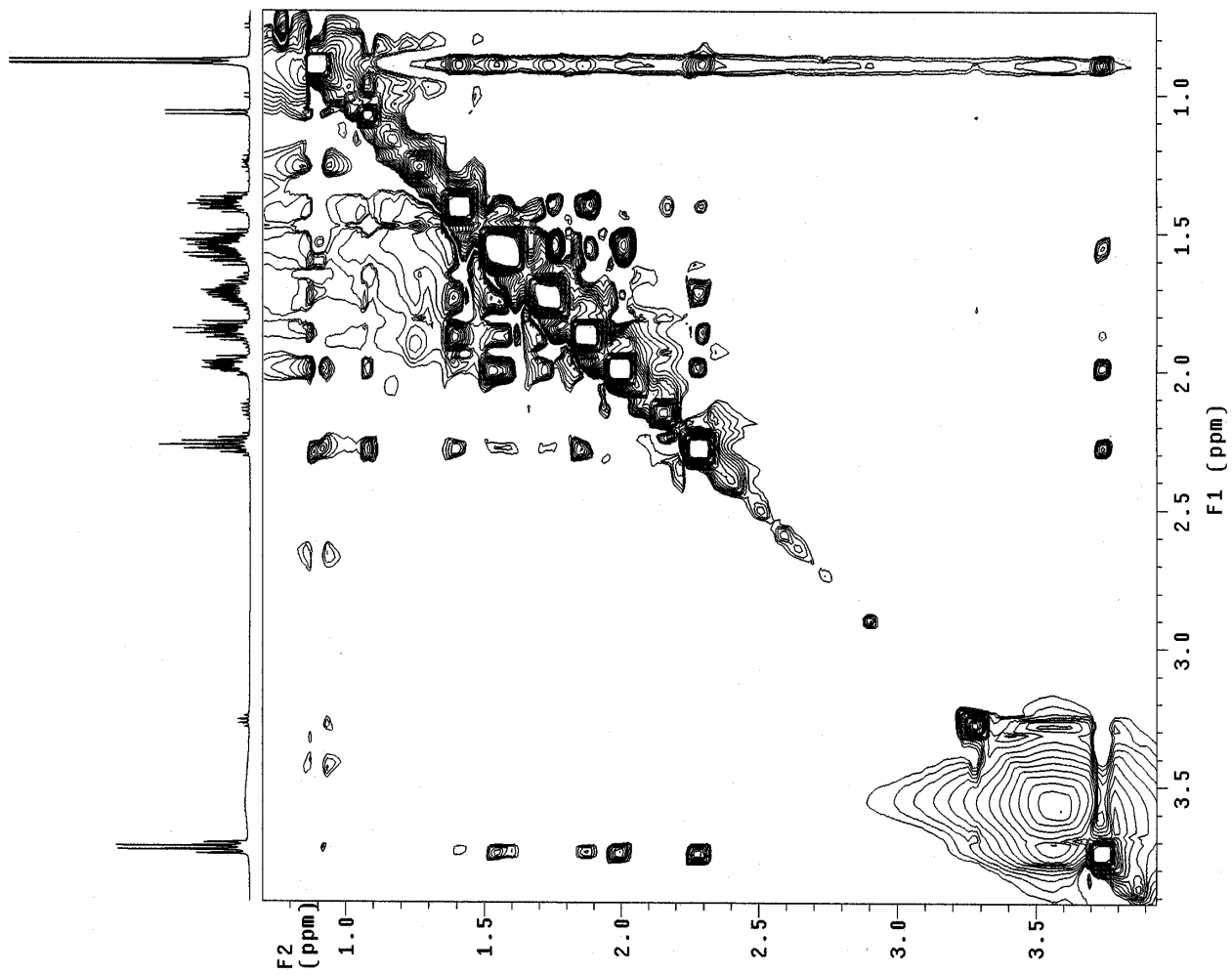
Pulse Sequence: noesy



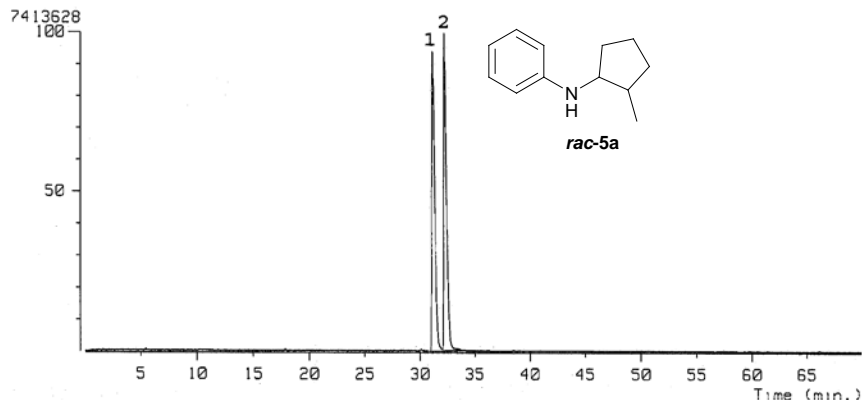
UNAH. Instituto de Quimica.
Dr.-A-Cabrera/Laura-R.P
Clave: Reacc352
No. Registro: 996
Experimento: Nocsy
Disolvente: CDCl3
Varianinova-500MHz (G)
Abr11-14-2009
Pulse Sequence: nocsy



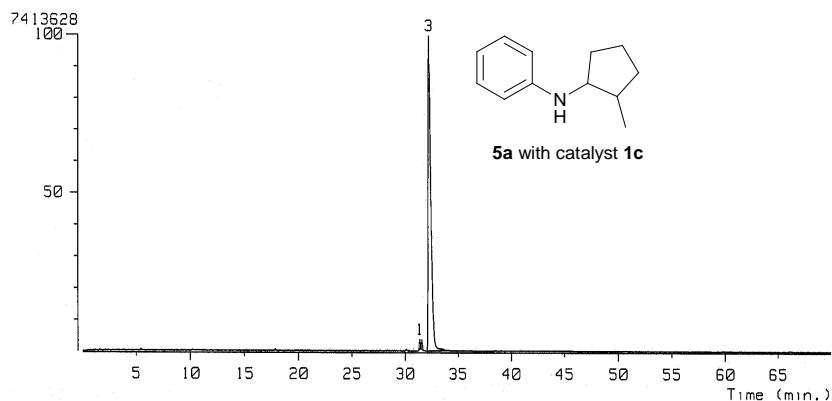
5a
NOE-Expansions



[TIC]
 Data : Dr-Cabrera-Armando-962 Date : 16-Feb-120 16:27
 Sample: 405 G reacc 321-rac AX505HA
 Note :
 Inlet : GC Ion Mode : EI+
 Ion Species : Normal Ion TIC Range : m/z 5 to 650

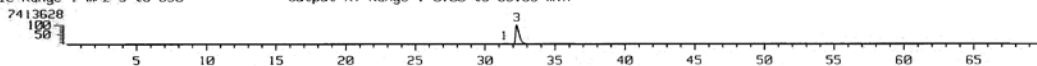


[TIC]
 Data : Dr-Cabrera-Armando-870 Date : 23-Nov-107 12:41
 Sample: 2324 G Reacc 352 AX505HA
 Note :
 Inlet : GC Ion Mode : EI+
 Ion Species : Normal Ion TIC Range : m/z 5 to 650

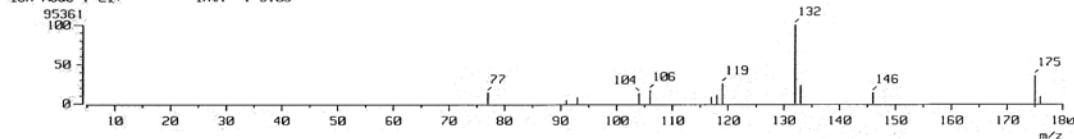


No.	RT[min]	Area	Area%	Height	Height%	Width[sec]	INTEG
1	31.35	135.68	1.06	22.01	2.95	5.79	BB
2	31.55	135.08	1.05	20.51	2.75	6.18	BB
3	32.24	12542.16	97.89	704.79	94.31	16.71	BB

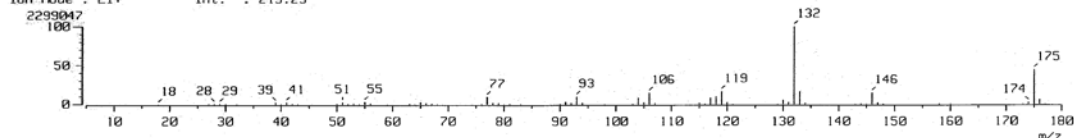
[TIC]
 Data : Dr-Cabrera-Armando-870 Date : 23-Nov-107 12:41
 Sample: 2324 G Reacc 352 AX505HA
 Note :
 Inlet : GC Ion Mode : EI+
 Ion Species : Normal Ion (MF-Linear) Output RT Range : 0.00 to 69.99 min
 TIC Range : m/z 5 to 650



[Mass Spectrum]
 RT : 31.55 min Scan# : 2367-2368-23 Temp : 0.0 deg.C
 Ion Mode : EI+ Int. : 9.09

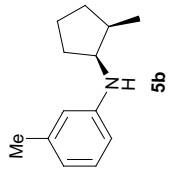
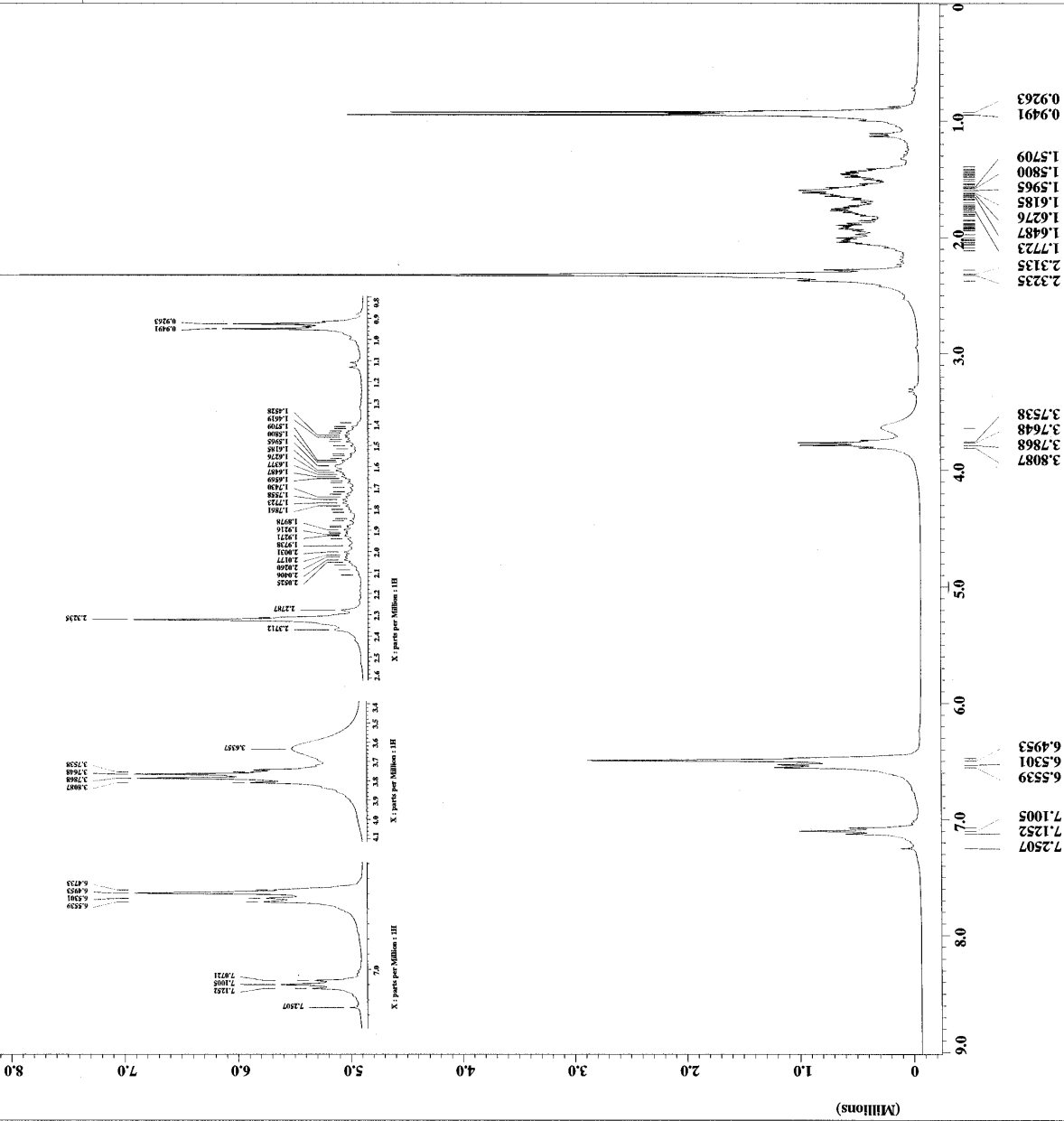


[Mass Spectrum]
 RT : 32.24 min Scan# : 2419-2403-2555 Temp : 0.0 deg.C
 Ion Mode : EI+ Int. : 219.25





Reacc507-2F1011-1H-9.jdf



```

Filename = Reacc507-2F1011-1H-9.
Author = Cabrera
Experiment = single_pulse.exp
Sample_id =
CURPROG =
PULPROG = zgpg30
Date_ Acquired = 30-NOV-2008 01:58:16
Revision_time = 11-NOV-2008 09:10:14
Current_time = 11-NOV-2008 09:12:23
Comment = Single Pulse Experiment
Data format = 1D COMPLEX
Dim_size = 16384
Dim_title =
Dim_units = 1H
Dimensions = X
Site = Eclipse+ 300
Spectrometer = DELTA_NMR
Field_strength = 7.0586013[T] (300 [MEZ]
X_acq_duration = 3.6339712[s]
X_domain = 1H
X_freq = 300.52965592[MEZ]
X_offset = 5[ppm]
X_points = 16384
X_prescans = 0
X_resolution = 0.27518105[Hz]
X_sweep = 4.50856628[kHz]
Clipped = FALSE
Mod_return = 1
Scans = 32
Total_scans = 32
X_90_width = 9.5[us]
X_acq_time = 3.6339712[s]
X_angle = 45[deg]
X_pulse = 4.75[us]
initial_wait = 3[s]
Phase_preset = 15
Recv_start = 1[s]
Relaxation_delay = 19.6[dc]
Temp_set = 2[us]
Unblank_time = 2[us]
  
```



```

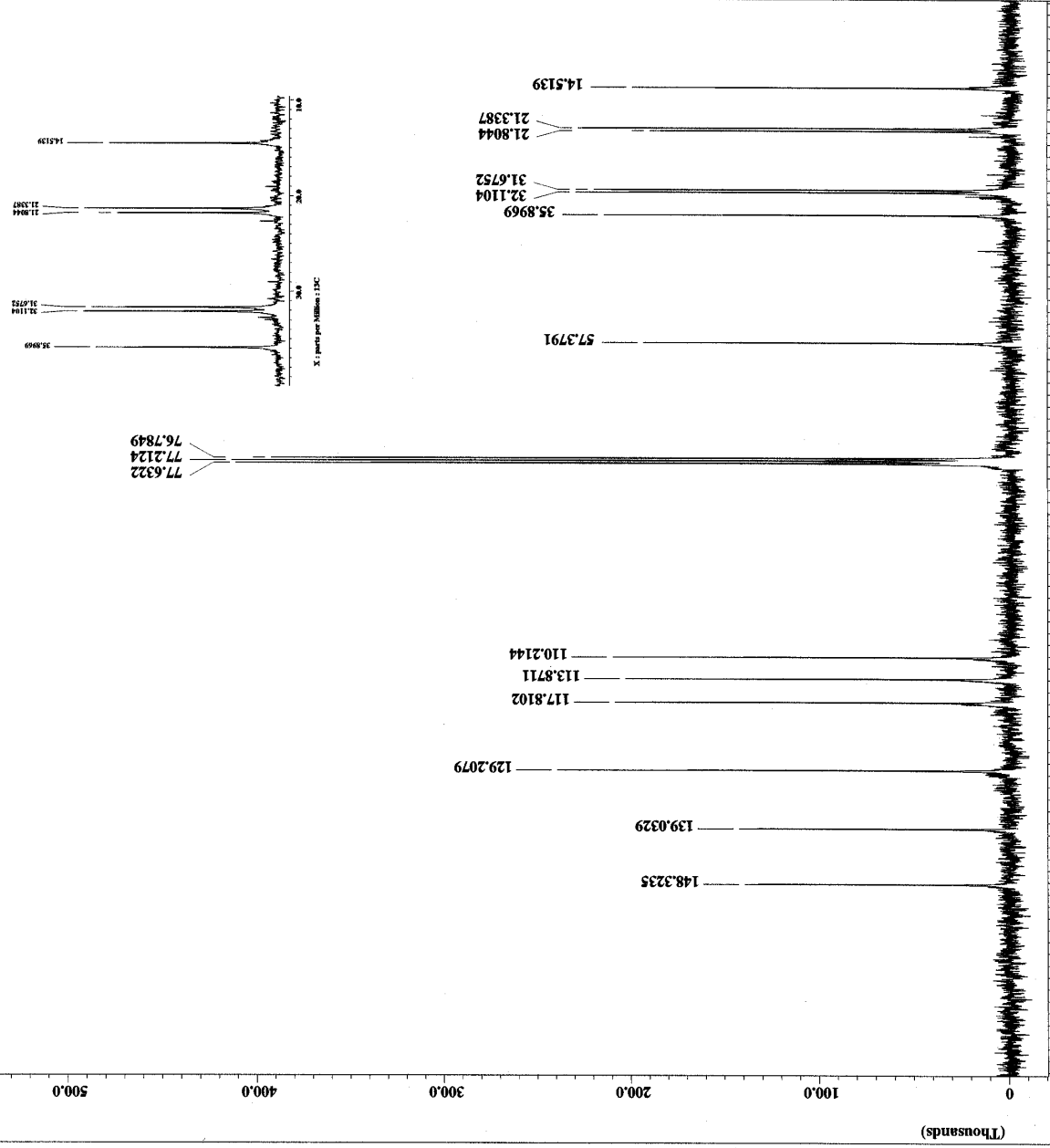
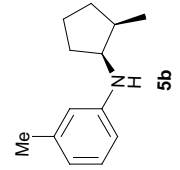
File Name      = Reacc507-2F1011-13C-5
Author        = Cabrera
Experiment    = single_pulse_dec
Sample ID     = Laura
Solvent      = CHLOROFORM-D
Creation time = 30-AUG-2008 01:55:08
Revision time = 11-NOV-2008 09:03:55
Current time  = 11-NOV-2008 09:04:48

Comment       = Single Pulse with Bro
Data format   = 1D COMPLEX
Data size     = 32768
Dim. title    = 13C
Dim. units    = [ppm]
Dimensions    = X
Site          = Eclipse+ 300
Spectrometer  = DELTA_NMR

Field strength = 7.0586013[T] (300[MHZ]
X acq duration = 1.7334272[s]
X domain       = 13C
X freq         = 75.56823426[MHZ]
X offset       = 100[ppm]
X points       = 32768
X prescans     = 4
X resolution   = 0.57689184[Hz]
X sweep       = 18.90359168[MHZ]
Irr domain    = 1H
Irr freq      = 300.52965592[MHZ]
Irr offset    = 5[ppm]
Clipped       = FALSE
Mod_return    = 1
Scans         = 2626
Total_scans   = 2626

X_90_width    = 11.3[us]
X_acq_time    = 1.7334272[s]
X_angle       = 30[deg]
X_pulse       = 3.76666667[us]
Initial_wait  = 1[s]
Phase_preset  = 3[us]
Recvr_gain    = 30
Relaxation_delay = 1[s]
Temp_get      = 20.9[dc]
Unblank_time  = 2[us]

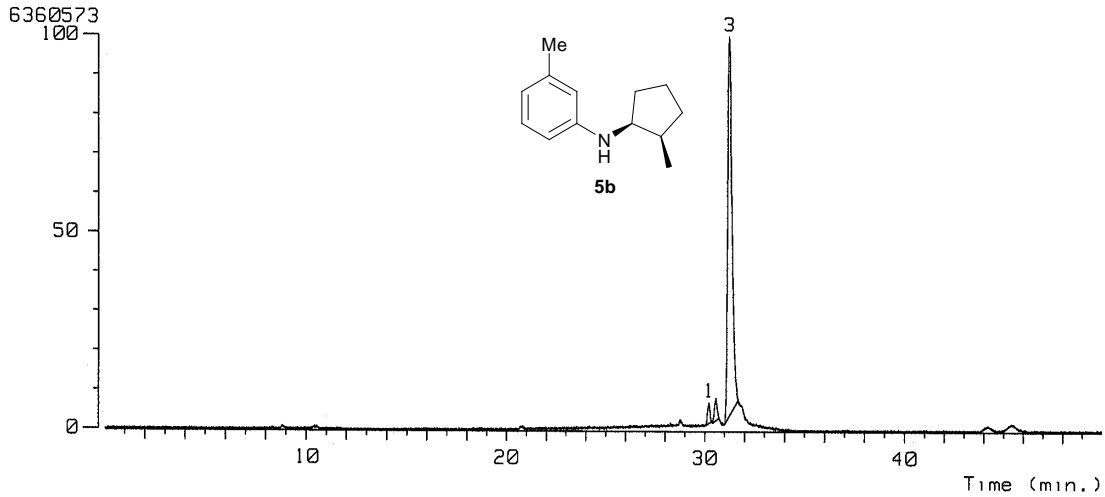
```



[TIC]

Data : Dr-Cabrera-Armando-151
Sample: 2570 reacc 507 JeolAX505HA
Note : 5 horas
Inlet : GC
Ion Species : Normal Ion

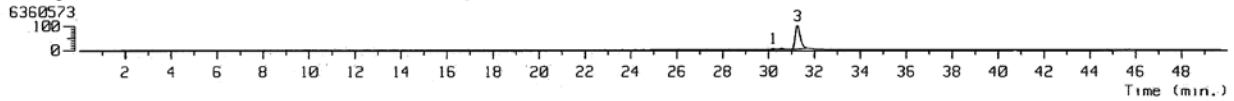
Date : 30-Oct-120 12:40
Ion Mode : EI+
TIC Range : m/z 33 to 650



No.	RT [min]	Area	Area%	Height	Height%	Width [sec]	INTEG
1	30.20	255.25	2.70	31.90	4.91	7.51	BB
2	30.56	317.29	3.36	34.27	5.28	8.69	BB
3	31.25	8876.89	93.94	583.43	89.81	14.29	BB

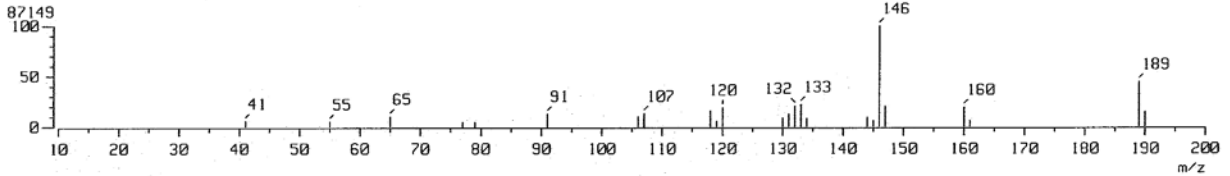
[TIC]

Data : Dr-Cabrera-Armando-151 Date : 30-Oct-120 12:40
Sample: 2570 reacc.507 JeolAX505HA
Note : 5 horas
Inlet : GC Ion Mode : EI+
Ion Species : Normal Ion [MF-Linear]
TIC Range : m/z 33 to 650 Output RT Range : 0.00 to 49.99 min



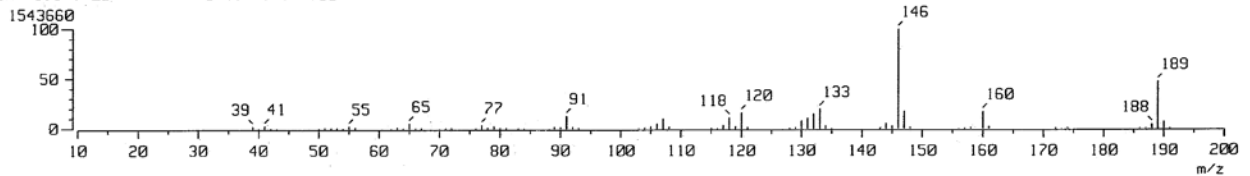
[Mass Spectrum]

RT : 30.56 min Scan#: 2631-2617-2645 Temp : 0.0 deg.C
Ion Mode : EI+ Int. : 8.31



[Mass Spectrum]

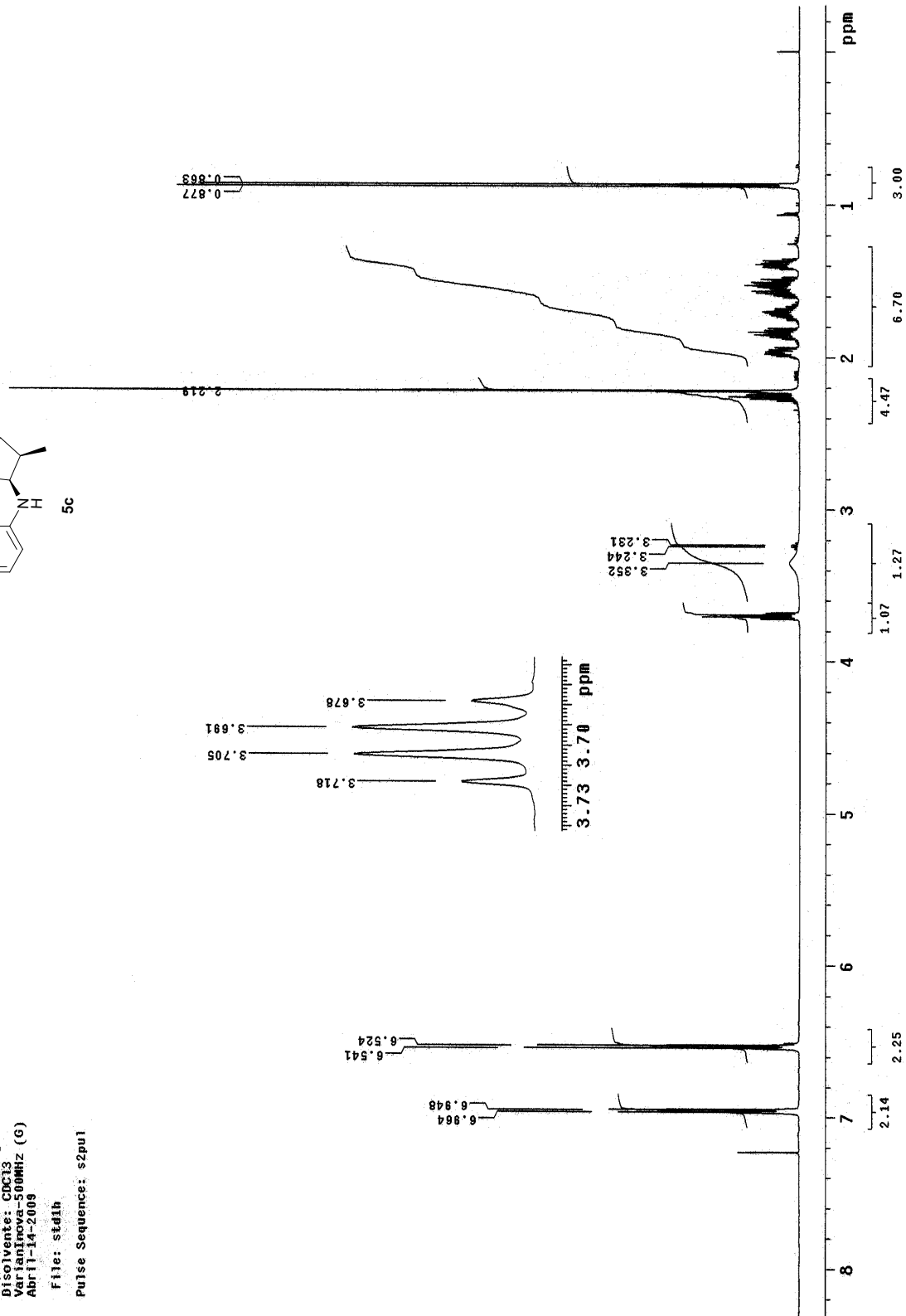
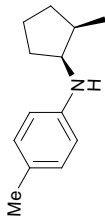
RT : 31.25 min Scan#: 2691-2670-2727 Temp : 0.0 deg.C
Ion Mode : EI+ Int. : 147.22



UNAM. Instituto de Química.
 Dr-A-Cabrera/Laura-K.P
 Clave: Reacc325129
 No. Registro: 337
 Experimento: H₂Ogeno
 Disolvente: CDCl₃
 Volumen: 0.5 ml
 Fecha: 14-04-2009

File: std1h

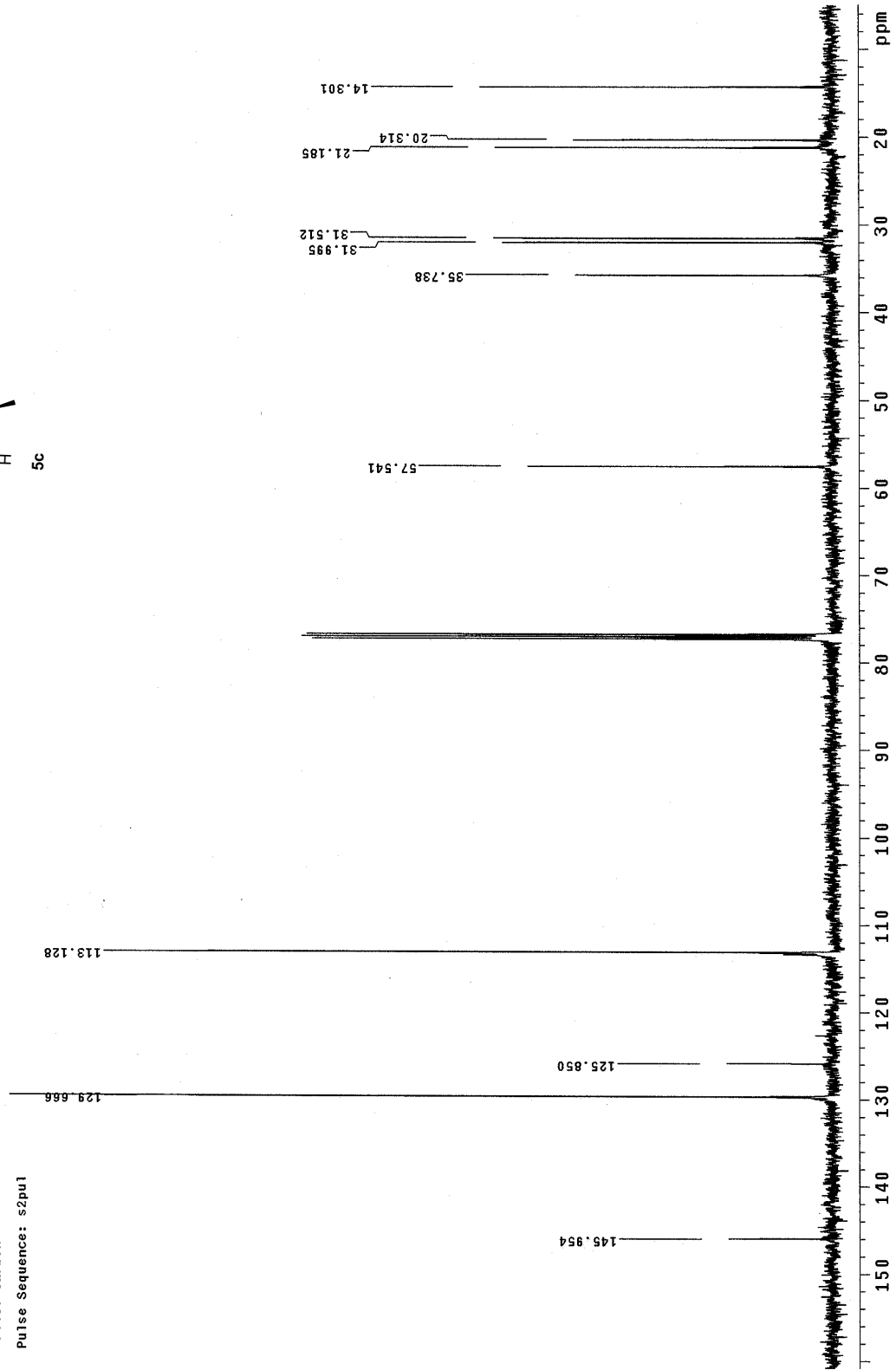
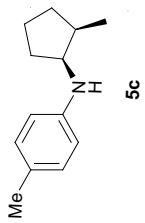
Pulse Sequence: s2pu1



UNAM - Instituto de Química
Dr-A-cabrera/laura-R.P
Clave: Reacc32579
No. registro: 33
Fecha: 10/03/12
Disolvente: CDCl3
UnityInova-125.71MHZ (G)
Abril-14-2009

File: Carbon

Pulse Sequence: s2pu1



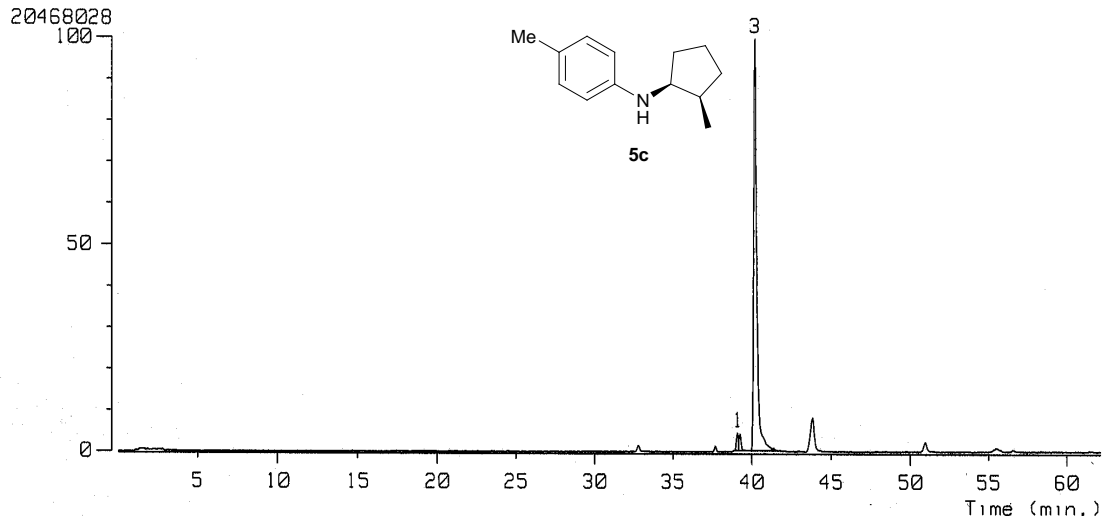
[TIC]

Data : Dr-Cabrera-Armando-882
Sample: 2566 G Reacc 325 AX505HA
Note :

Date : 06-Dec-107 12:14

Inlet : GC
Ion Species : Normal Ion

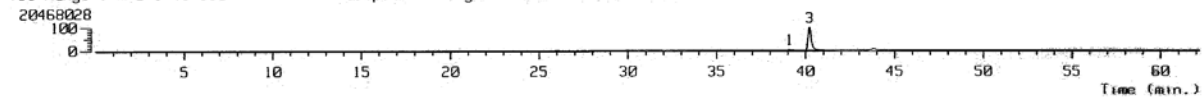
Ion Mode : EI+
TIC Range : m/z 5 to 650



No.	RT [min]	Area	Area%	Height	Height%	Width [sec]	INTEG
1	39.09	683.64	2.42	80.41	3.83	7.98	BV
2	39.25	651.28	2.30	76.03	3.62	8.04	VB
3	40.21	26923.40	95.28	1941.51	92.54	13.02	BB

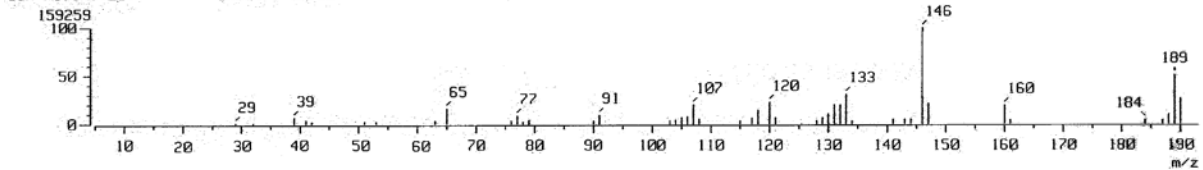
[TIC]

Data : Dr-Cabrera-Armando-882 Date : 06-Dec-107 12:14
Sample: 2566 G Reacc 325 AX505HA
Note :
Inlet : GC Ion Mode : EI+
Ion Species : Normal Ion [MF-Linear]
TIC Range : m/z 5 to 650 Output RT Range : 0.00 to 62.19 min



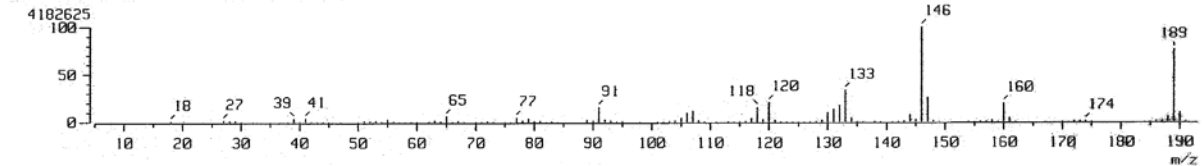
[Mass Spectrum]

RT : 39.25 min Scan# : 2941-2916-2962 Temp : 0.0 deg.C
Ion Mode : EI+ Int. : 15.19



[Mass Spectrum]

RT : 40.21 min Scan# : 3013-2993-3123 Temp : 0.0 deg.C
Ion Mode : EI+ Int. : 398.89





Reacc321-y-1H-3.jdf

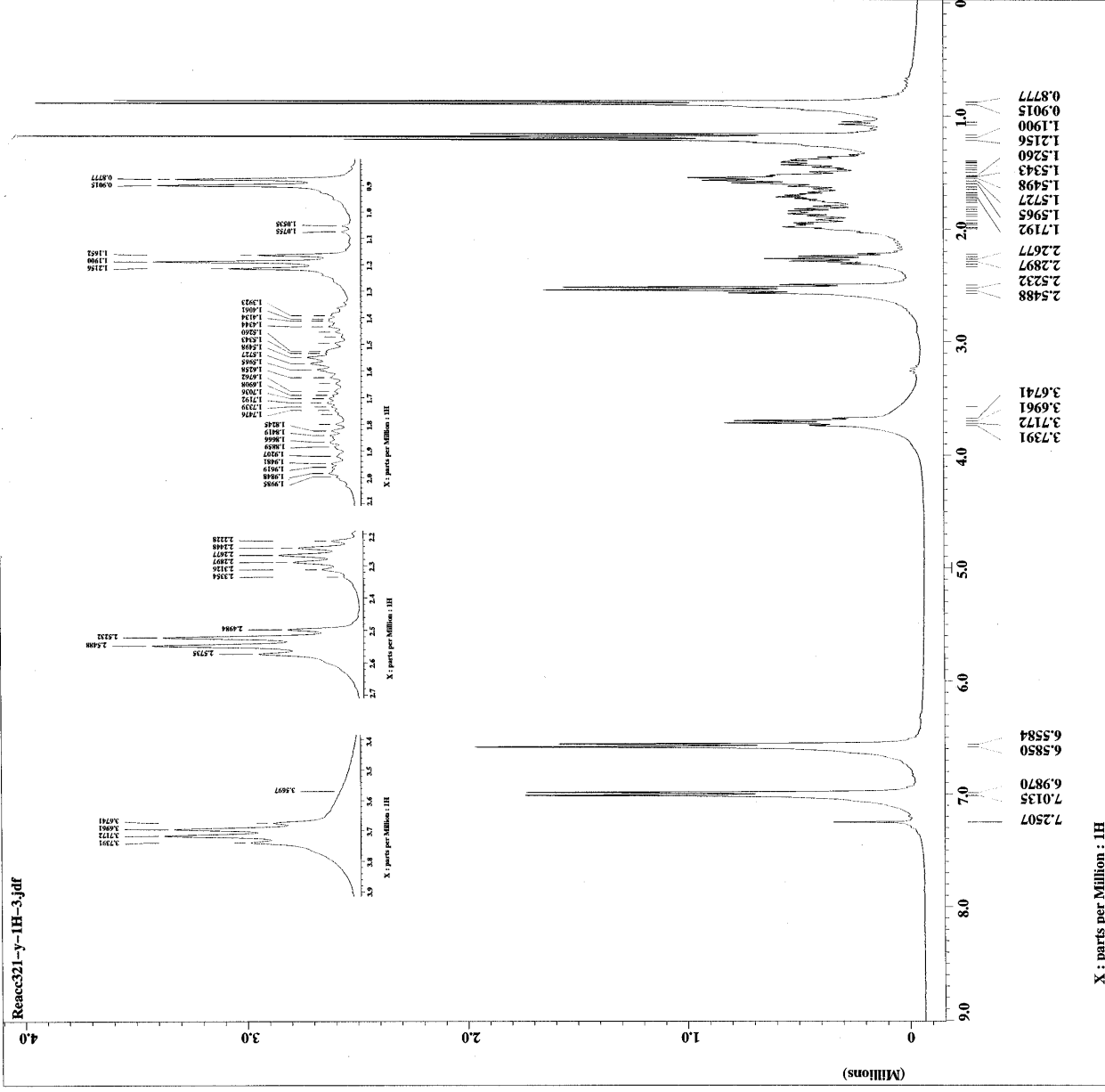
```

Filename = Reacc321-y-1H-3.jdf
Author = Cabrera
Experiment = single_pulse.exp
Sample_id = Laura
Solvent = CHLOROFORM-D
Creation_time = 4-AUG-2008 23:12:15
Revision_time = 17-OCT-2008 07:42:20
Current_time = 17-OCT-2008 07:47:01

Comment = Single Pulse Experiment
Data_format = 1D COMPLEX
Dir_size = 16384
Dir_title = 1H
Dir_units = [ppm]
Dimensions = X
Site = Eclipse+ 300
Spectrometer = DELTA_NMR

Field_strength = 7.0586013 [T] (300 [MHz]
X_acq_duration = 3.6339712 [s]
X_gain = 1H
X_freq = 300.52965592 [MHz]
X_offset = 5 [ppm]
X_points = 16384
X_prescans = 0
X_resolution = 0.27518105 [Hz]
X_sweep = 4.50856628 [kHz]
Clipped = FALSE
Mod_return = 1
Scans = 32
Total_scans = 32

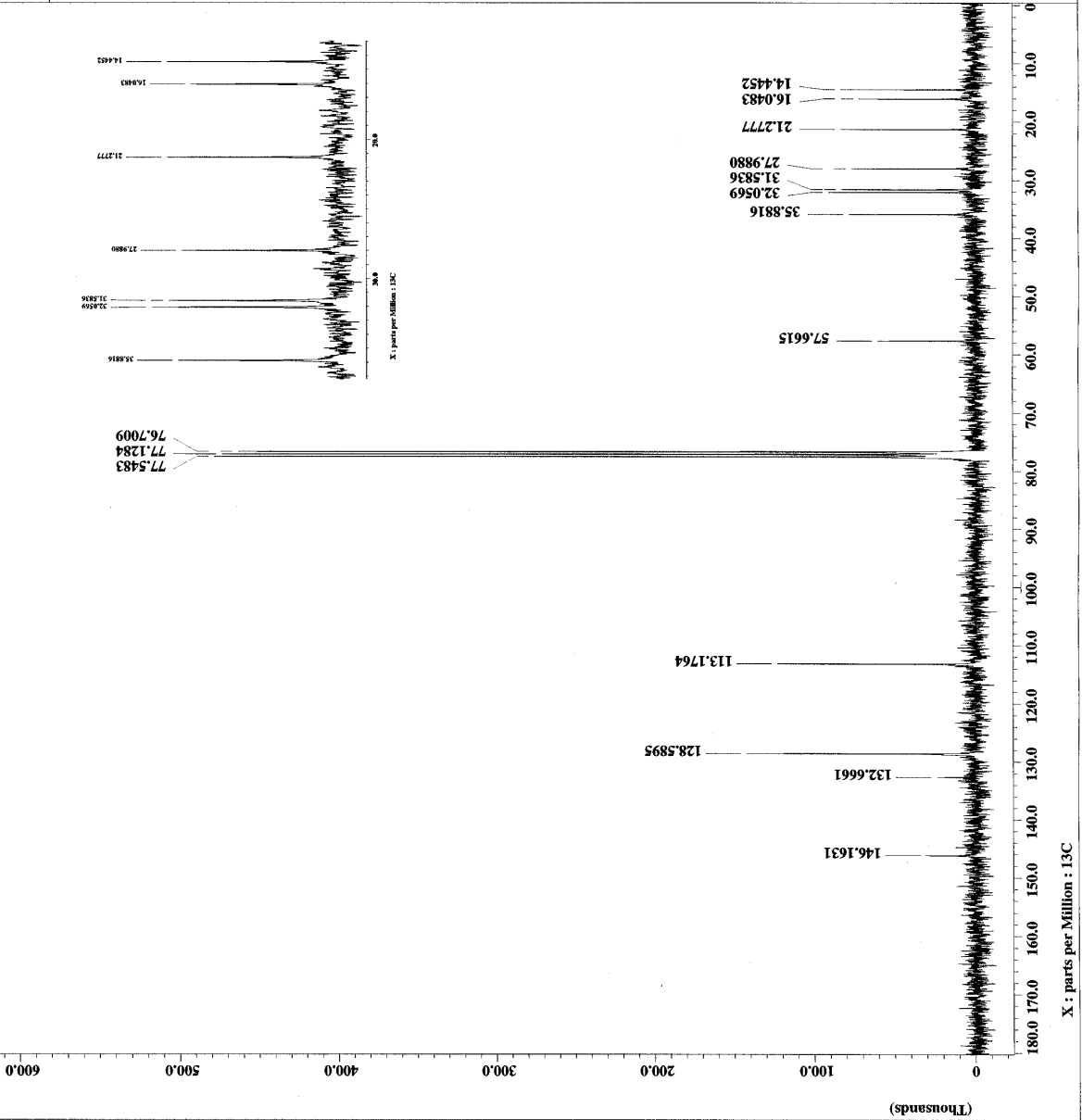
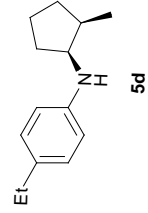
X_90_width = 9.5 [us]
X_acq_time = 3.6339712 [s]
X_angle = 45 [deg]
X_pulse = 4.75 [us]
Initial_wait = 1 [s]
Phase_preset = 3 [us]
Recvr_gain = 19
Relaxation_delay = 1 [s]
Temp_get = 19.3 [dC]
Tmbank_time = 2 [us]
  
```



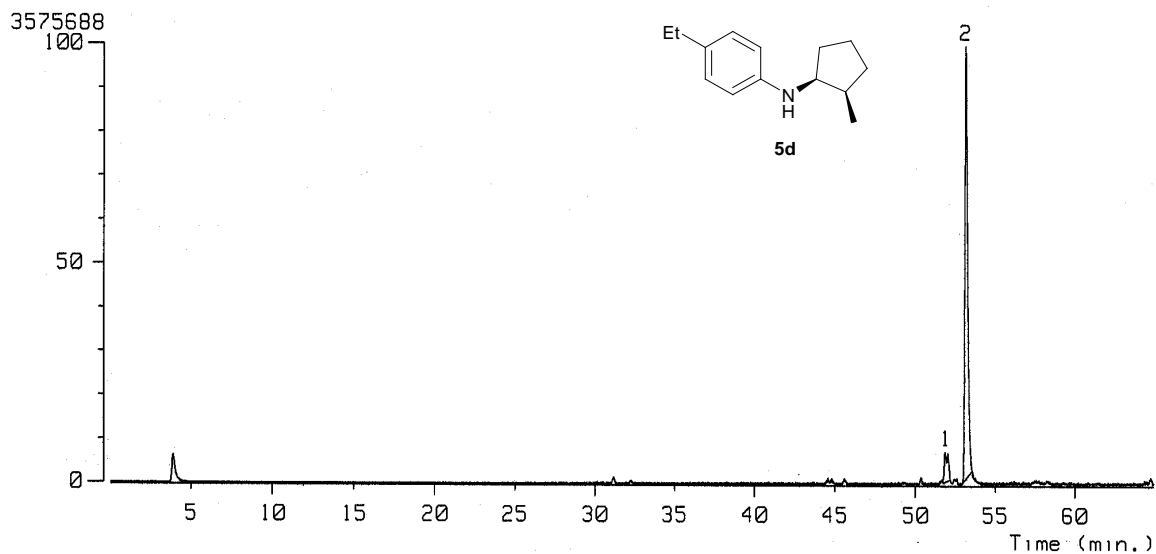


Reacc321-y-13C-5.jdf

Filename = Reacc321-y-13C-5.jdf
Author = Cabrera
Experiment = single_pulse_dec
Sample_id = Laura
Solvent = CHLOROFORM-D
Creation_time = 5-AUG-2008 00:43:29
Revision_time = 17-OCT-2008 08:58:40
Current_time = 17-OCT-2008 08:59:38
Comment = Single Pulse with BEO
Data format = 1D_COMPLEX
Data size = 32768
Dir title = 13C
Dim units = [ppm]
Dimensions = X
Site = Eclipse+ 300
Spectrometer = DELTA_NMR
Field_strength = 7.0586013 [T] (300 [MHz])
X_acq_duration = 1.7334272 [s]
X_domain = 13C
X_freq = 75.56823426 [MHz]
X_offset = 100 [ppm]
X_points = 32768
X_prescans = 4
X_resolution = 0.57689184 [Hz]
X_sweep = 18.90339188 [MHz]
Xrr_domain = 30.52965592 [MHz]
Xrr_freq = 5 [ppm]
Xrr_offset = FALSE
Clipped = 1
Mod return = 1976
Total_scans = 1976
X_90_width = 11.3 [us]
X_acq_time = 1.7334272 [s]
X_angle = 30 [deg]
X_pulse = 3.76666667 [us]
Initial_wait = 1 [s]
Phase_preset = 3 [us]
Recvr_gain = 30
Relaxation_delay = 1 [s]
Temp_get = 21 [dC]
Unblank_time = 2 [us]

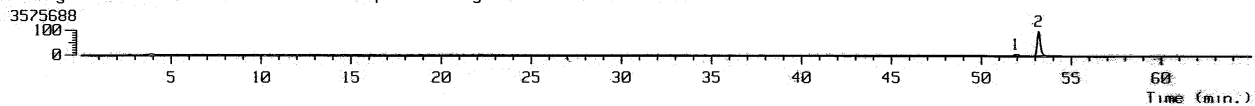


[TIC]
 Data : Dr-Cabrera-Armando-137 Date : 17-Oct-120 10:08
 Sample: 2435 G2 Reac 321 y JeolAX505HA
 Note : 5 horas
 Inlet : GC Ion Mode : EI+
 Ion Species : Normal Ion TIC Range : m/z 33 to 650

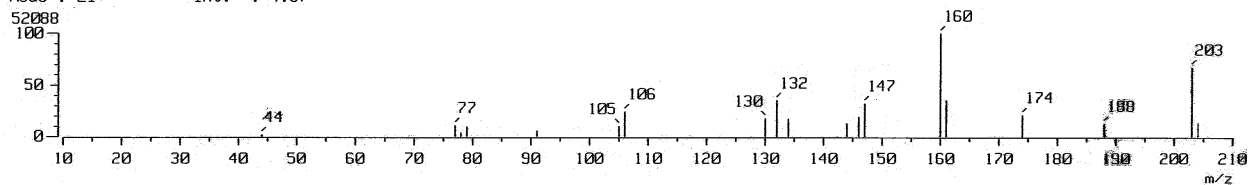


No.	RT [min]	Area	Area%	Height	Height%	Width [sec]	INTEG
1	51.86	376.07	8.32	22.97	6.39	15.37	BB
2	53.16	4143.46	91.68	336.27	93.61	11.57	BB

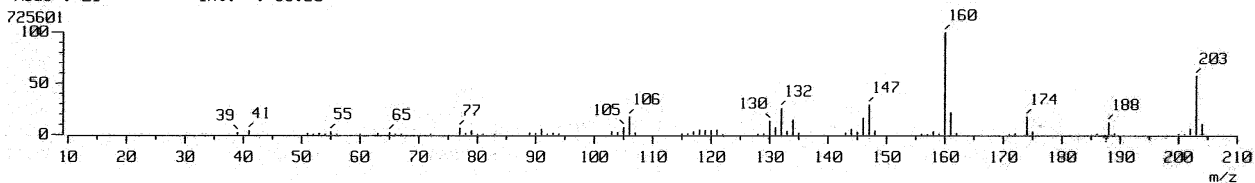
[TIC]
 Data : Dr-Cabrera-Armando-137 Date : 17-Oct-120 10:08
 Sample: 2435 G2 Reac 321 y JeolAX505HA
 Note : 5 horas
 Inlet : GC Ion Mode : EI+
 Ion Species : Normal Ion [MF-Linear]
 TIC Range : m/z 33 to 650 Output RT Range : 0.00 to 65.00 min



[Mass Spectrum]
 RT : 51.86 min Scan# : 4465-4453-4491 Temp : 0.0 deg.C
 Ion Mode : EI+ Int. : 4.97

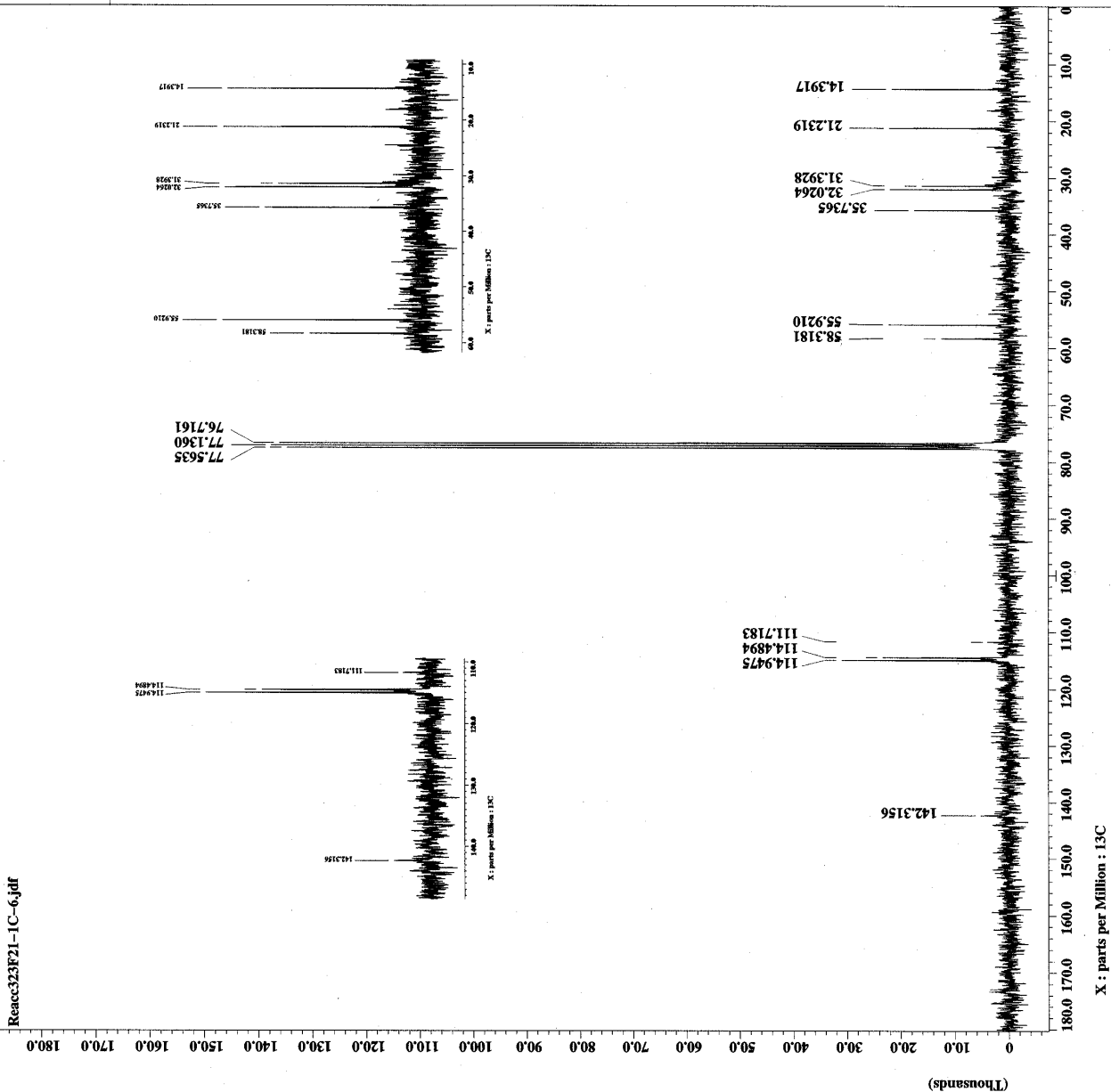
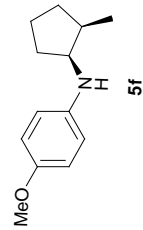


[Mass Spectrum]
 RT : 53.16 min Scan# : 4577-4557-4609 Temp : 0.0 deg.C
 Ion Mode : EI+ Int. : 69.20





Reacc323F21-1C-6.jdf
= Cabrera
= single_pulse_dec
= Laura
= CHLOROFORM-D
= 12-DEC-2007 14:18:40
= 11-DEC-2008 12:55:12
= 11-DEC-2008 12:57:26
= Single Pulse with Bro
= ID COMPLEX
= 32768
= 13C
= [ppm]
= X
= 300
= DELTA_MMR
Spectrometer
Field strength = 7.0586013 [T] (300 [MHZ])
X acq_duration = 1.7334272 [s]
X domain = 13C
X freq = 75.56823426 [MHZ]
X offset = 100 [ppm]
X points = 32768
X prescans = 4
X resolution = 0.57689184 [MHZ]
X sweep = 18.90359168 [MHZ]
Irr domain = IH
Irr freq = 300.52965592 [MHZ]
Irr offset = 5 [ppm]
Clipped = TRDE
Mod_return = 1
Total_scans = 4000
X_90_width = 11.3 [us]
X_acq_time = 1.7334272 [s]
X_angle = 30 [deg]
X_pulse = 3.76666667 [us]
X_pulse_wait = 1 [s]
X_release_pulse = 3 [us]
X_relaxation = 30 [s]
Relaxation_delay = 1 [s]
Temp_set = 17.7 [dC]
Umbank_time = 2 [us]



[TIC]

Data : Dr-Cabrera-Armando-878
Sample: 2389 G Reacc 323 AX505HA

Date : 28-Nov-107 11:49

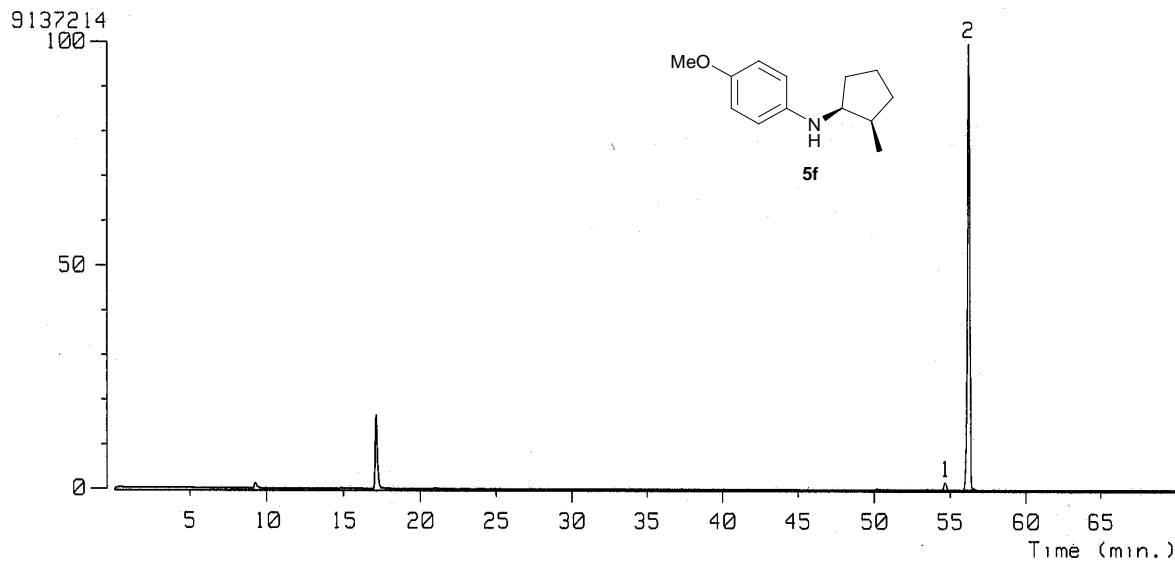
Note :

Inlet : GC

Ion Mode : EI+

Ion Species : Normal Ion

TIC Range : m/z 5 to 650



No.	RT[min]	Area	Area%	Height	Height%	Width[sec]	INTEG
1	54.66	181.96	2.02	14.44	1.63	11.83	BB
2	56.28	8829.38	97.98	871.36	98.37	9.51	BB

[TIC]

Data : Dr-Cabrera-Armando-878

Date : 28-Nov-107 11:49

Sample: 2389 G Reacc 323 AX505HA

Note :

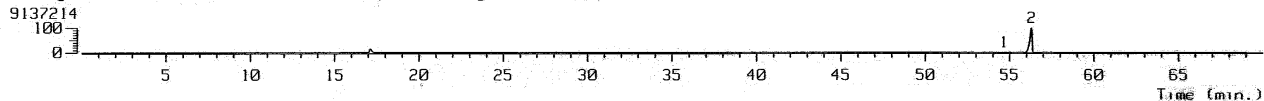
Inlet : GC

Ion Mode : EI+

Ion Species : Normal Ion [MF-Linear]

TIC Range : m/z 5 to 650

Output RT Range : 0.00 to 69.99 min



[Mass Spectrum]

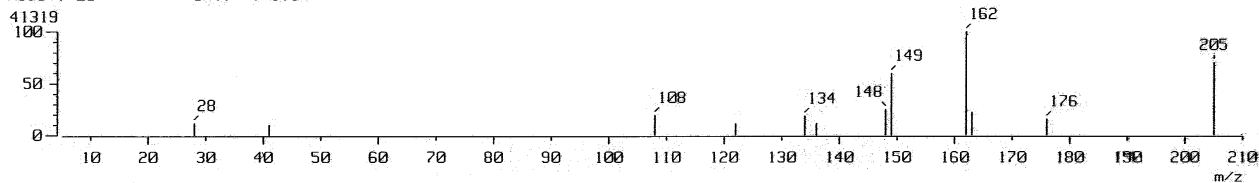
RT : 54.66 min

Scan# : 4100-4085-4121

Temp : 0.0 deg.C

Ion Mode : EI+

Int. : 3.94



[Mass Spectrum]

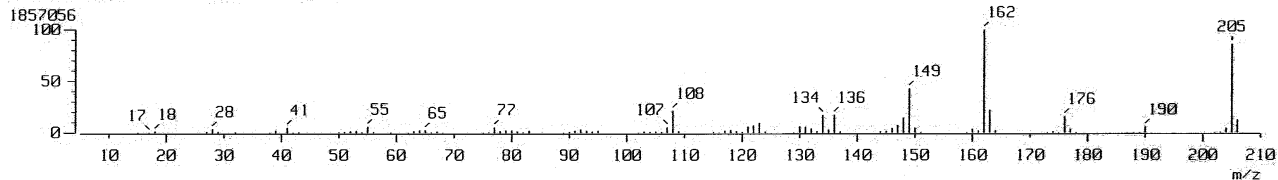
RT : 56.28 min

Scan# : 4222-4188-4266

Temp : 0.0 deg.C

Ion Mode : EI+

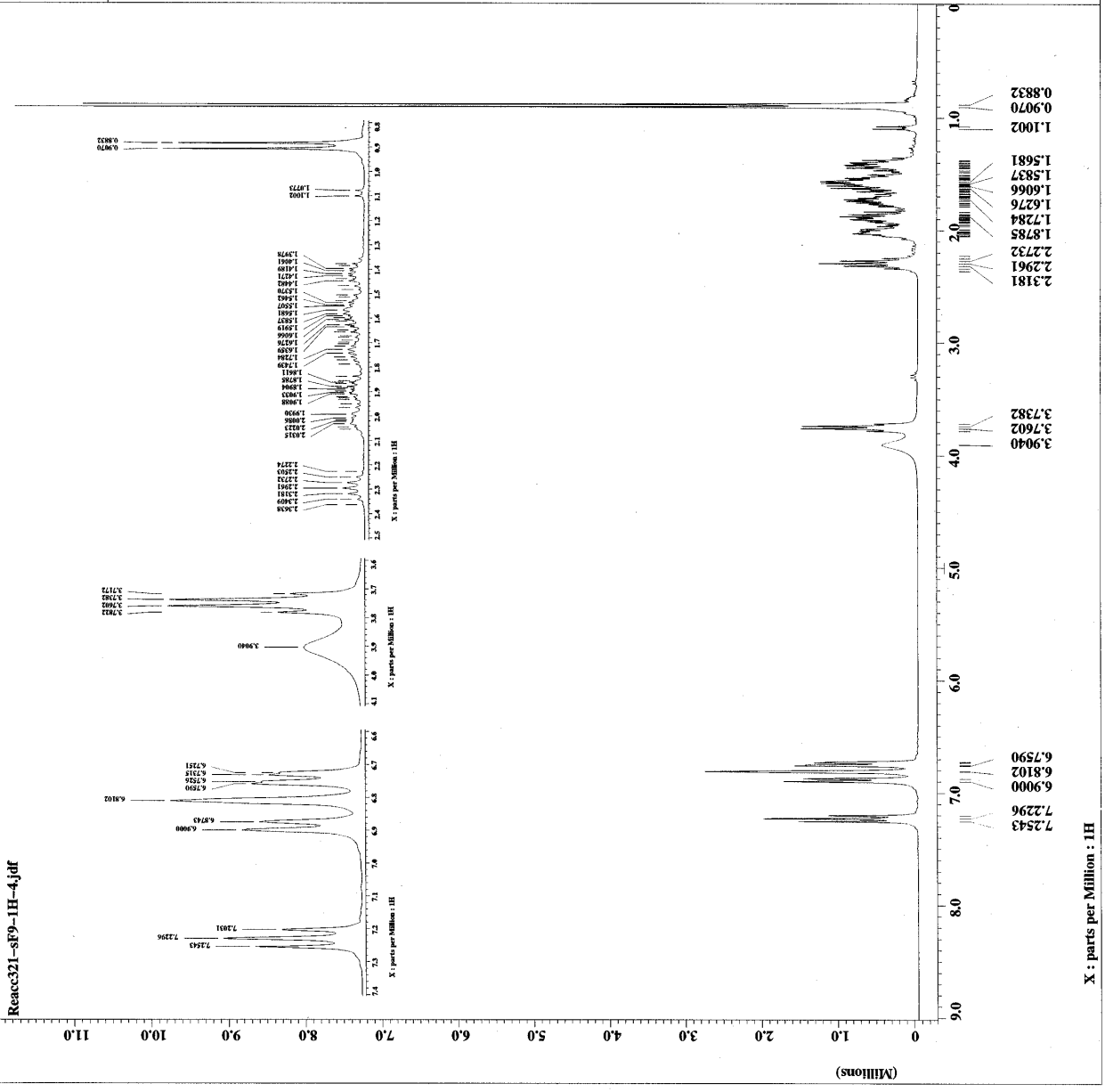
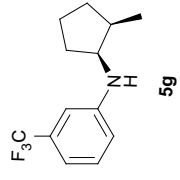
Int. : 177.10





```

Reacc321-sf9-1H-4.jdf
= Cabrea
= single_pulse.exp
= Laura
= CHLOROFORM-D
= 8-AUG-2008 08:02:11
= 20-OCT-2008 16:16:52
= 20-OCT-2008 16:17:56
= Single Pulse Experiment
= 1D COMPLEX
= 16384
= 1H
= [ppm]
= X
= Eclipse+ 300
= DELTA_NMR
Field_strength = 7.0586013[T] (300[MHz]
X_acq_duration = 3.6339712[s]
X_domain = 1H
X_freq = 300.52965592[MHz]
X_offset = 5[ppm]
X_points = 16384
X_prescans = 0
X_resolution = 0.27518105[Hz]
X_sweep = 4.50856628[MHz]
Clipped = FALSE
Mod_return = 1
Scans = 32
Total_scans = 32
X_90_width = 9.5[us]
X_acq_time = 3.6339712[s]
X_angle = 45[deg]
X_pulse = 4.75[us]
X_pulse_wait = 3[us]
X_pulse_offset = 3[us]
X_pulse_phase = 171
Relaxation_delay = 1[s]
Temp_get = 19.5[dc]
Dabblank_time = 2[us]
  
```

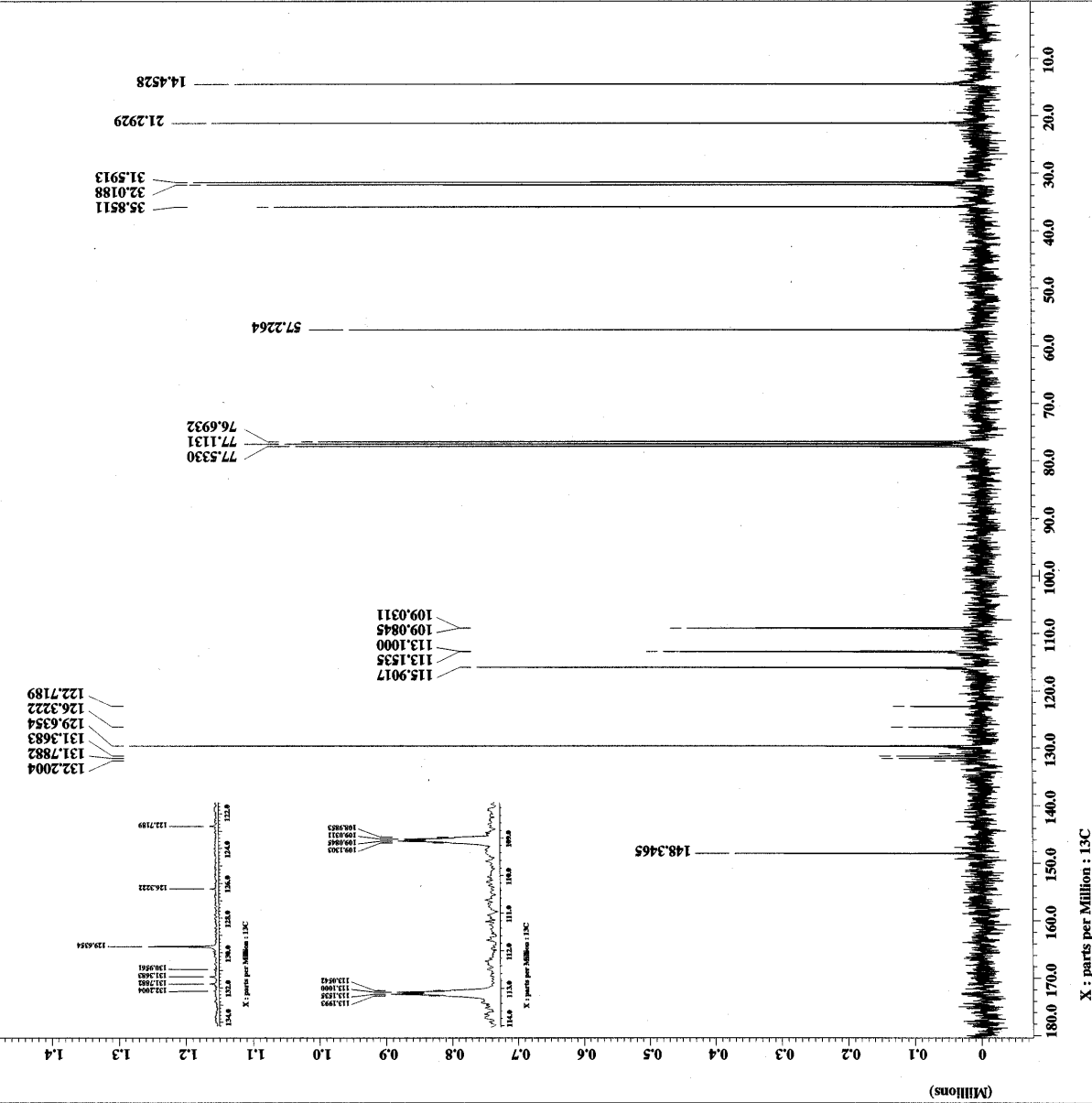


Reacc321-sf9-1H-4.jdf

X: parts per Million : 1H

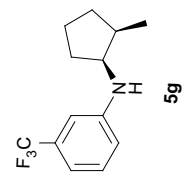


Reacc321-sf9-13C-3.jdf



```

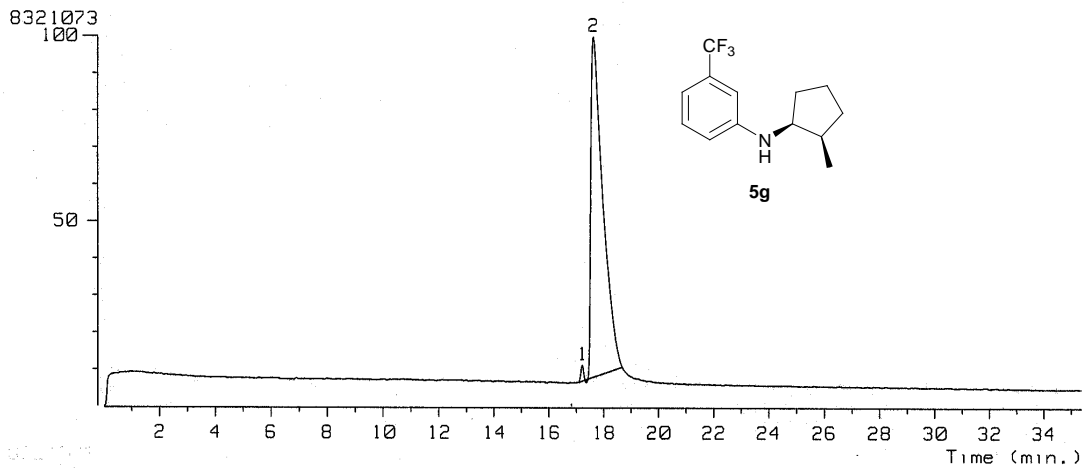
Filename = Reacc321-sf9-13C-3.jdf
Author = Cabrera
Experiment = single_pulse_dec
Sample_id = Laura
Solvent = CHLOROFORM-D
Creation_time = 8-AUG-2008 08:13:29
Revision_time = 20-OCT-2008 16:25:04
Current_time = 20-OCT-2008 16:25:52
Comment = Single Pulse with Bro
Data_format = AD_COMPLEX
Dir_size = 32768
Dir_title = 13C
Dir_units = [ppm]
Dir_extensions = Eclipse+ 300
Spectrometer = DELTA_NMR
Field_strength = 7.0586013[T] (300[MHZ]
X_acq_duration = 1.7334272[s]
X_domain = 13C
X_freq = 75.56823426[MHZ]
X_offset = 100[Dppm]
X_points = 32768
X_prescans = 4
X_resolution = 0.57689184[MHZ]
X_sweep = 18.90359168[MHZ]
Irr_domain = IR
Irr_freq = 300.52965592[MHZ]
Irr_offset = 5[Dppm]
Clipped = FALSE
Mod_return = 201
Total_scans = 221
X_90_width = 11.3[us]
X_acq_time = 1.7334272[s]
X_ang1 = 30[deg]
X_pulse = 3.76666667[us]
Initial_wait = 1[s]
Phase_preset = 3[us]
Recvr_gain = 30
Relaxation_delay = 1[s]
Temp_get = 21.3[degC]
Unblank_time = 2[us]
  
```



[TIC]

Data : Dr-Cabrera-Armando-505
Sample: 857 G Rx-321-s AX505HA
Note :
Inlet : GC
Ion Species : Normal Ion

Date : 26-Mar-120 10:16
Ion Mode : EI+
TIC Range : m/z 5 to 650



No.	RT [min]	Area	Area%	Height	Height%	Width [sec]	INTEG
1	17.22	211.29	0.95	33.69	4.41	5.89	BV
2	17.64	22132.42	99.05	729.55	95.59	28.49	VB

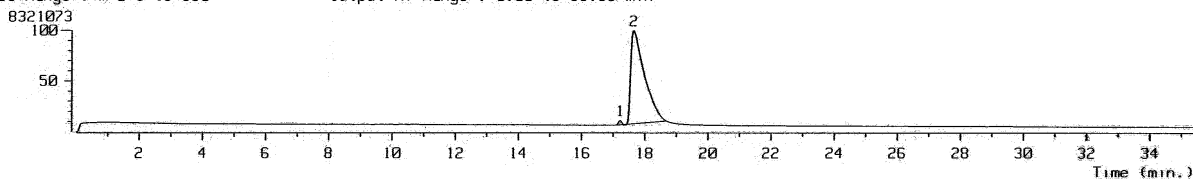
[TIC]

Data : Dr-Cabrera-Armando-505
Sample: 857 G Rx-321-s AX505HA
Note :
Inlet : GC
Ion Species : Normal Ion [MF-Linear]
TIC Range : m/z 5 to 650

Date : 26-Mar-120 10:16

Ion Mode : EI+

Output RT Range : 0.00 to 35.38 min

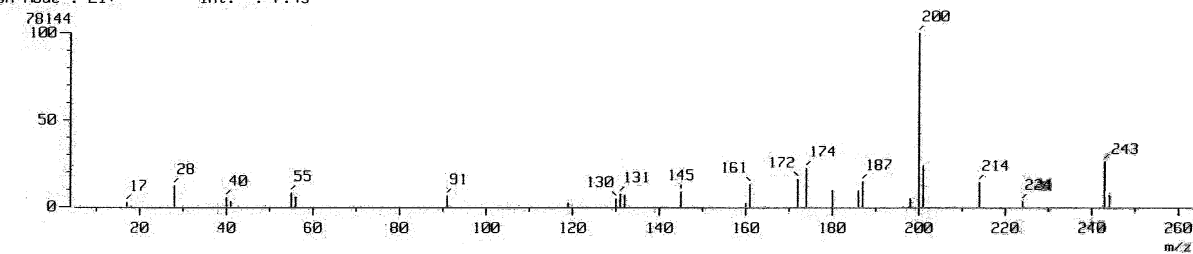


[Mass Spectrum]

RT : 17.22 min
Ion Mode : EI+

Scan# : 1292-1283-1402
Int. : 7.45

Temp : 0.0 deg.C

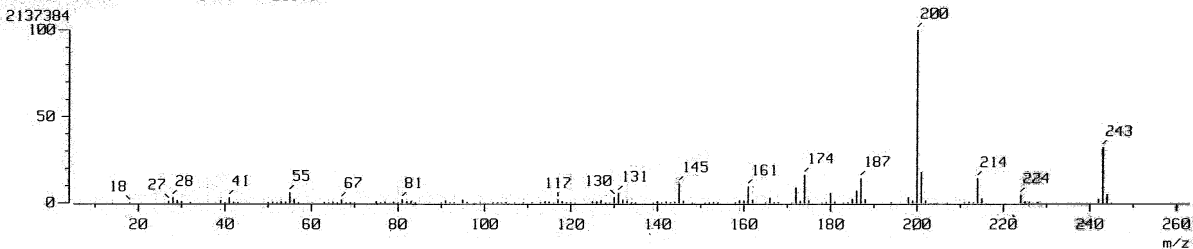


[Mass Spectrum]

RT : 17.64 min
Ion Mode : EI+

Scan# : 1324-1283-1402
Int. : 203.84

Temp : 0.0 deg.C



INSTITUTO DE QUIMICA, UNAM/EHS

Dr. A. Cabrera/Laura P. P.

Clave. Reacc514

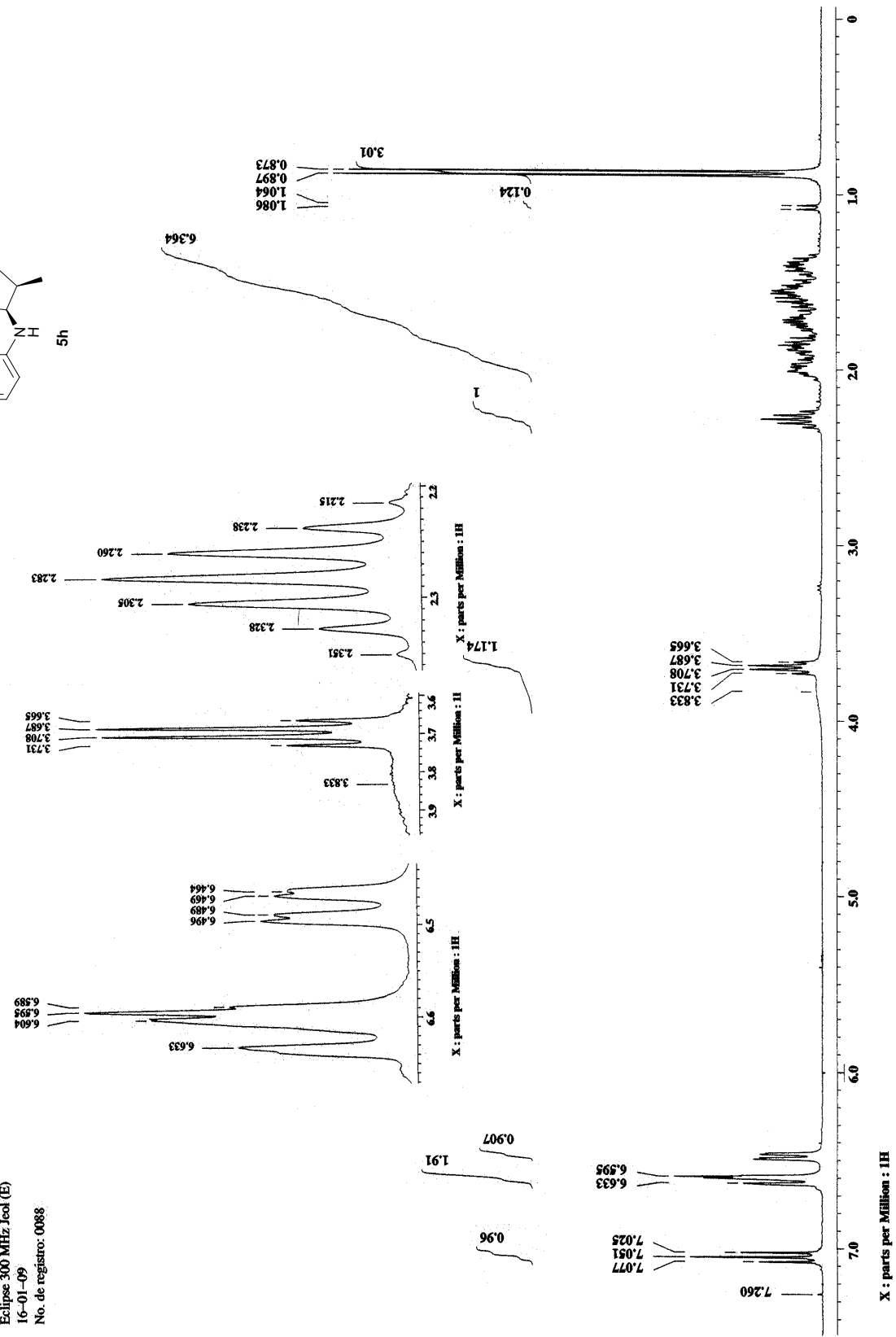
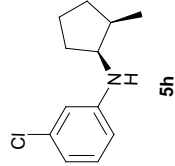
Disolvente: CDCl₃

Hidrogeno-1

Eclipse 300 MHz Teol (E)

16-01-09

No. de registro: 0088



INSTITUTO DE QUIMICA, UNAM/EHS

Dr. A. Cabrera/Laura R. P.

Clave: Reacc514-2

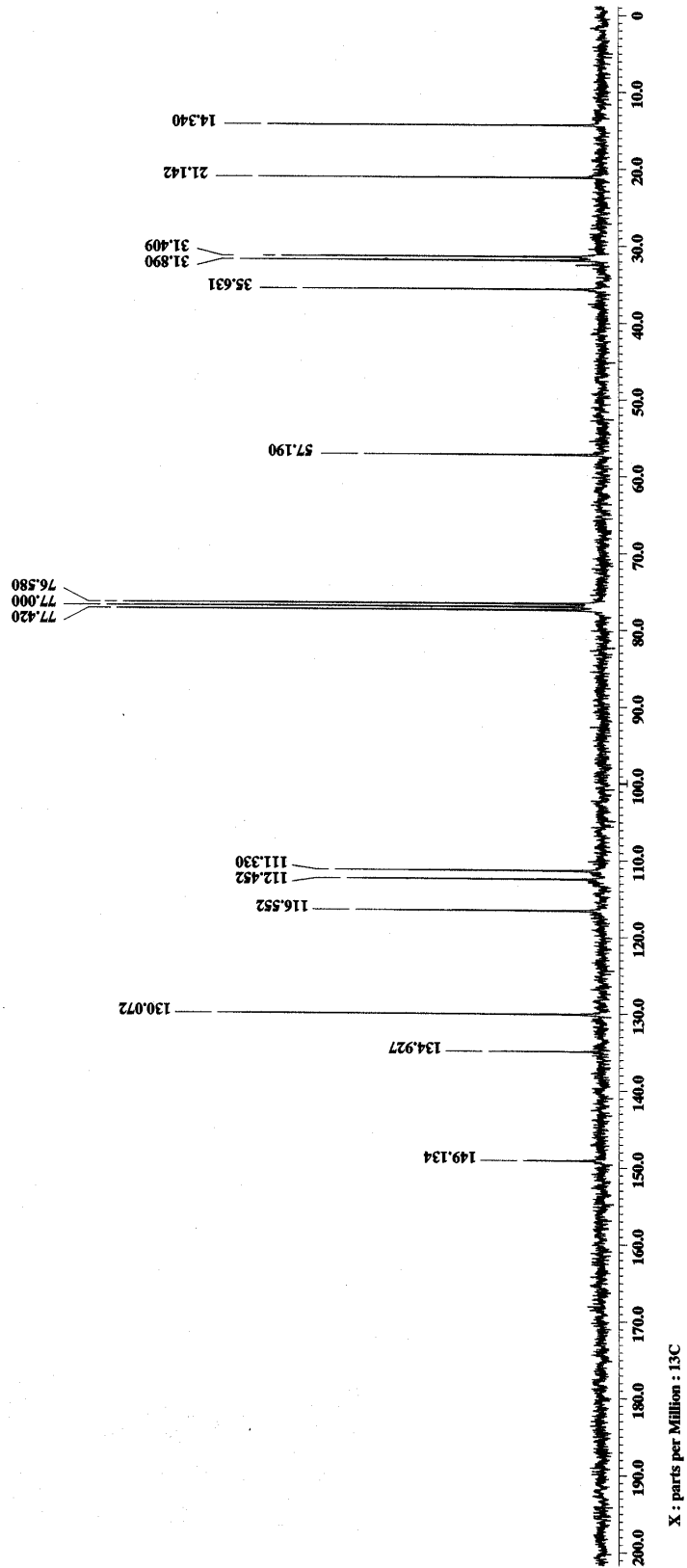
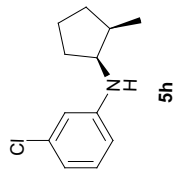
Disolvente: CDCl₃

Carbono-13

Eclipse 300 MHz Jeol (E)

26-01-09

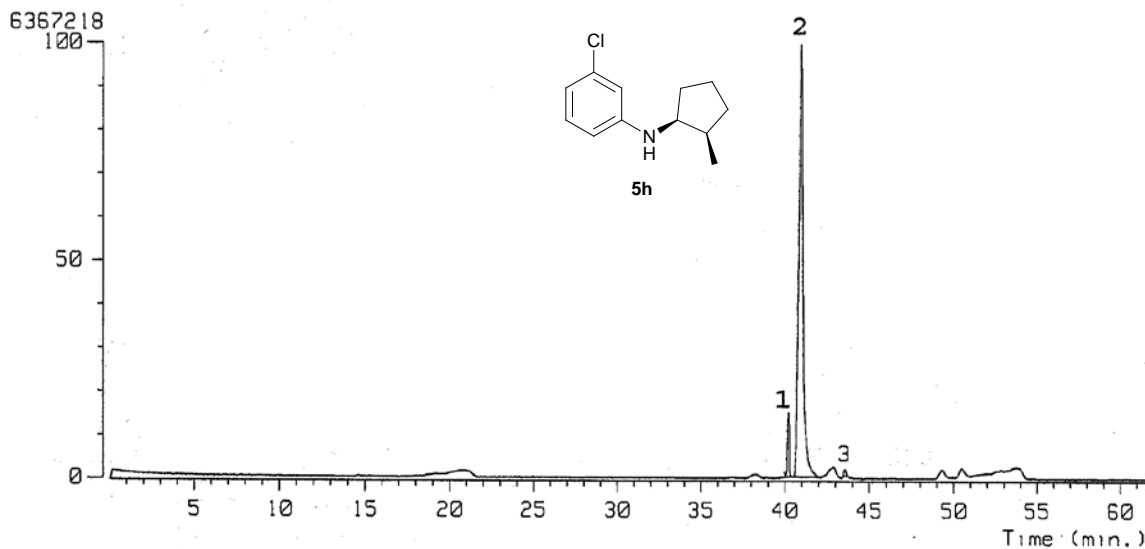
No. de registro: 0261



[TIC]

Data : Dr-Cabrera-Armando-922
Sample: 57 G reacc 514 AX505HA
Note :
Inlet : GC
Ion Species : Normal Ion

Date : 14-Jan-120 12:05
Ion Mode : EI+
TIC Range : m/z 5 to 650



No.	RT [min]	Area	Area%	Height	Height%	Width [sec]	INTEG
1	40.17	474.94	4.23	55.11	8.27	16.16	VB
2	41.00	10678.02	95.09	602.60	90.43	16.64	BB
3	43.55	6.68	0.68	8.67	1.30	8.30	BB

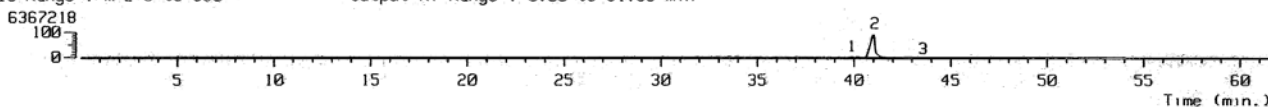
[TIC]

Data : Dr-Cabrera-Armando-922
Sample: 57 G reacc 514 AX505HA
Note :
Inlet : GC
Ion Species : Normal Ion [MF-Linear]
TIC Range : m/z 5 to 650

Date : 14-Jan-120 12:05

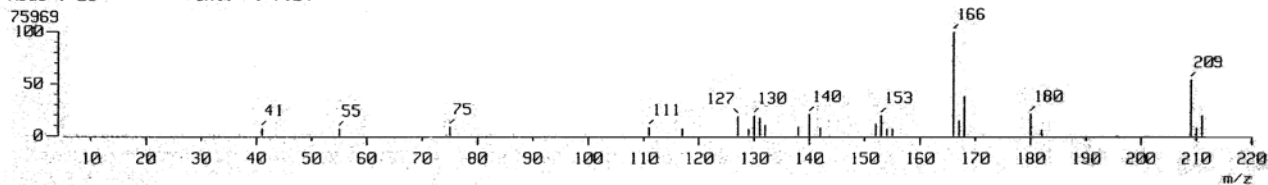
Ion Mode : EI+

Output RT Range : 0.00 to 61.56 min



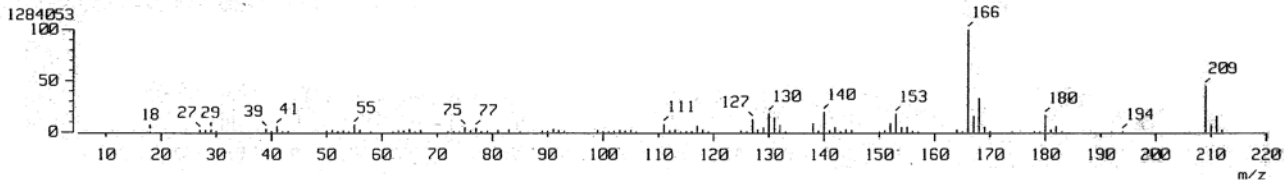
[Mass Spectrum]

RT : 40.17 min
Scan# : 3010-2977-3024
Ion Mode : EI+
Int. : 7.24
Temp : 0.0 deg.C

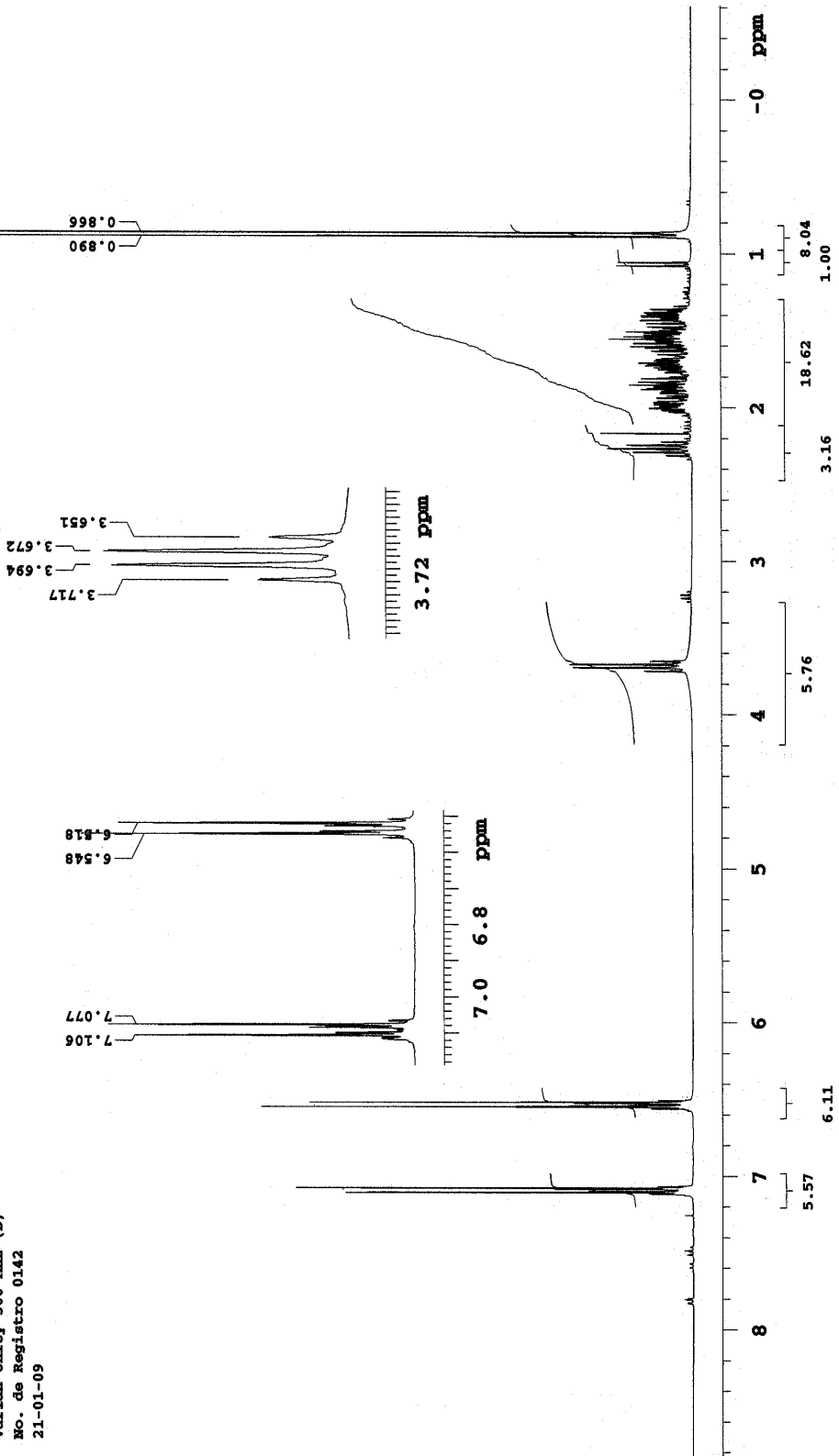
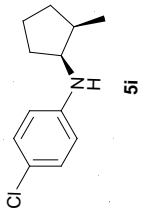


[Mass Spectrum]

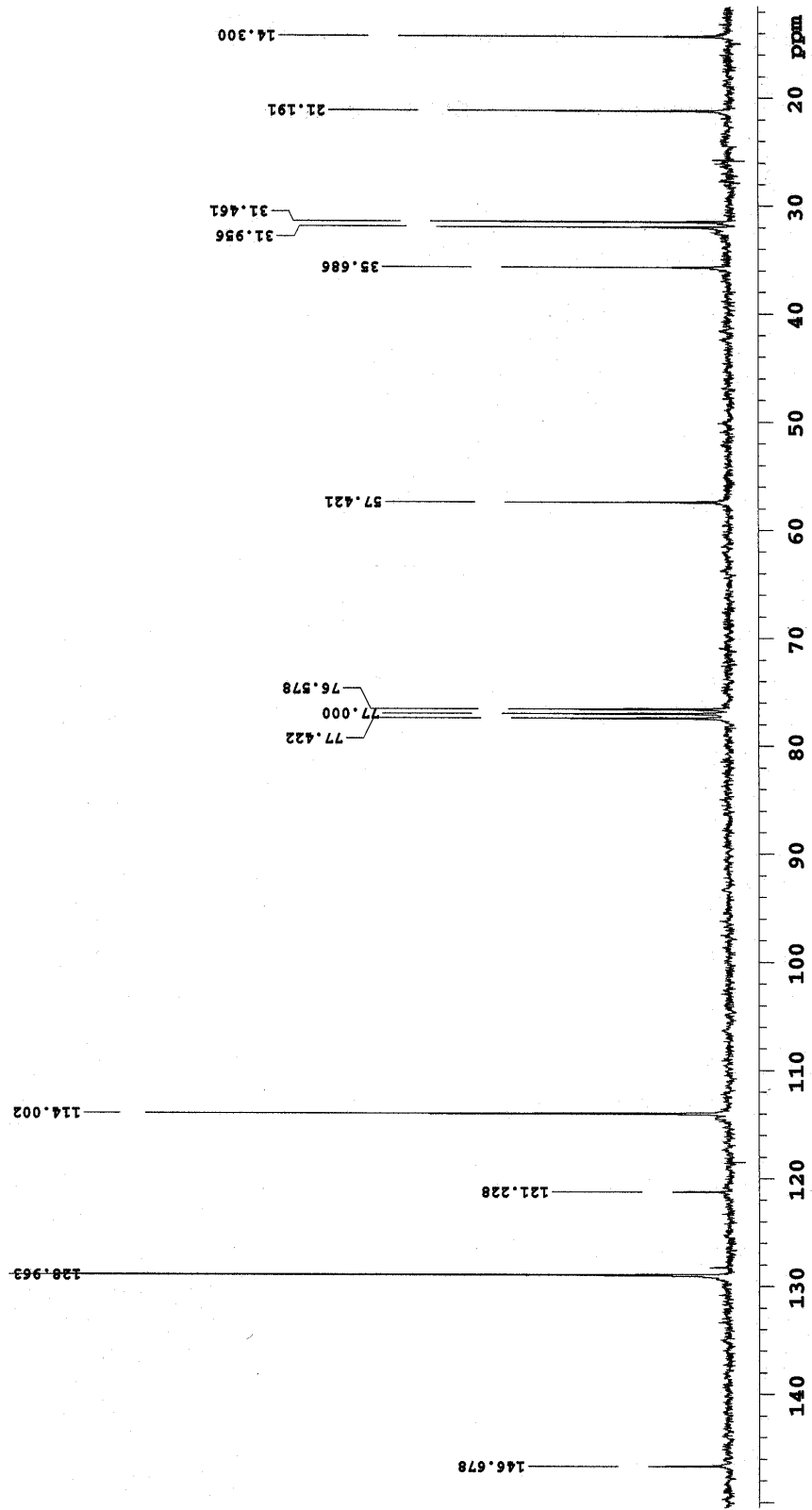
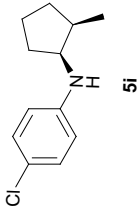
RT : 41.00 min
Scan# : 3072-3035-3154
Ion Mode : EI+
Int. : 122.46
Temp : 0.0 deg.C



U.N.A.M. Instituto de Química ICH
 Dr. A. Cabrera/Laura R. P
 Clave: Reacc512-(S)BIN
 Disolvente: CDCl3
 Experimento 1E
 Varian Unity 300 MHz (D)
 No. de Registro 0142
 21-01-09



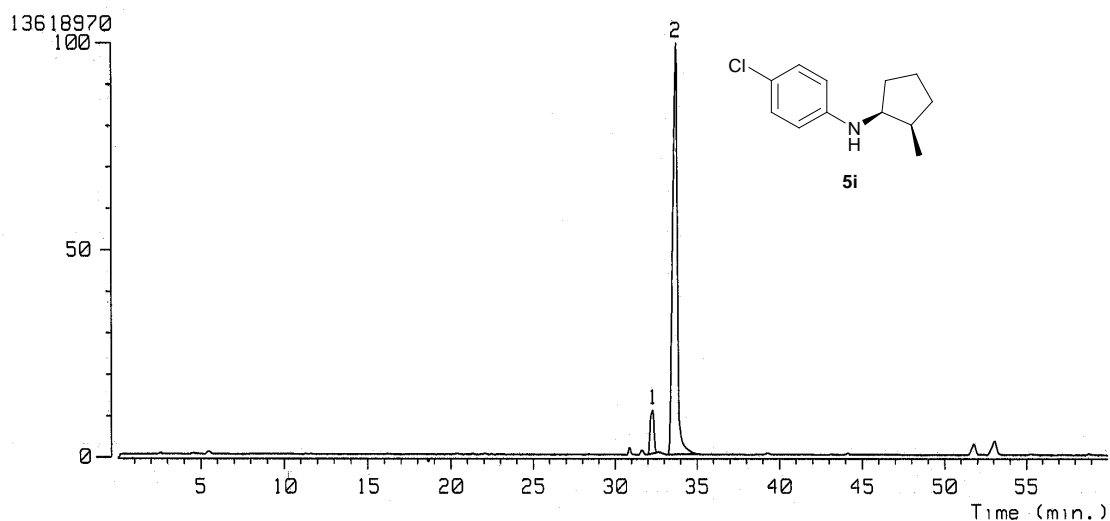
U.N.A.M. Instituto de Química ICH
Dr. A. Cabrera/Laura R. P
Clave: Reacc512-(S)BIN
Disolvente: CDCl₃
Experimento 13C
Varian Unity 75 MHz (D)
No. de Registro 0142
21-01-09



[TIC]

Data : Dr-Cabrera-Armando-918
Sample: 32 G Reacc 512 AX505HA
Note :
Inlet : GC
Ion Species : Normal Ion

Date : 12-Jan-108 13:12
Ion Mode : EI+
TIC Range : m/z 5 to 650



No.	RT [min]	Area	Area%	Height	Height%	Width [sec]	INTEG
1	32.30	2149.23	7.73	134.85	9.47	14.97	BB
2	33.79	25647.76	92.27	1288.72	90.53	18.69	BB

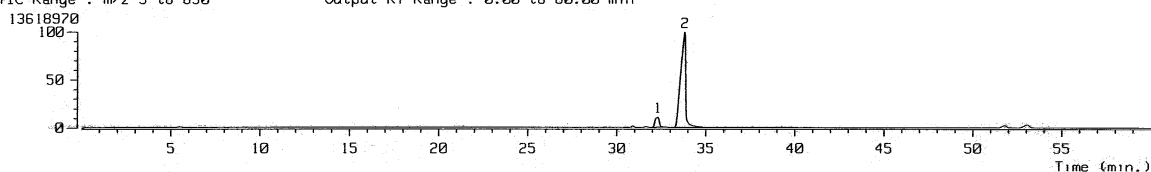
[TIC]

Data : Dr-Cabrera-Armando-918
Sample: 32 G Reacc 512 AX505HA
Note :
Inlet : GC
Ion Species : Normal Ion [MF-Linear]

Date : 12-Jan-108 13:12

Ion Mode : EI+

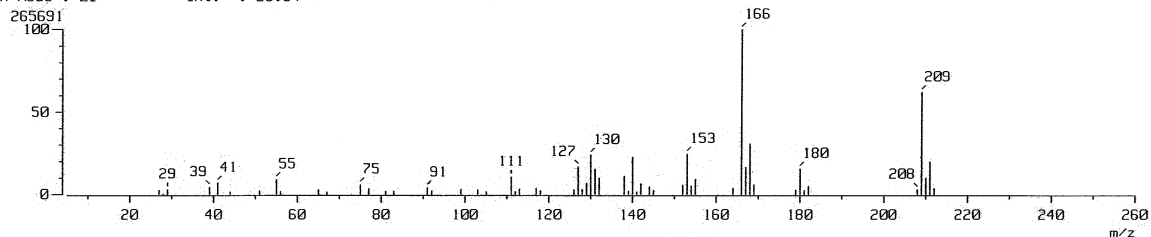
TIC Range : m/z 5 to 650
Output RT Range : 0.00 to 60.00 min



[Mass Spectrum]

RT : 32.30 min
Scan# : 2420-2396-2439
Ion Mode : EI+
Int. : 25.34

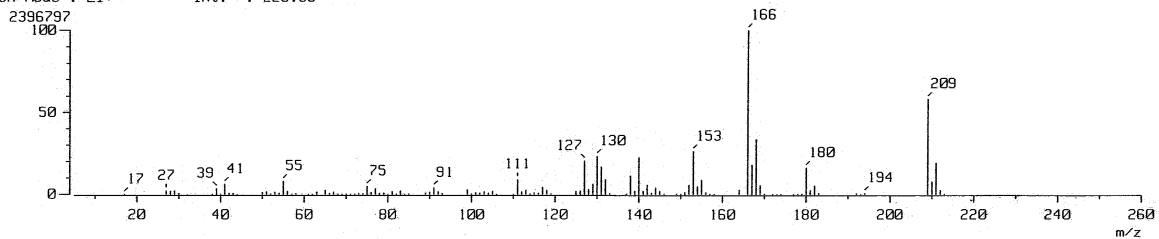
Temp : 0.0 deg.C



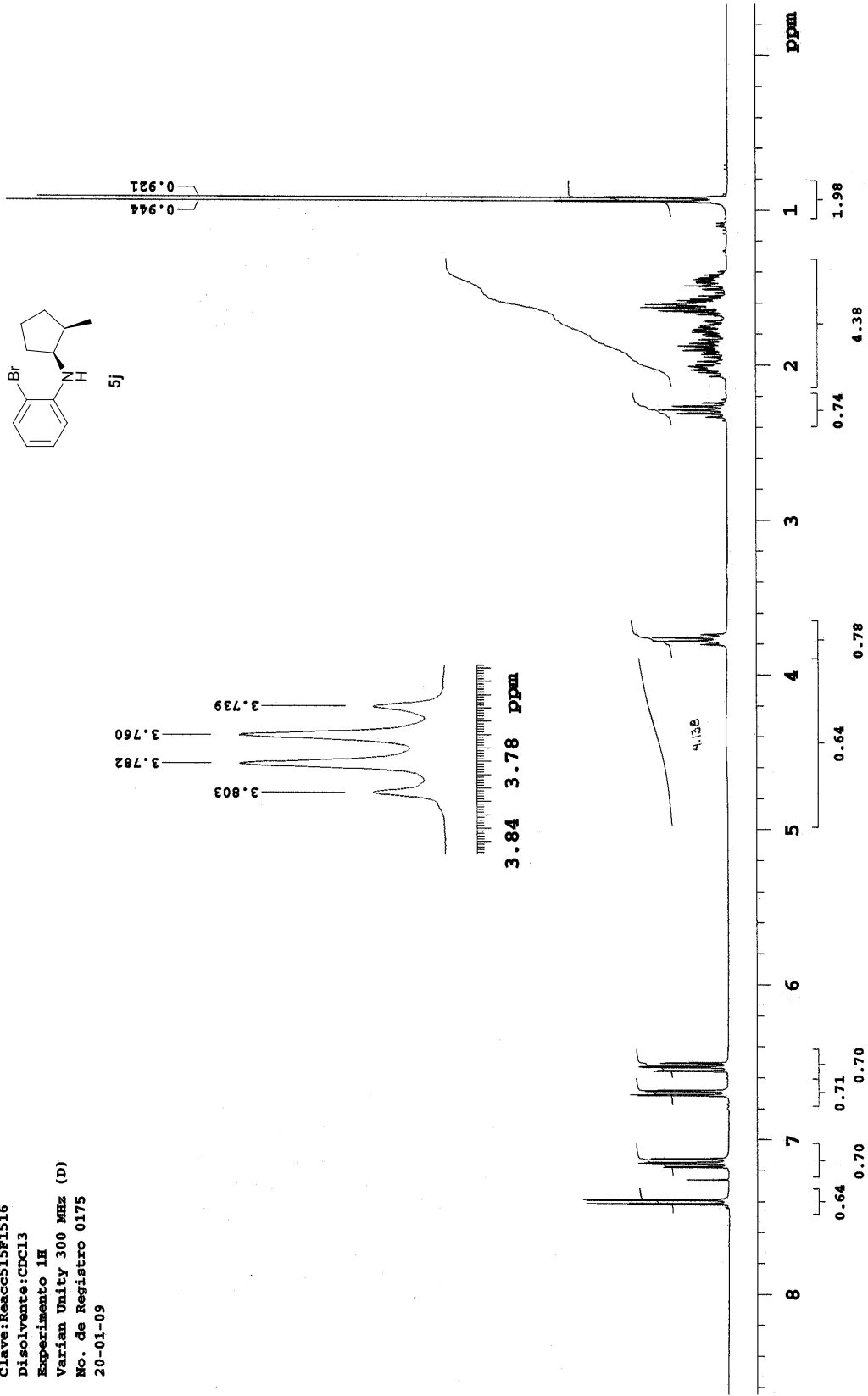
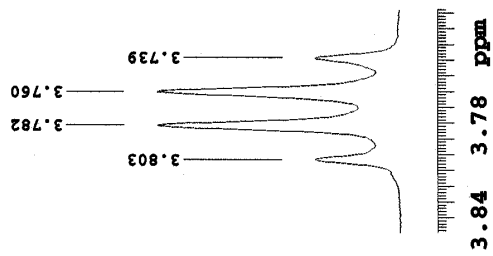
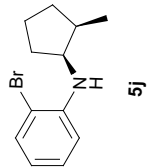
[Mass Spectrum]

RT : 33.79 min
Scan# : 2532-2481-2625
Ion Mode : EI+
Int. : 228.58

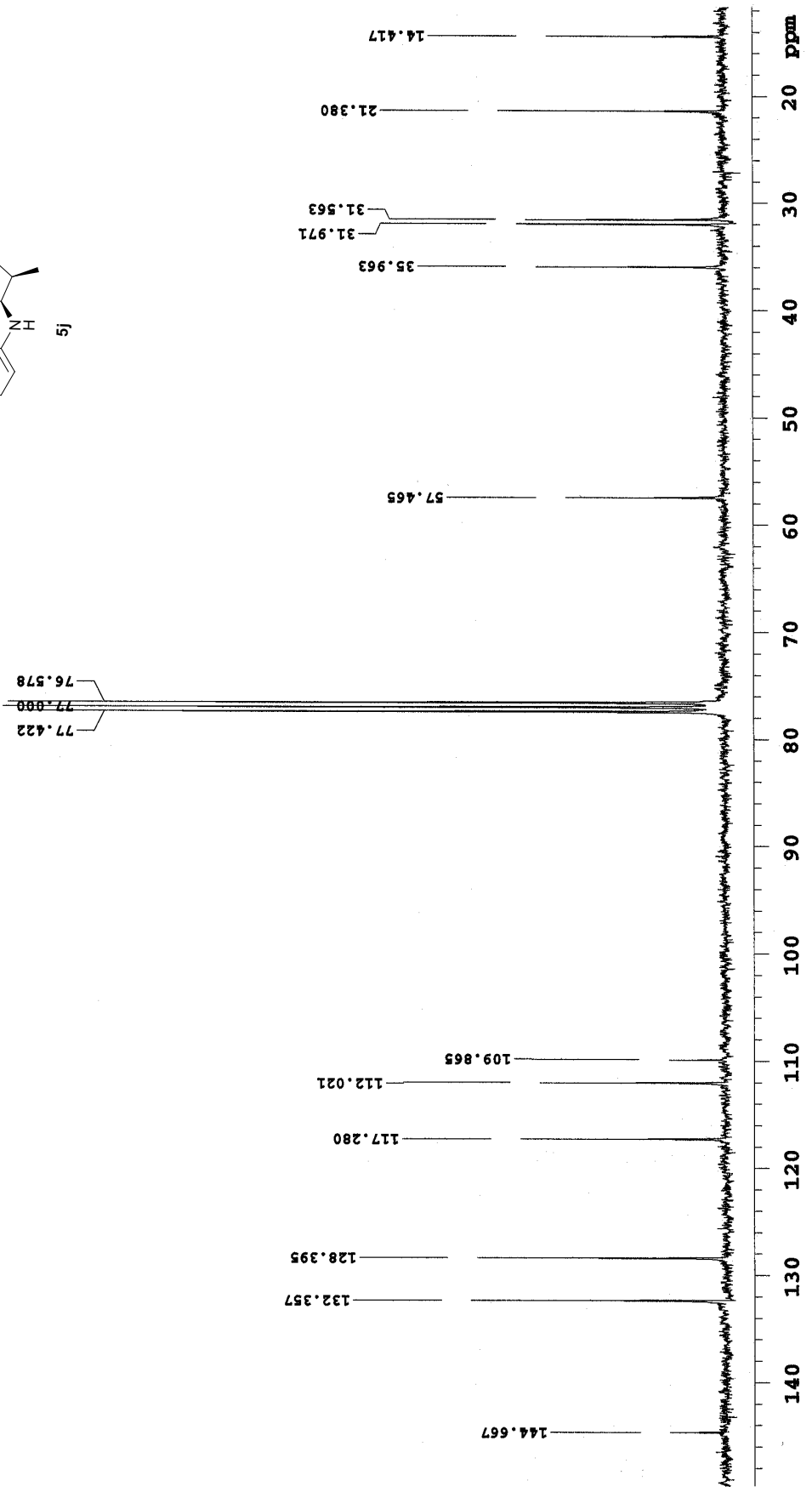
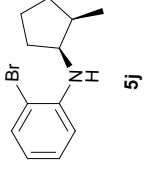
Temp : 0.0 deg.C



U.N.A.M. Instituto de Química ICQ
 Dr. A. Cabrera/Laura R. P
 Clave:Reacc515F1516
 Disolvente:CDCl3
 Experimento 1H
 Varian Unity 300 MHz (D)
 No. de Registro 0175
 20-01-09



U.N.A.M. Instituto de Química ICH
Dr. A. Cabrera/Laura R. P
Clave:Reacc515F1516
Disolvente:CDCl3
Experimento 13C
Varian Unity 75 MHz (D)
No. de Registro 0175
20-01-09



[TIC]

Data : Dr-Cabrera-Armando-924

Date : 15-Jan-120 14:48

Sample: 88 G reacc 515 AX505HA

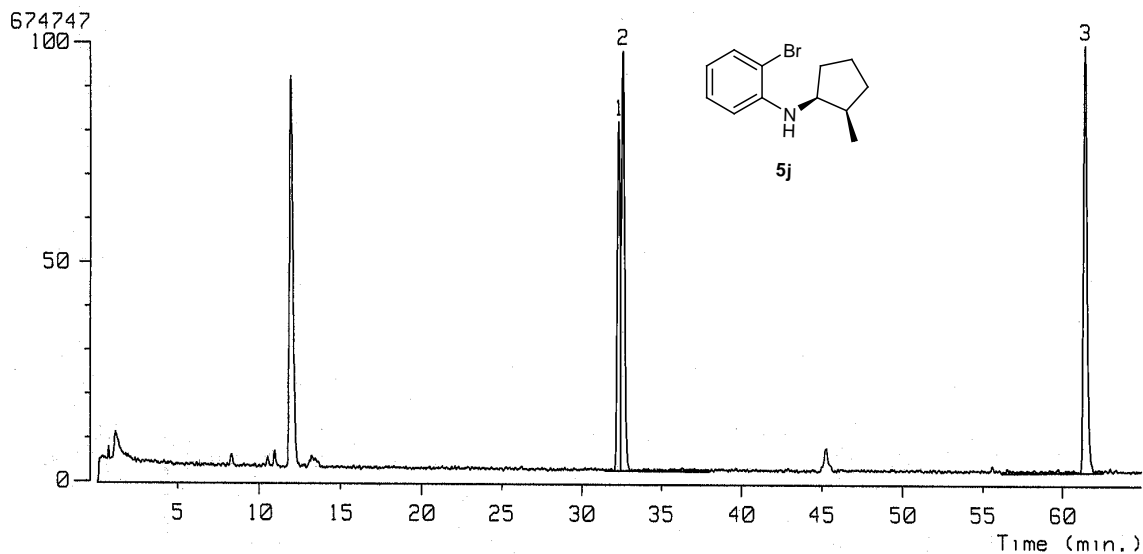
Note :

Inlet : GC

Ion Mode : EI+

Ion Species : Normal Ion

TIC Range : m/z 5 to 650



No.	RT[min]	Area	Area%	Height	Height%	Width[sec]	INTEG
1	32.32	631.92	24.50	51.16	29.19	11.60	BV
2	32.59	902.94	35.01	61.51	35.09	13.78	VB
3	61.44	1044.29	40.49	62.61	35.72	15.66	BB

[TIC]

Data : Dr-Cabrera-Armando-924

Date : 15-Jan-120 14:48

Sample: 88 G reacc 515 AX505HA

Note :

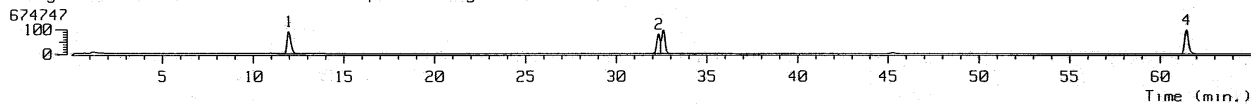
Inlet : GC

Ion Mode : EI+

Ion Species : Normal Ion [MF-Linear]

TIC Range : m/z 5 to 650

Output RT Range : 0.00 to 64.99 min



[Mass Spectrum]

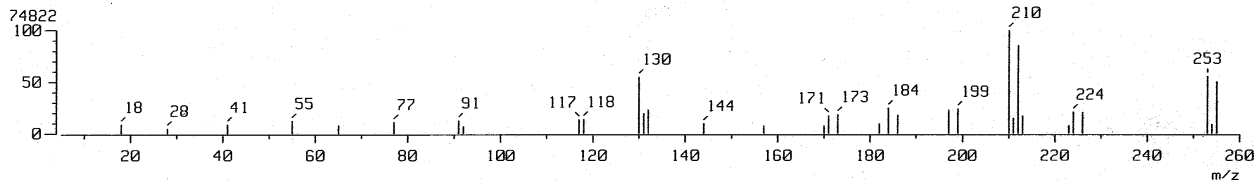
RT : 32.32 min

Scan# : 2422-2356-2046

Temp : 0.0 deg.C

Ion Mode : EI+

Int. : 7.14



[Mass Spectrum]

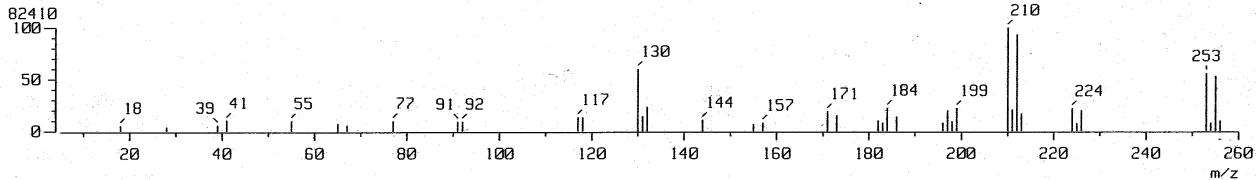
RT : 32.59 min

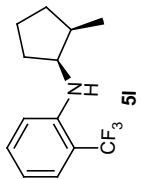
Scan# : 2442-2356-2046

Temp : 0.0 deg.C

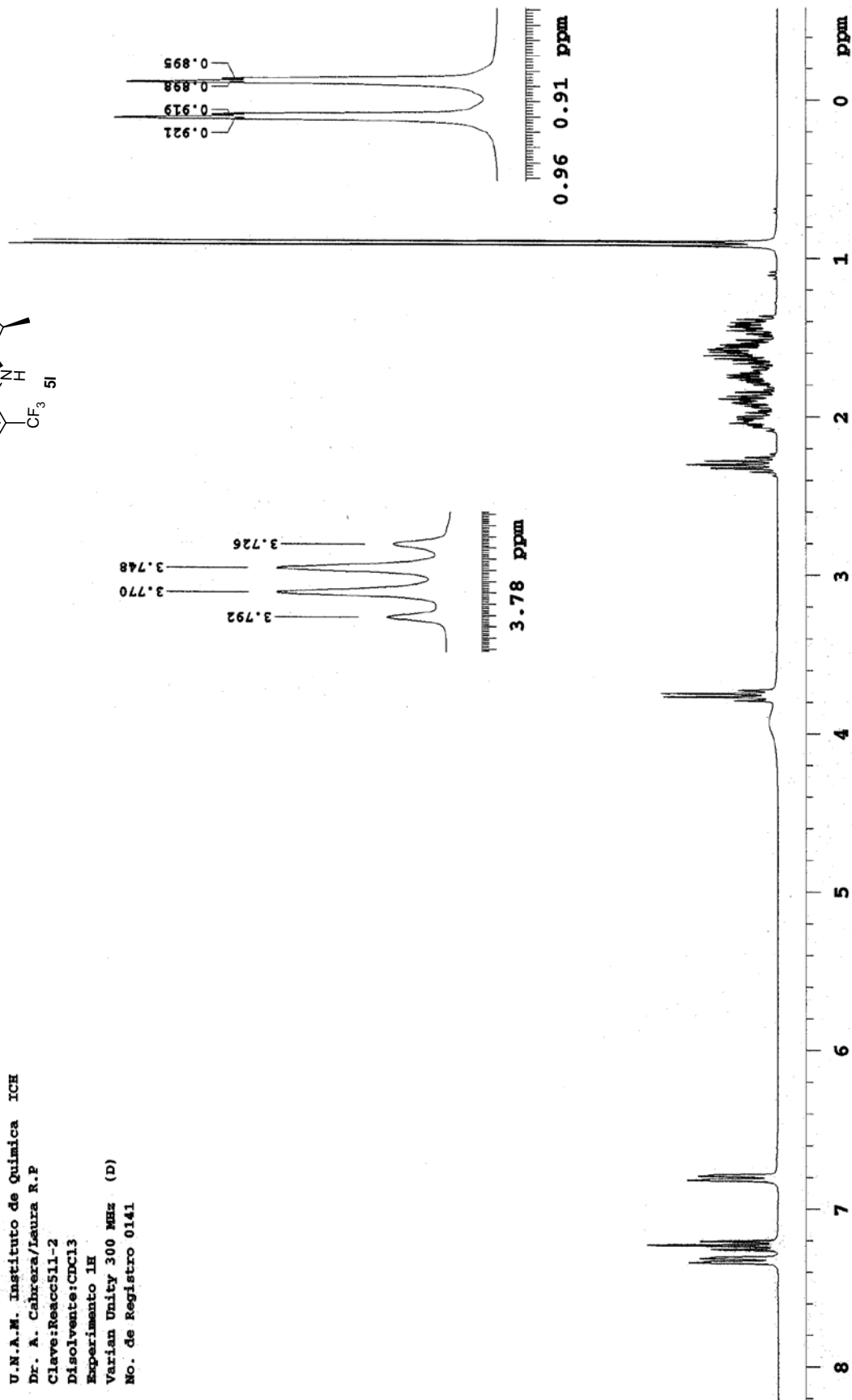
Ion Mode : EI+

Int. : 7.86





U.N.A.M. Instituto de Química ICH
 Dr. A. Cabrera/Laura R.P
 Clave:Reacc511-2
 Disolvente:CDCl3
 Experimento 1H
 Varian Unity 300 MHz (D)
 No. de Registro 0141



[TIC]

Data : Dr-Cabrera-Armando-916

Date : 09-Jan-108 07:58

Sample: 31 G Reacc 511 AX505HA

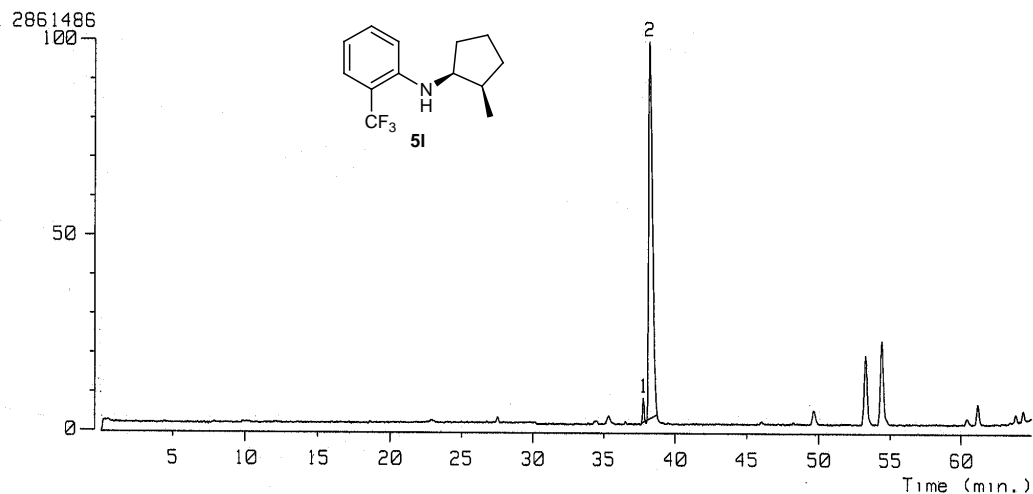
Note :

Inlet : GC

Ion Mode : EI+

Ion Species : Normal Ion

TIC Range : m/z 5 to 650



No.	RT[min]	Area	Area%	Height	Height%	Width[sec]	INTEG
1	37.77	112.96	2.27	16.64	5.94	6.37	BV
2	38.21	4858.82	97.73	263.57	94.06	17.31	VB

[TIC]

Data : Dr-Cabrera-Armando-916

Date : 09-Jan-108 07:58

Sample: 31 G Reacc 511 AX505HA

Note :

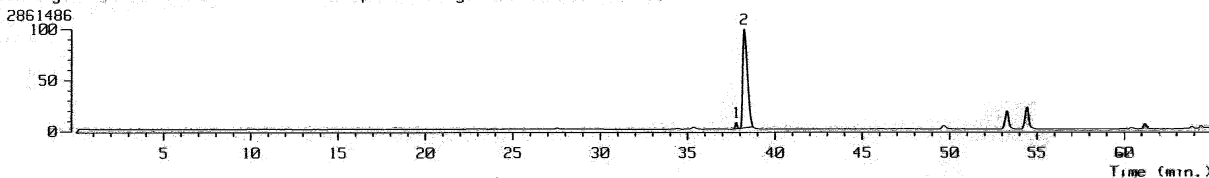
Inlet : GC

Ion Mode : EI+

Ion Species : Normal Ion [MF-Linear]

TIC Range : m/z 5 to 650

Output RT Range : 0.00 to 64.99 min



[Mass Spectrum]

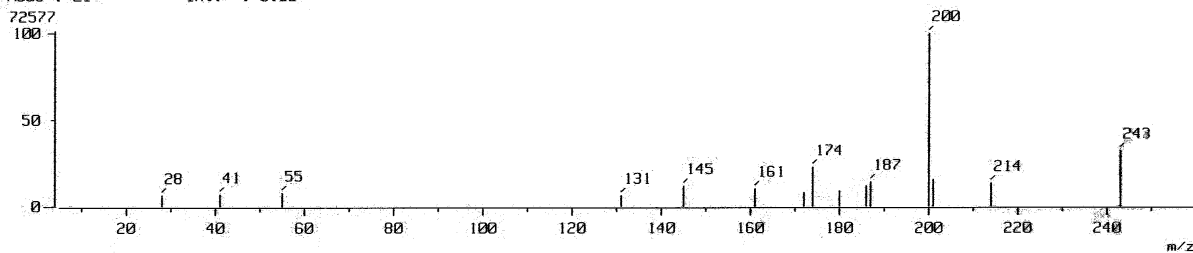
RT : 37.77 min

Scan# : 2830-2821-2903

Temp : 0.0 deg.C

Ion Mode : EI+

Int. : 6.82



[Mass Spectrum]

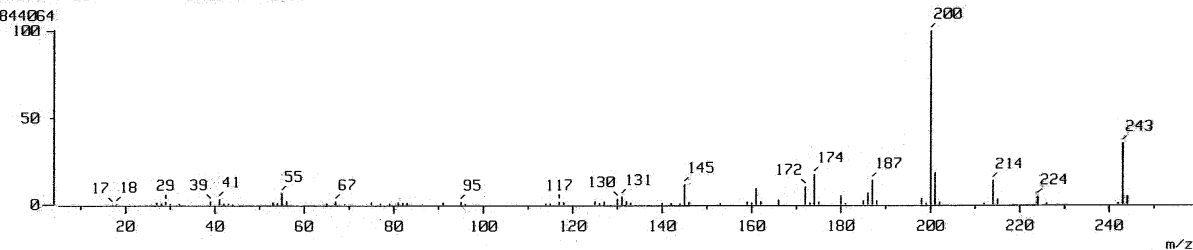
RT : 38.21 min

Scan# : 2863-2821-2903

Temp : 0.0 deg.C

Ion Mode : EI+

Int. : 79.91



INSTITUTO DE QUIMICA, UNAM/EHS

Dr. A. Cabrera / Laura R. P.

Clave: Reacc:309

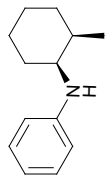
Disolvente: CDCl₃

Hidrogeno-1

Eclipse 300 MHz Jeol (E)

27-01-09

Nº. de registro: 0286



3.523
3.512
3.500
3.488
3.478

3.641

0.934

X : parts per Million : 1H

3.5 3.6 3.7

3.5 3.6 3.7

3.5 3.6 3.7

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3.5 3.6 3.7

3.5 3.6 3.7

3.5 3.6 3.7

INSTITUTO DE QUIMICA, UNAM/EHS

Dr. A. Cabrera/Laura R. P.

Clave: Reacc309

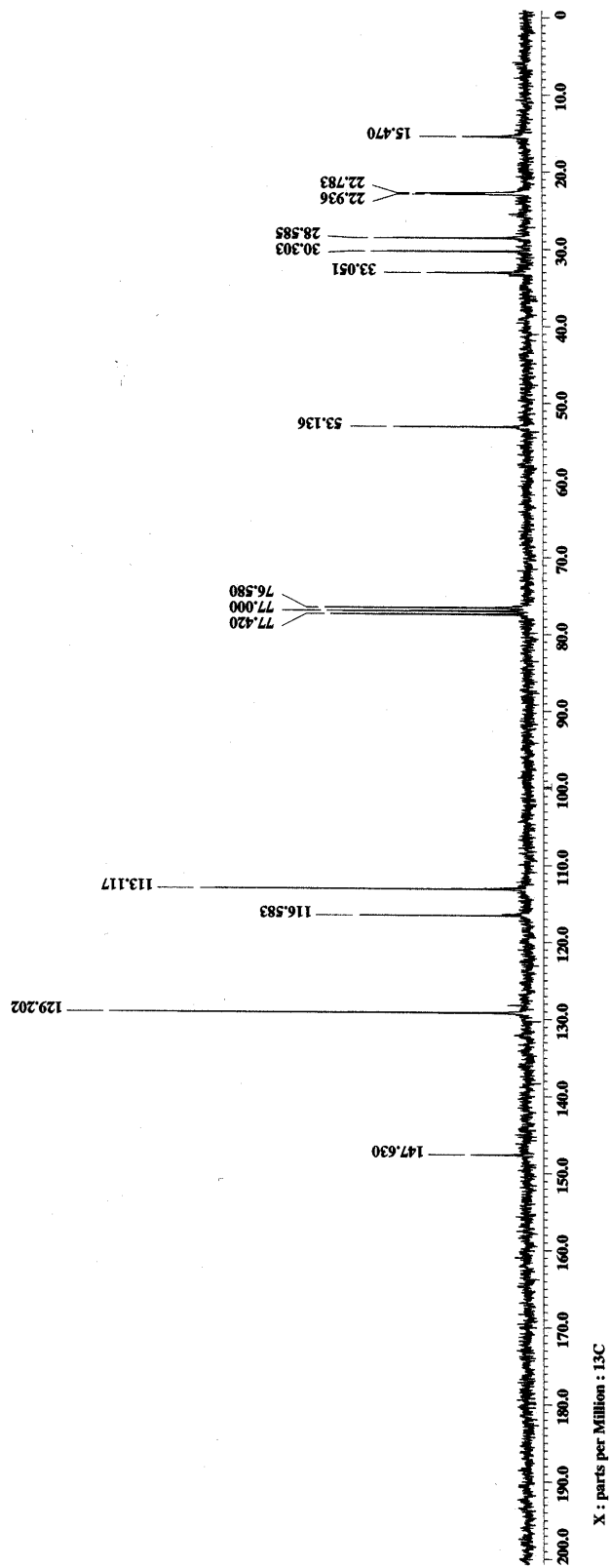
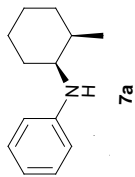
Disolvente: CDCl₃

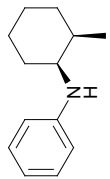
Carbono-13

Eclipse 300 MHz Jeol (E)

27-01-09

No. de registro: 0286





cis-6a
(Table 2, entry 11)

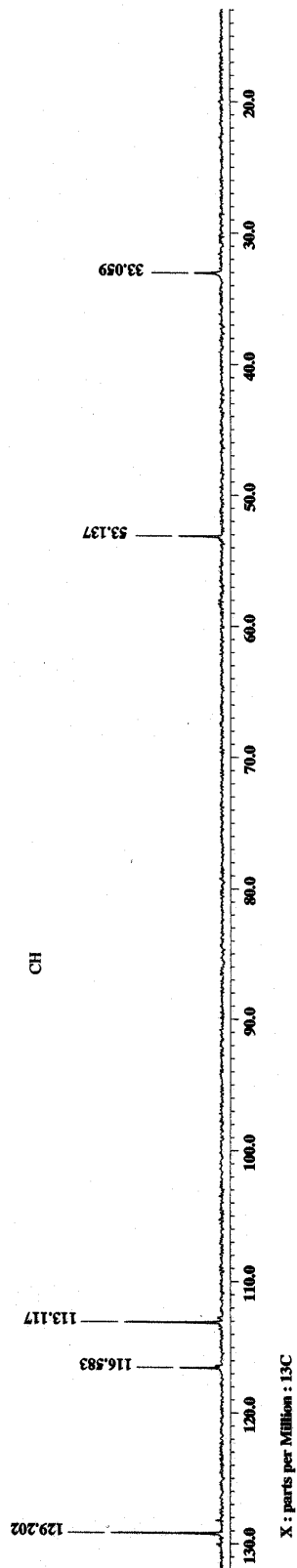
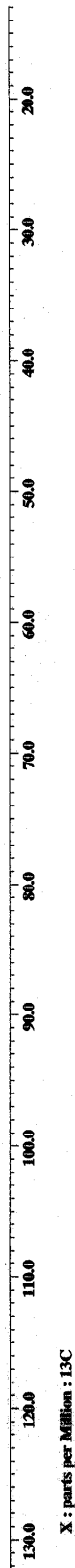
129.201
113.116
116.581
113.117

CH, CH3

33.057
30.309
28.591
22.942
22.782
15.476

CH2

INSTITUTO DE QUIMICA, UNAM/EHS
Dr. A. Cabrea/Laura R. P.
Clave: Reac309
Disolvente: CDCl3
DEPT
Echigo: 300 MHz, Teol (E)
27-01-09
No. de registre: 0286



CH

[TIC]

Data : Dr-Dr-Cabrera-Armando-841

Date : 23-Oct-107 15:37

Sample: 2118 G reacc 309 AX505HA

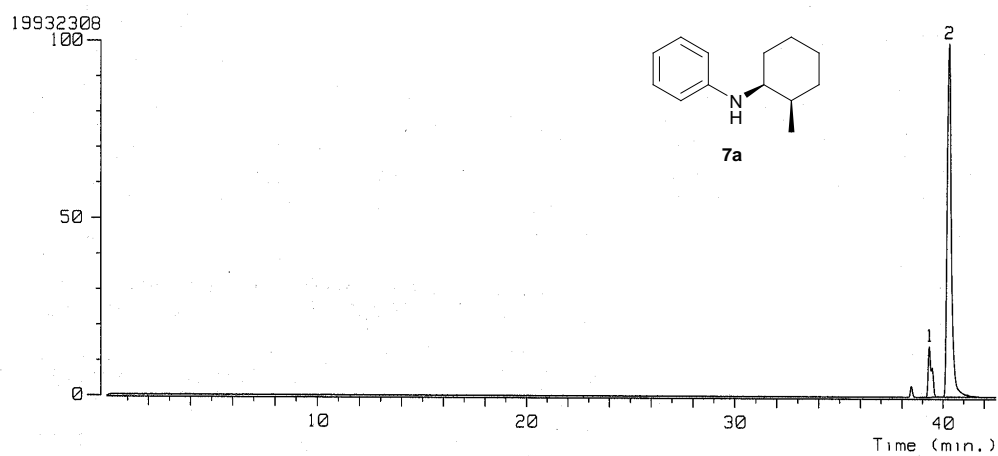
Note :

Inlet : GC

Ion Mode : EI+

Ion Species : Normal Ion

TIC Range : m/z 5 to 650



No.	RT[min]	Area	Area%	Height	Height%	Width[sec]	INTEG
1	39.34	3215.02	10.67	270.85	12.52	11.15	BB
2	40.35	26924.89	89.33	1892.75	87.48	13.36	BB

[TIC]

Data : Dr-Dr-Cabrera-Armando-841

Date : 23-Oct-107 15:37

Sample: 2118 G reacc 309 AX505HA

Note :

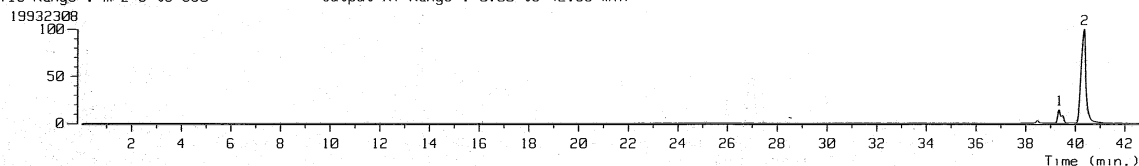
Inlet : GC

Ion Mode : EI+

Ion Species : Normal Ion [MF-Linear]

TIC Range : m/z 5 to 650

Output RT Range : 0.00 to 42.58 min



[Mass Spectrum]

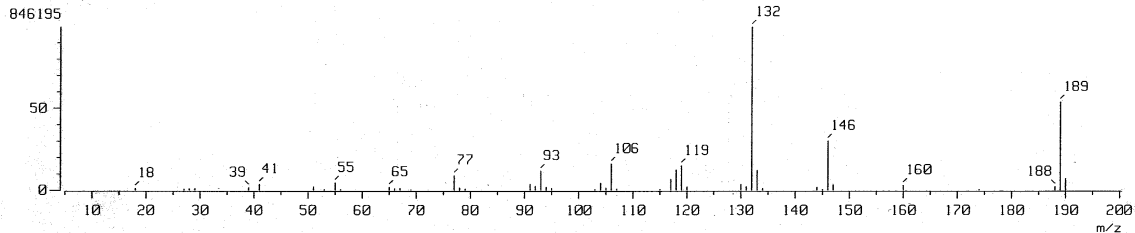
RT : 39.34 min

Scan# : 2951-2938-2980

Temp : 0.0 deg.C

Ion Mode : EI+

Int. : 81.10



[Mass Spectrum]

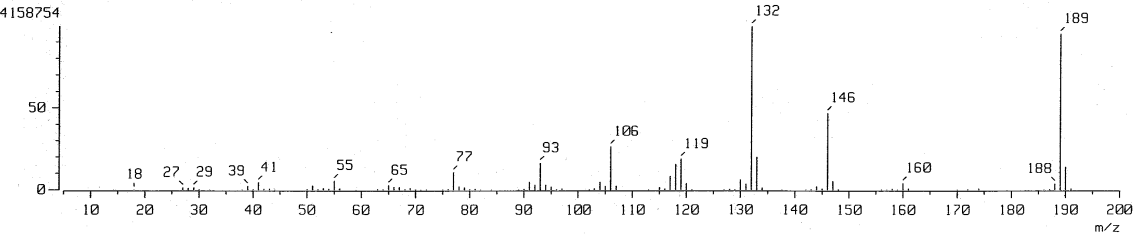
RT : 40.35 min

Scan# : 3027-2998-3125

Temp : 0.0 deg.C

Ion Mode : EI+

Int. : 398.54





Reacc327F24-1H-4.jdf

```

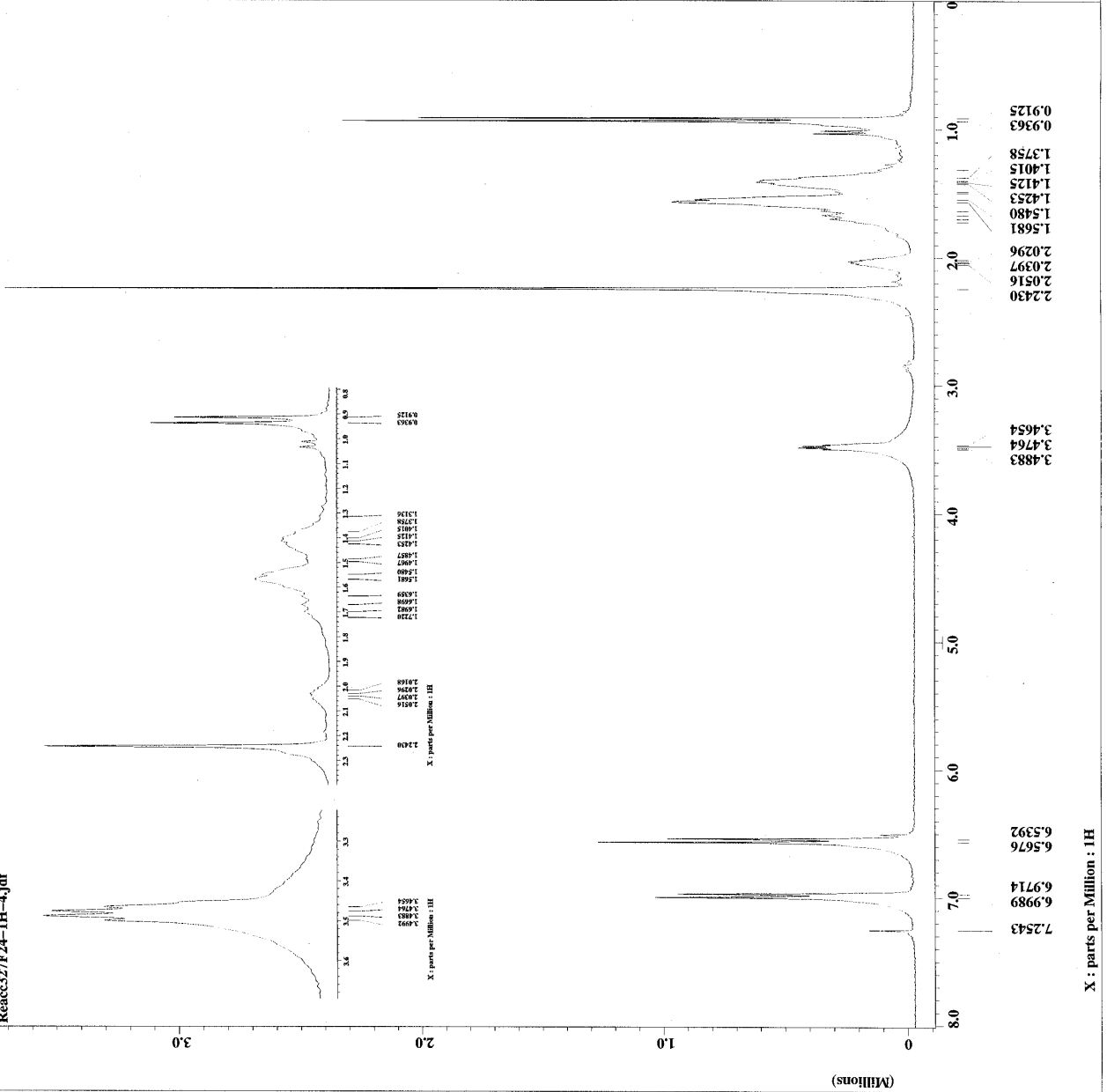
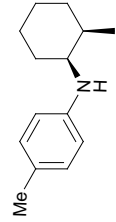
Filename = Reacc327F24-1H-4.jdf
Author = Cabrera
Experiment = Single_pulse_exp
Solvent = CHLOROFORM-D
Creation_time = 14-DEC-2007 18:16:05
Revision_time = 19-DEC-2007 16:57:53
Current_time = 19-DEC-2007 16:59:30

Comment = Single Pulse Experiment
Data_format = 1D COMPLEX
Dim_size = 16384
Dim_title = 1H
Dim_units = [ppm]
Dimensions = X
Site = Eclipse+ 300
Spectrometer = DELTA_NMR

Field_strength = 7.0586013[T] (300 MHz)
X_acq_duration = 3.6339712[s]
X_domain = 1H
X_freq = 300.52965592[MHz]
X_offset = 5[ppm]
X_points = 16384
X_prescans = 0
X_resolution = 0.27518105[Hz]
X_sweep = 4.50856628[MHz]
Clipped = TRUE
Mod_return = 1
Scans = 32
Total_scans = 32

X_90_width = 9.5[us]
X_acq_time = 3.6339712[s]
X_angle = 45[deg]
X_pulse = 1.75[us]
Initial_wait = 3[s]
Phase_preset = 19
Relaxation_delay = 4[s]
Temp_get = 16.3[dc]
Unblank_time = 2[us]

```



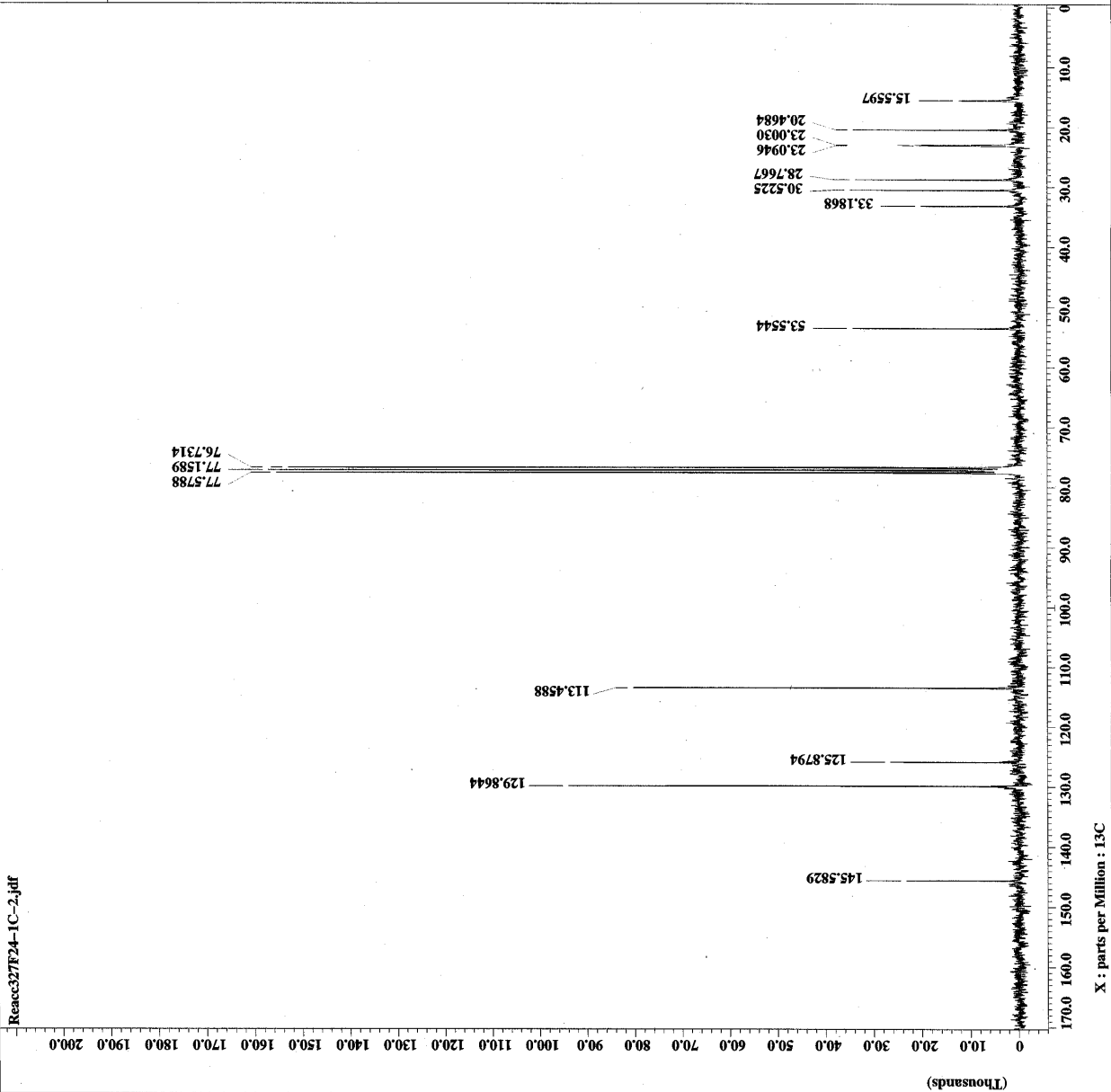
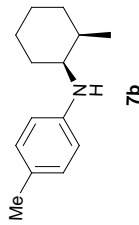
```

File name      = Reacc327F24-1C-2.jdf
Author        =
Experiment    =
Exp. date     =
Sample ID     = Laura_pulse_dec
Solvent       = CHLOROFORM-D
Creation time  = 15-DEC-2007 00:21:45
Revision time = 20-DEC-2007 08:55:59
Current time  = 20-DEC-2007 09:00:07

Comment       = Single Pulse with Bro
Data format   = 1D COMPLEX
Dir. size     = 32768
Dir. title    = 13C
Dir. units    = [ppm]
Dimensions    = x
Site          = Eclipse+ 300
Spectrometer = DELTA_NMR

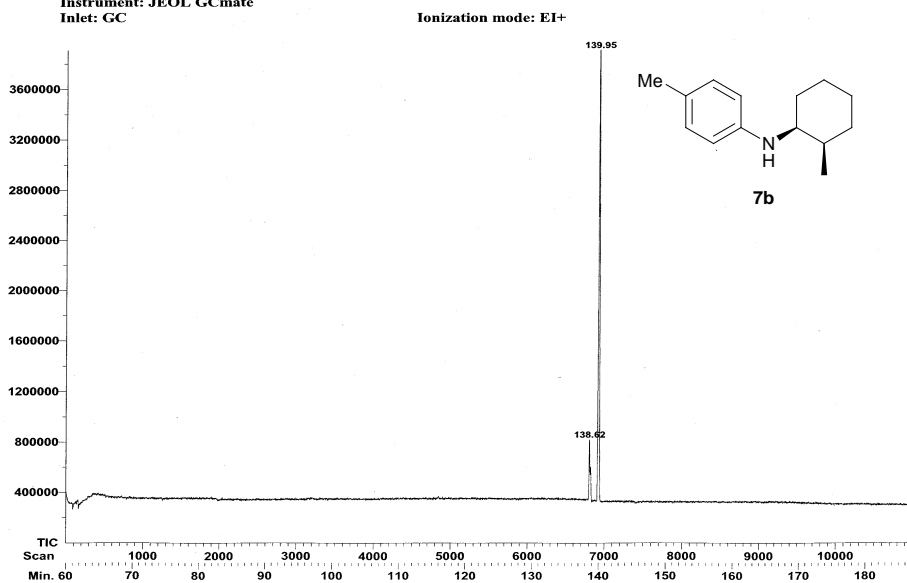
Field strength = 7.0586013[T] (300 [MHz]
X_acq_duration = 1.7334272[s]
X_domain       = 13C
X_freq         = 75.56823426 [MHz]
X_offset       = 30 [ppm]
X_resolution   = 32768
X_points       = 4
X_sweep        = 0.57689184 [Hz]
X_resolution   = 18.90359168 [kHz]
Irr_domain    = 1H
Irr_sweep     = 18.90359168 [kHz]
Irr_freq      = 300.52965592 [MHz]
Irr_offset    = 5 [ppm]
Clipped       = TRUE
Mod_return    = 1
Scans         = 8000
Total_scans   = 8000

X_90_width    = 11.3 [us]
X_acq_time    = 1.7334272 [s]
X_angle       = 30 [deg]
X_pulse       = 3.76666667 [us]
Initial_wait  = 1 [s]
Phase_preset = 3 [us]
Recvr_gain    = 30
Relaxation_delay = 1 [s]
Temp_get      = 17.8 [dc]
Unblank_time  = 2 [us]
  
```



File: 1236-Reac327-2
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 06-02-2009 (Time Run: 08:50:34)



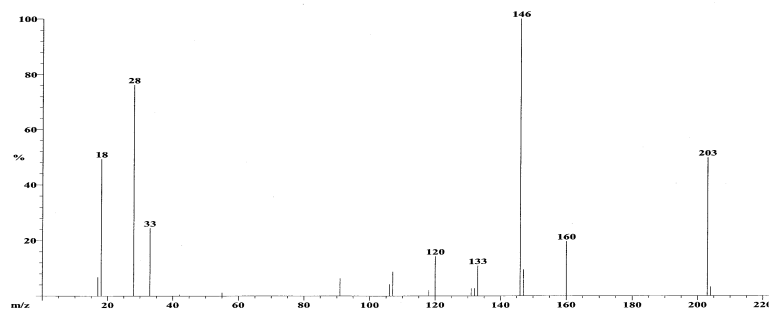
File: 1236-Reac327-2
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 06-02-2009 (Time Run: 08:50:34)

Ionization mode: EI+

Scan: 6805 R.T.: 138.62
 Base: m/z 146; 5.1%FS TIC: 838064

#Ions: 19



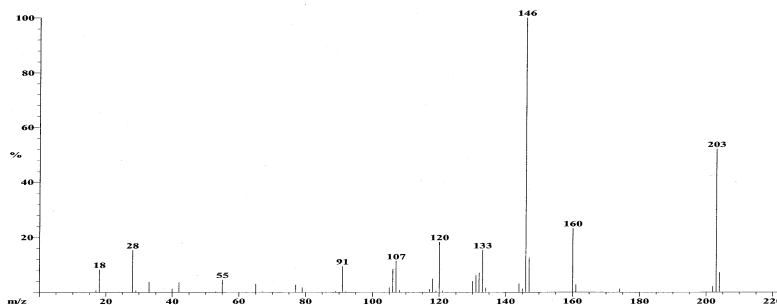
File: 1236-Reac327-2
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 06-02-2009 (Time Run: 08:50:34)

Ionization mode: EI+

Scan: 6921 R.T.: 139.95
 Base: m/z 146; 27%FS TIC: 3925680

#Ions: 43

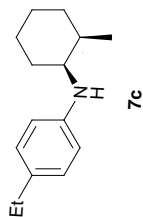


Peak Table

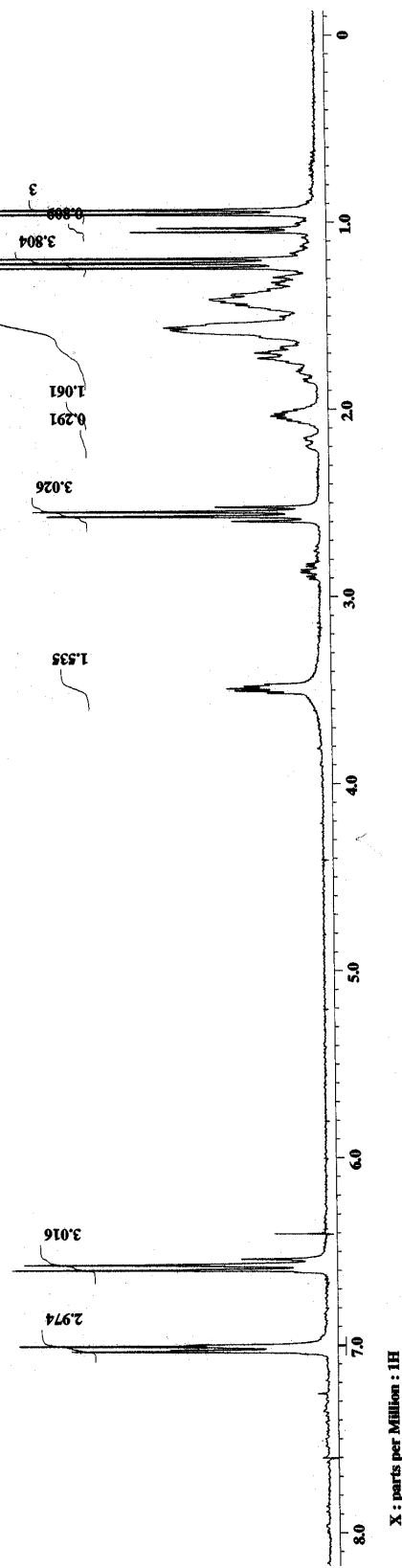
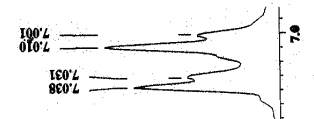
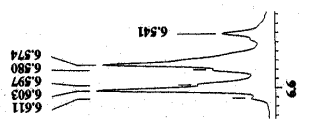
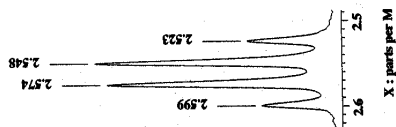
Peak Label	RIC	Scan Range	Baseline at	Total Area	Background	Peak Area
138.62	TIC	6793-6829	6784-6788	14388850	8469663	5919187
139.95	TIC	6905-6941	6895-6899	49880378	8273759	41606619

UNAM, INSTITUTO DE QUIMICA, apg
 Dr. Armando Cabrera / Laura R. P.
 Clave: Reacc. 507-Et
 Disolvente: CDCl₃

¹H
 Eclipse: 300 MHz Jeol (E)
 10-12-08
 No. Reg. 4116



1.245
 1.220
 1.195
 0.959
 0.936



UNAM, INSTITUTO DE QUIMICA, apg

Dr. Armando Cabezas / Laura R. P.

Clave: Reacc. 507-B

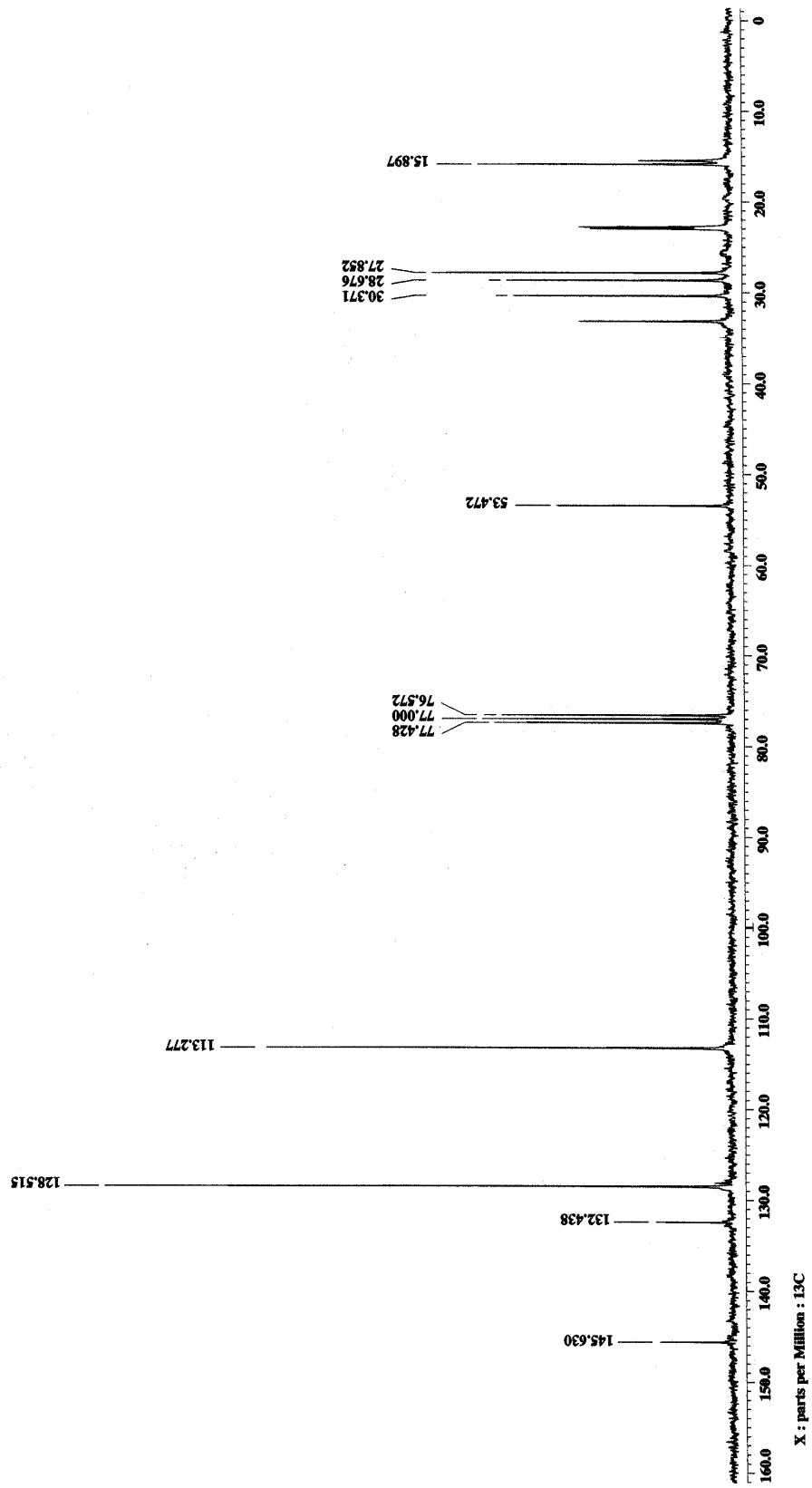
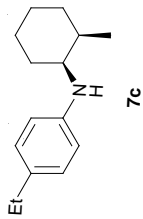
Disolvente: CDCl₃

13C

Eclipse: 900 MHz Isol (E)

10-12-08

No. Reg. 4116



[TIC]

Data : Dr-Cabrera-Armando-515

Date : 02-Apr-120 09:44

Sample: 799 G Rx 131 AX505HA

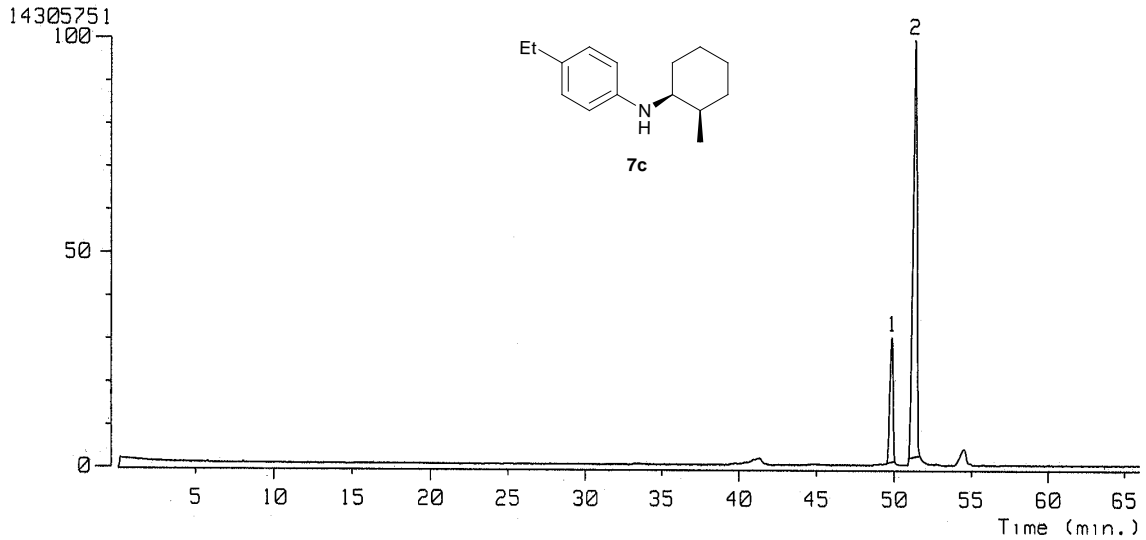
Note :

Inlet : GC

Ion Mode : EI+

Ion Species : Normal Ion

TIC Range : m/z 5 to 650



No.	RT [min]	Area	Area%	Height	Height%	Width [sec]	INTEG
1	49.87	5331.06	19.06	396.18	23.05	12.64	BV
2	51.44	22645.32	80.94	1322.93	76.95	16.07	VB

[TIC]

Data : Dr-Cabrera-Armando-515

Date : 02-Apr-120 09:44

Sample: 799 G Rx 131 AX505HA

Note :

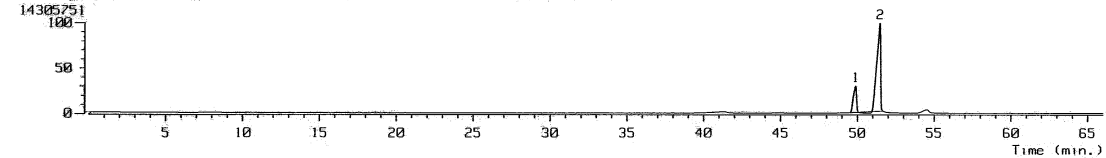
Inlet : GC

Ion Mode : EI+

Ion Species : Normal Ion [MF-Linear]

TIC Range : m/z 5 to 650

Output RT Range : 0.00 to 65.99 min



[Mass Spectrum]

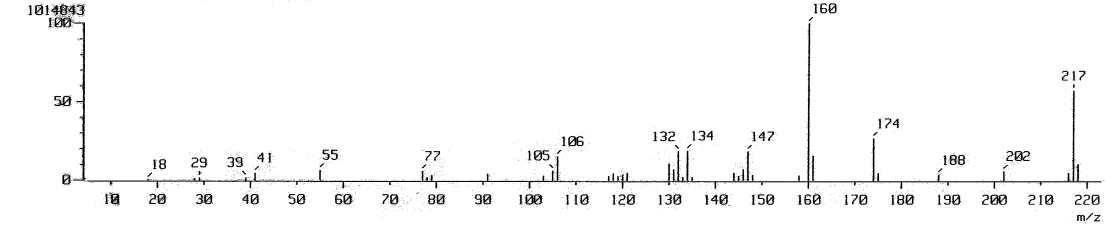
RT : 49.87 min

Scan# : 3741-3716-3876

Temp : 0.0 deg.C

Ion Mode : EI+

Int. : 96.78



[Mass Spectrum]

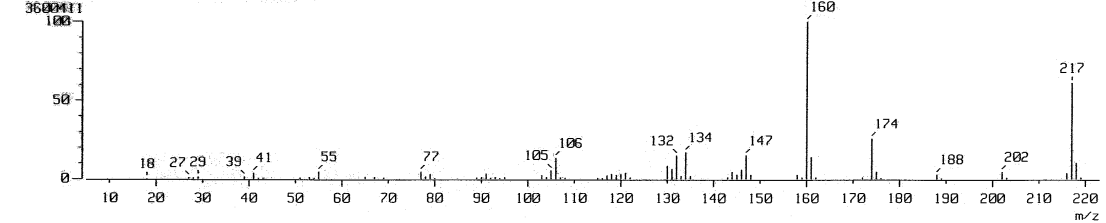
RT : 51.44 min

Scan# : 3859-3716-3876

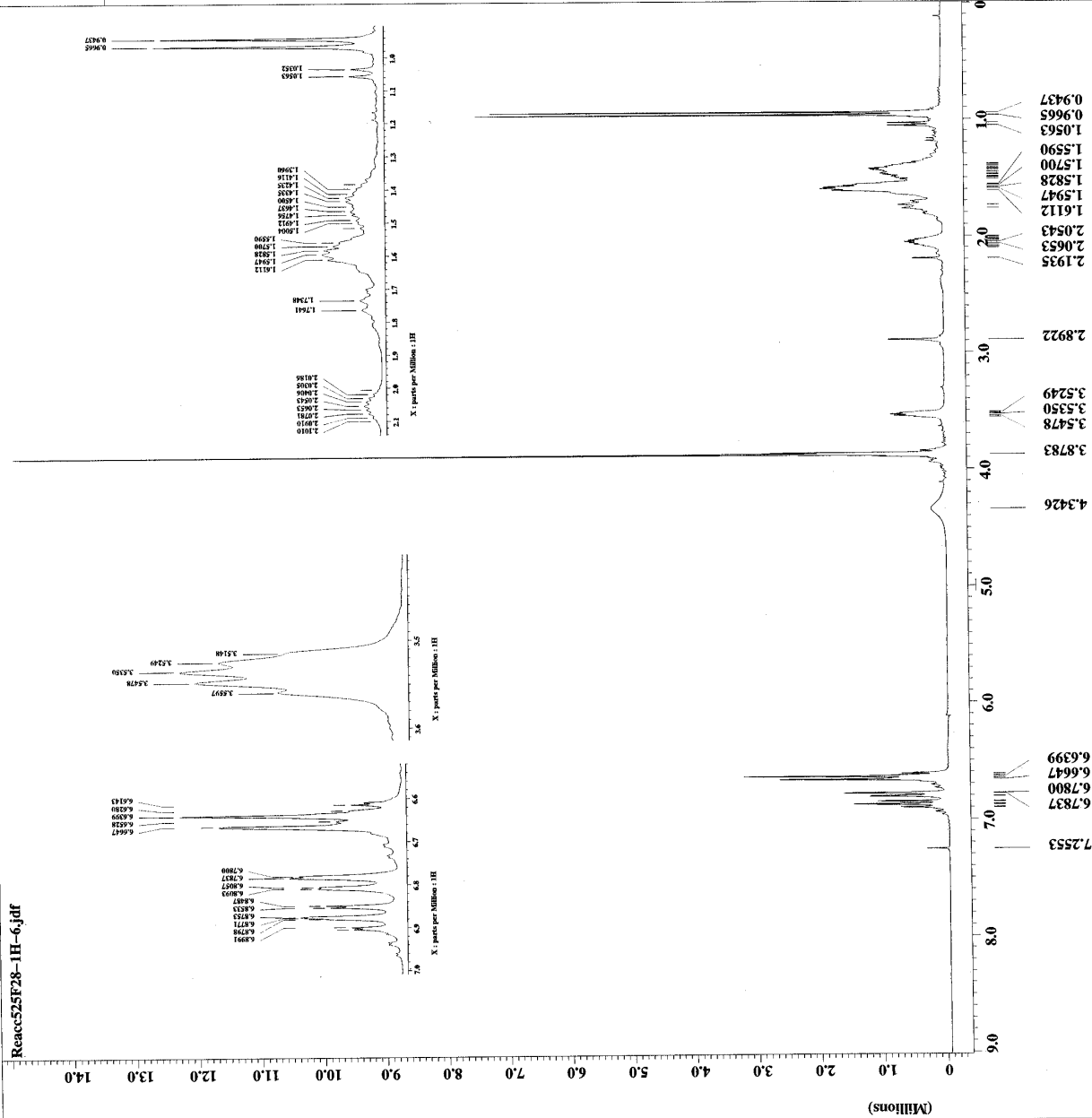
Temp : 0.0 deg.C

Ion Mode : EI+

Int. : 343.36



Reacc525F28-1H-6.jdf



JEOL

```

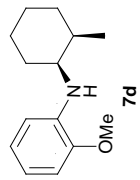
Filename = Reacc525F28-1H-6.jdf
Author = Cabrera
Experiment = single_pulse.exp
Sample_id =
SOLVENT = CDCl3
Creation_time = 8-DEC-2008 00:57:24
Revision_time = 19-FEB-2009 09:19:13
Current_time = 19-FEB-2009 09:19:30

Comment = Single Pulse Experiment
Data_format = 1D COMPLEX
Dir_name = 16384
Dir_title = 1H
Dim_units = [ppm]
Dimensions = X
Site = Eclipse+ 300
Spectrometer = DELTA_NMR

Field_strength = 7.0586013[T] (300[MHZ]
X_acq_duration = 3.6339712[s]
X_domain = 1H
X_freq = 300.52965592[MHZ]
X_offset = 5[ppm]
X_points = 16384
X_prescans = 0
X_resolution = 0.27518105[Hz]
X_sweep = 4.50856628[MHZ]
Clipped = FALSE
Mod_return = 1
Total_scans = 3

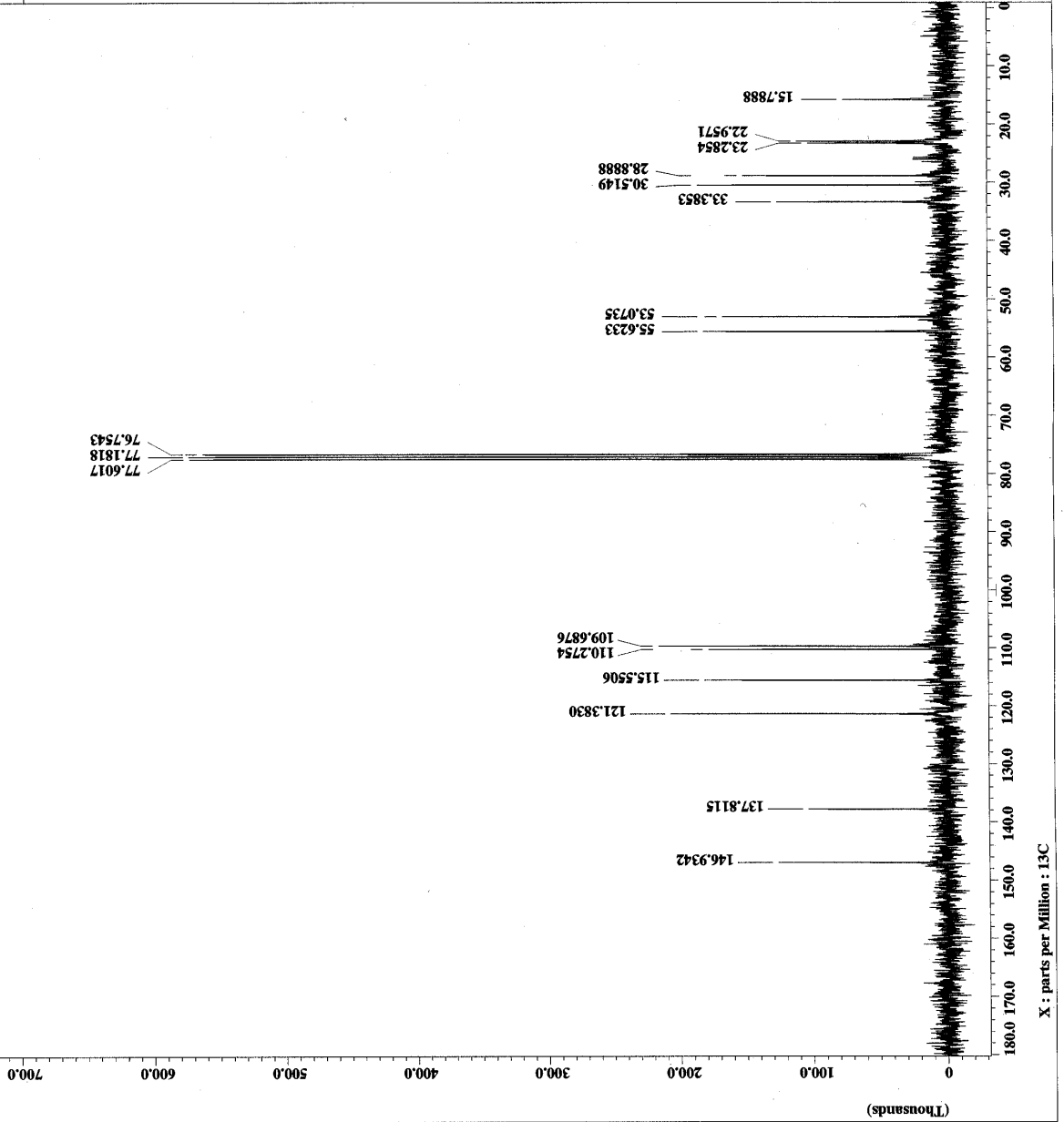
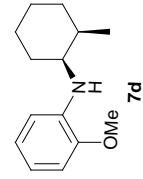
X_90_width = 9.5[us]
X_acq_time = 3.6339712[s]
X_angle = 45[deg]
X_pulse = 4.75[us]
Initial_wait = 1[s]
Phase_preset = 3[us]
Recvr_gain = 16
Relaxation_delay = 1[s]
Temp_get = 18.9[dc]
Unblank_time = 2[us]

```



```

Filename = Reacc525F28-13C-2.jdf
Author = Cabrera
Experiment = single_pulse_dec
Sample_id = laura
Solvent = CDCl3
Creation_time = 8-DEC-2008 01:40:57
Revision_time = 19-FEB-2009 09:45:59
Current_time = 19-FEB-2009 09:47:20
Comment = Single Pulse with Bro
Data_format = 1D COMPLEX
Pulse_program = zgpg30
Pulse_sequence = 13C
Pulse_width = 32768
Pulse_delay = 13C
Pulse_offset = 13C
Pulse_phase = X
Pulse_modulation = ECLIPSE+ 300
Spectrometer = DELTA_NMR
Field_strength = 7.0586013[T] (300[MHZ]
Acq_duration = 1.7334272[s]
F1_domain = 13C
F2_domain = 13C
F1_freq = 75.56823426[MHZ]
F2_freq = 300.52965592[MHZ]
F1_offset = 32768
F2_offset = 4
F1_resolution = 0.57689184[Hz]
F2_resolution = 18.90359168[MHz]
Sweep_rate = 18
Irr_domain = 1H
Irr_freq = 300.52965592[MHZ]
Irr_offset = 51ppm
Clipped = FALSE
Mod_return = 1
Scans = 320
Total_scans = 930
X_90_width = 11.3[us]
X_90_time = 1.7334272[s]
X_acq_time = 30[sec]
X_pulse = 3.76666667[us]
Initial_wait = 1[s]
Phase_preset = 3[us]
Recvr_gain = 30
Relaxation_delay = 1[s]
Temp_set = 20.6[degC]
Unblank_time = 2[us]
  
```

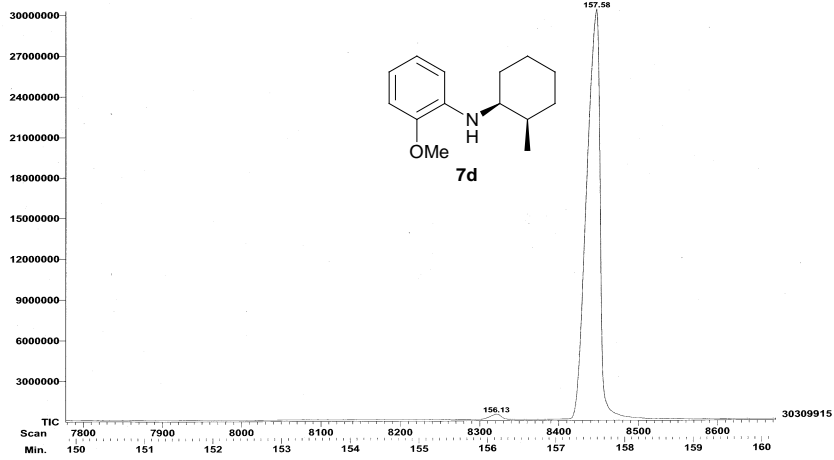


X : parts per Million : 13C

File: 1209-Reac525
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-28-2009 (Time Run: 15:26:08)

Ionization mode: EI+



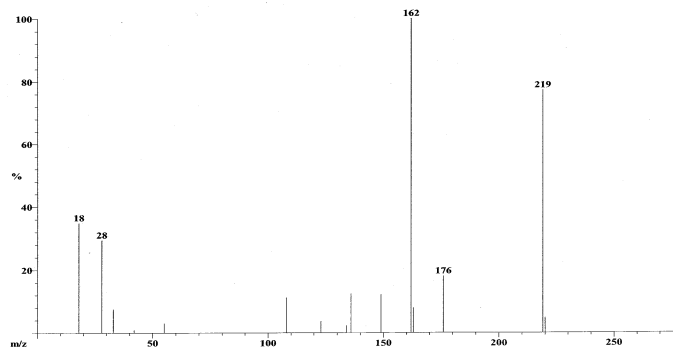
File: 1209-Reac525
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-28-2009 (Time Run: 15:26:08)

Ionization mode: EI+

Scan: 8321 R.T.: 156.13
 Base: m/z 162; 3.5%FS TIC: 474176

#Ions: 16



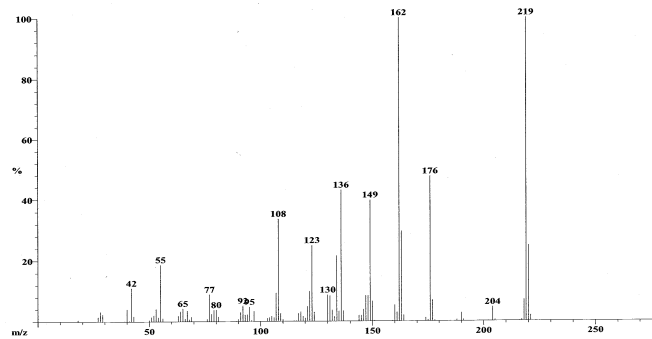
File: 1209-Reac525
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-28-2009 (Time Run: 15:26:08)

Ionization mode: EI+

Scan: 8447 R.T.: 157.58
 Base: m/z 219; 99.6%FS TIC: 30437712

#Ions: 109



Peak Table

Peak Label	RIC	Scan Range	Baseline at	Total Area	Background	Peak Area
156.13	TIC	8301-8337	8339-8343	6642961	2632589	4010372
157.58	TIC	8421-8457	8469-8473	397293413	12381400	384912013

INSTITUTO DE QUIMICA, UNAM / EHS

Dr. A. Cabrera/Laura R. P.

Clave: reacc341

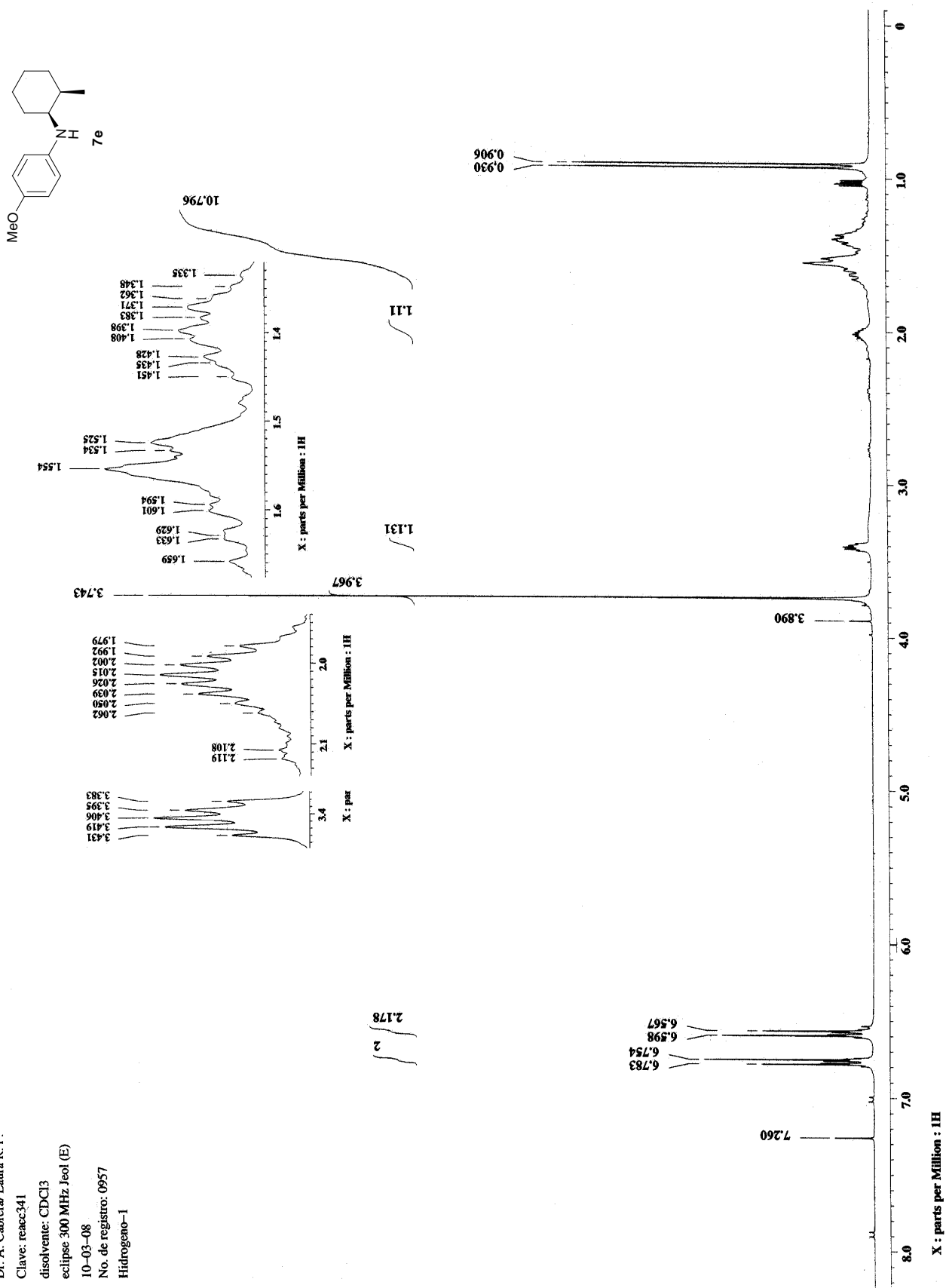
disolvente: CDCl₃

eclipse 300 MHz Jeol (E)

10-03-08

No. de registro: 0957

Hidrogeno-1



INSTITUTO DE QUIMICA, UNAM / EHS

Dr. A. Cabrera/ Laura R. P.

Clave: Reacc341

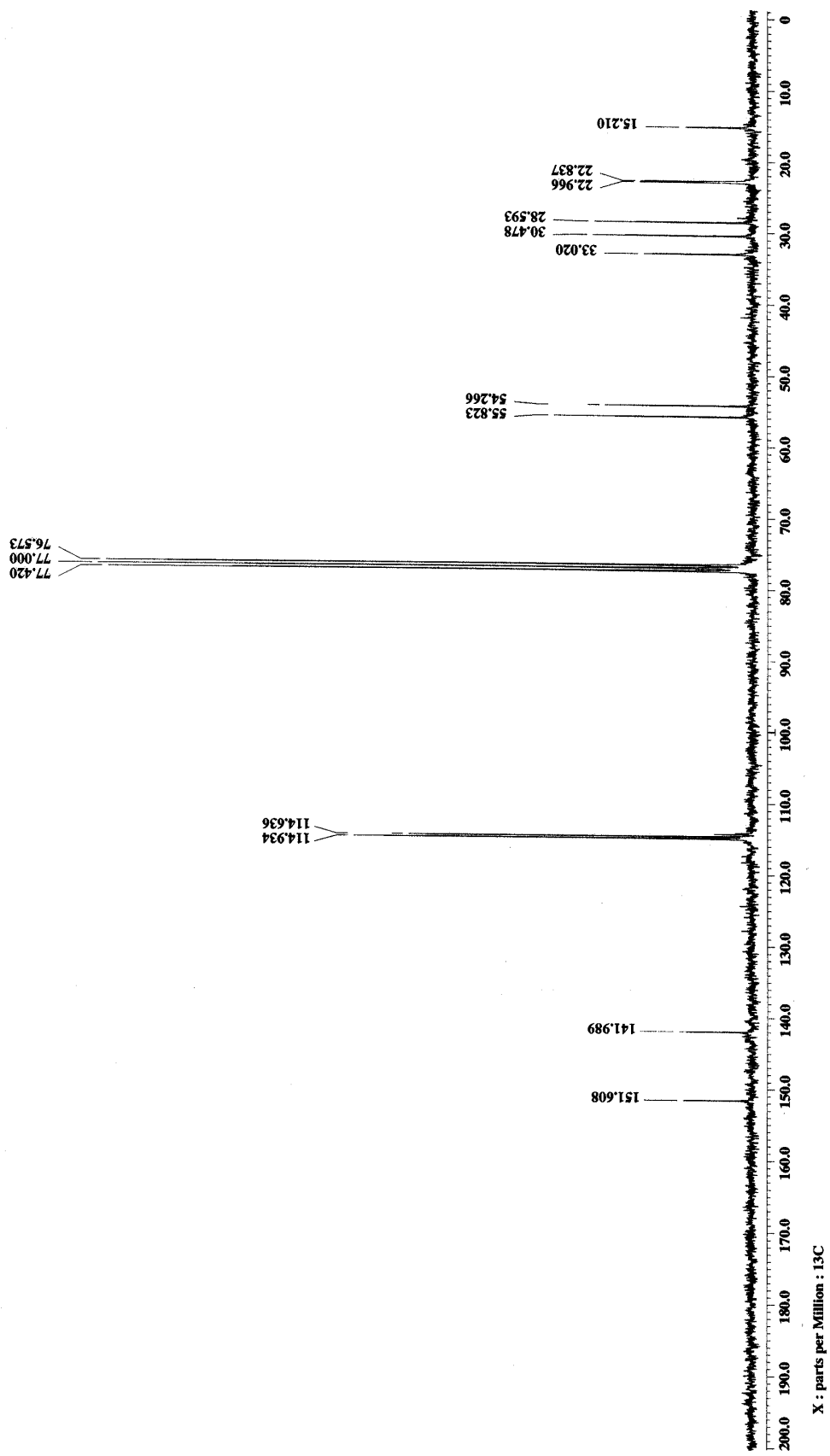
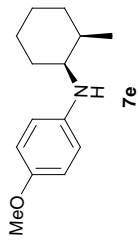
Disolvente: CDCl₃

Carbono-13

Eclipse 300 MHz Jeol (E)

10-03-08

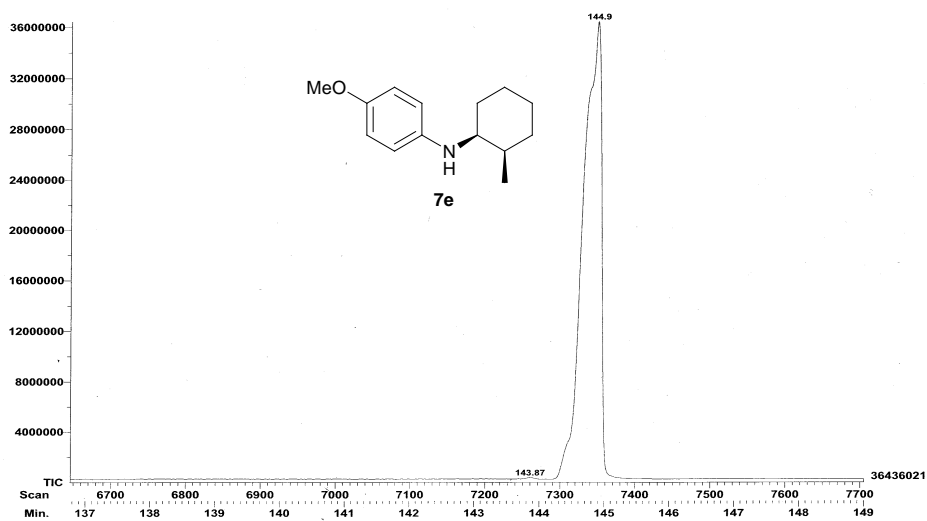
No. de registro: 0957



File: 1214-REACC341
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 06-01-2009 (Time Run: 11:53:06)

Ionization mode: EI+



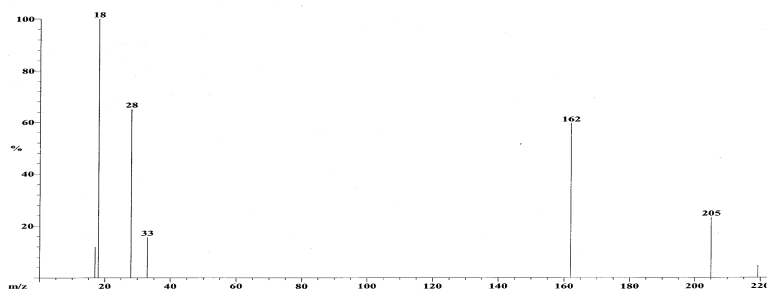
File: 1214-REACC341
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 06-01-2009 (Time Run: 11:53:06)

Ionization mode: EI+

Scan: 7260 R.T.: 143.87
 Base: m/z 18; 3.3%FS TIC: 383712

#Ions: 6



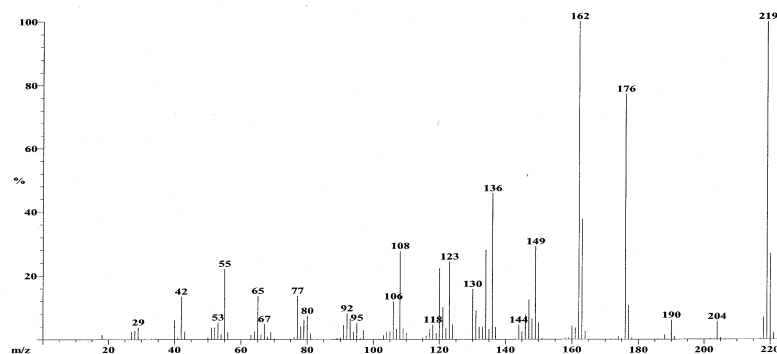
File: 1214-REACC341
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 06-01-2009 (Time Run: 11:53:06)

Ionization mode: EI+

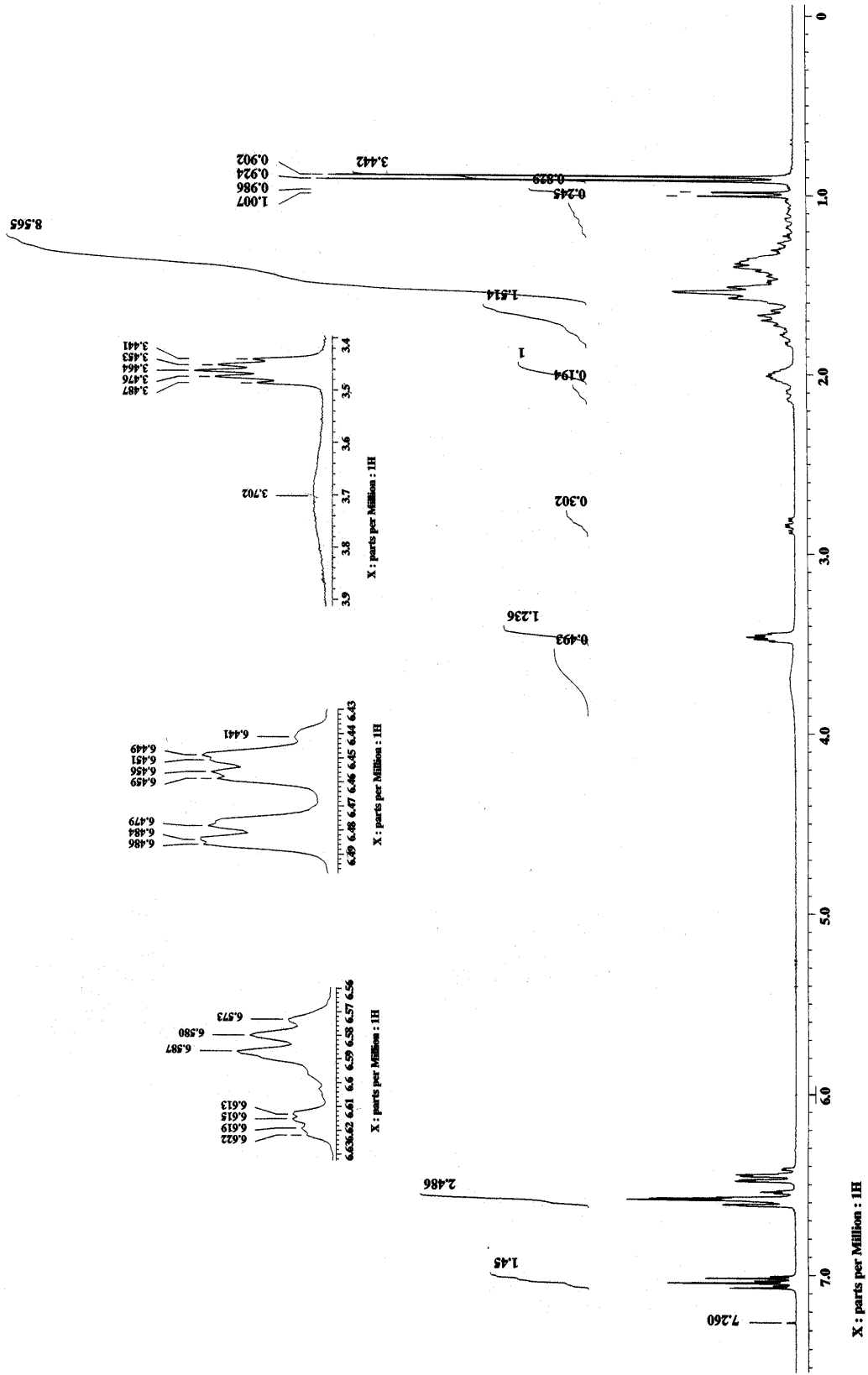
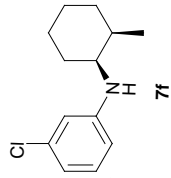
Scan: 7350 R.T.: 144.9
 Base: m/z 219; 99.6%FS TIC: 36824000

#Ions: 124



Peak Label	RIC	Scan Range	Baseline at	Total Area	Background	Peak Area
143.87	TIC	7247-7283	7235-7239	7461277	6447096	1014181
144.9	TIC	7324-7360	7372-7376	637084535	10078603	627005932

INSTITUTO DE QUIMICA, UNAM/EHS
 Dr. A. Cabrera/Laura R. P.
 Clave: Reacc519F22
 Disolvente: CDCl3
 Hidrogeno-1
 Eclipse 300 MHz Jeol (E)
 30-01-09
 No. de registro: 0343



INSTITUTO DE QUIMICA, UNAM/EHS

Dr. A. Cabrera/Laura R. P.

Clave: Reacc519F22

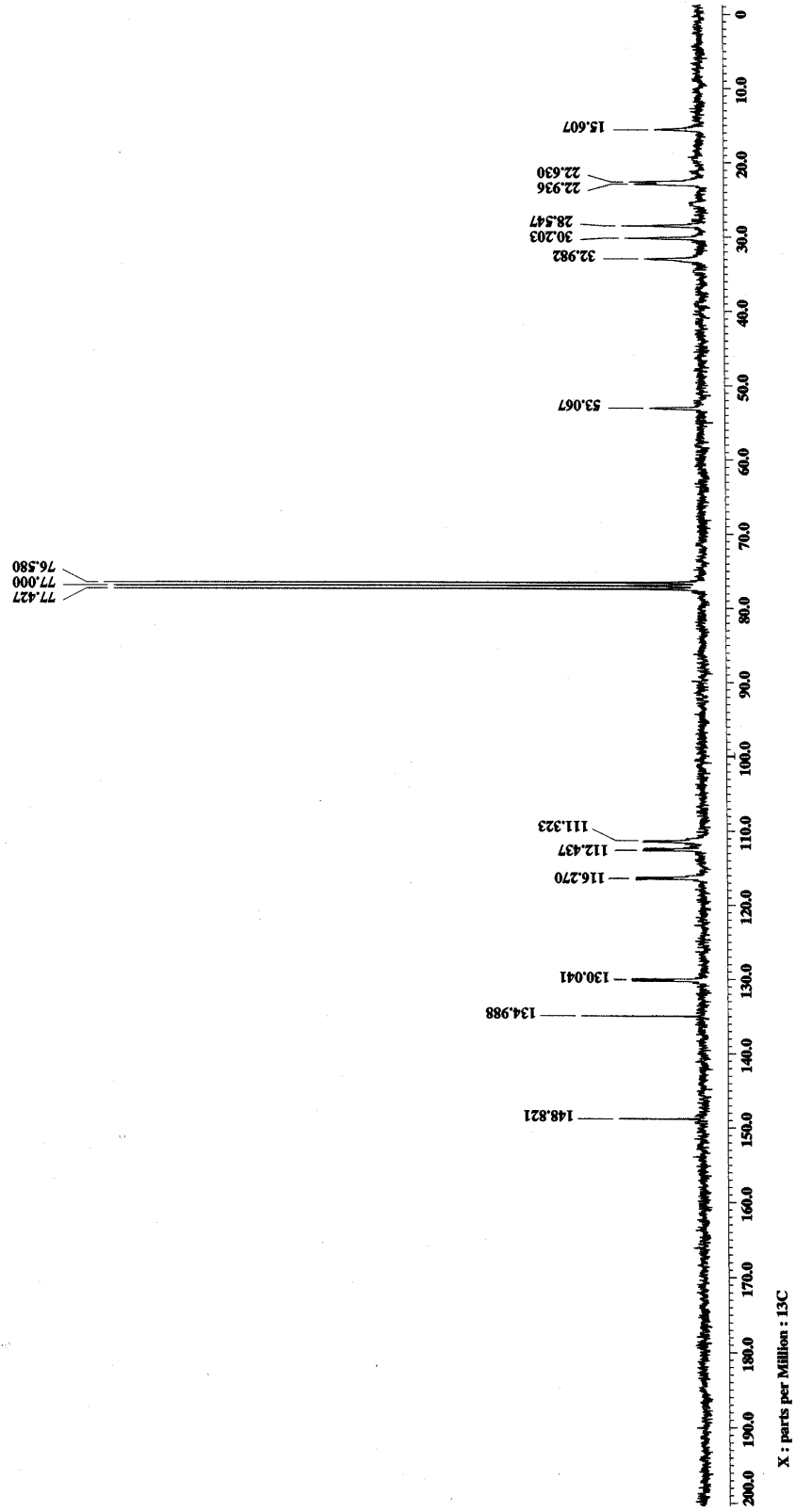
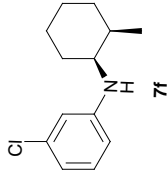
Disolvente: CDCl₃

Carbono-13

Eclipse 300 MHz Jeol (E)

30-01-09

No. de registro: 0343



[TIC]

Data : Dr-Cabrera-Armando-944
Sample: 219 G reacc 519 AX505HA

Date : 29-Jan-120 15:20

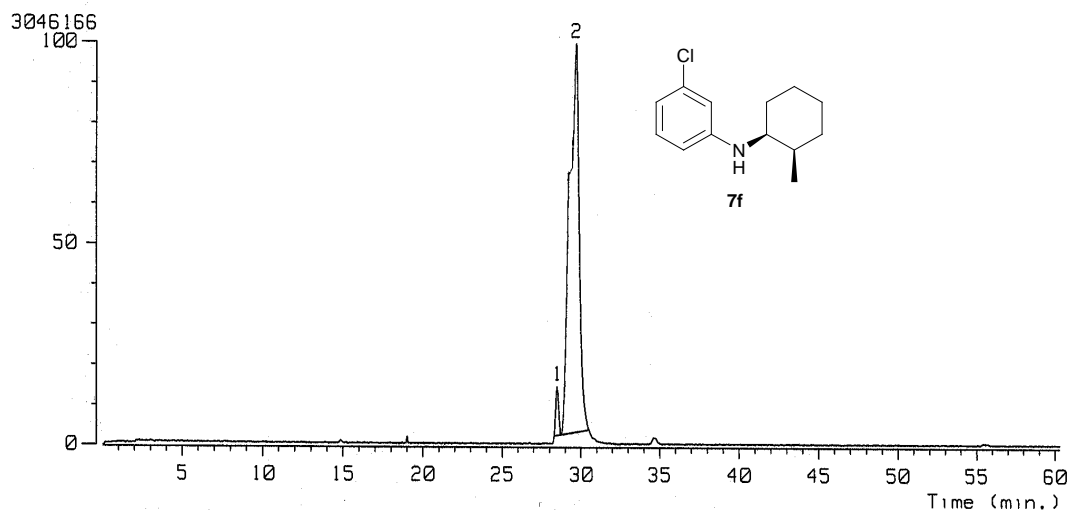
Note :

Inlet : GC

Ion Mode : EI+

Ion Species : Normal Ion

TIC Range : m/z 5 to 650



No.	RT [min]	Area	Area%	Height	Height%	Width [sec]	INTEG
1	28.48	443.20	3.40	35.05	11.12	11.87	BV
2	29.72	12604.04	96.60	280.19	88.88	42.24	VB

[TIC]

Data : Dr-Cabrera-Armando-944
Sample: 219 G reacc 519 AX505HA

Date : 29-Jan-120 15:20

Note :

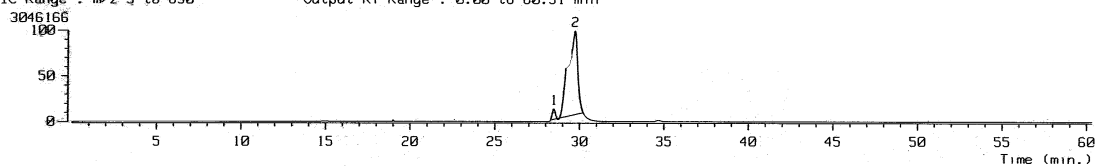
Inlet : GC

Ion Mode : EI+

Ion Species : Normal Ion [MF-Linear]

TIC Range : m/z 5 to 650

Output RT Range : 0.00 to 60.31 min



[Mass Spectrum]

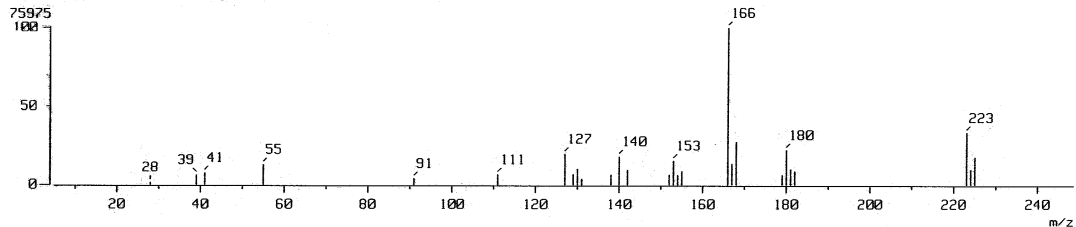
RT : 28.48 min

Scan# : 2134-2120-2264

Temp : 0.0 deg.C

Ion Mode : EI+

Int. : 7.19



[Mass Spectrum]

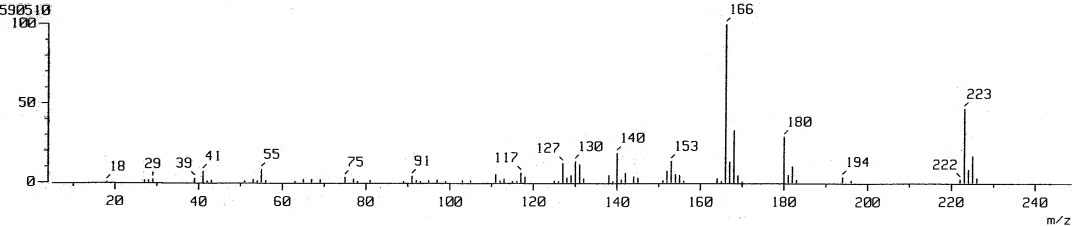
RT : 29.72 min

Scan# : 2227-2120-2264

Temp : 0.0 deg.C

Ion Mode : EI+

Int. : 56.32



Reacc362-1H-6.jdf



```

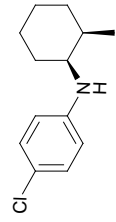
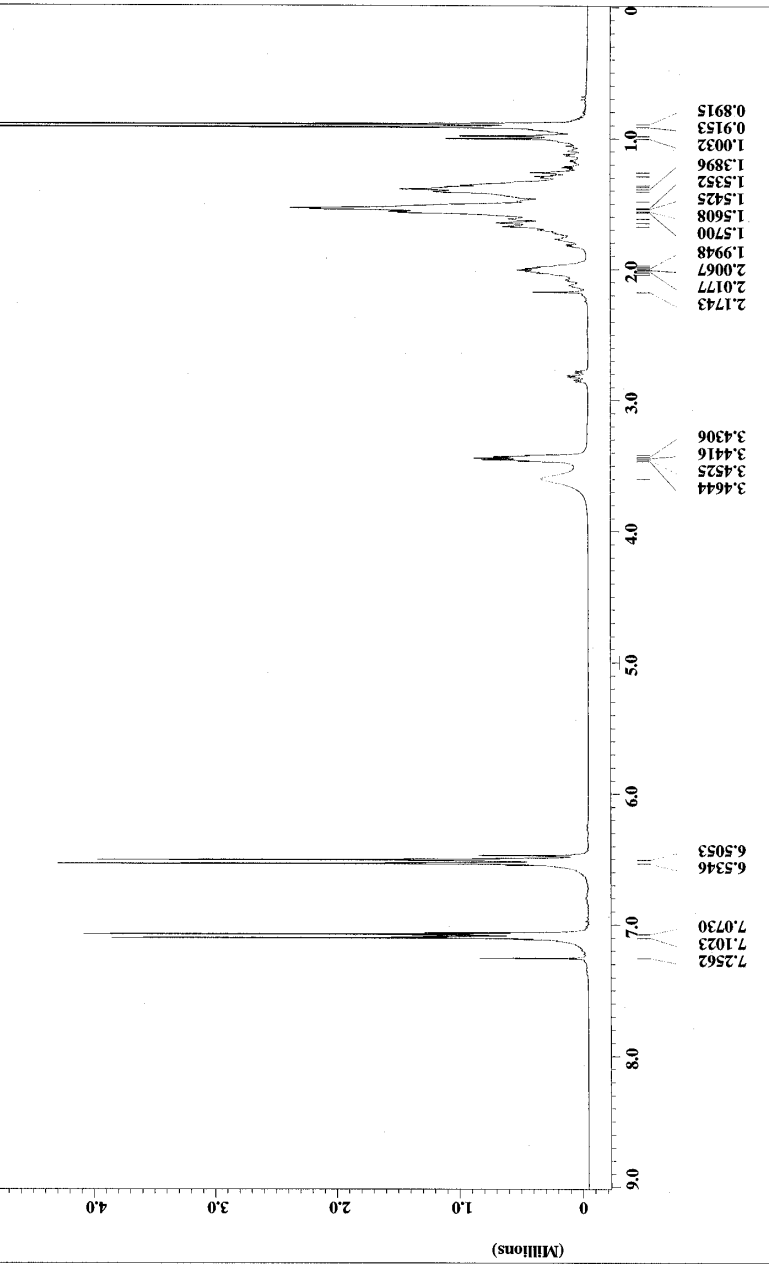
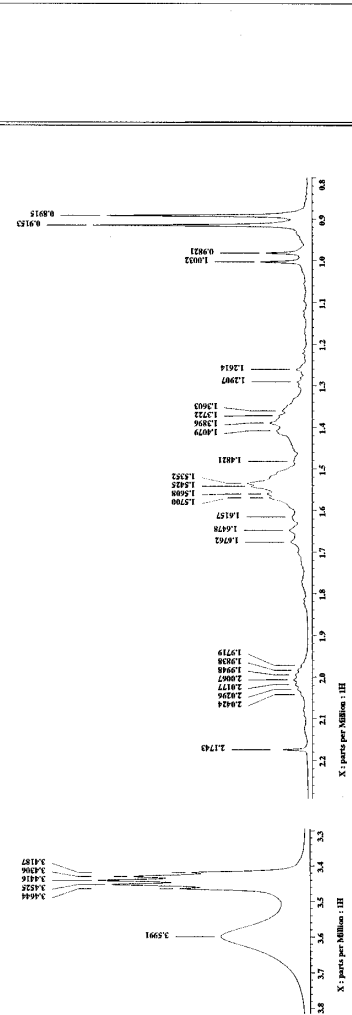
Filename = Reacc362-1H-6.jdf
Author = Cabrera
Experiment = single_pulse.exp
Sample_id = Laura
Solvent = CHLOROFORM-D
Creation_time = 11-FEB-2008 22:21:01
Revision_time = 19-DEC-2008 13:44:01
Current_time = 19-DEC-2008 13:45:08

Comment = Single Pulse Experiment
Data_format = 1D COMPLEX
Dir_size = 16384
Dir_title = 1H
Dim_units = [ppm]
Dimensions = X
Site = Eclipse+ 300
Spectrometer = DELTA_NMR

Field_strength = 7.0586013[T] (300[MHz]
X_acq_duration = 3.6339712[s]
X_domain = 1H
X_freq = 300.52965592[MHz]
X_offset = 5[ppm]
X_points = 16384
X_prescans = 0
X_resolution = 0.27518105[Hz]
X_sweep = 4.50856628[MHz]
Clipped = FALSE
Mod_return = 1
Scans = 32
Total_scans = 32

X_90_width = 9.5[us]
X_acq_time = 3.6339712[s]
X_angle = 45[deg]
X_pulse = 4.75[us]
Initial_wait = 1[s]
Phase_preset = 3[us]
Recvr_gain = 17
Relaxation_delay = 1[s]
Temp_get = 19.5[dC]
Unblank_time = 2[us]

```



79

X: parts per Million : 1H

Reacc362-13C-4.jdf



```

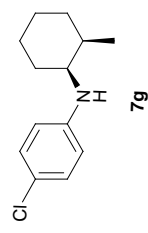
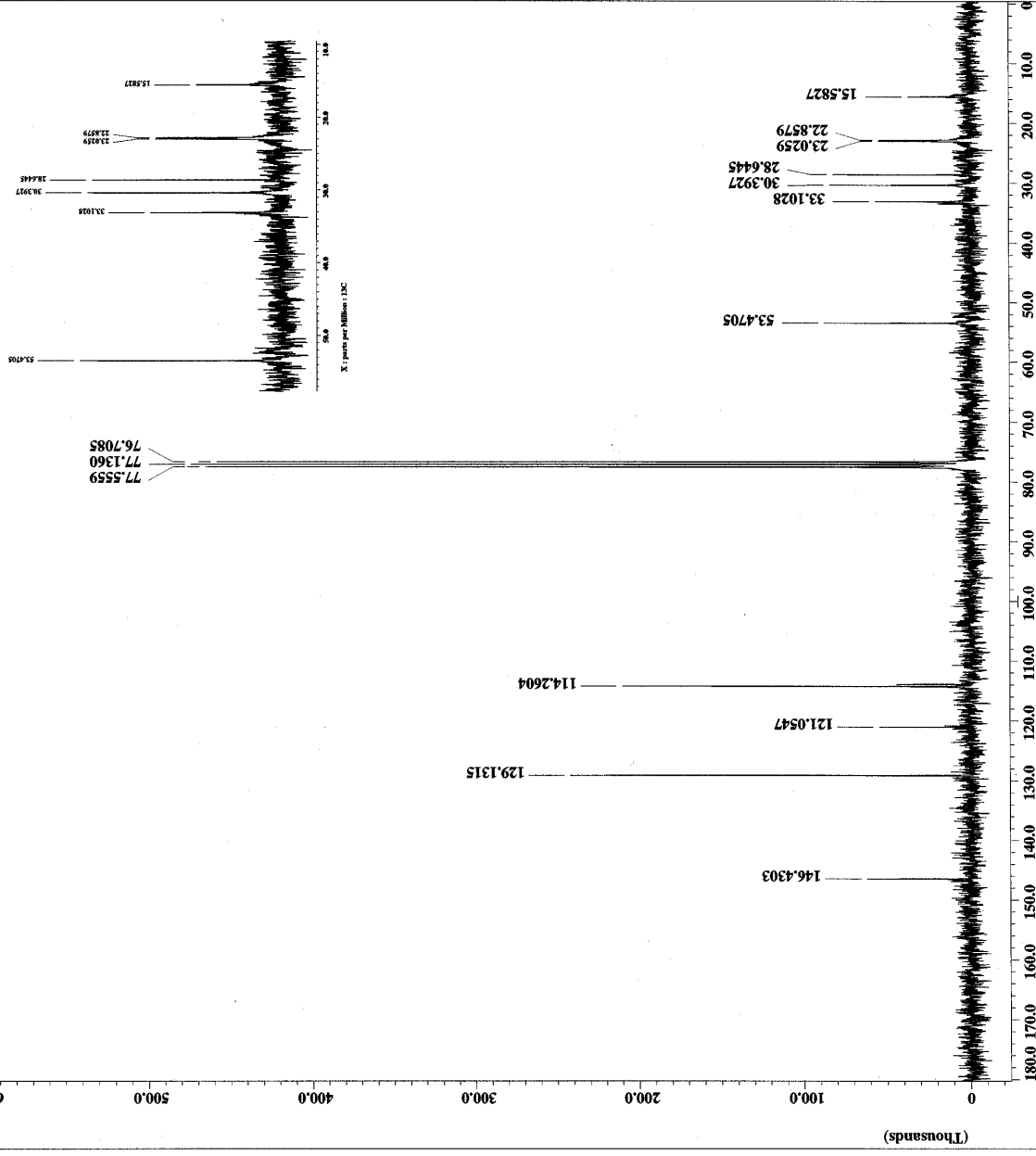
File name      = Reacc362-13C-4.jdf
Author        = Cabrera
Experiment    = single_pulse_dec
Sample ID     = Laura
Solvent       = CHLOROFORM-D
Creation time = 11-FEB-2008 23:22:45
Revision time = 19-DEC-2008 13:47:17
Current time  = 19-DEC-2008 13:48:11

Comment       = Single Pulse with Bro
Data format   = ID COMPLEX
Data size     = 32768
Dim title     = 13C
Dim units     = [ppm]
Dimensions    = X
Site          = Eclipse 300
Spectrometer  = DELTA_NMR

Field strength = 7.0586013 [T] (300 [MHz])
X_acq duration = 1.7334272 [s]
X_domain       = 75.6823426 [MHz]
X_freq         = 101 [ppm]
X_offset       = 32768
X_points       = 4
X_resolution   = 0.57689184 [Hz]
X_sweep        = 18.90359168 [MHz]
Irr_domain     = 1H
Irr_freq       = 300.52965592 [MHz]
Irr_offset     = 5 [ppm]
Clipped        = FALSE
Mod_return     = 1
Scans          = 1329
Total_scans    = 1329

X_90 width    = 11.3 [us]
X_acq time    = 1.7334272 [s]
X_angle       = 30 [deg]
X_pulse       = 3.76666667 [us]
Initial_wait  = 1 [s]
Phase_preset  = 3 [us]
Recvr_gain    = 30
Relaxation_delay = 1 [s]
Temp_get      = 21.2 [dC]
Unblank_time  = 2 [us]

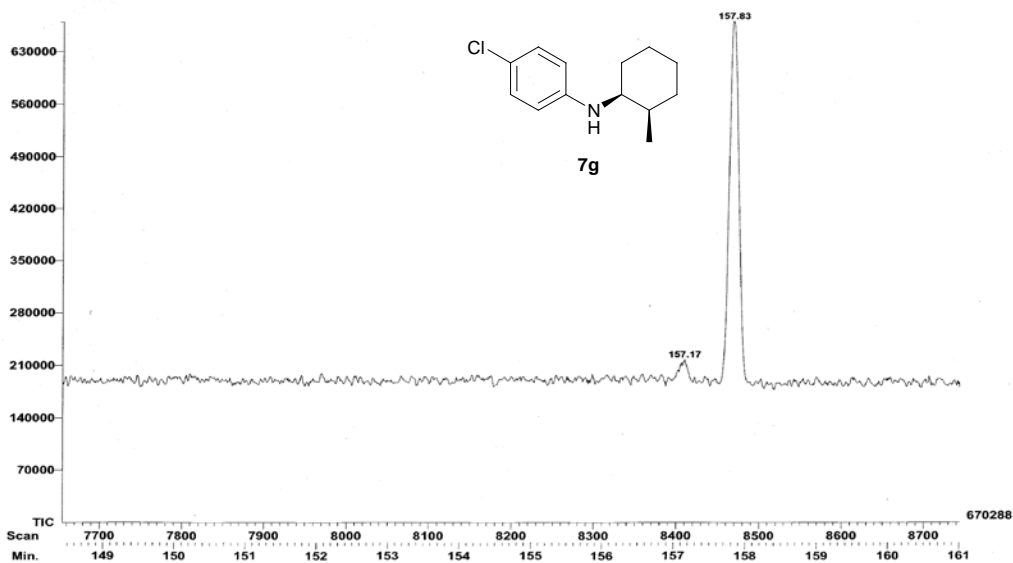
```



File: 1211-REACC362
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-28-2009 (Time Run: 19:16:44)

Ionization mode: EI+



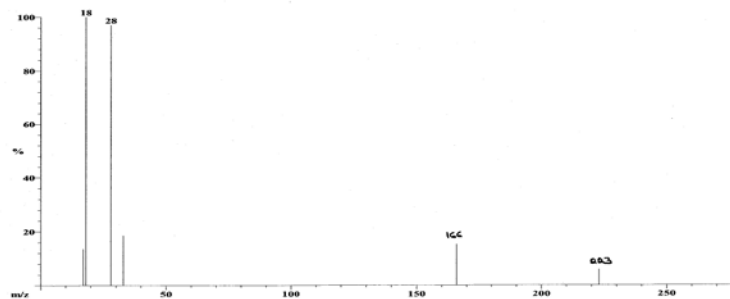
File: 1211-REACC362
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-28-2009 (Time Run: 19:16:44)

Ionization mode: EI+

Scan: 8410 R.T.: 157.17
 Base: m/z 18; 2%FS TIC: 212576

#Ions: 6



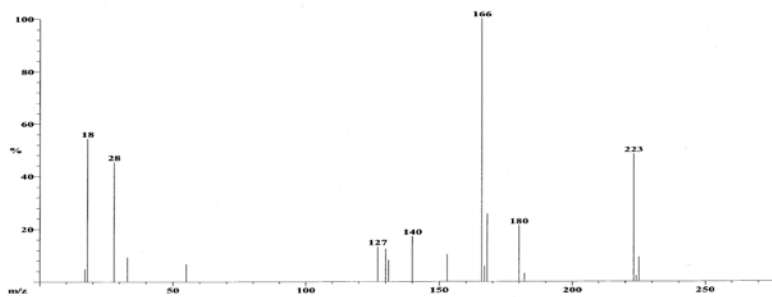
File: 1211-REACC362
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-28-2009 (Time Run: 19:16:44)

Ionization mode: EI+

Scan: 8468 R.T.: 157.83
 Base: m/z 166; 4%FS TIC: 671136

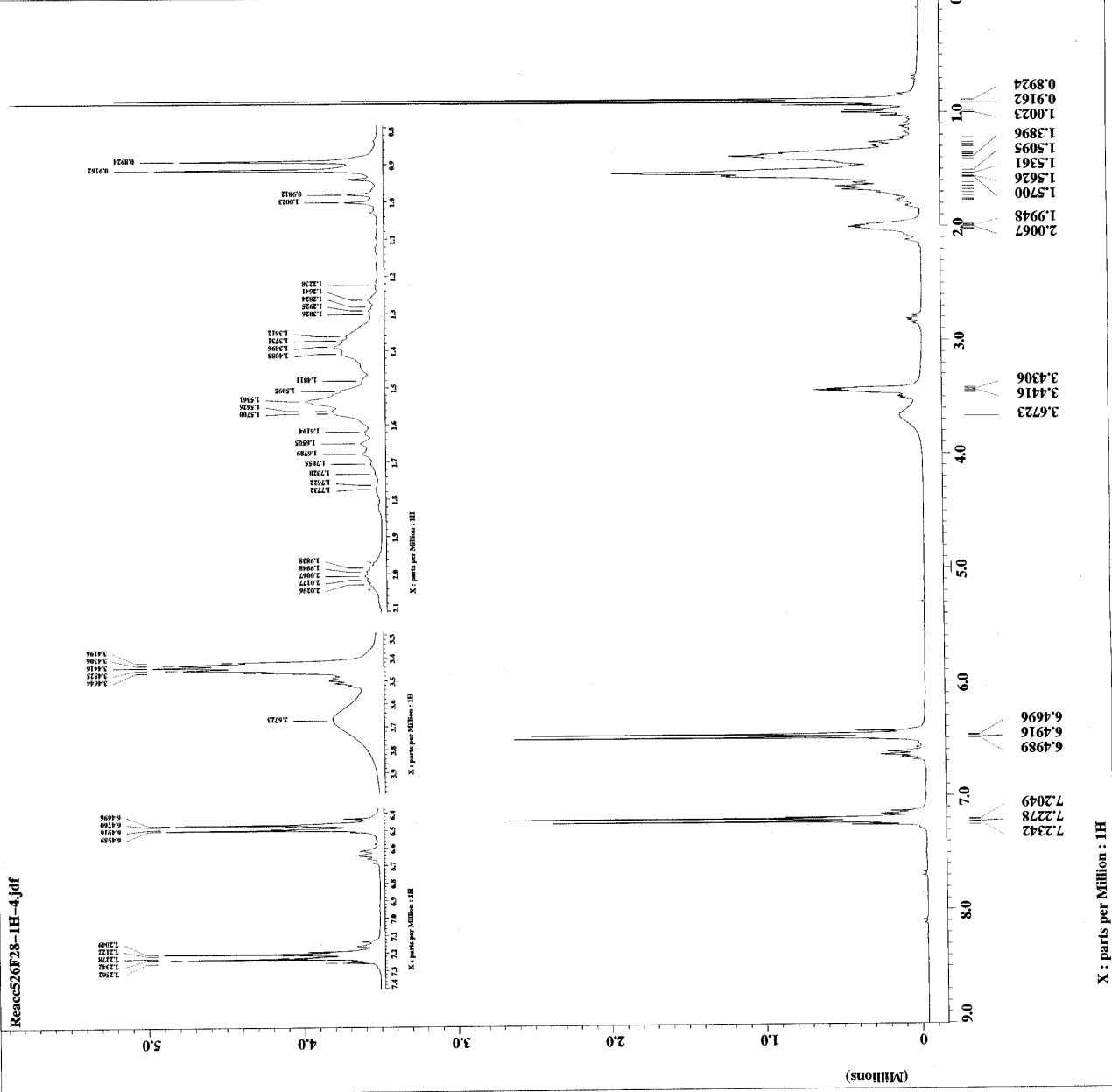
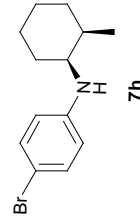
#Ions: 18



Peak Table

Peak Label	RIC	Scan Range	Baseline at	Total Area	Background	Peak Area
157.17	TIC	8397-8423	8385-8389	3629883	3379220	250663
157.83	TIC	8455-8481	8490-8494	7812256	3305930	4506326

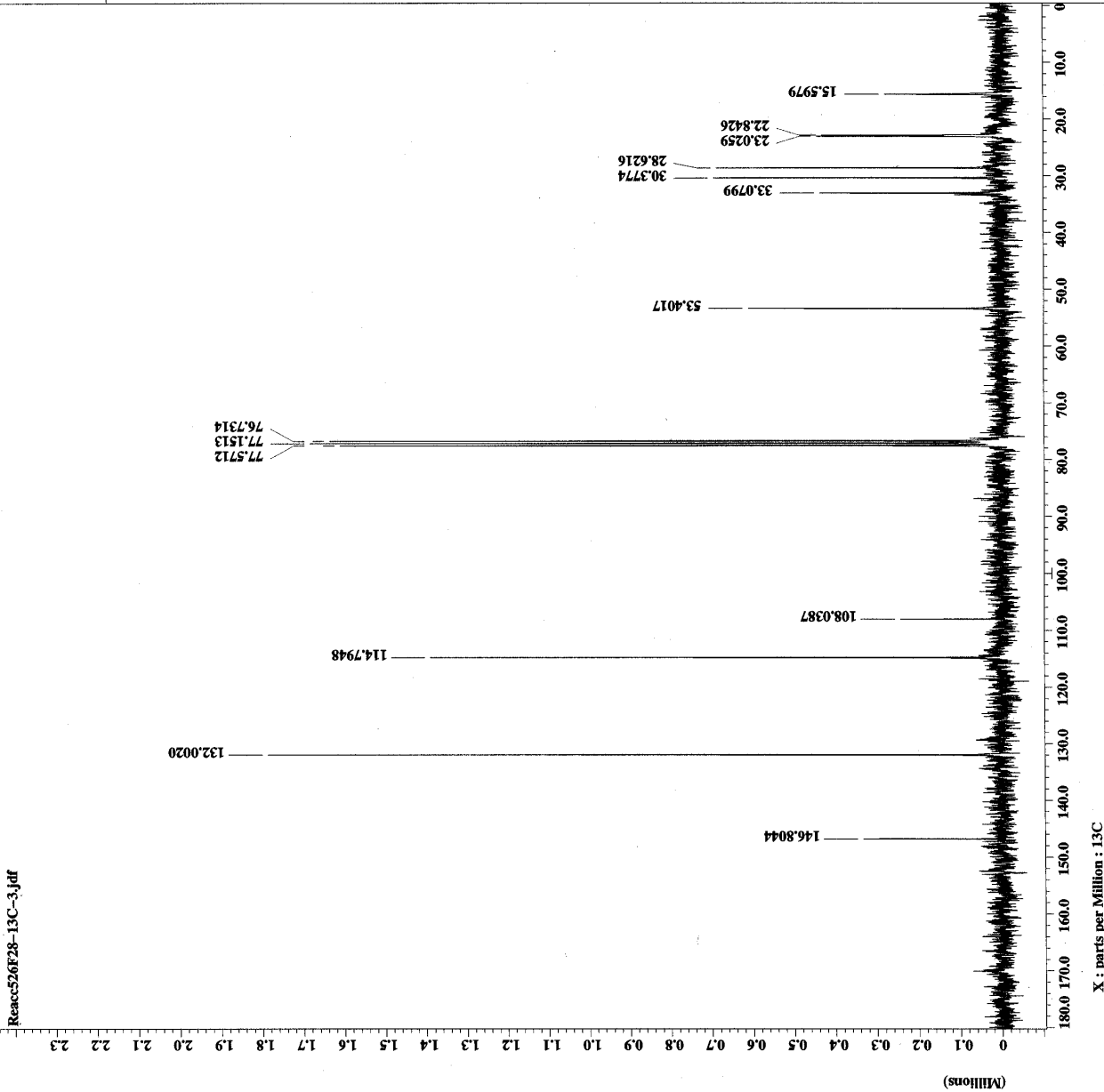
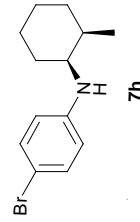
File Name = Reacc526F28-1H-4.jdf
Author = Cabrera
Experiment = single_pulse.exp
Sample_id = Laura
Solvent = CHLOROFORM-D
Creation_time = 9-DEC-2008 02:27:29
Revision_time = 20-FEB-2009 11:34:52
Current_time = 20-FEB-2009 11:36:25
Comment = Single Pulse Experiment
Data_format = 1D COMPLEX
Dim_size = 16384
Dim_title = 1H
Dim_units = [ppm]
Dimensions = X
Site = Eclipse+ 300
Spectrometer = DELTA_NMR
Field_strength = 7.0586013 [T] (300 [MHZ])
X_domain = 1H
X_acq_duration = 3.6339712 [s]
X_freq = 300.52965592 [MHZ]
X_offset = 7 [ppm]
X_points = 16384
X_prescans = 0
X_resolution = 0.27518105 [Hz]
X_sweep = 4.50856628 [kHz]
C1_type = FALSE
Mod_return = 1
Scans = 32
Total_scans = 32
X_90_width = 9.5 [us]
X_acq_time = 3.6339712 [s]
X_angle = 45 [deg]
X_pulse = 4.75 [us]
Initial_wait = 1 [s]
Phase_preset = 3 [us]
Recvr_gain = 17
Relaxation_delay = 1 [s]
Temp_get = 18.9 [dC]
Unblank_time = 2 [us]



X : parts per Million : 1H

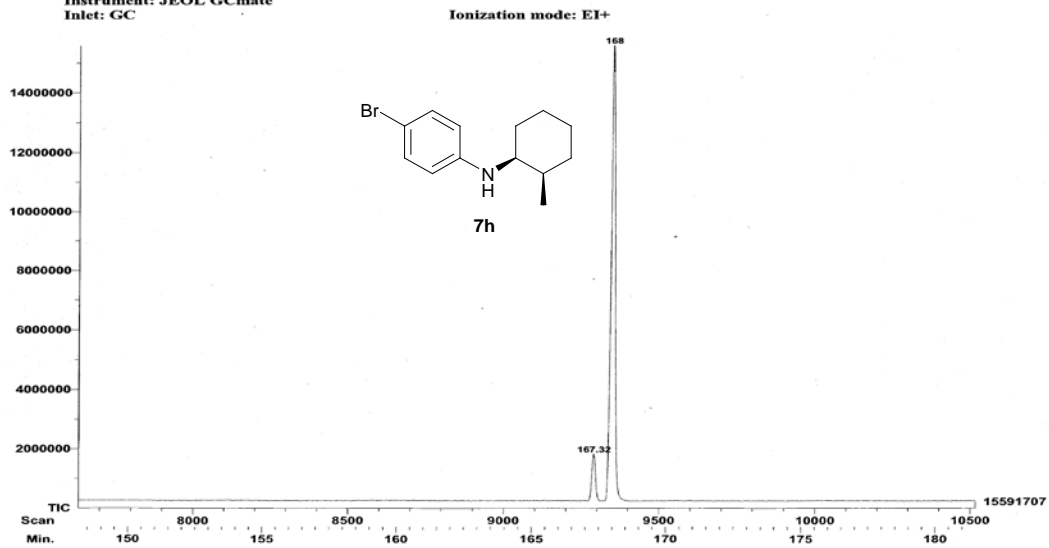


React526F28-13C-3.jdf
Filename = React526F28-13C-3.jdf
Author = Cabrera
Experiment = single_pulse_dec
Sample_id = Laura
Solvent = CHLOROFORM-D
Creation_Time = 9-DEC-2008 02:35:35
Revision_Time = 20-FEB-2009 10:41:54
Current_Time = 20-FEB-2009 11:19:36
Comment = Single Pulse with Bro
Data_format = ID COMPLEX
Data_size = 32768
Data_title = 13C
Data_units = ppm
Dimensions = 1
Spectrometer = Eclipse+ 300
Spectrometer = DELTA_NMR
Field_strength = 7.05860131[F] (300 [MHz]
X_acq_duration = 1.7334272[s]
X_domain = 13C
X_freq = 75.56823426[MHz]
X_offset = 100 [ppm]
X_points = 32768
X_prescans = 4
X_resolution = 0.57689184 [Hz]
X_sweep = 18.90359168 [kHz]
Irr_domain = 1H
Irr_freq = 300.52965592 [MHz]
Irr_offset = 5 [ppm]
Clipped = FALSE
Mod_return = 1
Scans = 150
Total_scans = 150
X_90_width = 11.3 [us]
X_acq_time = 1.7334272 [s]
X_angle = 30.734272 [s]
X_delay = 3.76696667 [us]
X_pulse_wait = 1 [s]
Initial_wait = 3 [us]
Phase_preset = 30
Recvr_gain = 30
Relaxation_delay = 1 [s]
Temp_get = 20.9 [dC]
Unblank_time = 2 [us]



File: 1204-Reac526
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-27-2009 (Time Run: 14:07:47)

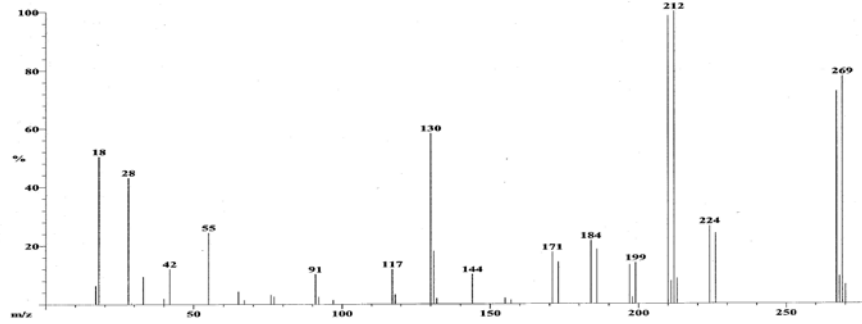


File: 1204-Reac526
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-27-2009 (Time Run: 14:07:47)

Scan: 9290 R.T.: 167.32
 Base: m/z 212; 5.3%FS TIC: 1814032

#Ions: 41

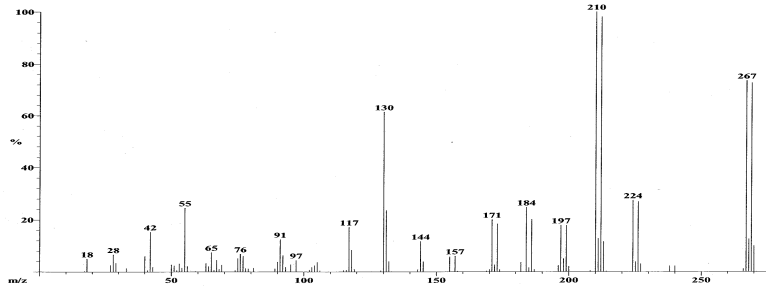


File: 1204-Reac526
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-27-2009 (Time Run: 14:07:47)

Scan: 9349 R.T.: 168
 Base: m/z 210; 41%FS TIC: 15824944

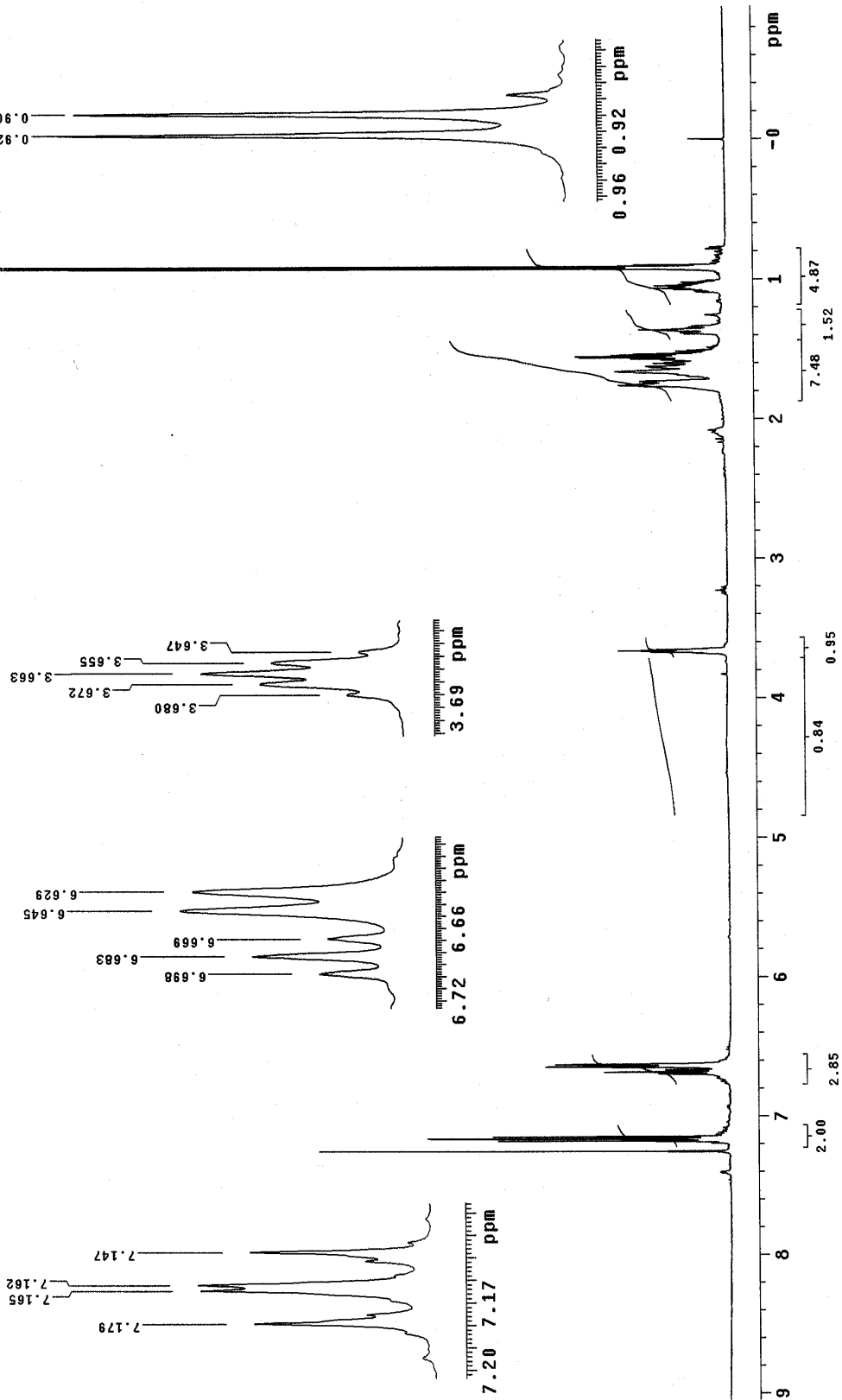
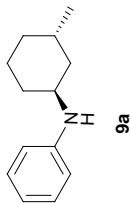
#Ions: 103



Peak Table

Peak Label	RIC	Scan Range	Baseline at	Total Area	Background	Peak Area
157.17	TIC	8397-8423	8385-8389	3629883	3379220	250663
157.83	TIC	8455-8481	8490-8494	7812256	3305930	4506326

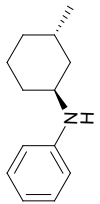
U.N.A.M. Instituto de Química ICH
 Dr. A. Cabrera/Laura R. P
 Clave:Reacc52F30
 Disolvente:CDCl3
 Experimento:IH
 Varian Inova IH 500 MHz (G)
 No. de registro 0834
 20-05-09
 Pulse Sequence: s2pu1



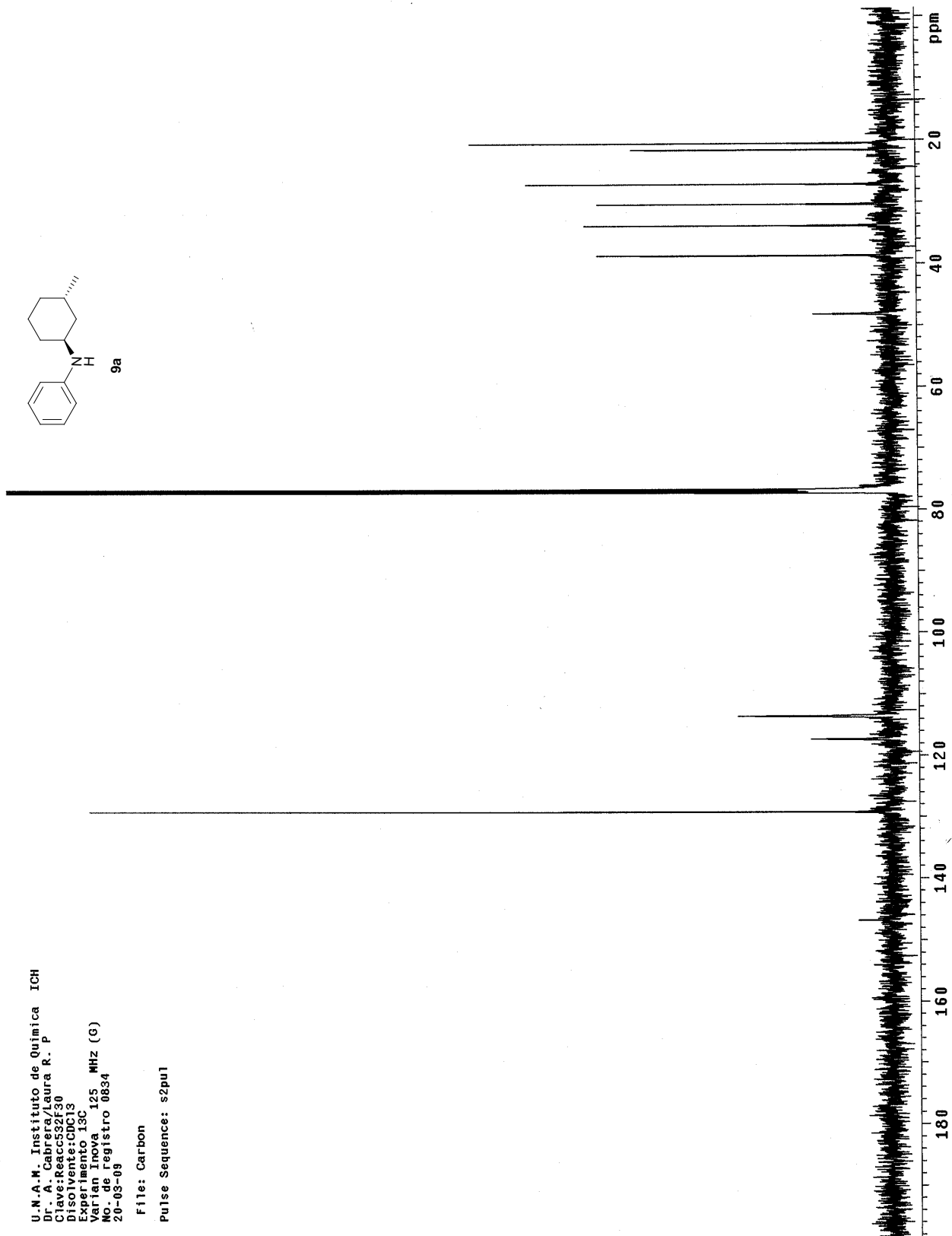
U.N.A.M. Instituto de Química ICH
Dr. A. Cabera/Laura R. P.
Clave: Peacc52f30
Disolvente: CDCl3
Experimento: 13C
Varian Inova 125 MHz (G)
No. de registro 0834
20-03-09

File: Carbon

Pulse Sequence: s2pul

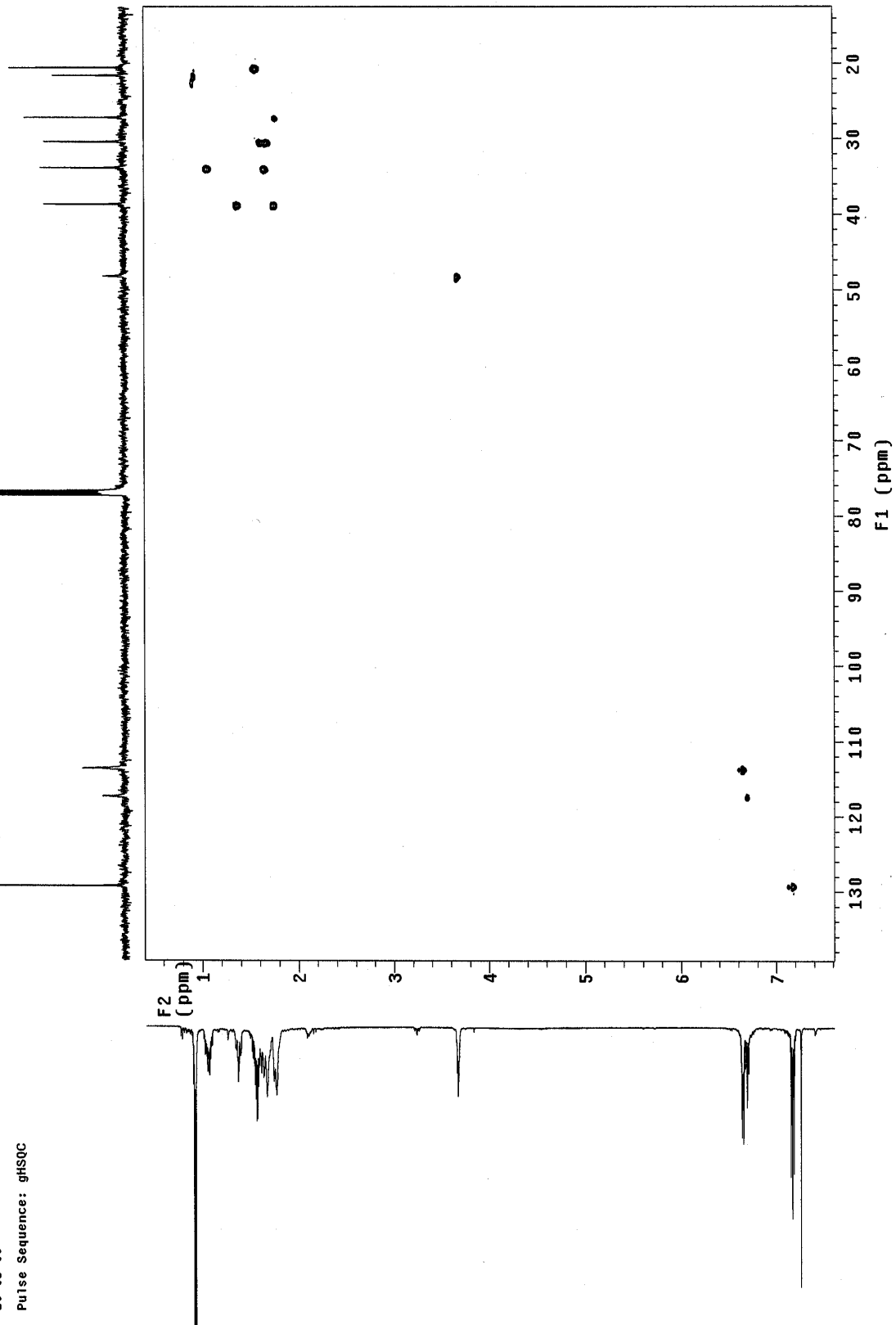
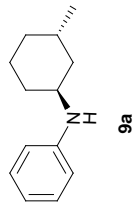


9a

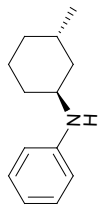


U.N.A.M. Instituto de Química ICH
Dr. A. Cabreriz/Saura R. P
Clave:Reacc552130
Solvente:CDCl3
Experimento:HSQC
Variate:Proa 500 MHz (G)
Número de Registro 0834
20-03-09

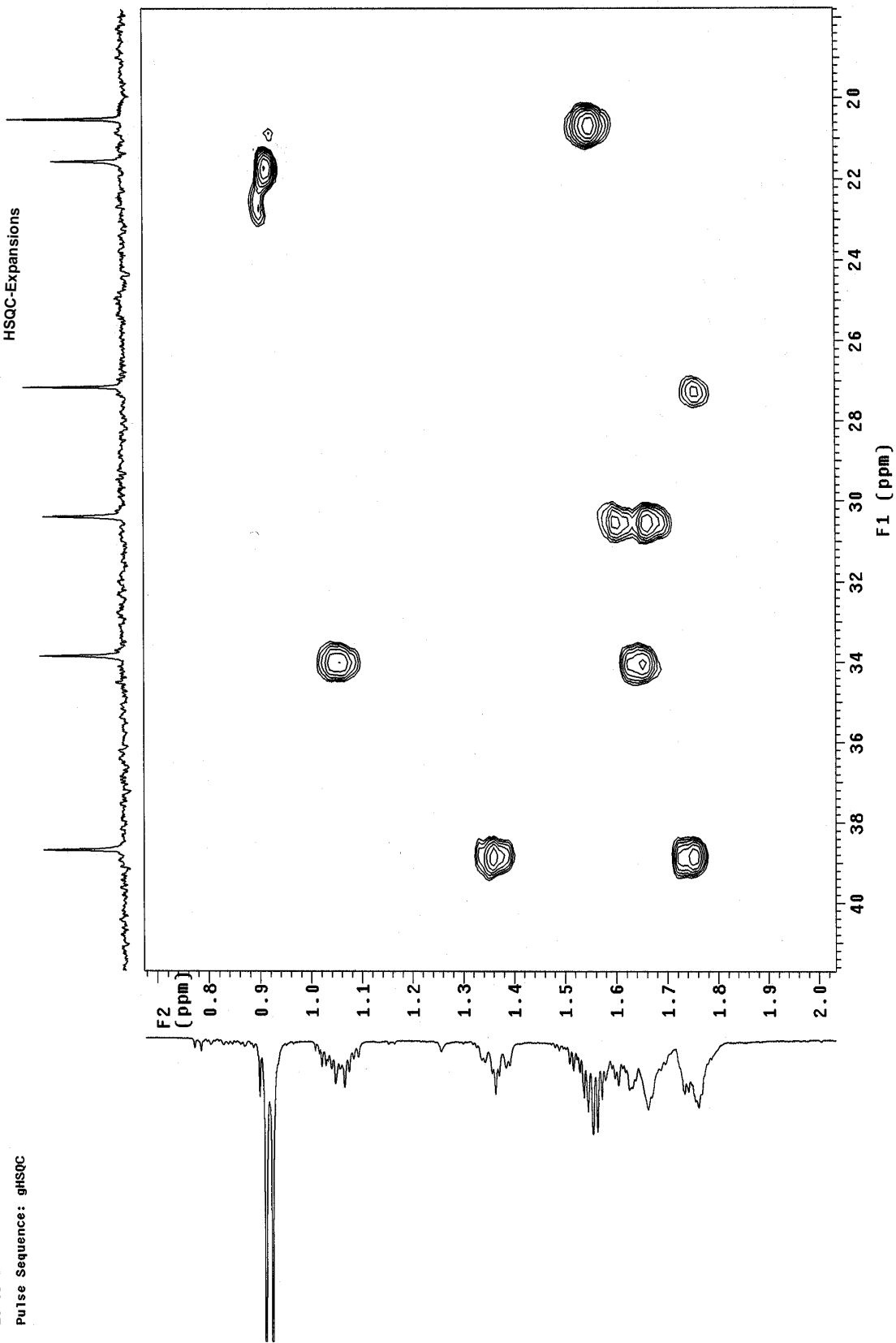
Pulse Sequence: gHSQC



U.N.A.M. Instituto de Química ICH
Dr. A. Cabrera/Laura R. P
Clave:Reacc32f30
Disolvente:CDCl3
Experimento HSQC
Marian Inoya_500_MHz (G)
No. de Registro 0834
20-05-03
Pulse Sequence: gHSQC

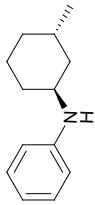


HSQC-Expansions

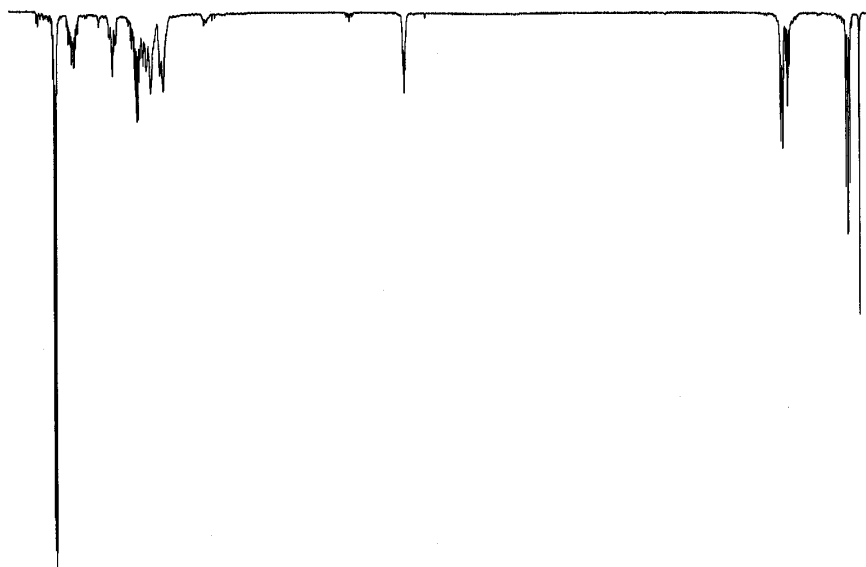
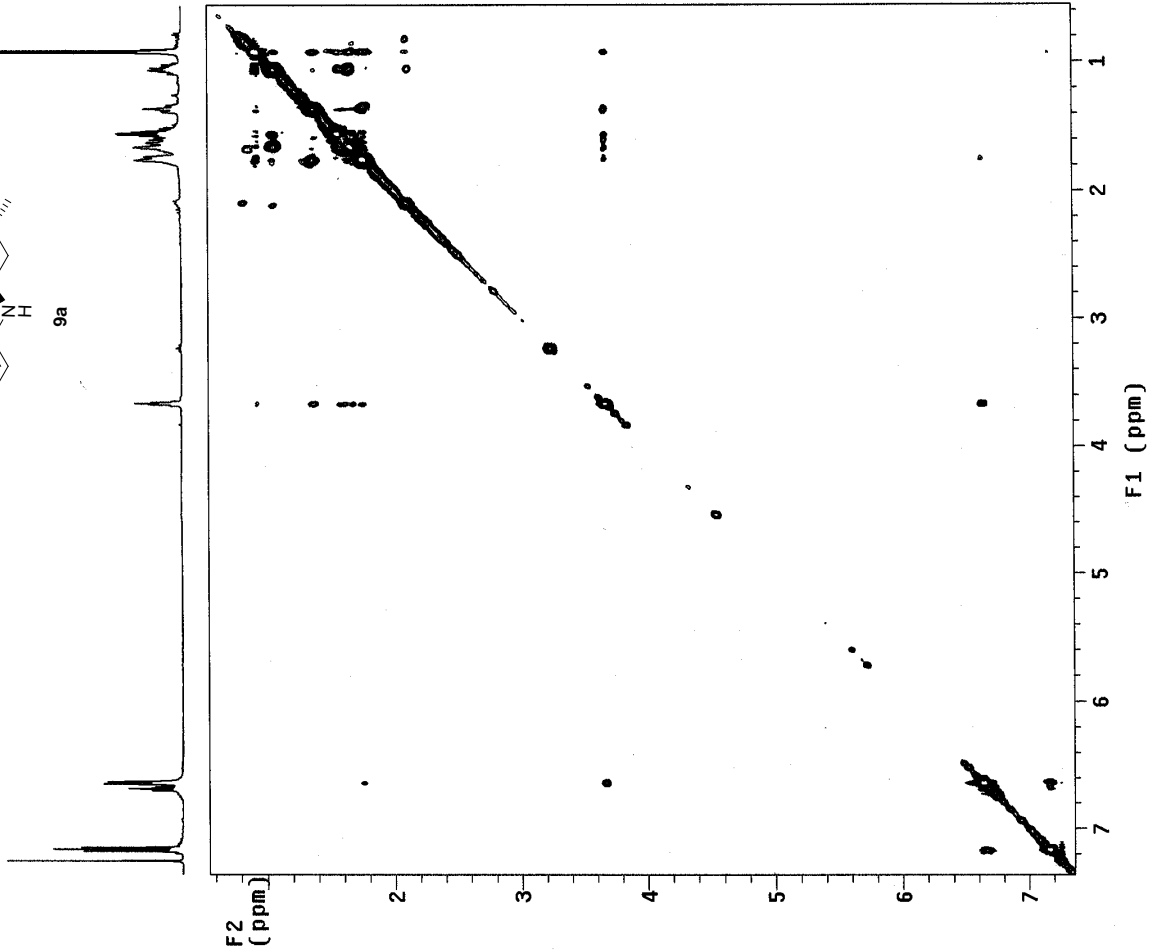


U.N.A.M. Instituto de Quimica ICH
Dr. A. Cabrerizo/Laura R. P
Clave: Reacc52/20
Disolvente: CDCl3
Experimento: NMR15
Varian Proya 500 MHz (G)
No. de registro 0834
20-03-09

Pulse Sequence: noesy

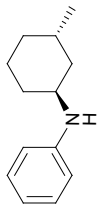


9a



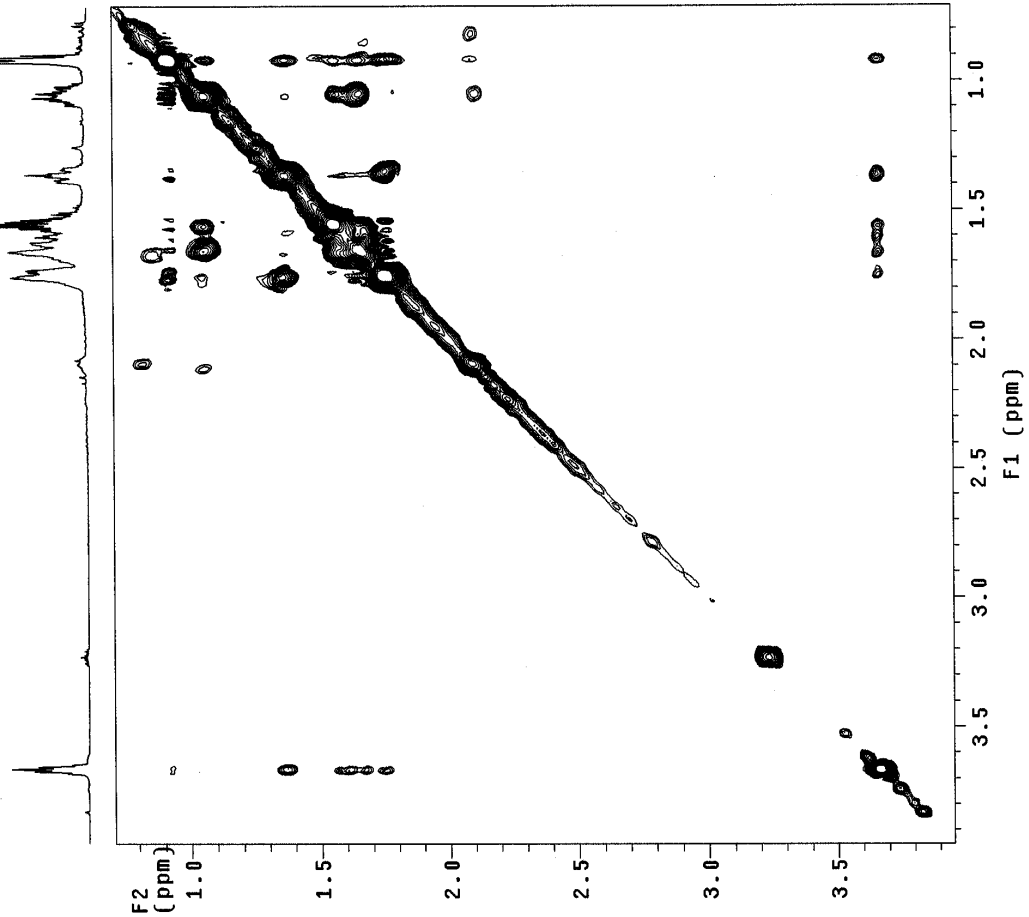
U.N.A.M. Instituto de Química ICH
Dr. A. Cabrera/Laura R. P
Clave:Reacc52F30
Disolvente:CDCl3
Experimento NOESY
Varian Inova 500 MHz (G)
No. de registro 0834
20-03-09

Pulse Sequence: noesy



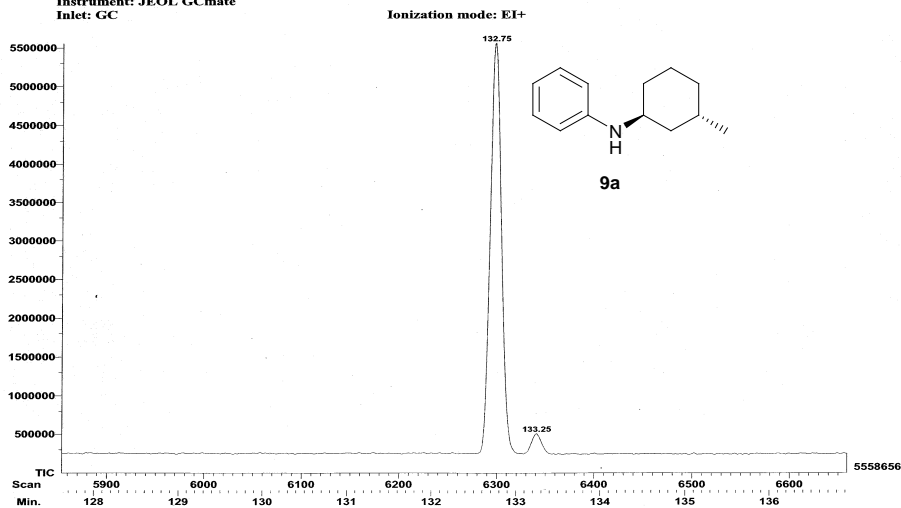
9a

NOESY-Expansions



File: REACC 532
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-26-2009 (Time Run: 13:24:55)

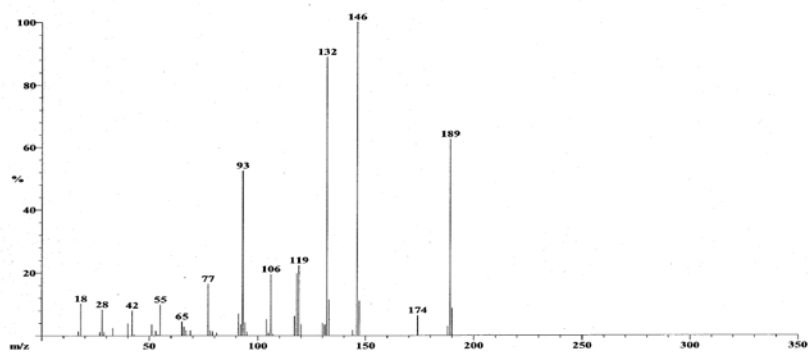


File: REACC 532
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-26-2009 (Time Run: 13:24:55)

Scan: 6298 R.T.: 132.75
 Base: m/z 146; 25%FS TIC: 5622400

#Ions: 48

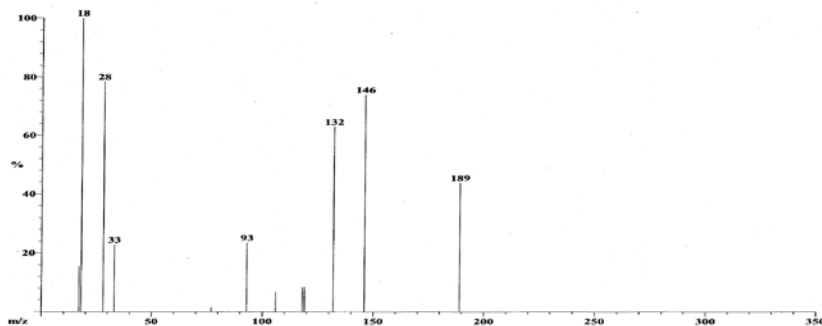


File: REACC 532
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-26-2009 (Time Run: 13:24:55)

Scan: 6341 R.T.: 133.25
 Base: m/z 18; 2.7%FS TIC: 496576

#Ions: 12



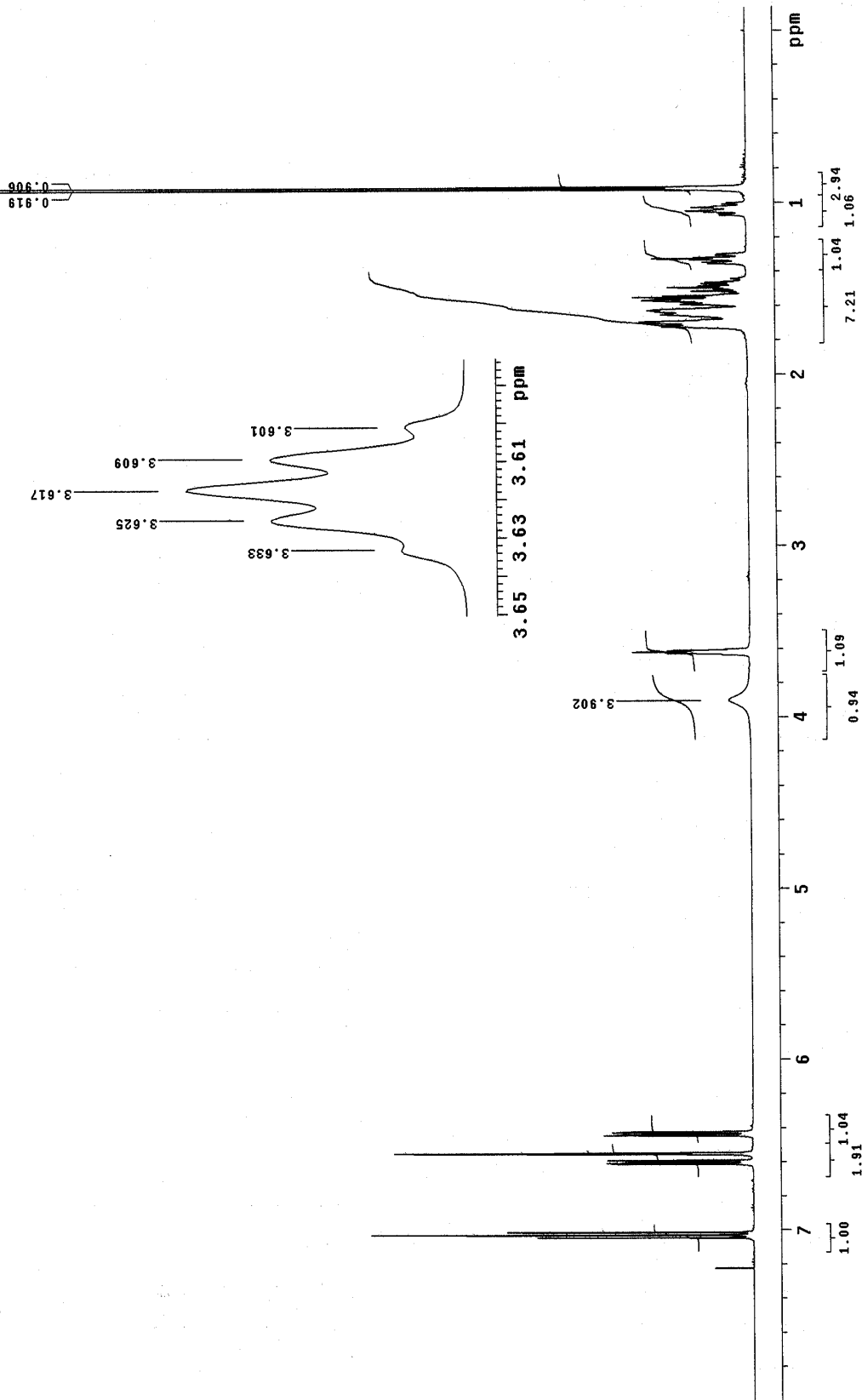
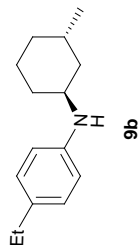
Peak Table

Peak Label	RIC	Scan Range	Baseline at	Total Area	Background	Peak Area
132.75	TIC	6286-6310	6274-6278	51692778	4175630	47517148
133.25	TIC	6329-6353	6360-6364	6197332	4020182	2177150

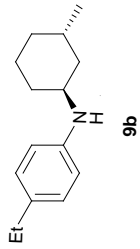
UNAM, Instituto de Quimica. (H. Rios)
 Dr-A-Cabrera/L-537F27
 Clave: seacc-86
 No. Registro: 86
 Experimento: Hidrogeno
 Disolvente: CDCl3
 Variacion: 500MHZ (G)
 Marzo-25-2008

File: std1h

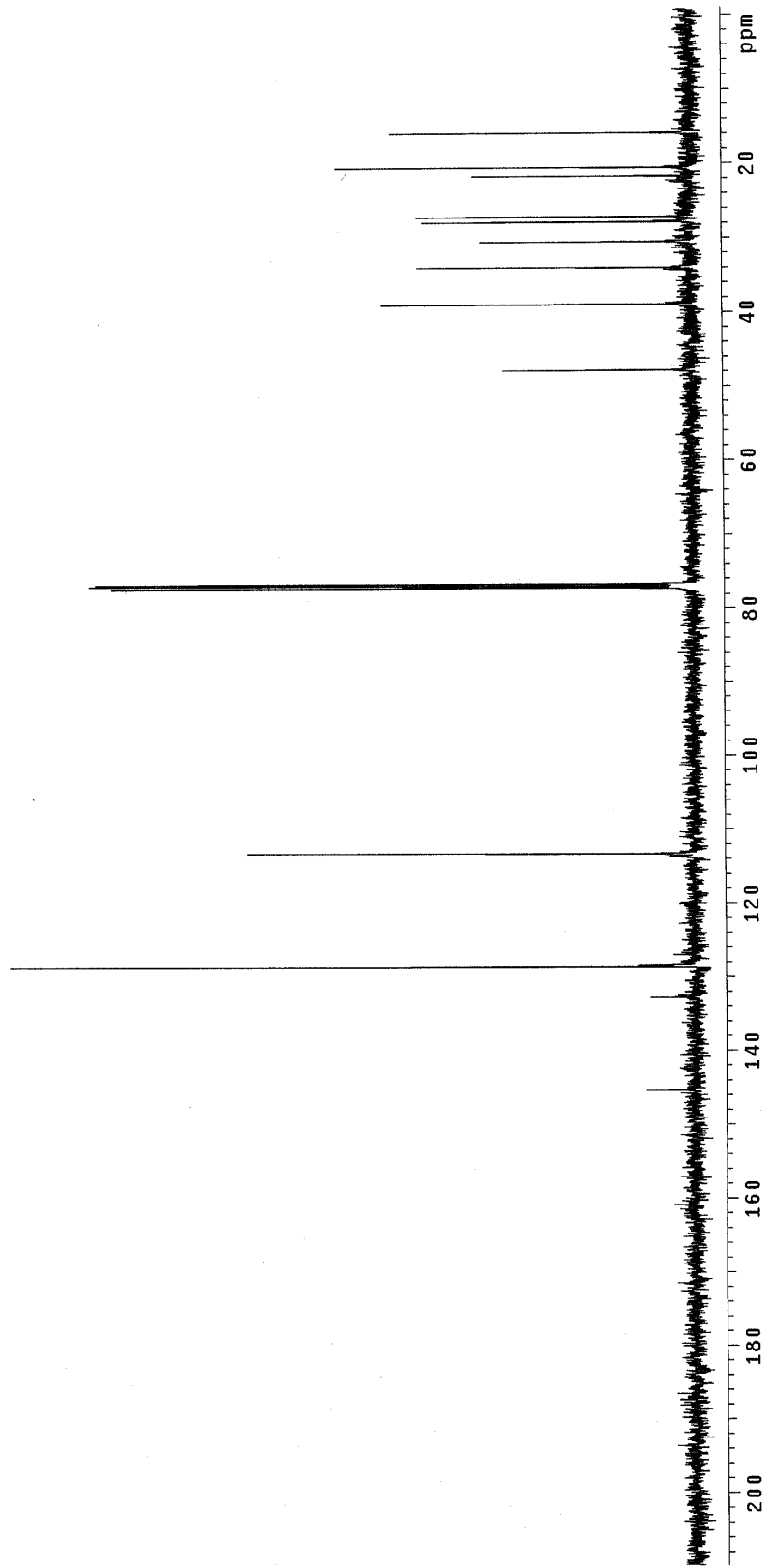
Pulse Sequence: s2pu1



U.N.A.M. Instituto de Química ICH
Dr. A. Cabrera/Laura RP
Clave : Reacc 538F2627
Disolvente: CDCl3
Experimento : 13C
Varian Inova 125 MHz
No. de Registro 1039
23-04-09

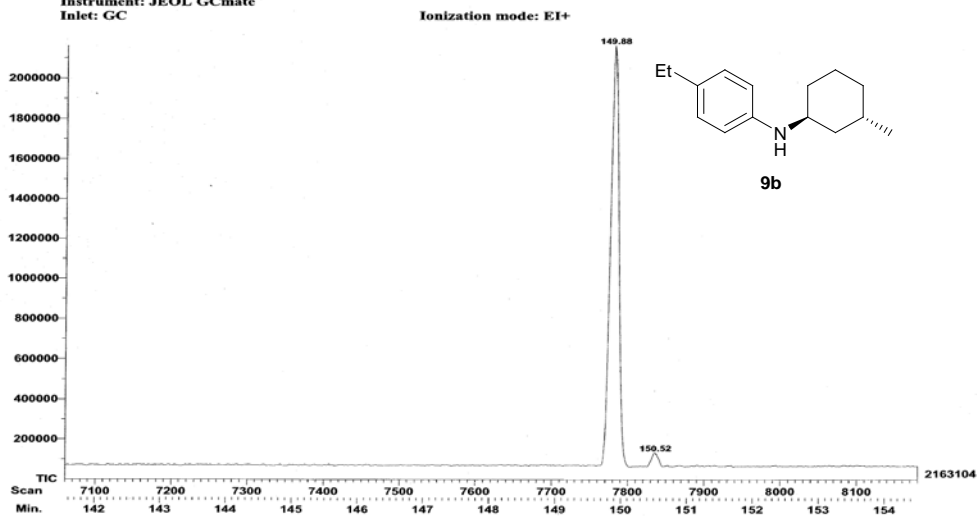


File: Carbon
Pulse Sequence: s2pul



File: 1039-REACC538F
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-25-2009 (Time Run: 12:20:07)

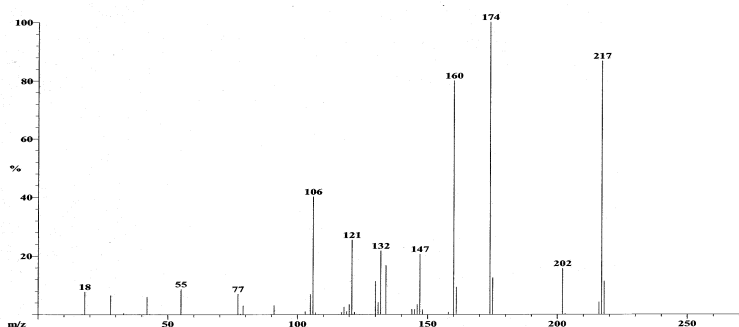


File: 1039-REACC538F
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-25-2009 (Time Run: 12:20:07)

Scan: 7780 R.T.: 149.88
 Base: m/z 174; 9.8%FS TIC: 2192880

#Ions: 3

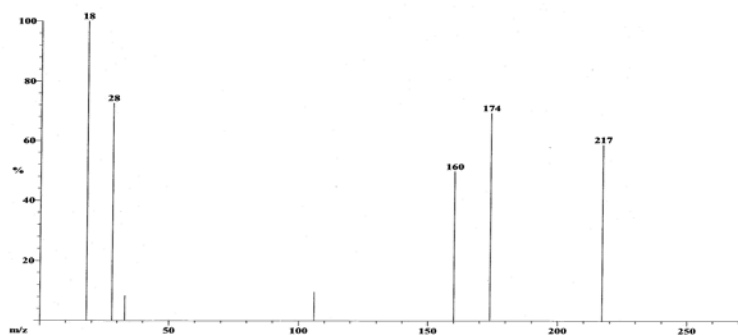


File: 1039-REACC538F
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-25-2009 (Time Run: 12:20:07)

Scan: 7836 R.T.: 150.52
 Base: m/z 18; .9%FS TIC: 135792

#Ions: 7



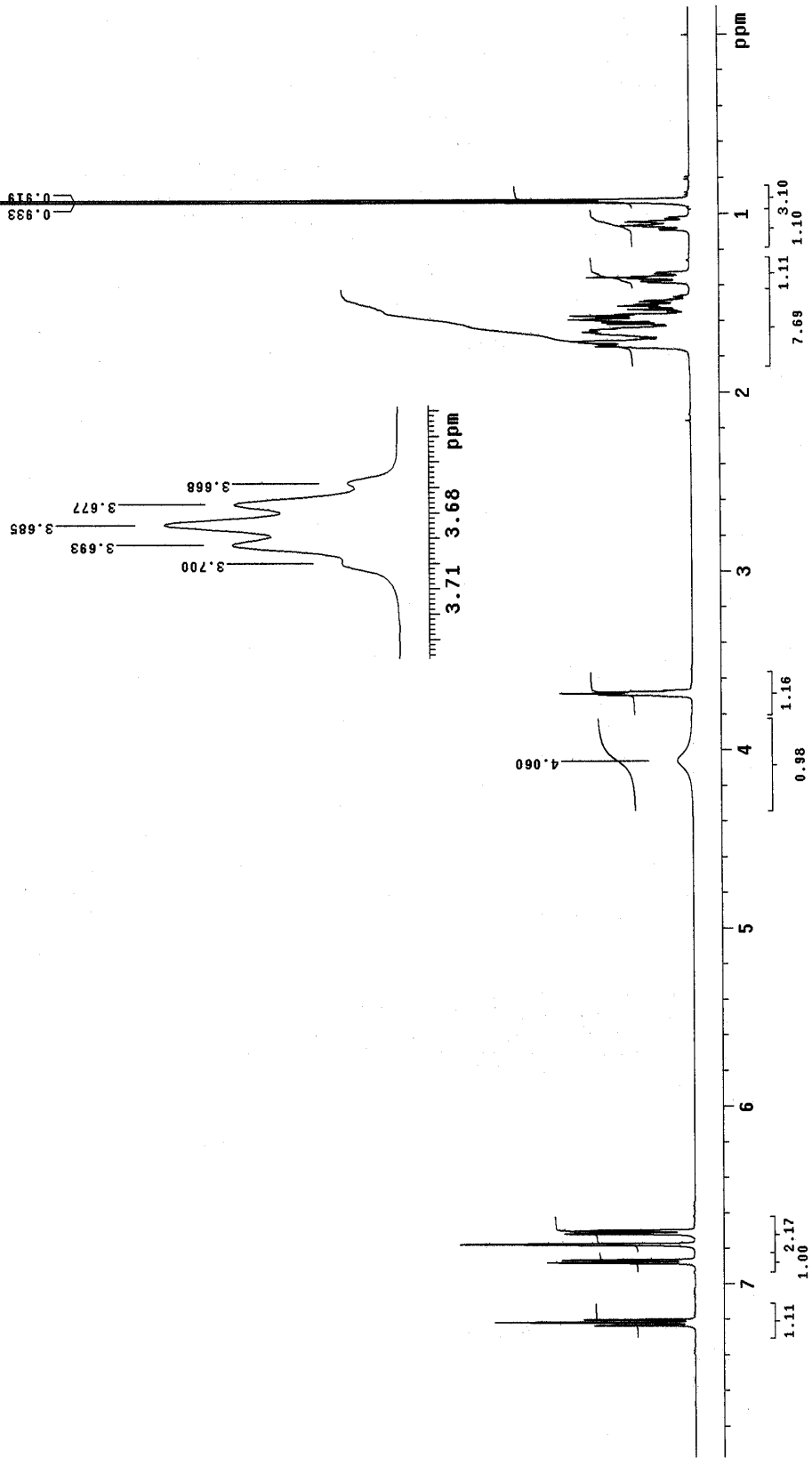
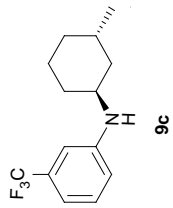
Peak Table

Peak Label	RIC	Scan Range	Baseline at	Total Area	Background	Peak Area
149.88	TIC	7768-7792	7801-7805	19318740	1071222	18247518
150.52	TIC	7824-7848	7856-7860	1571026	1086492	484534

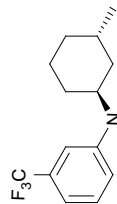
UNAM. Instituto de Quimica. (H. Rios)
Dr-A-cabrera/Laura-R.P
Clave: Reacc55F2526
No. Registro: 906
Experimento: Hidrogeno
Disolvente: CDCl3
VarianInova-500MHz (G)
Marzo-25-2009

File: std1h

Pulse Sequence: s2pul

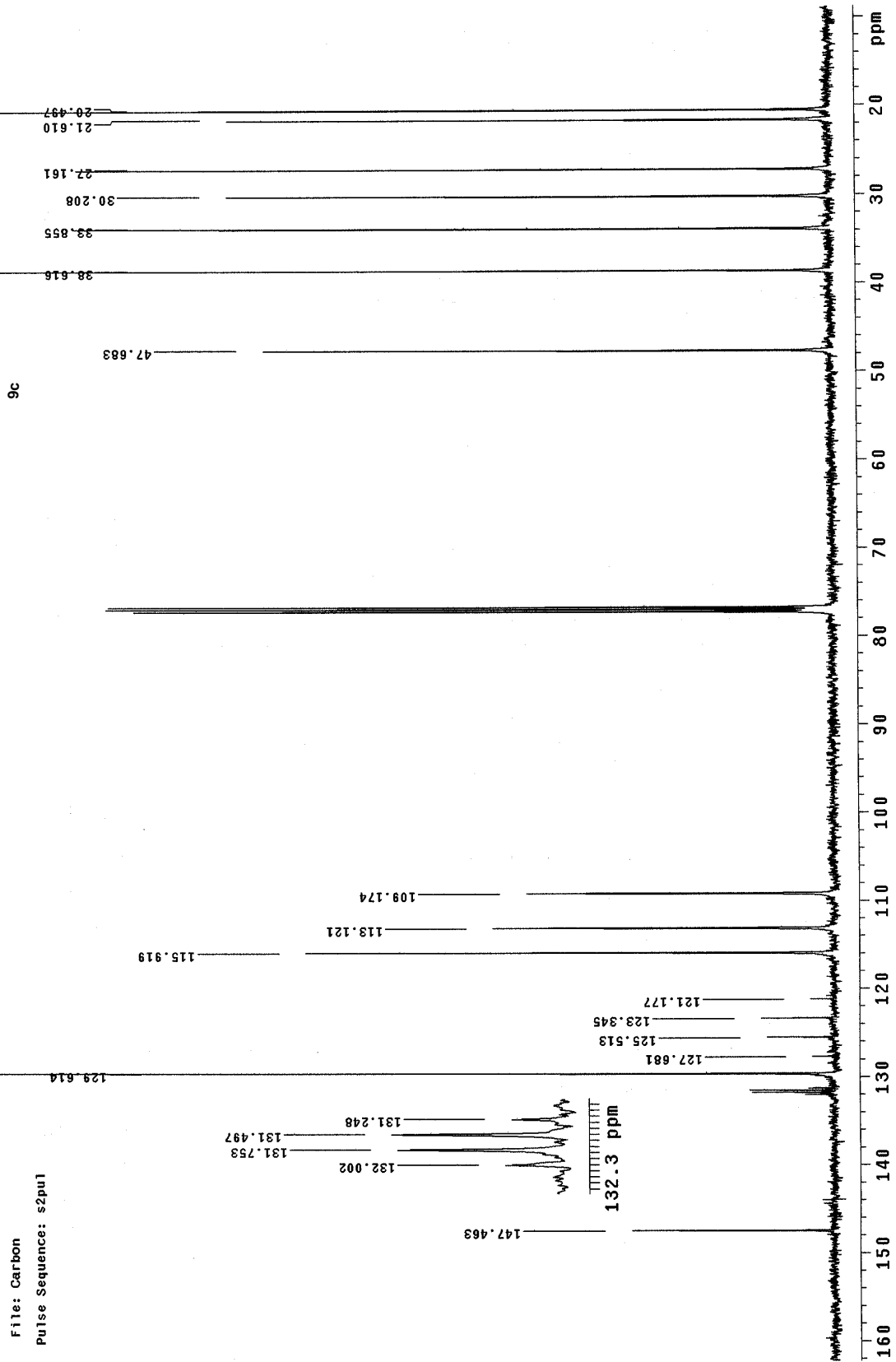


UNAM, Instituto de Química (H. Ríos)
Dr-A-Cabrera/Laura-R.P
Clave: Reacc535F2526
No. registro: 906
Expirante: C13
Disolvente: CDCl3
UnityInova-125.71MHz (G)
Marzo-26-2009



File: Carbon

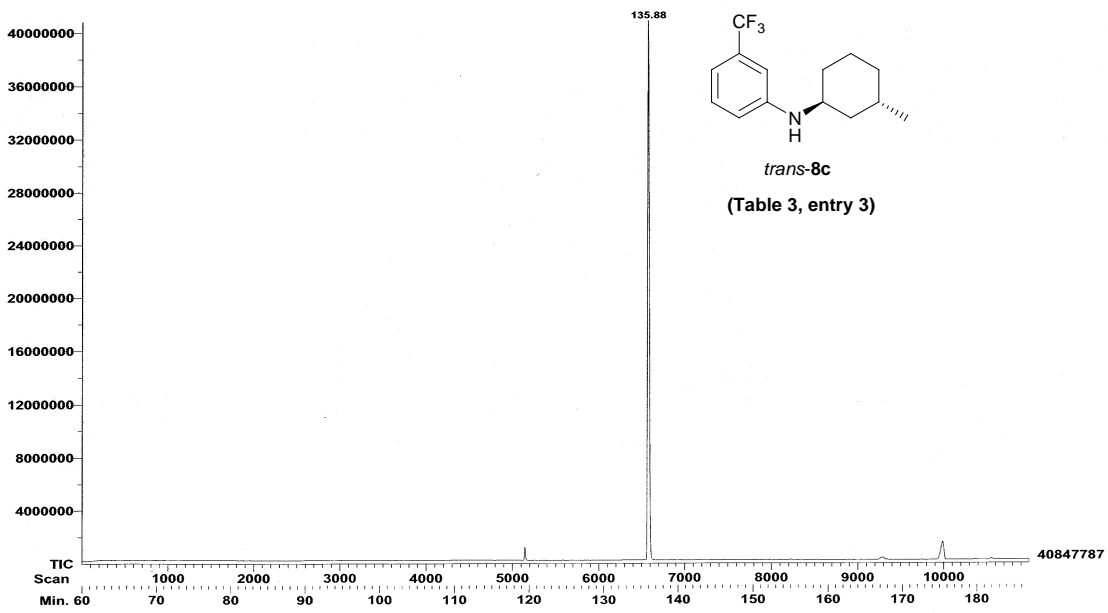
Pulse Sequence: s2pul



File: 1183-reac-535
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-25-2009 (Time Run: 19:05:23)

Ionization mode: EI+



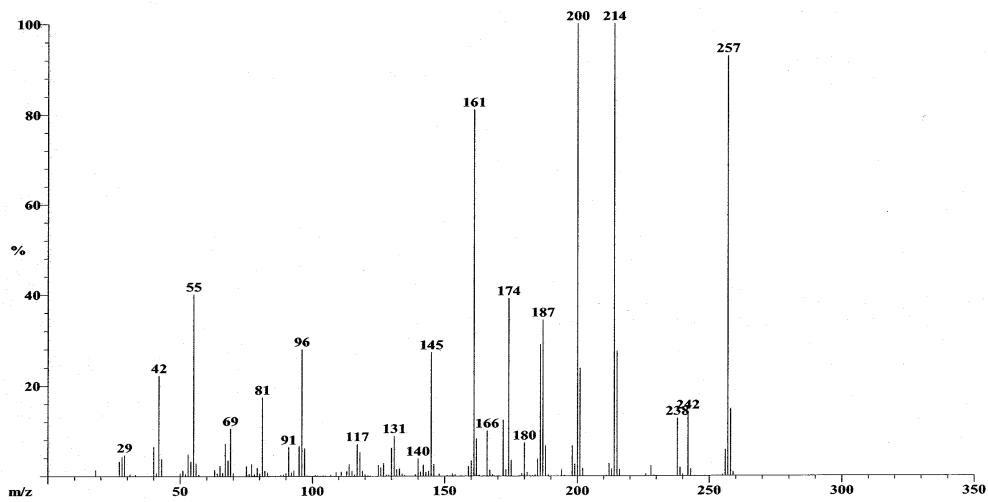
File: 1183-reac-535
 Sample: Dr-Cabrera
 Instrument: JEOL GCmate
 Inlet: GC

Date Run: 05-25-2009 (Time Run: 19:05:23)

Ionization mode: EI+

Scan: 6569 R.T.: 135.88
 Base: m/z 214; 99.6%FS TIC: 40759472

#Ions: 163



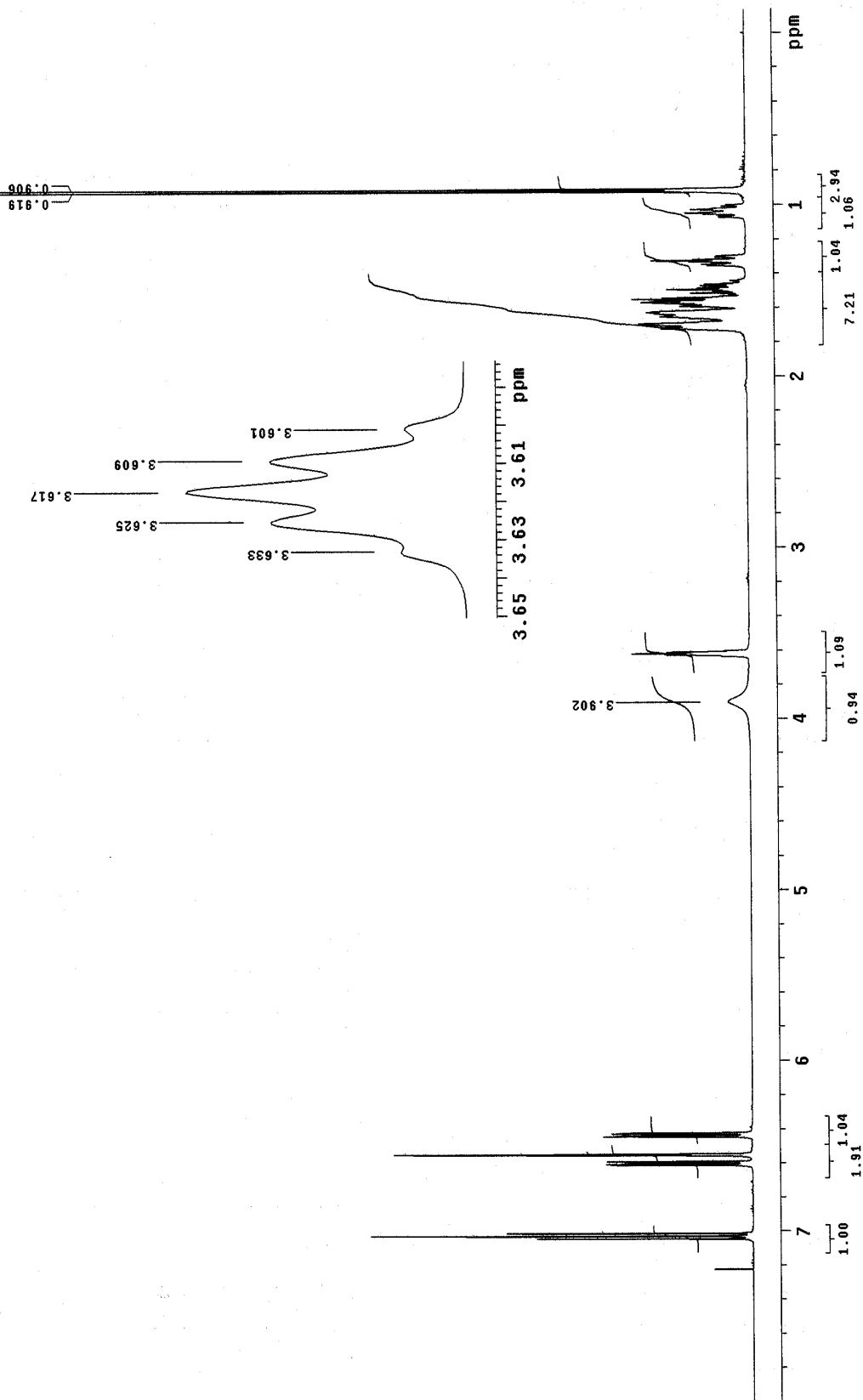
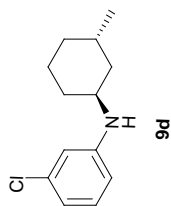
Peak Table

Peak Label	RIC	Scan Range	Baseline at	Total Area	Background	Peak Area
135.88	TIC	6557-6581	6547-6551	478247767	4150451	474097316

UNAM, Instituto de Quimica. (H. Rios)
Dr-A-Cabrera/LC97F27
Citeve: Itacc-597F27
Co. Registro: 866
Experiments: Hidrogeno
Disolvente: CDCl3
Varianova-500MHZ (G)
Marzo-25-2008

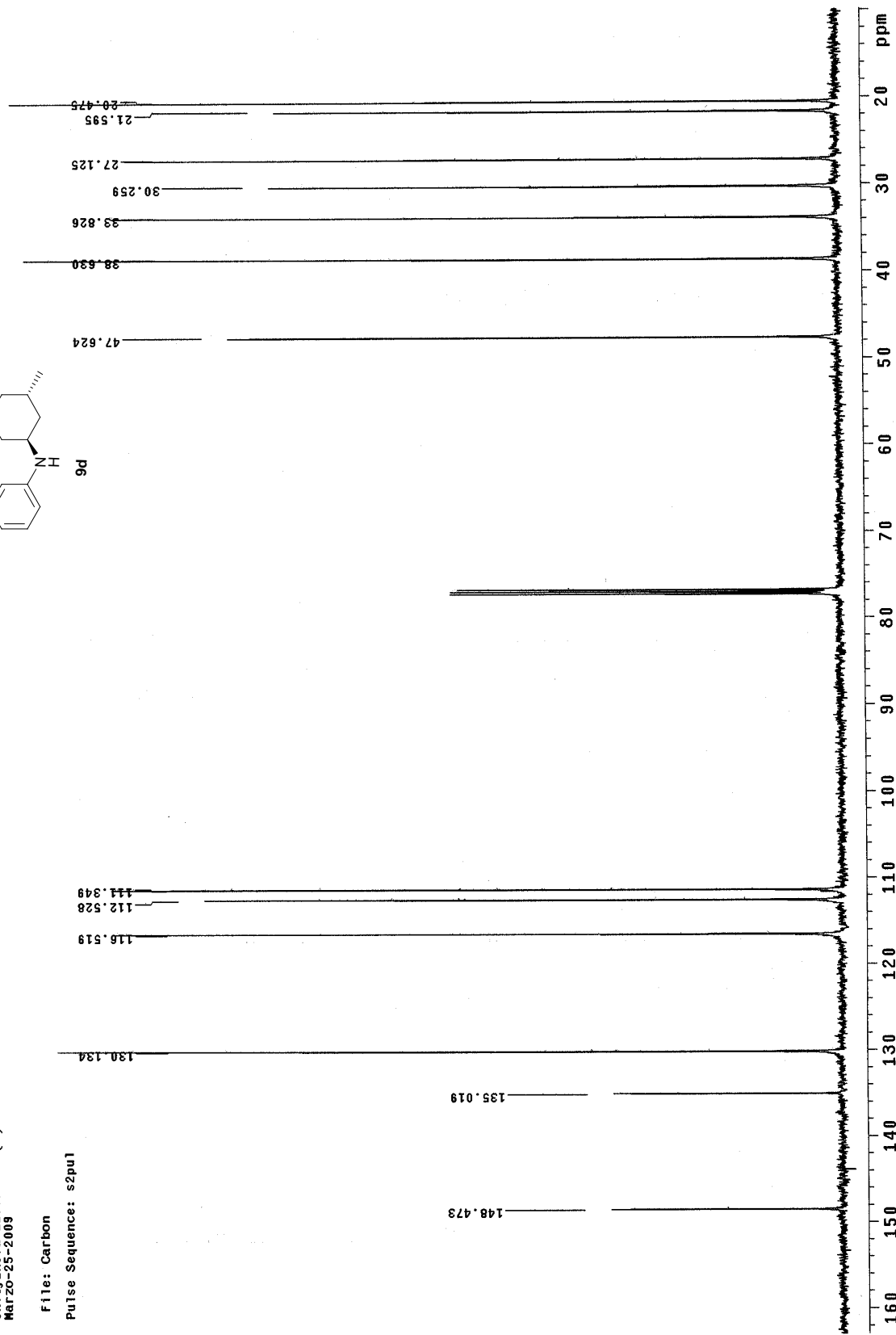
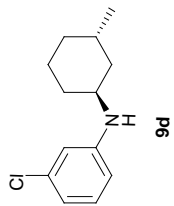
File: stdih

Pulse Sequence: s2pu1



UNAM - Instituto de Quimica (H. Rios)
Dr-A-Cabrera/Laura-R.P
Clave: Reacc53727
No. registro: 866
Experimento: C13
Disolvente: CDCl3
UnityInova-125.71MHz (G)
Marzo-25-2009

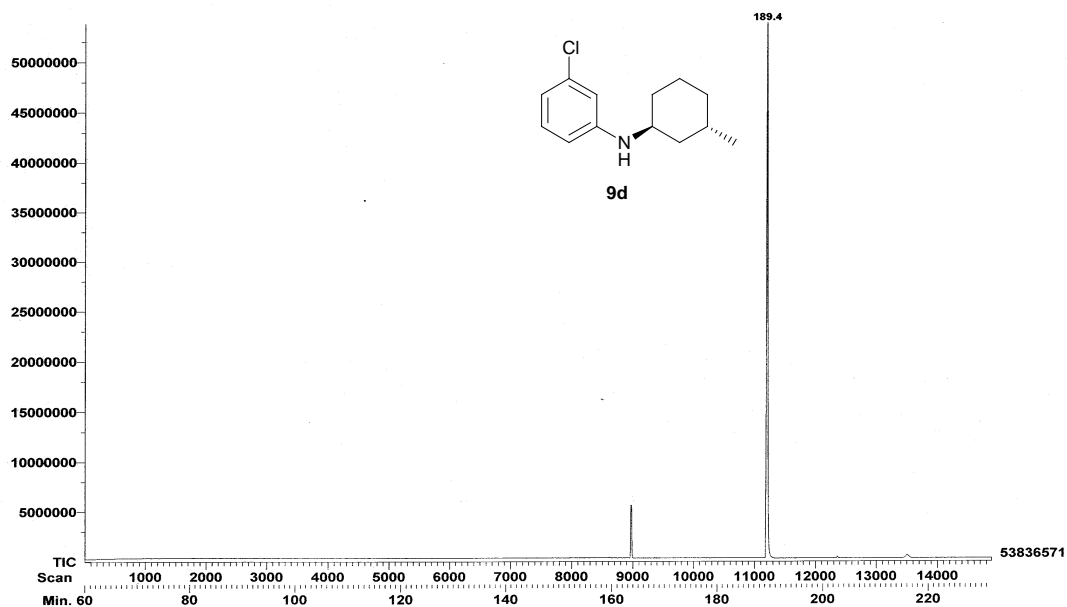
File: Carbon
Pulse Sequence: s2pu1



File: REACC-537
Sample: Dr-Cabrera
Instrument: JEOL GCmate
Inlet: GC

Date Run: 05-26-2009 (Time Run: 09:02:42)

Ionization mode: EI+



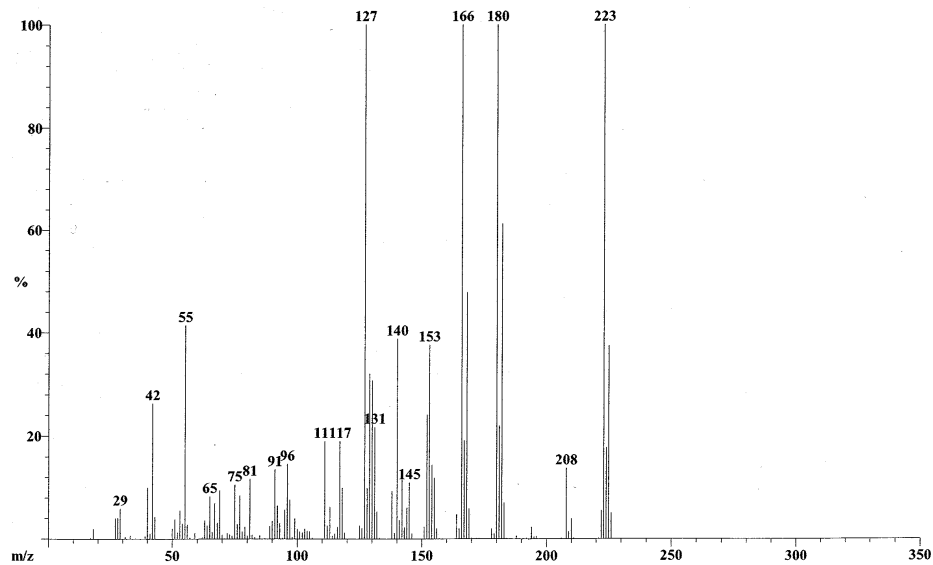
File: REACC-537
Sample: Dr-Cabrera
Instrument: JEOL GCmate
Inlet: GC

Date Run: 05-26-2009 (Time Run: 09:02:42)

Ionization mode: EI+

Scan: 11201 R.T.: 189.4
Base: m/z 223; 99.6%FS TIC: 53914192

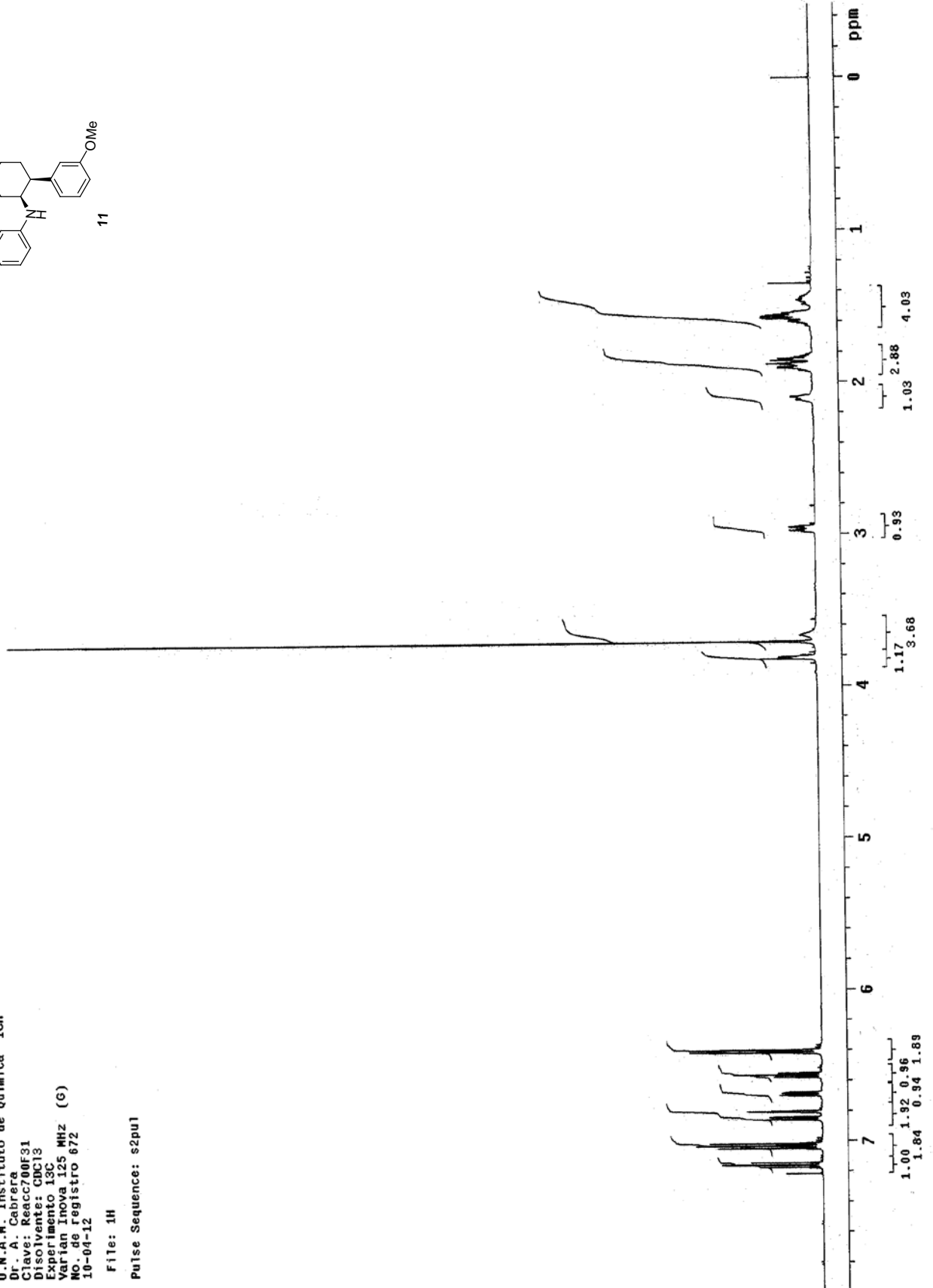
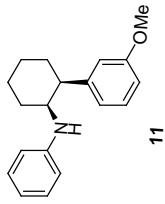
#Ions: 153



U.N.A.M. Instituto de Química ICH
Dr. A. Cabrera
Clave: Reacc700F31
Disolvente: CDCl3
Experimento: 13C
Varian Inova 125 MHz (G)
No. de registro 672
10-04-12

File: 1H

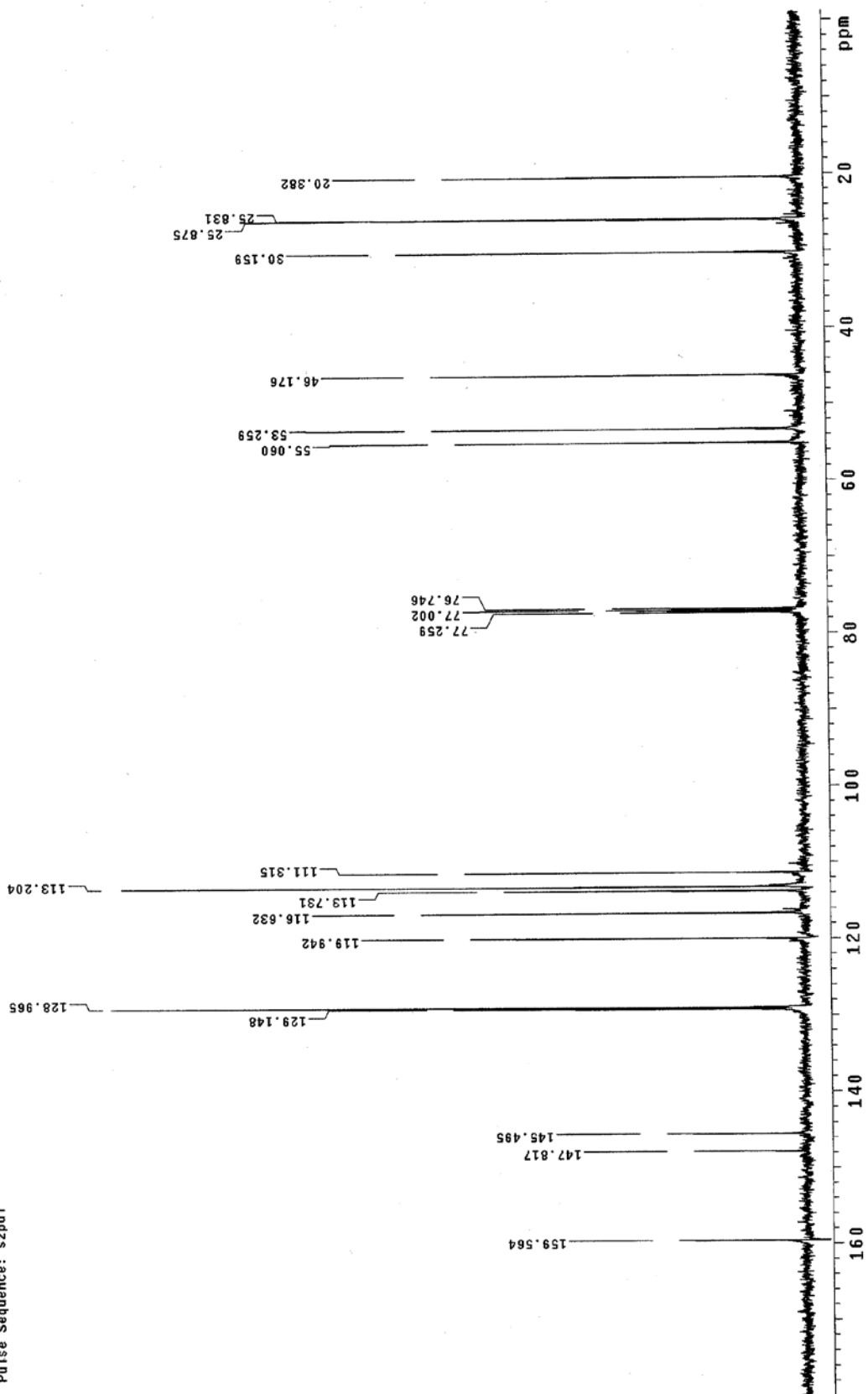
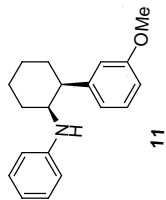
Pulse Sequence: s2pu1



U.N.A.M. Instituto de Química ICH
Dr. A. Cabrera
Clave: Reacc700F31
Disolvente: CDCl3
Experimento: 13C
Varian Inova 125 MHz (G)
No. de registro 672
10-04-12

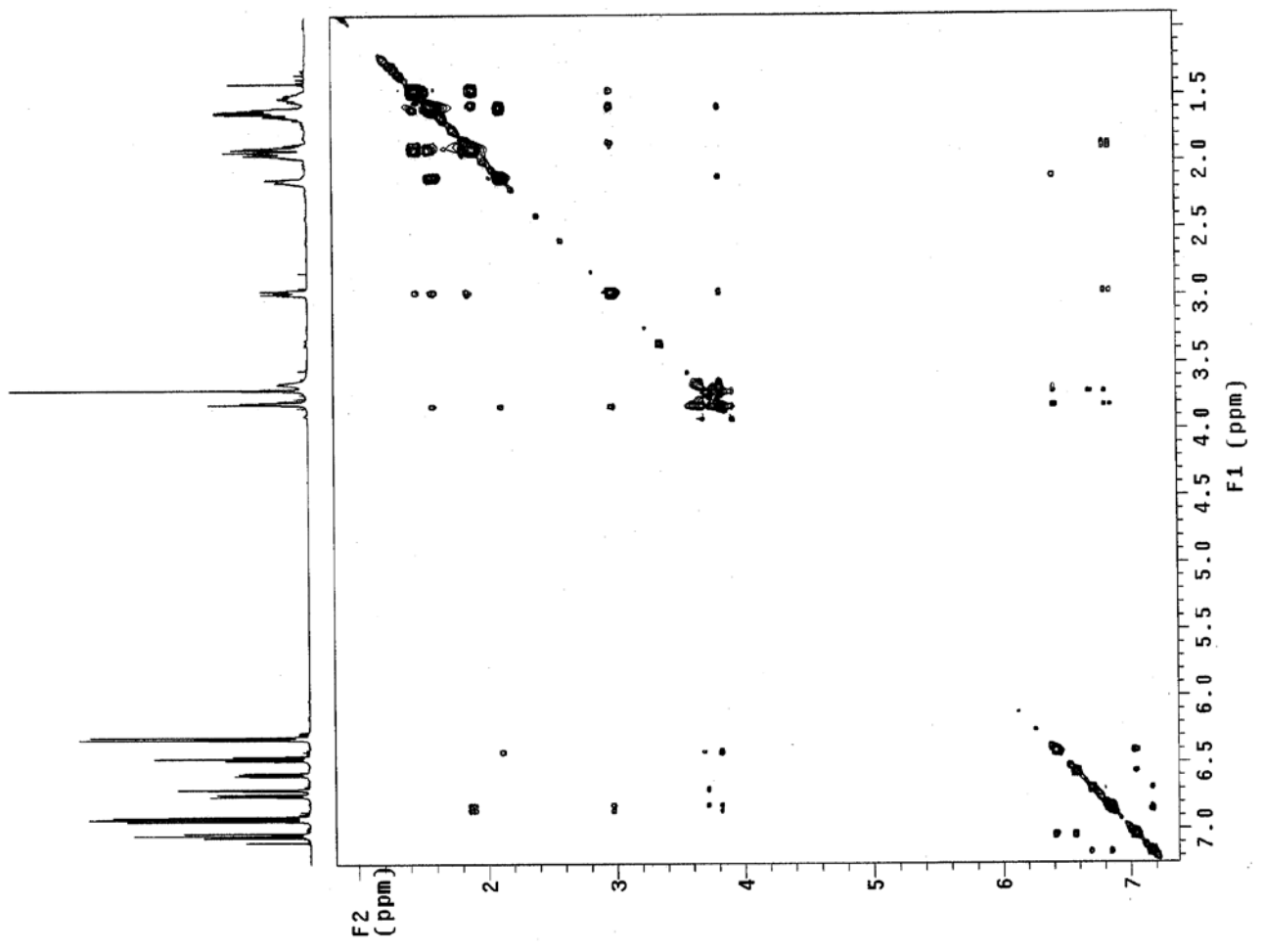
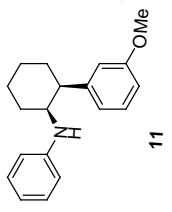
File: 13C

Pulse Sequence: s2pu1

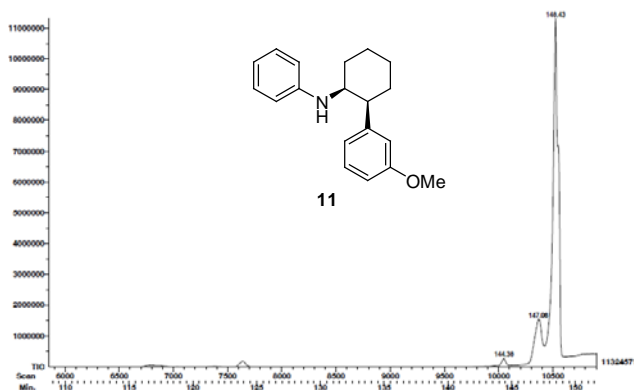


U.N.A.M. Instituto de Quimica ICH
 D.A. Cabrera
 Clave: Reacc700F31
 Disolvente: CDCl3
 Experimento: NQESY
 Varian Inova 500 MHz (G)
 No. de registro: 672
 10-04-12

Pulse Sequence: noesy

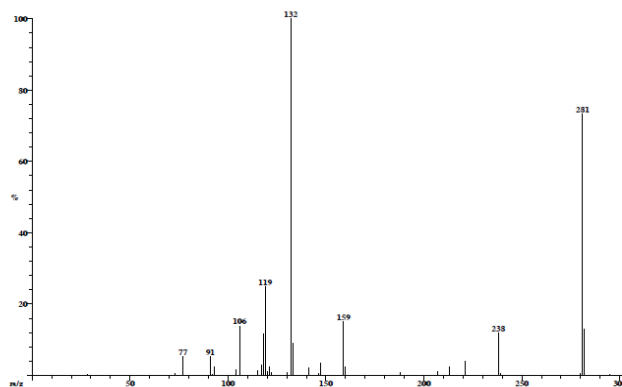


File: REACC -700F31-2
 Sample: Dr Laura Rubio
 Instrument: JEOL GCmate
 Inlet: GC
 Ionization mode: EI+
 Date Run: 07-06-2012 (Time Run: 16:05:15)



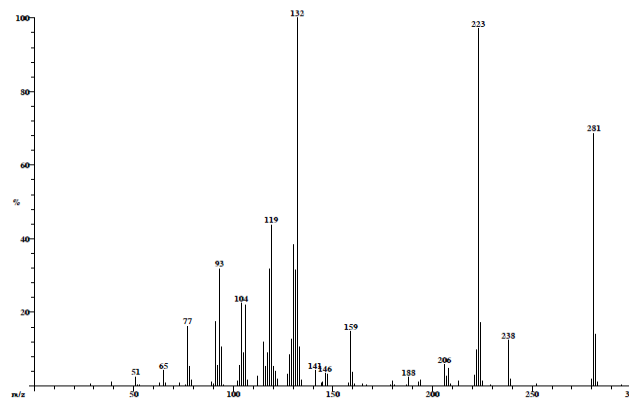
File: REACC -700F31-2
 Sample: Dr Laura Rubio
 Instrument: JEOL GCmate
 Inlet: GC
 Ionization mode: EI+
 Date Run: 07-06-2012 (Time Run: 16:05:15)

Scan: 10366 R.T.: 147.08
 Base: m/z 132; 11.3%FS TIC: 1557808



File: REACC -700F31-2
 Sample: Dr Laura Rubio
 Instrument: JEOL GCmate
 Inlet: GC
 Ionization mode: EI+
 Date Run: 07-06-2012 (Time Run: 16:05:15)

Scan: 10525 R.T.: 148.43
 Base: m/z 132; 35%FS TIC: 11463152

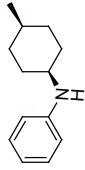


Peak	Compound Name	Peak	RIC	Scan Range	Baseline at	Total Area	Background	Peak Area	Percentage	
147.88		147.88	TIC	10452-10465	10455	16561208	15102734	1458474	9.80298967	Total Area
148.4		148.4	TIC	10518-10531	10518, 10532	1411140616	127721241	13419375	90.1970103	14877849
148.43		148.43	TIC	0-0	0	0	0	0	0	
										Total Percent
										100

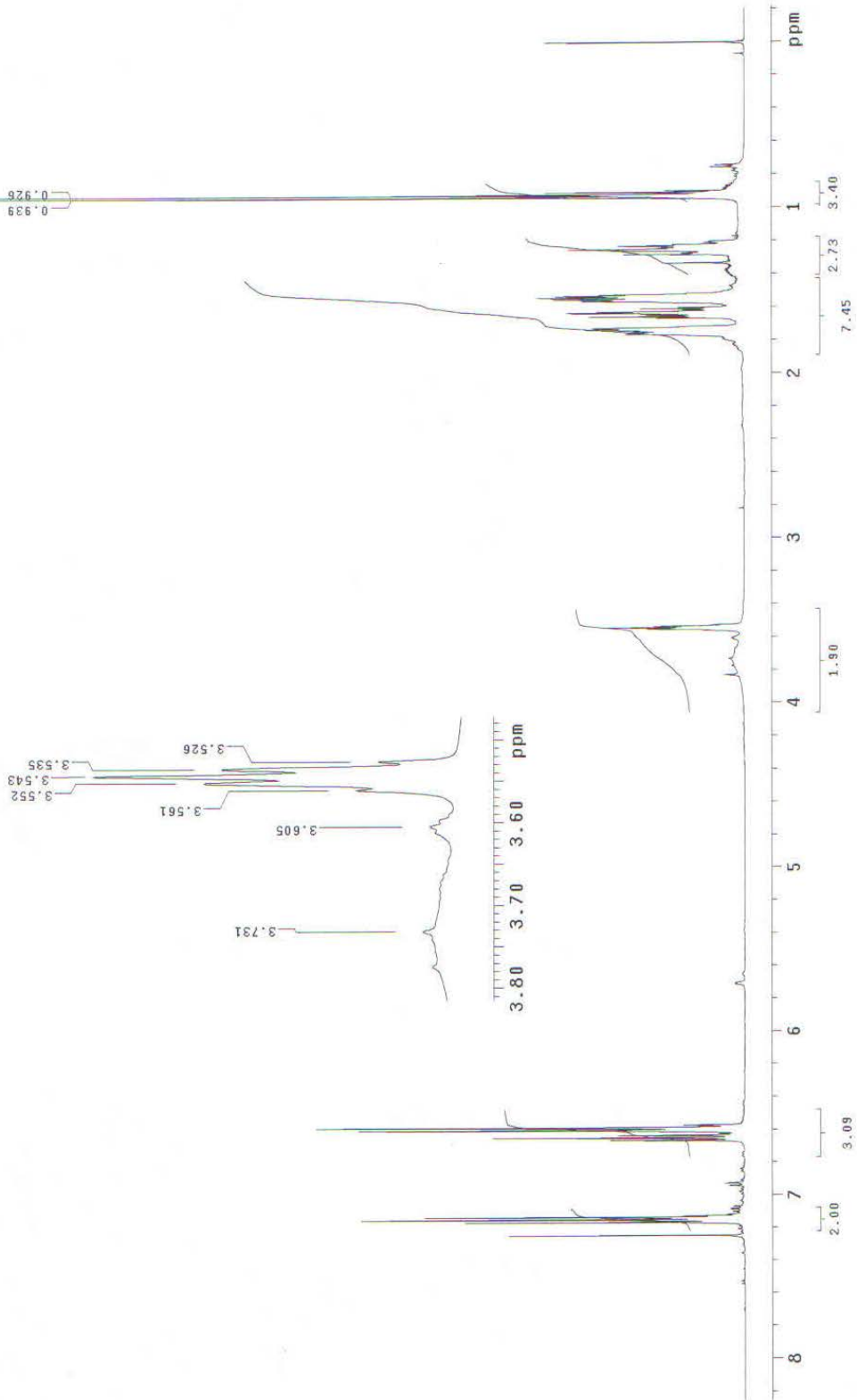
UNAM - Instituto de Química. (H. Ríos)
Dr-A-Cabrera
Clave: Reac547F26
No. Registro: 908
Experimento: RMN-1H
Solvente: CDCl3
Varian300MHz (G)
26-Abr11-2012

File: std1h

Pulse Sequence: s2pu1



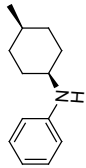
13



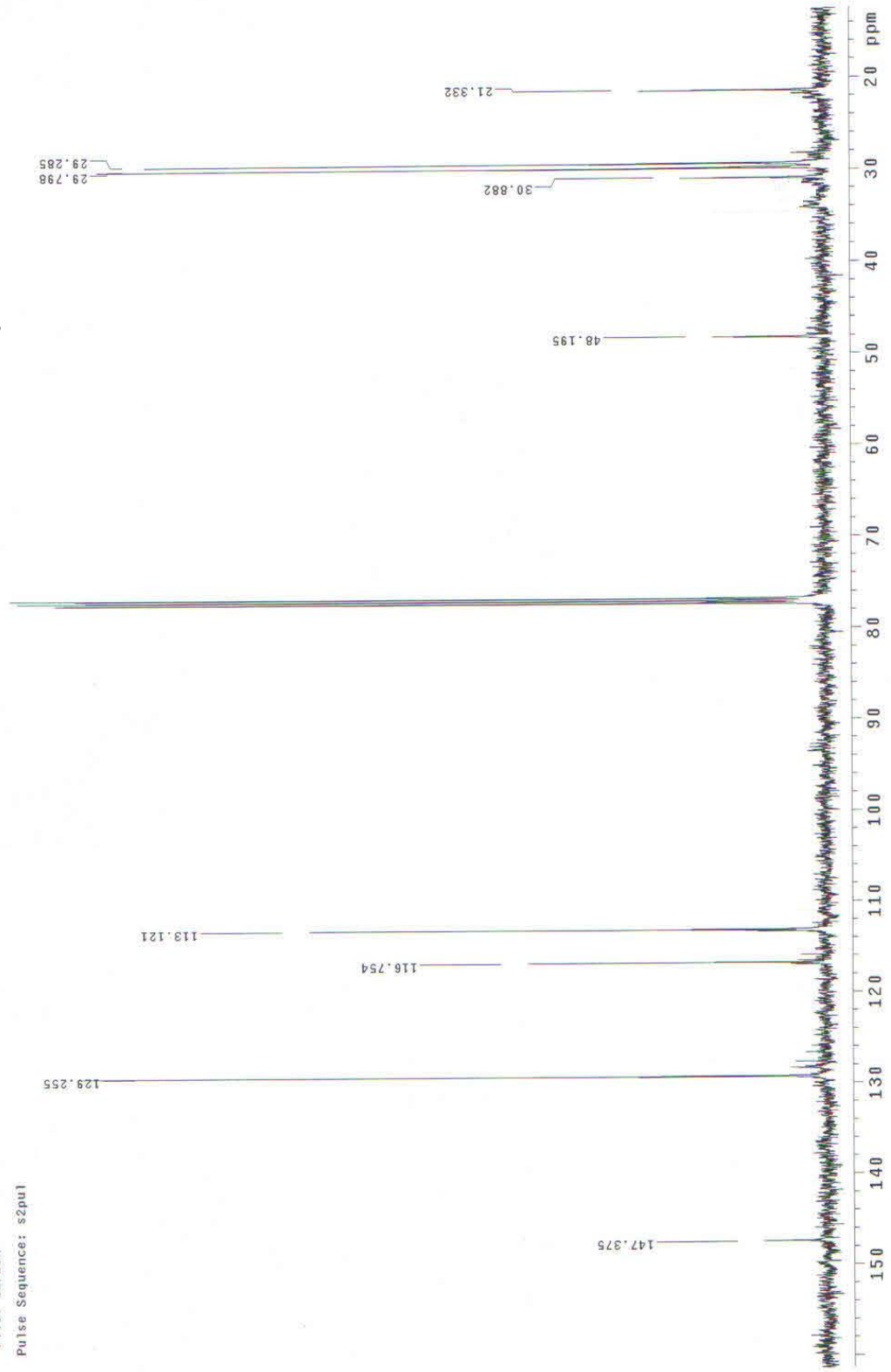
UNAM, Instituto de Química (H. Ríos)
Dr-ACabrera
Clave: Reac547F26
No. registro: RM08
Tipo muestra: RM08
Disolvente: CDCl3
Unidad/Inova-125.71MHz (G)
26-Abr11-2012

File: Carbon

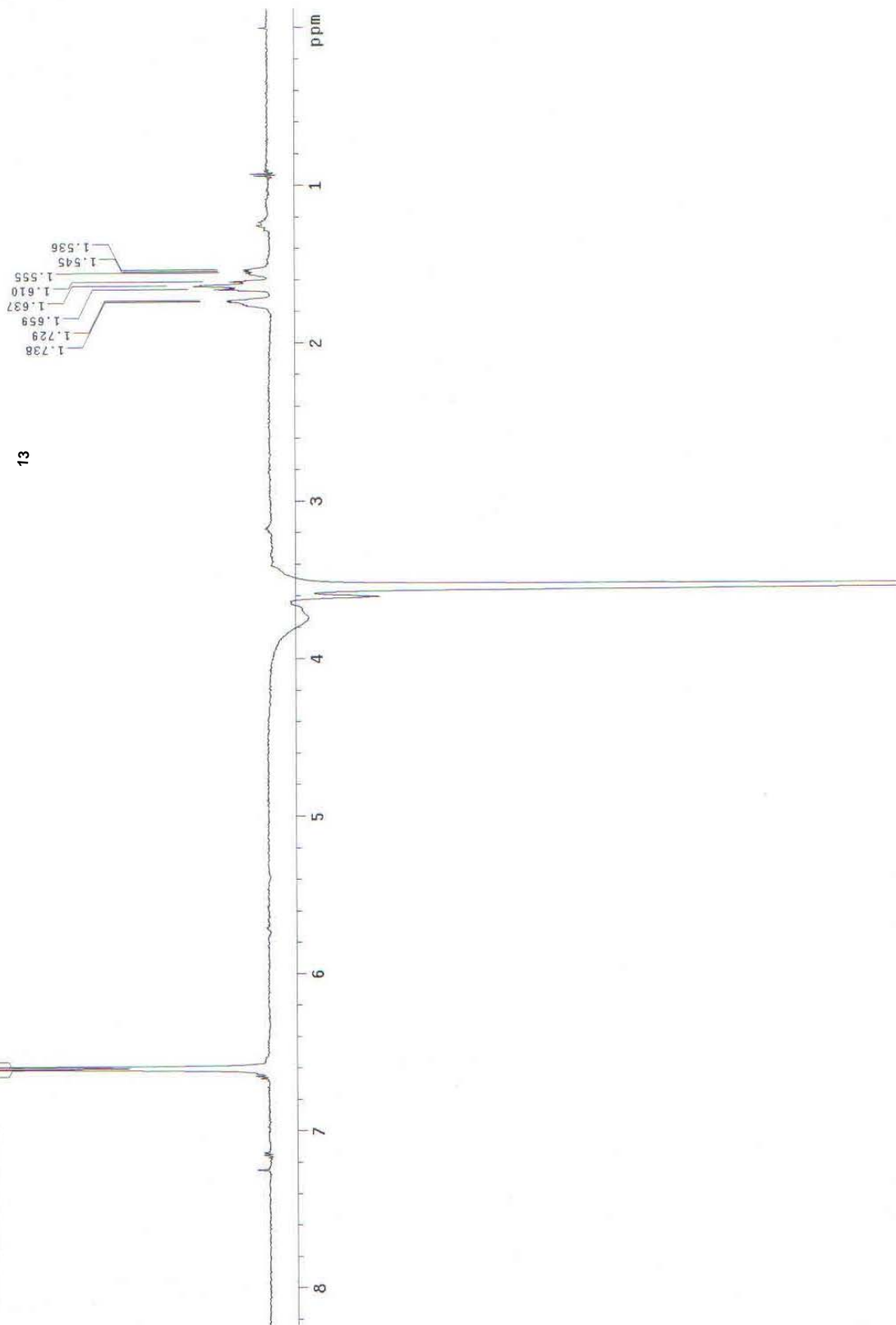
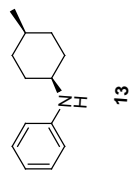
Pulse Sequence: s2pu1



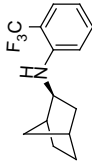
13



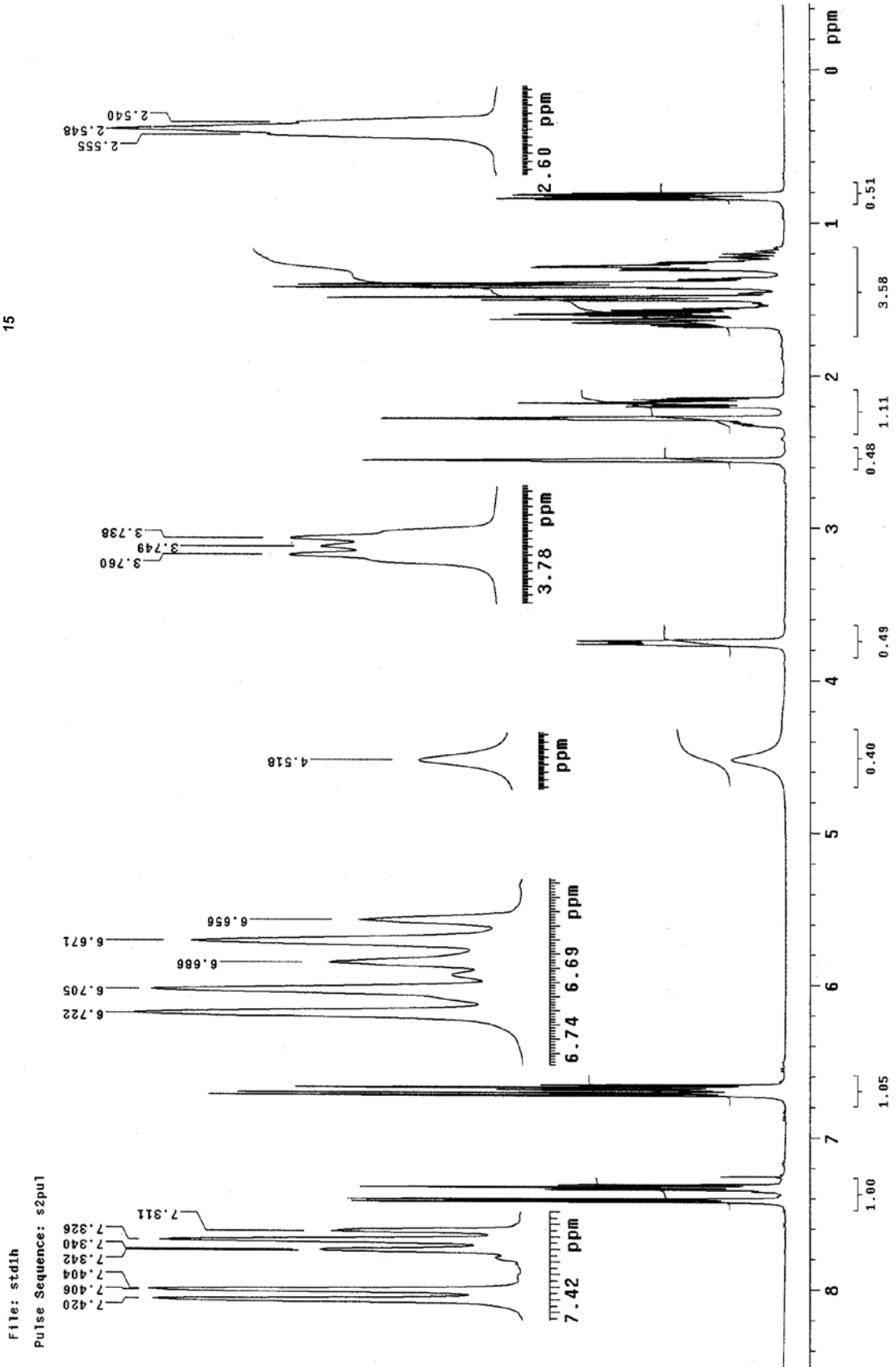
UNAM. Instituto de Química. (H. Ríos)
 Dr-A-Cabrera
 Clave: Reac547F26
 No. Registro: 308
 Experimento: MDE-Diff (Irrad. 3.54 ppm)
 Solvente: $CDCl_3$
 Sarian300MHz (G)
 28-Abr11-2012
 Pulse Sequence: cyclenoe



U.N.A.M. Instituto de Química ICH
 Dr. A. Cabrera
 Clave: Reac-725F12
 Disolvente: CDCl3
 Experiment: H
 Varian Inova 500 MHz (G)
 M. de Registro 1192
 24-05-12



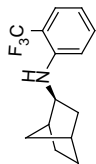
15



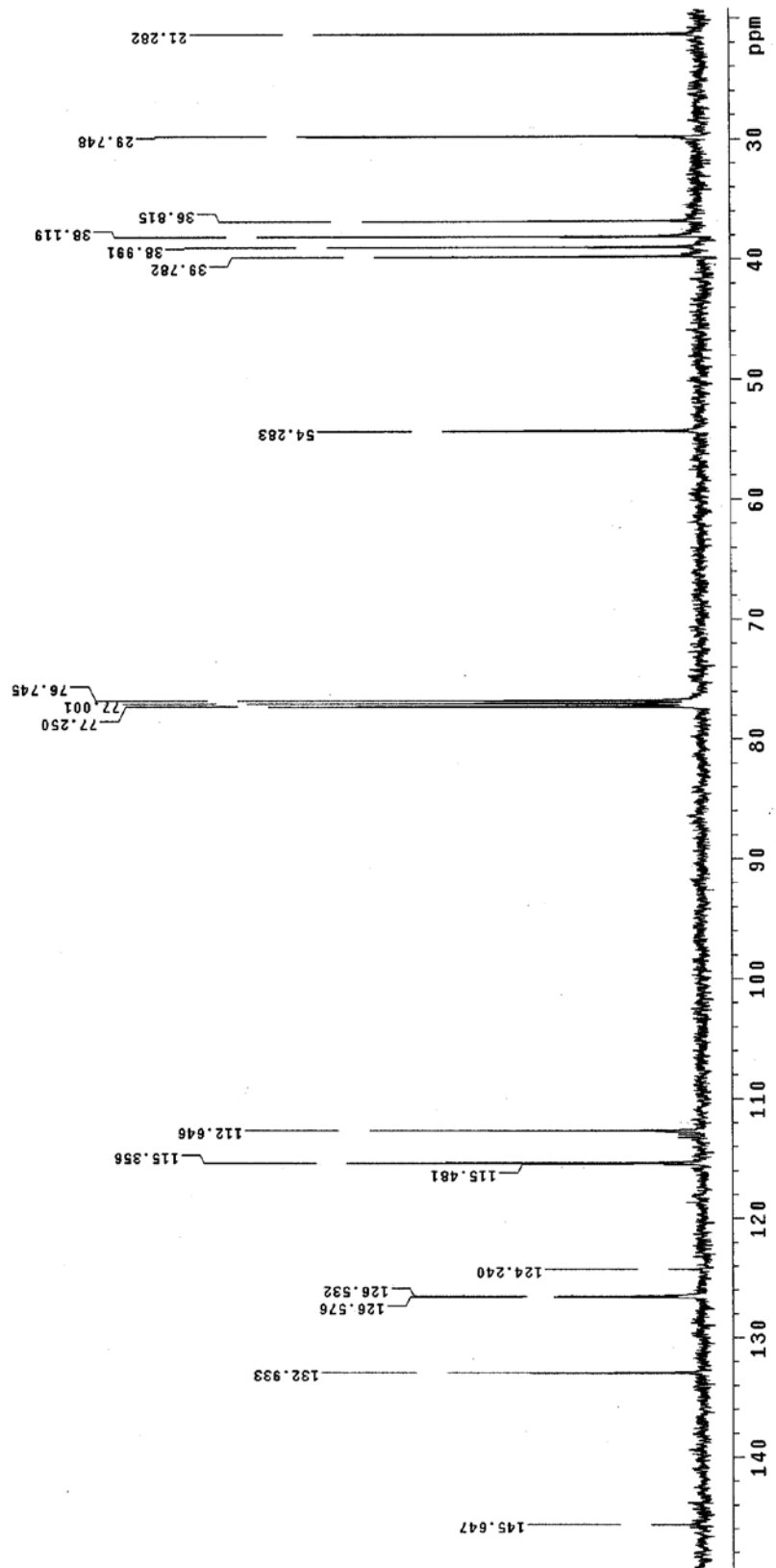
U.N.A.M. Instituto de Quimica ICH
Dr. A. Cabrera
Clave: Reacc 725F12
Disolvente: CDCl3
Experimento 13C
Varian Inova 125 MHz (G)
No. de registro 1192
24-05-12

File: 13C

Pulse Sequence: s2pul

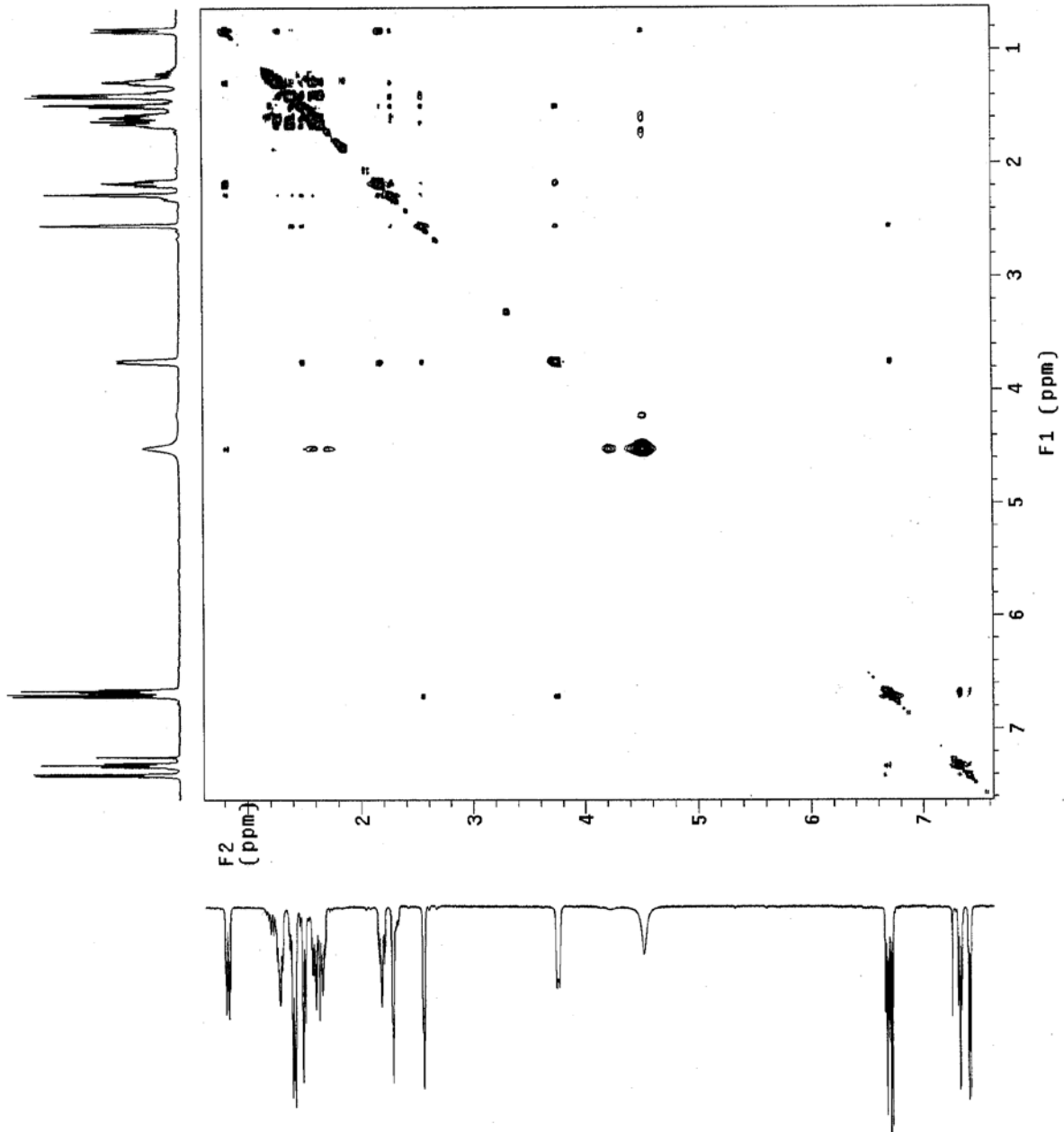
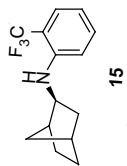


15



U.N.A.M. Instituto de Quimica ICH
Dr. A. Cabrera
Clave: Reacc 725F12
Disolvente: CDCl3
Experimento: NOESY
Varian Inova 500 MHz (G)
No. de registro 1192
11-06-12

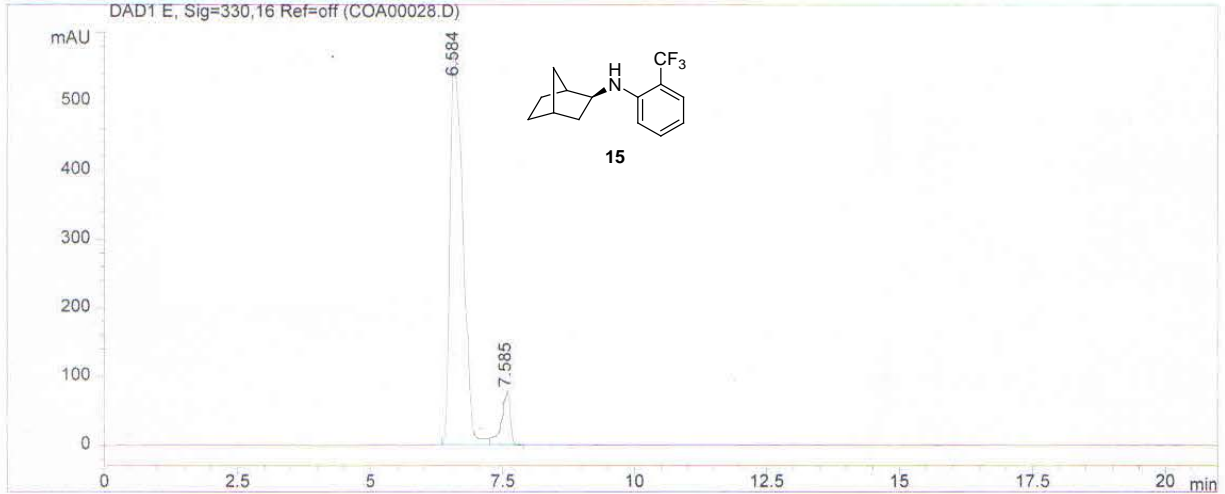
Pulse Sequence: noesy



Data File C:\HPCHEM\1\DATA\COA00028.D
Chiralpak IA Hex/iPrOH/ 99/1 flujo 0.5 mL/min

Sample Name: Reacc733F9Quiral

=====
Injection Date : 01/06/12 8:38:50 PM
Sample Name : Reacc725F9Quiral Vial : 1
Acq. Operator : Erendira Garcia
Method : C:\HPCHEM\1\METHODS\QUIRAL.M
Last changed : 01/06/12 7:11:45 PM by Erendira Garcia
(modified after loading)
=====



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000

Signal 1: DAD1 E, Sig=330,16 Ref=off
Results obtained with enhanced integrator!

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.584	BV	0.2941	1.23782e4	568.26147	93.2883
2	7.585	VP	0.1931	890.56598	76.88500	6.7117

Totals : 1.32688e4 645.14648

=====
*** End of Report ***

ee = 86.57 ~ 87%