

Supporting information

Carbon nitride nanoparticles as ultrasensitive fluorescent probe for the detection of α -glucosidase activity and inhibitor screening

Feng-Na Guo, Yi-Ting Wang, Na Wu, Li-Xia Feng, Hui-Chao Zhang, Ting Yang*,

Jian-Hua Wang

Research Center for Analytical Sciences, Department of Chemistry, College of Sciences, Box 332, Northeastern University, Shenyang 110819, China

Corresponding Author

*E-mail: yangting@mail.neu.edu.cn (T. Yang);

Tel: +86 24 83688944; Fax: +86 24 83687659

Table of contents

Fig. S1. UV-vis absorption spectra and fluorescence emission spectra under different excitation wavelengths of CNNPs.....	S-3
Fig. S2. The effects of pH, Xe-lamp irradiation time and ionic strength on the fluorescence intensity of CNNPs.....	S-4
Fig. S3. Changes of fluorescence emission spectra of CNNPs in the presence of different compositions.....	S-5
Fig. S4. Zeta potential of CNNPs and pNP at pH 7.0	S-6
Fig. S5. UV-vis absorption spectra of CNNPs, pNP and CNNPs+pNP.....	S-7
Fig. S6. The effects of pH and incubation time on the detection system of pNP.....	S-8
Fig. S7. The effect of pH on the detection system of α -glucosidase.....	S-9
Fig. S8. The effects of the concentration of pNPG and incubation time on the detection system of α -glucosidase.....	S-10
Table S1. Comparison of the methods for the detection of α -glucosidase.....	S-11
Reference.....	S-12

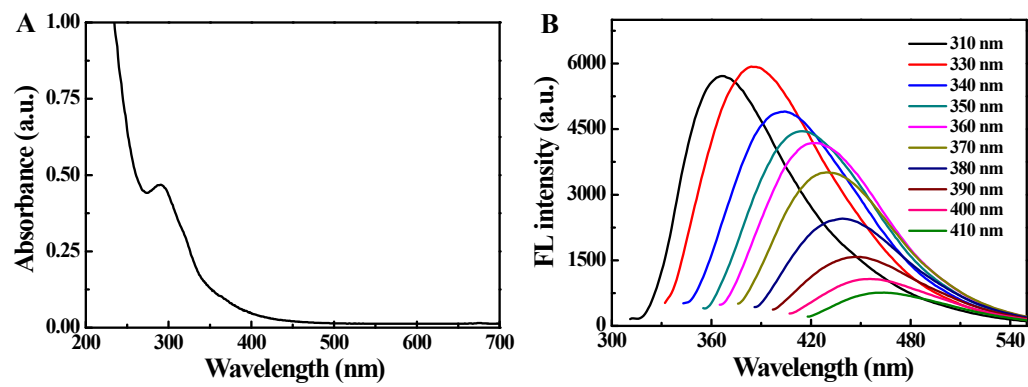


Fig. S1 (A) UV-vis absorption spectra of CNNPs. (B) Fluorescence emission spectra under different excitation wavelengths.

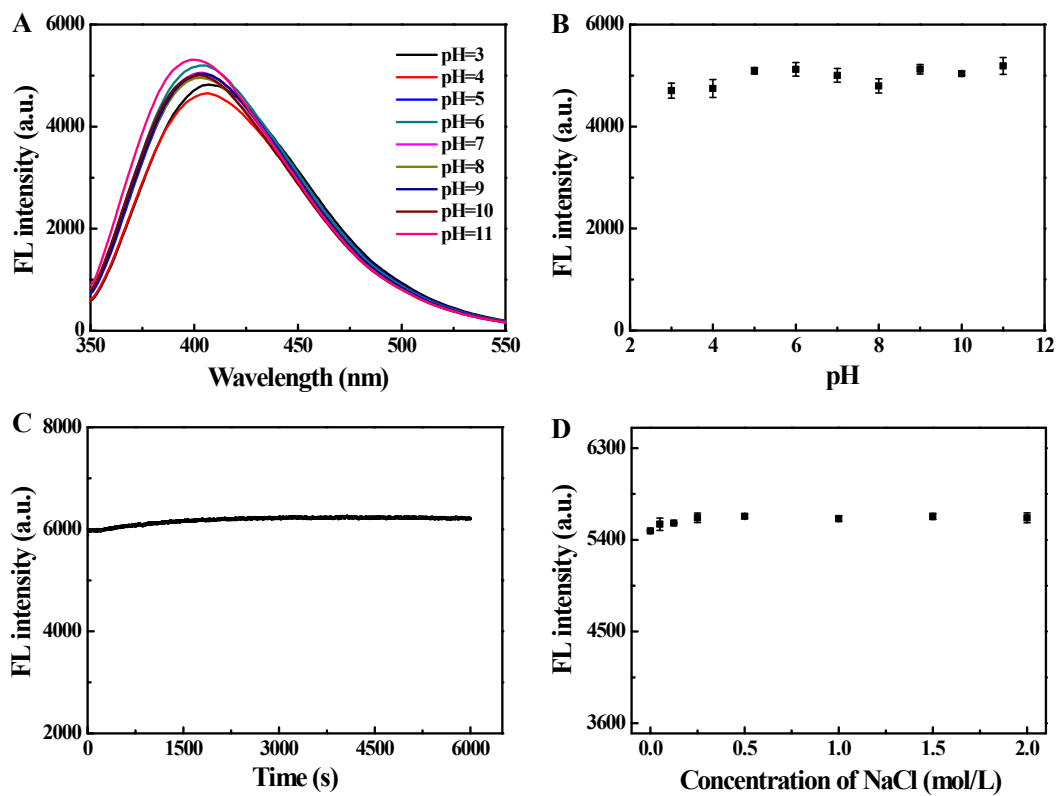


Fig. S2 (A) The fluorescence emission spectra of CNNPs under different pH conditions. The FL intensity of CNNPs as the function of pH (B), Xe-lamp irradiation time (C) and ionic strength (D).

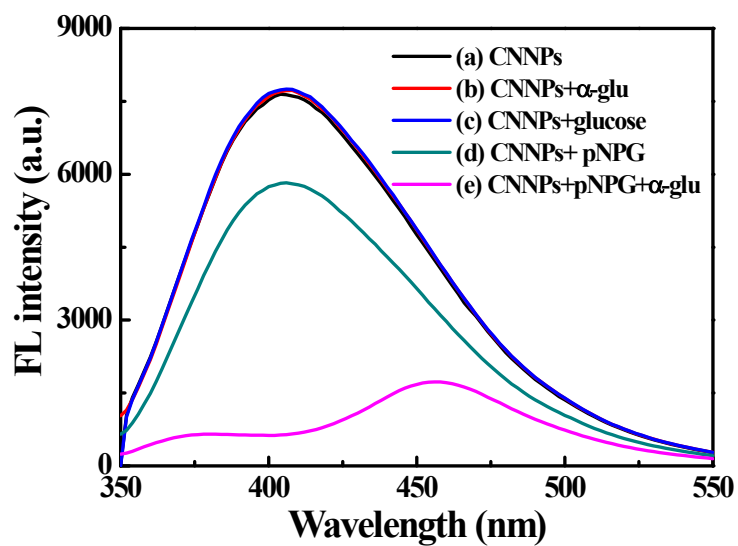


Fig. S3 Changes of fluorescence emission spectra of CNNPs in the presence of different compositions: (a) CNNPs alone, (b) CNNPs and α -glu (100 U/L), (c) CNNPs and glucose (5 mM), (d) CNNPs and pNPG (200 μ M), (e) CNNPs, α -glu and pNPG.

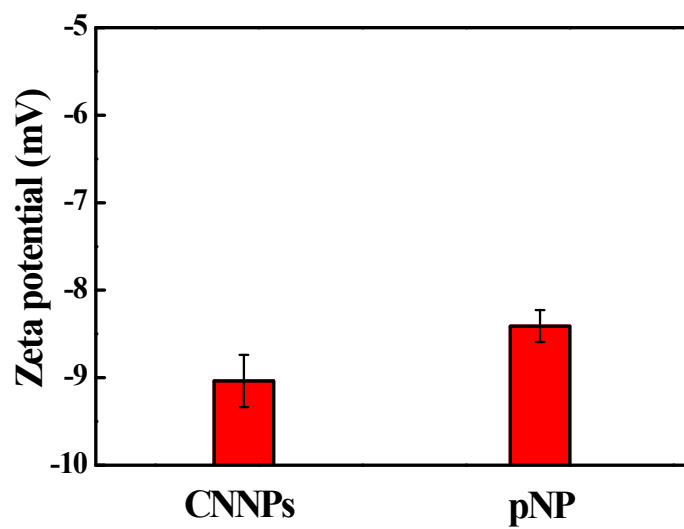


Fig. S4 Zeta potential of CNNPs and pNP at pH 7.0.

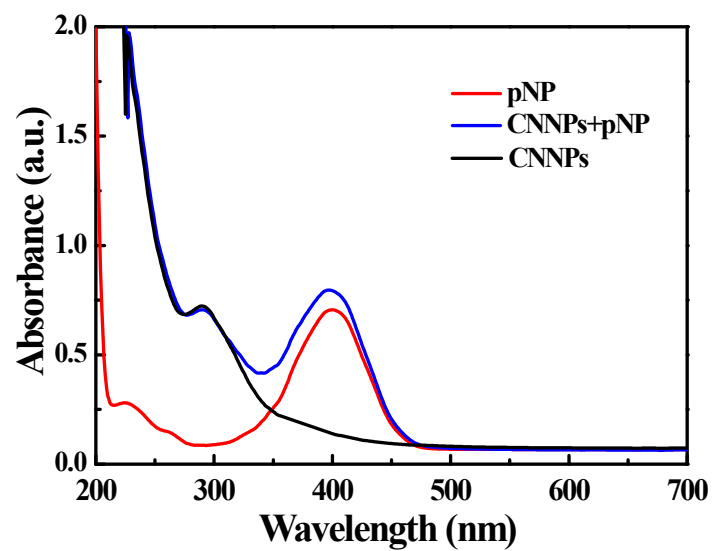


Fig. S5 UV-vis absorption spectra of CNNPs, pNP and CNNPs + pNP.

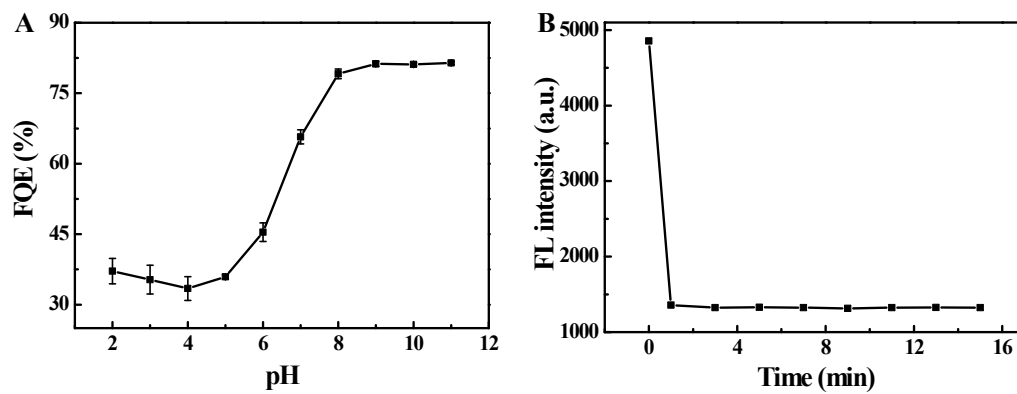


Fig. S6 (A) Fluorescence quenching efficiency (FQE) of pNP (100 μ M) for CNNPs versus pH values. (B) FL intensity of CNNPs in the presence of pNP (100 μ M) versus incubation time.

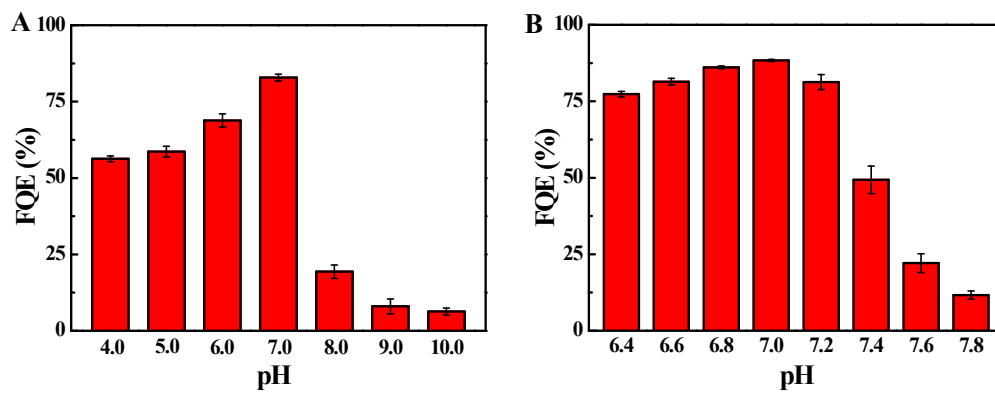


Fig. S7 Fluorescence quenching efficiency (FQE) of CNNPs versus pH values (A) varying from 4.0 to 10.0 and (B) varying from 6.4 to 7.8 in the presence of pNPG (200 μ M) and α -glucosidase (100 U/L).

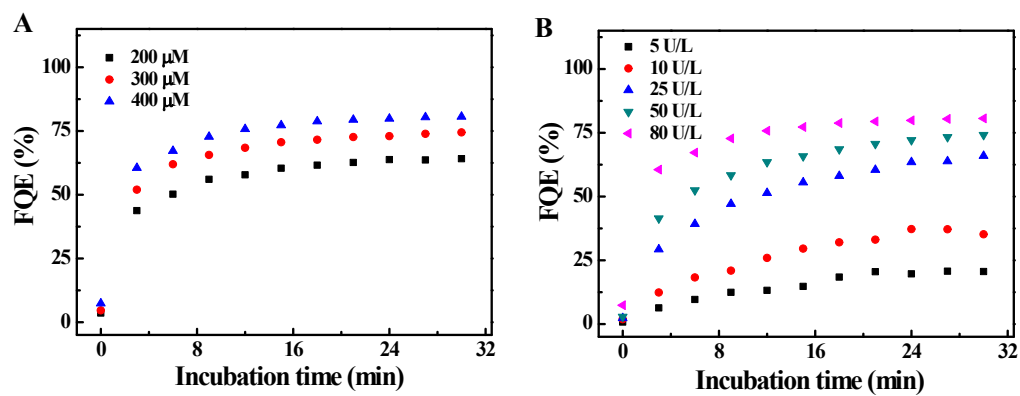


Fig. S8 (A) Fluorescence quenching efficiency (FQE) of CNNPs versus incubation time in the presence of α -glucosidase (80U/L) and different amounts of pNPG (200, 300, 400 μ M). (B) Fluorescence quenching efficiency of CNNPs versus incubation time in the presence of pNPG (200 μ M) and different concentrations of α -glucosidase (5, 10, 25, 50, 80 U/L).

Table S1. Comparison of the methods for the detection of α -glucosidase.

Methods	Probes	Linear range (U/L)	Detection limit (U/L)	Reference
Colorimetric assay	Gold nanorods	2.5 - 45	0.5	[S1]
Colorimetric assay	Unmodified gold nanoparticles	2.5 - 50	1	[S2]
Colorimetric assay	gold Nanoparticles	50 - 1100	4	[S3]
Electrochemical assay	AuNPs modified with ATP aptamer and pAPG	10 - 1300	5	[S4]
Electrochemical assay	AgNPs/DA and MNPs/pAPG with PBA/GE	0 - 1100	40	[S5]
Ratiometric fluorescent assay	Dual-color carbon dots	130 - 6700	36	[S6]
Ratiometric Fluorescent assay	MnO ₂ -OPD	200 - 8000	30	[S7]
Fluorescent assay	F-PDA- CoOOH	2 - 80	1.65	[S8]
Fluorescent assay	SiQDs-MnO ₂	20 - 2500	7	[S9]
Fluorescent assay	N, B-CDs	10 - 1000	3	[S10]
Fluorescent assay	PBA-CQDs	1.14 - 17.35	0.33	[S11]
Fluorescent assay	Supramolecular Self-assembly	2.2 - 60	0.66	[S12]
Fluorescent assay	Cationic Conjugated Polymers	10 - 1300	5	[S13]
Fluorescent assay	Carbon nitride nanoparticles	1.25 - 10	0.17	This work

Reference

- [S1] Xin Cheng, Yan Huang, Chao Yuan, et al. Colorimetric detection of α -glucosidase activity based on the etching of gold nanorods and its application to screen anti-diabetic drugs[J]. *Sensor. Actuat. B: Chem.*, 2019, 282: 838 - 843.
- [S2] Hongxia Chen, Jiangjiang Zhang, Heng Wu, et al. Sensitive colorimetric assays for α -glucosidase activity and inhibitor screening based on unmodified gold nanoparticles[J]. *Anal. Chim. Acta*, 2015, 87: 92 - 98.
- [S3] Juan Zhang, Ying Liu, Jun Lv, et al. A colorimetric method for α -glucosidase activity assay and its inhibitor screening based on aggregation of gold nanoparticles induced by specific recognition between phenylenediboronic acid and 4-aminophenyl- α -D-glucoopyranoside[J]. *Nano. Res.*, 2015, 8 (3): 920 - 930.
- [S4] Jinlong Li, Guangwu He, Bei Wang, et al. Fabrication of reusable electrochemical biosensor and its application for the assay of α -glucosidase activity[J]. *Anal. Chim. Acta*, 2018, 1026: 140 - 146.
- [S5] Juan Zhang, Ying Liu, Xiaonan Wang, et al. Electrochemical assay of α -glucosidase activity and the inhibitor screening in cell medium[J]. *Biosens. Bioelectron.*, 2015, 74: 666 - 672
- [S6] Xia Cheng, Jian Xu, Lin Wang, et al. A redox modulated ratiometric fluorometric method based on the use of dual-color carbon dots for determination of the activity of enzymes participating in ascorbic acid-related reactions[J]. *Microchim. Acta*, 2019, 186: 818
- [S7] Menglan Shi, Yao Cen, Guanhong Xu, et al. Ratiometric fluorescence monitoring of α -glucosidase activity based on oxidase-like property of MnO_2 nanosheet and its application for inhibitor screening [J]. *Anal. Chim. Acta*, 2019, 1077: 225 - 231.
- [S8] Heng Zhang, Zhen Wang, Xiaoqing Yang, et al. The determination of α -glucosidase activity through a nano fluorescent sensor of F-PDA-CoOOH[J]. *Anal. Chim. Acta*, 2019, 1080: 170 - 177
- [S9] Jinying Liu, Xinhe Duan, Mengke Wang, et al. A label-free fluorescent sensor based on silicon quantum dots- MnO_2 nanosheets for the detection of α -glucosidase and its inhibitor[J]. *Analyst*, 2019, 144: 7398 - 7405.
- [S10] Shan Huang, Erli Yang, Jiandong Yao, et al. Carbon dots doped with nitrogen and boron as ultrasensitive fluorescent probes for determination of α -glucosidase activity and its inhibitors in water samples and living cells[J]. *Microchim. Acta*, 2018, 185: 394
- [S11] Hang Ao, Hui Feng, Xiaolu Huang, et al. A reversible fluorescence nanoswitch based on dynamic covalent B-O bonds using functional carbon quantum dots and its application for α -glucosidase activity monitoring[J]. *J. Mater. Chem. C*, 2017, 5: 2826 - 2832.
- [S12] Cong Tang, Zhaosheng Qian, Yinjie Qian, et al. A fluorometric and real-time assay for α -glucosidase activity through supramolecular self-assembly and its application for inhibitor screening[J]. *Sensor. Actuat. B: Chem.*, 2017, 245: 282 - 289.
- [S13] Ali Cao, Yanli Tang, and Yue Liu. Novel fluorescent biosensor for α -glucosidase inhibitor screening based on cationic conjugated Polymers[J]. *Appl. Mater. Interfaces*, 2012, 4: 3773 - 3778.