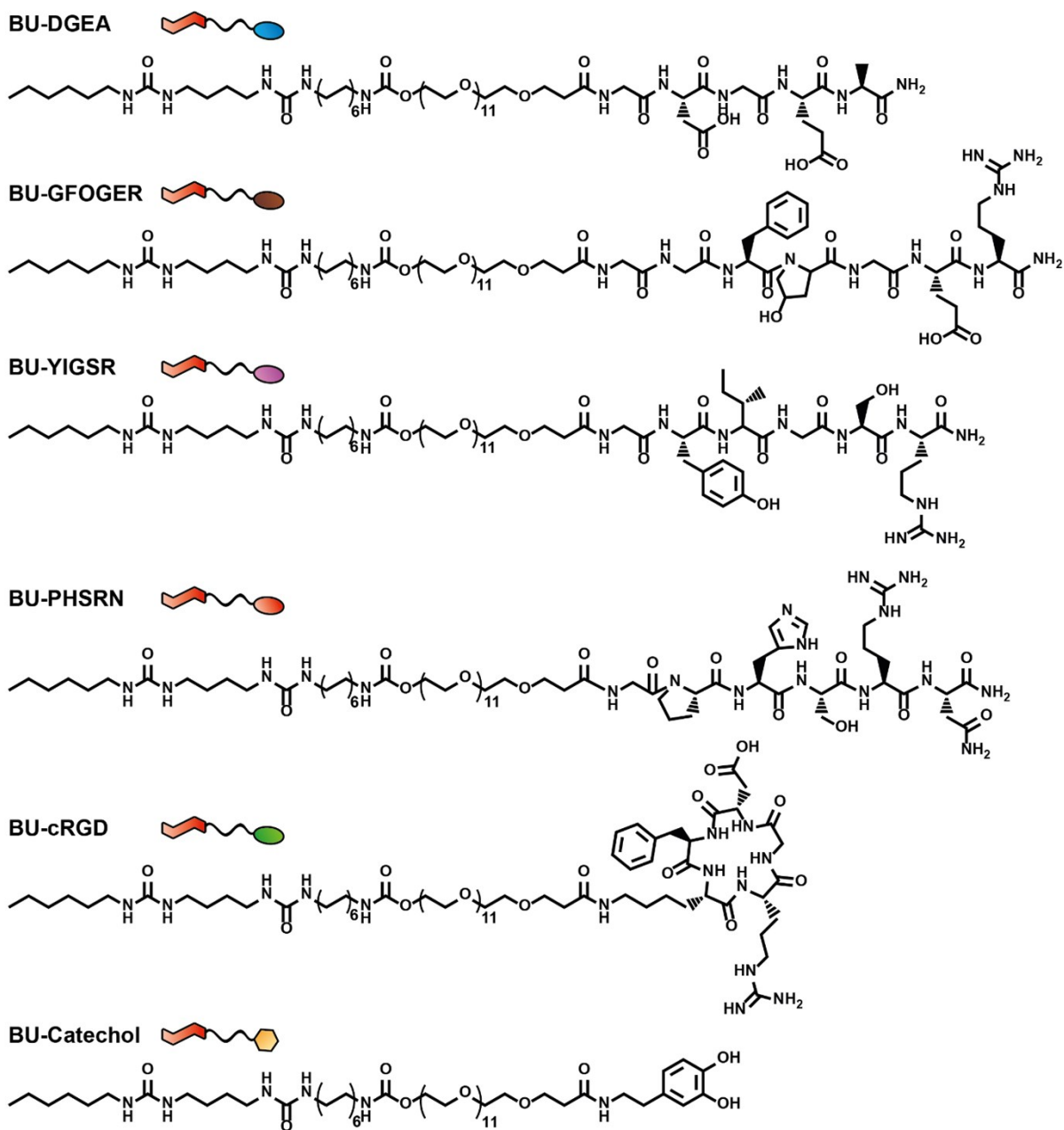


Supporting Information

Biomaterial screening of protein coatings and peptide additives: towards a simple synthetic mimic of a complex natural coating for a bio-artificial kidney

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Scheme S1. Chemical structures and schematic representations of BU-additives.

Table S1. ECM Library composition. “+” present, “-” absent in coating combination, *Not required for DoE analysis, performed as control. Sole presence of PCL-BU is termed Pristine.

Run	Collagen type IV	Laminin	Fibronectin	Collagen type I	L-DOPA
1	-	-	-	-	+
2	+	-	-	-	-
3	-	+	-	-	-
4	+	+	-	-	+
5	-	-	+	-	-
6	+	-	+	-	+
7	-	+	+	-	+
8	+	+	+	-	-
9	-	-	-	+	-
10	+	-	-	+	+
11	-	+	-	+	+
12	+	+	-	+	-
13	-	-	+	+	+
14	+	-	+	+	-
15	-	+	+	+	-
16	+	+	+	+	+
17*	-	-	-	-	-

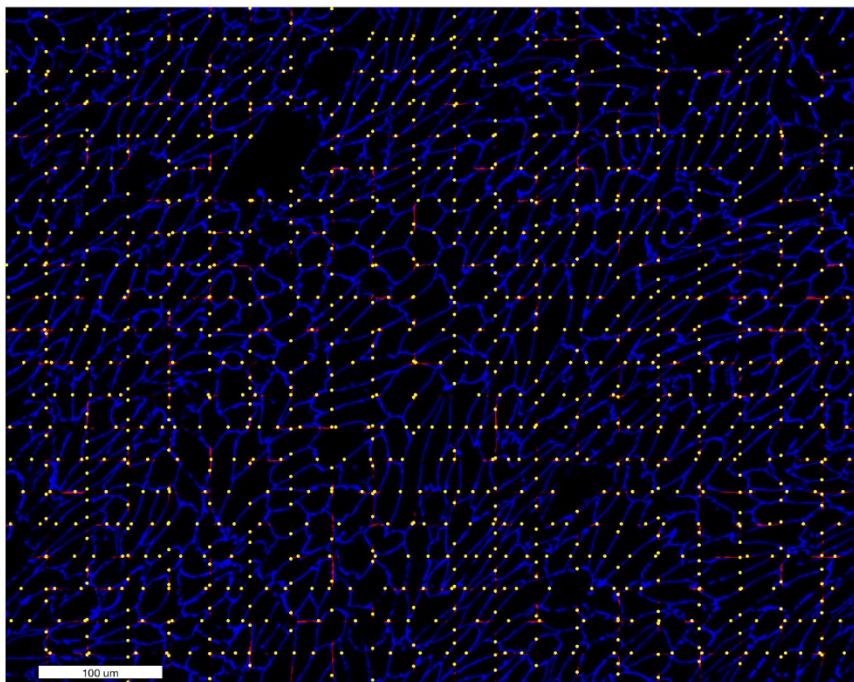


Fig. S1 Example of quantification method ZO-1 intersections.

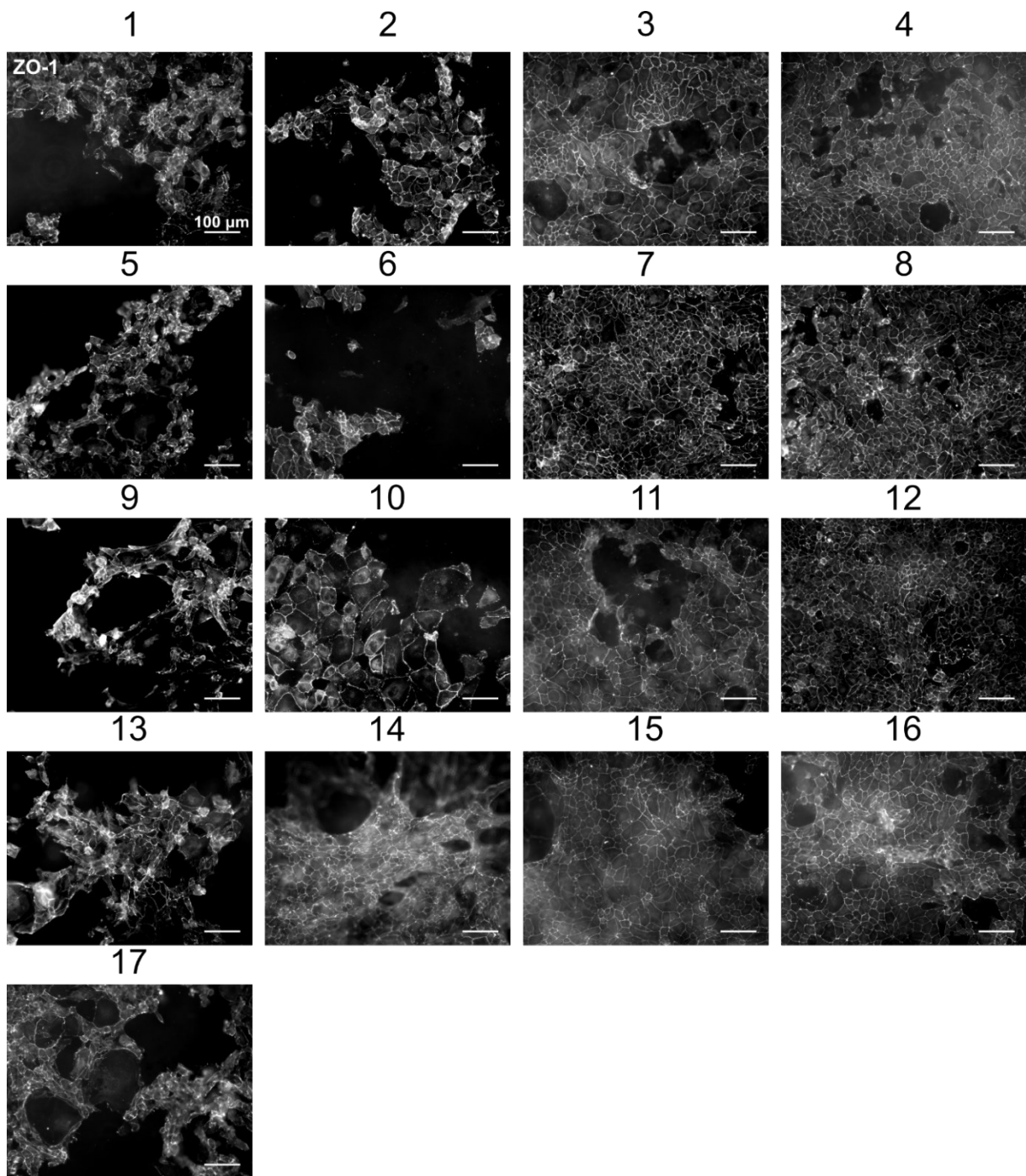


Fig. S2 Response of HK-2 to all ECM library conditions. HK-2 cells cultured for 3 weeks on biomaterial conditions listed in Table S1. Representative images of cells stained for Zona Occludens-1 (ZO-1), scale bar is 100 μ m, n=9.

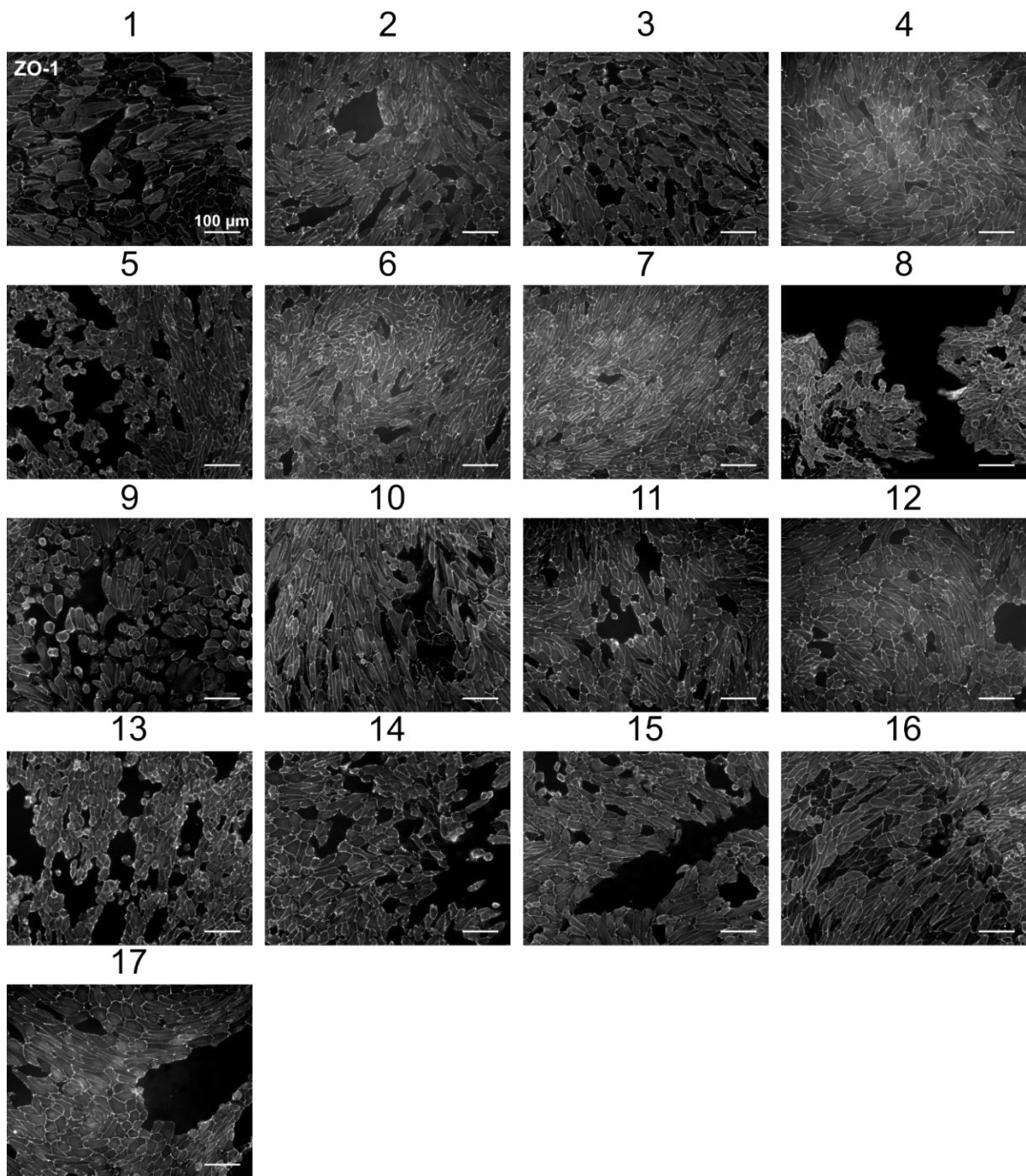


Fig. S3 Response of RPTEC to all ECM library conditions. RPTEC cells cultured for 3 weeks on biomaterial conditions listed in Table S1. Representative images of cells stained for Zona Occludens-1 (ZO-1), scale bar is 100 μm , n=9.

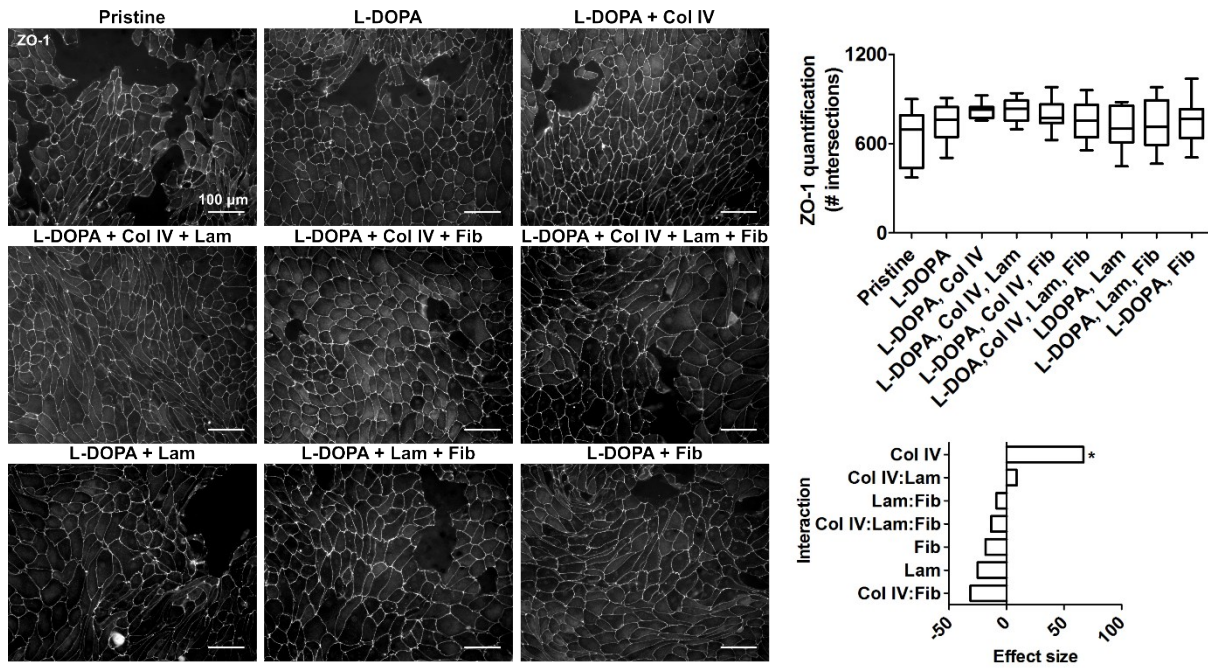
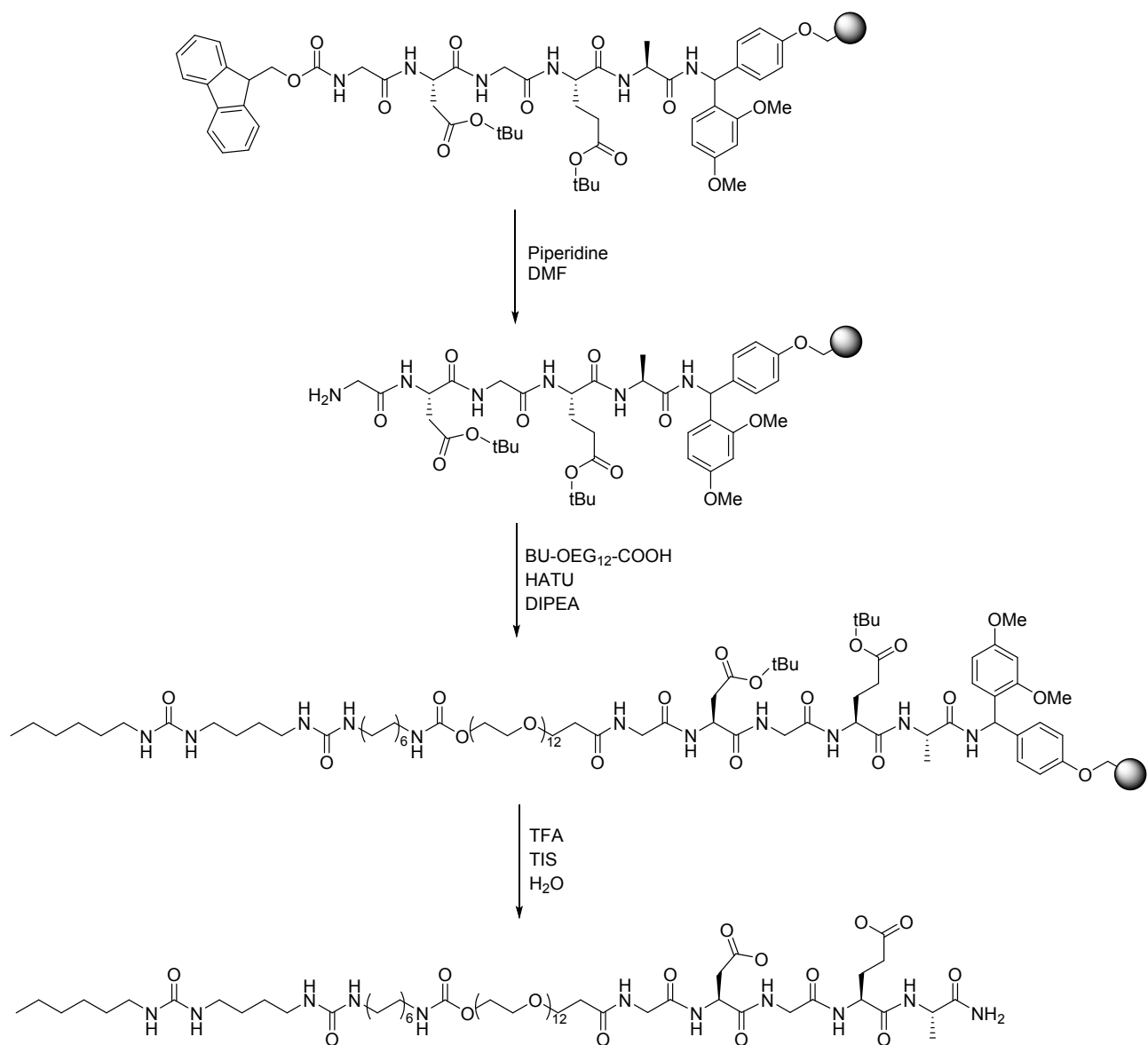


Fig. S4 Response of RPTEC to ECM proteins with L-DOPA constant. RPTEC cells cultured on biomaterial conditions coated with L-DOPA, Collagen type IV (Col IV), Laminin (Lam) or Fibronectin (Fib) or left pristine (no coating on PCL-BU) for 3 weeks. Representative images of cells stained for Zona Occludens-1 (ZO-1; Left), scale bar are 100 μm. Quantification of ZO-1 (Right top), effect size components (Right bottom), $n=3$, $*p\leq 0.05$.

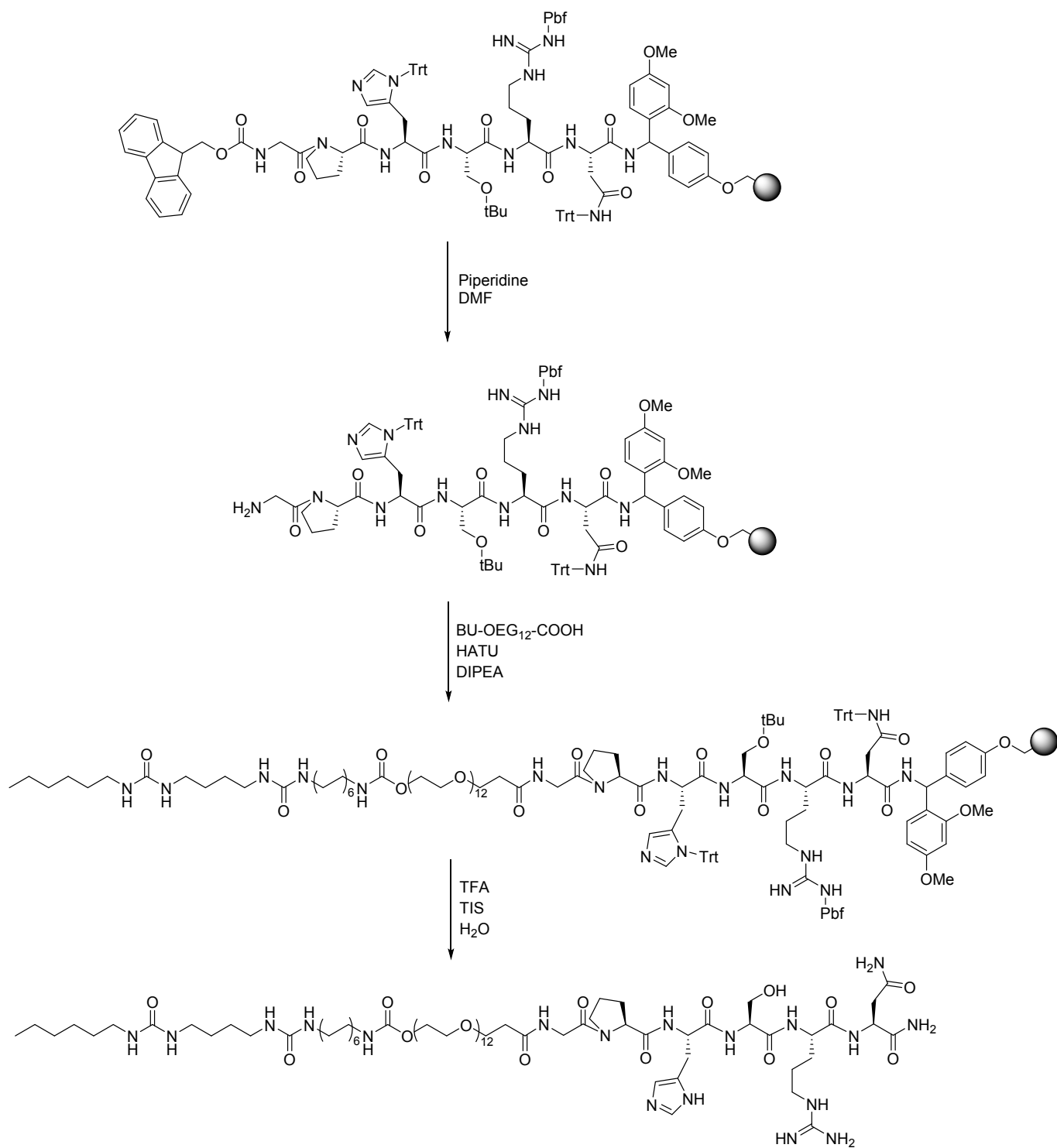
Synthesis bis-urea additives

BU-(OEG)₁₂-peptide

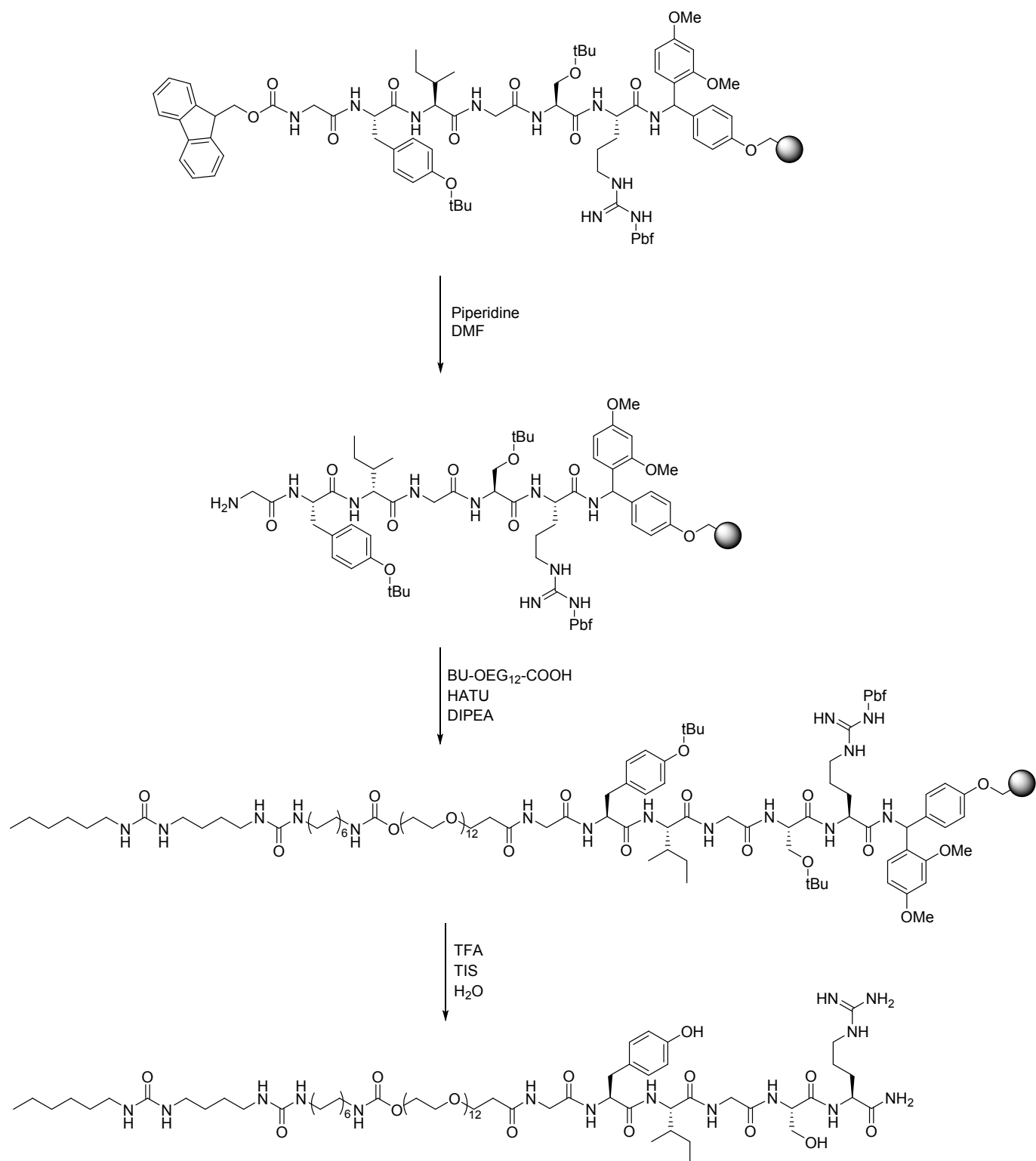
All chemicals were purchased from Sigma-Aldrich unless otherwise stated. Fmoc-Gly-Asp(OtBu)-Gly-Glu(OtBu)-Ala (40 mg, 0.05 mmol), Fmoc-Gly-Pro-His(Trt)-Ser(tBu)-Arg(Pbf)-Asn(Trt) (96 mg, 0.05 mmol), Fmoc-Gly-Tyr(tBu)-Ile-Gly-Ser(tBu)-Arg(Pbf) (62 mg, 0.05 mmol), or Fmoc-Gly-Gly-Phe-Hyp(tBu)-Gly-Glu(OtBu)-Arg(Pbf) (55 mg, 0.05 mmol) on rink amide resin (0.66 mmol/g) was deprotected 2x15 min with a 20 v/v% piperidine in DMF mixture. BU-OEG₁₂-COOH (108 mg; 0.1 mmol) was dissolved in 4 mL DMF. *N,N*-Diisopropylethylamine (44 μ l, 0.25 mmol) and HATU (38 mg, 0.1 mmol) were dissolved in 2 mL DMF, respectively, added to the BU-OEG₁₂-COOH mixture, and stirred for 30 min. After pre-activation, the reaction mixture was added to peptides on resin and reacted overnight under shaking conditions at room temperature. The BU-OEG₁₂-peptides were cleaved from the resin in a 95:2.5:2.5 v/v% TFA:TIS:H₂O mixture. The cleavage mixture was precipitated three times in ice-cold Et₂O. The precipitate was collected, dissolved in a mixture of water/acetonitrile, and freeze-dried yielding 58 mg BU-OEG₁₂-GDGEA (76 %), 49 mg BU-OEG₁₂-GPHSRN (52%), 37 mg BU-OEG₁₂-GYIGSR (43%), and 53 mg BU-OEG₁₂-GGFOGER (58 %) as fluffy white solids. BU-OEG₁₂-GDGEA: LC-MS(ESI) t_r = 6.10 min, m/z calcd (C₆₈H₁₂₇N₁₁O₂₆) 1513.90; found 505.67 [M+3H]³⁺, 758.00 [M+2H]²⁺, 1514.67 [M+H]⁺. BU-OEG₁₂-GPHSRN: LC-MS(ESI) t_r = 4.36 min, m/z calcd (C₇₈H₁₄₄N₁₈O₂₅) 1733.06; found 434.42 [M+4H]⁴⁺, 578.92 [M+3H]³⁺, 867.58 [M+2H]²⁺, 1733.06 [M+H]⁺. BU-OEG₁₂-GYIGSR: LC-MS(ESI) t_r = 4.65 min, m/z calcd (C₈₀H₁₄₇N₁₅O₂₅) 1718.07; found 573.92 [M+3H]³⁺, 860.42 [M+2H]²⁺, 1718.92 [M+H]⁺. BU-OEG₁₂-GGFOGER: LC-MS(ESI) t_r = 4.65 min, m/z calcd (C₈₃H₁₄₈N₁₆O₂₇) 1801.07; found 473.33 [M+4Na]⁴⁺, 601.58 [M+3H]³⁺, 901.58 [M+2H]²⁺, 1802.92 [M+H]⁺.



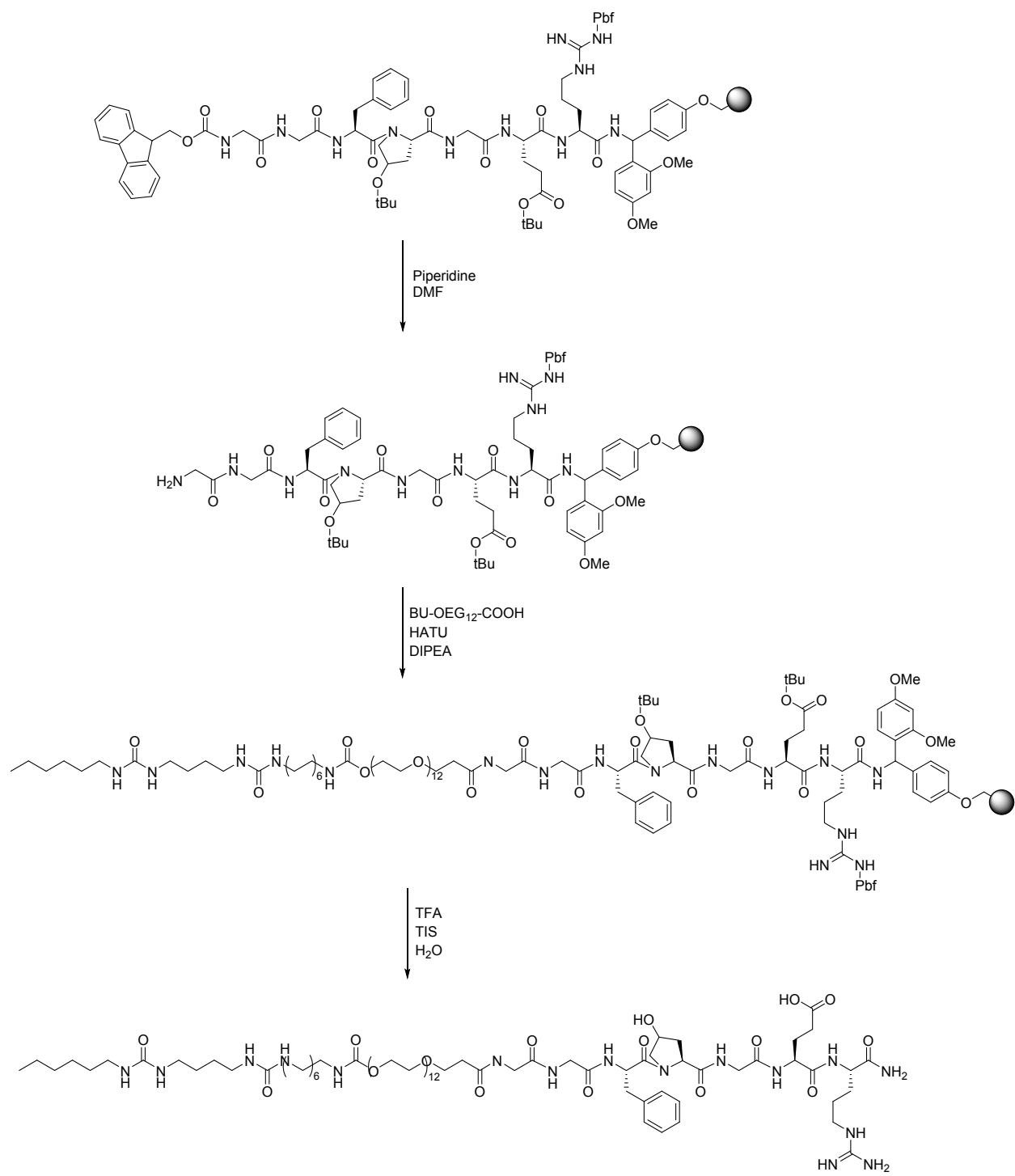
Scheme S2 Synthesis of BU-DGEA.



Scheme S3 Synthesis of BU-PHSRN.



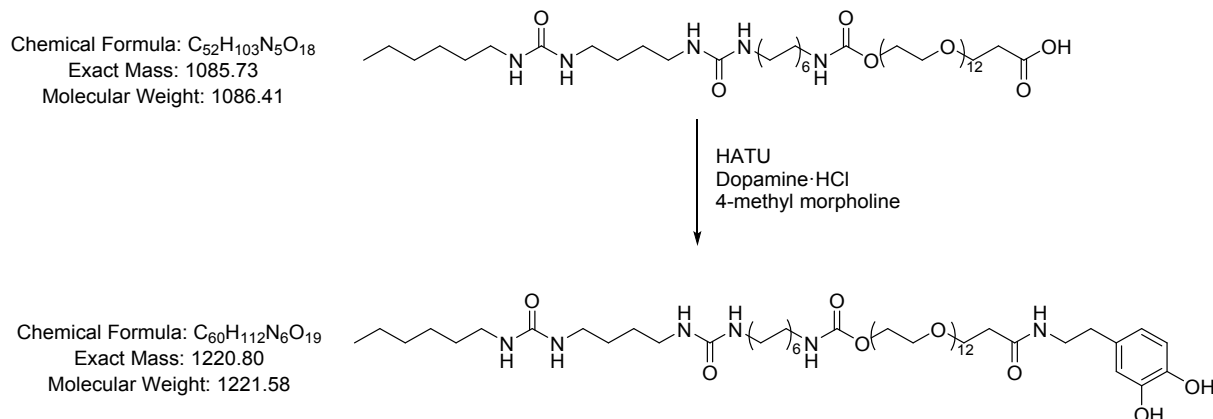
Scheme S4 Synthesis of BU-YIGSR.



Scheme S5 Synthesis of BU-GFOGER.

BU-(OEG)₁₂-Catechol

BU-OEG₁₂-COOH (100 mg; 0.09 mmol; SyMO-Chem) was dissolved in 4 mL DMF under argon. *N*-methylmorpholine (60 μ l, 0.54 mmol) and HATU (50 mg, 0.14 mmol) were added to the reaction mixture and stirred for 30 min. After pre-activation, 3,4-dihydroxyphenethylamine hydrochloride (26 mg, 0.14 mmol) was added and the reaction was stirred overnight under argon. The reaction mixture was precipitated three times in a 1% (v/v) solution of formic acid in water. The precipitate was collected, dissolved in mixture of water/acetonitrile and freeze-dried yielding 92 mg (84%) of the product as a fluffy white solid.



Scheme S6 Synthesis of BU-Catechol.

¹H NMR (400 MHz, DMSO) δ = 8.72 (bs, 1H), 8.63 (bs, 1H), 7.84 (NH), 7.16 (NH), 6.61 (d, *J* = 8.0 Hz, 1H), 6.56 (d, *J* = 2.0 Hz, 1H), 6.43 (dd, *J* = 8.0, 2.1 Hz, 1H), 5.73 (NH), 4.02 (bs, 2H), 3.60-3.40 (m, 48H), 3.17 (m, 2H), 2.94 (m, 10H), 2.50 (m, 2H), 2.28 (t, *J* = 6.44 Hz, 2H), 1.40-1.20 (m, 32H), 0.86 (t, *J* = 6.79 Hz, 3H)

¹³C NMR (100 MHz, DMSO) δ = 169.84, 158.08, 156.13, 145.03, 143.49, 130.20, 119.17, 115.92, 115.44, 69.78, 69.72, 69.67, 69.52, 68.91, 66.85, 62.98, 39.52, 36.17, 34.68, 31.07, 30.07, 30.03, 29.40, 29.09, 29.03, 28.85, 28.76, 27.61, 26.44, 26.26, 26.09, 22.11, 13.94.

LC-MS(ESI) *t_r* = 5.29 min, *m/z* calcd (C₆₀H₁₁₂N₆O₁₉) 1220.80; found 611.42 [M+2H]²⁺, 1221.50 [M+H]⁺.

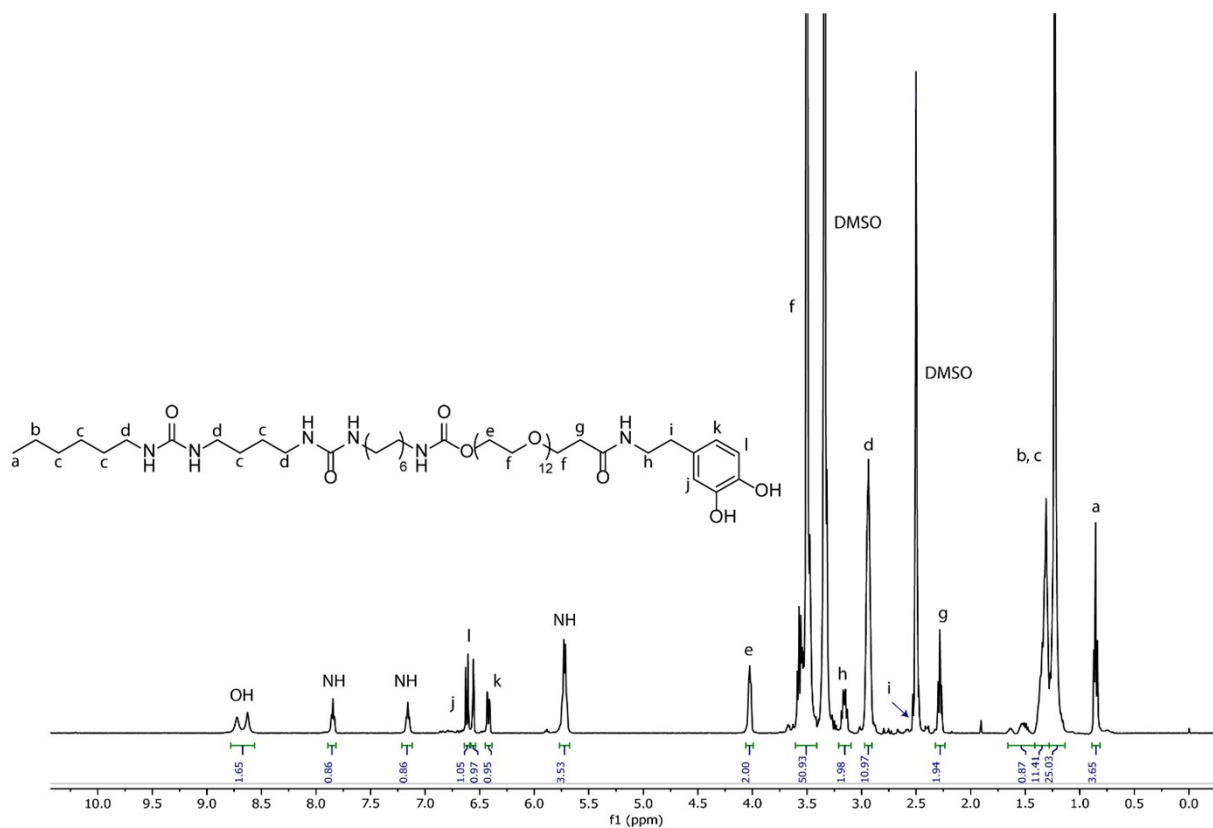


Fig. S5 ¹H-NMR spectrum of BU-Catechol.

Table S2 BU-additive library composition. “+” present, “-” absent in combination. Sole presence of PCL-BU is termed Pristine. 1 mol% per BU-peptide and/or 5 mol% per BU-catechol was/were employed.

Run	BU-GFOGER	BU-YIGSR	BU-cRGD	BU-DGEA	BU-Catechol	BU-PHSRN
1	-	-	-	-	-	-
2	+	-	-	-	-	+
3	-	+	-	-	-	+
4	+	+	-	-	-	-
5	-	-	+	-	-	+
6	+	-	+	-	-	-
7	-	+	+	-	-	-
8	+	+	+	-	-	+
9	-	-	-	+	-	+
10	+	-	-	+	-	-
11	-	+	-	+	-	-
12	+	+	-	+	-	+
13	-	-	+	+	-	-
14	+	-	+	+	-	+
15	-	+	+	+	-	+
16	+	+	+	+	-	-
17	-	-	-	-	+	+
18	+	-	-	-	+	-
19	-	+	-	-	+	-
20	+	+	-	-	+	+
21	-	-	+	-	+	-
22	+	-	+	-	+	+
23	-	+	+	-	+	+
24	+	+	+	-	+	-
25	-	-	-	+	+	-
26	+	-	-	+	+	+
27	-	+	-	+	+	+
28	+	+	-	+	+	-
29	-	-	+	+	+	+
30	+	-	+	+	+	-
31	-	+	+	+	+	-
32	+	+	+	+	+	+

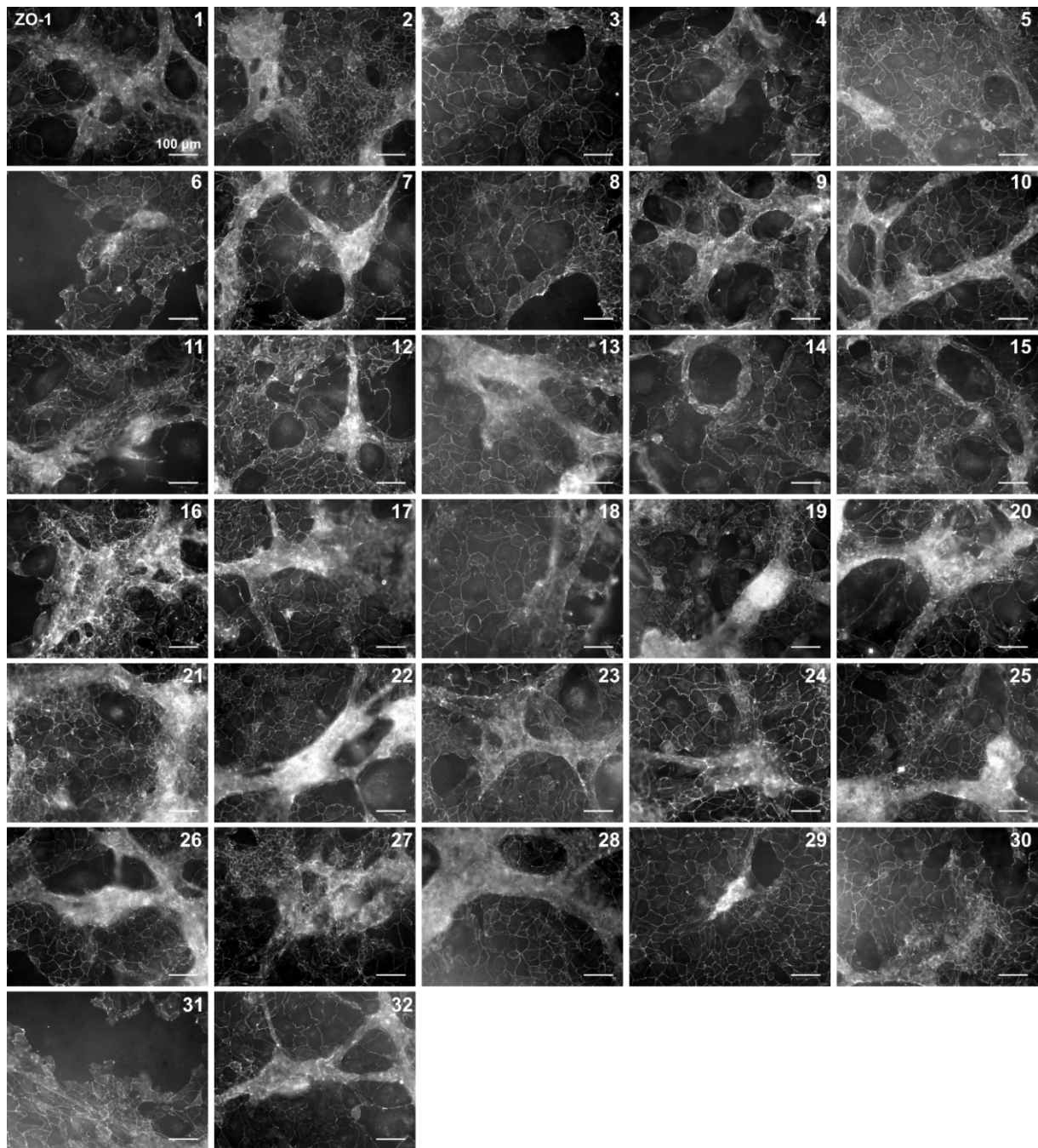


Fig. S6 HK-2 response to all BU-additive library conditions. HK-2 cells cultured for 3 weeks on biomaterial conditions listed in Table S2. Representative images of cells stained for ZO-1, scale bar is 100 μm , n=6.

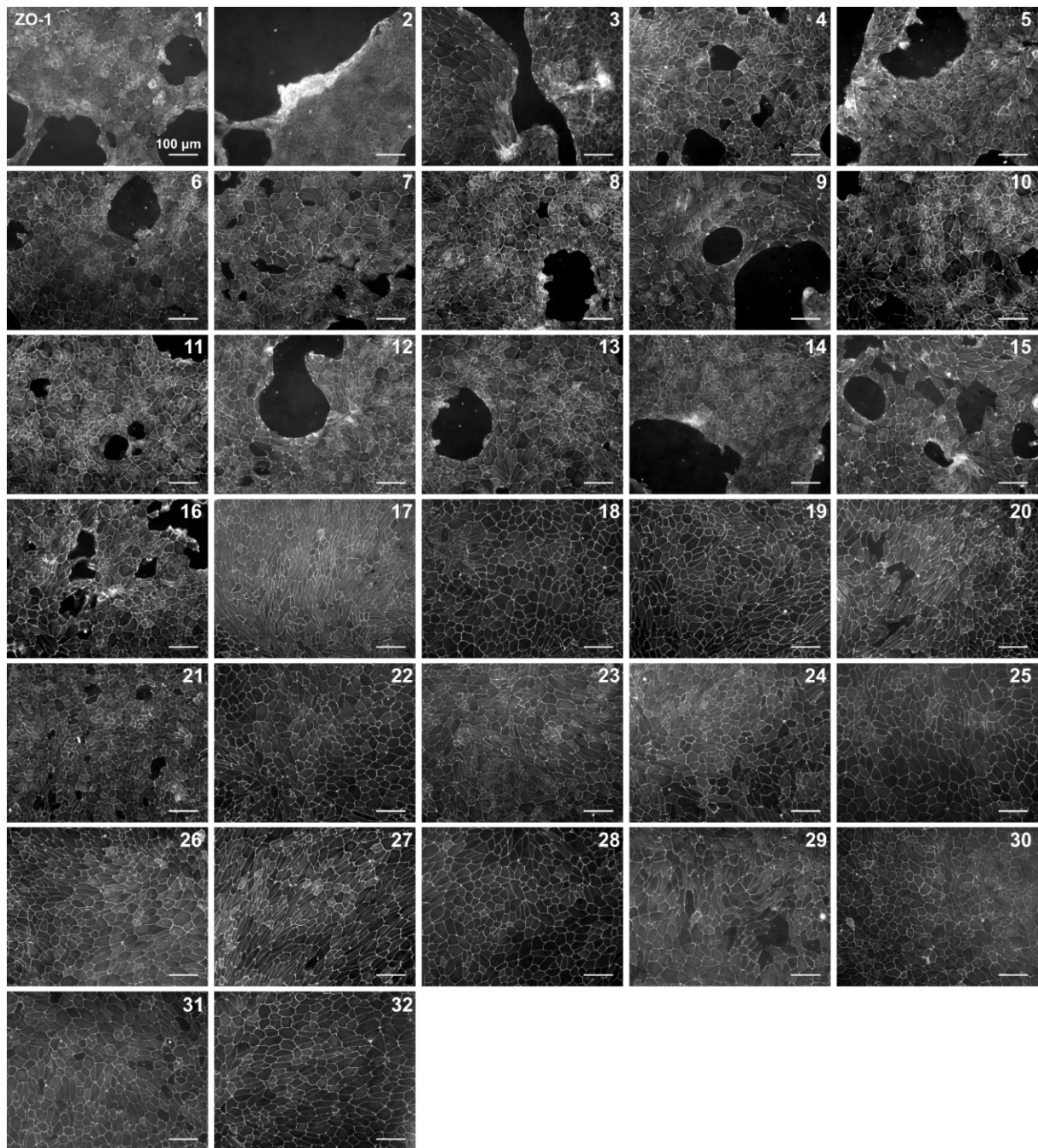


Fig. S7 RPTEC response to all BU-additive library conditions. RPTEC cells cultured for 3 weeks on biomaterial conditions listed in Table S2. Representative images of cells stained for Zona Occludens-1 (ZO-1), scale bar is 100 μm , n=6.

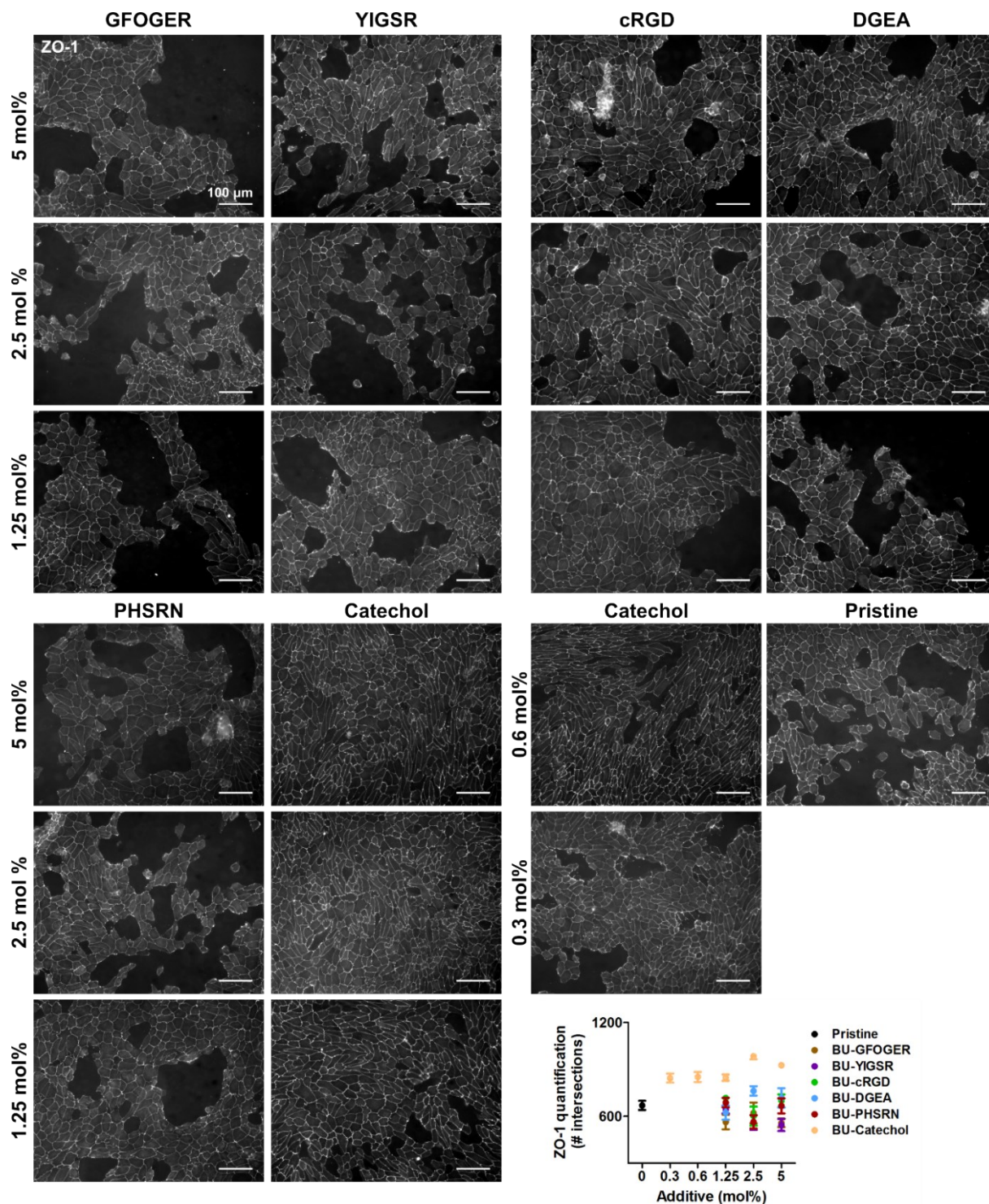


Fig. S8 Concentration range of BU-additives. RPTEC cells cultured for 3 weeks on biomaterials containing different concentrations of BU-additives. Representative images of cells stained for Zona Occludens-1 (ZO-1), and subsequently quantified, n=3.

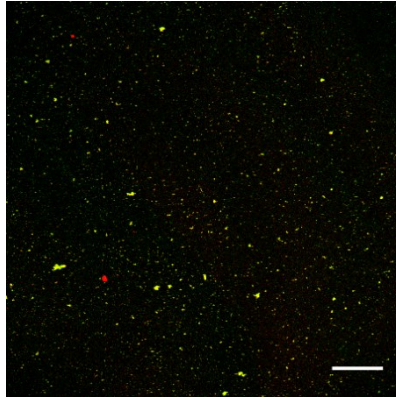


Fig. S9 Staining of L-DOPA with Collagen type IV and Laminin coating before cell culture. Laminin (green), Collagen type IV (red), scale bar is 100 μm .

Table S3. List of employed antibodies.

Antibody against	Supplier, catalogue number	Dilution
ZO-1	BD Biosciences, 610966	1:200
α Tubulin	Abcam, ab18251	1:200
Collagen type IV	Abcam ab6586	1:100
Laminin gamma 1	Abcam, ab17792	1:50
Megalin	Abcam, ab76969	1:200
Phalloidin-atto4888	Sigma-Aldrich, 49409	1:500
anti-mouse IgG (H+L) - AF555	Molecular probes, A21424	1:200
anti-mouse IgG (H+L) - AF647	Molecular probes, A21236	1:200
anti-rabbit IgG (H+L) - AF555	Molecular probes, A21428	1:200
anti-rabbit IgG (H+L) - AF647	Molecular probes, A21244	1:200
anti-Rat IgG (H+L) - AF488	Invitrogen, A11006	1:200
anti-Rat IgG (H+L) - AF555	Molecular Probes, A21434	1:200