

## Supporting Information

### Ni-Catalyzed Asymmetric Reduction of $\alpha$ -Keto- $\beta$ -Lactams via DKR Enabled by Proton Shuttling

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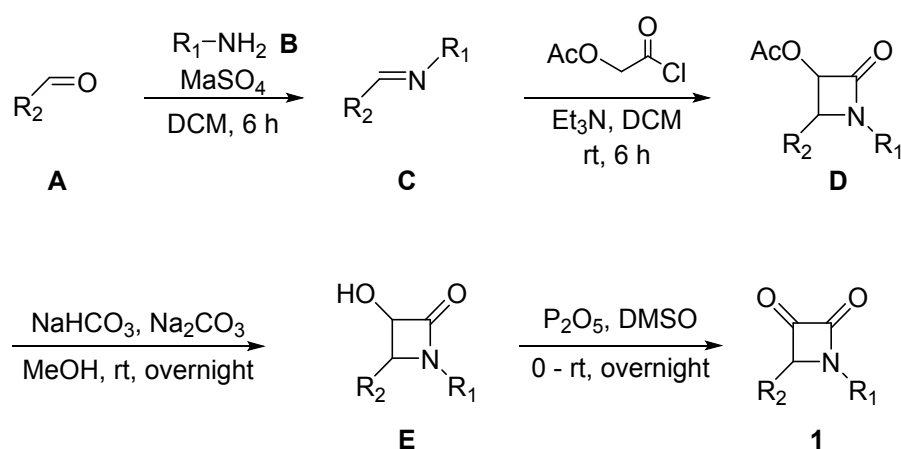
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## 1. General Information.

Unless otherwise mentioned, all experiments were carried out under an atmosphere of argon or using standard Schlenk techniques. Solvents and reagents were purchased from commercial suppliers and used without further purification. Column Chromatography was performed with silica gel Merck 60 (300-400 mesh). NMR spectra were recorded on a Bruker DPX 400 spectrometer at 400 MHz for  $^1\text{H}$  NMR, 101 MHz for  $^{13}\text{C}$  NMR and a Bruker DPX 600 spectrometer at 600 MHz for  $^1\text{H}$  NMR, 151 MHz for  $^{13}\text{C}$  NMR.  $\text{CDCl}_3$  and  $d^6$ -DMSO was the solvent used for the NMR analysis, with tetramethylsilane (TMS) as the internal standard. Chemical shifts are reported in ppm and coupling constants are given in Hz. Chemical shifts were reported relative to TMS (0.00 ppm) for  $^1\text{H}$  NMR and relative to  $\text{CDCl}_3$  (77.0 ppm) for  $^{13}\text{C}$  NMR. HPLC analysis was carried out on Agilent 1260 Series instrument using a chiral stationary phase IA or OD-3. PE refers to petroleum ether, and EA refers to ethyl acetate.

## 2. General Procedures for the Synthesis of Substrates.



**Scheme S1. Synthetic Routes of Substrates**

**Compound C**<sup>[1]</sup>: To a solution of aldehydes **A** (20 mmol, 1.0 equiv.), primary amines **B** (20 mmol, 1.0 equiv.) in DCM (50 mL) was added  $\text{MgSO}_4$  (40 mmol, 2.0 equiv.). The mixture was stirred at room temperature for 2-16 hours until the start materials was consumed completely, the mixture was filtered and the filter liquor was concentrated under vacuum. The crude product was obtained and used into the next step directly.

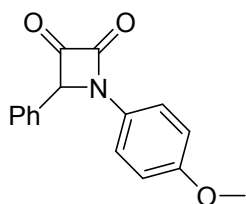
**Compound D**<sup>[1a]</sup>: To a solution of crude **C** (20 mmol, 1.0 equiv.) in DCM (50 mL) was added 2-chloro-2-oxoethyl acetate (30 mmol, 1.5 equiv.) and the mixture was cooled to 0 °C.  $\text{Et}_3\text{N}$  (60 mmol, 3.0 equiv.) was added slowly by syringe and then the mixture was stirred at room temperature for 6 hours until the start materials was consumed completely. The mixture was quenched by 20 mL  $\text{H}_2\text{O}$ , and then extracted by DCM (50 mL\*2). The combined organic layers were washed by  $\text{NH}_4\text{Cl}$  and brine, dried over anhydrous  $\text{Na}_2\text{SO}_4$ , filtered and concentrated under vacuum. The crude product was purified by chromatography ( $\text{SiO}_2$ , PE/EA from 20:1 to 4:1). Compound **D** was obtained as a light-yellow solid (57 - 84% yield).

**Compound E**<sup>[1a]</sup>: To a solution of **D** (10 mmol, 1.0 equiv.) in MeOH (30 mL) was added saturated  $\text{NaHCO}_3$  aqueous solution (15 mL), then  $\text{Na}_2\text{CO}_3$  (0.3 equiv.) was added into the mixture and the mixture was stirred at room temperature overnight until the start materials was consumed completely. The mixture was quenched by 20 mL  $\text{H}_2\text{O}$ , and the obtained clear liquid was concentrated under vacuum to remove most

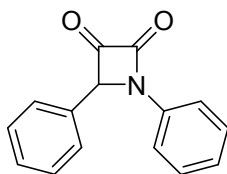
MeOH. After the above process, the mixture was extracted by DCM (50 mL\*2), the combined organic layer was washed by NH<sub>4</sub>Cl and brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under vacuum. The crude product was used into the next step without further purification.

**Compound 1** <sup>[1b]</sup>: To a solution of **E** (10 mmol, 1.0 equiv.) in DMSO (20 mL) was cooled to 0 °C and added P<sub>2</sub>O<sub>5</sub> (15 mol, 1.5 equiv.) under Ar. The mixture was stirred at room temperature overnight until the start material **E** was consumed completely. The mixture was quenched by saturated NaHCO<sub>3</sub> aqueous solution (20 mL) carefully, and extracted by DCM (50 mL\*2). The combined organic layer was washed by NH<sub>4</sub>Cl and brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under vacuum to give the crude product. The crude product was purified by chromatography (SiO<sub>2</sub>, PE/EA from 15:1 to 2:1). Compound **1** was obtained as a yellow solid (56 - 82% yield).

#### Characterization Data of Substrate

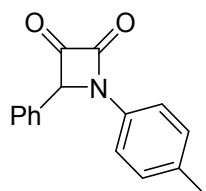


**1-(4-methoxyphenyl)-4-phenylazetidine-2,3-dione (1a)**: This is a known compound.<sup>[1b]</sup> Yellow powder, 2.1 g, 78% yield. <sup>1</sup>H NMR (600 MHz, Chloroform-*d*) δ 7.52 – 7.35 (m, 5H), 7.35 – 7.30 (m, 2H), 6.88 (dd, *J* = 9.0, 1.6 Hz, 2H), 5.55 (s, 1H), 3.78 (s, 3H). <sup>13</sup>C NMR (151 MHz, Chloroform-*d*) δ 190.6, 160.0, 158.0, 131.7, 129.9, 129.5, 129.4, 126.4, 119.8, 114.8, 74.9, 55.5. HRMS (ESI<sup>+</sup>), *m/z* 268.0965 ([*M*+*H*]<sup>+</sup>), calcd for C<sub>16</sub>H<sub>14</sub>NO<sub>3</sub><sup>+</sup>: 268.0968.

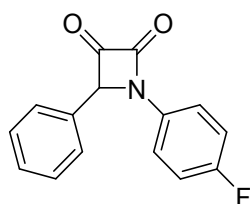


**1,4-diphenylazetidine-2,3-dione (1b)**: This is a known compound.<sup>[1b]</sup> White powder, 1.66 g, 70% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.51 (dd, *J* = 7.8, 1.7 Hz, 2H),

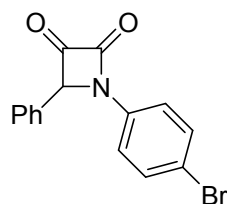
7.45 – 7.28 (m, 7H), 7.22 (t,  $J = 7.4$  Hz, 1H), 5.60 (s, 1H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  190.9, 160.6, 136.4, 131.6, 129.6, 129.6, 129.5, 126.6, 126.3, 118.3, 74.9. HRMS (ESI<sup>+</sup>),  $m/z$  238.0859 ( $[\text{M}+\text{H}]^+$ ), calcd for  $\text{C}_{15}\text{H}_{12}\text{NO}_2^+$ : 238.0863.



**4-phenyl-1-(*p*-tolyl) azetidione (1c):** This is a new compound. Yellow powder, 2.05 g, 82% yield.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.45 – 7.28 (m, 7H), 7.21 – 7.12 (m, 2H), 5.56 (s, 1H), 2.32 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  190.9, 160.4, 136.8, 134.0, 131.7, 130.1, 129.5, 129.4, 126.3, 118.3, 74.9, 21.2. HRMS (ESI<sup>+</sup>),  $m/z$  252.1015 ( $[\text{M}+\text{H}]^+$ ), calcd for  $\text{C}_{16}\text{H}_{14}\text{NO}_2^+$ : 252.1019.

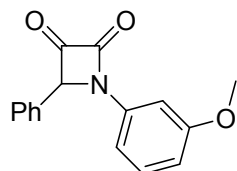


**1-(4-fluorophenyl)-4-phenylazetidione (1d):** This is a known compound. <sup>[1a]</sup> Yellow powder, 1.58 g, 62% yield.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.54 – 7.46 (m, 2H), 7.46 – 7.38 (m, 3H), 7.35 – 7.28 (m, 2H), 7.11 – 7.02 (m, 2H), 5.58 (s, 1H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  190.5, 160.3, 161.7, 159.3 (d,  $J = 248.5$  Hz), 132.7, 132.7 (d,  $J = 3.0$  Hz), 131.3, 129.7, 129.6, 126.3, 120.0, 119.9 (d,  $J = 8.1$  Hz), 116.7, 116.5 (d,  $J = 23.2$  Hz). 75.1. HRMS (ESI<sup>+</sup>),  $m/z$  256.0764 ( $[\text{M}+\text{H}]^+$ ), calcd for  $\text{C}_{15}\text{H}_{11}\text{FNO}_2^+$ : 256.0768.

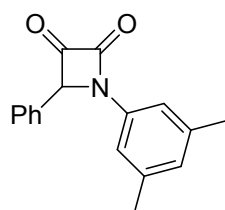


**1-(4-bromophenyl)-4-phenylazetidione (1e):** This is a known compound. <sup>[4]</sup> Yellow powder, 1.85 g, 59% yield.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.52 – 7.45

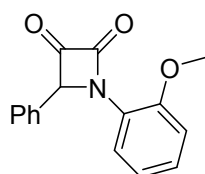
(m, 2H), 7.45 – 7.36 (m, 5H), 7.34 – 7.28 (m, 2H), 5.58 (s, 1H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 190.4, 160.6, 135.3, 132.7, 131.1, 129.8, 129.6, 126.3, 119.7, 119.7, 74.9. HRMS (ESI<sup>+</sup>), *m/z* 315.9964 ([M+H]<sup>+</sup>), calcd for C<sub>15</sub>H<sub>11</sub>BrNO<sub>2</sub><sup>+</sup>: 315.9968.



**1-(3-methoxyphenyl)-4-phenylazetidine-2,3-dione (1f):** This is a new compound. Yellow powder, 1.50 g, 56% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.44 – 7.37 (m, 3H), 7.35 – 7.29 (m, 2H), 7.25 – 7.20 (m, 2H), 6.96 – 6.90 (m, 1H), 6.80 – 6.74 (m, 1H), 5.58 (s, 1H), 3.78 (s, 3H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 190.9, 160.7, 160.4, 137.4, 131.6, 130.4, 129.6, 129.5, 126.3, 112.6, 110.6, 104.1, 75.1, 55.4. HRMS (ESI<sup>+</sup>), *m/z* 268.0965 ([M+H]<sup>+</sup>), calcd for C<sub>16</sub>H<sub>14</sub>NO<sub>3</sub><sup>+</sup>: 268.0968.

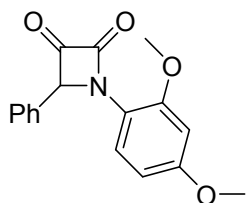


**1-(3,5-dimethylphenyl)-4-phenylazetidine-2,3-dione (1g):** This is a new compound. Yellow powder, 1.96 g, 0.74% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.45 – 7.34 (m, 3H), 7.35 – 7.28 (m, 2H), 7.15 – 7.09 (m, 2H), 6.86 (dt, *J* = 1.7, 0.8 Hz, 1H), 5.57 (s, 1H), 2.28 (s, 6H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 191.0, 160.6, 139.5, 136.3, 131.8, 129.4, 128.6, 126.2, 116.0, 74.9, 21.4. HRMS (ESI<sup>+</sup>), *m/z* 266.1171 ([M+H]<sup>+</sup>), calcd for C<sub>17</sub>H<sub>16</sub>NO<sub>2</sub><sup>+</sup>: 266.1176.

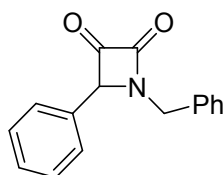


**1-(2-methoxyphenyl)-4-phenylazetidine-2,3-dione (1h):** This is a known compound. <sup>[4]</sup> Yellow powder, 1.52 g, 57% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 8.06 (dd, *J*

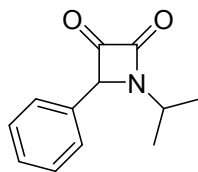
= 7.9, 1.7 Hz, 1H), 7.35 – 7.28 (m, 3H), 7.24 – 7.19 (m, 3H), 7.08– 6.98 (m, 1H), 6.89– 6.83 (m, 1H), 5.95 (s, 1H), 3.66 (s, 3H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 193.6, 162.3, 149.8, 133.6, 129.1, 129.0, 128.4, 126.4, 124.3, 123.2, 121.4, 112.3, 77.7, 55.6. HRMS (ESI<sup>+</sup>), *m/z* 268.0965 ([M+H]<sup>+</sup>), calcd for C<sub>16</sub>H<sub>14</sub>NO<sub>3</sub><sup>+</sup>: 268.0968.



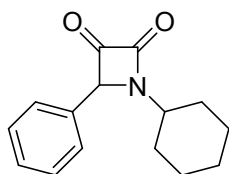
**1-(2,4-dimethoxyphenyl)-4-phenylazetidine-2,3-dione (1i):** This is a new compound. Yellow powder, 1.75 g, 59% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.93 (d, *J* = 8.8 Hz, 1H), 7.36 – 7.28 (m, 3H), 7.25 – 7.19 (m, 2H), 6.54 (dd, *J* = 8.8, 2.6 Hz, 1H), 6.40 (d, *J* = 2.6 Hz, 1H), 5.88 (s, 1H), 3.79 (s, 3H), 3.67 (s, 3H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 193.5, 162.0, 159.9, 151.4, 133.5, 129.0, 129.0, 126.5, 124.2, 117.7, 105.0, 99.7, 77.4, 55.6. HRMS (ESI<sup>+</sup>), *m/z* 298.1070 ([M+H]<sup>+</sup>), calcd for C<sub>17</sub>H<sub>16</sub>NO<sub>4</sub><sup>+</sup>: 298.1074.



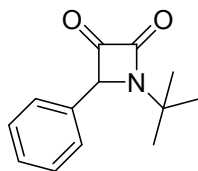
**1-benzyl-4-phenylazetidine-2,3-dione (1j):** This is a known compound. <sup>[5]</sup> White powder, 1.43 g, 57% yield. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.44 – 7.14 (m, 10H), 5.29 (s, 1H), 5.02 (d, *J* = 15.3 Hz, 1H), 4.37 (d, *J* = 15.4 Hz, 1H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 195.0, 164.4, 134.7, 133.1, 129.5, 129.1, 128.8, 128.3, 127.6, 73.8, 45.4. HRMS (ESI<sup>+</sup>), *m/z* 252.1015 ([M+H]<sup>+</sup>), calcd for C<sub>16</sub>H<sub>14</sub>NO<sub>2</sub><sup>+</sup>: 252.1019.



**1-isopropyl-4-phenylazetidine-2,3-dione (1k):** This is a known compound. <sup>[1a]</sup> White powder, 1.14 g, 56% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.48 – 7.36 (m, 3H), 7.34 – 7.25 (m, 2H), 5.13 (s, 1H), 4.19 (p, *J* = 6.7 Hz, 1H), 1.42 (d, *J* = 6.7 Hz, 3H), 1.17 (d, *J* = 6.7 Hz, 3H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  193.5, 163.9, 133.8, 129.5, 129.2, 127.1, 73.3, 46.6, 21.4, 20.1. HRMS (ESI<sup>+</sup>), *m/z* 204.1017 ([M+H]<sup>+</sup>), calcd for C<sub>12</sub>H<sub>14</sub>NO<sub>2</sub><sup>+</sup>: 204.1019.

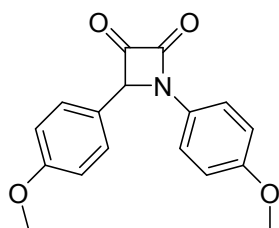


**1-cyclohexyl-4-phenylazetidine-2,3-dione (1l):** This is a known compound. <sup>[4]</sup> White powder, 1.85 g, 76% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.48 – 7.36 (m, 3H), 7.34 – 7.24 (m, 2H), 5.13 (s, 1H), 3.94 – 3.75 (m, 1H), 2.09 – 1.99 (m, 1H), 1.92 – 1.77 (m, 2H), 1.75 – 1.67 (m, 2H), 1.65 – 1.55 (m, 1H), 1.39 – 1.03 (m, 4H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  193.5, 164.0, 133.9, 129.5, 129.2, 127.1, 77.3, 73.5, 54.3, 31.6, 30.4, 25.0, 24.9. HRMS (ESI<sup>+</sup>), *m/z* 244.1328 ([M+H]<sup>+</sup>), calcd for C<sub>15</sub>H<sub>18</sub>NO<sub>2</sub><sup>+</sup>: 244.1332.

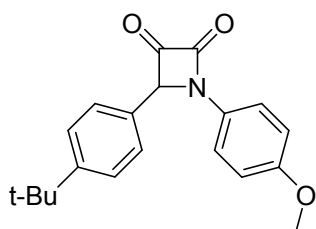


**1-(tert-butyl)-4-phenylazetidine-2,3-dione (1m):** This is a new compound. white powder, 1.56 g, 72% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.46 – 7.36 (m, 3H), 7.35 – 7.26 (m, 2H), 5.15 (s, 1H), 1.41 (s, 9H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  193.9, 163.7, 135.0, 129.4, 129.2, 127.0, 74.1, 56.8, 28.4. HRMS (ESI<sup>+</sup>), *m/z* 218.1173 ([M+H]<sup>+</sup>), calcd for C<sub>13</sub>H<sub>16</sub>NO<sub>2</sub><sup>+</sup>: 218.1176.

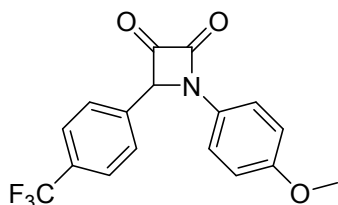




**1,4-bis(4-methoxyphenyl) azetidine-2,3-dione (1n):** This is a known compound. <sup>[1b]</sup> Yellow powder, 2.1 g, 72% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.46 (d, *J* = 9.1 Hz, 2H), 7.29 – 7.21 (m, 2H), 6.96 – 6.85 (m, 4H), 5.51 (s, 1H), 3.79 (s, 6H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 191.5, 160.6, 160.2, 157.9, 129.9, 127.8, 123.6, 119.8, 114.9, 114.7, 74.6, 55.5, 55.4. HRMS (ESI<sup>+</sup>), *m/z* 298.1070 ([M+H]<sup>+</sup>), calcd for C<sub>17</sub>H<sub>16</sub>NO<sub>4</sub><sup>+</sup>: 298.1074.

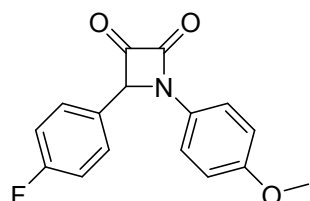


**4-(4-(tert-butyl)phenyl)-1-(4-methoxyphenyl)azetidine-2,3-dione (1o):** This is a new compound. Yellow powder, 2.4 g, 75% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.52 – 7.45 (m, 2H), 7.43 – 7.38 (m, 2H), 7.26 – 7.20 (m, 2H), 6.92 – 6.84 (m, 2H), 5.53 (s, 1H), 3.79 (s, 3H), 1.30 (s, 9H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 191.10, 160.12, 157.94, 152.71, 130.03, 128.61, 126.39, 126.09, 119.78, 114.74, 55.51, 34.74, 31.22. HRMS (ESI<sup>+</sup>), *m/z* 324.1591 ([M+H]<sup>+</sup>), calcd for C<sub>20</sub>H<sub>22</sub>NO<sub>3</sub><sup>+</sup>: 324.1594.

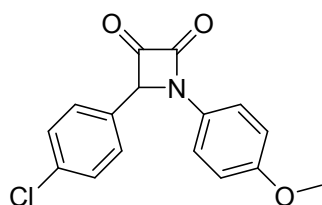


**1-(4-methoxyphenyl)-4-(4-(trifluoromethyl)phenyl)azetidine-2,3-dione (1p):** This is a new compound. Yellow powder, 1.87 g, 56% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.67 (d, *J* = 8.1 Hz, 2H), 7.49 – 7.38 (m, 4H), 6.96 – 6.86 (m, 2H),

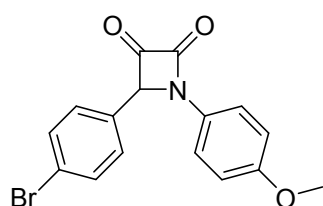
5.62 (s, 1H), 3.80 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  189.2, 159.5, 158.2, 135.7, 131.8, 131.5, 129.6, 126.6, {126.5, 126.4, 126.4, 126.4 (q,  $J = 3.5$  Hz)}, 125.0, 122.3, 119.7, 114.9, 74.2, 55.5. HRMS (ESI<sup>+</sup>),  $m/z$  336.0836 ([M+H]<sup>+</sup>), calcd for  $\text{C}_{17}\text{H}_{13}\text{F}_3\text{NO}_3^+$ : 336.0842.



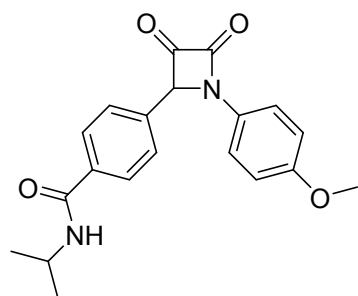
**4-(4-fluorophenyl)-1-(4-methoxyphenyl) azetidine-2,3-dione (1q):** This is a new compound. Yellow powder, 1.9 g, 67% yield.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.46 – 7.41 (m, 2H), 7.34 – 7.27 (m, 2H), 7.14 – 7.05 (m, 2H), 6.92 – 6.86 (m, 2H), 5.55 (s, 1H), 3.79 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  190.5, 164.6, 162.1 (d,  $J = 250.5$  Hz), 159.8, 158.1, 129.8, 128.3, 128.2 (d,  $J = 9.1$  Hz), 127.6 (d,  $J = 3.0$  Hz), 119.7, 116.7, 116.5 (d,  $J = 22.2$  Hz), 114.8, 74.1, 55.5. HRMS (ESI<sup>+</sup>),  $m/z$  286.0871 ([M+H]<sup>+</sup>), calcd for  $\text{C}_{16}\text{H}_{13}\text{FNO}_3^+$ : 286.0874.



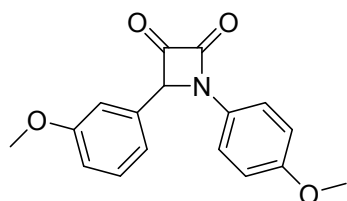
**4-(4-chlorophenyl)-1-(4-methoxyphenyl) azetidine-2,3-dione (1r):** This is a new compound. Yellow powder, 1.81 g, 60% yield.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.41 (dd,  $J = 18.8, 8.8$  Hz, 4H), 7.32 – 7.20 (m, 3H), 6.89 (d,  $J = 9.1$  Hz, 2H), 5.53 (s, 1H), 3.80 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  190.0, 159.7, 158.1, 135.6, 130.3, 129.7, 129.7, 127.7, 119.7, 114.9, 55.5. HRMS (ESI<sup>+</sup>),  $m/z$  302.0575 ([M+H]<sup>+</sup>), calcd for  $\text{C}_{16}\text{H}_{13}\text{ClNO}_3^+$ : 302.0578.



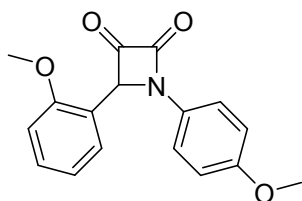
**4-(4-bromophenyl)-1-(4-methoxyphenyl) azetidine-2,3-dione (1s):** This is a known compound. <sup>12</sup> Yellow powder, 1.93 g, 56% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.60 – 7.51 (m, 2H), 7.47 – 7.36 (m, 2H), 7.25 – 7.14 (m, 2H), 6.95 – 6.85 (m, 2H), 5.51 (s, 1H), 3.80 (s, 3H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 189.9, 159.7, 158.1, 132.7, 130.8, 129.7, 127.9, 123.7, 119.7, 114.9, 55.6. HRMS (ESI<sup>+</sup>), *m/z* 346.0070 ([M+H]<sup>+</sup>), calcd for C<sub>16</sub>H<sub>13</sub>BrNO<sub>3</sub><sup>+</sup>: 346.0073.



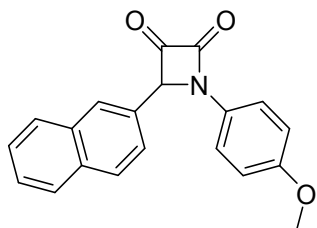
**4-(tert-butyl)-1-(4-methoxyphenyl)azetidine-2,3-dione (1t):** This is a new compound. Yellow powder, 1.4 g, 40% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.8 (d, *J* = 8.1 Hz, 2H), 7.4 (dd, *J* = 25.5, 8.6 Hz, 4H), 6.9 (d, *J* = 9.0 Hz, 2H), 6.1 (d, *J* = 7.4 Hz, 1H), 5.6 (s, 1H), 4.3 (dq, *J* = 13.3, 6.6 Hz, 1H), 3.8 (s, 3H), 1.2 (dd, *J* = 6.5, 1.8 Hz, 6H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 189.8, 165.9, 159.7, 158.1, 136.2, 134.9, 129.7, 128.0, 126.5, 119.7, 114.8, 74.5, 55.5, 42.1, 22.8. HRMS (ESI<sup>+</sup>), *m/z* 353.1490 ([M+H]<sup>+</sup>), calcd for C<sub>20</sub>H<sub>21</sub>N<sub>2</sub>O<sub>4</sub><sup>+</sup>: 353.1496.



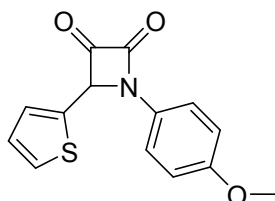
**4-(3-methoxyphenyl)-1-(4-methoxyphenyl)azetidine-2,3-dione (1u):** This is a new compound. Yellow powder, 1.9 g, 65% yield. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.51 – 7.40 (m, 2H), 7.31 (t, *J* = 8.0 Hz, 1H), 6.97 – 6.76 (m, 5H), 5.51 (s, 1H), 3.79 (s, 3H), 3.78 (s, 3H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 190.5, 160.4, 160.0, 158.0, 133.3, 130.6, 129.9, 119.7, 118.6, 114.9, 114.8, 112.0, 55.5, 55.4. HRMS (ESI<sup>+</sup>), *m/z* 298.1070 ([M+H]<sup>+</sup>), calcd for C<sub>17</sub>H<sub>16</sub>NO<sub>4</sub><sup>+</sup>: 298.1074.



**4-(2-methoxyphenyl)-1-(4-methoxyphenyl)azetidine-2,3-dione (1v):** This is a new compound. Yellow powder, 2.1 g, 72% yield.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.48 – 7.42 (m, 2H), 7.38 – 7.30 (m, 1H), 7.30 – 7.23 (m, 1H), 7.01 – 6.92 (m, 1H), 6.92 – 6.87 (m, 1H), 6.87 – 6.81 (m, 2H), 5.71 (s, 1H), 3.81 (s, 3H), 3.76 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  192.2, 160.8, 158.2, 157.6, 131.1, 130.3, 129.9, 121.1, 120.2, 119.4, 114.6, 111.6, 72.4, 55.9, 55.5. HRMS (ESI<sup>+</sup>),  $m/z$  298.1070 ([M+H]<sup>+</sup>), calcd for C<sub>17</sub>H<sub>16</sub>NO<sub>4</sub><sup>+</sup>: 298.1074.

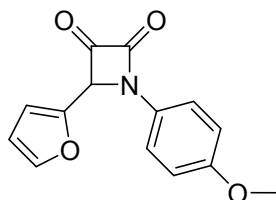


**1-(4-methoxyphenyl)-4-(naphthalen-2-yl)azetidine-2,3-dione (1w):** This is a new compound. Yellow powder, 2.6 g, 82% yield.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.92 – 7.77 (m, 4H), 7.57 – 7.46 (m, 4H), 7.41 – 7.34 (m, 1H), 6.91 – 6.82 (m, 2H), 5.70 (s, 1H), 3.76 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  190.6, 160.1, 158.0, 133.7, 133.3, 130.0, 129.6, 129.3, 128.0, 127.9, 127.0, 126.4, 123.0, 119.8, 114.8, 55.5. HRMS (ESI<sup>+</sup>),  $m/z$  318.1119 ([M+H]<sup>+</sup>), calcd for C<sub>20</sub>H<sub>16</sub>NO<sub>3</sub><sup>+</sup>: 318.1125.

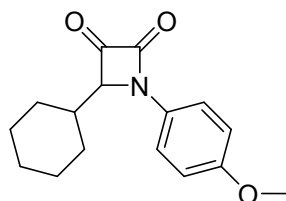


**1-(4-methoxyphenyl)-4-(thiophen-2-yl)azetidine-2,3-dione (1x):** This is a known compound.<sup>[3]</sup> Yellow powder, 1.91 g, 70% yield.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.54 – 7.47 (m, 2H), 7.36 (dd,  $J = 5.1, 1.2$  Hz, 1H), 7.16 (ddd,  $J = 3.6, 1.3, 0.6$  Hz, 1H),

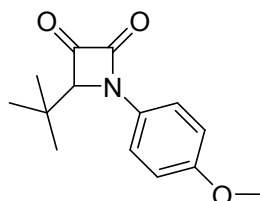
7.05 (dd,  $J = 5.1, 3.6$  Hz, 1H), 6.93 – 6.85 (m, 2H), 5.82 (s, 1H), 3.80 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  189.6, 159.6, 158.1, 134.7, 129.6, 127.8, 127.3, 127.2, 119.7, 114.8, 70.6, 55.5. HRMS (ESI<sup>+</sup>),  $m/z$  274.0529 ([M+H]<sup>+</sup>), calcd for  $\text{C}_{14}\text{H}_{12}\text{NO}_3\text{S}^+$ : 274.0532.



**4-(furan-2-yl)-1-(4-methoxyphenyl)azetidine-2,3-dione (1y):** This is a known compound.<sup>[4]</sup> Yellow powder, 1.54 g, 60% yield.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.6 – 7.5 (m, 2H), 7.5 – 7.4 (m, 1H), 7.0 – 6.8 (m, 2H), 6.6 (d,  $J = 3.3$  Hz, 1H), 6.4 (dd,  $J = 3.3, 1.9$  Hz, 1H), 5.6 (s, 1H), 3.8 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  189.3, 159.8, 158.1, 145.3, 144.3, 130.0, 119.3, 114.7, 111.6, 111.0, 68.1, 55.5.



**4-cyclohexyl-1-(4-methoxyphenyl)azetidine-2,3-dione (1z):** This is a new compound. Yellow powder, 1.93 g, 71% yield.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.60 – 7.45 (m, 2H), 7.02 – 6.91 (m, 2H), 4.56 (d,  $J = 4.0$  Hz, 1H), 3.84 (s, 3H), 2.21 – 2.07 (m, 1H), 1.87 – 1.62 (m, 5H), 1.54 – 0.77 (m, 5H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  195.4, 159.8, 158.0, 130.1, 119.3, 114.8, 55.6, 36.8, 28.9, 27.1, 26.1, 25.9, 25.6. HRMS (ESI<sup>+</sup>),  $m/z$  274.1432 ([M+H]<sup>+</sup>), calcd for  $\text{C}_{16}\text{H}_{20}\text{NO}_3^+$ : 274.1438.

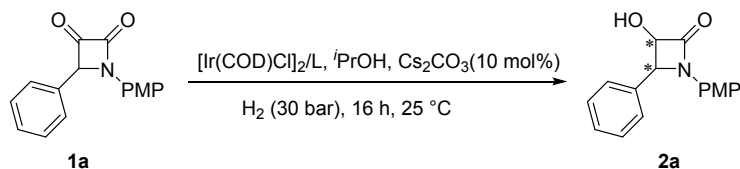


**4-(tert-butyl)-1-(4-methoxyphenyl)azetidine-2,3-dione (1aa):** This is a known compound.<sup>[4]</sup> Yellow powder, 1.85 g, 75% yield.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.5 – 7.4 (m, 2H), 7.0 (d,  $J = 9.0$  Hz, 2H), 4.6 (s, 1H), 3.8 (s, 3H), 1.1 (s, 9H).  $^{13}\text{C}$  NMR

(101 MHz, Chloroform-*d*)  $\delta$  195.7, 161.0, 158.1, 129.7, 121.5, 114.5, 80.4, 55.6, 35.2, 27.0.

### 3. Detailed Optimization of the Reaction Conditions.

Table S1. Evaluation of the effects of ligands in the Ir-Catalyzed Hydrogenation of **1a**<sup>a</sup>



entry	L	conv. (%) <sup>b</sup>	dr <sup>c</sup>	ee (%) <sup>d</sup>
1	<i>f</i> -amphox	>99	>20:1	1
2	<i>f</i> -amphol	42	>20:1	2
3	<i>f</i> -ampha	26	>20:1	13

<sup>a</sup> Reaction conditions: 0.1 mmol of substrate **1a**, 0.25 mol % [Ir(COD)Cl]<sub>2</sub>, 0.5 mol % ligand, 10 mol % Cs<sub>2</sub>CO<sub>3</sub>, *i*PrOH (1.0 mL). <sup>b</sup> Conversions were determined by <sup>1</sup>H NMR spectroscopy. <sup>c</sup> Diastereomeric ratio (dr) were determined by <sup>1</sup>H NMR spectroscopy. <sup>d</sup> Enantiomeric excesses (ee) were determined by HPLC analysis using a chiral stationary phase.

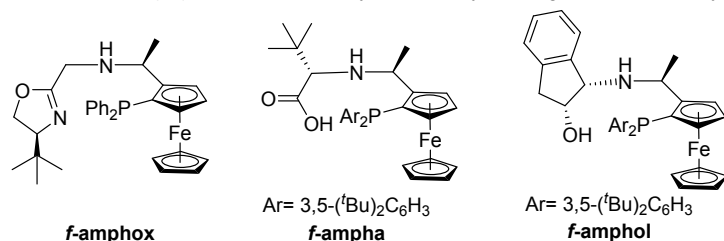
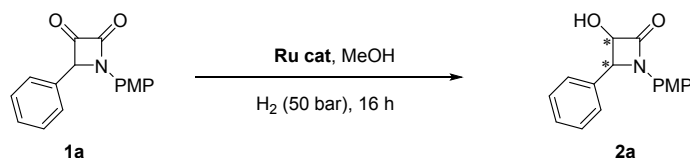


Table S2. Evaluation of the effects of additive in the Ru-Catalyzed Hydrogenation of **1a**<sup>a</sup>



entry	Ru cat	additive	temp. (°C)	conv.% <sup>b</sup>	dr <sup>c</sup>	ee (%) <sup>d</sup>
1	RuCl <sub>2</sub> (Ph-C <sub>3</sub> -TunePhos)(DMF) <sub>2</sub>	-	50	>95	>20:1	5
2	Ru(OAc) <sub>2</sub> (Ph-C <sub>3</sub> -TunePhos)	-	50	>95	>20:1	8
3	RuCl <sub>2</sub> (Ph-C <sub>3</sub> -TunePhos)(DMF) <sub>2</sub>	CeCl <sub>3</sub> ·7H <sub>2</sub> O	25	>95	>20:1	7
4	RuCl <sub>2</sub> (Ph-C <sub>3</sub> -TunePhos)(DMF) <sub>2</sub>	<i>p</i> -TsOH·H <sub>2</sub> O	25	56	>20:1	10
5	RuCl <sub>2</sub> (Ph-C <sub>3</sub> -TunePhos)(DMF) <sub>2</sub>	AcOH	25	53	>20:1	33

<sup>a</sup> Reaction conditions: **1a** (0.1 mmol), [Ru] (1 mol%), additive (10 mol%) in MeOH (1 mL) for 16 h. <sup>b</sup> Conversions were determined by <sup>1</sup>H NMR spectroscopy. <sup>c</sup> Diastereomeric ratio (dr) were determined by <sup>1</sup>H NMR spectroscopy. <sup>d</sup> Enantiomeric excesses (ee) were determined by HPLC analysis using a chiral stationary phase.

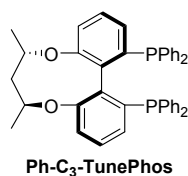
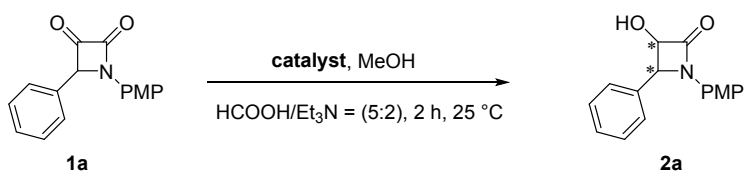


Table S3. Evaluation of the effects of additive in the Ru-Catalyzed Hydrogenation of **1a**<sup>a</sup>



entry	catalyst	conv.(%) <sup>b</sup>	dr <sup>c</sup>	ee (%) <sup>d</sup>
1	<b>cat.1</b>	>95	>20:1	0
2	<b>cat.2</b>	>95	>20:1	3
3	<b>cat.3</b>	>95	>20:1	2
4	<b>cat.4</b>	>95	>20:1	1

<sup>a</sup> Reaction conditions: **1a** (0.1 mmol), catalyst (1 mol%), HCOOH/Et<sub>3</sub>N = (5:2) (25  $\mu$ L), MeOH (1 mL), 2 h. <sup>b</sup> Conversions were determined by <sup>1</sup>H NMR spectroscopy. <sup>c</sup> Diastereomeric ratio (dr) were determined by <sup>1</sup>H NMR spectroscopy. <sup>d</sup> Enantiomeric excesses (ee) were determined by HPLC analysis using a chiral stationary phase.

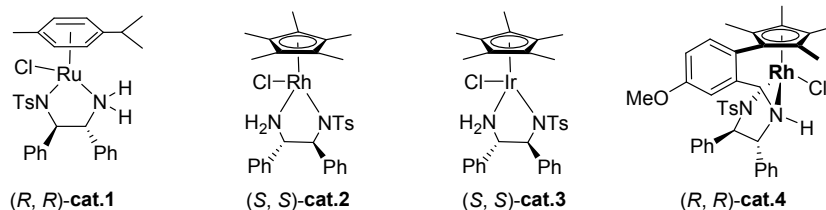
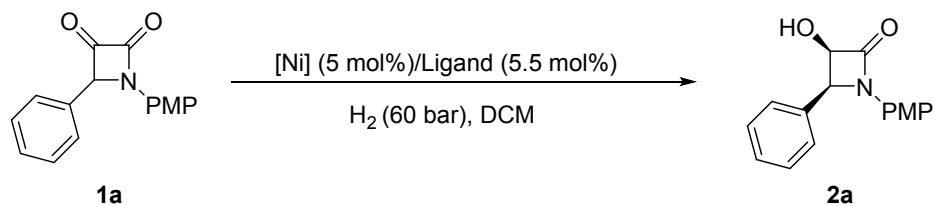


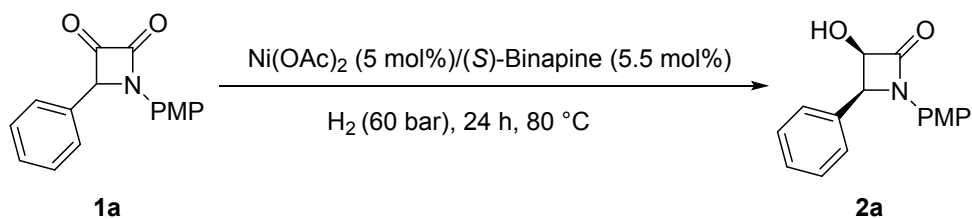


Table S4. Evaluation of the effects of ligands in the Ni-Catalyzed Hydrogenation of **1a**<sup>a</sup>

entry	[Ni]	Ligand	T (°C)	time (h)	conv. (%) <sup>b</sup>	dr <sup>c</sup>	ee (%) <sup>d</sup>
1	Ni(OAc) <sub>2</sub>	(S)-Ph-BPE	80	24	<5	--	--
2	Ni(OAc) <sub>2</sub>	(S,S)-Me-DuPhos	80	24	<5	--	--
3	Ni(OAc) <sub>2</sub>	(S)-SegPhos	80	24	<5	--	--
4	Ni(OAc) <sub>2</sub>	(R)-Binap	80	24	<5	--	--
5	Ni(OAc) <sub>2</sub>	(R, S)-JosiPhos	80	24	18	5:1	23
6	Ni(OAc) <sub>2</sub>	(R, R)-QuinoxP*	80	24	19	>20:1	77
7	Ni(OAc) <sub>2</sub>	(S)-Binapine	80	24	35	>20:1	95
8	Ni(OAc) <sub>2</sub>	(R <sub>c</sub> , Sp)-DuanPhos	80	24	41	>20:1	79
9	Ni(OAc) <sub>2</sub>	(S)-Binapine	65	36	17	>20:1	90
10	Ni(OAc) <sub>2</sub>	(R <sub>c</sub> , Sp)-DuanPhos	65	36	50	>20:1	85
11	Ni(OAc) <sub>2</sub>	(S)-Binapine	75	48	43	>20:1	95
12	Ni(OAc) <sub>2</sub>	(R <sub>c</sub> , Sp)-DuanPhos	75	48	47	>20:1	82
13	Ni(OAc) <sub>2</sub>	(S)-Binapine	100	24	58	>20:1	95
14	Ni(OAc) <sub>2</sub>	(R <sub>c</sub> , Sp)-DuanPhos	100	24	87	>20:1	31
15	Ni(OAc) <sub>2</sub>	(S)-Binapine	100	36	85	>20:1	37
16	Ni(OAc) <sub>2</sub>	(S)-Binapine	100	72	>95	>20:1	12
17	Ni(OTf) <sub>2</sub>	(S)-Binapine	100	48	95	4:1	rac
18	Ni(COD) <sub>2</sub>	(S)-Binapine	100	48	23	>20:1	94
19	Ni(acac) <sub>2</sub>	(S)-Binapine	100	48	89	>20:1	58

<sup>a</sup> All reactions were carried out with a Ni(OAc)<sub>2</sub>/(S)-Binapine/substrate (0.1 mmol) ratio of 1:1.1:20 on 0.1 mmol scale. The catalyst was pre-complexed in DCM (0.1 mL for each reaction vial). <sup>b</sup> Conversions were determined by <sup>1</sup>H NMR spectroscopy. <sup>c</sup> Diastereomeric ratio (dr) were determined by <sup>1</sup>H NMR spectroscopy <sup>d</sup> Enantiomeric excesses (ee) were determined by HPLC analysis using a chiral stationary phase.

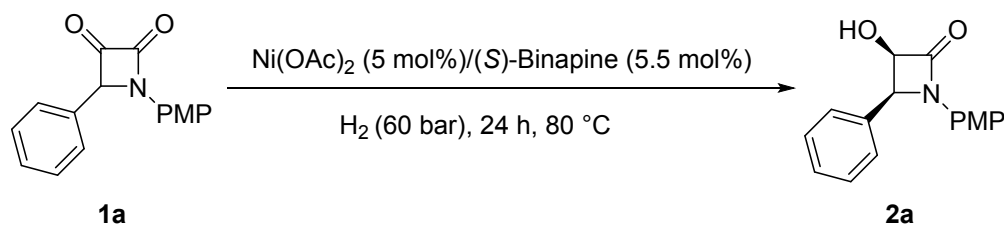
Table S5. Evaluation of the effects of solvents in the Ni-Catalyzed Hydrogenation of **1a**<sup>a</sup>



entry	solvent	conv. (%) <sup>b</sup>	dr <sup>c</sup>	ee (%) <sup>d</sup>
1	DCM	42	>20:1	95
2	THF	35	>20:1	93
3	toluene	23	>20:1	96
4	dioxane	38	>20:1	80
5	EA	30	>20:1	92
6	MeCN	18	>20:1	74
7	hexane	<5	--	--
8	MeOH	messy	--	--
9	IPA	messy	--	--
10	CF <sub>3</sub> COOH	messy	--	--

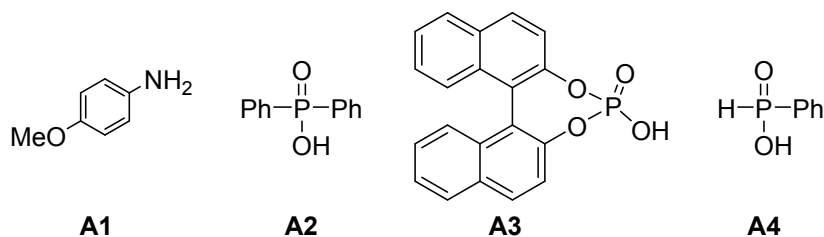
<sup>a</sup> All reactions were carried out with a Ni(OAc)<sub>2</sub>/(*S*)-Binapine/substrate ratio of 1:1.1:20 on 0.1 mmol scale. The catalyst was pre-complexed in DCM (0.1 mL for each reaction vial). <sup>b</sup> Conversions were determined by <sup>1</sup>H NMR spectroscopy. <sup>c</sup> Diastereomeric ratio (dr) were determined by <sup>1</sup>H NMR spectroscopy. <sup>d</sup> Enantiomeric excesses (ee) were determined by HPLC analysis using a chiral stationary phase.

Table S6. Evaluation of the effects of additives in the Ni-Catalyzed DKR of **1a**<sup>a</sup>



entry	additive	conv.(%) <sup>b</sup>	dr <sup>c</sup>	ee(%) <sup>d</sup>
1	DBU	messy	--	--
2	K <sub>2</sub> CO <sub>3</sub>	messy	--	--
3	NaBAR <sub>F</sub>	84	3:1	45
4	AgOTf	>95	9:1	3
5	AlCl <sub>3</sub>	<5	--	--
6 <sup>e</sup>	TsOH•H <sub>2</sub> O	56	>20:1	79
7	TsOH•H <sub>2</sub> O	>95	>20:1	8
8	AcOH	56	>20:1	45
9 <sup>e</sup>	BF <sub>3</sub> •OEt <sub>2</sub>	>95	1.3:1	3
10 <sup>f</sup>	<b>A1</b>	messy	--	--
11 <sup>f</sup>	<b>A2</b>	75	>20:1	65
12 <sup>f</sup>	<b>A3</b>	7	>20:1	25
13 <sup>f</sup>	<b>A4</b>	>95	>20:1	81
14	<b>A4</b>	>95	>20:1	83.5
15 <sup>g</sup>	<b>A4</b>	85	>20:1	93

<sup>a</sup> All reactions were carried out with a Ni(OAc)<sub>2</sub>/(*S*)-Binapine/substrate ratio of 1:1.1:20 on 0.1 mmol scale. Unless otherwise mentioned, the additives were 1.0 equiv. The catalyst was pre-complexed in DCM (0.1 mL for each reaction vial). Another 0.9 mL DCM for each entry <sup>b</sup> Conversions were determined by <sup>1</sup>H NMR spectroscopy. <sup>c</sup> Diastereomeric ratios (dr) were determined by <sup>1</sup>H NMR spectroscopy <sup>d</sup> Enantiomeric excesses (ee) were determined by HPLC analysis using a chiral stationary phase. <sup>e</sup> 0.1 equiv of additive was added. <sup>f</sup> 2.0 equiv of additive was added. <sup>g</sup> 0.5 equiv of additive was added.



**Table S7.** Further evaluation of the effects of solvents in the Ni-Catalyzed DKR of **1a**<sup>a</sup>

$\text{Ph}-\text{P}(=\text{O})(\text{OH})_2$  (100 mol%)

$\text{Ni}(\text{OAc})_2$  (5 mol%)/(*S*)-Binapine (5.5 mol%)

$\text{H}_2$  (60 bar), 24 h, 80 °C

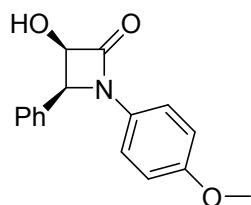
entry	solvent	conv. (%) <sup>b</sup>	dr <sup>c</sup>	ee(%) <sup>d</sup>
1	DCM	92	>20:1	86
2	THF	>95	>20:1	90.5
3	toluene	95	>20:1	92
<b>4<sup>e</sup></b>	<b>toluene</b>	<b>&gt;95</b>	>20:1	<b>91</b>
5	$\text{CHCl}_3$	92	>20:1	86
6	hexane	80	>20:1	39
7	MTBE	>95	>20:1	90

<sup>a</sup> All reactions were carried out with a  $\text{Ni}(\text{OAc})_2$ /*S*-Binapine/substrate ratio of 1:1.1:20 on 0.1 mmol scale. The additive was 1.0 equiv. of the substrate. The catalyst was pre-complexed in DCM (0.1 mL for each reaction vial), 0.9 mL other solvent for each. <sup>b</sup> Conversions were determined by <sup>1</sup>H NMR spectroscopy. <sup>c</sup> Diastereomeric ratios (dr) were determined by <sup>1</sup>H NMR spectroscopy. <sup>d</sup> Enantiomeric excesses (ee) were determined by HPLC analysis using a chiral stationary phase. <sup>e</sup> The reaction time was 36 h.

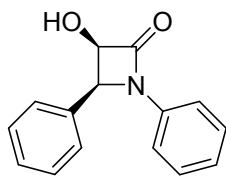
#### 4. General Procedure for the Asymmetric Hydrogenation via DKR.

A stock solution was prepared by mixing Ni(OAc)<sub>2</sub> (9.0 mg, 0.05 mmol) with (*S*)-Binapine (40 mg, 0.055 mmol) in a 1:1.1 molar ratio in DCM (1 mL at room temperature for 30 min in an argon-filled glovebox. An aliquot of the catalyst solution (0.1 mL, 0.005 mmol) was transferred by syringe into the vials charged with different substrates (0.1 mmol for each) and additive **A4** (0.1 mmol for each) in anhydrous toluene (0.9 mL). The vials were subsequently transferred into an autoclave into which hydrogen gas was charged. The reaction was then stirred under H<sub>2</sub> (60 bar) at 80 °C for 36 h. The hydrogen gas was released slowly and carefully after cooled the autoclave to room temperature. The reaction solution was concentrated and the residue was passed through a short column of silica gel (eluent: EA:PE = 5:1 to 2:1) to remove the metal complex and the additive. The ee values of compounds **2** were determined by HPLC analysis on a chiral stationary phase. The physical data were identical in all respect to those previously reported.

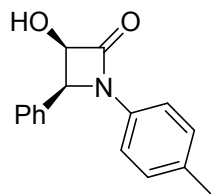
#### Characterization Data of 3-hydroxy- $\beta$ -Lactams



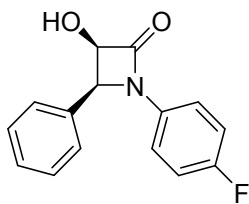
**(3*R*,4*S*)-3-hydroxy-1-(4-methoxyphenyl)-4-phenylazetidin-2-one (2a):** This is a known compound. <sup>[3]</sup> White solid; 23 mg, 87%; 91% ee; [ $\alpha$ ]<sub>D</sub><sup>20</sup> = +81.1 (c 1.0, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 85:15; flow rate = 1.0 mL/min; UV detection at 254 nm; t<sub>R</sub> = 12.1 min (major), 19.3 min (minor). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.45 – 7.27 (m, 7H), 6.80 (d, *J* = 9.0 Hz, 2H), 5.26 (d, *J* = 5.2 Hz, 1H), 5.18 (dd, *J* = 8.7, 5.2 Hz, 1H), 3.75 (s, 3H), 2.61 (d, *J* = 8.4 Hz, 1H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  165.5, 156.5, 133.2, 130.5, 129.1, 128.9, 127.5, 118.9, 114.4, 77.2, 62.3, 55.5. HRMS (ESI<sup>+</sup>), *m/z* 270.1120 ([M+H]<sup>+</sup>), calcd for C<sub>16</sub>H<sub>16</sub>NO<sub>3</sub><sup>+</sup>: 270.1125.



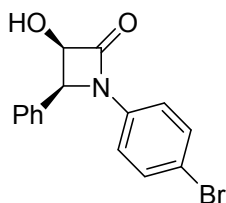
**(3R,4S)-3-hydroxy-1,4-diphenylazetidin-2-one (2b):** This is a known compound. <sup>[11]</sup> White solid, 22 mg, 90% yield, 89% ee,  $[\alpha]_D^{20} = +92.8$  (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 11.9$  min (major), 12.8 min (minor). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.45 – 7.37 (m, 3H), 7.34 (dd,  $J = 7.3, 1.5$  Hz, 4H), 7.30 – 7.24 (m, 2H), 7.12 – 7.05 (m, 1H), 5.30 (d,  $J = 5.3$  Hz, 1H), 5.19 (dd,  $J = 8.3, 5.3$  Hz, 1H), 2.89 (d,  $J = 8.4$  Hz, 1H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  166.2, 137.0, 133.0, 129.2, 129.2, 129.0, 127.5, 124.6, 117.6, 77.1, 62.2. HRMS (ESI<sup>+</sup>),  $m/z$  240.1016 ([M+H]<sup>+</sup>), calcd for C<sub>15</sub>H<sub>14</sub>NO<sub>2</sub><sup>+</sup>: 240.1019.



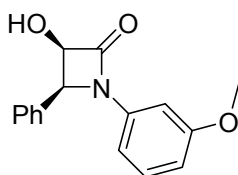
**(3R,4S)-3-hydroxy-4-phenyl-1-(*p*-tolyl)azetidin-2-one (2c):** This is a known compound. <sup>[11]</sup> White solid, 22 mg, 87% yield, 92% ee,  $[\alpha]_D^{20} = +58.6$  (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 85:15; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 8.8$  min (major), 15.3 min (minor). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.45 – 7.29 (m, 5H), 7.25 – 7.19 (m, 2H), 7.06 (d,  $J = 8.3$  Hz, 2H), 5.27 (d,  $J = 5.3$  Hz, 1H), 5.18 (s, 1H), 2.95 (s, 1H), 2.27 (s, 3H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  166.2, 134.5, 134.3, 133.2, 129.7, 129.1, 128.9, 127.5, 117.5, 77.1, 62.2, 20.9. HRMS (ESI<sup>+</sup>),  $m/z$  254.1171 ([M+H]<sup>+</sup>), calcd for C<sub>16</sub>H<sub>16</sub>NO<sub>2</sub><sup>+</sup>: 254.1176.



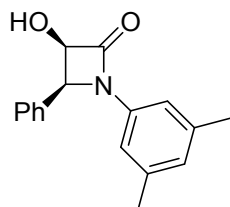
**(3R,4S)-1-(4-fluorophenyl)-3-hydroxy-4-phenylazetidin-2-one (2d):** This is a known compound. <sup>[1a]</sup> White solid, 23 mg, 90% yield, 87% ee, [ $\alpha$ ]<sup>20</sup><sub>D</sub> = +108.4 (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 85:15; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R$  = 8.1 min (major), 10.0 min (minor). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.47 – 7.27 (m, 7H), 7.01 – 6.88 (m, 2H), 5.30 – 5.15 (m, 2H), 3.53 (d,  $J$  = 7.8 Hz, 1H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  166.3, 160.6, 158.2 (d,  $J$  = 245.4 Hz), 133.2 (d,  $J$  = 3.0 Hz), 132.8, 129.1, 129.0, 127.6, 119.1, 119.0 (d,  $J$  = 8.1 Hz), 116.2, 115.9 (d,  $J$  = 22.2 Hz), 77.2, 62.6. HRMS (ESI<sup>+</sup>),  $m/z$  258.0922 ([M+H]<sup>+</sup>), calcd for C<sub>15</sub>H<sub>13</sub>FNO<sub>2</sub><sup>+</sup>: 258.0925.



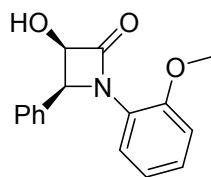
**(3R,4S)-1-(4-bromophenyl)-3-hydroxy-4-phenylazetidin-2-one (2e):** This is a new compound. White solid, 27 mg, 85% yield, 84% ee, [ $\alpha$ ]<sup>20</sup><sub>D</sub> = +112.4 (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 85:15; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R$  = 9.5 min (major), 15.2 min (minor). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.46 – 7.34 (m, 5H), 7.33 – 7.27 (m, 2H), 7.24 – 7.16 (m, 2H), 5.27 (d,  $J$  = 5.3 Hz, 1H), 5.20 (dd,  $J$  = 7.6, 5.2 Hz, 1H), 3.06 (d,  $J$  = 8.3 Hz, 1H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  166.3, 135.9, 132.5, 132.2, 129.2, 129.1, 127.5, 119.1, 117.3, 77.3, 62.4. HRMS (ESI<sup>+</sup>),  $m/z$  318.0122 ([M+H]<sup>+</sup>), calcd for C<sub>15</sub>H<sub>13</sub>BrNO<sub>2</sub><sup>+</sup>: 318.0124.



**(3*R*,4*S*)-3-hydroxy-1-(3-methoxyphenyl)-4-phenylazetidin-2-one (2f):** This is a known compound. <sup>[11]</sup> White solid, 22 mg, 82% yield, 84% ee,  $[\alpha]_D^{20} = +60.0$  (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak OD-3 column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 12.2$  min (minor), 14.8 min (major). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.45 – 7.36 (m, 3H), 7.36 – 7.31 (m, 2H), 7.15 (t,  $J = 8.2$  Hz, 1H), 7.05 (t,  $J = 2.3$  Hz, 1H), 6.80 (dd,  $J = 8.1, 2.0$  Hz, 1H), 6.64 (dd,  $J = 8.3, 2.5$  Hz, 1H), 5.29 (d,  $J = 5.4$  Hz, 1H), 5.18 (d,  $J = 5.4$  Hz, 1H), 3.75 (s, 3H), 2.56 (s, 1H). <sup>13</sup>C NMR (151 MHz, Chloroform-*d*)  $\delta$  166.1, 160.2, 138.1, 132.9, 130.0, 129.2, 129.0, 127.4, 110.4, 109.8, 103.6, 77.1, 62.3, 55.3. HRMS (ESI<sup>+</sup>),  $m/z$  270.1120 ( $[M+H]^+$ ), calcd for C<sub>16</sub>H<sub>16</sub>NO<sub>3</sub><sup>+</sup>: 270.1125.

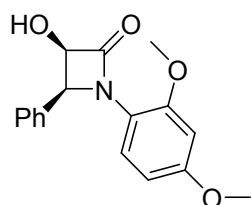


**(3*R*,4*S*)-1-(3,5-dimethylphenyl)-3-hydroxy-4-phenylazetidin-2-one (2g):** This is a new compound. White solid, 22 mg, 81% yield, 90% ee,  $[\alpha]_D^{20} = +86.6$  (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak OD-3 column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 8.2$  min (minor), 9.6 min (major). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.45 – 7.36 (m, 3H), 7.33 (dd,  $J = 7.9, 1.7$  Hz, 2H), 7.00 – 6.95 (m, 2H), 6.73 (s, 1H), 5.28 (d,  $J = 5.3$  Hz, 1H), 5.15 (d,  $J = 5.4$  Hz, 1H), 2.61 (s, 1H), 2.23 (s, 6H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  166.2, 139.0, 136.9, 133.2, 129.1, 128.8, 127.3, 126.4, 115.2, 76.9, 62.1, 21.3. HRMS (ESI<sup>+</sup>),  $m/z$  268.1328 ( $[M+H]^+$ ), calcd for C<sub>17</sub>H<sub>18</sub>NO<sub>2</sub><sup>+</sup>: 268.1332.

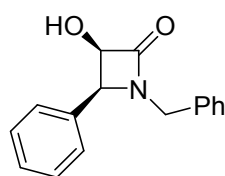




**(3*R*,4*S*)-3-hydroxy-1-(2-methoxyphenyl)-4-phenylazetidin-2-one (2h):** This is a new compound. White solid, 23 mg, 85% yield, 89% ee,  $[\alpha]_D^{20} = +49.7$  (c 0.3, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 14.5$  min (major), 16.3 min (minor). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  8.02 – 7.94 (m, 1H), 7.38 – 7.33 (m, 2H), 7.33 – 7.27 (m, 3H), 7.12 – 7.06 (m, 1H), 6.98 – 6.93 (m, 1H), 6.80 (d, *J* = 8.2 Hz, 1H), 5.66 (d, *J* = 5.3 Hz, 1H), 5.21 (d, *J* = 5.3 Hz, 1H), 3.59 (s, 3H), 2.75 (s, 1H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  167.9, 150.5, 135.1, 128.8, 128.4, 127.3, 126.3, 125.3, 123.3, 121.1, 112.1, 78.1, 65.9, 55.5. HRMS (ESI<sup>+</sup>), *m/z* 270.1120 ([*M*+*H*]<sup>+</sup>), calcd for C<sub>16</sub>H<sub>16</sub>NO<sub>3</sub><sup>+</sup>: 270.1125.

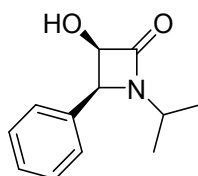


**(3*R*,4*S*)-1-(2,4-dimethoxyphenyl)-3-hydroxy-4-phenylazetidin-2-one (2i):** This is a new compound. White solid, 26 mg, 88% yield, 91% ee,  $[\alpha]_D^{20} = +47.0$  (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 85:15; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 14.1$  min (major), 19.8 min (minor). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.77 (dd, *J* = 8.7, 1.5 Hz, 1H), 7.36 (dd, *J* = 8.3, 6.4 Hz, 2H), 7.33 – 7.27 (m, 3H), 6.46 (dd, *J* = 8.8, 2.6 Hz, 1H), 6.37 (d, *J* = 2.6 Hz, 1H), 5.57 (d, *J* = 5.2 Hz, 1H), 5.20 (dd, *J* = 8.0, 5.1 Hz, 1H), 3.75 (s, 3H), 3.62 (s, 3H), 2.69 (d, *J* = 8.2 Hz, 1H). <sup>13</sup>C NMR (151 MHz, Chloroform-*d*)  $\delta$  167.8, 158.5, 152.4, 135.0, 128.8, 128.4, 127.5, 124.7, 118.4, 104.5, 99.8, 78.1, 65.6, 55.5. HRMS (ESI<sup>+</sup>), *m/z* 300.1227 ([*M*+*H*]<sup>+</sup>), calcd for C<sub>17</sub>H<sub>18</sub>NO<sub>4</sub><sup>+</sup>: 300.1230.

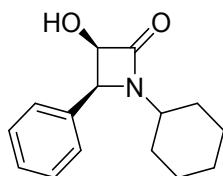


**(3*R*,4*S*)-1-benzyl-3-hydroxy-4-phenylazetidin-2-one (2j):** This is a known compound. <sup>[8]</sup> White solid, 18 mg, 70% yield, 88% ee,  $[\alpha]_D^{20} = +94.0$  (c 0.2, CHCl<sub>3</sub>);

The enantiomeric excess was determined by HPLC on Chiralpak OD-3 column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R$  = 10.0 min (major), 12.9 min (minor).  $^1\text{H}$  NMR (600 MHz, Chloroform-*d*)  $\delta$  7.41 – 7.33 (m, 3H), 7.32 – 7.23 (m, 5H), 7.12 (dd,  $J$  = 7.5, 2.0 Hz, 2H), 5.03 (d,  $J$  = 4.9 Hz, 1H), 4.82 (d,  $J$  = 14.8 Hz, 1H), 4.63 (d,  $J$  = 4.8 Hz, 1H), 3.86 (d,  $J$  = 14.8 Hz, 1H).  $^{13}\text{C}$  NMR (151 MHz, Chloroform-*d*)  $\delta$  169.5, 134.8, 133.5, 128.9, 128.8, 128.6, 128.1, 127.9, 78.0, 62.0, 44.2. HRMS (ESI<sup>+</sup>),  $m/z$  254.1172 ([M+H]<sup>+</sup>), calcd for C<sub>16</sub>H<sub>16</sub>NO<sub>2</sub><sup>+</sup>: 254.1176.

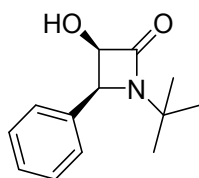


**(3R,4S)-3-hydroxy-1-isopropyl-4-phenylazetidin-2-one (2k):** This is a known compound. <sup>[1a]</sup> White solid, 16 mg, 77% yield, 91% ee,  $[\alpha]_D^{20} = +35.7$  (c 0.7, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak OD-3 column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R$  = 7.9 min (major), 9.6 min (minor).  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.50 – 7.32 (m, 5H), 4.97 (d,  $J$  = 5.0 Hz, 1H), 4.80 (d,  $J$  = 4.9 Hz, 1H), 3.96 (br, 1H), 3.85 – 3.45 (m, 1H), 1.27 (d,  $J$  = 6.8 Hz, 3H), 1.06 (d,  $J$  = 6.7 Hz, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  169.6, 135.2, 128.6, 128.5, 128.2, 76.6, 61.7, 45.0, 21.3, 20.2. HRMS (ESI<sup>+</sup>),  $m/z$  206.1174 ([M+H]<sup>+</sup>), calcd for C<sub>12</sub>H<sub>16</sub>NO<sub>2</sub><sup>+</sup>: 206.1176.

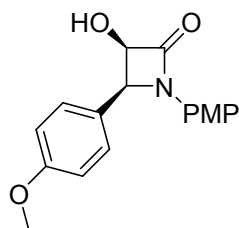


**(3R,4S)-1-cyclohexyl-3-hydroxy-4-phenylazetidin-2-one (2l):** This is a new compound. White solid, 18 mg, 72% yield, 94% ee,  $[\alpha]_D^{20} = +55.2$  (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R$  = 17.7 min

(major), 19.2 min (minor).  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.58 – 7.31 (m, 5H), 4.96 (dd,  $J = 7.5, 4.9$  Hz, 1H), 4.81 (d,  $J = 4.9$  Hz, 1H), 3.92 – 3.77 (m, 1H), 3.52 – 3.33 (m, 1H), 2.09 – 1.86 (m, 1H), 1.88 – 1.68 (m, 2H), 1.68 – 1.48 (m, 3H), 1.35 – 1.21 (m, 1H), 1.21 – 1.11 (m, 2H), 1.11 – 0.98 (m, 1H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  169.6, 135.3, 128.6, 128.6, 128.2, 76.6, 61.8, 52.6, 31.5, 30.5, 25.2, 24.9, 24.9. HRMS (ESI<sup>+</sup>),  $m/z$  246.1485 ([M+H]<sup>+</sup>), calcd for C<sub>15</sub>H<sub>20</sub>NO<sub>2</sub><sup>+</sup>: 246.1489.

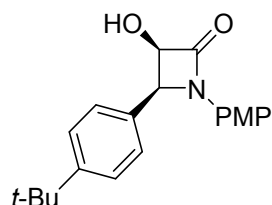


**(3R,4S)-1-(tert-butyl)-3-hydroxy-4-phenylazetidin-2-one (2m):** This is a new compound. White solid, 15 mg, 68% yield, 88% ee,  $[\alpha]_D^{20} = -7.6$  (c 0.7, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak OD-3 column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 7.0$  min (major), 9.9 min (minor).  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.46 – 7.33 (m, 5H), 4.90 (dd,  $J = 7.8, 5.1$  Hz, 1H), 4.86 (d,  $J = 5.1$  Hz, 1H), 2.81 (d,  $J = 7.9$  Hz, 1H), 1.29 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  168.8, 136.0, 128.7, 128.6, 128.0, 75.9, 61.9, 54.4, 28.2. HRMS (ESI<sup>+</sup>),  $m/z$  220.1330 ([M+H]<sup>+</sup>), calcd for C<sub>13</sub>H<sub>18</sub>NO<sub>2</sub><sup>+</sup>: 220.1332.



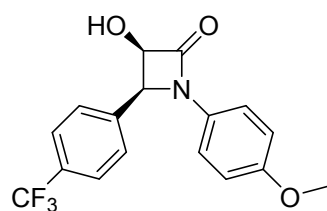
**(3R,4S)-3-hydroxy-1,4-bis(4-methoxyphenyl)azetidin-2-one (2n):** This is a known compound. <sup>[2]</sup> White solid, 27 mg, 91% yield, 92% ee,  $[\alpha]_D^{20} = +115.4$  (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 85:15; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 18.3$  min (major), 20.6 min (minor).  $^1\text{H}$  NMR (600 MHz, Chloroform-*d*)  $\delta$  7.31 – 7.21 (m, 5H),

6.94 (d,  $J = 8.3$  Hz, 2H), 6.80 (d,  $J = 8.7$  Hz, 2H), 5.22 (d,  $J = 5.2$  Hz, 1H), 5.14 (s, 1H), 3.80 (s, 3H), 3.75 (s, 3H), 2.59 (s, 1H).  $^{13}\text{C}$  NMR (151 MHz, Chloroform- $d$ )  $\delta$  165.6, 160.1, 156.4, 130.6, 128.8, 124.8, 118.9, 114.6, 114.4, 77.1, 61.9, 55.5, 55.3. HRMS (ESI $^+$ ),  $m/z$  300.1227 ([M+H] $^+$ ), calcd for  $\text{C}_{17}\text{H}_{18}\text{NO}_4^+$ : 300.1230.



**(3R,4S)-4-(4-(tert-butyl) phenyl)-3-hydroxy-1-(4-methoxyphenyl) azetidin-2-one**

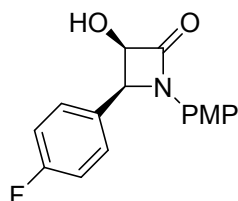
**(2o):** This is a known compound. <sup>[6]</sup> White solid, 28 mg, 88% yield, 87% ee,  $[\alpha]_{\text{D}}^{20} = +58.0$  (c 0.5,  $\text{CHCl}_3$ ); The enantiomeric excess was determined by HPLC on Chiralpak OD-3 column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_{\text{R}} = 9.6$  min (minor), 12.3 min (major).  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  7.37 (d,  $J = 8.4$  Hz, 2H), 7.25 – 7.18 (m, 4H), 6.88 (d,  $J = 9.0$  Hz, 2H), 6.06 (d,  $J = 7.0$  Hz, 1H), 5.26 (d,  $J = 5.1$  Hz, 1H), 5.14 (dd,  $J = 6.9, 5.1$  Hz, 1H), 3.68 (s, 3H), 1.26 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO}-d_6$ )  $\delta$  166.8, 156.0, 150.5, 132.3, 131.2, 128.2, 125.4, 118.7, 114.9, 77.3, 62.0, 55.7, 34.7, 31.6. HRMS (ESI $^+$ ),  $m/z$  326.1748 ([M+H] $^+$ ), calcd for  $\text{C}_{20}\text{H}_{24}\text{NO}_3^+$ : 326.1751.



**(3R,4S)-3-hydroxy-1-(4-methoxyphenyl)-4-(4-(trifluoromethyl) phenyl) azetidin-**

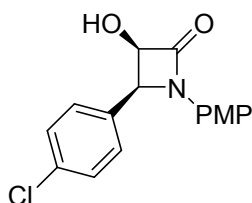
**2-one (2p):** This is a new compound. White solid, 28 mg, 82% yield, 90% ee,  $[\alpha]_{\text{D}}^{20} = +112.0$  (c 0.5,  $\text{CHCl}_3$ ); The enantiomeric excess was determined by HPLC on Chiralpak OD-3 column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_{\text{R}} = 10.5$  min (minor), 14.0 min (major).  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  7.74 (d,  $J = 8.1$  Hz, 2H), 7.49 (d,  $J = 8.1$  Hz, 2H), 7.22 (d,  $J = 9.0$  Hz, 2H), 6.90 (d,  $J =$

9.0 Hz, 2H), 6.24 (d,  $J = 7.1$  Hz, 1H), 5.45 (d,  $J = 5.1$  Hz, 1H), 5.24 (dd,  $J = 7.1, 5.1$  Hz, 1H), 3.69 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  166.5, 156.2, 140.5, 131.0, 129.2, { 129.3, 128.9, 128.6, 128.3 (q,  $J = 31.8$  Hz)} {128.8, 126.1, 123.4, 120.7 (q,  $J = 273.2$  Hz)} {125.6, 125.6, 125.5, 125.5 (q,  $J = 4.0$  Hz)} 118.6, 115.0, 77.5, 61.7, 55.7. HRMS (ESI<sup>+</sup>),  $m/z$  338.0993 ([M+H]<sup>+</sup>), calcd for  $\text{C}_{17}\text{H}_{15}\text{F}_3\text{NO}_3^+$ : 338.0999.



**(3R,4S)-4-(4-fluorophenyl)-3-hydroxy-1-(4-methoxyphenyl)azetidin-2-one (2q):**

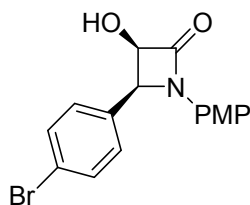
This is a known compound. <sup>[7]</sup> White solid, 24 mg, 85% yield, 94% ee,  $[\alpha]_D^{20} = +102.2$  (c 0.5,  $\text{CHCl}_3$ ); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 18.8$  min (major), 20.8 min (minor).  $^1\text{H}$  NMR (400 MHz,  $\text{Chloroform-}d$ )  $\delta$  7.35 – 7.29 (m, 2H), 7.27 – 7.22 (m, 2H), 7.13 – 7.03 (m, 2H), 6.86 – 6.75 (m, 2H), 5.23 (d,  $J = 5.1$  Hz, 1H), 5.19 (d,  $J = 5.4$  Hz, 1H), 3.75 (s, 3H), 3.63 (d,  $J = 7.0$  Hz, 1H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{Chloroform-}d$ )  $\delta$  166.2, 164.2, 161.7 (d,  $J = 249.5$  Hz), 156.6, 130.3, 129.5, 129.4 (d,  $J = 8.1$  Hz), 129.1, 129.1 (d,  $J = 3.0$  Hz), 118.9, 116.1, 115.9 (d,  $J = 22.2$  Hz), 114.5, 76.9, 61.9, 55.5. HRMS (ESI<sup>+</sup>),  $m/z$  288.1026 ([M+H]<sup>+</sup>), calcd for  $\text{C}_{16}\text{H}_{15}\text{FNO}_3^+$ : 288.1030.



**(3R,4S)-4-(4-chlorophenyl)-3-hydroxy-1-(4-methoxyphenyl)azetidin-2-one (2r):**

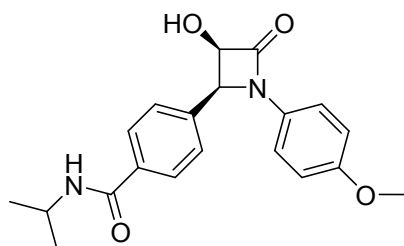
This is a new compound. White solid, 24 mg, 81% yield, 94% ee,  $[\alpha]_D^{20} = +126.8$  (c 0.5,  $\text{CHCl}_3$ ); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 19.6$  min (major), 22.2 min (minor).  $^1\text{H}$  NMR (400 MHz,  $\text{Chloroform-}d$ )  $\delta$  7.36 (d,

$J = 8.5$  Hz, 2H), 7.30 – 7.20 (m, 4H), 6.86 – 6.75 (m, 2H), 5.20 (d,  $J = 6.9$  Hz, 2H), 3.75 (s, 3H), 3.71 (s, 1H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  166.0, 156.6, 134.7, 131.9, 130.2, 129.2, 129.1, 118.9, 114.5, 61.9, 55.5. HRMS (ESI<sup>+</sup>),  $m/z$  304.0733 ([M+H]<sup>+</sup>), calcd for C<sub>16</sub>H<sub>15</sub>ClNO<sub>3</sub><sup>+</sup>: 304.0735.



**(3*R*,4*S*)-4-(4-bromophenyl)-3-hydroxy-1-(4-methoxyphenyl)azetidin-2-one (2s):**

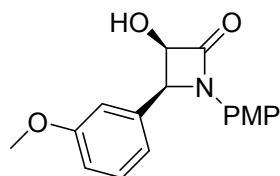
This is a new compound. White solid, 30 mg, 86% yield, 90% ee,  $[\alpha]_{\text{D}}^{20} = +112.4$  (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak OD-3 column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_{\text{R}} = 13.1$  min (minor), 15.7 min (major).  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.57 – 7.43 (m, 2H), 7.30 – 7.17 (m, 4H), 6.86 – 6.75 (m, 2H), 5.19 (s, 2H), 3.75 (s, 3H), 3.64 (s, 1H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  165.9, 156.6, 132.5, 132.1, 130.2, 129.4, 122.9, 118.9, 114.5, 62.0, 55.5. HRMS (ESI<sup>+</sup>),  $m/z$  348.0226 ([M+H]<sup>+</sup>), calcd for C<sub>16</sub>H<sub>15</sub>BrNO<sub>3</sub><sup>+</sup>: 348.0230.



**4-((2*S*,3*R*)-3-hydroxy-1-(4-methoxyphenyl)-4-oxoazetidin-2-yl)-N-**

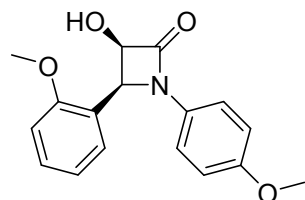
**isopropylbenzamide (2t):** This is a new compound. White solid, 15 mg, 45% yield, 78% ee,  $[\alpha]_{\text{D}}^{20} = +62.0$  (c 0.3, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 85:15; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_{\text{R}} = 19.5$  min (minor), 23.6 min (major).  $^1\text{H}$  NMR (600 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  8.2 (d,  $J = 7.7$  Hz, 1H), 7.8 (d,  $J = 8.1$  Hz, 2H), 7.3 (d,  $J = 8.1$  Hz, 2H), 7.2 (d,  $J = 8.9$  Hz, 2H), 6.9 (d,  $J = 8.9$  Hz, 2H), 6.2 (s, 1H), 5.4 (d,  $J = 5.0$  Hz, 1H), 5.2 (d,  $J = 5.0$  Hz, 1H), 4.1 (dq,  $J = 13.5, 6.6$  Hz, 1H), 3.7 (s, 3H), 1.1 (dd,  $J = 6.5, 2.7$  Hz,

7H).  $^{13}\text{C}$  NMR (151 MHz,  $\text{DMSO-}d_6$ )  $\delta$  166.7, 165.7, 156.1, 138.4, 135.0, 131.1, 128.2, 127.6, 118.7, 114.9, 77.5, 62.1, 55.7, 41.4, 22.8. HRMS (ESI<sup>+</sup>),  $m/z$  355.1648 ([M+H]<sup>+</sup>), calcd for  $\text{C}_{20}\text{H}_{23}\text{N}_2\text{O}_4^+$ : 355.1652.



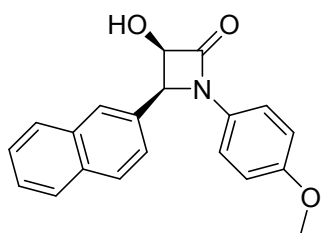
**(3R,4S)-3-hydroxy-4-(3-methoxyphenyl)-1-(4-methoxyphenyl)azetididin-2-one (2u):**

This is a new compound. White solid, 26 mg, 85% yield, 87% ee,  $[\alpha]_D^{20} = +96.8$  (c 0.5,  $\text{CHCl}_3$ ); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 85:15; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R$  = 14.5 min (major), 21.5 min (minor).  $^1\text{H}$  NMR (400 MHz,  $\text{Chloroform-}d$ )  $\delta$  7.39 – 7.20 (m, 3H), 6.97 – 6.81 (m, 3H), 6.83 – 6.72 (m, 2H), 5.25 – 5.10 (m, 2H), 3.75 (d,  $J$  = 10.3 Hz, 6H), 3.35 (d,  $J$  = 8.5 Hz, 1H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{Chloroform-}d$ )  $\delta$  166.0, 160.1, 156.5, 135.0, 130.5, 130.2, 119.7, 118.9, 114.4, 114.2, 113.3, 77.1, 62.4, 55.5, 55.3. HRMS (ESI<sup>+</sup>),  $m/z$  300.1227 ([M+H]<sup>+</sup>), calcd for  $\text{C}_{17}\text{H}_{18}\text{NO}_4^+$ : 300.1230.



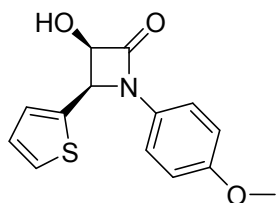
**(3R,4S)-3-hydroxy-4-(2-methoxyphenyl)-1-(4-methoxyphenyl)azetididin-2-one (2v):**

This is a new compound. White solid, 24 mg, 80% yield, 82% ee,  $[\alpha]_D^{20} = +44.8$  (c 0.5,  $\text{CHCl}_3$ ); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 85:15; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R$  = 12.5 min (major), 23.2min (minor).  $^1\text{H}$  NMR (400 MHz,  $\text{Chloroform-}d$ )  $\delta$  7.36 – 7.26 (m, 3H), 7.18 (dd,  $J$  = 7.5, 1.7 Hz, 1H), 7.00 – 6.90 (m, , 2H), 6.84 – 6.75 (m, 2H), 5.49 (d,  $J$  = 5.2 Hz, 1H), 5.18 (dd,  $J$  = 9.4, 5.2 Hz, 1H), 3.87 (s, 3H), 3.74 (s, 3H), 3.08 (d,  $J$  = 9.4 Hz, 1H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{Chloroform-}d$ )  $\delta$  166.1, 157.7, 156.3, 130.9, 129.9, 128.8, 121.6, 121.1, 118.7, 114.4, 111.3, 77.4, 59.2, 55.9, 55.5. HRMS (ESI<sup>+</sup>),  $m/z$  300.1227 ([M+H]<sup>+</sup>), calcd for  $\text{C}_{17}\text{H}_{18}\text{NO}_4^+$ : 300.1230.



**(3*R*,4*S*)-3-hydroxy-1-(4-methoxyphenyl)-4-(naphthalen-2-yl)azetidin-2-one (2w):**

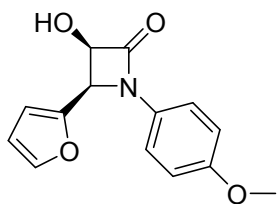
This is a new compound. White solid, 28 mg, 92% yield, 88% ee,  $[\alpha]_D^{20} = +110.0$  (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 85:15; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 16.5$  min (major), 24.1 min (minor). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.94 – 7.80 (m, 4H), 7.51 (dt, *J* = 6.3, 3.4 Hz, 2H), 7.41 (dd, *J* = 8.6, 1.8 Hz, 1H), 7.30 – 7.19 (m, 2H), 6.92 – 6.82 (m, 2H), 6.15 (d, *J* = 7.1 Hz, 1H), 5.47 (d, *J* = 5.1 Hz, 1H), 5.24 (dd, *J* = 7.1, 5.1 Hz, 1H), 3.66 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.8, 156.1, 133.3, 133.2, 131.3, 128.2, 128.1, 128.0, 127.5, 126.7, 126.5, 126.2, 118.7, 114.9, 77.6, 62.5, 55.7. HRMS (ESI<sup>+</sup>), *m/z* 338.0993 ([*M*+*H*]<sup>+</sup>), calcd for C<sub>20</sub>H<sub>18</sub>NO<sub>3</sub><sup>+</sup>: 320.1281.



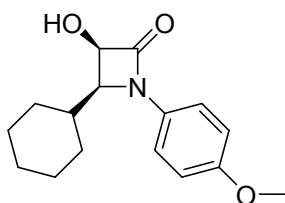
**(3*R*,4*R*)-3-hydroxy-1-(4-methoxyphenyl)-4-(thiophen-2-yl) azetidin-2-one (2x):**

This is a known compound. <sup>[3]</sup> White solid, 24 mg, 88% yield, 87% ee,  $[\alpha]_D^{20} = +60.4$  (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak OD-3 column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 19.2$  min (minor), 20.5 min (major). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.51 (dd, *J* = 5.0, 1.3 Hz, 1H), 7.30 – 7.23 (m, 2H), 7.18 (dd, *J* = 3.6, 1.3 Hz, 1H), 7.03 (dd, *J* = 5.1, 3.5 Hz, 1H), 6.94 – 6.83 (m, 2H), 6.33 (s, 1H), 5.62 (d, *J* = 4.9 Hz, 1H), 5.18 (d, *J* = 4.5 Hz, 1H), 3.69 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.5, 156.2, 138.7, 131.0, 130.1, 128.2, 127.2, 118.8, 114.9, 77.3, 58.7, 55.7. HRMS (ESI<sup>+</sup>), *m/z* 276.0685 ([*M*+*H*]<sup>+</sup>), calcd for C<sub>14</sub>H<sub>14</sub>NO<sub>3</sub>S<sup>+</sup>: 276.0689.

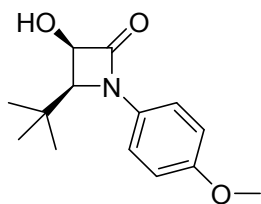




**(3*R*,4*R*)-4-(furan-2-yl)-3-hydroxy-1-(4-methoxyphenyl)azetidin-2-one (2y):** This is a known compound.<sup>[12]</sup> White solid, 22 mg, 85% yield, 82% ee,  $[\alpha]_D^{20} = +115.5$  (c 0.8, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak IA column, hexane: isopropanol = 85:15; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 11.9$  min (major), 18.9 min (minor). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  7.7 (dd,  $J = 1.7, 0.7$  Hz, 1H), 7.3 – 7.2 (m, 2H), 6.9 – 6.9 (m, 2H), 6.5 – 6.4 (m, 2H), 6.3 (d,  $J = 7.3$  Hz, 1H), 5.4 (d,  $J = 5.0$  Hz, 1H), 5.2 (dd,  $J = 7.3, 5.0$  Hz, 1H), 3.7 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  166.6, 156.1, 149.3, 143.9, 131.3, 118.4, 114.9, 111.1, 110.6, 77.3, 56.5, 55.7.



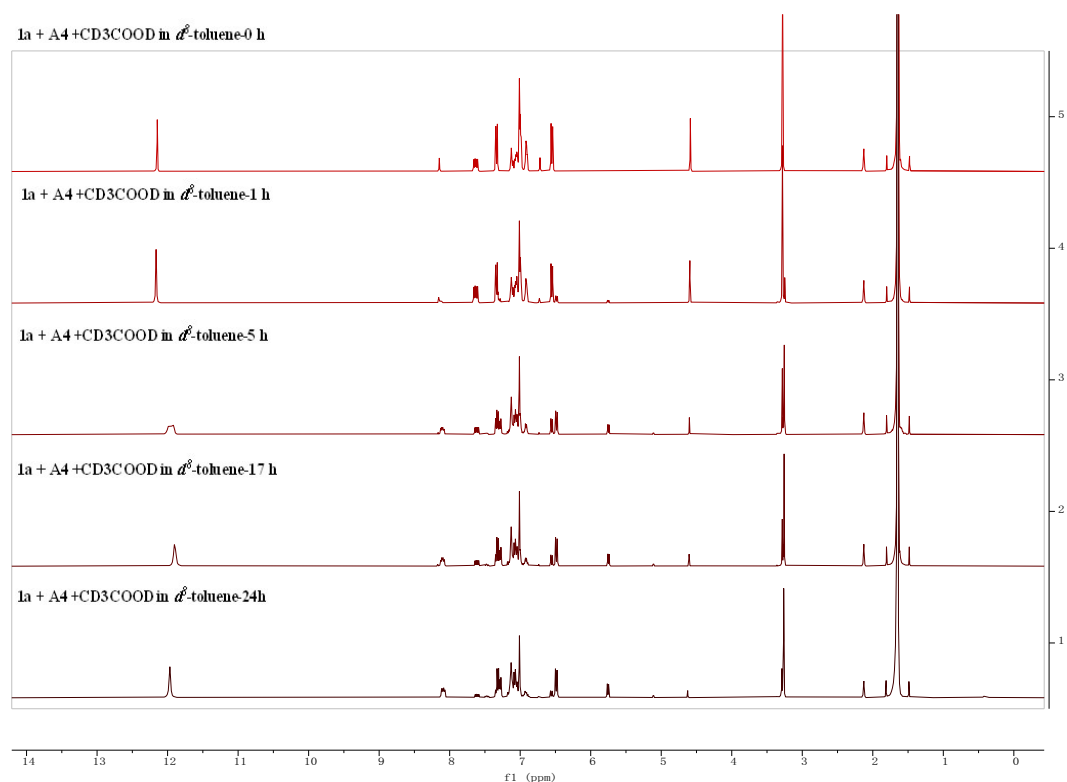
**(3*R*,4*S*)-4-cyclohexyl-3-hydroxy-1-(4-methoxyphenyl)azetidin-2-one (2z):** This is a new compound. White solid, 25 mg, 91% yield, 90% ee,  $[\alpha]_D^{20} = +28.2$  (c 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak OD-3 column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 6.6$  min (minor), 8.5 min (major). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.42 – 7.28 (m, 2H), 6.92 – 6.80 (m, 2H), 5.07 (t,  $J = 5.1$  Hz, 1H), 4.73 (d,  $J = 6.1$  Hz, 1H), 4.05 (dd,  $J = 7.3, 5.2$  Hz, 1H), 3.79 (s, 3H), 1.90 – 1.81 (m, 1H), 1.76 – 1.59 (m, 4H), 1.35 – 1.07 (m, 6H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  168.5, 156.6, 131.2, 119.9, 114.2, 75.0, 63.4, 55.5, 38.5, 30.2, 29.6, 26.3, 26.2, 26.0. HRMS (ESI<sup>+</sup>),  $m/z$  276.1591 ([M+H]<sup>+</sup>), calcd for C<sub>16</sub>H<sub>22</sub>NO<sub>3</sub><sup>+</sup>: 276.1594.



**(3*R*,4*S*)-4-(tert-butyl)-3-hydroxy-1-(4-methoxyphenyl)azetidin-2-one (2aa):** This is a known compound.<sup>[13]</sup> White solid, 17.5 mg, 70% yield, 85% ee,  $[\alpha]_D^{20} = +20.0$  (c 0.3, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on Chiralpak OD-3 column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 254 nm;  $t_R = 7.9$  min (minor), 10.9 min (major). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.3 – 7.3 (m, 2H), 6.9 – 6.8 (m, 2H), 5.1 (t,  $J = 5.7$  Hz, 1H), 4.4 (d,  $J = 6.0$  Hz, 1H), 4.1 (d,  $J = 5.3$  Hz, 1H), 3.8 (s, 3H), 1.1 (s, 9H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  169.1, 157.0, 130.3, 122.0, 114.2, 75.6, 67.9, 55.5, 34.9, 27.2.

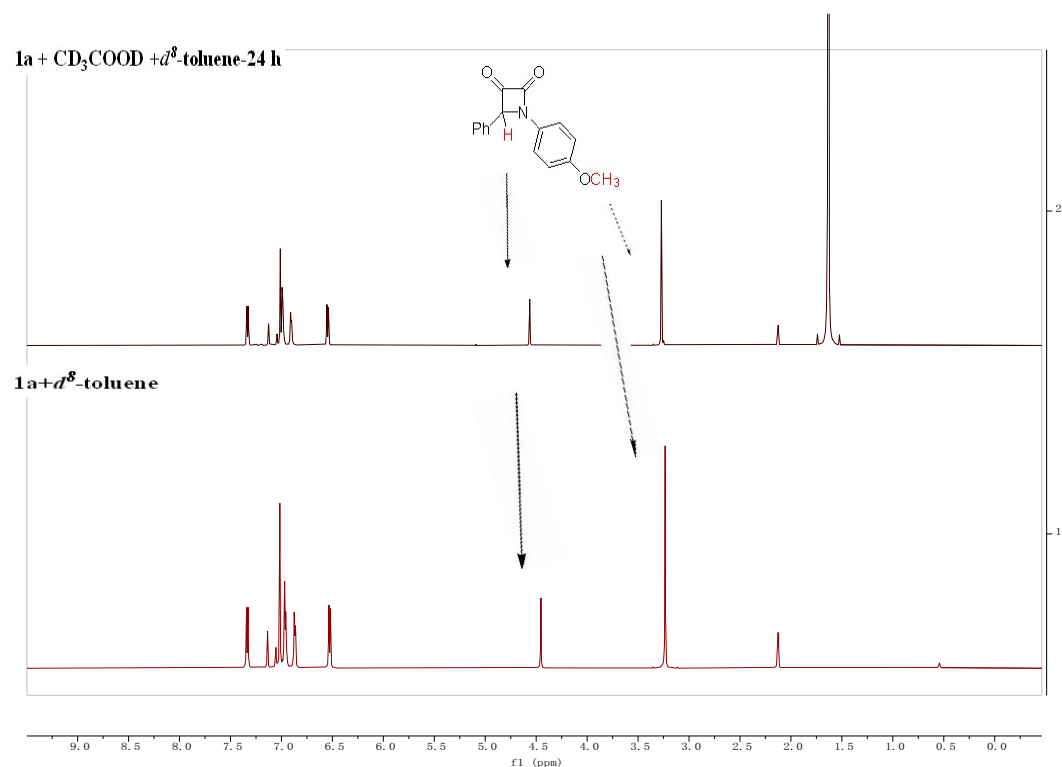
## 5. Deuterium-labelling Experiments.

**Deuterium-labelling experiments in the presence of phenylphosphinic acid (A4):** **1a** (0.05 mmol, 1.0 equiv), **A4** (phenylphosphinic acid, 0.05 mmol, 1.0 equiv), CD<sub>3</sub>COOD (50 μL, 0.87 mmol, 17.5 equiv) and *d*<sup>8</sup>-toluene (0.5 mL) were added into 5 NMR tubes, and the mixture was stirred at 80 °C for 0 h, 1 h, 5 h, 17 h and 24 h respectively. The deuteration ratios were determined by <sup>1</sup>H-NMR spectroscopy (Figure S1).



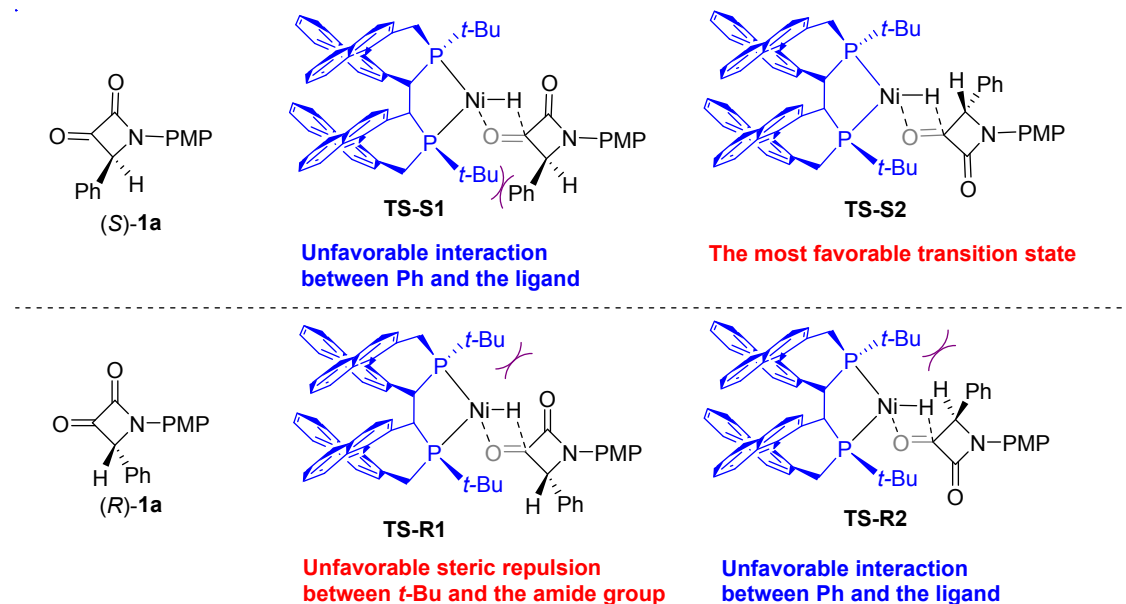
**Figure S1.** <sup>1</sup>H-NMR spectra of the deuteration experiments in the presence of phenylphosphinic acid (**A4**)

**Deuterium-labelling experiments in the absence of phenylphosphinic acid (A4):**  
**1a** (0.05 mmol, 1.0 equiv), CD<sub>3</sub>COOD (50 μL, 0.87 mmol, 17.5 equiv) and *d*<sup>8</sup>-toluene (0.5 mL) were added into **NMR tube 1**, and the mixture was stirred at 80 °C for 24 h.  
**1a** (0.05 mmol, 1.0 equiv) and *d*<sup>8</sup>-toluene (0.5 mL) were added into **NMR tube 1**.  
The deuteration ratio was determined by <sup>1</sup>H-NMR spectroscopy (Figure S2).  
The <sup>1</sup>H-NMR spectroscopy showed both integral ratio of the **H** and O-**CH**<sub>3</sub> in was 1:3.



**Figure S2.** <sup>1</sup>H-NMR spectra of the deuteration experiment in the absence of phenylphosphinic acid (**A4**).

## 6. Rationale for the Origin of Diastereoselectivity and Enantioselectivity.

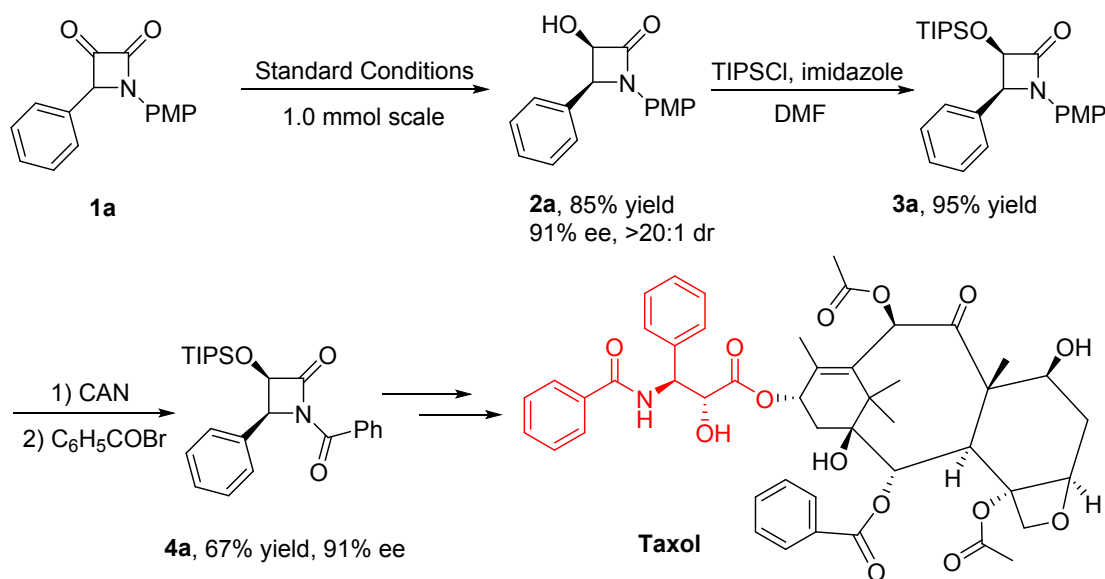


**Figure S3.** Rationale for the origin of diastereoselectivity and enantioselectivity.

According to the previous mechanistic investigations on nickel-catalyzed asymmetric hydrogenation of our group and Wanbin Zhang's group (see: (a) Y. Liu, Z. Yi, X. Yang, H. Wang, C. Yin, M. Wang, X.-Q. Dong, X. Zhang, *ACS Catal.* **2020**, *10*, 11153-11161; (b) Y. Hu, J. Chen, B. Li, Z. Zhang, I. D. Gridnev, W. Zhang, *Angew. Chem., Int. Ed.* **2020**, *59*, 5371-5375. (c) B. Li, J. Chen, Z. Zhang, I. D. Gridnev, W. Zhang, *Angew. Chem., Int. Ed.* **2019**, *58*, 7329-7334.), nickel hydride was identified as the key catalytic species in the current reaction.

As can be seen from Figure S3 above, the origin of diastereoselectivity was the energy difference between **TS-S1** and **TS-S2**, and **TS-R1** and **TS-R2**, the nickel hydride species attacks the carbonyl group from the opposite side of the  $\alpha$ -phenyl group due to the steric repulsion between the phenyl group and the ligand. The origin of enantioselectivity (rate difference observed in the reduction of the *(S)*- vs. the *(R)*- enantiomer of **1a**) was caused by the unfavorable steric interactions between the *t*-Bu group and the amide group of **(R)-1a** in **TS-R1**, and **TS-S2** is the most favorable transition state (**(S)-1a** matches better with the catalyst than **(R)-1a**).

## 7. Synthetic Applications.

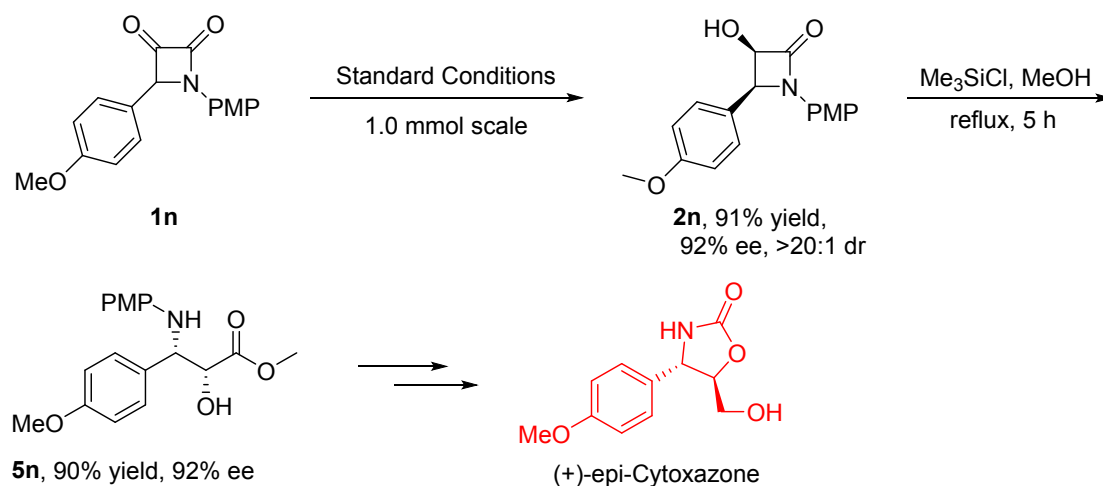


**Compound 2a:** A stock solution was made by mixing Ni(OAc)<sub>2</sub> (9.0 mg, 0.05 mmol) with (*S*)-Binapine (40 mg, 0.055 mmol) in a 1:1.1 molar ratio in DCM (1.0 mL) at room temperature for 30 min in an argon-filled glovebox. The catalyst solution (1.0 mL, 0.005 mmol) was transferred by syringe into the vials charged with substrate **1a** (267 mg, 1.0 mmol) in anhydrous toluene (9.0 mL). The vials were subsequently transferred into an autoclave into which hydrogen gas was charged. The reaction was then stirred under H<sub>2</sub> (50 atm) at 80 °C for 36 h. The hydrogen gas was released slowly and carefully. The reaction solution was concentrated and the residue was purified by column chromatography on silica gel (eluent: PE: EA = 5:1 to 2:1). The ee value of compound **2a** was determined by HPLC analysis on a chiral stationary phase. Pure compound **2a** (215 mg, 85% yield, 91% ee) was afforded after column chromatography.

**Compound 3a** <sup>[10]</sup>: To a solution of **2a** (0.85 mmol, 215 mg, 1.0 equiv.) in anhydrous DMF (1.0 mL) was cooled to 0 °C, imidazole was added (cat. 0.01 equiv.) into the mixture and TIPSCl (1.5 mmol, 340 μL, 1.5 equiv.) was added slowly by syringe. TLC indicated that **2a** was consumed completely. The mixture was cooled to room temperature and quenched by 1 mL H<sub>2</sub>O, the mixture was extracted by DCM (20 mL for 2 times), and the combined organic layer was washed by NaHCO<sub>3</sub>, brine

successively. The obtained organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under vacuum. The residue was purified by chromatography (Si<sub>2</sub>O, PE/EA from 20:1 to 5:1). Compound **3a** was obtained as a light yellow solid (340 mg, 95% yield).

**Compound 4a** <sup>[10]</sup>: (step 1) To a solution of **3a** (0.81 mmol, 340 mg, 1.0 equiv.) in CH<sub>3</sub>CN (5 mL) was cooled to -5 °C, CAN (2.67 mmol, 1.01g, 3.3 equiv.) in 3 mL water was added dropwise. The reaction mixture was stirred for 45 min until TLC indicated the consumption of the starting material. Then the mixture was diluted with EtOAc (10 mL) and washed with saturated aqueous NaHCO<sub>3</sub> (2×10 mL), water (2×5 mL), saturated sodium metabisulfite (2×5 mL) and brine, and then the organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. The crude product was chromatographed on silica gel with 40% EtOAc in hexanes to give the deprotected lactam. (step 2) To a solution of the deprotected lactam in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (1.0 mL) was cooled to 0 °C, triethylamine (285 μL, 1.62 mmol, 2 equiv.) and benzoyl chloride (137 μL, 0.89 mmol, 1.1 equiv.) were added. The mixture was then stirred at room temperature for 3 hours, diluted with EtOAc (10 mL), washed with saturated aqueous NaHCO<sub>3</sub> and brine, and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. The crude product was purified by chromatography (15% EtOAc in hexane) to give the β-lactam **4a** (225 mg, 67% yield, 91% ee). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 8.09 – 7.98 (m, 2H), 7.63 – 7.54 (m, 1H), 7.51 – 7.44 (m, 2H), 7.43 – 7.28 (m, 5H), 5.43 (d, *J* = 6.1 Hz, 1H), 5.25 (d, *J* = 6.1 Hz, 1H), 1.04 – 0.94 (m, 3H), 0.94 – 0.83 (m, 18H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 166.3, 165.5, 133.8, 133.4, 132.1, 129.9, 128.4, 128.2, 128.2, 76.6, 61.2, 17.5, 17.4, 11.7.



**Compound 2n:** A stock solution was made by mixing  $\text{Ni}(\text{OAc})_2$  (9.0 mg, 0.05 mmol) with (*S*)-Binapine (40 mg, 0.055 mmol) in a 1:1.1 molar ratio in DCM (1.0 mL) at room temperature for 30 min in an argon-filled glovebox. The catalyst solution (1.0 mL, 0.005 mmol) was transferred by syringe into the vial charged with substrate **1n** (297 mg, 1.0 mmol) in anhydrous toluene (9.0 mL). The vial was subsequently transferred into an autoclave into which hydrogen gas was charged. The reaction was then stirred under  $\text{H}_2$  (50 atm) at 80 °C for 36 h. The hydrogen gas was released slowly and carefully. The reaction solution was concentrated and the residue was purified by column chromatography on silica gel (eluent: EA: PE = 5:1 to 2:1). The ee value of compound **2n** was determined by HPLC analysis on a chiral stationary phase. Pure compound **2n** (269 mg, 91% yield, 92% ee) was afforded after column chromatography.

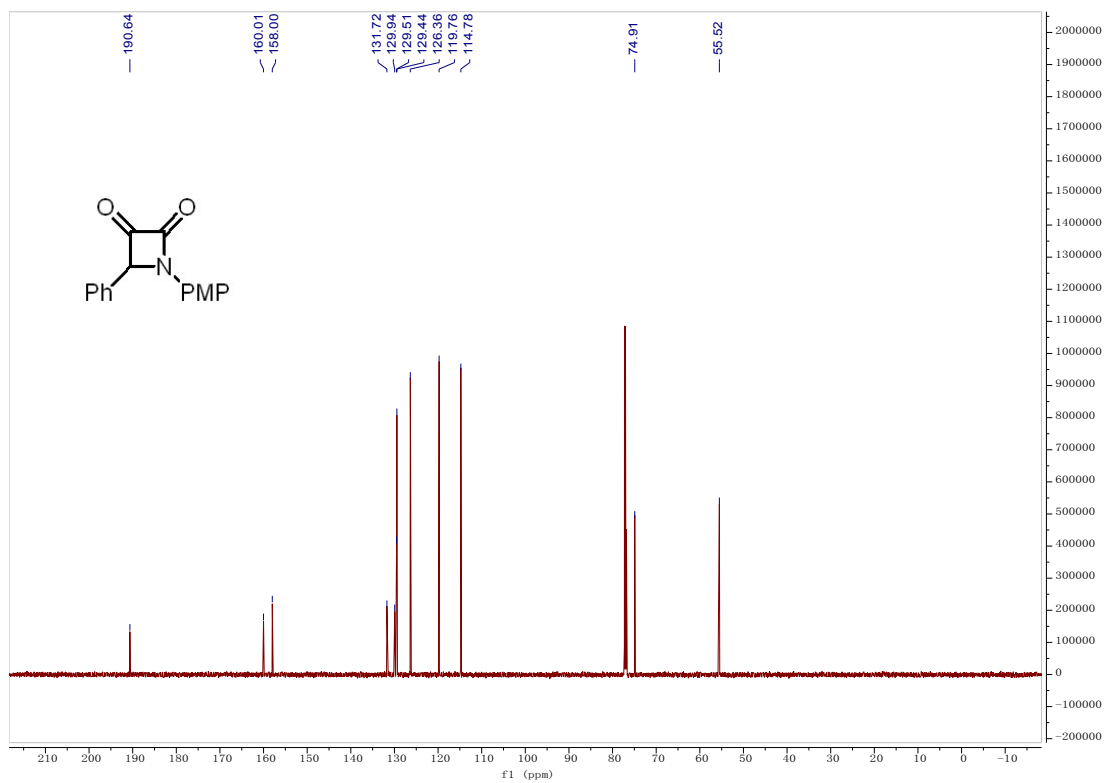
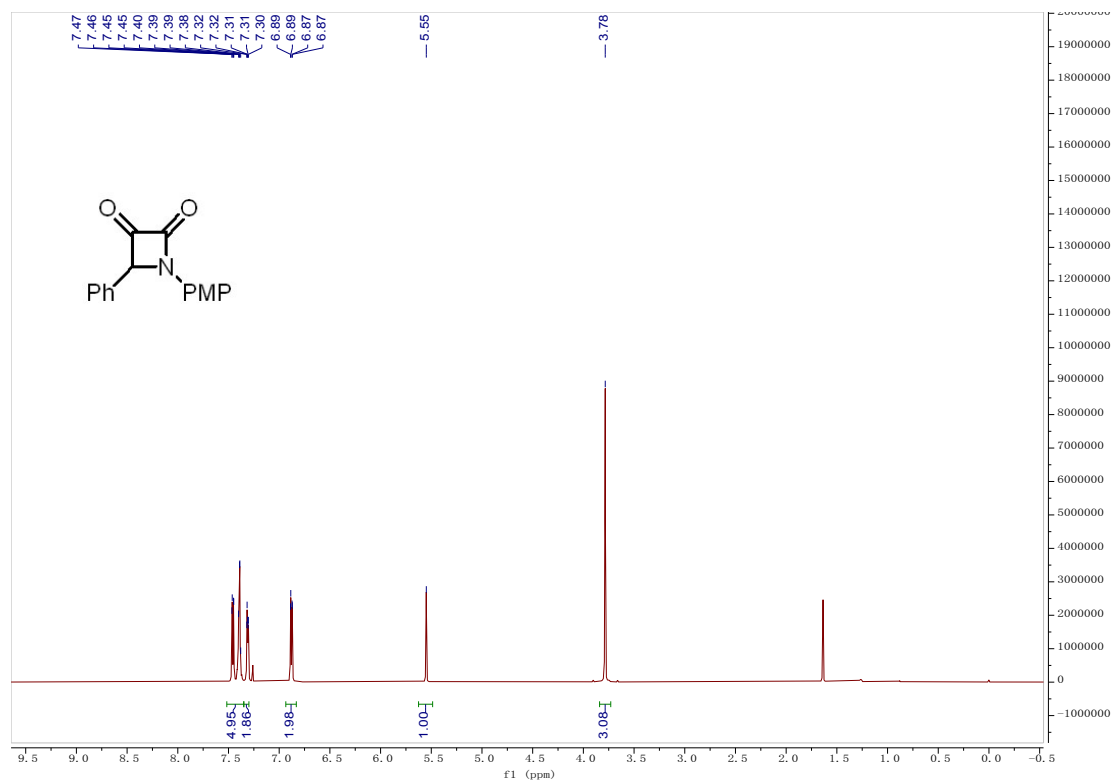
**Compound 5n** <sup>[9]</sup>: To a solution of **2n** (0.9 mmol, 269 mg, 1.0 equiv.) in anhydrous MeOH (2.0 mL) was cooled to 0°C,  $\text{Me}_3\text{SiCl}$  (2.7 mmol, 340  $\mu\text{L}$ , 3.0 equiv.) was added slowly by syringe. TLC indicated that some **2n** was remained and  $\text{Me}_3\text{SiCl}$  (1.8 mmol, 227  $\mu\text{L}$ , 2.0 equiv.) was added into the mixture. The mixture was warmed to room temperature and refluxed for 5 hours. After the starting materials was consumed completely the mixture was cooled to room temperature and quenched by 1 mL  $\text{H}_2\text{O}$ , the mixture was extracted by EtOAc (10 mL for 2 times), and the combined organic layer was washed by  $\text{NaHCO}_3$ , brine successively. The obtained organic layer was dried over anhydrous  $\text{Na}_2\text{SO}_4$ , filtered and concentrated under vacuum. The residue was purified by chromatography ( $\text{Si}_2\text{O}$ , PE/EA from 20:1 to 6:1). Compound **5n** was



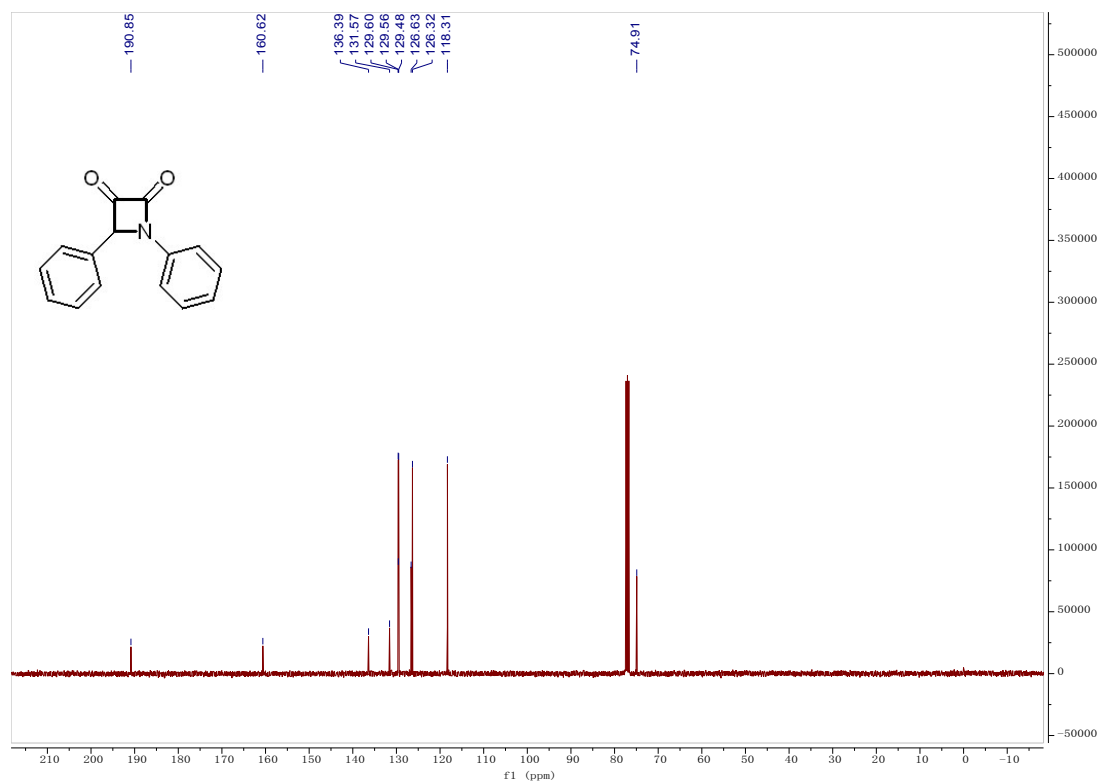
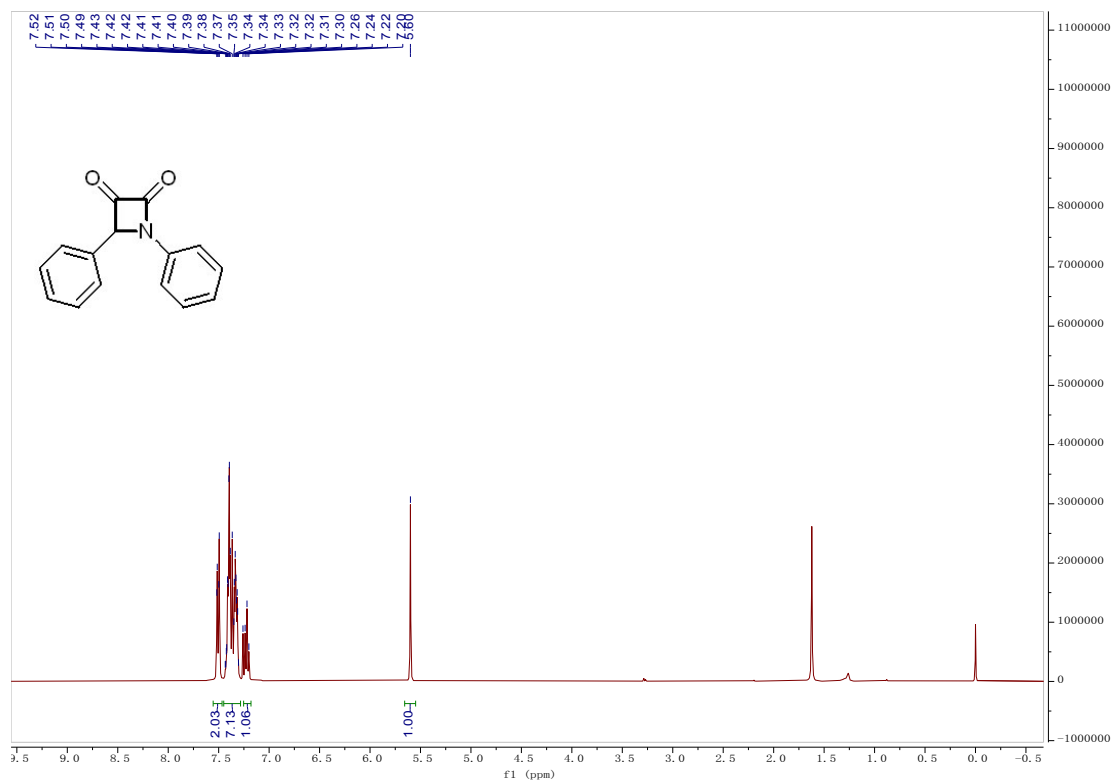
obtained as a light yellow solid (265 mg, 90% yield, 92% ee).  $^1\text{H}$  NMR (600 MHz, Chloroform-*d*)  $\delta$  7.33 – 7.22 (m, 2H), 6.90 – 6.82 (m, 2H), 6.73 – 6.65 (m, 2H), 6.56 – 6.46 (m, 2H), 4.79 (d,  $J = 2.9$  Hz, 1H), 4.47 (s, 1H), 4.43 (d,  $J = 2.8$  Hz, 1H), 3.75 (d,  $J = 2.9$  Hz, 6H), 3.67 (s, 3H), 3.21 (s, 1H).  $^{13}\text{C}$  NMR (151 MHz, Chloroform-*d*)  $\delta$  173.5, 159.0, 152.3, 140.5, 131.4, 128.1, 115.4, 114.7, 114.1, 74.8, 59.5, 55.7, 55.2, 52.9.

## 8. NMR Spectroscopic Data.

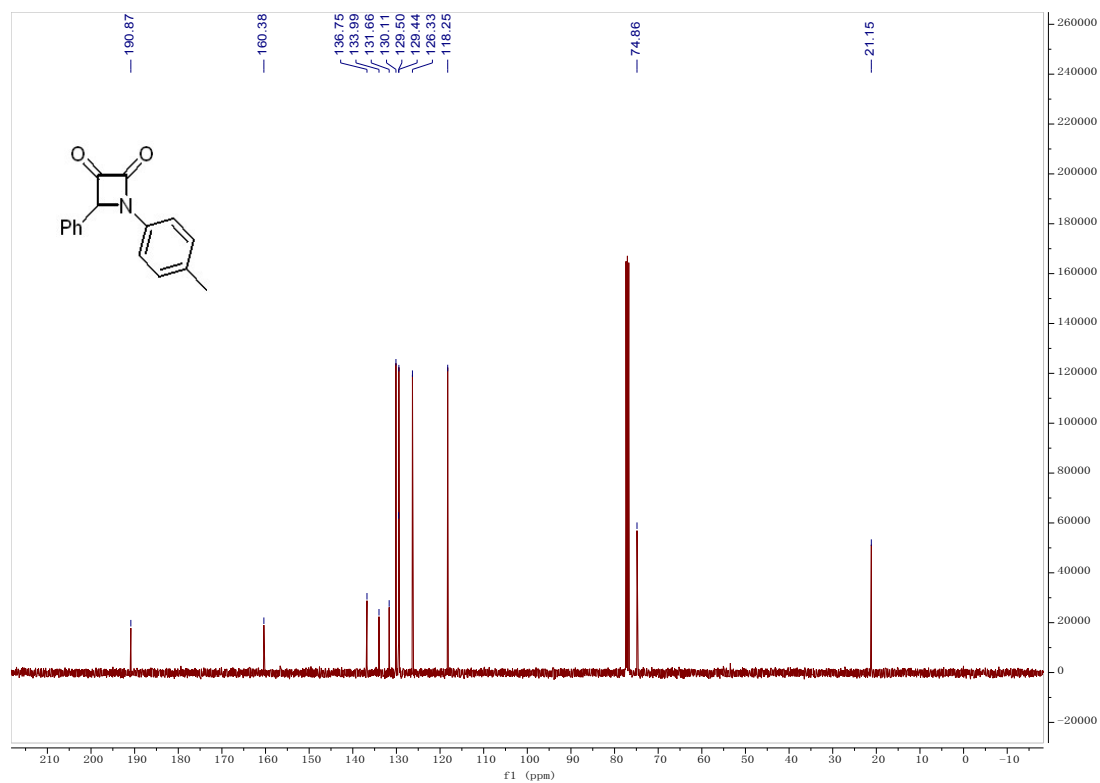
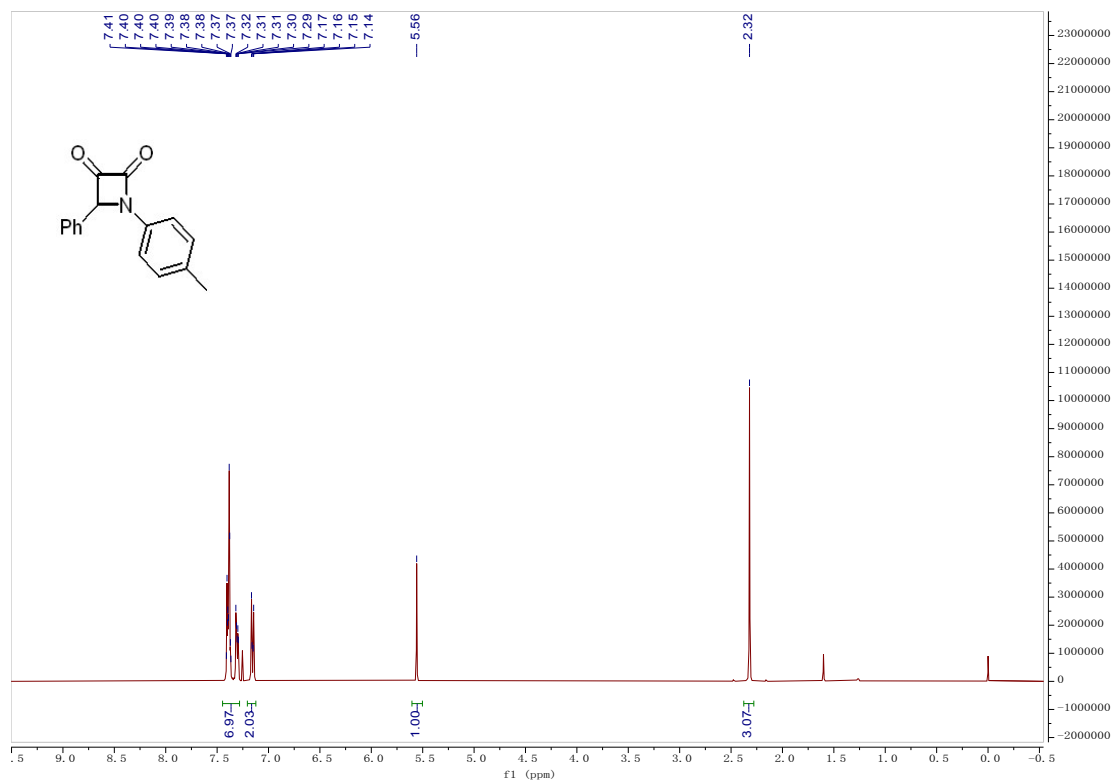
### $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 1a



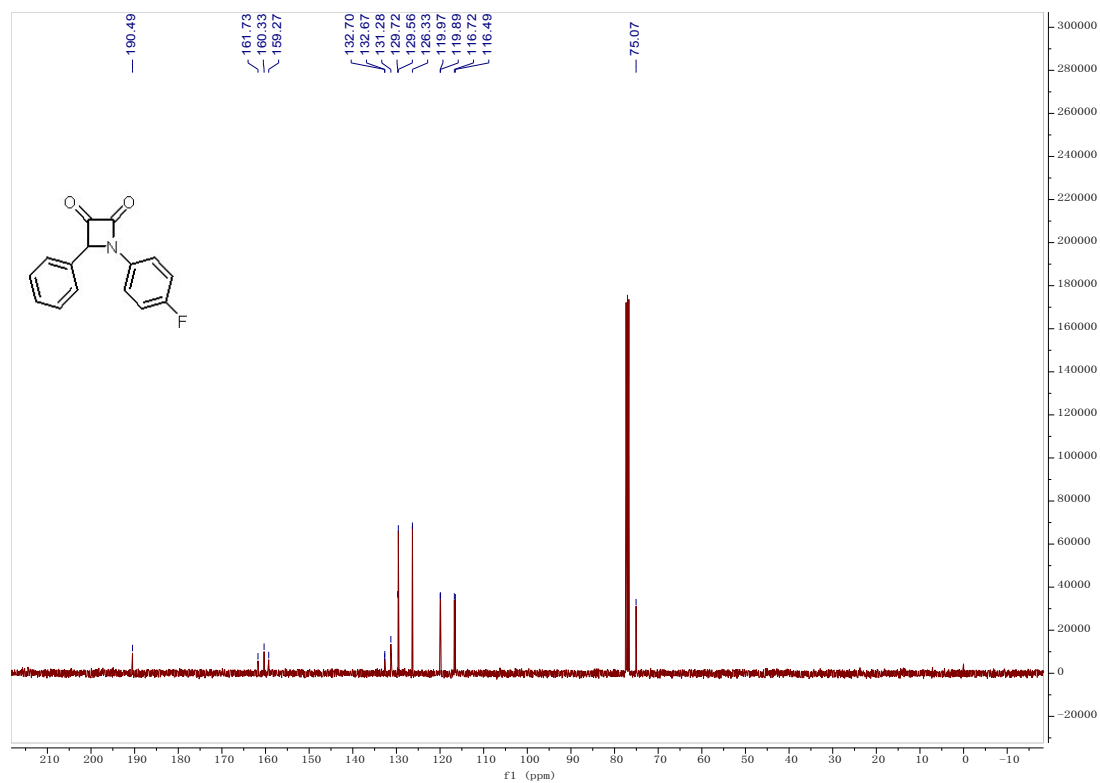
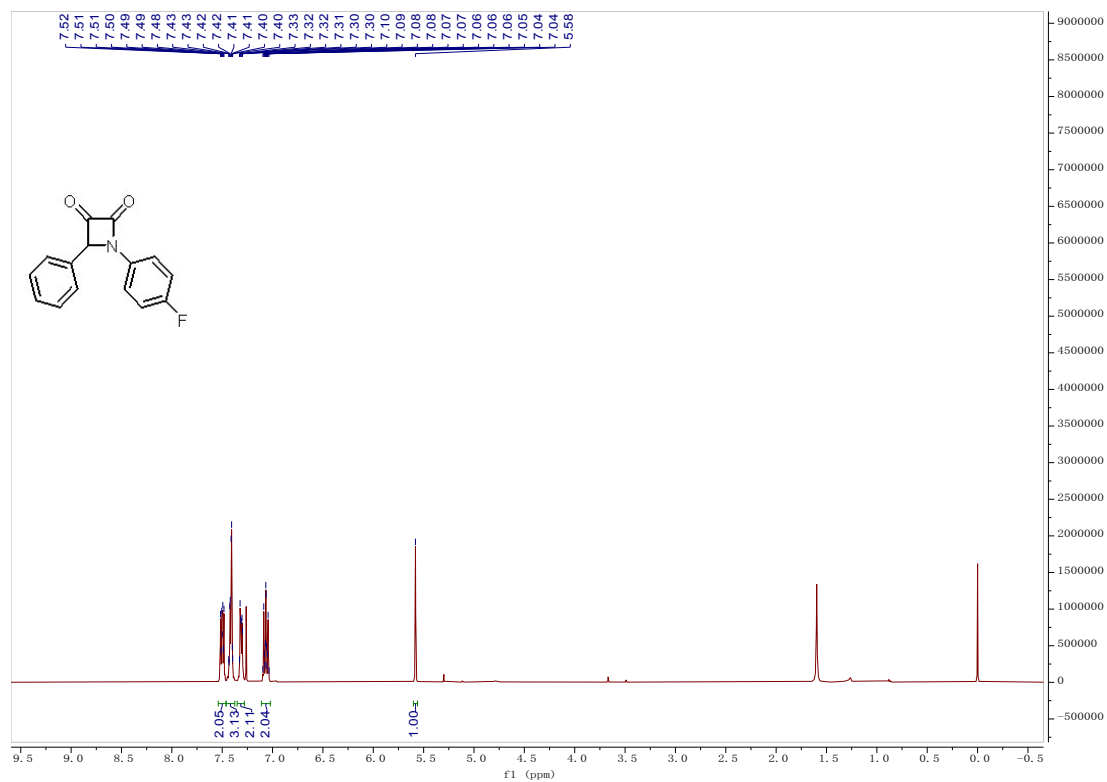
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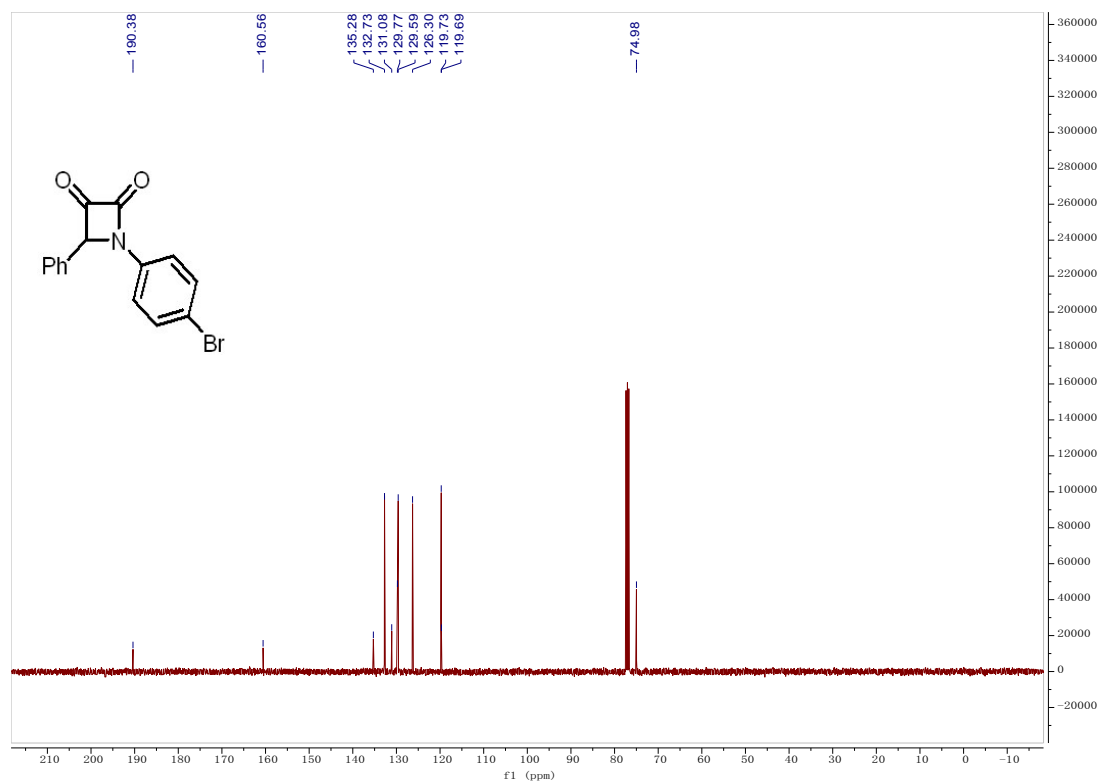
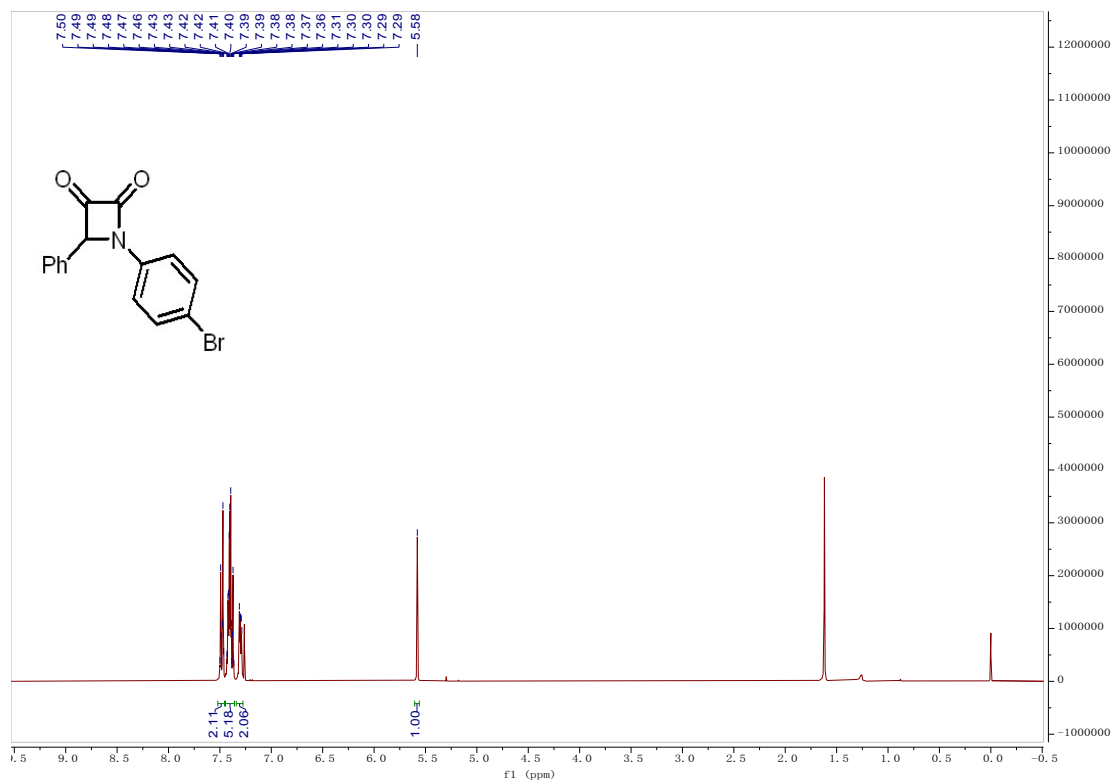
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1c



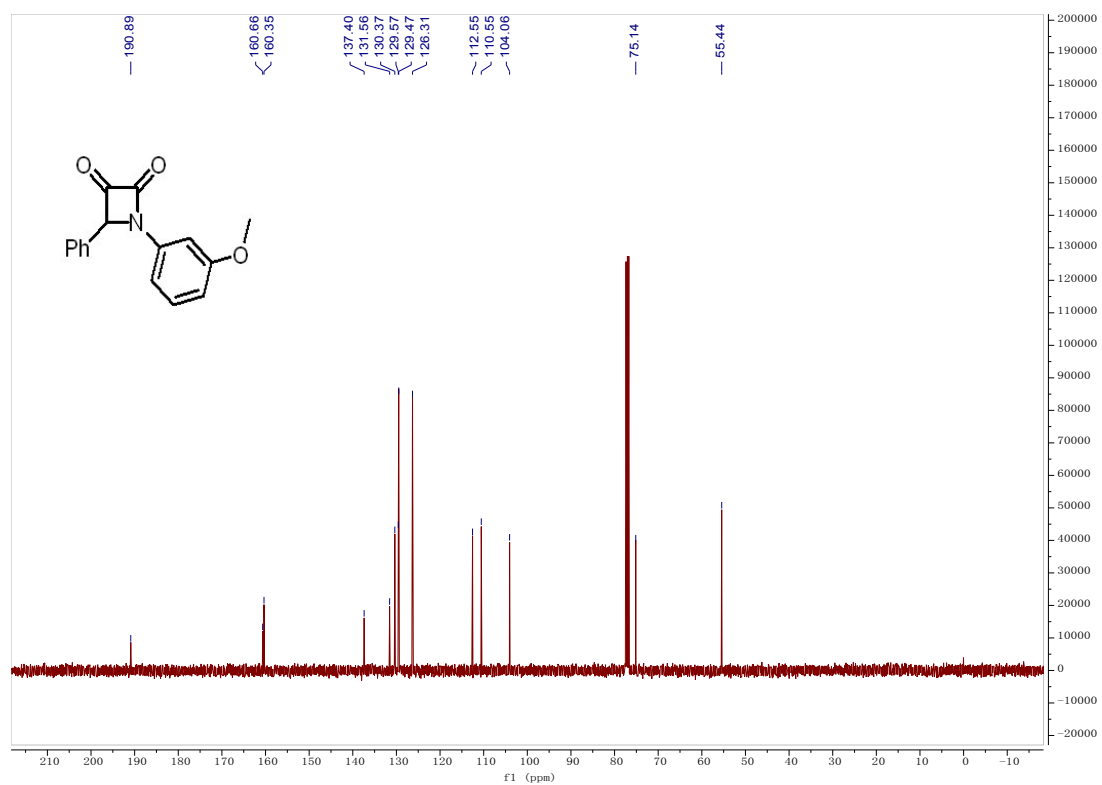
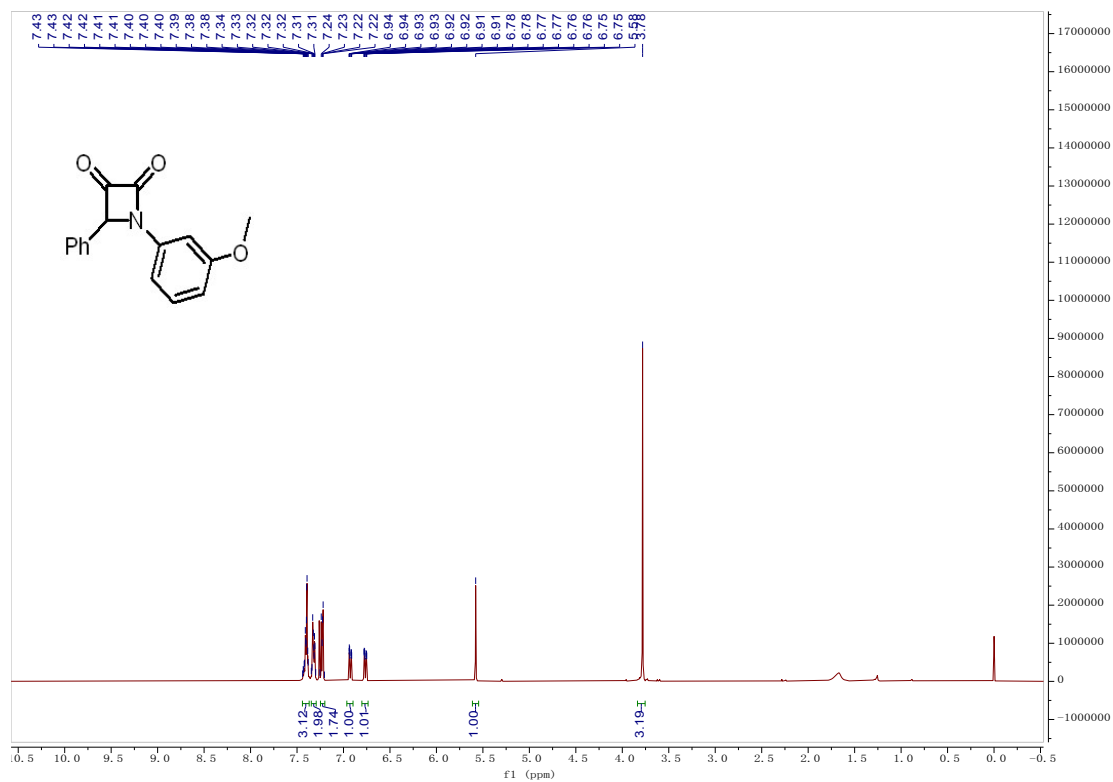
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1d



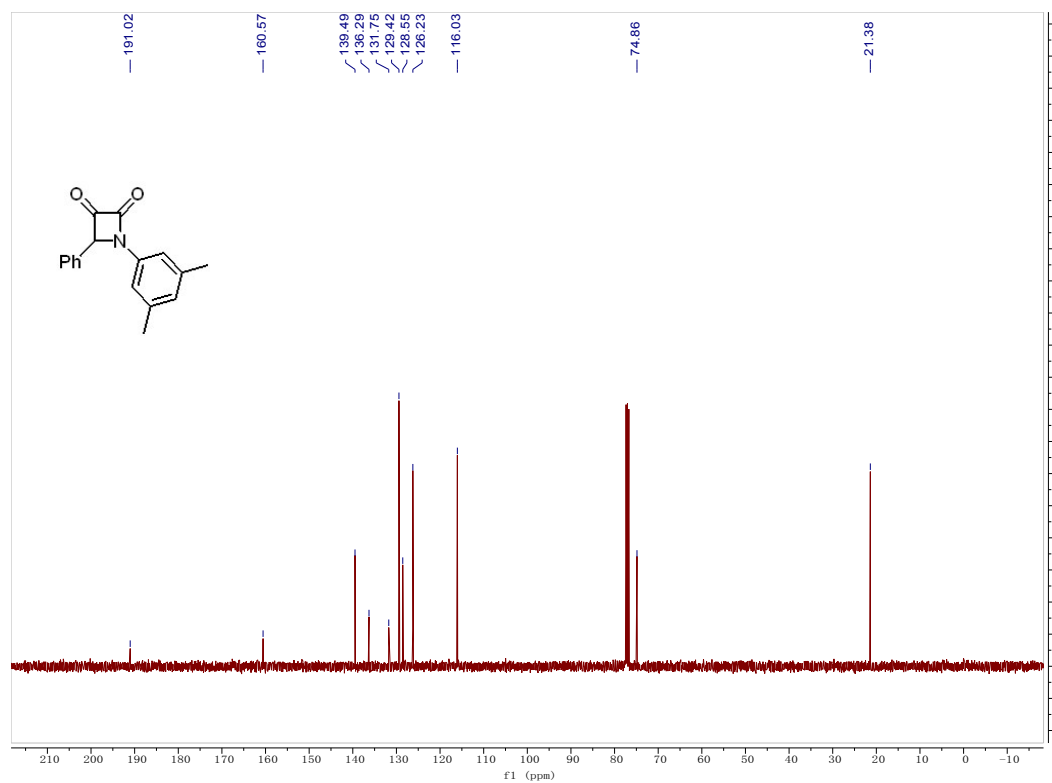
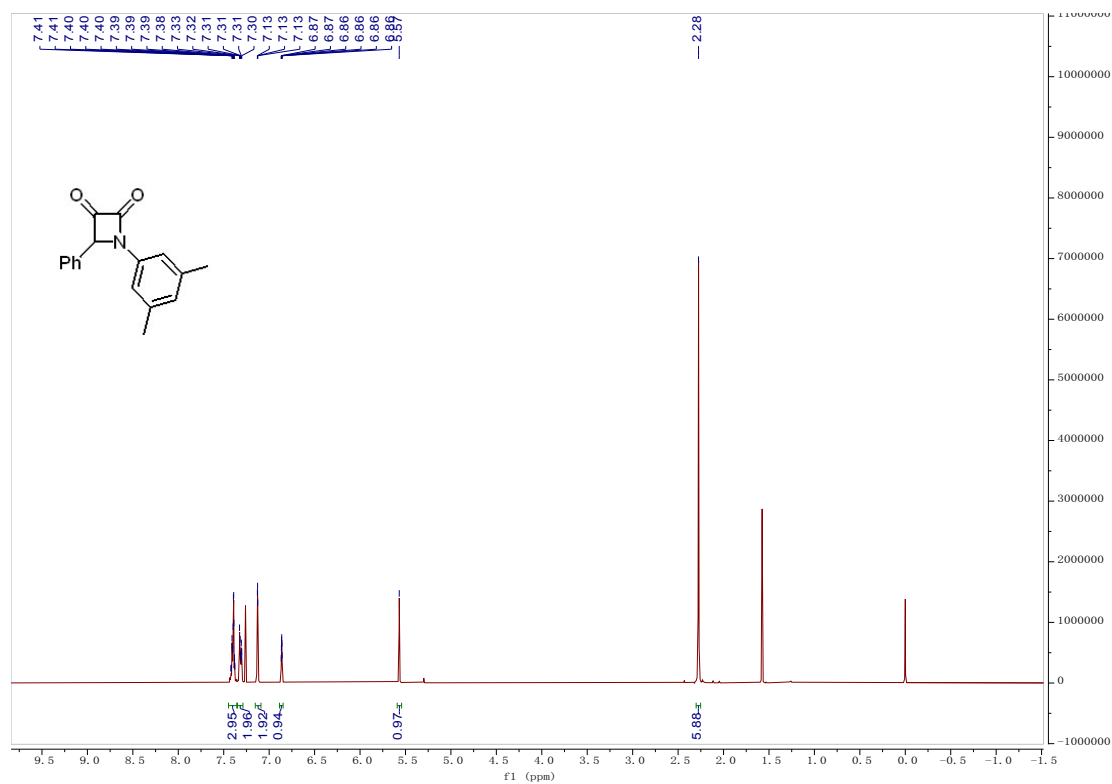
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# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1f

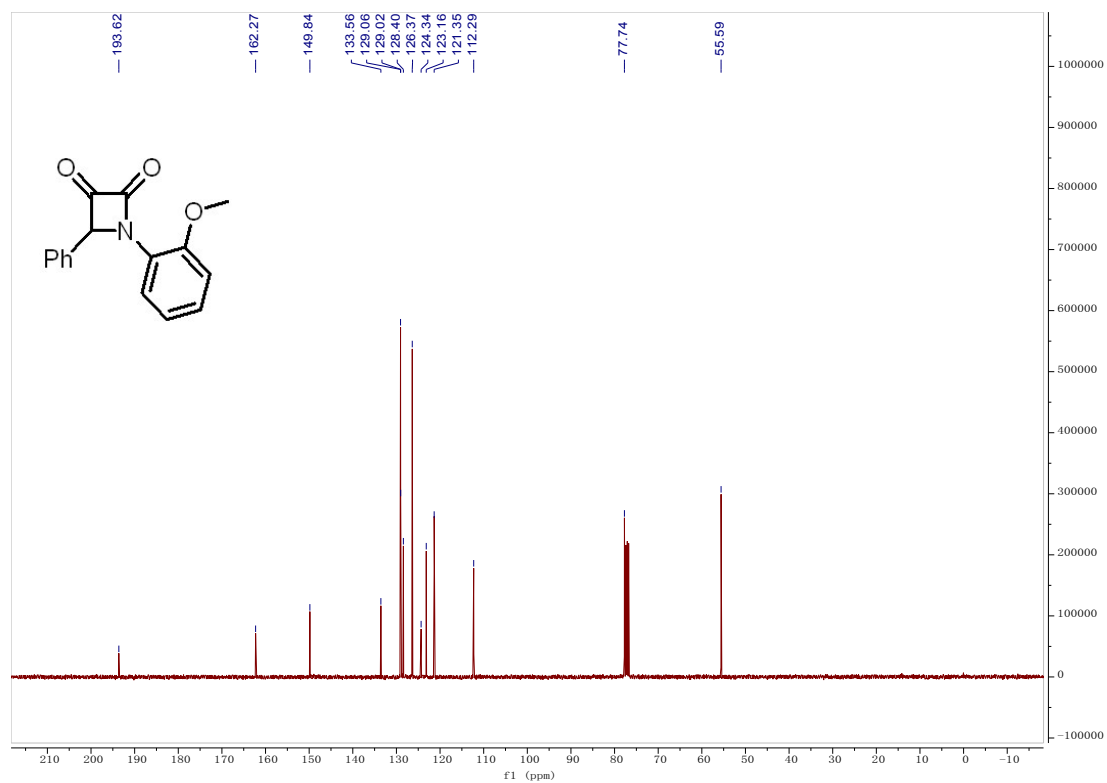
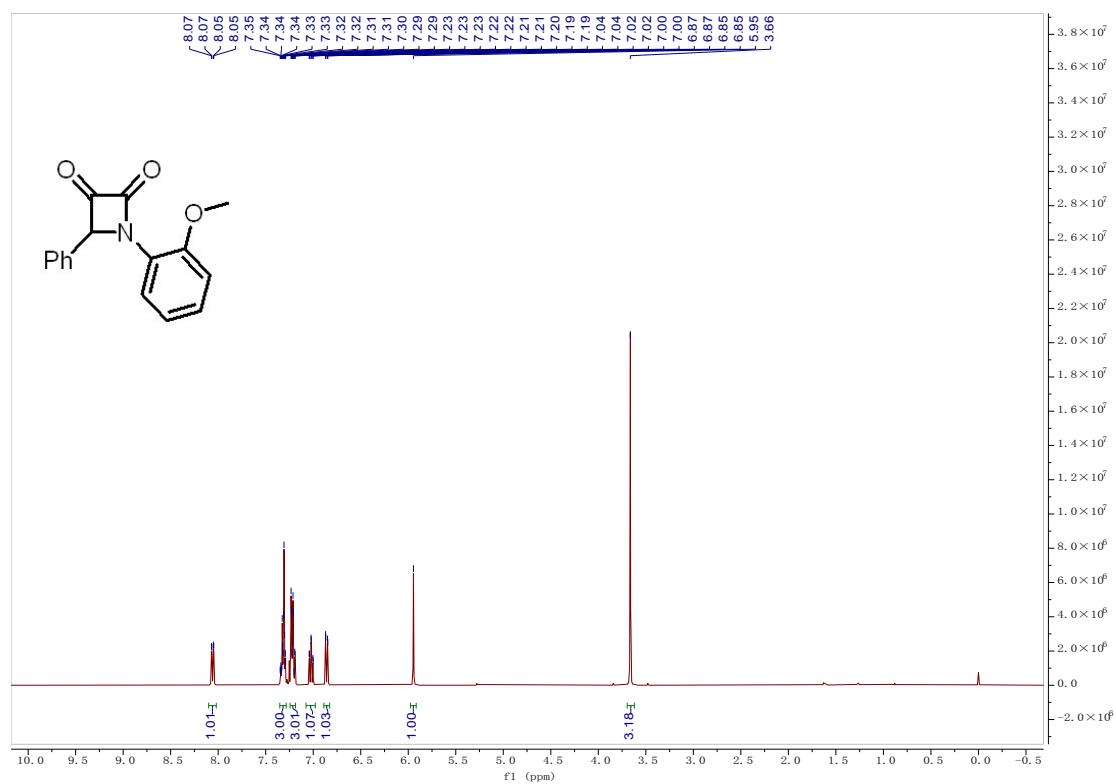


# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 1g

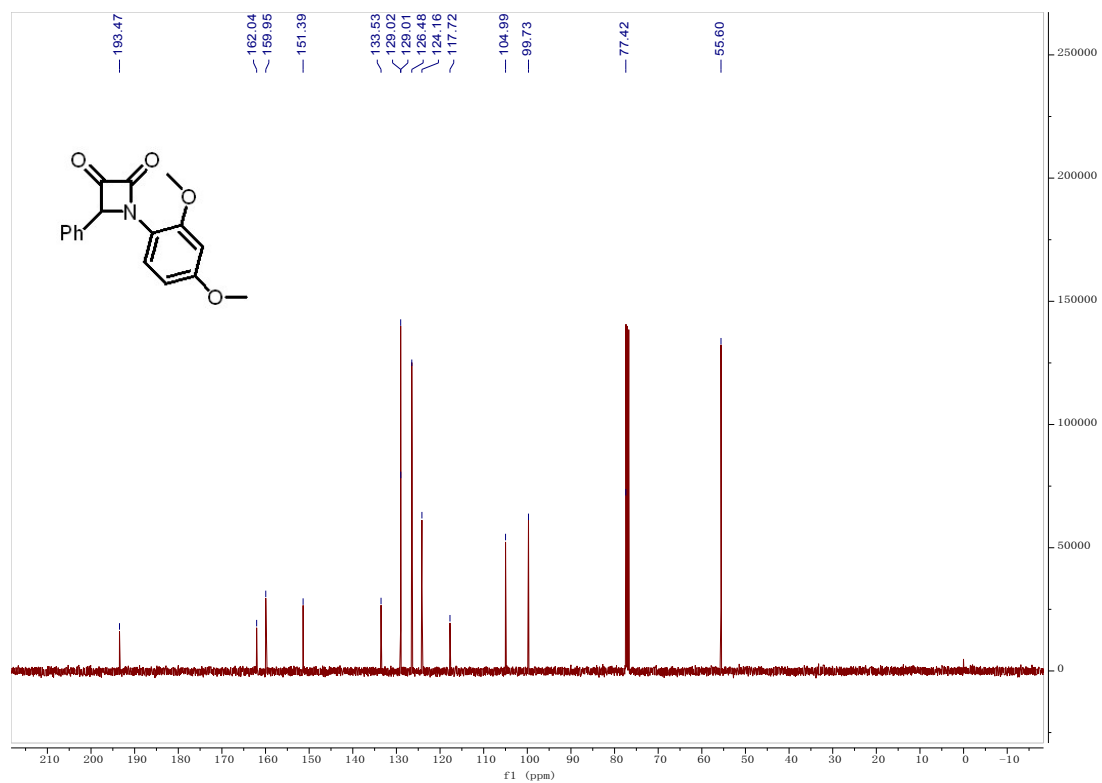
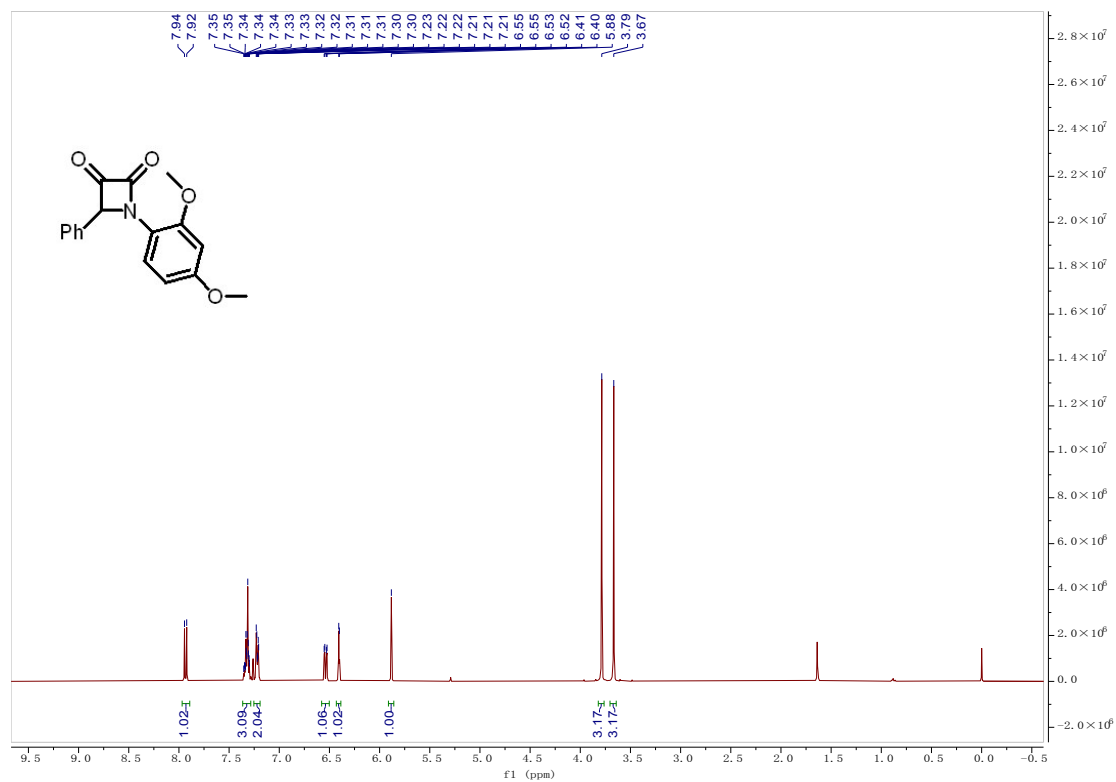




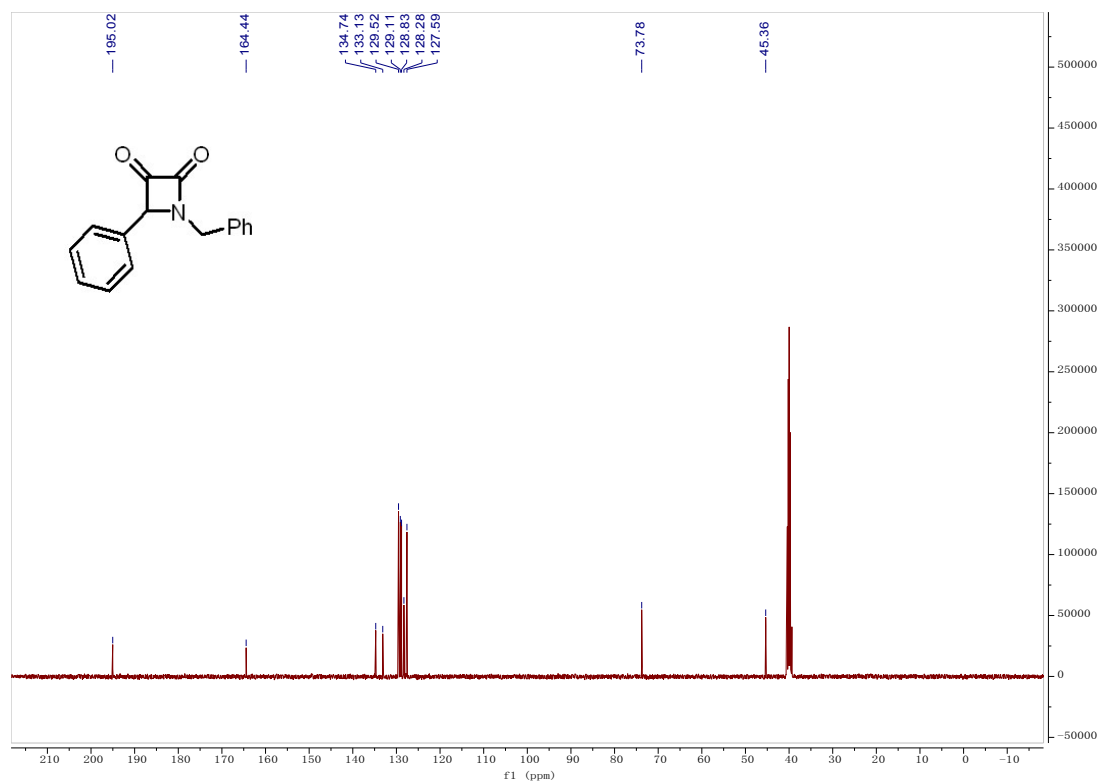
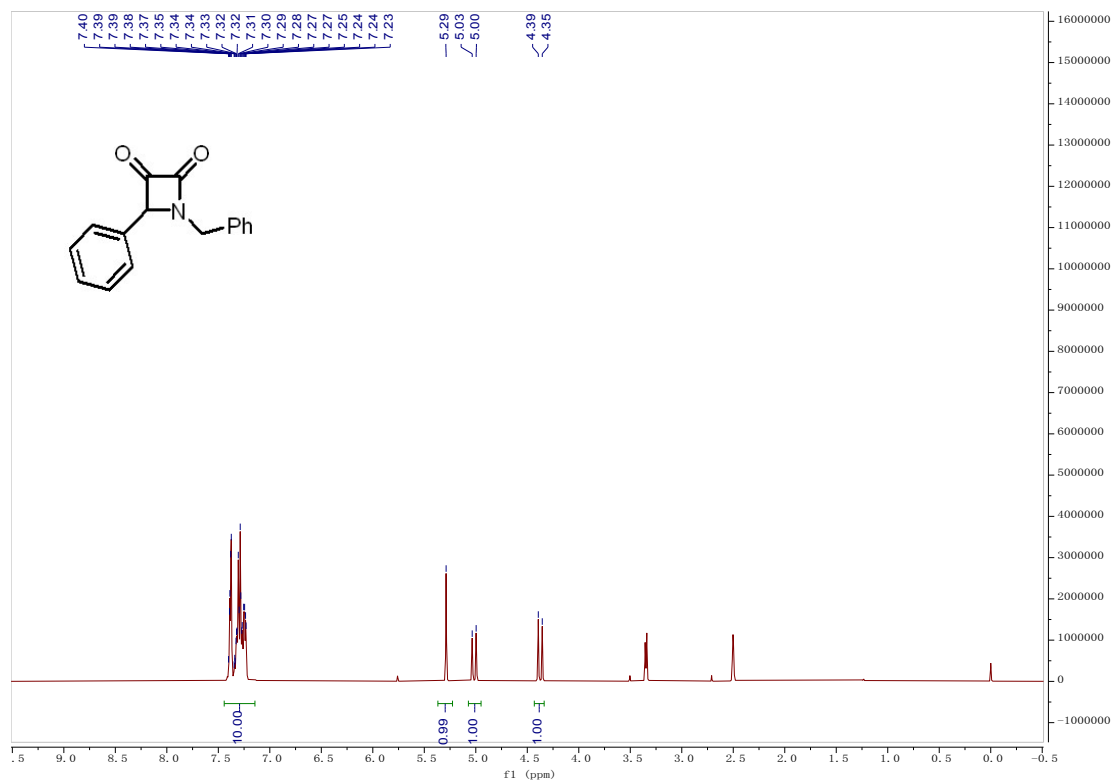
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1h



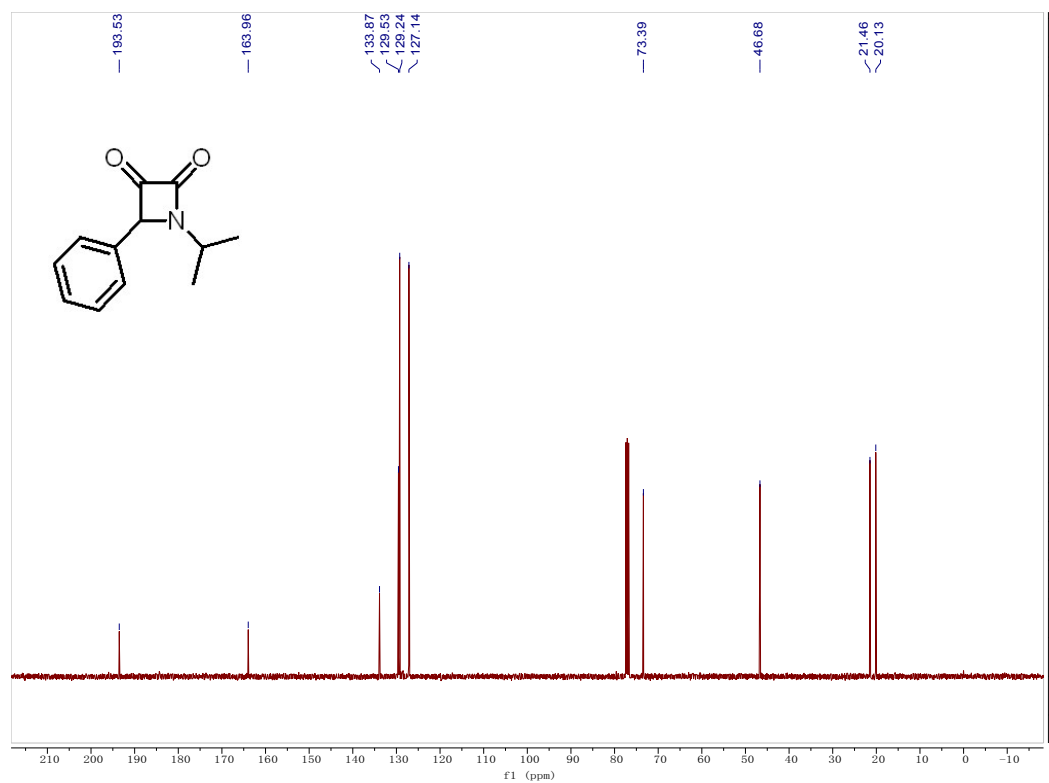
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1i



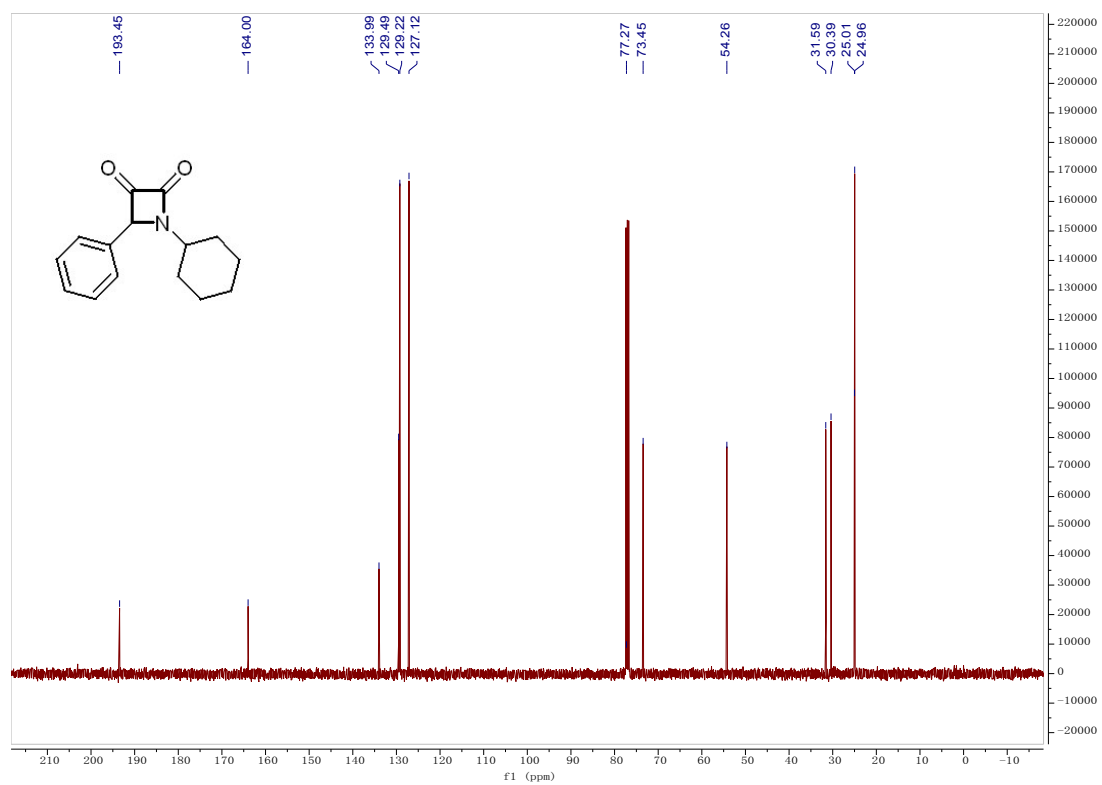
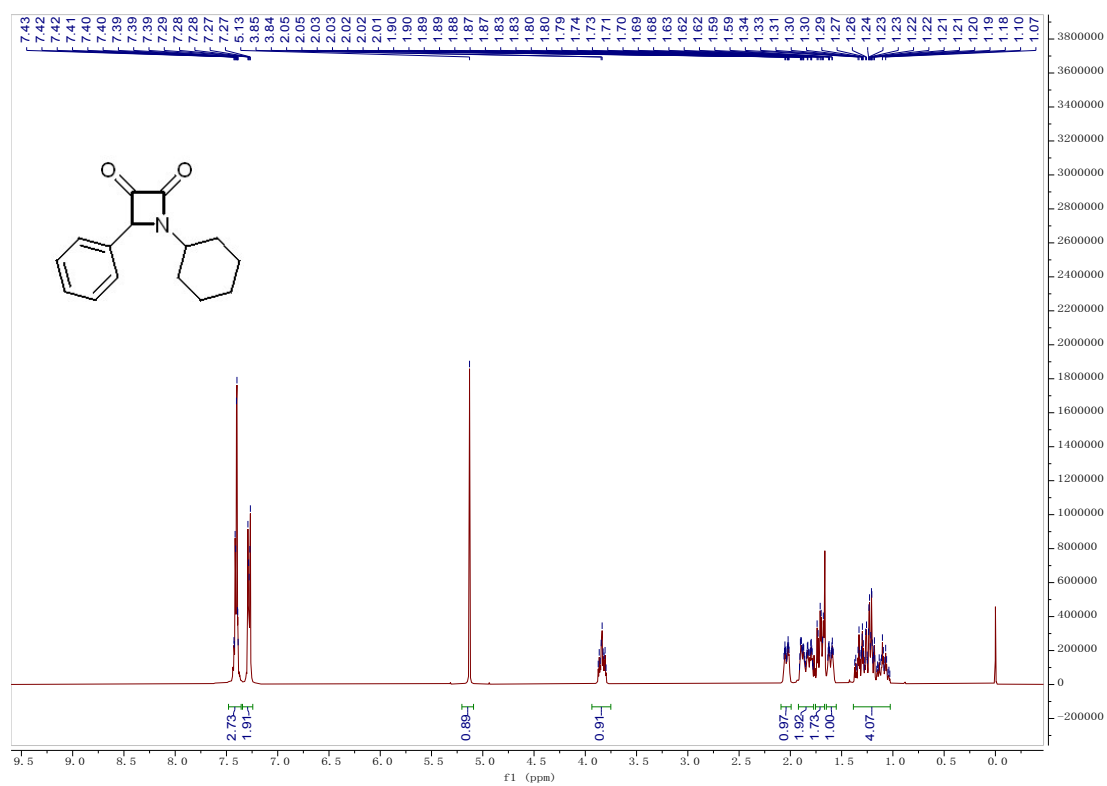
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1j



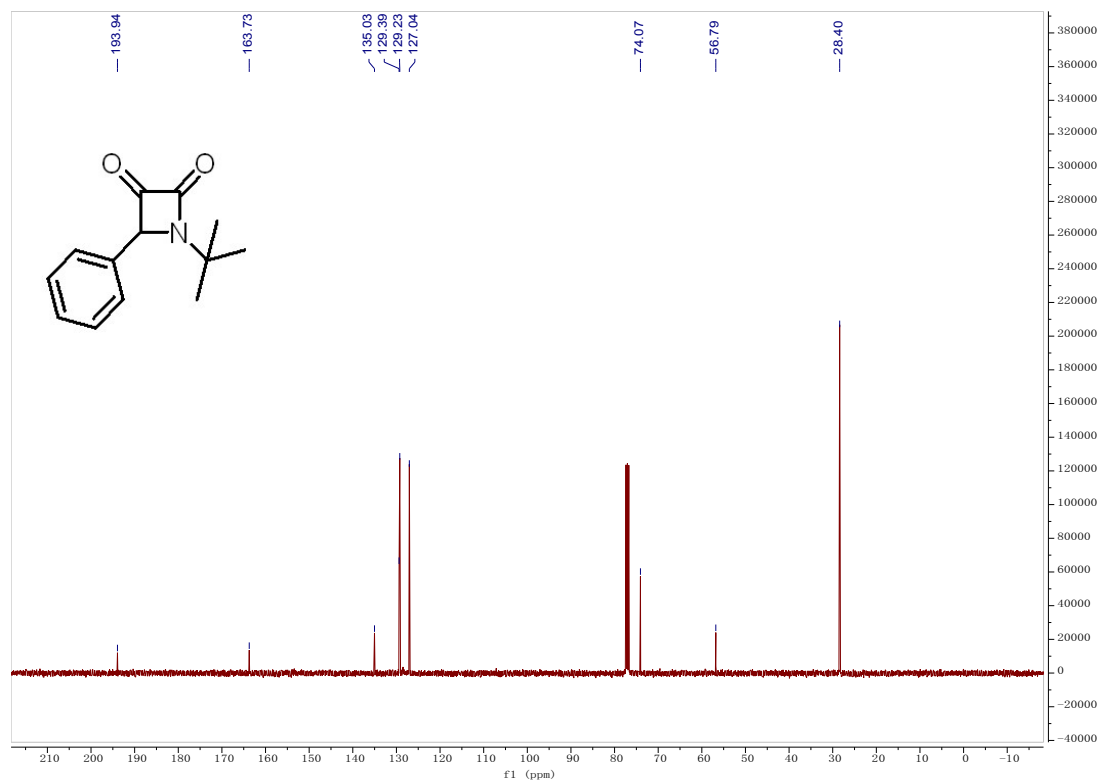
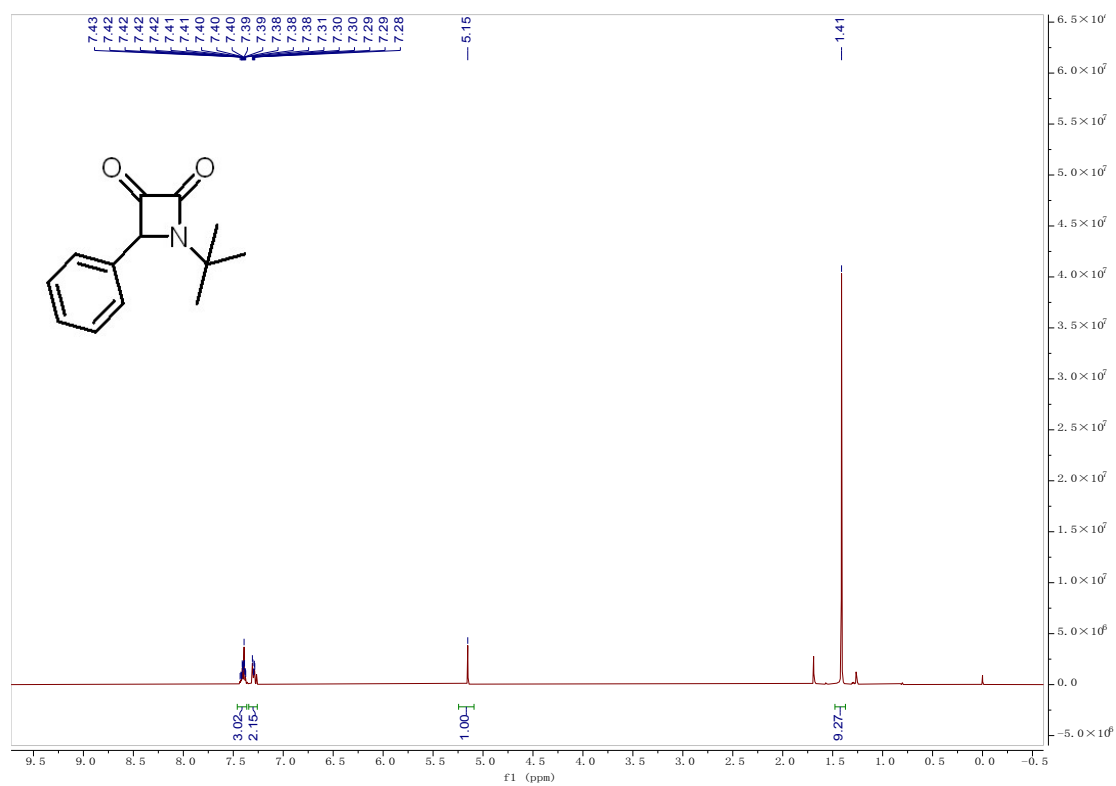
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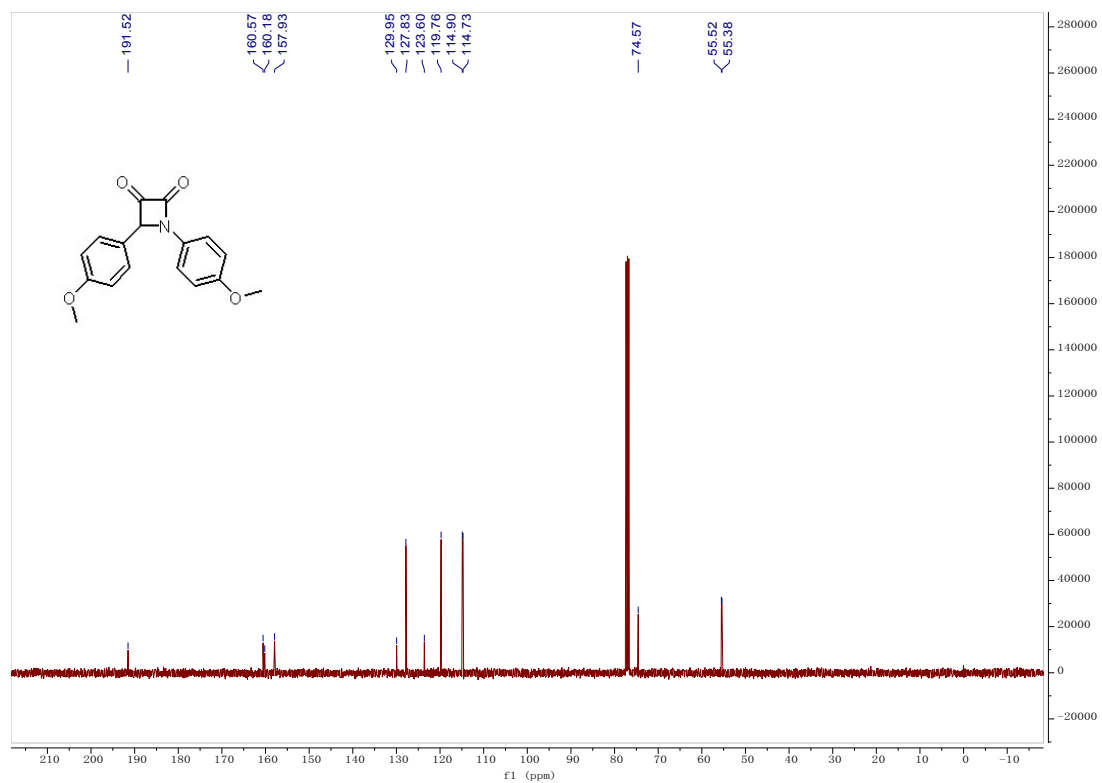
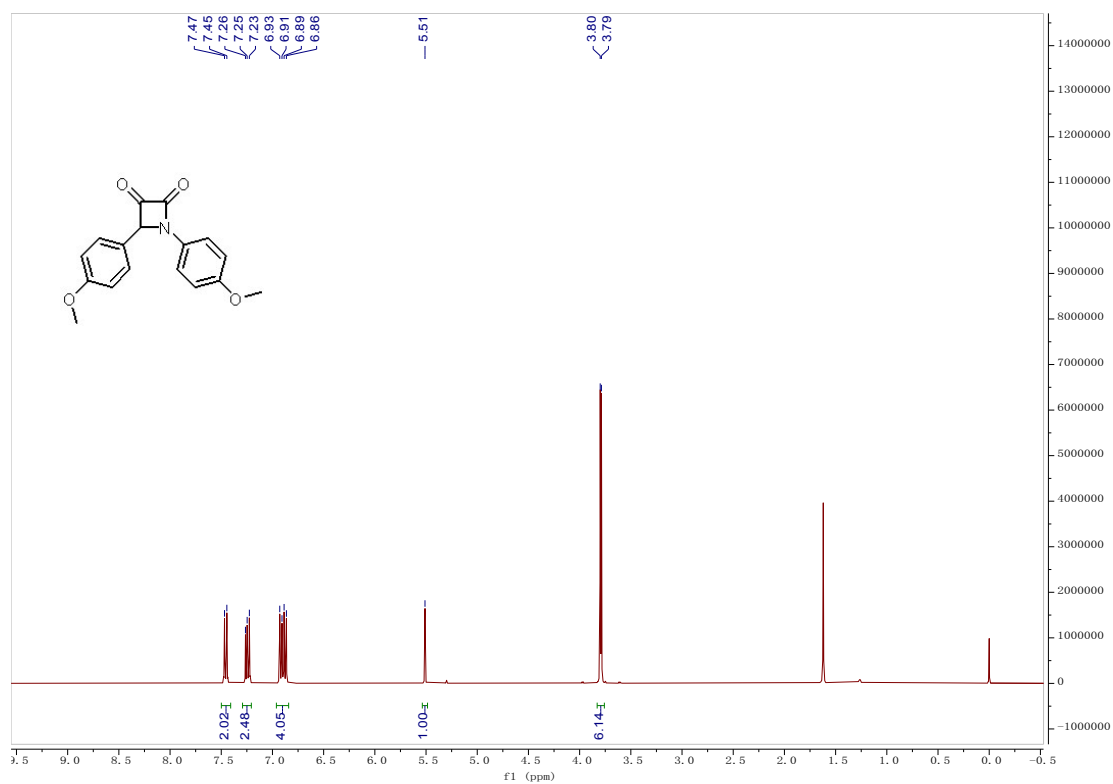
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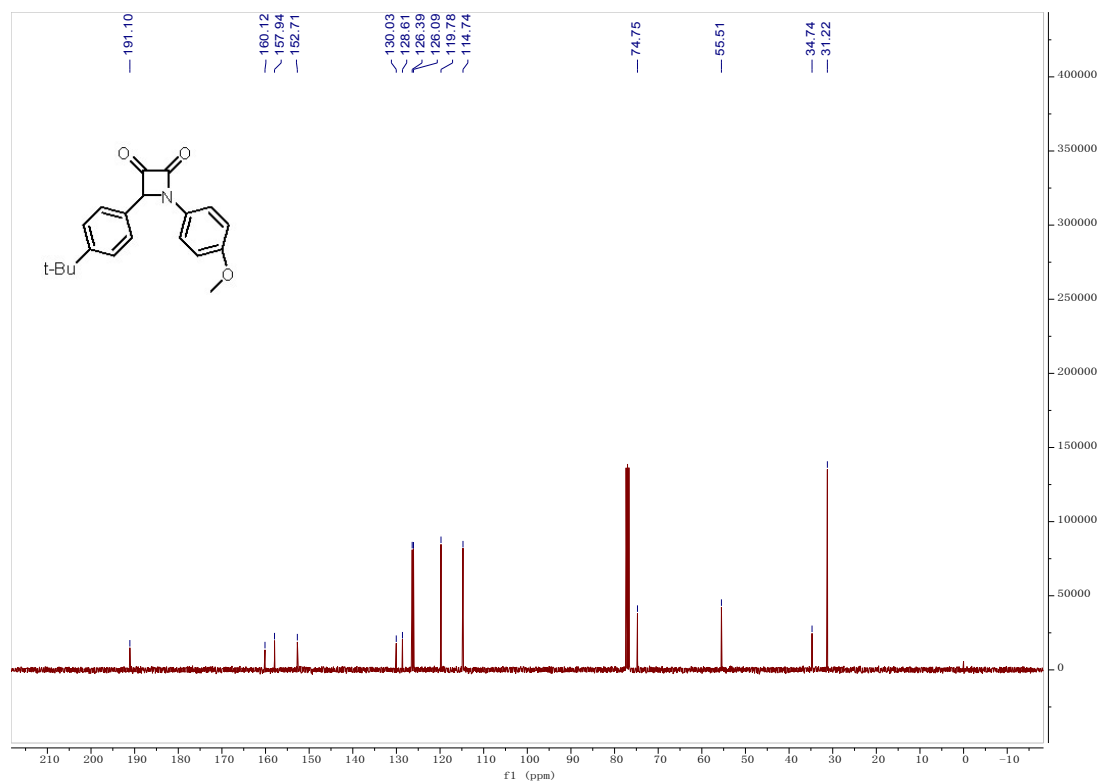
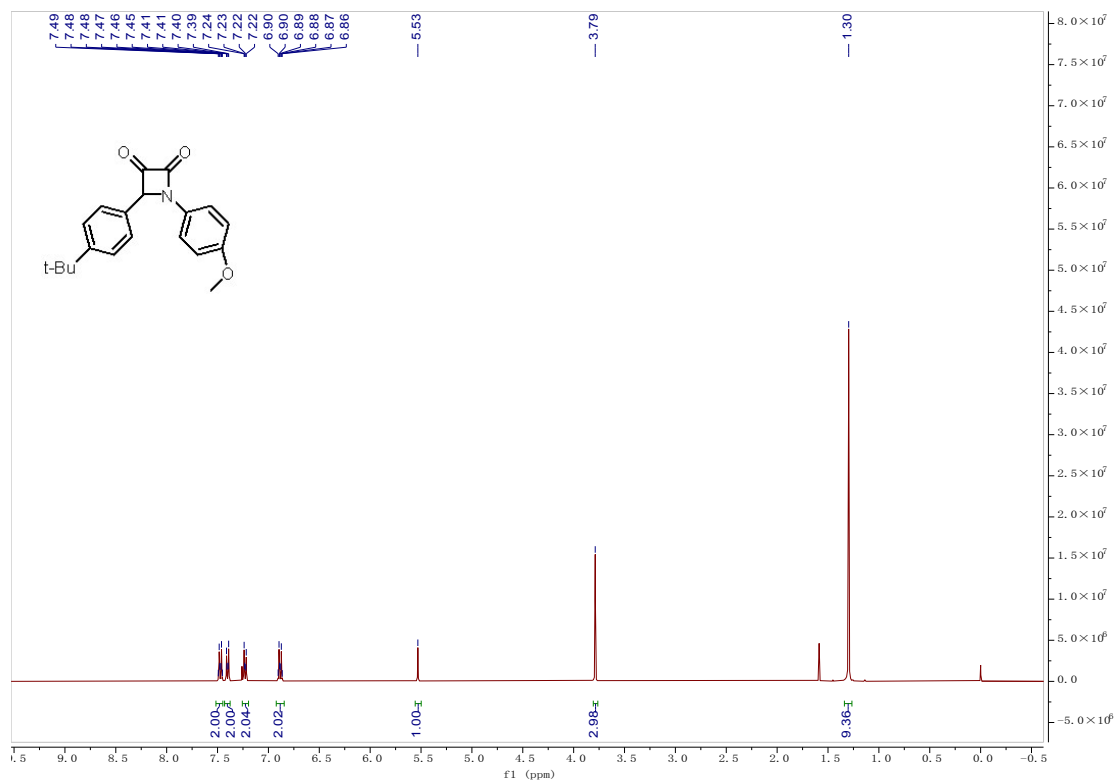
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1m



# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1n

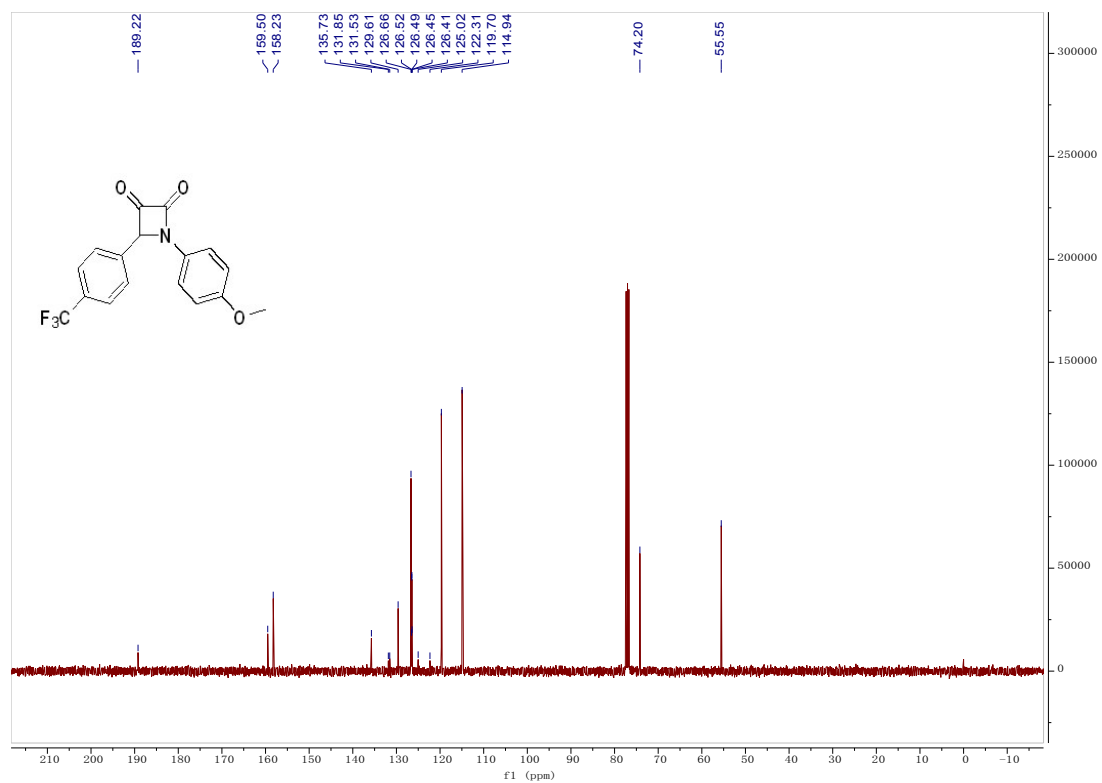
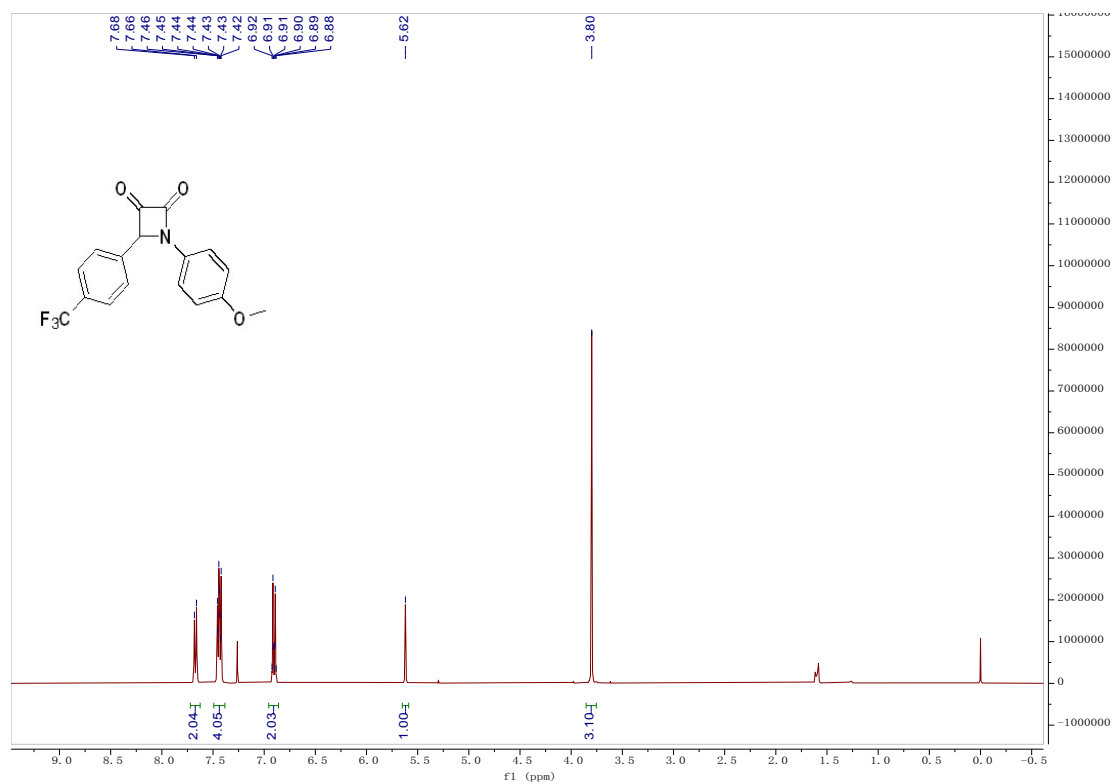


# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1o

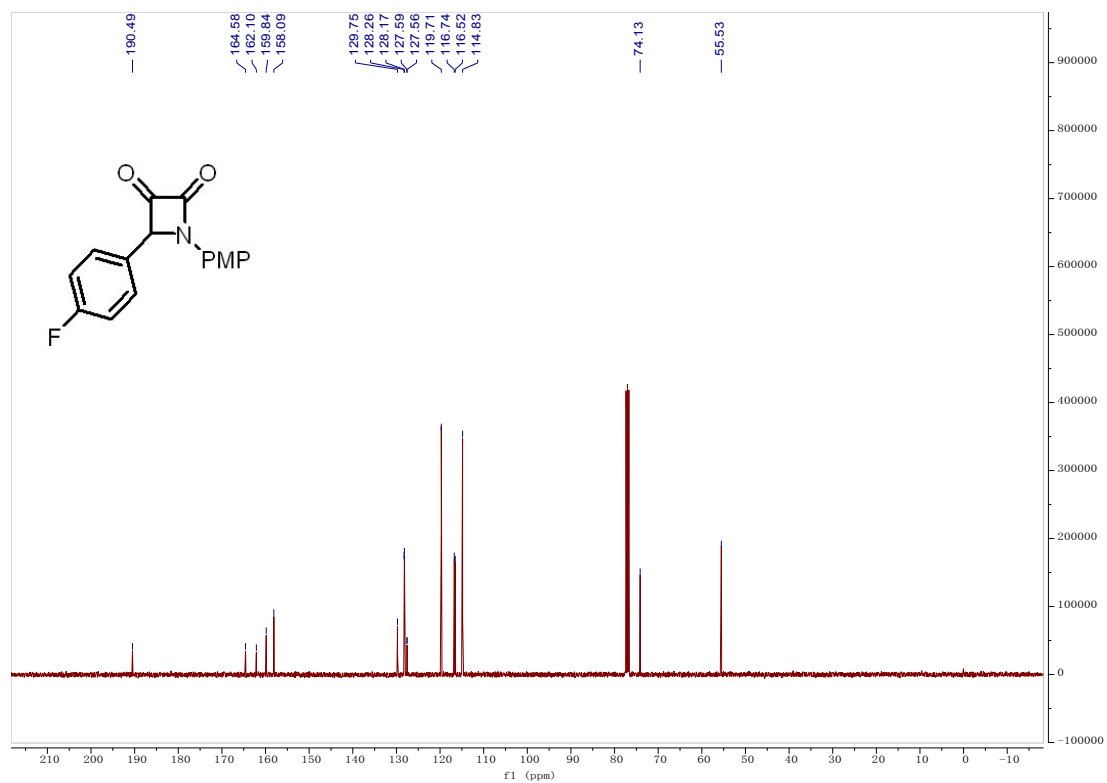
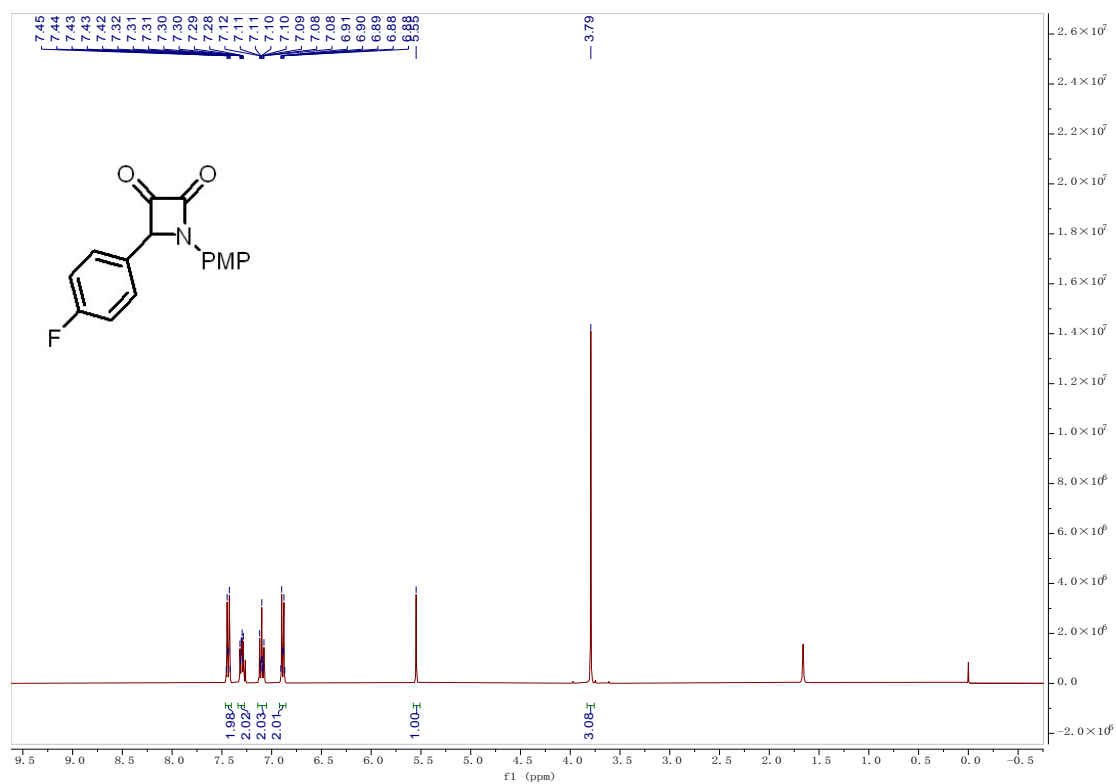




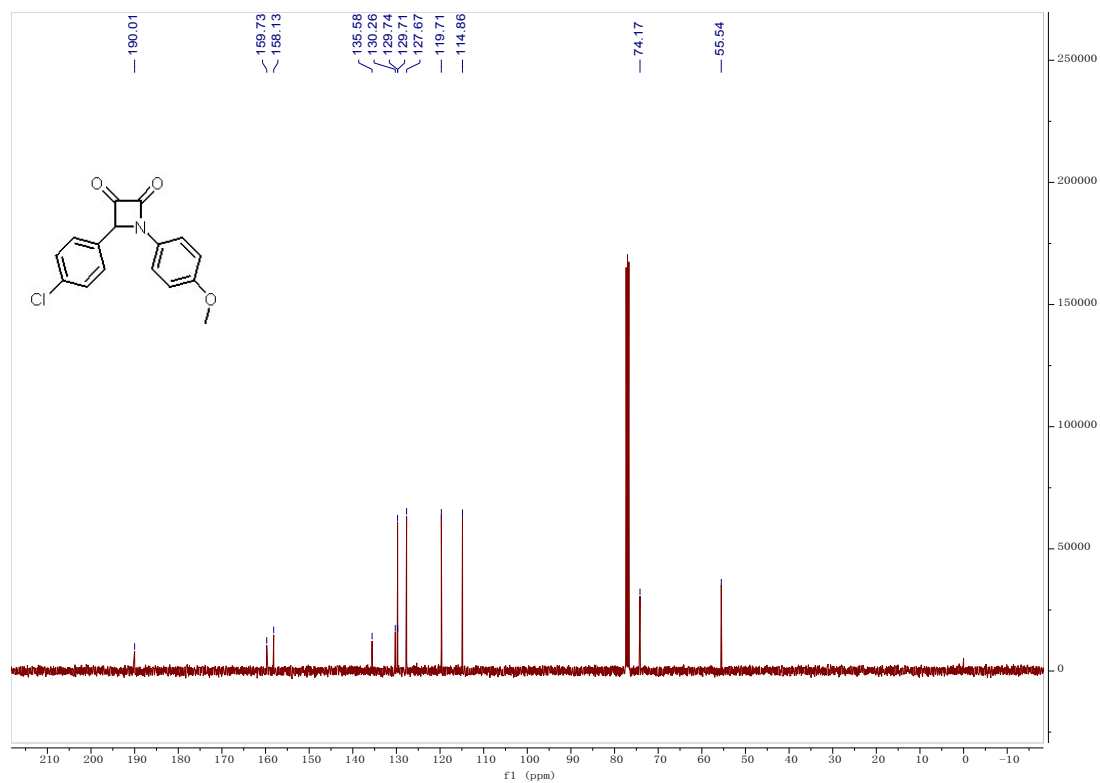
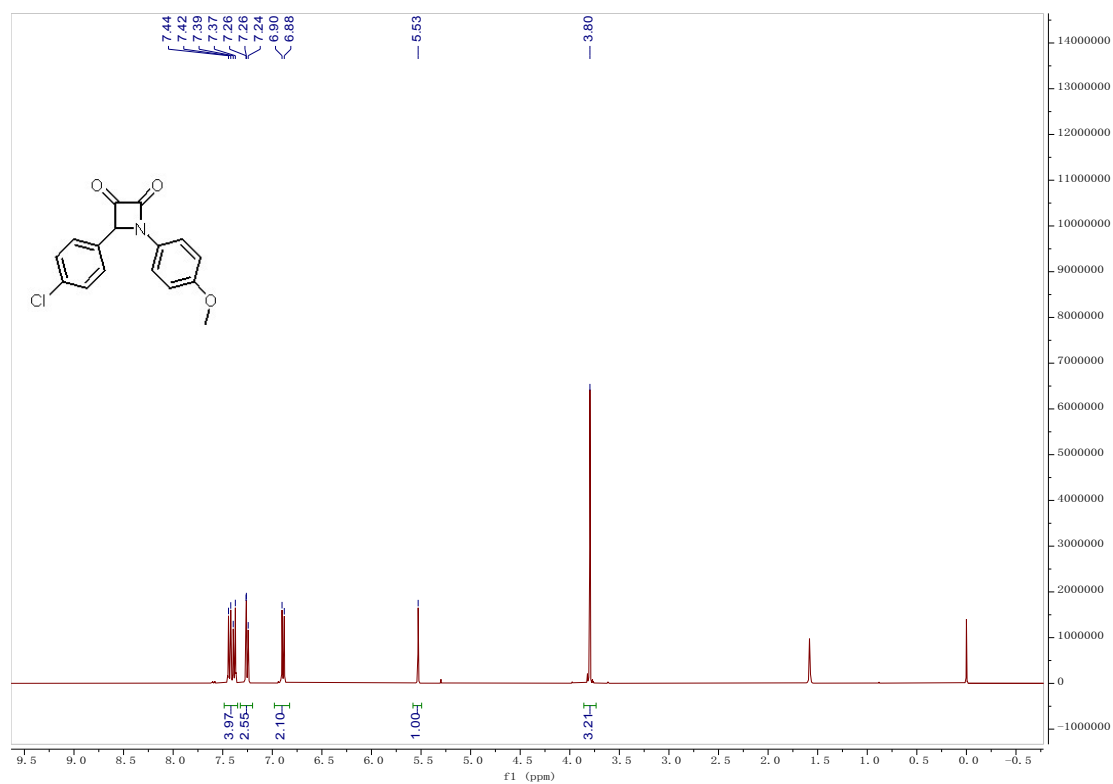
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1p



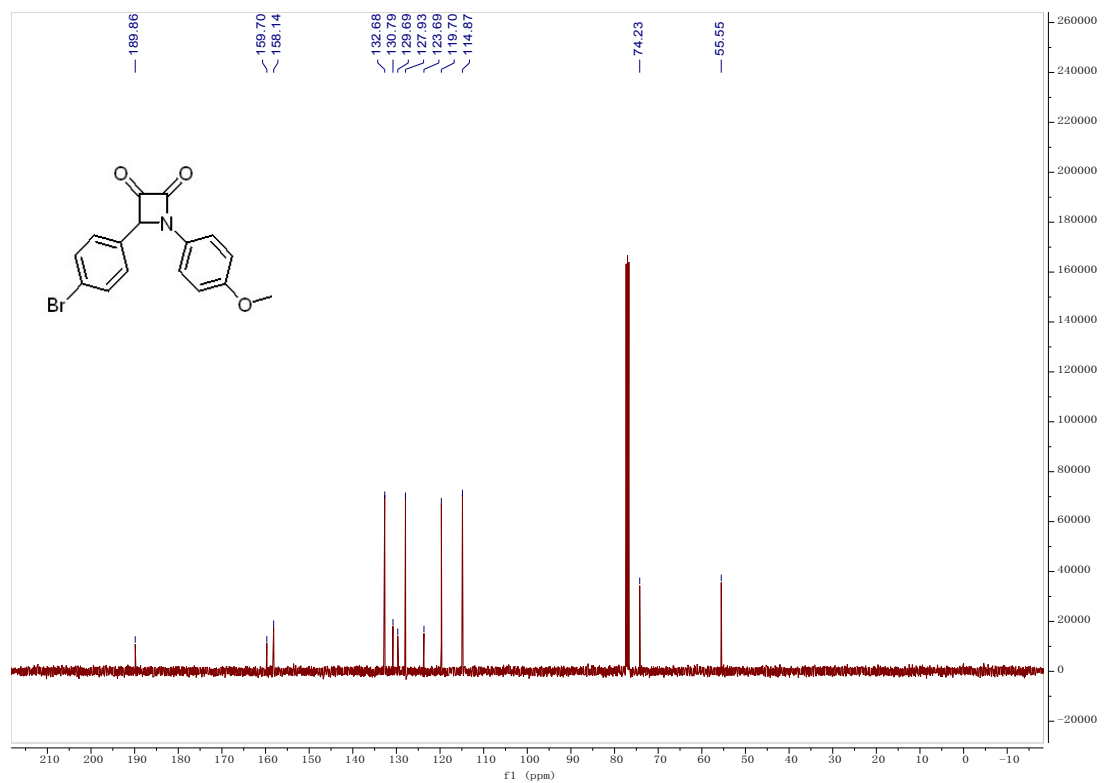
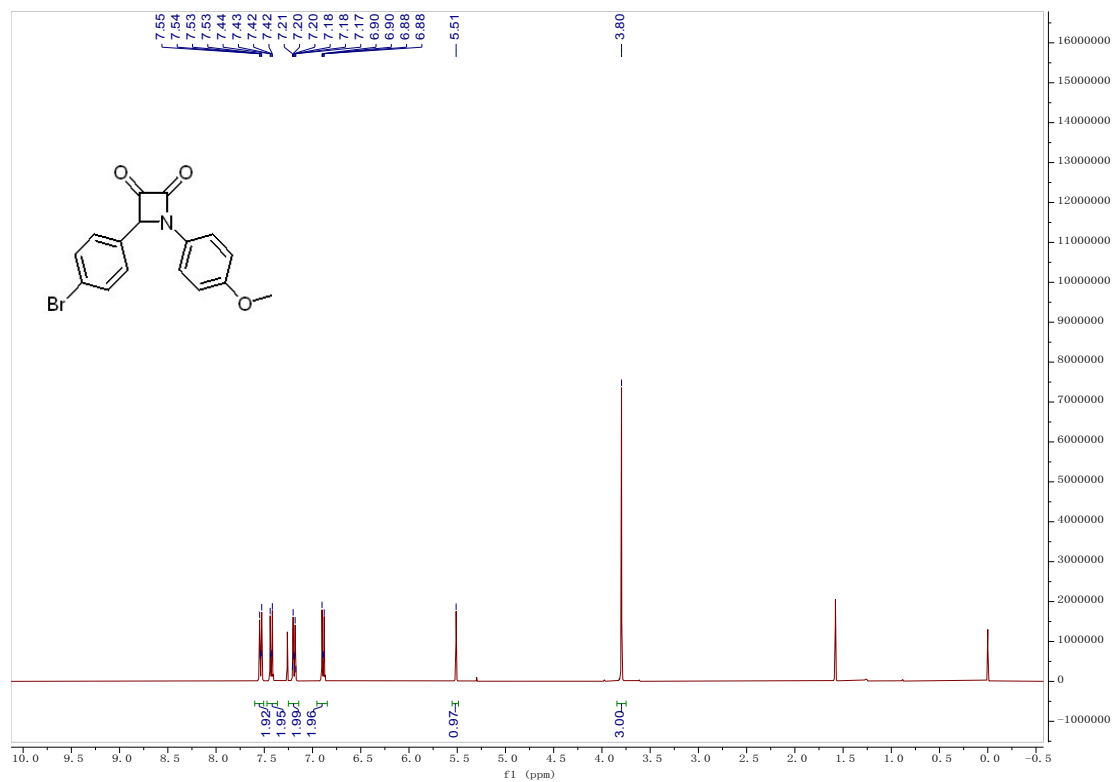
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1q



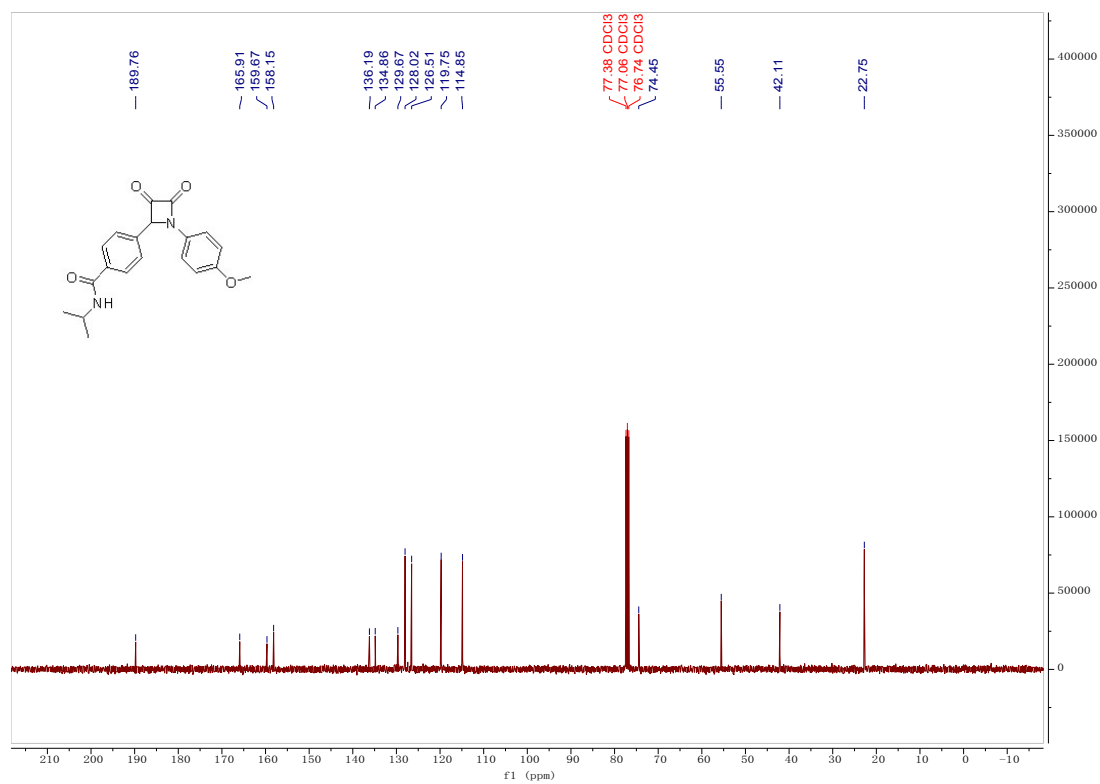
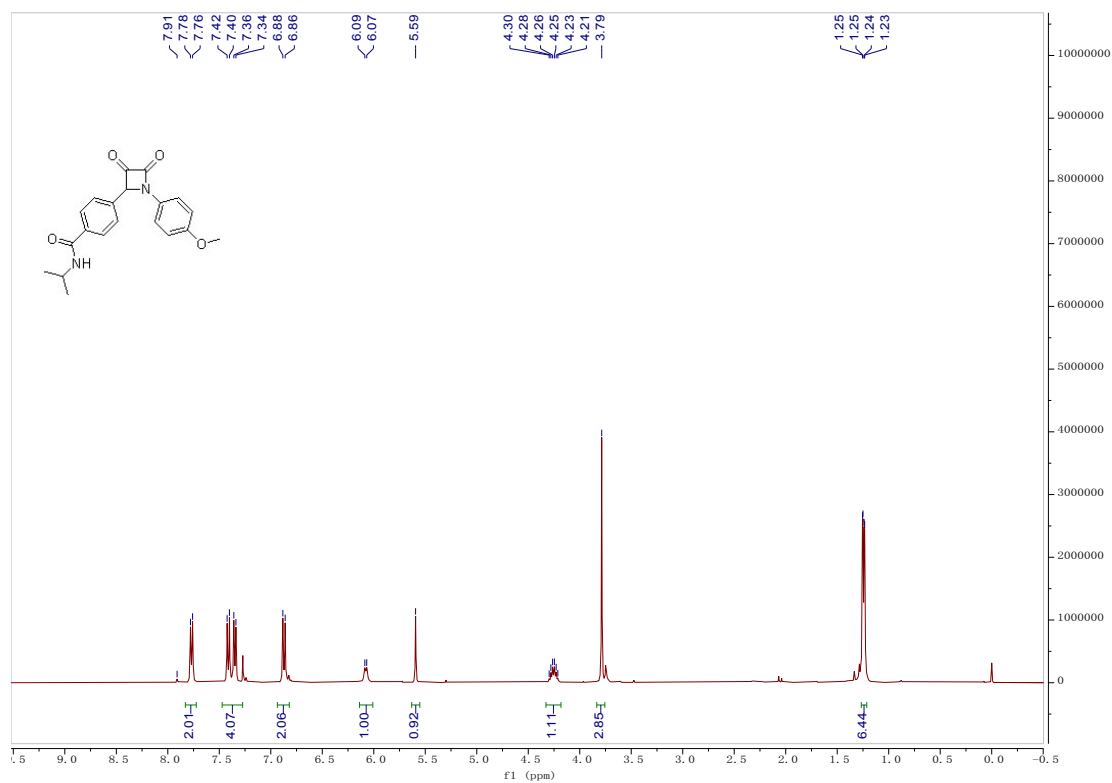
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 1r



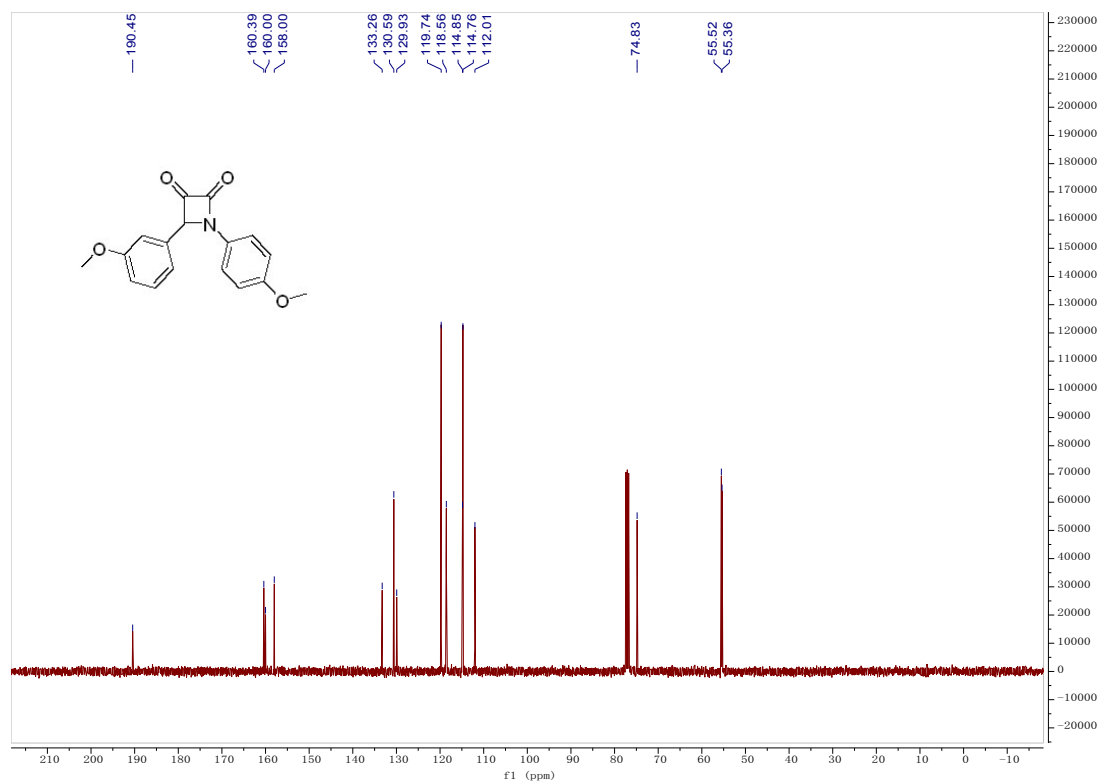
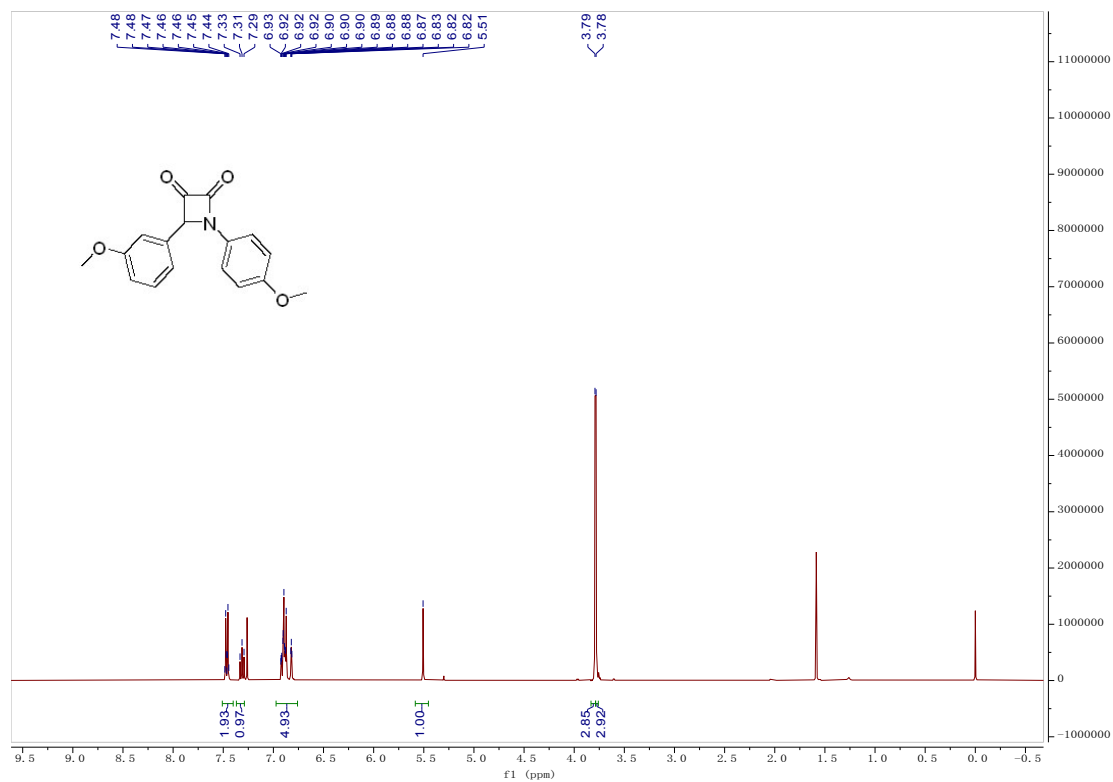
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1s



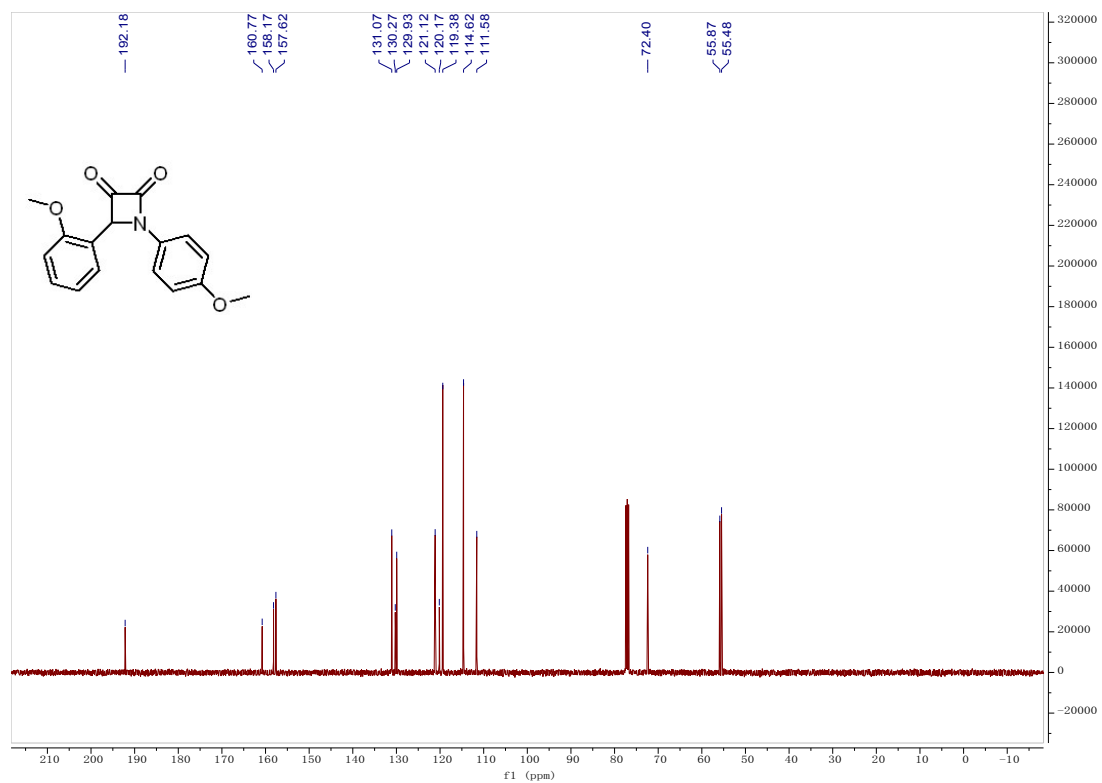
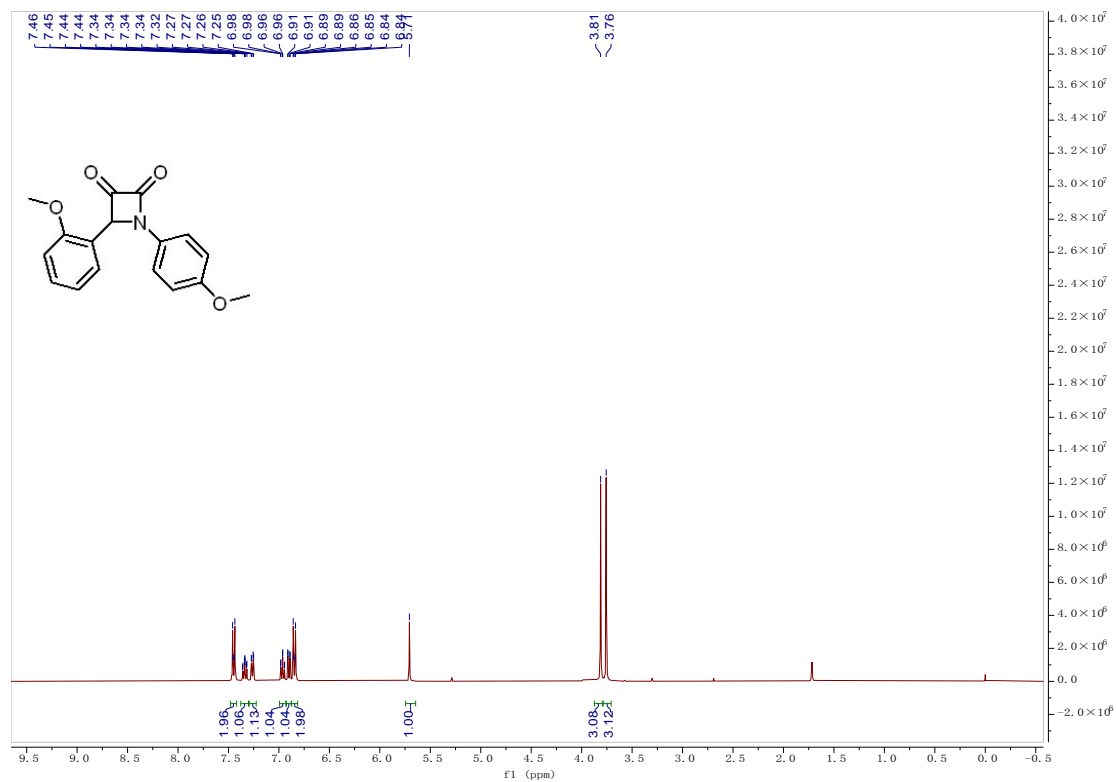
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1t



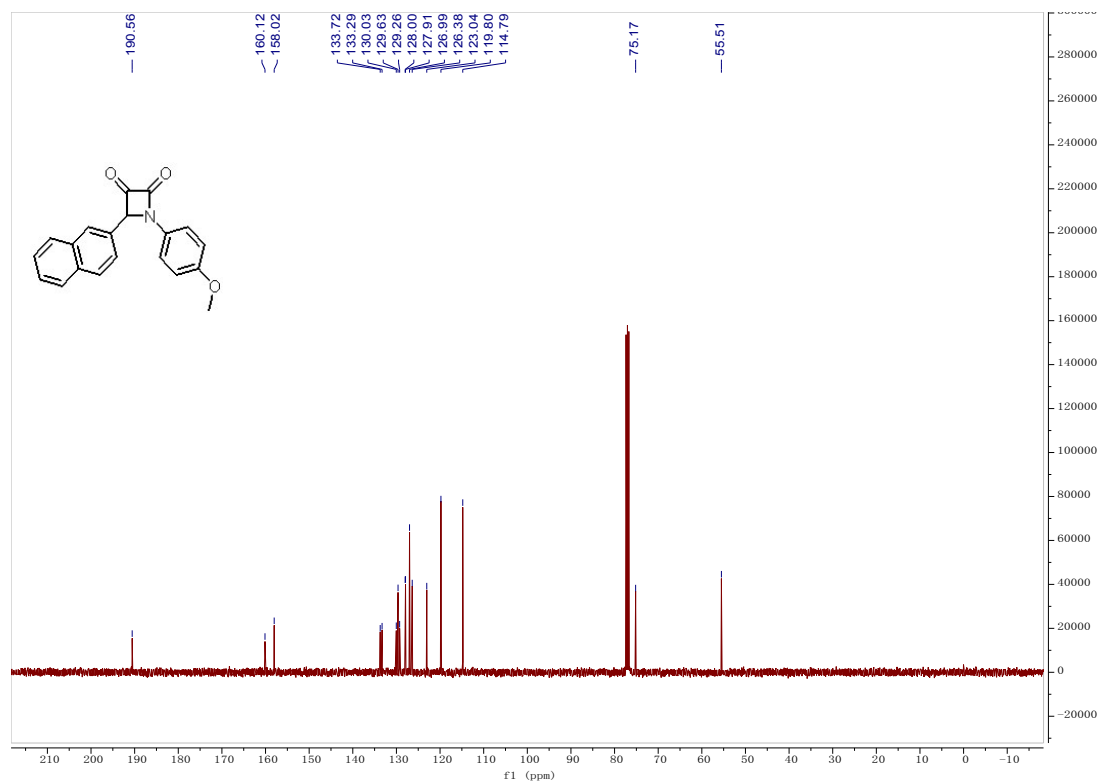
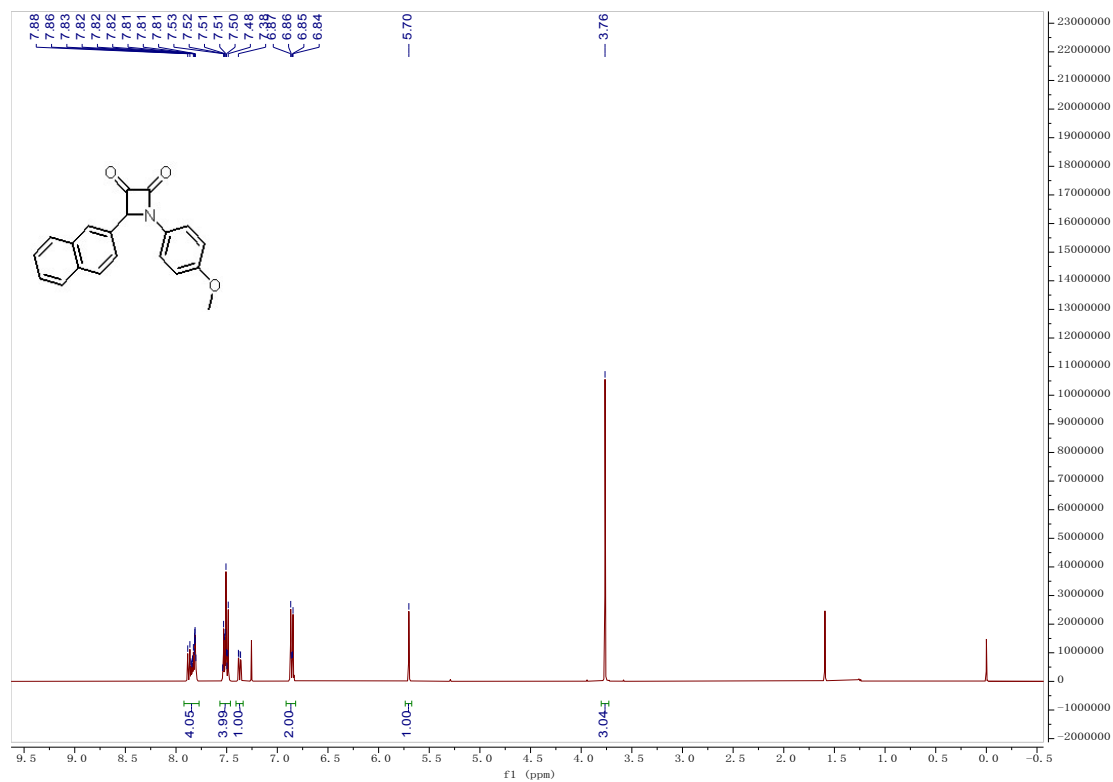
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 1u



# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1v

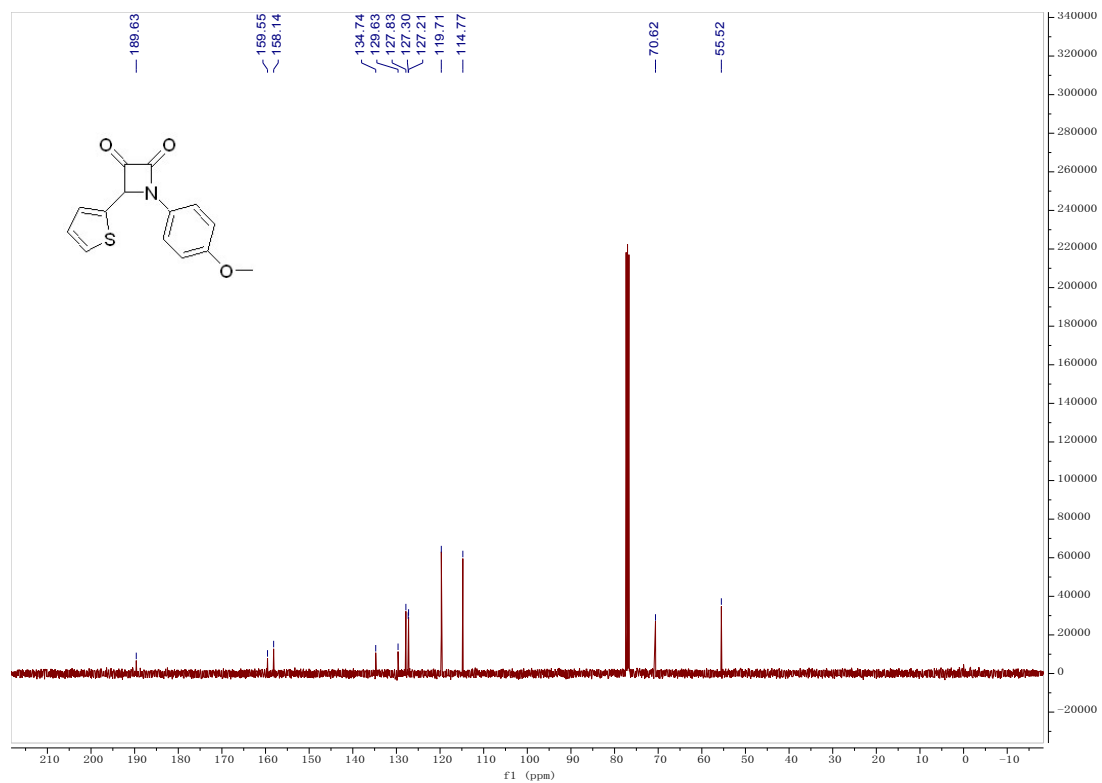
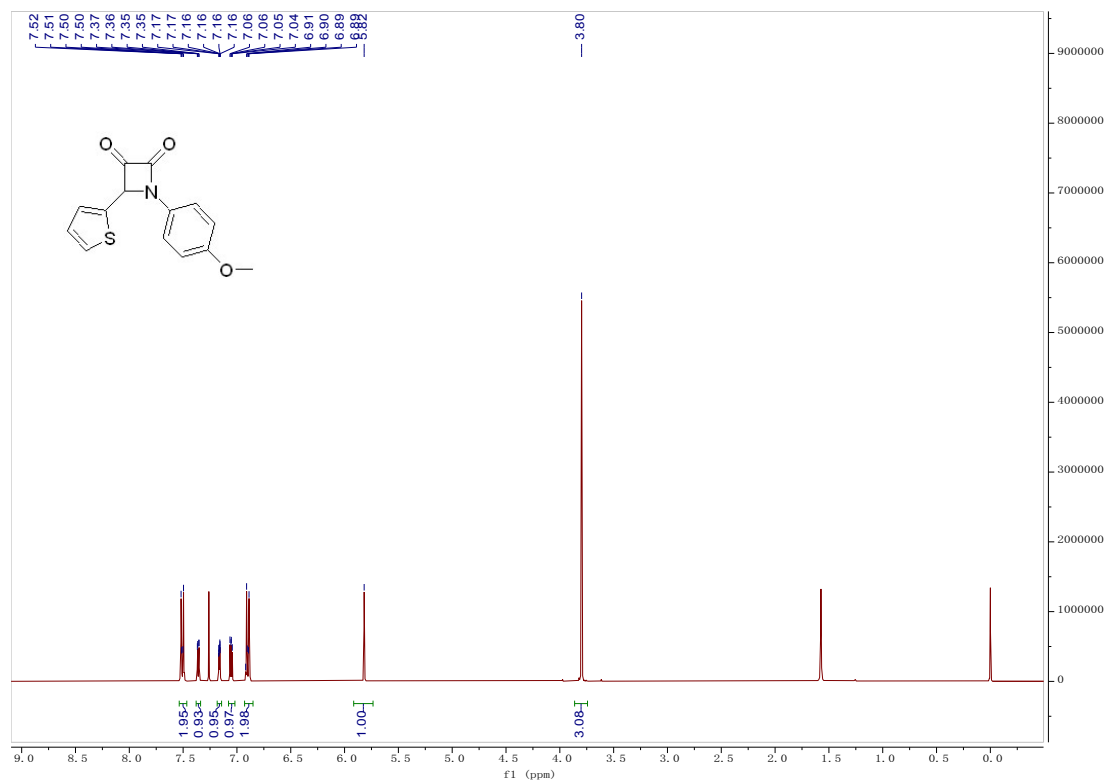


# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 1w

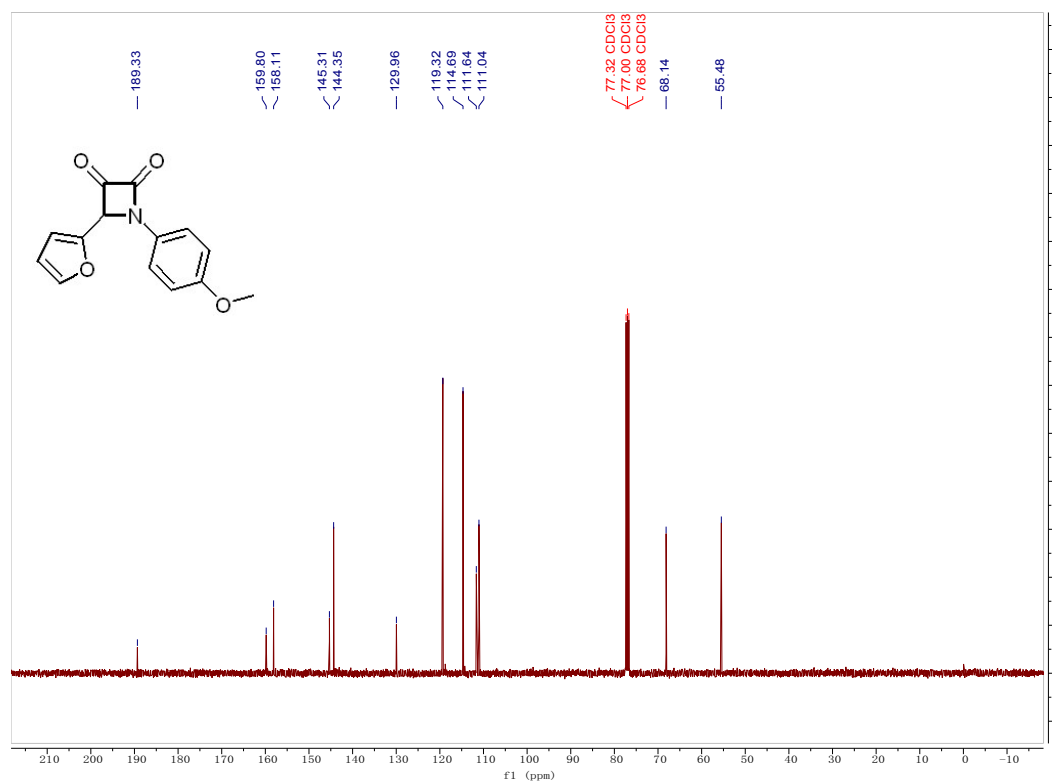
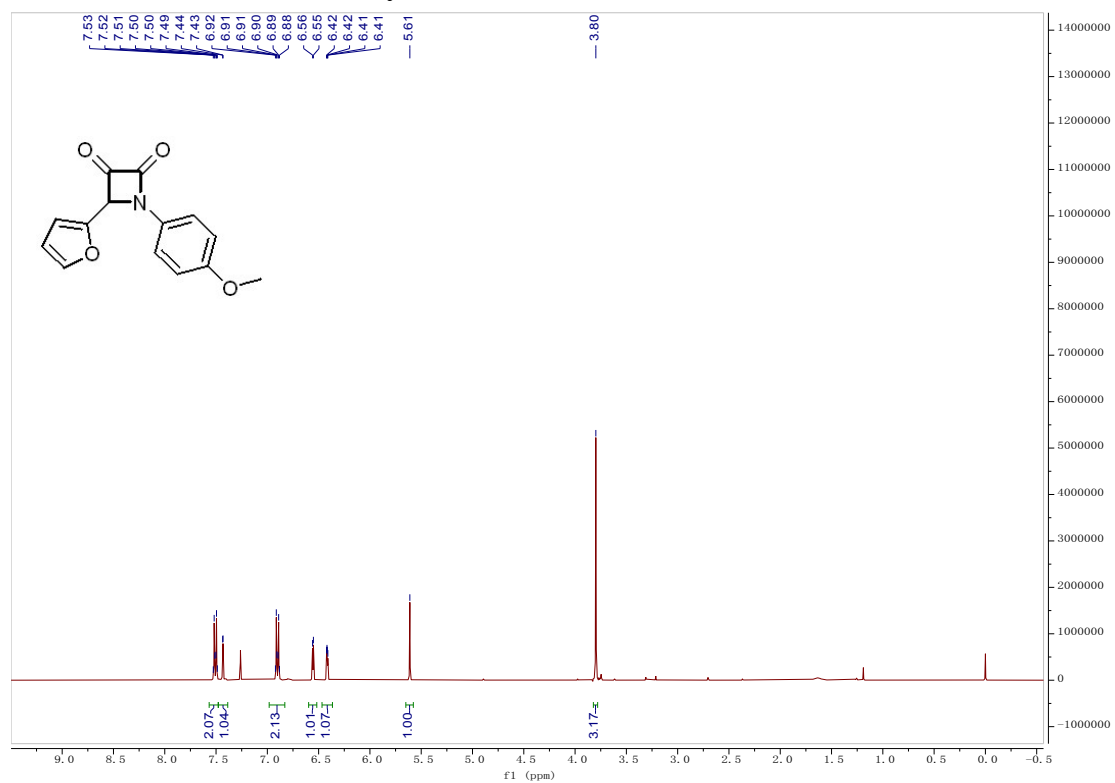




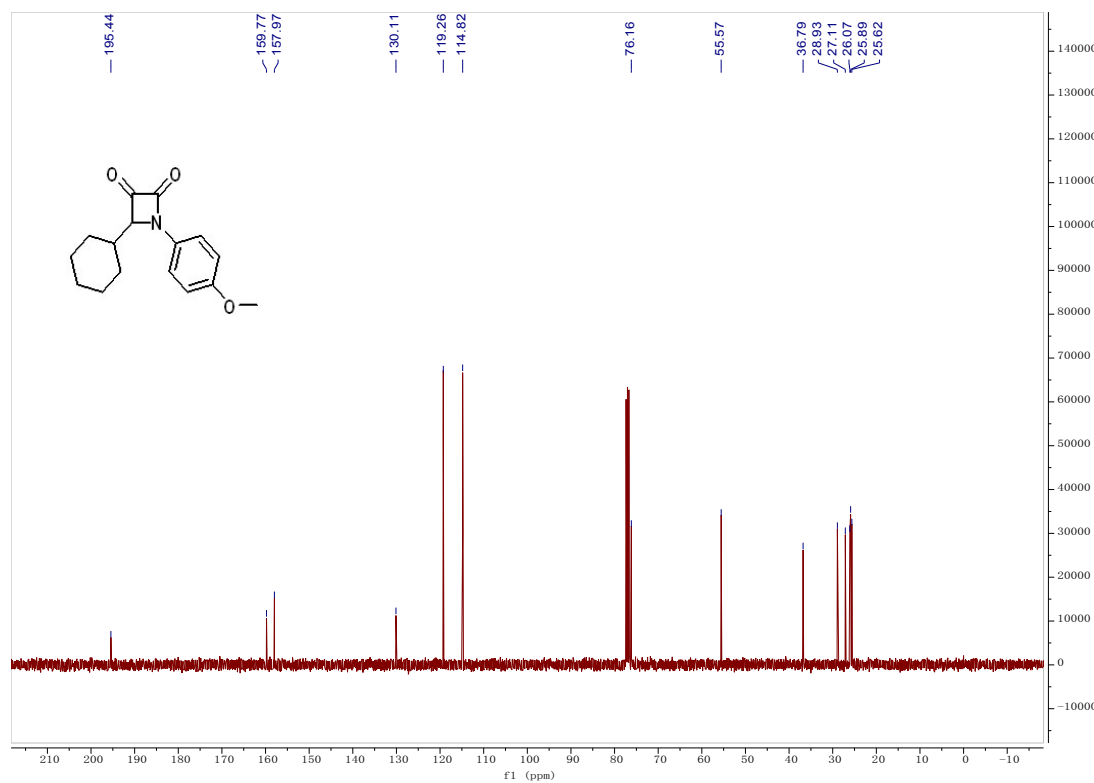
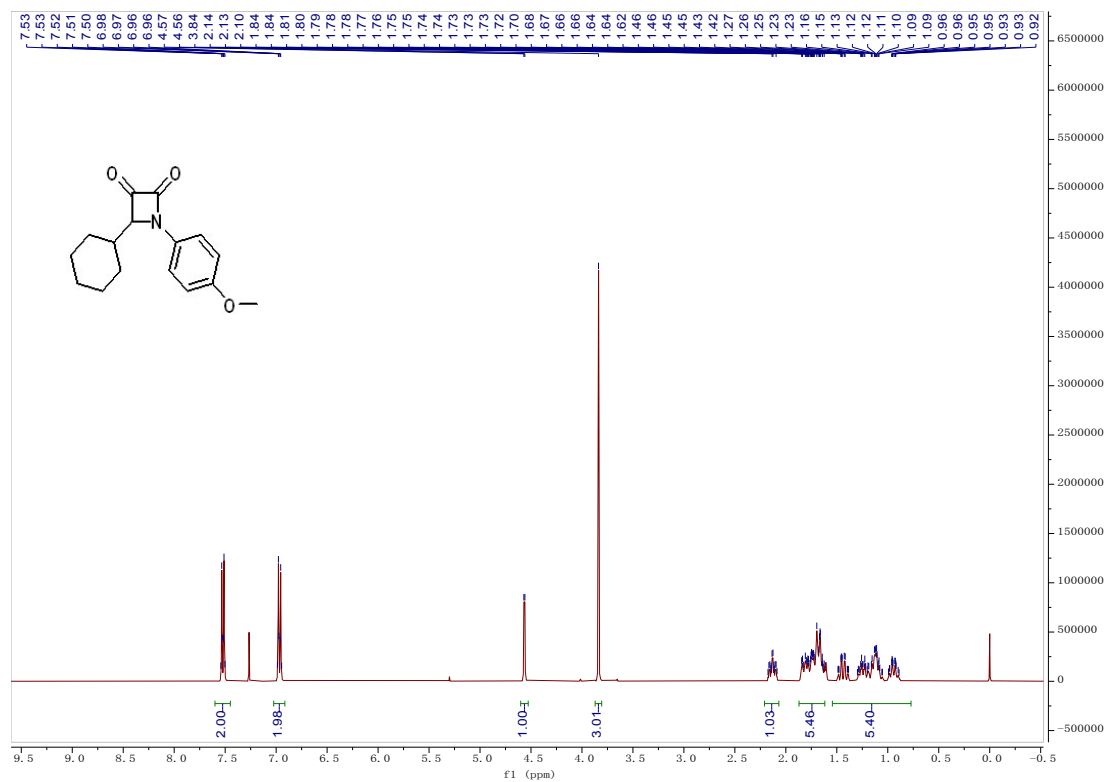
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 1x



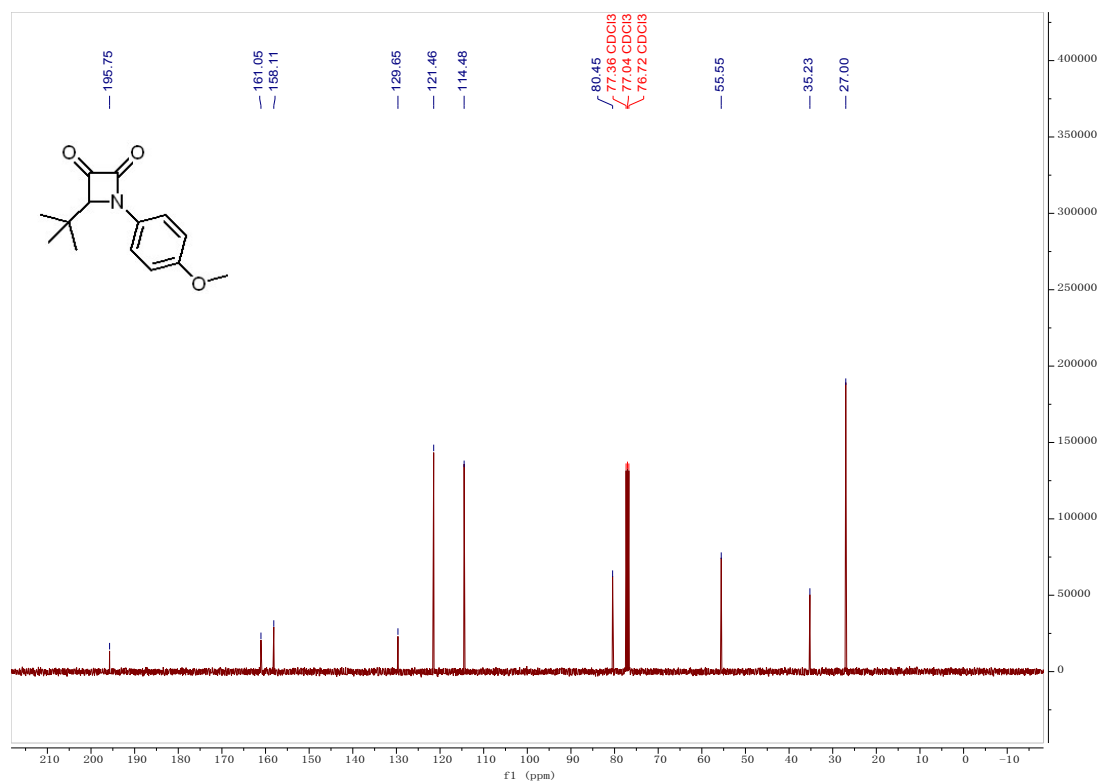
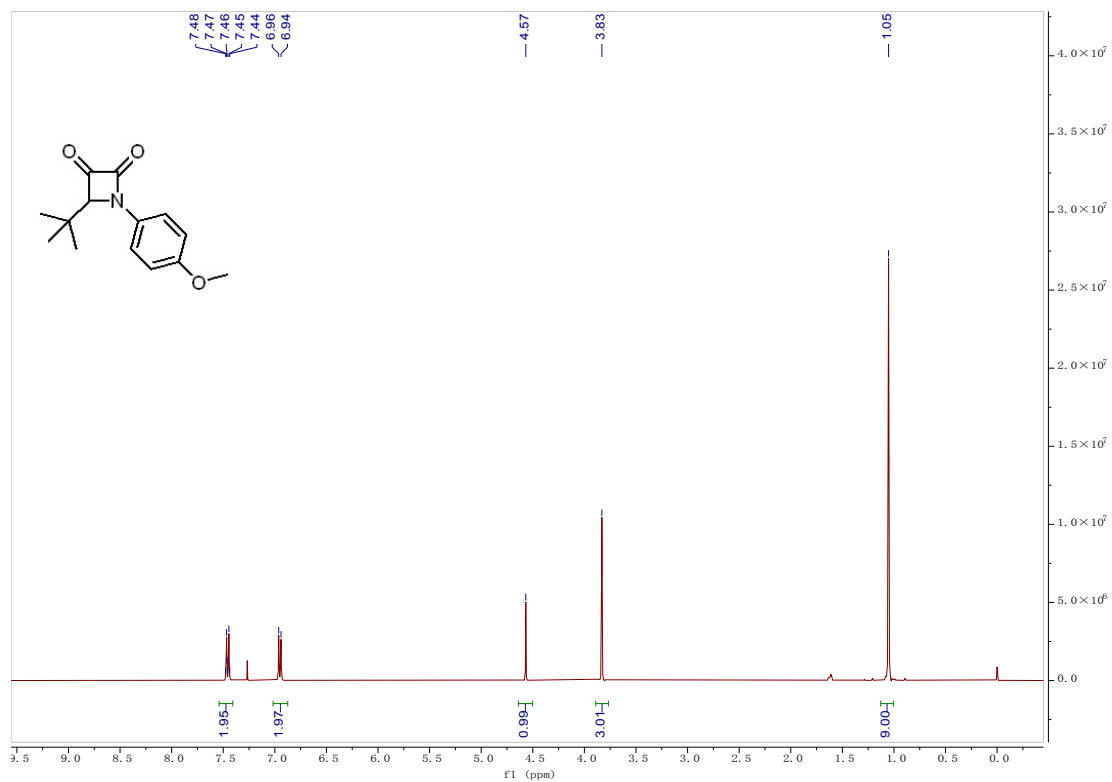
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1y



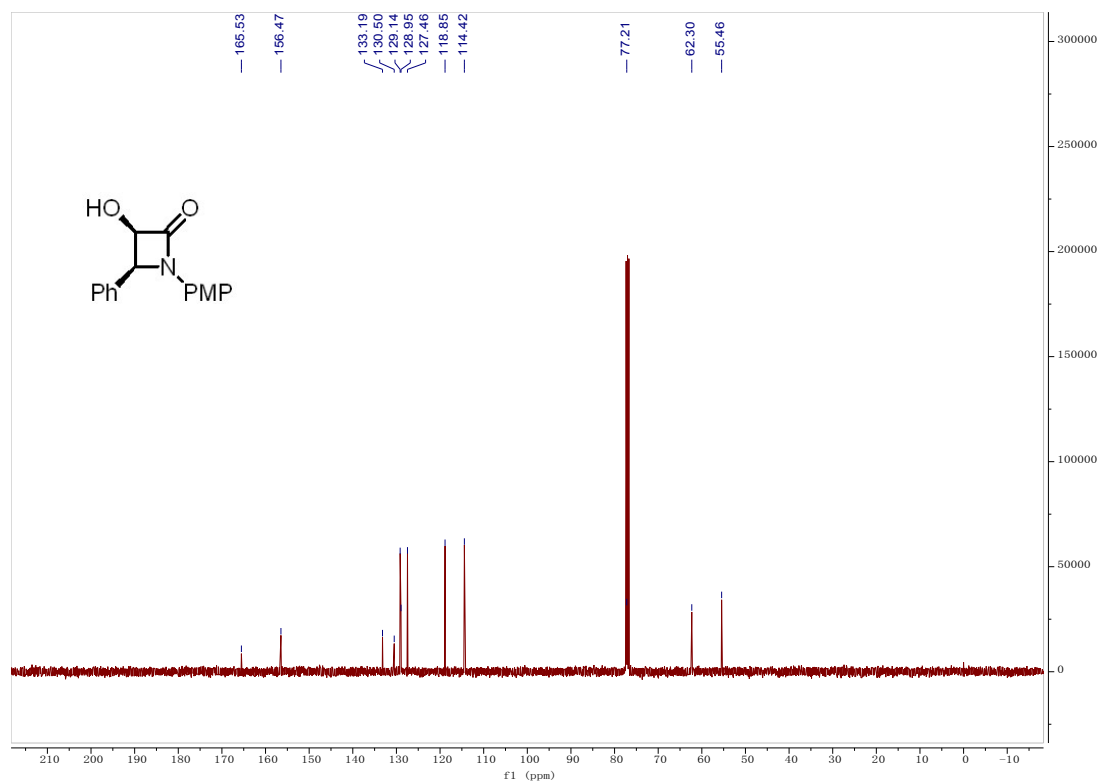
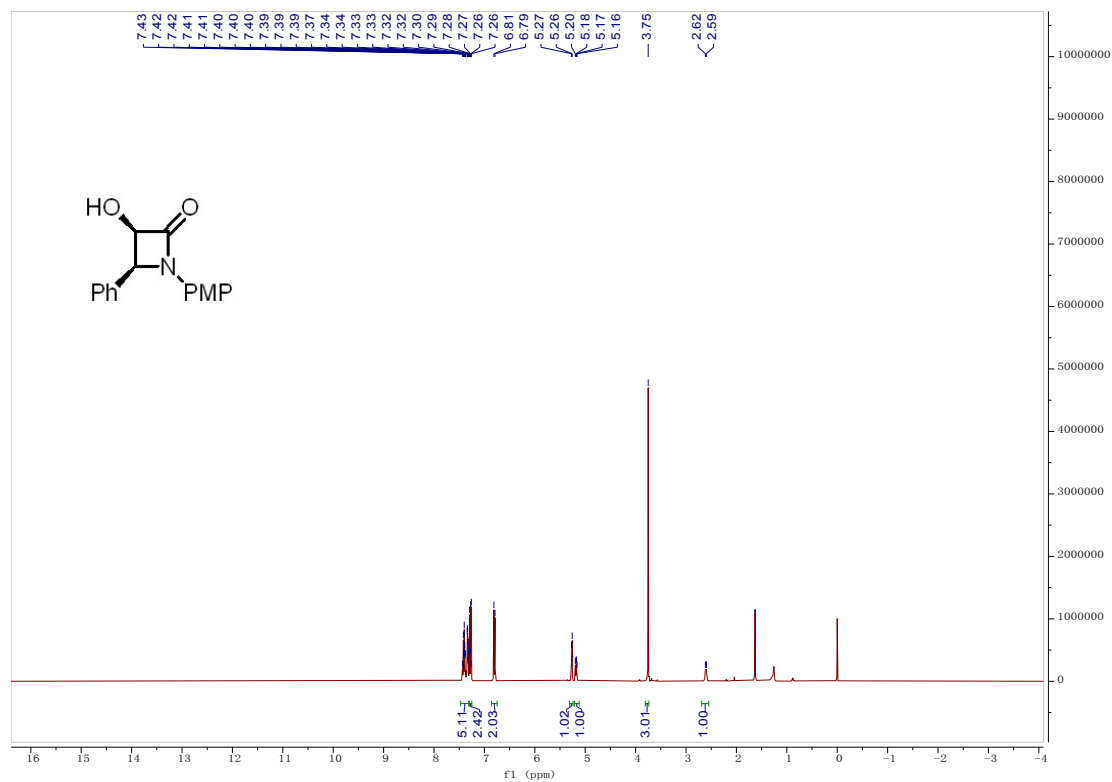
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 1z



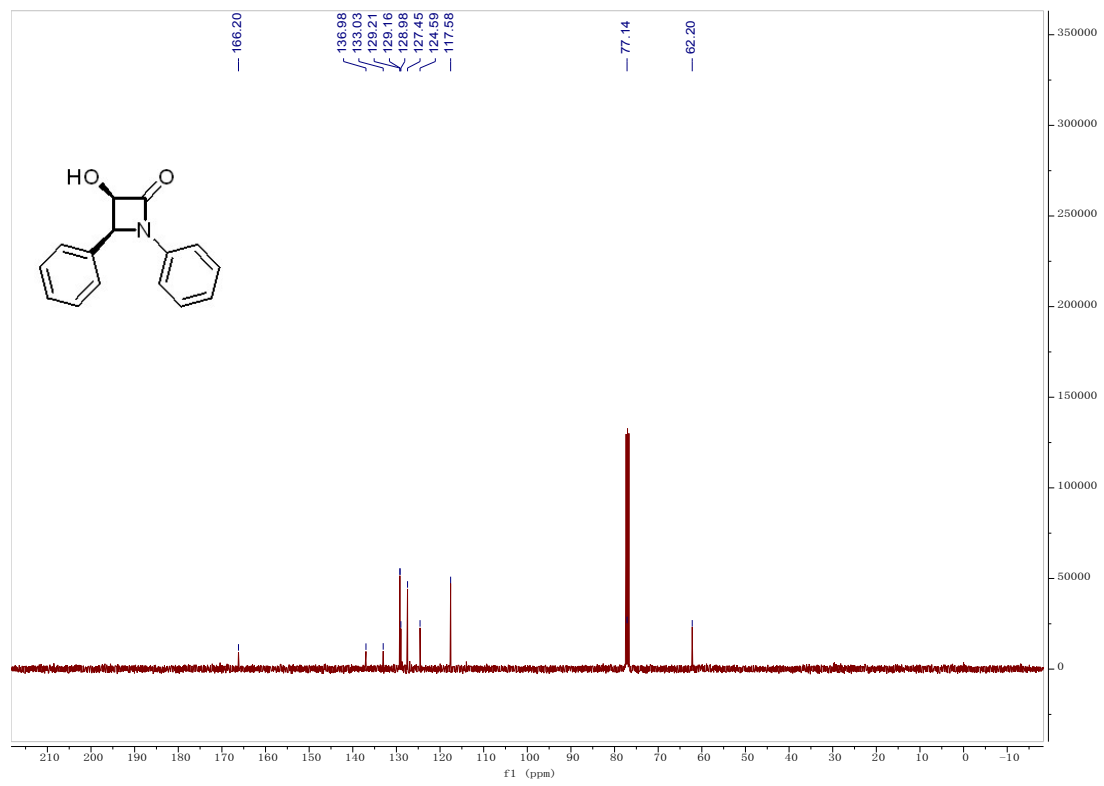
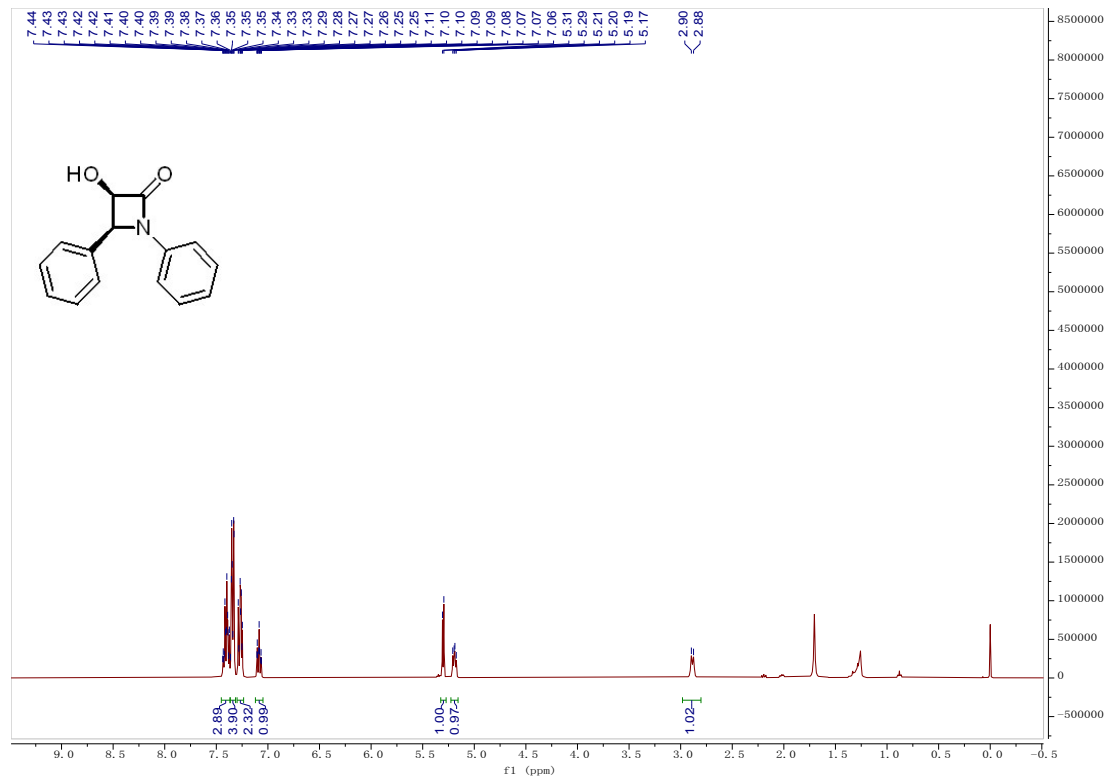
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 1aa



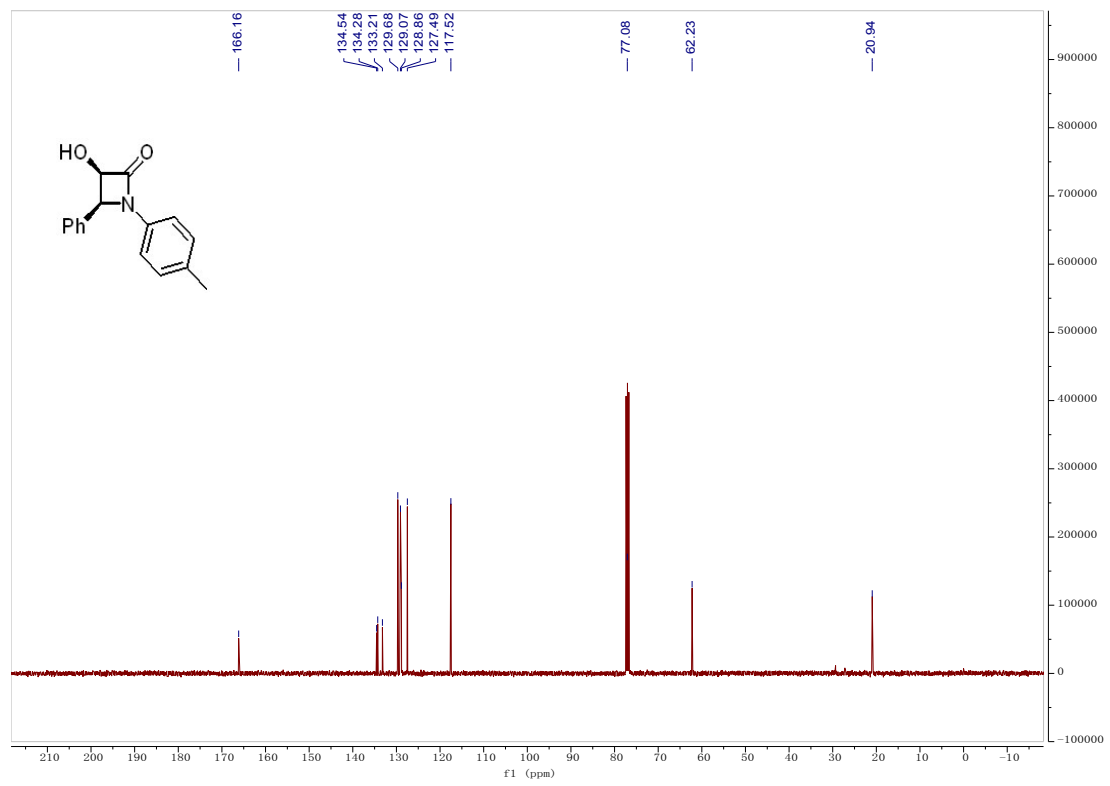
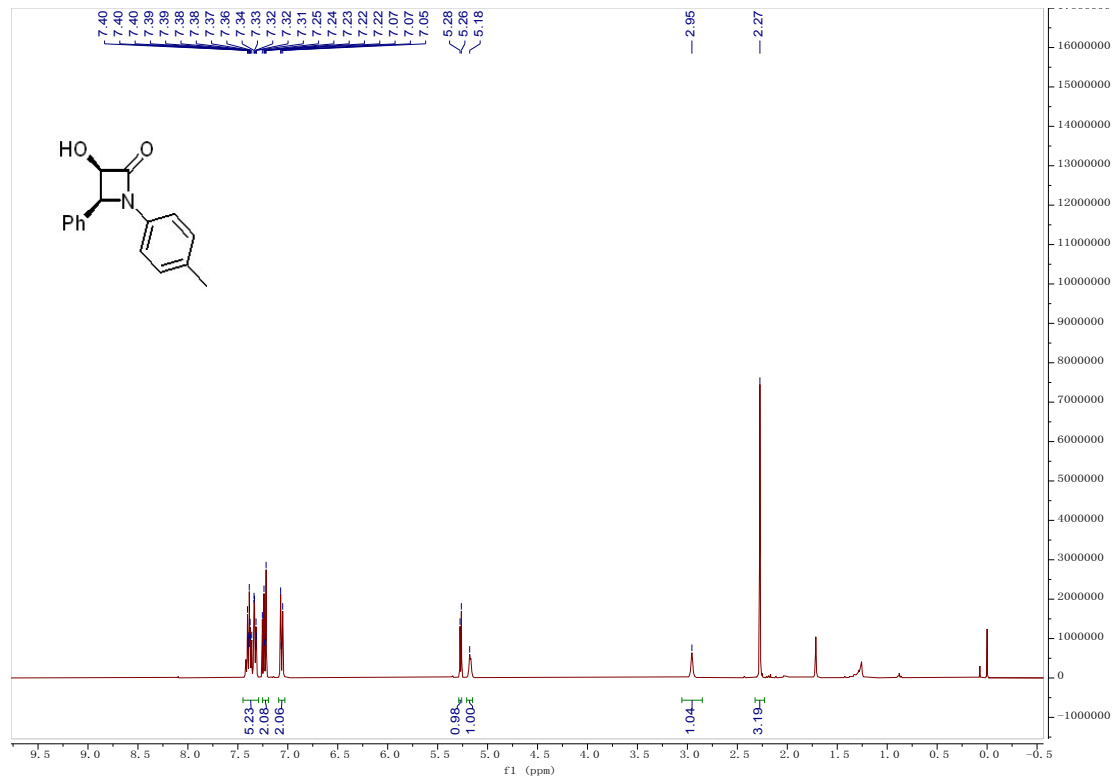
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2a



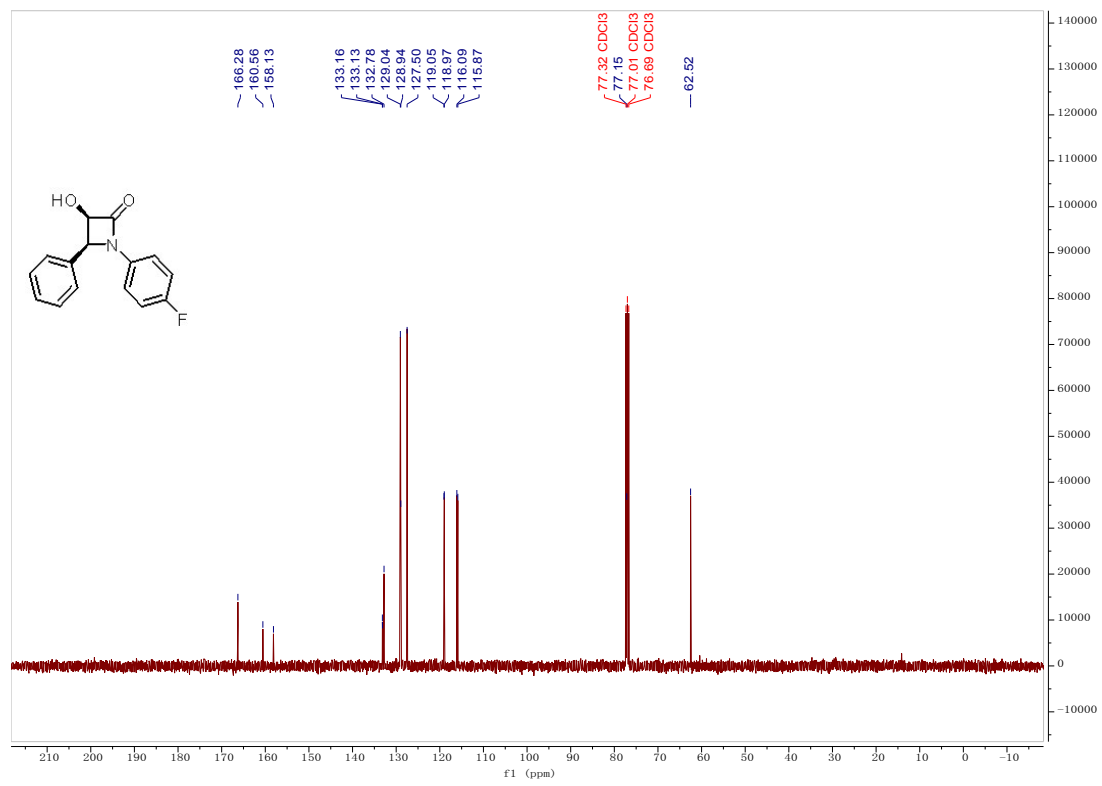
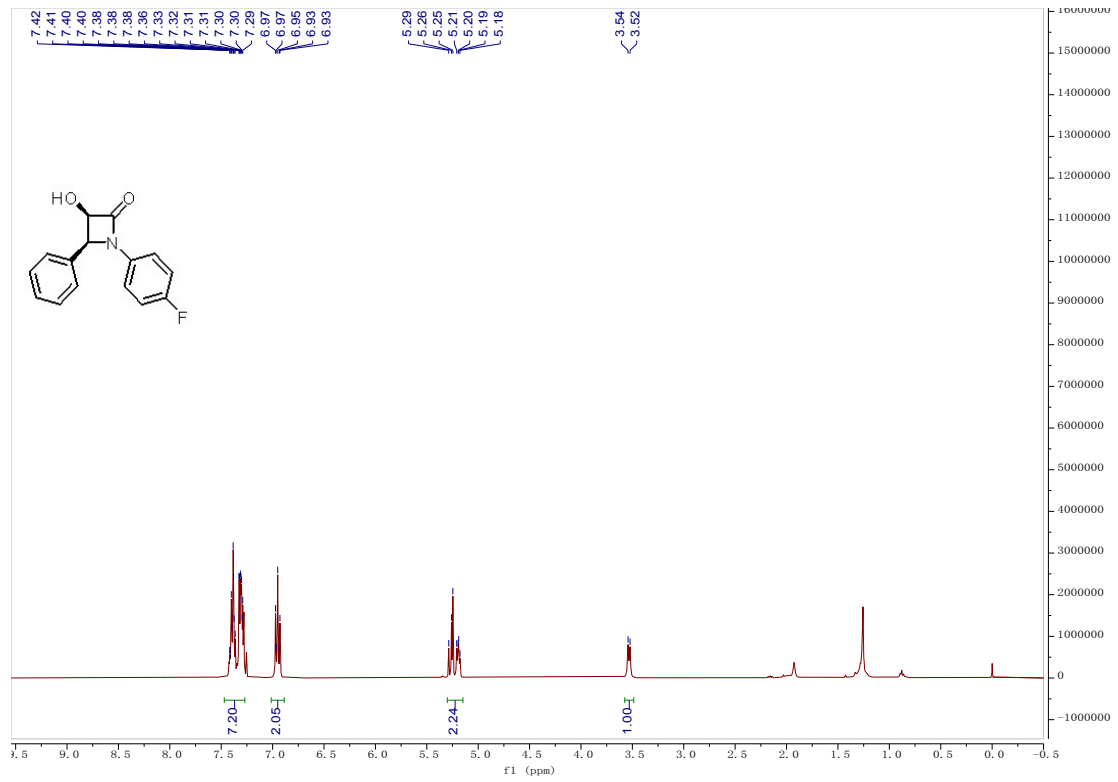
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2b



# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2c

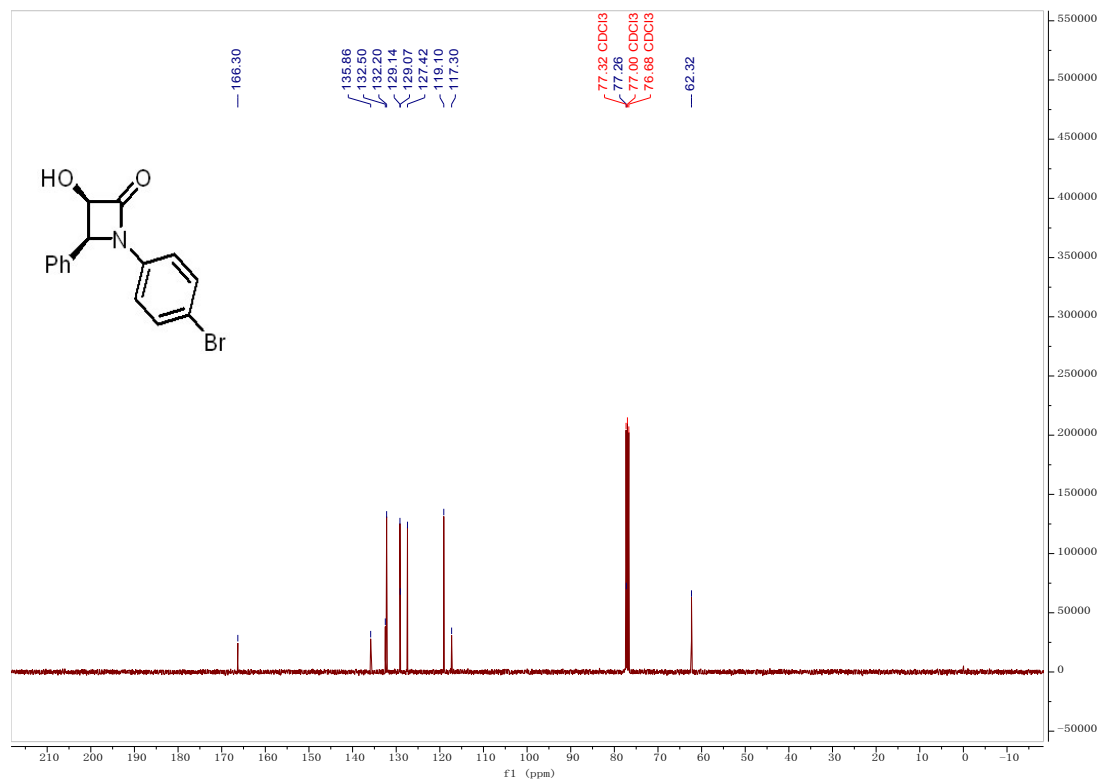
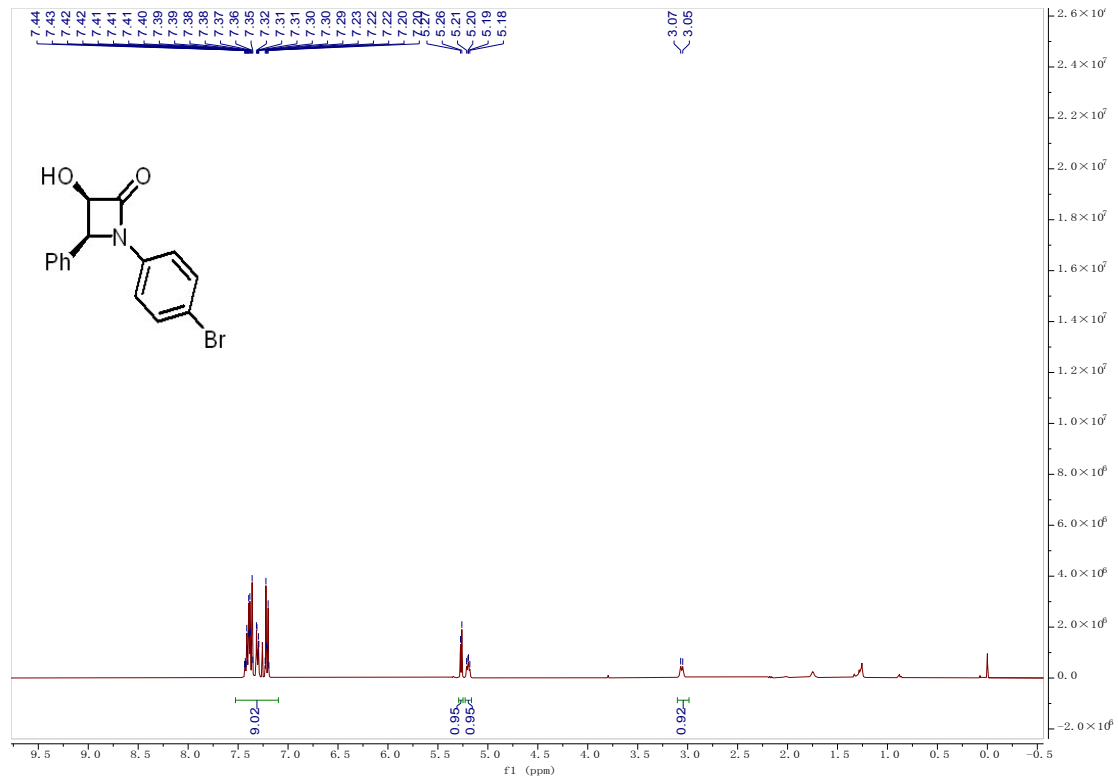


# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2d

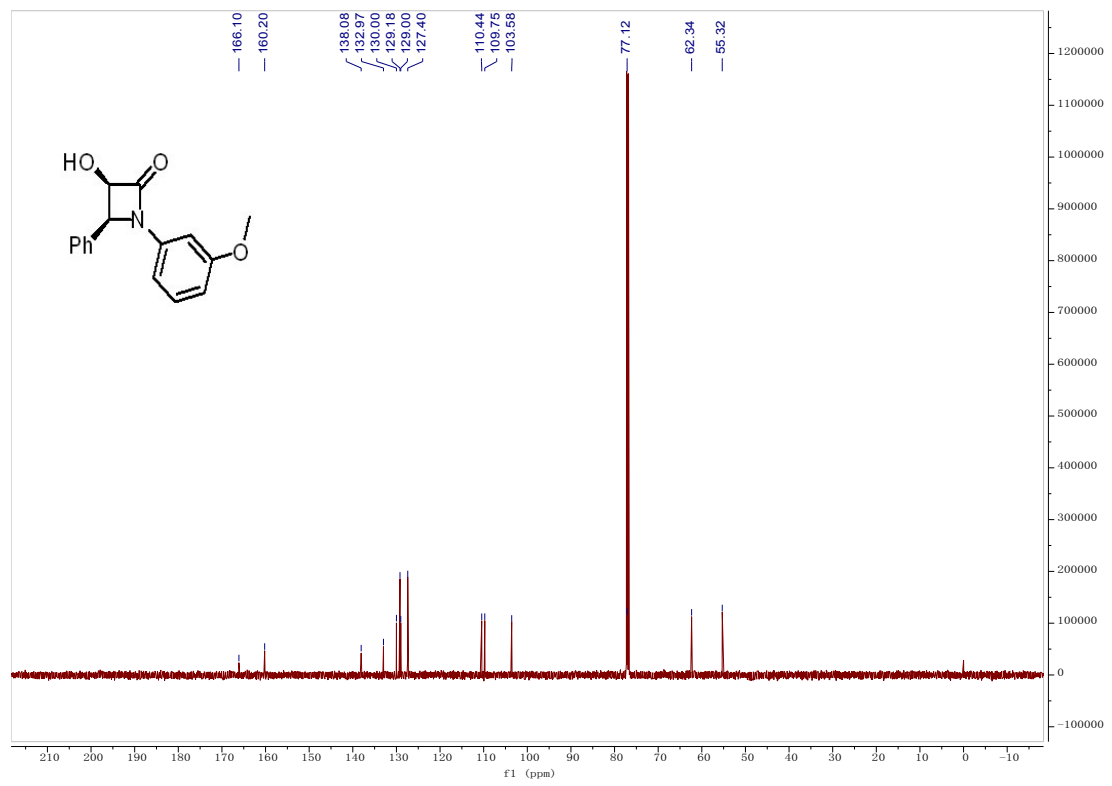
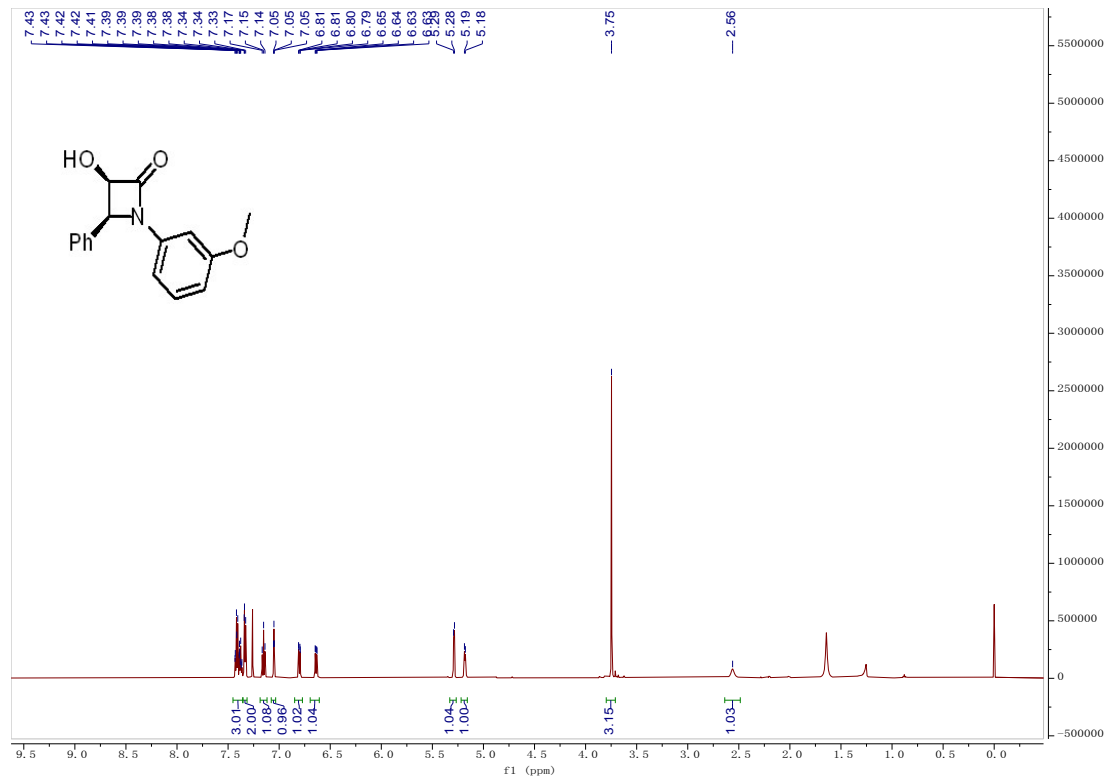




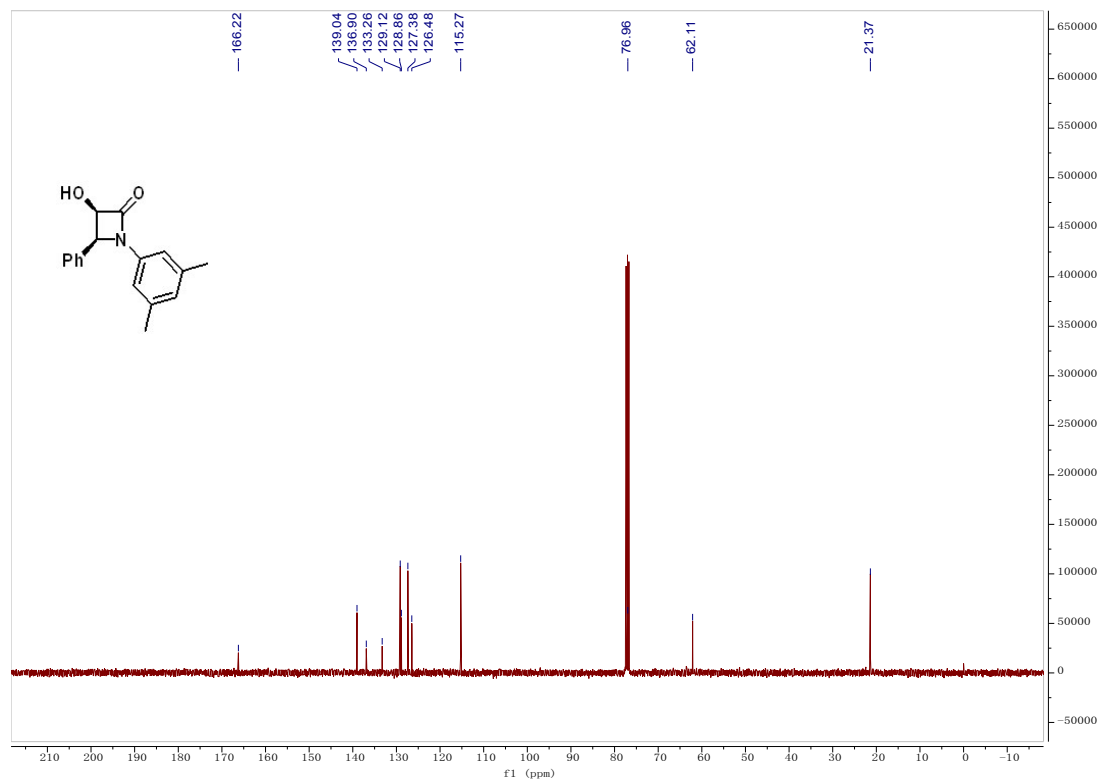
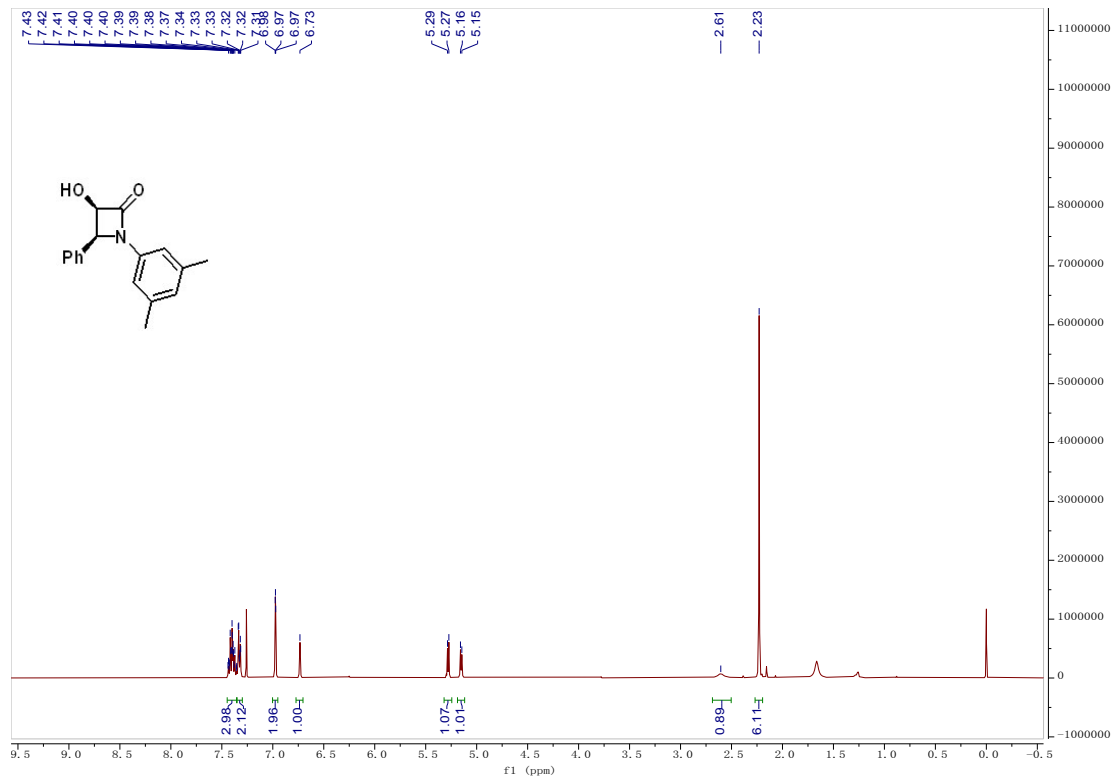
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2e



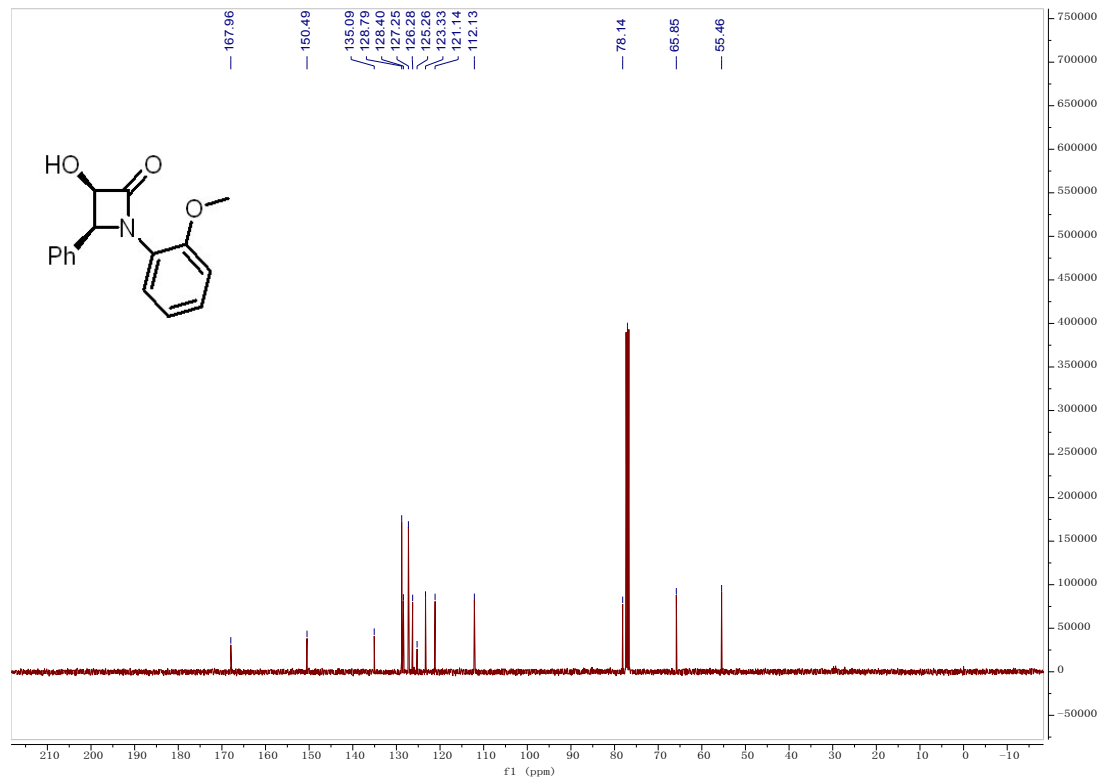
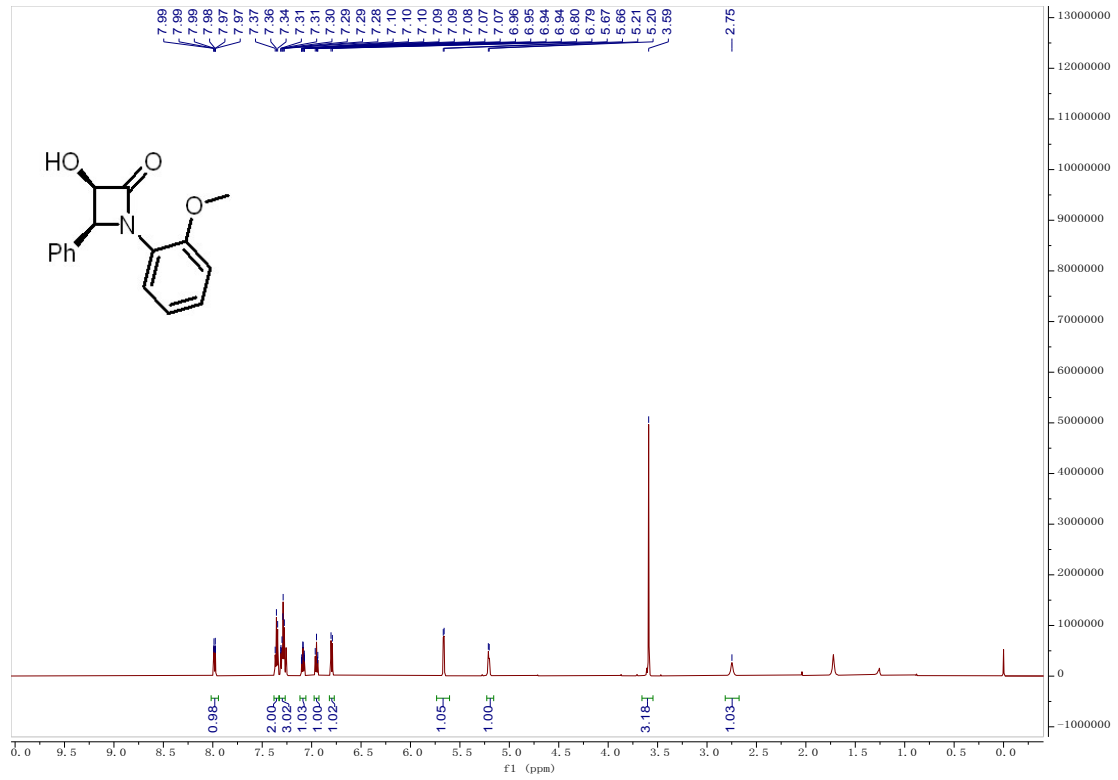
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2f



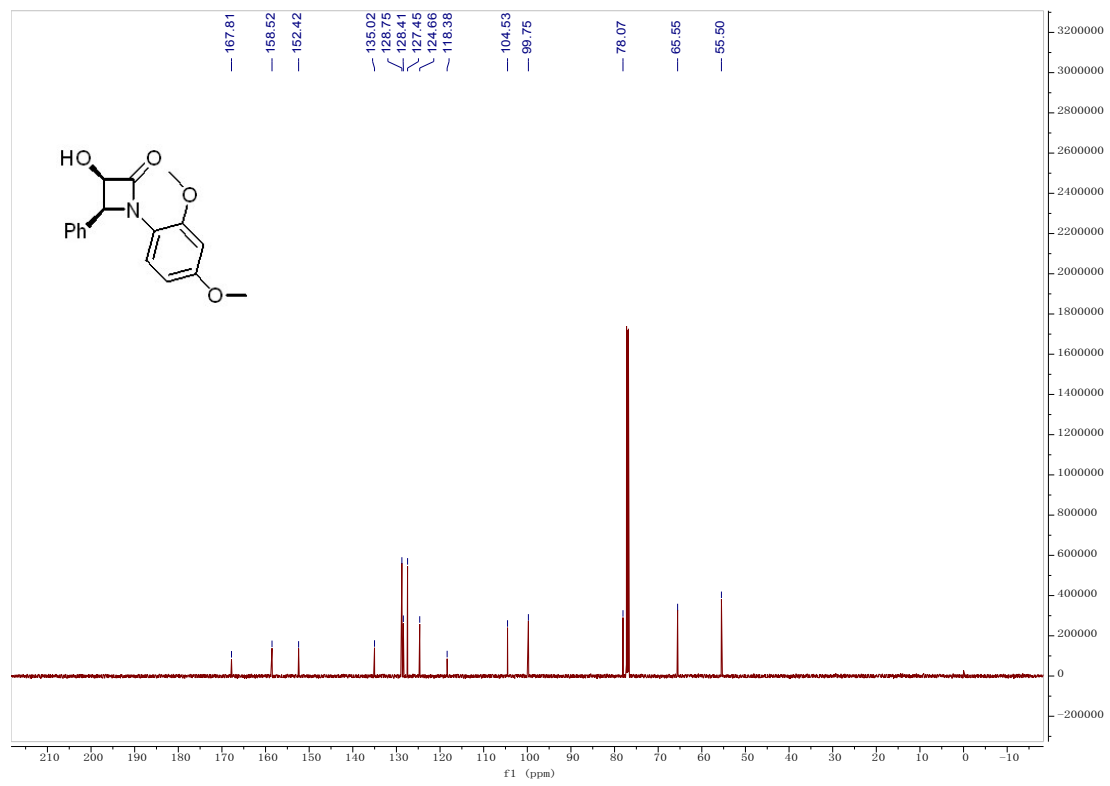
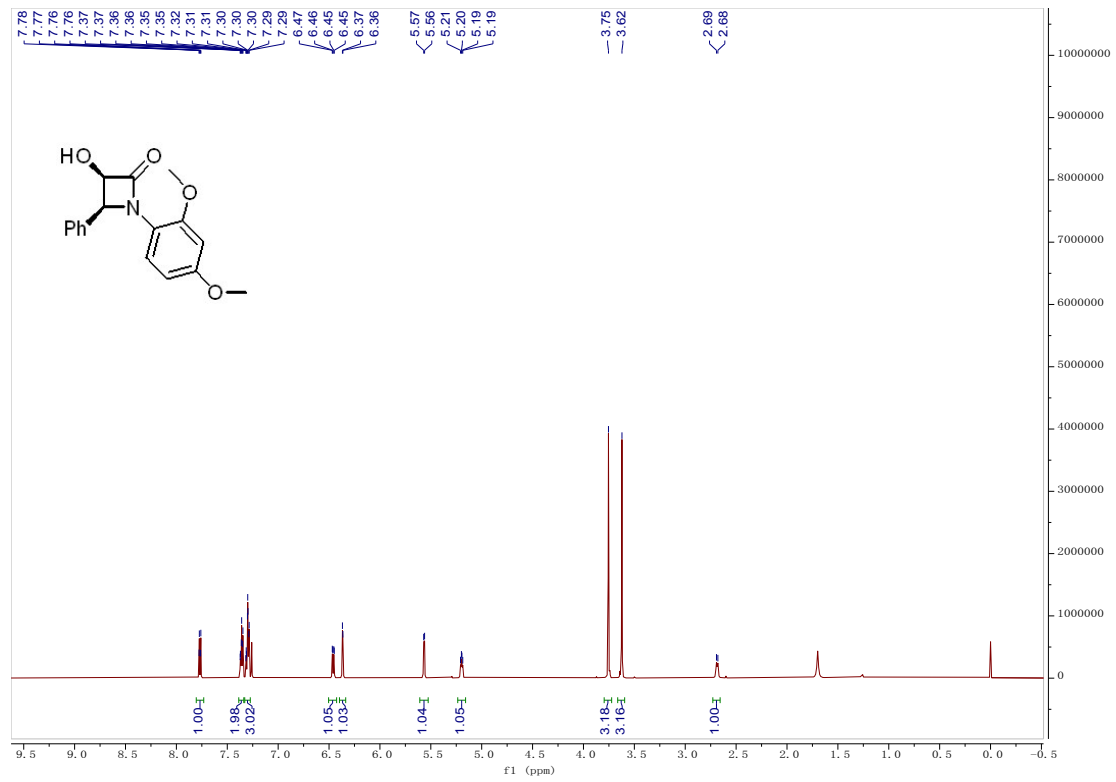
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2g



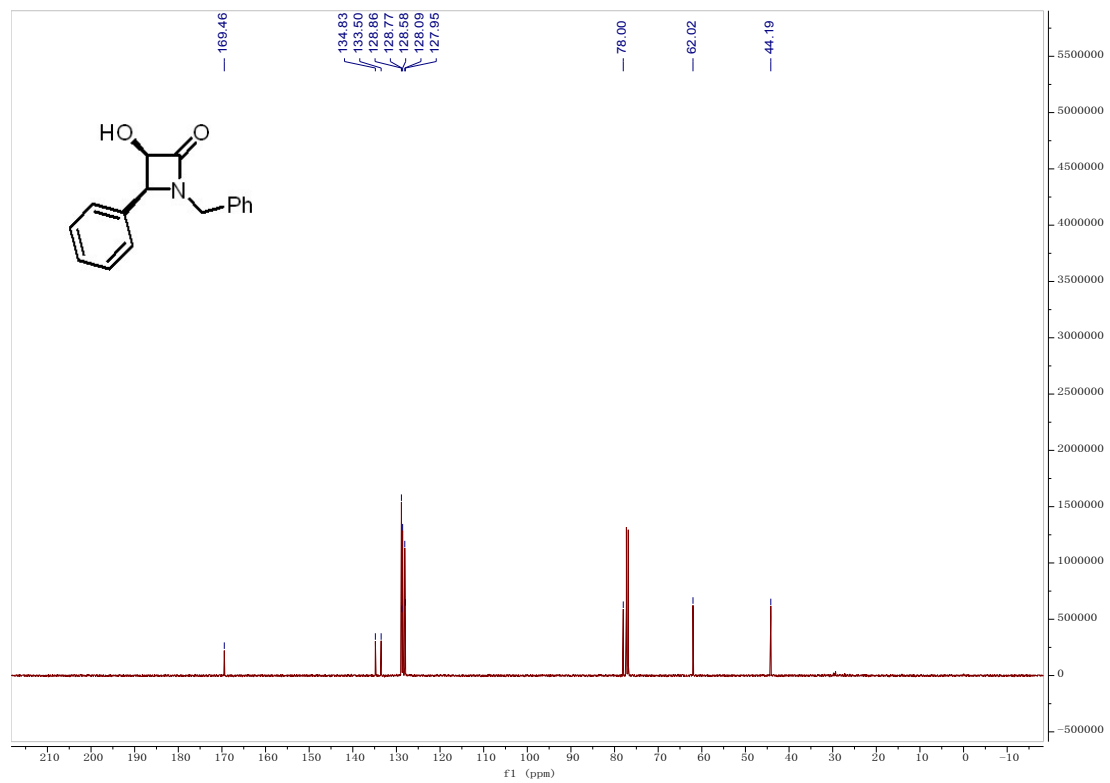
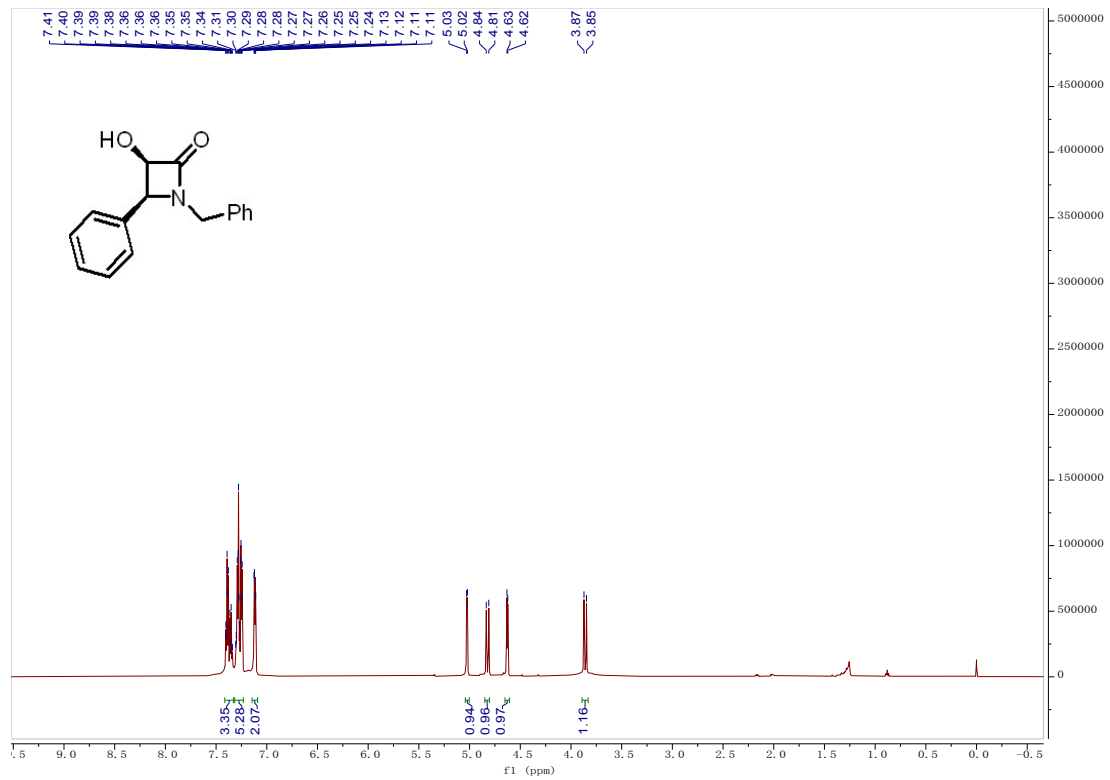
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2h



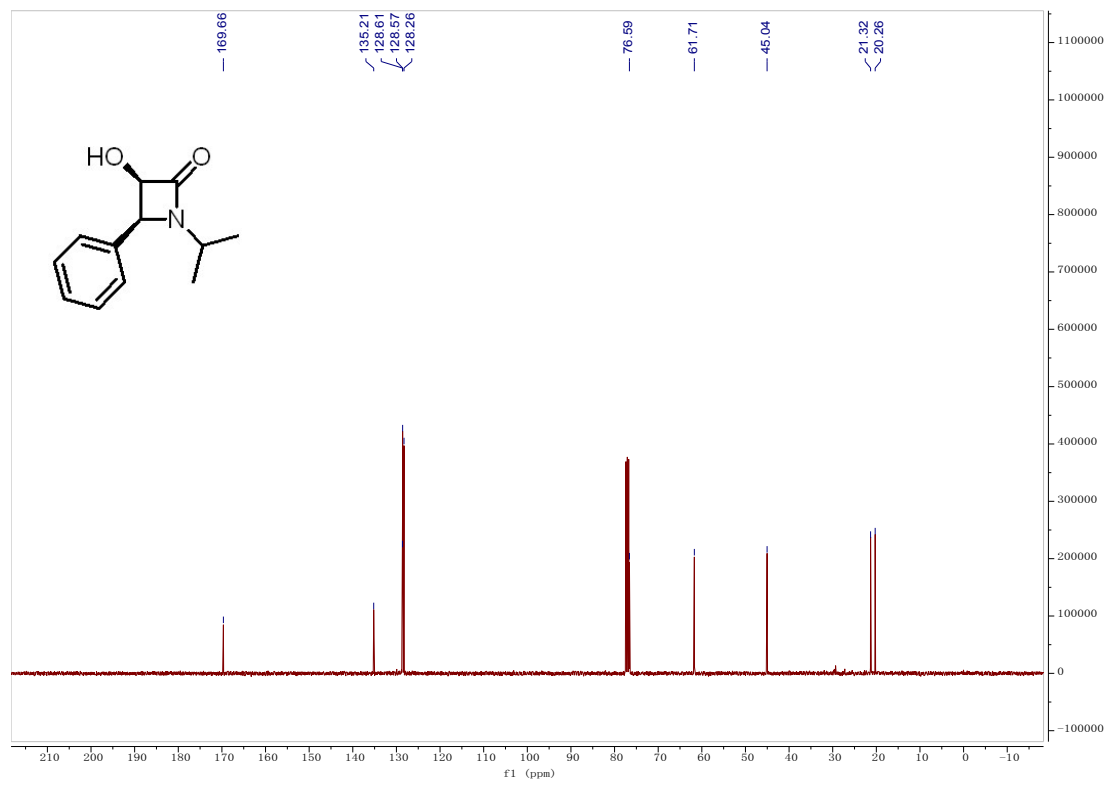
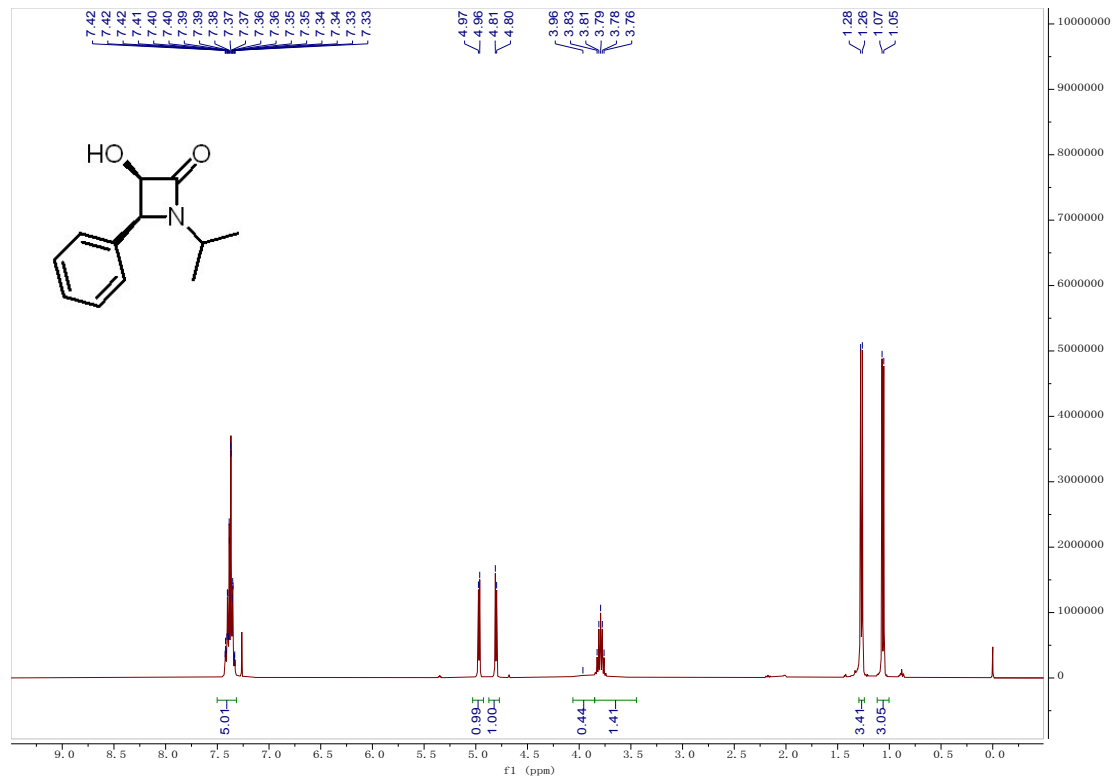
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2i



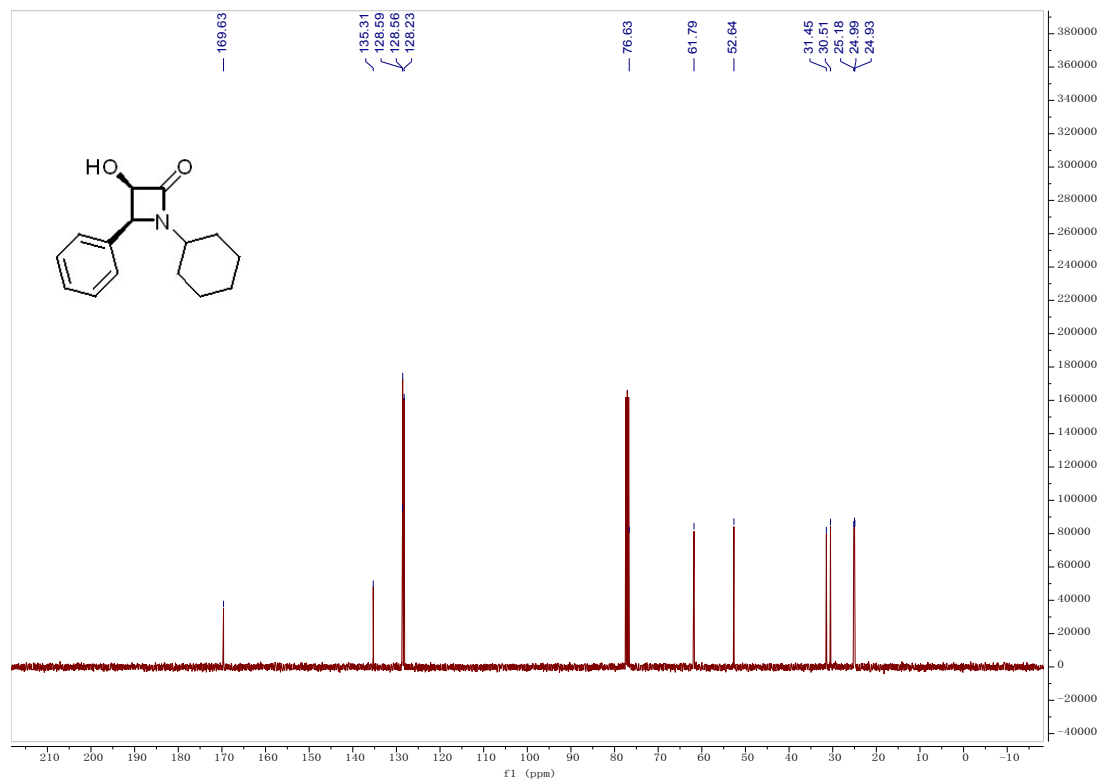
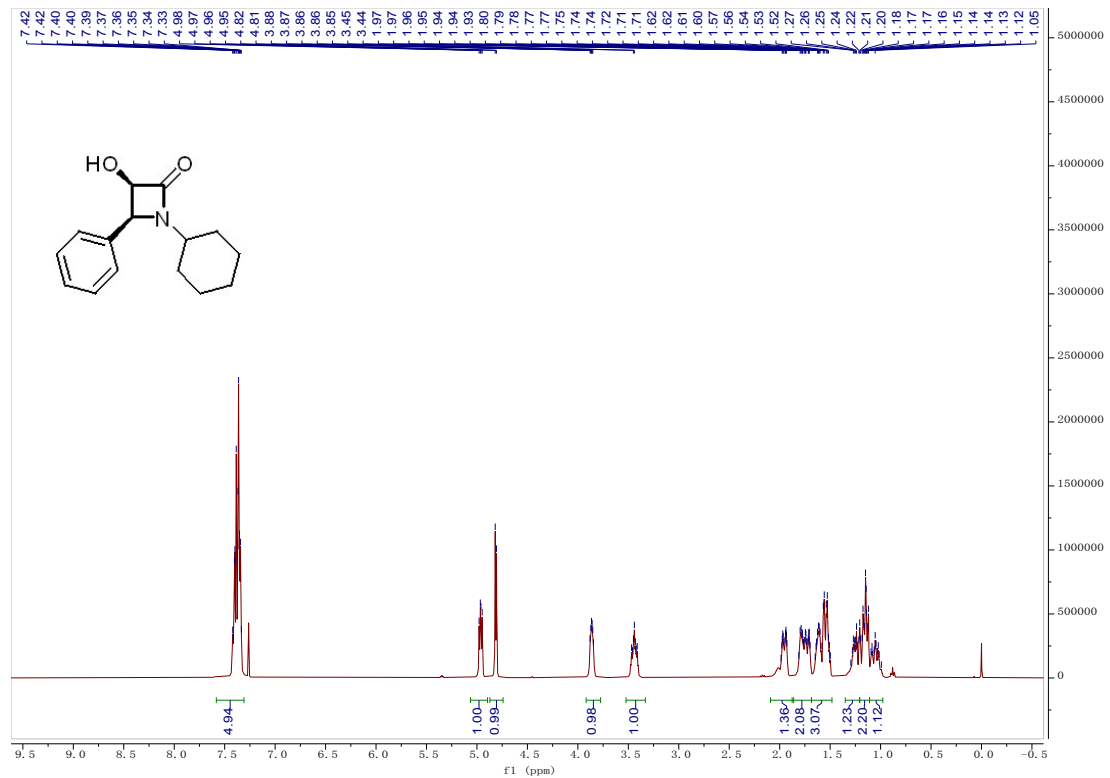
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2j



# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2k

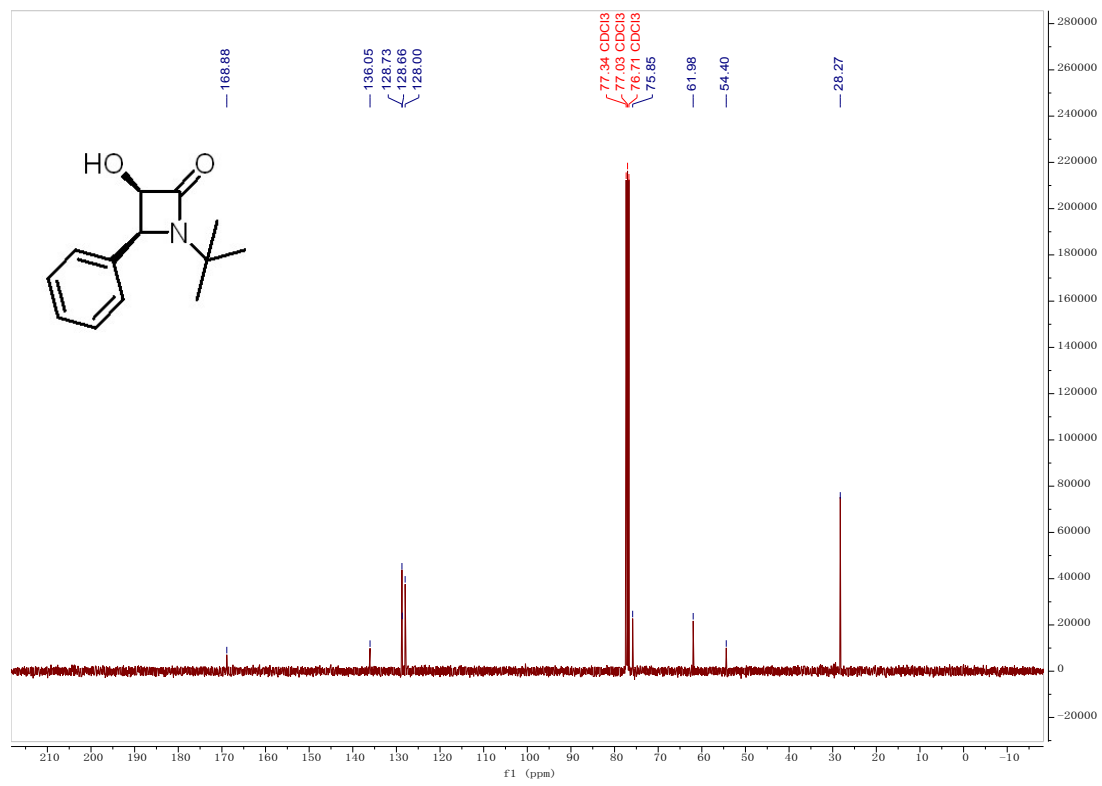
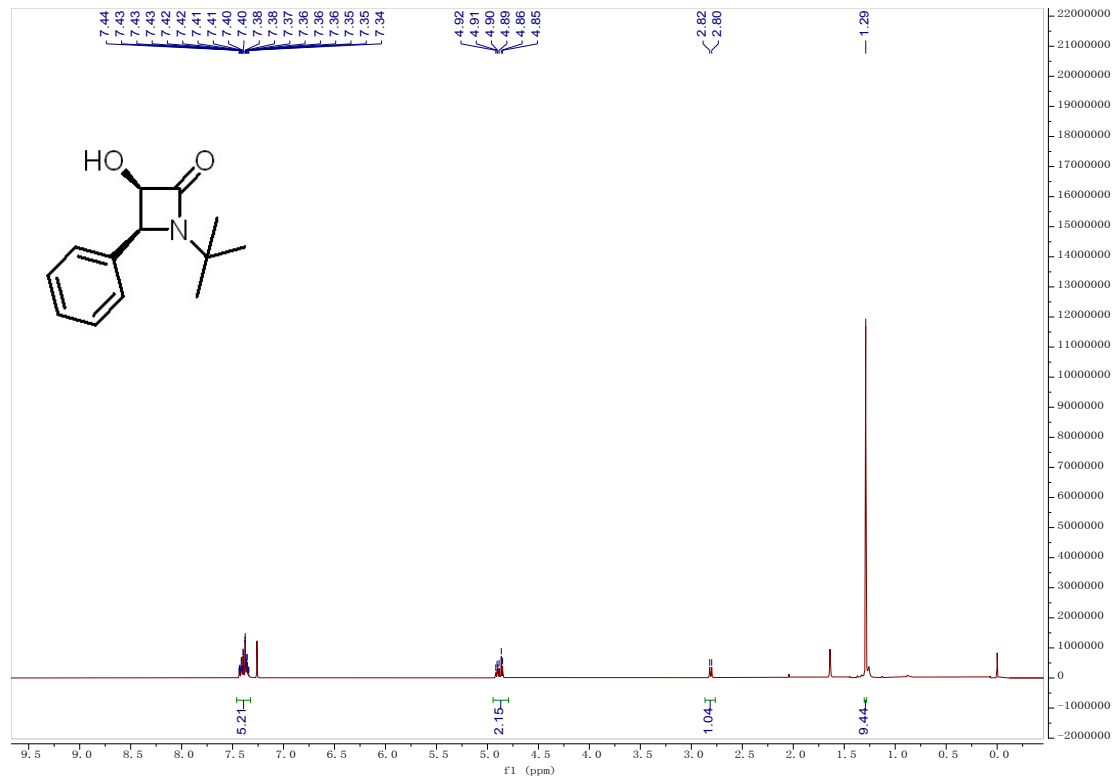


# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2l

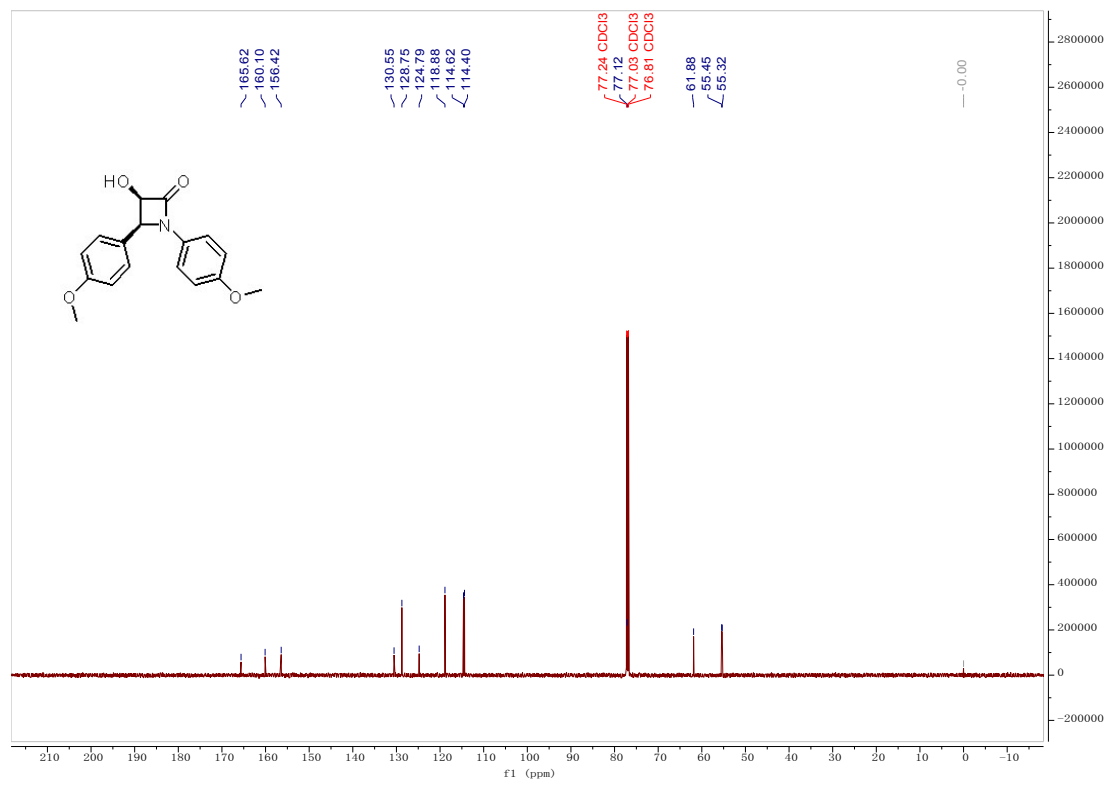
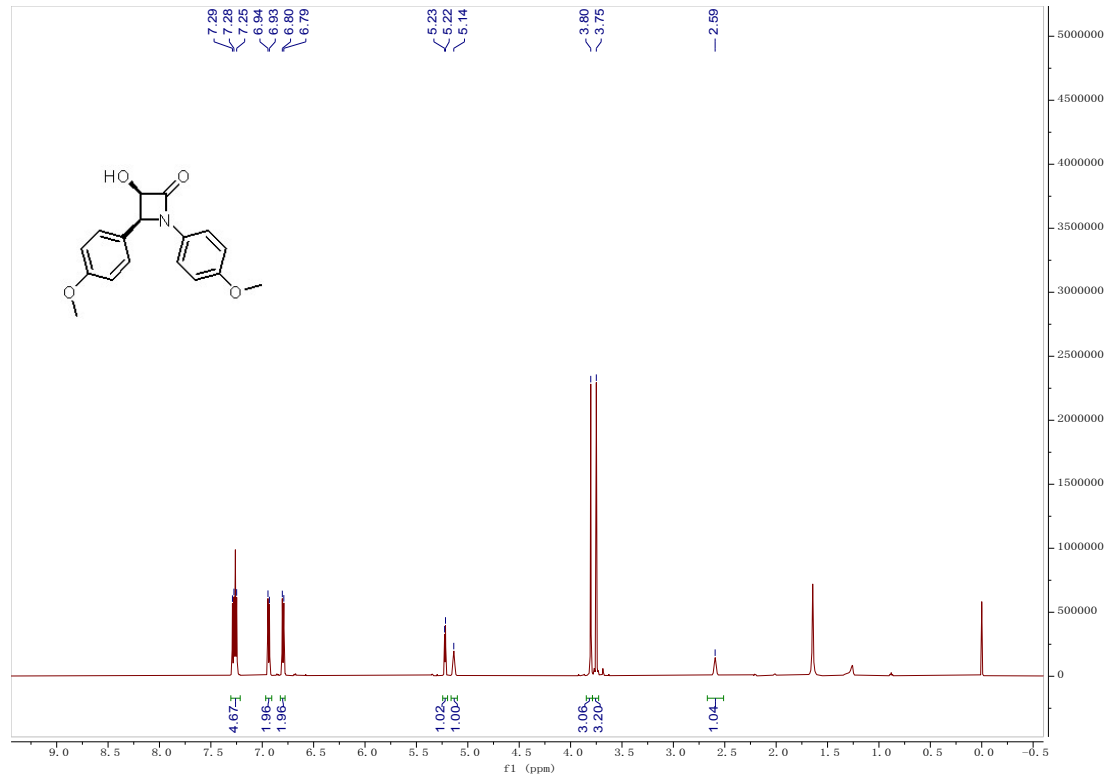




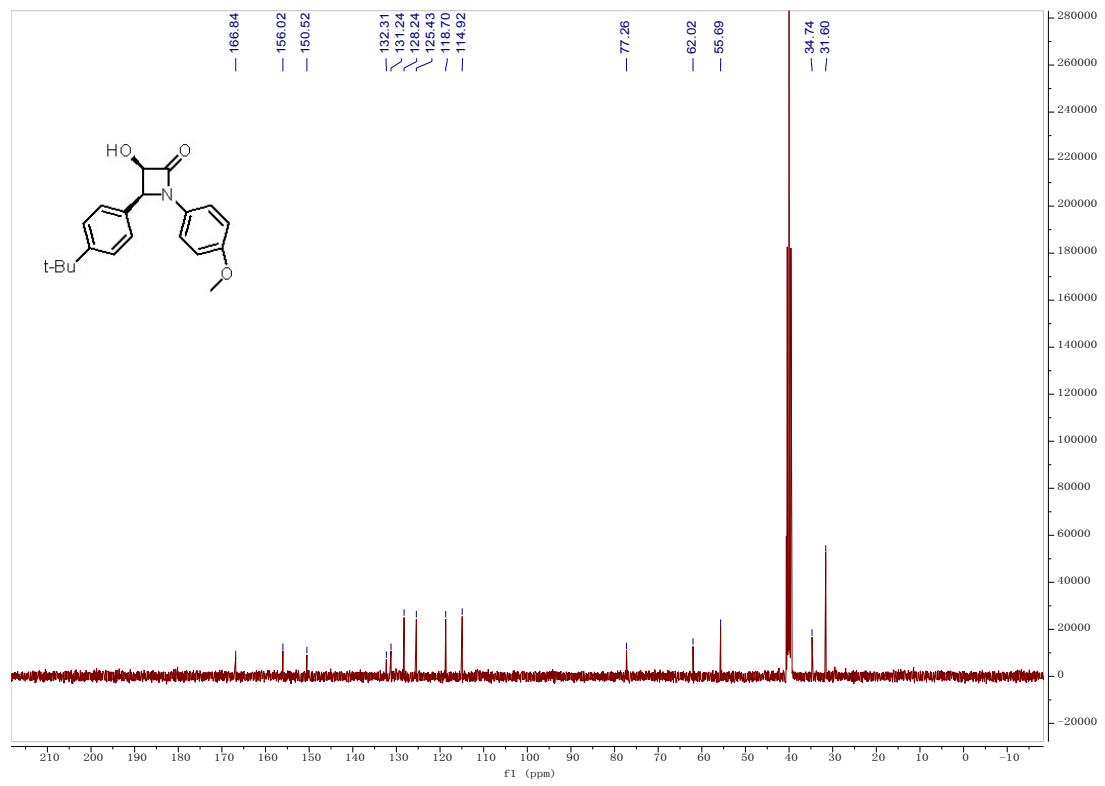
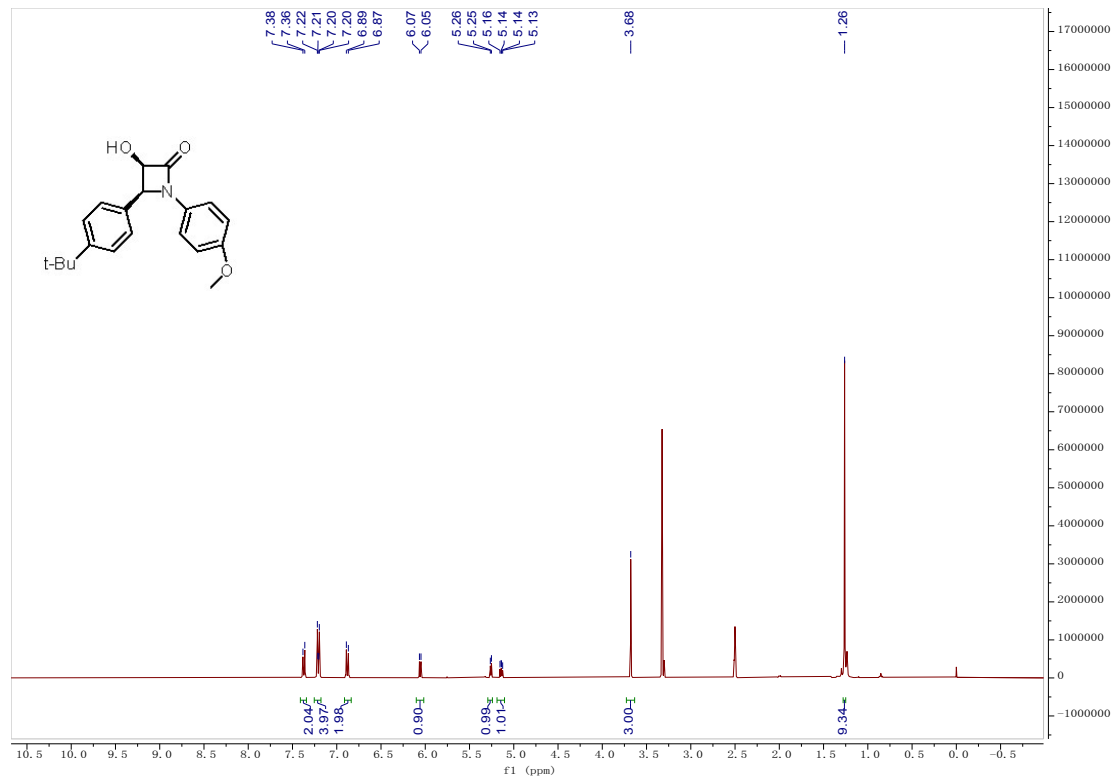
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2m



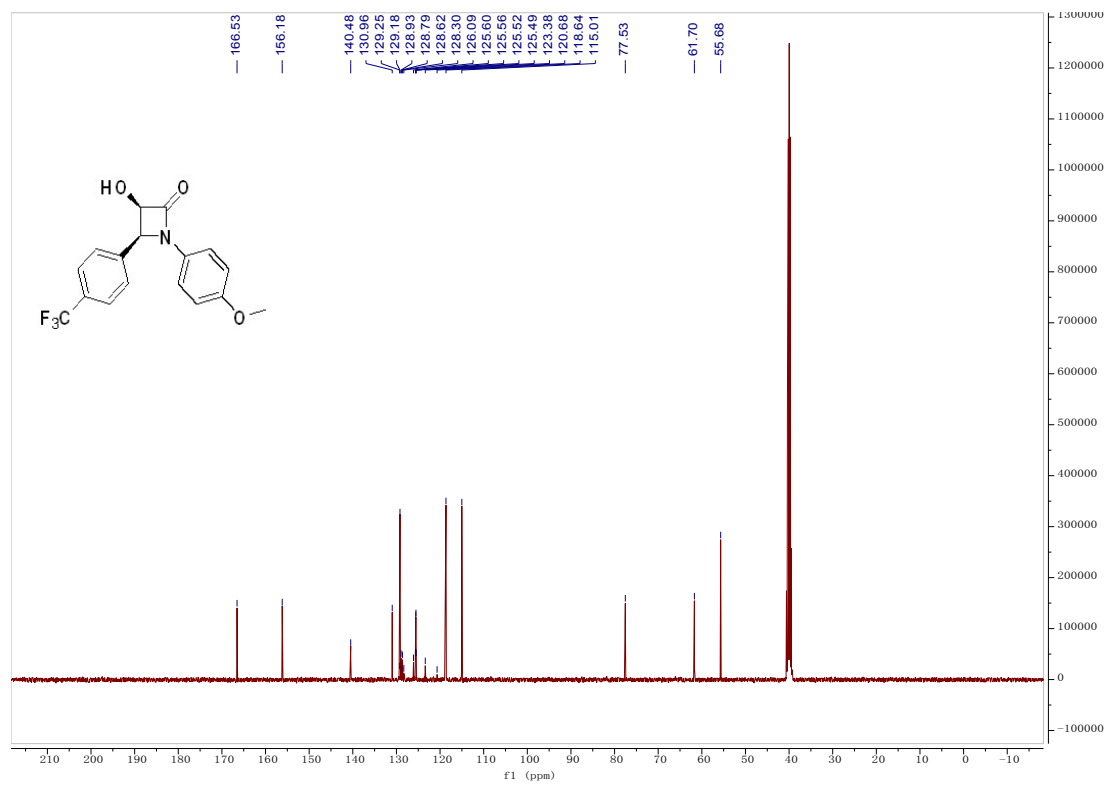
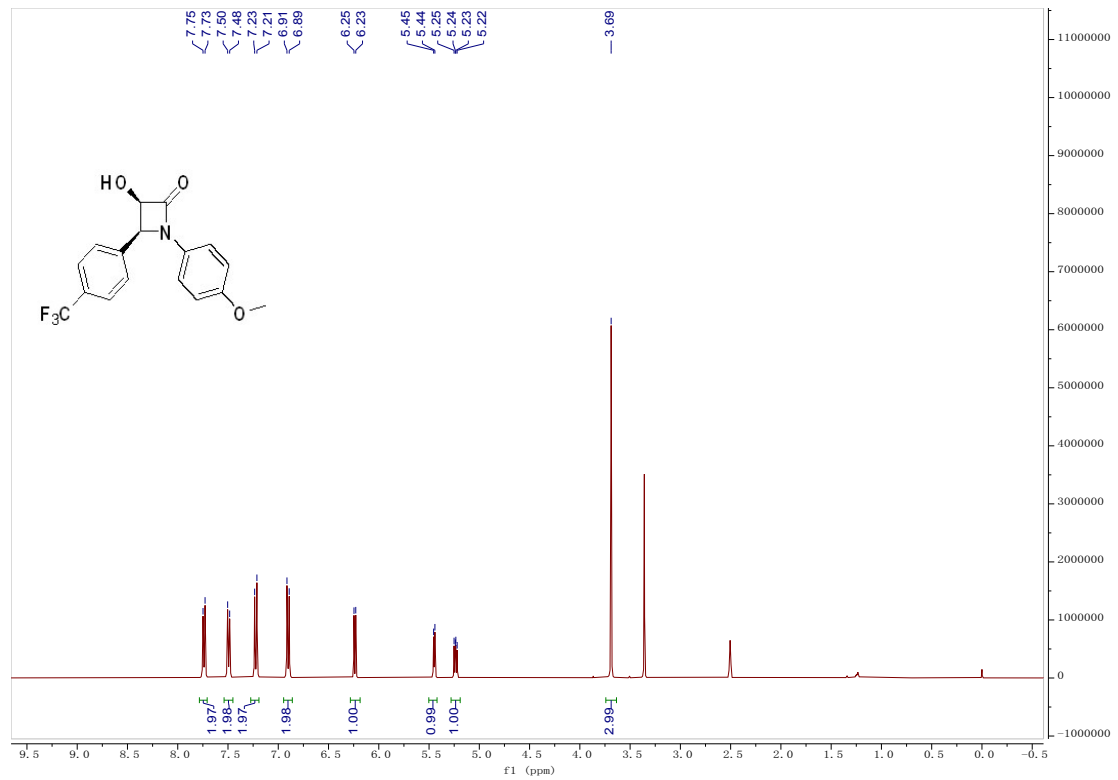
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2n



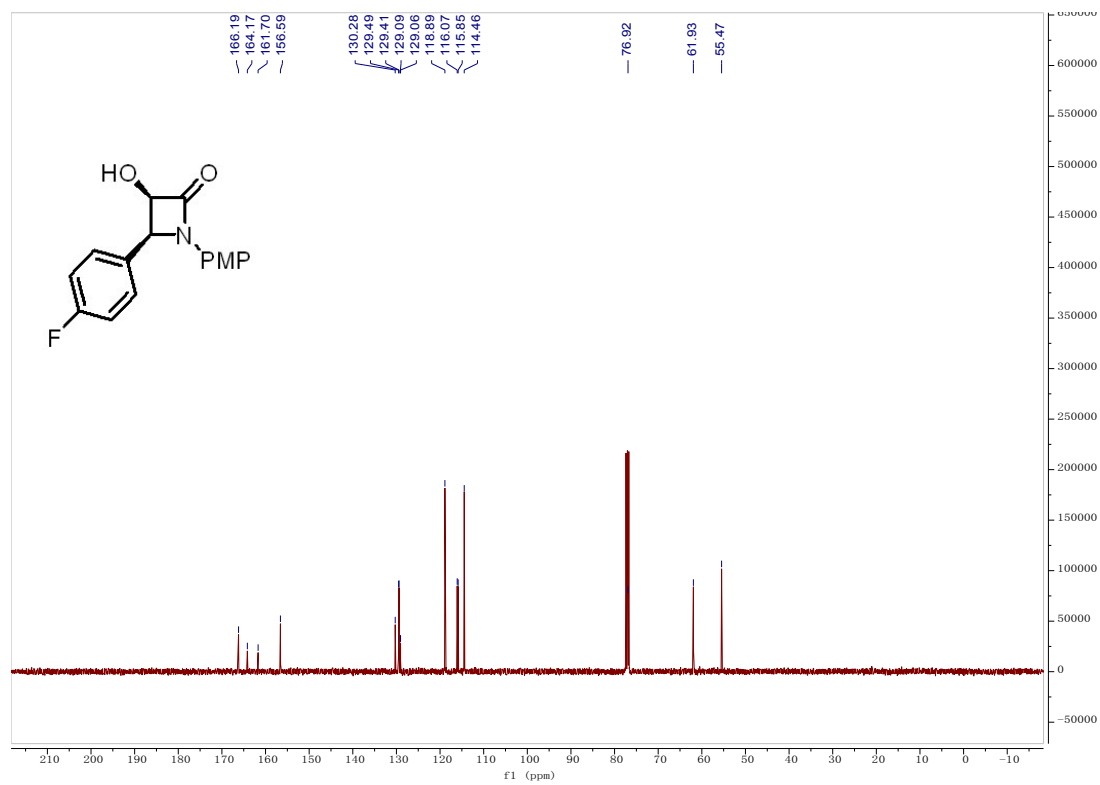
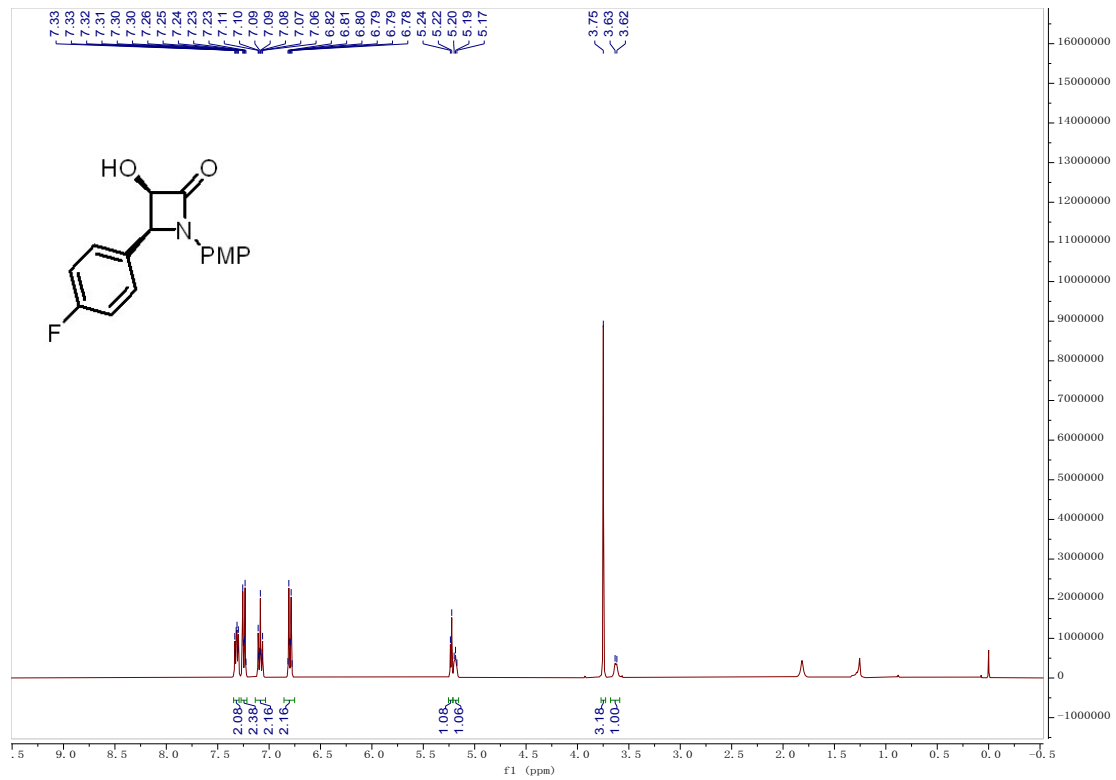
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2o



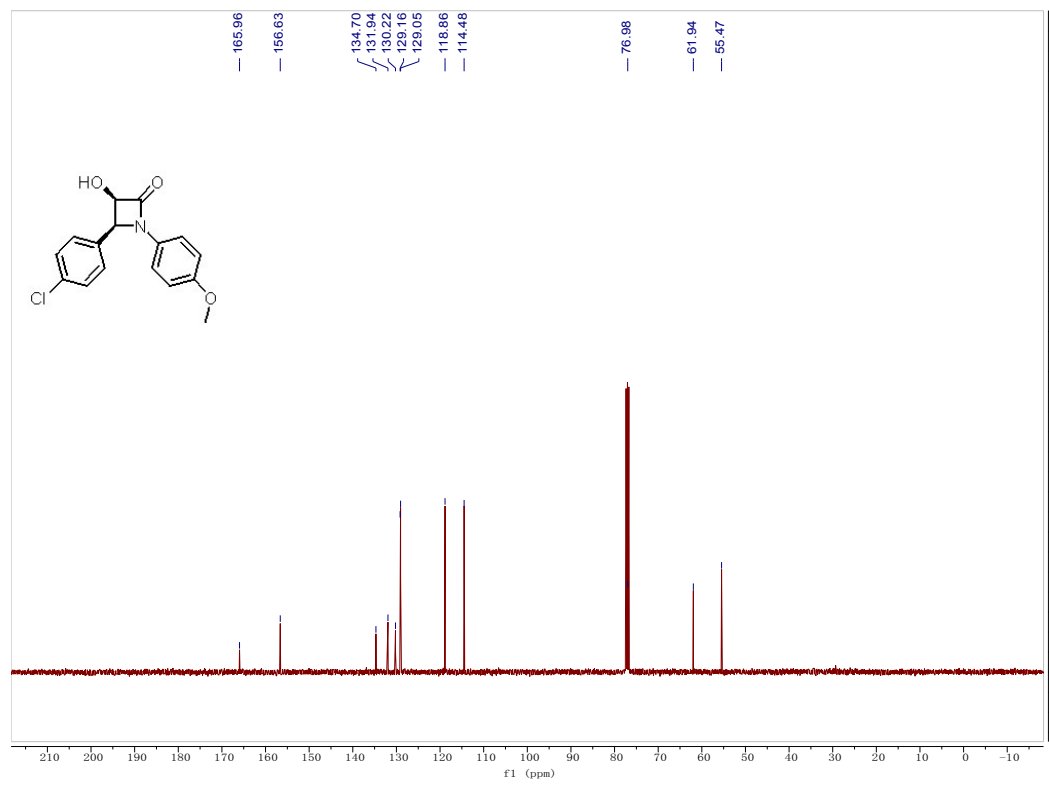
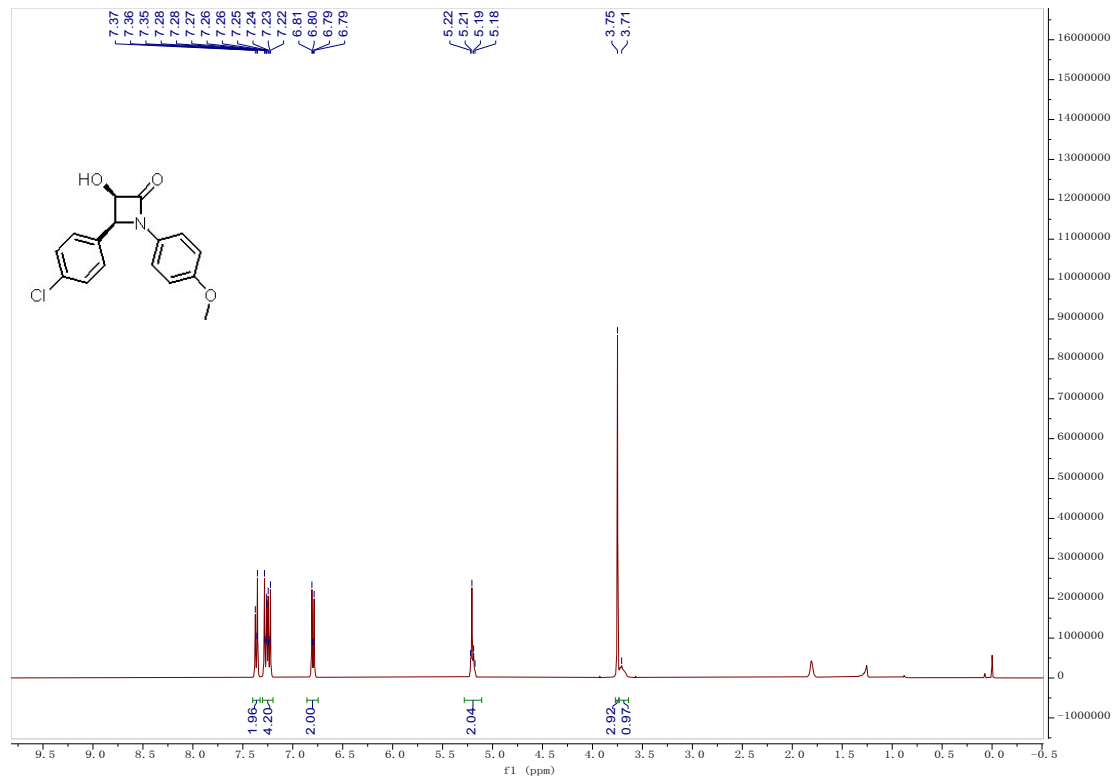
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2p



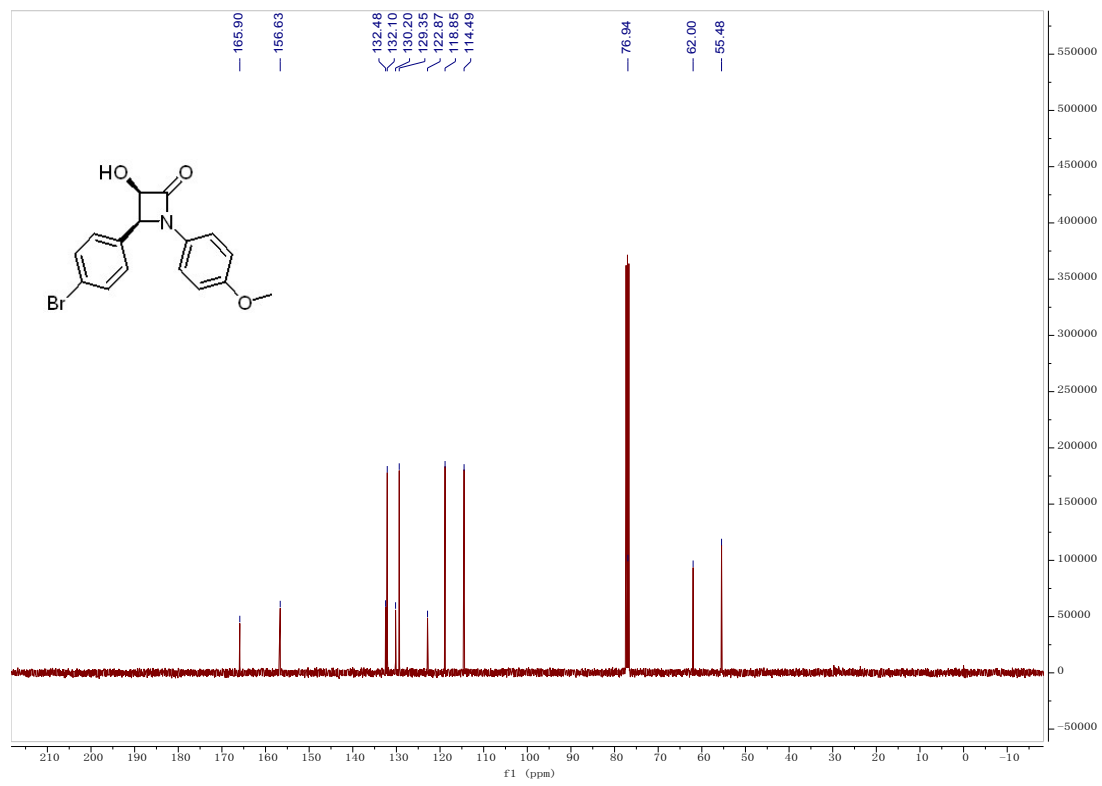
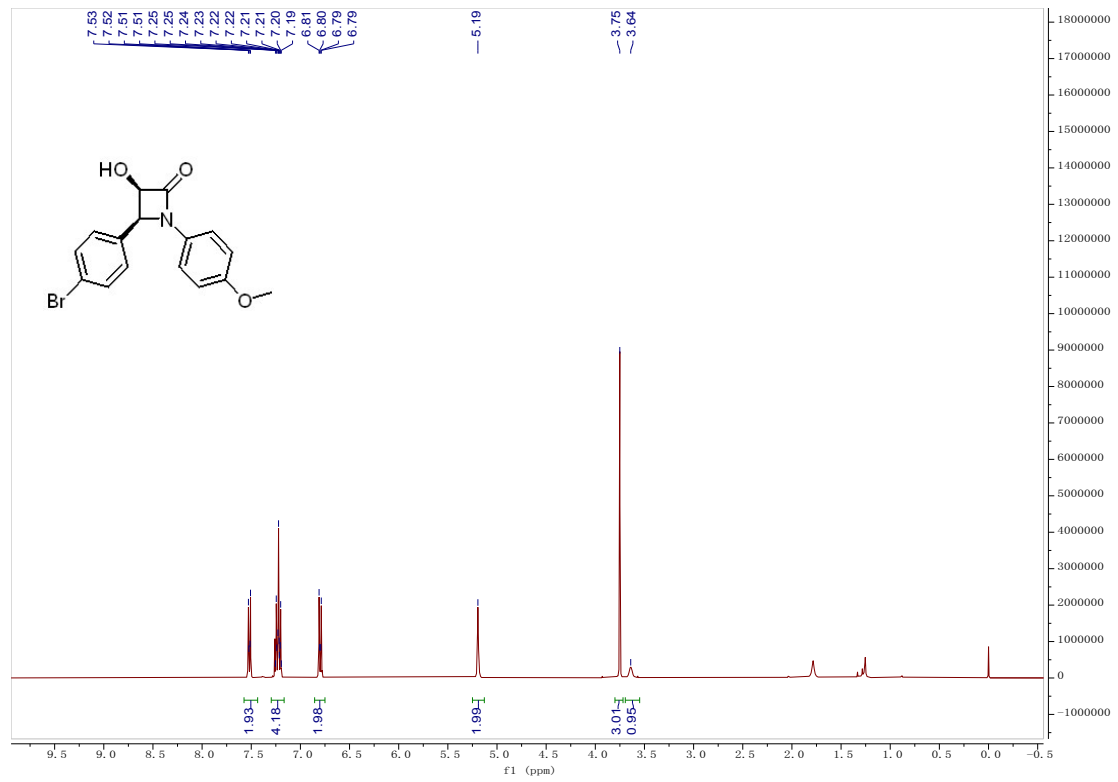
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2q



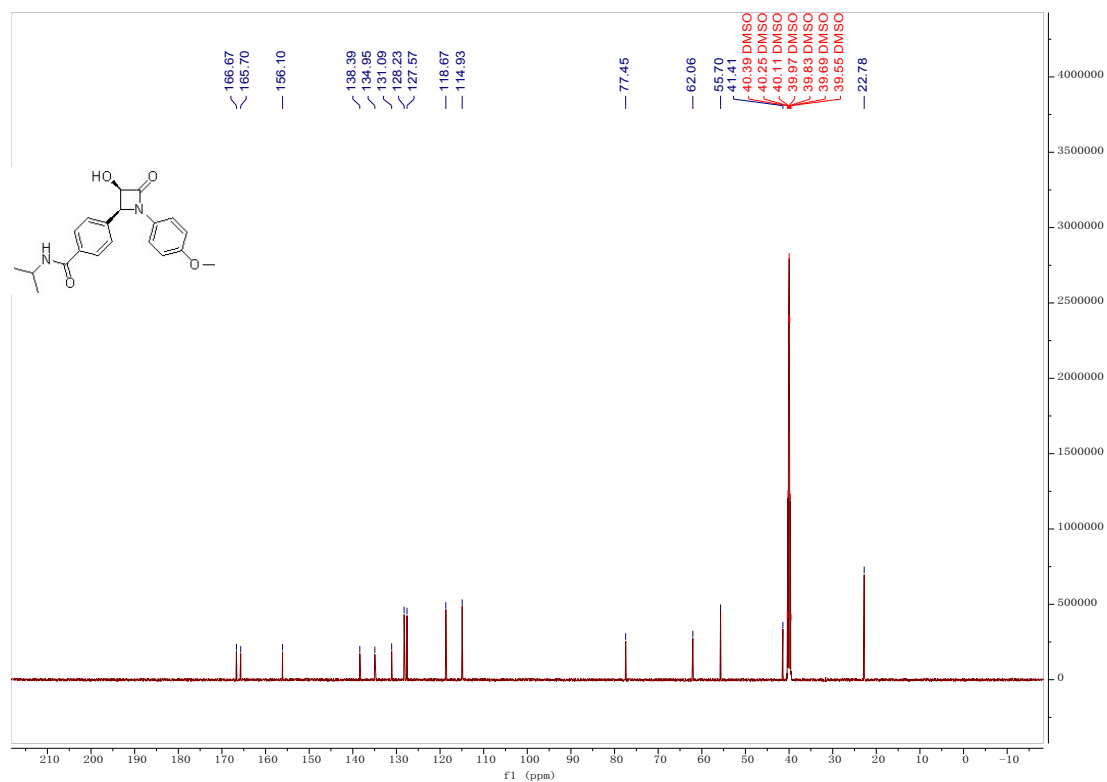
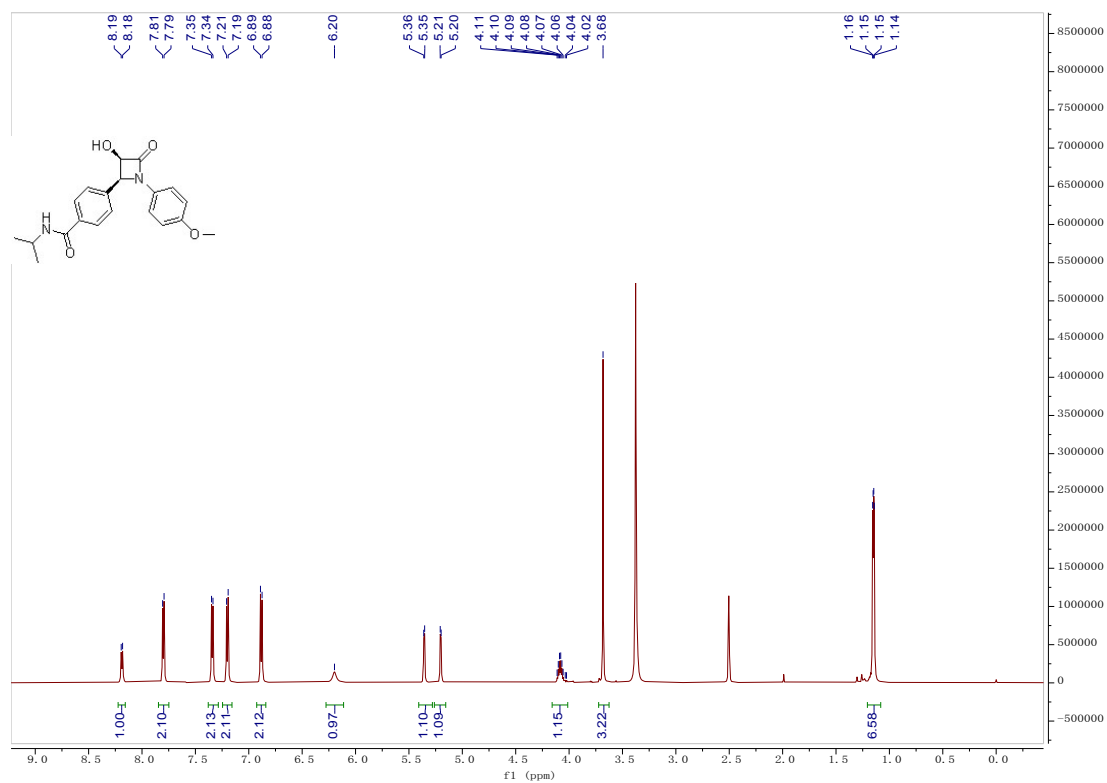
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2r



# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2s

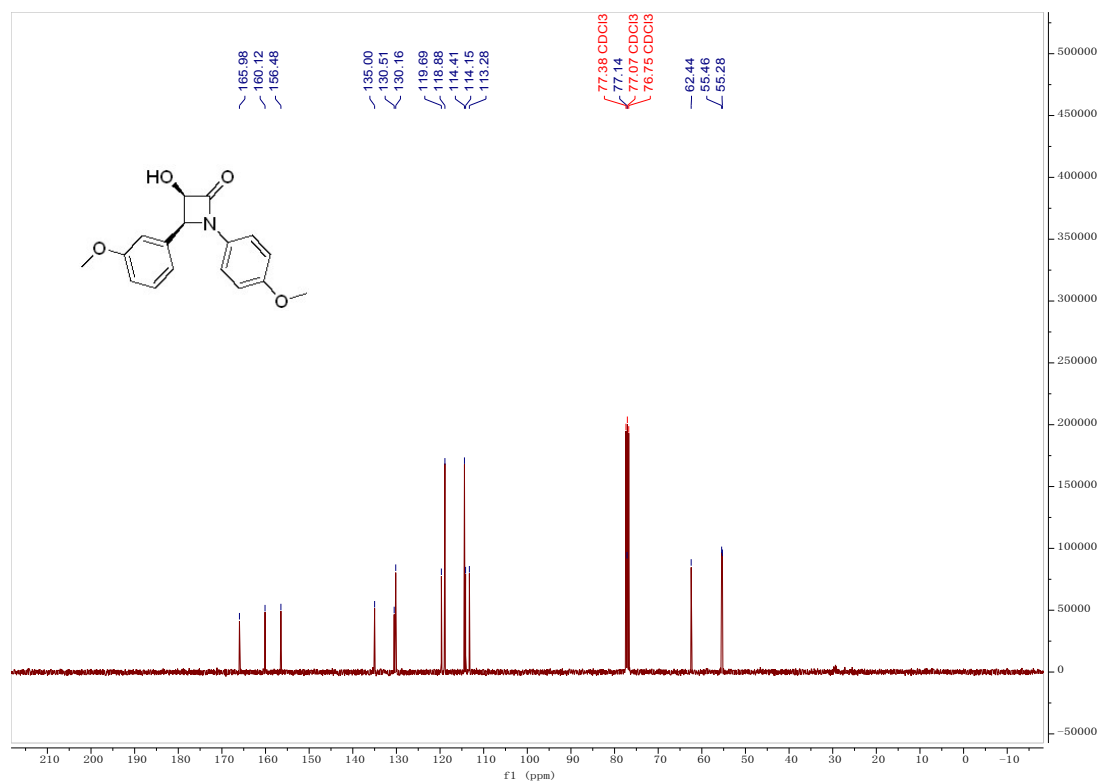
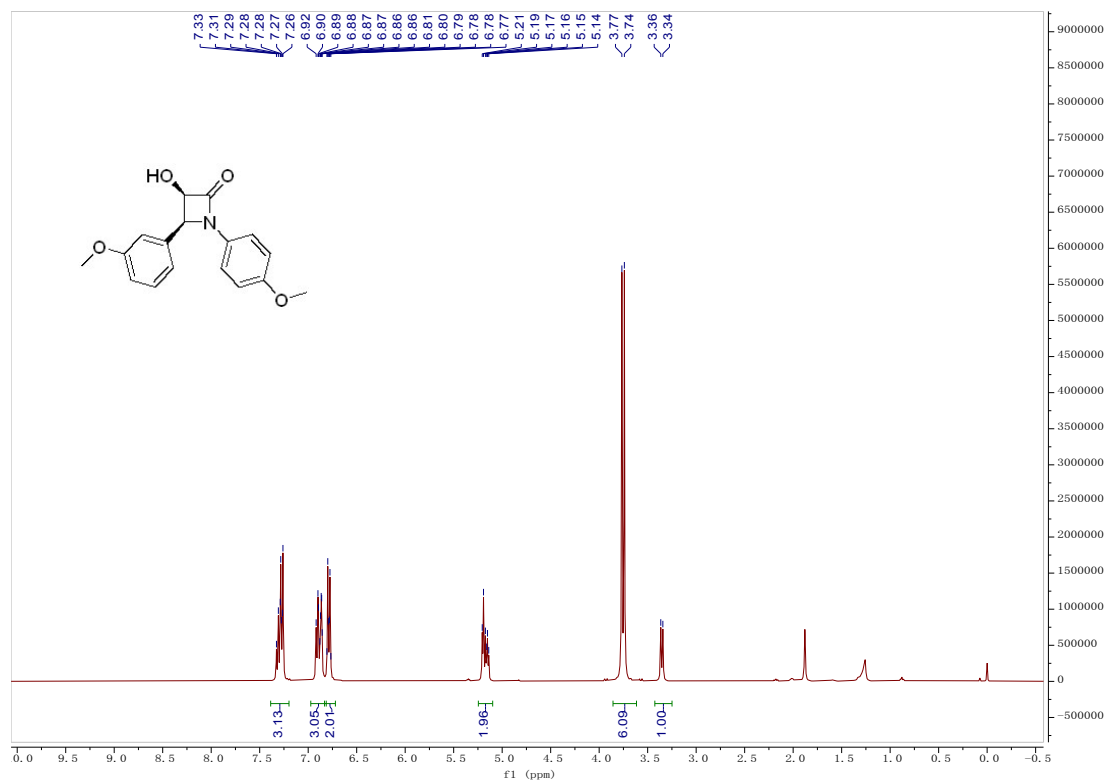


# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2t

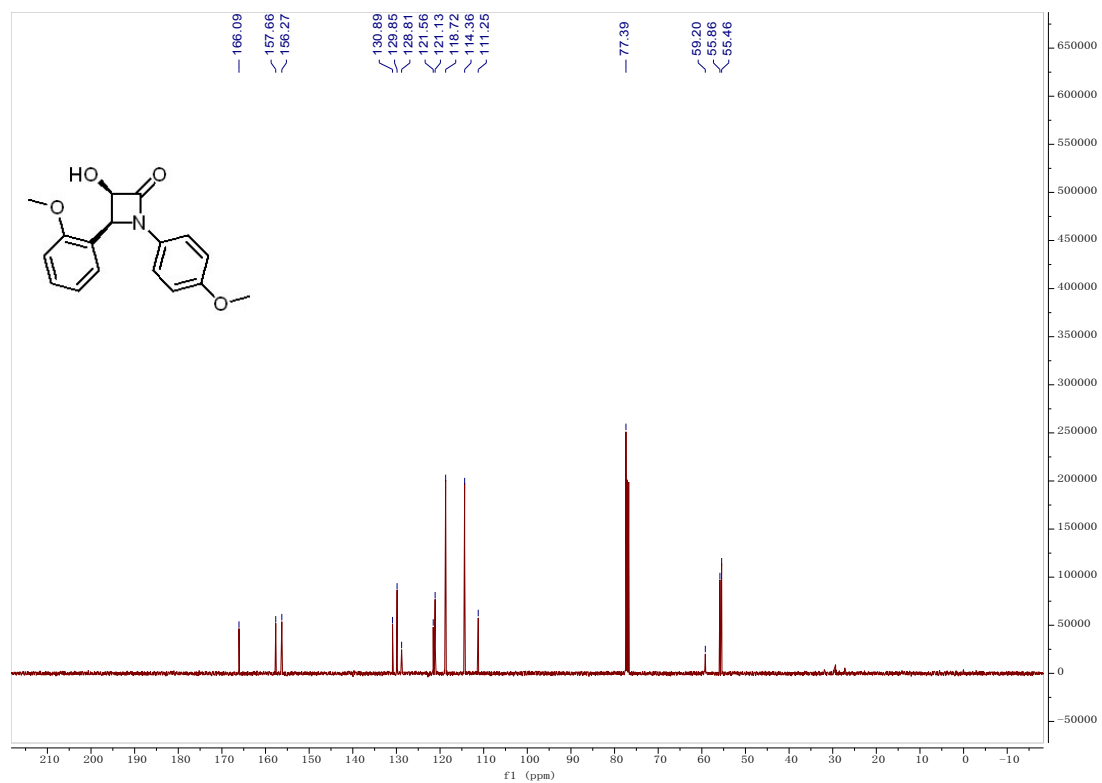
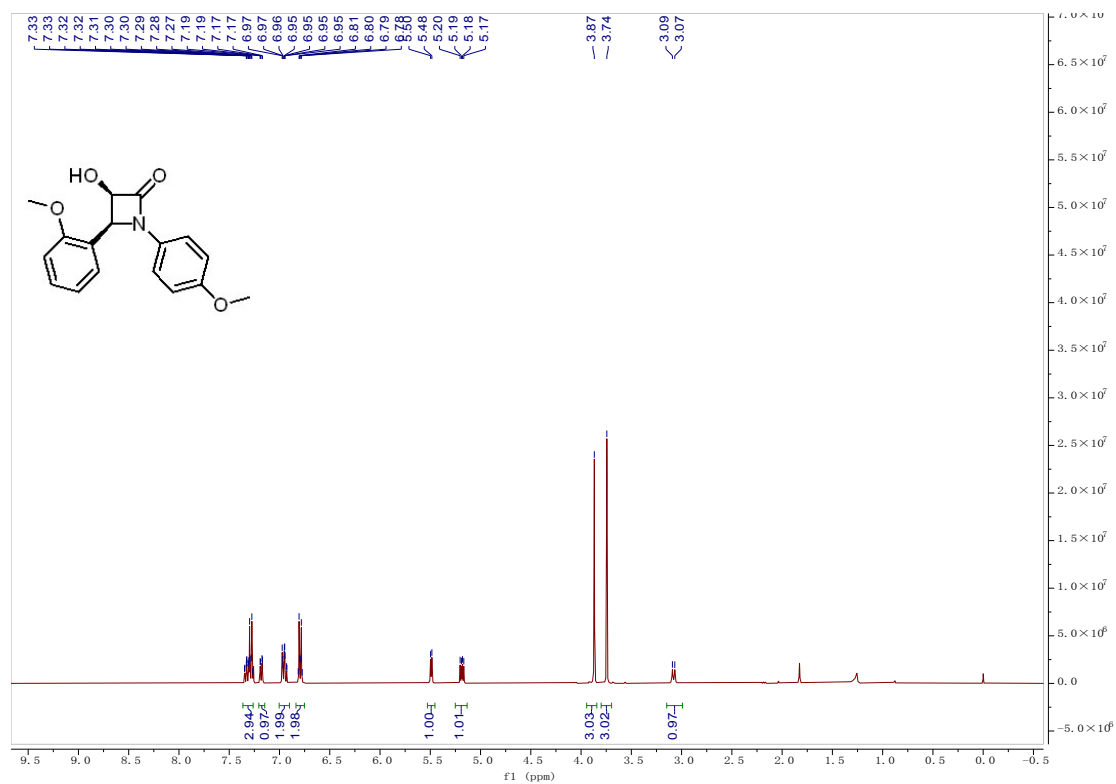




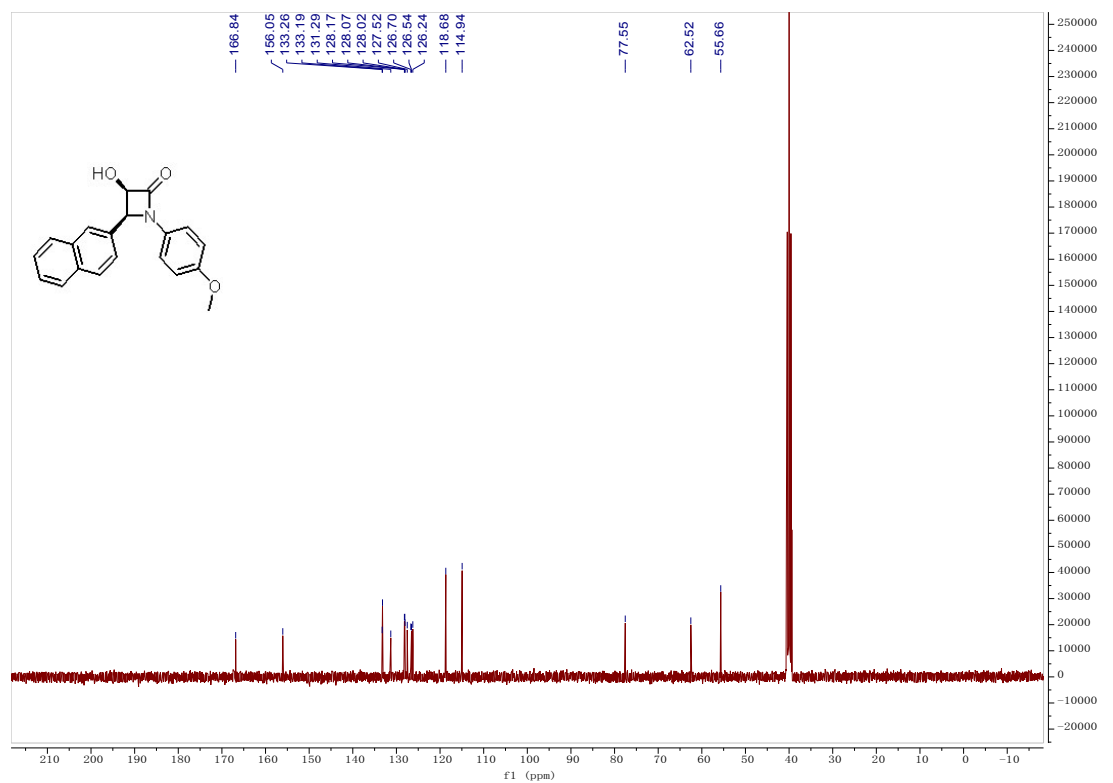
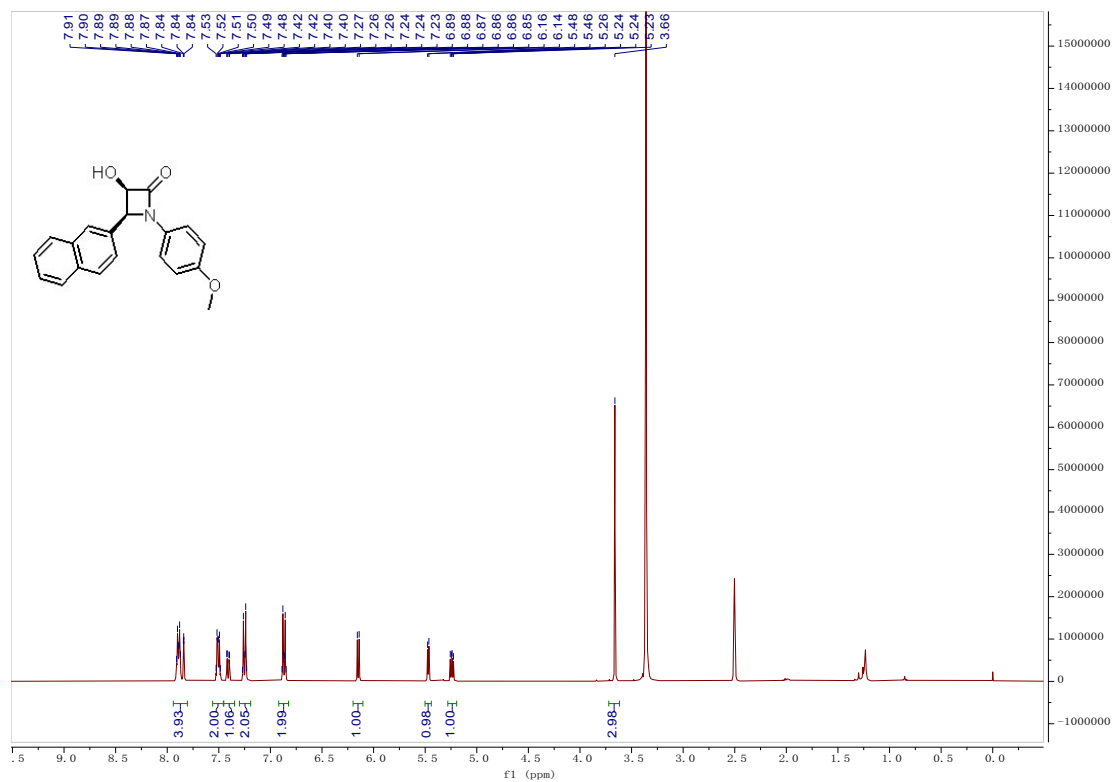
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2u



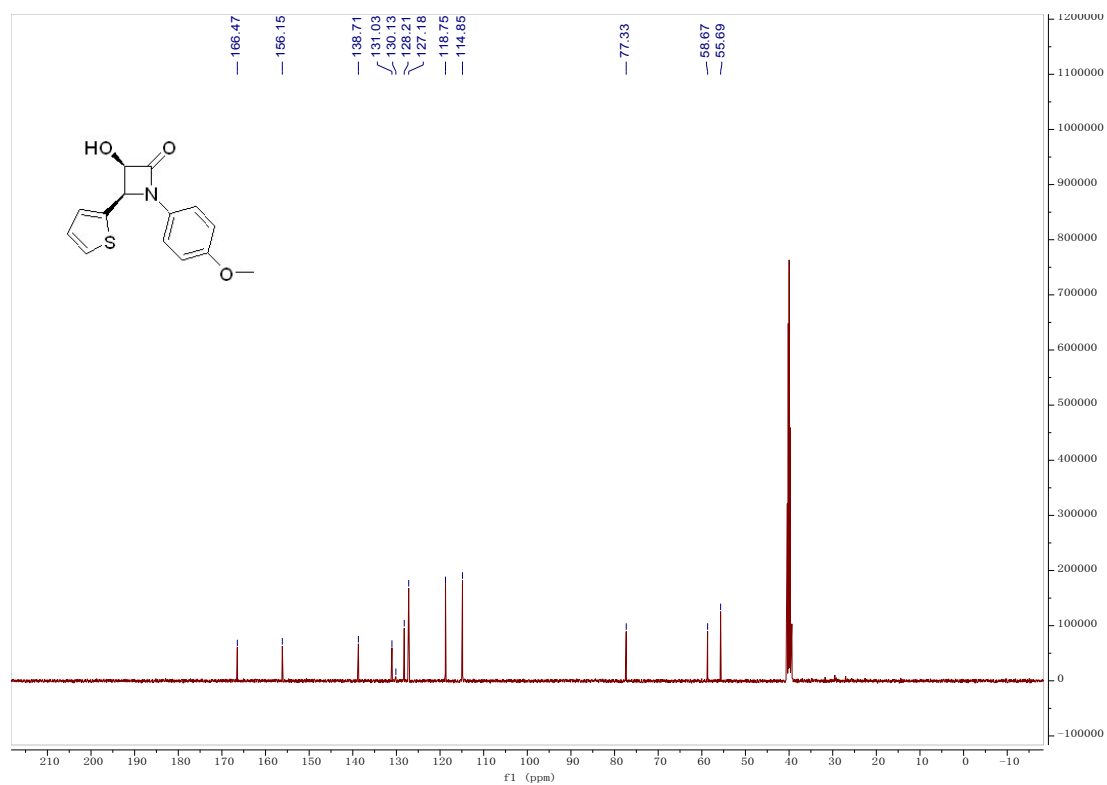
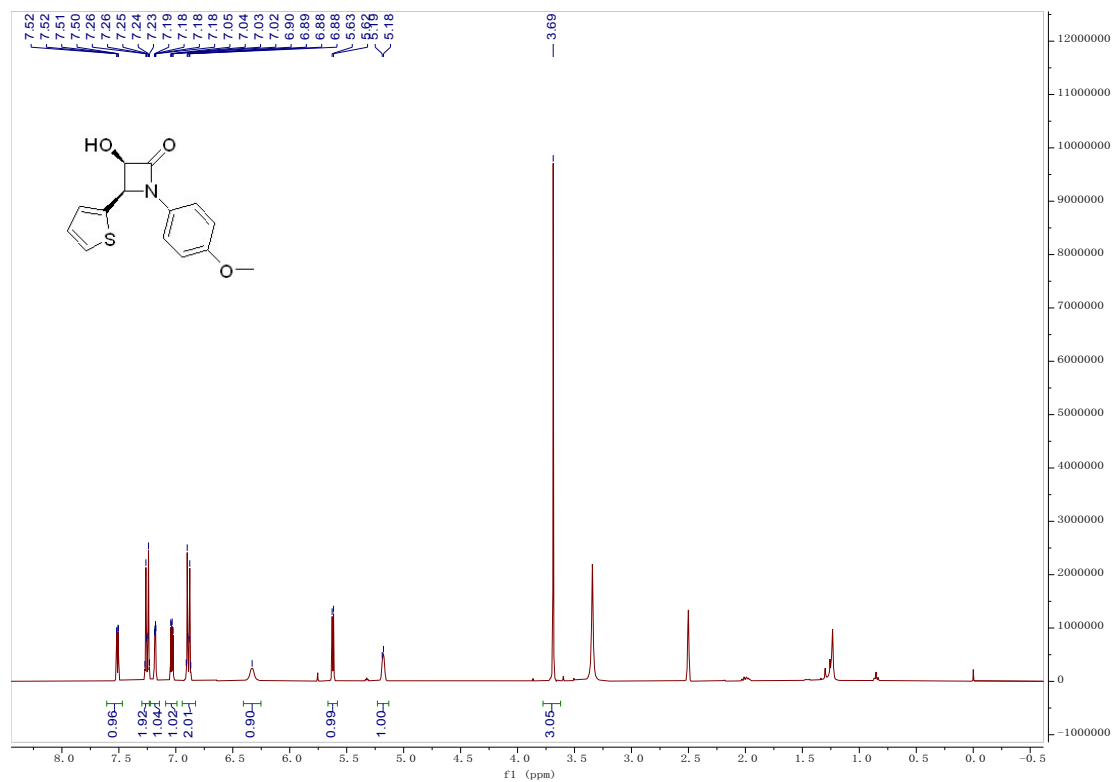
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2v



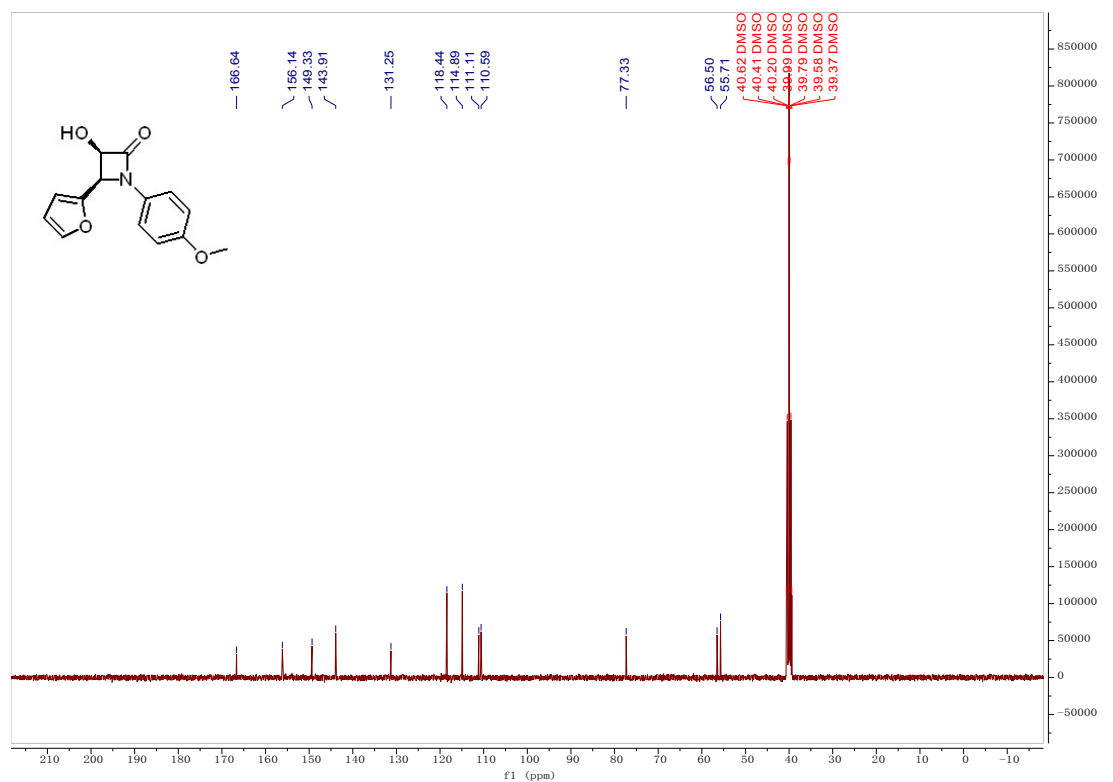
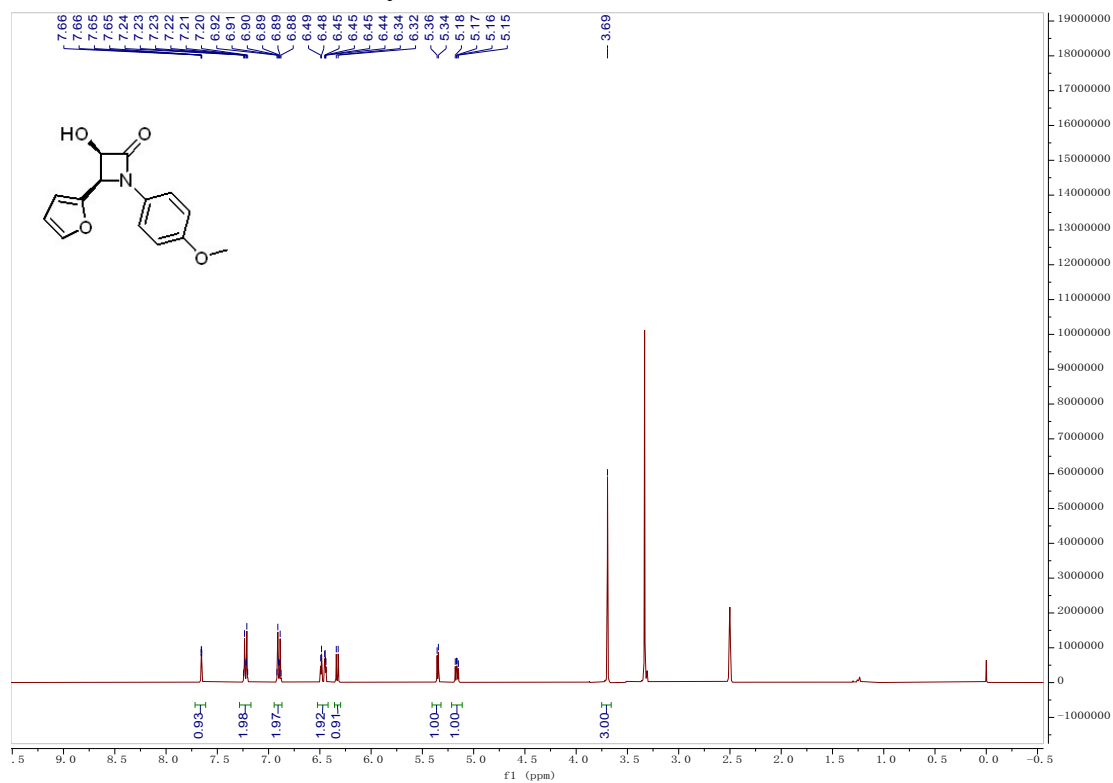
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2w



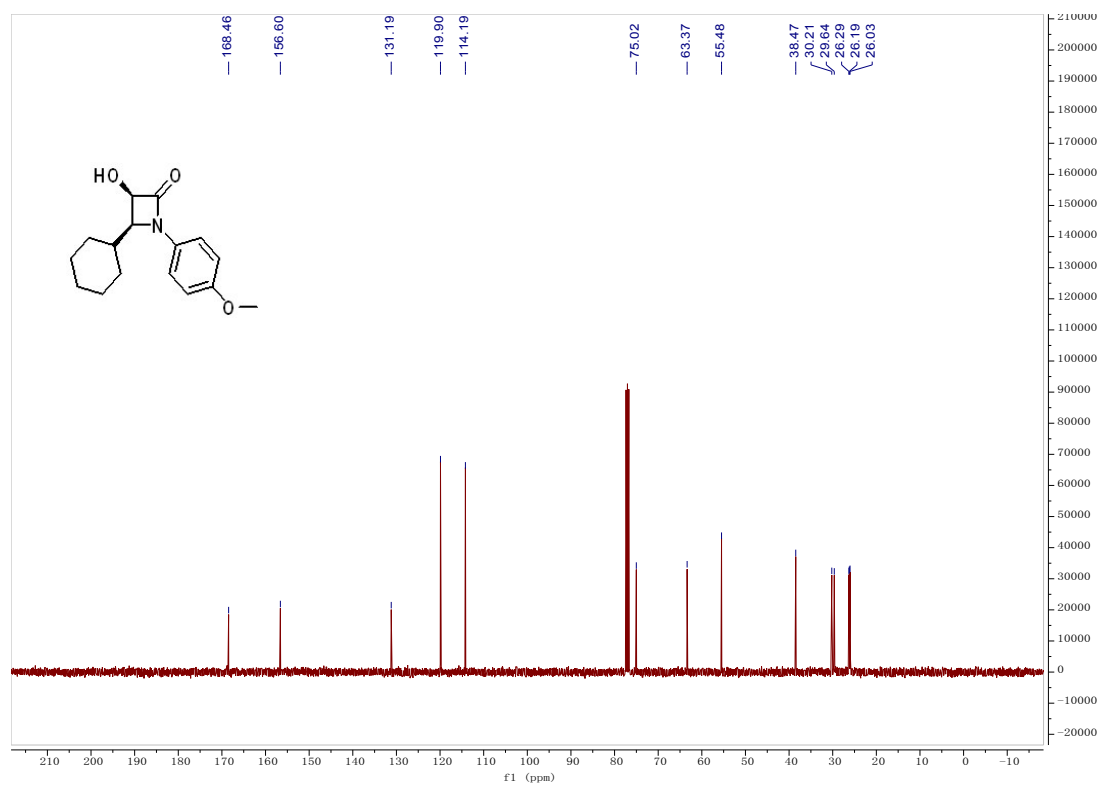
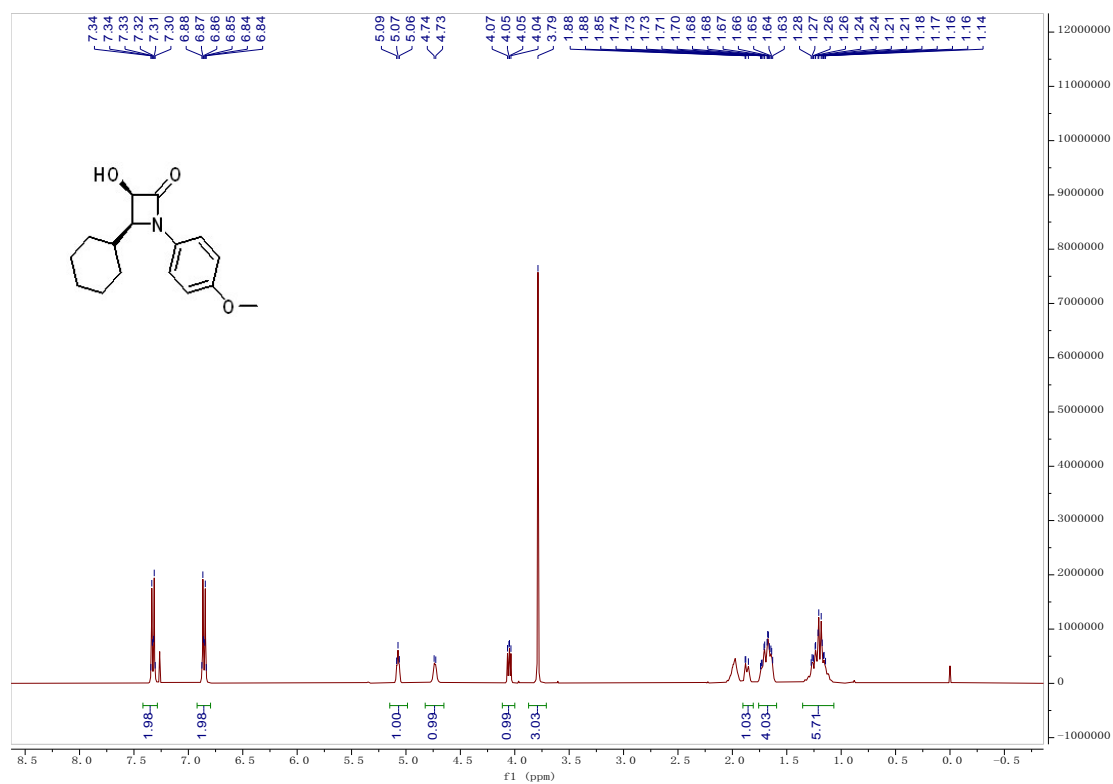
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2x



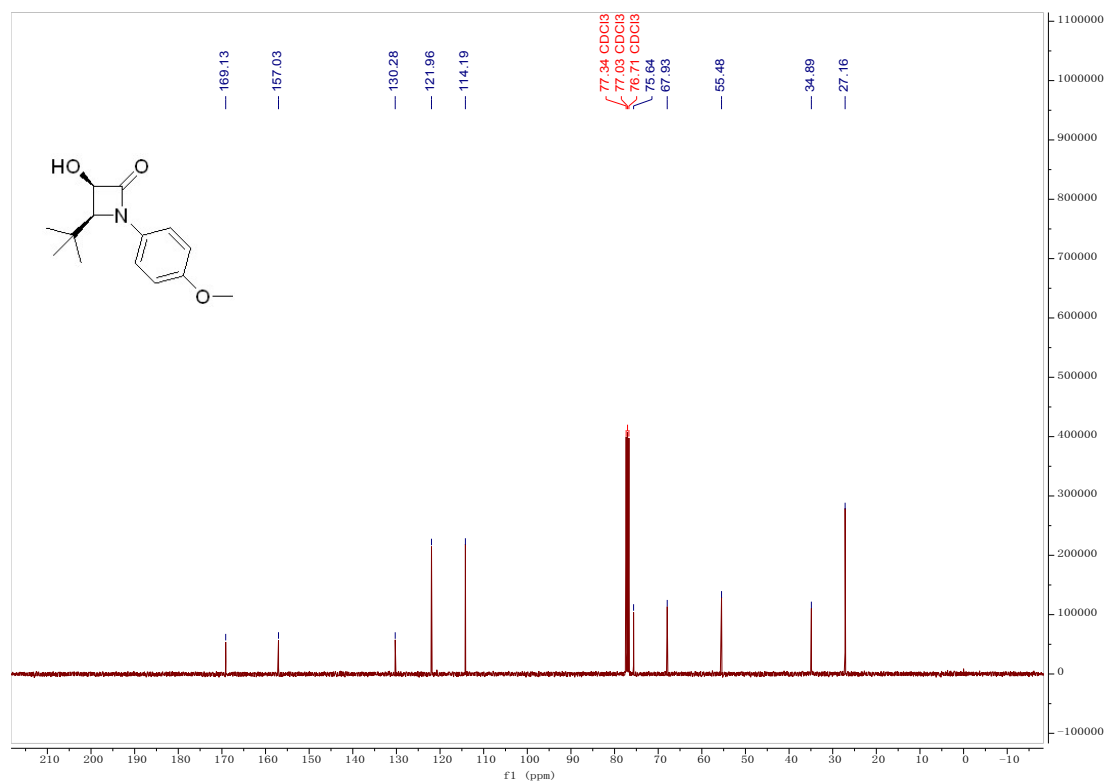
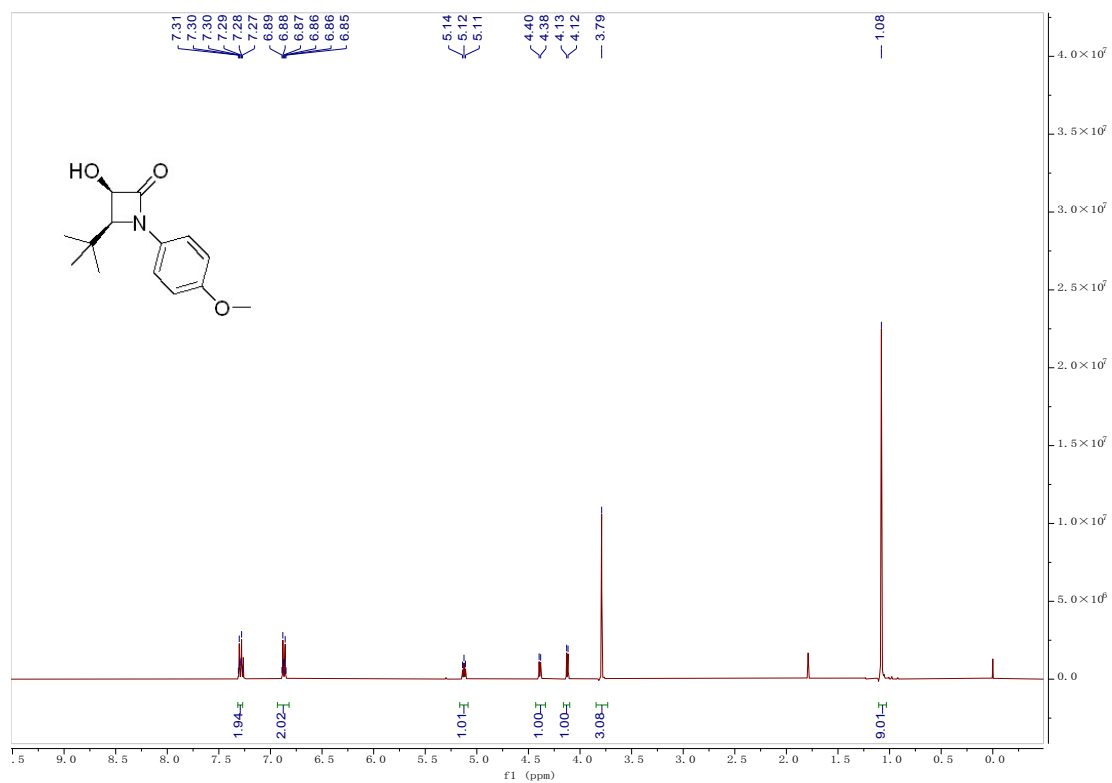
# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2y



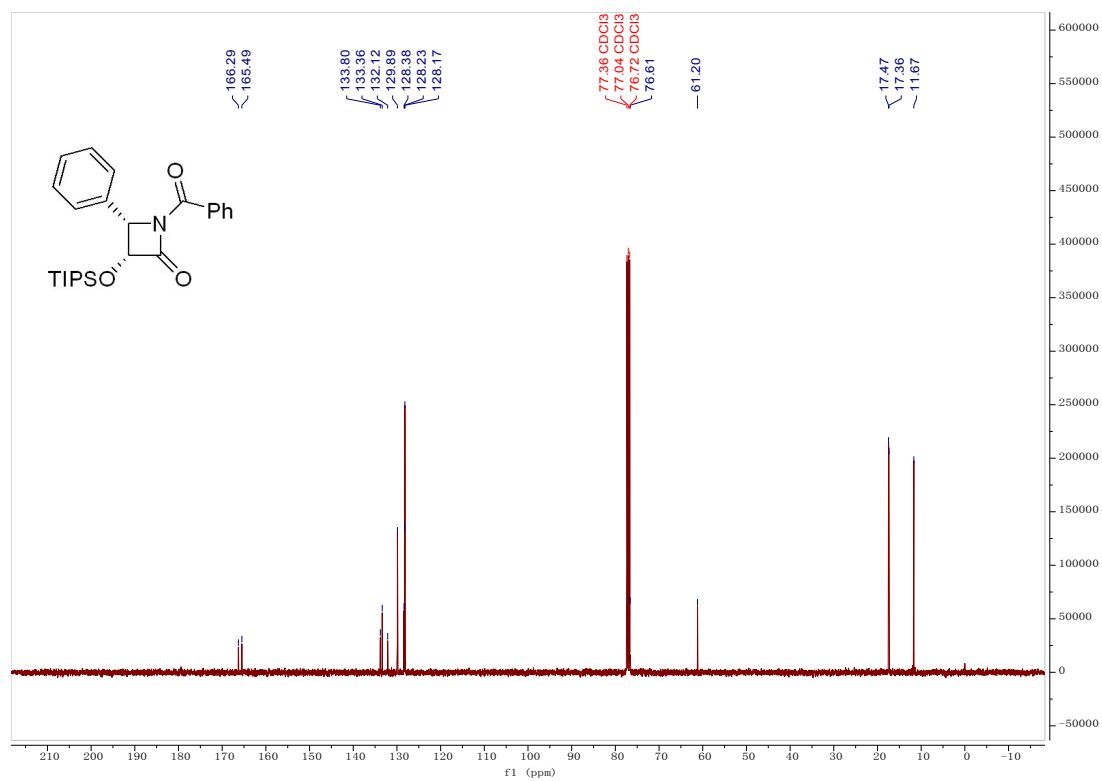
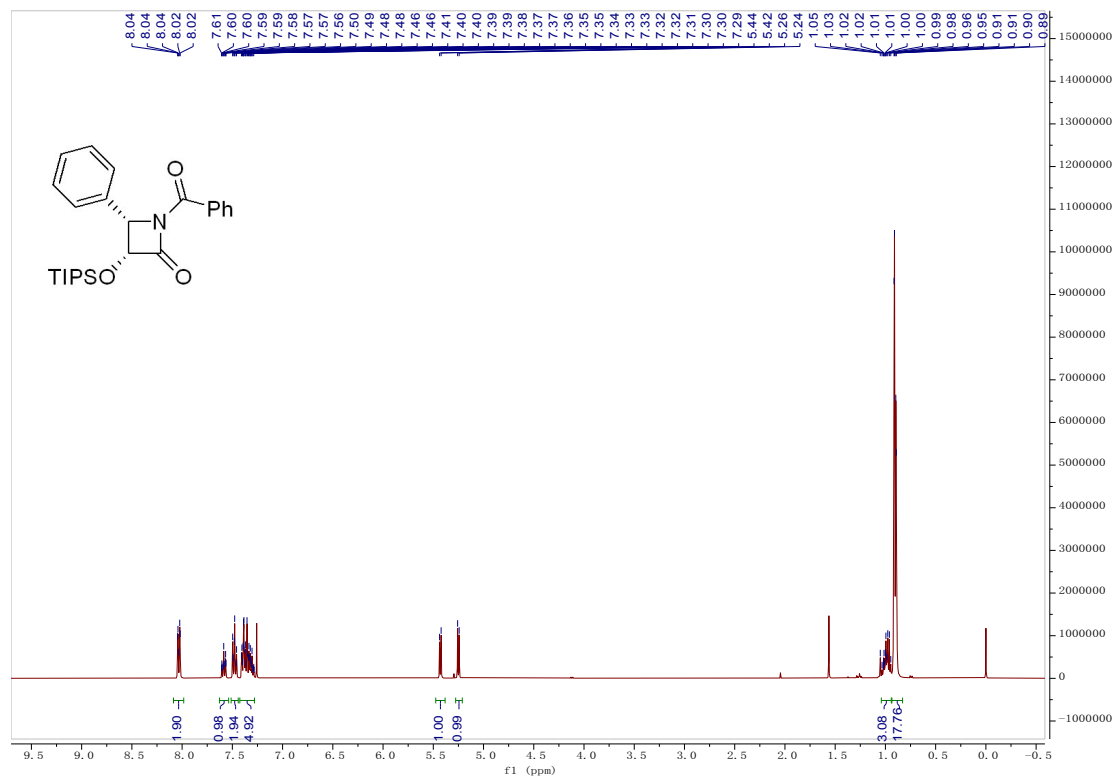
# $^1\text{H}$ NMR and $^{13}\text{C}$ NMR of 2z



# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 2aa

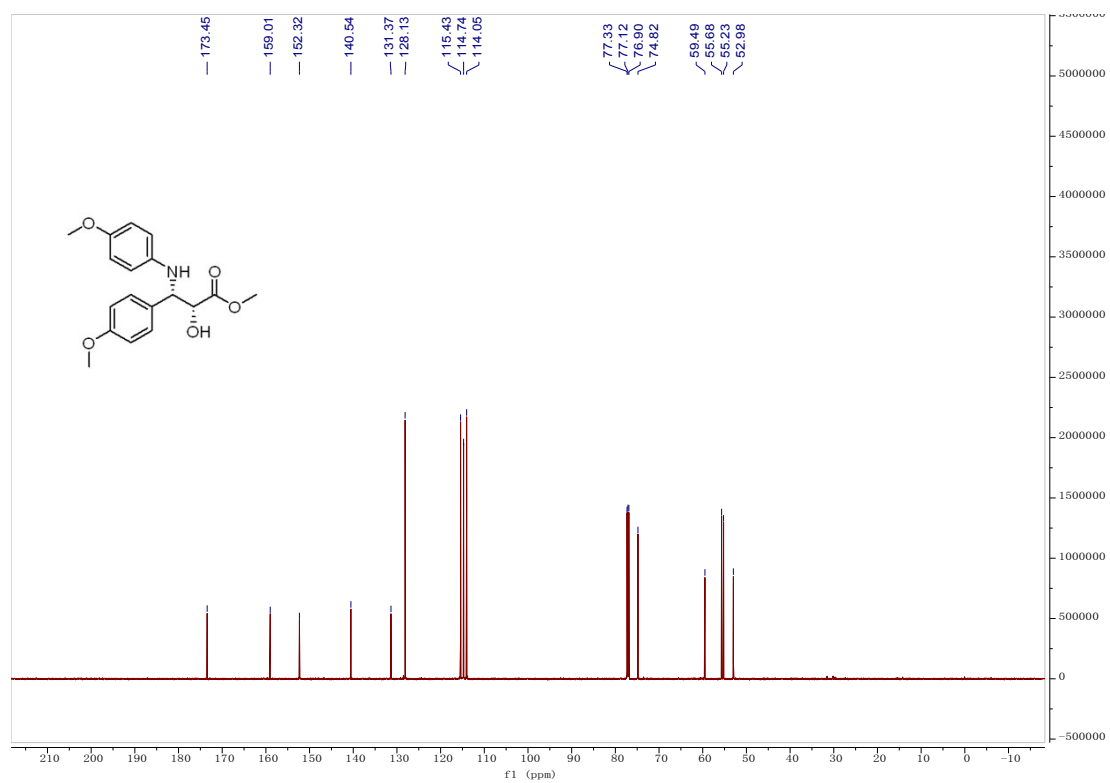
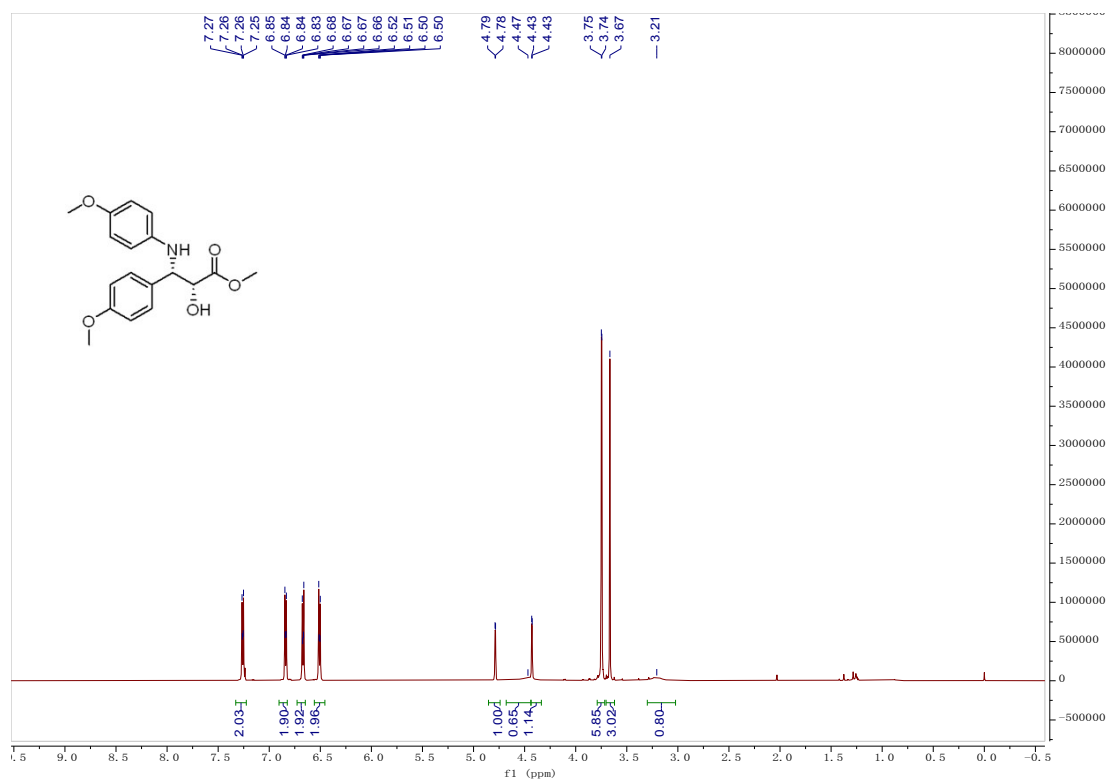


# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 4a





# <sup>1</sup>H NMR and <sup>13</sup>C NMR of 5n



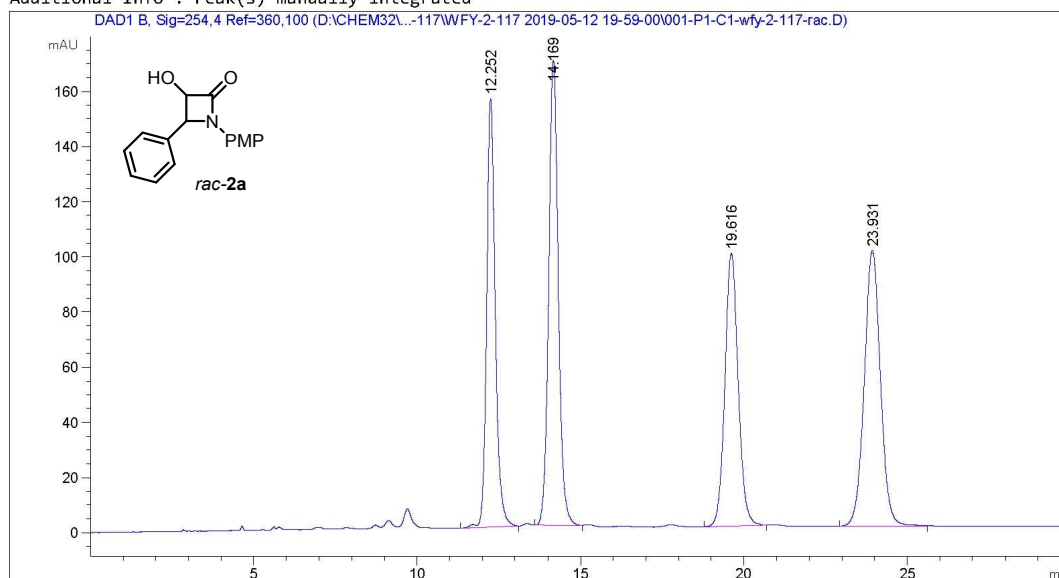


## 9. HPLC Spectra of the Products.

Data File D:\CHEM32\...\WFY-2-117\WFY-2-117 2019-05-12 19-59-00\001-P1-C1-wfy-2-117-rac.D  
 Sample Name: wfy-2-117-rac

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    1
Acq. Instrument : 1260-DAD                    Location  : P1-C-01
Injection Date  : 5/12/2019 19:59:51         Inj       :    1
                                           Inj Volume: 3.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 2.000 µl
Acq. Method     : D:\Chem32\1\Data\WFY\wfy-2-117\wfy-2-117 2019-05-12 19-59-00\IA-85-15-1ml-30min.M
Last changed    : 1/7/2019 21:25:37 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WFY\wfy-2-117\wfy-2-117 2019-05-12 19-59-00\IA-85-15-1ml-30min.M (Sequence Method)
Last changed    : 4/19/2020 22:41:24 by SYSTEM
                 (modified after loading)
Additional Info  : Peak(s) manually integrated
  
```



### Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.252	VB R	0.2749	2826.83594	155.51921	22.6851
2	14.169	BB	0.3112	3427.30908	168.54640	27.5039
3	19.616	BB	0.4302	2783.29785	98.95164	22.3357
4	23.931	BB	0.5299	3423.74170	99.87918	27.4753

Data File D:\CHEM32\...\Wfy-3-35-COMPLEX 2019-09-09 15-50-15\003-P1-A5-wfy-3-DCM complex.D  
Sample Name: wfy-3-DCM complex

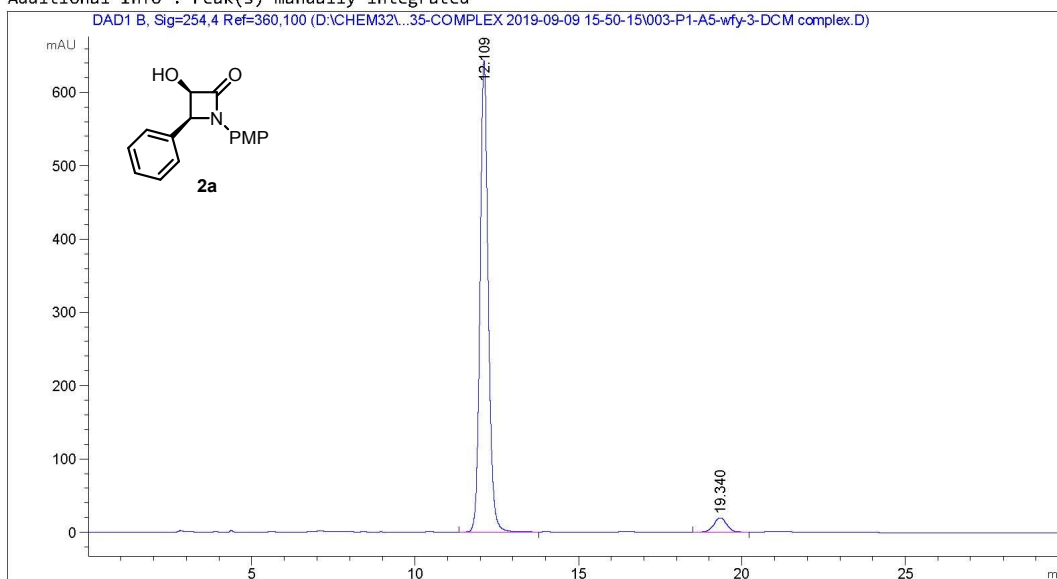
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Acq. Operator	: SYSTEM	Seq. Line	: 3
Acq. Instrument	: 1260-DAD	Location	: P1-A-05
Injection Date	: 9/9/2019 16:37:24	Inj	: 1
		Inj Volume	: 3.000 µl

Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl

Acq. Method	: D:\Chem32\1\Data\Wfy\wfy-3-35\wfy-3-35-complex 2019-09-09 15-50-15\IA-85-15-1ml-30min.M
Last changed	: 1/7/2019 21:25:37 by SYSTEM
Analysis Method	: D:\Chem32\1\Data\Wfy\wfy-3-35\wfy-3-35-complex 2019-09-09 15-50-15\IA-85-15-1ml-30min.M (Sequence Method)
Last changed	: 4/19/2020 22:40:02 by SYSTEM (modified after loading)

Additional Info : Peak(s) manually integrated



=====  
Area Percent Report  
=====

Sorted By : Signal  
Multiplier : 1.0000  
Dilution : 1.0000  
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

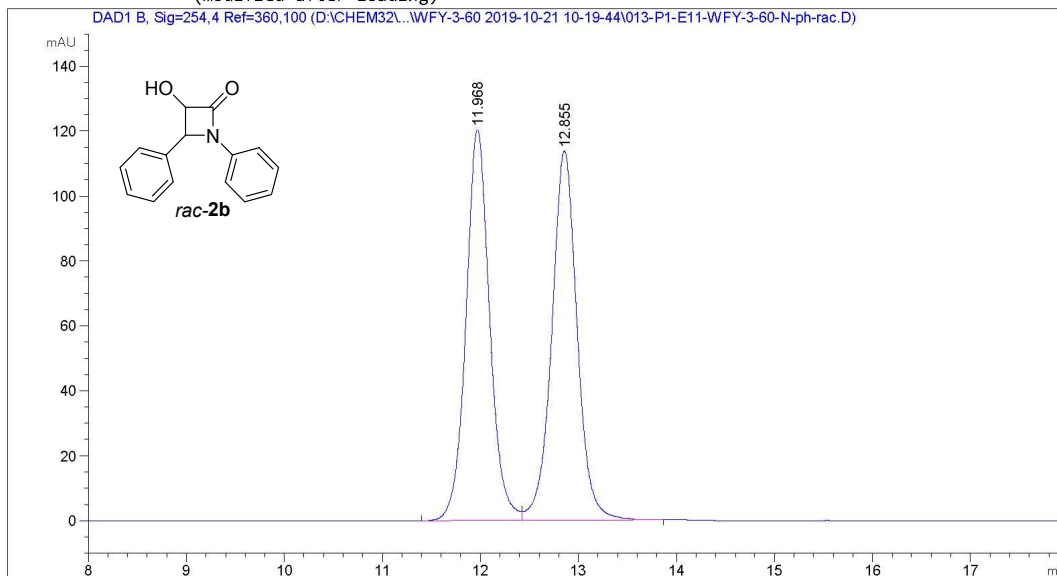
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.109	BB	0.2652	1.12109e4	643.78705	95.3865
2	19.340	BB	0.4181	542.23633	19.76000	4.6135

Totals : 1.17532e4 663.54705

Data File D:\CHEM32\...FY-3-60\WFY-3-60 2019-10-21 10-19-44\013-P1-E11-WFY-3-60-N-ph-rac.D  
 Sample Name: WFY-3-60-N-ph-rac

```
=====
Acq. Operator   : SYSTEM                      Seq. Line :   13
Acq. Instrument : 1260-DAD                   Location  : P1-E-11
Injection Date  : 10/21/2019 15:55:45       Inj       :    1
                                           Inj Volume: 3.000 µl

Acq. Method     : D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-90-10-1.0ml-
                  25min.M
Last changed    : 3/16/2019 16:37:28 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-90-10-1.0ml-
                  25min.M (Sequence Method)
Last changed    : 11/15/2019 16:31:39 by SYSTEM
                  (modified after loading)
=====
```



Area Percent Report

```
Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
```

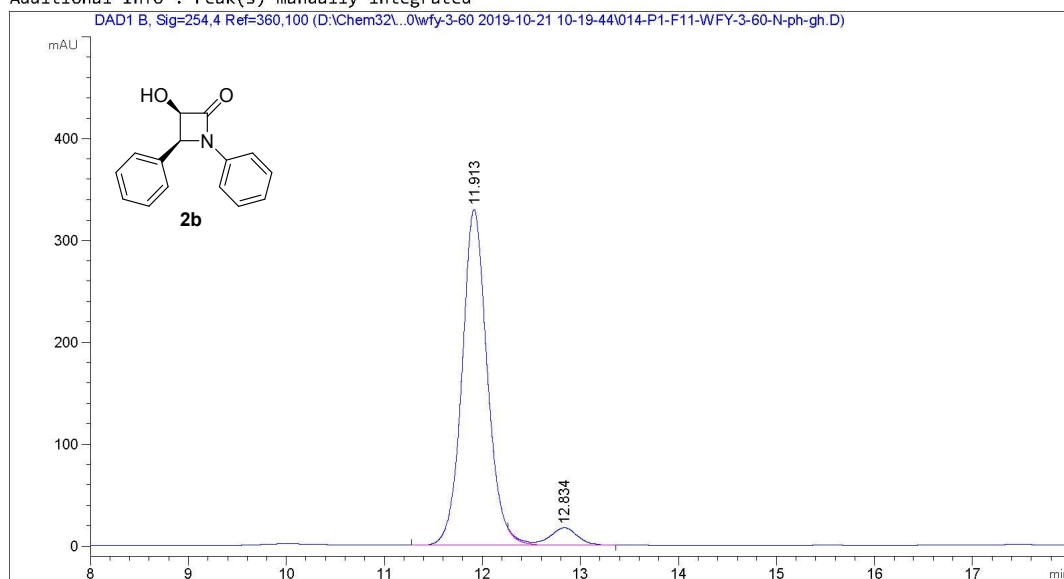
Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.968	BV	0.2536	2017.66260	120.31926	49.6550
2	12.855	VB	0.2718	2045.69629	113.74475	50.3450

Totals : 4063.35889 234.06401

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :   14
Acq. Instrument : 1260-DAD                    Location  : P1-F-11
Injection Date  : 10/21/2019 16:21:40        Inj       :    1
                                           Inj Volume: 3.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 10.000 µl
Acq. Method     : D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-90-10-1.0ml-
                25min.M
Last changed    : 3/16/2019 16:37:28 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-90-10-1.0ml-
                25min.M (Sequence Method)
Last changed    : 11/15/2019 16:32:19 by SYSTEM
                (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution      :      1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

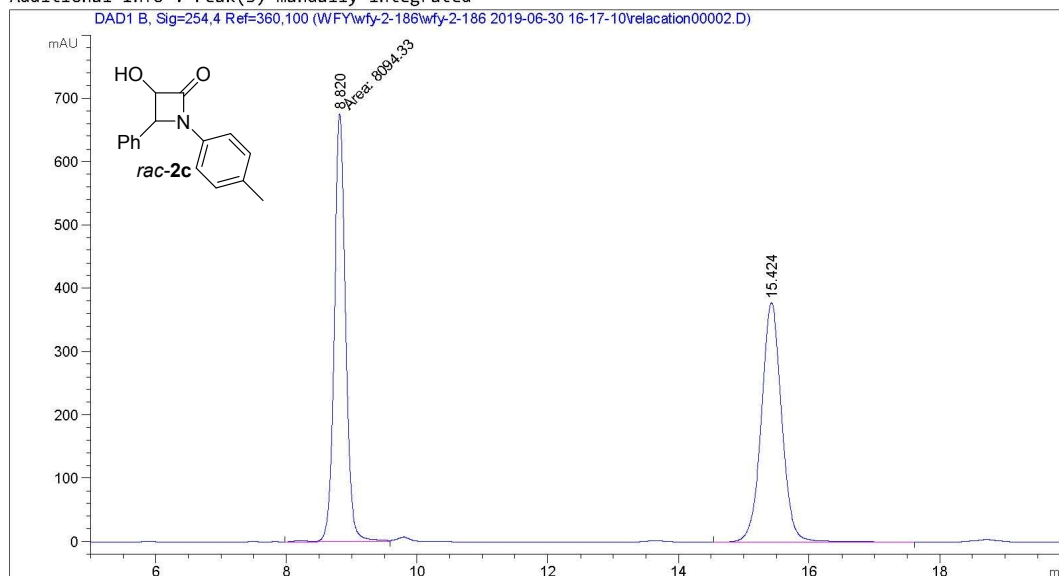
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.913	BV R	0.2708	5905.14404	329.88065	94.5105
2	12.834	VB E	0.3011	342.98813	17.01727	5.4895

Totals :                                  6248.13217    346.89791

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    2
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P1-E-01
Injection Date  : 6/30/2019 4:28:26 PM      Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 3.000 µl
Acq. Method     : D:\ChemStation\1\Data\WfY\wfy-2-186\wfy-2-186 2019-06-30 16-17-10\IA-85-15-
                  1ML-30 min.M
Last changed    : 6/26/2019 8:16:18 PM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\WfY\wfy-2-186\wfy-2-186 2019-06-30 16-17-10\IA-85-15-
                  1ML-30 min.M (Sequence Method)
Last changed    : 12/18/2019 8:25:53 PM by SYSTEM
                  (modified after loading)
  
```

Additional Info : Peak(s) manually integrated



Area Percent Report

```

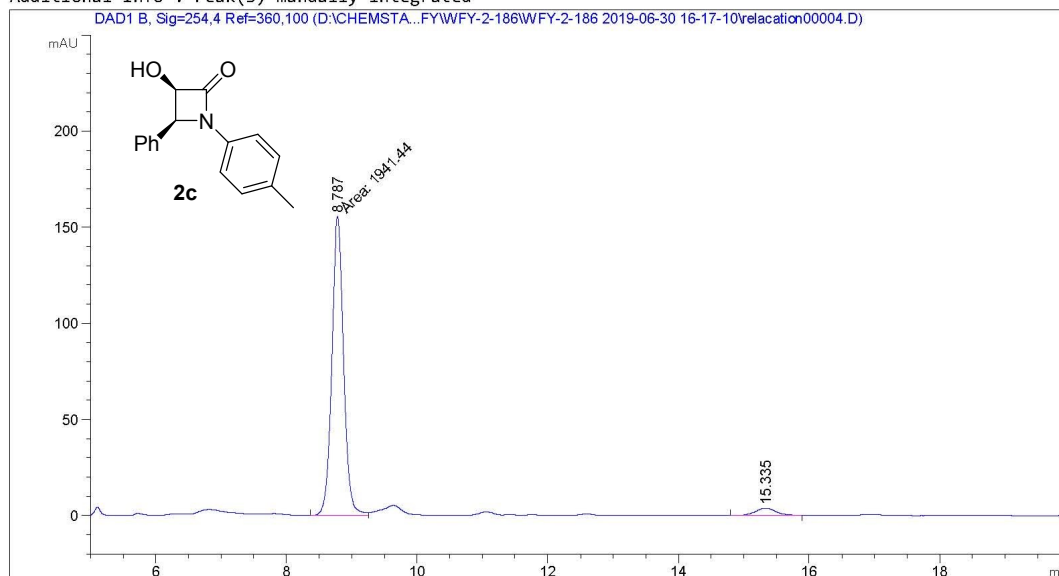
Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.820	MF	0.1995	8094.32568	676.24884	50.0021
2	15.424	BB	0.3260	8093.64795	377.72263	49.9979

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    4
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P1-E-03
Injection Date  : 6/30/2019 5:30:16 PM      Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 10.000 µl
Acq. Method     : D:\ChemStation\1\Data\WfY\wfy-2-186\wfy-2-186 2019-06-30 16-17-10\IA-85-15-
                  1ML-30 min.M
Last changed    : 6/26/2019 8:16:18 PM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\WfY\wfy-2-186\wfy-2-186 2019-06-30 16-17-10\IA-85-15-
                  1ML-30 min.M (Sequence Method)
Last changed    : 12/18/2019 8:24:30 PM by SYSTEM
                  (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.787	MF	0.2081	1941.43738	155.46277	96.0785
2	15.335	BB	0.3267	79.24133	3.68805	3.9215





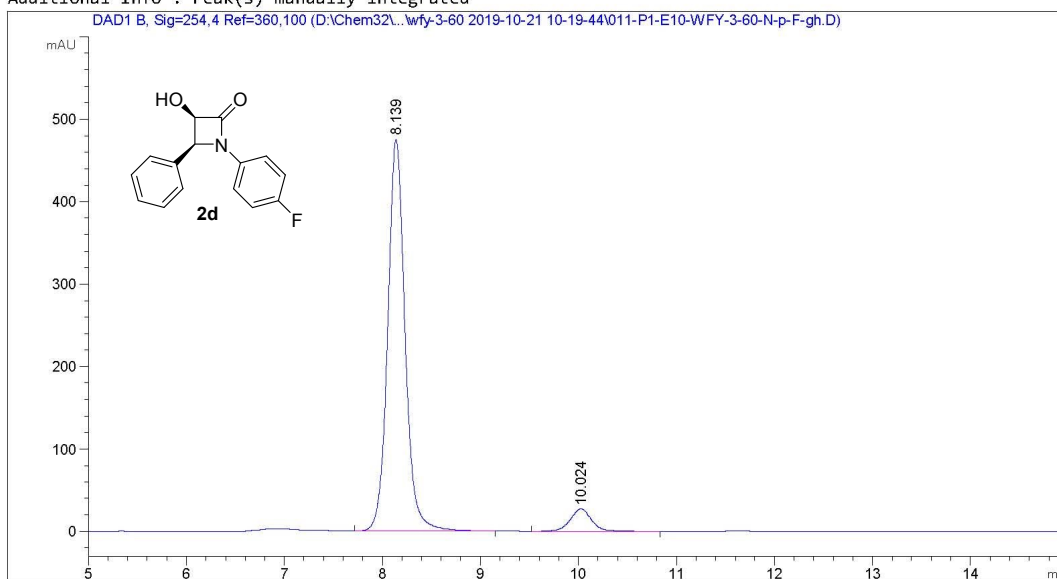
=====

Acq. Operator	: SYSTEM	Seq. Line	: 11
Acq. Instrument	: 1260-DAD	Location	: P1-E-10
Injection Date	: 10/21/2019 15:14:25	Inj	: 1
		Inj Volume	: 3.000 µl

Different Inj Volume from Sample Entry! Actual Inj Volume : 10.000 µl

Acq. Method	: D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-30min.M
Last changed	: 1/7/2019 21:25:37 by SYSTEM
Analysis Method	: D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-30min.M (Sequence Method)
Last changed	: 11/15/2019 16:29:33 by SYSTEM (modified after loading)

Additional Info : Peak(s) manually integrated



=====  
Area Percent Report  
=====

Sorted By : Signal  
Multiplier : 1.0000  
Dilution : 1.0000  
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.139	BB	0.1850	5822.56787	475.07040	93.5051
2	10.024	BB	0.2258	404.43726	27.14475	6.4949

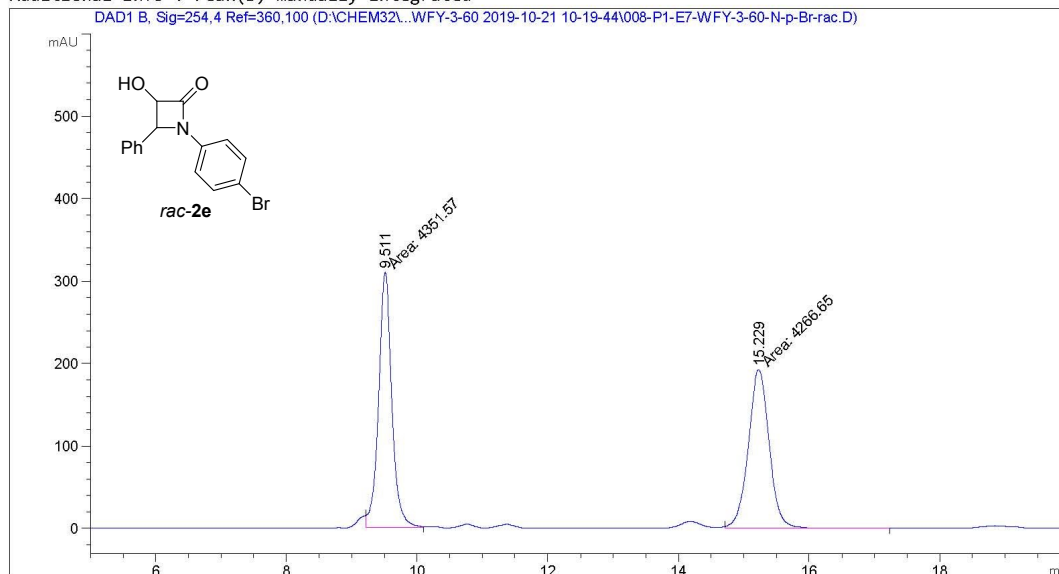
Totals : 6227.00513 502.21515

Data File D:\CHEM32\...Y-3-60\Wfy-3-60 2019-10-21 10-19-44\008-P1-E7-WFY-3-60-N-p-Br-rac.D  
 Sample Name: WFY-3-60-N-p-Br-rac

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    8
Acq. Instrument : 1260-DAD                    Location  : P1-E-07
Injection Date  : 10/21/2019 13:41:37        Inj       :    1
                                           Inj Volume: 3.000 µl

Acq. Method    : D:\Chem32\1\Data\Wfy\Wfy-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-
                30min.M
Last changed   : 1/7/2019 21:25:37 by SYSTEM
Analysis Method : D:\Chem32\1\Data\Wfy\Wfy-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-
                30min.M (Sequence Method)
Last changed   : 11/15/2019 16:26:42 by SYSTEM
                (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

=====
Sorted By      : Signal
Multiplier     : 1.0000
Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.511	FM	0.2340	4351.57227	309.97708	50.4927
2	15.229	FM	0.3691	4266.65186	192.66867	49.5073

Totals : 8618.22412 502.64575

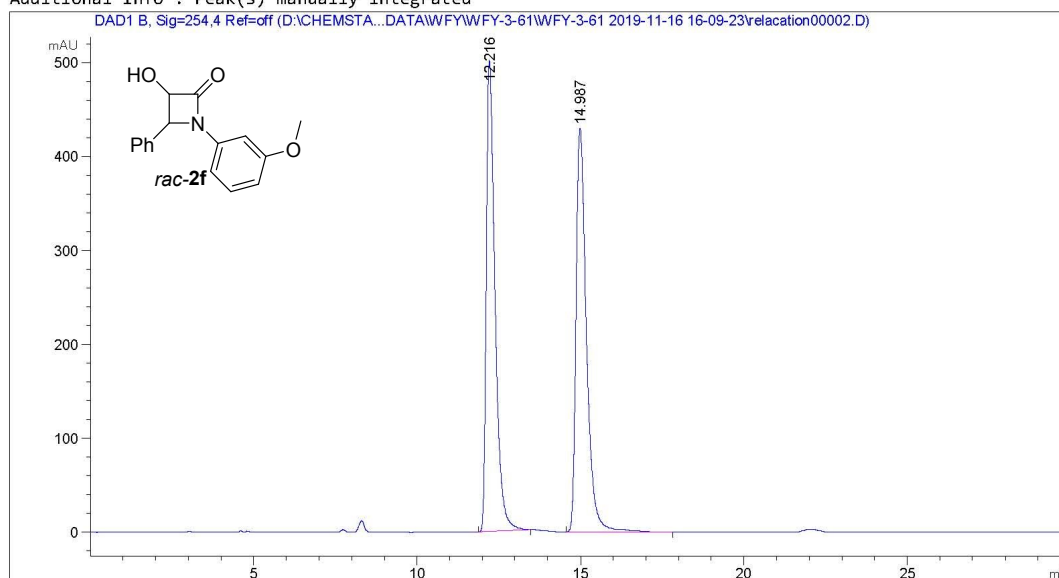


Data File D:\CHEMSTA...\1\DATA\WfY\WfY-3-61\WfY-3-61 2019-11-16 16-09-23\relacation00002.D  
 Sample Name: wfy-3-61-N-MeO-rac

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    2
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P1-E-01
Injection Date  : 11/16/2019 4:20:40 PM      Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl
Acq. Method    : D:\ChemStation\1\Data\WfY\wfy-3-61\wfy-3-61 2019-11-16 16-09-23\OD-3-90-10-
                1ML-30min.M
Last changed   : 6/28/2019 8:29:21 AM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\WfY\wfy-3-61\wfy-3-61 2019-11-16 16-09-23\OD-3-90-10-
                1ML-30min.M (Sequence Method)
Last changed   : 11/19/2019 9:43:05 PM by SYSTEM
                (modified after loading)
  
```

Additional Info : Peak(s) manually integrated



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

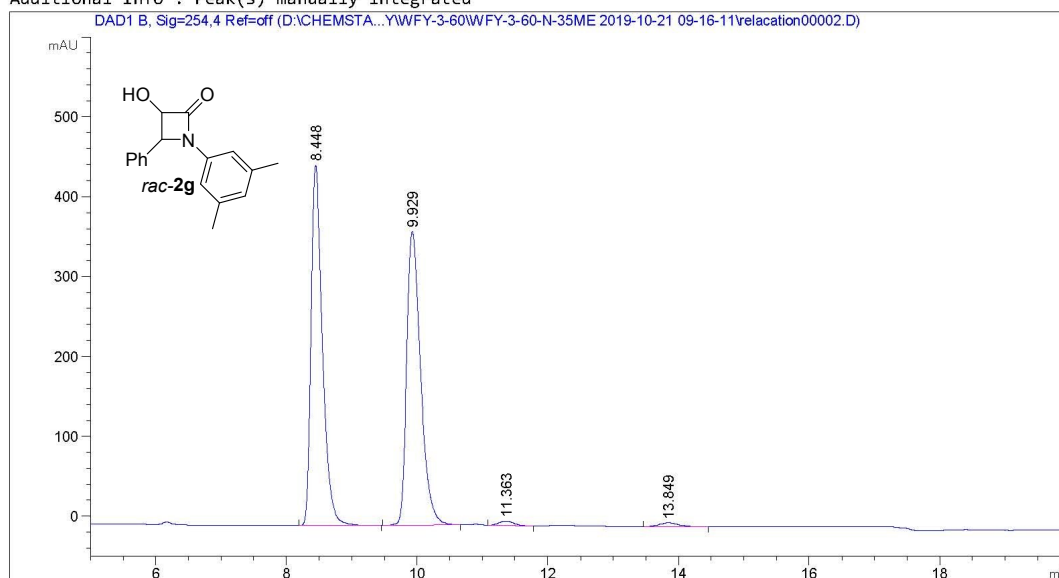
Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.216	BB	0.2782	9287.84180	501.23859	49.4806
2	14.987	BB	0.3336	9482.82031	429.53882	50.5194



```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    2
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P2-C-01
Injection Date  : 10/21/2019 9:27:25 AM      Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 3.000 µl
Acq. Method    : D:\ChemStation\1\Data\WFY\wfy-3-60\wfy-3-60-N-35Me 2019-10-21 09-16-11\OD-3
               : -90-10-1ML-30min.M
Last changed   : 6/28/2019 8:29:21 AM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\WFY\wfy-3-60\wfy-3-60-N-35Me 2019-10-21 09-16-11\OD-3
               : -90-10-1ML-30min.M (Sequence Method)
Last changed   : 11/14/2019 6:37:36 PM by SYSTEM
               : (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

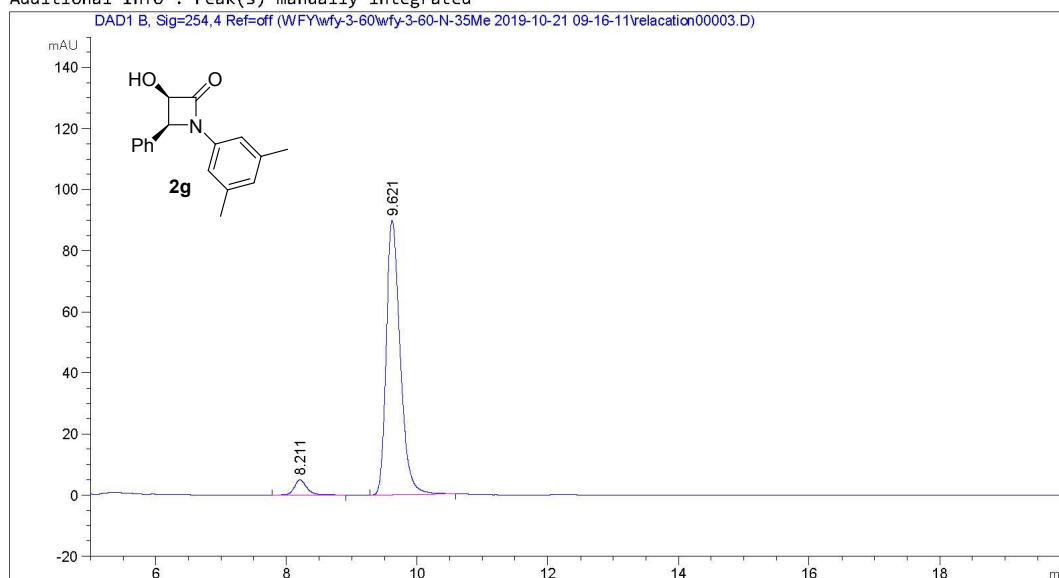
Sorted By      : Signal
Multiplier    : 1.0000
Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.448	BB	0.1855	5471.21289	451.12543	49.4161
2	9.929	BB	0.2241	5422.83057	367.69009	48.9791
3	11.363	BB	0.2457	88.44357	5.61560	0.7988

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    3
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P2-C-02
Injection Date  : 10/21/2019 9:58:16 AM      Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl
Acq. Method     : D:\ChemStation\1\Data\WFY\wfy-3-60\wfy-3-60-N-35Me 2019-10-21 09-16-11\OD-3
                  -90-10-1ML-30min.M
Last changed    : 6/28/2019 8:29:21 AM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\WFY\wfy-3-60\wfy-3-60-N-35Me 2019-10-21 09-16-11\OD-3
                  -90-10-1ML-30min.M (Sequence Method)
Last changed    : 11/14/2019 6:39:01 PM by SYSTEM
                  (modified after loading)
Additional Info  : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.211	BB	0.2059	68.27014	5.04445	4.8913
2	9.621	BB	0.2264	1327.46912	89.83994	95.1087



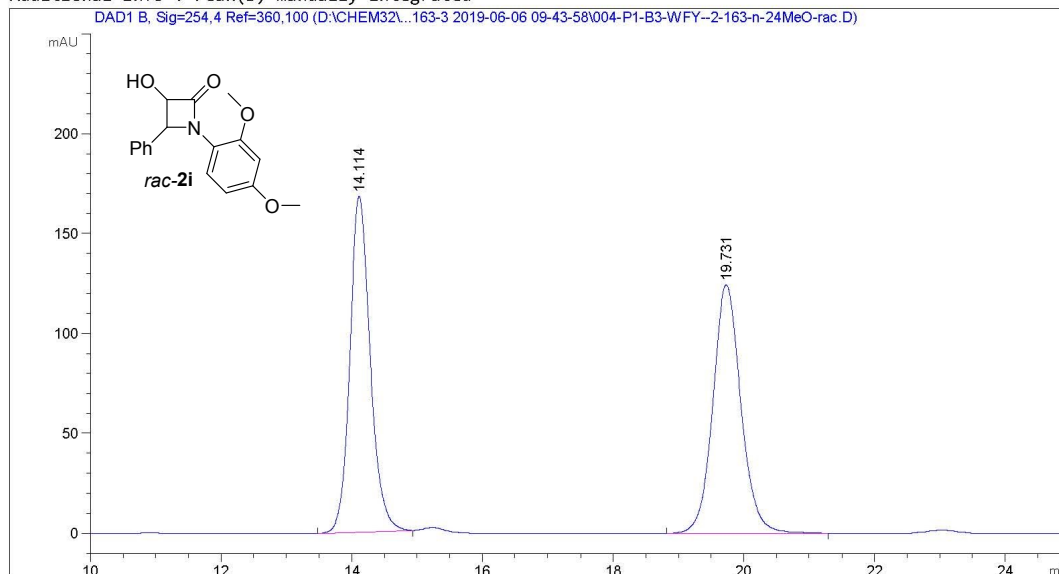




```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    4
Acq. Instrument : 1260-DAD                    Location  : P1-B-03
Injection Date  : 6/6/2019 11:01:58          Inj       :    1
                                           Inj Volume: 3.000 µl

Acq. Method    : D:\Chem32\1\Data\WFY\wfy-2-163\wfy-2-163-3 2019-06-06 09-43-58\IA-85-15-1ml
                -30min.M
Last changed   : 1/7/2019 21:25:37 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WFY\wfy-2-163\wfy-2-163-3 2019-06-06 09-43-58\IA-85-15-1ml
                -30min.M (Sequence Method)
Last changed   : 12/18/2019 21:08:51 by SYSTEM
                (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



=====  
 Area Percent Report  
 =====

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution      :      1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

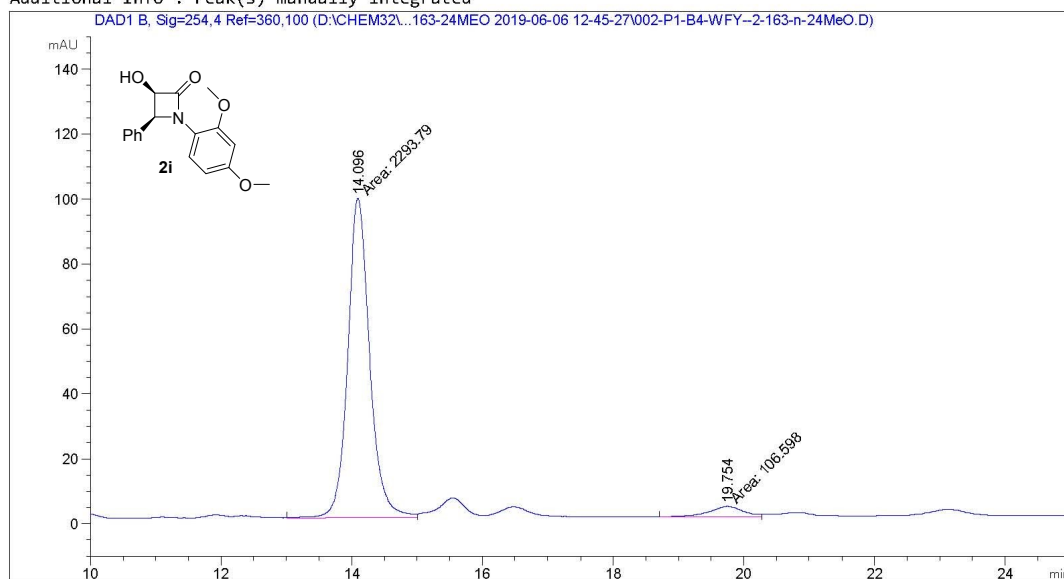
Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.114	BB	0.3278	3626.81567	168.06200	49.5057
2	19.731	BB	0.4550	3699.23828	124.36620	50.4943

Totals :                    7326.05396   292.42819

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    2
Acq. Instrument : 1260-DAD                    Location  : P1-B-04
Injection Date  : 6/6/2019 13:01:44          Inj       :    1
                                           Inj Volume: 3.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 10.000 µl
Acq. Method     : D:\Chem32\1\Data\WFY\wfy-2-163\wfy-2-163-24MeO 2019-06-06 12-45-27\IA-85-15
                  -1ml-30min.M
Last changed    : 1/7/2019 21:25:37 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WFY\wfy-2-163\wfy-2-163-24MeO 2019-06-06 12-45-27\IA-85-15
                  -1ml-30min.M (Sequence Method)
Last changed    : 12/18/2019 21:07:20 by SYSTEM
                  (modified after loading)
Additional Info  : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.096	MF	0.3885	2293.79004	98.40395	95.5591
2	19.754	MF	0.5817	106.59768	3.05441	4.4409

Totals : 2400.38772 101.45835

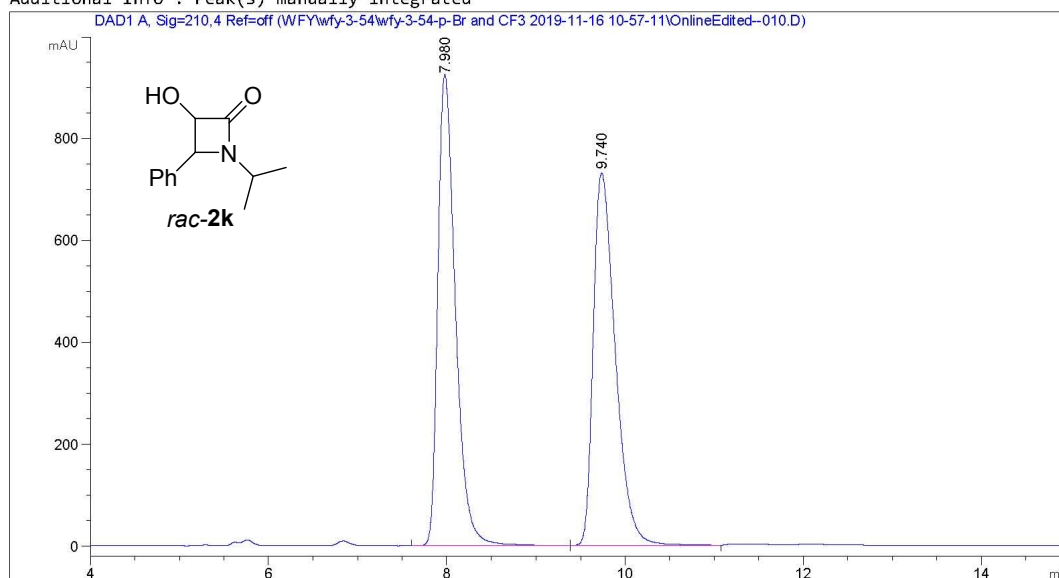




=====

Acq. Operator	: SYSTEM	Seq. Line	: 10
Sample Operator	: SYSTEM		
Acq. Instrument	: LC	Location	: P1-C-07
Injection Date	: 11/16/2019 2:15:39 PM	Inj	: 1
		Inj Volume	: 1.000 µl
		Different Inj Volume from Sample Entry!	Actual Inj Volume : 5.000 µl
Acq. Method	: D:\ChemStation\1\Data\Wfy\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-11\OD-3-90-10-1ML-30min.M		
Last changed	: 11/16/2019 12:28:44 PM by SYSTEM		
Analysis Method	: D:\ChemStation\1\Data\Wfy\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-11\OD-3-90-10-1ML-30min.M (Sequence Method)		
Last changed	: 11/19/2019 9:27:35 PM by SYSTEM		
	(modified after loading)		

Additional Info : Peak(s) manually integrated



=====  
Area Percent Report  
=====

Sorted By : Signal  
Multiplier : 1.0000  
Dilution : 1.0000  
Use Multiplier & Dilution Factor with ISTDs

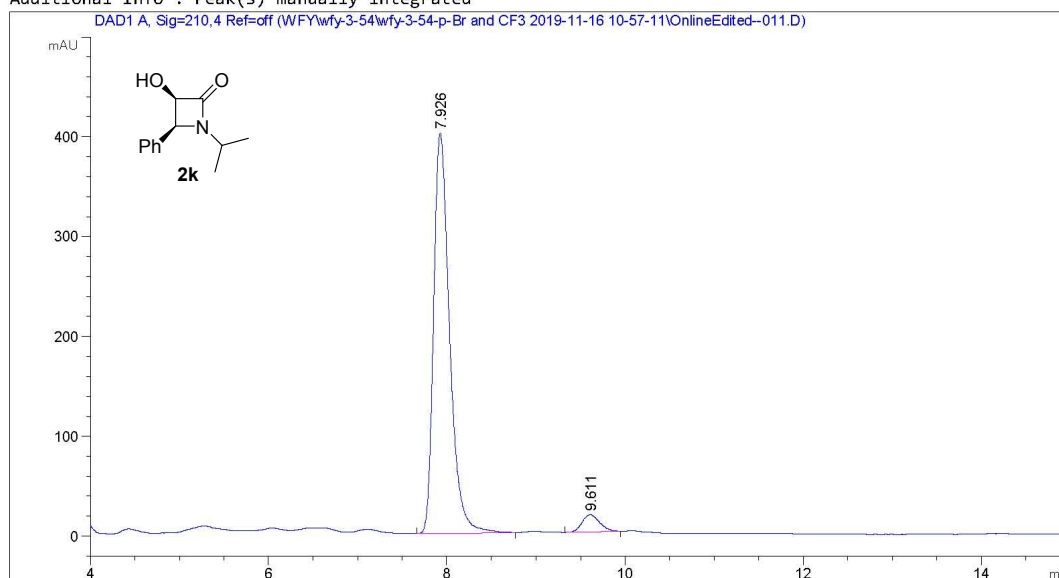
Signal 1: DAD1 A, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.980	BB	0.2087	1.25994e4	926.00726	50.2652
2	9.740	BB	0.2628	1.24664e4	731.55688	49.7348

=====

Acq. Operator	: SYSTEM	Seq. Line	: 11
Sample Operator	: SYSTEM		
Acq. Instrument	: LC	Location	: P1-C-08
Injection Date	: 11/16/2019 2:36:33 PM	Inj	: 1
		Inj Volume	: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 8.000 µl			
Acq. Method	: D:\ChemStation\1\Data\Wfy\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-11\OD-3-90-10-1ML-30min.M		
Last changed	: 11/16/2019 12:28:44 PM by SYSTEM		
Analysis Method	: D:\ChemStation\1\Data\Wfy\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-11\OD-3-90-10-1ML-30min.M (Sequence Method)		
Last changed	: 11/19/2019 9:28:29 PM by SYSTEM		
	(modified after loading)		

Additional Info : Peak(s) manually integrated



=====  
Area Percent Report  
=====

Sorted By : Signal  
Multiplier : 1.0000  
Dilution : 1.0000  
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.926	BB	0.1880	4944.41602	400.47427	95.4395
2	9.611	BB	0.2116	236.26707	17.26842	4.5605



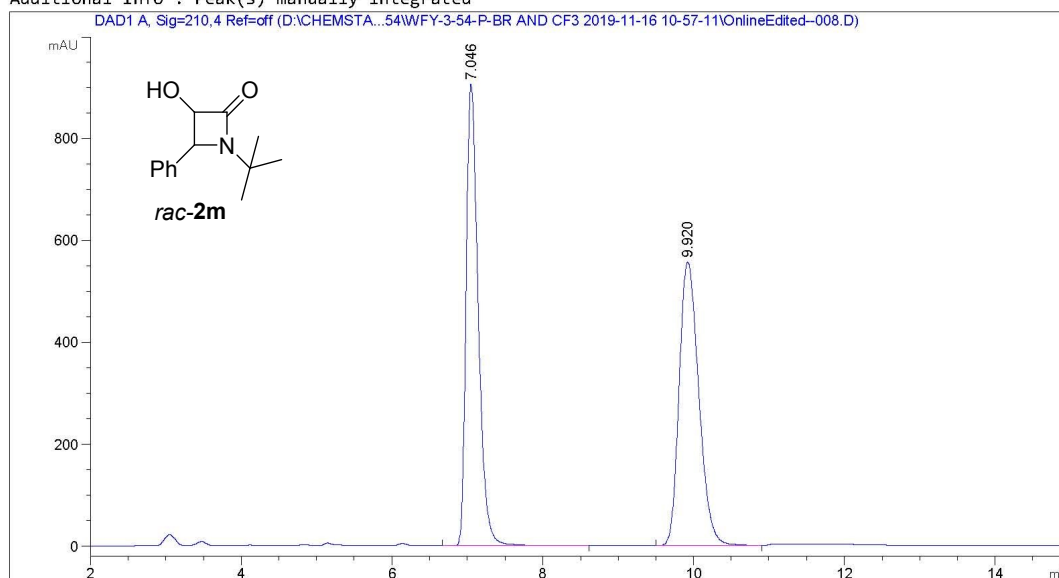




```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    8
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P1-C-05
Injection Date  : 11/16/2019 1:33:48 PM      Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl
Acq. Method     : D:\ChemStation\1\Data\Wfy\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-
                  11\OD-3-90-10-1ML-30min.M
Last changed    : 11/16/2019 12:28:44 PM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\Wfy\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-
                  11\OD-3-90-10-1ML-30min.M (Sequence Method)
Last changed    : 11/19/2019 9:24:45 PM by SYSTEM
                  (modified after loading)
  
```

Additional Info : Peak(s) manually integrated



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 A, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.046	BV R	0.1718	1.00922e4	906.98041	50.1066
2	9.920	BB	0.2826	1.00492e4	556.80591	49.8934

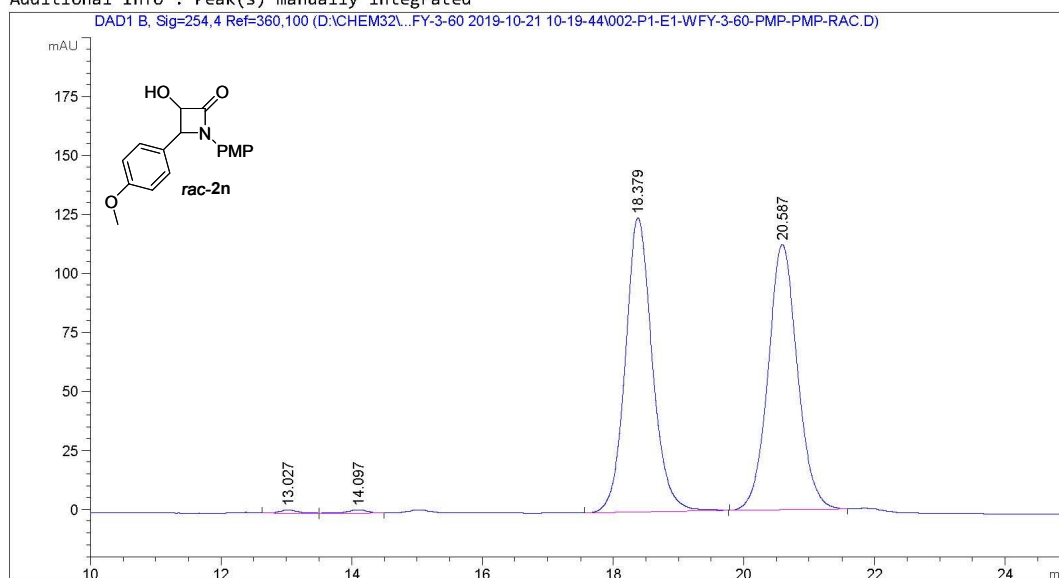


Data File D:\CHEM32\...-3-60\WfY-3-60 2019-10-21 10-19-44\002-P1-E1-WfY-3-60-PMP-PMP-RAC.D  
 Sample Name: WfY-3-60-PMP-PMP-RAC

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    2
Acq. Instrument : 1260-DAD                    Location  : P1-E-01
Injection Date  : 10/21/2019 10:35:59        Inj       :    1
                                                    Inj Volume: 3.000 µl

Acq. Method     : D:\Chem32\1\Data\WfY\WfY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-
30min.M
Last changed    : 1/7/2019 21:25:37 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WfY\WfY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-
30min.M (Sequence Method)
Last changed    : 11/15/2019 16:15:32 by SYSTEM
(modified after loading)
Additional Info  : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

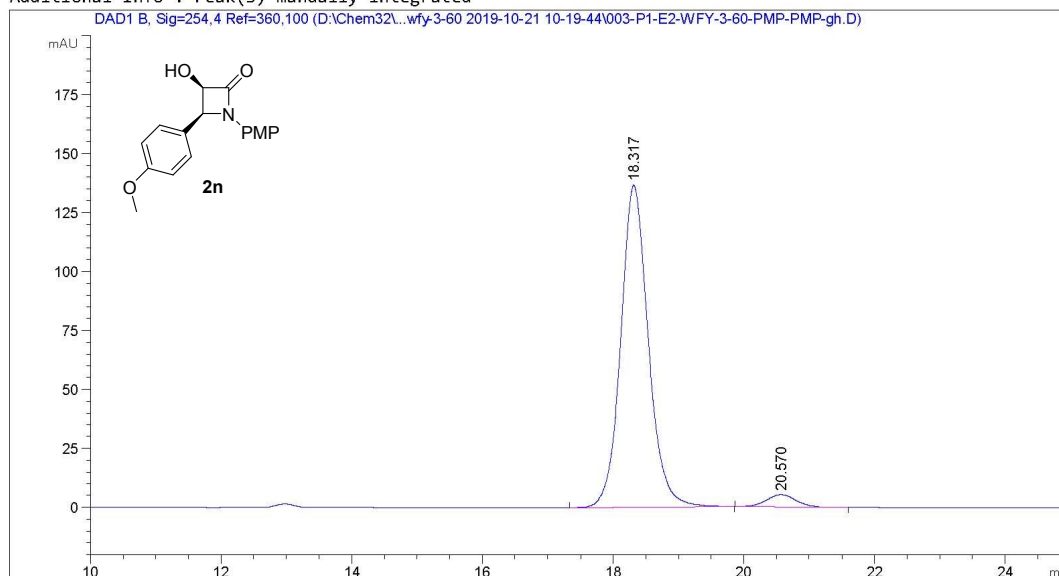
Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.027	BB	0.2796	24.73135	1.32593	0.3481
2	14.097	BB	0.2996	26.97620	1.31347	0.3797
3	18.379	BB	0.4339	3568.28564	124.67841	50.2229
4	20.587	BB	0.4738	3484.90698	112.36911	49.0493

Data File D:\Chem32\...Y-3-60\wfy-3-60 2019-10-21 10-19-44\003-P1-E2-WFY-3-60-PMP-PMP-gh.D  
 Sample Name: WFY-3-60-PMP-PMP-gh

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    3
Acq. Instrument : 1260-DAD                    Location  : P1-E-02
Injection Date  : 10/21/2019 11:06:55        Inj       :    1
                                                Inj Volume: 3.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 10.000 µl
Acq. Method     : D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-30min.M
Last changed    : 1/7/2019 21:25:37 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-30min.M (Sequence Method)
Last changed    : 11/15/2019 16:15:32 by SYSTEM
                (modified after loading)
Additional Info  : Peak(s) manually integrated
  
```



=====  
 Area Percent Report  
 =====

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.317	BB	0.4438	4002.24438	136.59238	95.9853
2	20.570	BB	0.4814	167.40054	5.28678	4.0147

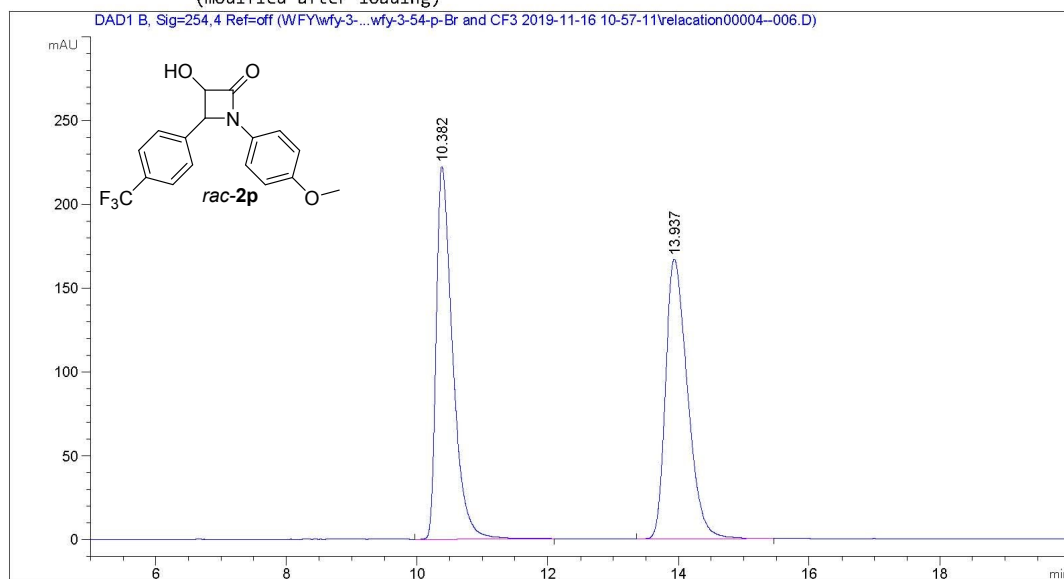
Totals :                    4169.64493 141.87916







```
=====
Acq. Operator   : SYSTEM                      Seq. Line :    6
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P1-C-03
Injection Date  : 11/16/2019 12:52:00 PM      Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl
Acq. Method    : D:\ChemStation\1\Data\WFY\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-
11\OD-3-90-10-1ML-30min.M
Last changed   : 11/16/2019 12:28:44 PM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\WFY\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-
11\OD-3-90-10-1ML-30min.M (Sequence Method)
Last changed   : 11/19/2019 9:07:39 PM by SYSTEM
                (modified after loading)
=====
```



=====  
Area Percent Report  
=====

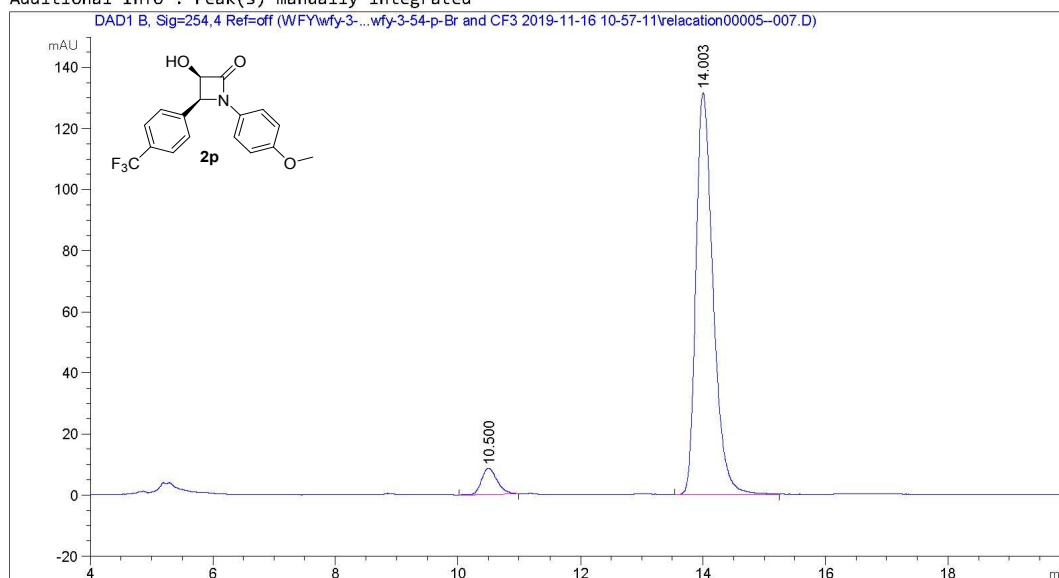
```
Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Use Multiplier & Dilution Factor with ISTDs
```

Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.044	BB	0.1174	10.27355	1.23083	0.1320
2	10.382	BB	0.2637	3887.44653	222.60713	49.9403
3	13.937	BB	0.3457	3886.46045	166.89842	49.9277

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    7
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P1-C-04
Injection Date  : 11/16/2019 1:12:54 PM      Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl
Acq. Method     : D:\ChemStation\1\Data\WFY\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-
                  11\OD-3-90-10-1ML-30min.M
Last changed    : 11/16/2019 12:28:44 PM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\WFY\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-
                  11\OD-3-90-10-1ML-30min.M (Sequence Method)
Last changed    : 11/19/2019 9:33:25 PM by SYSTEM
                  (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=off

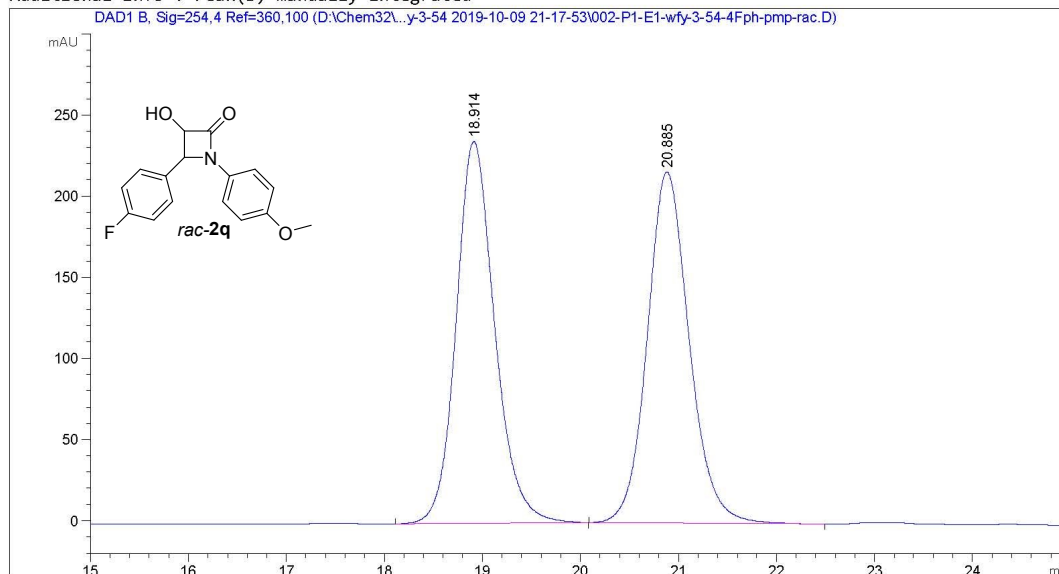
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.500	BB	0.2656	149.71025	8.57843	5.5519
2	14.003	BB	0.2980	2546.85449	131.45151	94.4481

Data File D:\Chem32\...3-54\wfy-3-54 2019-10-09 21-17-53\002-P1-E1-wfy-3-54-4Fph-pmp-rac.D  
 Sample Name: wfy-3-54-4Fph-pmp-rac

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    2
Acq. Instrument : 1260-DAD                   Location  : P1-E-01
Injection Date  : 10/9/2019 21:29:09        Inj       :    1
                                           Inj Volume: 3.000 µl

Acq. Method     : D:\Chem32\1\Data\Wfy\wfy-3-54\wfy-3-54 2019-10-09 21-17-53\IA-90-10-1.0ml-
                 30min.M
Last changed    : 7/24/2019 08:50:24 by SYSTEM
Analysis Method : D:\Chem32\1\Data\Wfy\wfy-3-54\wfy-3-54 2019-10-09 21-17-53\IA-90-10-1.0ml-
                 30min.M (Sequence Method)
Last changed    : 11/14/2019 18:51:36 by SYSTEM
                 (modified after loading)
Additional Info  : Peak(s) manually integrated
  
```



=====  
 Area Percent Report  
 =====

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

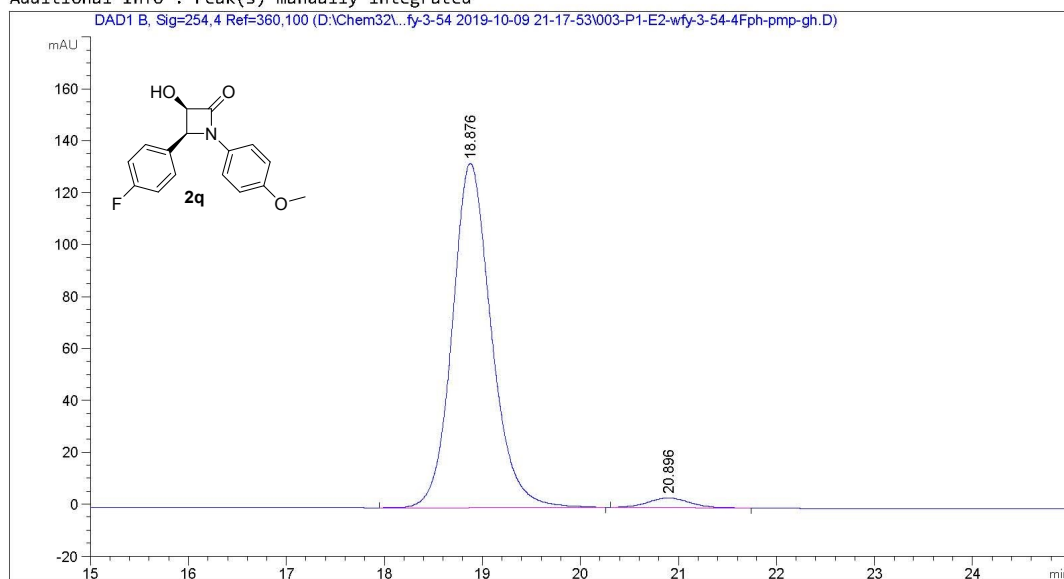
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.914	BB	0.4141	6413.76074	235.21341	49.9667
2	20.885	BB	0.4524	6422.32031	216.30305	50.0333

Totals :                    1.28361e4    451.51646

Data File D:\Chem32\...-3-54\wfy-3-54 2019-10-09 21-17-53\003-P1-E2-wfy-3-54-4Fph-pmp-gh.D  
 Sample Name: wfy-3-54-4Fph-pmp-gh

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    3
Acq. Instrument : 1260-DAD                    Location  : P1-E-02
Injection Date  : 10/9/2019 22:00:04         Inj       :    1
                                           Inj Volume: 3.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl
Acq. Method     : D:\Chem32\1\Data\Wfy\wfy-3-54\wfy-3-54 2019-10-09 21-17-53\IA-90-10-1.0ml-
                 30min.M
Last changed    : 7/24/2019 08:50:24 by SYSTEM
Analysis Method : D:\Chem32\1\Data\Wfy\wfy-3-54\wfy-3-54 2019-10-09 21-17-53\IA-90-10-1.0ml-
                 30min.M (Sequence Method)
Last changed    : 11/14/2019 18:52:03 by SYSTEM
                 (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.876	BB	0.4200	3659.78003	132.60747	97.1604
2	20.896	BB	0.4458	106.95877	3.67228	2.8396

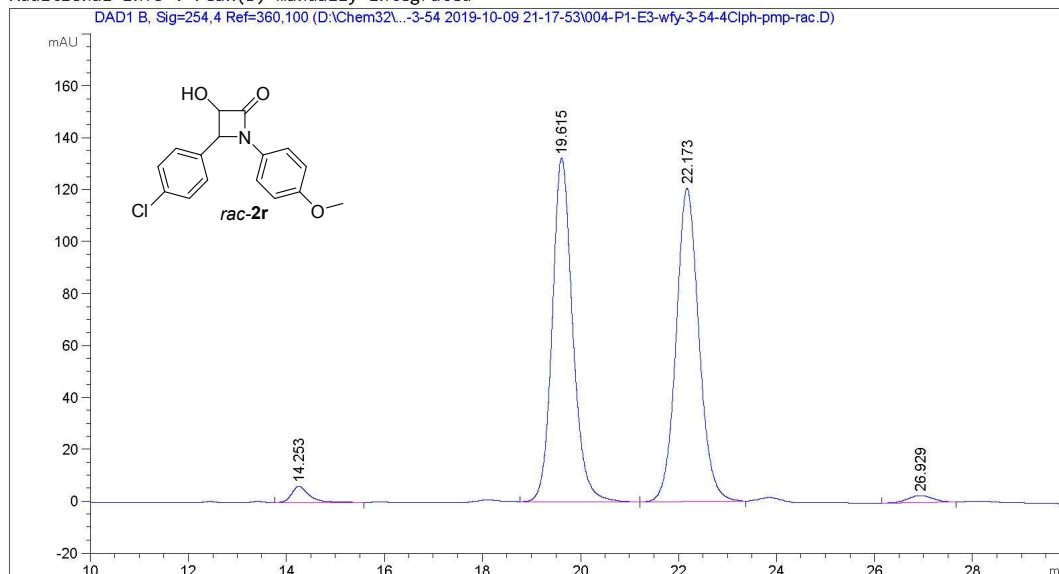
Totals : 3766.73880 136.27974

Data File D:\Chem32\...-54\wfy-3-54 2019-10-09 21-17-53\004-P1-E3-wfy-3-54-4Clph-pmp-rac.D  
 Sample Name: wfy-3-54-4Clph-pmp-rac

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    4
Acq. Instrument : 1260-DAD                    Location  : P1-E-03
Injection Date  : 10/9/2019 22:30:58         Inj       :    1
                                           Inj Volume: 3.000 µl

Acq. Method     : D:\Chem32\1\Data\Wfy\wfy-3-54\wfy-3-54 2019-10-09 21-17-53\IA-90-10-1.0ml-
                 30min.M
Last changed    : 7/24/2019 08:50:24 by SYSTEM
Analysis Method : D:\Chem32\1\Data\Wfy\wfy-3-54\wfy-3-54 2019-10-09 21-17-53\IA-90-10-1.0ml-
                 30min.M (Sequence Method)
Last changed    : 11/14/2019 18:53:01 by SYSTEM
                 (modified after loading)
Additional Info  : Peak(s) manually integrated
  
```



=====  
 Area Percent Report  
 =====

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

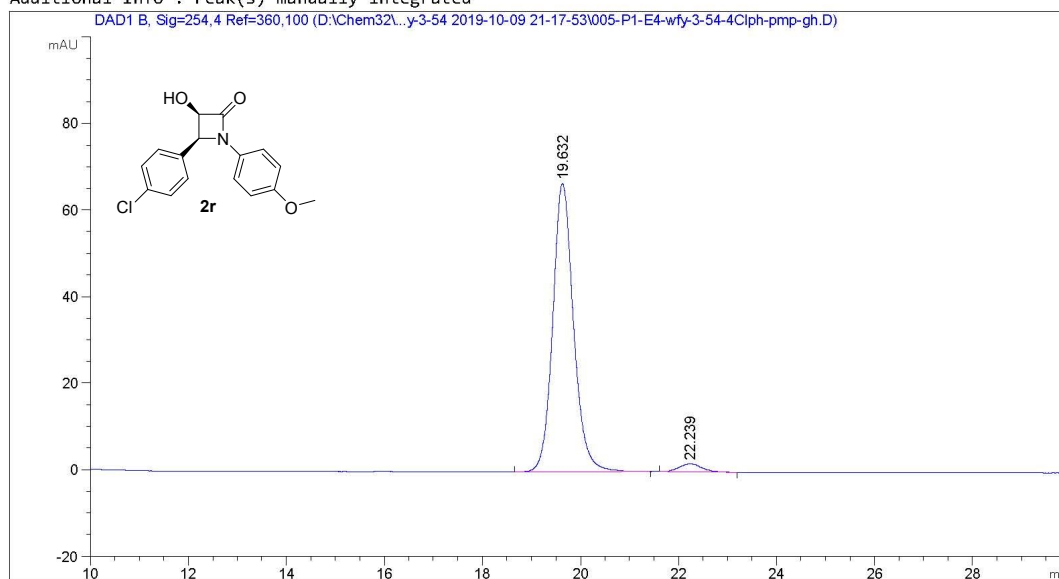
Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.253	BB	0.3633	151.60417	6.15692	1.9001
2	19.615	BB	0.4403	3837.85669	132.36440	48.1020
3	22.173	BB	0.4924	3893.10571	120.66217	48.7944
4	26.929	BB	0.5075	96.01798	2.73532	1.2034

Data File D:\Chem32\...3-54\wfy-3-54 2019-10-09 21-17-53\005-P1-E4-wfy-3-54-4Clph-pmp-gh.D  
 Sample Name: wfy-3-54-4Clph-pmp-gh

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    5
Acq. Instrument : 1260-DAD                    Location  : P1-E-04
Injection Date  : 10/9/2019 23:01:54         Inj       :    1
                                           Inj Volume: 3.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl
Acq. Method     : D:\Chem32\1\Data\WfY\wfy-3-54\wfy-3-54 2019-10-09 21-17-53\IA-90-10-1.0ml-
                 30min.M
Last changed    : 7/24/2019 08:50:24 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WfY\wfy-3-54\wfy-3-54 2019-10-09 21-17-53\IA-90-10-1.0ml-
                 30min.M (Sequence Method)
Last changed    : 11/14/2019 18:54:22 by SYSTEM
                 (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

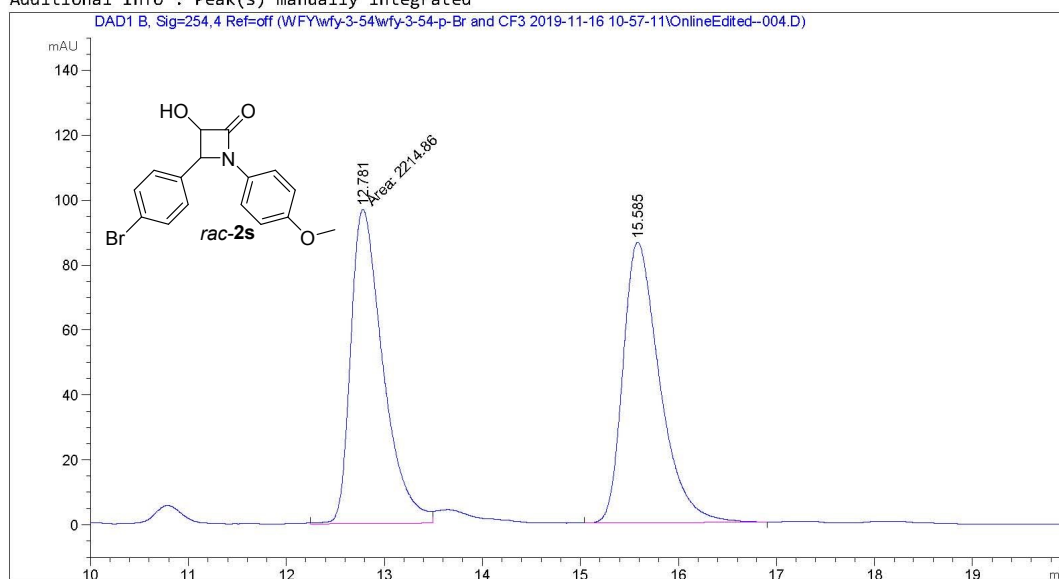
Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.632	BB	0.4484	1965.79492	66.59377	97.0672
2	22.239	BB	0.4506	59.39445	1.87994	2.9328

Totals :                    2025.18938   68.47372

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    4
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P1-C-01
Injection Date  : 11/16/2019 12:10:12 PM    Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 10.000 µl
Acq. Method     : D:\ChemStation\1\Data\WFY\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-
                  11\OD-3-90-10-1ML-30min.M
Last changed    : 11/16/2019 12:28:44 PM by SYSTEM
                  (modified after loading)
Analysis Method : D:\ChemStation\1\Data\WFY\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-
                  11\OD-3-90-10-1ML-30min.M (Sequence Method)
Last changed    : 11/19/2019 9:05:48 PM by SYSTEM
                  (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

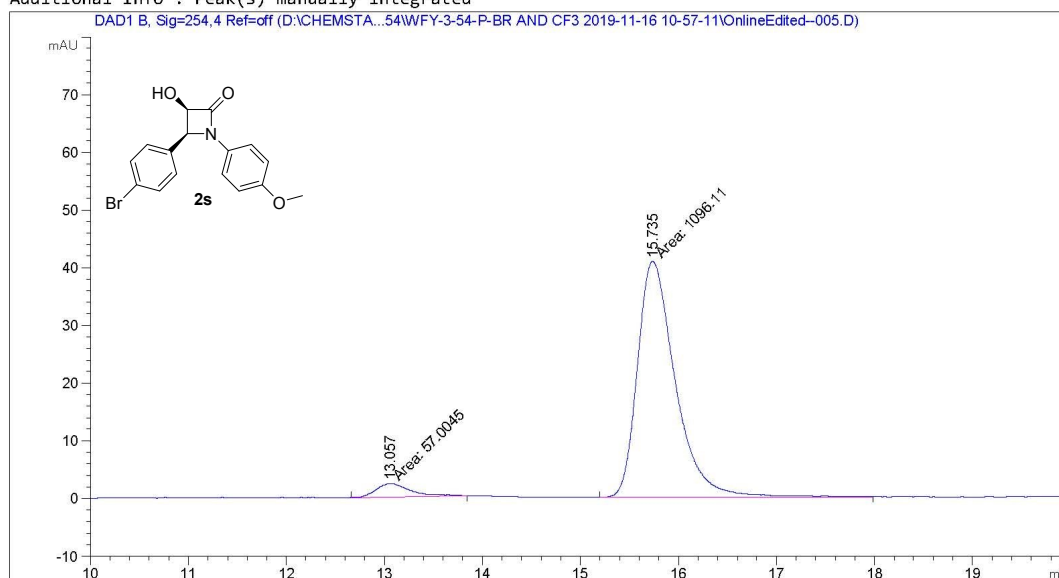
Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.781	MM	0.3814	2214.85864	96.79659	49.6119
2	15.585	BB	0.3958	2249.51392	86.39938	50.3881

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    5
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P1-C-02
Injection Date  : 11/16/2019 12:31:06 PM    Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 3.000 µl
Acq. Method     : D:\ChemStation\1\Data\Wfy\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-
                  11\OD-3-90-10-1ML-30min.M
Last changed    : 11/16/2019 12:28:44 PM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\Wfy\wfy-3-54\wfy-3-54-p-Br and CF3 2019-11-16 10-57-
                  11\OD-3-90-10-1ML-30min.M (Sequence Method)
Last changed    : 11/19/2019 9:04:36 PM by SYSTEM
                  (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.057	MM	0.4088	57.00448	2.32401	4.9435
2	15.735	MM	0.4467	1096.11255	40.89233	95.0565



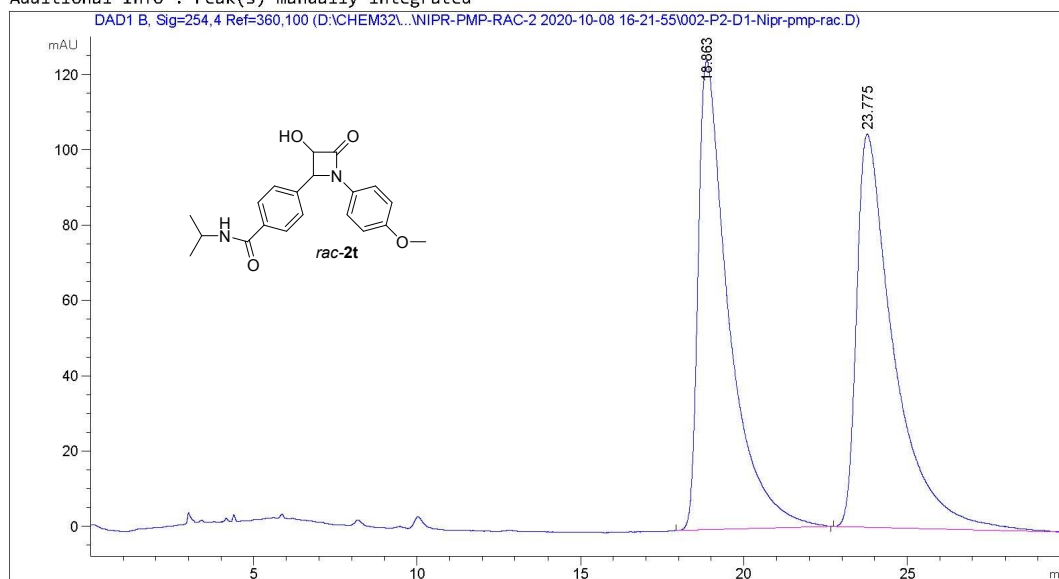
=====

Acq. Operator	: SYSTEM	Seq. Line	: 2
Acq. Instrument	: 1260-DAD	Location	: P2-D-01
Injection Date	: 10/8/2020 16:28:13	Inj	: 1
		Inj Volume	: 3.000 µl

Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl

Acq. Method	: D:\Chem32\1\Data\WfY\wfy-4-paper2\Nipr-pmp-rac-2 2020-10-08 16-21-55\IA-85-15-1ml-30min.M
Last changed	: 1/7/2019 21:25:37 by SYSTEM
Analysis Method	: d:\Chem32\1\Data\WfY\wfy-4-paper2\Nipr-pmp-rac-2 2020-10-08 16-21-55\IA-85-15-1ml-30min.M (Sequence Method)
Last changed	: 10/9/2020 08:54:53 by SYSTEM (modified after loading)

Additional Info : Peak(s) manually integrated



=====  
Area Percent Report  
=====

Sorted By : Signal  
Multiplier : 1.0000  
Dilution : 1.0000  
Use Multiplier & Dilution Factor with ISTDs

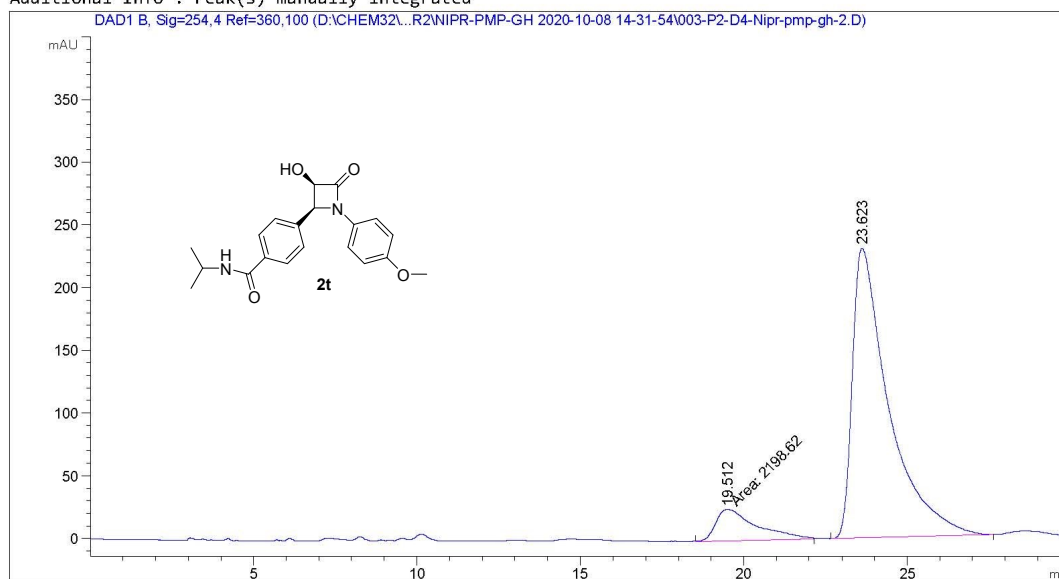
Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.863	BB	0.9134	8067.21631	124.59436	49.5297
2	23.775	BBA	1.1272	8220.42383	104.33062	50.4703

Totals : 1.62876e4 228.92498

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    3
Acq. Instrument : 1260-DAD                    Location  : P2-D-04
Injection Date  : 10/8/2020 15:14:01         Inj       :    1
                                           Inj Volume: 3.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 8.000 µl
Acq. Method    : D:\Chem32\1\Data\WfY\wfy-4-paper2\Nipr-pmp-gh 2020-10-08 14-31-54\IA-85-15-
                1ml-30min.M
Last changed   : 1/7/2019 21:25:37 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WfY\wfy-4-paper2\Nipr-pmp-gh 2020-10-08 14-31-54\IA-85-15-
                1ml-30min.M (Sequence Method)
Last changed   : 10/8/2020 16:20:01 by SYSTEM
                (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.512	MM	1.4493	2198.61914	25.28300	10.9829
2	23.623	BB	1.0702	1.78199e4	230.95465	89.0171

Totals :                    2.00185e4    256.23765

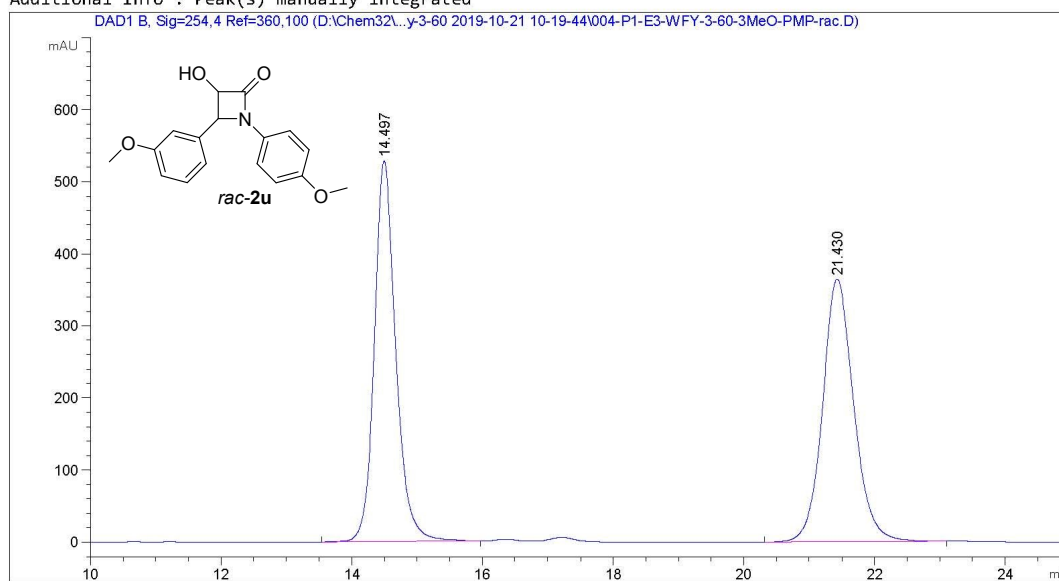
Data File D:\Chem32\...3-60\wfy-3-60 2019-10-21 10-19-44\004-P1-E3-WFY-3-60-3MeO-PMP-rac.D  
 Sample Name: WFY-3-60-3MeO-PMP-rac

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    4
Acq. Instrument : 1260-DAD                    Location  : P1-E-03
Injection Date  : 10/21/2019 11:37:51        Inj       :    1
                                           Inj Volume: 3.000 µl

Acq. Method     : D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-
30min.M
Last changed    : 1/7/2019 21:25:37 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-
30min.M (Sequence Method)
Last changed    : 11/15/2019 16:17:57 by SYSTEM
                 (modified after loading)

Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

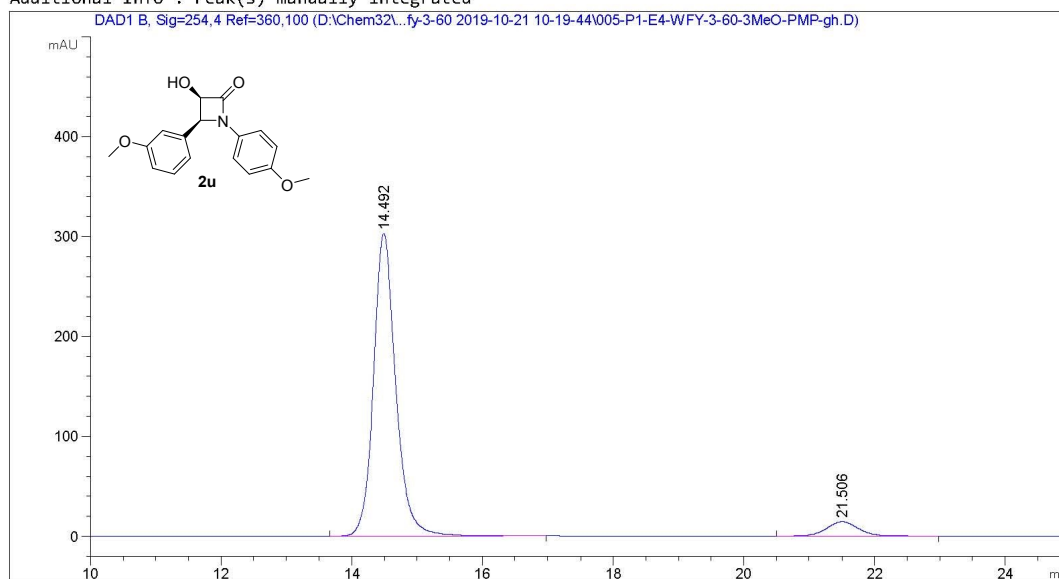
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.497	BB	0.3409	1.19822e4	527.68707	49.9497
2	21.430	BB	0.5007	1.20064e4	364.09576	50.0503

Totals :                                    2.39886e4    891.78284

Data File D:\Chem32\...-3-60\wfy-3-60 2019-10-21 10-19-44\005-P1-E4-WFY-3-60-3MeO-PMP-gh.D  
 Sample Name: WFY-3-60-3MeO-PMP-gh

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    5
Acq. Instrument : 1260-DAD                    Location  : P1-E-04
Injection Date  : 10/21/2019 12:08:47        Inj       :    1
                                           Inj Volume: 3.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 10.000 µl
Acq. Method     : D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-
                                           30min.M
Last changed    : 1/7/2019 21:25:37 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-
                                           30min.M (Sequence Method)
Last changed    : 11/15/2019 16:18:37 by SYSTEM
                                           (modified after loading)
Additional Info  : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.492	BB	0.3559	7166.12158	302.98111	93.4674
2	21.506	BB	0.5172	500.85095	14.63854	6.5326

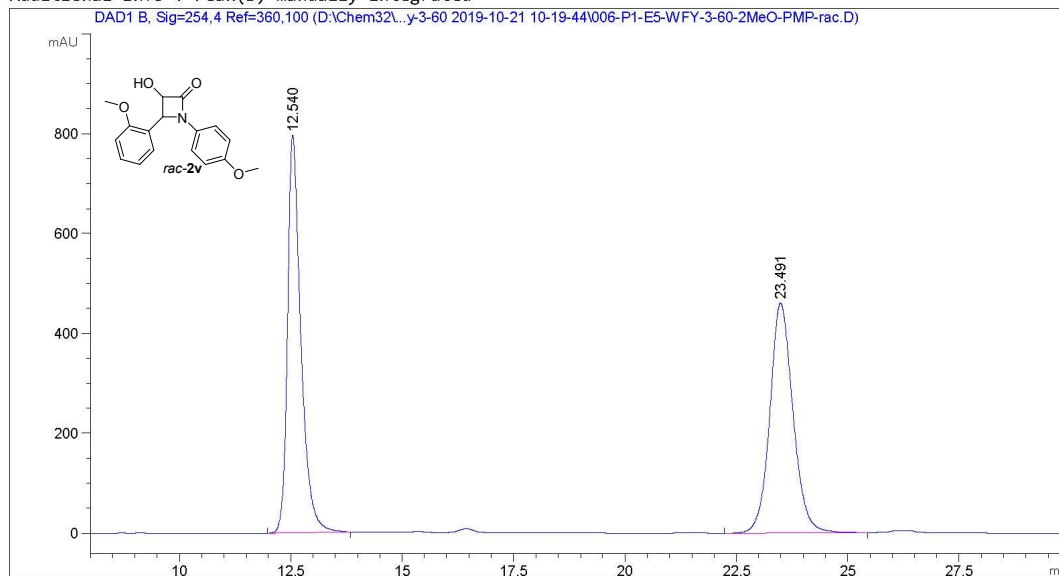
Totals : 7666.97253 317.61965

Data File D:\Chem32\...3-60\wfy-3-60 2019-10-21 10-19-44\006-P1-E5-WFY-3-60-2MeO-PMP-rac.D  
 Sample Name: WFY-3-60-2MeO-PMP-rac

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    6
Acq. Instrument : 1260-DAD                    Location  : P1-E-05
Injection Date  : 10/21/2019 12:39:44        Inj       :    1
                                           Inj Volume: 3.000 µl

Acq. Method     : D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-
                                           30min.M
Last changed    : 1/7/2019 21:25:37 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WFY\WFY-3-60\wfy-3-60 2019-10-21 10-19-44\IA-85-15-1ml-
                                           30min.M (Sequence Method)
Last changed    : 11/15/2019 16:20:12 by SYSTEM
                                           (modified after loading)
Additional Info  : Peak(s) manually integrated
  
```



=====  
 Area Percent Report  
 =====

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution      :      1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.540	BB	0.3067	1.64155e4	795.71454	49.7787
2	23.491	BB	0.5459	1.65615e4	460.17523	50.2213

Totals :                                    3.29771e4  1255.88977







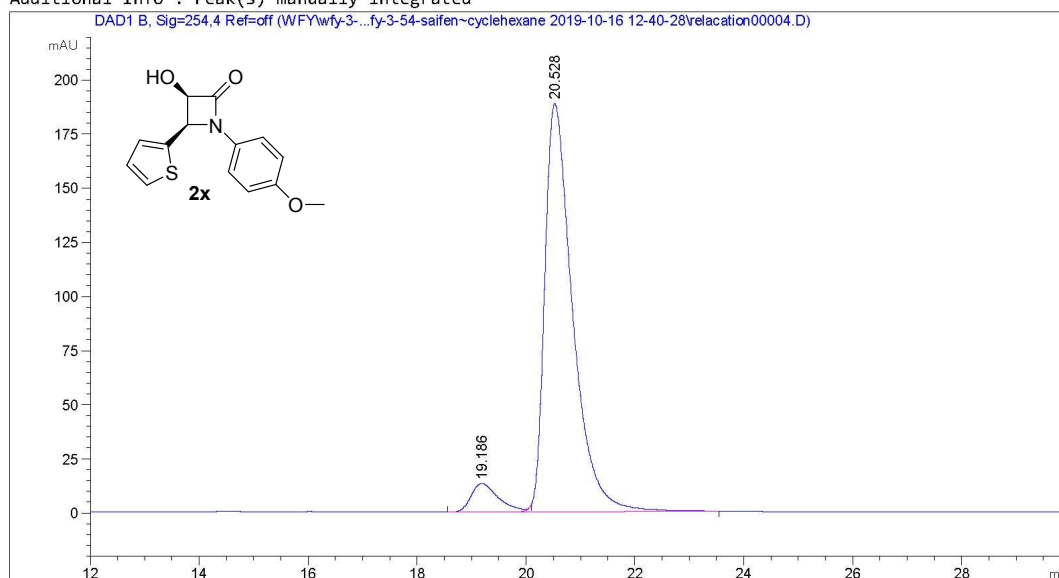




=====

Acq. Operator	: SYSTEM	Seq. Line	: 4
Sample Operator	: SYSTEM		
Acq. Instrument	: LC	Location	: P2-C-03
Injection Date	: 10/16/2019 1:53:26 PM	Inj	: 1
		Inj Volume	: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl			
Acq. Method	: D:\ChemStation\1\Data\WFY\wfy-3-54\wfy-3-54-saifen~cyclohexane 2019-10-16 12-40-28\OD-3-90-10-1ML-30min.M		
Last changed	: 6/28/2019 8:29:21 AM by SYSTEM		
Analysis Method	: D:\ChemStation\1\Data\WFY\wfy-3-54\wfy-3-54-saifen~cyclohexane 2019-10-16 12-40-28\OD-3-90-10-1ML-30min.M (Sequence Method)		
Last changed	: 11/14/2019 2:39:57 PM by SYSTEM		
	(modified after loading)		

Additional Info : Peak(s) manually integrated



=====  
Area Percent Report  
=====

Sorted By : Signal  
Multiplier : 1.0000  
Dilution : 1.0000  
Use Multiplier & Dilution Factor with ISTDs

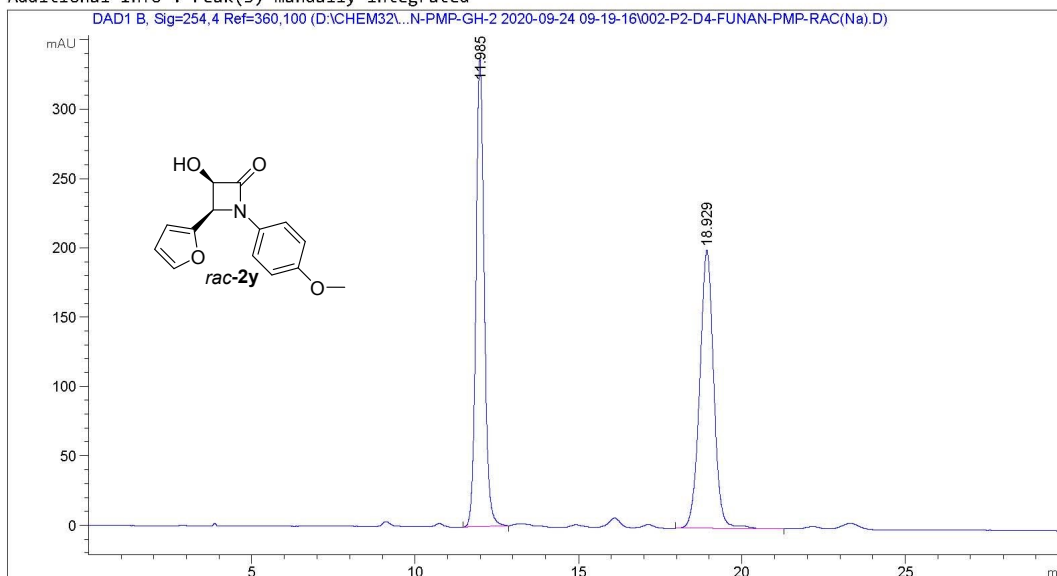
Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.186	BV E	0.5213	453.74026	13.12718	6.3606
2	20.528	VB R	0.5354	6679.85205	188.52869	93.6394

Data File D:\CHEM32\...\R2\FUNAN-PMP-GH-2 2020-09-24 09-19-16\002-P2-D4-FUNAN-PMP-RAC(Na).D  
 Sample Name: FUNAN-PMP-RAC(Na)

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    2
Acq. Instrument : 1260-DAD                    Location  : P2-D-04
Injection Date  : 9/24/2020 09:35:29         Inj       :    1
                                           Inj Volume: 3.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl
Acq. Method     : D:\Chem32\1\Data\WFY\wfy-4-paper2\FUNAN-PMP-GH-2 2020-09-24 09-19-16\IA-85-15-1ml-30min.M
Last changed    : 1/7/2019 21:25:37 by SYSTEM
Analysis Method : D:\Chem32\1\Data\WFY\wfy-4-paper2\FUNAN-PMP-GH-2 2020-09-24 09-19-16\IA-85-15-1ml-30min.M (Sequence Method)
Last changed    : 10/8/2020 16:15:27 by SYSTEM
                 (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.985	BB	0.2652	5874.34375	337.19861	49.5997
2	18.929	BB	0.4510	5969.17188	200.68852	50.4003

Totals : 1.18435e4 537.88713

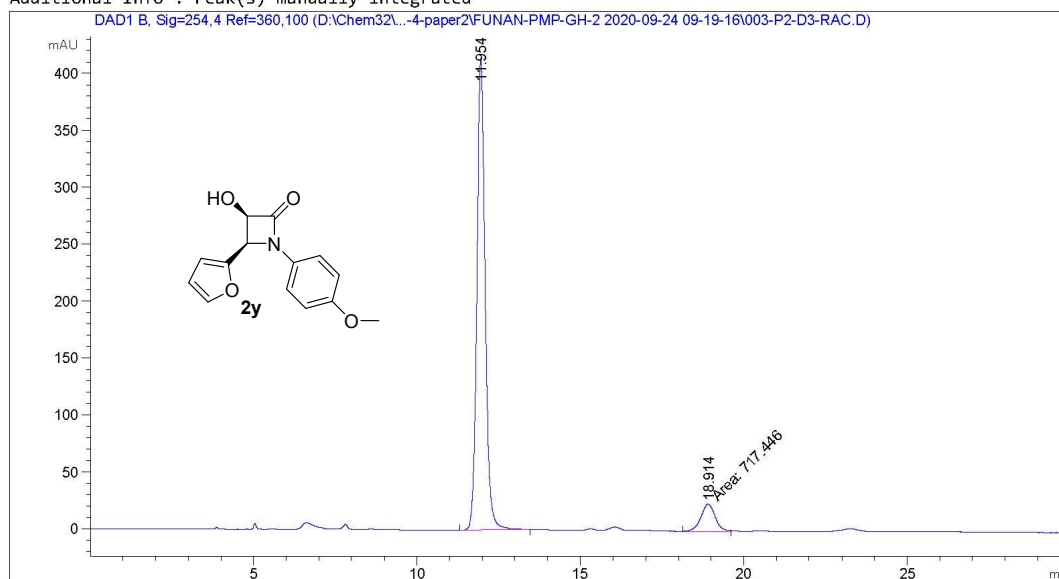
=====

Acq. Operator	: SYSTEM	Seq. Line	: 3
Acq. Instrument	: 1260-DAD	Location	: P2-D-03
Injection Date	: 9/24/2020 10:06:22	Inj	: 1
		Inj Volume	: 3.000 µl

Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl

Acq. Method	: D:\Chem32\1\Data\WFY\wfy-4-paper2\FUNAN-PMP-GH-2 2020-09-24 09-19-16\IA-85-15-1ml-30min.M
Last changed	: 1/7/2019 21:25:37 by SYSTEM
Analysis Method	: D:\Chem32\1\Data\WFY\wfy-4-paper2\FUNAN-PMP-GH-2 2020-09-24 09-19-16\IA-85-15-1ml-30min.M (Sequence Method)
Last changed	: 10/8/2020 16:15:27 by SYSTEM (modified after loading)

Additional Info : Peak(s) manually integrated



=====  
Area Percent Report  
=====

Sorted By : Signal  
Multiplier : 1.0000  
Dilution : 1.0000  
Use Multiplier & Dilution Factor with ISTDs

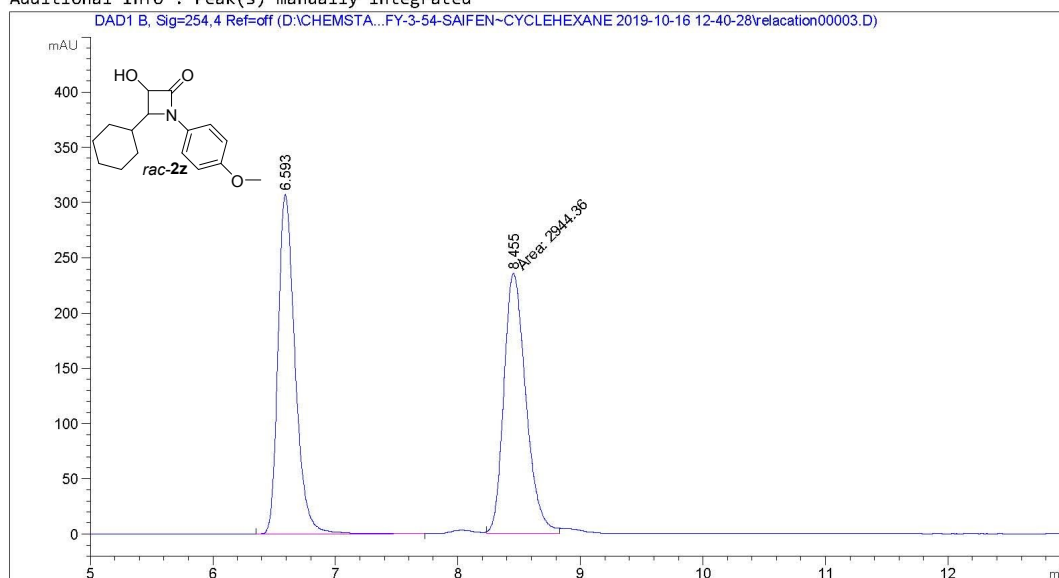
Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.954	BB	0.2665	7231.65381	412.65378	90.9745
2	18.914	MM	0.4986	717.44562	23.98012	9.0255

Totals : 7949.09943 436.63390

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    3
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P2-C-02
Injection Date  : 10/16/2019 1:22:35 PM      Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl
Acq. Method     : D:\ChemStation\1\Data\Wfy\wfy-3-54\wfy-3-54-saifen~cyclehexane 2019-10-16
                                           12-40-28\OD-3-90-10-1ML-30min.M
Last changed    : 6/28/2019 8:29:21 AM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\Wfy\wfy-3-54\wfy-3-54-saifen~cyclehexane 2019-10-16
                                           12-40-28\OD-3-90-10-1ML-30min.M (Sequence Method)
Last changed    : 11/14/2019 2:42:46 PM by SYSTEM
                                           (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

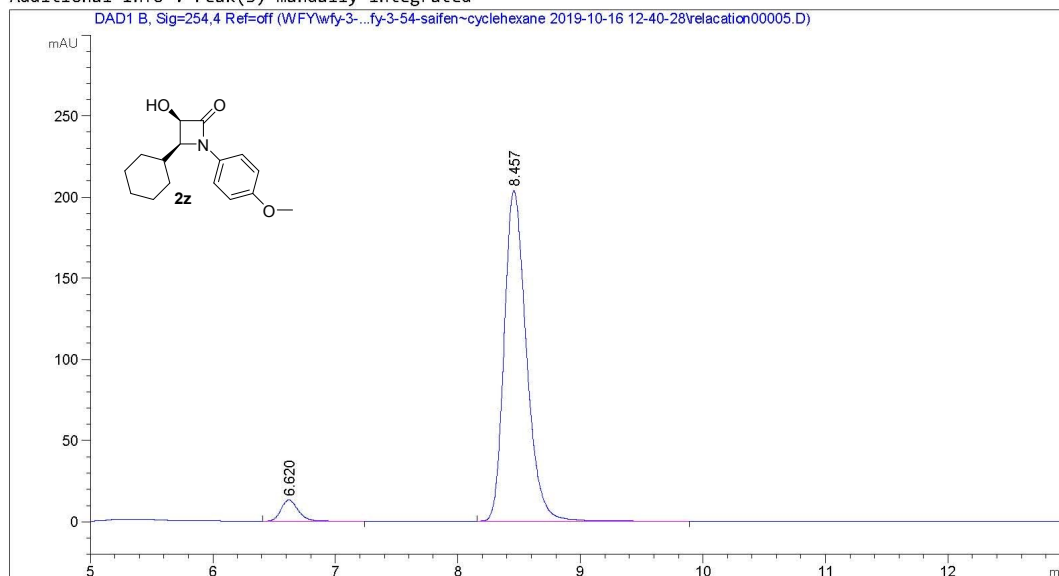
Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.593	BB	0.1465	2964.00635	307.38565	50.1662
2	8.455	FM	0.2082	2944.36230	235.64851	49.8338

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    5
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P2-C-04
Injection Date  : 10/16/2019 2:24:17 PM      Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl
Acq. Method     : D:\ChemStation\1\Data\Wfy\wfy-3-54\wfy-3-54-saifen~cyclohexane 2019-10-16
                  12-40-28\OD-3-90-10-1ML-30min.M
Last changed    : 6/28/2019 8:29:21 AM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\Wfy\wfy-3-54\wfy-3-54-saifen~cyclohexane 2019-10-16
                  12-40-28\OD-3-90-10-1ML-30min.M (Sequence Method)
Last changed    : 11/14/2019 2:44:23 PM by SYSTEM
                  (modified after loading)
  
```

Additional Info : Peak(s) manually integrated



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.620	BB	0.1489	129.83894	13.18489	4.8267
2	8.457	BB	0.1926	2560.18066	203.73653	95.1733

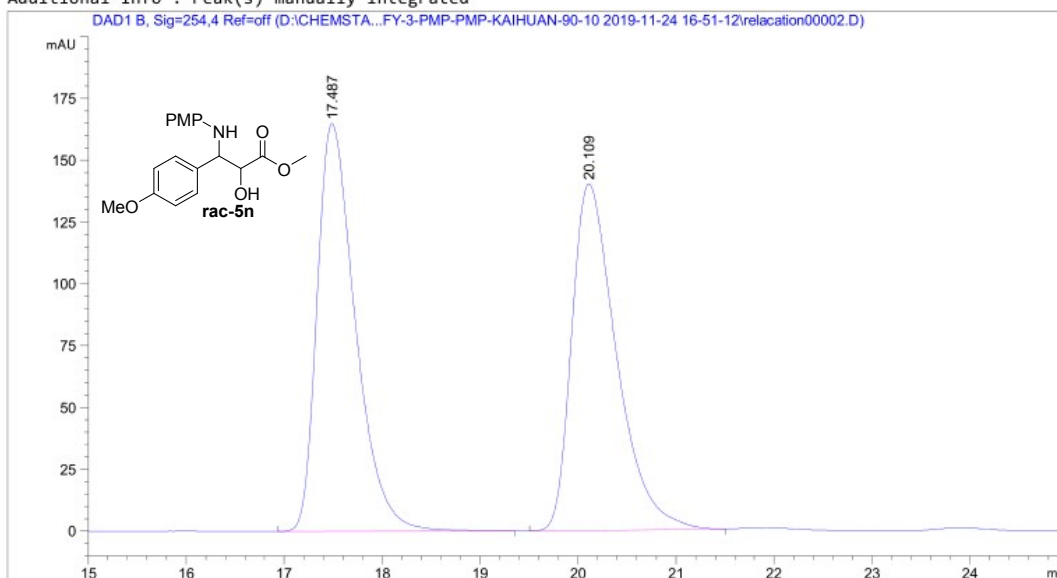






```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    2
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P1-C-08
Injection Date  : 11/24/2019 5:02:28 PM      Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl
Acq. Method     : D:\ChemStation\1\Data\WFY\WFY-3-PMPPMP-KAIHUAN\WFY-3-PMP-PMP-KAIHUAN-90-10
                 2019-11-24 16-51-12\OD-3-90-10-1ML-45min.M
Last changed    : 6/27/2019 12:37:27 PM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\WFY\WFY-3-PMPPMP-KAIHUAN\WFY-3-PMP-PMP-KAIHUAN-90-10
                 2019-11-24 16-51-12\OD-3-90-10-1ML-45min.M (Sequence Method)
Last changed    : 12/18/2019 9:04:07 PM by SYSTEM
                 (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



Area Percent Report

```

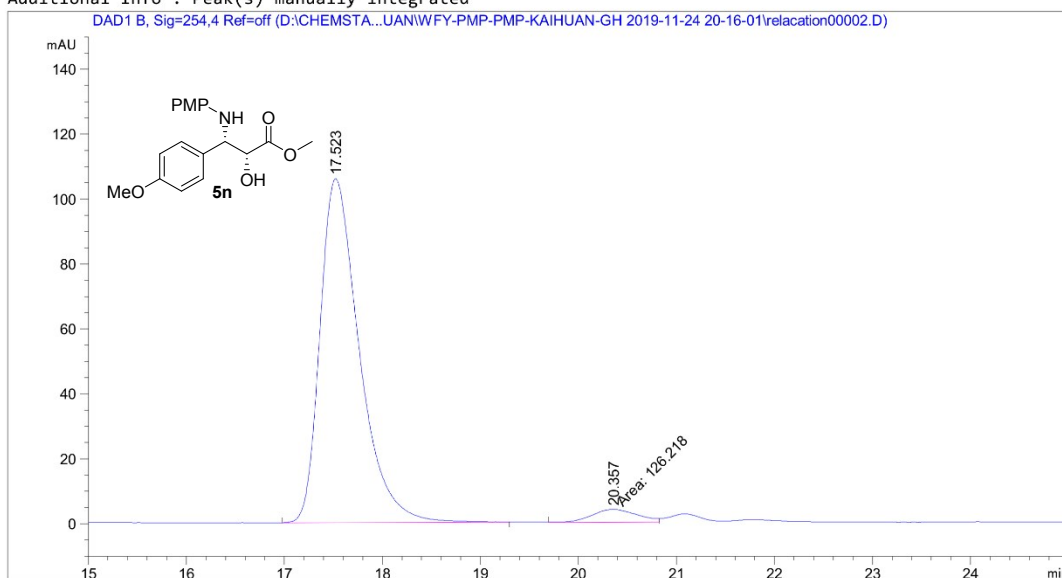
Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.487	BB	0.4133	4488.58545	165.02647	50.2252
2	20.109	BB	0.4842	4448.33887	140.20222	49.7748

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    2
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P1-C-09
Injection Date  : 11/24/2019 8:27:15 PM      Inj       :    1
                                           Inj Volume: 1.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 5.000 µl
Acq. Method    : D:\ChemStation\1\Data\WFY\WFY-3-PMPPMP-KAIHUAN\WFY-PMP-PMP-KAIHUAN-GH 2019-
11-24 20-16-01\OD-3-90-10-1ML-30min.M
Last changed   : 6/28/2019 8:29:21 AM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\WFY\WFY-3-PMPPMP-KAIHUAN\WFY-PMP-PMP-KAIHUAN-GH 2019-
11-24 20-16-01\OD-3-90-10-1ML-30min.M (Sequence Method)
Last changed   : 12/18/2019 9:06:26 PM by SYSTEM
                (modified after loading)
Additional Info : Peak(s) manually integrated
  
```



=====  
 Area Percent Report  
 =====

```

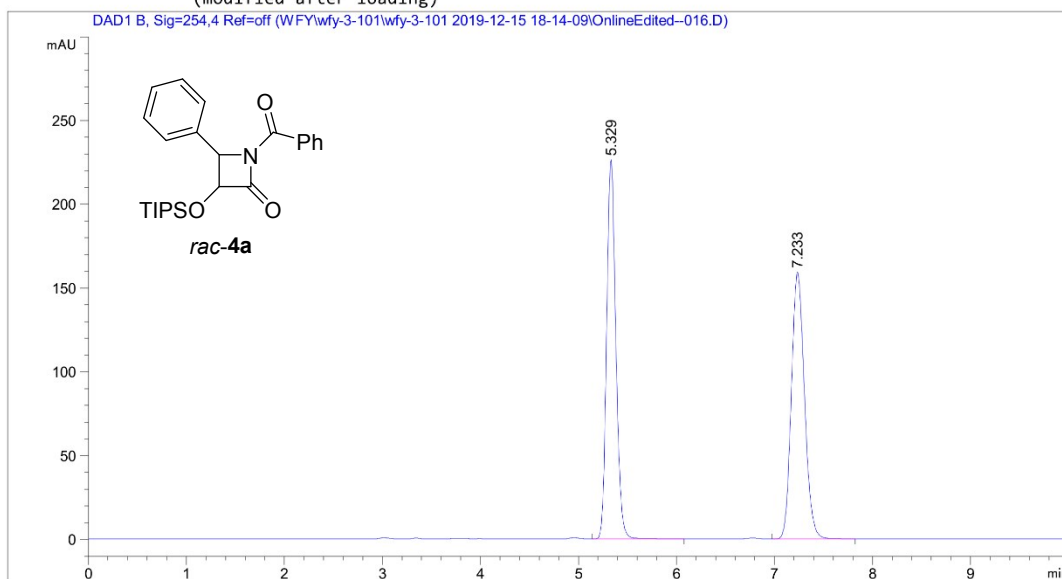
Sorted By      : Signal
Multiplier     : 1.0000
Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.523	BB	0.4197	2937.01978	105.86768	95.8796
2	20.357	MF	0.5393	126.21805	3.90050	4.1204

```

=====
Acq. Operator   : SYSTEM                      Seq. Line : 16
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P1-A-06
Injection Date  : 12/16/2019 12:54:42 AM    Inj       : 1
                                           Inj Volume: 2.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 3.000 µl
Acq. Method     : D:\ChemStation\1\Data\WfY\wfy-3-101\wfy-3-101 2019-12-15 18-14-09\OD-3-90-10-1ML-10min.M
Last changed    : 6/27/2019 12:37:27 PM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\WfY\wfy-3-101\wfy-3-101 2019-12-15 18-14-09\OD-3-90-10-1ML-10min.M (Sequence Method)
Last changed    : 12/18/2019 9:14:31 PM by SYSTEM
                (modified after loading)
  
```



=====  
 Area Percent Report  
 =====

```

Sorted By       : Signal
Multiplier      : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

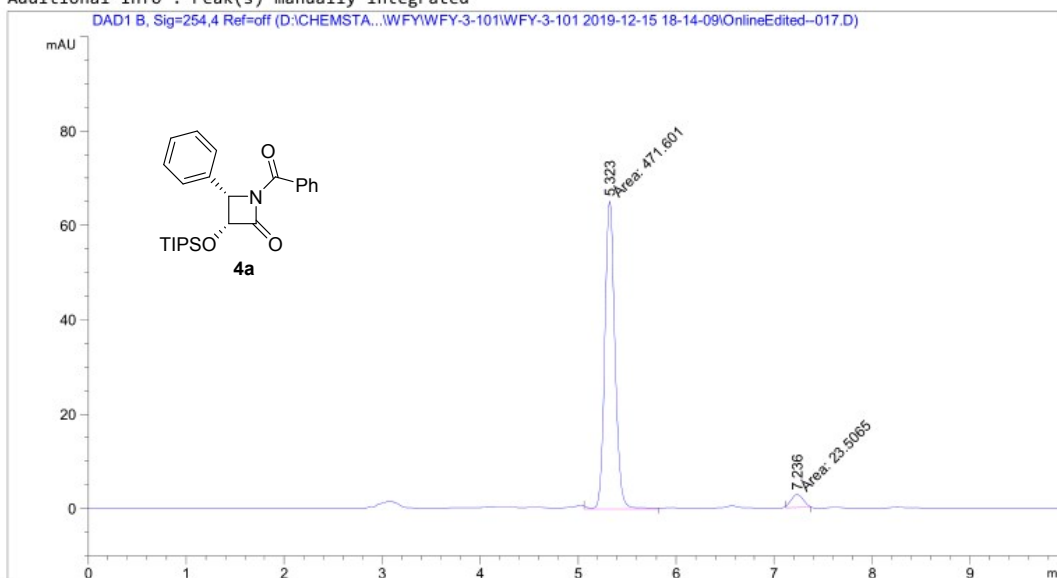
Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.329	BB	0.1002	1477.27869	226.32774	50.0182
2	7.233	BB	0.1442	1476.20435	159.22878	49.9818

Totals :                            2953.48303    385.55652

```

=====
Acq. Operator   : SYSTEM                      Seq. Line : 17
Sample Operator : SYSTEM
Acq. Instrument : LC                        Location  : P1-A-07
Injection Date  : 12/16/2019 1:05:36 AM      Inj       : 1
                                           Inj Volume: 2.000 µl
Different Inj Volume from Sample Entry! Actual Inj Volume : 8.000 µl
Acq. Method     : D:\ChemStation\1\Data\WfY\wfy-3-101\wfy-3-101 2019-12-15 18-14-09\OD-3-90-10-1ML-10min.M
Last changed    : 6/27/2019 12:37:27 PM by SYSTEM
Analysis Method : D:\ChemStation\1\Data\WfY\wfy-3-101\wfy-3-101 2019-12-15 18-14-09\OD-3-90-10-1ML-10min.M (Sequence Method)
Last changed    : 12/18/2019 9:13:44 PM by SYSTEM
                 (modified after loading)
Additional Info  : Peak(s) manually integrated
  
```



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 B, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.323	MM	0.1203	471.60080	65.35562	95.2522
2	7.236	MM	0.1420	23.50647	2.75968	4.7478

## 10. References.

- [1] a) N. Piens, H. Goossens, D. Hertsen, S. Deketelaere, L. Crul, L. Demeurisse, J. De Moor, E. Van den Broeck, K. Mollet, K. Van Hecke, V. Van Speybroeck, M. D'Hooghe, *Chemistry* **2017**, *23*, 18002-18009; b) A. L. Shaikh, B. K. Banik, *Helv. Chim. Acta* **2012**, *95*, 839-844.
- [2] S. S. Bari, M. S. Magtoof, A. Bhalla, *Monatshefte für Chemie - Chemical Monthly* **2010**, *141*, 987-991.
- [3] Y. Yang, M. Drolet, M. M. Kayser, *Tetrahedron: Asymmetry* **2005**, *16*, 2748-2753.
- [4] Y. Kawashima, M. Satoh, Y. Hatada, F. Hazato, Y. Nakashima, K. Sota, Taisho Pharmaceutical Co., Ltd., Japan . **1988**, p. 20 pp.
- [5] G. Cremonesi, P. Dalla Croce, C. La Rosa, *Helv. Chim. Acta* **2005**, *88*, 1580-1588.
- [6] J. A. Carr, T. F. Al-Azemi, T. E. Long, J.-Y. Shim, C. M. Coates, E. Turos, K. S. Bisht, *Tetrahedron* **2003**, *59*, 9147-9160.
- [7] Y. Paik, C. Yang, B. Metaferia, S. Tang, S. Bane, R. Ravindra, N. Shanker, A. A. Alcaraz, S. A. Johnson, J. Schaefer, R. D. O'Connor, L. Cegelski, J. P. Snyder, D. G. I. Kingston, *J. Am. Chem. Soc.* **2007**, *129*, 361-370.
- [8] L. Decuyper, J. Franceus, S. Dhaene, M. Debruyne, K. Vandoorne, N. Piens, G. Dewitte, T. Desmet, M. D'Hooghe, *ACS Omega* **2018**, *3*, 15235-15245.
- [9] R. K. Mishra, C. M. Coates, K. D. Revell, E. Turos, *Org. Lett.* **2007**, *9*, 575-578.
- [10] Y. Paik, C. Yang, B. Metaferia, S. Tang, S. Bane, R. Ravindra, N. Shanker, A. A. Alcaraz, S. A. Johnson, J. Schaefer, R. D. O'Connor, L. Cegelski, J. P. Snyder, D. G. Kingston, *J. Am. Chem. Soc.* **2007**, *129*, 361-370.
- [11] Yadav, Ram Naresh et al, *J. Indian Chem. Soc.*, **2018**, *95*, 1381-1384.
- [12] Anand, Naveen et al, *Tetrahedron: Asymmetry*, **2007**, *18*, 1059-1069.
- [13] Kayser, Margaret M. et al, *Can. J. Chem.*, **2002**, *80*, 796-800.