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Selective Nickel-catalyzed Fluoroalkylations of Olefins

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1. General comments

All reactions were carried out under an atmosphere of dry argon using standard Schlenk technique, Ace pressure tube or 300 mL autoclave equipped with 10 mL vials. Chemicals were purchased from *Aldrich*, *TCl*, *Alfa*, or *ABCR*. 3,3-Dimethylbutene was frozen and vacuumed for 3 times to get rid of H₂O and O₂ and stored in a Schlenk tube or stored in the fridge of the glovebox as received. Allylbenzene was distilled at 40 mbar with a 70 °C oil bath and stored in a Schlenk tube. Perfluorobenzene was transferred to a Schlenk tube in glovebox as received. Other solvents were dried and degassed and stored in ©Aldrich Sure/Stor flasks or Schlenk tubes under argon. NaO(*t*-Bu), NaH, and TMEDA were stored in glovebox as received. Other chemicals were used as received. Products were purified by fast chromatography using silica gel 60, 0.063-0.2 mm, 70-230 mesh (Merck). Multiplets of NMR were assigned as s (singlet), d (doublet), t (triplet), q (quartet), p (pentet), dd (doublet of doublets), dt (doublet of triplets), td (triplet of doublets), ddt (doublet of doublet of triplets), dtt (doublet of triplet of triplets), br.s (broad singlet), and m (multiplet).

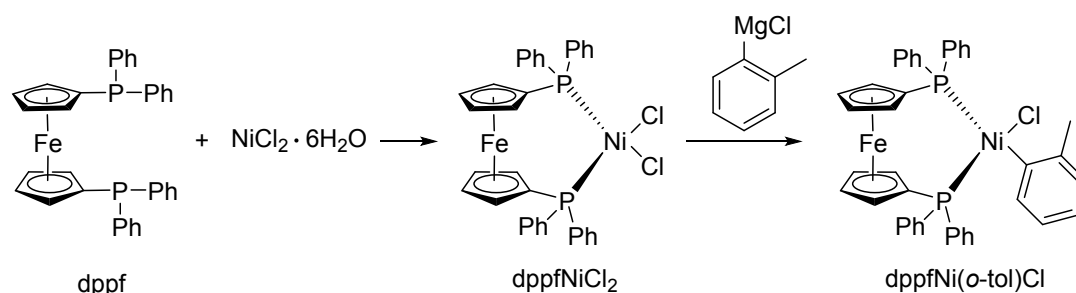
2. Analytical methods

NMRs were recorded on Bruker AV 300, AV 400, and Fourier 300 spectrometers at room temperature. For quantitative ¹⁹F NMR a puls delay of 30s are required. NMR chemical shifts are reported in parts per million (ppm) downfield from TMS. ¹³C and ¹H NMR spectra were referenced to signals of deuterio solvents and residual protiated solvents, respectively. EPR spectra were recorded on a Bruker EMX CW-micro spectrometer (X-band) with a microwave power of Ca 6.9 mW and modulation frequency and amplitude of 100 kHz and up to 5 G, respectively. For monitoring the EPR spectra at low temperature, the EPR spectrometer was equipped with a temperature controller and liquid N₂ cryostat. The g values were calculated using $h\nu = g B_0\beta$ equation being h = Planck's constant, ν = frequency, B_0 = resonance field and β = Bohr magneton. High resolution mass spectra were recorded on a MAT 95XP ThermoFisher Mass Spectrometer using Electrospray Ionization mode. Electron impact (EI) mass spectra were recorded on AMD 402 mass spectrometer (70 eV). The data are given as mass units per charge (m/z).

3. General procedures

3.1 Synthesis of dppfNi(*o*-tol)Cl

dppfNi(*o*-tol)Cl was synthesized using the slightly modified method from a literature.^[1]



Step 1: NiCl₂·6H₂O (8.5 mmol, 2.02 g) and EtOH (25 mL) were placed in an argon flushed 100 mL round Schlenk. Then the septum was removed, and dppf (8.5 mmol, 4.712 g) was added in one portion. The flask was fitted with

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a reflux condenser, and the mixture was refluxed for 30 min with an 80 °C oil bath and then cooled to room temperature. Once cool, the flask was chilled to 0 °C with an ice bath for 10 min, after which the solid was collected by vacuum filtration, washed twice with EtOH (20 mL) and twice with ether (20 mL). Drying under vacuum yielded dppfNiCl₂ (4.98 g, 85 %) as a deep green powder.

Step 2: dppfNi(*o*-tol)Cl (6.81 mmol, 4.658 g) and 180 mL CH₂Cl₂ were placed in an argon flushed 100 mL round Schlenk. The solution was cooled to 0 °C with an ice bath, and *o*-tolylmagnesium chloride (6.81 mmol, 0.945 M in THF, 7.21 mL) was added dropwise with vigorous stirring. Near the end of the addition, the solution began to change in color from green to orange. The solution was stirred for 15 min at 0 °C, after which the solution was evaporated to dryness under reduced pressure. MeOH (25 mL) was added and the mixture was stirred for 5 min to form a suspension. After the suspension was cooled to 0 °C, the solvent was filtered out. The residue was washed with two portions of cold MeOH (10 mL), and dried under high vacuum to yield dppfNi(*o*-tol)Cl (4.63 g, 92 %) as a fine, bright yellow powder.

dppfNi(*o*-tol)Cl: ¹H NMR (400 MHz, CD₂Cl₂) δ = 8.23 – 8.12 (m, 4H), 8.02 – 7.93 (m, 2H), 7.51 – 7.38 (m, 7H), 7.27 (td, *J* = 8.3, 2.0 Hz, 2H), 7.21 – 7.15 (m, 1H), 6.99 (t, *J* = 7.4 Hz, 1H), 6.76 (td, *J* = 8.2, 2.6 Hz, 2H), 6.66 – 6.54 (t, 2H), 6.43 (t, *J* = 7.4 Hz, 1H), 6.30 (t, *J* = 6.8 Hz, 1H), 6.10 (d, *J* = 7.1 Hz, 1H), 5.15 (s, 1H), 4.54 (m, 1H), 4.25 (s, 1H), 4.19 (s, 1H), 4.02 (d, *J* = 10.1 Hz, 2H), 3.52 (m, 1H), 3.33 (m, 1H), 2.44 (s, 3H).

³¹P NMR (162 MHz, CD₂Cl₂) δ = 29.51 (d, *J* = 25.9 Hz, 1P), 12.12 (d, *J* = 25.9 Hz, 1P).

3.2 Synthesis of perfluoroalkylated alkenes

Step 1: *For the reactions of C₁₀F₂₁I*: 0.2 mmol C₁₀F₂₁I, 5 mol% dppfNi(*o*-tol)Cl and 2 equiv NaO(*t*-Bu) were placed in an Ace pressure tube. After flushed with argon, 0.5 mL perfluorobenzene and 2 equiv alkene was added. *For the reactions of other perfluoro reagents except CF₃Br and CF₃I*: 5 mol% dppfNi(*o*-tol)Cl and 2 equiv NaO(*t*-Bu) were placed in an Ace pressure tube. After flushed with argon, 1 mL perfluorobenzene, 0.5 mmol perfluoro reagent and 2 equiv alkene was added. *For the reactions of CF₃I*: 5 mol% dppfNi(*o*-tol)Cl and 2 equiv NaO(*t*-Bu) were placed in an Ace pressure tube. After flushed with argon, 2 equiv alkene and 1 mL perfluorobenzene (or DCE) stock solution containing 0.5 mmol CF₃I were added.

Step 2: Then the tube was sealed and heated for 16 h at specific temperature. After cooling to room temperature, 20 μL 1,4-difluorobenzene and 4 mL heptane or pentane were added to the reaction mixture. Then 250 μL of the solution was taken to measure the conversion and yield by ¹⁹F NMR. Then, the products were isolated by fast chromatography using silica gel 60, 0.063-0.2 mm, 70-230 mesh (Merck) (Heptane or pentane elution).

3.3 Synthesis of 3,3-dimethyl-trifluoromethyl butene using CF₃Br

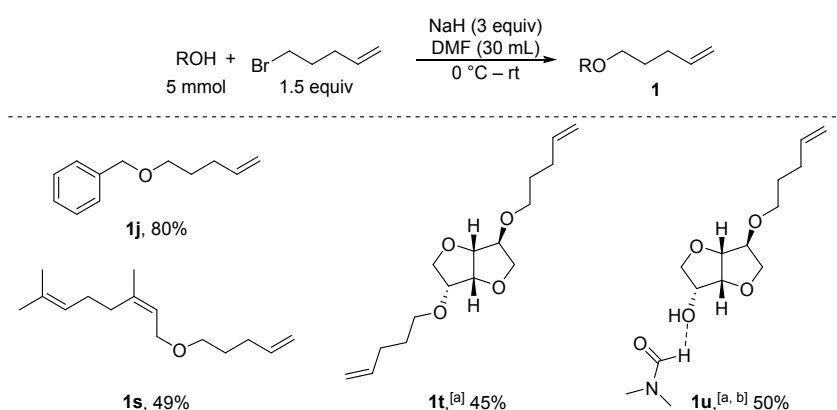
The reaction was carried out in a Parr Instruments 4560 series 300 mL autoclave containing an alloy plate with wells for five 10 mL glass vials. Under argon atmosphere, 0.5 mmol 3,3-dimethylbutene, 5 mol% dppfNi(*o*-tol)Cl and 2 equiv NaO(*t*-Bu), 1 mL perfluorobenzene and a magnetic stirring bar were placed in the vial, which were then capped with a septum equipped with an inlet needle and flushed with argon. The vial was placed in an alloy plate, which was then placed in the autoclave. Once sealed, the autoclave was purged three times with 15 bar nitrogen and then pressurized with 5 bar CF₃Br and 40 bar N₂ at room temperature and heated at 50 °C for 16 h. It was then cooled to room temperature and vented to discharge the excess gas. 20 μL 1,4-difluorobenzene and 4 mL pentane were then added to the reaction mixture. And 250 μL of the pentane solution was taken and mixed with

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350 μL CDCl_3 to measure the conversion and yield by ^{19}F NMR.

3.4 Synthesis of alkenes

In this project, we synthesized 4 alkenes which are not commercially available using slightly modified procedures from literature.^[2] The structure of these olefins are shown in Scheme S1. Firstly, the corresponding alcohol (0.5 mmol), NaH (3 equiv) and a magnetic stirring bar were placed in a 100 mL round Schlenk. After flushed with argon, DMF (30 mL) was added. Then the flask was cooled to 0 °C using an ice bath. Once cool, 5-bromopentene was added dropwise using a syringe pump. Thereafter, the reaction could warm slowly to rt and was further stirred overnight. After isolated by fast chromatography using silica gel 60, 0.063-0.2 mm, 70-230 mesh (Merck) (heptane and ethyl acetate elution), the pure products were afforded in 45% to 80% yields.



Scheme S1. Synthesis of the alkenes. Reactions were performed under Ar, yields were of the isolated products. [a] These two products were obtained by one reaction. [b] As indicated by NMR, The -OH group of the product was supposed to be protected by DMF. It will release -OH group again during the perfluoroalkylation reaction.

4. Kinetic time course

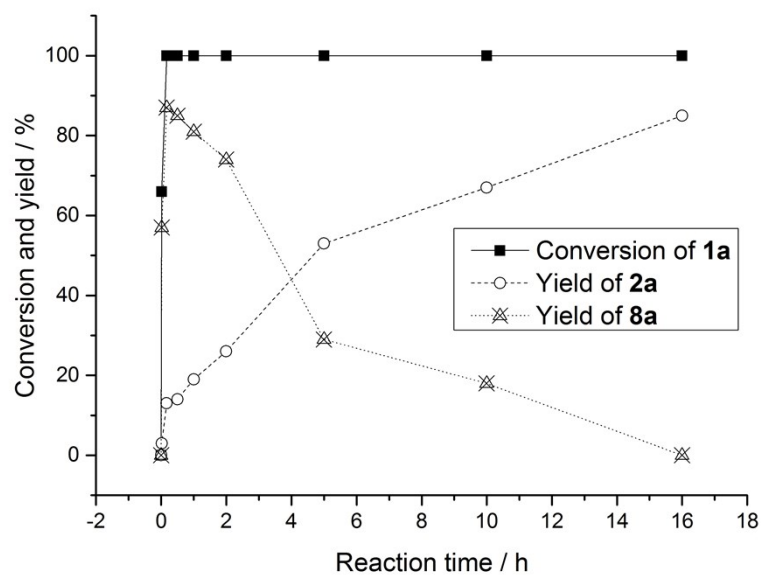


Figure S1. Kinetic time course of the nickel-catalyzed perfluoroalkylation of 3,3-dimethylbutene. Reaction conditions: $C_{10}F_{21}I$ (0.2 mmol), **1a** (0.5 mL), $dppfNi(o-tol)Cl$ (5 mol%), $NaO(t-Bu)$ (2 equiv), 50 °C, Ar, conversions and yields were determined by ^{19}F NMR using 1,4-difluorobenzene as the standard. The conversion of **1a** was completed within only 10 mins. There were two inflection points of the yield of **2a**. The first inflection point at 10 mins was caused by the ending of direct perfluoroalkylation. The second inflection point at 5 h was caused by the low concentration of **8a**.

5. In situ EPR

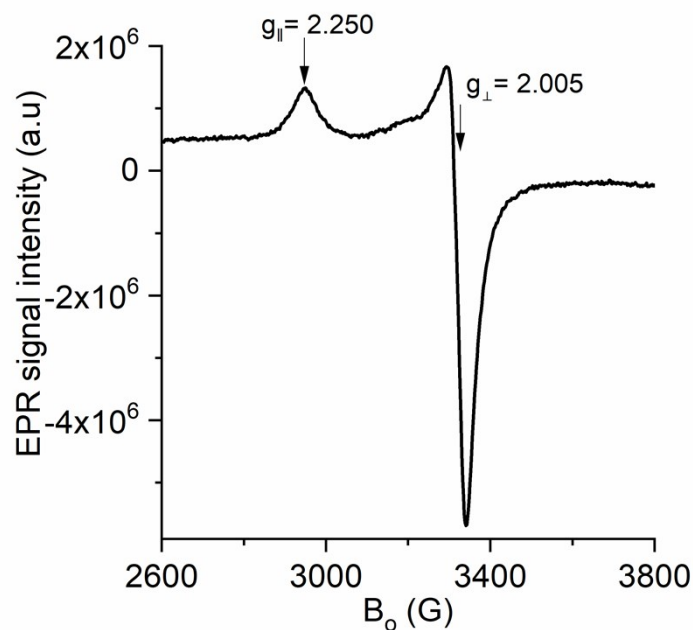


Figure S2. EPR spectrum of the reaction mixture **1a** (0.5 mL), dppfNi(o-tol)Cl (5 mol%) and NaO(*t*-Bu) (2 equiv) measured at 100 K under Ar. The spectrum shows an axial EPR signal due to the formation of Ni(I) species (**A**).

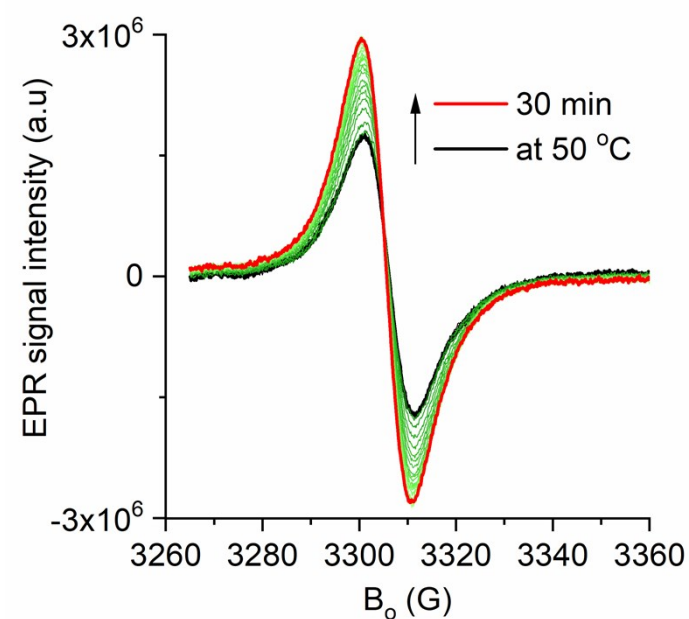
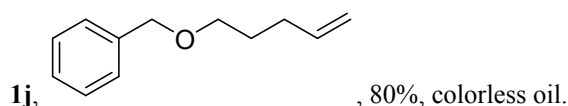


Figure S3. In situ EPR spectra with time of the reaction mixture $C_{10}F_{21}I$ (0.2 mmol), **1a** (0.5 mL), dppfNi(o-tol)Cl (5 mol%) and NaO(*t*-Bu) (2 equiv) measured at 50 °C under Ar. The spectra show EPR signal at $g = 2.005$ without hyperfine splitting due to the formation of organic radicals (**B** and **C**). The Ni(I) signal is not observable at reaction temperature due to the short relaxation time.^[3]

6. Characterization data

6.1 Characterization data of alkenes

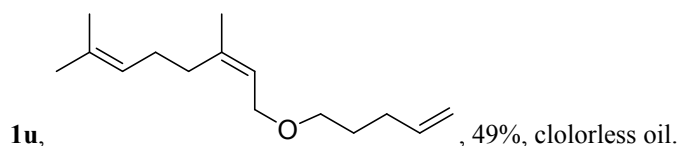


¹H NMR (300 MHz, CDCl₃): δ = 7.38 – 7.03 (m, 5H), 5.74 (ddt, J = 17.0, 10.2, 6.7 Hz, 1H), 5.06 – 4.74 (m, 2H), 4.41 (s, 2H), 3.40 (t, J = 6.5 Hz, 2H), 2.20 – 1.94 (m, 2H), 1.75 – 1.43 (m, 2H).

¹³C NMR (75 MHz, CDCl₃): δ = 138.57, 138.22, 128.27, 127.54, 127.42, 114.66, 72.82, 69.65, 30.30, 28.92.

MS (EI, 70 eV): m/z (%) = 176.1 (7) [M]⁺, 175.1 (14) [M-H]⁺, 107.1 (13), 91.1 (100), 79.1 (5), 65.1 (9), 55.1 (3).

TLC (silica gel 60 F₂₅₄): Ethyl acetate/heptane = 1:10, R_f = 0.24 (KMnO₄ stain).



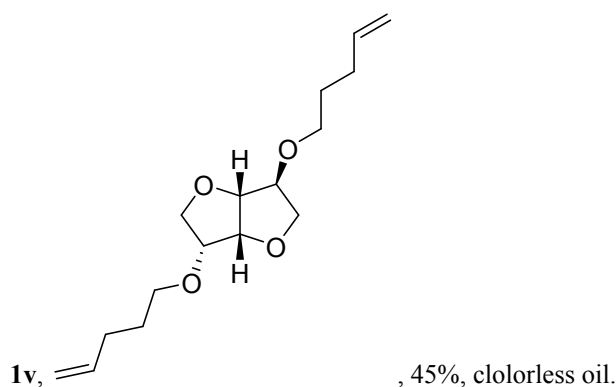
¹H NMR (300 MHz, CDCl₃): δ = 5.89 (ddt, J = 16.8, 9.9, 6.5 Hz, 1H), 5.43 (t, J = 6.9 Hz, 1H), 5.25 – 4.94 (m, 3H), 4.01 (d, J = 6.9 Hz, 2H), 3.49 (t, J = 6.6 Hz, 2H), 2.26 – 2.07 (m, 6H), 1.87 – 1.64 (m, 9H).

¹³C NMR (75 MHz, CDCl₃): δ = 140.04, 138.28, 131.76, 123.86, 122.02, 114.56, 69.50, 66.95, 32.20, 30.34, 28.97, 26.67, 25.62, 23.41, 17.56.

HRMS (EI): [M]⁺ calcd for C₁₅H₂₆O₁, 222.19782; found, 222.19802.

TLC (silica gel 60 F₂₅₄): Ethyl acetate/heptane = 1:10, R_f = 0.67.

TLC (silica gel 60 F₂₅₄): Ethyl acetate/heptane = 1:2, R_f = 0.76.

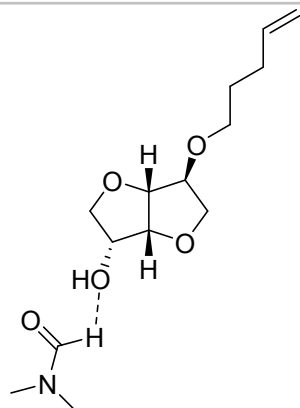


¹H NMR (300 MHz, CDCl₃): δ = 5.91 – 5.61 (m, 2H), 5.11 – 4.83 (m, 4H), 4.66 – 4.39 (m, 2H), 4.05 – 3.79 (m, 5H), 3.74 – 3.35 (m, 5H), 2.10 (m, 4H), 1.78 – 1.51 (m, 4H).

¹³C NMR (75 MHz, CDCl₃): δ = 138.03, 137.93, 114.83, 114.72, 86.23, 84.35, 80.32, 80.08, 73.37, 70.02, 69.68, 68.83, 30.09, 30.05, 28.82, 28.74.

MS (EI, 70 eV): m/z (%) = 282.2 (2) [M]⁺, 179.1 (12), 84.1 (42), 69.1 (100).

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1w, 50%, colorless oil.

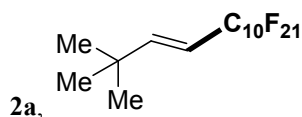
¹H NMR (300 MHz, CDCl₃): δ = 7.91 (s, 1H), 5.73 (ddt, J = 14.2, 10.6, 7.0 Hz, 1H), 5.11 – 4.77 (m, 2H), 4.60 (t, J = 3.5 Hz, 1H), 4.36 (d, J = 3.1 Hz, 1H), 4.20 (s, 1H), 3.98 – 3.66 (m, 5H), 3.66 – 3.25 (m, 3H), 2.88 (s, 3H), 2.80 (s, 3H), 2.04 (q, J = 6.9 Hz, 2H), 1.63 (p, J = 6.9 Hz, 2H).

¹³C NMR (75 MHz, CDCl₃): δ = 162.38, 137.78, 114.46, 88.10, 80.16, 79.67, 76.06, 75.65, 69.74, 69.50, 36.25, 31.15, 29.78, 28.54.

MS (EI, 70 eV): m/z (%) = 214.2 (1) [M-DMF]⁺, 159.1 (14), 128.1 (14), 85.1 (24), 84.1 (60), 69.1 (100), 68.1 (73), 67.2 (21).

6.2 Characterization data of perfluorodecylated alkenes

Notes: For the ¹³C NMR (F coupled), peaks for the C of perfluoroalkyl chain and C adjacent to perfluoroalkyl chain (in most cases) are too broad to be assigned (not shown in the following data).



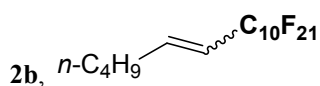
2a, colorless oil.

¹H NMR (300 MHz, CDCl₃): δ = 6.39 (dt, J = 16.0, 2.2 Hz, 1H), 5.48 (dt, J = 16.0, 12.0 Hz, 1H), 1.08 (s, 9H).

¹⁹F NMR (282 MHz, CDCl₃): δ = -81.03 (t, J = 10.0 Hz, 3F), -111.31 (q, J = 12.3 Hz, 2F), -120.65 – -124.69 (14F), -126.33 (br.s 2F).

TLC (silica gel 60 F₂₅₄): pentane, R_f = 0.9, KMnO₄ stain.

HRMS (EI): [M]⁺ calcd for C₁₆H₁₁F₂₁, 602.05199; found, 602.05119.



2b, *n*-C₄H₉, colorless oil, mixture of *E* and *Z* isomers (84:16).

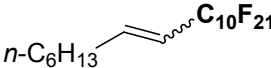
¹H NMR (300 MHz, CDCl₃): δ = 6.52 – 5.99 (m, =CHCF₂- (*E/Z*), 1H), 5.74 – 5.27 (m, -CH=CHCF₂- (*E/Z*), 1H), 2.47 – 1.99 (m, -CH₂-CH= (*E/Z*), 2H), 1.50 – 1.12 (m, -CH₂-, 4H), 0.92 (t, J = 7.2 Hz, -CH₃, 3H).

¹³C NMR (75 MHz, CDCl₃): δ = 143.29, 116.76, 77.00, 31.69, 30.07, 22.03, 13.74.

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.80 (t, J = 10.0 Hz, -CF₃, 3F), -106.67 (q, J = 13.7, 13.3 Hz, =CHCF₂-, *Z*-isomer), -111.22 (q, J = 11.2, 10.0 Hz, =CHCF₂-, *E*-isomer), -121.48 (br.s, -CF₂-, 2F), -121.84 (br.s, -CF₂-, 8F) - 122.72 (br.s, -CF₂-, 2F), -123.56 (br.s, =CHCF₂CF₂-, *E*-isomer), -123.74 (br.s, =CHCF₂CF₂-, *Z*-isomer), -126.14 (br.s, -CF₂-, 2F).

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HRMS (EI): [M]⁺ calcd for C₁₆H₁₁F₂₁, 602.05199; found, 602.05121.

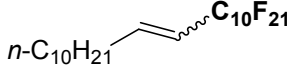
2c, , yellow sticky oil, mixture of *E* and *Z* isomers (87:13).

¹H NMR (300 MHz, CDCl₃): δ = 6.58 – 5.98 (m, =CHCF₂- (*E/Z*), 1H), 5.74 – 5.26 (m, -CH=CHCF₂- (*E/Z*), 1H), 2.41 – 2.08 (m, -CH₂-CH= (*E/Z*), 2H), 1.44 (m, -CH₂-, 2H), 1.29 (br.s, -CH₂-, 6H), 0.89 (t, *J* = 6.9 Hz, -CH₃, 3H).

¹³C NMR (75 MHz, CDCl₃): δ = 143.22 (t, *J* = 9.1 Hz, =CHCF₂-), 116.74 (t, *J* = 22.9 Hz, -CH=CHCF₂-), 76.94, 31.96, 31.48, 28.58, 27.90, 22.46, 13.84.

¹⁹F NMR (282 MHz, CDCl₃): δ = -81.16 (t, *J* = 9.9 Hz, -CF₃, 3F), -106.86 (q, *J* = 13.2 Hz, =CHCF₂-, *Z*-isomer), -111.46 (q, *J* = 11.7, 12.8 Hz, =CHCF₂-, *E*-isomer), -121.65 (br.s, -CF₂-, 2F), -122.05 (br.s, -CF₂-, 8F), -122.96 (br.s, -CF₂-, 2F), -123.77 (br.s, =CHCF₂CF₂-, *E*-isomer), -123.94 (br.s, =CHCF₂CF₂-, *Z*-isomer), -126.41 (br.s, -CF₂-, 2F).

HRMS (EI): [M]⁺ calcd for C₁₈H₁₅F₂₁, 630.08329; found, 630.08205.

2d, , colorless oil, mixture of *E* and *Z* isomers (88:12).

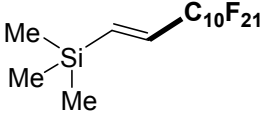
¹H NMR (300 MHz, CDCl₃): δ = 6.55 – 6.04 (m, =CHCF₂- (*E/Z*), 1H), 5.94 – 5.34 (m, -CH=CHCF₂- (*E/Z*), 1H), 2.50 – 2.09 (m, -CH₂-CH= (*E/Z*), 2H), 1.56 – 1.38 (m, -CH₂-, 2H), 1.28 (br.s, -CH₂-, 14H), 0.96 – 0.74 (t, *J* = 6.9 Hz, -CH₃, 3H).

¹³C NMR (75 MHz, CDCl₃): δ = 143.27 (t, *J* = 9.0 Hz, =CHCF₂-), 116.94 (t, *J* = 23.0 Hz, -CH=CHCF₂-), 32.07, 32.00, 29.67, 29.61, 29.42, 29.06, 28.08, 22.74, 13.98.

¹⁹F NMR (282 MHz, CDCl₃): δ = -81.31 (t, *J* = 10.0 Hz, -CF₃, 3F), -106.94 (q, *J* = 13.5 Hz, =CHCF₂-, *Z*-isomer), -111.56 (q, *J* = 11.4, 10.4 Hz, =CHCF₂-, *E*-isomer), -121.71 (br.s, -CF₂-, 2F), -122.12 (br.s, -CF₂-, 8F), -123.04 (br.s, -CF₂-, 2F), -123.82 (br.s, =CHCF₂CF₂-, *E*-isomer), -124.01 (br.s, =CHCF₂CF₂-, *Z*-isomer), -126.52 (br.s, -CF₂-, 2F).

MS (EI, 70 eV): *m/z* (%) = 131.1 (17), 111.2 (31), 98.1 (30), 97.1 (61), 85.1 (46), 84.1 (61), 83.1 (58), 71.1 (64), 70.1 (91), 69.1 (95), 57.1 (100), 56.1 (92), 55.1 (64).

HRMS (EI): [M]⁺ calcd for C₂₂H₂₃F₂₁, 686.14589; found, 686.14522.

2e, , light yellow oil.

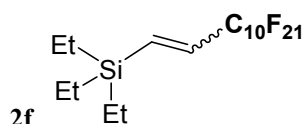
¹H NMR (300 MHz, CDCl₃): δ = 6.57 (dt, *J* = 19.0, 2.3 Hz, =CHCF₂-, 1H), 5.87 (dt, *J* = 19.1, 11.0 Hz, -CH=CHCF₂-, 1H), 0.00 (s, -CH₃, 9H).

¹³C NMR (75 MHz, CDCl₃): δ = 145.99 (t, *J* = 5.9 Hz, -C=CHCF₂-), 132.04 (t, *J* = 24.1 Hz, -C=CHCF₂-), 0.00.

¹⁹F NMR (282 MHz, CDCl₃): δ = -81.12 (CF₃, 3F), -113.43 (q, *J* = 11.6, 11.2 Hz, alkenyl- CF₂, 2F), -120.90 – -124.77 (m, 14F), -126.40 (br.s, 2F). -81.12 (t, *J* = 9.9 Hz, -CF₃, 3F), -113.43 (q, *J* = 11.6, 11.2 Hz, =CHCF₂-, 2F), -121.64 (br.s, -CF₂-, 2F), -122.03 (br.s, -CF₂-, 8F), -122.94 (br.s, -CF₂-, 2F), -123.67 (br.s, =CHCF₂CF₂-, 2F), -126.40 (br.s, -CF₂-, 2F).

MS (EI, 70 eV): *m/z* (%) = 603.1 (27) [M-Me]⁺, 214.1 (4), 184.1 (10), 153.1 (77), 131.0 (27), 103.1 (17), 77.1 (100), 53.1 (15).

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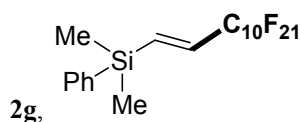


2f, colorless oil, mixture of *E* and *Z* isomers (84:16).

¹H NMR (300 MHz, CDCl₃): δ = 6.67 (dt, *J* = 19.2, 2.3 Hz, =CHCF₂- (*E*), 0.8H), 6.49 – 6.28 (m, -CH=CHCF₂- (*Z*), 0.4H), 6.05 (dt, *J* = 19.2, 10.9 Hz, -CH=CCF₂- (*E*), 0.8H), 0.96 (t, *J* = 7.9 Hz, -CH₃, 9H), 0.66 (q, *J* = 8.4 Hz, -CH₂-, 6H).

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.98 (t, *J* = 10.0 Hz, -CF₃, 3F), -110.46 (q, *J* = 12.1 Hz, alkenyl- CF₂ (*Z*), 0.4F), -113.29 (q, *J* = 11.4, alkenyl- CF₂ (*E*), 1.6F), -121.22 – -123.80 (14F), -126.26 (br.s, 2F).

MS (EI, 70 eV): *m/z* (%) = 631.2 (100) [M-Et]⁺, 169.0 (19), 167.1 (19), 131.0 (23), 119.0 (26), 105.1 (49), 95.1 (33), 77.1 (22), 69.1 (27).



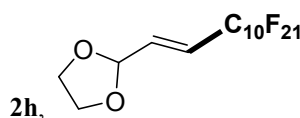
2g, colorless oil.

¹H NMR (300 MHz, CDCl₃): δ = 7.52 – 7.46 (m, -Ph, 2H), 7.43 – 7.35 (m, -Ph, 3H), 6.85 (dt, *J* = 19.0, 2.3 Hz, =CHCF₂-, 1H), 6.07 (dt, *J* = 19.0, 10.9 Hz, -CH=CCF₂-, 1H), 0.44 (s, -CH₃, 6H).

¹³C NMR (75 MHz, CDCl₃): δ = 142.05 (t, *J* = 5.6 Hz, -C=CHCF₂-), 135.73, 133.77, 131.30 (t, *J* = 24.0 Hz, -C=CHCF₂-), 129.72, 128.09, -3.32.

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.83 (t, *J* = 10.0 Hz, -CF₃, 3F), -113.24 (q, *J* = 12.0 Hz, =CHCF₂-, 2F), -121.44 (br.s, -CF₂-, 2F), -121.84 (br.s, -CF₂-, 10F), -122.73 (br.s, -CF₂-, 2F), -123.39 (br.s, -CF₂-, 2F), -126.17 (br.s, -CF₂-, 2F).

MS (EI, 70 eV): *m/z* (%) = 665.1 (29) [M-Me]⁺, 584.1 (22), 215.1 (51), 165.1 (100), 139.0 (49), 135.1 (25), 115.1 (24), 77.1 (64).



2h, colorless sticky oil.

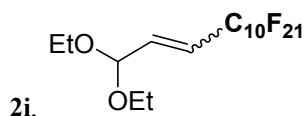
¹H NMR (300 MHz, CDCl₃): δ = 6.37 (ddt, *J* = 15.8, 4.2, 2.1 Hz, 1H), 6.05 (dt, *J* = 16.1, 12.1 Hz, 1H), 5.47 (m, 1H), 4.08 – 3.84 (m, 4H).

¹³C NMR (75 MHz, CDCl₃): δ = 137.94, 120.45, 100.57, 65.11.

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.79 (t, *J* = 10.0 Hz, 3F), -112.52 (q, *J* = 12.1 Hz, alkenyl- CF₂, 2F), -120.13 – -124.31 (m, 14F), -126.12 (2F).

HRMS (EI): [M-H]⁺ calcd for C₁₅H₆O₂F₂₁, 617.00270; found, 616.99916.

TLC (silica gel 60 F₂₅₄): Ethyl acetate/heptane = 1:10, R_f = 0.49 (KMnO₄ stain).



2i, colorless oil, mixture of *E* and *Z* isomers (87:13).

¹H NMR (300 MHz, CDCl₃): δ = 6.39 – 6.32 (m, =CHCF₂- (*E*)), 6.20 – 6.12 (m, =CHCF₂- (*Z*)), 6.12 – 5.90 (m, -CH=CHCF₂- (*E*)), 5.78 – 5.55 (m, -CH=CHCF₂- (*Z*)), 5.33 – 5.25 (m, -CH(OEt)₂ (*Z*)), 5.11 – 5.02 (m, -CH(OEt)₂

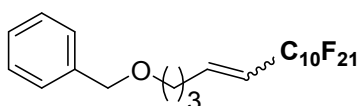
SUPPORTING INFORMATION

(*E*)), 3.76 – 3.44 (m, $-CH_2-$, 4H), 1.22 (t, $J = 7.1$ Hz, $-CH_3$, 6H).

^{13}C NMR (75 MHz, CDCl_3): $\delta = 139.28$ (t, $J = 8.7$ Hz, $-C=CHCF_2-$), 120.24 (t, $J = 23.5$ Hz, $-C=CHCF_2-$), 98.15, 77.00, 61.16, 15.06.

^{19}F NMR (282 MHz, CDCl_3): $\delta = -81.02$ (t, $J = 10.0$ Hz, CF_3 , 3F), -106.89 (q, $J = 13.7, 13.1$ Hz, $=CHCF_2-$ (*Z*)), -112.41 (q, $J = 12.4$ Hz, $=CHCF_2-$ (*E*)), -121.59 (br.s, $-CF_2-$, 2F), -121.96 (br.s, $-CF_2-$, 10F), -122.87 (br.s, $-CF_2-$, 2F), -123.46 (br.s, $-CF_2-$, 2F), -126.31 (br.s, $-CF_2-$, 2F).

MS (EI, 70 eV): m/z (%) = 603.1 (48) $[\text{M}-\text{EtO}]^+$, 575.1 (100), 238.1 (5), 218.9 (8), 169.0 (24), 131.1 (47), 106.1 (66), 69.0 (54).

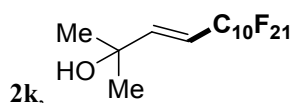


2j, colorless oil, mixture of *E* and *Z* isomers (82:18).

^1H NMR (300 MHz, CDCl_3): $\delta = 7.37 - 7.28$ (m, -Ph, 5H), 6.49 – 5.41 (m, $-CH=CHCF_2-$, *E/Z*, 2H), 4.52 (s, $\text{Ph}-CH_2-$, 2H), 3.54 – 3.49 (t, $J = 6.2$ Hz, $-OCH_2CH_2-$, 2H), 2.45 – 2.30 (m, $-CH_2-$, 2H), 1.83 – 1.64 (m, $-CH_2-$, 2H).

^{19}F NMR (282 MHz, CDCl_3): $\delta = -80.98$ (t, $J = 10.0$ Hz, $-CF_3$, 3F), -106.81 (q, $J = 13.5$ Hz, $=CHCF_2-$ (*Z*)), -111.39 (q, $J = 11.5, 10.5$ Hz, $=CHCF_2-$ (*E*)), -121.53 (br.s, $-CF_2-$, 2F), -121.92 (br.s, $-CF_2-$, 8F), -122.82 (br.s, $-CF_2-$, 2F), -123.57 and -123.78 (br.s, $=CHCF_2CF_2-$ (*E/Z*), 2F), -126.27 (br.s, $-CF_2-$, 2F).

HRMS (EI): $[\text{M}]^+$ calcd for $\text{C}_{22}\text{H}_{15}\text{O}_1\text{F}_{21}$, 694.07821; found, 694.07752.

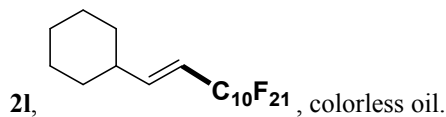


2k, colorless oil.

^1H NMR (300 MHz, CDCl_3): $\delta = 6.71 - 6.41$ (m, $=CHCF_2-$, 1H), 5.87 (dt, $J = 15.4, 11.5$ Hz, $-CH=CHCF_2-$, 1H), 1.54 (s, $-\text{OH}$), 1.51 (s, $-CH_3$, 6H).

^{19}F NMR (282 MHz, CDCl_3): $\delta = -80.87$ (t, $J = 10.0$ Hz, $-CF_3$, 3F), -111.49 (q, $J = 12.2$ Hz, $=CHCF_2-$, 2F), -121.44 (br.s, $-CF_2-$, 2F), -121.87 (br.s, $-CF_2-$, 8F), -122.76 (br.s, $-CF_2-$, 2F), -123.56 (br.s, $-CF_2-$, 2F), -126.19 (br.s, $-CF_2-$, 2F).

HRMS (EI): $[\text{M}-\text{Me}]^+$ calcd for $\text{C}_{14}\text{H}_6\text{O}_1\text{F}_{21}$, 589.00778; found, 589.00799.



2l, colorless oil.

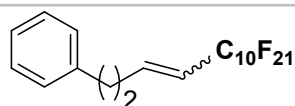
^1H NMR (300 MHz, CDCl_3): $\delta = 6.35$ (ddt, $J = 15.9, 6.7, 2.2$ Hz, $=CHCF_2-$, 1H), 5.72 – 5.22 (m, $-CH=CHCF_2-$, 1H), 2.20 – 2.04 (m, 1H), 1.79 – 1.65 (m, 4H), 1.37 – 1.08 (m, 6H).

^{13}C NMR (75 MHz, CDCl_3): $\delta = 148.16$ (t, $J = 8.8$ Hz, $-C=CHCF_2-$), 114.64 (t, $J = 23.0$ Hz, $-C=CHCF_2-$), 40.22, 31.69, 25.87, 25.65.

^{19}F NMR (282 MHz, CDCl_3): $\delta = -81.08$ (t, $J = 10.0$ Hz, $-CF_3$, 3F), -111.42 (q, $J = 12.1$ Hz, $=CHCF_2-$, 2F), -121.55 (br.s, $-CF_2-$, 2F), -122.01 (br.s, $-CF_2-$, 8F), -122.91 (br.s, $-CF_2-$, 2F), -123.78 (br.s, $=CHCF_2CF_2-$, 2F), -126.37 (br.s, $-CF_2-$, 2F).

MS (EI, 70 eV): m/z (%) = 628.1 (9) $[\text{M}]^+$, 609.1 (15), 589.1 (1), 567.1 (9), 169.0 (6), 131.0 (8), 103.1 (14), 82.1 (100), 81.1 (32), 69.1 (25), 67.1 (52).

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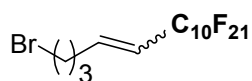
2m, colorless oil, mixture of *E* and *Z* isomers (83:17).

¹H NMR (300 MHz, CDCl₃): δ = 7.64 – 6.84 (m, -Ph, 5H), 6.70 – 6.39 (m, =CHCF₂- (*E*)), 6.21 – 6.03 (m, =CHCF₂- (*Z*)), 5.99 – 5.82 (m, -CH=CHCF₂- (*Z*)), 5.69 – 5.33 (m, -CH=CHCF₂- (*E*)), 2.81 – 2.74 (m, PhCH₂-, 2H), 2.72 – 2.64 (m, -CH₂CH= (*Z*)), 2.57 – 2.43 (m, -CH₂CH= (*E*)).

¹³C NMR (75 MHz, CDCl₃): δ = 142.07 (t, *J* = 8.9 Hz, -C=CHCF₂-), 140.36, 128.52, 128.35, 126.29, 117.62 (t, *J* = 22.9 Hz, -C=CHCF₂-), 34.40, 33.74.

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.98 (t, *J* = 10.0 Hz, -CF₃, 3F), -106.98 (q, *J* = 13.3 Hz, =CHCF₂- (*Z*)), -111.47 (q, *J* = 11.5, 10.6 Hz, =CHCF₂- (*E*)), -121.55 (br.s, -CF₂-, 2F), -121.94 (br.s, -CF₂-, 8F), -122.84 (br.s, -CF₂-, 2F), -123.61 and -123.83 (br.s, =CHCF₂CF₂- (*E/Z*), 2F), -126.27 (br.s, -CF₂-, 2F).

HRMS (EI): [M]⁺ calcd for C₂₀H₁₁F₂₁, 650.05199; found, 650.05178.



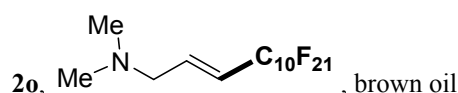
2n, colorless oil, mixture of *E* and *Z* isomers (78:22).

¹H NMR (300 MHz, CDCl₃): δ = 6.37 (dtt, *J* = 16.2, 7.1, 2.3 Hz, =CHCF₂- (*E*)), 6.15 – 6.05 (m, =CHCF₂- (*Z*)), 5.76 – 5.50 (m, -CH=CHCF₂- (*E/Z*), 1H), 3.19 (t, *J* = 6.7 Hz, BrCH₂-, 2H), 2.49 – 2.31 (m, -CH₂CH= (*E/Z*), 2H), 1.98 (p, *J* = 6.8 Hz, BrCH₂CH₂-, 2H).

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.74 (t, *J* = 10.0 Hz, -CF₃, 3F), -106.79 (q, *J* = 13.1 Hz, =CHCF₂- (*Z*)), -111.44 (q, *J* = 11.9, 11.2 Hz, =CHCF₂- (*E*)), -121.40 (br.s, -CF₂-, 2F), -121.78 (br.s, -CF₂-, 8F), -122.68 (br.s, -CF₂-, 2F), -123.39 and -123.63 (br.s, =CHCF₂CF₂- (*E/Z*), 2F), -126.09 (br.s, -CF₂-, 2F).

MS (EI, 70 eV): *m/z* (%) = 631.2 (100), 169.0 (19), 167.1 (19), 131.0 (23), 119.0 (26), 105.1 (49), 95.1 (33), 77.1 (22), 69.1 (27).

TLC (silica gel 60 F₂₅₄): Ethyl acetate/heptane = 1:10, R_f = 0.71.



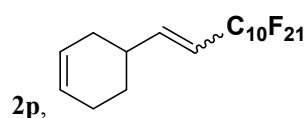
2o, brown oil

¹H NMR (300 MHz, CDCl₃): δ = 6.38 – 6.47 (m, =CHCF₂-, 1H), 5.88 – 5.74 (m, -CH=CHCF₂-, 1H), 3.06 (s, -CH₂-, 2H), 2.24 (s, -Me, 6H).

¹³C NMR (75 MHz, CDCl₃): δ = 140.17 (t, *J* = 8.9 Hz, -C=CHCF₂-), 118.89 (t, *J* = 23.3 Hz, C=CHCF₂-), 60.11, 45.23.

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.97 (t, *J* = 10.0 Hz, -CF₃, 3F), -111.75 (q, *J* = 11.1, 10.2 Hz, =CHCF₂-, 2F), -121.57 (br.s, -CF₂-, 2F), -121.94 (br.s, -CF₂-, 8F), -122.84 (br.s, -CF₂-, 2F), -123.58 (br.s, =CHCF₂CF₂-, 2F), -126.28 (br.s, -CF₂-, 2F).

MS (EI, 70 eV): *m/z* (%) = 603.1 (56) [M]⁺, 602.1 (66) [M-H]⁺, 584.1 (100), 484.1 (1), 355.1 (1), 284.1 (1), 134.1 (23), 84.1 (24), 58.1 (44).



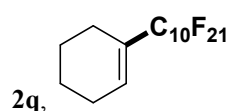
2p, colorless oil, mixture of *E* and *Z* isomers (89:11).

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¹H NMR (300 MHz, CDCl₃): δ = 6.42 (ddt, *J* = 15.9, 7.0, 2.2 Hz, =CHCF₂- (*E*)), 5.83 – 5.53 (m, -CH=CHCF₂- (*E*) and -CH₂CH=CHCH₂-), 5.15 – 4.97 (m, =CHCF₂- (*Z*) and -CH=CHCF₂- (*Z*)), 2.50 – 2.40 (m, 1H), 2.22 – 2.04 (m, 3H), 1.97 – 1.79 (m, 2H), 1.51 – 1.41 (m, 1H).

¹³C NMR (75 MHz, CDCl₃): δ = 147.19 (t, *J* = 8.7 Hz, -C=CHCF₂-), 127.04, 125.02, 115.39 (t, *J* = 22.9 Hz, -C=CHCF₂-), 36.21, 30.07, 27.47, 24.28.

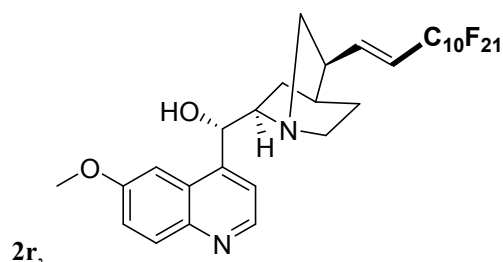
¹⁹F NMR (282 MHz, CDCl₃): δ = -81.03 (t, *J* = 10.0 Hz, -CF₃, 3F), -109.61 (br.s, =CHCF₂- (*Z*)), -111.44 (q, *J* = 12.0 Hz, =CHCF₂- (*E*)), -121.52 (br.s, -CF₂-, 2F), -121.97 (br.s, -CF₂-, 8F), -122.88 (br.s, -CF₂-, 2F), -123.70 (br.s, =CHCF₂CF₂-, 2F), -126.31 (br.s, -CF₂-, 2F).



¹H NMR (300 MHz, CDCl₃): δ = 6.26 (tt, *J* = 3.9, 1.7 Hz, 1H), 2.15 – 2.02 (m, 4H), 1.66 – 1.53 (m, 4H).

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.80 (t, *J* = 9.9 Hz, 3F), -113.42 (s, alkenyl- CF₂-, 2F), -121.92 (12F), -122.73 (br.s, 2F), -126.14 (br.s, 2F).

HRMS (EI): [M-H]⁺ calcd for C₁₆H₈F₂₁, 599.02852; found, 599.02776.

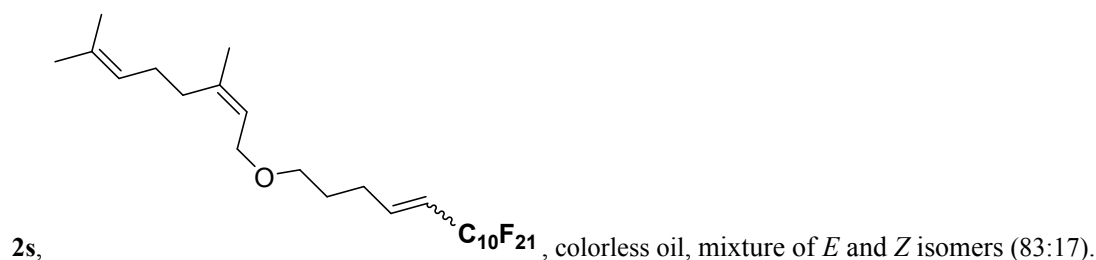


, light brown sticky oil, mixture of *E* and *Z* isomers (81:19).

¹H NMR (300 MHz, CDCl₃): δ = 8.73 (d, *J* = 4.5 Hz, -ArH, 1H), 8.01 (d, *J* = 9.2 Hz, -ArH, 1H), 7.51 – 7.32 (m, -ArH, 2H), 7.25 (s, -ArH, 1H), 6.73 – 6.51 (m, =CHCF₂- (*E* and *Z*), 1H), 6.31 (br.s, -OH, 1H), 6.13 – 5.56 (m, -CH=CHCF₂- (*E* and *Z*), 1H), 3.93 (s, -OCH₃, 3H), 3.45 (s, 1H), 3.33 (m, 1H), 3.06 (m, 2H), 2.82 (m, 2H), 2.57 – 2.12 (m, 2H), 2.04 – 1.83 (m, 1H), 1.58 (d, *J* = 9.3 Hz, 3H).

¹³C NMR (75 MHz, CDCl₃): (only the signals of *E* isomer are shown) δ = 158.28, 147.31, 144.67, 141.73, 132.01, 126.73, 122.04, 100.41, 60.16, 55.51, 50.44, 49.92, 49.81, 48.92, 38.46, 27.36, 25.91.

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.95 (t, *J* = 10.0 Hz, -CF₃, 3F), -106.21 (q, *J* = 11.1, 10.2 Hz, =CHCF₂- (*Z*)), -111.37 (q, *J* = 11.5, 10.9 Hz, =CHCF₂- (*E*)), -121.45 (br.s, -CF₂-, 2F), -121.93 (br.s, -CF₂-, 8F), -122.82 (br.s, -CF₂-, 2F), -123.48 (br.s, =CHCF₂CF₂-, (*E*)), -123.75 (br.s, =CHCF₂CF₂-, (*Z*)), -126.25 (br.s, -CF₂-, 2F).



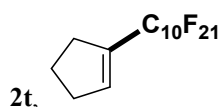
¹H NMR (300 MHz, CDCl₃): δ = 6.58 – 6.01 (m, 1H), 5.93 – 5.48 (m, 1H), 5.34 (m, 1H), 5.17 – 4.99 (m, 1H), 3.94 (d, *J* = 7.5 Hz, 2H), 3.42 (t, *J* = 6.2 Hz, 2H), 2.46 – 2.20 (m, 2H), 2.08 (m, 2H), 1.75 (s, 3H), 1.67 (s, 3H),

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1.60 (s, 3H), 1.42 – 0.99 (m, 4H).

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.87 (t, J = 10.0 Hz, 3F), -106.77 (q, J = 13.3 Hz, alkenyl- CF₂ (Z), 0.3F), -111.34 (q, J = 11.5, 10.7 Hz, alkenyl- CF₂ (E), 1.7F), -121.50 – -123.53 (14F), -126.20 (br.s, 2F).

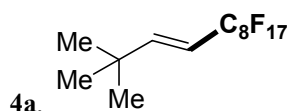
HRMS (EI): [M]⁺ calcd for C₂₅H₂₅O₁F₂₁, 740.15646; found, 740.15590.



¹⁹F NMR (282 MHz, CDCl₃): δ = -80.93 (t, J = 9.9 Hz, 3F), -110.27 (br.s, alkenyl- CF₂-, 2F), -120.93 (br.s, -CF₂-, 2F), -121.87 (br.s, -CF₂-, 8F), -122.35 (br.s, -CF₂-, 2F), -122.82 (br.s, =CHCF₂CF₂-, 2F), -126.25 (br.s, -CF₂-, 2F).

6.3 Characterization data of other perfluoroalkylated alkenes

Notes: For the ¹³C NMR (F coupled), peaks for the C of perfluoroalkyl chain and C adjacent to perfluoroalkyl chain (in most cases) are too broad to be assigned (not shown in the following data).

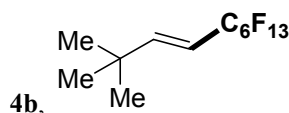


¹H NMR (300 MHz, CDCl₃): δ = 6.40 (dt, J = 16.0, 2.2 Hz, =CHCF₂-, 1H), 5.49 (dt, J = 16.0, 11.9 Hz, -CH=CHCF₂-, 1H), 1.08 (s, -CH₃, 9H).

¹³C NMR (75 MHz, CDCl₃): δ = 152.93 (t, J = 8.3 Hz, -C=CHCF₂-), 112.47 (t, J = 23.0 Hz, -C=CHCF₂-), 33.46, 28.34.

¹⁹F NMR (282 MHz, CDCl₃): δ = -81.18 (t, J = 10.1 Hz, -CF₃, 3F), -111.41 (q, J = 12.2 Hz, =CHCF₂-, 2F), -121.61 (br.s, -CF₂-, 2F), -122.18 (m, -CF₂-, 4F), -122.99 (br.s, -CF₂-, 2F), -123.89 (br.s, =CHCF₂CF₂-, 2F), 126.42 (m, -CF₂-, 2F).

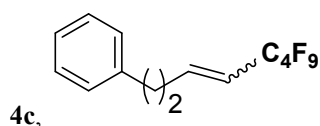
HRMS (EI) [M]⁺ calcd for C₁₄H₁₁F₁₇, 502.05838; found, 502.05844.



¹H NMR (300 MHz, CDCl₃): δ = 6.42 (dt, J = 16.0, 2.2 Hz, =CHCF₂-, 1H), 5.51 (dt, J = 16.0, 11.9 Hz, -CH=CHCF₂-, 1H), 1.11 (s, -CH₃, 9H).

¹⁹F NMR (282 MHz, CDCl₃): δ = -81.05 (tt, J = 10.1, 2.5 Hz, -CF₃, 3F), -111.29 (q, J = 12.3 Hz, =CHCF₂-, 2F), -121.75 (m, -CF₂-, 2F), -123.05 (m, -CF₂-, 2F), -123.82 (m, -CF₂-, 2F), -126.34 (m, -CF₂-, 2F).

MS (EI, 70 eV): m/z (%) = 402.1 (2) [M]⁺, 401.1 (13) [M-H]⁺, 387.1 (2), 295.0 (1), 131.0 (1), 83.1 (5), 69.1 (10), 57.0 (100).

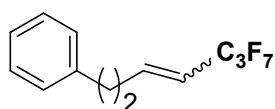


mixture of *E* and *Z* isomers (81:19).

SUPPORTING INFORMATION

¹H NMR (300 MHz, CDCl₃): δ = 7.46 – 7.29 (m, Ar-*H*, 2H), 7.28 – 7.14 (m, Ar-*H*, 3H), 6.47 (dtt, J = 15.9, 6.8, 2.3 Hz, =CHCF₂- (*E*)), 6.17 (dtt, J = 12.1, 7.3, 2.4 Hz, =CHCF₂- (*Z*)), 5.82 – 5.38 (m, -CH=CHCF₂- (*E* and *Z*), 1H), 2.79 (m, PhCH₂-, 2H), 2.74 – 2.62 (m, -CH₂CH= (*Z*)), 2.61 – 2.49 (m, -CH₂CH= (*E*)).

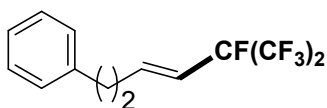
¹⁹F NMR (282 MHz, CDCl₃): δ = -79.93 – -83.15 (m, -CF₃, 3F), -105.71 – -108.92 (m, =CHCF₂- (*Z*)), -111.55 – -111.68 (m, =CHCF₂- (*E*)), -124.38 – -124.54 (m, =CHCF₂CF₂- (*E*)), -124.64 – -124.74 (m, =CHCF₂CF₂- (*Z*)), -125.75 – -125.89 (m, -CF₂-, 2F).



4d, mixture of *E* and *Z* isomers (80:20).

¹H NMR (300 MHz, CDCl₃): δ = 7.45 – 7.31 (m, Ar-*H*, 2H), 7.31 – 7.18 (m, Ar-*H*, 3H), 6.50 (dtt, J = 15.9, 6.8, 2.3 Hz, =CHCF₂- (*E*)), 6.20 (dtt, J = 12.0, 7.3, 2.3 Hz, =CHCF₂- (*Z*)), 5.81 – 5.45 (m, -CH=CHCF₂- (*E* and *Z*), 1H), 2.81 (q, J = 6.9 Hz, PhCH₂-, 2H), 2.76 – 2.65 (m, -CH₂CH= (*Z*)), 2.65 – 2.49 (m, -CH₂CH= (*E*)).

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.43 (q, J = 8.7 Hz, -CF₃, 3F), -107.76 – -108.08 (m, =CHCF₂- (*Z*)), -112.20 – -112.78 (m, =CHCF₂- (*E*)), -127.93 (s, =CHCF₂CF₂- (*E*)), -128.18 (s, =CHCF₂CF₂- (*Z*)).



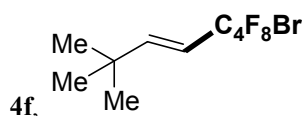
4e,

¹H NMR (300 MHz, CDCl₃): δ = 7.50 – 7.29 (m, Ar-*H*, 2H), 7.29 – 7.13 (m, Ar-*H*, 3H), 6.40 (dt, J = 15.7, 6.9 Hz, =CHCF₂-, 1H), 5.73 – 5.34 (m, -CH=CHCF₂-, 1H), 2.81 (t, J = 7.6 Hz, PhCH₂-, 2H), 2.64 – 2.40 (m, -CH₂CH=, 2H).

¹³C NMR (75 MHz, CDCl₃): δ = 140.47 and 140.33 (d, J = 10.3 Hz, -CH₂CH=), 140.40, 128.48, 128.40, 126.23, 114.73 (d, J = 16.9 Hz, =CHCF₂-), 34.58, 34.05.

¹³C NMR Dept135 (75 MHz, CDCl₃): δ = (+): 140.41 (d, J = 10.3 Hz, -CH₂CH=), 128.49, 128.41, 126.24, 114.74 (d, J = 17.0 Hz, =CHCF₂-). (-): 34.59, 34.06.

¹⁹F NMR (282 MHz, CDCl₃): δ = -77.51 (d, J = 7.7 Hz, -CF(CF₃)₂, 6F), -182.89 – -189.09 (m, -CF(CF₃)₂, 1F).

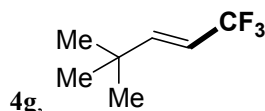


4f,

¹H NMR (300 MHz, CDCl₃): δ = 6.31 (dt, J = 16.0, 2.2 Hz, =CHCF₂-, 1H), 5.41 (dt, J = 16.0, 11.9 Hz, -CH=CHCF₂-, 1H), 1.01 (s, -CH₃, 9H).

¹⁹F NMR (282 MHz, CDCl₃): δ = -63.02 – -63.30 (m, -CF₂Br, 2F), -111.31 (q, J = 12.2 Hz, =CHCF₂-, 2F), -116.26 – -117.04 (m, -CF₂-, 2F), -122.44 – -123.19 (m, -CF₂-, 2F).

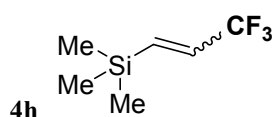
6.4 Characterization data of trifluoromethylated alkenes



4g,

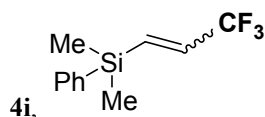
¹⁹F NMR (282 MHz, CDCl₃): δ = -67.30 (t, J = 11.0 Hz, -CF₃, 3F).

SUPPORTING INFORMATION



mixture of *E* and *Z* isomers (84:16).

¹⁹F NMR (282 MHz, CDCl₃): δ = -62.35 (s, -CF₃ (*Z*)), -67.43 (s, -CF₃ (*E*)).



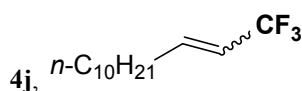
mixture of *E* and *Z* isomers (84:16).

¹H NMR (300 MHz, CDCl₃): δ = 7.42 – 7.31 (m, Ar-*H*, 2H), 7.29 – 7.14 (m, Ar-*H*, 3H), 6.59 (dq, *J* = 18.9, 2.0 Hz, =CHCF₂- (*E*)), 6.35 – 6.06 (m, =CHCF₂- (*Z*)), 5.97 – 5.79 (m, -CH=CHCF₂- (*E*)), 5.68 – 5.52 (m, -CH=CHCF₂- (*Z*)), 0.27 (s, -CH₃, 6H).

¹³C NMR (101 MHz, CDCl₃): δ = 143.32 (q, *J* = 5.7 Hz, -CH=CHCF₂- (*Z*)), 140.76 (q, *J* = 4.8 Hz, -CH=CHCF₂- (*E*)), 137.77 (*C* of -Ph, 1C), 135.77 (*C* of -Ph, 2C), 134.43 (q, *J* = 34.0 Hz, =CHCF₂-, 1C), 131.67 (*C* of -Ph, 1C), 130.06 (*C* of -Ph, 2C), 124.13 (q, *J* = 271.6 Hz, -CF₃, 1C), -1.36 (-CH₃, 2C).

¹⁹F NMR (282 MHz, CDCl₃): δ = -61.37 – -61.42 (m, -CF₃ (*Z*)), -66.49 (dd, *J* = 5.9, 2.0 Hz, -CF₃ (*E*)).

MS (EI, 70 eV): *m/z* (%) = 200.1 (21) [M]⁺, 91.1 (100).



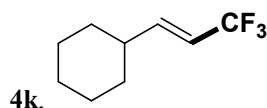
mixture of *E* and *Z* isomers (87:13).

¹H NMR (300 MHz, CDCl₃): δ = 6.54 – 6.28 (m, =CHCF₂- (*E*)), 5.98 (dt, *J* = 11.6, 7.9 Hz, =CHCF₂- (*Z*)), 5.74 – 5.45 (m, -CH=CHCF₂-, 1H), 2.43 – 2.21 (m, -CH₂CH= (*Z*)), 2.22 – 2.06 (m, -CH₂CH= (*E*)), 1.52 – 1.36 (m, -CH₂-, 2H), 1.27 (br.s, -CH₂-, 14H), 0.89 (t, *J* = 6.7 Hz, -CH₃, 3H).

¹³C NMR (101 MHz, CDCl₃): δ = 143.20 (q, *J* = 5.5 Hz, -CH=CHCF₂- (*Z*)), 140.81 (q, *J* = 6.5 Hz, -CH=CHCF₂- (*E*)), 123.13 (q, *J* = 269.0 Hz, -CF₃), 118.25 (q, *J* = 33.1 Hz, =CHCF₂- (*E*)), 118.20 (q, *J* = 33.2 Hz, =CHCF₂- (*Z*)), 31.90, 31.45, 29.58, 29.52, 29.36, 29.32, 29.03, 27.96, 22.68, 14.09.

¹⁹F NMR (282 MHz, CDCl₃): δ = -58.09 (dt, *J* = 8.7, 2.3 Hz, -CF₃ (*Z*)), -63.86 – -64.00 (m, -CF₃ (*E*)).

MS (EI, 70 eV): *m/z* (%) = 236.3 (1) [M]⁺, 208.2 (9), 193.1 (13), 179.1 (15), 165.1 (20), 151.1 (30), 137.1 (18), 123.1 (11), 111.1 (27), 97.1 (58), 83.1 (56), 69.1 (89), 57.1 (100).

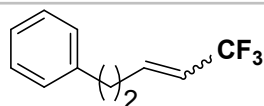


¹H NMR (300 MHz, CDCl₃): δ = 6.37 – 6.11 (m, =CHCF₂-, 1H), 5.64 – 5.31 (m, -CH=CHCF₂-, 1H), 2.11 – 1.91 (m, -CH(CH₂)₂-, 1H), 1.77 – 1.53 (m, -CH₂-, 4H), 1.34 – 0.92 (m, -CH₂-, 6H).

¹⁹F NMR (282 MHz, CDCl₃): δ = -63.78 (dt, *J* = 6.6, 2.1 Hz, -CF₃, 3F).

MS (EI, 70 eV): *m/z* (%) = 178.1 (2) [M]⁺, 163.1 (4), 136.1 (6), 115.1 (8), 95.1 (7), 82.1 (100), 67.1 (72), 54.1 (14).

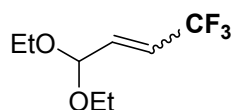
SUPPORTING INFORMATION



4l, mixture of *E* and *Z* isomers (84:16).

¹⁹F NMR (282 MHz, CDCl₃): δ = -59.08 (s, -CF₃ (*Z*)), -65.00 (s, -CF₃ (*E*)).

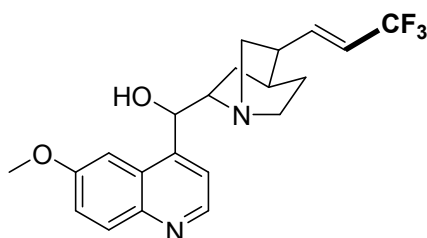
MS (EI, 70 eV): *m/z* (%) = 229.9 (<1) [M]⁺, 215.1 (100) [M-Me]⁺, 153.1 (8), 133.1 (67), 121.1 (23), 105.1 (11), 91.1 (7), 77.1 (31).



4m,

¹⁹F NMR (282 MHz, CDCl₃): δ = -64.01 – -64.43 (m, -CF₃ (*Z*)), -65.18 – -65.80 (m, -CF₃ (*E*)).

MS (EI, 70 eV): *m/z* (%) = 198.0 (<1) [M]⁺, 197.1 (1) [M-H]⁺, 153.1 (77) [M-EtO]⁺, 125.1 (100), 103.1 (6), 77.0 (6).



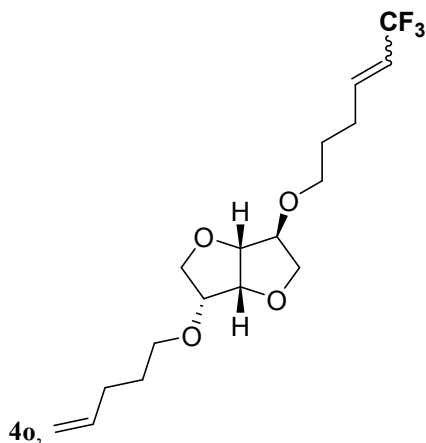
4n,

¹H NMR (300 MHz, CDCl₃): δ = 8.67 (d, *J* = 4.5 Hz, 1H), 7.96 (d, *J* = 9.2 Hz, 1H), 7.38 (d, *J* = 4.5 Hz, 1H), 7.32 (dd, *J* = 9.3, 2.7 Hz, 1H), 7.15 (s, 1H), 6.34 (dd, *J* = 11.6, 10.2 Hz, 1H), 6.22 (s, 1H), 5.76 – 5.53 (m, 1H), 3.87 (s, 3H), 3.38 – 3.12 (m, 1H), 3.07 – 2.58 (m, 5H), 2.31 – 2.10 (m, 1H), 1.86 – 1.76 (m, 2H), 1.58 – 1.43 (m, 3H).

¹³C NMR (101 MHz, CDCl₃): δ = 158.22, 147.35, 144.70, 143.92 (q, *J* = 4.1 Hz, -CH=CHCF₂-), 141.86, 132.08, 126.70, 123.13 (q, *J* = 271.9 Hz, -CF₃), 121.96, 119.01 (q, *J* = 33.3 Hz, =CHCF₂-), 100.57, 60.20, 55.63, 49.88, 49.80, 34.80, 27.64, 25.83, 21.62.

¹⁹F NMR (282 MHz, CDCl₃): δ = -57.79 (dd, *J* = 8.6, 1.8 Hz, -CF₃).

MS (EI, 70 eV): *m/z* (%) = 391.3 (<1) [M-H]⁺, 375.2 (100) [M-OH]⁺, 347.1 (4), 253.1 (7), 204.1 (11), 184.0 (8), 154.1 (4), 136.0 (3), 77.0 (1).

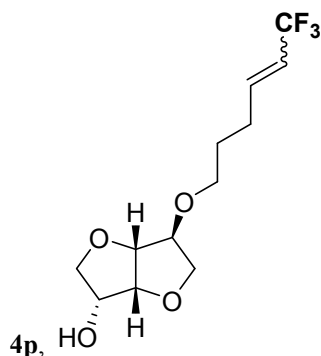


4o,

¹⁹F NMR (282 MHz, CDCl₃): δ = -58.42 (-CF₃ (*Z*)), -64.33 (-CF₃ (*E*)).

SUPPORTING INFORMATION

MS (EI, 70 eV): m/z (%) = 350.2 (5) [M]⁺, 264.2 (8), 136.1 (13), 117.1 (26), 97.1 (13), 84.1 (46), 69.1 (100).

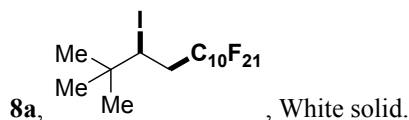


¹⁹F NMR (282 MHz, CDCl₃): δ = -58.43 (-CF₃ (Z)), -64.35 (-CF₃ (E)).

MS (EI, 70 eV): m/z (%) = 282.2 (23) [M]⁺, 136.1 (37), 117.1 (44), 97.1 (24), 86.1 (36), 85.1 (22), 84.1 (51), 77.1 (21), 69.1 (100), 68.1 (31), 58.1 (20).

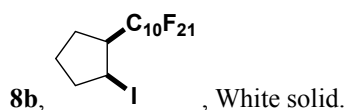
6.5 Characterization data of others

Notes: For the ¹³C NMR (F coupled), peaks for the C of perfluoroalkyl chain and C adjacent to perfluoroalkyl chain (in most cases) are too broad to be assigned (not shown in the following data).



¹H NMR (300 MHz, CDCl₃): δ = 4.18 (dd, *J* = 8.5, 2.0 Hz, -CHI-, 1H), 3.05 – 2.66 (m, -CH₂CF₂-, 2H), 1.11 (s, -Me, 9H).

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.80 (t, *J* = 10.0 Hz, -CF₃, 3F), -113.85 – -116.41 (m, -CH₂CF₂-, 2F), -121.81 (br.s, -CF₂-, 10F), -122.73 (br.s, -CF₂-, 2F), -123.66 (br.s, -CF₂-, 2F), -126.14 (br.s, -CF₂-, 2F).



¹H NMR (300 MHz, CDCl₃): δ = 4.50 (dt, *J* = 6.8, 4.5 Hz, -CHI-, 1H), 3.40 – 2.92 (m, -CH₂CF₂-, 1H), 2.36 – 1.65 (m, -CH₂-, 6H).

¹³C NMR (101 MHz, CDCl₃): δ = 53.68 (dd, *J* = 21.6, 19.7 Hz), 41.01, 26.35, 25.35, 18.65.

¹⁹F NMR (282 MHz, CDCl₃): δ = -80.85 (t, *J* = 10.0 Hz, -CF₃, 3F), -114.05 – -120.85 (dd, *J* = 1643.1, 274.4 Hz, -CH₂CF₂-, 2F, the right line is overlapped with the signal of -120.88), -120.88 (br.s, -CF₂-, 2F), -121.78 (br.s, -CF₂-, 10F), -122.75 (br.s, -CF₂-, 2F), -126.17 (br.s, -CF₂-, 2F).

7. References

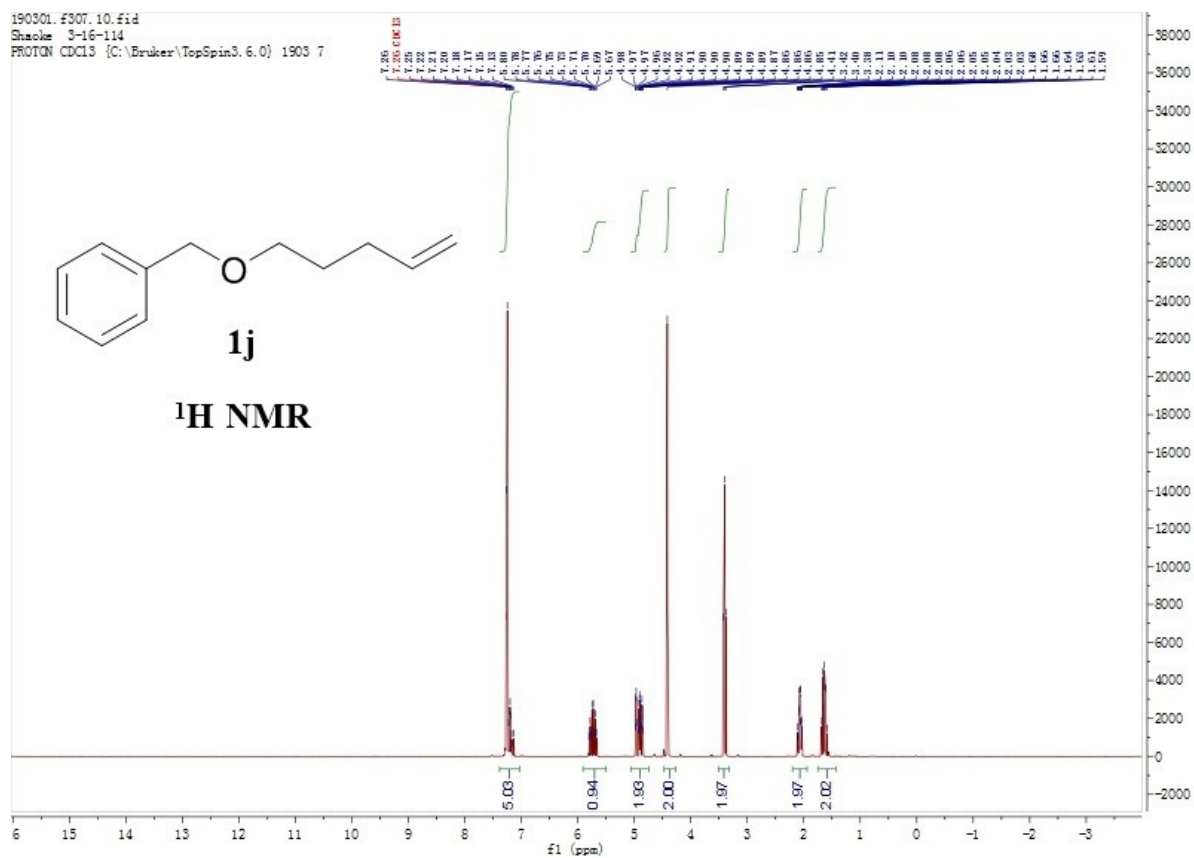
- [1] E. A. Standley, S. J. Smith, P. Muller and T. F. Jamison, *Organometallics* **2014**, 33, 2012-2018.
- [2] L. Legnani, G. Prina-Cerai, T. Delcaillau, S. Willems and B. Morandi, *Science* **2018**, 362, 434-439.

SUPPORTING INFORMATION

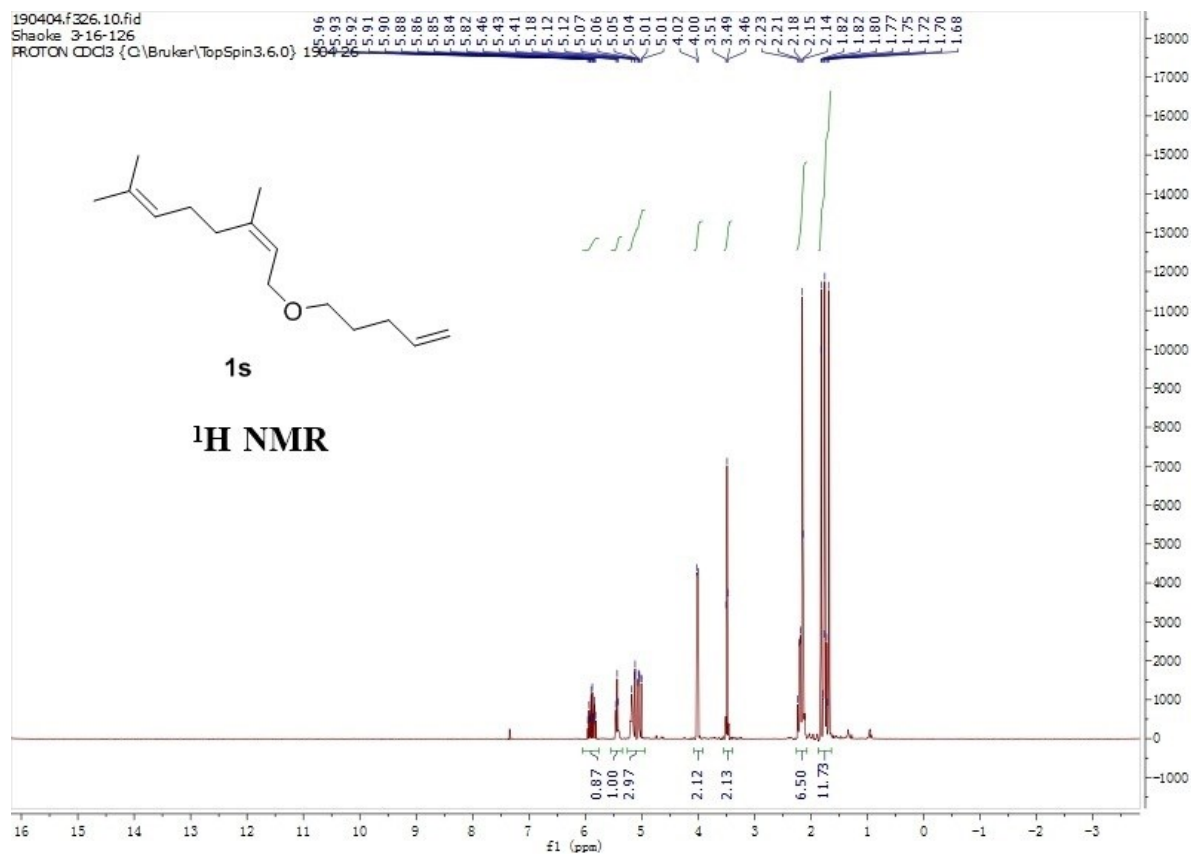
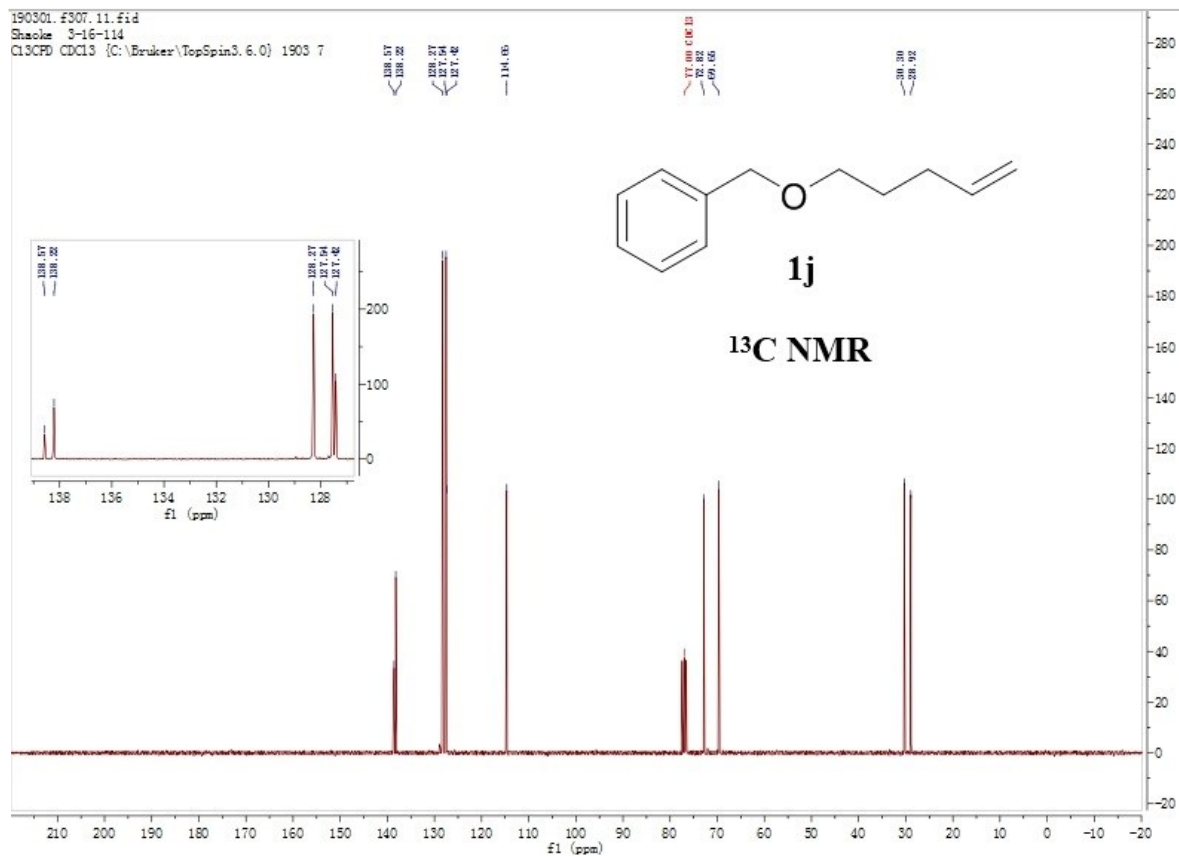
[3] J. Rabeah, J. Radnik, V. Briois, D. Maschmeyer, G. Stochniol, S. Peitz, H. Reeker, C. La Fontaine and A. Brückner, *ACS Catal.* **2016**, 6, 8224-8228.

8. Spectra

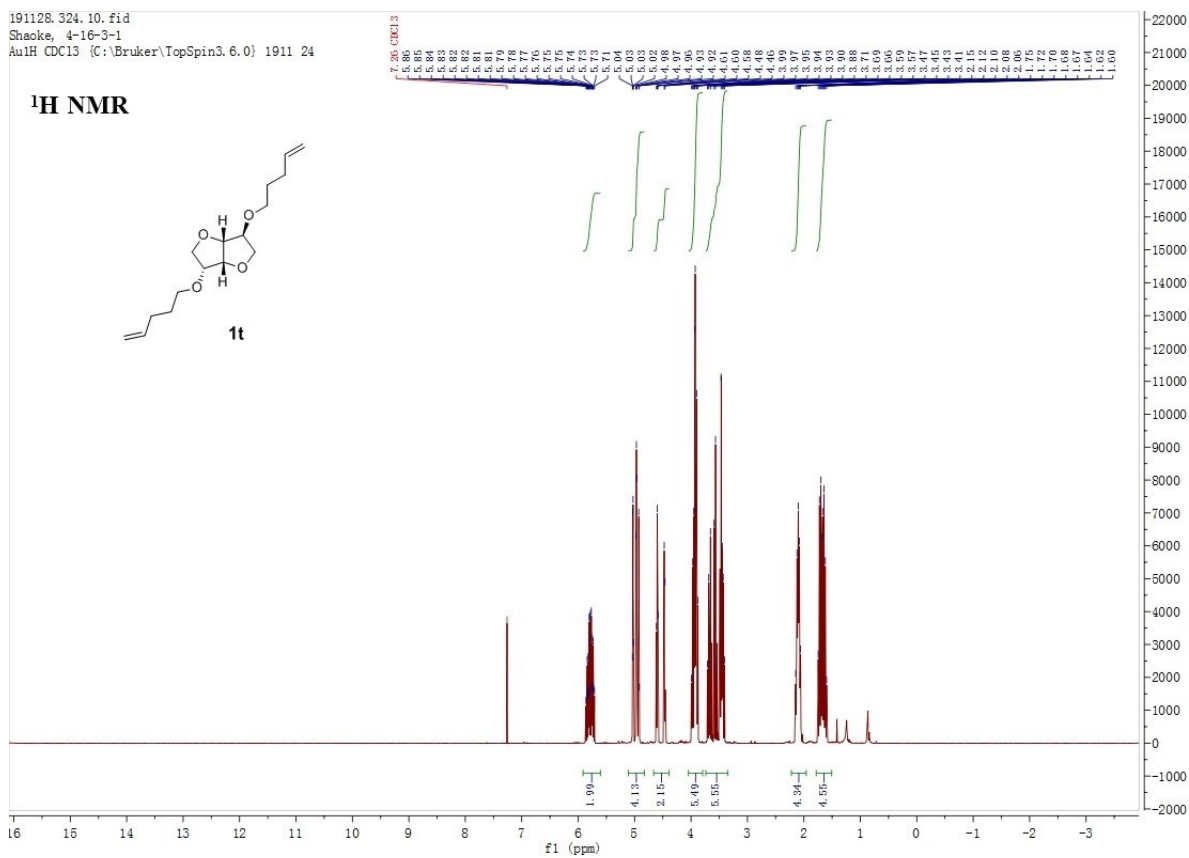
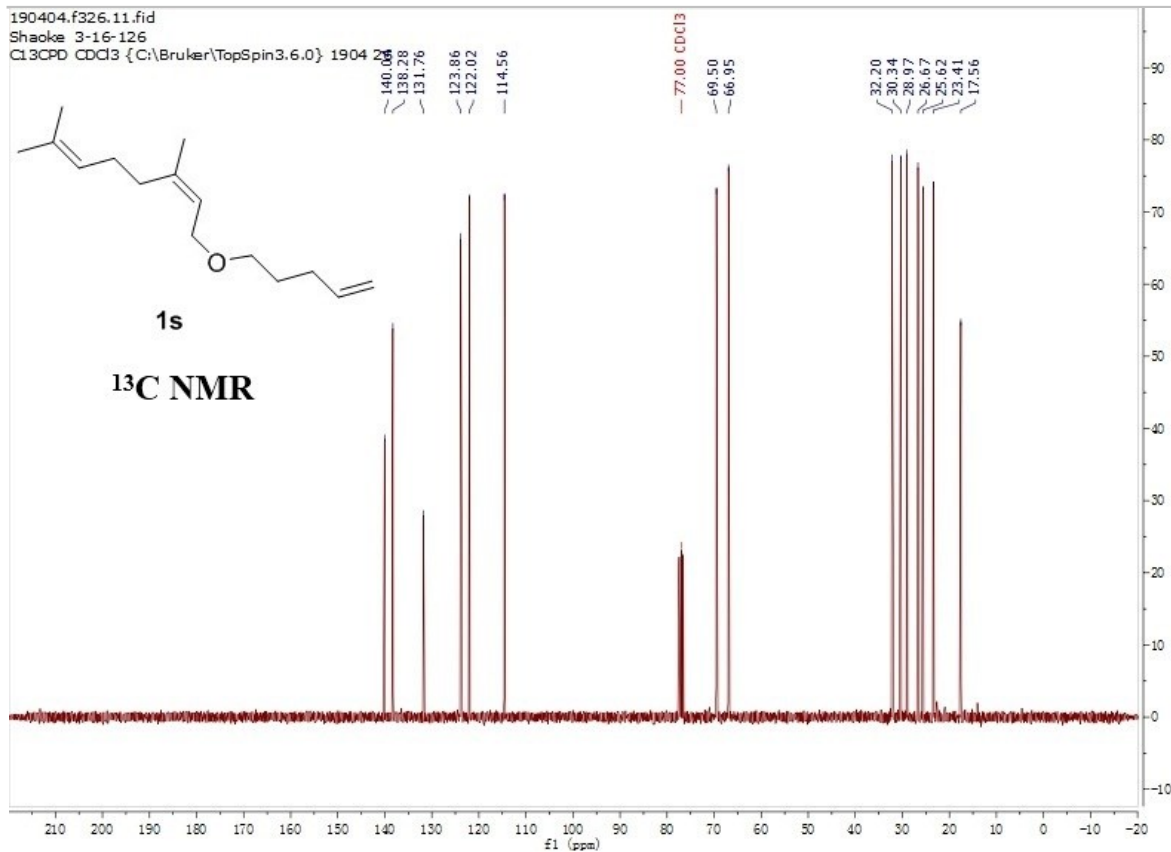
8.1 Spectra of alkenes



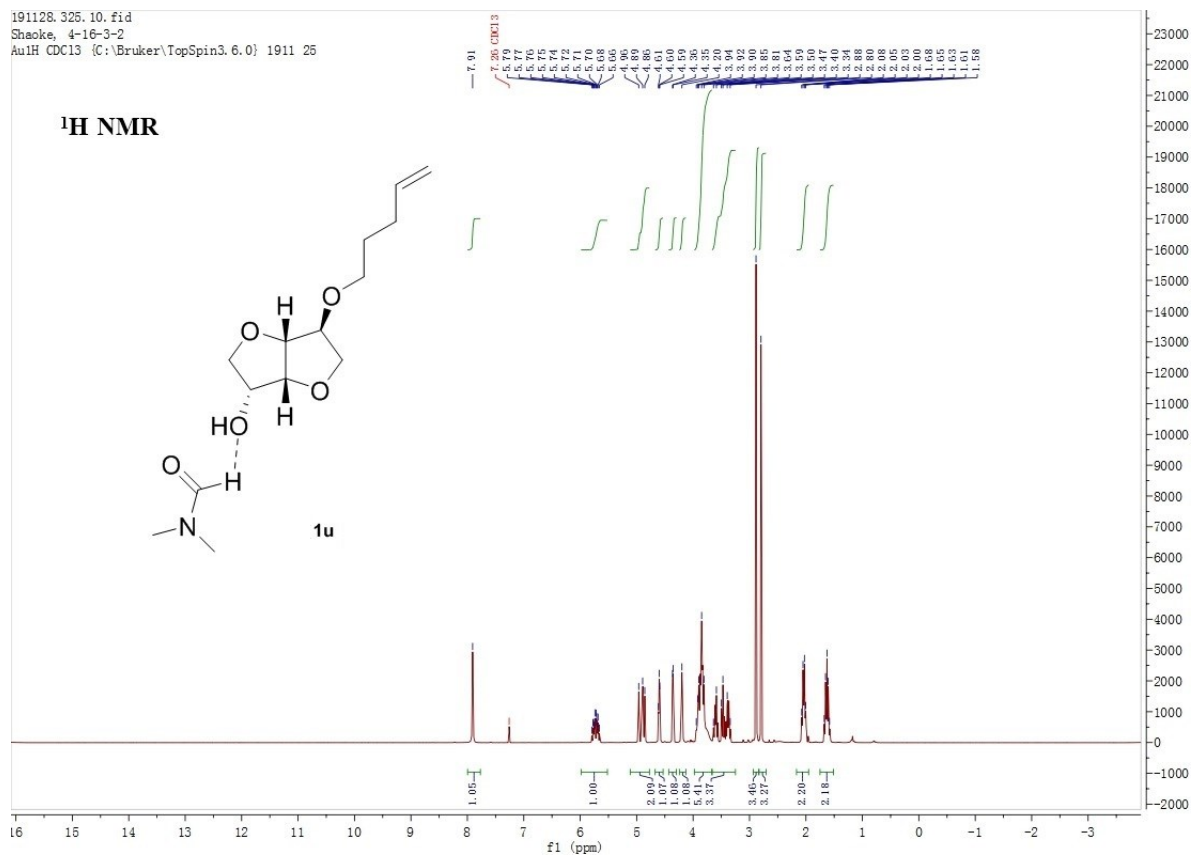
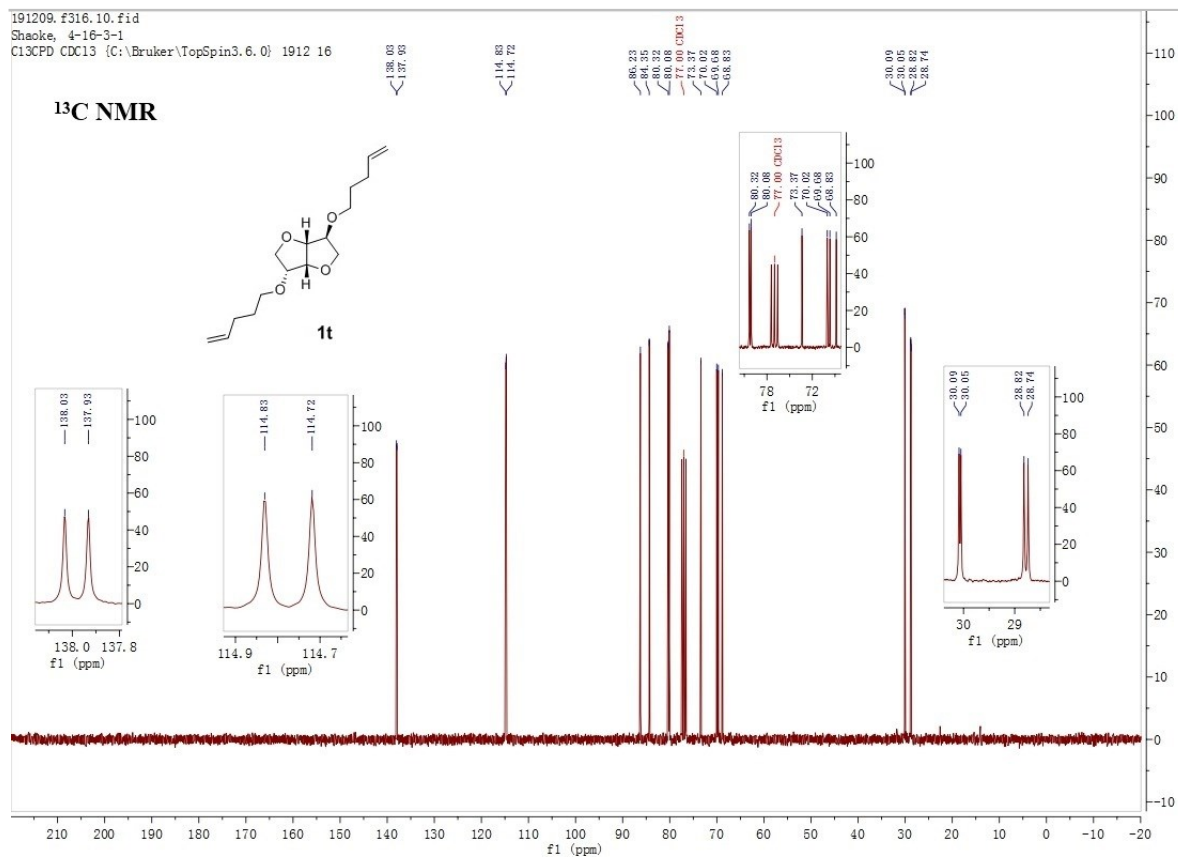
SUPPORTING INFORMATION



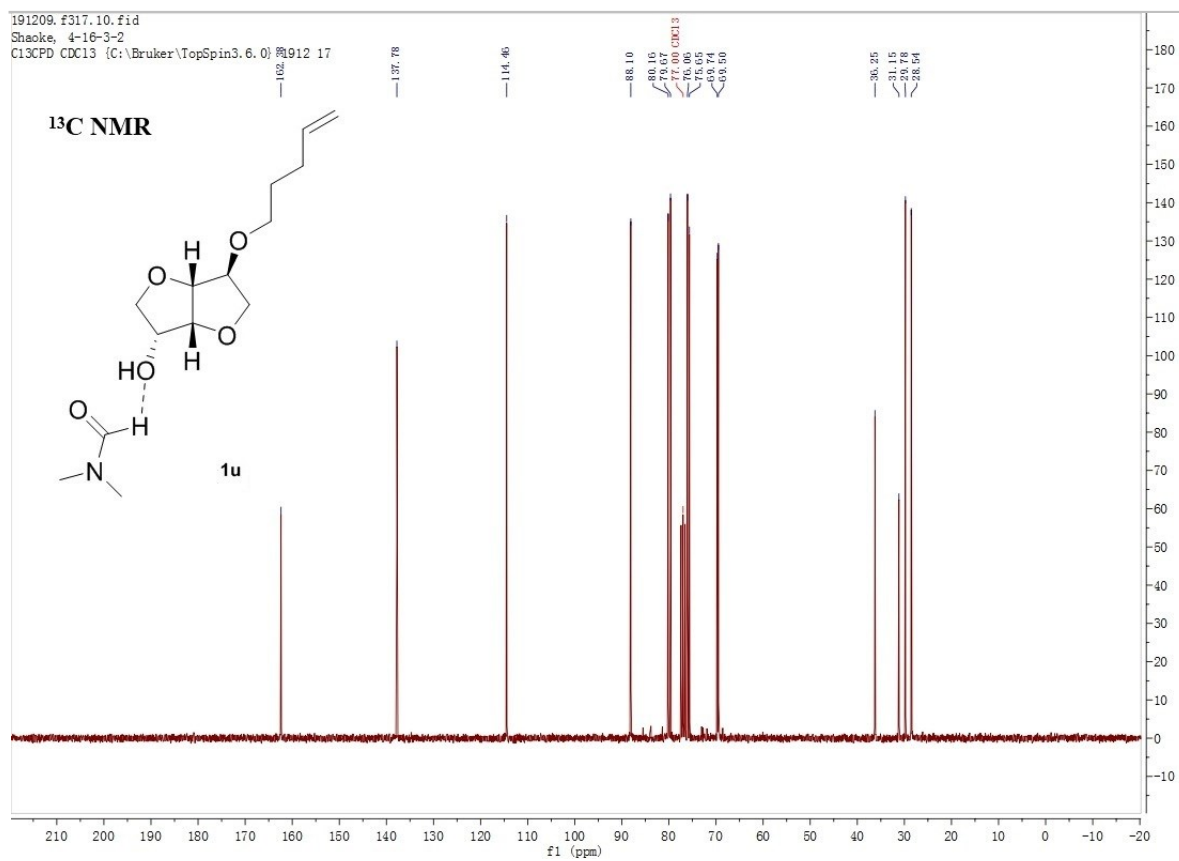
SUPPORTING INFORMATION



SUPPORTING INFORMATION

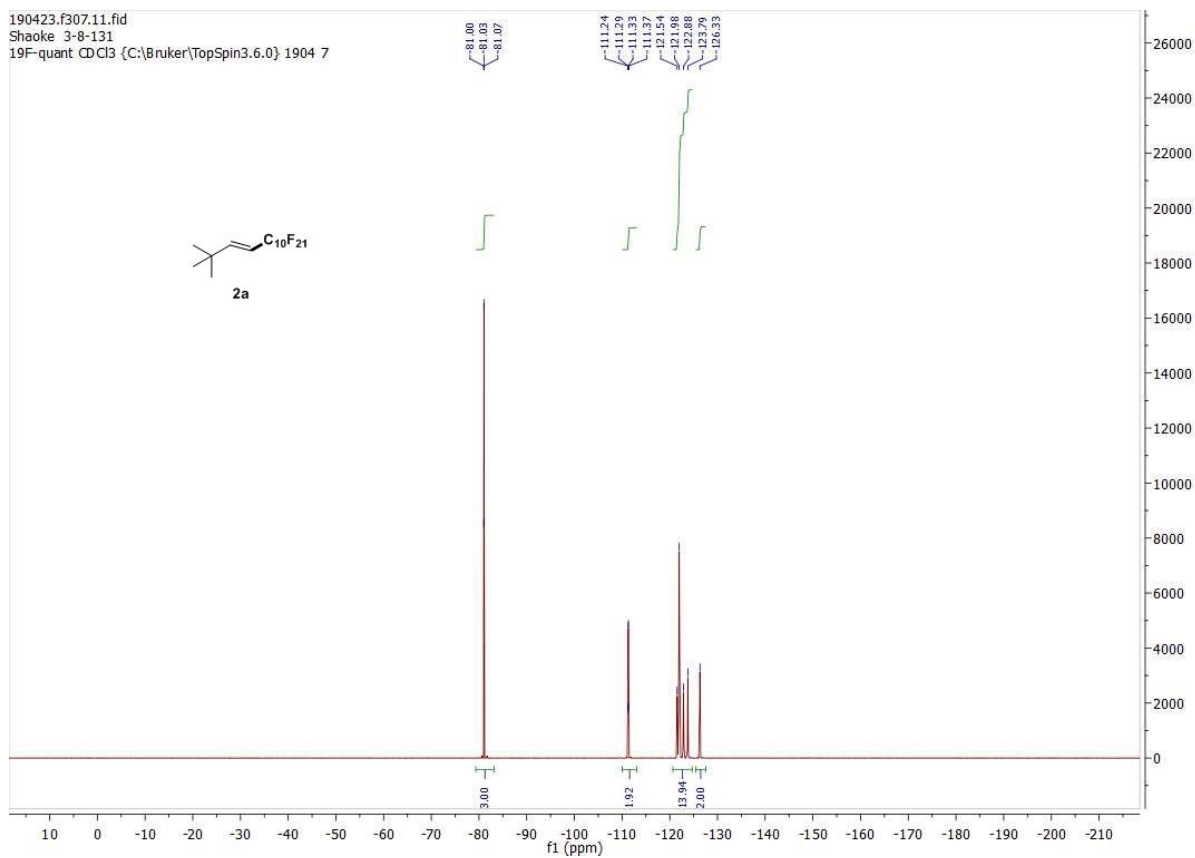
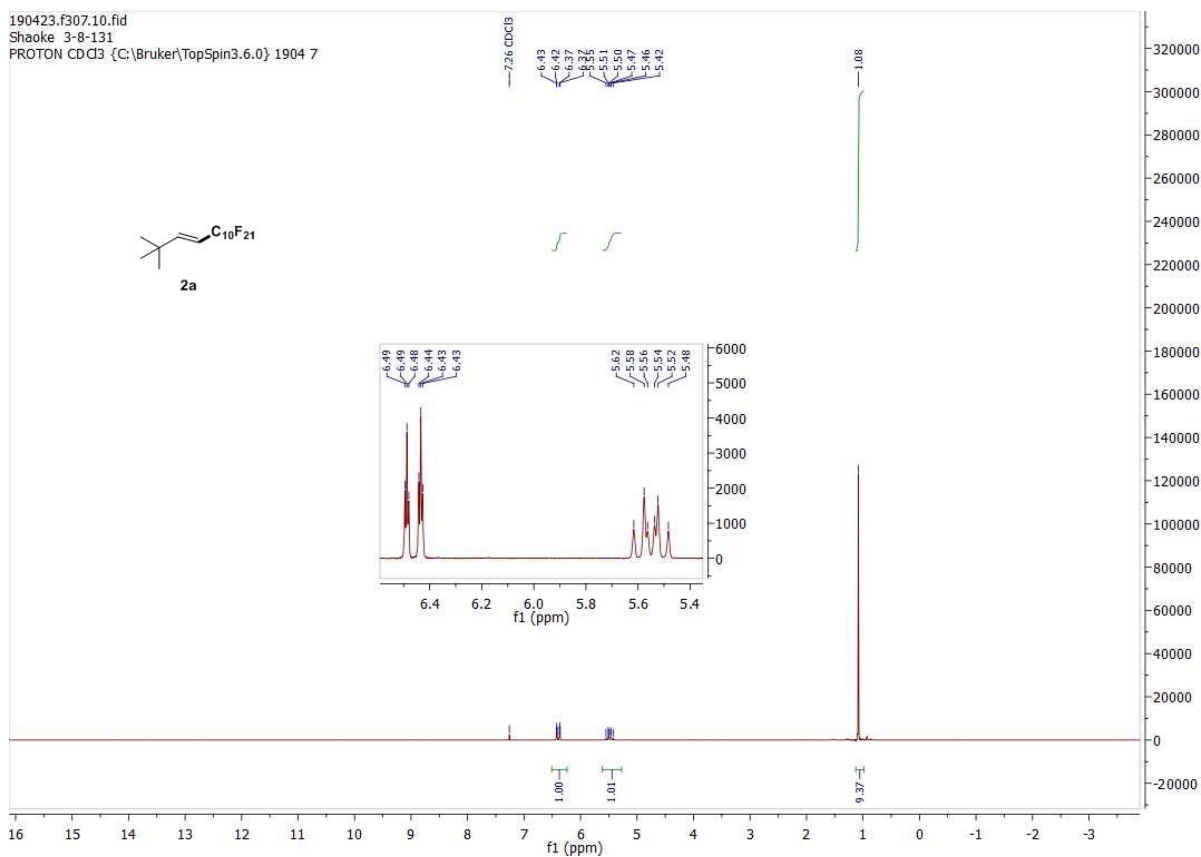


SUPPORTING INFORMATION

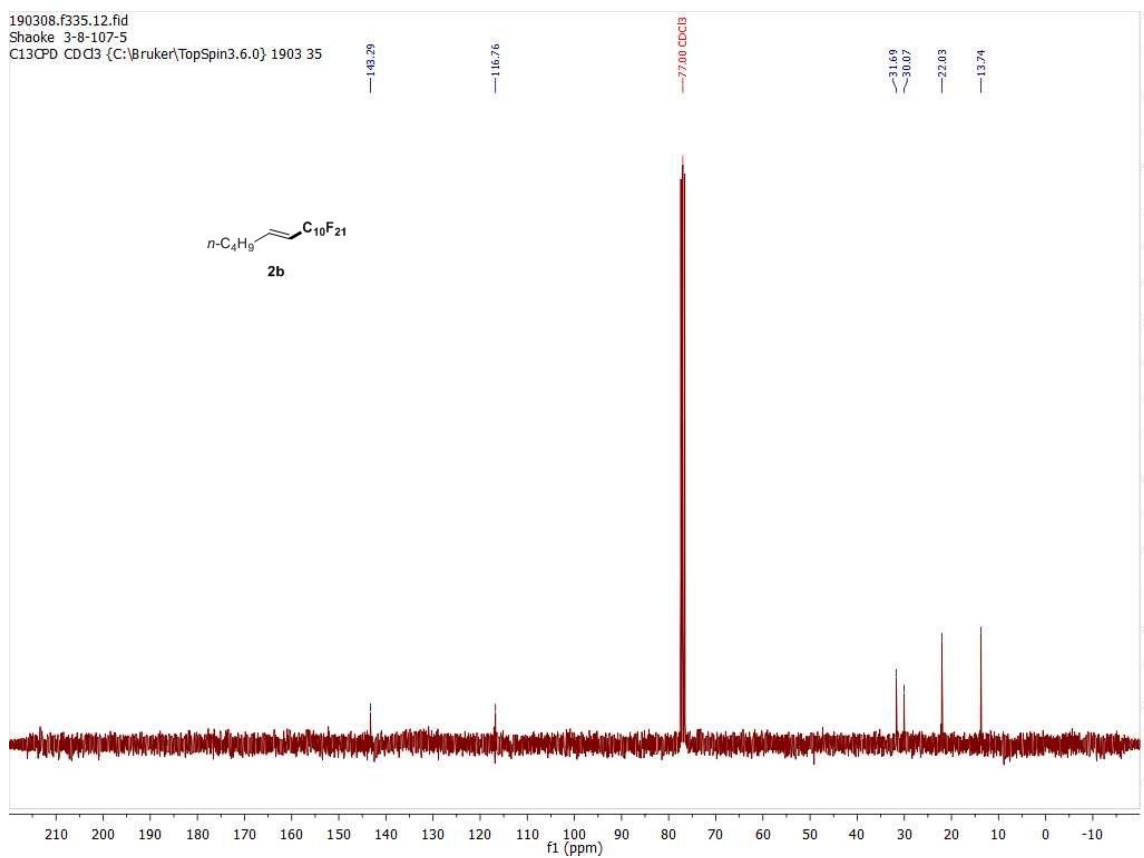
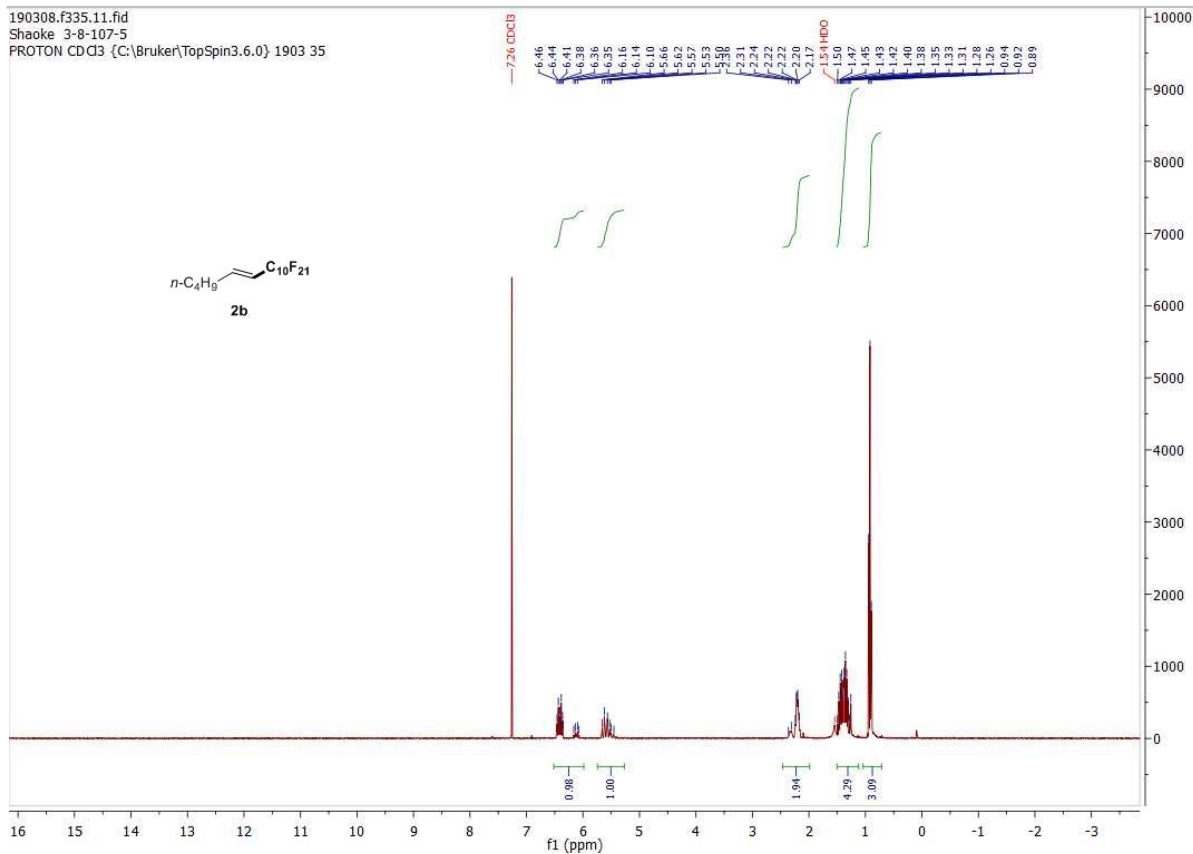


SUPPORTING INFORMATION

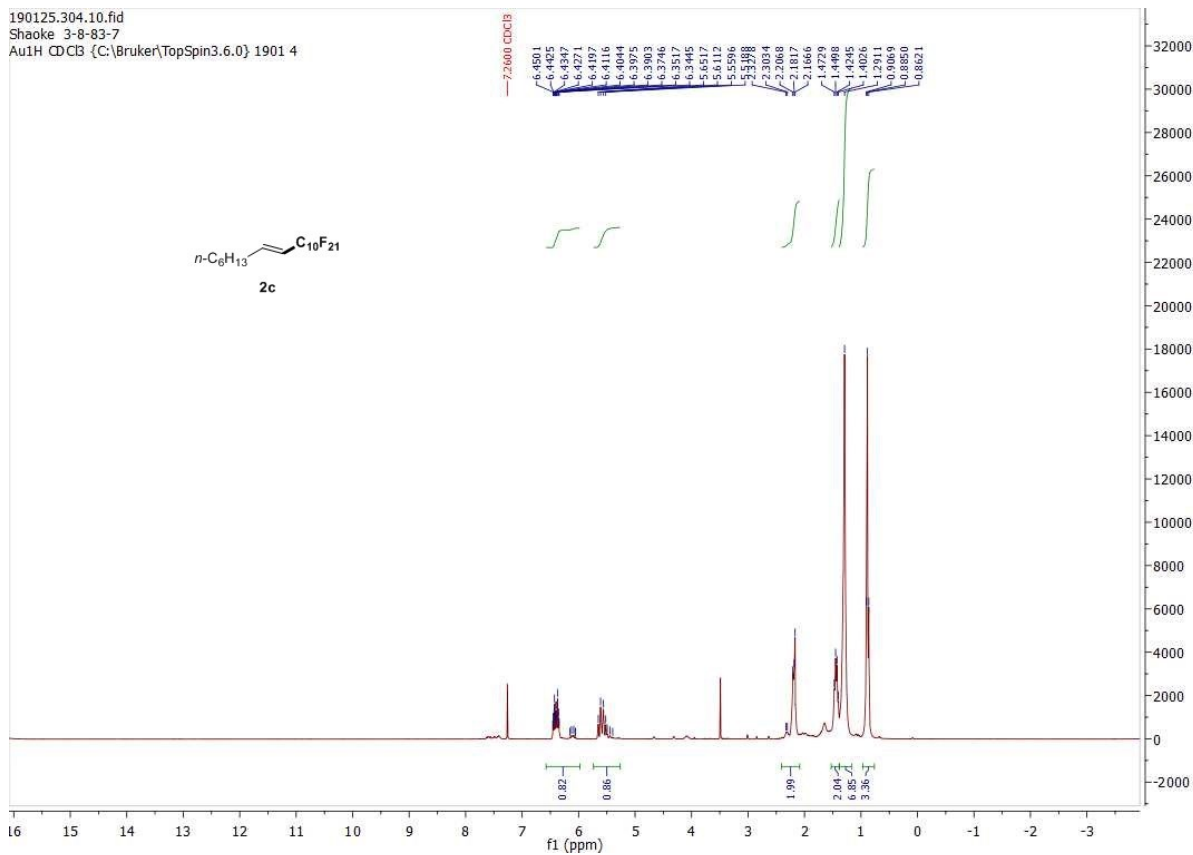
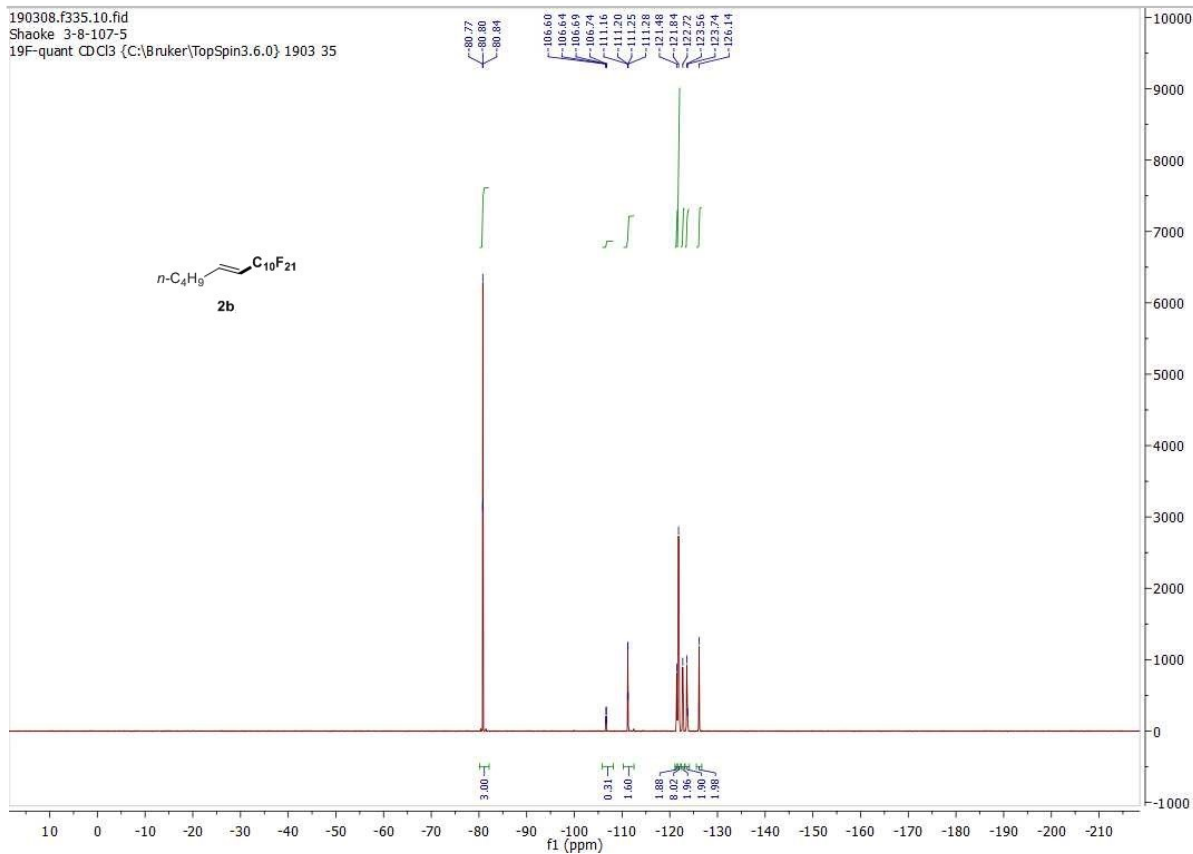
8.2 Spectra of perfluorodecylated alkenes



SUPPORTING INFORMATION

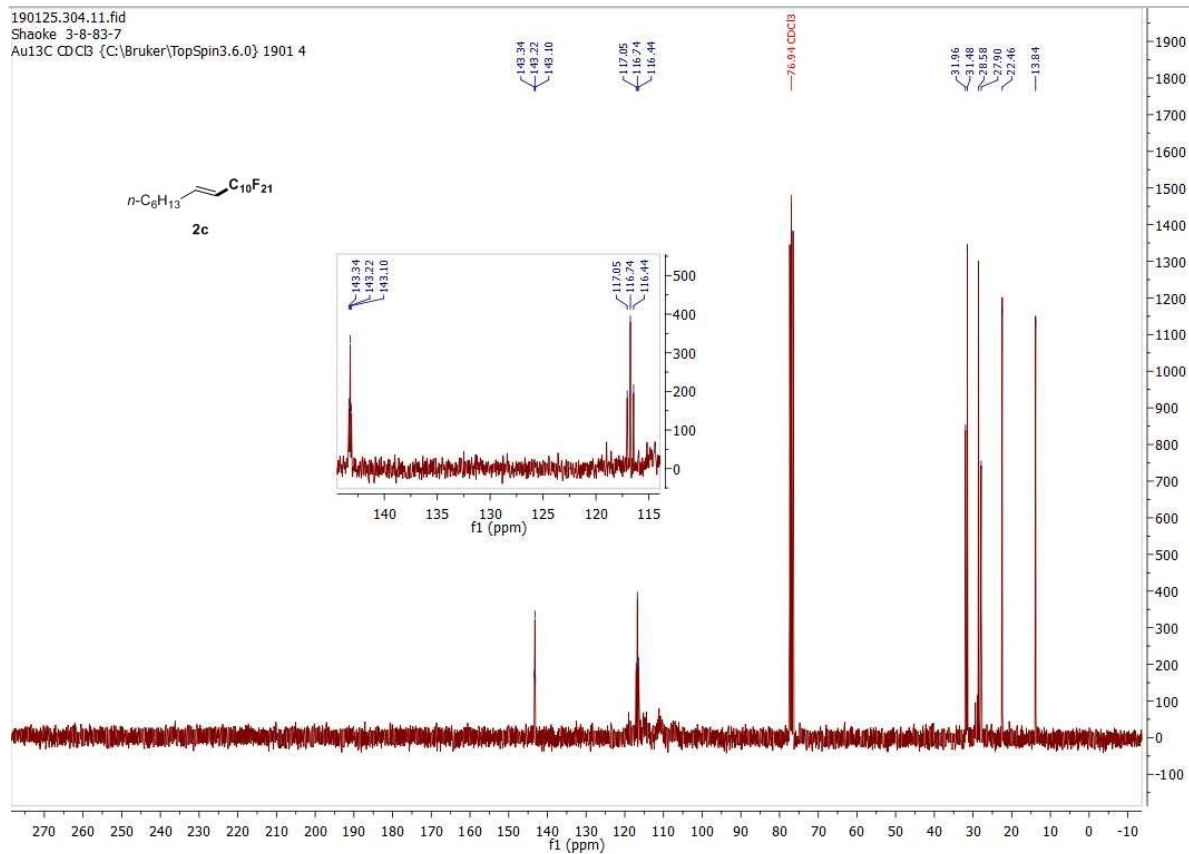


SUPPORTING INFORMATION

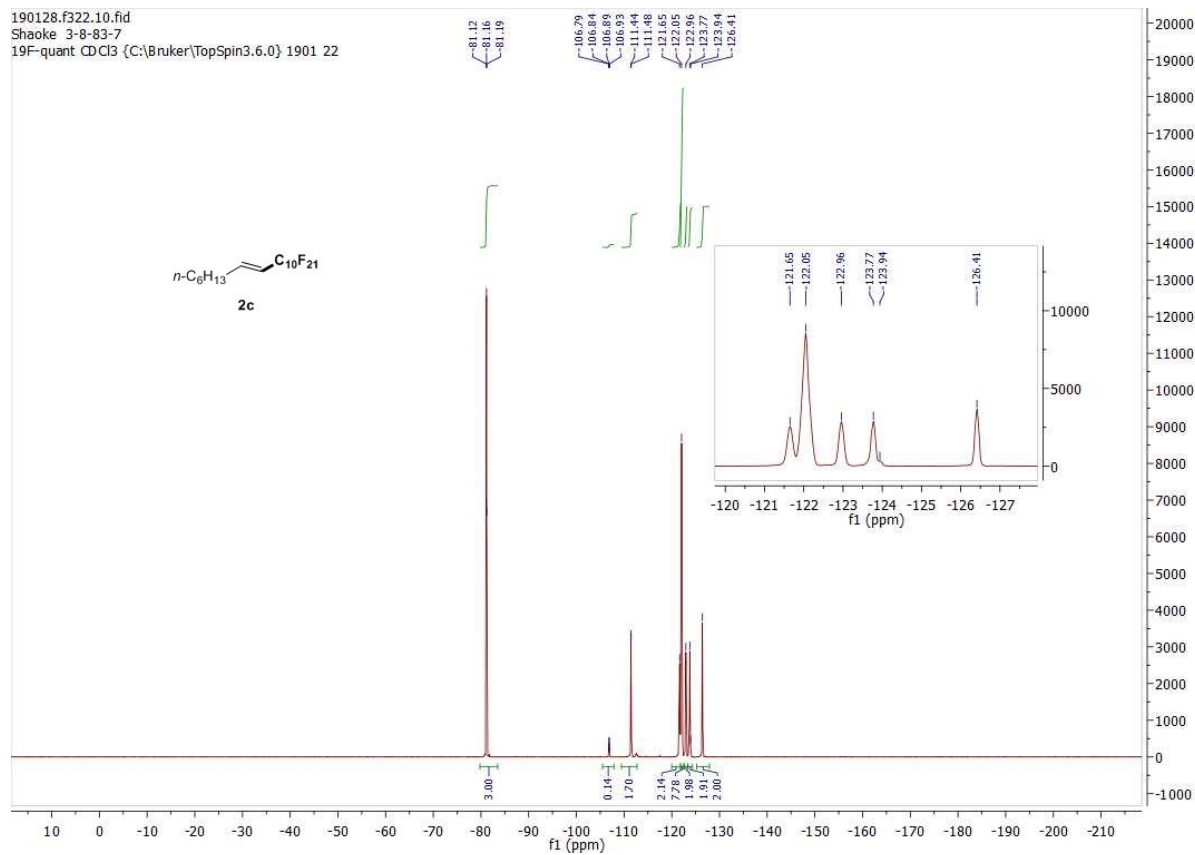


SUPPORTING INFORMATION

190125.304.11.fid
 Shaoke 3-8-83-7
 Au13C CDCl3 {C:\Bruker\TopSpin3.6.0} 1901 4

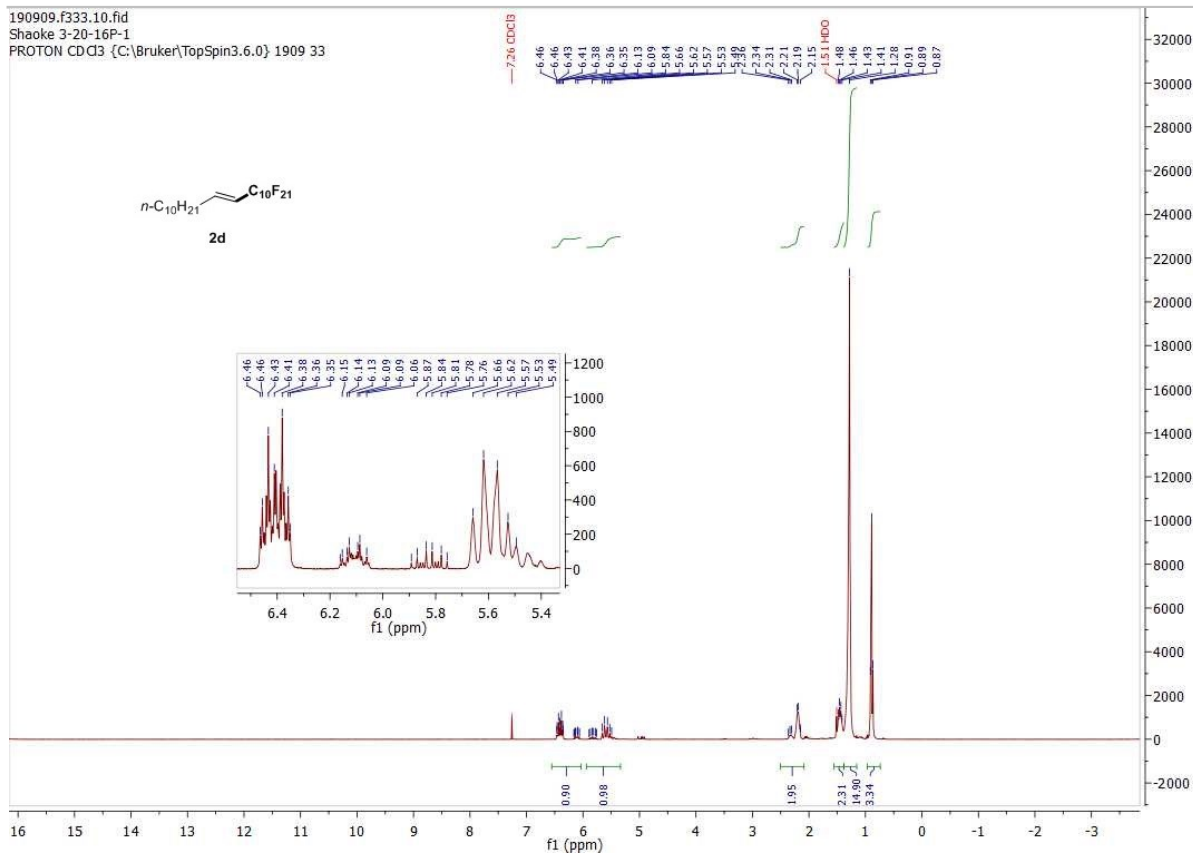


190128.f322.10.fid
 Shaoke 3-8-83-7
 19F-quant CDCl3 {C:\Bruker\TopSpin3.6.0} 1901 22

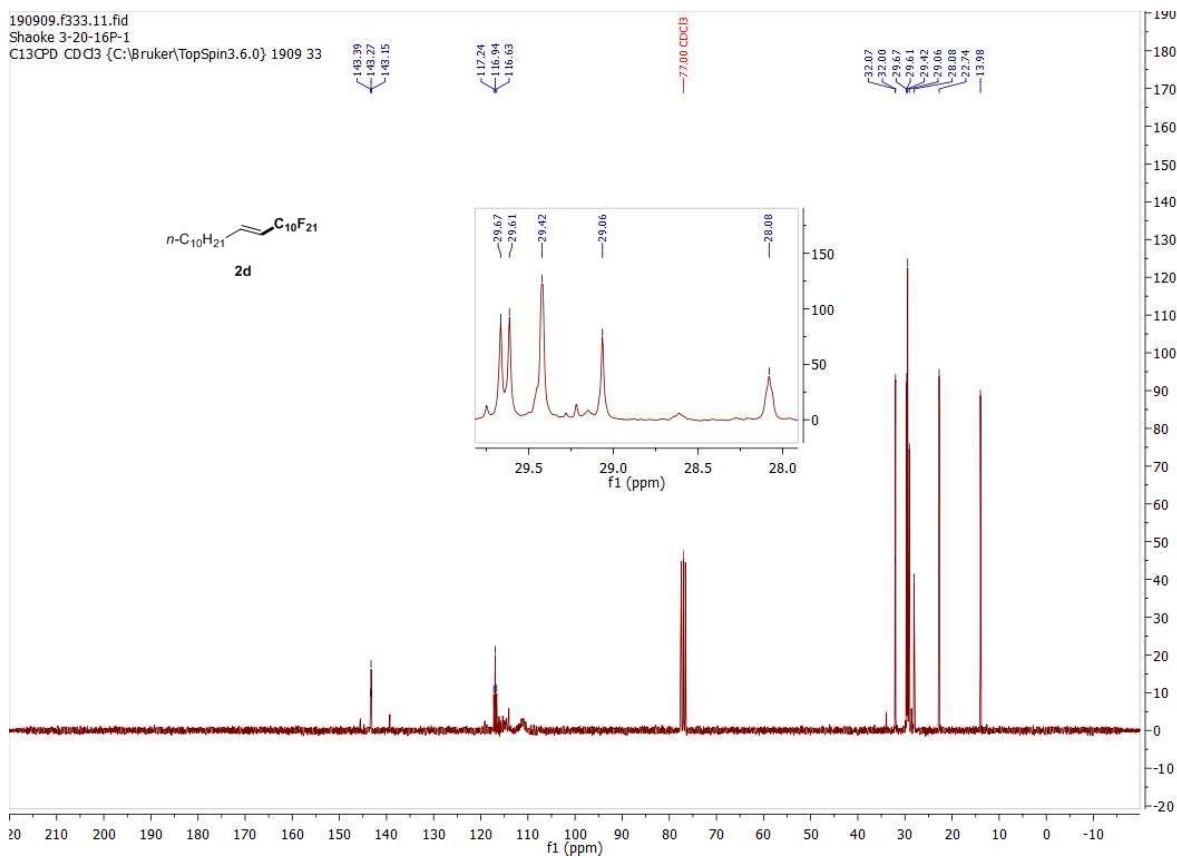


SUPPORTING INFORMATION

190909.f333.10.fid
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 PROTON CDCl₃ {C:\Bruker\TopSpin3.6.0} 1909 33

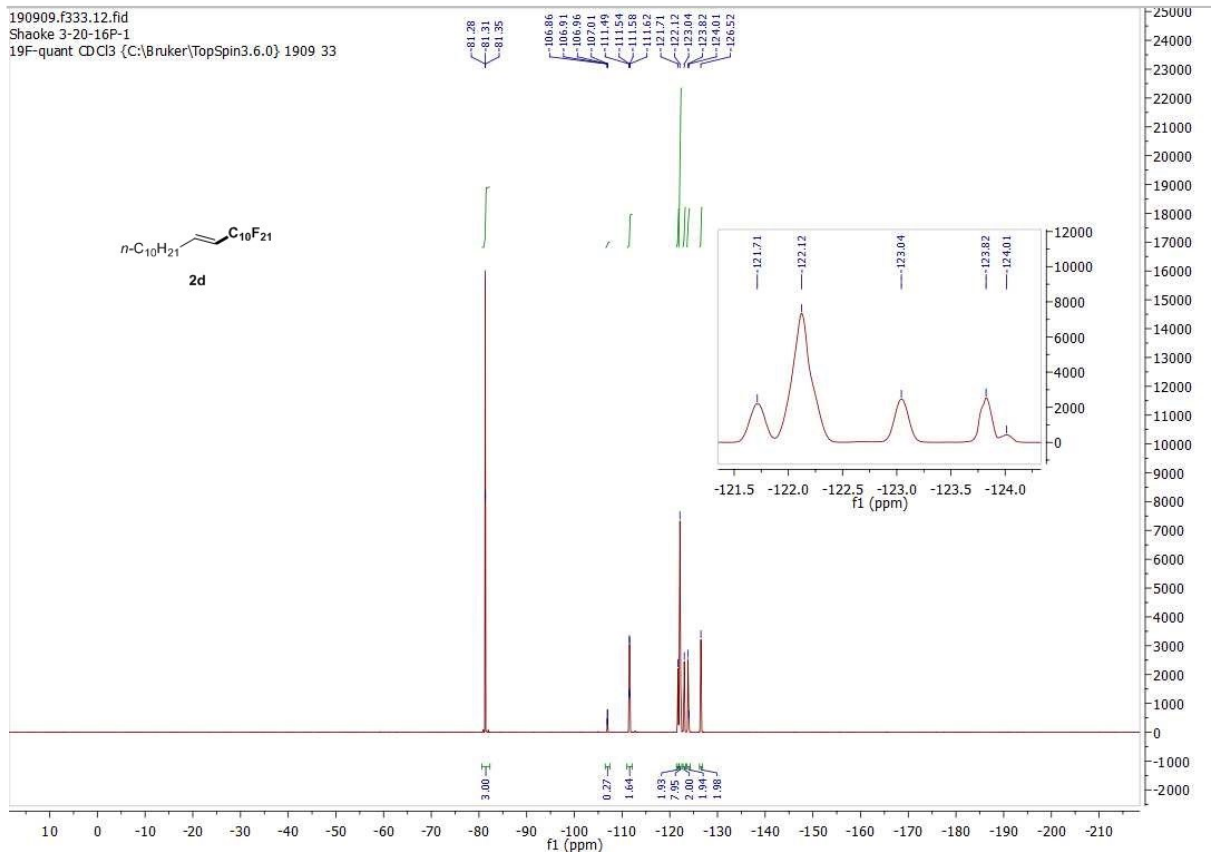


190909.f333.11.fid
 Shaok 3-20-16P-1
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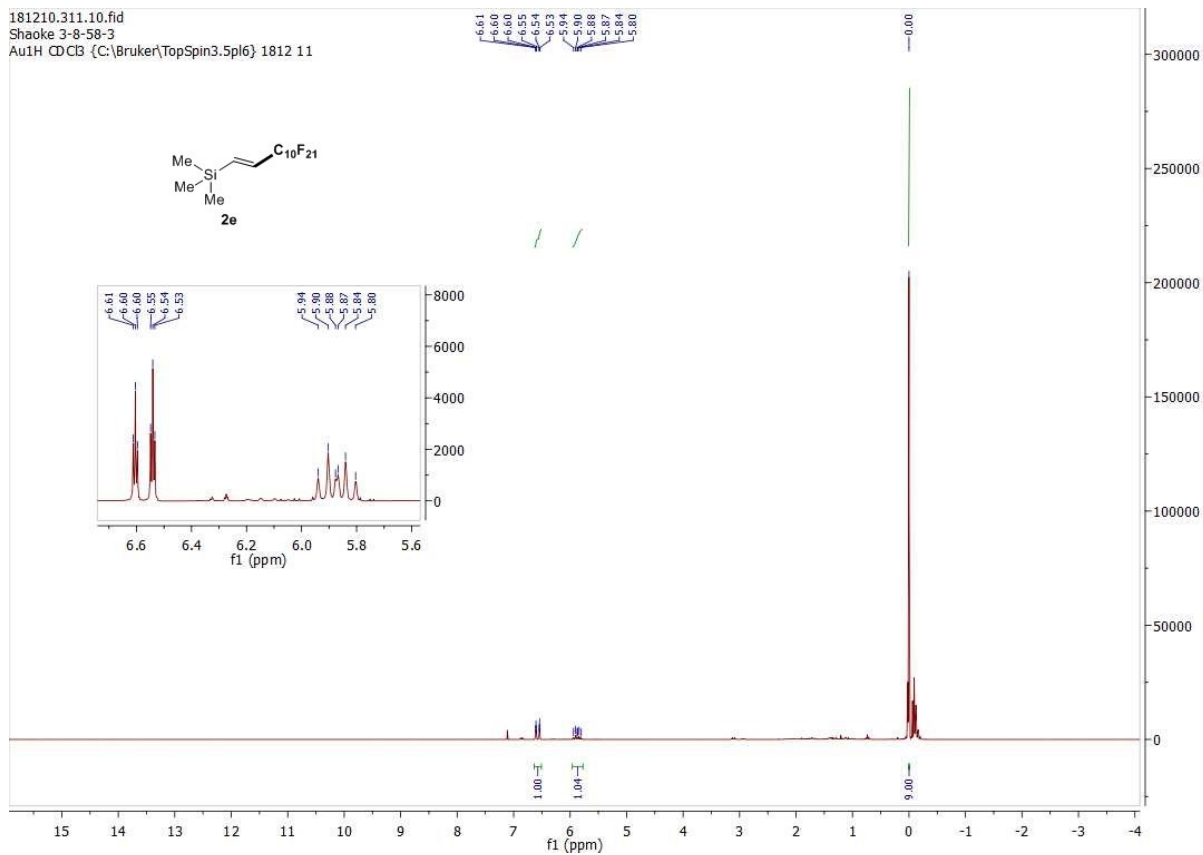


SUPPORTING INFORMATION

190909.f333.12.fid
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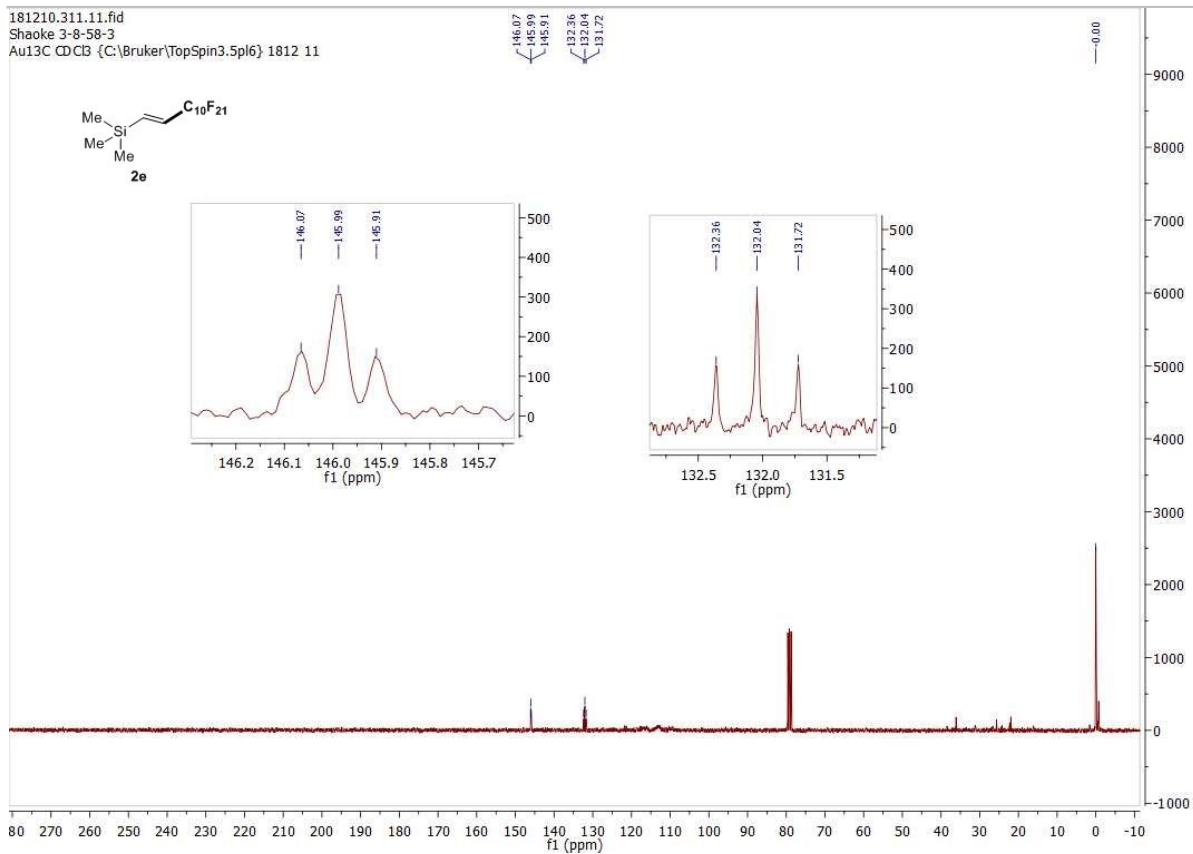


181210.311.10.fid
 Shaoke 3-8-58-3
 Au1H CDCl3 {C:\Bruker\TopSpin3.5pl6} 1812 11

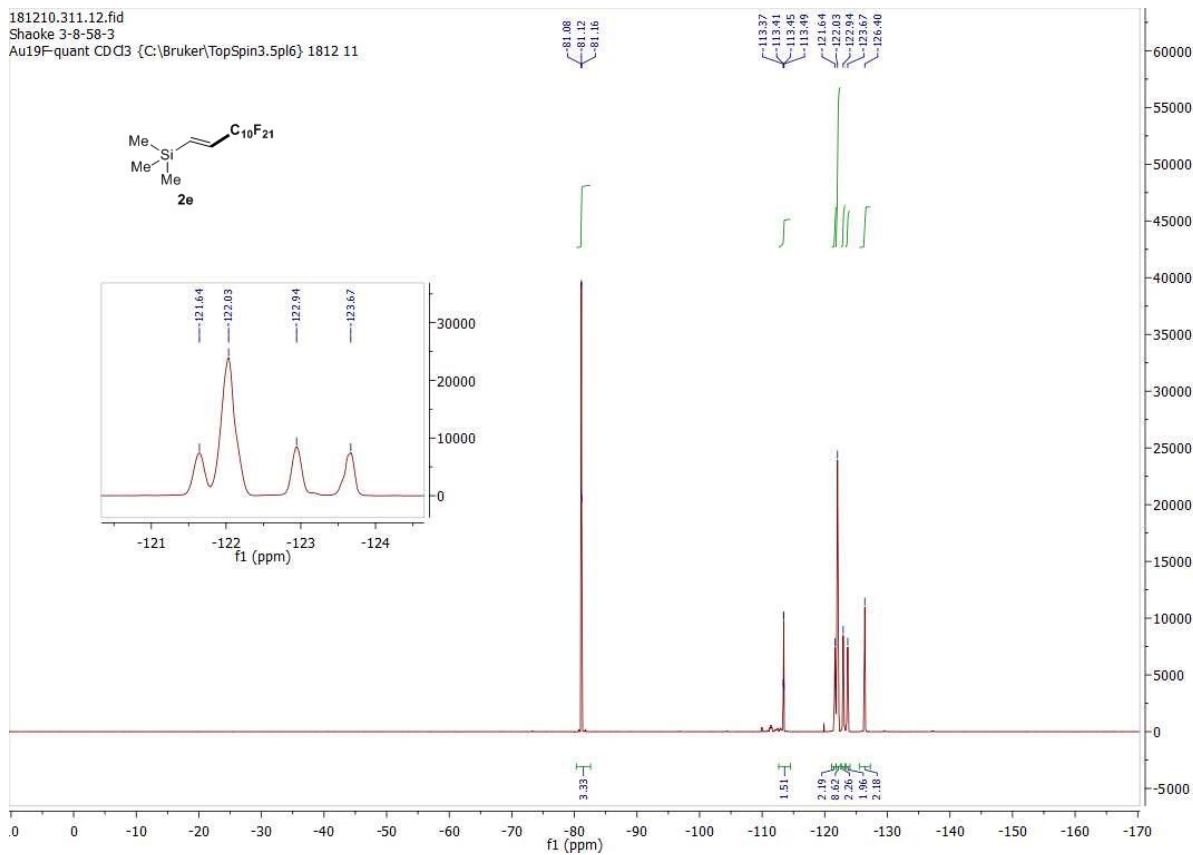


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181210.311.11.fid
 Shaoke 3-8-58-3
 Au13C CDCl3 {C:\Bruker\TopSpin3.5pl6} 1812 11

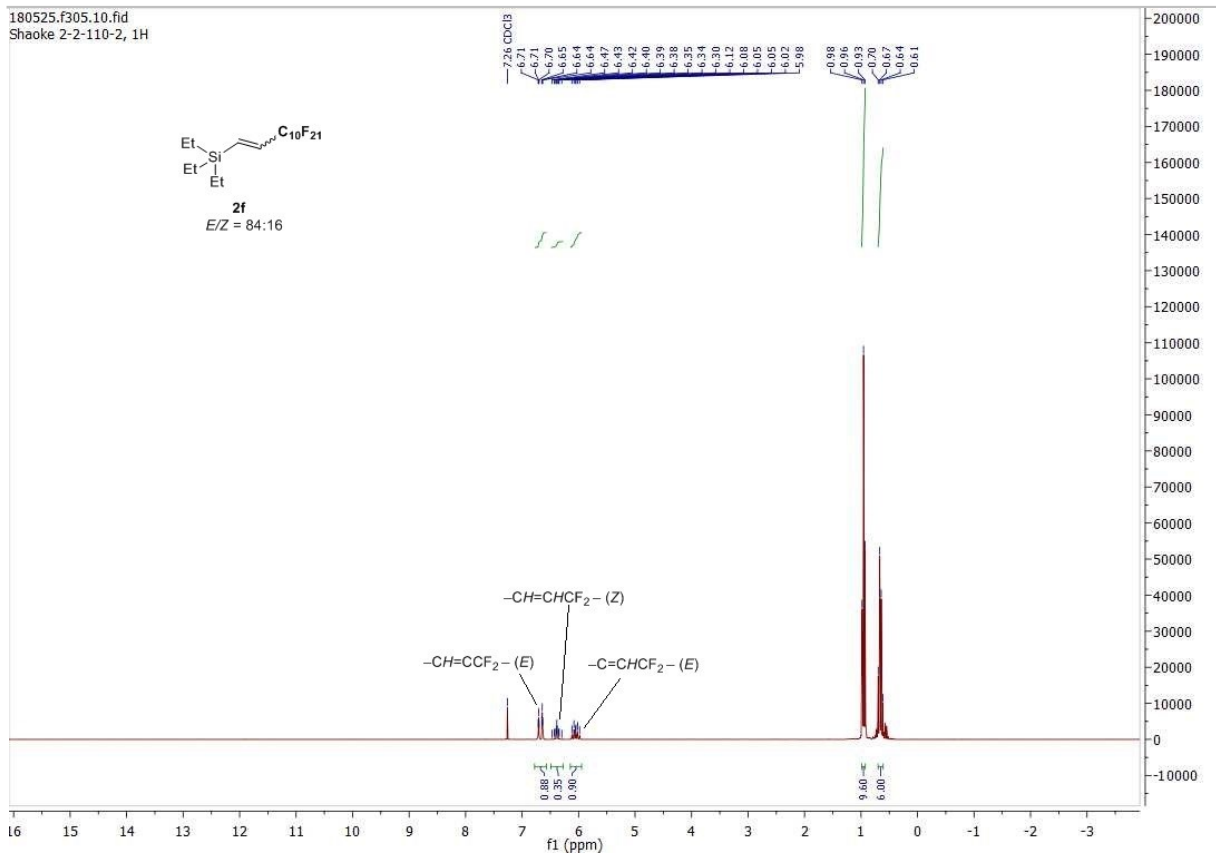


181210.311.12.fid
 Shaoke 3-8-58-3
 Au19F-quant CDCl3 {C:\Bruker\TopSpin3.5pl6} 1812 11

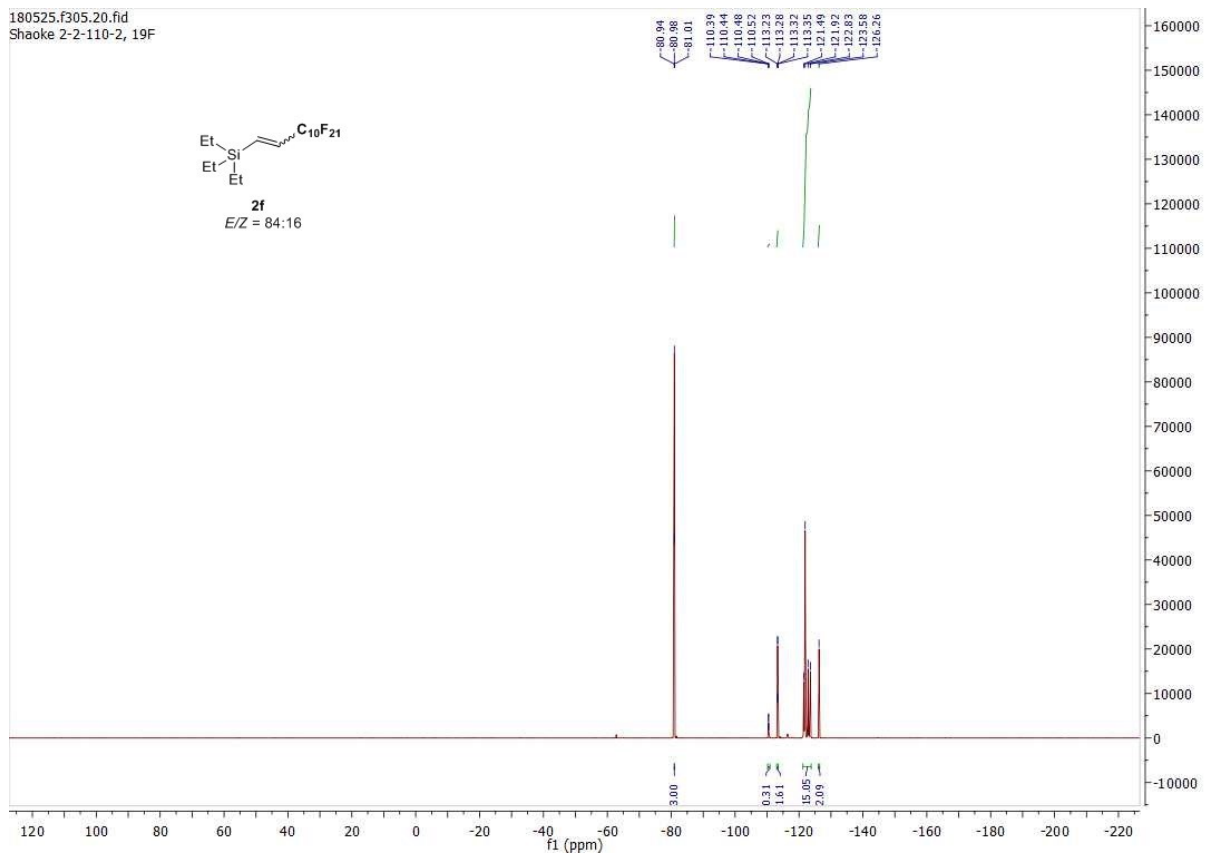


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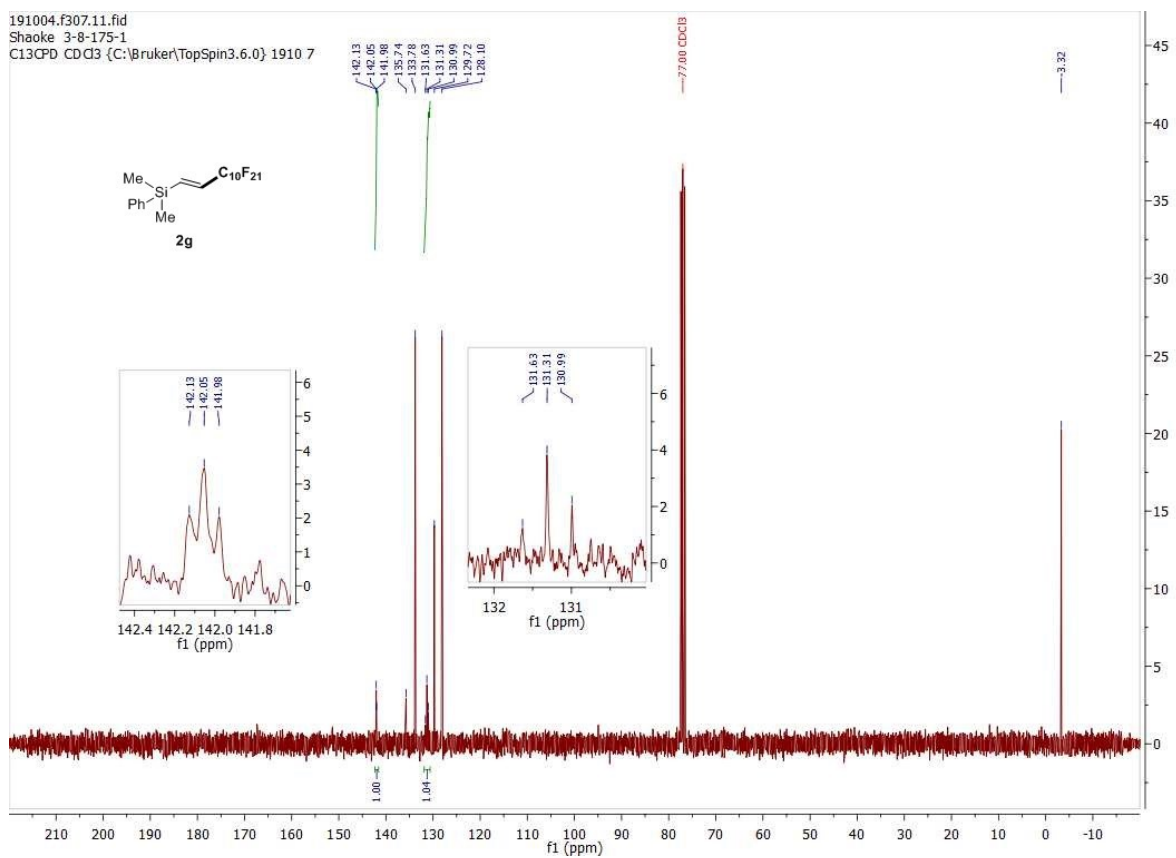
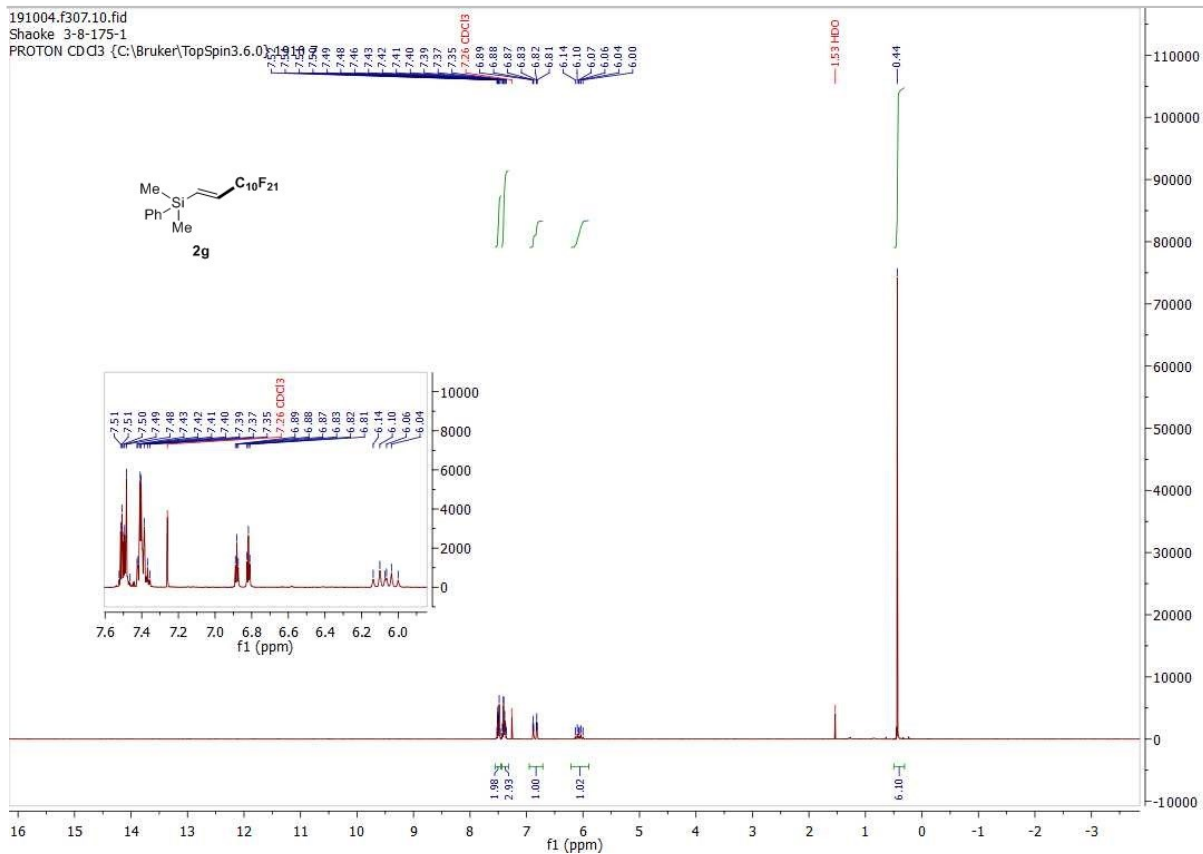
180525.F305.10.fid
Shaoke 2-2-110-2, 1H



180525.F305.20.fid
Shaoke 2-2-110-2, 19F

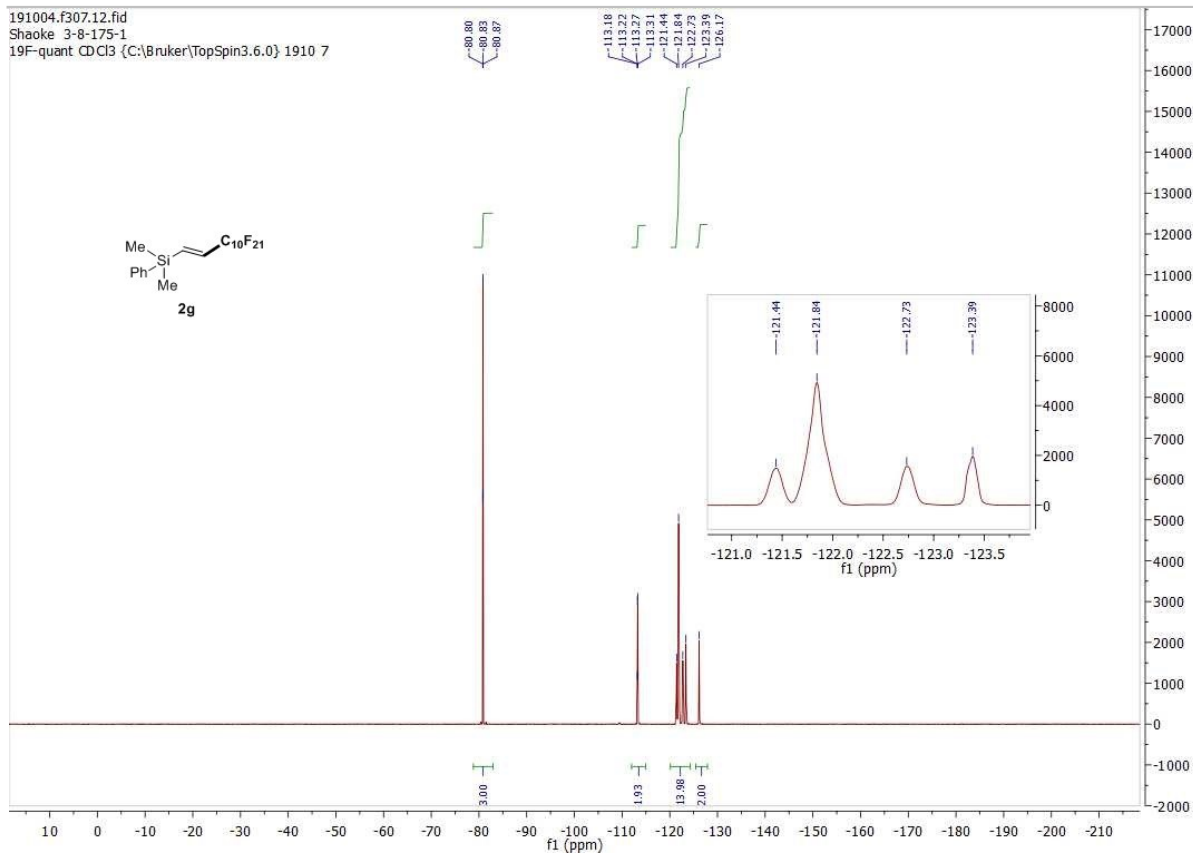


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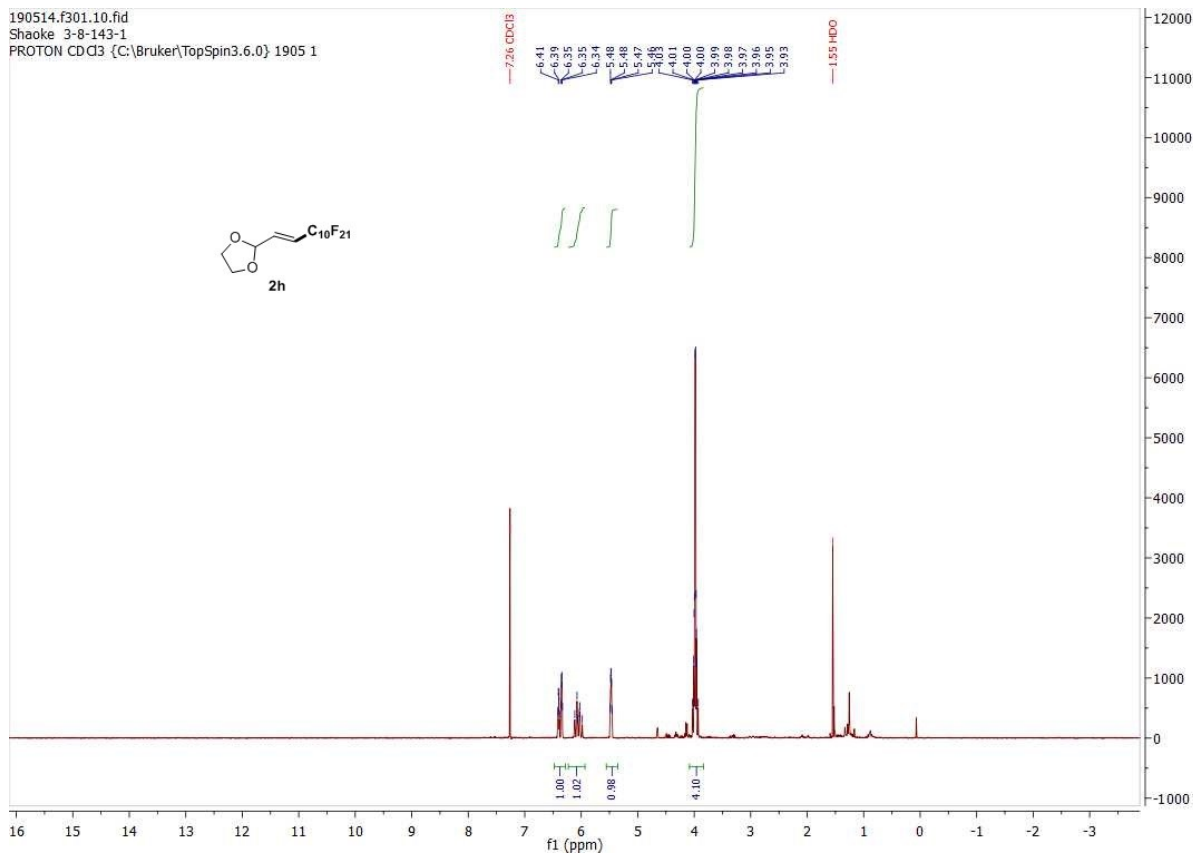


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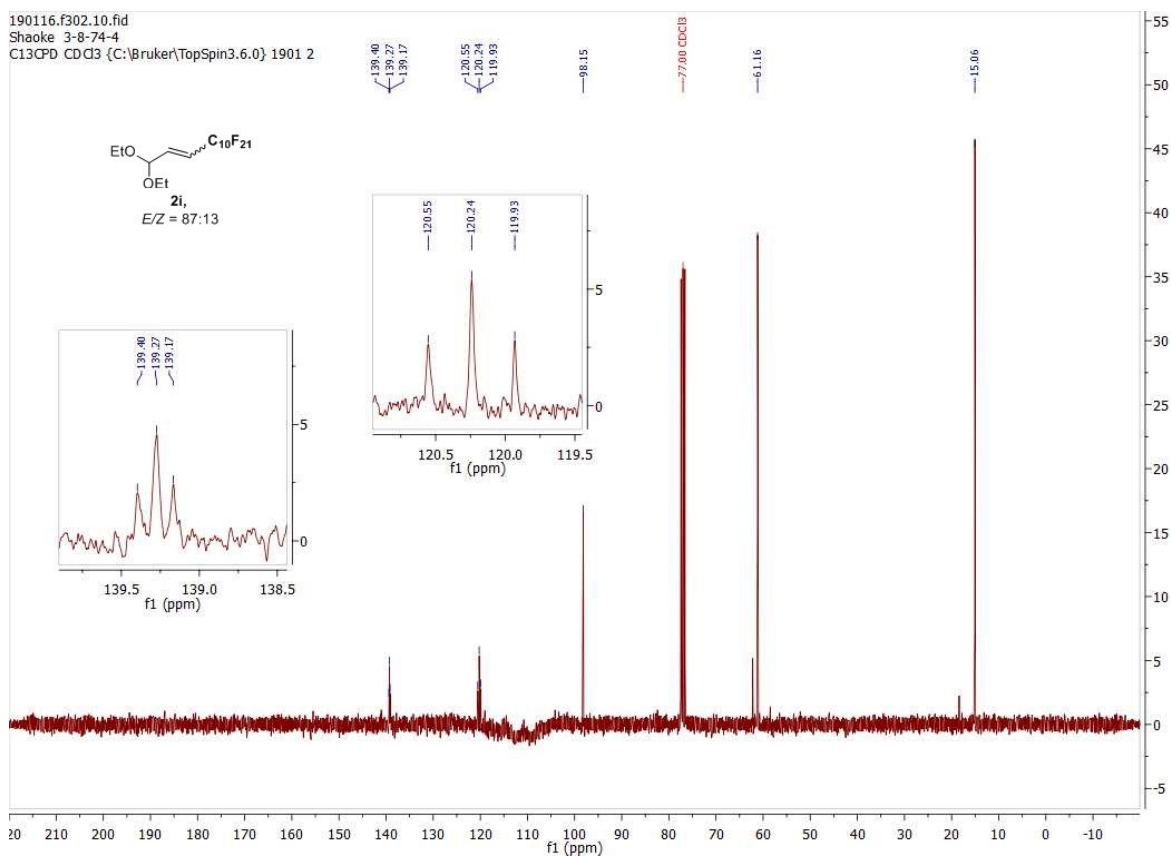
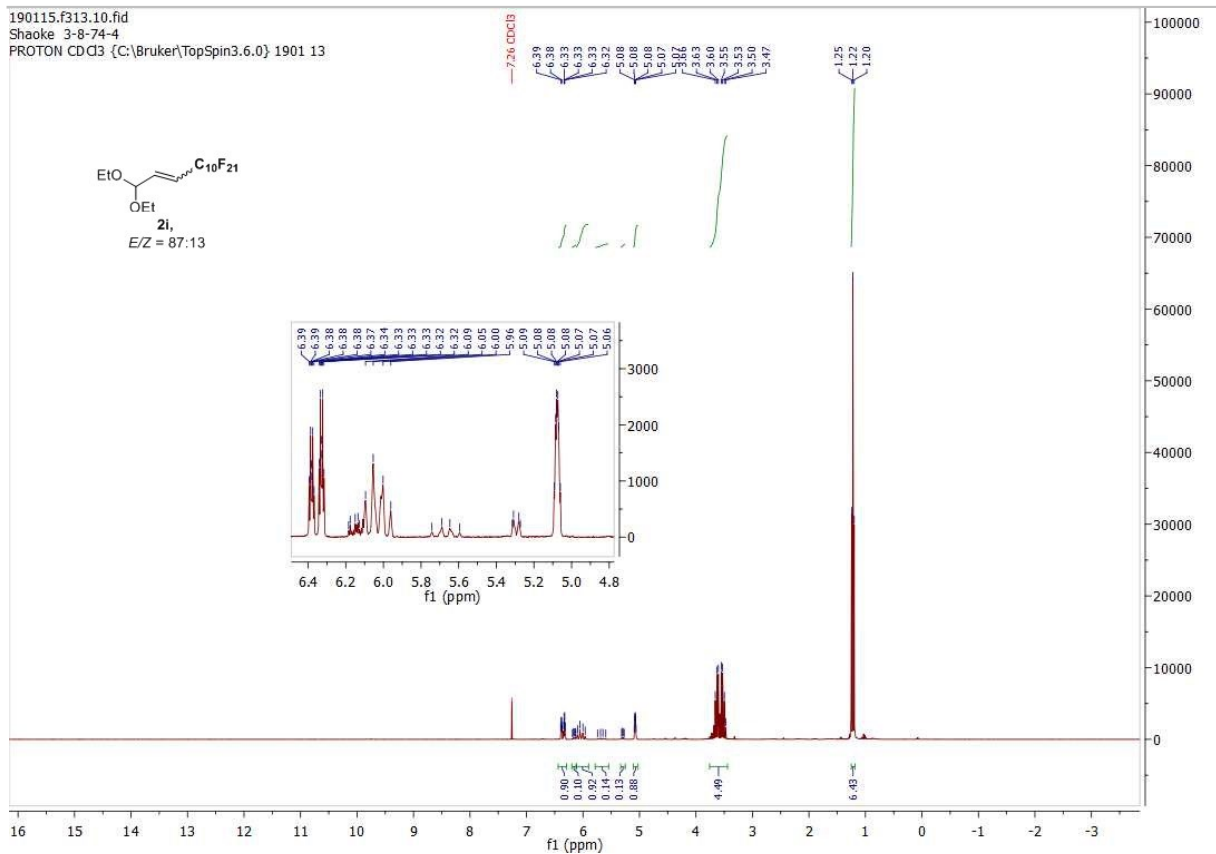
191004.f307.12.fid
 Shaoke 3-8-175-1
 19F-quant CDCl3 {C:\Bruker\TopSpin3.6.0} 1910 7



190514.f301.10.fid
 Shaoke 3-8-143-1
 PROTON CDCl3 {C:\Bruker\TopSpin3.6.0} 1905 1

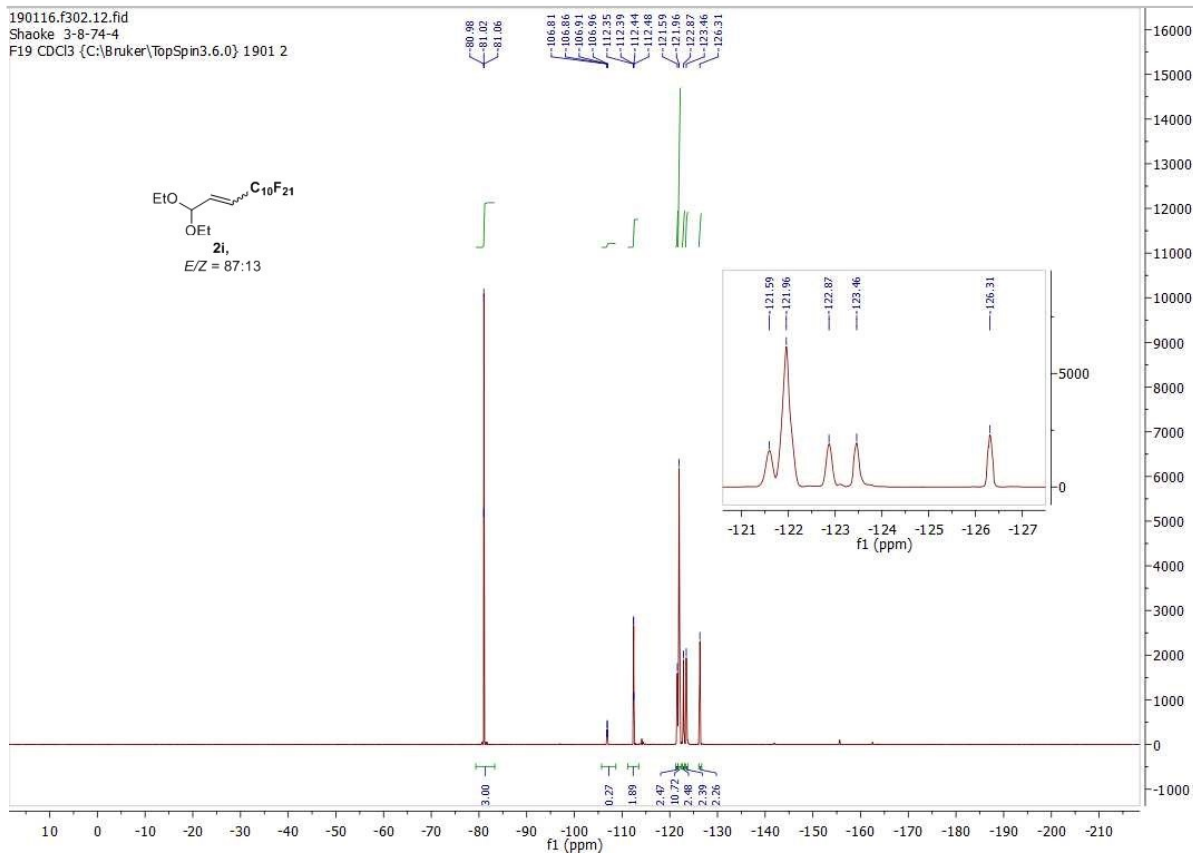


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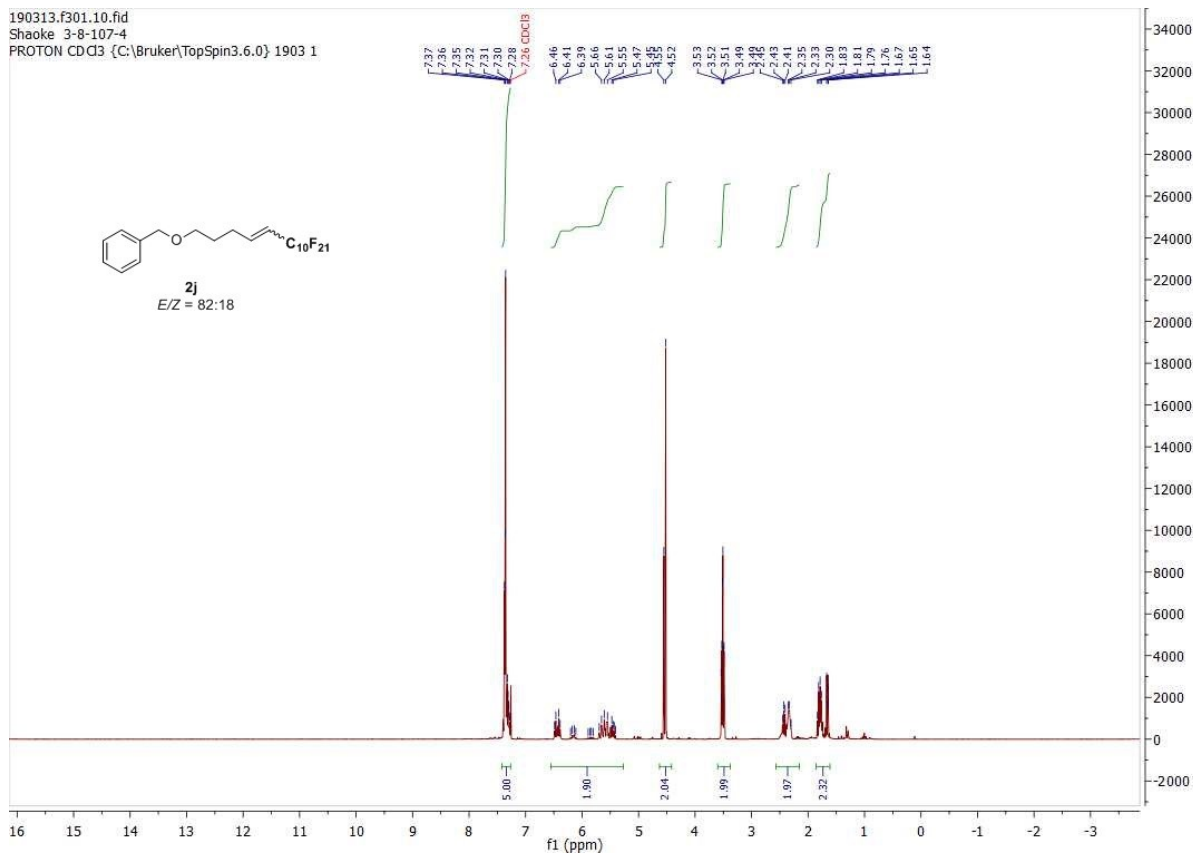


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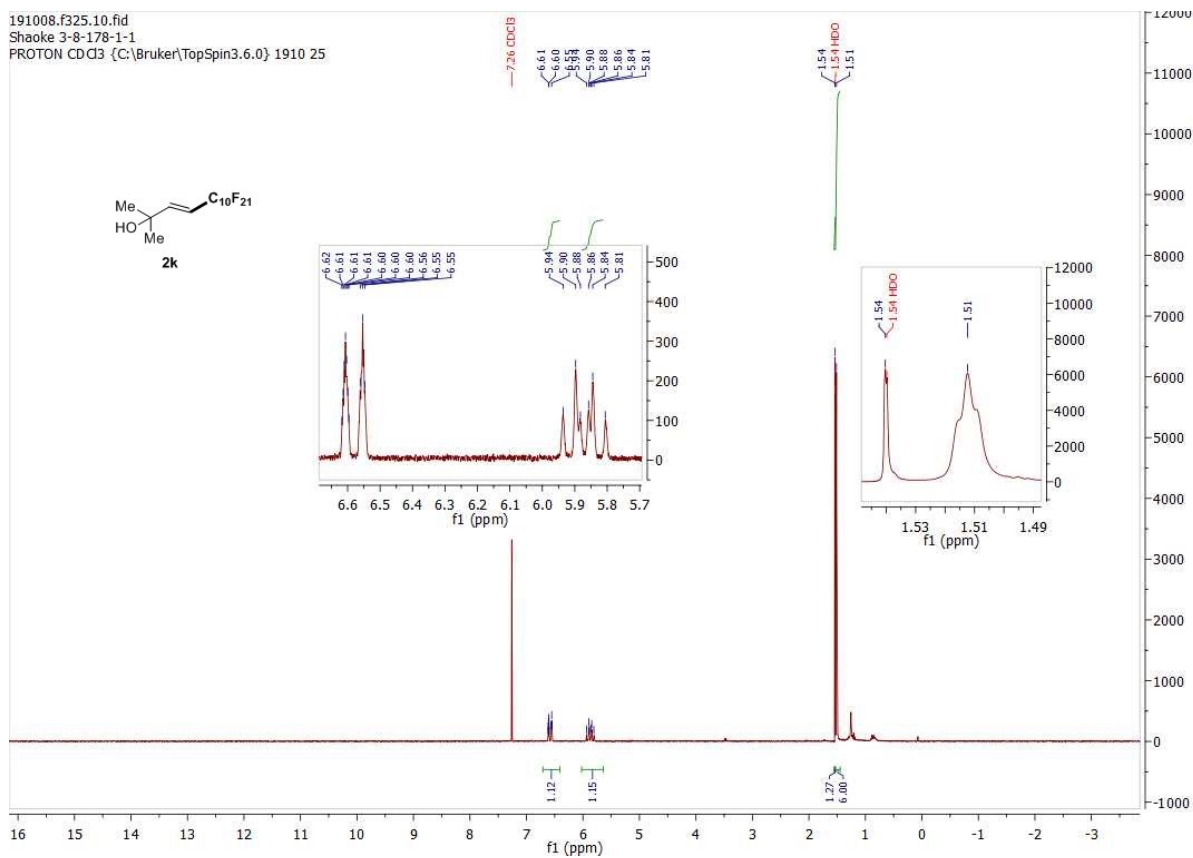
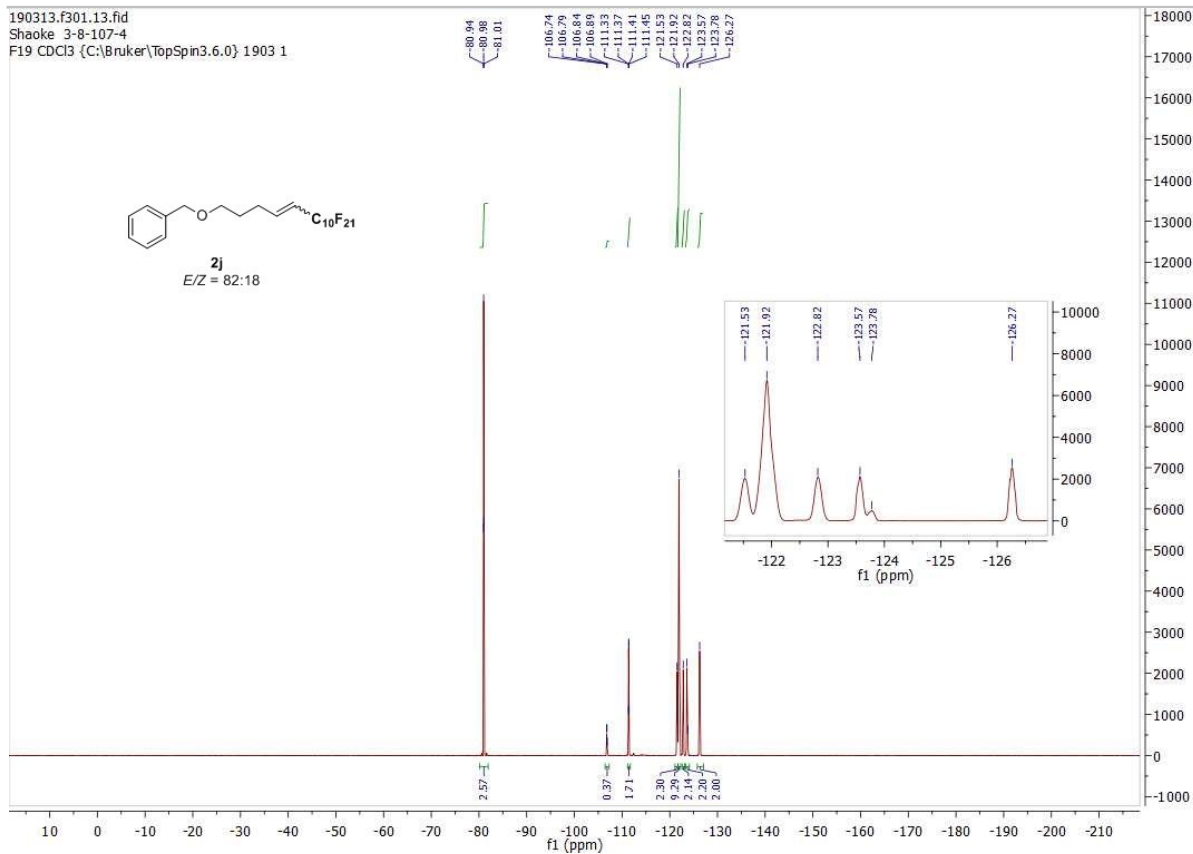
190116.f302.12.fid
 Shaoke 3-8-74-4
 F19 CDCl3 {C:\Bruker\TopSpin3.6.0} 1901 2



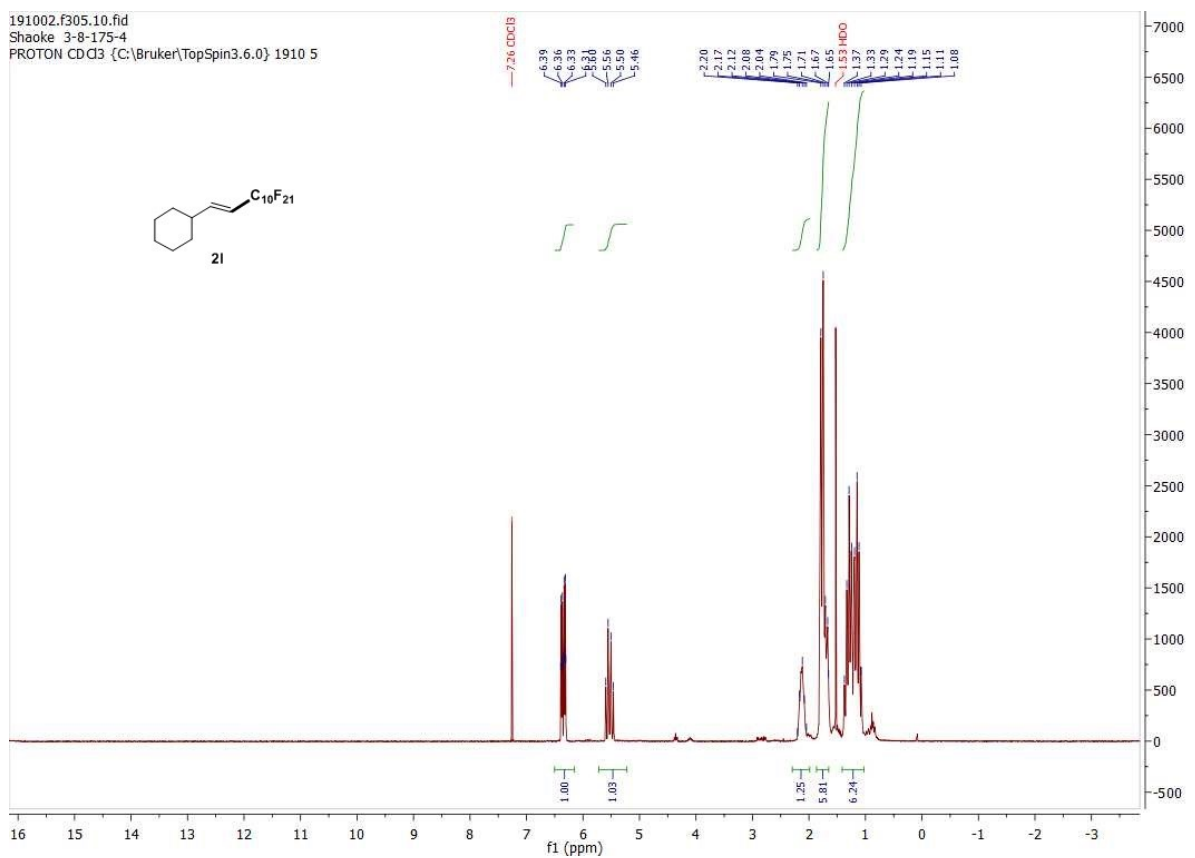
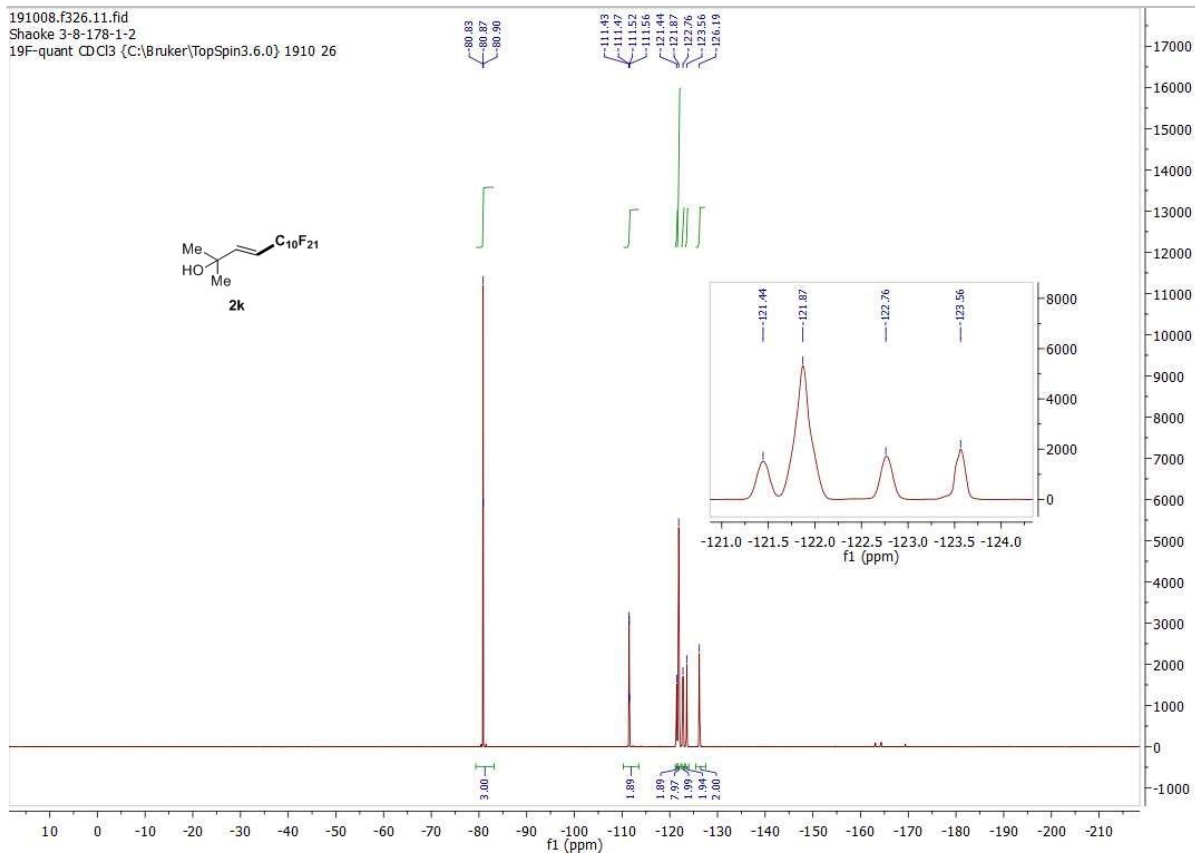
190313.f301.10.fid
 Shaoke 3-8-107-4
 PROTON CDCl3 {C:\Bruker\TopSpin3.6.0} 1903 1



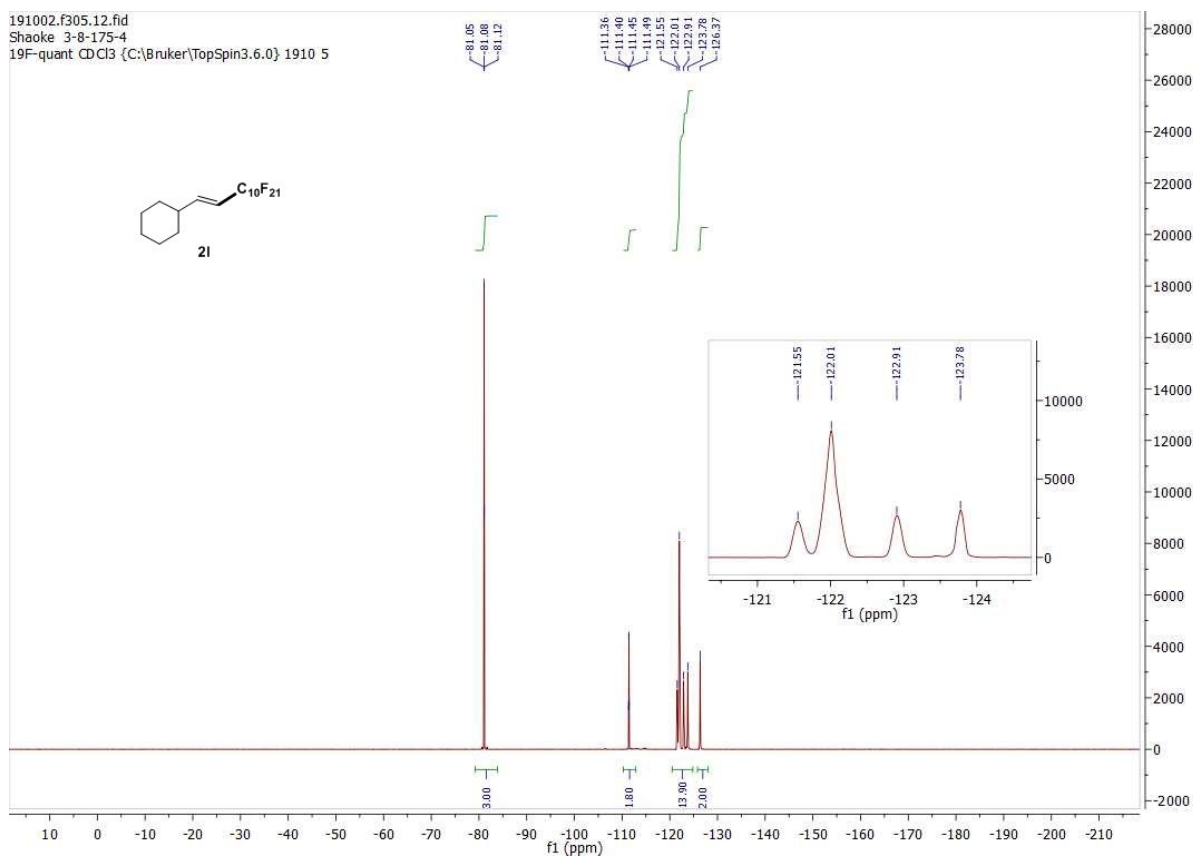
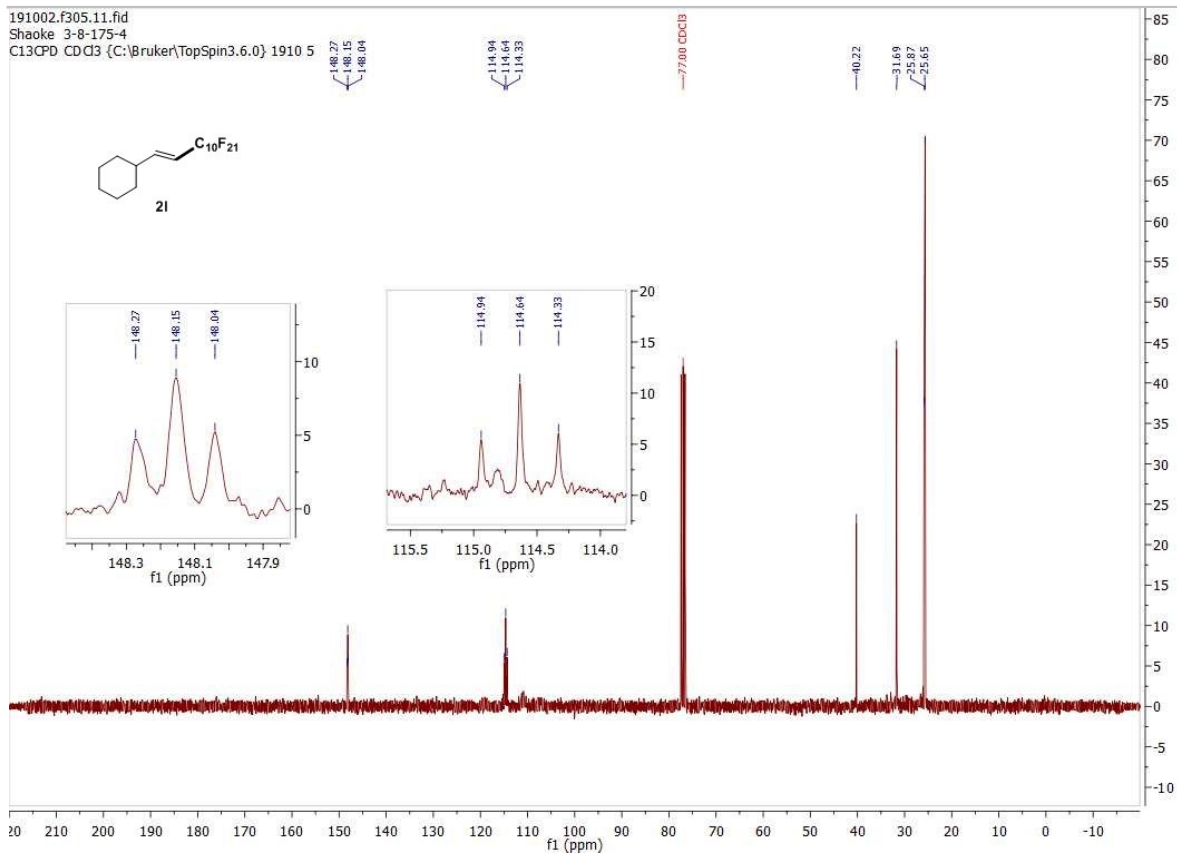
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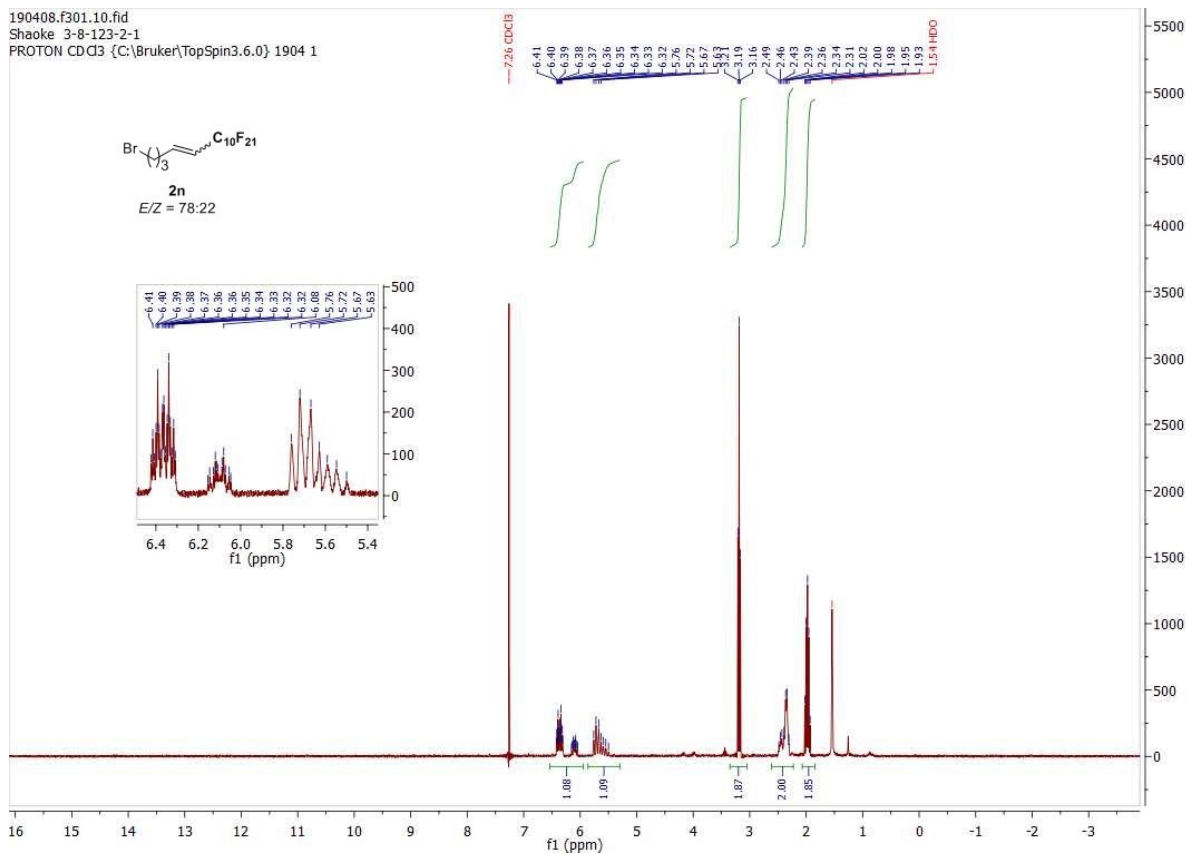
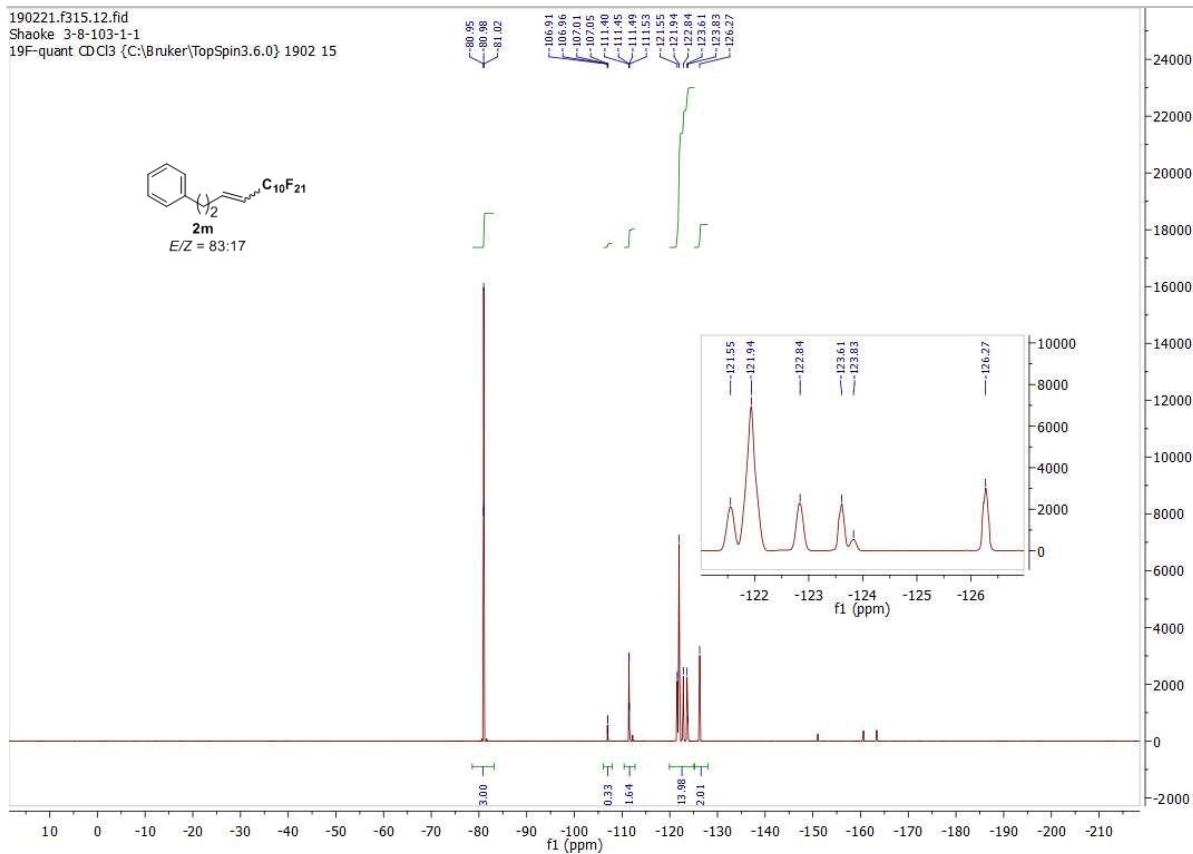
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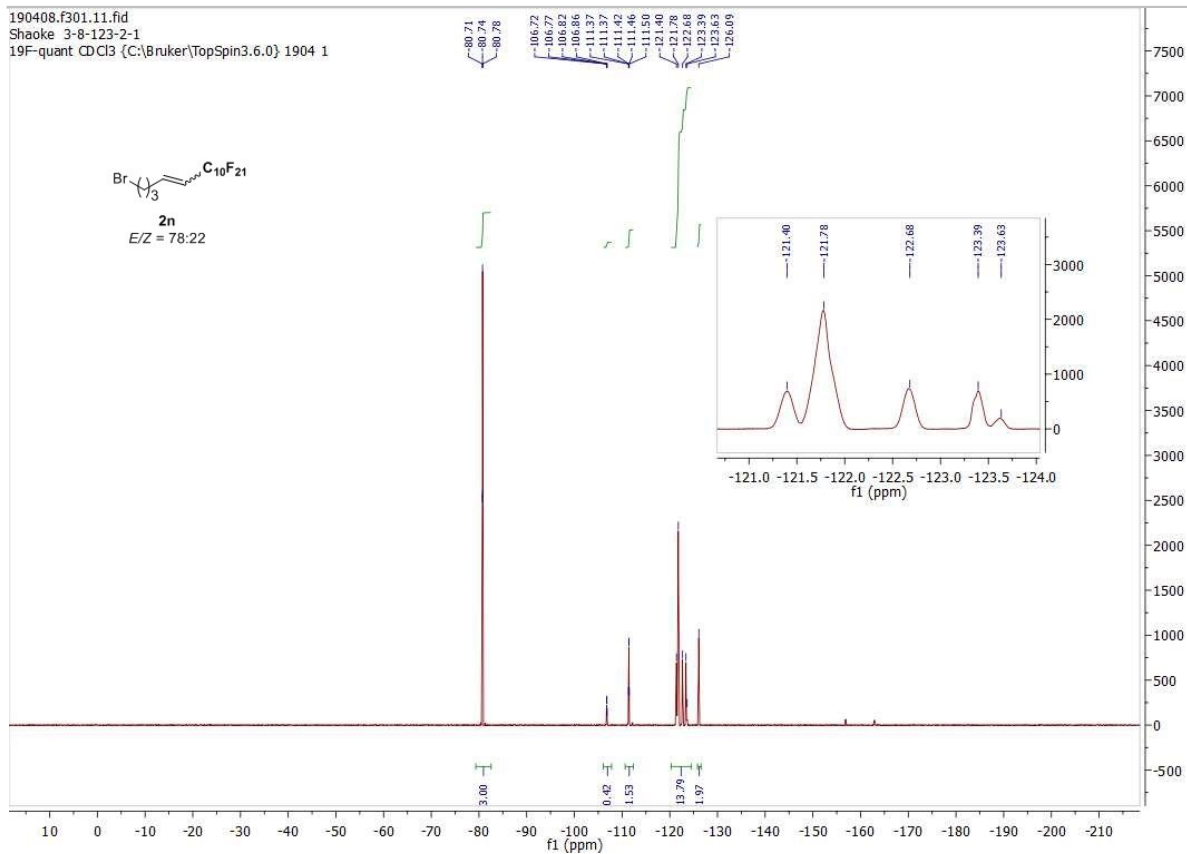


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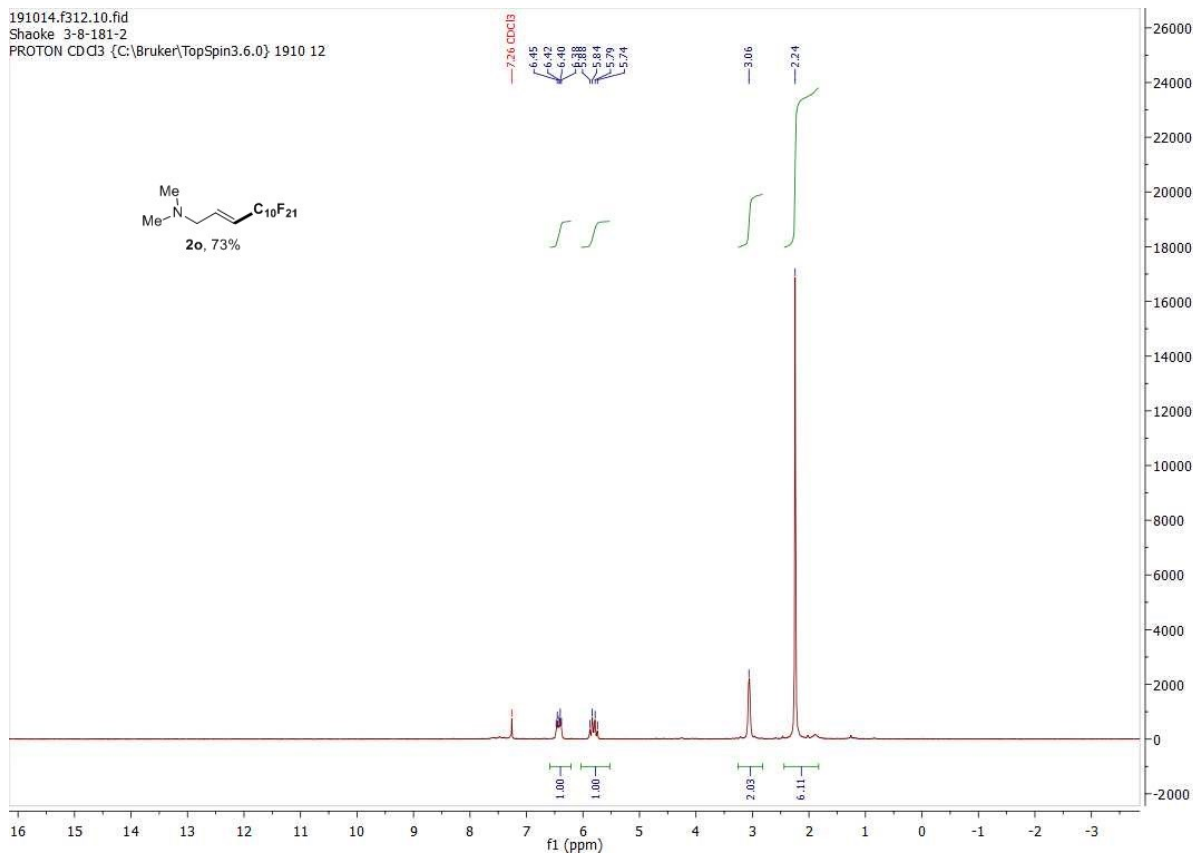


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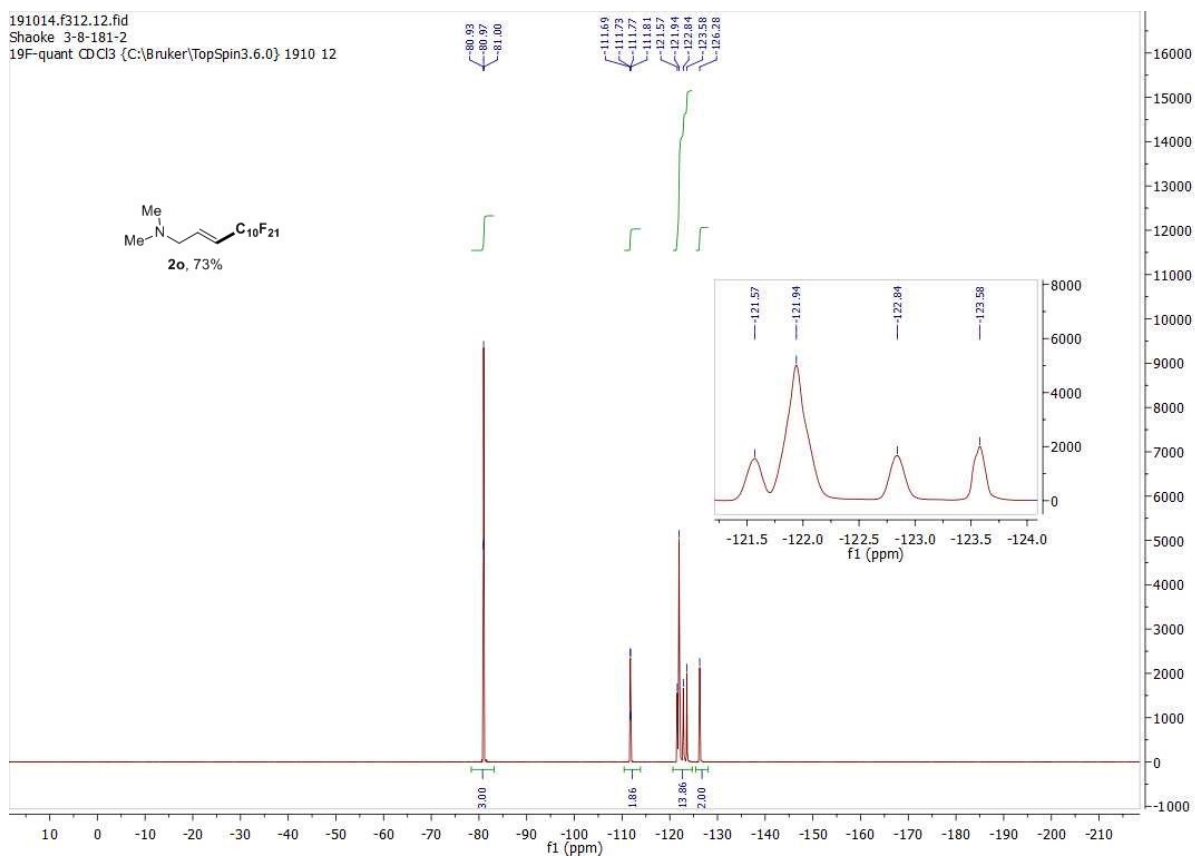
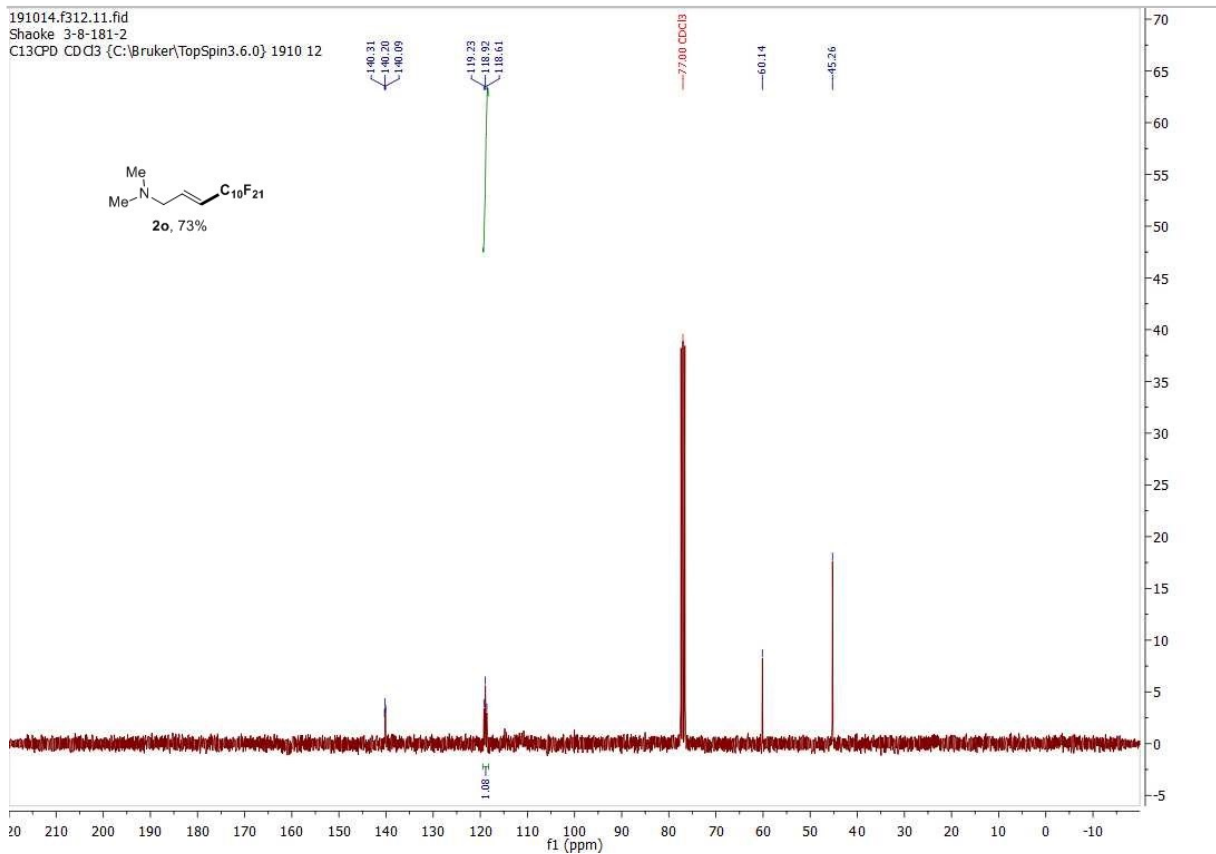
190408.f301.11.fid
 Shaoke 3-8-123-2-1
 19F-quant CDCl3 {C:\Bruker\TopSpin3.6.0} 1904 1



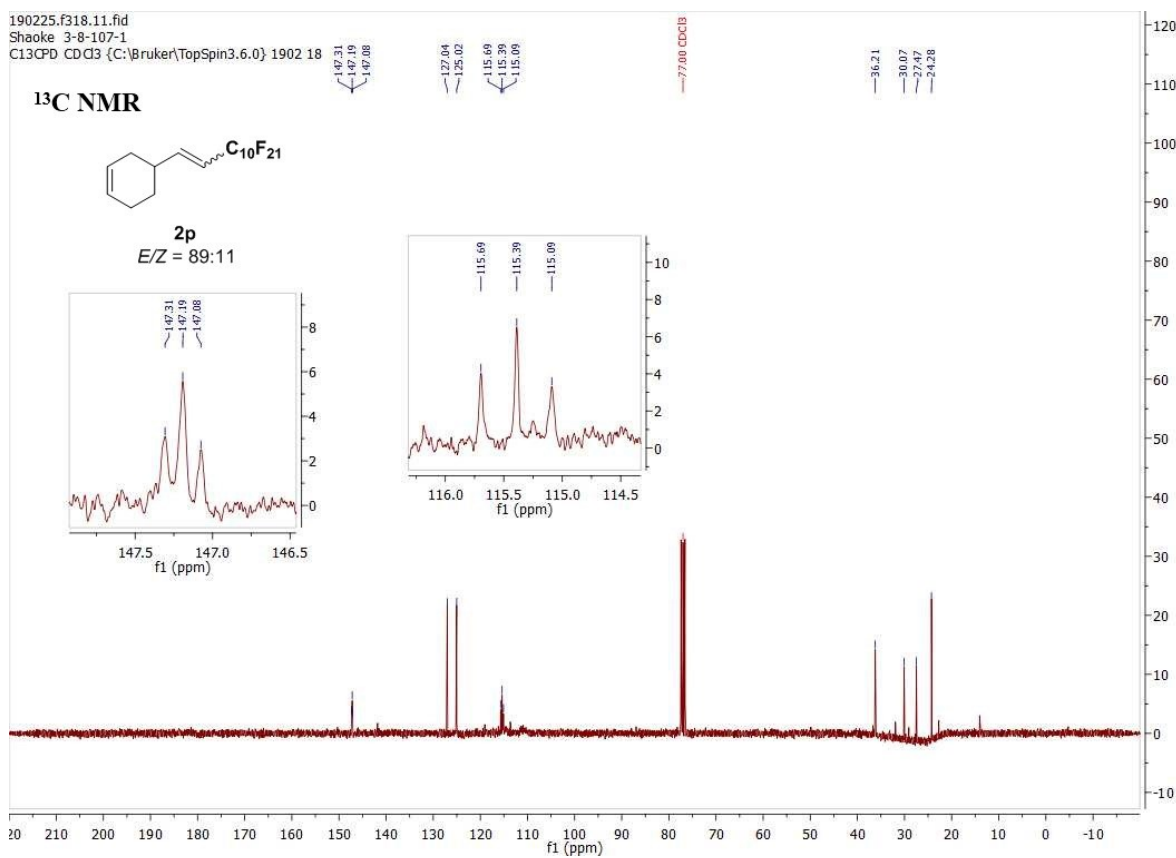
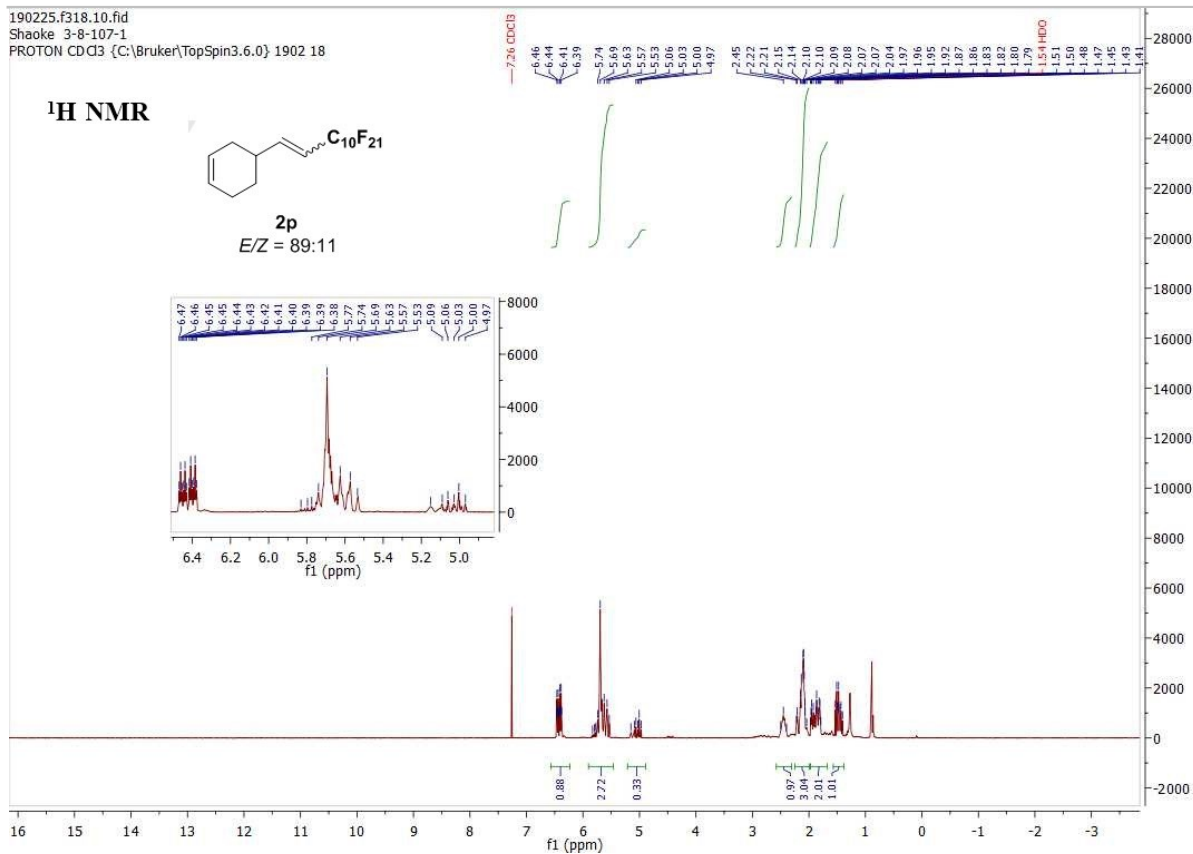
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 Shaoke 3-8-181-2
 PROTON CDCl3 {C:\Bruker\TopSpin3.6.0} 1910 12



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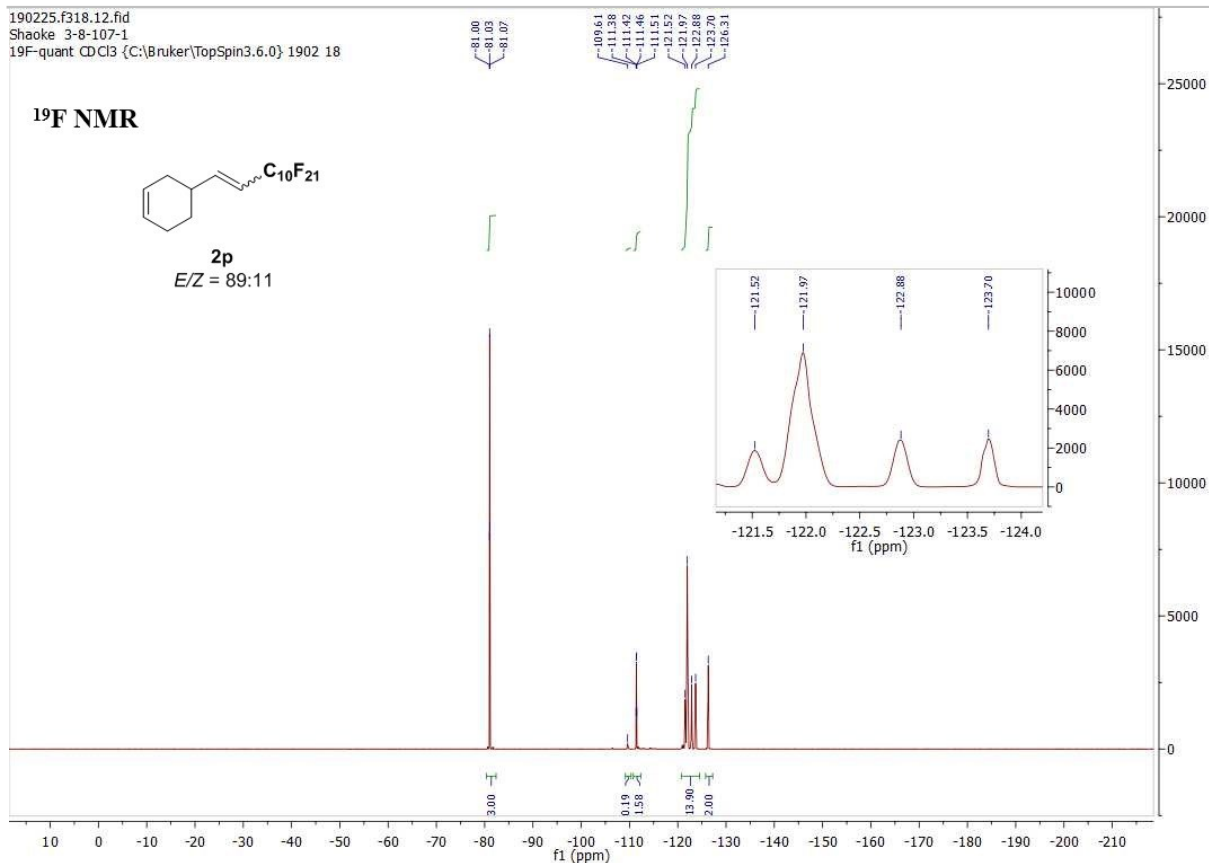
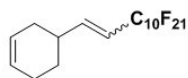
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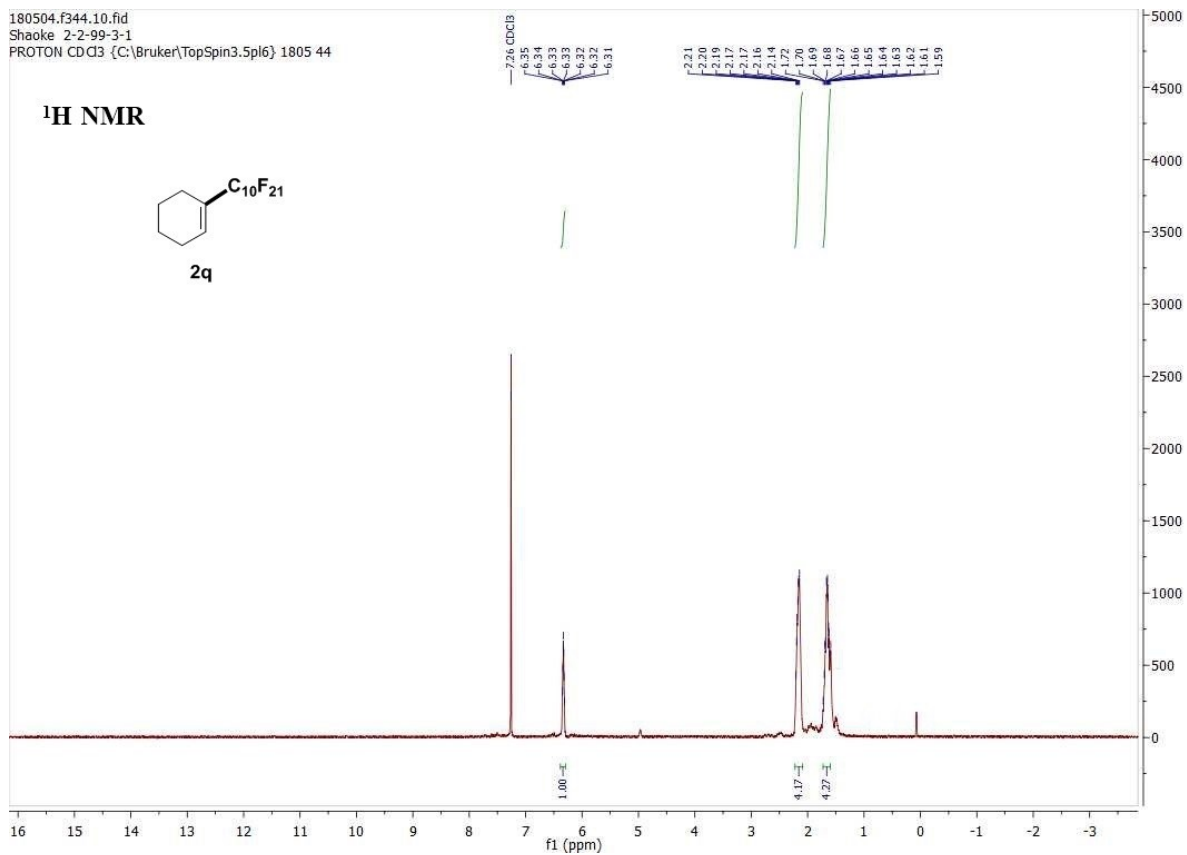
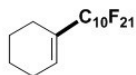
190225.f318.12.fid
 Shaoke 3-8-107-1
 19F-quant CDCl3 {C:\Bruker\TopSpin3.6.0} 1902 18

¹⁹F NMR

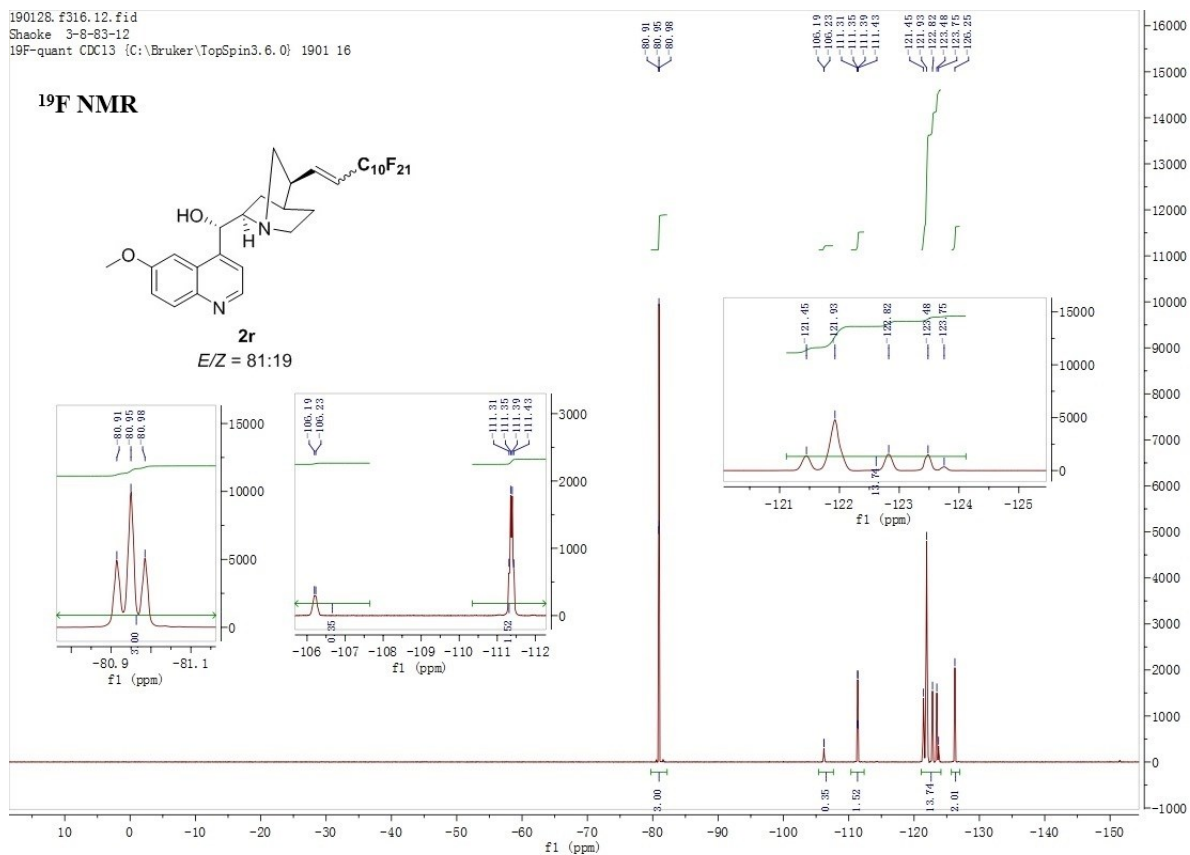
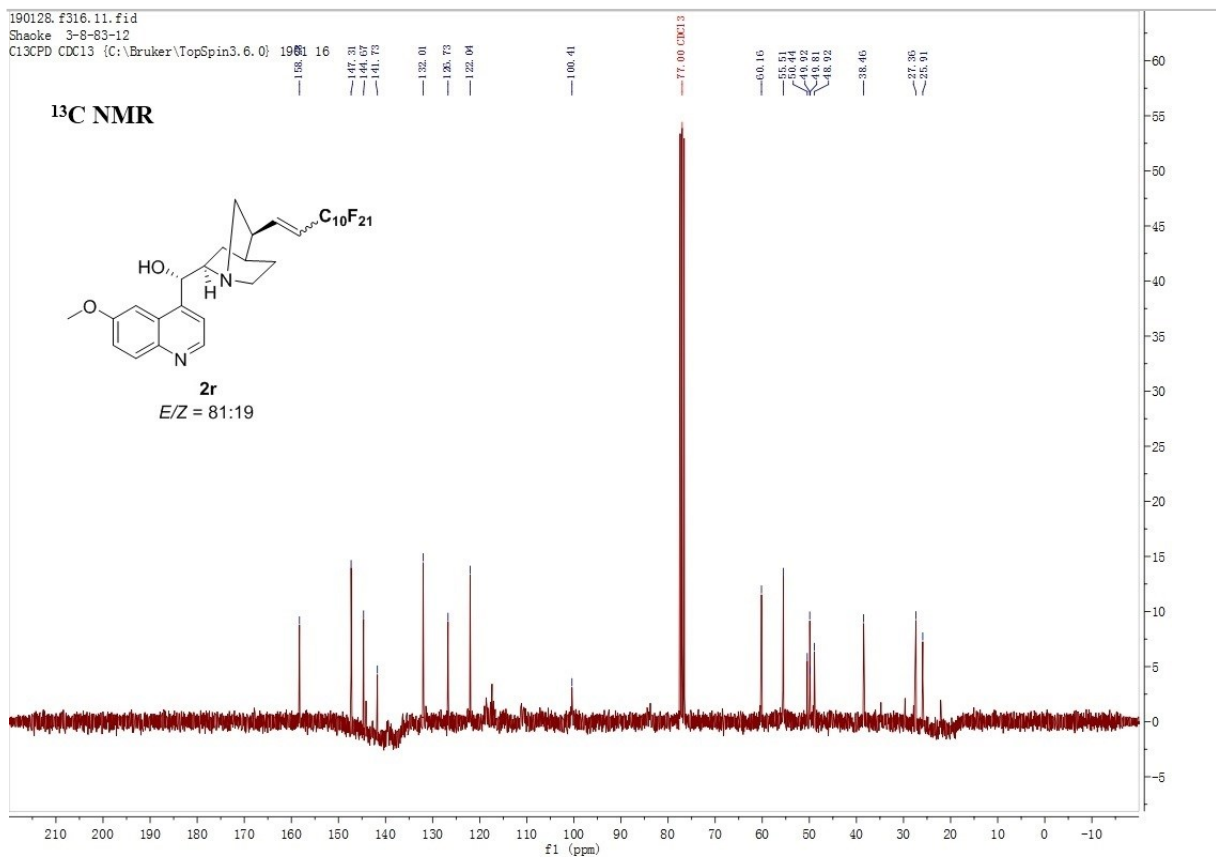


180504.f344.10.fid
 Shaoke 2-2-99-3-1
 PROTON CDCl3 {C:\Bruker\TopSpin3.5pl6} 1805 44

¹H NMR

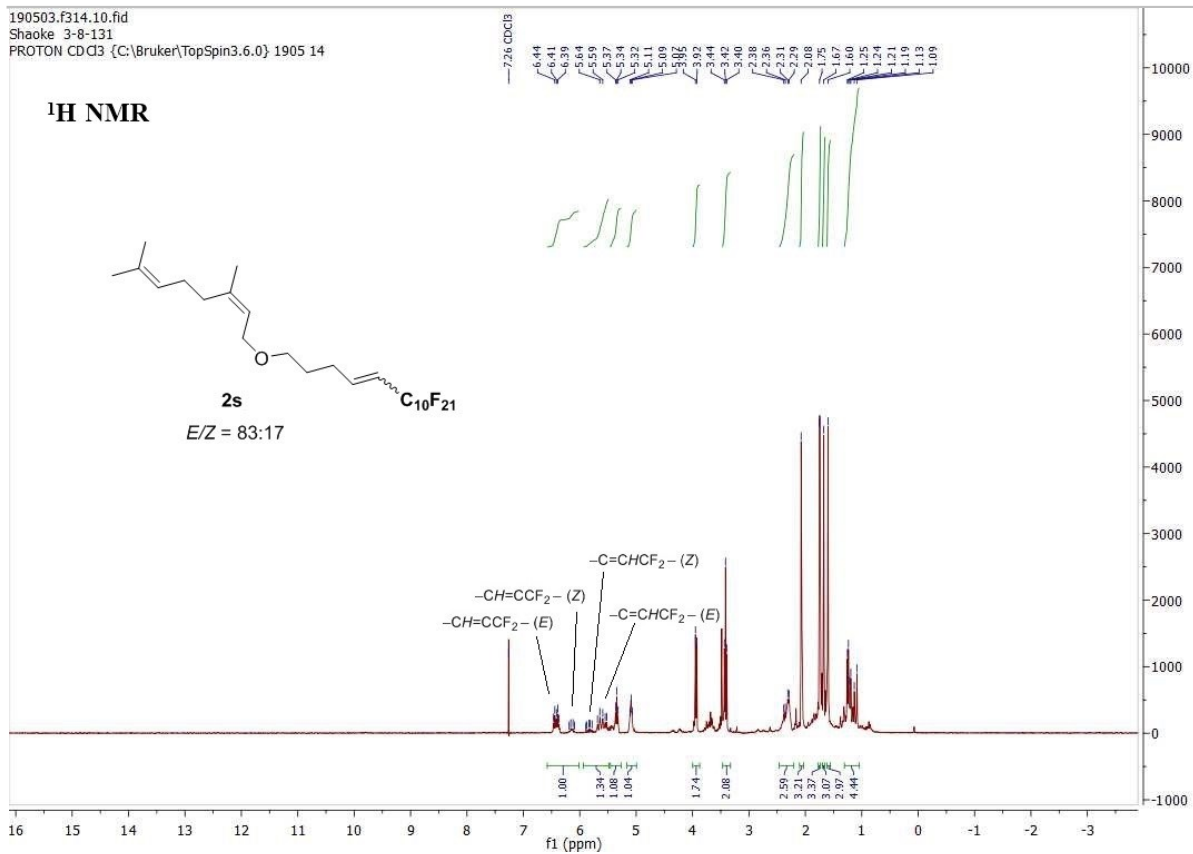


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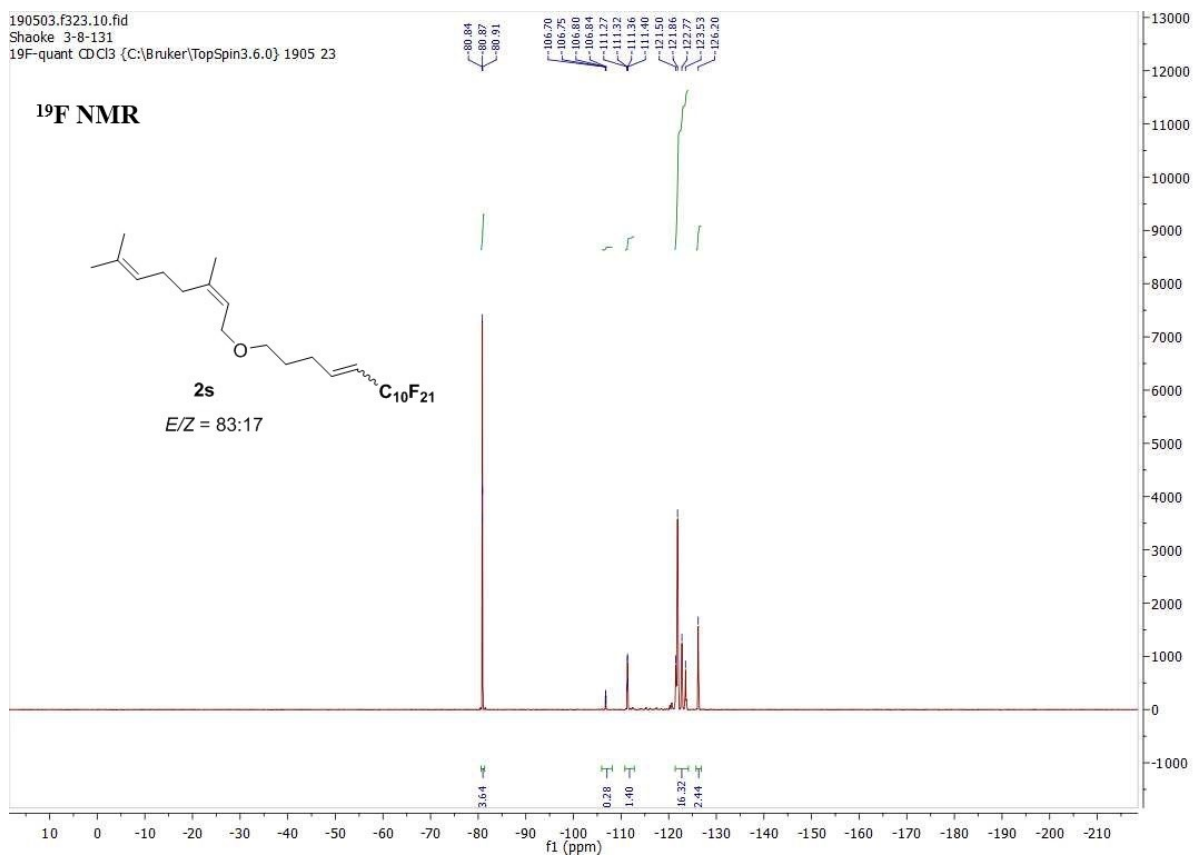


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190503.F314.10.fid
 Shaok 3-8-131
 PROTON CDCl₃ {C:\Bruker\TopSpin3.6.0} 1905 14



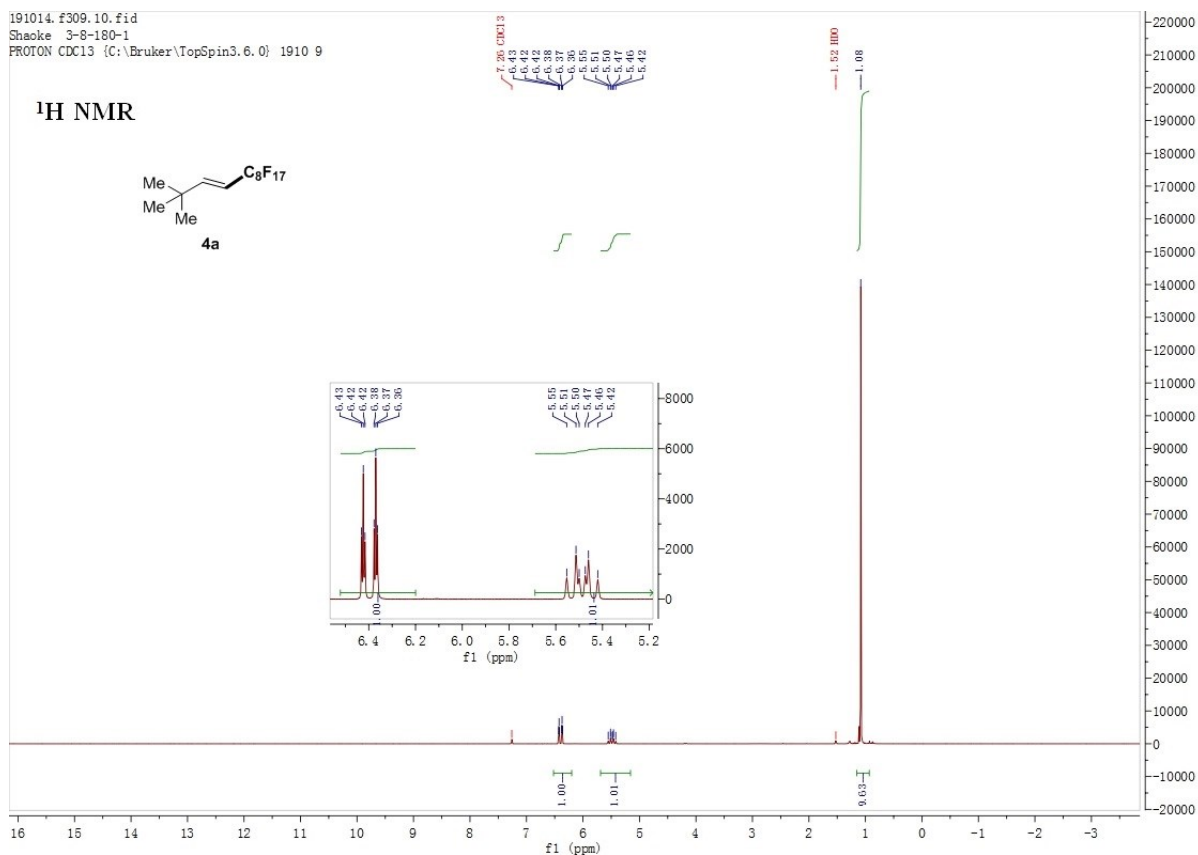
190503.F323.10.fid
 Shaok 3-8-131
¹⁹F-quant CDCl₃ {C:\Bruker\TopSpin3.6.0} 1905 23



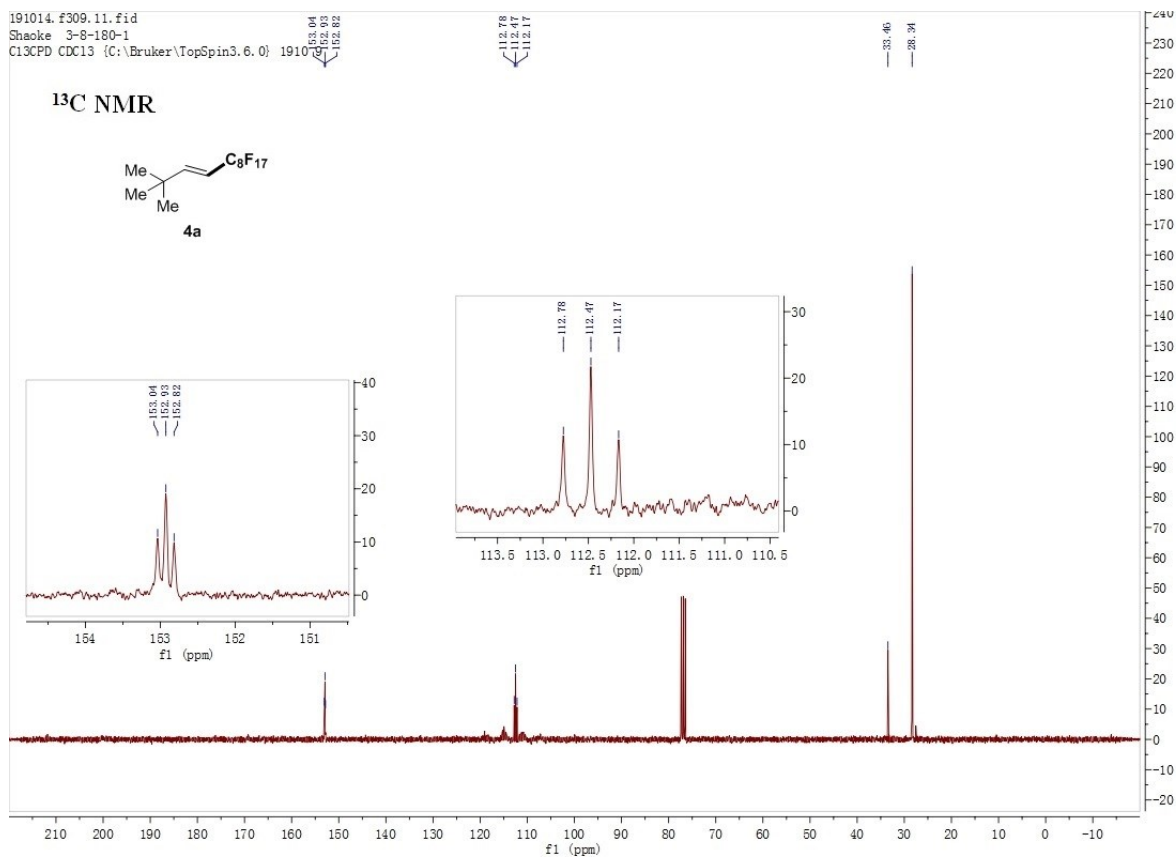
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8.3 Spectra of other perfluoroalkylated alkenes

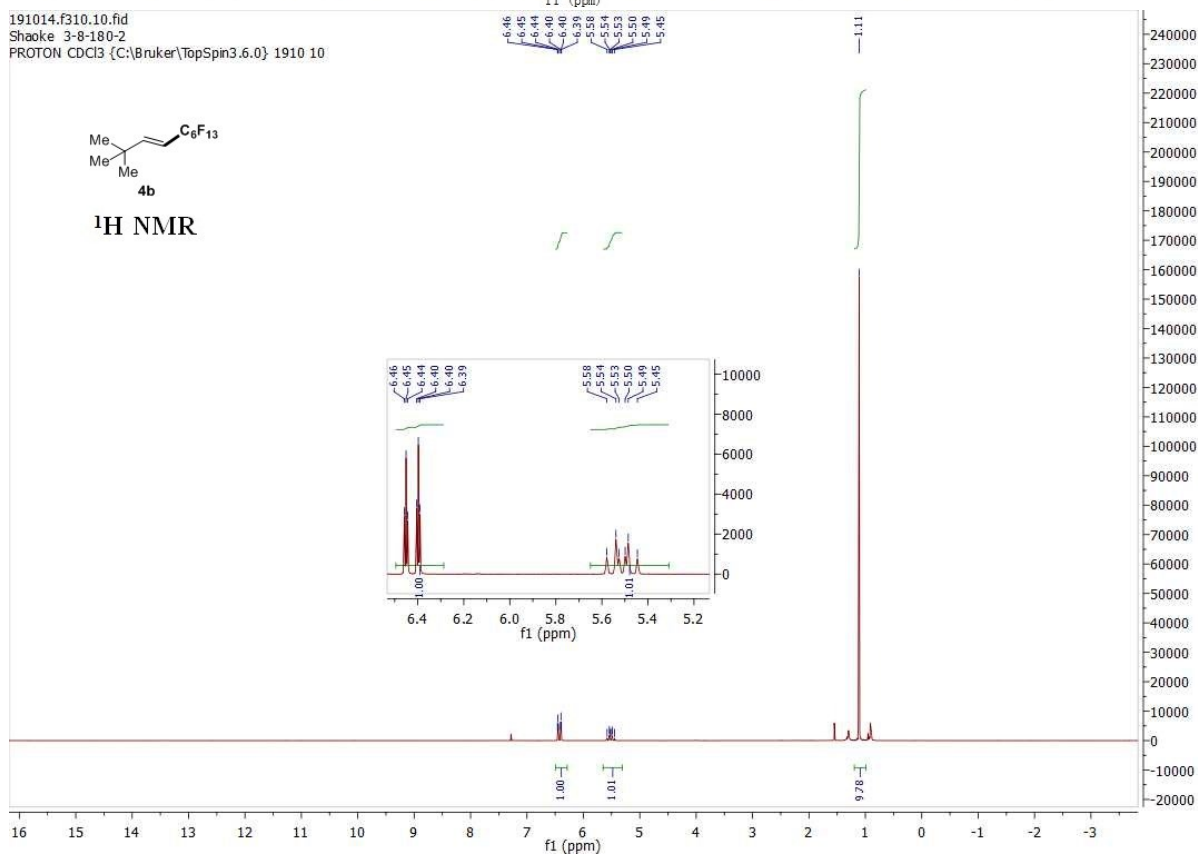
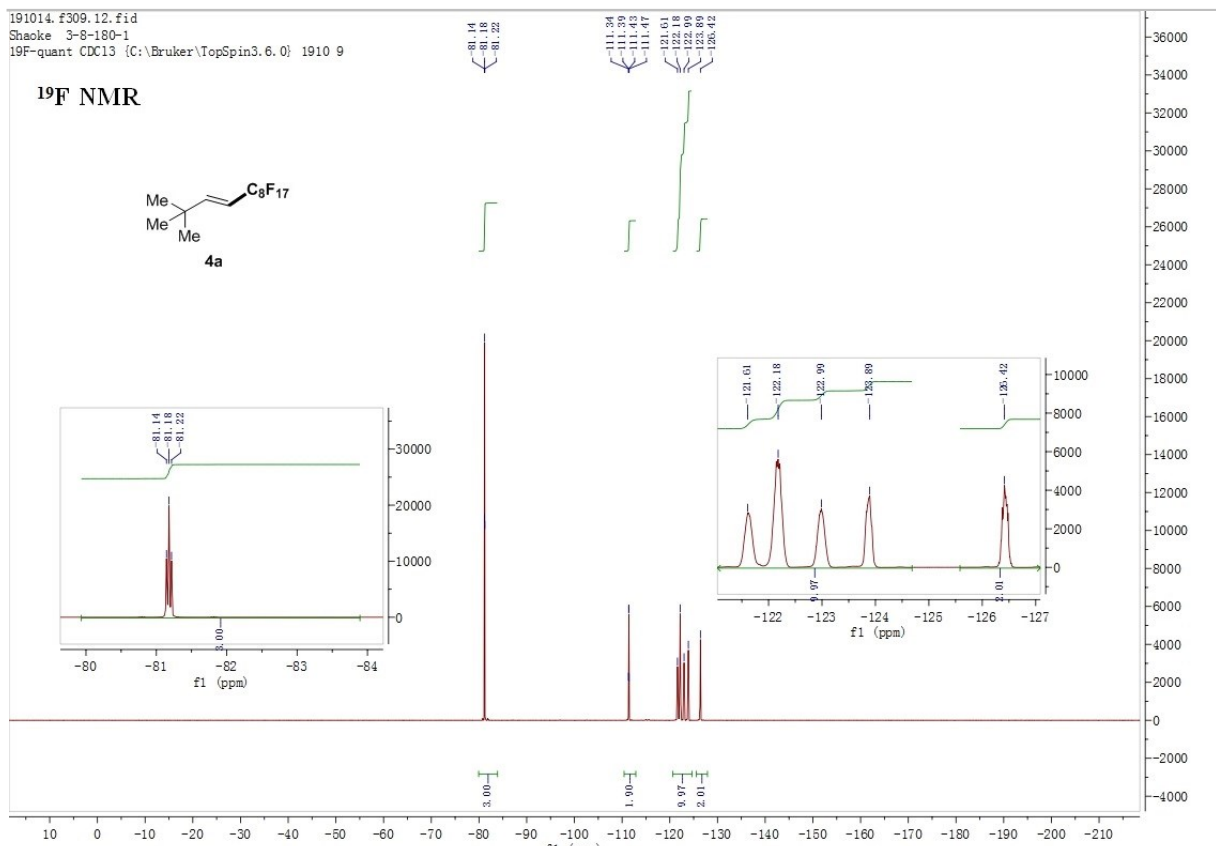
191014.f309.10.fid
Shaoke 3-8-180-1
PROTON CDC13 {C:\Bruker\TopSpin3.6.0} 1910 9



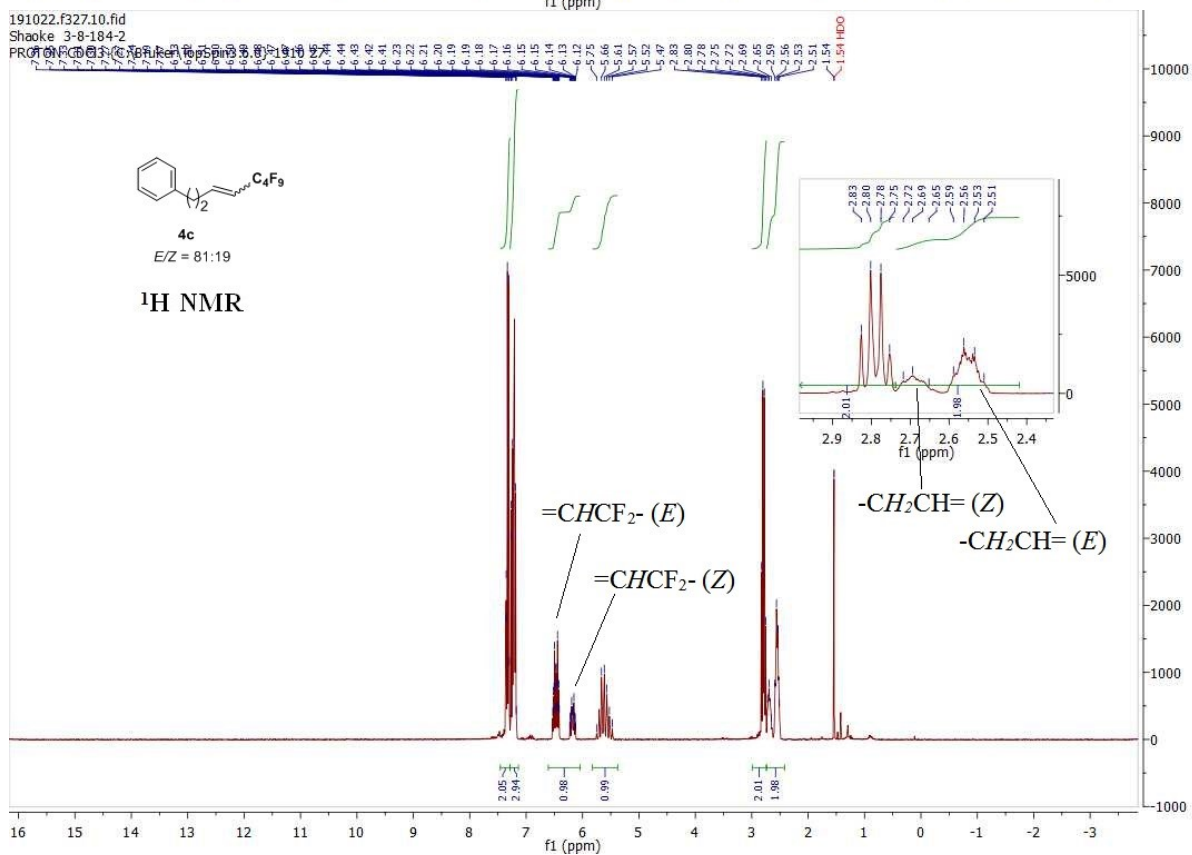
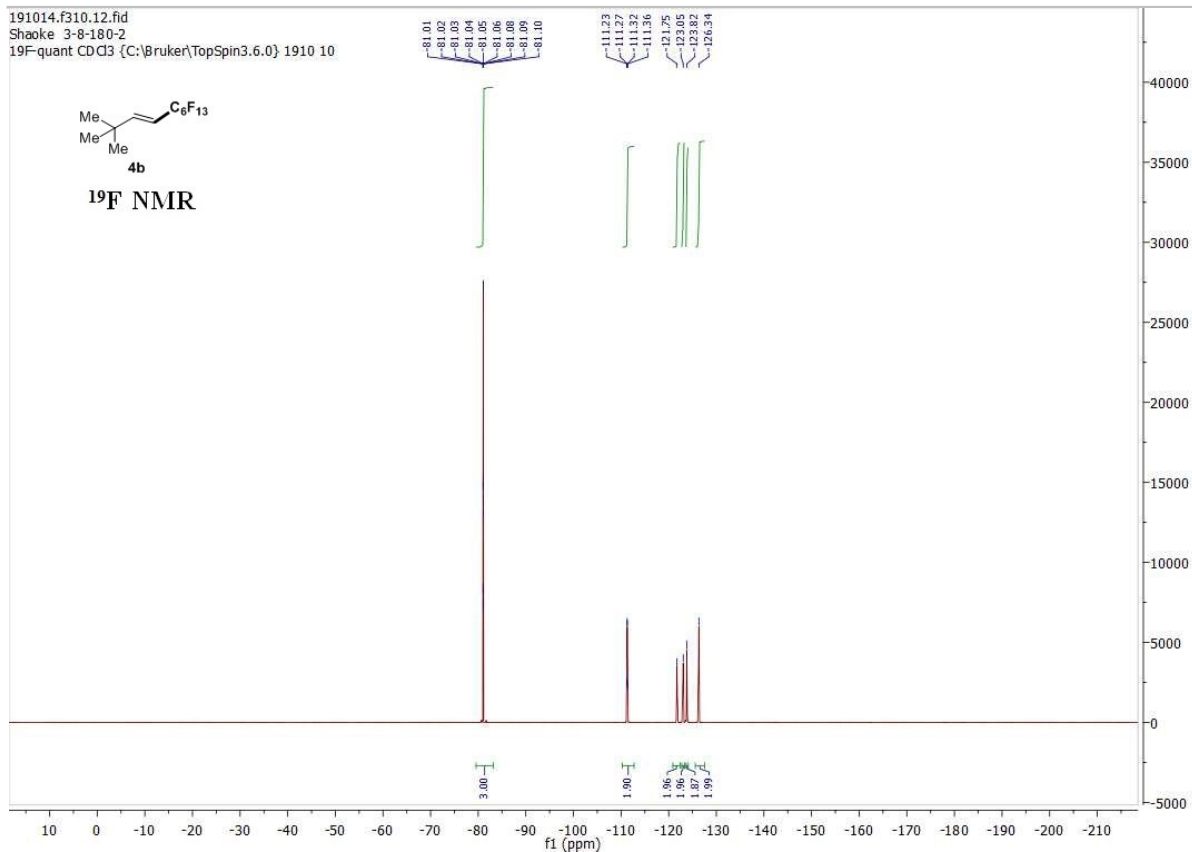
191014.f309.11.fid
Shaoke 3-8-180-1
C13CPD CDC13 {C:\Bruker\TopSpin3.6.0} 1910 9



SUPPORTING INFORMATION

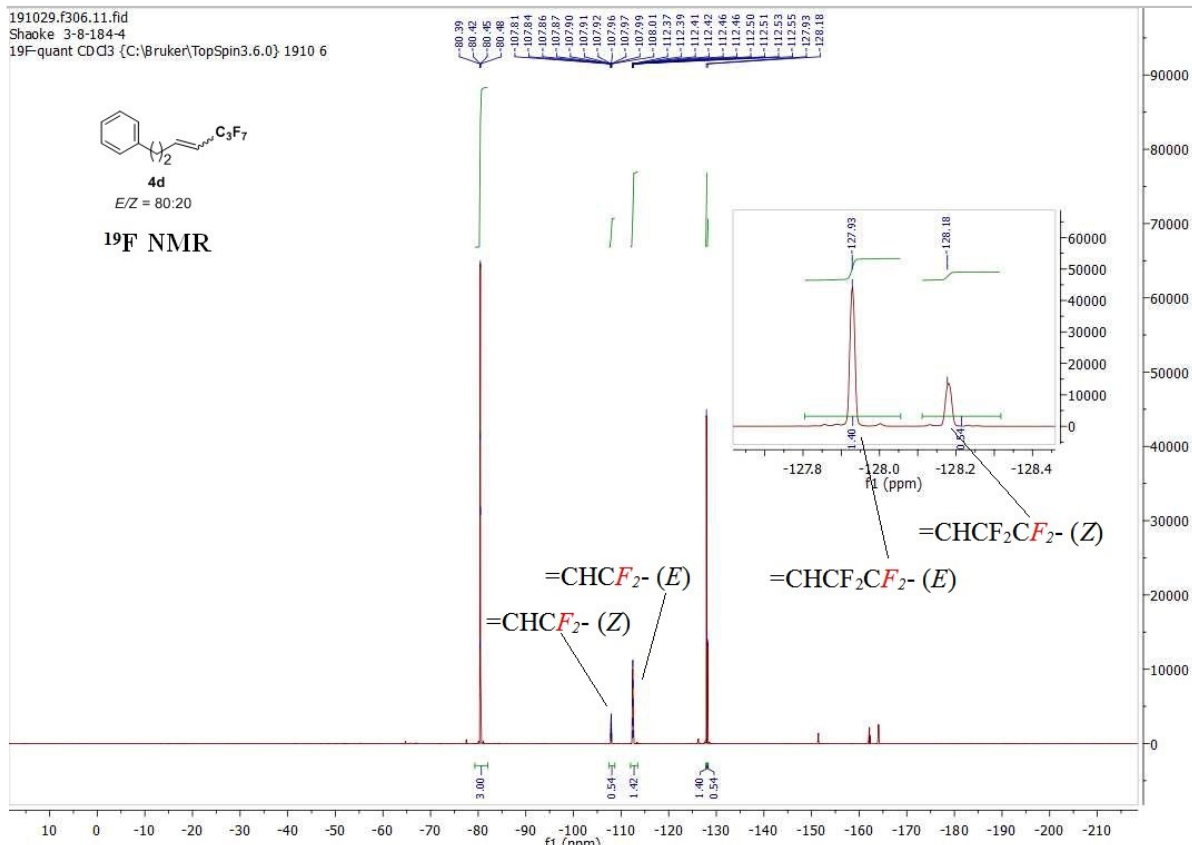


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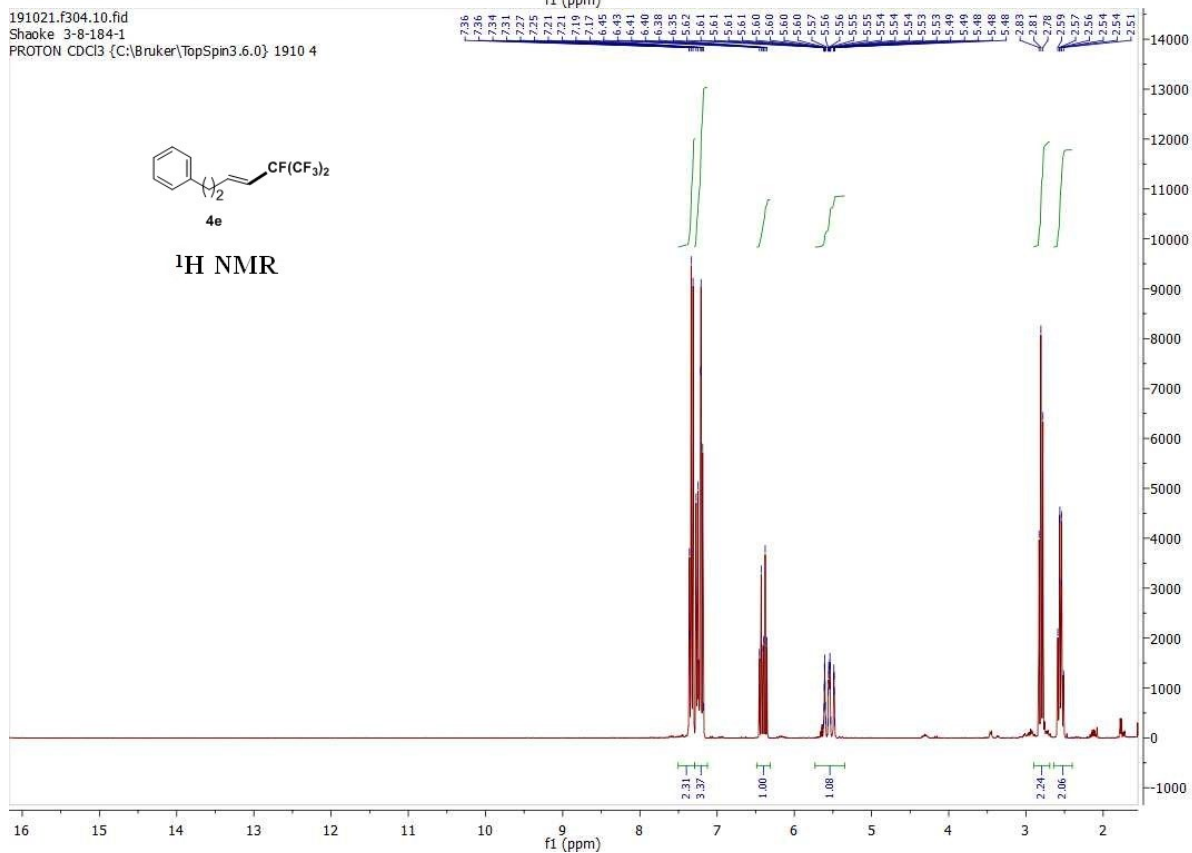


SUPPORTING INFORMATION

191029.f306.11.fid
 Shaoke 3-8-184-4
 19F-quant CDCl3 {C:\Bruker\TopSpin3.6.0} 1910 6

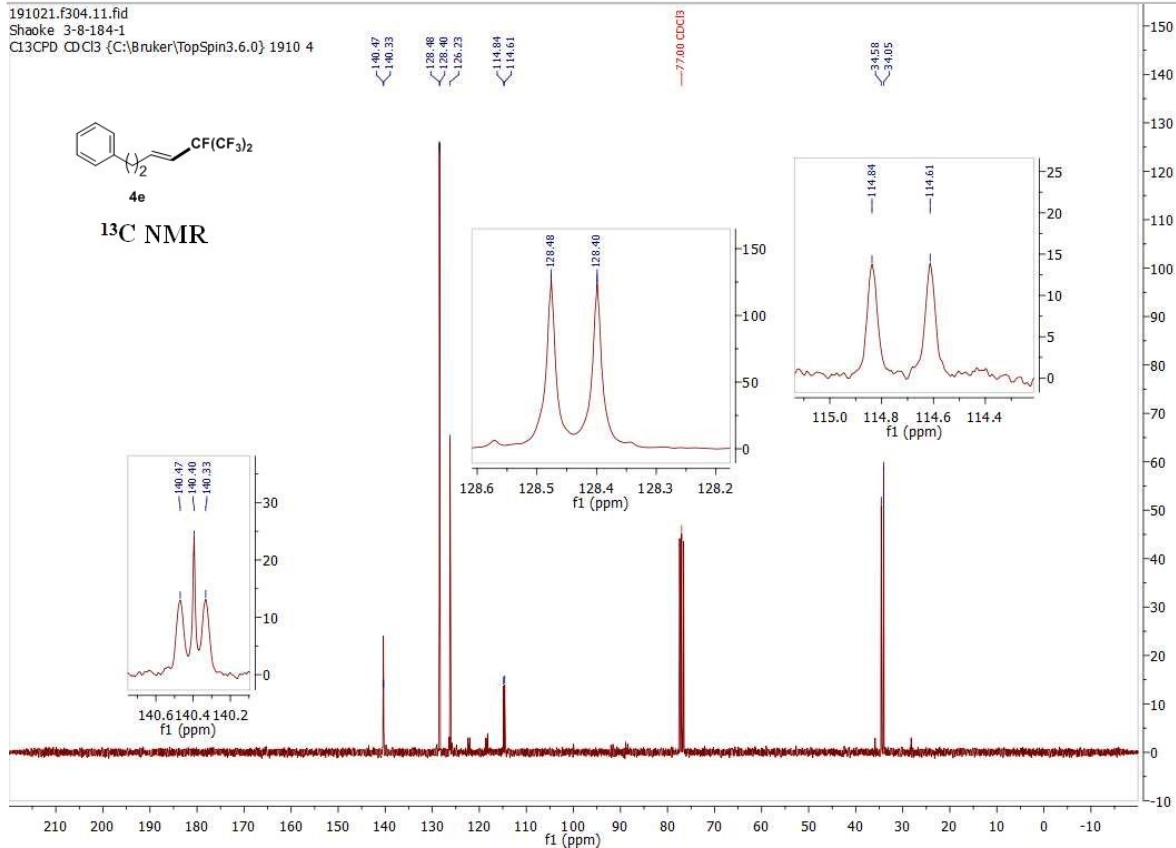


191021.f304.10.fid
 Shaoke 3-8-184-1
 PROTON CDCl3 {C:\Bruker\TopSpin3.6.0} 1910 4

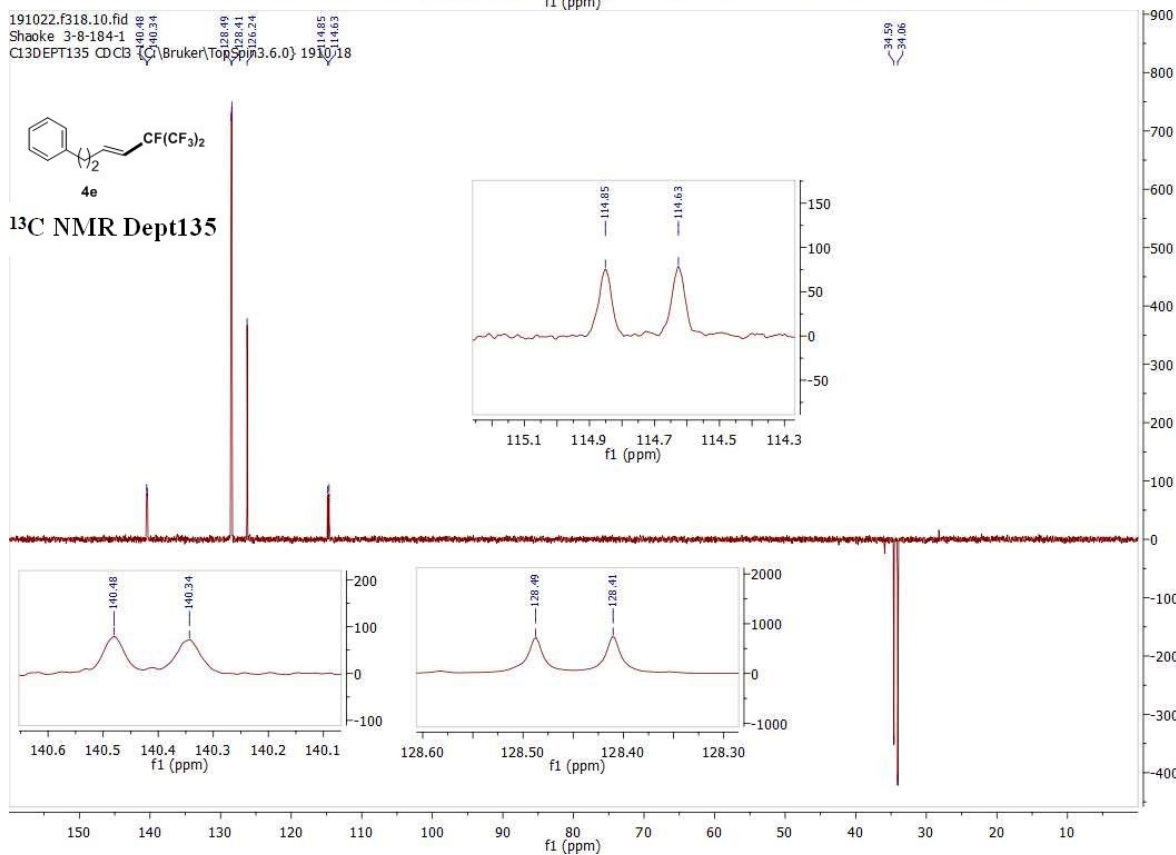


SUPPORTING INFORMATION

191021.f304.11.fid
 Shaoke 3-8-184-1
 C13CPD CDCl3 (C:\Bruker\TopSpin3.6.0) 1910 4

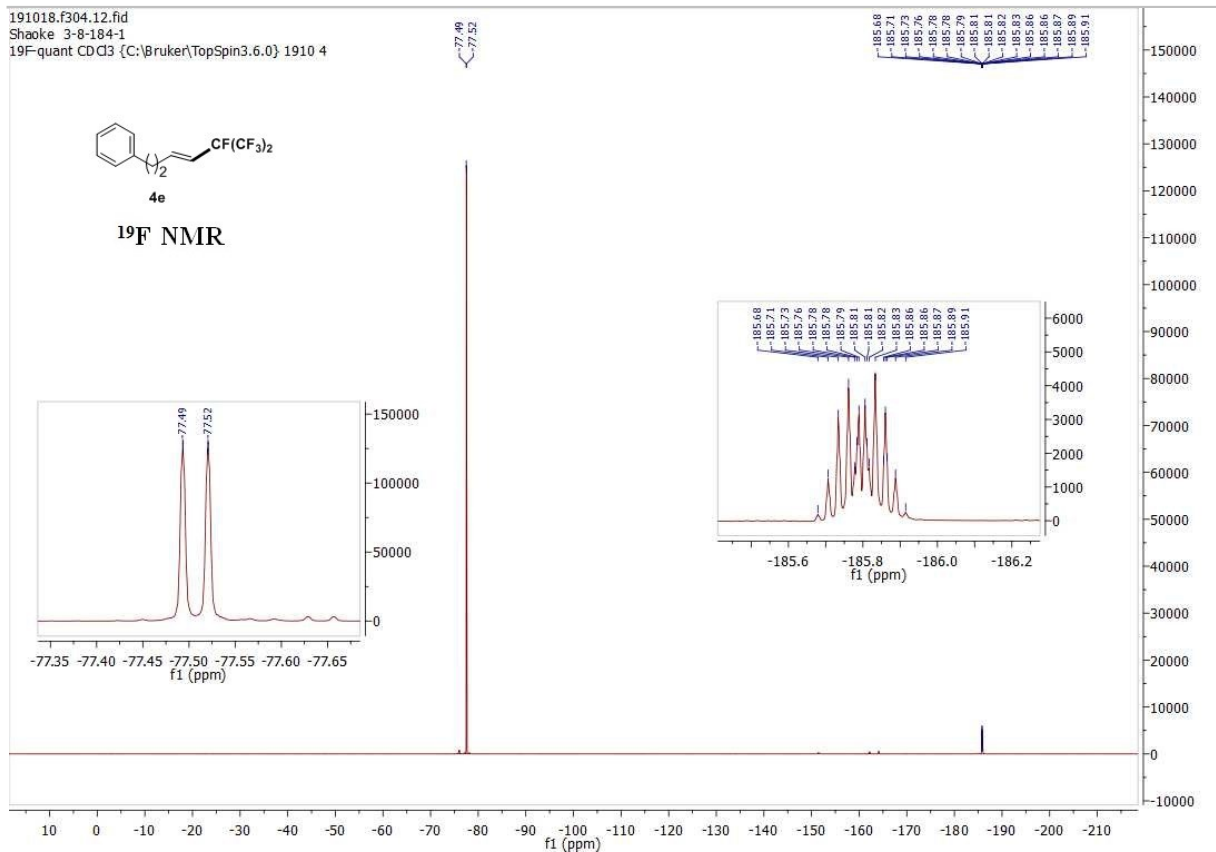


191022.f318.10.fid
 Shaoke 3-8-184-1
 C13DEPT135 CDCl3 (C:\Bruker\TopSpin3.6.0) 1910 18

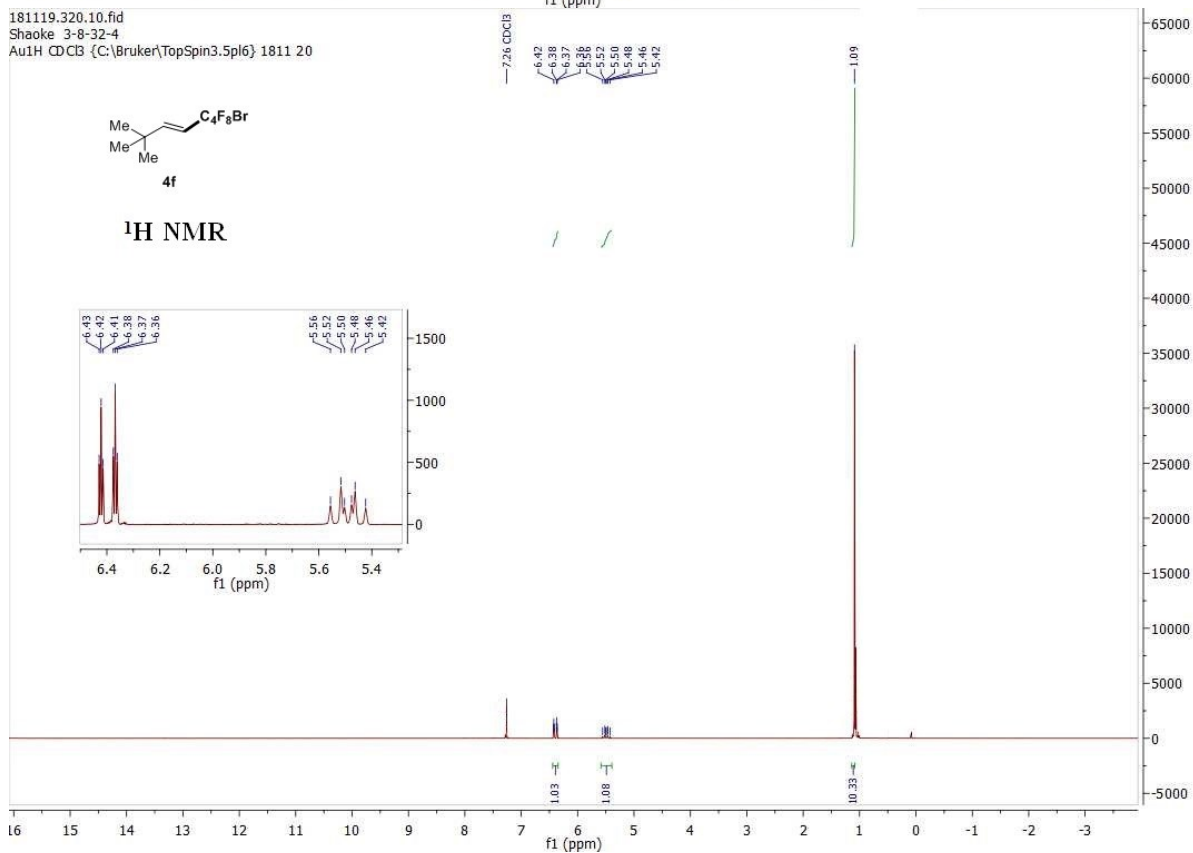


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191018.f304.12.fid
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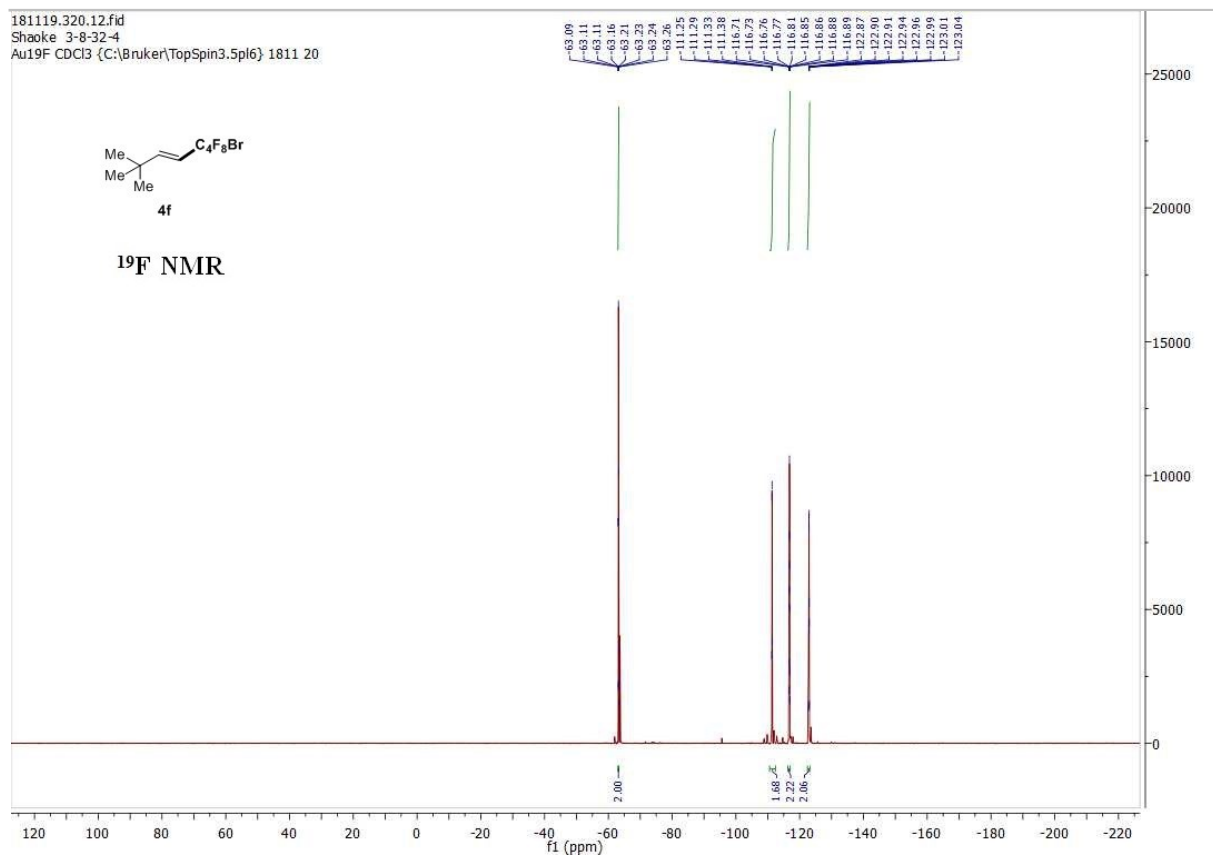


181119.320.10.fid
 Shaoke 3-8-32-4
 Au1H CDCl3 {C:\Bruker\TopSpin3.5pl6} 1811 20

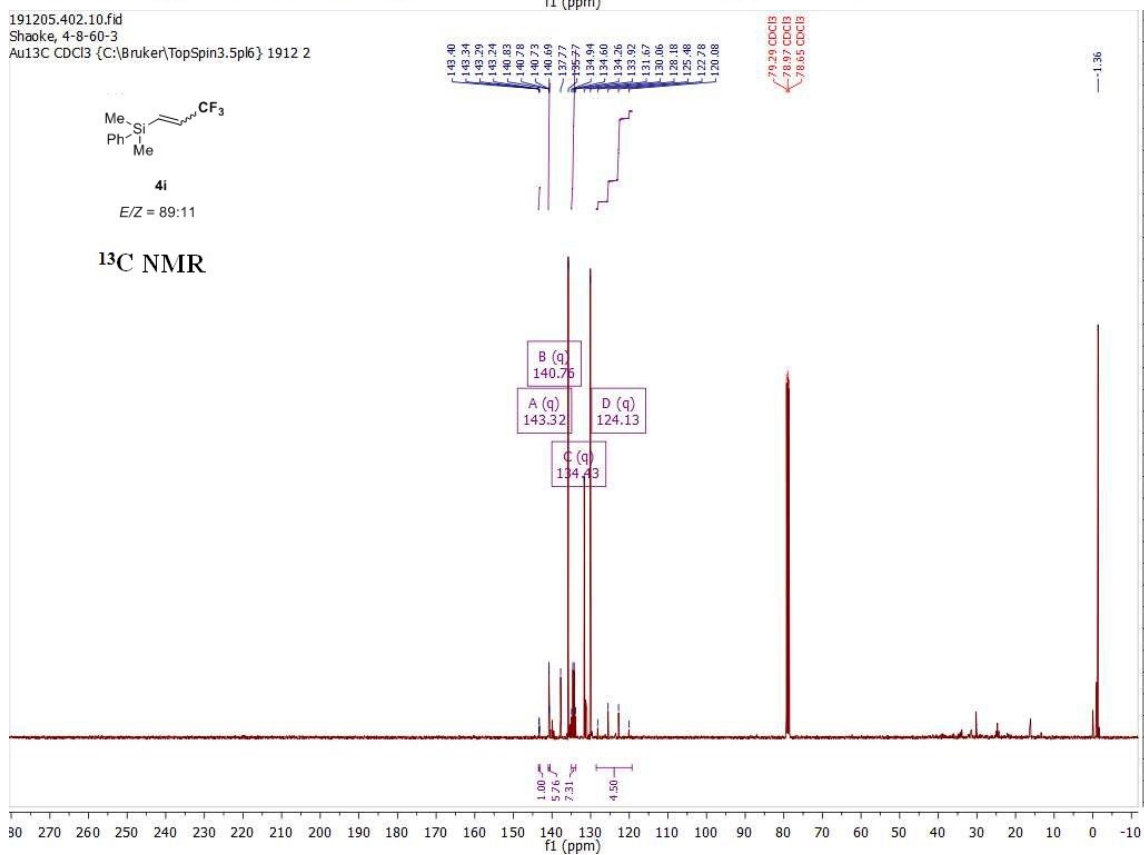
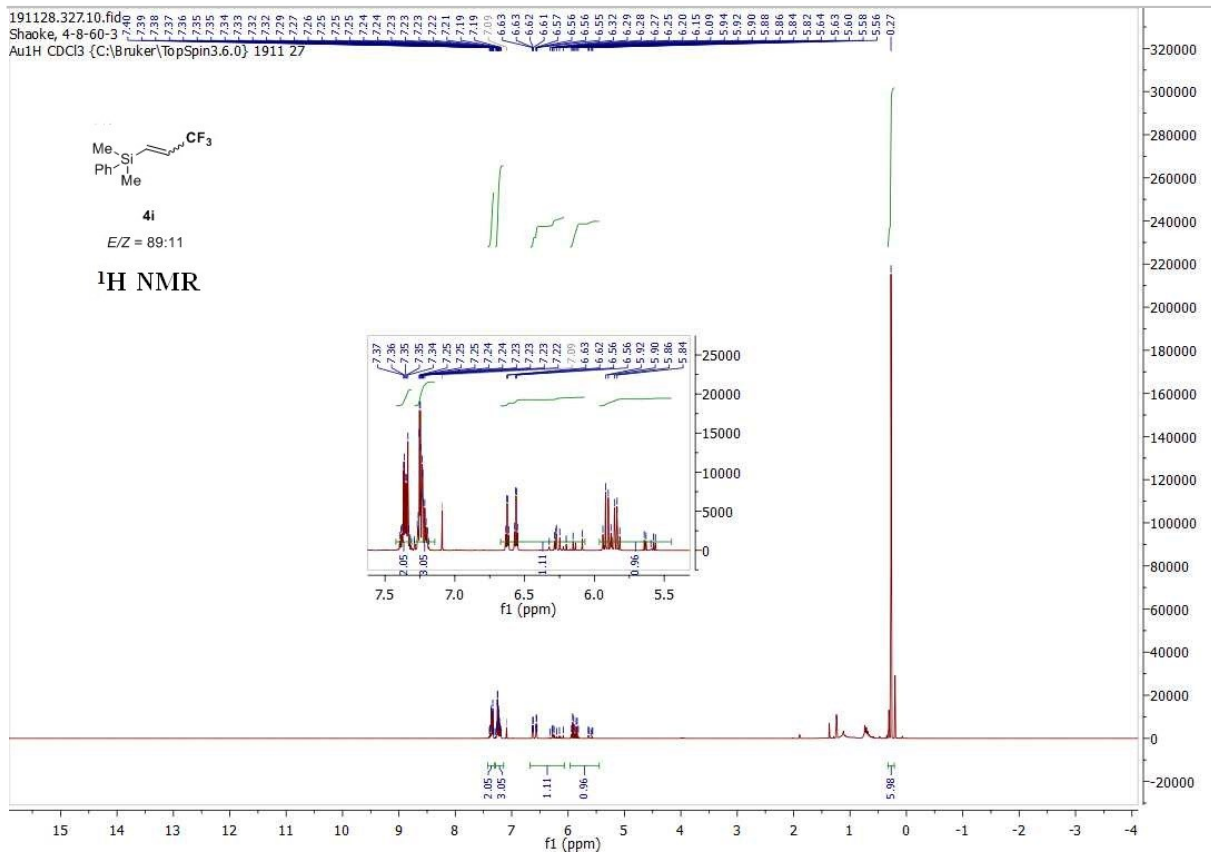


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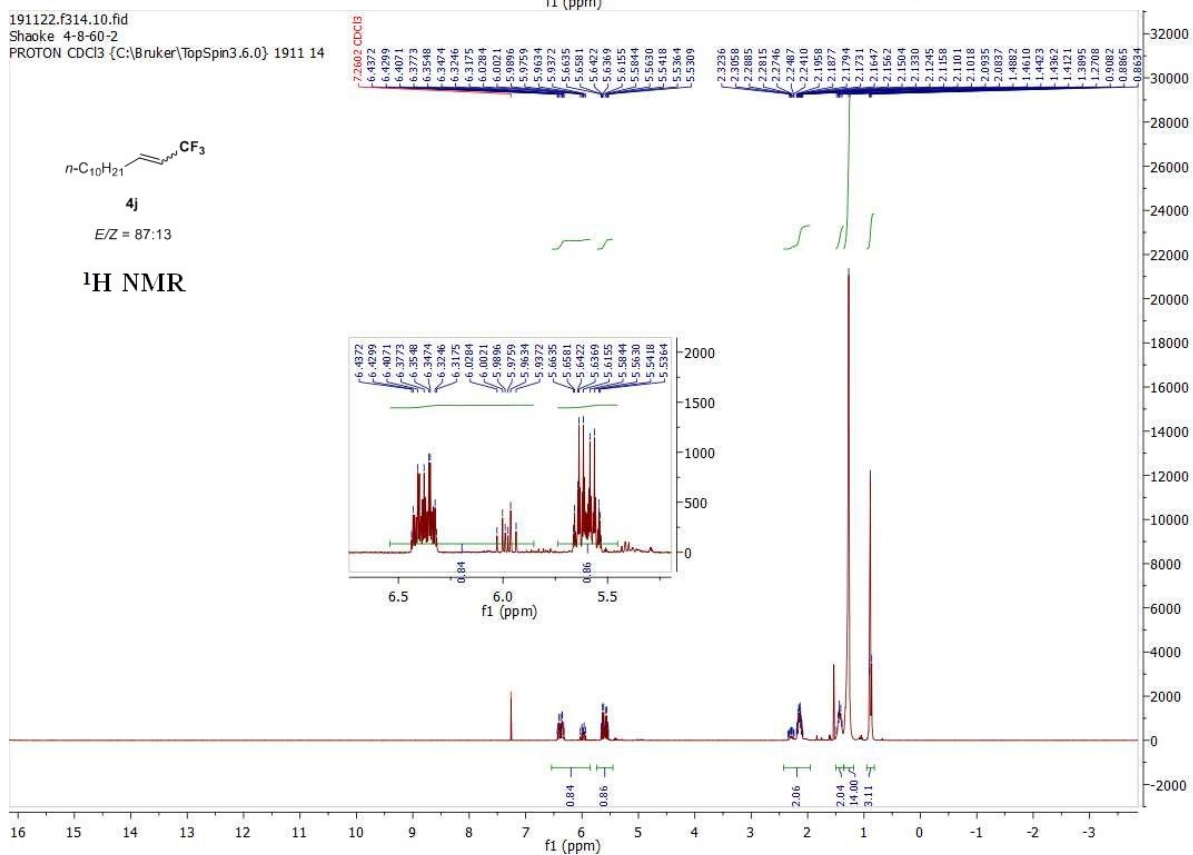
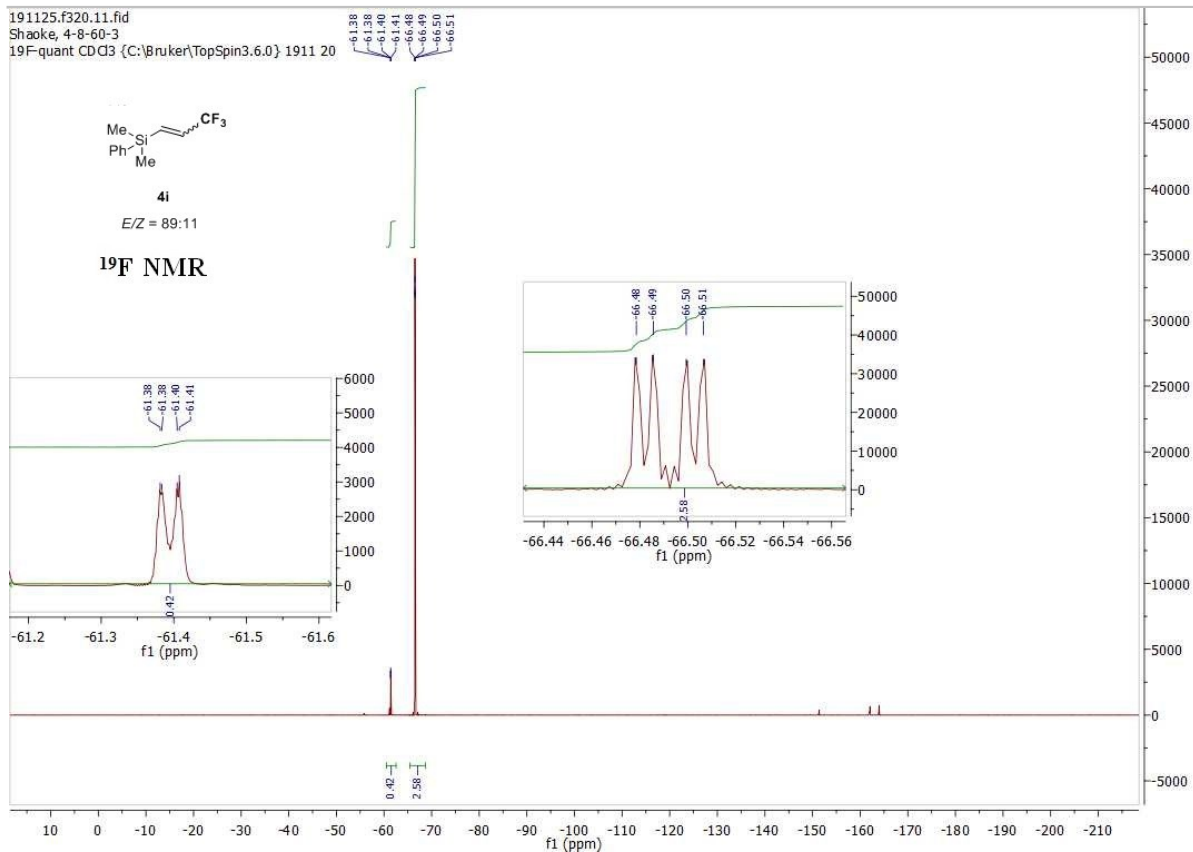
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Shaoke 3-8-32-4
Au19F CDCl3 {C:\Bruker\TopSpin3.5pl6} 1811 20



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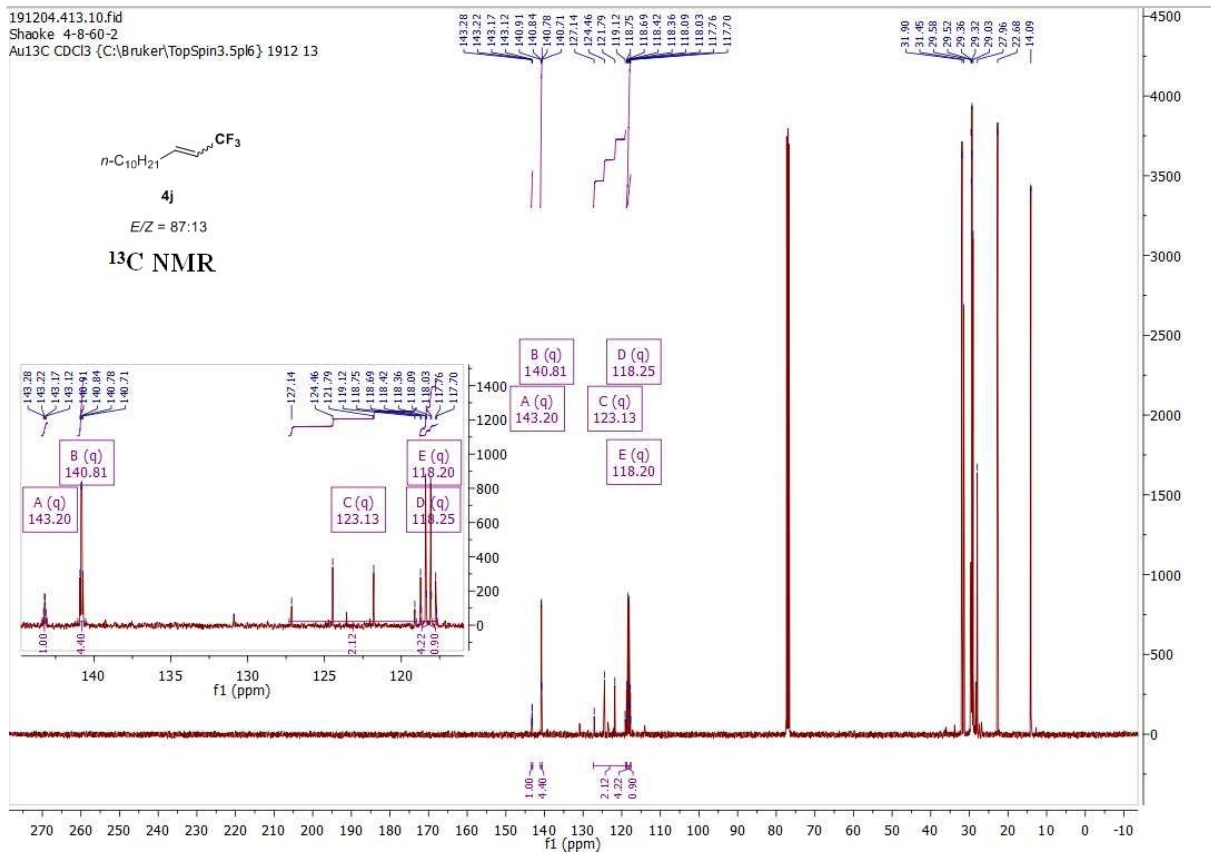


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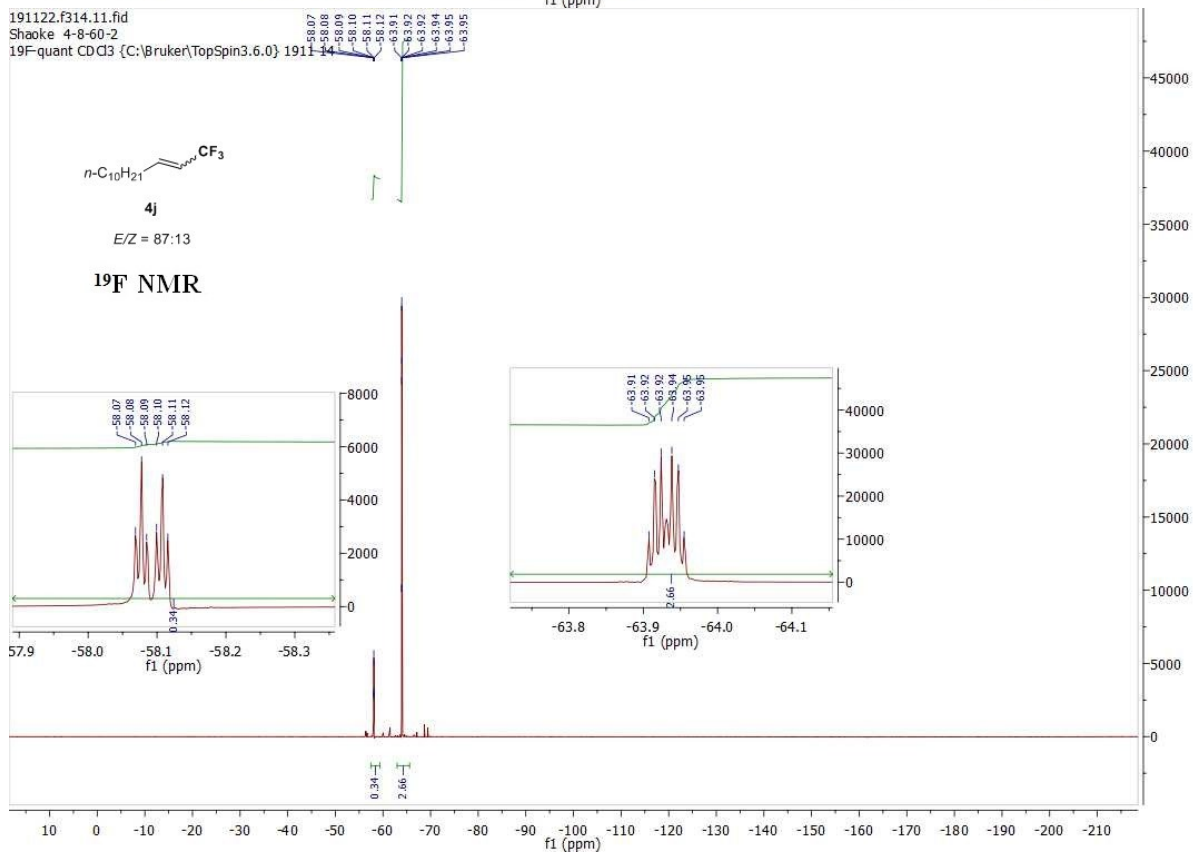


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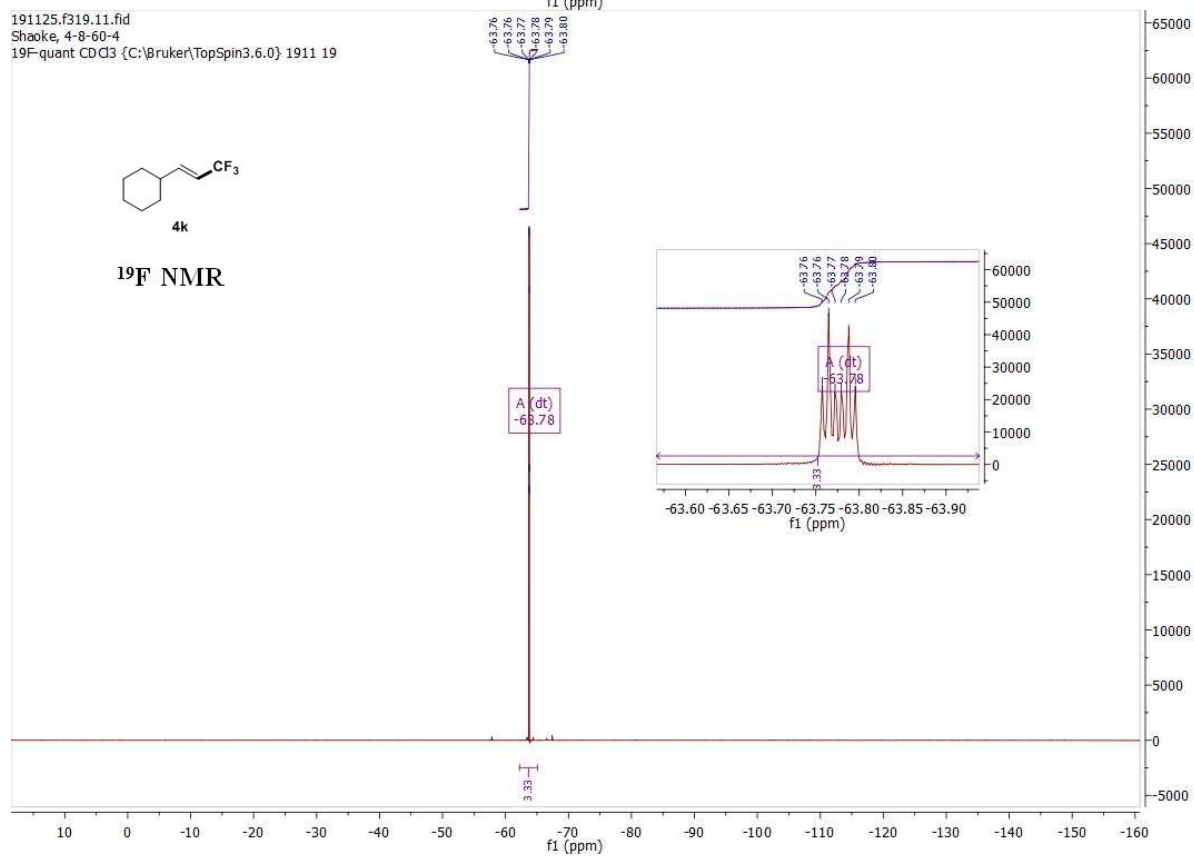
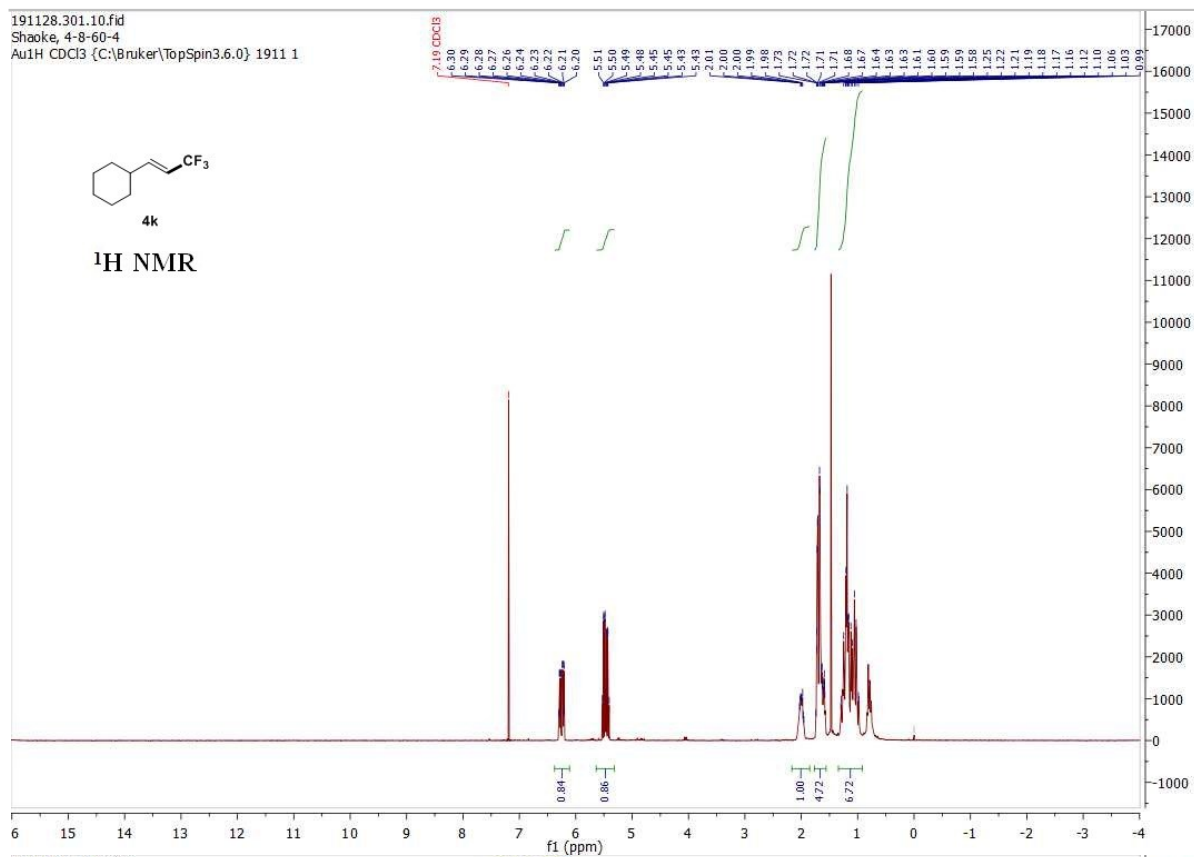
191204.413.10.fid
 Shaoke 4-8-60-2
 Au13C CDCl3 {C:\Bruker\TopSpin3.5pl6} 1912 13



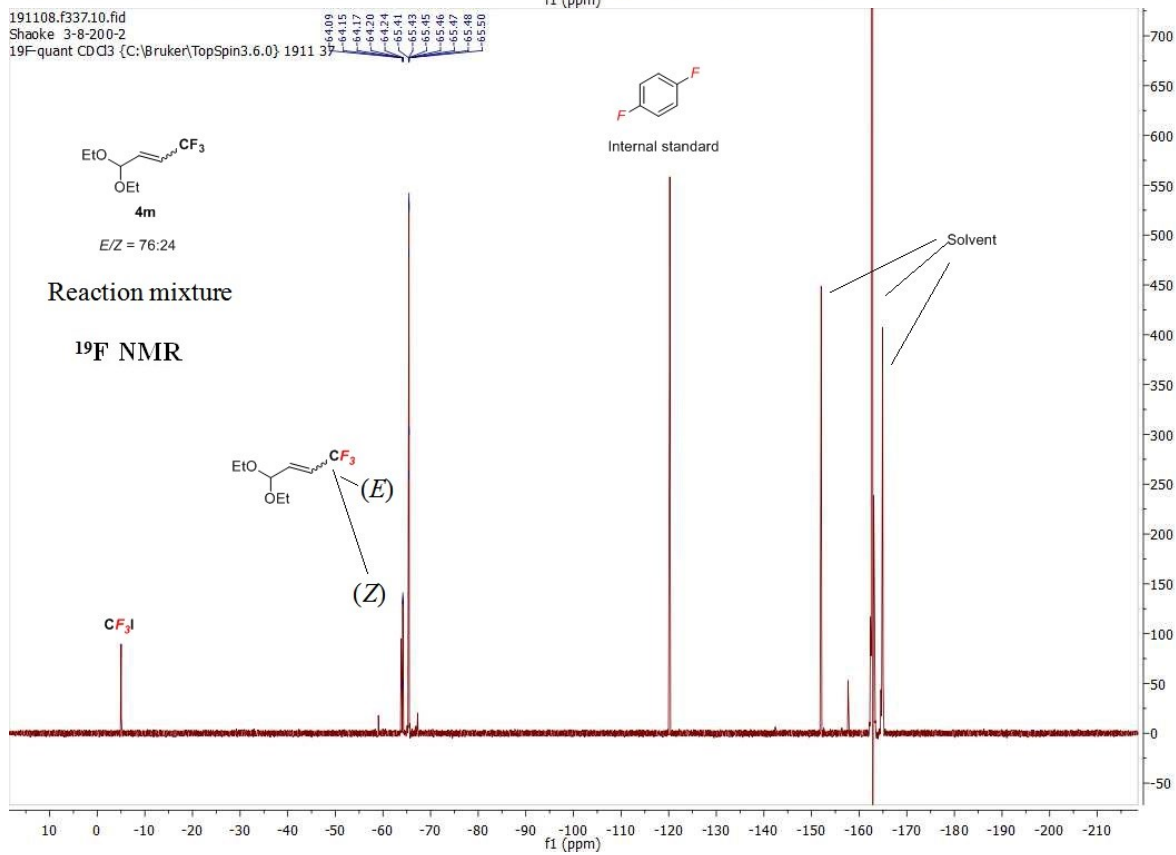
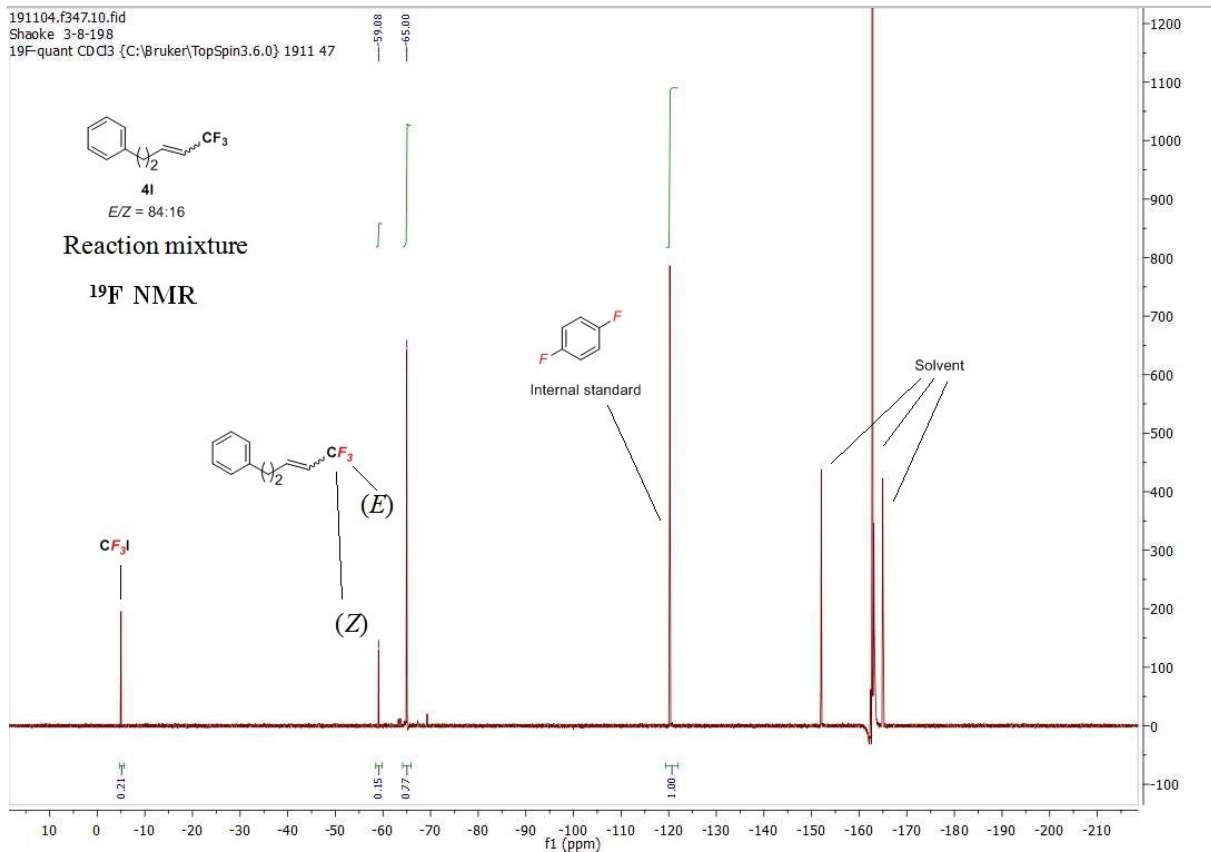
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 Shaoke 4-8-60-2
 19F-quant CDCl3 {C:\Bruker\TopSpin3.6.0} 1911



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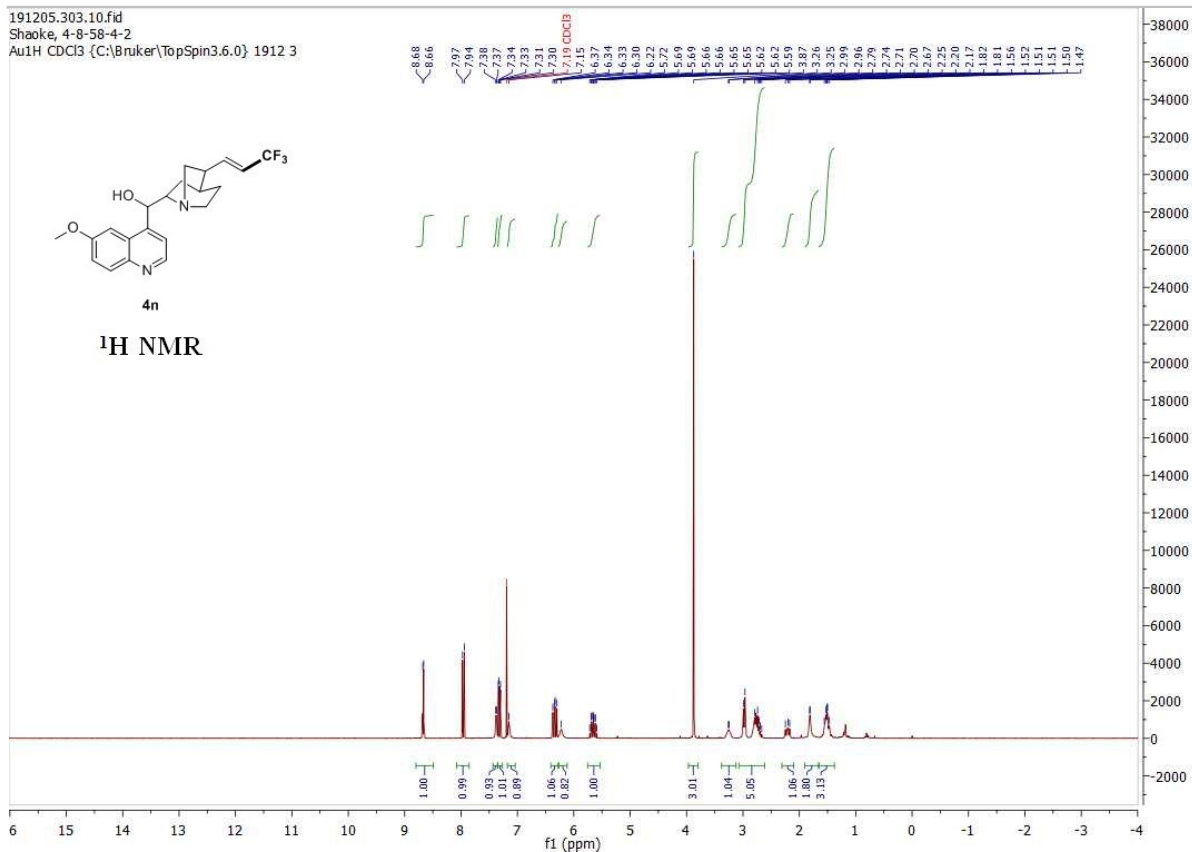


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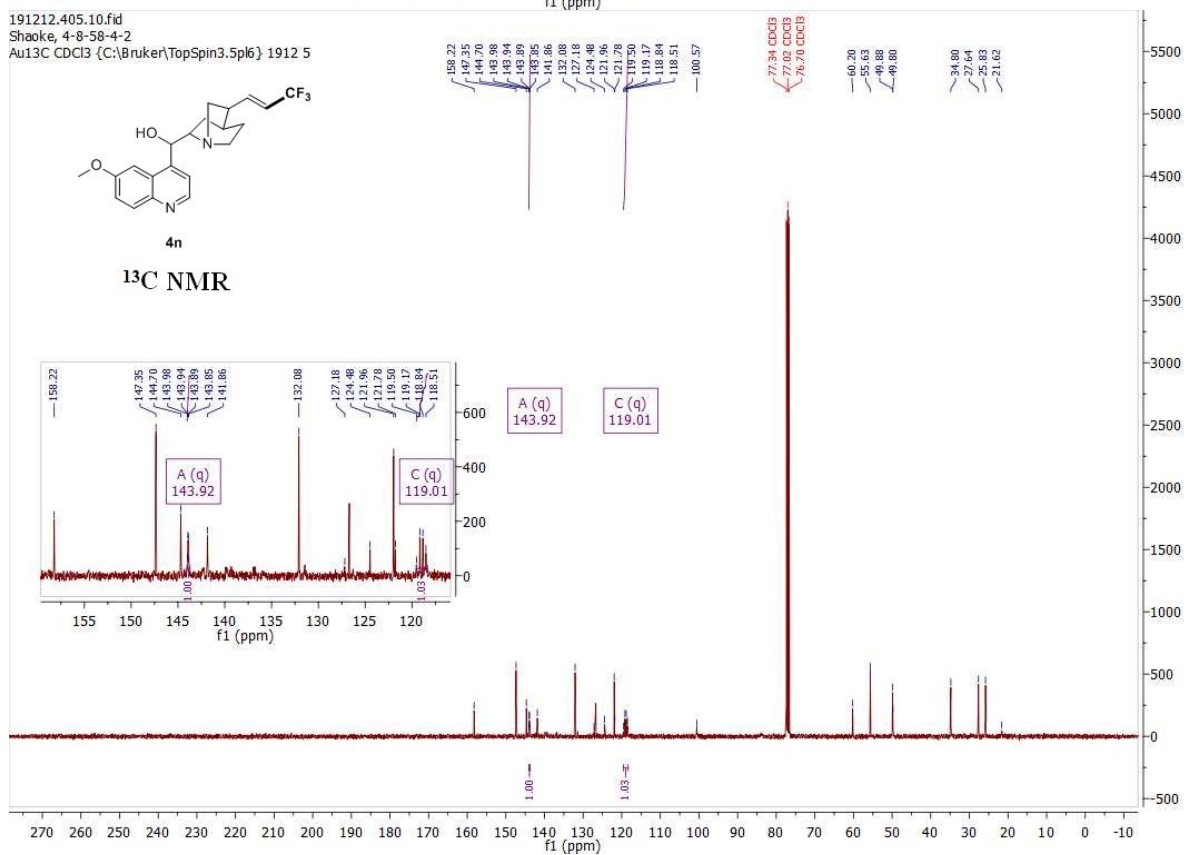


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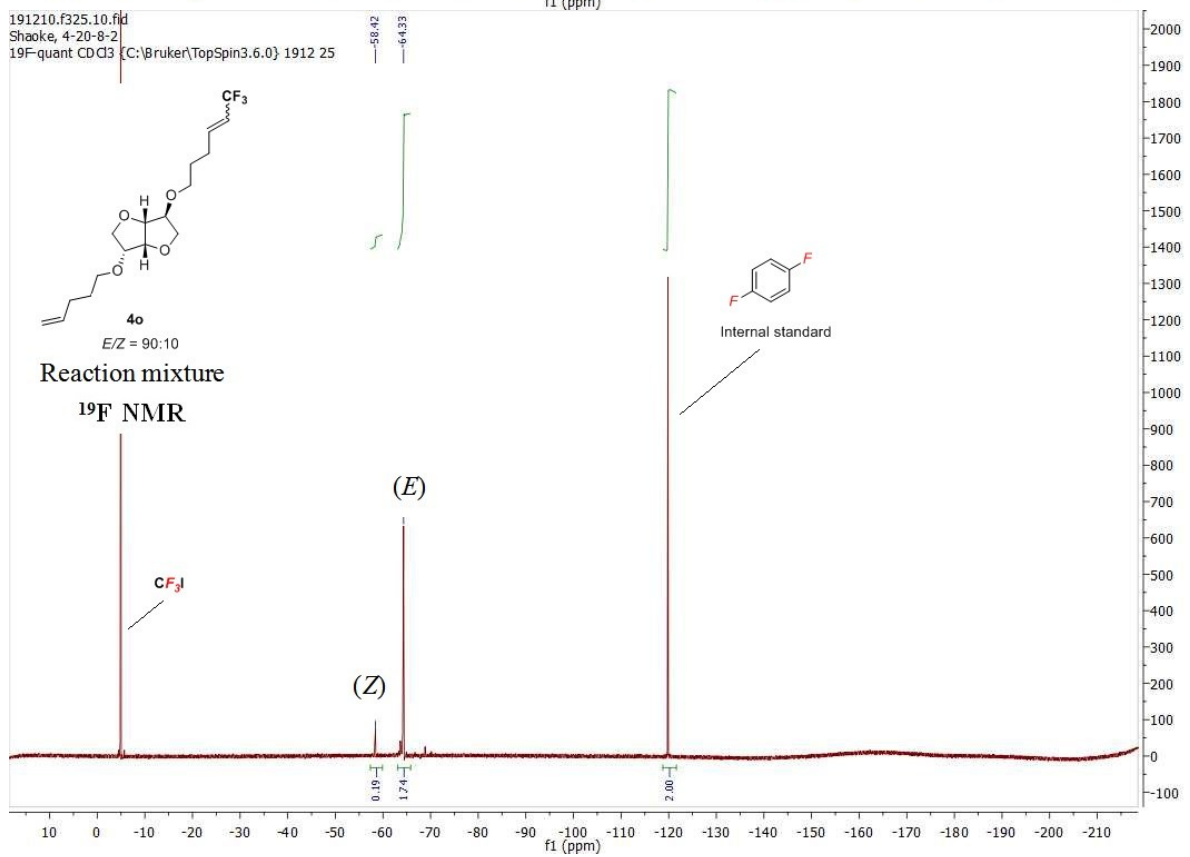
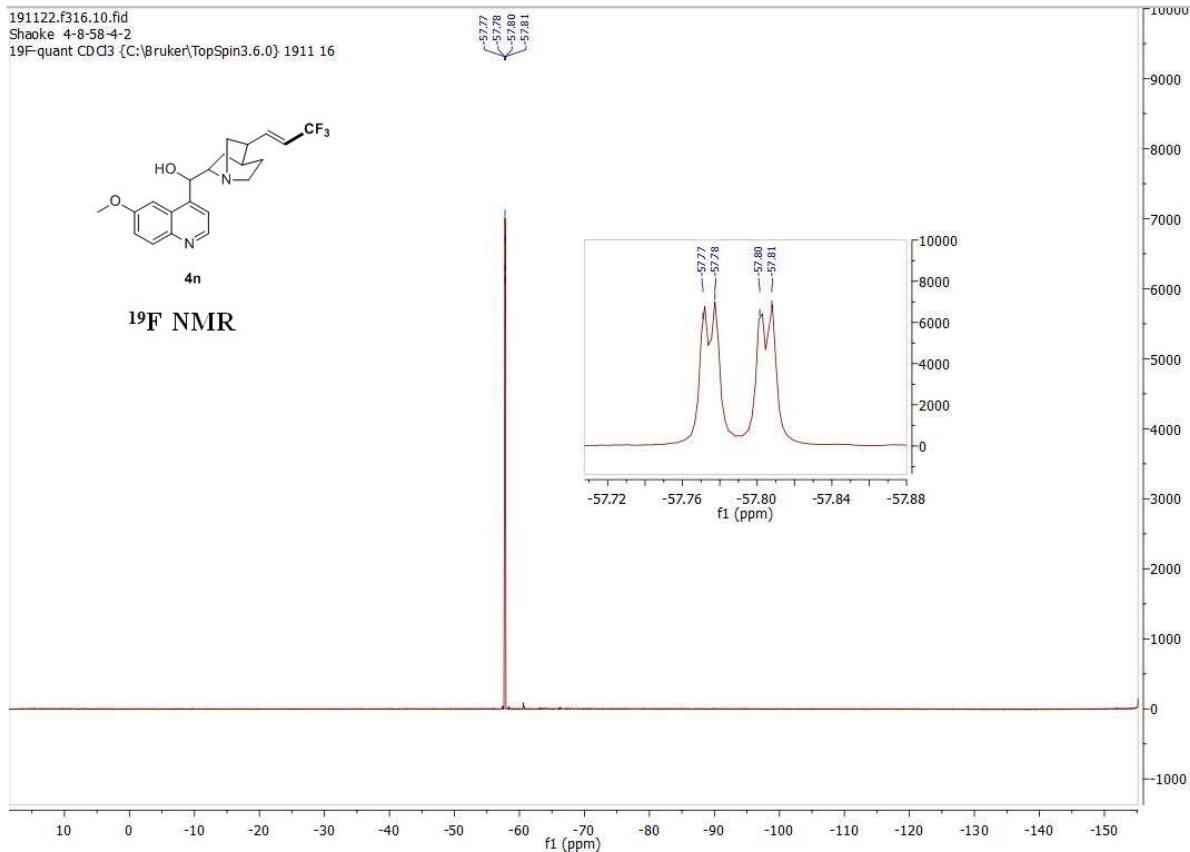
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 Shaoke, 4-8-58-4-2
 Au1H CDCl3 {C:\Bruker\TopSpin3.6.0} 1912 3



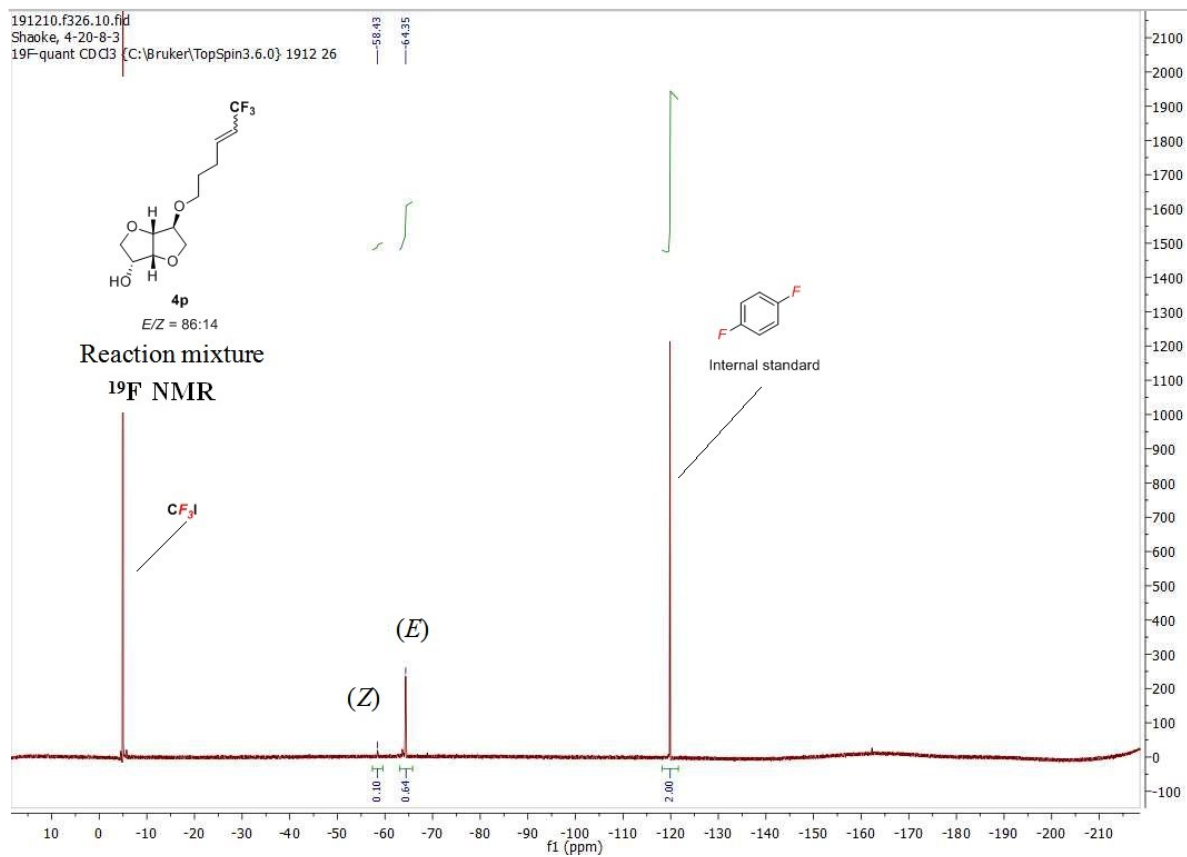
191212.405.10.fid
 Shaoke, 4-8-58-4-2
 Au13C CDCl3 {C:\Bruker\TopSpin3.5pl6} 1912 5



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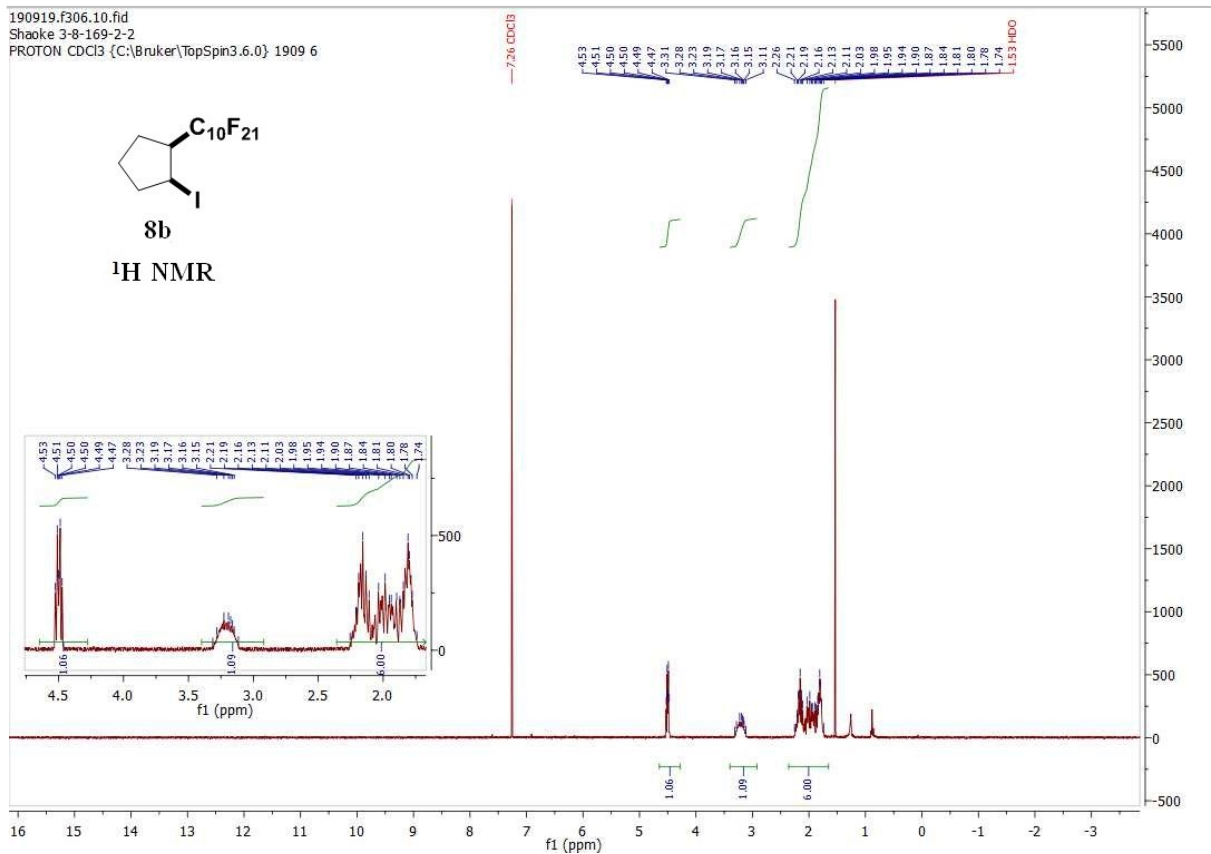


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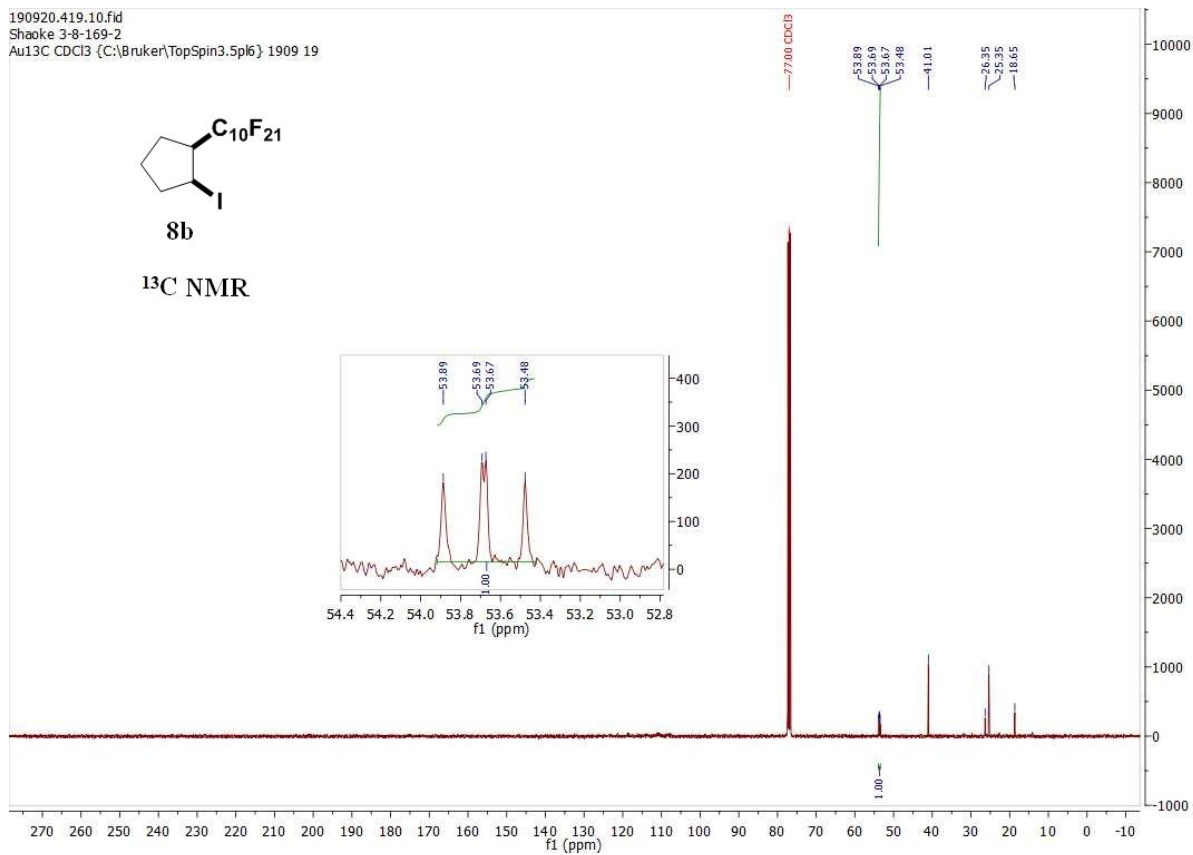


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190919.f306.10.fid
 Shaake 3-8-169-2-2
 PROTON CDCl₃ {C:\Bruker\TopSpin3.6.0} 1909 6



190920.419.10.fid
 Shaake 3-8-169-2
 Au13C CDCl₃ {C:\Bruker\TopSpin3.5pl6} 1909 19



SUPPORTING INFORMATION

