Supporting Information

Reversible Stimuli-Responsive Chromism of a Cyclometallated Platinum(II) Complex

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S1. General experimental procedures

Unless stated otherwise, all chemicals were purchased from commercial sources (Sigma Aldrich UK, Acros UK, VWR UK or Fluorochem UK) and used without further purification. Dry solvents were obtained by means of a "Glass Contour" brand solvent purification system, where solvents were passed through filter columns and dispensed under an argon atmosphere. Flash column chromatography was performed using Geduran[®] Si60 (40-63 mm, Merck, Germany) as the stationary phase, and thin-layer chromatography (TLC) was performed on pre-coated silica gel plates (0.25 mm thick, 60F254, Merck, Germany) and observed under UV light (I_{max} 254 nm). Mass spectrometry was performed using a Bruker MicrOTOF II or Bruker MicrOTOF11 spectrometer for ESI-HRMS. ¹H and ¹³C NMR spectra were recorded on Bruker Ultrashield 400 MHz, Bruker Ascend 500 MHz equipped with a DCH cryoprobe, and Bruker AVANCE III HD at a constant temperature of 25 °C, unless otherwise stated. ¹H, ¹³C, and ¹⁹⁵Pt chemical shifts are reported in parts per million (ppm) from low to high field. ¹H and ¹³C values are referenced to the literature values for chemical shifts of residual nondeuterated solvent, with respect to tetramethylsilane. ¹⁹⁵Pt is referenced externally to K₂PtCl₆ at 0 ppm. Diffusion ordered spectroscopy (DOSY), Nuclear overhauser effect spectroscopy (NOESY) and Exchange spectroscopy (EXSY) were recorded on Bruker AVANCE III HD 500 MHz. Standard abbreviations indicating multiplicity are used as follows: bs (broad singlet), d (doublet), dd (doublet of doublets), m (multiplet), q (quartet), s (singlet), t (triplet), tt (triplet of triplets), J (coupling constant). All spectra were analysed using MestReNova software. NMR tubes, precision glassware and glass syringes were dried under vacuum before use. Fluorescence spectra were recorded by using a Shimadzu RF-5300PC spectrophotometer. UV-vis spectroscopy was performed on a Varian Cary 50 Scan UV Visible Spectrophotometer. Non-deuterated anhydrous solvents were used directly as commercially obtained anhydrous solvents, or were redistilled under reduced pressure from analytical-grade solvents.

Complexes 1-Pt and 1-Pd were synthesised as previously described. All structural and characterisation data matched literature values.^[S1]



Figure S1 ¹H NMR spectrum of complex 1-Pt in CDCl₃.

S2. Solid State Characterisation

S2.1 Visual characterisation

Ambient light UV 365 nm



Figure S2 Photographs of the complex 1-Pt under ambient light and under UV light (365 nm).

S2.2 Solid State Emission Spectroscopy





S2.3 Crystallography

Crystal structures of complexes 1-Pt and 1-Pd were previously published.^[S1]

S3. Solution-Phase Behaviour

S3.1 ¹H NMR dilution experiments

¹H NMR dilution experiments of complexes **1**-Pt or **1**-Pd in CDCl₃ were previously published.^[51]

¹H NMR dilution experiments of complexes **1**-Pt in CD₃OD was performed via following procedures:

Stock solutions of complex **1**-Pt was dissolved in the CD₃OD at high concentrations. A known volume of the stock solution was added to a Wilmad screw-cap NMR tube containing 350 µL of solvent. ¹H NMR spectra were recorded. Further amounts of the stock solution were added and spectra were obtained. The procedure was repeated at several different concentrations.



Figure S4 Partial ¹H NMR spectra of dilution experiments (500 MHz, CD₃OD, 298 K) for complex **1**-Pt and binding isotherms generated based on peaks around 7.7 ppm (black dot) and 6.4 ppm (red dot). The data is fitted to a 1:1 binding isotherm (solid line).





Partial ¹H NMR spectra (500 MHz, CD₃OD, 298 K) of complex **1**-Pt at different concentrations.



S3.2 UV-vis dilution experiments



(A) UV-vis absorption spectra of dilution experiments (MeOH, 298 K) for complex 1-Pt. (B) Linear response in absorption at 475 nm. (C) Non-linear response in absorption at 570 nm.







¹H NMR spectra (500 MHz, 298 K) of complex **1**-Pt over time in degassed methanol-*d*₄.

S3.4 Images of complex 1-Pt over time



Figure S9 Images of complex 1-Pt in degassed anhydrous methanol in a Schlenk tube after standing for different period of time.



S3.5 Time dependant UV-vis spectroscopy

Figure S10UV-vis spectra of 1-Pt (0.25 mM) showing the changing absorbance over time for (A) the first 90 mins
after adding solvent and (B) 5–24 hours after adding solvent. The change in absorbance at 475 nm and
570 nm is shown on the small graphs to the right.

S3.6 Variable temperature UV-vis spectroscopy



Figure S11 (A) Temperature-dependant UV-vis spectra of complex 1-Pt in CHCl₃ (0.2 mM) 24 hours after preparation. (B) Photographs of complex 1-Pt in CHCl₃ 24 hours after preparation under ambient light showing no thermochromic response.

S3.7 Solution-Phase Emission Spectroscopy

Solutions of **1**-Pt in polar solvents ([**1**-Pt] = 0.05 mM) showed strong emission at ~620 nm (480 nm excitation) (Figure S12, ESI⁺), which are red-shifted compared to the two emission peaks at 520 nm and 550 nm observed in apolar solvents, indicating the presence of different species in polar solvents. Further increasing the concentration of **1**-Pt in polar solvents (Figures S13–S14, ESI⁺) allows the appearance of new broad peaks at 650–750 nm, corresponding to metal-metal-to-ligand charge transfer (MMLCT).



Figure S12 Emission spectrum of complex **1**-Pt in different solvents (5×10^{-5} M) excited at 470 nm.



Figure S13Emission spectrum of complex 1-Pt in different solvents (5×10^{-4} M) excited at 570 nm. Broad peaks
corresponding to MMLCT were observed in polar solvents (MeOH, EtOH, acetonitrile) but not in less
polar or apolar solvents (Et₂O, CHCl₃, acetone), suggesting 1-Pt tended to aggregate in polar solvents.



Figure S14 Concentration-dependent fluorescence emission spectrum of complex 1-Pt in methanol excited at 570 nm. The intensity of the broad peaks corresponding to MMLCT increased upon increasing the concentration of 1-Pt from 1×10^{-5} mM to 5×10^{-4} mM. But further increase of concentration to 10^{-3} mM would slightly quench the fluorescence.

S4. Further characterisation of complex 1-Pt in MeOH

S4.1 ¹H and ¹⁹⁵Pt NMR spectroscopy









Comparison of the partial ¹H NMR spectra (500 MHz, 298 K) of complex 1-Pt in CDCl₃ (bottom) and CD₃OD (top). The single peaks in CDCl₃ split into 3 distinct signals in CD₃OD.



Figure S17Partial ¹⁹⁵Pt NMR spectra (500 MHz, 298 K) of complex 1-Pt in CDCl₃ (top) and CD₃OD (bottom). Despite
the inherent broadness, the single peak in CDCl₃ is seen to split into 3 distinct signals in CD₃OD.

S4.2 Synthesis and characterisation of possible cationic diPt complex.

Scheme S1 Synthesis of cationic diPt complex.



The cationic diPt complex $[2-Pt_2]^+BF_4^-$ was synthesised according to a modified procedure of literature.^[52] Complex 1-Pt (0.15 mmol, 0.099 g), complex LPtCl (0.15 mmol, 0.113 g) were dissolved in CH₂Cl₂ (40 mL) in a round bottom flask wrapped with foil, yielding solution 1. Another flask wrapped with foil was charged AgBF₄ (0.18 mmol, 0.035 g) and MeOH (15 mL), giving solution 2. The obtained solution 2 was added dropwise into solution 1 at room temperature. Precipitate was generate immediately and the mixture changed to blue quickly. The mixture was stirred overnight at r.t. After filtration, the filtrate was concentrated. The raw powder was collected and purified by column chromatography (CH_2CI_2 :MeOH = 15:1). The dark blue powder was collected and dried under vacuum (60% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.90 (d, J = 2.7 Hz, 2H), 7.23 (dd, J = 8.9, 2.8 Hz, 2H), 7.20 - 7.12 (m, 4H), 7.07 (d, J = 8.9 Hz, 2H), 6.91 (d, J = 8.9 Hz, 2H), 6.76 (dd, J = 8.2, 7.0 Hz, 1H), 6.68 - 6.60 (m, 3H), 6.46 (d, J = 7.6 Hz, 2H), 3.98 - 3.69 (m, 8H), 2.70 (s, 3H), 1.74 (m, 8H), 1.54 - 1.27 (m, 24H), 1.02 – 0.85 (m, 12H). ¹³C NMR (101 MHz, CDCl₃) δ 163.12, 159.83, 159.37, 154.15, 154.06, 140.08, 139.34, 138.35, 138.04, 124.29, 123.77, 123.47, 123.32, 122.44, 122.20, 120.46, 120.32, 69.12, 69.02, 31.88, 31.75, 29.41, 29.15, 25.86, 25.67, 22.81, 22.73, 16.29, 14.26, 14.21.







¹H NMR spectra (500 MHz, 298 K) of cationic diPt complex [2-Pt₂]⁺BF₄⁻ in CDCl₃.



Figure S19 ¹³C NMR spectra (500 MHz, 298 K) of the cationic diPt complex [2-Pt₂]+BF₄⁻ in CDCl₃.



Figure S20 Images of the cationic diPt complex [**2**-Pt₂]⁺BF₄⁻ in CH₂Cl₂ (DCM) and methanol upon heating and cooling. These solvatochromic and thermochromic properties are consistent with those of **1**-Pt, again suggesting the presence of the cationic [**2**-Pt₂]⁺ complex in the solution of **1**-Pt in methanol.









¹H NMR spectra (500 MHz, 298 K) of complex **1**-Pt and the cationic [**2**-Pt₂]⁺ complex in CD₃OD.



$\mathfrak{3.9}$ 9.7 9.5 9.3 9.1 8.9 8.7 8.5 8.3 8.1 7.9 7.7 7.5 7.3 7.1 6.9 6.7 6.5 6.3 6. δ (ppm)

Figure S23

¹H NMR spectra (500 MHz, 298 K) of complex **1**-Pt and the cationic $[2-Pt_2]^+$ complex in DMSO- d_6 .



Figure S24 ¹H NMR spectra (500 MHz, 298 K) of complex 1-Pt and the cationic [2-Pt₂]⁺ complex in acetonitrile-d₃.



Figure S25 ¹H NMR spectra (500 MHz, 298 K) of complex 1-Pt and the cationic [2-Pt₂]⁺ complex in acetone-*d*₆.

S4.3 2D NMR spectroscopy















NOESY spectra of **1**-Pt in methanol- d_4 .





 1 H- 1 H EXSY spectra of **1**-Pt in methanol- d_{4} (298 K, t_{mix} = 2s).



Figure S33 Electrospray ionisation mass spectrometry of complex 1-Pt from (A) a solution in CH_2Cl_2 and (B) a solution in MeOH.



Figure S34High resolution electrospray ionisation mass spectrometry of the independently synthesised cationic
 $[2-Pt_2]^+BF_4^-$ complex from a solution in CH_2Cl_2 .



 Figure S35
 High resolution electrospray ionisation mass spectrometry of the independently synthesised cationic

 [2-Pt₂]*BF₄⁻ complex from a solution in MeOH.

S4.5 Species of 1-Pt in MeOH solution

The ¹H and ¹⁹⁵Pt NMR spectra of 1-Pt in methanol- d_4 show three distinct groups of peaks, suggesting there are three different chemical environments for Pt centre and some protons.

The independently synthesised cationic $[2-Pt_2]^+BF_4^-$ complex displays overlapped peaks with those of **1**-Pt in methanol- d_4 (Figure S22), suggesting **1**-Pt in methanol is partially transformed into cationic $[2-Pt_2]^+$ species. Consistent overlapped NMR signals are also observed in the 2D DOSY spectra of **1**-Pt and the cationic $[2-Pt_2]^+$ complex (Figures S26–29). The presence of free thiol ligand (Figure S27), which is detached from **1**-Pt, further confirms the presence of the cationic $[2-Pt_2]^+$ complex. This is further supported by the ¹H-¹H NOESY and EXSY spectra (Figures S31-32), in which correlations and exchange are observed for the three CH₃ (from the thiol ligand) protons.

S5. References

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- [S2] V. V. Sivchik, E. V. Grachova, A. S. Melnikov, S. N. Smirnov, A. Yu. Ivanov, P. Hirva, S. P. Tunik, and I. O. Koshevoy, *Inorg. Chem.*, 2016, **55**, 3351–3363.