

## Supporting Information

### Luminescent Naphthalimide-Tagged Ruthenium(II)-Arene Complexes: Cellular Imaging, Photocytotoxicity and Transferrin Binding

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## Stability Studies

The stability of [RuLCl] and [RuLPTA] was determined by <sup>1</sup>H NMR spectroscopy in DMSO-d<sub>6</sub> after 24 h. No additional peaks were observed during the time period (Figure S6, S7). Integration of the peaks for both the complexes in DMSO-d<sub>6</sub> remain consistent with the reported spectra, suggesting higher stability in the solvent. However, the two chlorides attached to the Ru(II) ion in [RuLCl] can potentially undergo hydrolysis in an aqueous medium. To monitor anticipated hydrolysis, we monitored the time-dependent UV-vis spectral changes of the complexes at 298 K in a 4 mM NaCl-aqueous medium. The spectra displayed a decrease in the band centered at 360 nm, while an increase in the band at 440 nm (Figure S8a). However, when the experiment was carried out in 100 mM NaCl aqueous medium, no apparent spectral changes were observed and was found to be stable in such condition (Figure S8b). These data suggests that the complex possibly could remain stable in the bloodstream i.e., in high [Cl<sup>-</sup>] (~100 mM) and will plausibly get aquated when it passes into the cell with low [Cl<sup>-</sup>] (~4 mM). However, [RuLPTA] complex didn't show any noticeable spectral changes of the band at 440 nm in the 4 mM or 100 mM NaCl aqueous solution, while showed some changes in lower wavelength bands ~280 nm (Figure S4c and S4d). This may be due to enhanced hydrophilicity imparted by Ru-bound PTA ligand.

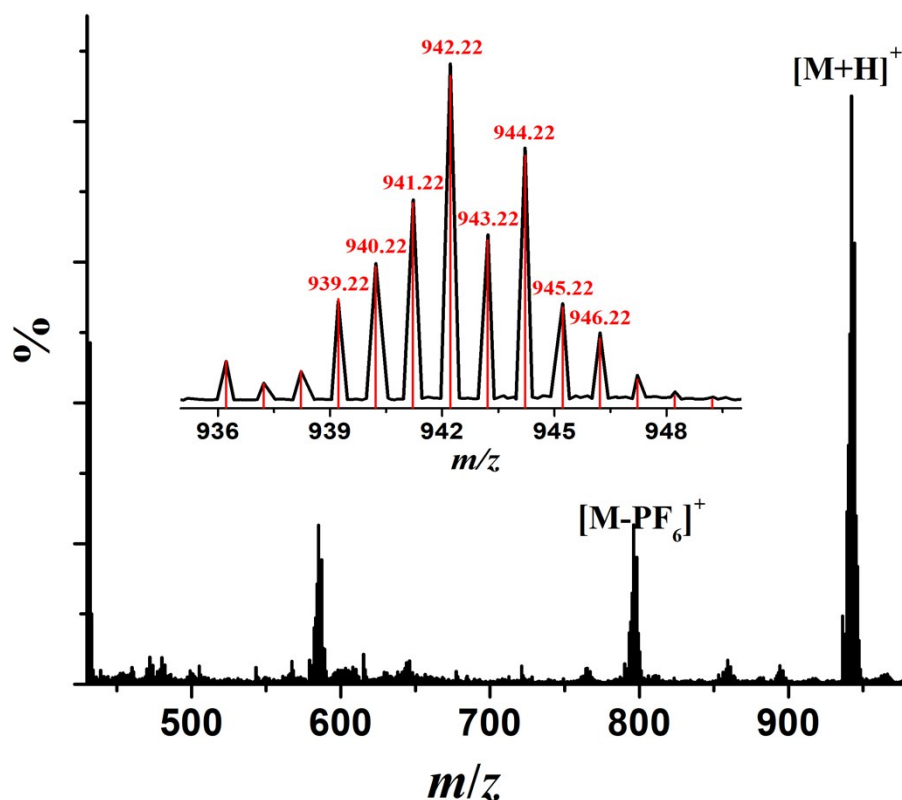
### Molecular Modeling

Molecular docking of [RuLCl] and [RuLPTA] was performed using Autodock-Vina with the crystal structure of human serum transferrin proteins (hTf), PDB IDs: 2HAV and 3QYT, which are apo- and holoprotein, respectively.<sup>S1</sup> For docking, 2HAV was aligned against 3QYT. Both the protein structures were initialized for docking by removing the water molecules or any other non-protein moieties present in the crystal structure. The 2HAV and 3QYT were protonated based on physiological pH value *i.e.* pH=7.2. AutoDockTool (ADT) was used to merge the non-polar hydrogen atoms into their parent heavy-atoms and the system was neutralized using Gasteiger charges.<sup>S2</sup> The crystal structure of [RuLCl] and [RuLPTA] was initially optimized at the PM6 level of theory, using the MOPAC program.<sup>S3</sup> The atoms corresponding to aromatic rings in the ligand molecules, as well as atoms bonded with Ru atom, were fixed during docking procedure while aliphatic chains in the ligand were allowed to rotate providing a wide range of conformational flexibility.

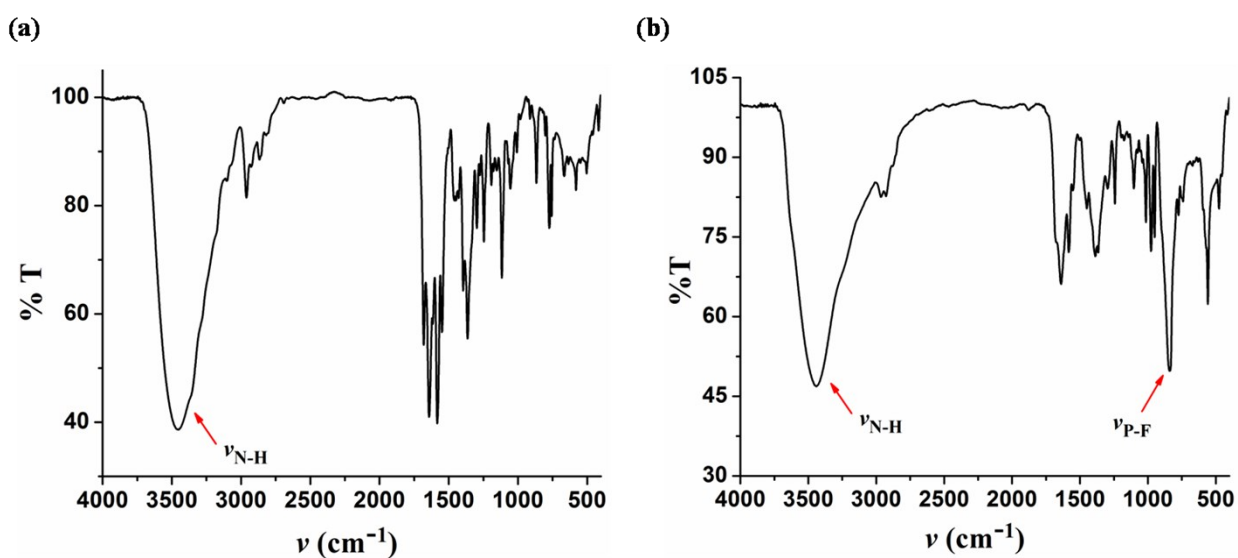
### References

1. O. Trott and A. J. Olson, *J. Comput. Chem.*, 2010, **31**, 455-461.
2. G. M. Morris, R. Huey, W. Lindstrom, M. F. Sanner, R. K. Belew, D. S. Goodsell, A. J. Olson, *J. Computational Chemistry*, 2009, **16**, 2785-2791.
3. MOPAC2016, Version: 16.060W, Stewart, J. J. P. *Stewart Computational Chemistry*. Available online: <http://OpenMOPAC.net>.

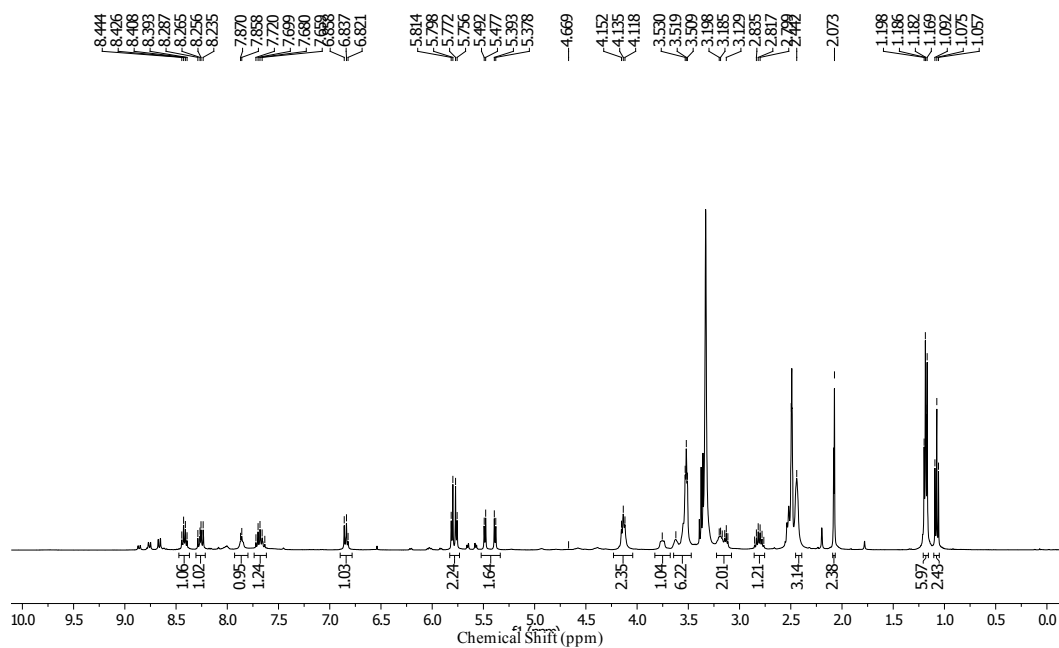




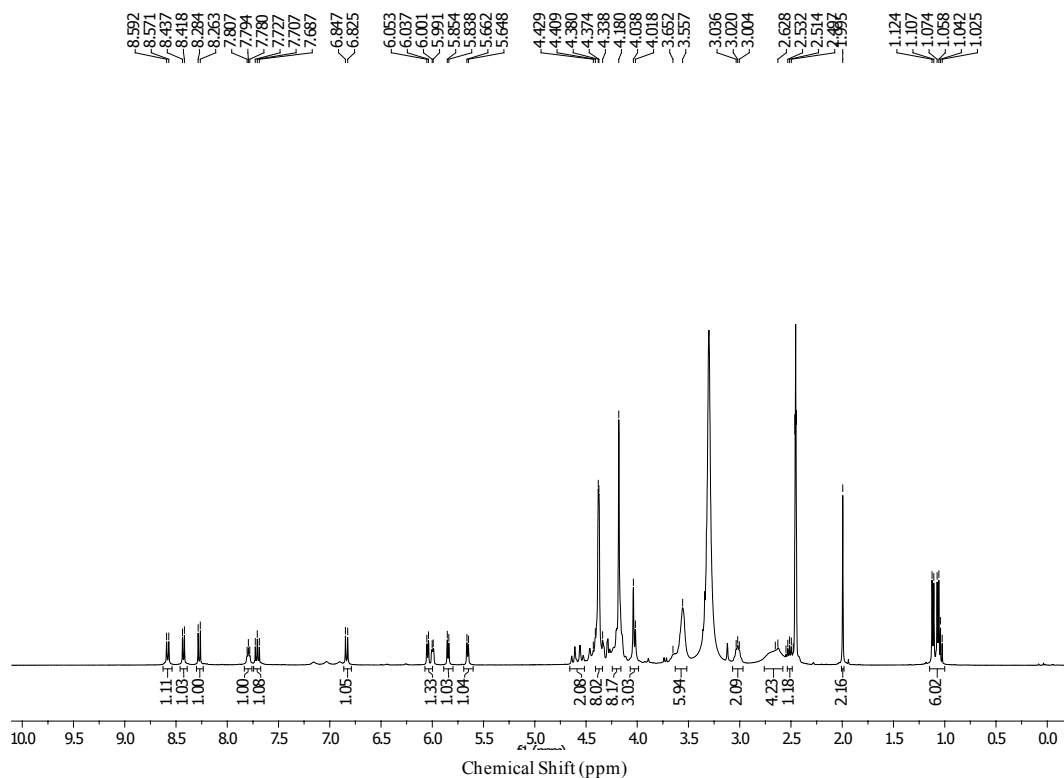
**Figure S2.** ESI-MS of the complex **[RuLPTA]** in  $\text{CH}_2\text{Cl}_2$ . Inset shows  $m/z$  ( $[\text{M}+\text{H}]^+$ ) calc. for  $[\text{C}_{36}\text{H}_{51}\text{N}_7\text{O}_3\text{ClP}_2\text{F}_6\text{Ru}]^+$ : 942.22 (experimentally and theoretically) with matching isotopic distribution pattern.



**Figure S3.** FTIR spectra of the complex (a) **[RuLCl]** and (b) **[RuLPTA]** in KBr phase, characteristic stretching frequencies were mentioned in the spectra.



**Figure S4.**  $^1\text{H}$  NMR spectra of  $[\text{RuLCl}]$  in  $\text{DMSO-d}_6$  at 298 K (400 MHz) using TMS as reference.



**Figure S5**  $^1\text{H}$  NMR spectra of  $[\text{RuLPTA}]$  in  $\text{DMSO-d}_6$  at 298 K (400 MHz) using TMS as reference.

**Table S1.** Selected crystallographic data for the complex [RuLCl]

Parameters	[RuLCl]
Empirical formula	C <sub>31</sub> H <sub>40</sub> Cl <sub>4</sub> N <sub>4</sub> O <sub>3</sub> Ru
Formula weight	759.54
Temperature/ <i>K</i>	273.15
Crystal system	monoclinic
Space group	<i>P</i> 2 <sub>1</sub> / <i>c</i>
<i>a</i> /Å	20.4731(12)
<i>b</i> /Å	19.8145(11)
<i>c</i> /Å	8.2261(5)
$\alpha$ /°	90
$\beta$ /°	91.100(2)
$\gamma$ /°	90
Volume/Å <sup>3</sup>	3336.4(3)
<i>Z</i>	4
$\rho_{\text{calc}}$ /g/cm <sup>3</sup>	1.512
$\mu$ /mm <sup>-1</sup>	0.828
<i>F</i> (000)	1560.0
Crystal size/mm <sup>3</sup>	0.2 × 0.16 × 0.12
Radiation	M <sub>o</sub> K $\alpha$ ( $\lambda$ = 0.71073)
2 $\Theta$ range for data collection/°	5.688 to 56.534
Index ranges	-27 ≤ <i>h</i> ≤ 27, -26 ≤ <i>k</i> ≤ 26, -10 ≤ <i>l</i> ≤ 10
Reflections collected	52151
Independent reflections	8256 [ <i>R</i> <sub>int</sub> = 0.0479, <i>R</i> <sub>sigma</sub> = 0.0313]
Data/restraints/parameters	8256/0/391
Goodness-of-fit on <i>F</i> <sup>2</sup>	1.080
<i>R</i> <sub><i>I</i></sub> <sup><i>a</i></sup> and <i>wR</i> <sub>2</sub> <sup><i>b</i></sup> [[ <i>I</i> ≥ 2 $\sigma$ ( <i>I</i> )]	0.0310, 0.0780
<i>R</i> <sub><i>I</i></sub> and <i>wR</i> <sub>2</sub> [all data]	0.0415, 0.0835
Largest diff. peak/hole / e Å <sup>-3</sup>	0.47/-0.54

<sup>*a*</sup>*R*<sub>1</sub> =  $\sum ||F_0| - |F_C|| / \sum |F_0|$ ; <sup>*b*</sup>*wR*<sub>2</sub> =  $\{\sum [w(F_0^2 - F_C^2)] / \sum [w(F_0^2)^2]\}^{1/2}$

**Table S2.** Selected bond lengths (Å) and angles (°) of the complex [RuLCl] with e.s.d.s. in parentheses.

[RuLCl]	Bond distance
Ru(1)-Cl(1)	2.4057(6)
Ru(1)-Cl(2)	2.4394(6)
Ru(1)-N(1)	2.1312(18)
Ru(1)-C(5)	2.194(2)
Ru(1)-C(3)	2.193(2)
Ru(1)-C(4)	2.165(2)
Ru(1)-C(6)	2.155(2)
Ru(1)-C(7)	2.173(2)
Ru(1)-C(2)	2.208(2)
[RuLCl]	Bond angles
Cl(2)-Ru(1)-Cl(1)	89.92(2)
N(1)-Ru(1)-Cl(1)	79.96(5)
N(1)-Ru(1)-Cl(2)	80.83(5)
N(1)-Ru(1)-C(5)	100.93(8)
N(1)-Ru(1)-C(3)	169.39(8)
N(1)-Ru(1)-C(4)	131.43(8)
N(1)-Ru(1)-C(6)	93.24(8)
N(1)-Ru(1)-C(7)	112.19(8)
N(1)-Ru(1)-C(2)	147.37(8)
C(5)-Ru(1)-Cl(1)	104.85(6)
C(5)-Ru(1)-Cl(2)	165.22(6)
C(5)-Ru(1)-C(2)	81.84(9)
C(3)-Ru(1)-Cl(1)	99.43(7)
C(3)-Ru(1)-Cl(2)	109.79(7)
C(3)-Ru(1)-C(5)	68.88(9)
C(3)-Ru(1)-C(2)	37.09(9)
C(4)-Ru(1)-Cl(1)	87.57(6)
C(4)-Ru(1)-Cl(2)	146.37(7)
C(4)-Ru(1)-C(5)	37.92(8)
C(4)-Ru(1)-C(3)	38.15(9)



C(4)-Ru(1)-C(7)	80.85(9)
C(4)-Ru(1)-C(2)	68.29(9)
C(6)-Ru(1)-Cl(1)	140.91(6)
C(6)-Ru(1)-Cl(2)	127.26(6)
C(6)-Ru(1)-C(5)	38.31(8)
C(6)-Ru(1)-C(3)	80.52(9)
C(6)-Ru(1)-C(4)	68.23(9)
C(6)-Ru(1)-C(7)	38.00(8)
C(6)-Ru(1)-C(2)	68.68(9)
C(7)-Ru(1)-Cl(1)	167.01(7)
C(7)-Ru(1)-Cl(2)	96.44(6)
C(7)-Ru(1)-C(5)	69.20(8)
C(7)-Ru(1)-C(3)	67.76(9)
C(7)-Ru(1)-C(2)	38.10(9)
C(2)-Ru(1)-Cl(1)	131.23(7)
C(2)-Ru(1)-Cl(2)	88.87(6)

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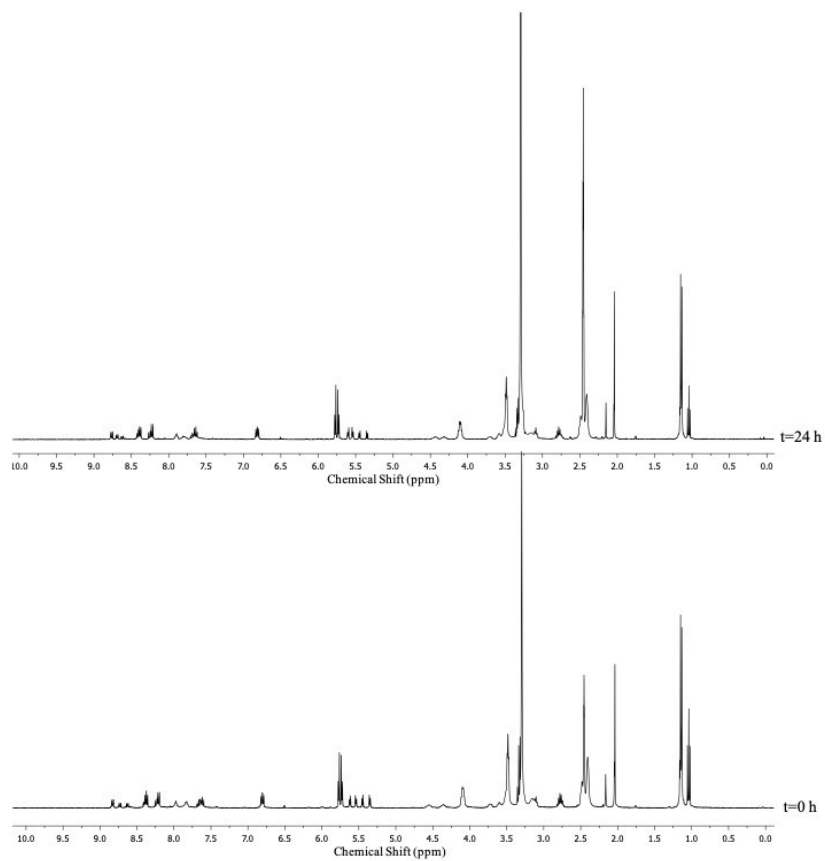


Figure S6.  $^1\text{H}$  NMR spectra showing the stability of  $[\text{RuLCl}]$  in  $\text{DMSO-d}_6$  over 24 h.

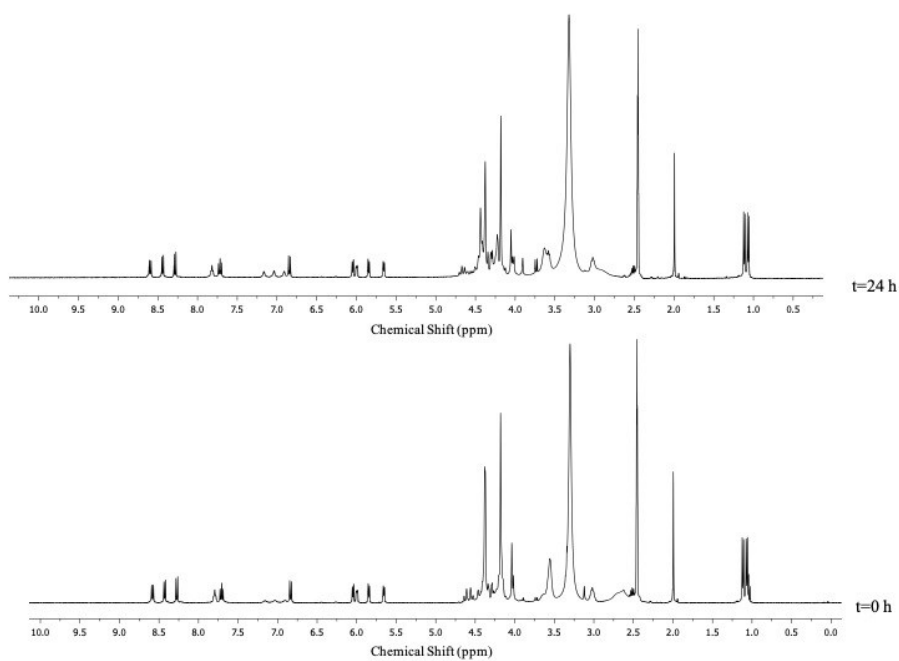
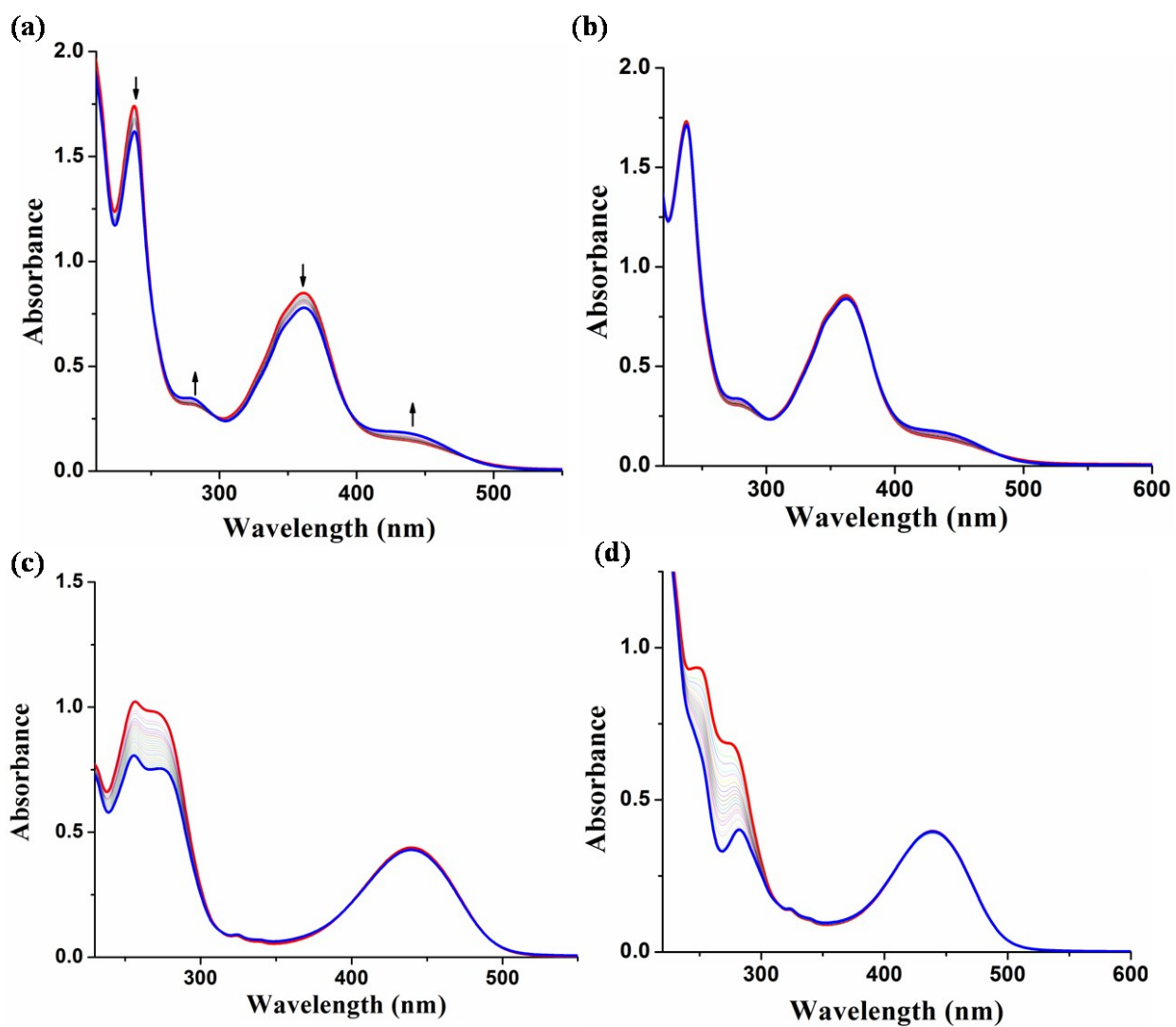
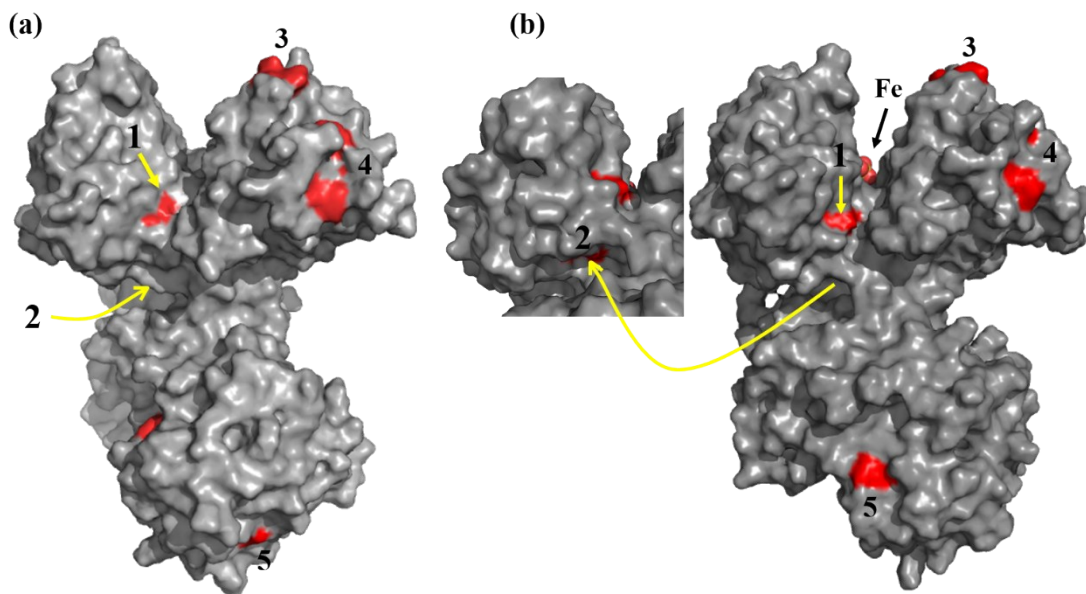


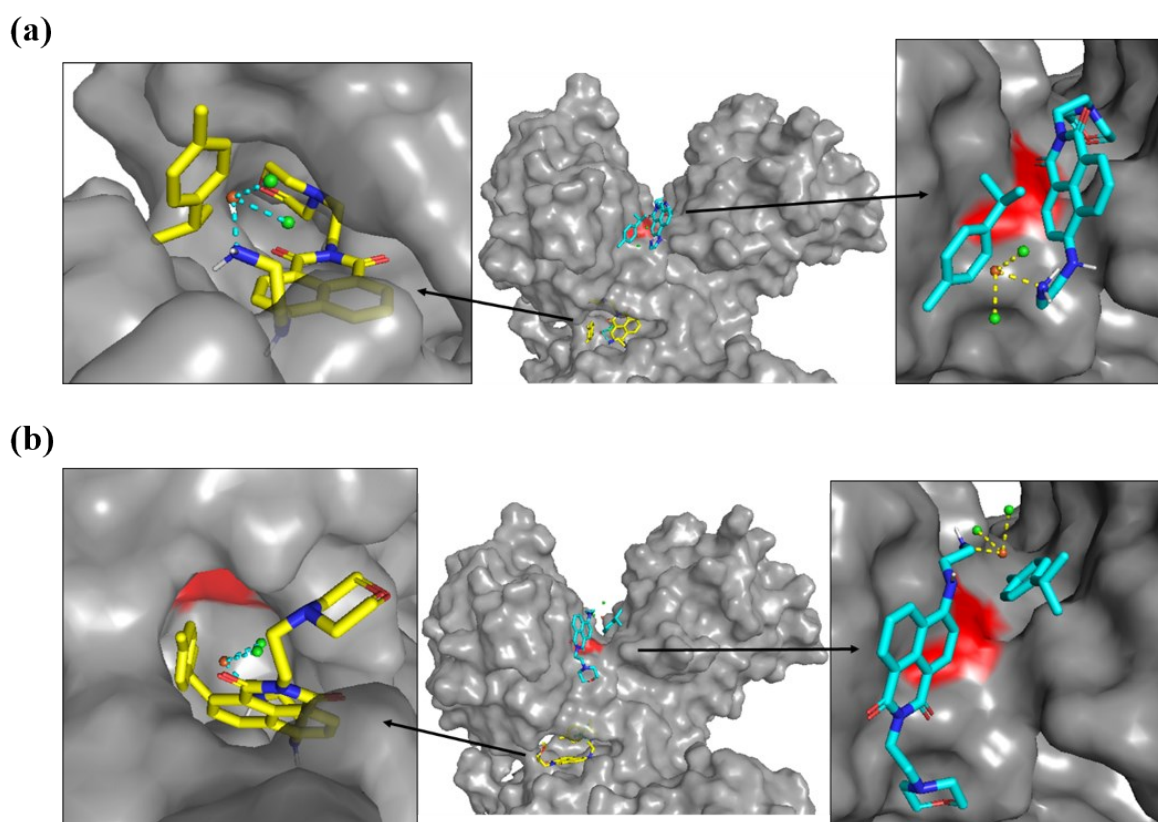
Figure S7.  $^1\text{H}$  NMR spectra showing the stability of  $[\text{RuLPTA}]$  in  $\text{DMSO-d}_6$  over 24 h.



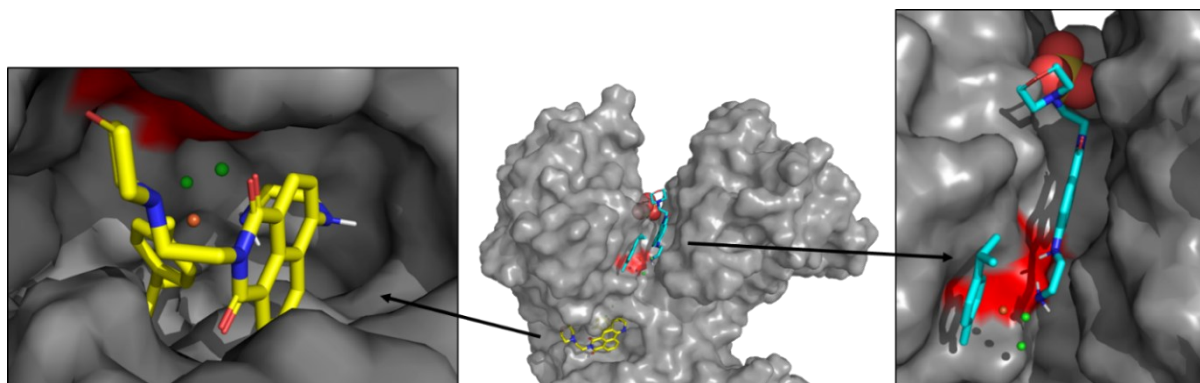
**Figure S8.** Time dependent absorption spectral changes of the complex [RuLCl] ( $54 \mu\text{M}$ ) in (a) 4 mM NaCl-H<sub>2</sub>O medium, (b) 100 mM NaCl-H<sub>2</sub>O medium and [RuLPTA] ( $79 \mu\text{M}$ ) in (c) 4 mM NaCl-H<sub>2</sub>O medium, (d) 100 mM NaCl-H<sub>2</sub>O medium for 2 h at 298 K.



**Figure S9.** Five designated sites used for molecular docking shown on the vdW surface of **a)** apo-hTf (PDB code: 2HAV) and **b)** holo-hTf (PDB code: 3QYT). Each site contains histidine residue(s) (red), SITE1: HIS207; SITE2: HIS242; SITE3: HIS273 and HIS25 SITE4: HIS289 and HIS14; SITE5: HIS578 and HIS535. SITE2 *i.e.* site containing HIS242 lies in a cavity located under the N-lobe and is not visible in 2HAV while it is slightly exposed in 3QYT.



**Figure S10.** Conformation of [RuLCl] docked into the SITE1 and SITE2 on apo-hTf (PDB code: 2HAV) with **a)** highest binding affinity and **b)** third largest binding affinity. Latter is illustrated because Ru of [RuLCl] is closer to HIS207 at SITE1 and HIS242 at SITE2.



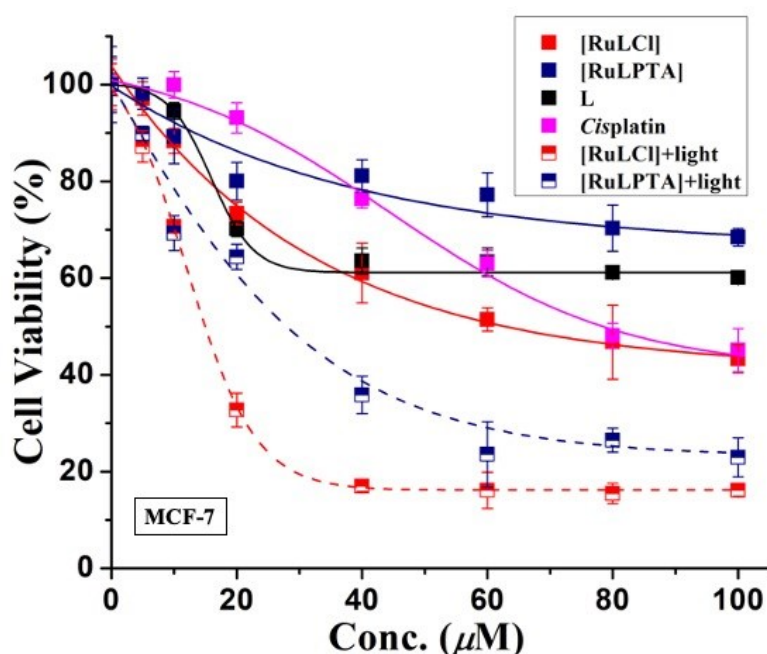
**Figure S11.** Conformation of [RuLCl] docked into the SITE1 and SITE2 on holo-hTf (PDB code: 3QYT) with largest binding affinity.

**Table S3:** Binding affinity of [RuLCl] at five different sites on apo-hTf (PDB code: 2HAV) and holo-hTf (PDB code: 3QYT) obtained using Autodock Vina.

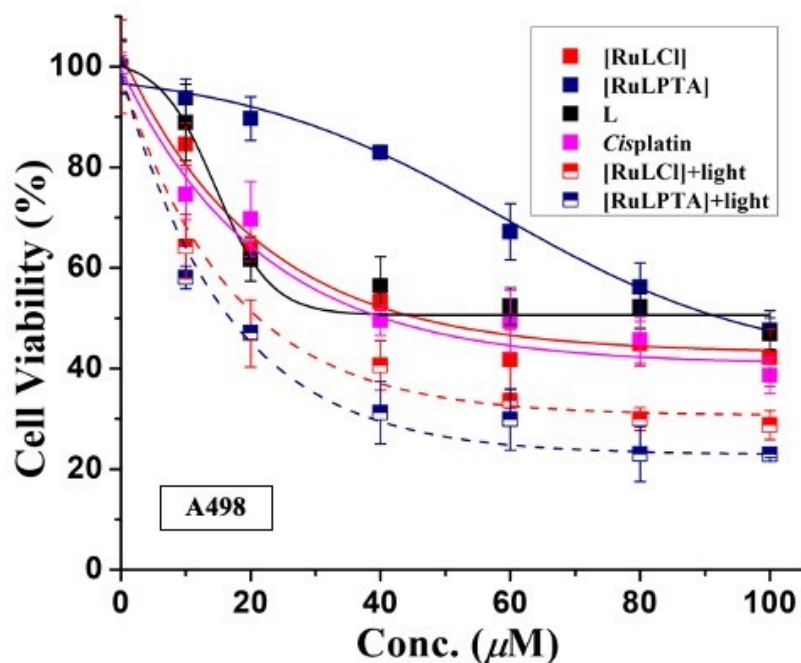
Poses	Binding Affinity (kcal/mol)									
	SITE1		SITE2		SITE3		SITE4		SITE5	
	2HAV	3QYT	2HAV	3QYT	2HAV	3QYT	2HAV	3QYT	2HAV	3QYT
<b>1</b>	-7.0	-9.3	-7.5	-6.9	-5.2	-5.6	-5.1	-4.4	-5.4	-5.6
<b>2</b>	-6.9	-9.1	-7.3	-6.9	-4.8	-5.4	-4.9	-4.3	-5.1	-5.6
<b>3</b>	-6.7	-9.1	-7.2	-6.8	-4.7	-5.4	-4.8	-4.2	-4.9	-5.4
<b>4</b>	-6.7	-8.6	-7.0	-6.8	-4.6	-5.3	-4.8	-4.2	-4.8	-5.1
<b>5</b>	-6.7	-8.5	-7.0	-6.7	-4.4	-5.3	-4.7	-4.1	-4.7	-5.1
<b>6</b>	-6.7	-8.5	-6.9	-6.7	-4.4	-5.3	-4.7	-4.1	-4.7	-5.1
<b>7</b>	-6.6	-8.4	-6.8	-6.6	-4.3	-5.3	-4.7	-4.1	-4.6	-5.1
<b>8</b>	-6.6	-8.1	-6.7	-6.4	-4.3	-5.2	-4.6	-4.0	-4.6	-5.0
<b>9</b>	-6.5	-8.1	-6.7	-6.4	-4.3	-5.2	-4.6	-3.9	-4.4	-5.0

**Table S4:** Binding affinity of [RuLPTA] at five different sites on apo-hTf (PDB code: 2HAV) and holo-hTf (PDB code: 3QYT) obtained using Autodock Vina.

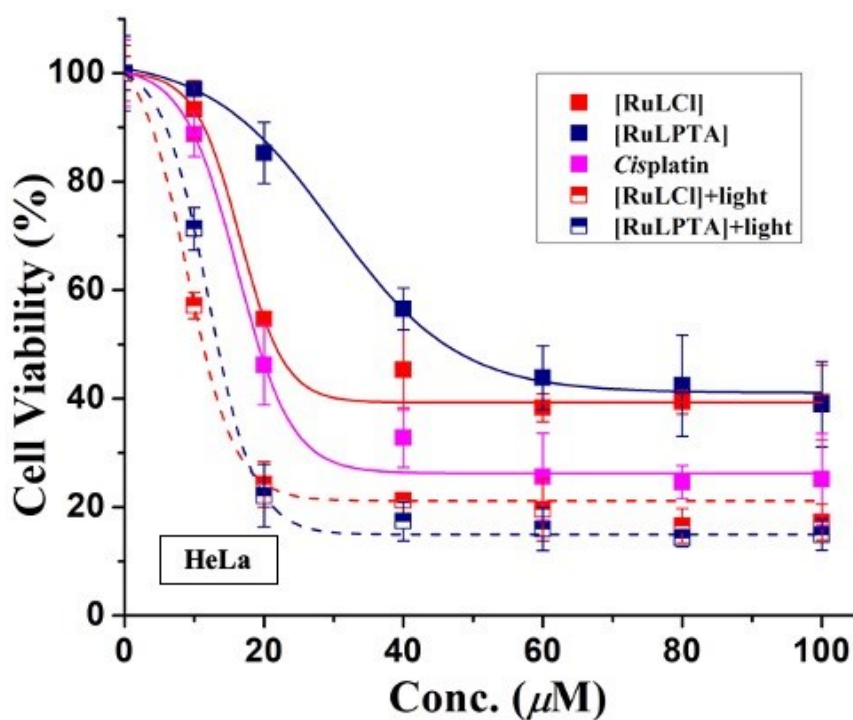
Poses	Binding Affinity (kcal/mol)									
	SITE1		SITE2		SITE3		SITE4		SITE5	
	2HAV	3QYT	2HAV	3QYT	2HAV	3QYT	2HAV	3QYT	2HAV	3QYT
1	-7.4	-9.1	-8.6	-8.7	-5.2	-5.4	-4.2	-4.8	-3.6	-6.2
2	-7.2	-9.0	-7.9	-8.6	-4.8	-5.0	-4.2	-4.7	-3.3	-6.0
3	-7.2	-8.8	-7.9	-8.5	-4.2	-4.9	-4.0	-4.7	-2.4	-6.0
4	-7.1	-8.2	-7.9	-8.4	-4.0	-4.7	-4.0	-4.6	-1.0	-5.9
5	-6.9	-8.2	-7.7	-8.3	-4.0	-4.6	-3.9	-4.6	--	-5.8
6	-6.9	-8.1	-7.4	-8.3	-3.5	-4.6	-3.8	-4.5	--	-5.8
7	-6.7	-7.9	-7.4	-8.2	-3.4	-4.5	-3.8	-4.5	--	-5.8
8	-6.7	-7.8	-7.2	-8.1	-3.3	-4.3	-3.6	-4.5	--	-5.8
9	-6.7	-7.8	-7.2	-7.8	-3.1	-4.3	-3.5	-4.5	--	-5.7



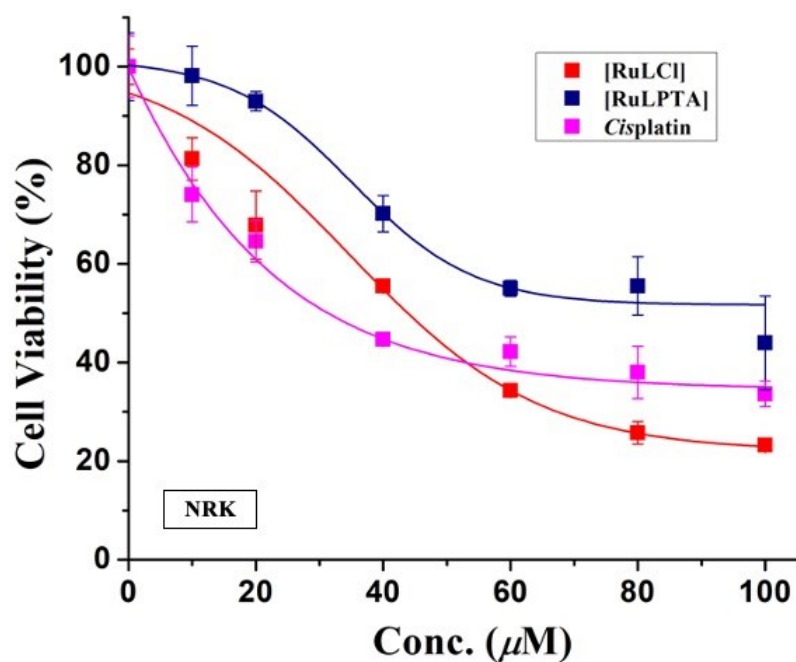
**Figure S12.** MTT assay plot for determination of IC<sub>50</sub> values in MCF-7 cancer cells on incubation for 24 h of the complexes [RuLCl], [RuLPTA], L, cisplatin (0-100 μM) in the dark and on light irradiation ( $\lambda_{irr}$  = 448 nm, 1030 mW @ 700mA from LuxeonStar LEDs) of the complexes for 1 h.



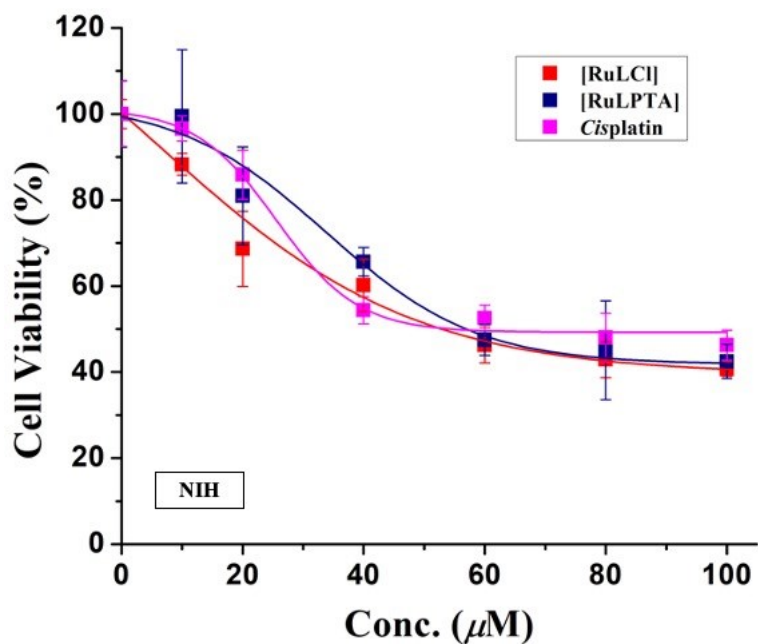
**Figure S13.** MTT assay plot for determination of IC<sub>50</sub> values in A498 cancer cells on incubation for 24 h of the complexes [RuLCl], [RuLPTA], L, cisplatin (0-100 μM) in the dark and on light irradiation ( $\lambda_{\text{irr}} = 448$  nm, 1030 mW @ 700mA from LuxeonStar LEDs) of the complexes for 1 h.



**Figure S14.** MTT assay plot for determination of IC<sub>50</sub> values in HeLa cancer cells on incubation for 24 h of the complexes [RuLCl], [RuLPTA], cisplatin (0-100 μM) in the dark and on light irradiation ( $\lambda_{\text{irr}} = 448$  nm, 1030 mW @ 700mA from LuxeonStar LEDs) of the complexes for 1 h.



**Figure S15.** MTT assay plot for determination of  $IC_{50}$  values in NRK cells on incubation for 24 h of the complexes [RuLCl], [RuLPTA], and cisplatin (0-100  $\mu$ M) in the dark.



**Figure S16.** MTT assay plot for determination of  $IC_{50}$  values in NIH cells on incubation for 24 h of the complexes [RuLCl], [RuLPTA], and cisplatin (0-100  $\mu$ M) in the dark.