

Supplementary information

New Pd-Fe ferrocenyl antiparasitic compounds with bioactive 8-hydroxyquinoline

ligands: a comparative study with their Pt-Fe analogues

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Table S1. Crystal data and structure refinement results for [Pd^{II}(L)(dppf)](PF₆) compounds, where HL= HL1, HL3 and HL4.

	[Pd ^{II} (L1)(dppf)](PF ₆)	[Pd ^{II} (L3)(dppf)](PF ₆)	[Pd ^{II} (L4)(dppf)](PF ₆)
Empirical formula	C ₄₃ H ₃₄ F ₆ FeNOP ₃ Pd [+ solvent]	C ₄₃ H ₃₂ Cl ₂ F ₆ FeNOP ₃ Pd	C ₄₃ H ₃₂ ClF ₆ FeINOP ₃ Pd
Formula weight	949.93	1018.75	1109.20
Temperature (K)	298 (2)	298 (2)	298 (2)
Wavelength (Å)	0.71073	1.54178	1.54178
Crystal system	Monoclinic	Monoclinic	Monoclinic
Space group	P2 ₁ /c	P2 ₁ /c	P2 ₁ /c
Unit cell dimensions			
a (Å)	10.6523 (4)	11.9767 (5)	12.1289 (2)
b (Å)	27.2163 (9)	36.8096 (16)	36.9940 (7)
c (Å)	16.1957 (6)	9.6399 (4)	9.7312 (2)
β (°)	101.297 (1)	110.311 (2)	110.7210 (10)
Volume (Å ³)	4604.4 (3)	3985.6 (3)	4083.92 (13)
Z, density (calculated, g cm ⁻³)	4, 1.3702	4, 1.698	4, 1.804
Absorption coefficient (mm ⁻¹)	0.865	9.501	14.607
F (000)	1912.0	2040.0	2180.0
Crystal shape/color	Rectangular prism/brown	Rectangular prism/brown	Prism/brown
Crystal size (mm ³)	0.185 × 0.222 × 0.383	0.026 × 0.095 × 0.223	0.052 × 0.062 × 0.175
θ-range (°) for data collection	2.9 to 25.41	3.94 to 68.40	4.78 to 68.35
Index ranges	-12 ≤ h ≤ 12, -32 ≤ k ≤ 32, -19 ≤ l ≤ 19	-14 ≤ h ≤ 14, -44 ≤ k ≤ 44, -11 ≤ l ≤ 8	-13 ≤ h ≤ 14, -44 ≤ k ≤ 44, -11 ≤ l ≤ 9
Reflections collected	71524	42643	30208
Independent reflections	8442	7286	7446
Observed reflections [I > 2σ(I)]	7530	6753	6800
Completeness (%)	99.5	99.4	99.3
Absorption correction	multi-scan	multi-scan	multi-scan
Max. and min. transmission	0.7457 and 0.6856	0.4785 and 0.7531	0.4319 and 0.7531
Refinement method	Full-matrix least-squares	Full-matrix least-squares	Full-matrix least-squares

	on F ²	on F ²	on F ²
Data/restraints/parameters	8442/402/564	7286/225/582	7446/281/581
Goodness-of-fit on F ²	1.093	1.020	1.094
Final R indices ^a [I>2σ(I)]	R1 = 0.0376, wR2 = 0.0880	R ₁ = 0.0255, wR ₂ = 0.0616	R ₁ = 0.0312, wR ₂ = 0.0744
R indices (all data)	R1 = 0.0440, wR2 = 0.0929	R ₁ = 0.0284, wR ₂ = 0.0632	R ₁ = 0.0351, wR ₂ = 0.0763
Largest diff. peak and hole (e.Å ⁻³)	0.58/-0.52	0.37/-0.35	0.59/-0.66

$$^a R_1 = \frac{\sum ||F_o| - |F_c||}{\sum |F_o|}, wR_2 = \left[\frac{\sum w(|F_o|^2 - |F_c|^2)^2}{\sum w(|F_o|^2)} \right]^{1/2}$$

Table S2. Mobile phases: Gradients used in the HPLC experiment

Tiempo (min)	% A	% B
0 – 3	100	0
3 – 6	100 – 75	0 – 25
6 – 9	75 – 66	25 – 34
9 – 20	66 – 0	34 – 100
20 – 27	0	100
27 – 30	0 – 100	100 – 0

Table S3. Main FTIR bands for HL1 (8-hydroxyquinoline), HL3 (5,7-dichloro-8-quinolinol) and their [Pd(L)(dppf)](PF₆) complexes (Pd-dppf-L) (cm⁻¹).

	HL1	Pd-dppf-L1	HL3	Pd-dppf-L3
v(O-H)	3204 (m)	-	3164 (m)	-
v(C=N)	1508 (vs)	1501 (vs)	1498 (s)	1492 (s)
v(C-N)	1286 (s)	1316 (m)	1275 (m)	1286 (vw)
v(C-O)	1059 (s)	1110 (m)	1051 (w)	1116 (w)
v: stretching; vs very strong; s strong; m medium; w weak; vw very weak				

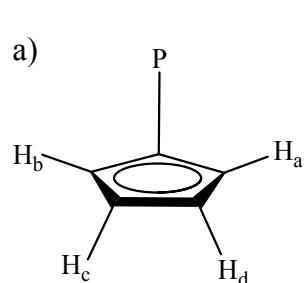
Table S4. Main FTIR bands for HL4 (5-chloro-7-iodo-8-quinolinol), HL5 (5,7-diiodo-8-quinolinol) and their [Pd(L)(dppf)](PF₆) complexes (Pd-dppf-L) (cm⁻¹).

	HL4	Pd-dppf-L4	HL5	Pd-dppf-L5
v(O-H)	3172 (m)	-	3428 (m)	-
v(C=N)	1490 (s)	1486 (m)	1483 (s)	1480 (m)
v(C-N)	1273 (m)	1311 (vw)	1269 (m)	1307 (vw)
v(C-O)	1043 (w)	1098 (m)	1043 (w)	1096 (m)
v: stretching; vs very strong; s strong; m medium; w weak; vw very weak				

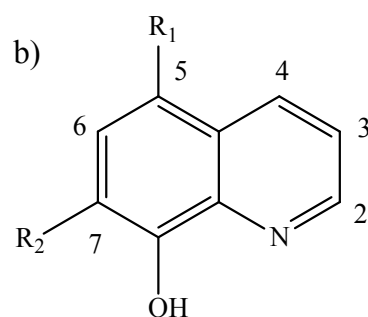
Table S5. Main FTIR bands for HL2 (5-nitro-8-quinolinol) and [Pd(L2)(dppf)](PF₆) (cm⁻¹).

	HL2	Pd-dppf-L2
v(O-H)	3172 (m)	-
v(C=N)	1515(s)	1508 (s)
v(N-O)_{as/s}	1511 (vs) /1289 (s)	1508(s) /1291 (vs)
v(C-N)	1273 (m)	1311 (vw)
v(C-O)	1043 (w)	1113 (vw)
v: stretching; vs very strong; s strong; m medium; w weak; vw very weak		

Table S6. ^1H NMR chemical shift values (δ , in ppm) of the ligands and the complexes in $\text{DMSO}-d_6$.



Cp ring (dppf)



8-hydroxyquinoline

Code	R ₁	R ₂	
HL1	H	H	8-hydroxyquinoline
HL2	NO ₂	H	5-nitro-8-quinolinol
HL3	Cl	Cl	5,7-dichloro-8-quinolinol
HL4	Cl	I	5-chloro-7-iodo-8-quinolinol
HL5	I	I	5,7-diiodo-8-quinolinol

H	δ_{H} (multiplicity, coupling constant (Hz))(integration)														
	HL1	Pd-dppf-L1	$\Delta\delta^a$	HL2	Pd-dppf-L2	$\Delta\delta^a$	HL3	Pd-dppf-L3	$\Delta\delta^a$	HL4	Pd-dppf-L4	$\Delta\delta^a$	HL5	Pd-dppf-L5	$\Delta\delta^a$
2	8.84 (dd, 4.1, 1.6) (1)	8.43 (d, 8.0) (1)	-0.41	9.02 (dd, 4.1, 1.2) (1)	9.31 (d, 8.9) (1)	0.29	9.00 (d, 3.3) (1)	8.55 (m)(1)	-0.45	8.97 (dd, 4.2, 1.4) (1)	8.54 (d, 8.5) (1)	-0.43	8.87 (dd, 4.1, 1.0) (1)	8.34 (br) (1)	-0.53
3	7.54 (dd, 8.3, 4.2) (1)	7.10 (m)(1) ^c	-0.44	7.89 (dd, 8.9, 4.1) (1)	7.42 (dd, 8.9, 5.0) (1)	-0.47	7.75 (dd, 8.5, 4.2) (1)	7.26 (m)(2)	-0.49	7.77 (dd, 8.6, 4.2) (1)	7.24 (m) (1)	-0.53	7.73 (dd, 8.5, 4.1) (1)	7.19 (br) (1)	-0.54
4	8.31 (dd, 8.3, 1.6) (1)	7.15 (m)(1)	-1.16	9.15 (dd, 8.9, 1.1) (1)	7.54 (m)(1) ^b	-1.61	8.50 (d, 8.5) (1)	NA	-	8.50 (dd, 8.5, 1.4) (1)	7.08 (br) (1)	-1.42	8.29 (dd, 8.6, 1.0) (1)	7.04 (br) (1)	-1.25
5	7.38 (dd,	7.10 (m)(1) ^c	-0.28	-	-	-	-	4	-	-	-	-	-	-	-

	8.2, 1.4) (1)														
6	7.44 (m) (1)	7.36 (t, 7.8) (1)	-0.08	8.55 (d, 8.8) (1)	8.49 (d, 9.3) (1)	-0.06	7.82 (s)(1)	NA	-	8.00 (s)(1)	7.92 (s) (1)	-0.08	8.34 (s) (1)	8.20 (br) (1)	-0.14
7	7.09 (dd, 7.4, 1.5) (1)	6.30 (d, 7.7) (1)	-0.79	7.20 (d, 8.8) (1)	6.22 (d, 9.3) (1)	-0.98	-	-	-	-	-	-	-	-	-
H _α	-	5.07 (s)(2)	1.13	-	4.73 (s)(4)	-	-	5.18 (br)(2)	-	-	4.72 (s)(4)	-	-	4.72 (br) (4)	-
		3.82 (s)(2)	-0.12												-
H _β	-	4.81 (s)(2)	0.54	-	4.60 (s)(2)	-	-	4.67 (br)(4)	-	-	4.47 (br) (2)	-	-	4.57 (br) (2)	-
		4.59 (s)(2)	0.32		4.30 (br) (2)	-		3.72 (br)(2)	-			4.29 (br) (3)		-	

^a: $\Delta\delta: \delta_{\text{complex}} - \delta_{\text{ligand}}$

^b: overlapped with PPh₂ protons signals

^c: protons 3 and 5 show the same chemical shift, integrating the signal for two protons.

NA: Not assigned.

Multiplicity: s: singlet, d: doublet, dd: doublet of doublets, t: triplet, m: multiplet, br: broad.

Table S7. Retention time values for precursors [MCl₂dppf], ligands HL4 and dppf and M-dppf-L4 compounds, M = Pd(II) or Pt(II) using DMSO as solvent at t = 0.

Compounds	RT(min)
[Pd(L4)(dppf)](PF ₆)	24.797
[Pt(L4)(dppf)](PF ₆)	24.433
[PdCl ₂ (dppf)]	22.165
[PtCl ₂ (dppf)]	21.807
Dppf	27.297
HL4	22.279

Table S8. Retention time (RT) values for M-dppf-L4 compounds, M = Pd(II) or Pt(II) in DMSO/10 mM Tris-HCl buffer solution pH 7.4 (4:1).

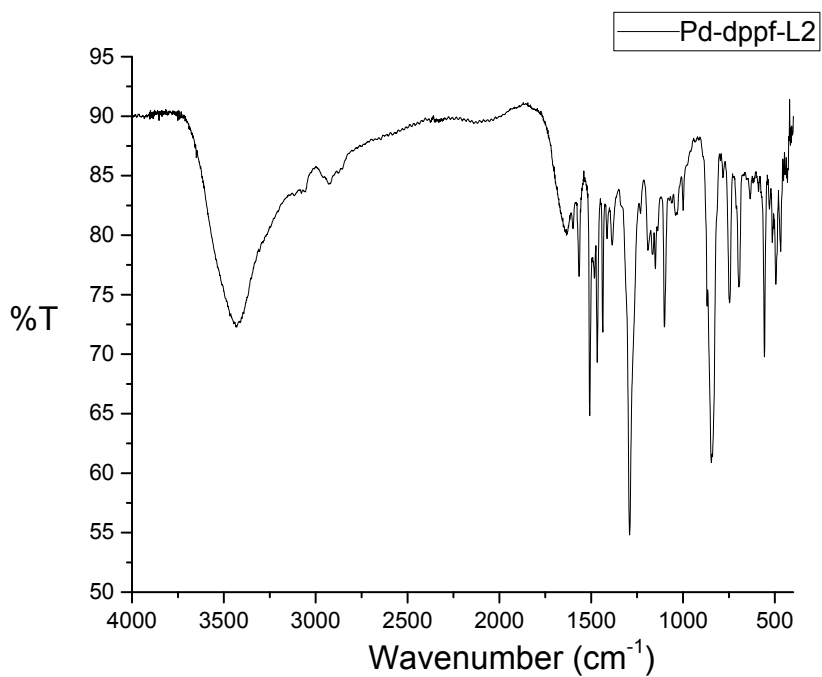
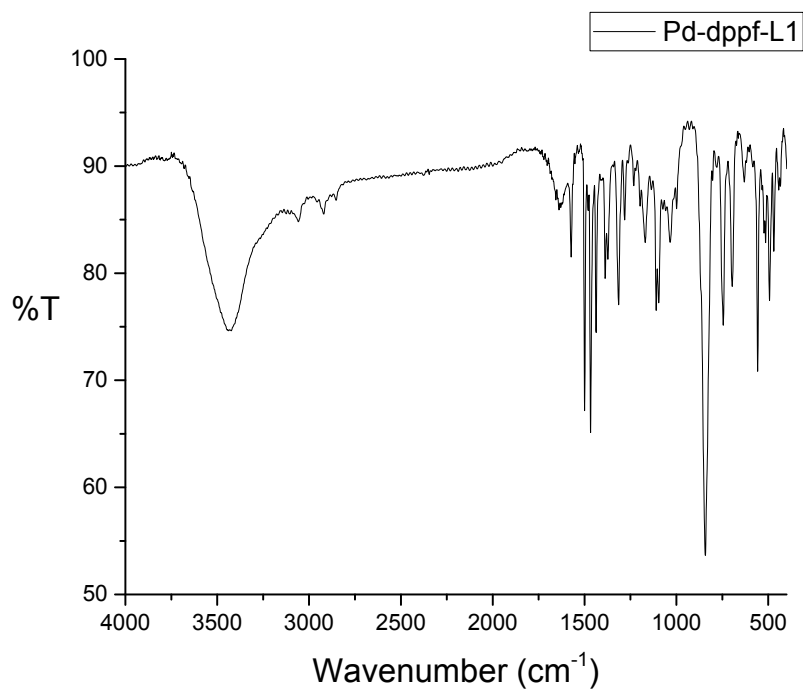
Time	RT (min)	
	Pd-dppf-L4	Pt-dppf-L4
t = 0	24.846	24.752
t = 24 h	24.862	24.713

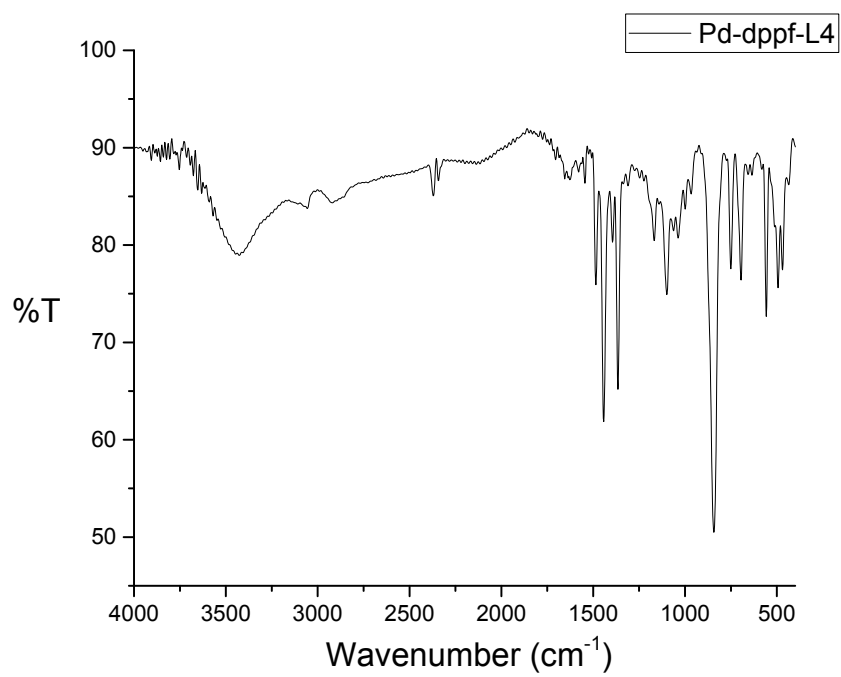
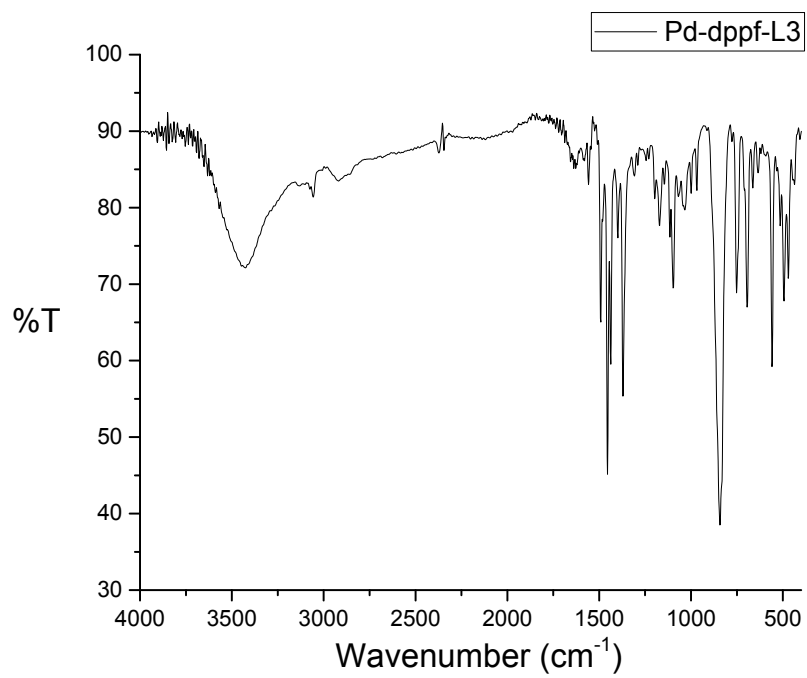
Table S9. Stern-Volmer constants of the Pd-dppf-L compounds and their Pt analogues for the competitive binding to {DNA-EB} adduct in 5 % DMSO/Tris HCl medium. Quenching % is included for each Pd-dppf-L compound.

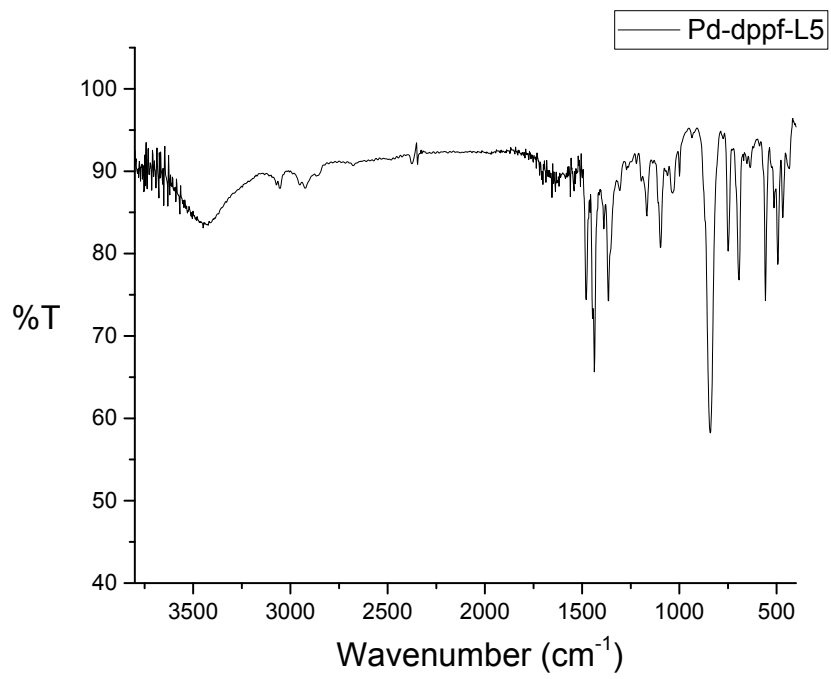
Compound	K _{SV} (M ⁻¹)	Log (K _{SV})	Quenching (%)	Compound	K _{SV} (M ⁻¹) ^a	Log (K _{SV}) ^a
Pd-dppf-L1	15957	4.2	60 (100 μM)	Pt-dppf-L1	2102	3.3
Pd-dppf-L2	6793	3.8	52 (110 μM)	Pt-dppf-L2	7744	3.9
Pd-dppf-L3	19525	4.3	75 (100μM)	Pt-dppf-L3	3599	3.6
Pd-dppf-L4	23471	4.4	77 (110μM)	Pt-dppf-L4	6053	3.8
Pd-dppf-L5	43942	4.6	77 (110μM)	Pt-dppf-L5	3505	3.5

^a Data from [10]

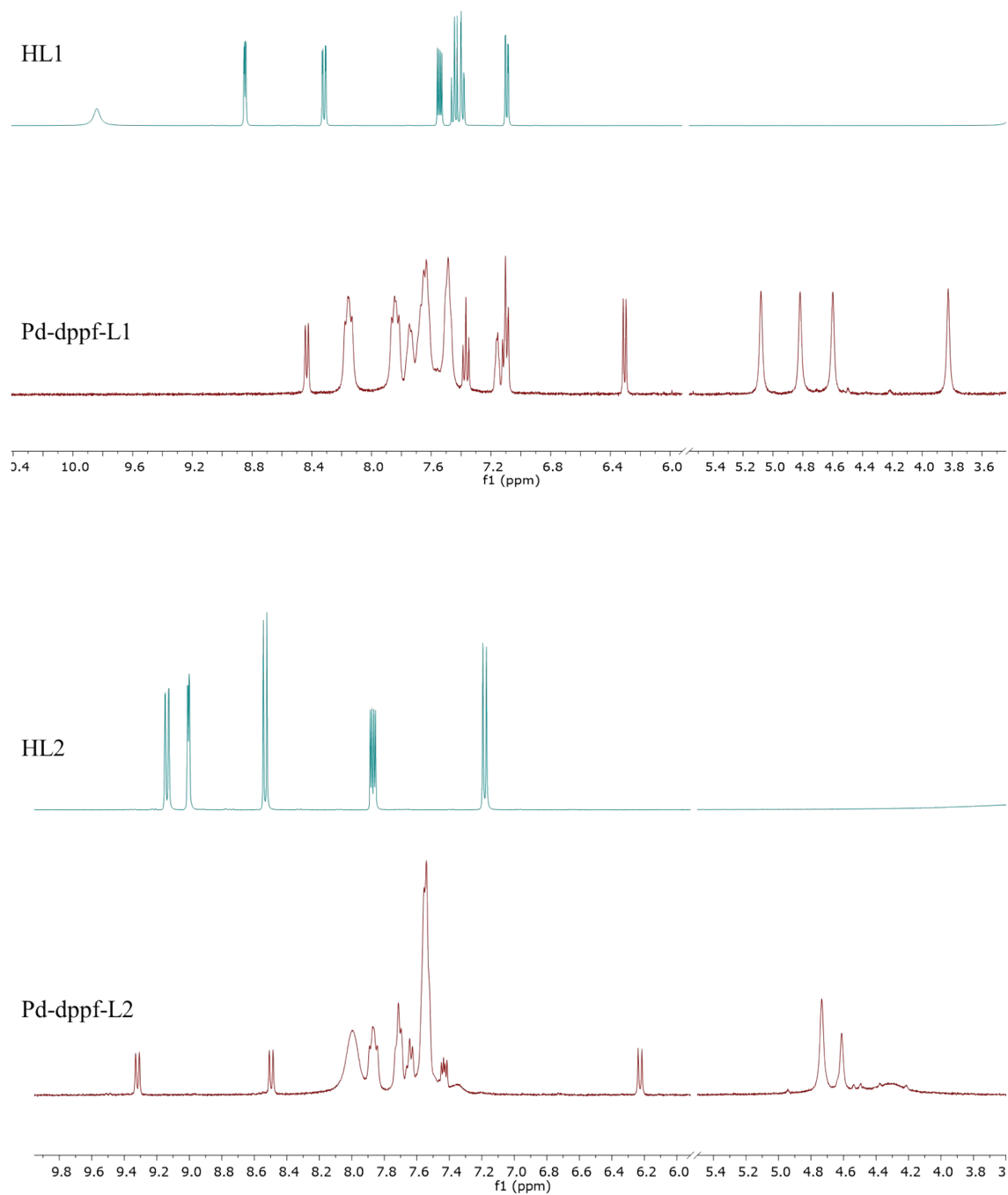
Figures S1. FTIR spectra in KBr pellets (4000 - 400 cm^{-1}) of complexes $[\text{Pd}(\text{L})(\text{dppf})](\text{PF}_6)$, where HL = HL1-HL5.

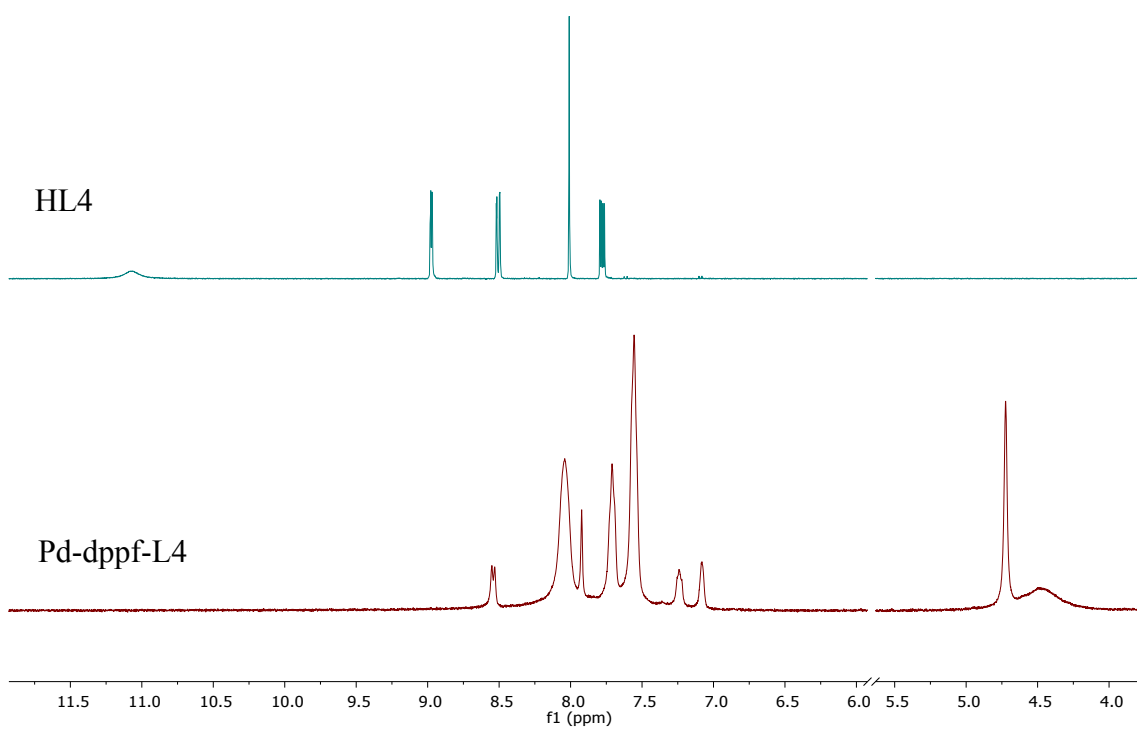
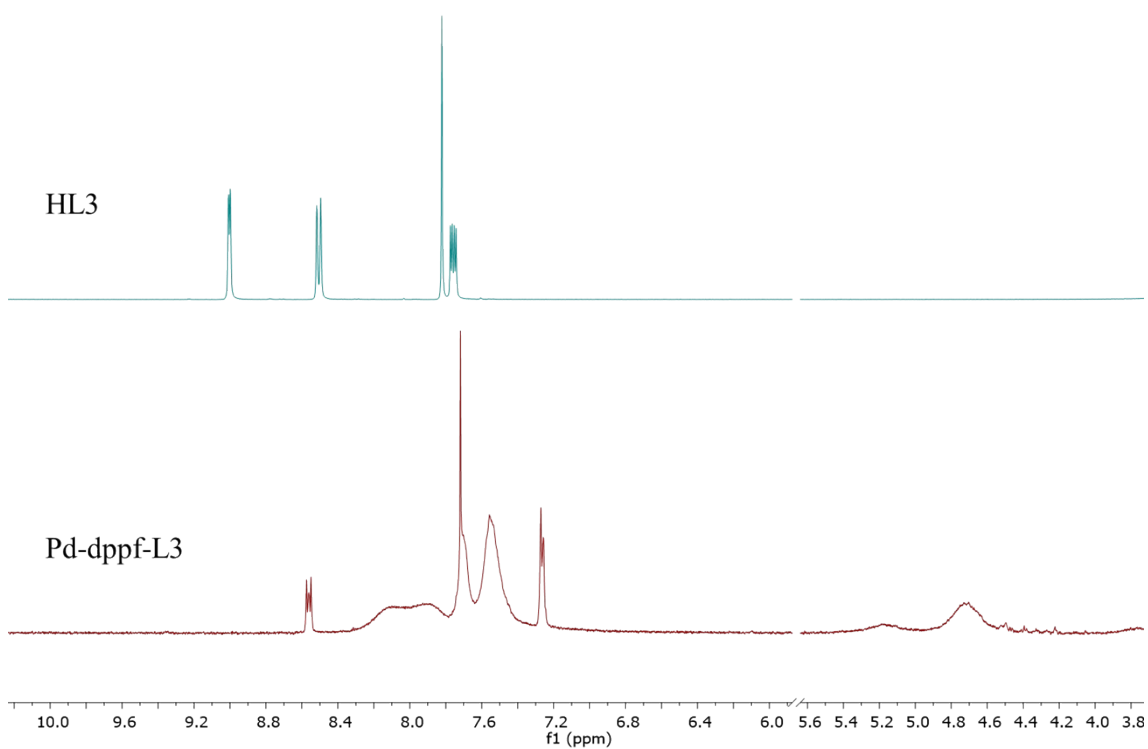






Figures S2. ^1H -NMR spectra of complexes $[\text{Pd}(\text{L})(\text{dppf})](\text{PF}_6)$ in $\text{DMSO-}d_6$ solution, where $\text{HL} = \text{HL1-HL5}$, compared with the respective HL ligands.





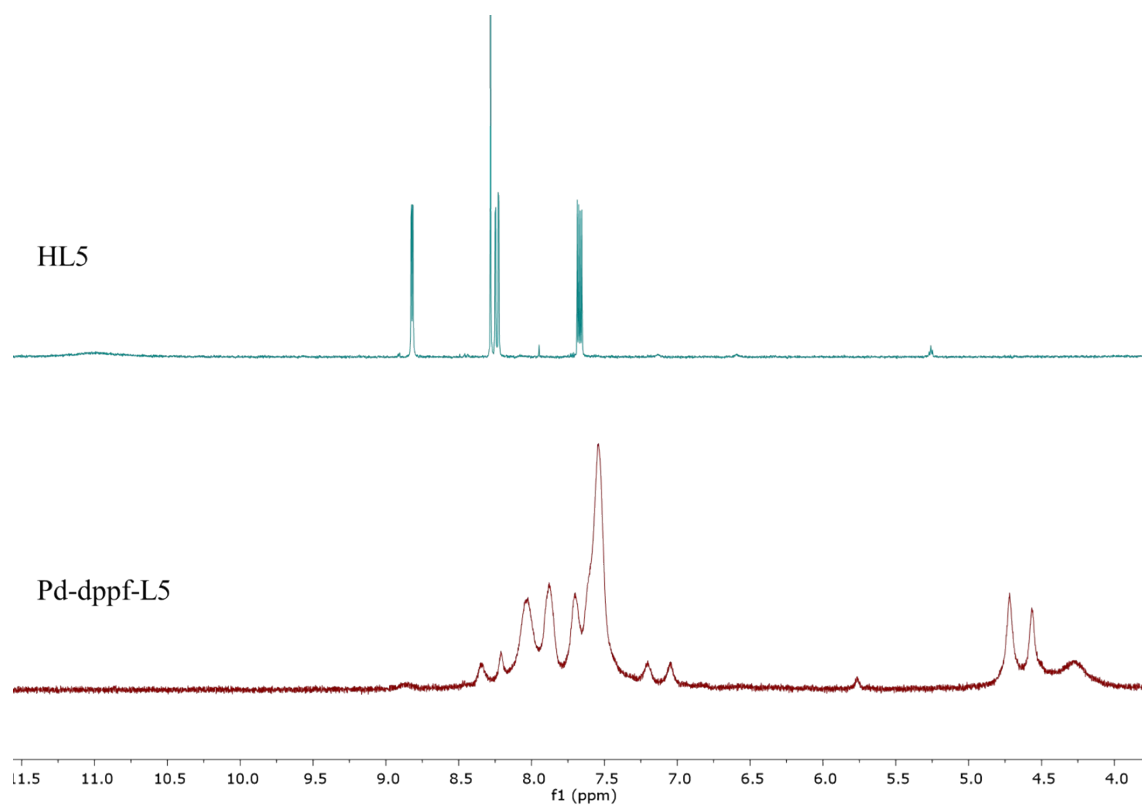


Figure S3. Chromatograms for Pd-dppf-L4 in DMSO/10 mM Tris-HCl buffer solution pH 7.4 (4:1) at t = 0 and t = 24 h.

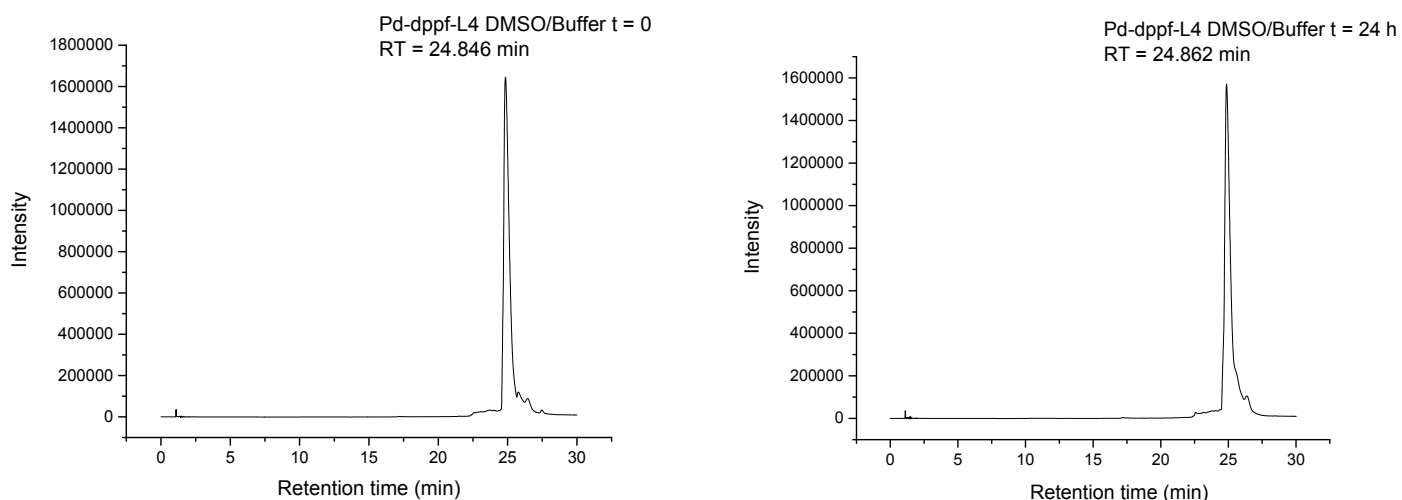


Figure S4. Chromatograms for Pt-dppf-L4 in DMSO/10 mM Tris-HCl buffer solution pH 7.4 (4:1) at t = 0 and t = 24 h

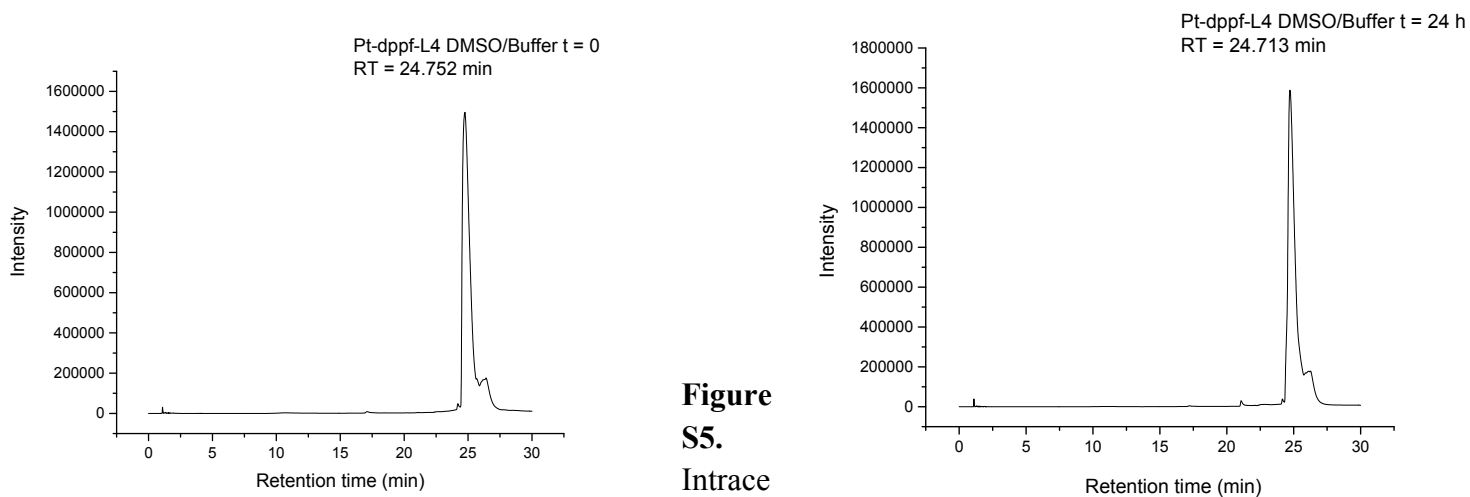
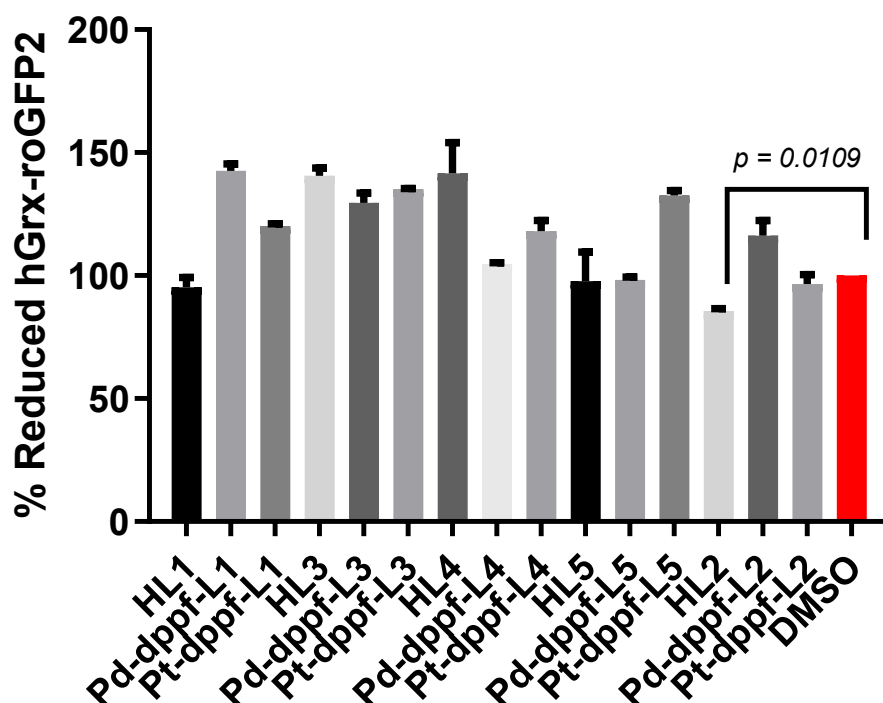


Figure S5.
Intracellular

redox state of bloodstream *T. brucei* treated with the compounds under study. The bloodstream form of *T. b. brucei* (5×10^5 parasites/mL) was incubated 24 h with each compound at concentrations closest to their corresponding EC_{50} values (see Table below). Thereafter, the intracellular redox state was assessed by measuring fluorescence intensity of the redox biosensor hGrx-roGFP2 expressed by the parasites. Controls included parasites treated with vehicle (DMSO 1%, v/v, red bar). For details in the protocol see section Material and Methods. The results are expressed as mean % biosensor reduction

± S.D (n = 3) with respect to the vehicle-treated parasites (100% biosensor reduction, red bar). The probability index for the compound displaying a significant oxidation of the redox biosensor is shown as inset compared to DMSO (unpaired t-Test).



Compound	IC ₅₀ <i>T. brucei</i> (μM)	Concentration tested in redox assay
HL1	12.4 ± 0.8	12.5 μM
Pd-dppf-L1	0.9 ± 0.2	1 μM
Pt-dppf-L1	0.3 ± 0.1	0.25 μM
HL3	2.5 ± 0.2	2.5 μM
Pd-dppf-L3	4.5 ± 0.1	2.5 μM
Pt-dppf-L3	0.22 ± 0.01	0.25 μM
HL4	2.4 ± 0.4	2.5 μM
Pd-dppf-L4	4.8 ± 0.5	2.5 μM
Pt-dppf-L4	0.14 ± 0.05	0.5 μM
HL5	3.0 ± 0.2	2.5 μM
Pd-dppf-L5	7 ± 4	0.01 μM
Pt-dppf-L5	0.22 ± 0.04	0.5 μM
HL2	0.8 ± 0.3	1 μM

Pd-dppf-L2	0.33 ± 0.09	0.25 μ M
Pt-dppf-L2	0.93 ± 0.03	1 μ M