

## Supplementary Information

### Synthesis of UCST-type zwitterionic polymer for efficiently recycling cellulase at room temperature

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## The $^1\text{H-NMR}$ characterization of PSPE

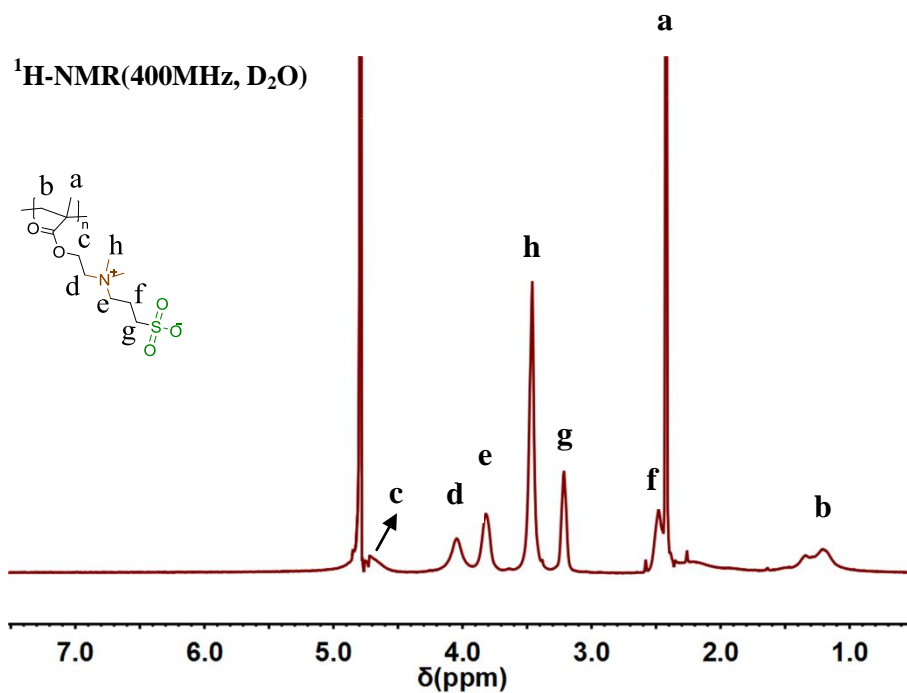
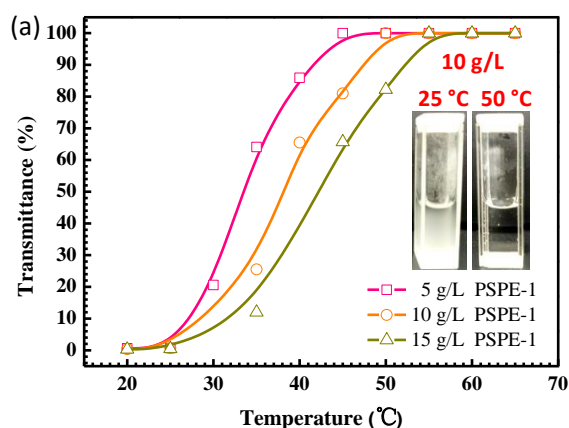
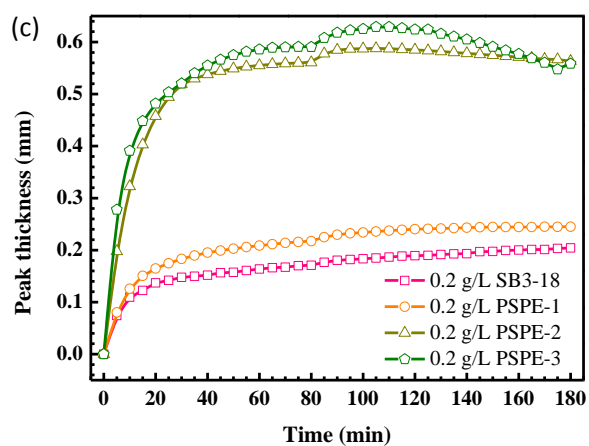
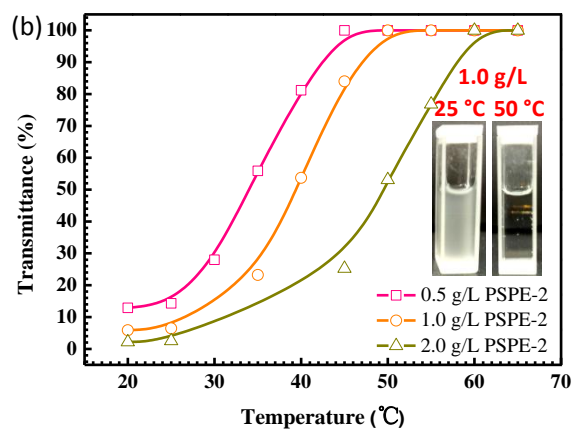


Fig. S1. The  $^1\text{H-NMR}$  spectra of PSPE.

PSPE with different molecular weights was prepared by free radical polymerization in aqueous solution, recorded as PSPE-1, PSPE-2 and PSPE-3. The structures were characterized by  $^1\text{H-NMR}$  (400 MHz,  $\text{D}_2\text{O}$ ), and the result was as follow (PSPE-3), a,  $\delta$ 2.42 ppm, b,  $\delta$ 1.20 ppm, c,  $\delta$ 4.71 ppm, d,  $\delta$ 4.04 ppm, e,  $\delta$ 3.82 ppm, f,  $\delta$ 2.48 ppm, g,  $\delta$ 3.21 ppm, h,  $\delta$ 3.46 ppm.

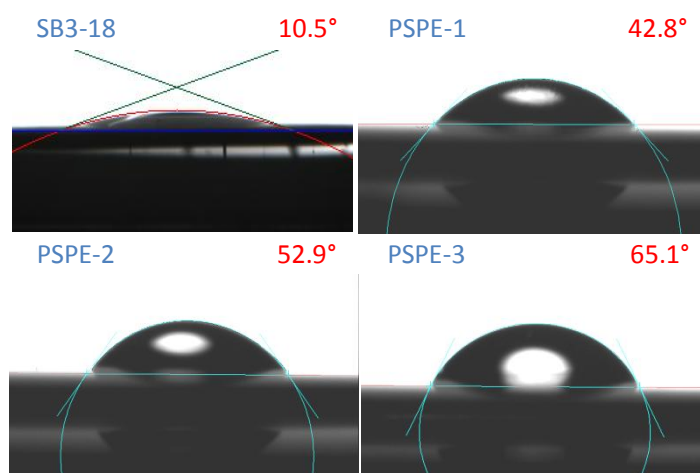
## UCST-responsive performances of PSPE





**Fig. S2.** UCST-responsive performances of PSPE in the acetate buffer solution ( pH 5.0, 50 mM ), (a) PSPE-1 (b) PSPE-2 (c) peak thickness of SB3-18 and PSPE.

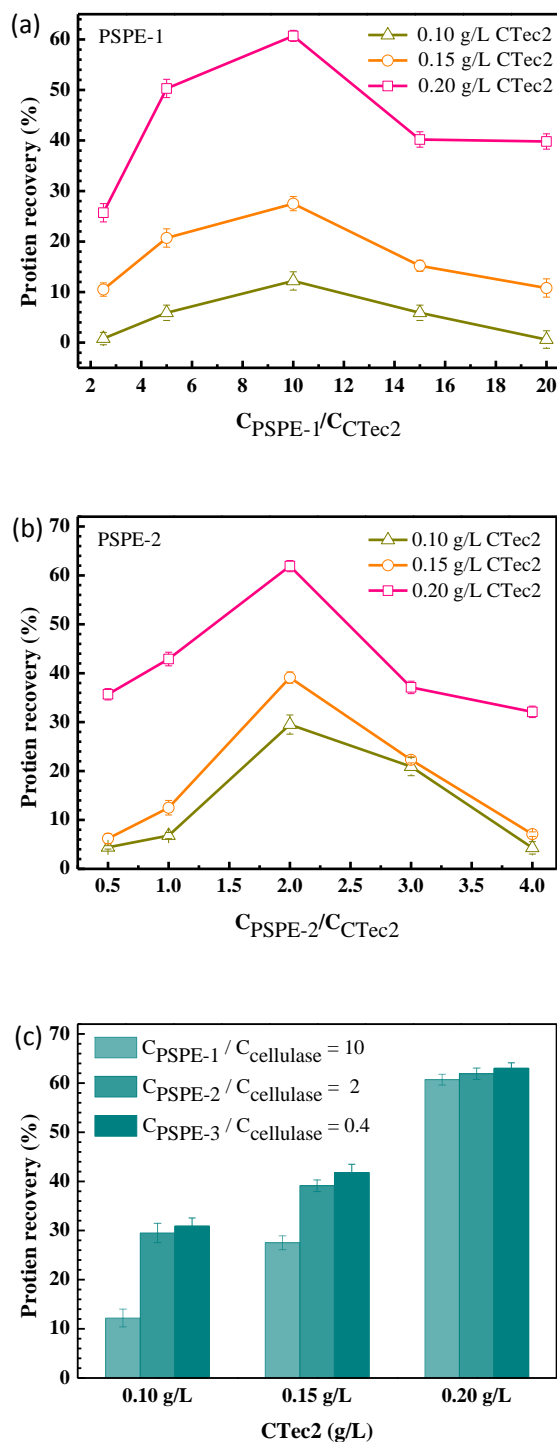
### Contact angles measurement



**Fig. S3.** The contact angles of water on SB3-18 and PSPE films.

The contact angles of distilled water on SB3-18 and PSPE-x (x = 1, 2, 3) films were measured according to the sessile drop method by using contact angle measuring instrument (OCA40Micro, Data physics, Germany).

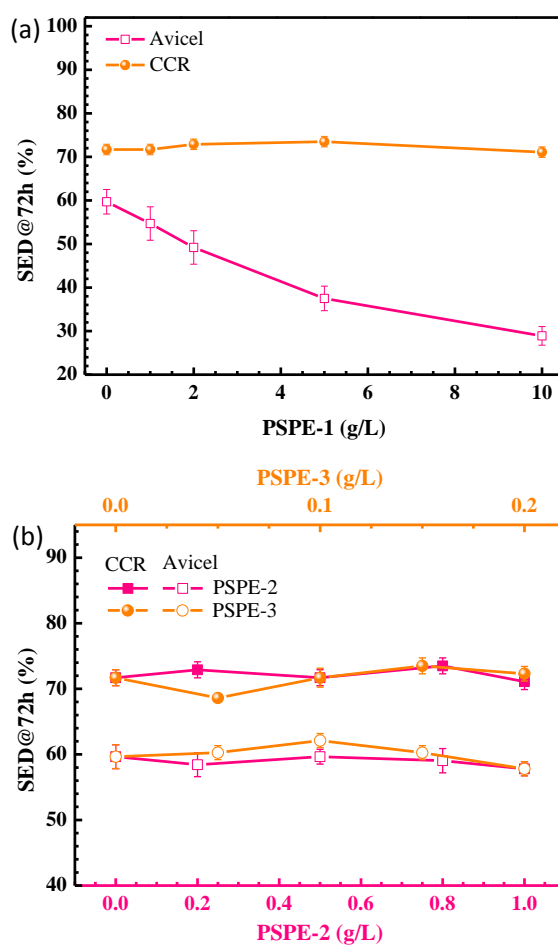
### Recovering cellulase at room temperature

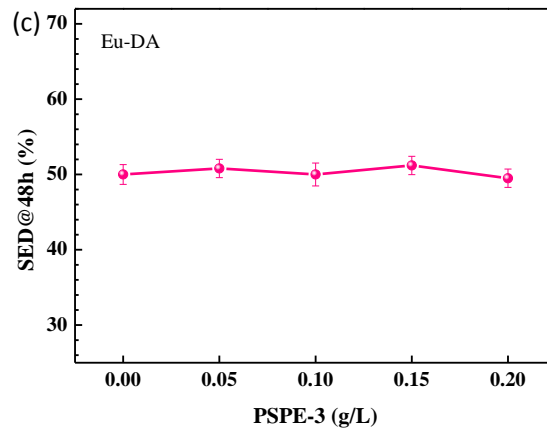


**Fig. S4.** The effect of concentration ratio upon the recovery of protein in cellulase in buffer solution (a) PSPE-1 (b) PSPE-2, (c) the effect of the molecular weight of PSPE and the concentration of cellulase upon the recovery of protein in cellulase in buffer solution.

## The effect PSPE on the enzymatic hydrolysis efficiency of Avicel and lignocelluloses

The conditions of enzymatic hydrolysis were as follows. 0.6 g Avicel or lignocelluloses, PSPE-x ( $x = 1, 2, 3$ ) and cellulase were added to the mixture of 30 mL buffer (pH 5.0, 50 mM). The enzyme load for Avicel, CCR and Eu-DA were 5, 10, 10 FPU/g glucan, respectively. The enzymatic hydrolysis of the substrate was carried out under the conditions of 2% (w/v) solid concentration at 50 °C at 150 rpm for 48 or 72 h in a shaker (DDHZ-300, Jiangsu Taicang Equipment Factory, China). Aliquots of 0.1 mL were obtained at 48 or 72 h for glucose analysis. The concentration of glucose was monitored by SBA-40E with its own  $H_2O_2$  electrode sensor (Institute of Biology of the Shandong Academy of Sciences, China), and the enzymatic hydrolysis efficiency of lignocelluloses (SED@h) was expressed by the yield of glucose. In addition, the blank experiment without PSPE-x was carried out for comparison, all experimental datas were the average of three parallel experiments, the data deviations were shown in the figures.

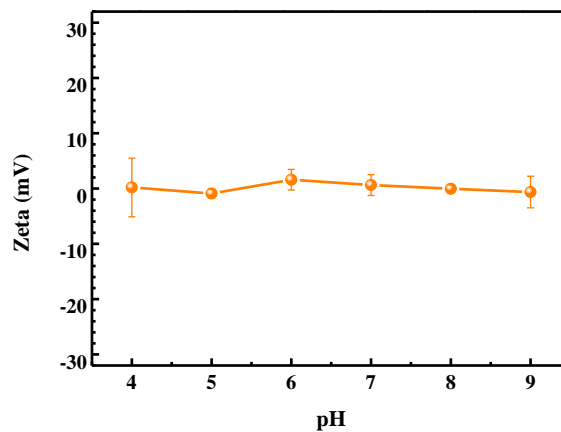




**Fig. S5.** (a) Effect of PSPE-1 on the enzymatic hydrolysis efficiency of Avicel and CCR, (b) effect of PSPE-2 and PSPE-3 on the enzymatic hydrolysis efficiency of Avicel and CCR, (c) effect of PSPE-3 on the enzymatic hydrolysis efficiency of Eu-DA. (solid concentration, 2% (w/v), pH 5.0, ionic strength, 50 mM, cellulase loading, 5 FPU/g glucan for Avicel, 10 FPU/g glucan for CCR, 10 FPU/g glucan for Eu-DA).

### The zeta potentials of PSPE

A zeta potential analyzer (Brookhaven Zeta Plus, Brookhaven, USA) was used to determine the zeta potentials of PSPE solution (1M NaCl).



**Fig. S6.** The zeta potentials of PSPE-2 in NaCl solution (1 M).