

Supporting information for

DES-N-Doped Oxygenated Carbon Dots Colloidal Solutions for Light Harvesting and Bio-imaging Applications

Krishnaiah Damarla,^{†,§} Sanjay Mehra,^{†,§} Tejwant Singh Kang,[‡] Sonam Yadav,^{†,§} Avinash Mishra^{†,§} and Arvind Kumar^{†,§,}*

[†]CSIR-Central Salt and Marine Chemicals Research Institute, G. B. Marg Bhavnagar-364002, Gujarat, India.

[§]Academy of Scientific and Innovative Research (AcSIR), Ghaziabad-201002, India

[‡]Department of Chemistry, UGC-centre for Advance Studies – II, Guru Nanak Dev University, Amritsar, 143005, India.

Corresponding Author e-mail: mailme_arvind@yahoo.com ; arvind@csmcri.res.in Tel.: +91-278-2567039. Fax: +91-278-2567562.

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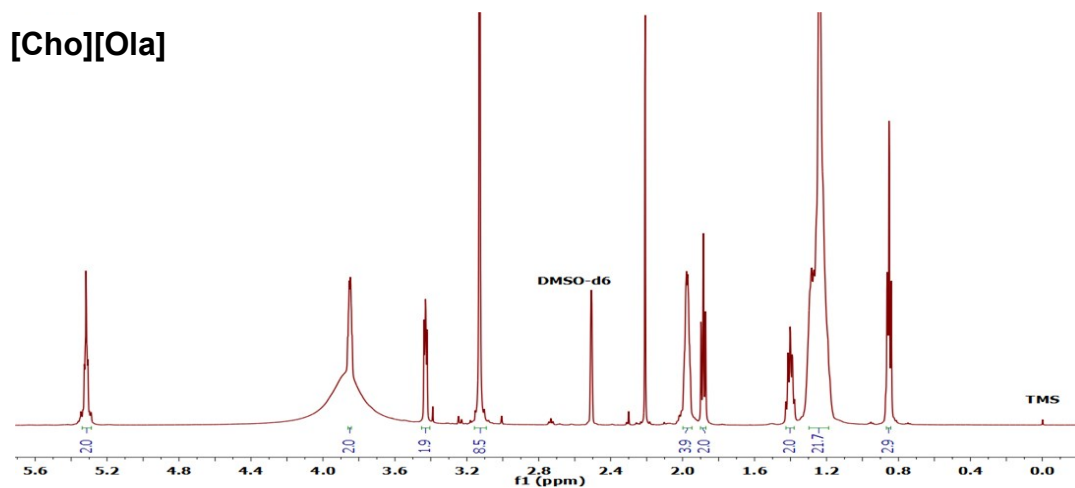


Fig. S1 ^1H NMR spectra of [Cho][Ola].

Choline Oleate, [Cho][Ola]: ^1H -NMR; 600MHz, DMSO-d6, δ_{H} 0.9 (t, 3H), 1.3 (m, 20H), 1.4 (q, 2H), 1.9 (t, 2H), 2.0 (q, 4H), 3.1 (s, 9H), 3.4 (t, 2H), 3.9 (m, 3H), 5.3 (d, 2H); ESI-MS: $[\text{C}_5\text{H}_{14}\text{NO}]^+$ m/z:104.10, $[\text{C}_{18}\text{H}_{33}\text{O}_2]^-$ m/z = 285.6.

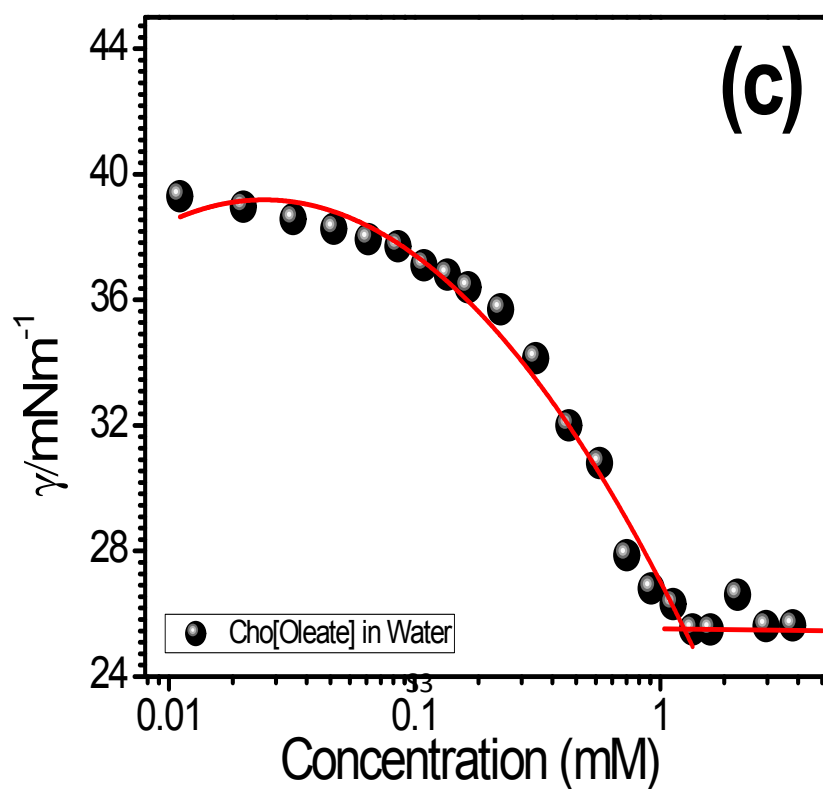


Fig. S2 Plot shows the Surface tension of [Cho][Ola] in aqueous system.

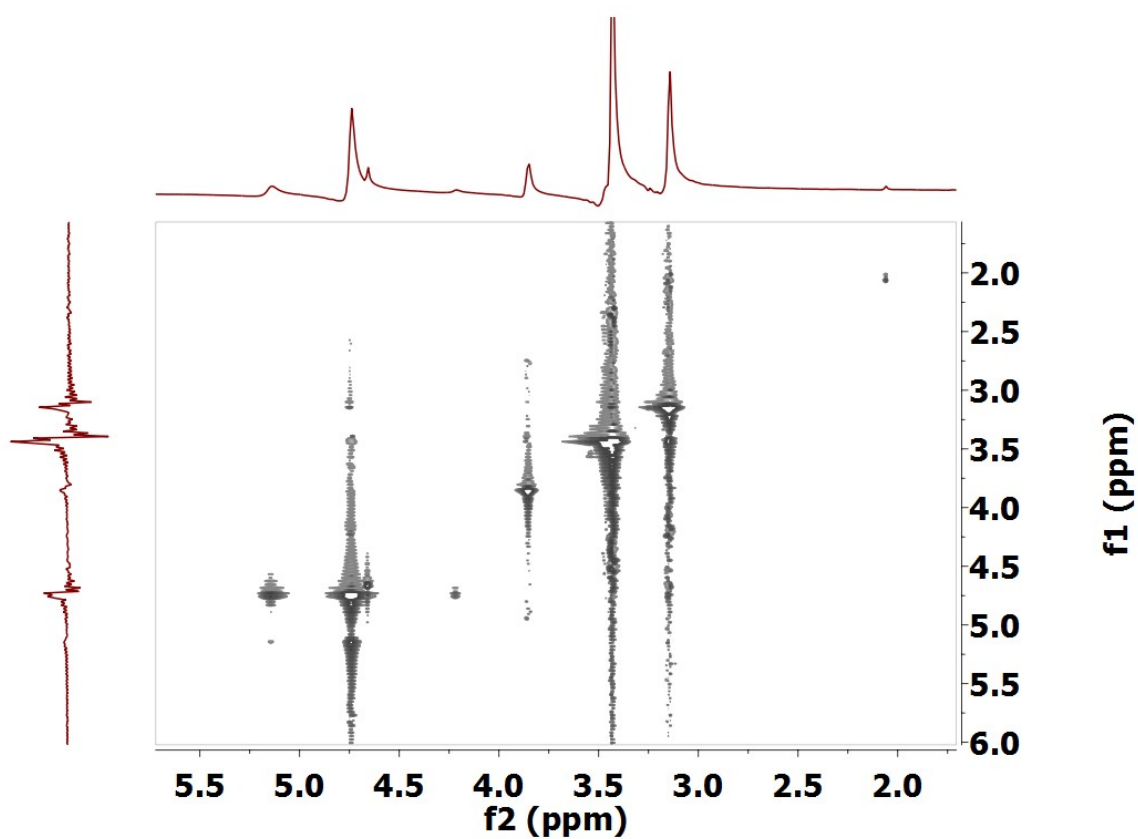


Fig. S3 2D NMR NOESY spectra spectra of [Cho][Ola] in choline chloride-ethylene glycol (1:2 mole ratio) deep eutectic solvent (DES).

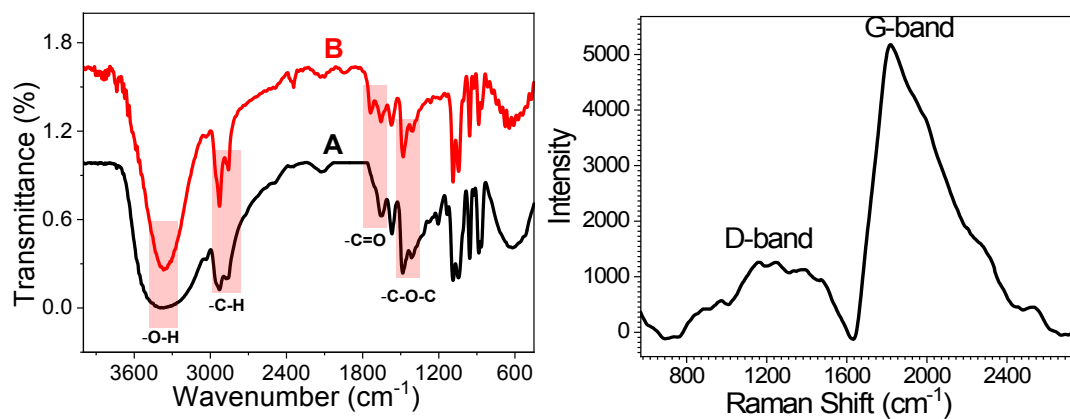


Fig. S4 (a) FT-IR spectra of (A) [Cho][Ola]-DES colloidal solutions and (B) N doped CDs-[Cho][Ola]-DES colloidal solutions, and (b) Raman spectra of N doped CDs-[Cho][Ola]-DES colloidal solutions.

Fig. S5 Fluorescence spectra, of N doped CDs-[Cho][Ola]-DES colloidal solution.

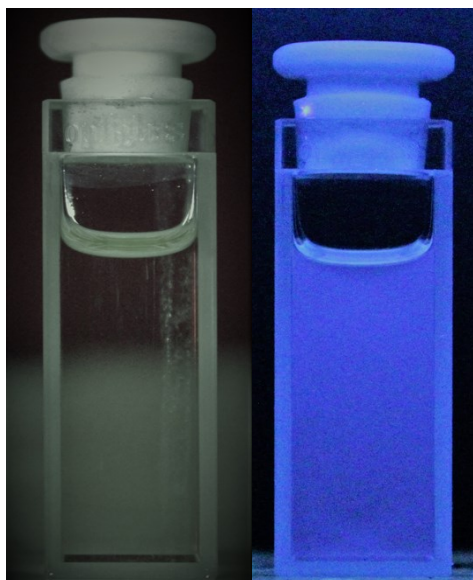


Fig. S6 Photographs of blue emission from N doped CDs dispersed in toluene.

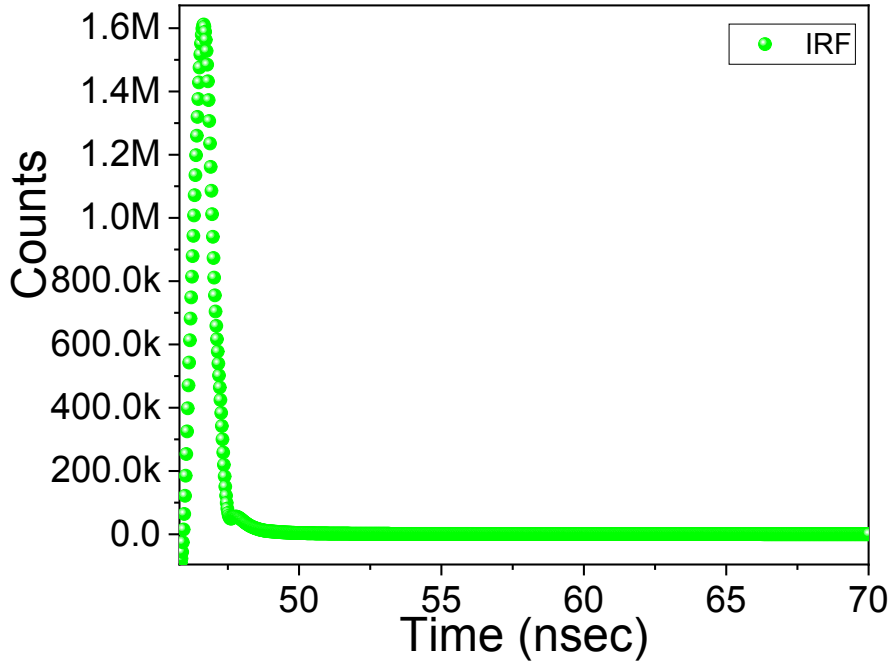


Fig. S7 TrPL life time IRF spectra.

Quantum efficiency calculations: Quantum yield (QY) of N doped CDs-[Cho][Ola] -DES colloidal solutions was calculated using Fluorescein dye as a standard having, QY = 0.79 (79%). We prepared 0.1M NaOH solution of dye to measure the QY and it was calculated using following equation:

$$\phi = \phi_R \times \frac{I}{I_R} \times \frac{A_R}{A} \times \frac{\eta^2}{\eta_R^2}$$

Where,

ϕ, ϕ_R = Quantum yield of sample and reference respectively,

I, I_R = Emission of the sample and reference respectively,

A, A_R = optical density of the sample and reference respectively,

η^2, η_R^2 = refractive index of the sample and reference respectively.

Table S1. Comparison of quantum yields of CDs vis. a vis. method of preparation.

S.No.	Type of material	Synthetic route	Quantum yield	Application	Reference/s
1	DES-N-CDs micelles	100°C	0.82	Light harvesting and bioimaging	Present study
2	N-CDs	Microwave @ 160 °C	0.93	pH response	1
3	N-CDs	Autoclave at 180 °C	0.47	Light harvesting	2
4	N-CDs	autoclave @ 180 °C	0.31	bioimaging	3
5	N-CDs	Autoclave at 200 °C	0.16	bioimaging	4
6	N-CDs	Microwave @ 180 °C	0.14	bioimaging	5
7	Co-doped CDs	Autoclave at 200 °C	0.73	Light harvesting	6
8	S-doped CDs	240 °C	0.67	Fe(III) ion detection	7
9	Silica functionalised CDs	240 °C	0.47	bioimaging	8
10	Polyamine doped CDs	< 200 °C	0.42	chemical sensing	9

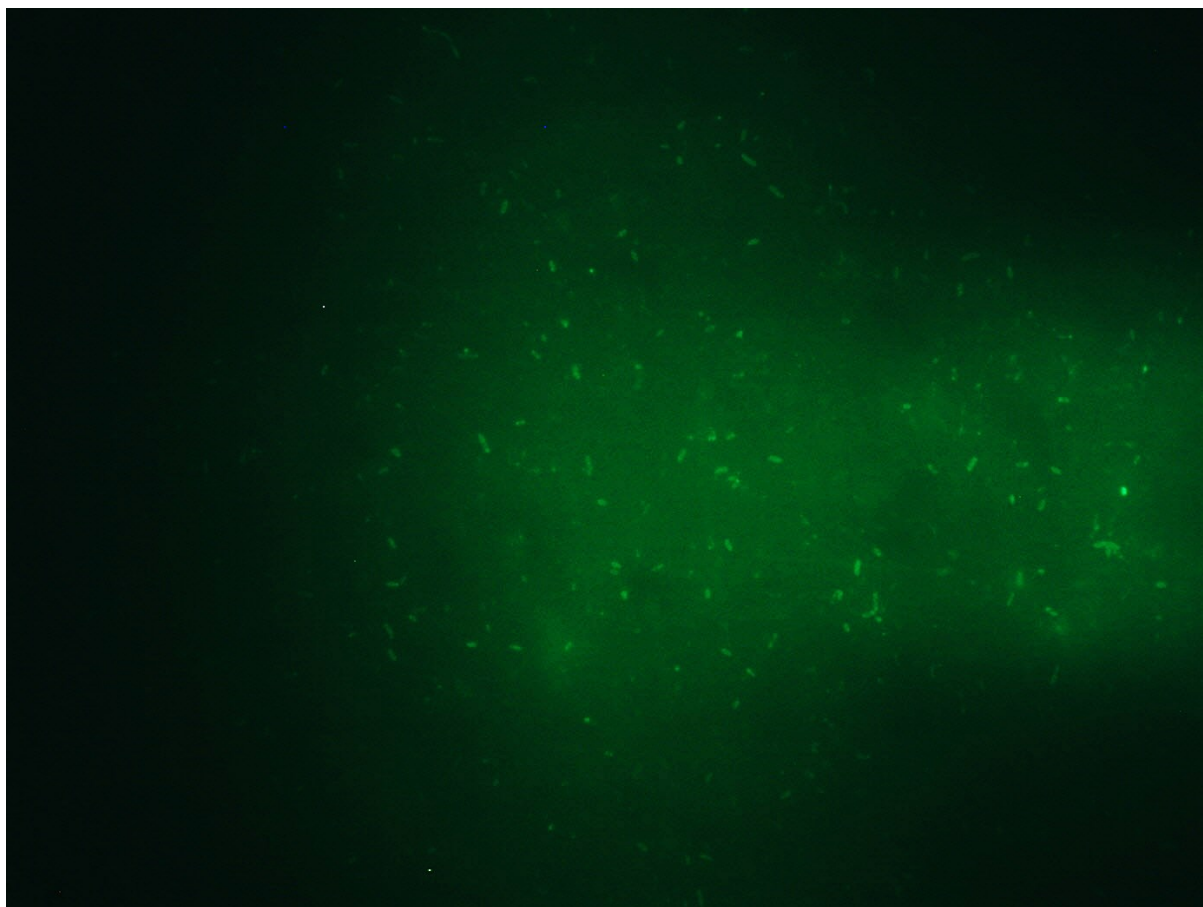


Fig. S8 Fluorescence microscopic images of *Escherichia coli* with the N-doped carbon dots under and through green filter.

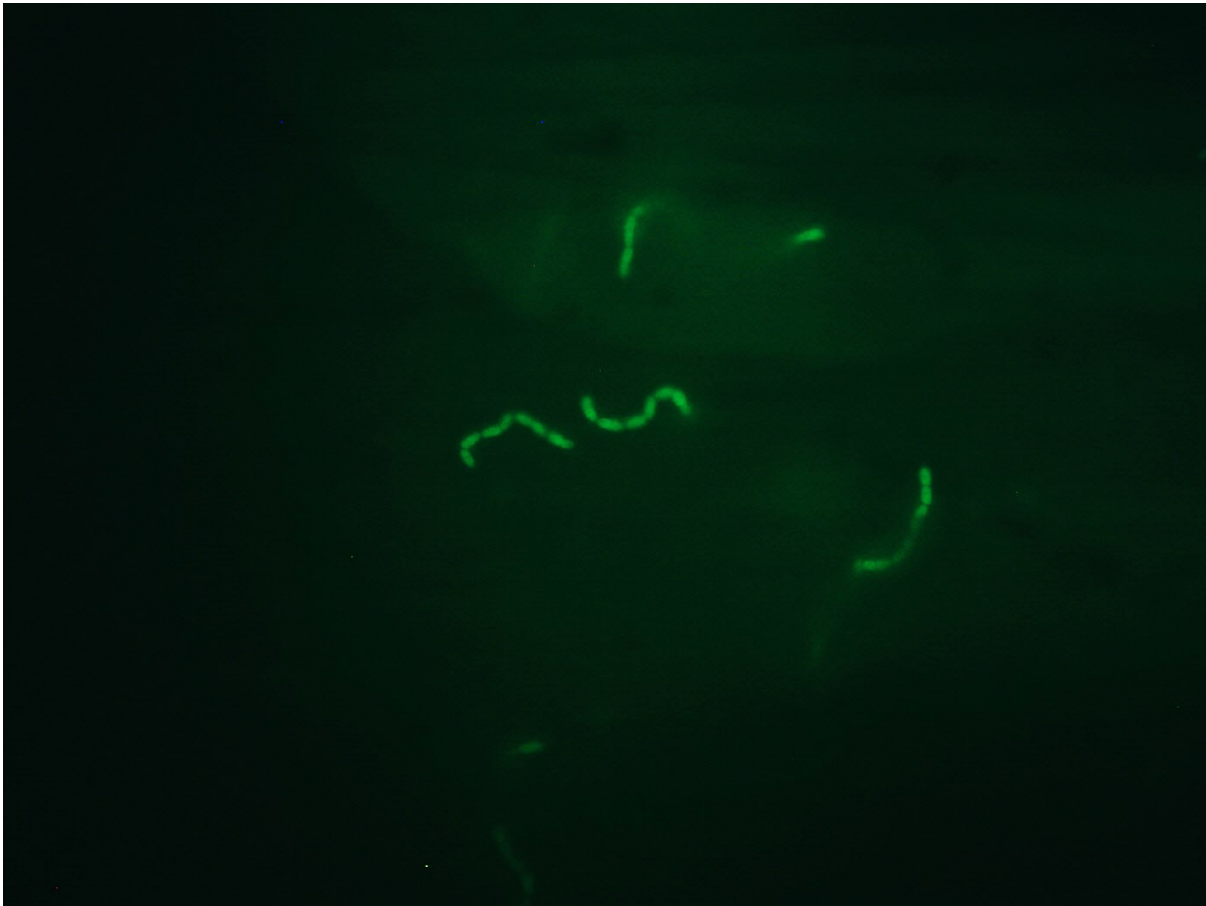


Fig. S9 Fluorescence microscopic images of *Bacillus cereus*, labeled with the N-doped carbon dots under and through green filter.

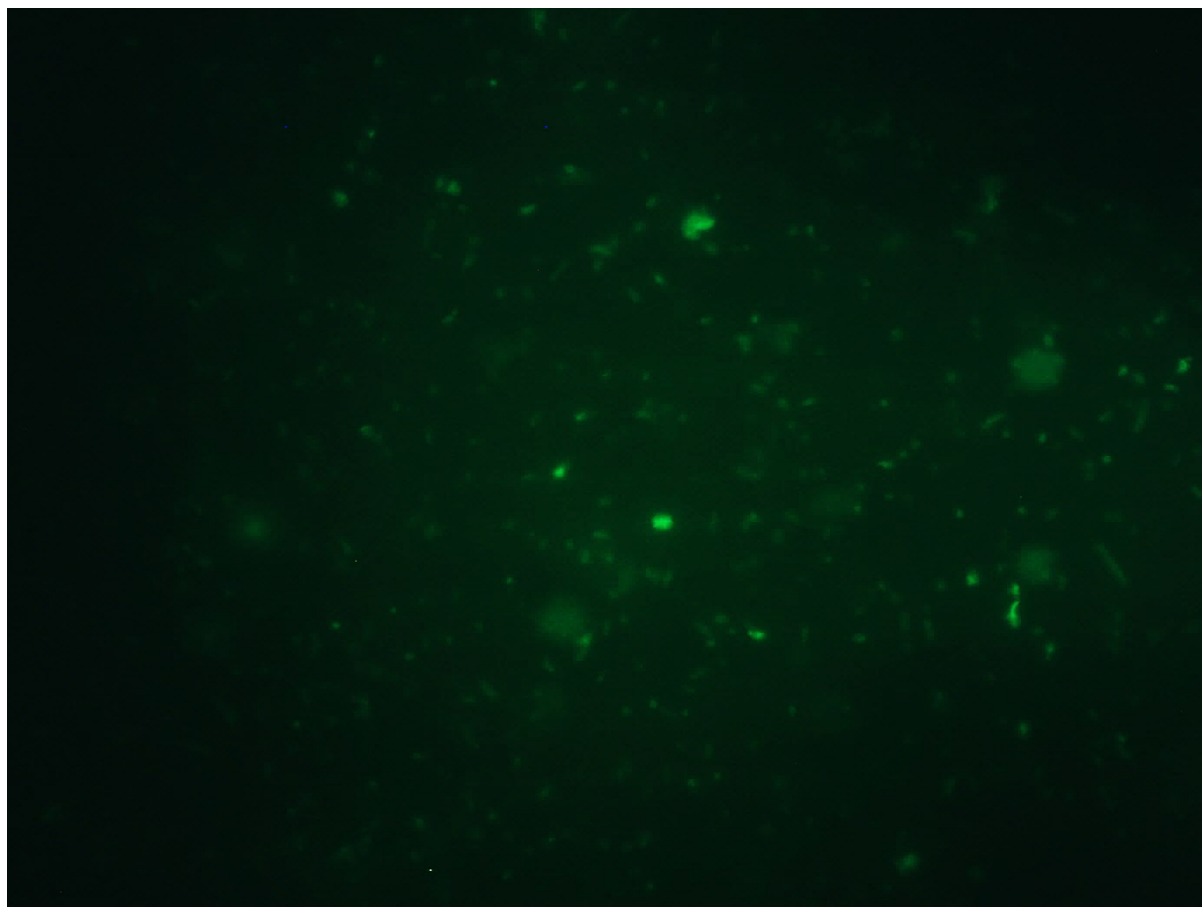


Fig. S10 Fluorescence microscopic images of *Vibrio owensii*, labeled with the N-doped carbon dots under and through green filter.

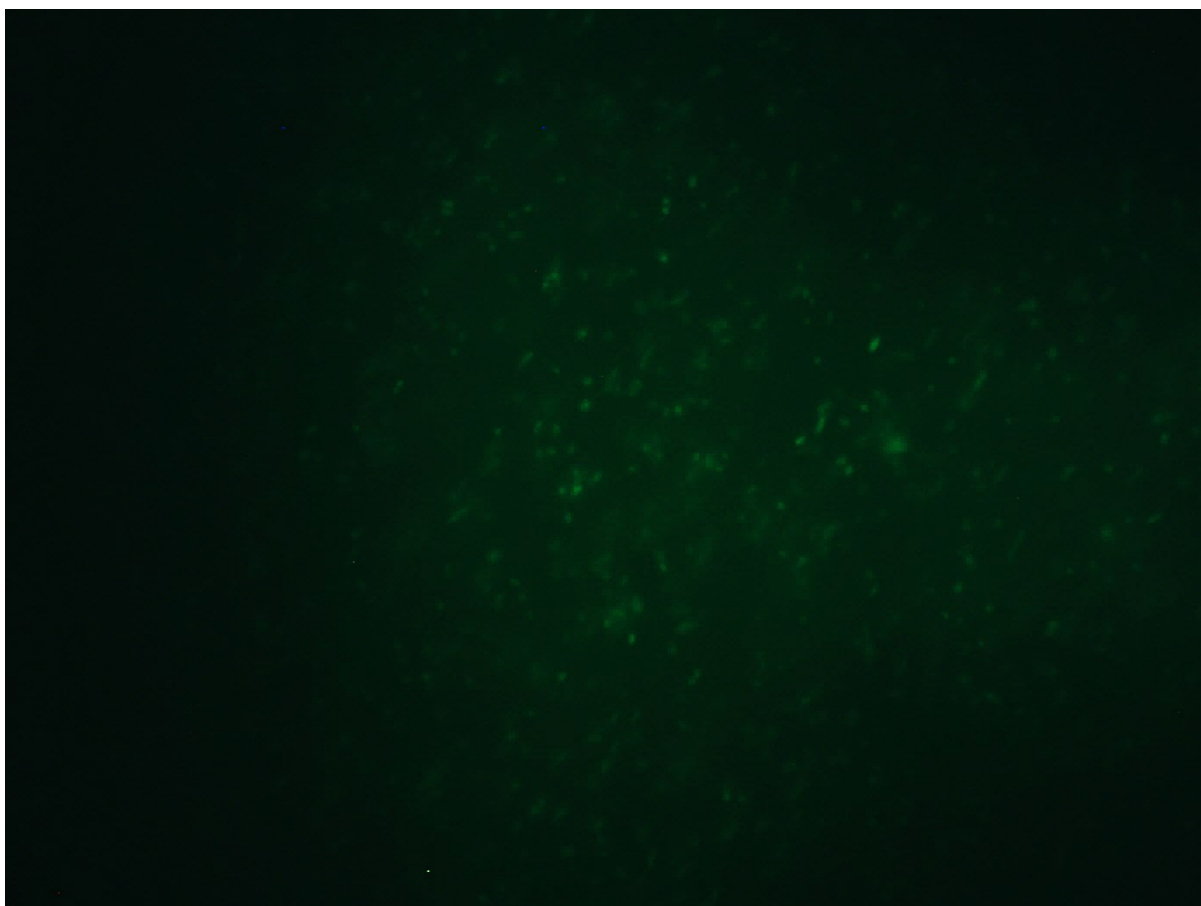


Fig. S11 Fluorescence microscopic images of *Vibrio alginolyticus* of OD₆₀₀ 0.6, labeled with the N-doped carbon dots under and through green filter.

References:

1. C. Zheng, X. An and J. Gong, *RSC Adv.*, 2015, **5**, 32319–32322.
2. M. Li, C. Yu, C. Hu, W. Yang, C. Zhao, S. Wang, M. Zhang, J. Zhao, X. Wang, J. Qiu, *Chem. Eng. J.*, 2017, **320**, 570-575.
3. H. Zhang, Y. Chen, M. Liang, L. Xu, S. Qi, H. Chen, and X. Chen, *Anal. Chem.* 2014, **86**, 19, 9846–9852.
4. Y. Song, C. Zhu, J. Song, H. Li, D. Du, and Y. Lin, *ACS Appl. Mater. Interfaces* 2017, **9**, 8, 7399–7405

5. T. N. J. I. Edisona, R. Atchudana, M. G. Sethuraman, J. -J. Shima and Y. R. Leea, *J. Photochem. Photobiol. B*, 2016, **161**, 154-161.
6. Y. Dong, H. Pang, H. B. Yang, C. Guo, J. Shao, Y. Chi, C. M. Li and T. Yu, *Angew. Chem. Int. Ed.* 2013, **52(30)**, 7800–7804.
7. Q. Liang, W. Ma, Y. Shi, Z. Li, X. Yang, *Carbon* 2013, **60**, 421–428.
8. M. J. Krysmann, A. Kelarakis, P. Dallas, E. P. Giannelis, *J. Am. Chem.Soc.* 2012, **134(2)**, 747–750.
9. X. Li, S. Zhang, S. A. Kulinich, Y. Liu, H. Zeng, *Sci Rep.* 2014, **4(1)**, 4976.