

Electronic Supplementary Information (ESI)
for

Ligand and Cu free *N*-arylation of indoles, pyrroles and benzylamines with aryl halides catalyzed by a Pd nanocatalyst

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Experimental

General Procedure

The starting materials and all solvents used were purchased from commercial sources. Anhydrous solvents were prepared using standard methods. TLC was performed on pre-coated aluminium plates of silica gel 60 F₂₅₄. TLC spots were visualized by UV light (254 nm). Photolysis was carried out in an immersion well cooled by chilled water using medium pressure Hg vapour lamp. ¹H and ¹³C-NMR spectra were recorded on Bruker 400 MHz NMR in solution of CDCl₃.

General experimental procedure for *N*-arylation

In a 25 mL round bottom flask fitted with a magnetic stirrer bar 1 mmol indole/pyrrole/benzyl amine, 1.1 mmol aryl halides and 2 mmol K₂CO₃ were added and after that in it 10 mL of 1:1 DMSO/DMF was added. Then in the reaction mixture Pd-NP (0.003%) catalyst was added and the whole system was then degassed for 10 minutes with Ar. The reaction mixture was then heated under argon atmosphere at 130 °C for 24 h. The completion of the reaction was monitored by TLC. After completion of the reaction, the reaction mixture was cooled, washed with water and the organic part extracted with ethyl acetate. Then the organic layer dried over anhydrous Na₂SO₄ and concentrated in *vacuo*. The crude product was purified by column chromatography using ethyl acetate and petroleum benzene as eluent to afford the desired product which was then characterized by standard spectroscopic techniques.

Three Phase Test:

To a 50 mL of round bottomed flask charged with magnetic stirrer bar, Wang Resin (0.625g) was added followed by the addition of anhydrous DCM (7.5 mL). Then the mixture was left to stand for 20 min. to allow it to swell, after which 4-iodobenzoic acid (0.697 g, 2.81 mmol), DCC (0.580 g, 2.81 mmol) and DMAP (0.069 g, 0.565 mmol) was added. The round bottom flask was capped with rubber septum and stirred for 4 days. After TLC indicated the complete disappearance of 4-iodobenzoic acid. Then the solid material was filtered and washed successively three times each with 5 mL of DMF, THF, MeOH, and finally with the CH₂Cl₂. The resin bound material was then dried *in vacuo* for 24 hours. The resin bound acid derivative was then characterized by Solid state NMR spectroscopy.

Compound Characterization

1-*p*-tolyl-1*H*-indole (3a)¹: ¹H NMR (400 MHz, CDCl₃) δ 7.68 (d, *J* = 7.6 Hz, 1H), 7.52 (d, *J* = 8.2 Hz, 1H), 7.38 (d, *J* = 8.3 Hz, 2H), 7.30 (t, *J* = 3.2 Hz, 3H), 7.13-7.23 (m, 2H), 6.66 (d, *J* = 3.2 Hz, 1H), 2.42 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 135.9, 130.1, 129.1, 128.1, 122.2, 121.0, 120.2, 110.5, 103.2, 21.0.

1-phenyl-1*H*-indole (3b)²: ¹H NMR (400 MHz, CDCl₃) δ 8.28 (d, *J* = 8.7 Hz, 2H), 7.84 (q, *J* = 8.7 Hz, 3H), 7.64-7.71 (m, 3H), 7.40 (d, *J* = 3.3 Hz, 1H), 7.20-7.24 (m, 1H), 6.75 (d, *J* = 4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 137.5, 130.3, 129.7, 128.0, 127.5, 126.5, 124.4, 122.4, 121.2, 120.4, 110.6, 103.6, 94.5.

1-(4-methoxyphenyl)-1*H*-indole (3c)³: ¹H NMR (400 MHz, CDCl₃) δ 7.70 (d, *J* = 8.1 Hz, 1H), 7.47 (d, *J* = 8.8 Hz, 1H), 7.42 (d, *J* = 9 Hz, 1H), 7.29 (d, *J* = 3.2 Hz, 1H), 7.15-7.24 (m, 2H), 7.04 (d, *J* = 8.9 Hz, 2H), 6.67 (d, *J* = 3.9 Hz, 1H), 3.89 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 158.3, 136.3, 132.9, 129.0, 128.3, 126.0, 122.2, 121.0, 120.1, 114.7, 110.4, 102.9, 55.6.

Methyl-4(1*H*-indol-1-yl) benzoate (3d)³: ¹H NMR (400 MHz, CDCl₃) δ 8.21 (d, *J* = 8.8 Hz, 2H), 7.59-7.72 (m, 4H), 7.38 (d, *J* = 3.2 Hz, 1H), 7.20-7.30 (m, 2H), 6.74 (d, *J* = 3.2 Hz, 1H), 3.97 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.4, 143.7, 135.4, 131.2, 129.8, 127.5, 127.4, 123.2, 122.8, 121.4, 120.9, 110.6, 104.9, 52.3.

5-methoxy-1-*p*-tolyl-1*H*-indole (3e)¹: ¹H NMR (400 MHz, CDCl₃) δ 7.46 (d, *J* = 8.8 Hz, 1H), 7.40 (d, *J* = 8 Hz, 2H), 7.33 (s, 1H), 7.31 (d, *J* = 2.8 Hz, 2H), 7.17 (d, *J* = 2.4 Hz, 1H), 6.90 (dd, *J* = 2.4 Hz and *J* = 9.2 Hz, 1H), 6.61 (d, *J* = 3.2 Hz, 1H), 3.90 (s, 3H), 2.45 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 154.5, 137.4, 136.1, 130.1, 129.7, 128.5, 124.0, 112.4, 111.3, 102.9, 102.7, 55.9, 21.0.

5-cyano-1-*p*-tolyl-1*H*-indole (3f)¹: ¹H NMR (400 MHz, CDCl₃) δ 8.02 (d, *J* = 1.2 Hz, 1H), 7.51 (d, *J* = 8.4 Hz, 1H), 7.42 (dd, *J* = 3.6 Hz and *J* = 7 Hz, 2H), 7.34 (s, 4H), 6.73 (d, *J* = 3.2 Hz, 1H), 2.45 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 137.6, 136.1, 130.5, 130.4, 128.7, 126.6, 125.0, 124.5, 120.6, 103.8, 103.2, 21.0.

5-bromo-1-*p*-tolyl-1*H*-indole (3g)⁴: ¹H NMR (400 MHz, CDCl₃) δ 7.85 (d, *J* = 2 Hz, 1H), 7.38-7.44 (m, 2H), 7.36 (d, *J* = 4 Hz, 3H), 7.33 (d, *J* = 2.8 Hz, 2H), 6.63 (d, *J* = 3.2 Hz, 1H), 2.8 (s,

3H); ^{13}C NMR (100 MHz, CDCl_3) δ 136.8, 134.8, 130.9, 130.3, 129.3, 125.1, 124.3, 123.5, 113.4, 112.0, 102.7, 21.1.

methyl 4-(5-methoxy-1*H*-indol-1-yl)benzoate (3h)³: ^1H NMR (400 MHz, CDCl_3) δ 8.19 (d, J = 8.4 Hz, 2H), 7.54-7.57 (m, 3H), 7.35 (d, J = 3.2 Hz, 1H), 7.16 (d, J = 2.4 Hz, 1H), 6.93 (dd, J = 2.8 Hz and J = 9 Hz, 1H), 6.66 (d, J = 3.2 Hz, 1H), 3.97 (s, 3H), 3.89 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 166.4, 154.9, 143.8, 131.3, 130.5, 127.8, 127.3, 112.8, 111.5, 104.7, 103.1, 55.8, 52.2.

methyl 4-(5-cyano-1*H*-indol-1-yl)benzoate (3i)³: ^1H NMR (400 MHz, CDCl_3) δ 8.24 (d, J = 8.4 Hz, 2H), 8.05 (s, 1H), 7.63 (d, J = 8.4 Hz, 1H), 7.57 (d, J = 8.4 Hz, 2H), 7.49 (d, J = 8 Hz, 2H), 6.98 (d, J = 2.4 Hz, 1H), 3.98 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 166.1, 142.5, 137.1, 131.5, 129.9, 129.4, 128.9, 126.8, 125.7, 123.9, 120.2, 111.4, 105.2, 104.1, 52.4.

1-(4-(1*H*-indol-1-yl)phenyl)ethanone (3j)²: ^1H NMR (400 MHz, CDCl_3) δ 8.13 (d, J = 8.8 Hz, 2H), 7.62-7.71 (m, 4H), 7.39 (d, J = 3.2 Hz, 1H), 7.21-7.29 (m, 2H), 6.74 (d, J = 3.2 Hz, 1H), 2.66 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 196.9, 143.8, 135.4, 134.6, 130.1, 129.8, 127.4, 123.3, 122.9, 121.4, 121.1, 110.6, 105.1, 26.6.

1-phenyl-1*H*-pyrrole (5a)⁵: ^1H NMR (400 MHz, CDCl_3) δ 7.44-7.49 (m, 4H), 7.29 (t, J = 6.4 Hz, 1H), 7.15 (s, 2H), 6.42 (s, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 140.8, 129.6, 125.6, 120.6, 119.4, 110.5.

1-*p*-tolyl-1*H*-pyrrole (5b)⁶: ^1H NMR (400 MHz, CDCl_3) δ 7.35 (d, J = 8.4 Hz, 2H), 7.28 (d, J = 8 Hz, 2H), 7.14 (t, J = 2 Hz, 2H), 6.42 (t, J = 2 Hz, 2H), 2.44 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 138.6, 135.4, 130.1, 120.6, 119.4, 110.1, 20.1.

1-(4-methoxyphenyl)-1*H*-pyrrole (5c)⁵: ^1H NMR (400 MHz, CDCl_3) δ 7.27-7.31 (m, 2H), 6.98 (t, J = 2.2 Hz, 2H), 6.90-6.95 (m, 2H), 6.31 (t, J = 2.2 Hz, 2H), 3.81 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 157.7, 134.7, 122.2, 119.7, 114.7, 109.9, 55.6.

methyl 4-(1*H*-pyrrol-1-yl)benzoate (5d)⁷: ^1H NMR (400 MHz, CDCl_3) δ 8.11 (d, J = 8.4 Hz, 2H), 7.45 (d, J = 8.8 Hz, 2H), 7.17 (t, J = 2 Hz, 2H), 6.40 (t, J = 2.2 Hz, 2H), 3.94 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 166.4, 144.0, 131.3, 126.9, 119.3, 119.0, 111.5, 52.1.

1-(4-(1*H*-pyrrol-1-yl)phenyl)ethanone (5e)⁵: ¹H NMR (400 MHz, CDCl₃) δ 8.03 (d, *J* = 8.8 Hz, 2H), 7.47 (d, *J* = 8.8 Hz, 2H), 7.17 (t, *J* = 1.8 Hz, 2H), 6.39 (t, *J* = 1.8 Hz, 2H), 2.61 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 196.8, 144.0, 134.0, 130.2, 119.3, 119.0, 111.6, 26.5.

***N*-benzylbenzenamine (7a)**⁸: ¹H NMR (400 MHz, CDCl₃) δ 7.42-7.45 (m, 4H), 7.38 (d, *J* = 6 Hz, 1H), 7.28 (t, *J* = 7.8 Hz, 2H), 6.83 (t, *J* = 7.6 Hz, 1H), 6.72 (d, *J* = 7.6 Hz, 2H), 4.39 (s, 2H), 4.06 (bs, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 148.0, 139.3, 129.2, 128.5, 127.4, 127.1, 117.5, 112.8, 48.2.

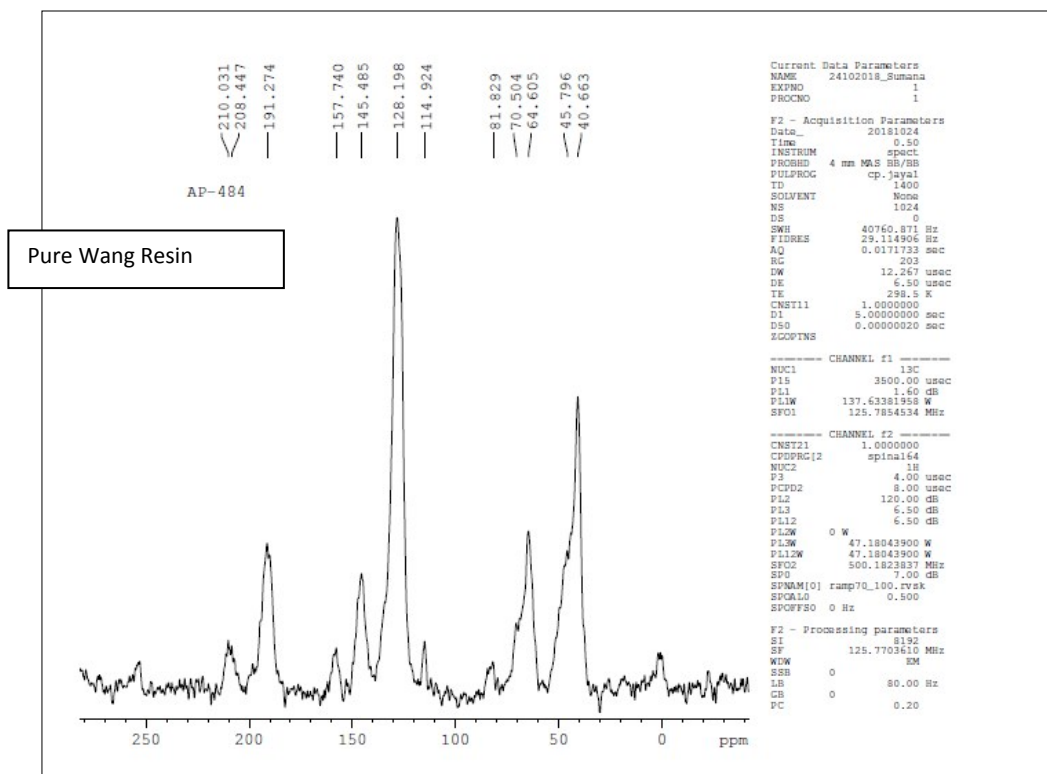
***N*-benzyl-4-methylbenzenamine (7b)**⁸: ¹H NMR (400 MHz, CDCl₃) δ 7.28-7.38 (m, 6H), 7.0 (t, *J* = 10 Hz, 2H), 6.58 (dd, *J* = 8 Hz and *J* = 12 Hz, 2H), 4.33 (s, 2H), 2.27 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 145.7, 139.5, 129.6, 128.5, 127.4, 127.0, 126.6, 112.9, 48.5, 20.3.

***N*-benzyl-4-methoxybenzenamine (7c)**⁸: ¹H NMR (400 MHz, CDCl₃) δ 7.26-7.40 (m, 5H), 6.78-6.82 (m, 2H), 6.62-6.66 (m, 2H), 4.30 (s, 2H), 3.76 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 152.4, 142.0, 139.4, 128.6, 127.6, 127.2, 114.9, 114.4, 55.8, 49.4.

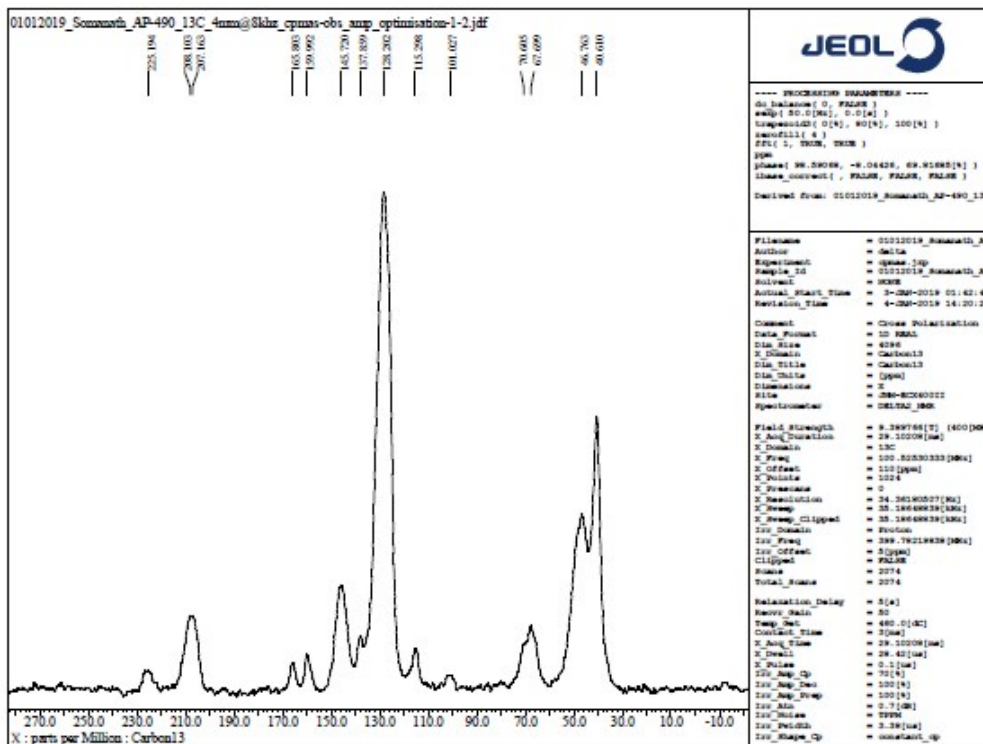
methyl 4-(benzylamino)benzoate (7d)⁹: ¹H NMR (400 MHz, CDCl₃) δ 7.78 (d, *J* = 8 Hz, 2H), 7.17-7.29 (m, 6H), 6.51 (d, *J* = 8 Hz, 2H), 4.61 (s, 1H), 4.29 (s, 2H), 3.75 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 167.3, 151.5, 138.2, 131.5, 128.7, 127.5, 127.4, 126.9, 118.6, 111.8, 51.5, 47.7.

References:

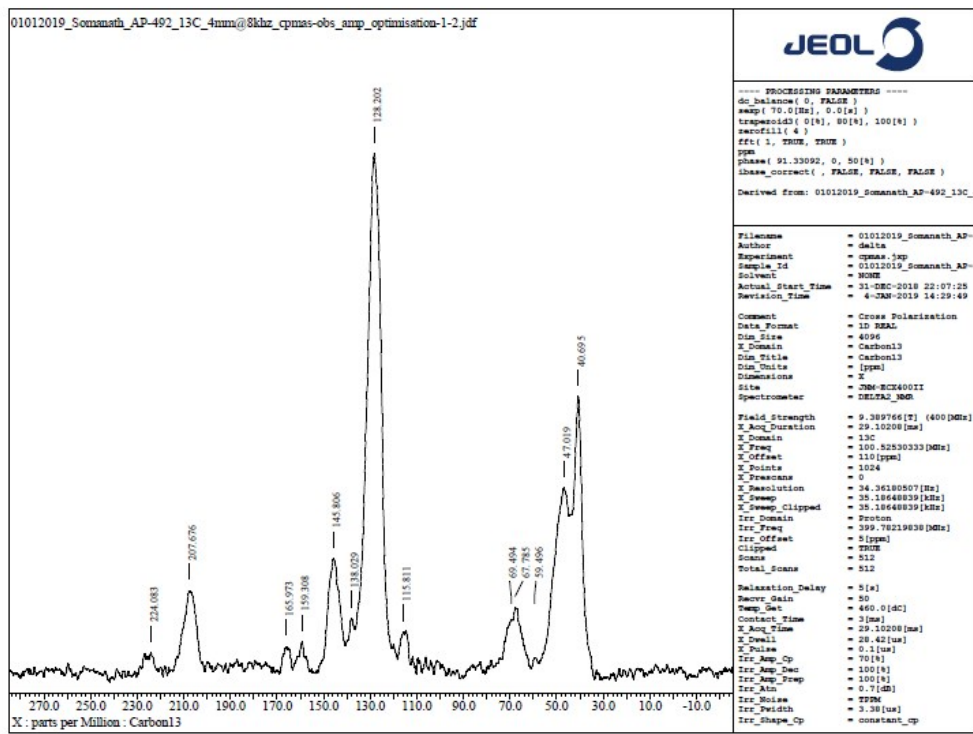
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Solid State ¹³C-NMR spectra of pure Wang Resin.



Solid State ¹³C-NMR spectra of Wang Resin anchored 4-iodobenzoic acid.



Solid State ^{13}C -NMR spectra of Wang Resin anchored 4-iodobenzoic acid after *N*-arylation.

NMR Spectra

