Electronic Supplementary Material (ESI) for New Journal of Chemistry. This journal is © The Royal Society of Chemistry and the Centre National de la Recherche Scientifique 2020

New Journal of Chemistry

Electronic Supplementary Information

Thermodynamics of the inclusion within cyclodextrins and structural evidences of Cu(Indomethacin)₂ and Zn(Indomethacin)₂ complexes in aqueous solution.

Adrián Ricardo Hipólito-Nájera^a, Norma Rodríguez-Laguna^a, Luis I. Reyes-García^a, Alberto Rojas-Hernández^b, Rodolfo Gómez-Balderas^{*a} and Rosario Moya-Hernández^{*a}

^a Laboratorio de Fisicoquímica Analítica, Unidad de Investigación Multidisciplinaria, Facultad de Estudios Superiores Cuautitlán, Universidad Nacional Autónoma de México, 54700, Cuautitlán Izcalli, Edo. Méx., México.

^b.Departamento de Química, Área de Química Analítica, UAM-Iztapalapa, Apdo. Postal 55–534, 09340 Iztapalapa, Cd. Méx., México

^c-Instituto de Química, Universidad Nacional Autónoma de México, Circuito exterior, Ciudad Universitaria, 04510 Coyoacán, Cd. Méx., México.

*To whom correspondence should be addressed

email: rodolfo.gomez@unam.mx, rosariomoya@gmail.com.

Tel. (+5255) 5623 1999, ext. 39420

Methodology for the determination of the soluble Cu(IMC)₂(H₂O)₂ by UV-Vis

Calibration Standards.

In 5 volumetric flasks of 10 mL, were added 0, 0.18, 0.88, 1.75 and 3.50 mL of Cu(II) 0.01 M and adding deionized water to the fill level. From these solutions 3 mL were poured in 5 different volumetric flasks of 5 mL each. Adding 0.5 mL of CuCl₂ 0.01 to reach at least 1.20×10^{-3} M of Cu(II); 0.5 mL of NH₃/NH₄⁺ 1 M at pH 8.4 buffer were added; 0.7 mL of EDTA 0.04 M prepared in NH₃/NH₄⁺ 1 M at pH 8.4 buffer were poured to the 5 mL flask. Finally, deionized water was added to the fill level at each calibration standard. Each calibration standard was prepared for triplicate.

Calibration curve.

The UV-Vis spectra of the previous calibrations standards were obtained. Fig S1 shows an example of the average spectra of each calibrations standards obtained for calibration curve. Fig S2 shows the calibration curve created from points at 730 nm from Fig. S1, fitting to a linear least squares showing slope and y-intercept with the determination coefficient (R^2) (Fig. S1 and S2 was used for LOD, LOQ, Table S1; and HP- β -CD experiments, Fig. S6, Table S5. For the rest of CDs experiments an independent calibration curve was constructed).¹ *LOD and LOQ of Cu(II)-EDTA method.*

To determine limit of detection (LOD) and limit of quantification (LOQ) 10 blank samples (n=10) were prepared with 0.5 mL of CuCl₂ 0.01 M, to reach at least 1.20×10^{-3} M of Cu(II) in final 5 mL; then was added 0.7 mL of EDTA 0.04M prepared in NH₃/NH₄⁺ 1 M at pH 8.4 buffer, finally was added NH₃/NH₄⁺ 1 M at pH 8.4 buffer to the to 5 mL volumetric flask fill level.

Average of blanks (M)	σ_0	σ_{0}	LOD (M)	LOQ (M)	
2.88×10 ⁻⁶	5.79×10 ⁻⁶	1.83×10 ⁻⁶	5.49×10 ⁻⁶	1.83×10 ⁻⁵	

Table S1. Values obtained for the determination of LOD and LOQ of Cu(II)-EDTA (n=10).



Fig. S1. Spectra obtained of calibration standards for determination of Cu(II)-EDTA.



Fig. S2. Calibration curve for determination of Cu(II)-EDTA in 1 M NH₃/NH₄⁺ buffer pH 8.4. The data were selected at 730 nm (n=3).

The UV-Vis spectra of the blanks were obtained. The concentrations of the blanks was determined using the equation of calibration curve Fig. S2, the average concentration in molarity (M) of blanks are shown in Table S1 with the results of standard deviation σ_0 ; calculated standard deviation: $\sigma'_0 = \sigma_0/\sqrt{n}$; n = number of blank samples, LOD= $3 \times \sigma'_0$ and LOQ = $10 \times \sigma'_0$.²

Experimental samples.

The different CDs (α -, γ -, HP- β - o HP- γ -CD) were prepared around 0.08 M (β -CD was prepared at 0.017 M) at 25 mL volumetric flasks in deionized water.

Different volumetric flasks of 5 mL were prepared adding a volume of solution of CD. In each volumetric flask, 3 mg of insoluble $Cu(IMC)_2(H_2O)_2$ were added. The CD solutions with insoluble $Cu(IMC)_2(H_2O)_2$ were left in agitation for 3 days. After this time, the solutions were filtrated (filtrate), 3 mL of the filtrate were taken and poured in a volumetric flask of 5 mL, then it was poured 0.5 mL of $CuCl_2 0.01$ M to reach at least 1.20×10^{-3} M of Cu(II); 0.5 mL of NH_3/NH_4^+ 1 M at pH 8.4 buffer were added then 0.7 mL of EDTA 0.04M prepared in NH_3/NH_4^+ 1 M at pH 8.4 buffer were poured to the 5 mL flask. Finally, NH_3/NH_4^+ 1 M at pH 8.4 buffer were poured to the 5 mL flask. Finally, NH_3/NH_4^+ 1 M at pH 8.4 buffer were poured to the 5 mL flask. Finally, NH_3/NH_4^+ 1 M at pH 8.4 buffer were poured to the 5 mL flask. Finally, NH_3/NH_4^+ 1 M at pH 8.4 buffer were poured to the 5 mL flask. Finally, NH_3/NH_4^+ 1 M at pH 8.4 buffer were poured to the 5 mL flask. Finally, NH_3/NH_4^+ 1 M at pH 8.4 buffer were poured to the 5 mL flask. Finally, NH_3/NH_4^+ 1 M at pH 8.4 buffer were poured to the 5 mL flask. Finally, NH_3/NH_4^+ 1 M at pH 8.4 buffer were poured to the 5 mL flask. Finally, NH_3/NH_4^+ 1 M at pH 8.4 buffer were poured to the 5 mL flask. Finally, NH_3/NH_4^+ 1 M at pH 8.4 buffer were poured to the 5 mL flask. Finally, NH_3/NH_4^+ 1 M at pH 8.4 buffer were poured to the 5 mL flask.

The UV-Vis spectra of the experimental samples were obtained. Fig S3-7 shows the spectra obtained following the method. With the points at 730 nm of each curve of Fig. S3-7, the soluble amount of Cu(II)-EDTA was determined with a calibration curve like Fig. S2, the concentration of Cu(II)-EDTA obtained with calibration curve for each experiment with different CDs, are presented in Tables S2-5. The amount of Cu(II)-EDTA in solution was related to the Cu(IMC)₂(H₂O)₂ solubility and is presented in Tables S2-5. The relation of Cu(IMC)₂(H₂O)₂ solubility with the amount of CD, this diagrams are called phase solubility diagrams showed in Fig. 9a) of the main article. Finally, Table S6 shows the percentage of the phase solubility diagrams with the average of the lower concentration or intrinsic solubility of Cu(IMC)₂(H₂O)₂ (S₀).



Fig. S3. Spectra obtained of Cu(II)-EDTA from experimental samples for the experiment with α -CD.

[α-CD]	Experimental sample* [Cu(II)-EDTA]	Filtrate [Cu(IMC) ₂ (H ₂ O) ₂]
0.000	(9.44 ± 22.50)×10 ⁻⁶	(1.57 ± 3.75)×10 ⁻⁵
0.032	(9.18 ± 1.78)×10 ⁻⁵	(1.53 ± 0.30)×10 ⁻⁴
0.047	$(1.61 \pm 0.25) \times 10^{-4}$	(2.69 ± 0.41)×10 ⁻⁴
0.079	$(2.16 \pm 0.34) \times 10^{-4}$	(3.60 ± 0.56)×10 ⁻⁴

Table S2. Concentration of Cu(II)-EDTA from calibration curve for the α -CD experiment (Fig. S3) and concentration of
 $Cu(IMC)_2(H_2O)_2$ in filtrate..



Fig. S4. Spectra obtained of Cu(II)-EDTA from experimental samples for the experiment with β-CD.

[β-CD]	Experimental sample* [Cu(II)-EDTA]	Filtrate [Cu(IMC) ₂ (H ₂ O) ₂]
0.000	(5.32 ± 2.10)×10 ⁻⁵	(8.87 ± 3.51)×10 ⁻⁵
0.005	$(1.06 \pm 0.26) \times 10^{-4}$	(1.77 ± 0.44)×10 ⁻⁴
0.008	$(1.30 \pm 0.32) \times 10^{-4}$	(2.17 ± 0.54)×10 ⁻⁴
0.010	(1.47 ± 0.36)×10 ⁻⁴	(2.45 ± 0.61)×10 ⁻⁴

Table S3. Concentration of Cu(II)-EDTA from calibration curve for the β -CD experiment (Fig. S4) and concentration of
 $Cu(IMC)_2(H_2O)_2$ in filtrate.



Fig. S5. Spectra obtained of Cu(II)-EDTA from experimental samples for the experiment with γ -CD.

[γ-CD]	Experimental sample* [Cu(II)-EDTA]	Filtrate [Cu(IMC) ₂ (H ₂ O) ₂]
0.000	(4.70 ± 8.10)×10 ⁻⁷	(7.84 ± 1.35)×10 ⁻⁷
0.021	(5.90 ± 0.60)×10 ⁻⁵	(9.83 ± 1.01)×10 ⁻⁵
0.027	(7.89 ± 0.61)×10 ⁻⁵	(1.32 ± 0.10)×10 ⁻⁴
0.032	(9.82 ± 0.65)×10 ⁻⁵	(1.64 ± 0.11)×10 ⁻⁴
0.037	(1.20 ± 0.74)×10 ⁻⁵	(1.99 ± 0.12)×10 ⁻⁴
A OLIVER AND A SHERE A REAL AND A SHERE A		

Table S4. Concentration of Cu(II)-EDTA from calibration curve for the γ -CD experiment (Fig. S5) and concentration of Cu(IMC)₂(H₂O)₂ in filtrate



Fig. S6. Spectra obtained of Cu(II)-EDTA from experimental samples for the experiment with HP- β -CD.

[HP-β-CD]	Experimental sample*	Filtrate [Cu(IMC) ₂ (H ₂ O) ₂]
	[Cu(II)-EDTA]	
0.000	(3.15 ± 1.49)×10 ⁻⁵	(3.88 ± 1.84)×10 ⁻⁵
0.029	(1.91 ± 0.15)×10 ⁻⁴	(2.35 ± 0.18)×10 ⁻⁴
0.043	$(2.88 \pm 0.14) \times 10^{-4}$	(3.55 ± 0.18)×10 ⁻⁴
0.058	(3.43 ± 0.14)×10 ⁻⁴	(4.24 ± 0.18)×10 ⁻⁴
0.072	(4.12 ± 0.14)×10 ⁻⁵	(5.08 ± 0.17)×10 ⁻⁴
Obtained by calibration sympo		

Table S5. Concentration of Cu(II)-EDTA from calibration curve for the HP- β -CD experiment (Fig. S6) and concentration of
 $Cu(IMC)_2(H_2O)_2$ in filtrate



Fig. S7. Spectra obtained of Cu(II)-EDTA from experimental samples for the experiment with HP- γ -CD.

[HP-γ-CD]	Experimental sample* [Cu(II)-EDTA]	Filtrate [Cu(IMC) ₂ (H ₂ O) ₂]
0.000	(8.27 ± 11.60)×10 ⁻⁶	(1.38 ± 1.95)×10 ⁻⁵
0.029	(4.01 ± 1.09)×10 ⁻⁵	$(6.69 \pm 0.18) \times 10^{-4}$
0.043	(6.11 ± 1.07)×10 ⁻⁵	$(1.02 \pm 0.18) \times 10^{-4}$
0.058	(7.98 ± 1.05)×10 ⁻⁵	(1.33 ± 0.17)×10 ⁻⁴
*Obtained by calibration curve		

Table S6. Concentration of Cu(II)-EDTA from calibration curve for the HP-γ-CD experiment (Fig. S7) and concentration of Cu(IMC)₂(H₂O)₂ in filtrate

[Cu(IMC) ₂ (H ₂ O) ₂]	S ₀ (M)	Highest concentration (M)	Increased solubility (%)
α-CD	(1.57 ± 3.75)×10 ⁻⁵	(3.60 ± 0.56)×10 ⁻⁴	1140.80 ± 69.12
β-CD	(8.87 ± 3.51)×10 ⁻⁵	(2.45 ± 0.61)×10 ⁻⁴	776.38 ± 71.77
γ-CD	(7.84 ± 1.35)×10 ⁻⁷	(1.99 ± 0.12)×10 ⁻⁴	630.61 ± 67.33
HP-β-CD	(3.88 ± 1.84)×10 ⁻⁵	(5.08 ± 0.17)×10 ⁻⁴	1609.80 ± 67.38
HP-γ-CD	(1.38 ± 1.95)×10 ⁻⁵	(1.33 ± 0.17)×10 ⁻⁴	421.46 ± 68.61
Average	(3.16 ± 2.12)×10 ⁻⁵		

Table S7. Intrinsic solubility (S₀), the highest concentration obtained in phase solubility diagrams and the percentage of increased solubility of Cu(IMC)₂(H2O)₂.

Methodology for the determination of the soluble Zn(IMC)₂(H₂O)₂ by UV-Vis

Calibration Standards.

A total of 5 calibration standard were prepared adding 0, 3, 6, 9 and 12 μ L of ZnCl₂ 0.01M in volumetric flasks of 5 mL, an amount of 0.5 mL of 4-(2pyridylazo) resorcinol (PAR) 1.00×10^{-3} M was added and a solution of phosphate 0.1 M pH 7 was added to the fill level in each calibration standard. All standards were prepared in triplicate.

Calibration Curve.

Calibration standards were measured by UV-Vis. The mean of the three spectra obtained for calibration curve are shown in Fig. S8 and the calibration curve constructed from points at 490 nm, from spectra of Fig. S8, against Zn(II)-PAR concentration is shown in Fig. S9, slope, y-intercept and the determination coefficient (R^2) were obtained from a linear least squares fit (1)(Fig. S9, was used for LOD and LOQ, Table S8; and HP- γ -CD experiments, Fig. S14 and Table S13. For the rest of CDs experiments an independent calibration curve was constructed).

LOD and LOQ of Zn(II)-PAR method.

To determine limit of detection (LOD) and limit of quantification (LOQ) 10 blank samples (n = 10) were prepared with an amount of 0.5 mL of PAR 1.00×10^{-3} M and a solution of phosphate 0.1 M pH 7 was added to the fill level in each blank samples volumetric flask of 5 mL.

The UV-Vis spectra of the blanks were obtained. The concentrations of the blanks was determined using the equation of calibration curve Fig. S9, the average concentration in molarity (M) of blanks are shown in Table S8 with the results of standard deviation σ_0 ; calculated standard deviation: $\sigma'_0 = \sigma_0/\sqrt{n}$; n = number of blank samples, LOD= $3 \times \sigma'_0$ and LOQ = $10 \times \sigma'_0$.²

Average of blanks (M)	σ_{0}	$\sigma^{'}{}_{0}$	LOD (M)	LOQ (M)
3.60×10 ⁻⁷	1.24×10 ⁻⁷	3.91×10 ⁻⁸	1.17×10 ⁻⁷	3.91×10 ⁻⁷

Table S8. Table of the values to obtain LOD and LOQ of Cu(II)-EDTA (n=10).



Fig. S8. Spectra obtained of calibration standards for determination of Zn(II)

Experimental samples.

The different CDs (α -, β -, γ -, HP- β - o HP- γ -CD) were prepared around 0.08 M (β -CD was prepared at 0.017M) at 25 mL volumetric flasks in deionized water.

In different volumetric flasks of 5 mL were added different amounts of CD 0.08 M adding deionized water to the fill level, then 3 mg of $Zn(IMC)_2(H_2O)_2$ were added on each

volumetric flask. These volumetric flasks were left in agitation by 3 days. After this time the solutions were filtrated (filtrate).

In another 5 mL volumetric flask, 0.1 mL (0.3 mL in case of α - and β -CD) of filtrate, 0.5 mL of PAR 1.00×10⁻³ M and a solution of phosphate 0.1 M pH 7 was added to the fill level for each experimental sample.

The experimental samples were measured by UV-Vis for triplicated. The mean of the three spectra obtained for experimental samples are shown in Fig. S10-S14. The amount of Zn(II)-PAR in solution was related to the soluble amount of Zn(IMC)₂ with the amount of CD ₂ (Table S9-S13), this diagrams are called phase solubility diagrams showed in Fig. 9b) of the main article. Finally, Table S14 shows the percentage of increment of Zn(IMC)₂(H₂O)₂ relating the highest level concentration obtained with CDs of the phase solubility diagrams with the average of the lower concentration or intrinsic solubility of Zn(IMC)₂(H₂O)₂ (S₀).



Fig. S9. Calibration curve for determination of Zn(II) with PAR in 0.1 M phosphate buffer pH 7. The data were selected at 490 nm.



Fig. S10. Spectra obtained of Zn(II)-PAR from experimental samples for the experiment with α -CD.

[α-CD]	Experimental sample* [Zn(II)-PAR]	Filtrate [Zn(IMC) ₂ (H ₂ O) ₂]
0.000	(3.56 ± 2.23)×10 ⁻⁶	(1.78 ± 1.12)×10 ⁻⁵
0.048	(1.42 ± 0.21)×10 ⁻⁵	(2.37 ± 0.36)×10 ⁻⁴
0.056	(1.62 ± 0.37)×10 ⁻⁵	$(2.70 \pm 0.37) \times 10^{-4}$
0.064	(1.98 ± 0.24)×10 ⁻⁵	$(3.30 \pm 0.40) \times 10^{-4}$
0.081	(2.40 ± 0.27)×10 ⁻⁵	$(4.00 \pm 0.45) \times 10^{-4}$
4. Obtained by calibratian arms	(n-2)	

Table S9. Concentration of Zn(II)-PAR from calibration curve for the α -CD experiment (Fig. S10) and concentration of $Zn(IMC)_2(H_2O)_2$ in filtrate.



Fig. S11. Spectra obtained of Zn(II)-PAR from experimental samples for the experiment with β-CD.

[β-CD]	Experimental sample* [Zn(II)-PAR]	Filtrate [Zn(IMC) ₂ (H ₂ O) ₂]
0.000	(3.08 ± 2.37)×10 ⁻⁶	(1.54 ± 1.35)×10 ⁻⁵
0.005	(1.07 ± 0.22)×10 ⁻⁵	$(1.79 \pm 0.11) \times 10^{-4}$
0.008	(1.31 ± 0.21)×10 ⁻⁵	$(2.19 \pm 0.11) \times 10^{-4}$
0.010	(1.78 ± 0.22)×10 ⁻⁵	$(2.96 \pm 0.11) \times 10^{-4}$

Table S10. Concentration of Zn(II)-PAR from calibration curve for the β -CD experiment (Fig. S11) and concentration of
Zn(IMC)₂(H₂O)₂ in filtrate.



Fig. S12. Spectra obtained of Zn(II)-PAR from experimental samples for the experiment with γ -CD.

[γ-CD]	Experimental sample* [Cu(II)-EDTA]	Filtrate [Cu(IMC) ₂ (H ₂ O) ₂]
0.000	(9.13 ± 10.70)×10 ⁻⁷	(4.56 ± 5.35)×10 ⁻⁵
0.021	(1.95 ± 1.03)×10 ⁻⁶	(9.77 ± 5.15)×10 ⁻⁵
0.032	(2.44 ± 1.01)×10 ⁻⁶	(1.22 ± 0.51)×10 ⁻⁴
0.037	(2.75 ± 1.00)×10 ⁻⁶	(1.38 ± 0.50)×10 ⁻⁴
0.043	(3.08 ± 0.99)×10 ⁻⁶	(1.54 ± 0.50)×10 ⁻⁴
0.048	(3.27 ± 0.98)×10 ⁻⁶	(1.63 ± 0.49)×10 ⁻⁴

Table S11. Concentration of Zn(II)-PAR from calibration curve for the γ -CD experiment (Fig. S12) and concentration of $Zn(IMC)_2(H_2O)_2$ in filtrate.



Fig. S13. Spectra obtained of Zn(II)-PAR from experimental samples for the experiment with HP-6-CD.

[HP-β-CD]	Experimental sample* [Zn(II)-PAR]	Filtrate [Zn(IMC) ₂ (H ₂ O) ₂]
0.000	(3.90 ± 3.22)×10 ⁻⁶	(9.75 ± 8.05)×10 ⁻⁶
0.029	(1.16 ± 0.28)×10 ⁻⁵	(5.82 ± 1.38)×10 ⁻⁴
0.044	(1.52 ± 0.27)×10 ⁻⁵	(7.59 ± 1.37)×10 ⁻⁴
0.051	(1.73 ± 0.28)×10 ^{−5}	(8.64 ± 1.40)×10 ⁻⁴

Table S12. Concentration of Zn(II)-PAR from calibration curve for the HP-6-CD experiment (Fig. S13) and concentration of $Zn(IMC)_2(H_2O)_2$ in filtrate.



Fig. S14. Spectra obtained of Zn(II)-PAR from experimental samples for the experiment with HP- γ -CD.

[HP-γ-CD]	Experimental sample* [Zn(II)-PAR]	Filtrate [Zn(IMC) ₂ (H ₂ O) ₂]
0.000	(1.43 ± 0.29)×10 ⁻⁵	(3.58 ± 1.45)×10 ⁻⁵
0.029	(1.07 ± 0.29)×10 ⁻⁵	(1.79 ± 0.15)×10 ⁻⁴
0.051	(1.55 ± 0.29)×10 ⁻⁵	(2.59 ± 0.14)×10 ⁻⁴
0.065	(1.87 ± 0.30)×10 ⁻⁵	(3.12 ± 0.15)×10 ⁻⁴
0.072	(2.12 ± 0.30)×10 ⁻⁵	(3.53 ± 0.15)×10 ⁻⁴

Table S13. Concentration of Zn(II)-PAR from calibration curve for the HP- γ -CD experiment (Fig. S14) and concentration of $Zn(IMC)_2(H_2O)_2$ in filtrate.

	C (DA)	Highest	Increased solubility
	$S_0(IVI)$	concentration (M)	(%)
α-CD	(1.78 ± 1.12)×10 ⁻⁵	$(4.00 \pm 0.45) \times 10^{-4}$	1608.36 ± 78.58
β-CD	(1.54 ± 1.35)×10 ⁻⁵	(2.96 ± 0.11)×10 ⁻⁴	1190.19 ± 77.88
γ-CD	(4.56 ± 5.35)×10 ⁻⁵	(1.63 ± 0.49)×10 ⁻⁴	655.41 ± 83.44
HP-β-CD	(9.75 ± 8.05)×10 ⁻⁶	(8.64 ± 1.40)×10 ⁻⁴	3474.07 ± 78.44
HP-γ-CD	(3.58 ± 1.45)×10 ⁻⁵	(3.53 ± 0.15)×10 ⁻⁴	1419.38 ± 77.90
Average	(2.49 ± 1.93)×10 ⁻⁵		
*Obtained by cal	libration curve.		

Table S14. Intrinsic solubility (S_0), the highest concentration obtained in phase solubility diagrams and the percentage of increased solubility of $Cu(IMC)_2(H_2O)_2$.

Ca-EDTA standard reaction

The results of the Ca-EDTA to check reliability of the equipment is reported in Fig. S15 and

Table S15.



Fig. S15. ITC titration of EDTA with Ca^{2+} at pH 5.5. 1.06mM EDTA and 10.1 mM Ca^{2+} in acetate buffer at 298.15 K.

Parameter	Experimental values	Reference values ³
Ν	1.030 ± 0.002	1.07 ± 0.05
logK₅	5.01 ± 0.02	5.08 ± 0.02
$\Delta H_{ m b}$ (kcal/mol)	1.76 ± 0.01	1.80 ± 0.07

Table S13. Experimental parameters of the Ca-EDTA, compared with the reference values.

References.

1. Miller JN, Miller JM. Estadística y quimiometría para química analítica [Internet]. Editor. Prentice Hall. 2002. p. 296.

 Magnusson O. Eurachem Guide: The Fitness for Purpose of Analytical Methods – A Laboratory Guide to Method Validation and Related Topics, [Internet]. Eurachem Guid.
 Available from: <u>http://www.eurachem.org/images/stories/Guides/pdf/valid.pdf</u>

3. Ràfols, C.; Bosch, E.; Barbas, R.; Prohens, R. The Ca2+–EDTA Chelation as Standard Reaction to Validate Isothermal Titration Calorimeter Measurements (ITC). *Talanta* 2016, 154, 354–359. https://doi.org/http://dx.doi.org/10.1016/j.talanta.2016.03.075.