

Porphyrin structure carbon dots under red light irradiation for bacterial inactivation

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Electronic Supplementary Information

Experimental details

ROS measurement

1, 3-diphenylisobenzofuran (DPBF) was chosen as ROS trapping agent to indicate the amount of ROS produced. 5.4 mg of DPBF was dissolved into 4 mL of ethanol to form the fresh indicator solution. The ROS measurement was achieved by following the next steps in dark. First, 2 mL with 50 μ L of DPBF stock solution was added to the UV cuvette, and covered with seal, then was scanned in UV-Vis absorption spectra recording from 400 to 440 nm. Subsequently, UV-Vis were measured every 1 min under laser irradiation (638-nm, 1 W cm^{-2}) for 20 min. The absorbance at 400–440 nm was recorded. 50 μ L of DPBF stock solution was added in 2 mL of DI as control group, and UV-Vis absorption spectra were measured at the same conditions.

Photothermal experiment

To evaluate the photothermal activity, 900 μ L of the DI with 100 μ L of CDs solution (a final concentration within 25 $\mu\text{g mL}^{-1}$) was transferred into a 24-well plate. The temperature of the solution (every minute) was recorded under the irradiation during 20 min with a 638-nm laser (0.1-1 W cm^{-2}).

Intracellular ROS detection in bacteria

The intracellular ROS level was measured by CDs mediated PDT using 2',7'-dichlorofluoresce in diacetate (DCFH-DA) through a ROS assay kit (Beyotime Reagent Co., China), according to previous studies^[1]. Briefly, *E.coil* (1.0×10^9) were washed three times with PBS and the cell density adjusted to 1.0×10^6 cfu mL^{-1} .

DCFH-DA (10 μM) was added to the bacterial suspension, and then incubated for 20 min at 25°C. The suspensions were washed thoroughly for three times with PBS to remove the DCFH-DA. Then, cleaned bacteria except control were exposed to 1 $\mu\text{g mL}^{-1}$ of CDs for 30 min in dark and irradiated under 638 nm laser (0.05W cm^{-2}) for 3 min. The fluorescence of DCFH-DA was observed under a laser scanning confocal microscope (LSCM) at 488 nm excitation and 525 nm emission wavelengths.

Cytotoxicity experiment

3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (MTS, Promega, (Beijing) Biotech Co., Ltd.) was involved to evaluate the cytotoxicity of CDs, and Bend.3 cells were chosen as model. Briefly, cells were seeded in a 96-well plate (100 cells/well) to adhere for 12 h at 37°C. After that, medium containing different concentrations of CDs (0, 5, 10, 15, 20, 25, 30, 40, 50 $\mu\text{g mL}^{-1}$) was added. After 24 h of incubation in the dark, the medium was washed thoroughly, and then replaced by 100 μL of fresh medium containing 20 μL of MTS. After incubation for another 1 h, shaking the plate for 3 min. Subsequently, the absorbance at 490 nm was measured using an ELISA plate reader. The cell viability was estimated based on the following equation: $\text{cell viability\%} = (\text{OD}_{\text{CDs}} - \text{OD}_{\text{control}}) / (\text{OD}_{\text{cell}} - \text{OD}_{\text{control}})$.

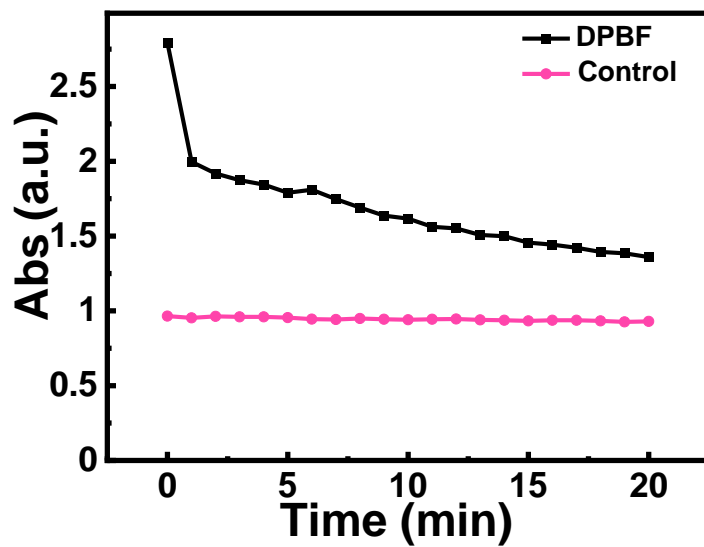


Fig. S1 Absorbance of DPBF at 422 nm under the 638-nm laser irradiation in the presence and absence of $10 \mu\text{g mL}^{-1}$ CDs as photosensitizer;

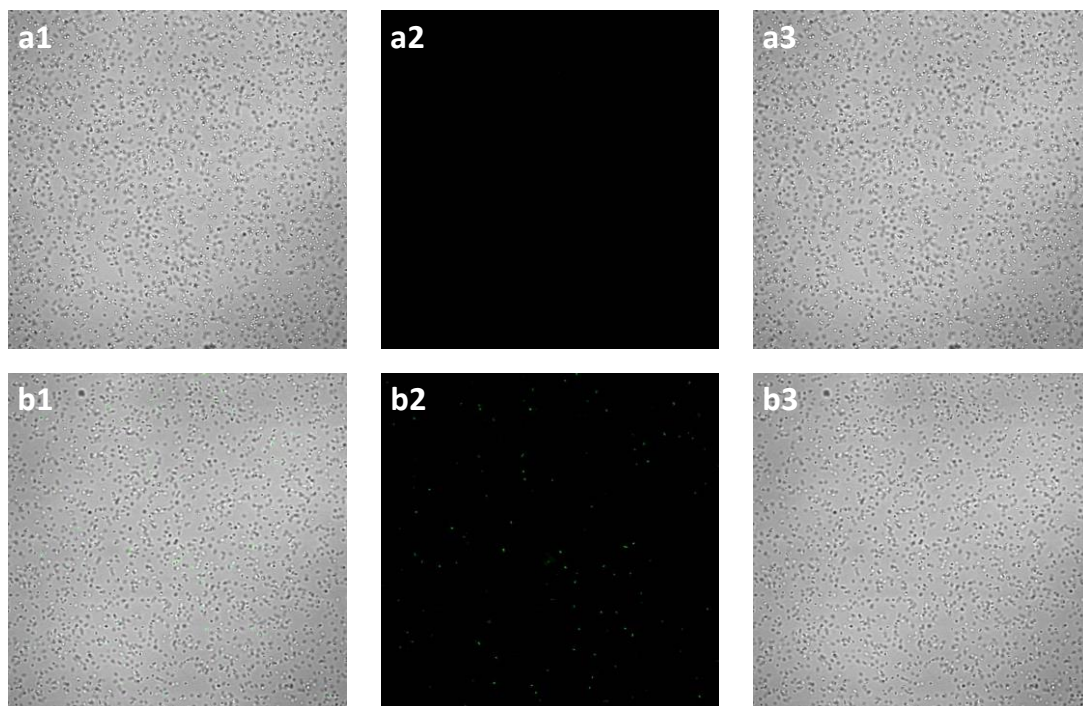


Fig. S2 Intracellular ROS detection in *E.coil* (a1-a3) control group without CDs incubation and (b1-b3) $1 \mu\text{g mL}^{-1}$ of CDs for 30 min in dark. (a1, b1) merged field, (a2, b2) $\lambda_{\text{ex}}=488 \text{ nm}$ and $\lambda_{\text{em}}=525 \text{ nm}$, (a3, b3) bright field.

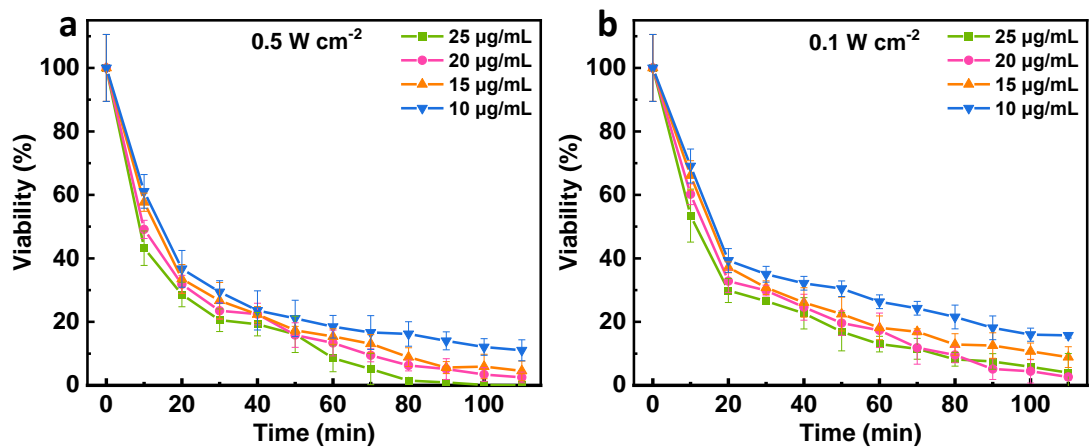


Fig. S3 Within the CDs concentration of 10-25 $\mu\text{g mL}^{-1}$, the bacteria inactivation by the 638-nm laser with different power densities of (a) 0.5 W cm^{-2} and (b) 0.1 W cm^{-2} .

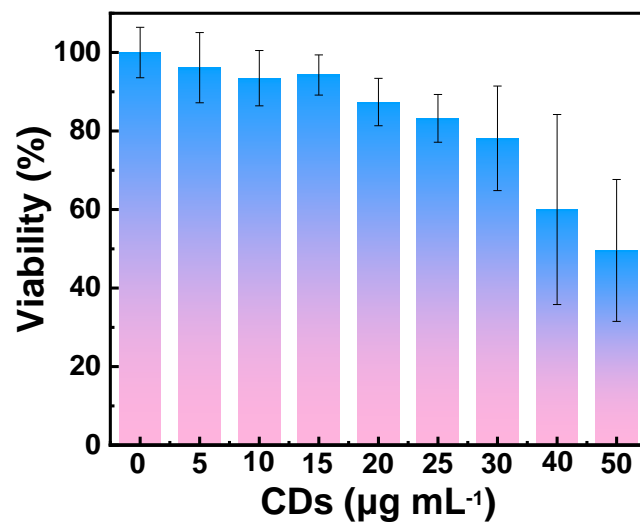


Fig. S4 Cell viability of Bend.3 with CDs (0-50 $\mu\text{g mL}^{-1}$) in the dark

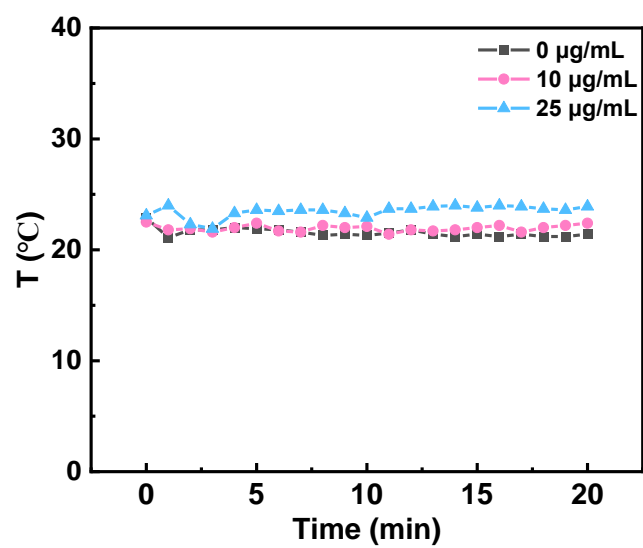


Fig. S5 Temperature measurement with CDs under the 1 W cm^{-2} of the 638-nm laser.

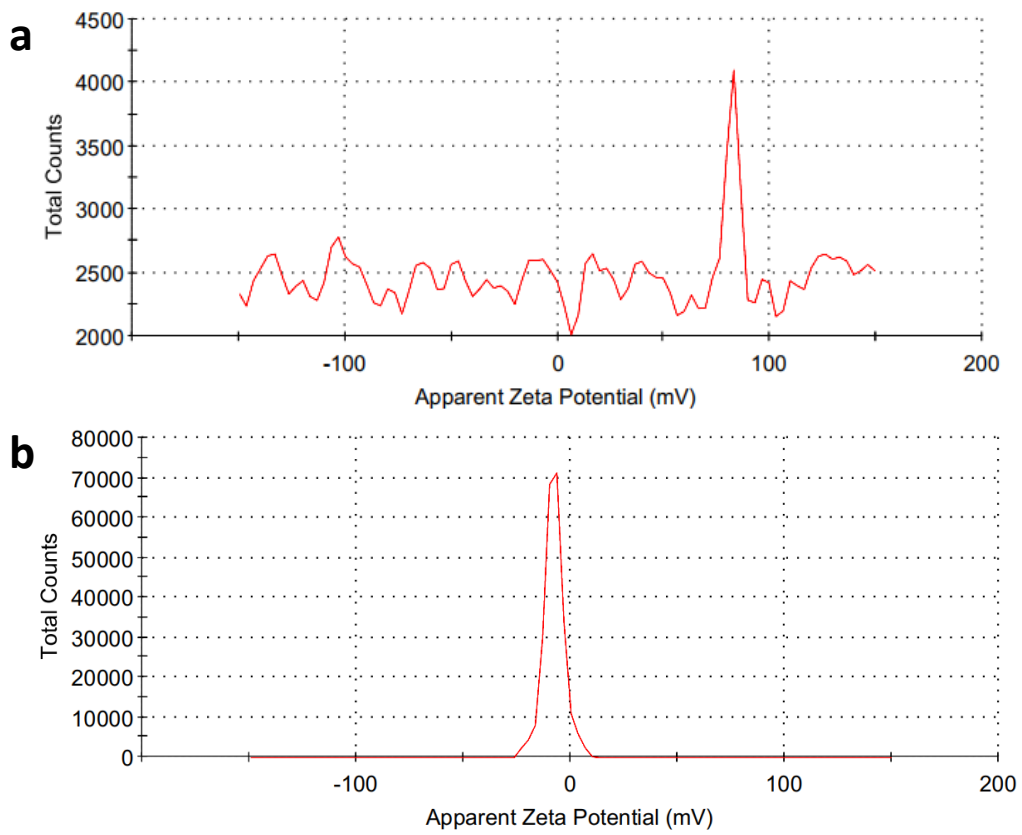


Fig. S6 Zeta potentials of (a) CDs and (b) *E.coli*.

References

1. Dwivedi S, Wahab R, Khan F, et al. Reactive oxygen species mediated bacterial biofilm inhibition via zinc oxide nanoparticles and their statistical determination [J]. PLoS One, 2014, 9(11): e111289.