

Supporting information

Deep eutectic solvent-based manganese molybdate nanosheets for sensitive and simultaneous detection of human lethal compounds: Comparing the electrochemical performances of M-molybdate (M= Mg, Fe, and Mn) electrocatalysts

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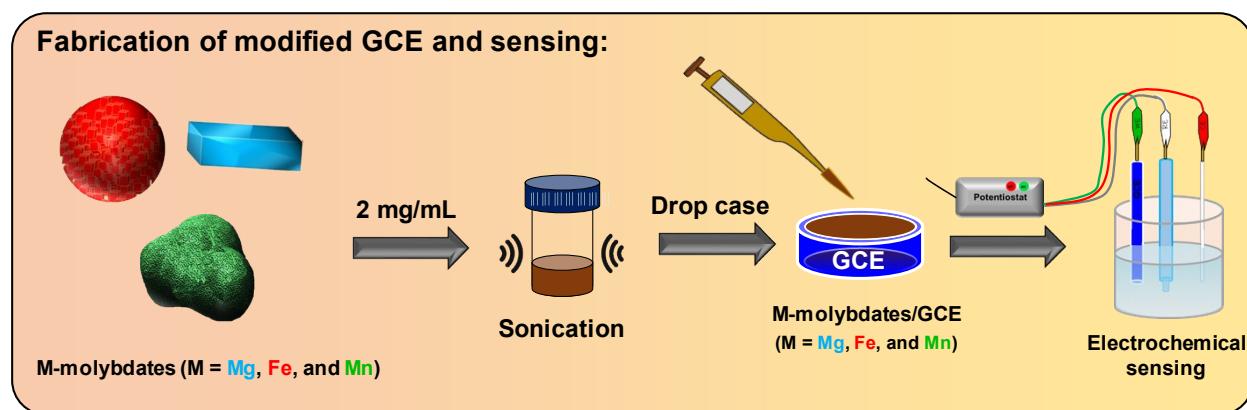
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Chemicals and Reagents

All the chemicals used in the study are of analytical grade and are received from Sigma-Aldrich and Merck. Ultrapure fresh water is obtained from a millipore water purification system (Milli-Q, specific resistivity $>18 \text{ M}\Omega\text{cm}$, S.A.; Molsheim, France) and is used in all the experiments. Sodium phosphate dibasic and sodium dihydrogen phosphate (Na_2HPO_4 and NaH_2PO_4) are utilized to prepare 0.05 M (pH 7) PB (phosphate buffer). All the electrochemical experiments are carried out using 0.05 M PB (pH 7) as the supporting electrolyte.

Fabrication of Modified Electrodes.



Scheme S1. Fabrication of M-molybdate (M- Mg, Fe and Mn) modified electrodes and sensing of HQ, Hg^{2+} and NO_2^-

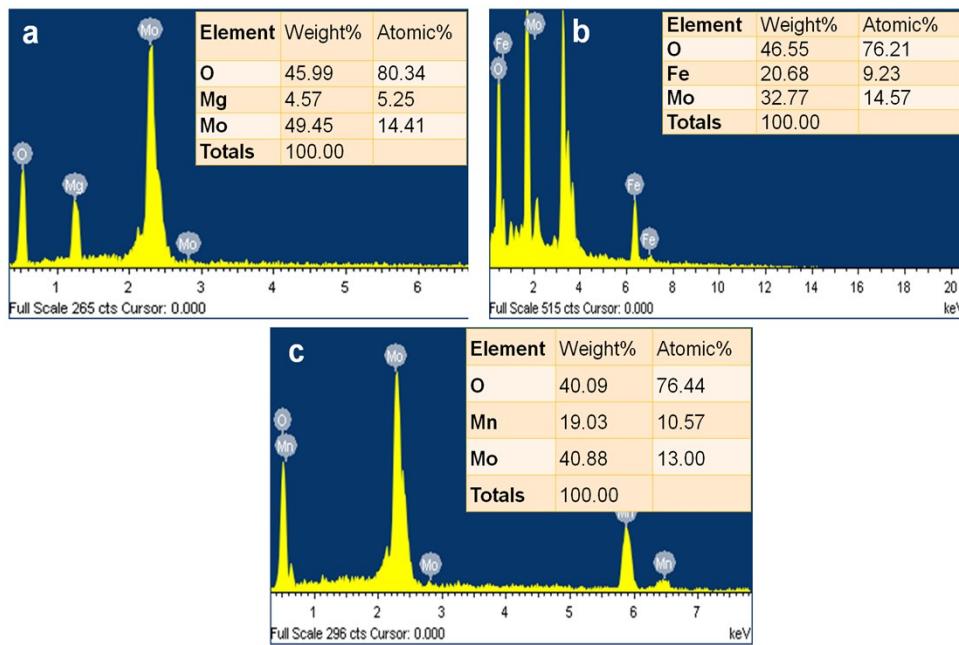


Fig. S1. EDS of **(a)** MgMoO₄ NRs **(b)** Fe₂(MoO₄)₃ YSs and **(c)** MnMoO₄ BC.

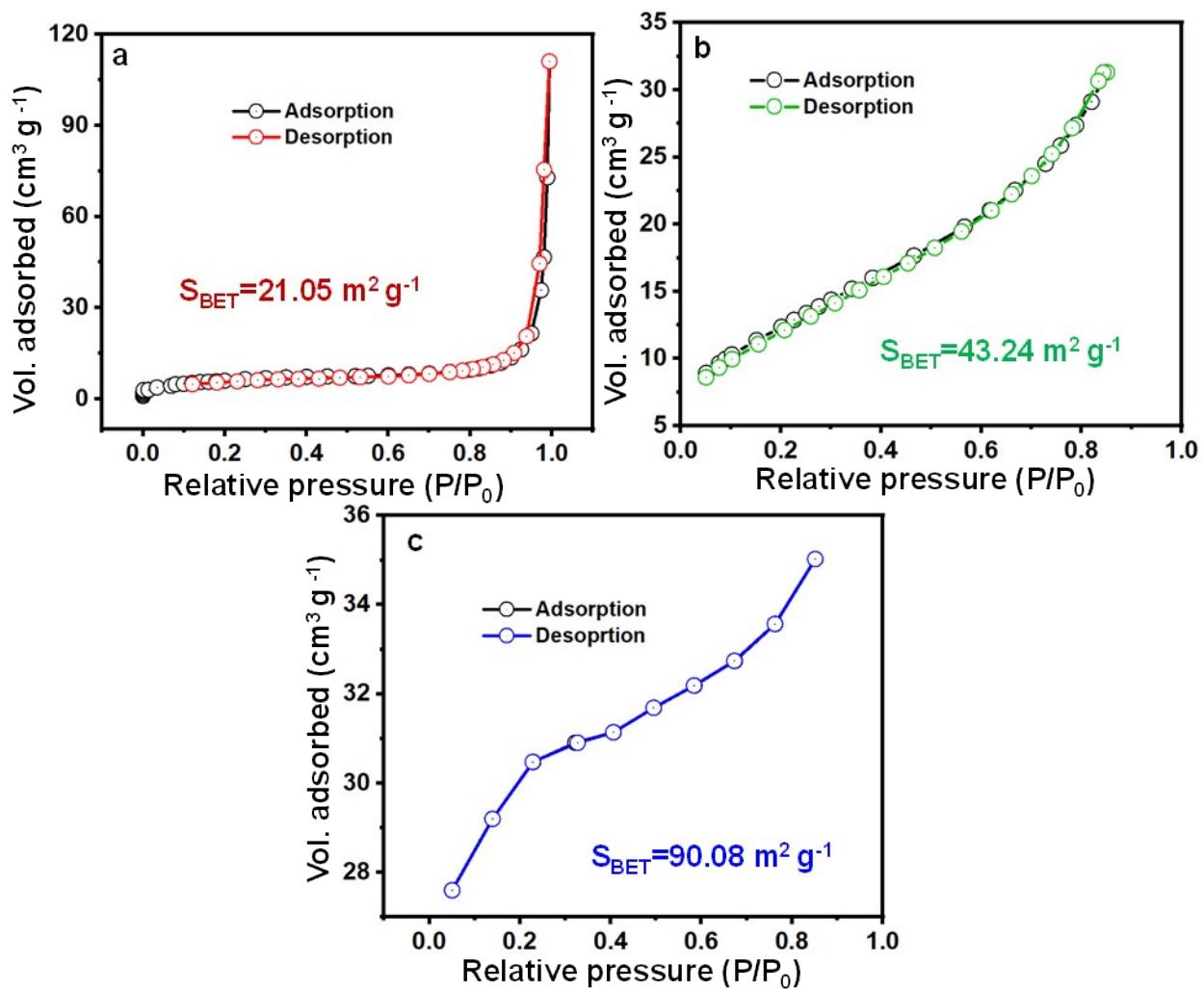


Fig. S2. BET isotherms of (a) MgMoO₄ (b) Fe₂(MoO₄)₃ and (c) MnMoO₄.

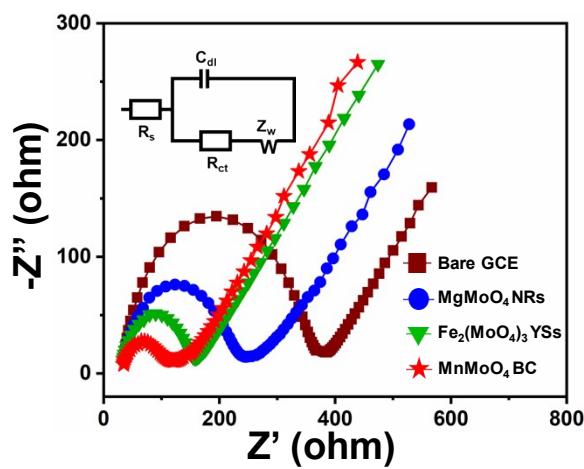


Fig. S3. Nyquist plot of various electrodes: bare GCE, MgMoO₄ NRs/GCE, Fe₂(MoO₄)₃ YSS/GCE and MnMoO₄ BC/GCE; inset: diagram of Randles equivalent circuit model.

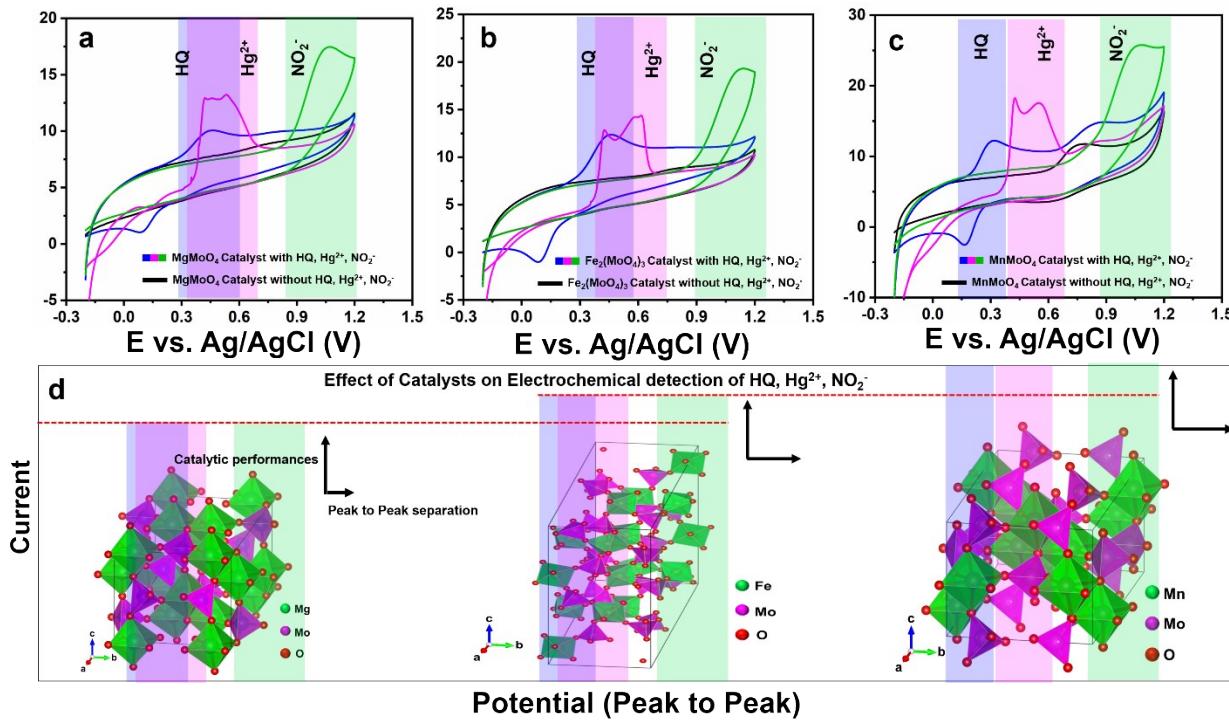


Fig.S4. CV profiles of background, unmodified and modified MgMoO₄ NRs (a), Fe₂(MoO₄)₃ YSS (b), and MnMoO₄ BC (c) electrodes towards 100 μ M of HQ, Hg²⁺ and NO₂⁻. All measurements were recorded in electrolyte: 0.05 M PB (pH 7.0) at a scan rate of 50 mV s⁻¹. (d), Crystal structures of MgMoO₄ (a), Fe₂(MoO₄)₃ (b), and MnMoO₄ with catalytic performances towards Electrochemical detection of HQ, Hg²⁺ and NO₂⁻.

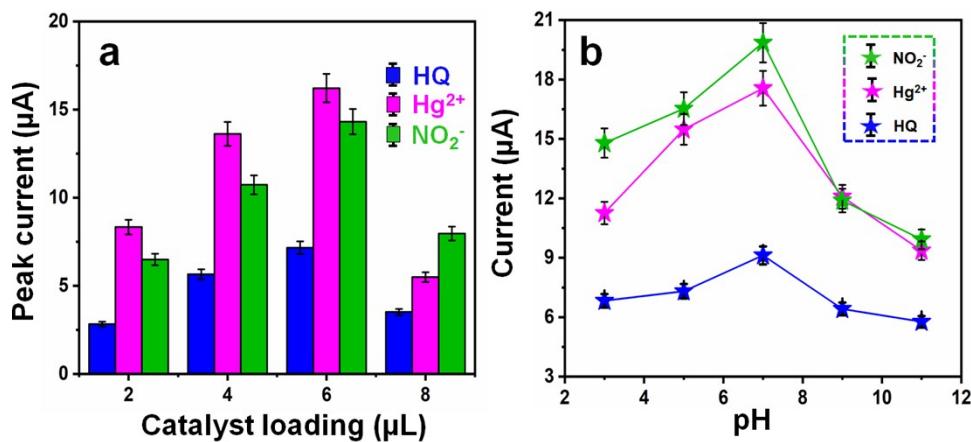


Fig. S5. (a) Effect of catalyst loading on GCEs (μ L) vs. relative peak current in presence of HQ, Hg^{2+} and NO_2^- . (b) Plot of MnMoO_4 BC/GCE obtained in supporting electrolyte at different pH containing 100 μM HQ, Hg^{2+} and NO_2^- (plot between current vs pH).

Real sample preparation

Face cream was obtained from local shop market. Tuna fish & Salami are brought from Zhnogshan fish market, Taipei, Taiwan. Certain amount of fishes is weighed for about 6g and is been homogenized with 3.0 mL of 0.1 mol L⁻¹ EDTA and merged with 2.0 mL in pH 7.0 McIavine buffer solution by utilizing HY-4 homogenizer model instrument with a lateral speed of 5000 rpm for nearly 30 mins. Afterwards it is shaken vigorously for almost 10 mins with 3 m mL of acetonitrile. Continuing, it is centrifuged at a speed of 5000 rpm for 10 mins, transferred to a clean polypropylene tube. Moreover, using 3.0 mL of aliquot acetonitrile fish residues were extracted. Later, both the extracts are combined further evaporated by drying in nitrogen atmosphere. Obtained dried extract is re-formed in 200 μL of methanol, centrifuged in 5000 rpm for 5 mins and then stored for further use in 40C dark area.

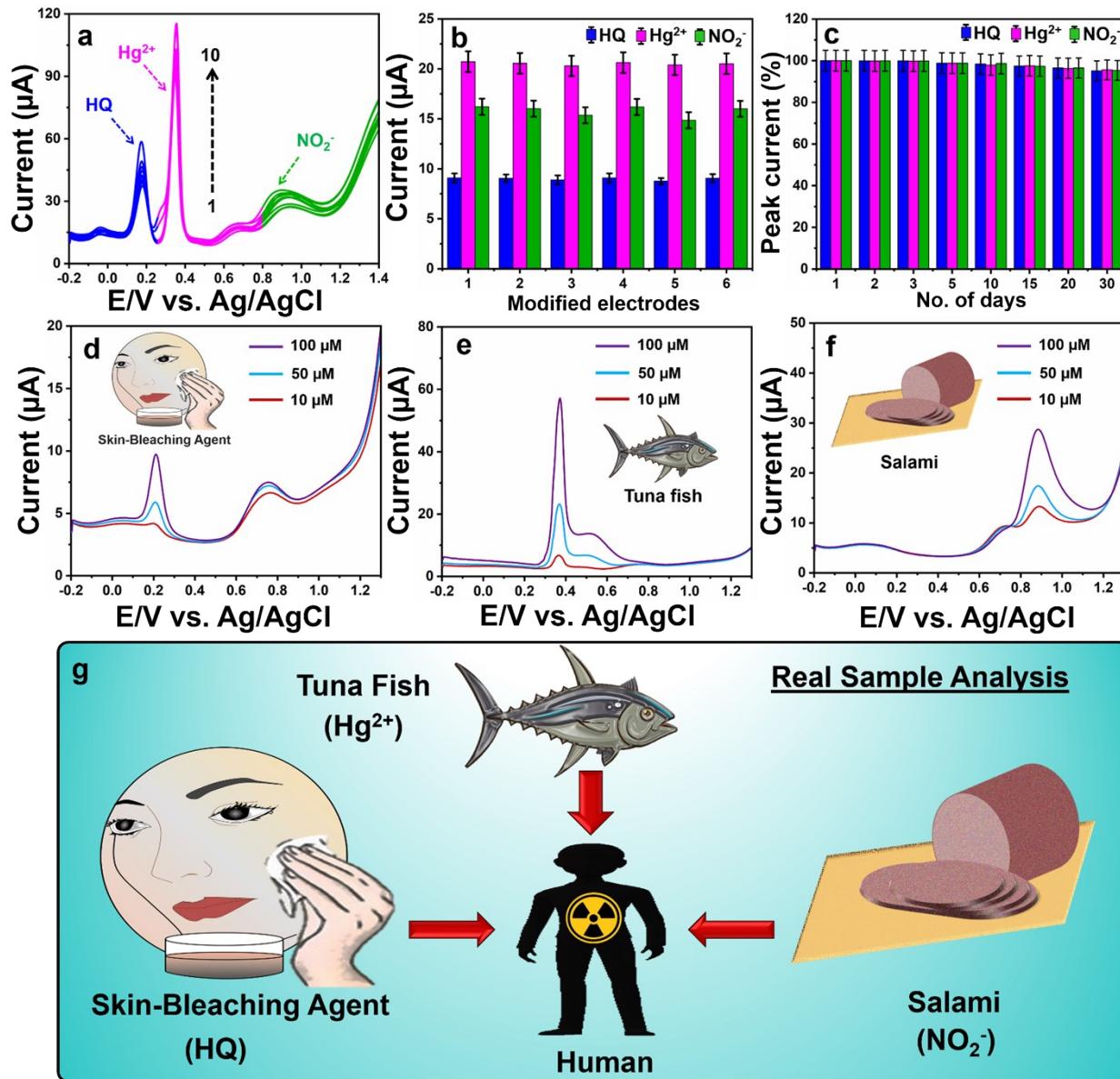


Fig. S6. **(a)** DPV profiles of selectivity of the MnMoO_4 modified GCE containing simultaneous addition HQ Hg^{2+} and NO_2^- in the presence of an excess concentration (1–9) of CC, RC, Cd^{2+} , Pb^{2+} , NaAc, ascorbic acid, NaCl, bisphenol A, and glucose. **(b and c)** Reproducibility and stability of the MnMoO_4 modified GCE in 0.05 M pH 7.0. DPV profiles of different real samples of the MnMoO_4 modified GCE containing **(d)** HQ, **(e)** Hg^{2+} and **(f)** NO_2^- . **(g)** Schematic representation of real-time analysis of analytes causing metabolic toxication.

Table S1. Structural, BET and BJH parameters of MgMoO₄, Fe₂(MoO₄)₃ and MnMoO₄.

Sample	Lattice parameters (Å)				Scherrer method	Lattice strain	Surface area (m ² /g)	Pore volume (cc/g)	Pore diameter (nm)
	a	b	c	β					
MgMoO ₄	10.15	9.24	7.03	105.9	27.55	0.0102	21.05	0.078	8.91
Fe ₂ (MoO ₄) ₃	15.70	9.23	18.2	125.2	22.95	0.0122	45.24	0.046	6.19
MnMoO ₄	10.46	9.51	7.14	106.2	13.45	0.0105	90.08	0.019	2.88

Table S2. Comparison of analytical confines for the determination of HQ at MnMoO₄ BC modified electrode with previous reports.

Electrode materials	Methods	Linear range (μM)	LOD (μM)	Interfering species	Ref
PANI/MnO ₂	DPV	0.2–100	0.13	CC and RC	S1
CD ^a /r-GO	DPV	0.5–1000	0.17	CC and RC	S2
AuNPs/Fe ₃ O ₄ - ^b APTES-GO	Amperometric i-t	3–137	1.1	CC	S3
GR ^c -La(OH) ₃	DPV	5–300	0.015	CC	S4
Pt-Au- ^d OSi@CS	Amperometric i-t	0.06–90.98	0.02	CC	S5
Graphene-chitosan	DPV	1.0–200	0.75	CC	S6
NA-COGH ^e	DPV	0.8–500	0.034	CC	S7
Polydopamine-RGO	DPV	1.0–2300	0.074	CC	S8
P-rGO	DPV	5-120	0.18	CC	S9
GMC	DPV	2-70	0.31	CC	S10
Nitrogen-doped CNT	DPV	20–1000	2.71	CC	S11
MnMoO₄ BC	DPV	0.9-591.2	0.026	Hg²⁺ and NO₂⁻	This work

*a- Carbon dot, b- (3-Aminopropyl) triethoxysilane, c- Graphene,**d- Organosilica@chitosan composites, e- Nanopore array derived cysteine oxide/gold hybrids.***Table S3.** Comparison of analytical limitations for the determination of Hg²⁺ at MnMoO₄ BC modified electrode with previous reports.

Electrode Materials	Methods	Linear Range	LOD	Interfering Species	Ref.
Pd _{1.5} / ^a PAC-900	DPV	0.24–7.5 μM	0.054 μM	Cd ²⁺ , Pb ²⁺ , and Cu ²⁺	S12
^b NH ₂ -CMS	SWASV ^c	0.4–1.2 μM	0.098 μM	Cd ²⁺ , Pb ²⁺ , and Cu ²⁺	S13
DNA Modified Fe ₃ O ₄ @Au NPs	SWV	10–100 nM	1.7 nM	Ag ⁺	S14
SnO ₂ /RGO	SWASV	0.4–1.2 μM	0.279 nM	Cd ²⁺ , Pb ²⁺ , and Cu ²⁺	S15
NG ^d	DPSV ^e	0.2–9 μM	0.05 μM	Cd ²⁺ , Pb ²⁺ , and Cu ²⁺	S16
MnFe ₂ O ₄ NAs ^f	DPV	100 nM–872.3 μM	0.14 nM	-	S17
^g Alk-Ti ₃ C ₂	SWASV	0.1–1.5 μM	0.130	Cd ²⁺ , Pb ²⁺ , and Cu ²⁺	S18
MnMoO₄ BC	DPV	0.9–591.2 μM	0.05 μM	HQ and NO₂⁻	This work

a- Porous activated carbons, b- Amino-functionalized carbon microsphere, c- Square wave anodic stripping voltammetry, d- N-doped graphene e- Differential pulse stripping voltammetry, f- Nano-agglomerates g- Alkaline intercalation of Ti₃C₂ MXene

Table S4. Comparison of analytical limitations for the determination of NO₂⁻ at MnMoO₄ BC modified electrode with previous reports.

Electrode Materials	Methods	Linear Range	LOD (μM)	Interfering Species	Ref.
La-MWCNTs	Chronoamperometry	0.4–710 μM	0.13	AA, DA and UA	S19
HOOC-MWCNT	DPV	100 μM to 0.7 mM	0.56	SO ₃ ²⁻	S20
Cu/MWCNT/RGO	SWV	0.1–75 μM	0.03	NO ₃ ⁻	S21
AgZEGE ^a	CV	100–1000 μM	0.005	NO ₃ ⁻	S22
G-PLA ^b	Multiple-pulse amperometry	0.5–250 μM	0.03	Uric Acid	S23
NRCu ^c /rGO	DPV	0.3–131.5 μM	0.1	HQ, CC and RS	S24
^d 2,4-DDMA-NiO/NP	SWVs	10.0–700.0 μM	5.0	TS and 4-CP ^e	S25
MnMoO₄ BC	DPV	0.9–453.1 μM	0.01	HQ and Hg²⁺	This work

a-Zeolite-expanded graphite-epoxy composite electrode, b-Polylactic acid containing graphene, c-Nanoraspberry-like copper, d- 2,4-dimethyl-N'-[1-(2,3-dihydroxyphenyl)methylidene]aniline, e-Thiosulfate and 4-chlorophenol

Table S5. Results of determination of HQ, Hg²⁺ and NO₂⁻ in face cream, tuna fish, and salami samples towards MnMoO₄ BC/GCE.

Samples	Added* (μM)	Detected (μM) ^a HPLC	Detection Rate (%) (Mean \pm RSD) (n=3)
Face cream	0	-	-
	10	09.86	98.60 \pm 0.017
	50	49.90	99.80 \pm 0.011
	100	99.70	99.70 \pm 0.005
Tuna fish	0	-	-
	10	09.73	97.30 \pm 0.014
	50	49.50	99.00 \pm 0.008
	100	99.17	99.17 \pm 0.019
Salami	0	-	-
	10	09.82	98.20 \pm 0.012
	50	49.27	98.54 \pm 0.018
	100	99.16	99.16 \pm 0.017

^aStandard addition method. Relative standard deviation deduced from three separate measurements.

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