

### Supplemental Information

## Tunable surface wettability and pH-responsive 2D structures from amphiphilic and amphoteric protein microfibrils

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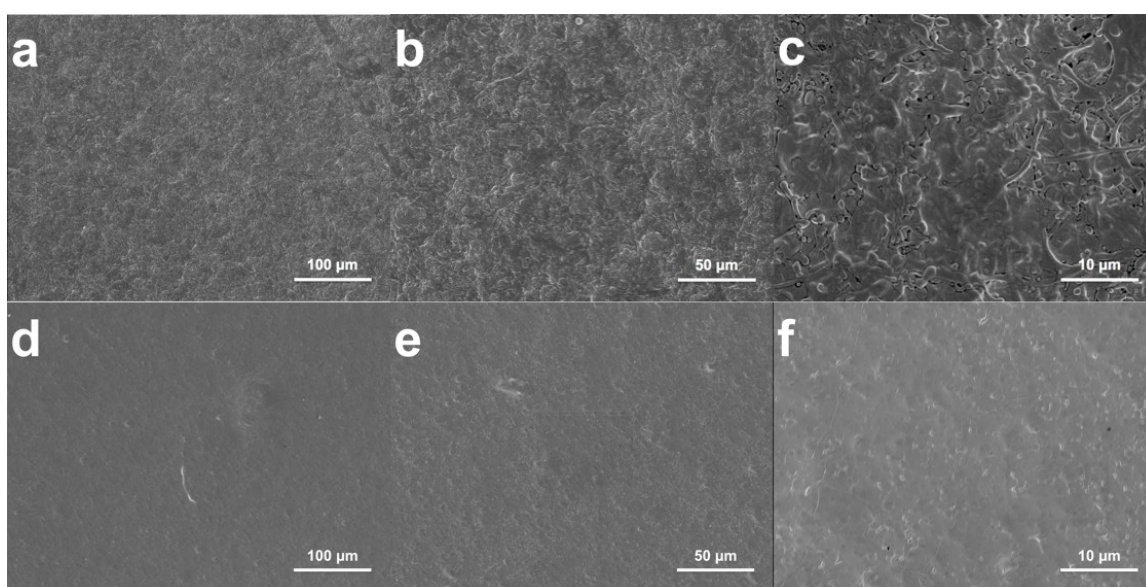


Figure S1. SEM images of film #1 (21 °C): a-c. top surface; d-f. bottom surface.

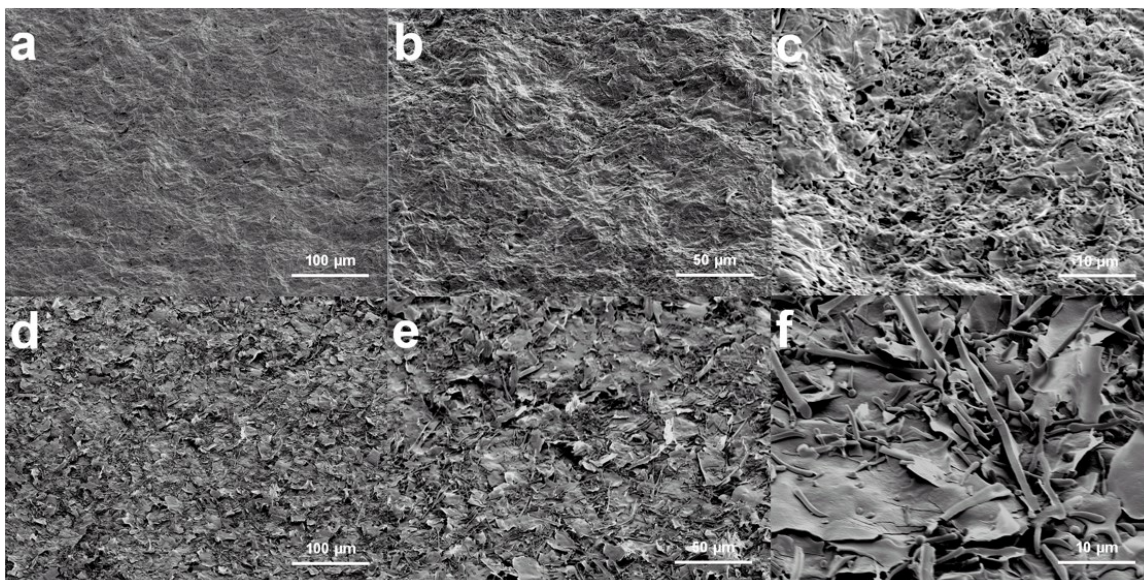


Figure S2. SEM images of film #2 (21 °C, vacuum): a-c. top surface; d-f. bottom surface.

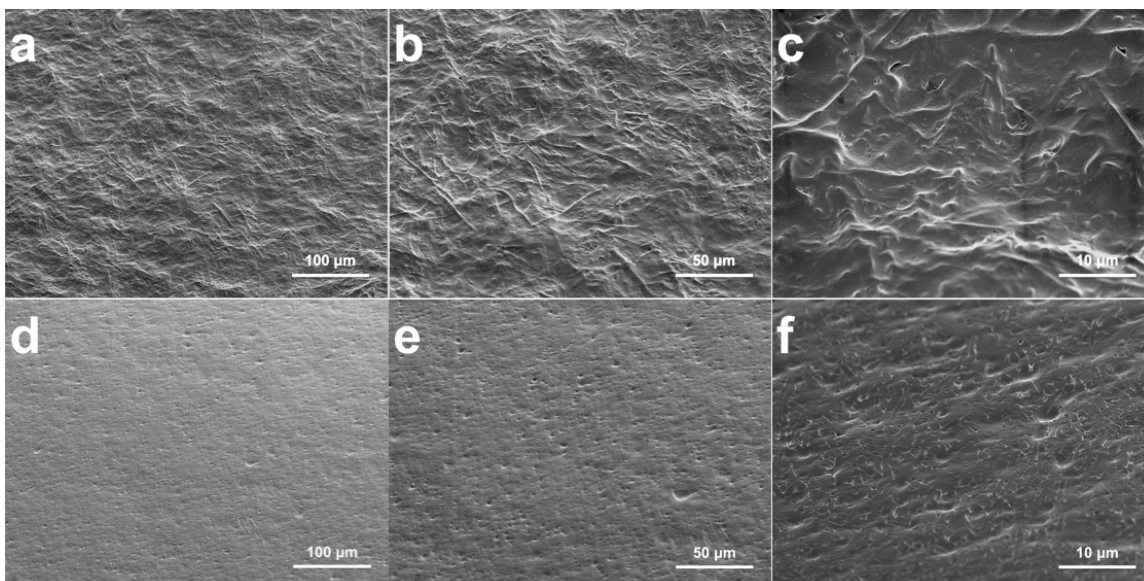


Figure S3. SEM images of film #3 (21 °C, vacuum, moisture): a-c. top surface; d-f. bottom surface.

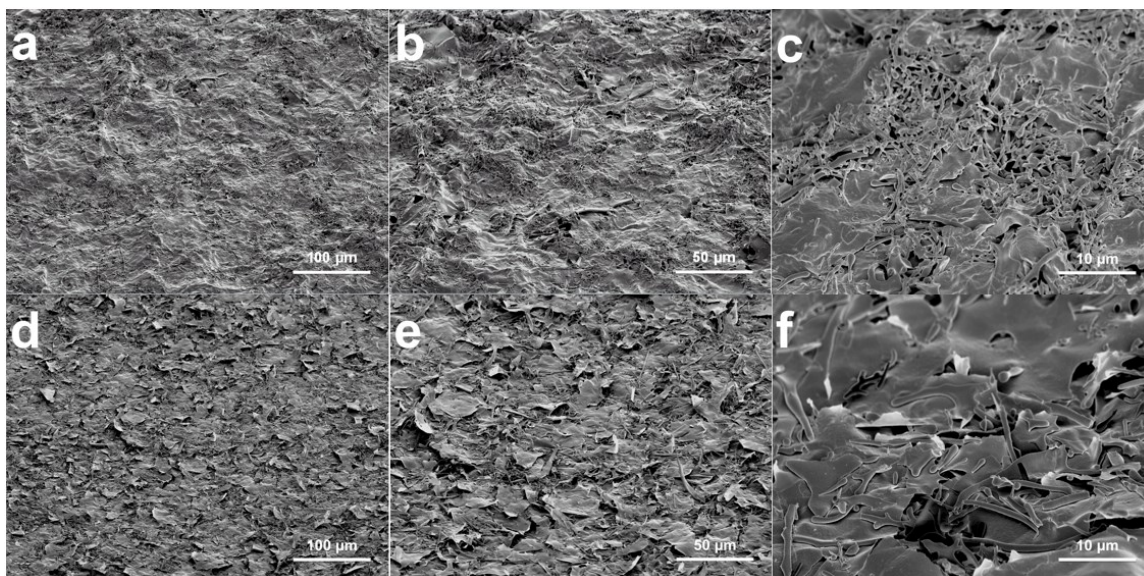


Figure S4. SEM images of film #4 (65 °C): a-c. top surface; d-f. bottom surface.

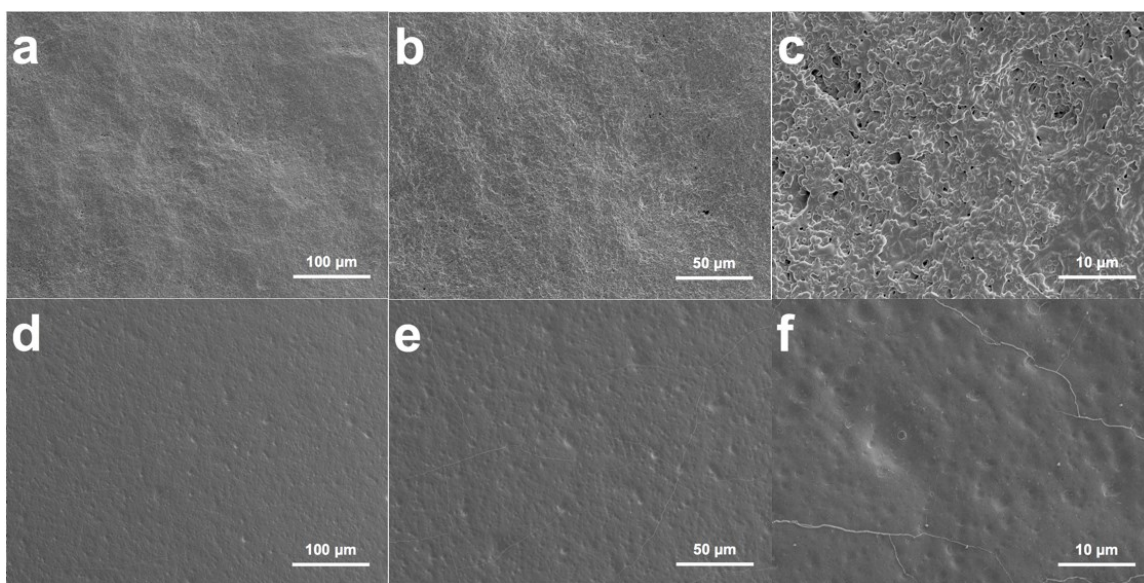


Figure S5. SEM images of film #5 (65 °C, vacuum, moisture): a-c. top surface; d-f. bottom surface.

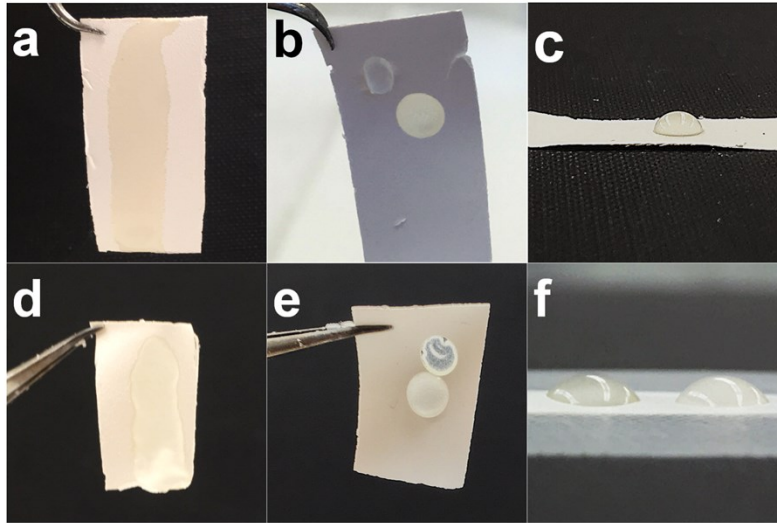
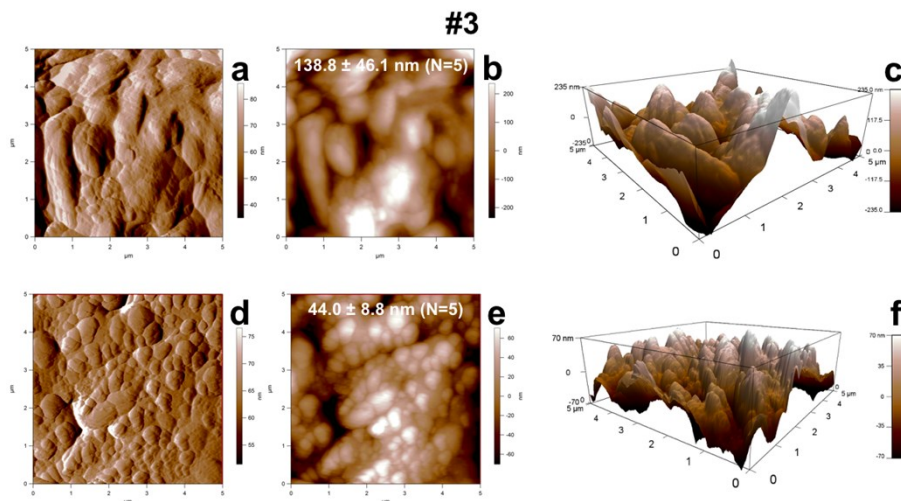


Figure S6. Water drops on top (a and d) and bottom (b-c and e-f) surfaces of film #2 (21 °C, vacuum, a-c) and #4 (65 °C, d-f) and: films were vertical in a-b and d-e and horizontal in c and f.



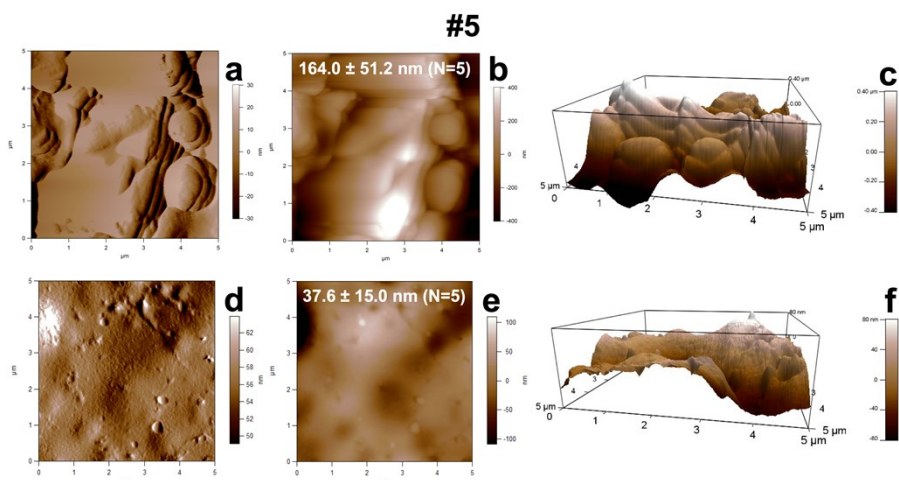


Figure S7. AFM amplitude (a, d), height (b, e) and 3D (c, f) images of the top (a-c) and bottom (d-f) surface of film #3 (21 °C, vacuum, moisture) and #5 (65 °C, vacuum, moisture).

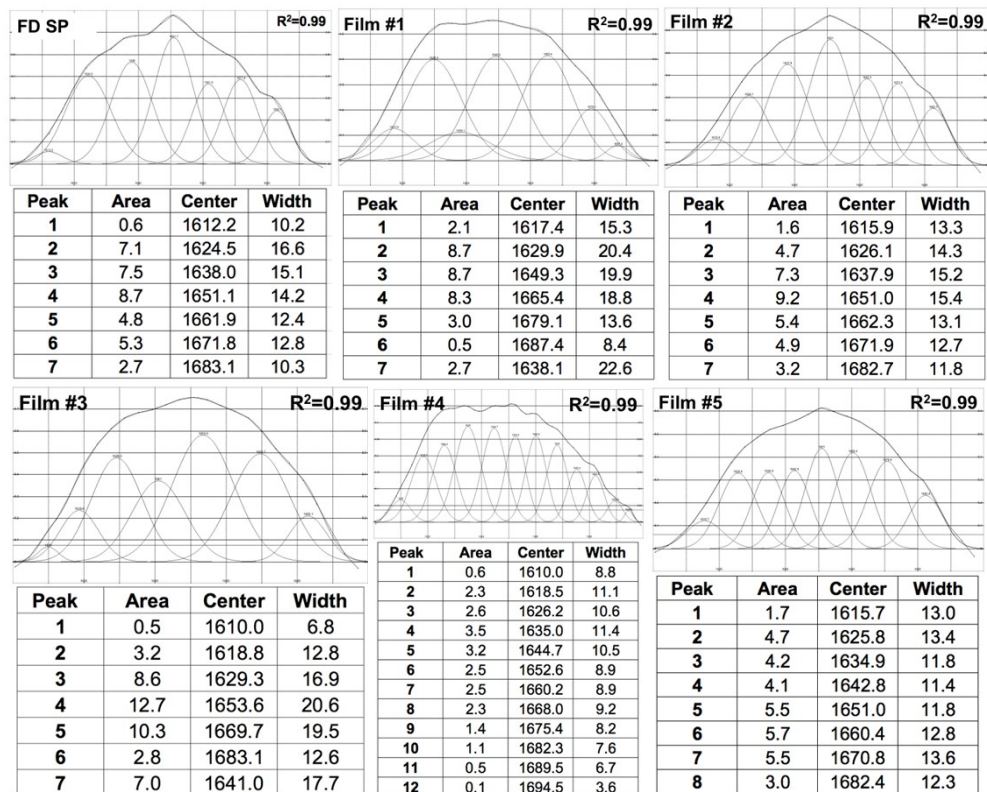


Figure S8. Curve-fitted amide I region (1600–1700  $\text{cm}^{-1}$ ) with secondary structure determination of FD SPs and films.

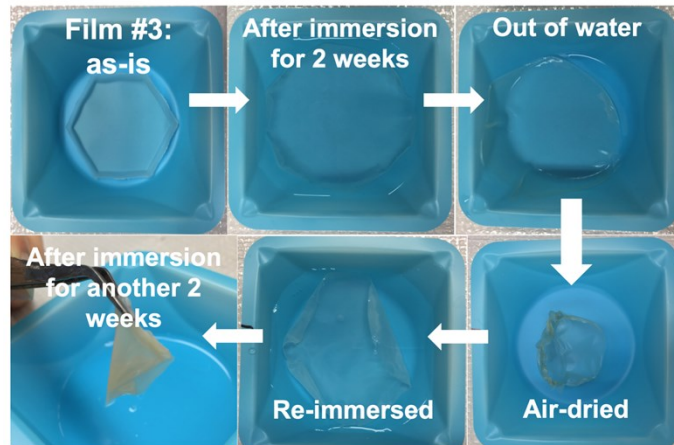


Figure S9. Film #3 was immersed in water for two weeks, air-dried and re-immersed in water for another two weeks.

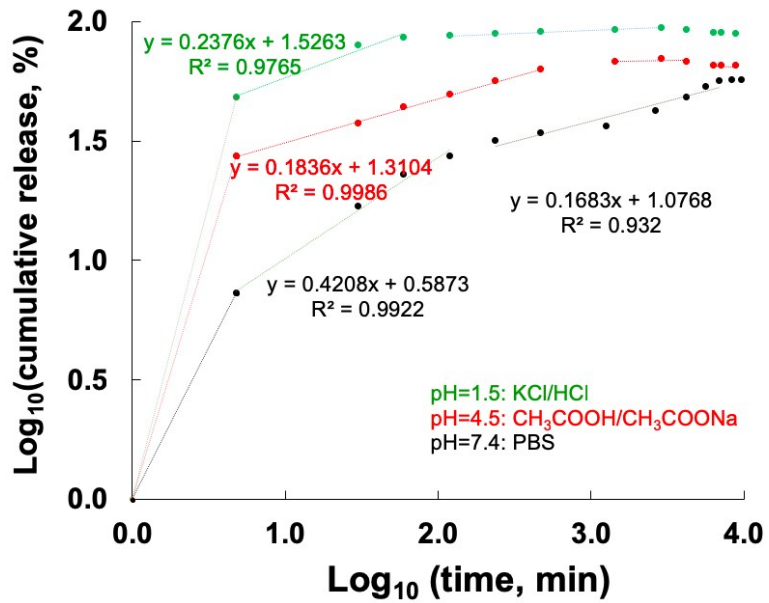


Figure S10. Korsmeyer-Peppas model fitting of MB release in three buffers.

Table S1. XRD parameters of SPs analyzed by peak fitting method

<b>FD SP</b> (CrI: 41.1%)	<b>2<math>\theta</math> (°)</b>	9.0	17.1	19.4	22.3	25.2
	<b>d<sub>hkl</sub> (Å)</b>	9.8	5.2	4.6	4.0	3.5
	<b>Area (%)</b>	<b>10.8</b>	<b>4.2</b>	<b>9.9</b>	<b>8.4</b>	<b>7.7</b>
<b>#1</b> (CrI: 53.6 %)	<b>2<math>\theta</math> (°)</b>	7.7	15.1	17.6	20.3	23.4
	<b>d<sub>hkl</sub> (Å)</b>	11.5	5.9	5.0	4.4	3.8
	<b>Area (%)</b>	<b>9.6</b>	<b>7.1</b>	<b>17.5</b>	<b>12.2</b>	<b>7.2</b>
<b>#2</b> (CrI: 49.8 %)	<b>2<math>\theta</math> (°)</b>	8.9	17.5	19.4	23.7	28.3
	<b>d<sub>hkl</sub> (Å)</b>	9.9	5.1	4.6	3.8	3.2
	<b>Area (%)</b>	<b>1.4</b>	<b>18.1</b>	<b>16.2</b>	<b>8.0</b>	<b>4.1</b>
<b>#3</b> (CrI: 52.3 %)	<b>2<math>\theta</math> (°)</b>	6.7	14.1	17.4	21.6	24.3
	<b>d<sub>hkl</sub> (Å)</b>	13.2	6.3	5.1	4.1	3.7
	<b>Area (%)</b>	<b>1.1</b>	<b>18.1</b>	<b>17.3</b>	<b>7.5</b>	<b>4.9</b>
<b>#4</b> (CrI: 55.6 %)	<b>2<math>\theta</math> (°)</b>	8.6	16.3	19.1	22.6	27.1
	<b>d<sub>hkl</sub> (Å)</b>	10.3	5.4	4.6	3.9	3.3
	<b>Area (%)</b>	<b>0.5</b>	<b>13.3</b>	<b>15.6</b>	<b>14.4</b>	<b>8.9</b>
<b>#5</b> (CrI: 56.2 %)	<b>2<math>\theta</math> (°)</b>	8.8	16.6	19.6	23.5	28.1
	<b>d<sub>hkl</sub> (Å)</b>	10.0	5.3	4.5	3.8	3.2
	<b>Area (%)</b>	<b>1.5</b>	<b>12.4</b>	<b>17.5</b>	<b>14.6</b>	<b>8.1</b>

Table S2. Moisture content (%) of films under different conditions (N=3)

Sample	150 °C, 0 % RH	21 °C, 30-35 % RH	21 °C, 65 % RH	21 °C, 85-90 % RH
<b>#1</b>	-12.0 ± 1.4	0.0	4.9 ± 0.3	40.3 ± 1.1
<b>#2</b>	-9.0 ± 1.1	0.0	1.2 ± 0.9	53.0 ± 1.7
<b>#3</b>	-9.9 ± 0.9	0.0	6.8 ± 0.4	44.0 ± 1.0
<b>#4</b>	-8.0 ± 0.3	0.0	2.9 ± 0.8	51.4 ± 5.4
<b>#5</b>	-9.4 ± 0.6	0.0	4.1 ± 0.5	38.9 ± 1.3

Table S3. S<sub>p</sub> of film #3 and 5 (N=3)

pH	5 min		1 week		2 weeks	
	#3	#5	#3	#5	#3	#5
<b>0</b>	1.50 ± 0.07	1.29 ± 0.03	1.49 ± 0.05	1.27 ± 0.03	1.31 ± 0.01	1.34 ± 0.01
<b>1</b>	2.24 ± 0.05	2.24 ± 0.04	2.28 ± 0.04	2.40 ± 0.03	2.24 ± 0.01	2.23 ± 0.03

<b>3</b>	1.17 ± 0.07	1.52 ± 0.01	1.27 ± 0.04	1.68 ± 0.03	1.17 ± 0.03	1.70 ± 0.01
<b>7</b>	1.38 ± 0.12	1.55 ± 0.09	1.57 ± 0.01	1.46 ± 0.03	1.57 ± 0.02	1.41 ± 0.04
<b>10</b>	1.87 ± 0.12	1.59 ± 0.01	4.13 ± 0.18	1.57 ± 0.04	4.63 ± 0.09	1.74 ± 0.01

Table S4.  $S_p$  of SP/MB films in buffers at 37 °C (N=3)

<b>pH</b>	<b>5 min</b>	<b>30 min</b>	<b>1 h</b>	<b>17 h</b>	<b>48 h</b>	<b>120 h</b>
<b>7.4</b>	1.23 ± 0.02	1.22 ± 0.03	1.16 ± 0.01	1.18 ± 0.04	1.19 ± 0.04	1.20 ± 0.04
<b>4.5</b>	1.18 ± 0.02	1.12 ± 0.05	1.14 ± 0.05	1.08 ± 0.00	1.08 ± 0.03	1.09 ± 0.06
<b>1.5</b>	1.38 ± 0.03	1.47 ± 0.04	1.47 ± 0.08	1.45 ± 0.04	1.49 ± 0.04	1.47 ± 0.05