

Supplementary Information

Molecular detection using aptamer-modified gold nanoparticles with an immobilized DNA brush for the prevention of nonspecific aggregation

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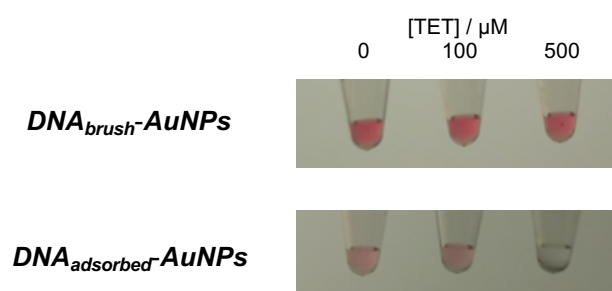


Fig. S1 Prevention of non-specific aggregation by tetracycline using DNA-brush. Tetracycline (TET) was added to $\text{DNA}_{\text{brush}}\text{-AuNPs}$ and $\text{DNA}_{\text{adsorbed}}\text{-AuNPs}$ solutions at the indicated concentrations.

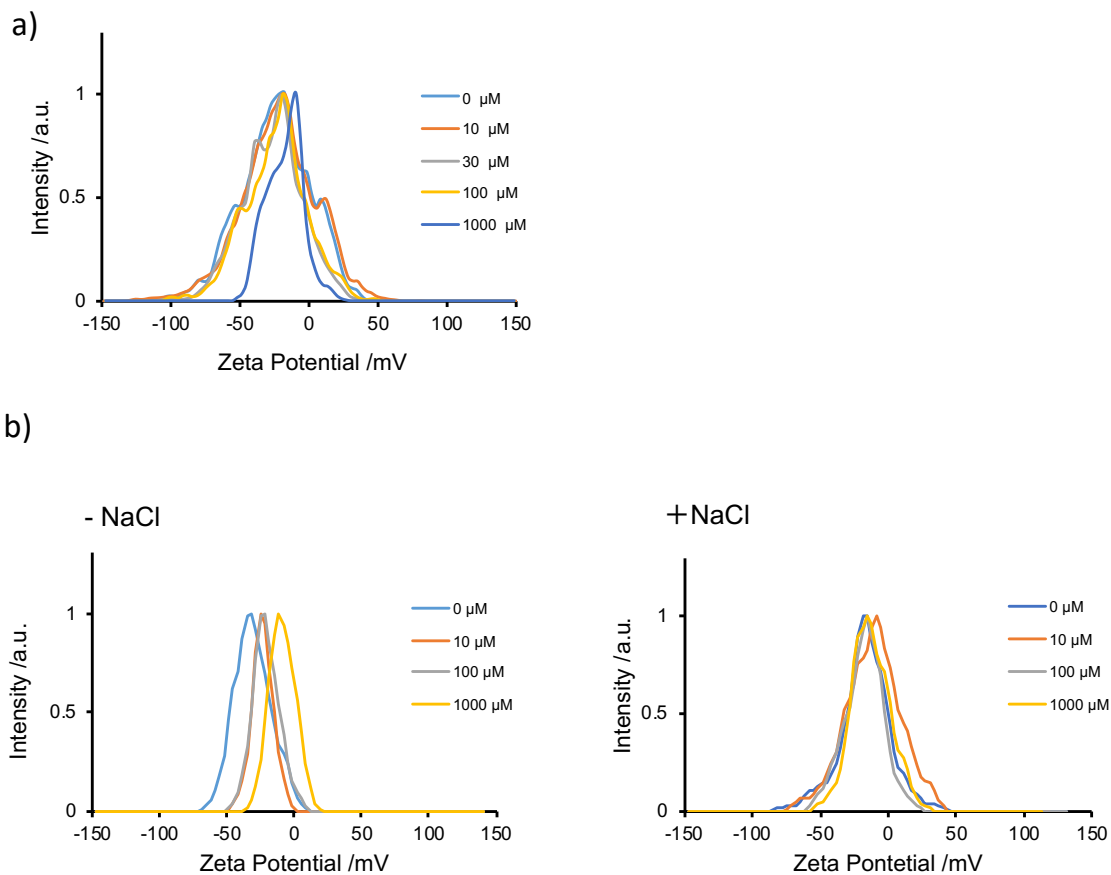


Fig. S2 Zeta potential of DNA_{brush}-AuNPs in the presence of KN. a) Zeta potential of DNA_{brush}-AuNPs in the presence of various concentrations of KN in TN buffer (20 mM Tris-HCl, pH 7.5 and 20 mM NaCl). Zeta-potential of 50 pM DNA_{brush}-AuNPs was measured using Zetasizer-Nano ZS (Malvern, Worcestershire, UK). b) Zeta potential of DNA_{brush}-AuNPs in the presence of various concentrations of KN in a Tris buffer (20 mM Tris-HCl, pH 7.5)(left). The solutions were taken, and 20 mM NaCl was added. Then the zeta potential values were measured (right).

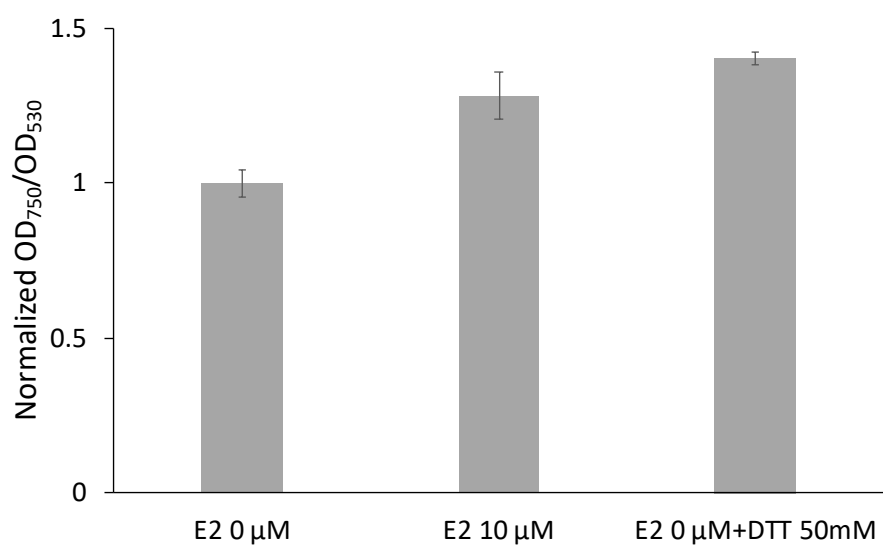


Fig. S3 Effect of DTT on Apt-DNA_{brush}-AuNPs. E2 (10 μM) or DTT (50 mM) was added to Apt-DNA_{brush}-AuNPs, and incubated at room temperature for 30 min. Then the reaction buffer was added and incubated for 60 min to induce AuNP aggregation. The degree of nanoparticle aggregation was evaluated by calculating the OD₇₅₀/OD₅₃₀ ratio. Normalized OD₇₅₀/OD₅₃₀ values are shown.

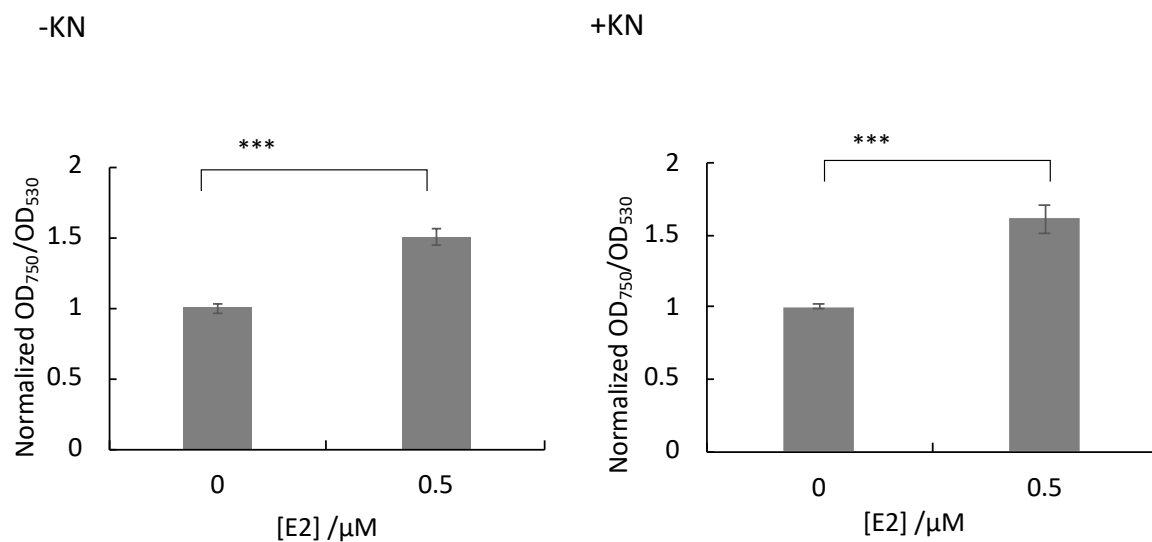


Fig. S4 Detection of 500 nM E2 with DNA_{brush}-Apt-AuNPs in the absence (left) or presence (right) of 30 μM KN was confirmed by another round of experiment. The normalized OD₇₅₀/OD₅₃₀ values were shown. OD ratios of the sample without E2 were defined as 1. The averaged values of three different samples tubes were shown. ***, p<0.005

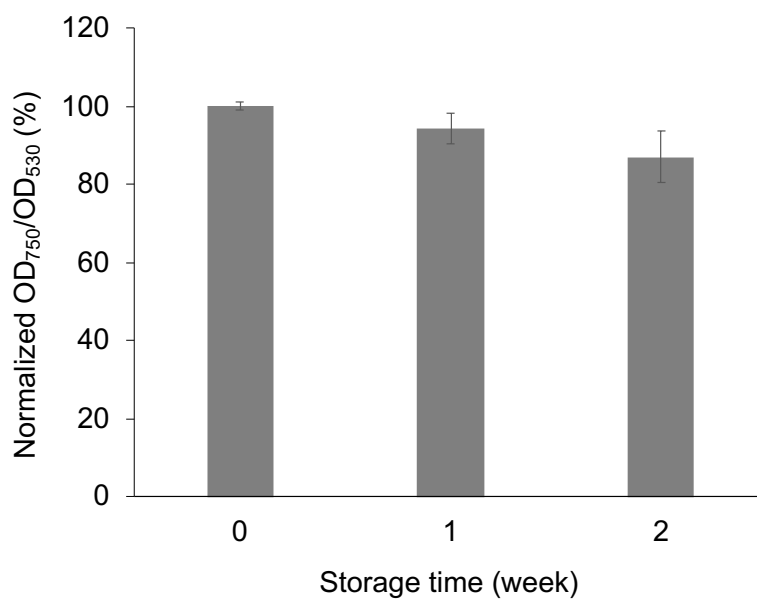


Fig. S5 Storage of DNA_{brush}-AuNPs. DNA_{brush}-AuNPs solution was stored at 4°C. After 1 or 2 weeks, Apt-DNA_{brush}-AuNPs were prepared for E2 detection. E2 (10 μM) was added to Apt-DNA_{brush}-AuNPs, and incubated at room temperature for 30 min. After the addition of the reaction buffer, the samples were incubated for 60 min to induce AuNP aggregation. The degree of nanoparticle aggregation was evaluated by calculating the OD₇₅₀/OD₅₃₀ ratio. Normalized OD₇₅₀/OD₅₃₀ values are shown using the value of 0 week storage with a fresh DNA_{brush}-AuNPs sample as 100%.

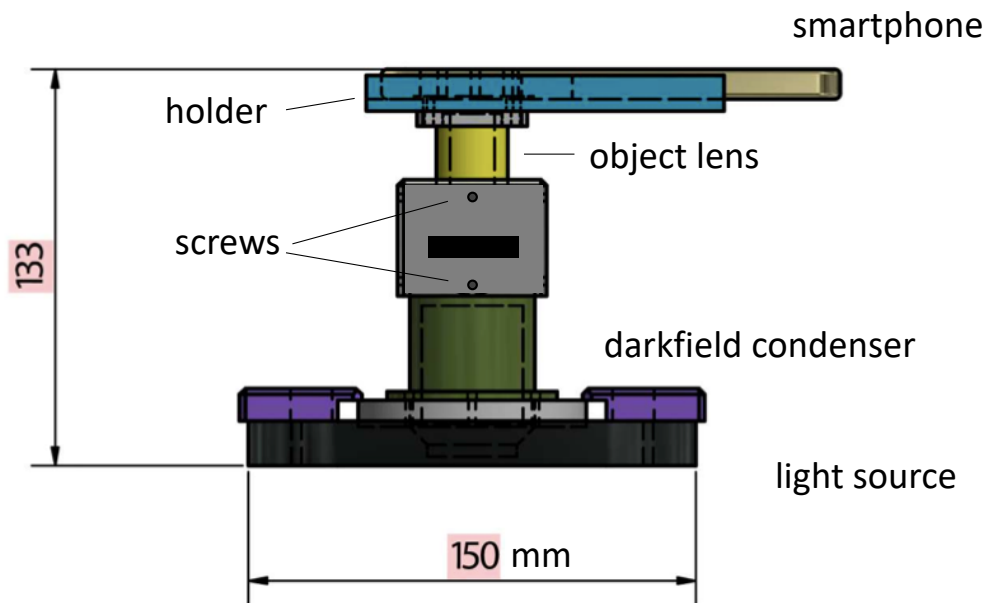


Fig. S6 Set up of a smartphone dark field microscope. The oil dark field condenser (U-DCW, Olympus) was attached to the light source (FL1400R, ANSMANN, Germany). The 10×object lens (CACHN10xIPC/0.25, Olympus) was attached with the smartphone (Huawei P30). The condenser and the lens were connected with the custom-made adapter with screws, by which the position was adjusted. The slide glass was inserted to the hole of the adapter. The cost of the total setup was approximately 1300 USD (condensor, 500; light source, 100; object lens, 200; smartphone, 500).

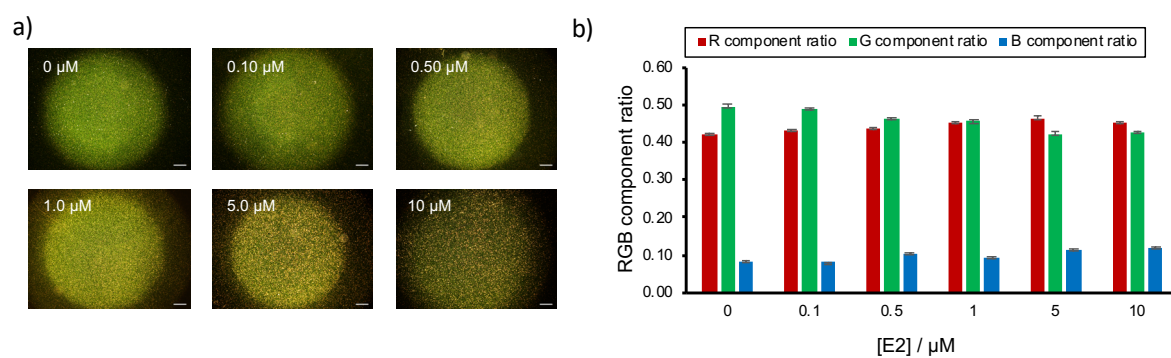


Fig. S7 Characterization of estradiol-induced nanoparticle aggregation by a conventional dark field microscope. a) Visualization of the analysis procedure of gold nanoparticle aggregation caused by different concentrations of estradiol (0, 0.1, 0.5, 1, 5 and 10 μM). For each concentration a smartphone microscope image was obtained. In each picture, three circular areas were subjected to RGB component ratio analysis. b) Results of the RGB component ratio analysis of the discs presented in a).