

SUPPORTING INFORMATION

Hydrogen Sulfide Sensing Using an Aurone-based Fluorescent Probe

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Materials and methods

ACS grade chemicals and solvents used as received without further purification. All the reactions were conducted under an air atmosphere. ^1H and ^{13}C NMR spectra were recorded on a JEOL AS (500 and 300 MHz) NMR instrument and chemical shifts were recorded in ppm with reference to the tetramethyl silane (TMS) chloroform-*d* or DMSO-*d*₆.¹ The following conventions are used for multiplicities: *s*, singlet; *d*, doublet; *t*, triplet; *m*, multiplet; *dd*, doublet of doublet; *dt*, doublet of triplet; *ddd*, doublet of doublet of doublet; *br*, broad. A Cary 630 FT-IR (Agilent Technologies) was used to collect IR spectra. Solid samples were used for collecting the IR-spectra. Fluorescence spectra were collected using a Hitachi F-4500 fluorescence spectrophotometer (Hitachi, Japan) with the slit width for excitation and emission set at 5 nm and photomultiplier voltage set at 700v. Excitation wavelength details are provided in-text as relevant. An HP-8452A Diode Array Spectrophotometer (Agilent Technologies Inc.) was used to record UV-visible absorption spectra. The reactions were monitored by Thin Layer Chromatography (TLC) on aluminum supported silica coated plates (Sorbent Technologies, Inc.). All extracts were concentrated under reduced pressure using a Buchi Rotary Evaporator. Products were purified by flash silica gel (32-63u) column chromatography.

General procedure for the synthesis of 4'-azido benzaldehyde derivatives (4)

In a 3-dram glass vial containing a magnetic stir bar, 1.0 mmol of fluorinated aromatic aldehyde and 1.5 mmol (1.5 equivalent) of sodium azide (NaN_3) were dissolved in 2 mL of dimethylsulfoxide (DMSO) and heated for 2-3 h at 70-90 °C (70 °C for multi-fluorosubstituted) in a sand bath. After the completion of the reaction, it was diluted with DI water and extracted with ethylacetate ($\times 4$). To remove residual DMSO the organic fraction was further washed with DI water and brine followed by drying over anhydrous MgSO_4 . The organic fraction was concentrated *in vacuo* and purified by flash column chromatography using 2-10% ethyl acetate: hexane to obtain the desired 4'-azido benzaldehydes.

General procedure for the synthesis of aurones (6a-6g)

In a 3-dram glass vial, benzofuranone (1 equivalent) was dissolved in 5 mL of glacial acetic acid containing a magnetic stir bar. To this solution 4'-azidobenzaldehyde (1.5 equivalent) and 2-3 drops (0.2 mL) of concentrated HCl were added and stirred at room temperature for up to 3 hours. In most

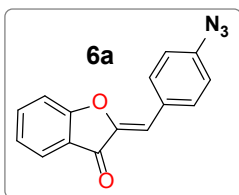
cases the reaction afforded a precipitate, normally after 30 min, indicating completion of the reaction. After the reaction was complete, it was poured into ice cold DI water. The precipitate obtained was filtered and the residue was washed multiple times with water and allowed to air dry. No further purification was required. For some of the auronones which did not precipitate out efficiently, the water diluted reaction mixture was neutralized with saturated NaHCO₃ and extracted with ethyl acetate, concentrated under reduced pressure using a Rotary Evaporator and purified by flash column chromatography.

General procedure for the synthesis of amino derivatives (7a-7g)

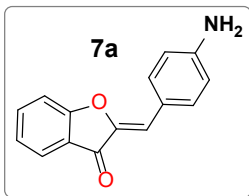
For the synthesis of the corresponding amines (**7a-7g**) of the probe candidates, 1 equivalent of the aurone (**6a-6g**) was mixed with an excess of NaHS (4 equivalents) in a 1-dram glass vial containing a magnetic stir bar. To the mixture, 1.5 mL of MeCN with a few drops of DI H₂O was added and stirred for 30 minutes (acetone can also be used as a solvent). The solution turned bright red. Once the reaction was complete based on the TLC, the mixture was concentrated under vacuum and subjected to flash column chromatography (eluted with 5% MeOH: DCM).

Characterization data

(Z)-2-(4-azidobenzylidene)benzofuran-3(2H)-one (6a): Reaction scale (1.80 mmol), Yield: 70% (331 mg); yellow solid, mp 128-129 °C. ¹H NMR (500 MHz, CDCl₃) δ 7.92 (d, *J* = 8.5 Hz, 2H), 7.81 (d, *J* = 7.5 Hz, 1H), 7.66 (t, *J* = 7.5 Hz, 1H), 7.33 (d, *J* = 8.0 Hz, 1H), 7.23 (t, *J* = 7.4 Hz, 1H), 7.10 (d, *J* = 8.5 Hz, 2H), 6.85 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 184.7, 166.1, 146.8, 141.6, 137.0, 133.2, 129.3, 124.8, 123.7, 121.8, 119.7, 113.0, 112.2. IR (neat) 2121, 1704, 1655, 1596, 1508, 1302, 1186, 1108, 829, 750 cm⁻¹.

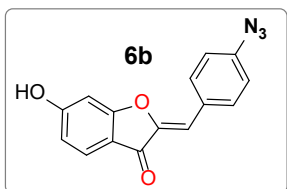


(Z)-2-(4-aminobenzylidene)benzofuran-3(2H)-one (7a): Reaction scale (0.114 mmol), Yield: 94% (25.4 mg); orange red solid, mp 173-175 °C. ¹H NMR (500 MHz, CDCl₃) δ 7.80 (d, *J* = 8.5 Hz, 1H), 7.77 (d, *J* = 8.5 Hz, 2H), 7.61 (ddd, *J* = 8.5, 7.0, 1.0 Hz, 1H), 7.31 (d, *J* = 8.0 Hz, 1H), 7.19 (t, *J* = 7.5 Hz, 1H), 6.87 (s, 1H), 6.72 (d, *J* = 8.5 Hz, 2H), 4.09 (s, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 184.4, 165.6, 148.7, 145.4, 136.3, 133.9, 124.5, 123.2, 122.5, 122.3, 115.1, 114.8, 112.9. IR (neat)



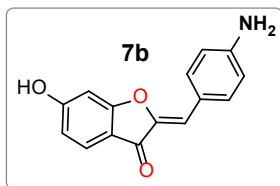
3439, 3312, 3216, 1683, 1577, 1518, 1488, 1302, 1182, 1100, 832, 754 cm^{-1} . HRMS (EI): m/z [$M + H$] calcd for $\text{C}_{15}\text{H}_{12}\text{NO}_2$ 238.0868, found 238.0871.

(Z)-2-(4-azidobenzylidene)-6-hydroxybenzofuran-3(2H)-one (6b): Reaction scale (0.114 mmol),



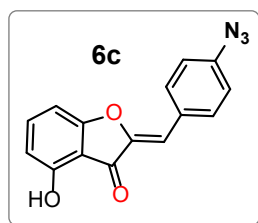
Yield: 94% (25.4 mg); faint orange red solid, mp 194-196 °C. ^1H NMR (500 MHz, DMSO- D_6) δ 11.23 (s, 1H), 7.99 (d, $J = 8.5$ Hz, 2H), 7.63 (d, $J = 8.5$ Hz, 1H), 7.24 (d, $J = 8.5$ Hz, 2H), 6.81 (s, 1H), 6.79 (d, $J = 2.0$ Hz, 1H), 6.72 (dd, $J = 8.5, 2.0$ Hz, 1H). ^{13}C NMR (125 MHz, DMSO- D_6) δ 181.2, 167.8, 166.5, 147.1, 140.5, 132.7, 128.9, 125.9, 119.6, 113.0, 112.8, 109.5, 98.6. IR (neat) 3074-2600 (br), 2117, 1681, 1577, 1506, 1458, 1294, 1112, 825 cm^{-1} .

(Z)-2-(4-aminobenzylidene)-6-hydroxybenzofuran-3(2H)-one (7b): Reaction scale (0.11 mmol),



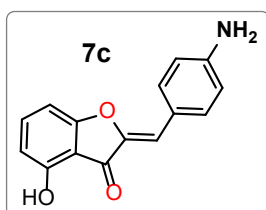
Yield: 92% (25.6 mg); orange red solid, mp > 260 °C. ^1H NMR (500 MHz, DMSO- D_6) δ 11.00 (s, 1H), 7.66 (d, $J = 8.5$ Hz, 2H), 7.57 (d, $J = 8.5$ Hz, 1H), 6.77 (d, $J = 2.0$ Hz, 1H), 6.69 (dd, $J = 8.5, 2.0$ Hz, 1H), 6.65 (s, 1H), 6.64 (d, $J = 9.0$ Hz, 3H), 5.94 (s, 2H). ^{13}C NMR (125 MHz, DMSO- D_6) δ 180.7, 166.9, 165.6, 151.1, 144.5, 133.3, 125.3, 119.0, 113.8, 113.6, 112.9, 112.5, 98.4. IR (neat) 3461, 3361, 3300-2500 (br), 1666, 1629, 1551, 1454, 1287, 1108, 821 cm^{-1} . HRMS (EI): m/z [$M + H$] calcd for $\text{C}_{15}\text{H}_{12}\text{NO}_3$ 254.0817, found 254.0813.

(Z)-2-(4-azidobenzylidene)-4-hydroxybenzofuran-3(2H)-one (6c): Reaction scale (0.30 mmol),



Yield: 90% (75 mg); red solid, mp 179-181 °C. ^1H NMR (500 MHz, DMSO- D_6) δ 11.16 (s, 1H), 7.98 (d, $J = 8.5$ Hz, 2H), 7.54 (t, $J = 8.0$ Hz, 1H), 7.23 (d, $J = 8.5$ Hz, 2H), 6.85 (d, $J = 8.0$ Hz, 1H), 6.77 (s, 1H), 6.64 (d, $J = 8.5$ Hz, 1H). ^{13}C NMR (76 MHz, DMSO- D_6) δ 181.2, 165.9, 157.2, 146.31, 140.5, 138.7, 132.7, 129.1, 119.7, 110.7, 109.2, 109.1, 102.4. IR (neat) 3283, 2125, 2028, 1693, 1652, 1592, 1458, 1316, 1134, 769 cm^{-1} .

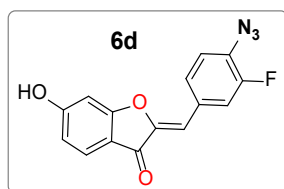
(Z)-2-(4-aminobenzylidene)-4-hydroxybenzofuran-3(2H)-one (7c): Reaction scale (24 mg, 0.083



mmol), Yield: 92% (20 mg); red solid, mp >260 °C. ^1H NMR (300 MHz, DMSO- D_6) δ 7.54 (d, $J = 8.5$ Hz, 2H), 6.99 (dd, $J = 8.5, 7.5$ Hz, 1H), 6.60 (d, $J = 8.5$ Hz, 2H), 6.25 (s, 1H), 5.86 (d, $J = 9.0$ Hz, 1H), 5.79 (d, $J = 7.5$ Hz, 1H), 5.63 (s, 2H). ^{13}C NMR (75 MHz, DMSO- D_6) δ 182.1, 173.9,

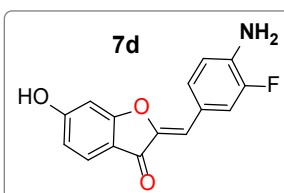
165.9, 149.7, 145.9, 136.4, 131.9, 120.4, 117.7, 113.8, 110.7, 107.5, 89.3. IR (neat) 3394-3275 (br), 1674, 1592, 1561, 1477, 1376, 1235, 1127, 1004, 803 cm^{-1} . HRMS (EI): m/z [M + H] calcd for $\text{C}_{15}\text{H}_{12}\text{NO}_3$ 254.0817, found 254.0819.

(Z)-2-(4-azido-3-fluorobenzylidene)-6-hydroxybenzofuran-3(2H)-one (6d): Reaction scale (0.27



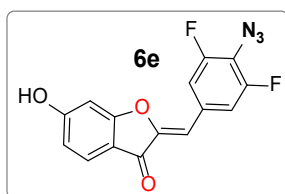
mmol), Yield: 76% (61 mg); red solid, mp 156 °C (decompose). ^1H NMR (500 MHz, DMSO- D_6) δ 11.24 (s, 1H), 7.80 (d, $J = 12.6$ Hz, 1H), 7.75 (d, $J = 8.6$ Hz, 1H), 7.59 (d, $J = 8.3$ Hz, 1H), 7.34 (t, $J = 8.6$ Hz, 1H), 6.78 (d, $J = 1.8$ Hz, 1H), 6.73 (s, 1H), 6.71 (dd, $J = 8.3, 1.8$ Hz, 1H). ^{13}C NMR (75 MHz, DMSO- D_6) δ 181.2, 167.9, 166.7, 153.5 (d, $^1J_{\text{C-F}} = 247.3$ Hz), 147.6, 130.3 (d, $^2J_{\text{C-F}} = 7.5$ Hz), 128.3 (d, $^3J_{\text{C-F}} = 3.5$ Hz), 128.2, 126.0, 121.8, 118.30 (d, $^2J_{\text{C-F}} = 20.1$ Hz), 113.8, 112.6, 108.43 (d, $^3J_{\text{C-F}} = 2.4$ Hz), 98.7. IR (neat) 3074-2653 (br), 2136, 2106, 1685, 1573, 1462, 1298, 1119, 851 cm^{-1} .

(Z)-2-(4-amino-3-fluorobenzylidene)-6-hydroxybenzofuran-3(2H)-one (7d): Reaction scale



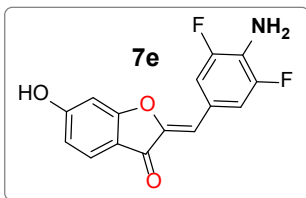
(21.5 mg, 0.069 mmol), Yield: 98% (18.4 mg); orange solid, mp >260 °C. ^1H NMR (500 MHz, DMSO- D_6) δ 7.64 (d, $J = 12.0$ Hz, 1H), 7.58 (dd, $J = 8.0, 3.0$ Hz, 1H), 7.50 (d, $J = 8.0$ Hz, 1H), 6.86 – 6.76 (m, 2H), 6.70–6.68 (m, 2H), 5.96 (s, 2H). ^{13}C NMR (125 MHz, DMSO- D_6) δ 180.8, 167.2, 166.0, 149.8 (d, $^1J_{\text{C-F}} = 237.2$ Hz), 145.3, 138.9 (d, $^2J_{\text{C-F}} = 12.8$ Hz), 129.1, 125.5, 119.5 (d, $^3J_{\text{C-F}} = 7.0$ Hz), 117.25 (d, $^2J_{\text{C-F}} = 18.8$ Hz), 115.7, 113.2, 112.8, 111.5, 98.5. IR (neat) 3478, 3372, 3041-2586 (br), 1674, 1566, 1521, 1454, 1294, 1112, 999 cm^{-1} . HRMS (EI): m/z [M + H] calcd for $\text{C}_{15}\text{H}_{11}\text{FNO}_3$ 272.07230, found 272.07228.

(Z)-2-(4-azido-3, 5-difluorobenzylidene)-6-hydroxybenzofuran-3(2H)-one (6e): Reaction scale



(0.33 mmol), Yield: 76% (79.3 mg); yellow solid, mp 165 °C (decompose). ^1H NMR (500 MHz, DMSO- D_6) δ 11.30 (s, 1H), 7.72 (d, $J = 9.5$ Hz, 2H), 7.61 (d, $J = 8.5$ Hz, 1H), 6.83 (s, 1H), 6.74 (s, 1H), 6.72 (dd, $J = 8.5, 2.0$ Hz, 1H). ^{13}C NMR (125 MHz, DMSO- D_6) δ 181.0, 167.9, 166.8, 154.7 (d, $^1J_{\text{C-F}} = 247.5$ Hz), 148.0, 129.6 (t, $^2J_{\text{C-F}} = 10.1$ Hz), 125.9, 116.0, 114.3 (d, $^2J_{\text{C-F}} = 19.6$ Hz), 113.2, 112.3, 107.3, 98.8. IR (neat) 3078, 2117, 1685, 1577, 1506, 1458, 1279, 1138, 1112, 1045, 832 cm^{-1} .

(Z)-2-(4-amino-3, 5-difluorobenzylidene)-6-hydroxybenzofuran-3(2H)-one (7e): Reaction scale

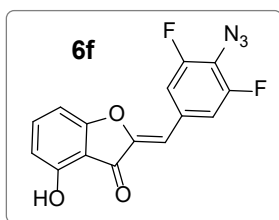


(31 mg, 0.098 mmol), Yield: 80% (22.7 mg); orange solid, mp >260 °C.

¹H NMR (300 MHz, DMSO-D₆) δ 7.62 – 7.49 (m, 3H), 6.81 (d, *J* = 2.0 Hz, 1H), 6.71 (s, 1H), 6.69 (dd, *J* = 7.5, 2.5 Hz, 1H), 6.00 (s, 1H). ¹³C NMR (75 MHz, DMSO-D₆) δ 180.9, 167.4, 150.4 (dd, *J*_{C-F} = 238.8, 10.0 Hz), 146.0, 128.0 (t, *J*_{C-F} = 16.5 Hz), 125.7, 117.85 (t, *J*_{C-F} = 9.1 Hz),

113.95 (d, *J*_{C-F} = 13.3 Hz), 113.9, 113.8, 113.0, 110.4, 98.7. IR (neat) 3469, 3368, 1678, 1581, 1532, 1454, 1324, 1287, 1115, 821, 769 cm⁻¹. HRMS (EI): *m/z* [M + H] calcd for C₁₅H₁₁FNO₃ 272.07230, found 272.08226.

(Z)-2-(4-azido-3,5-difluorobenzylidene)-4-hydroxybenzofuran-3(2H)-one (6f): Reaction scale

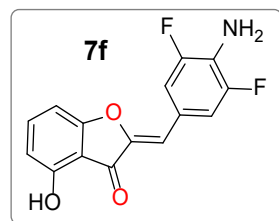


(0.36 mmol), Yield: 70% (80 mg); yellow solid, mp 165-170 °C. ¹H NMR

(300 MHz, DMSO-D₆) δ 11.27 (s, 1H), 7.75 (d, *J* = 10.0 Hz, 2H), 7.56 (t, *J* = 8.0 Hz, 1H), 6.92 (d, *J* = 8.0 Hz, 1H), 6.76 (s, 1H), 6.65 (d, *J* = 8.0 Hz, 1H). ¹³C NMR (125 MHz, DMSO-D₆) δ 180.9, 165.8, 157.2, 154.9 (d, ¹*J*_{C-F} = 245.9, 4.6 Hz), 147.2, 138.9, 129.68 (t, ³*J*_{C-F} = 9.8 Hz), 117.32 (t, ²*J*_{C-F}

= 14.0 Hz), 114.22 (dd, ²*J*_{C-F} = 19.2, 4.7 Hz), 111.0, 108.6, 106.8, 102.5. IR (neat) 3379, 2121, 1696, 1607, 1495, 1339, 1305, 1179, 1041, 799 cm⁻¹.

(Z)-2-(4-amino-3,5-difluorobenzylidene)-4-hydroxybenzofuran-3(2H)-one (7f): Reaction scale

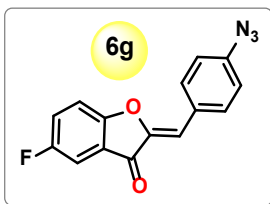


(13 mg, 0.041 mmol), Yield: 84% (10 mg); red solid, mp 222-225 °C. ¹H

NMR (500 MHz, DMSO-D₆) δ 7.44 (dd, *J* = 8.0, 2.1 Hz, 2H), 7.08 (t, *J* = 8.0 Hz, 1H), 6.33 (s, 1H), 6.00 (d, *J* = 8.0 Hz, 2H), 5.74 (s, 2H). ¹³C NMR (125 MHz, DMSO-D₆) δ 181.2, 165.8, 150.6 (dd, ¹*J*_{C-F} = 238.4, 9.8 Hz), 149.6 (d, *J*_{C-F} = 10.4), 147.4, 136.6, 126.3 (t, ²*J*_{C-F} = 11.2 Hz), 119.5, 112.7

(d, ²*J*_{C-F} = 16.7 Hz), 109.9, 104.9, 104.8, 104.7. IR (neat) 3346, 1689, 1596, 1566, 1566, 1480, 1354, 1235, 1115, 1000, 803 cm⁻¹. HRMS (EI): *m/z* [M + H] calcd for C₁₅H₁₀F₂NO₃ 290.06288, found 290.06290.

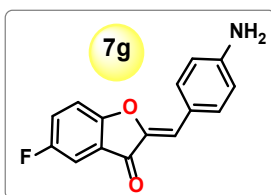
(Z)-2-(4-azidobenzylidene)-5-fluorobenzofuran-3(2H)-one (6g): Reaction scale (0.40 mmol),



Yield: 80% (90 mg); yellow solid, mp 137-138 °C. ^1H NMR (500 MHz, CDCl_3) δ 7.89 (d, $J = 8.5$ Hz, 2H), 7.45 (dd, $J = 7.0, 3.0$ Hz, 1H), 7.38 (td, $J = 8.5, 3.0$ Hz, 1H), 7.29 (dd, $J = 9.0, 3.5$ Hz, 1H), 7.10 (d, $J = 9.0$ Hz, 2H), 6.85 (s, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 183.9, 162.1, 159.02 (d, $^1J_{\text{C-F}} = 244.7$ Hz), 147.4, 141.9, 133.3, 129.0, 124.4 (d, $^2J_{\text{C-F}} = 26.1$ Hz),

122.5 (d, $^3J_{\text{C-F}} = 8.0$ Hz), 119.7, 114.21 (d, $^3J_{\text{C-F}} = 7.8$ Hz), 113.08, 110.36 (d, $^2J_{\text{C-F}} = 24.3$ Hz). IR (neat) 2128, 1707, 1655, 1598, 1484, 1305, 1268, 1190, 892, 832 cm^{-1} .

(Z)-2-(4-aminobenzylidene)-5-fluorobenzofuran-3(2H)-one (7g): Reaction scale (18 mg, 0.064



mmol), Yield: 73% (12 mg); red solid, mp 178-180 °C. ^1H NMR (500 MHz, DMSO-D_6) δ 7.74 (d, $J = 8.5$ Hz, 2H), 7.65 – 7.58 (m, 2H), 7.57 (dd, $J = 7.0, 2.4$ Hz, 1H), 6.88 (s, 1H), 6.66 (d, $J = 8.5$ Hz, 2H), 6.16 (s, 2H). ^{13}C NMR (76 MHz, DMSO-D_6) δ 181.64, 160.5, 158.18 (d, $^1J_{\text{C-F}} =$

240.9 Hz), 152.1, 144.3, 134.2, 123.7 (d, $^2J_{\text{C-F}} = 26.0$ Hz), 122.6 (d, $^3J_{\text{C-F}} = 8.0$ Hz), 118.7, 116.2, 114.72 (d, $^3J_{\text{C-F}} = 8.1$ Hz), 113.9, 109.48 (d, $^2J_{\text{C-F}} = 24.2$ Hz). IR (neat) 3420, 3320, 3216, 1689, 1633, 1562, 1514, 1480, 1272, 1156, 817, 762 cm^{-1} . HRMS (EI): m/z [M + H] calcd for $\text{C}_{15}\text{H}_{11}\text{FNO}_2$ 256.0774, observed 256.0071.

References

1. Gottlieb, H. E.; Kotlyar, V.; Nudelman, A. NMR Chemical Shifts of Common Laboratory Solvents as Trace Impurities. *J. Org. Chem.* **1997**, 62, 7512-7515.
2. Geissman, A. T.; Harborne, J. B. Anthochlor pigment. X. Aureusin and cernuoside. *J. Am. Chem. Soc.* **1955**, 77, 4622-4624.

UV-visible absorption and emission spectra

The 10 mM stock solution of all the 4'-azidoaurones **6a-g** and corresponding amines **7a-g** were prepared in DMSO. Unless otherwise stated 1X, pH = 7.4 PBS was used for the preparation of samples. All the UV-absorption and emission spectra were collected using a 50 μ M solution of the compounds in different solvent systems (90% DMSO/10% PBS, 5% DMSO in PBS, and 0.5% DMSO in PBS) in the presence or absence of CTAB.

Sample preparation for UV-Vis absorption

- a) **50 μ M solution of **6g** and **7g** in 0.5% DMSO in PBS**
15 μ L 10 mM stock solution of **6g** or **7g** in DMSO was diluted with 2985 μ L of PBS (1x pH = 7.4) in a 3 mL cuvette.
- b) **50 μ M solution of **6g** and **7g** in 5% DMSO in PBS**
150 μ L 1.0 mM stock solution of **6g** or **7g** in DMSO was diluted with 2850 μ L of PBS (1x pH = 7.4) in a 3 mL cuvette.
- c) **50 μ M solution of **6g** and **7g** in 90% DMSO in PBS**
150 μ L 1.0 mM stock solution of **6g** or **7g** in DMSO was diluted with 2550 μ L DMSO and 300 PBS (1x pH = 7.4) in a 3 mL cuvette.
- d) **50 μ M solution of **6g** v **7g** in 0.5% DMSO in PBS and 1mM CTAB (in PBS)**
To a 15 μ L 10 mM **6g** and **7g**, 600 μ L of CTAB (5 mM stock solution in PBS) and 2385 μ L of PBS in a 3 mL cuvette.

Similarly, for collecting the emission spectra the 50 μ M solutions of **6g** and **7g** in a 1.5 mL cuvette were excited at their respective absorption maxima (λ_{max}) obtained for the specific solvent condition e.g., **6g** and **7g** were excited at 405 and 460 nm respectively in 0.5% v/v DMSO in PBS with 1mM CTAB.

Effect of pH on spectral properties of **6g and **7g****

The effect of pH on the absorption and fluorescence behavior of probe **6g** and the corresponding amine **7g** in the presence or absence of 1 mM CTAB was studied. This study was done in 0.5% v/v DMSO in PBS (1X) solvent. For the study, five different PBS (1X) solutions of pH = 3, 4.8, 7.5, 9.15, and 11 were prepared by adjusting pH using a pH meter. Five different 10 mM stock solution of CTAB (pH = 3, 4.8, 7.5, 9.15, and 11) were prepared using the corresponding PBS (1X, pH = 3,

4.8, 7.5, 9.15, and 11) solutions. The UV-Vis absorption spectra of **6g** and **7g** were collected with and without the inclusion of 1mM CTAB in 0.5% v/v DMSO in PBS (1X) of the specific pH as described above.

Sample preparation (example)

1. 50 μ M solution of **6g** and **7g** in 0.5% DMSO in PBS (pH = 3) with 1mM CTAB

To a 15 μ L 10 mM stock solution of **6g** and **7g**, 600 μ L of CTAB (5 mM stock solution in PBS, pH =3) and 2385 μ L of PBS (pH = 3) were mixed in a 3 mL cuvette and excited at the corresponding λ_{max} to collect the emission spectra.

Similarly, the emission spectra of **6g** and **7g** at specific pH were collected by preparing and exciting their 50 μ M solution in a 1.5 mL cuvette at their corresponding absorption maxima (λ_{max}).

Determination of detection limit

Using the stock solutions of **7g** the emission spectra of its various concentrations (0.1 - 50 μ M) were collected in either 0.5% v/v DMSO in PBS/ 1 mM CTAB or 90% v/v DMSO/ 10% v/v PBS (1X, pH 7.4) solvent systems. Each solution of **7g** (0.1, 0.5, 1, 2, 5, 10, 15, 20, 25, 30, 40 and 50 μ M) prepared in a particular solvent system in a 1.5 mL cuvette was excited at the 465 nm. For each concentration, at least three independent trials were performed. The corresponding emission intensity observed at 560 nm was then plotted as a function of [**7g**] to obtain the linear range of detection. This generated the calibration curve for each solvent condition revealing the useful working range of **7g**.

Sample preparation in 0.5% DMSO in PBS (1X, pH = 7.4) with 1mM CTAB)

a) 0.1 μ M **7g** in 0.5% DMSO in PBS with 1mM CTAB

In a cuvette containing 3.0 μ L of **7g** (50 μ M stock in DMSO), 4.5 μ L DMSO, 300 μ L CTAB (5 mM in PBS) were added and diluted to the final volume of 1500 μ L with PBS (1192.5 μ L, 1X, pH = 7.4)

b) 0.5 μ M **7g** in 0.5% DMSO in PBS with 1mM CTAB

In a cuvette containing 7.5 μ L of **7g** (100 μ M stock in DMSO), 300 μ L CTAB (5 mM in PBS) were added and diluted to the final volume of 1500 μ L with PBS (1192.5 μ L)

c) 1.0 μ M **7g** in 0.5% DMSO in PBS with 1mM CTAB

In a cuvette containing 7.5 μ L of **7g** (200 μ M stock in DMSO), 300 μ L CTAB (5 mM in PBS) were added and diluted to the final volume of 1500 μ L with PBS (1192.5 μ L)

- d) 2.0 μ M 7g in 0.5% DMSO in PBS with 1mM CTAB
In a cuvette containing 3.0 μ L of 7g (1 mM stock in DMSO), 4.5 μ L DMSO, 300 μ L CTAB (5 mM in PBS) were added and diluted to the final volume of 1500 μ L with PBS (1192.5 μ L)
- e) 5.0 μ M 7g in 0.5% DMSO in PBS with 1mM CTAB
In a cuvette containing 7.5 μ L of 7g (1 mM stock in DMSO), 300 μ L CTAB (5 mM in PBS) were added and diluted to the final volume of 1500 μ L with PBS (1192.5 μ L)
- f) 10 μ M 7g in 0.5% DMSO in PBS with 1mM CTAB
In a cuvette containing 3.0 μ L of 7g (5 mM stock in DMSO), 4.5 μ L DMSO, 300 μ L CTAB (5 mM in PBS) were added and diluted to the final volume of 1500 μ L with PBS (1192.5 μ L)
- g) 15 μ M 7g in 0.5% DMSO in PBS with 1mM CTAB
In a cuvette containing 4.5 μ L of 7g (5 mM stock in DMSO), 3.0 μ L DMSO, 300 μ L CTAB (5 mM in PBS) were added and diluted to the final volume of 1500 μ L with PBS (1192.5 μ L)
- h) 20 μ M 7g in 0.5% DMSO in PBS with 1mM CTAB
In a cuvette containing 3.0 μ L of 7g (10 mM stock in DMSO), 4.5 μ L DMSO, 300 μ L CTAB (5 mM in PBS) were added and diluted to the final volume of 1500 μ L with PBS (1192.5 μ L)
- i) 25 μ M 7g in 0.5% DMSO in PBS with 1mM CTAB
In a cuvette containing 7.5 μ L of 7g (5 mM stock in DMSO), 300 μ L CTAB (5 mM in PBS) were added and diluted to the final volume of 1500 μ L with PBS (1192.5 μ L).
- j) 30 μ M 7g in 0.5% DMSO in PBS with 1mM CTAB
In a cuvette containing 4.5 μ L of 7g (10 mM stock in DMSO), 3.0 μ L DMSO, 300 μ L CTAB (5 mM in PBS) were added and diluted to the final volume of 1500 μ L with PBS (1192.5 μ L).
- k) 40 μ M 7g in 0.5% DMSO in PBS with 1mM CTAB
In a cuvette containing 6.0 μ L of 7g (10 mM stock in DMSO), 1.5 μ L DMSO, 300 μ L CTAB (5 mM in PBS) were added and diluted to the final volume of 1500 μ L with PBS (1192.5 μ L)

1) 50 μ M **7g** in 0.5% DMSO in PBS with 1mM CTAB

In a cuvette containing 7.5 μ L of **7g** (10 mM stock in DMSO), 300 μ L CTAB (5 mM in PBS) were added and diluted to the final volume of 1500 μ L with PBS (1192.5 μ L).

Reaction kinetics study

The time course reaction kinetics for the reduction of the azide (**6g**) to amine (**7g**) was performed by incubating 20 μ M **6g** with 100 μ M NaHS in either 0.5% v/v DMSO in PBS (1X, pH = 7.4) +/- 1 mM CTAB or 90% v/v DMSO/10% v/v PBS (1X, pH = 7.4) solvent systems. Samples were excited at 465 nm and emission spectra spanning from 480 to 700 nm were collected at 2-minute intervals.

Sample preparation (solvent system 0.5% v/v DMSO in PBS (1X, pH = 7.4)/ 1 mM CTAB):

In a cuvette, 3.0 μ L of **6g** (10 mM stock in DMSO) was added followed by 4.5 μ L DMSO, 300 μ L CTAB (5 mM in PBS), and 1042.5 μ L PBS (1X, pH = 7.4). To this solution, 150 μ L of NaHS (1 mM stock solution in PBS) was mixed and placed in the fluorescence spectrophotometer to monitor the progress of the reaction by exciting the sample at 465 nm. The emission spectra corresponding to the amine were collected every 2-minutes for 60 minutes. At least 3 trials were performed for each experiment. For the reaction that employed 90% v/v DMSO/10% v/v PBS (1X, pH = 7.4) solvent system, the emission spectra were collected every 2-minutes for 30 minutes

Selectivity study

To evaluate the reaction specificity, a series of fluorescence time courses were collected using the same methodology as described for the kinetics study. In place of NaHS, three potential sulfide-donors were substituted: L-cysteine, reduced glutathione, and 2-mercaptoethanol. All emission spectra reflect the reaction of 100 μ M sulfide-donor with 20 μ M **6g** in 0.5 % v/v DMSO, 99.5 % v/v PBS, and 1 mM CTAB. At least 3 trials were performed for each experiment.

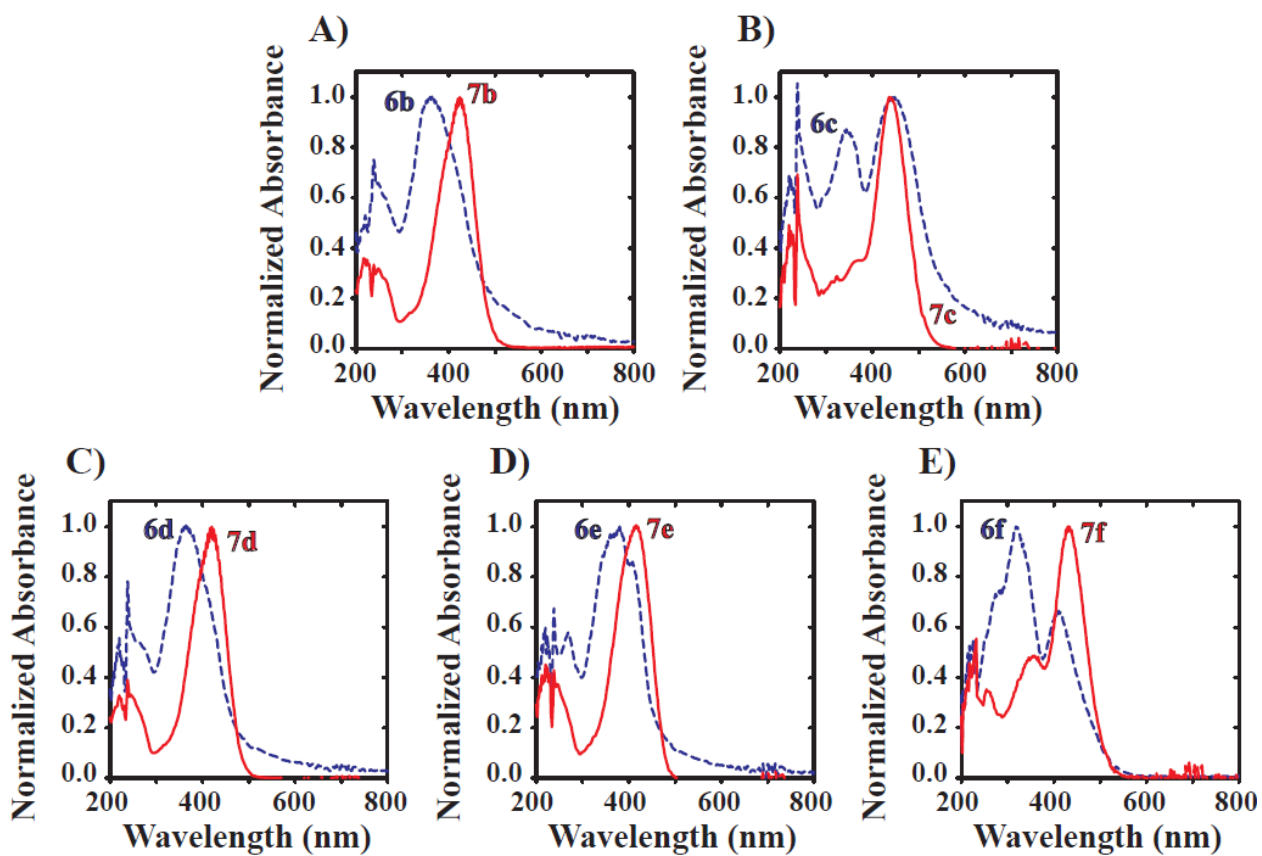


Figure S1. UV-visible Spectra Illustrating Spectral Overlap. Samples (50 μM) were prepared in 5% DMSO in PBS (v/v) for the study.

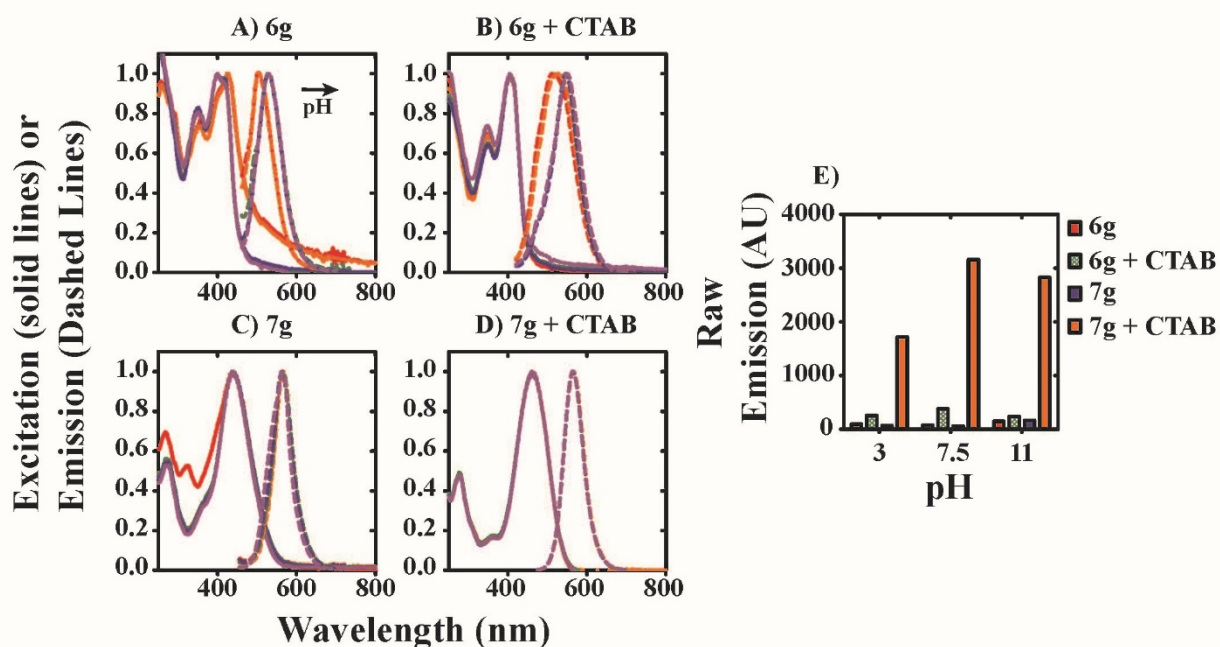


Figure S2. Effects of pH on 6g/7g Spectral Properties. Excitation spectra collected in 0.5 % DMSO/PBS buffer with pH adjusted to 3.0, 4.8, 7.5, 9.2, and 11.0 are represented as solid red, orange, green, blue, and purple lines, respectively. Corresponding emission spectra collected at pH = 3.0, 4.8, 7.5, 9.2, and 11.0 are represented as dashed red, orange, green, blue, and purple lines, respectively. Excitation/emission spectra were collected for (A) **6g** in the absence of CTAB, (B) **6g** in the presence of 1 mM CTAB, (C) **7g** in the absence of CTAB, and (D) **7g** in the presence of 1 mM CTAB. **6g** emission spectra were collected using excitation wavelengths of 425, 425, 405, 405, and 405 nm for conditions with pH = 3.0, 4.8, 7.5, 9.2, and 11.0, respectively, in the absence of CTAB. In the presence of CTAB, **6g** emission spectra were collected using an excitation wavelength equal to 405 nm at all pH values. **7g** emission spectra were generated using excitation wavelengths equal to 440 or 460 nm depending on whether 1 mM CTAB was absent or present, respectively. (E) Raw emission intensities plotted for **6g** without CTAB (red bars), **6g** with 1 mM CTAB (green bars), **7g** without CTAB (purple bars), and **7g** with 1 mM CTAB (orange bars) as a function of pH.

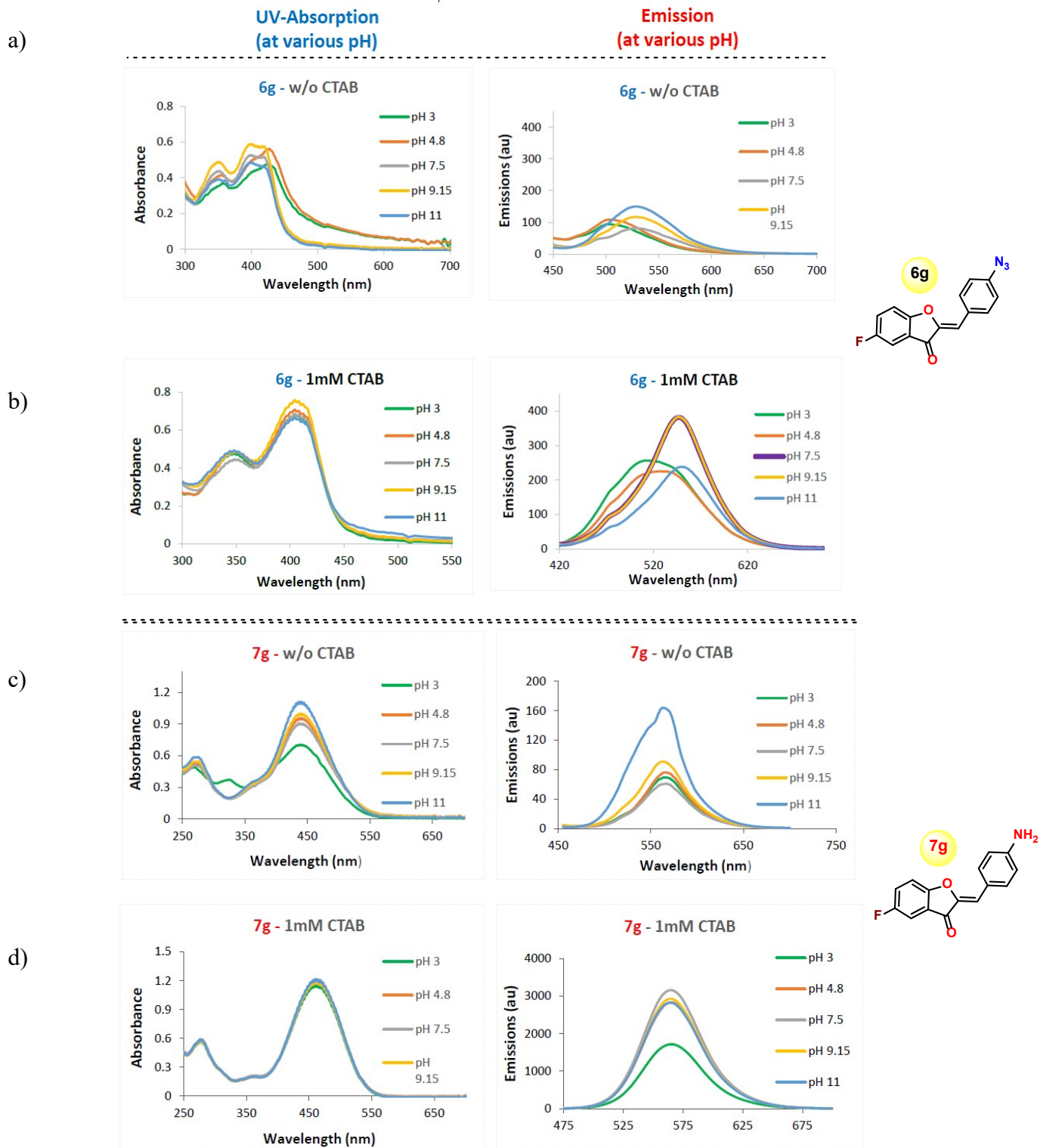
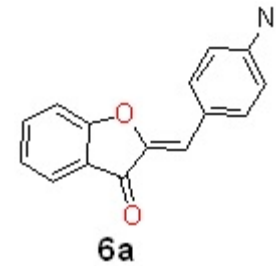


Figure S3. Effects of CTAB on Fluorescence Intensity of 6g/7g at various pH. Solvent system; 0.5%v/v DMSO in PBS.

AK_3_82_2_1H

7.926
7.909
7.815
7.801
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7.660
7.646
7.335
7.319
7.244
7.229
7.214
7.109
7.092
6.851

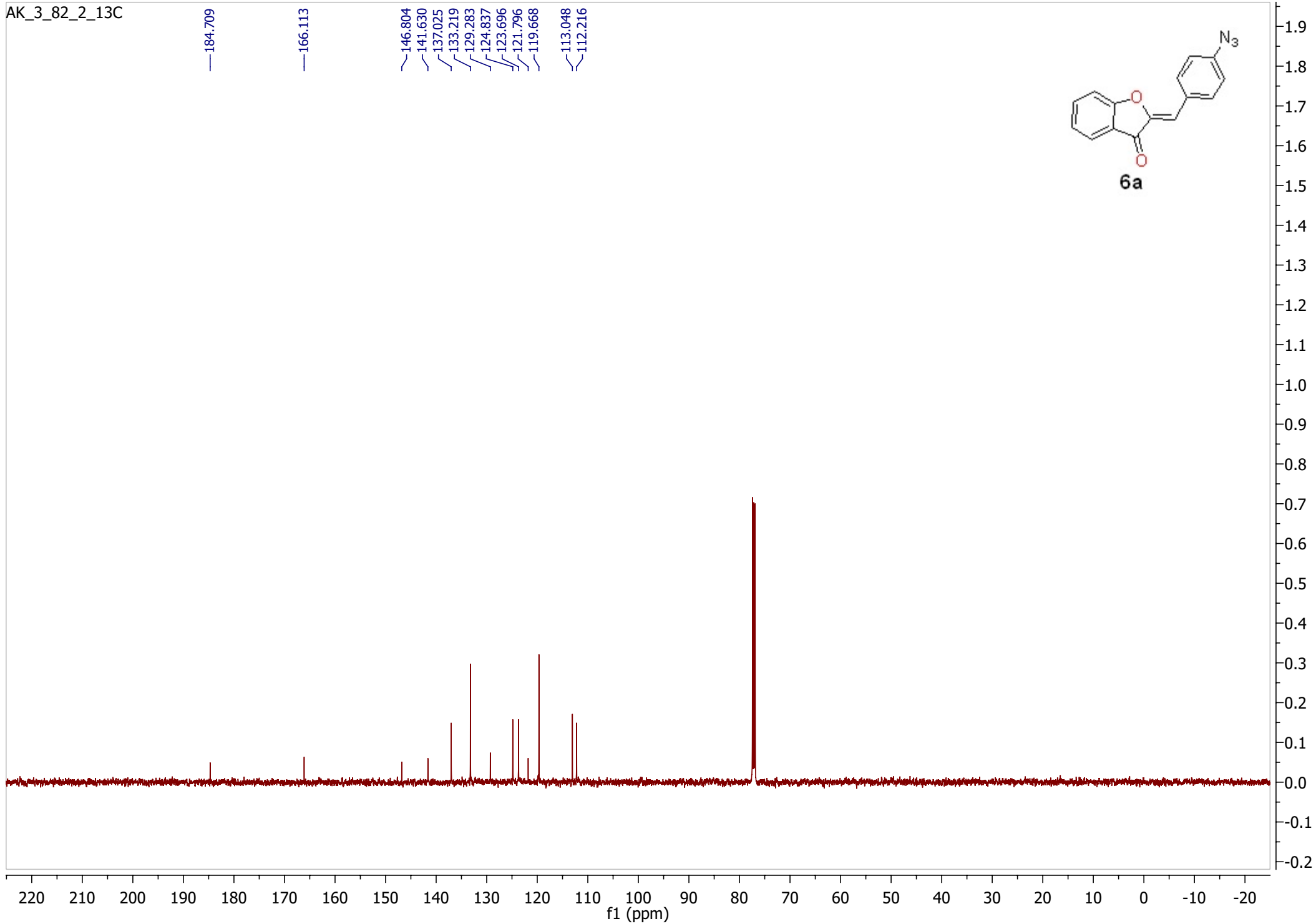


2.15
1.03
1.11
1.06
1.09
2.06
1.00

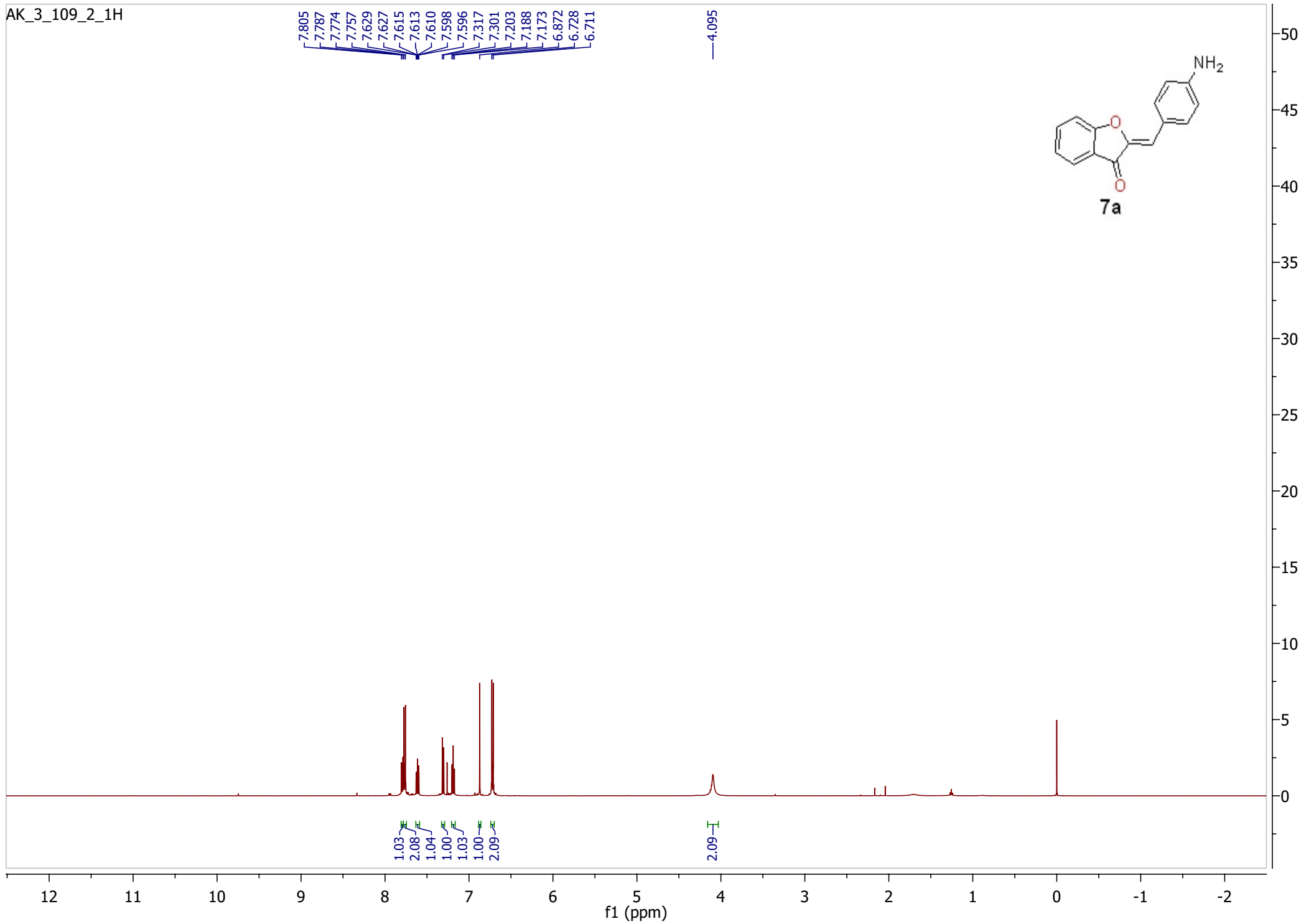
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f1 (ppm)

AK_3_82_2_13C

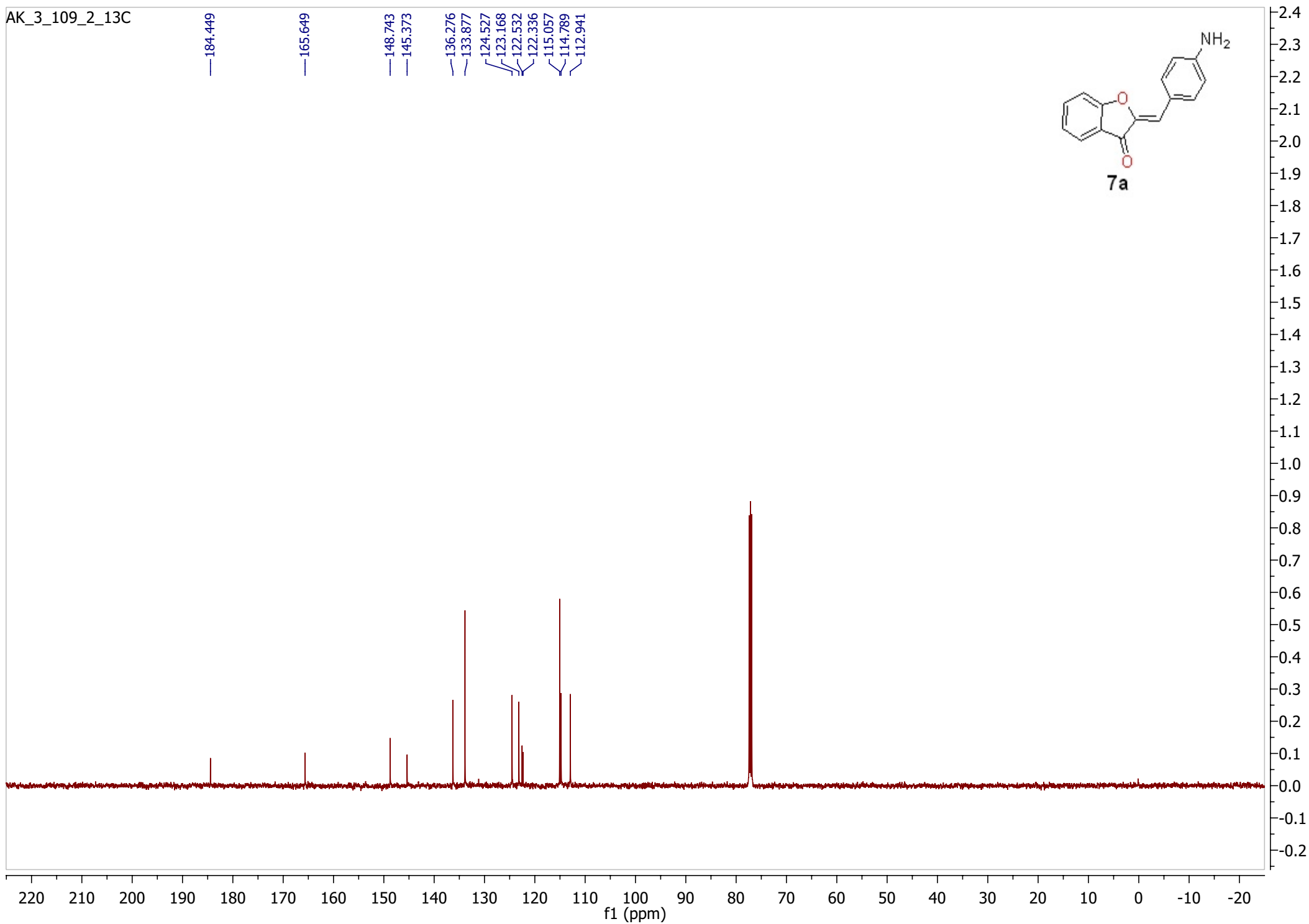
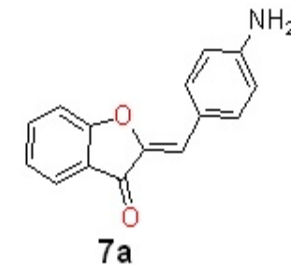


AK_3_109_2_1H



AK_3_109_2_13C

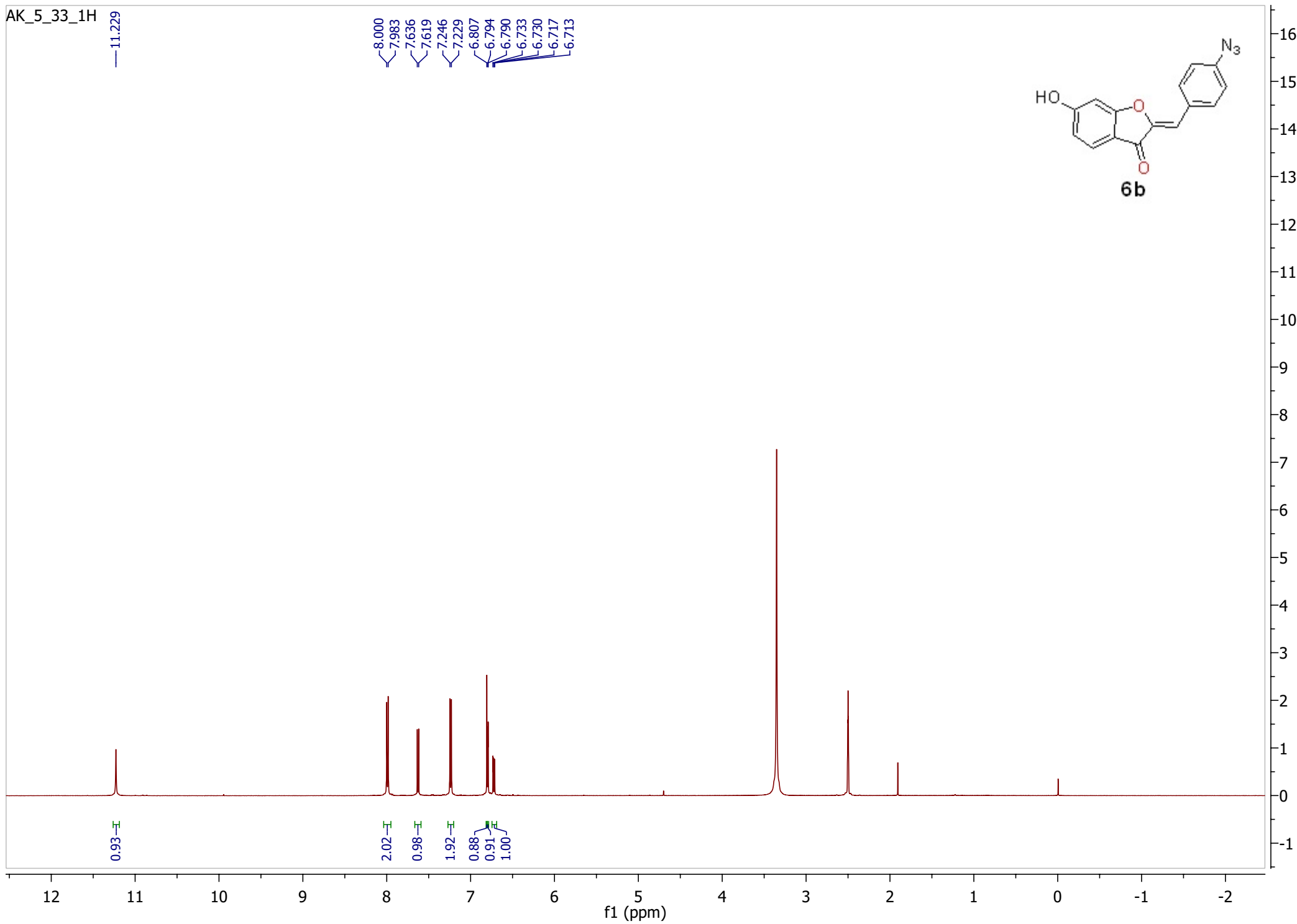
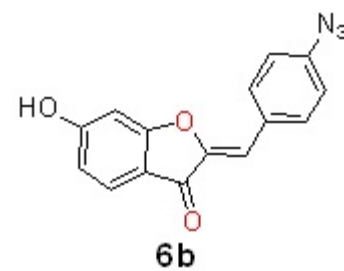
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—145.373
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—122.336
—115.057
—114.789
—112.941



AK_5_33_1H

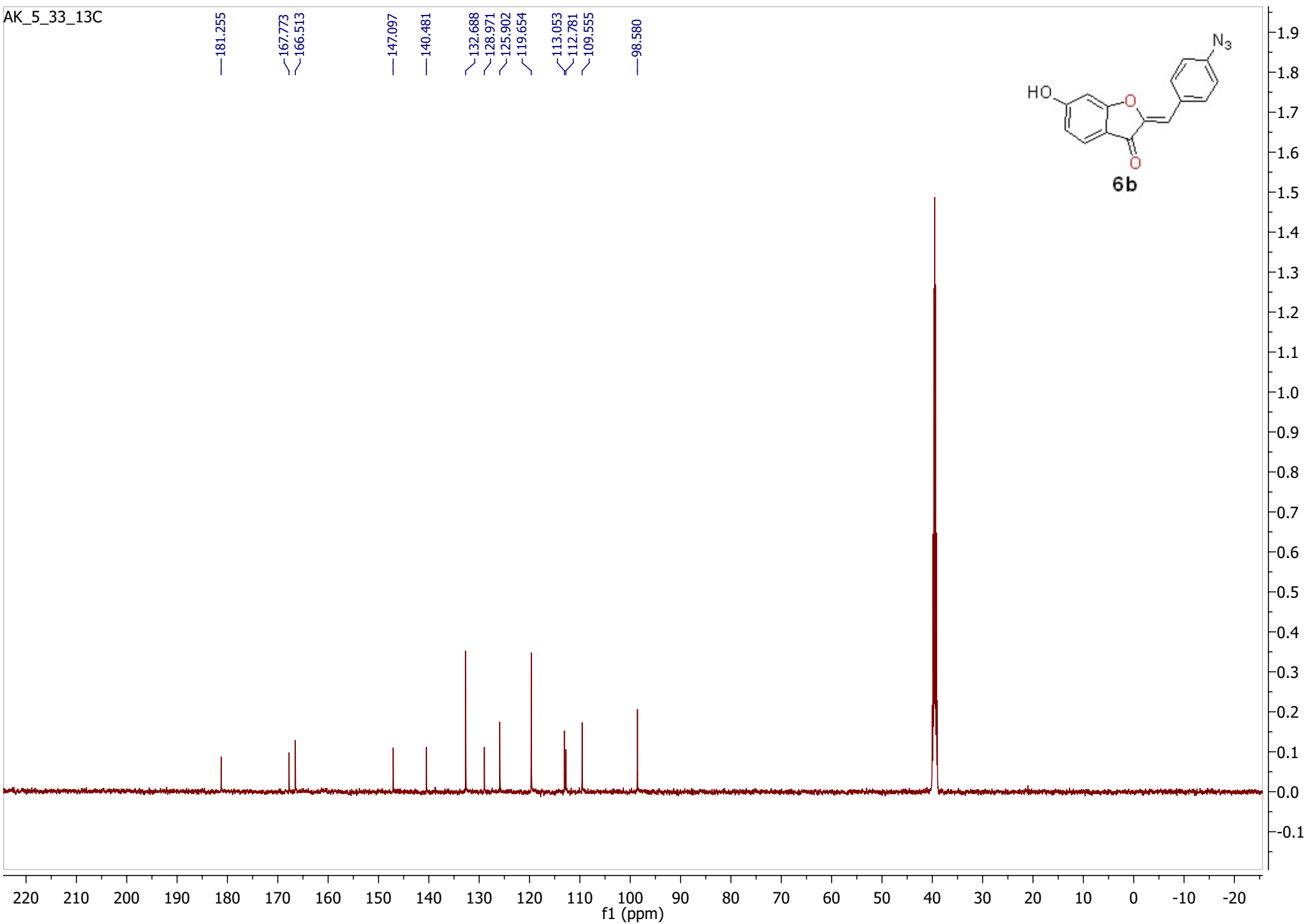
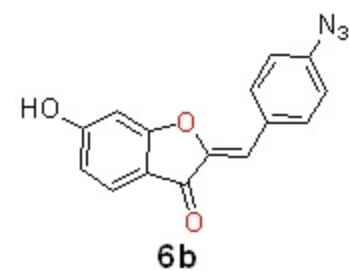
11.229

8.000
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7.246
7.229
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6.790
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6.730
6.717
6.713

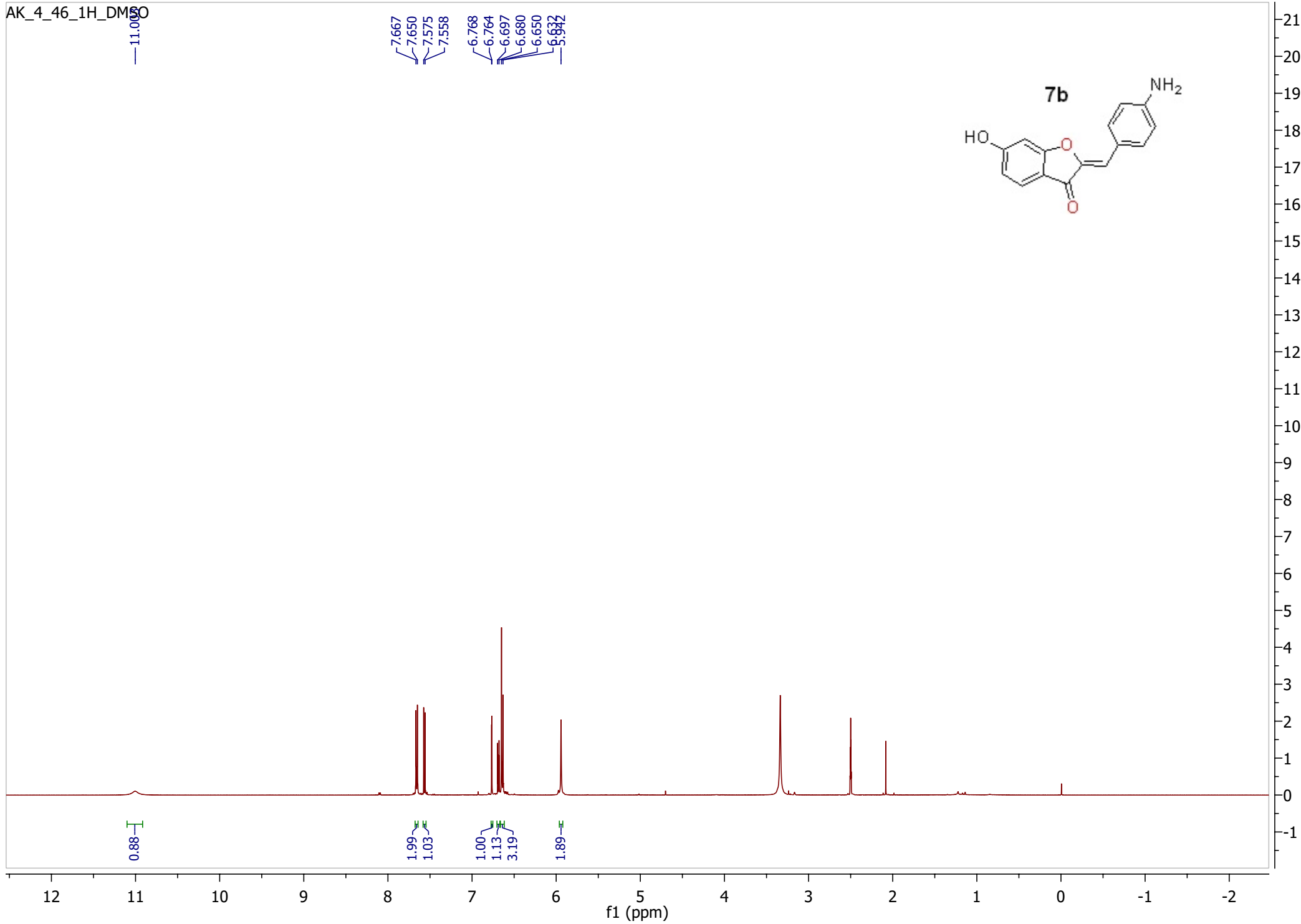


AK_5_33_13C

181.255
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147.097
140.481
132.688
128.971
125.902
119.654
113.053
112.781
109.555
98.580

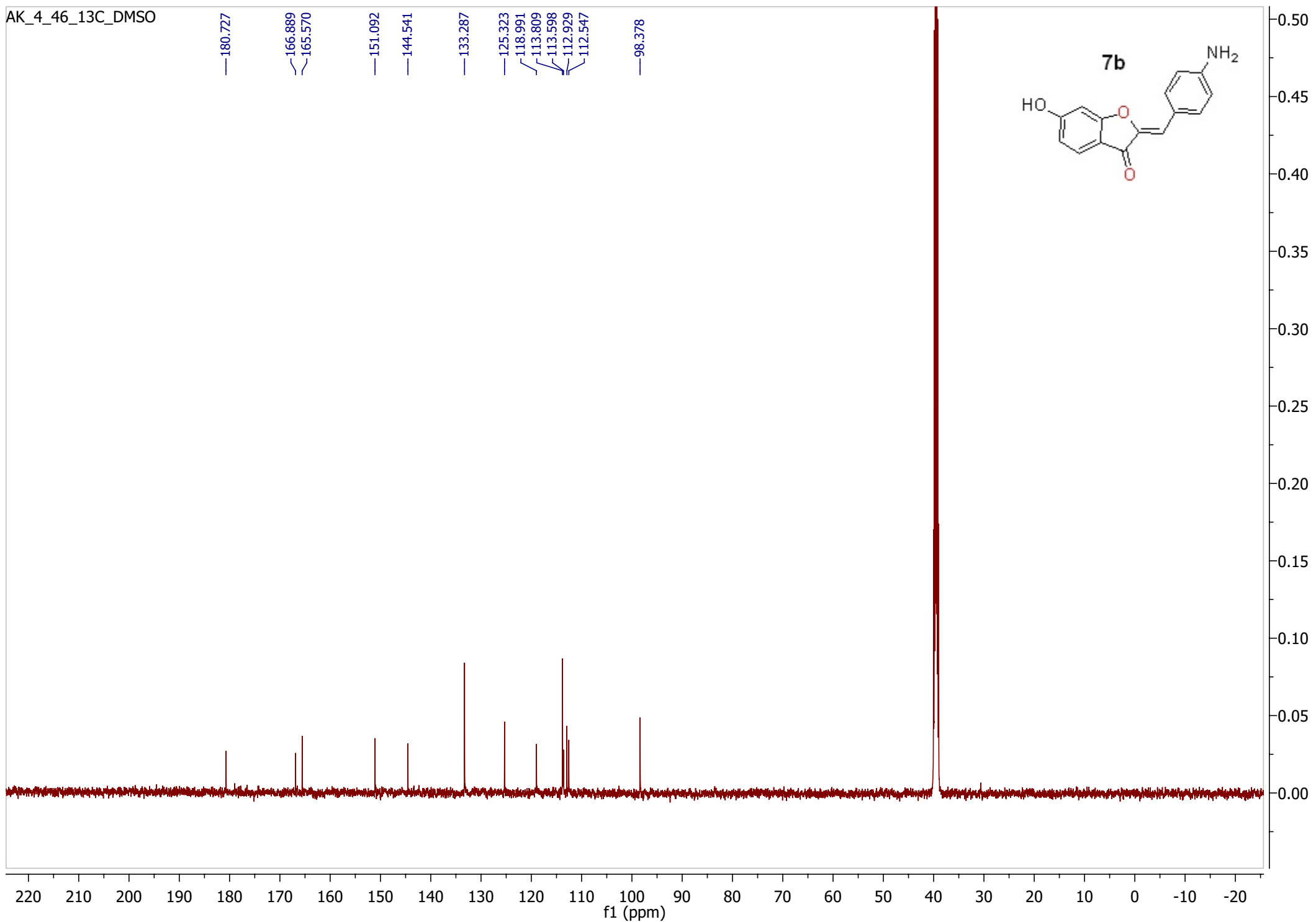
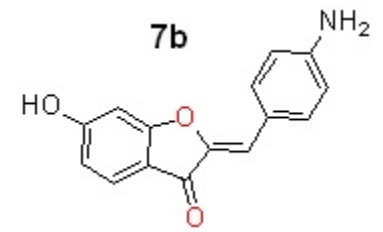


AK_4_46_1H_DMSO



AK_4_46_13C_DMSO

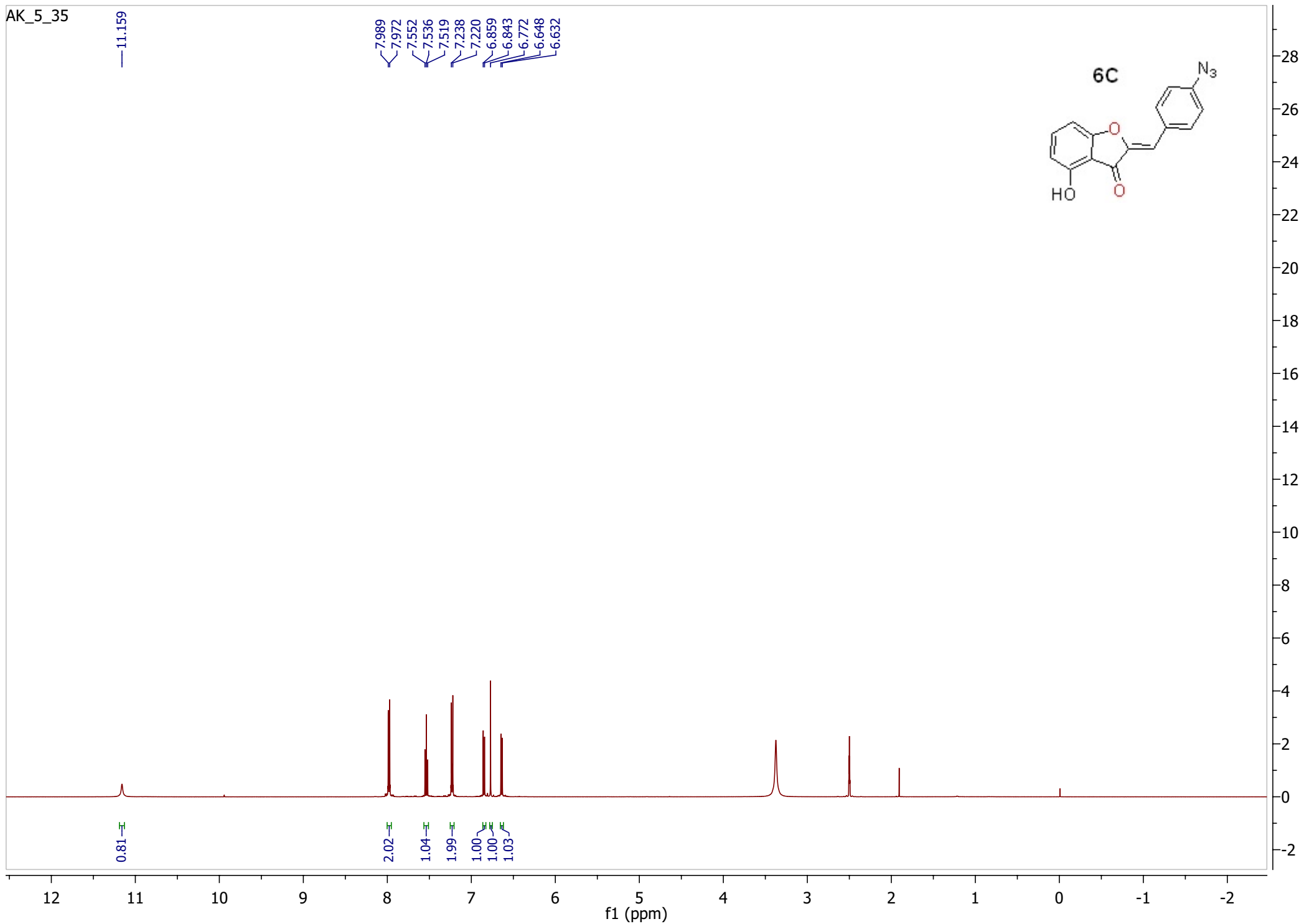
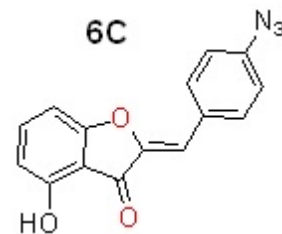
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98.378



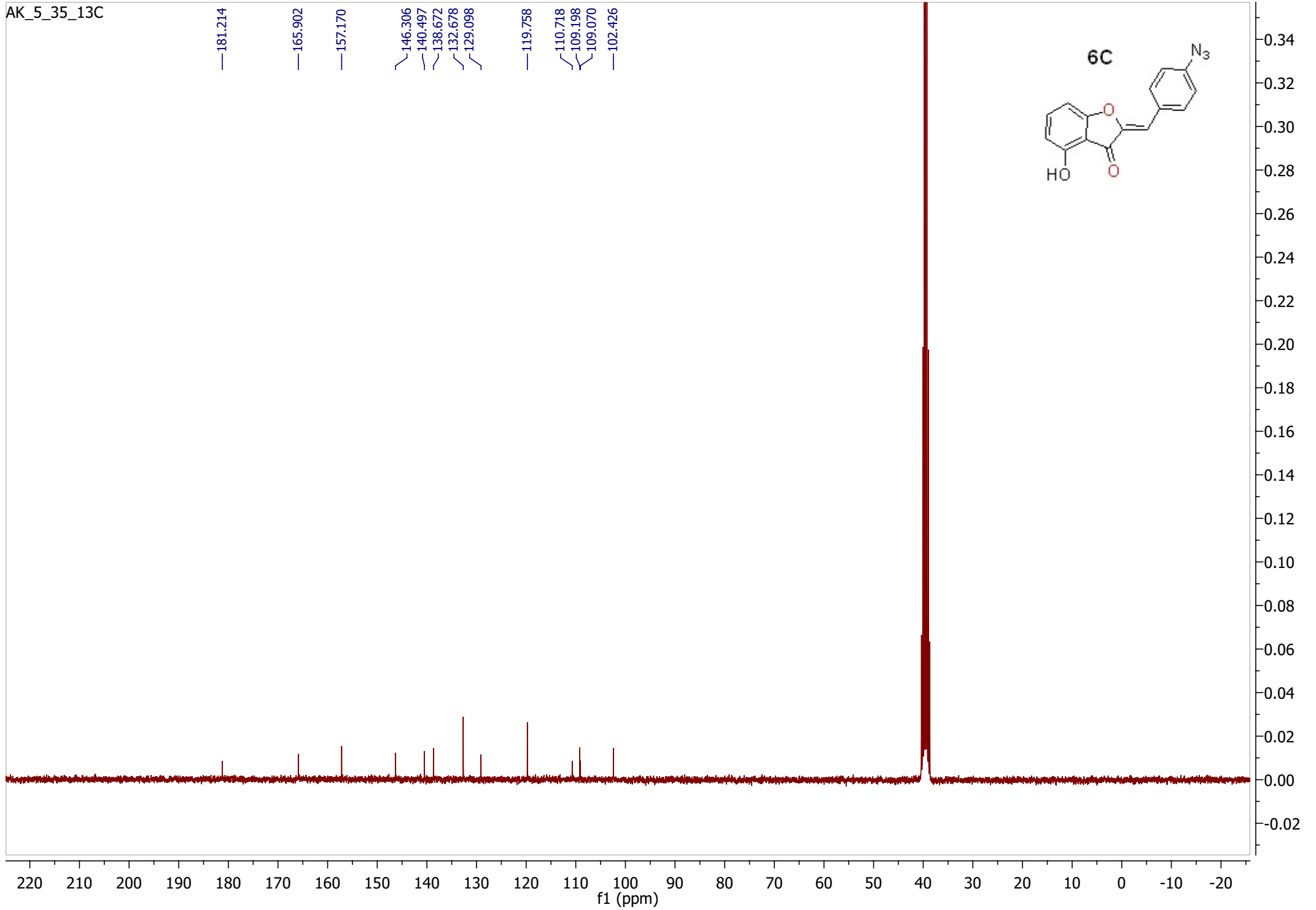
AK_5_35

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0.81

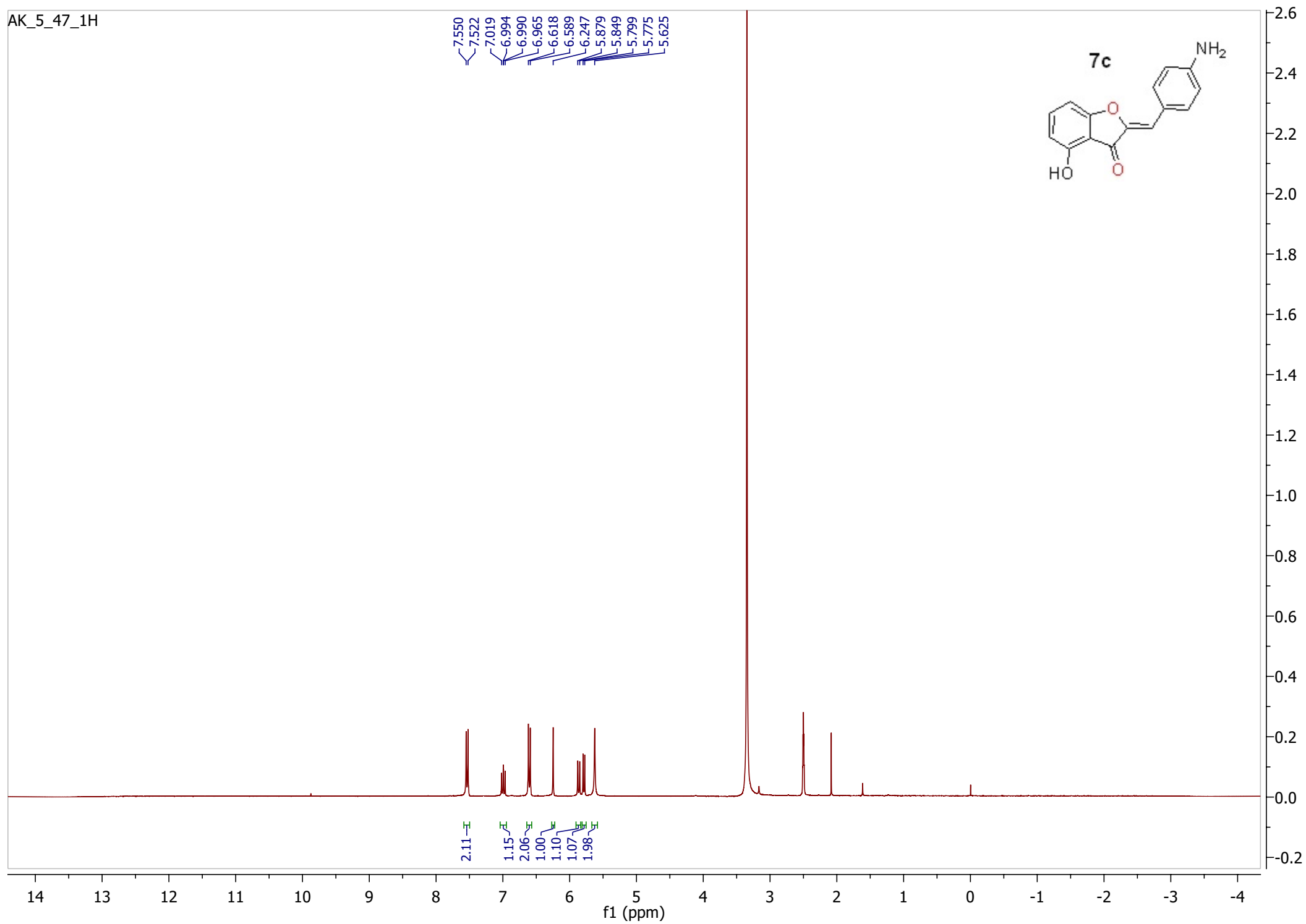
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7.519
7.238
7.220
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6.772
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6.632
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1.04
1.99
1.00
1.00
1.03



AK_5_35_13C



AK_5_47_1H



AK_5_47_13C

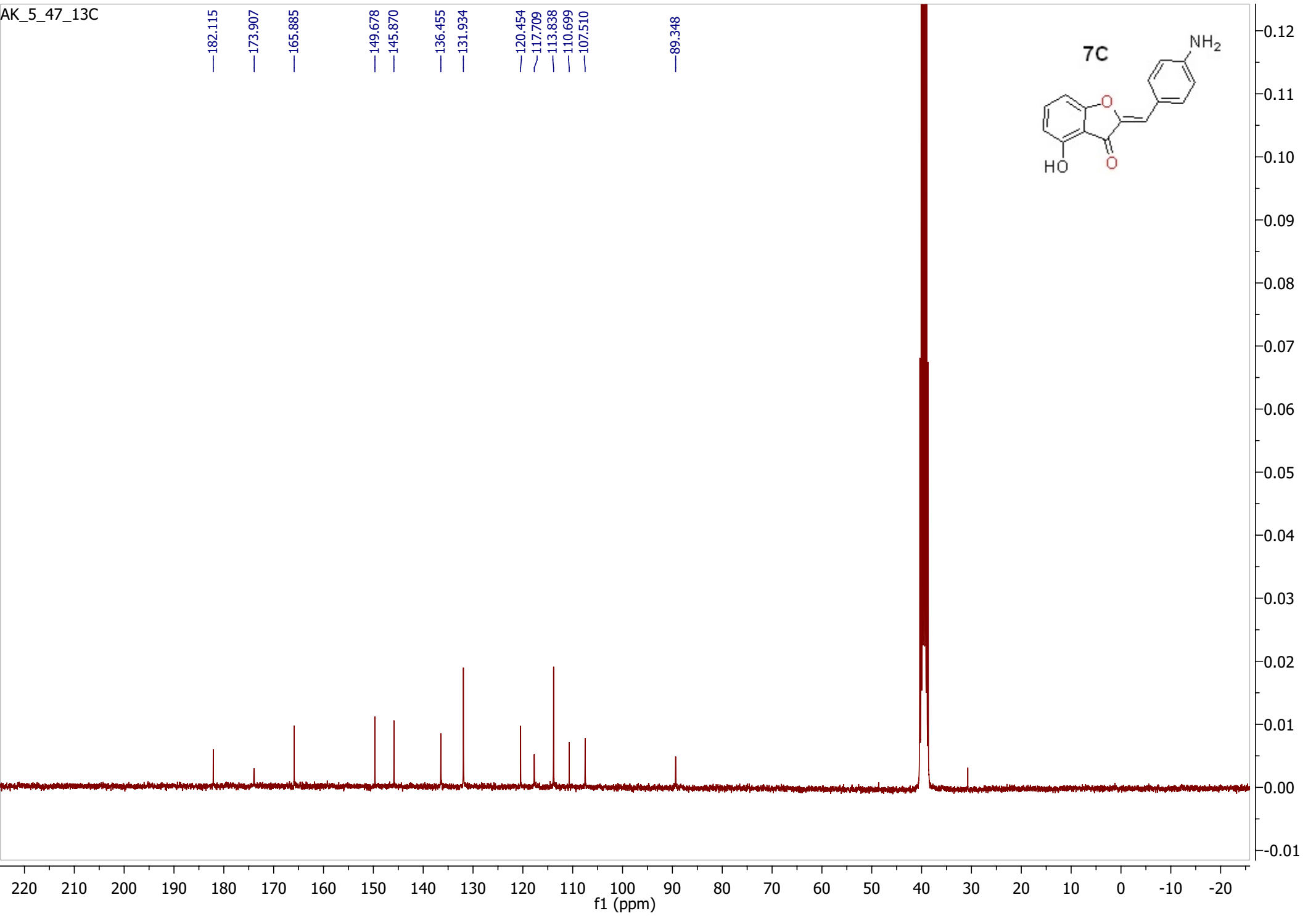
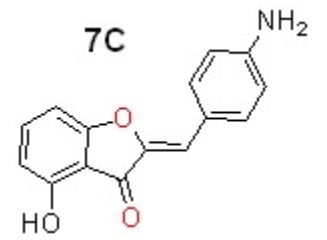
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—145.870

—136.455
—131.934

—120.454
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—113.838
—110.699
—107.510

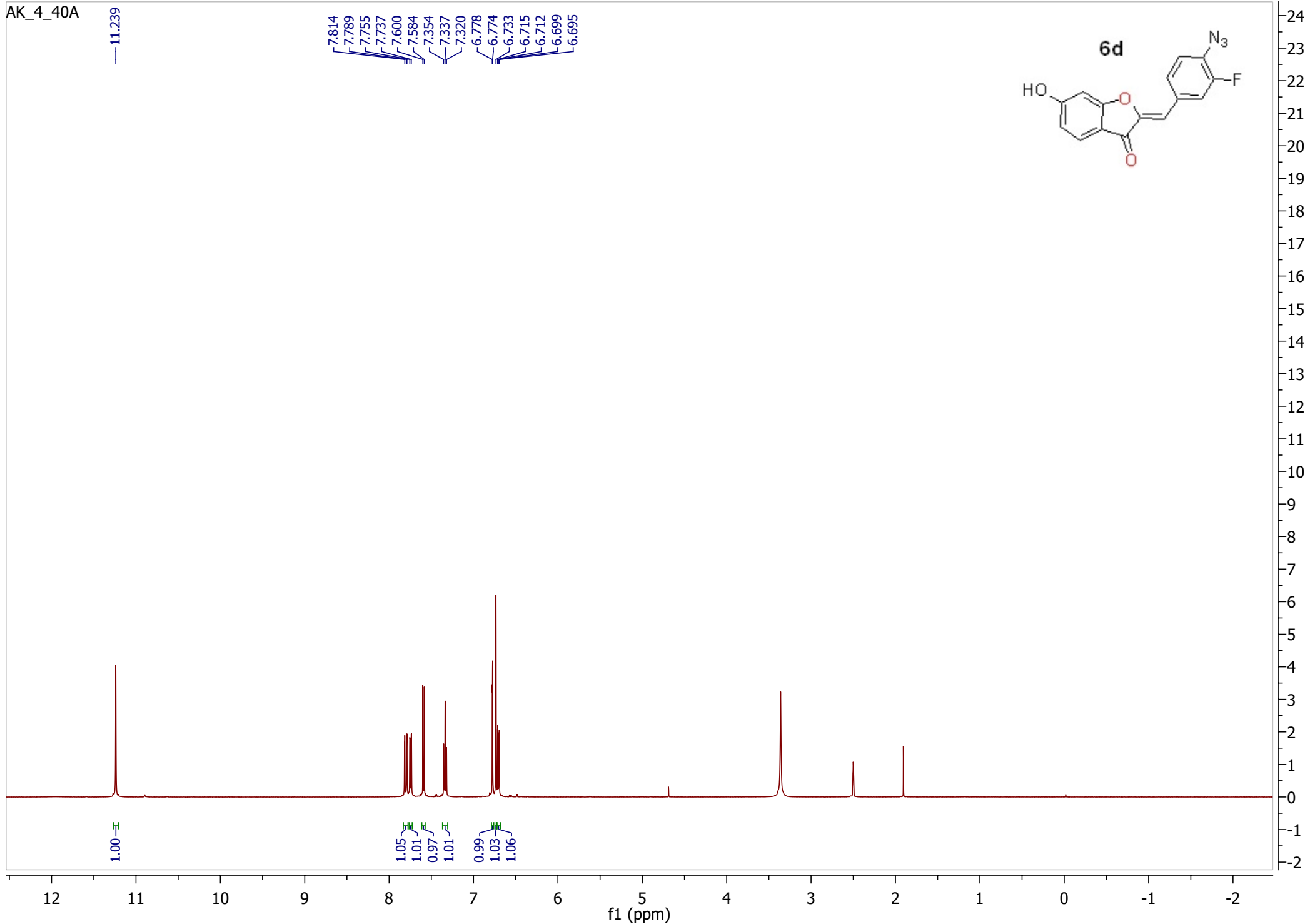
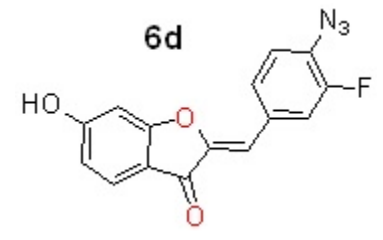
—89.348



AK_4_40A

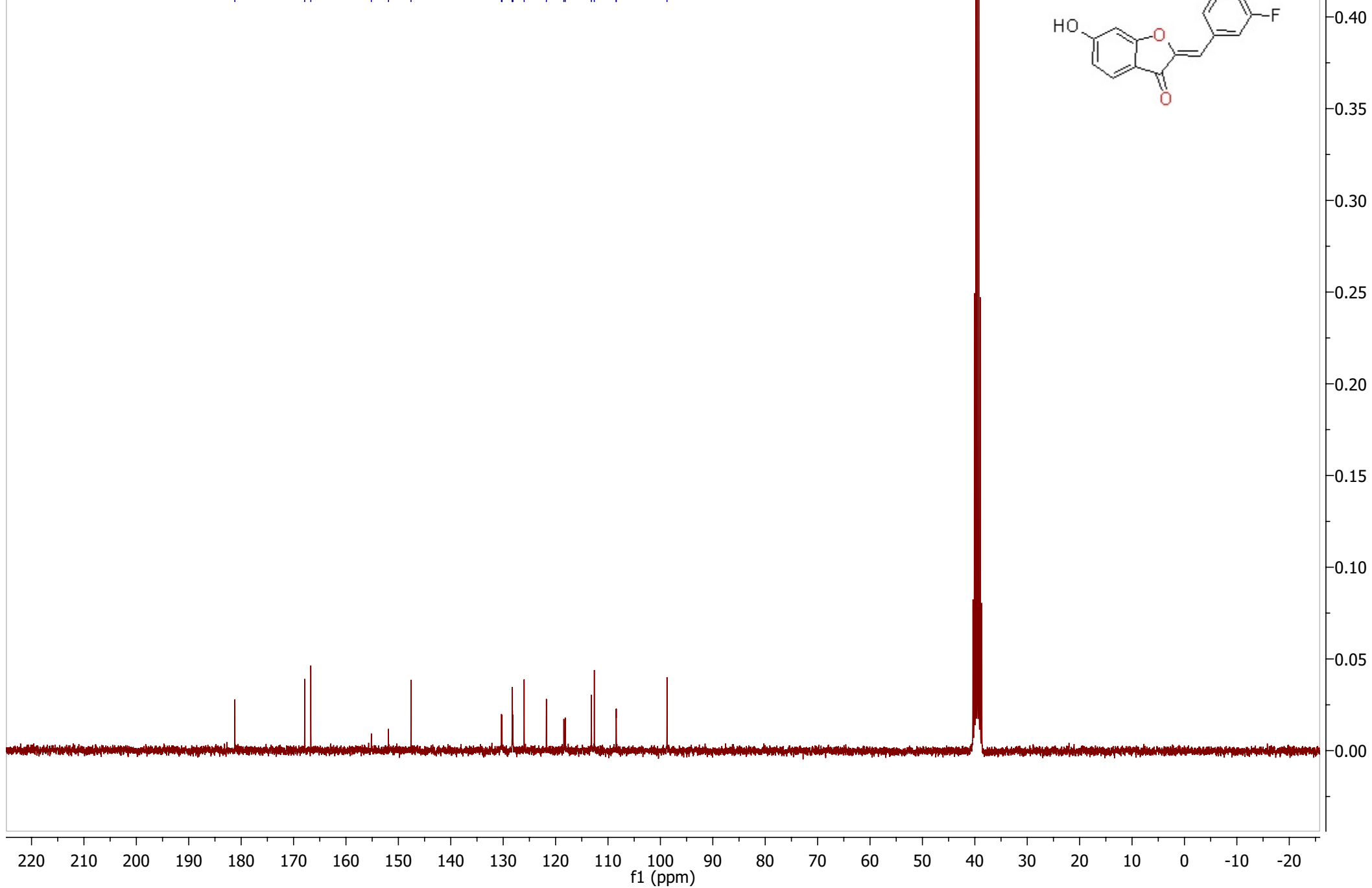
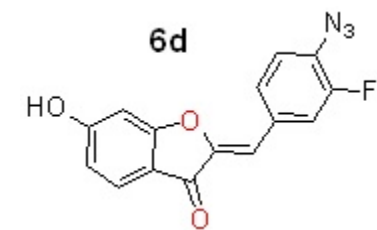
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6.733
6.715
6.712
6.699
6.695



AK_4_40A_13C

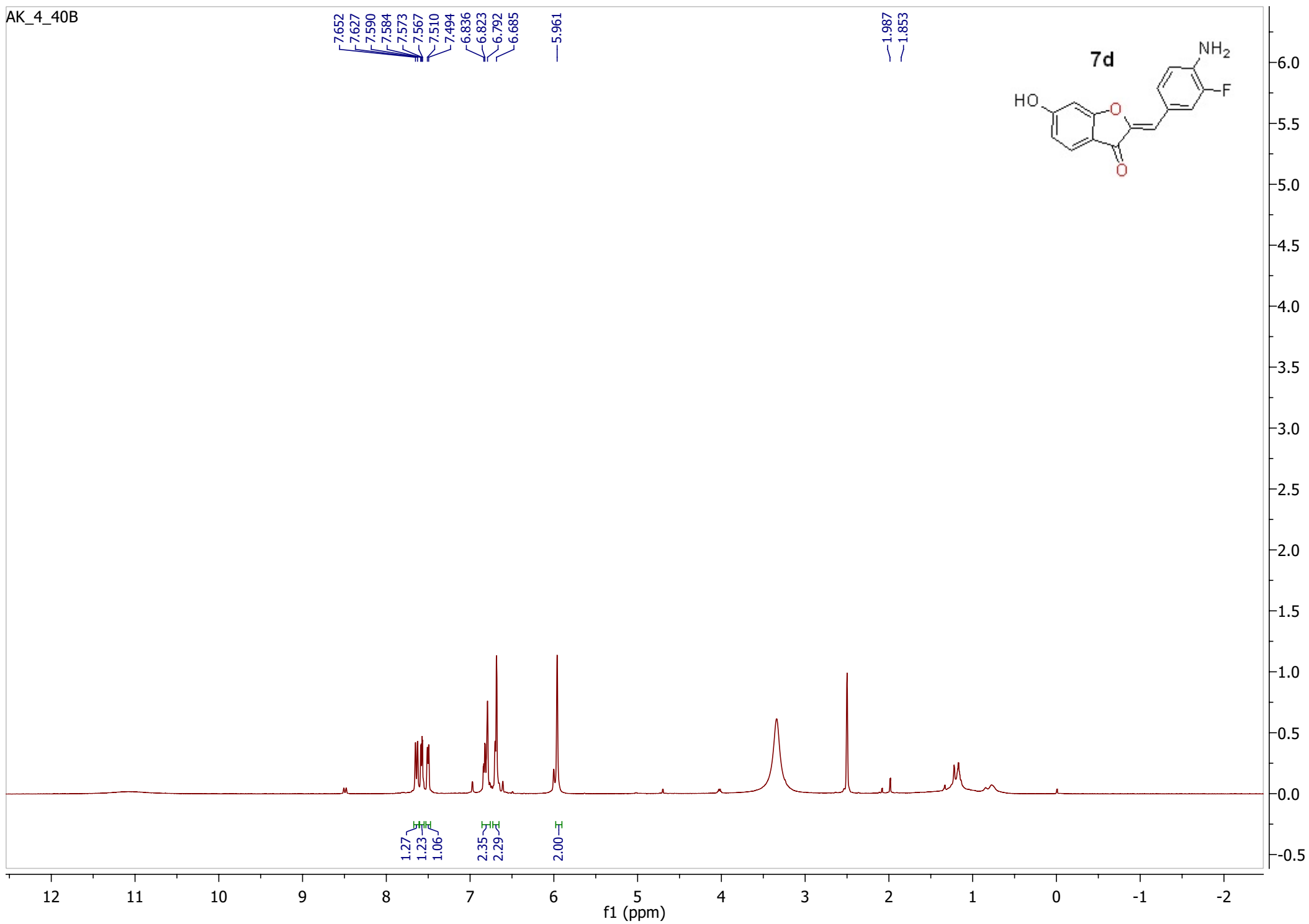
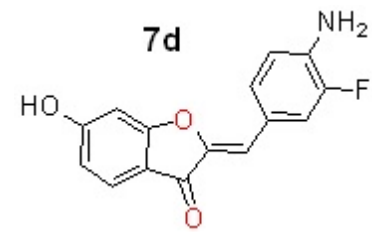
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108.416
98.742



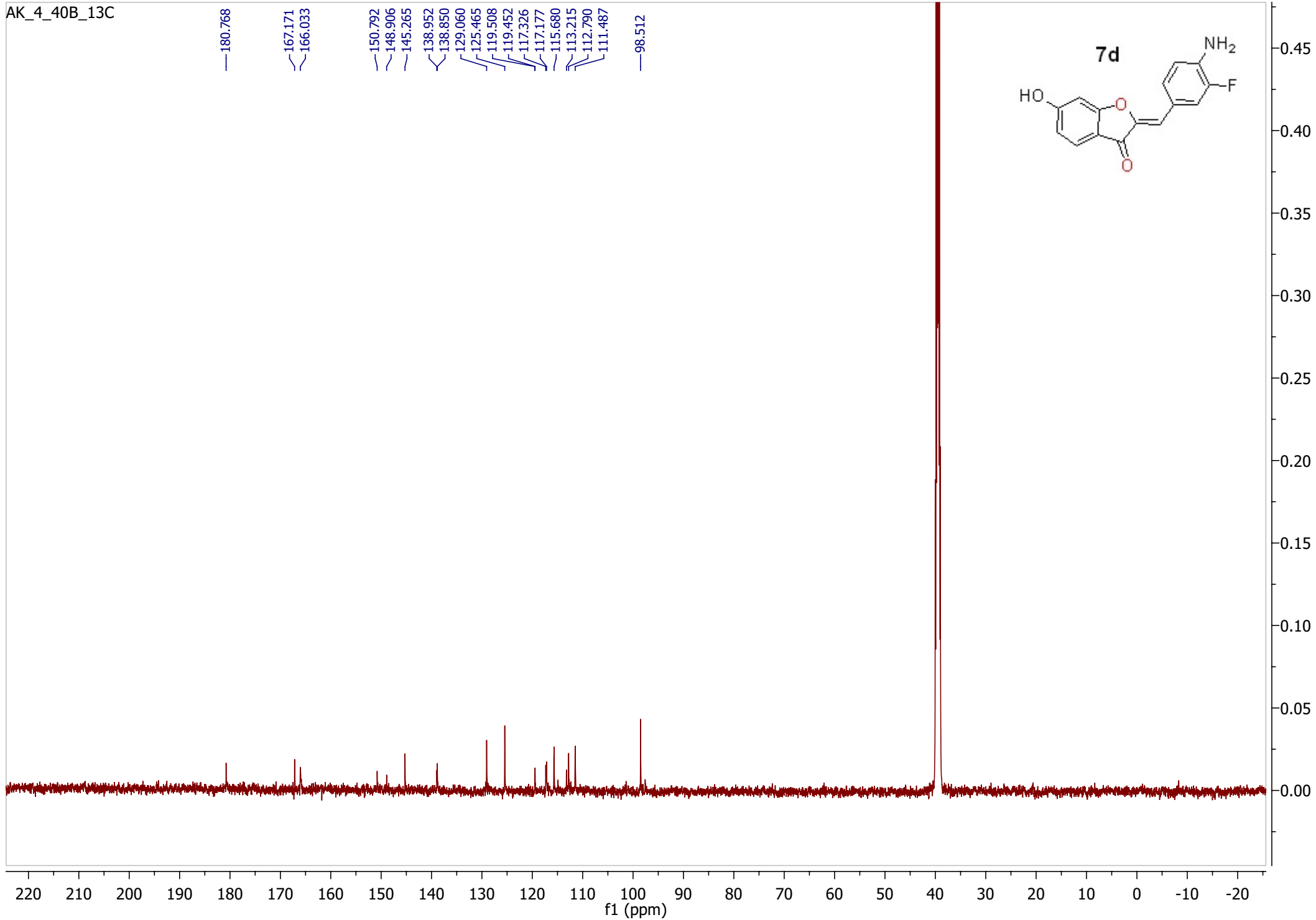
AK_4_40B

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6.823
6.792
6.685
5.961

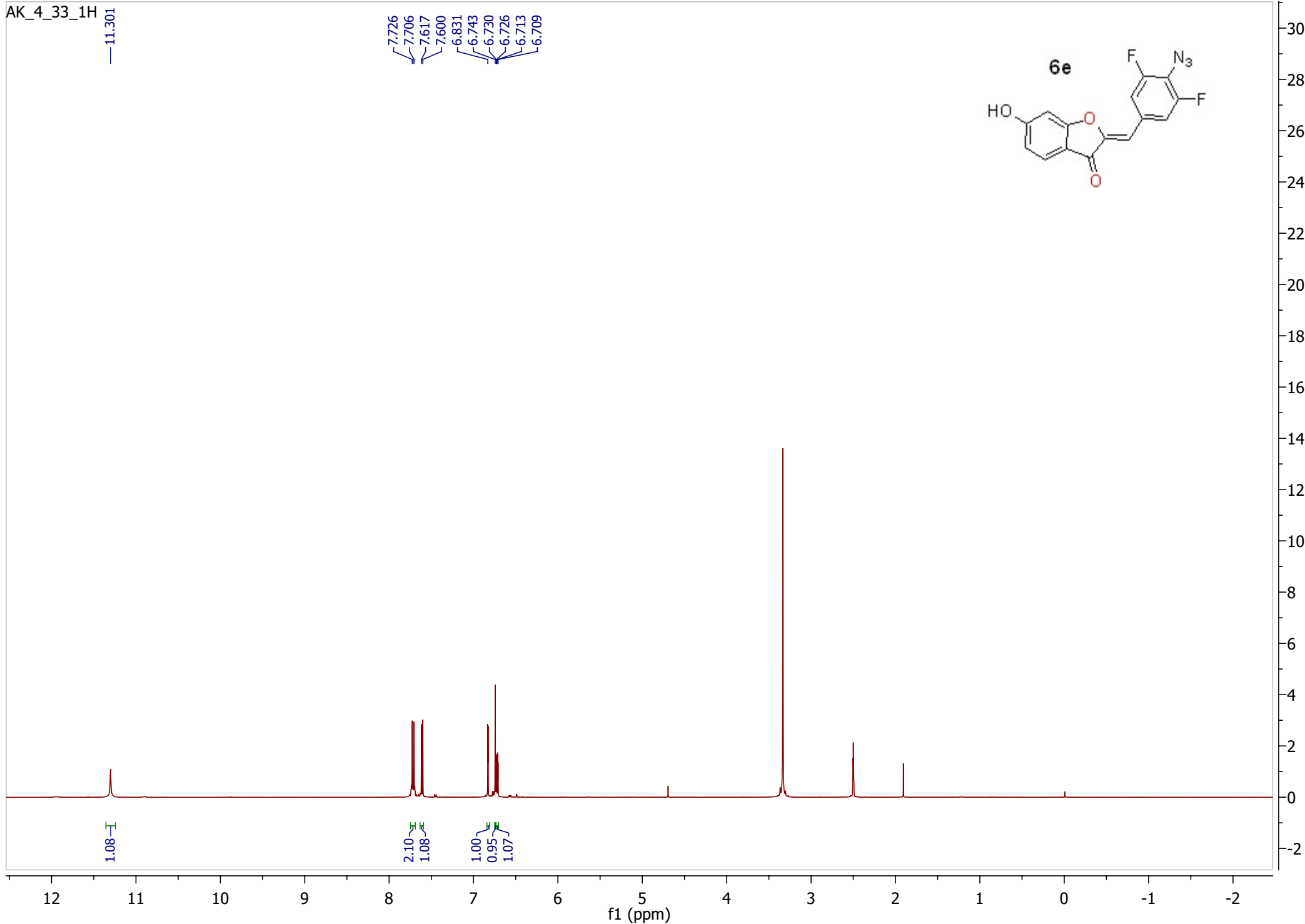
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1.853



AK_4_40B_13C

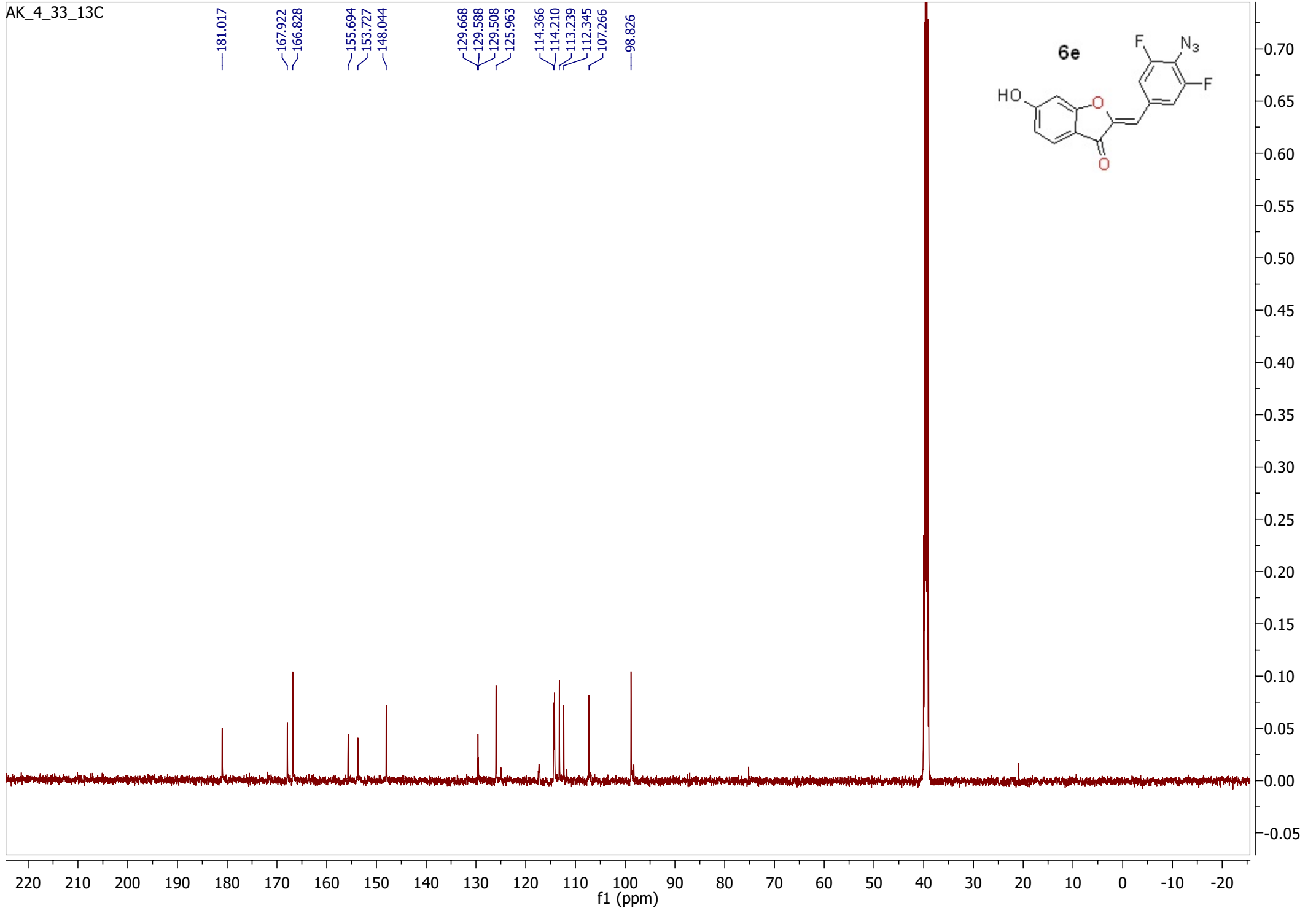
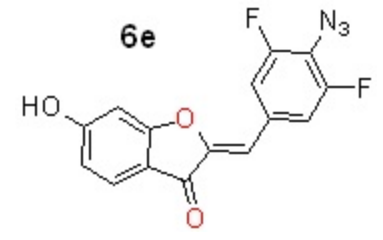


AK_4_33_1H



AK_4_33_13C

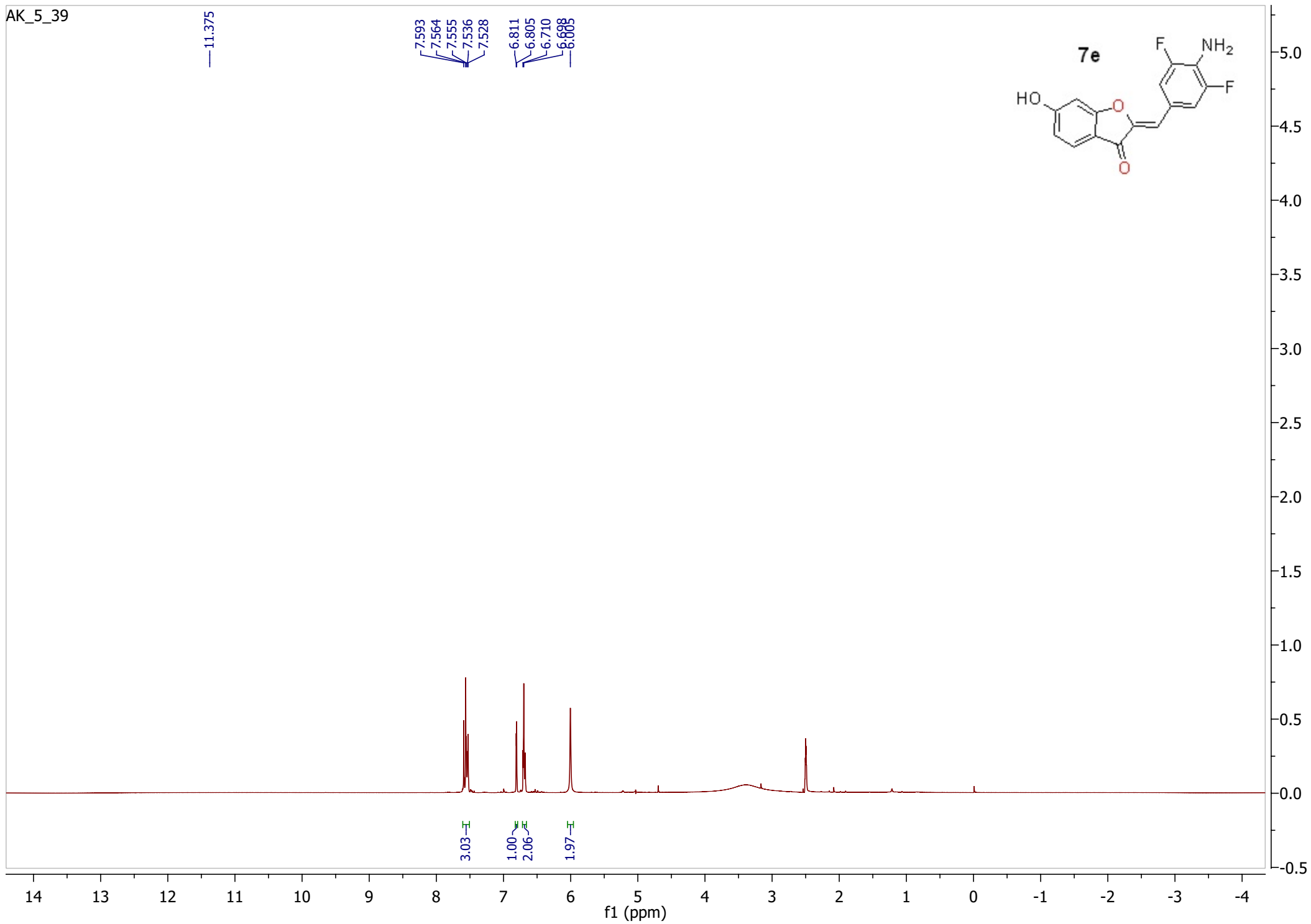
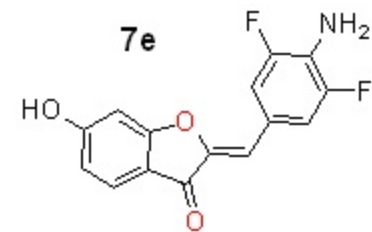
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107.266
98.826



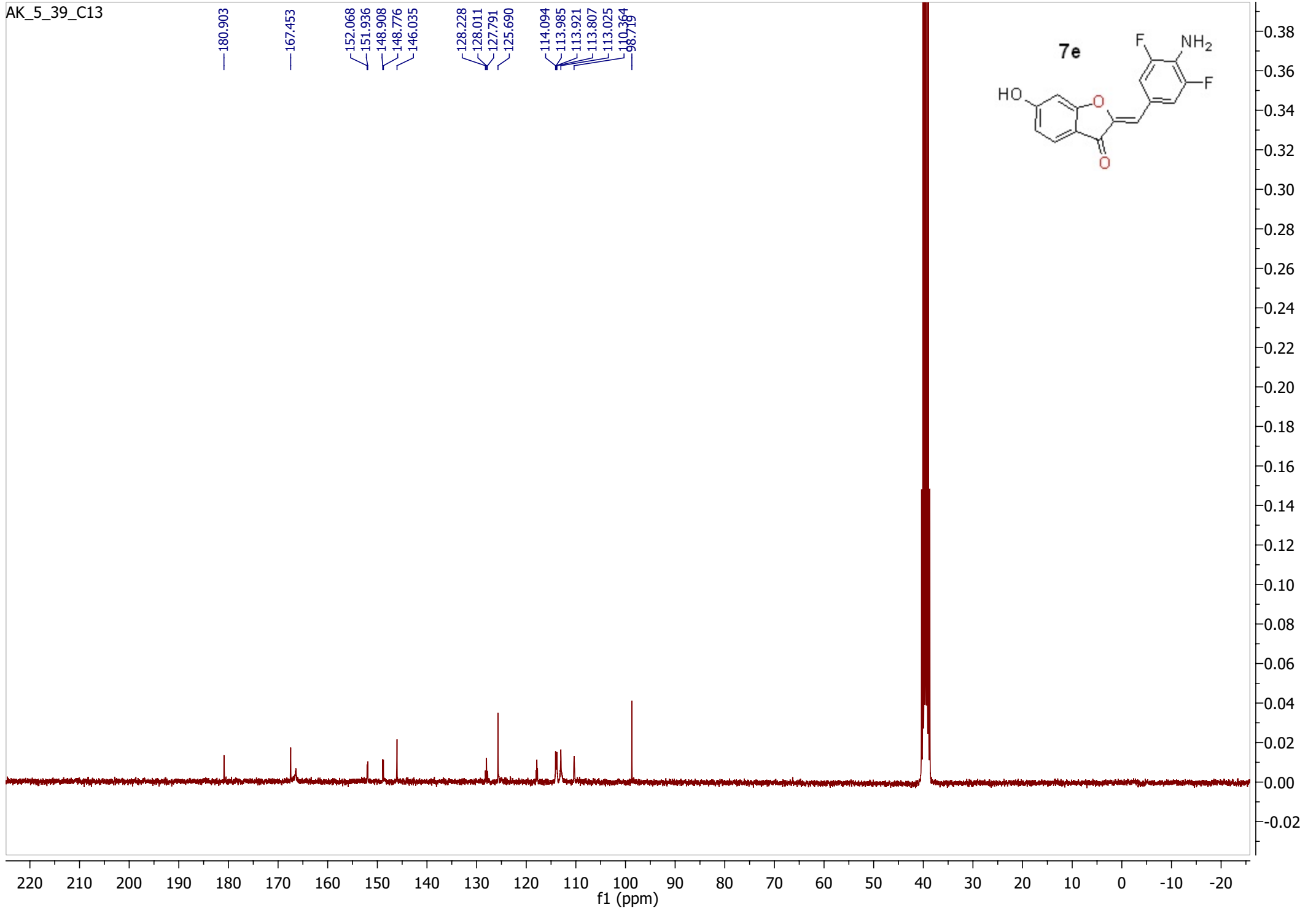
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6.710
6.688



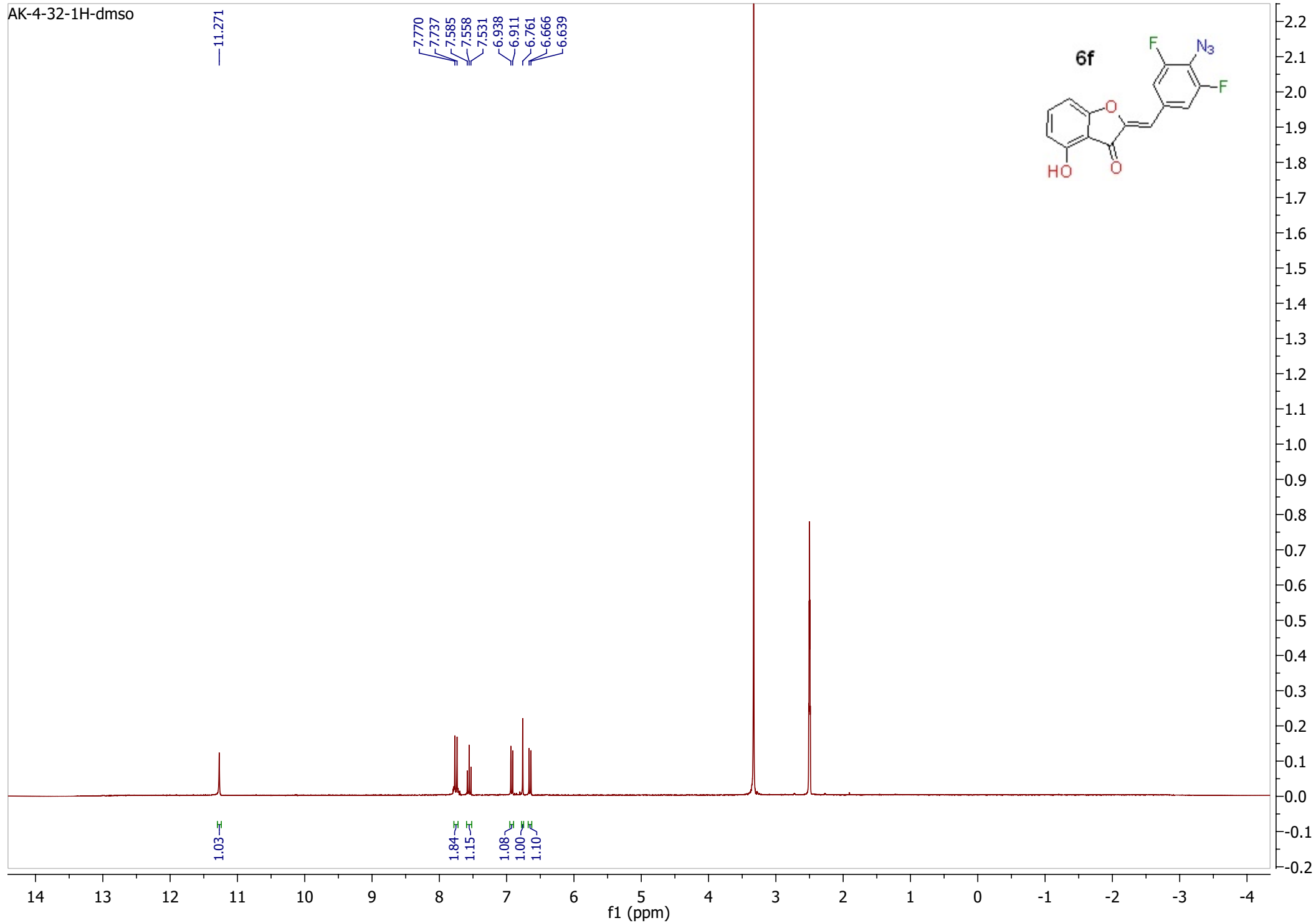
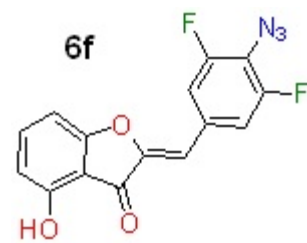
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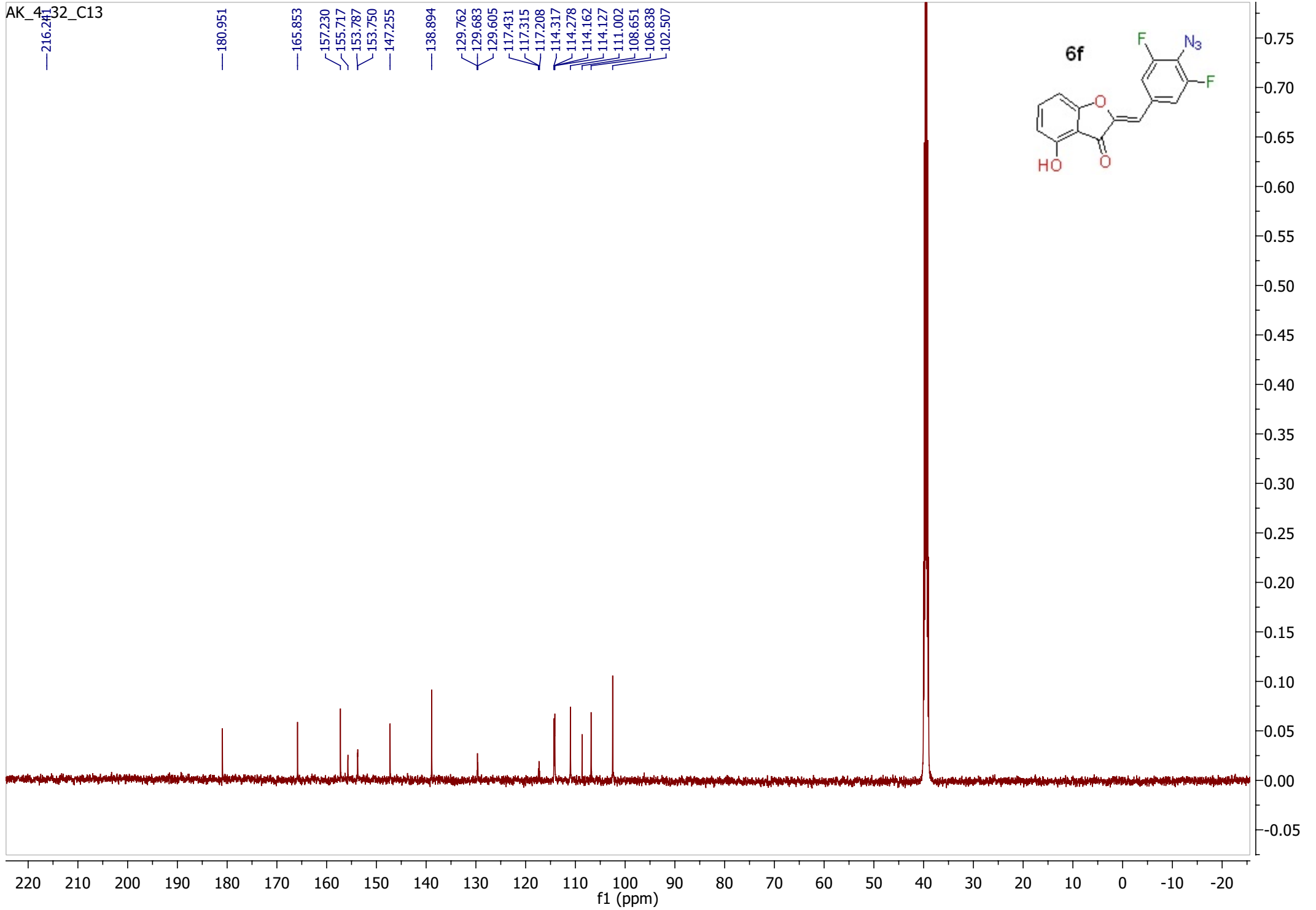
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1.03

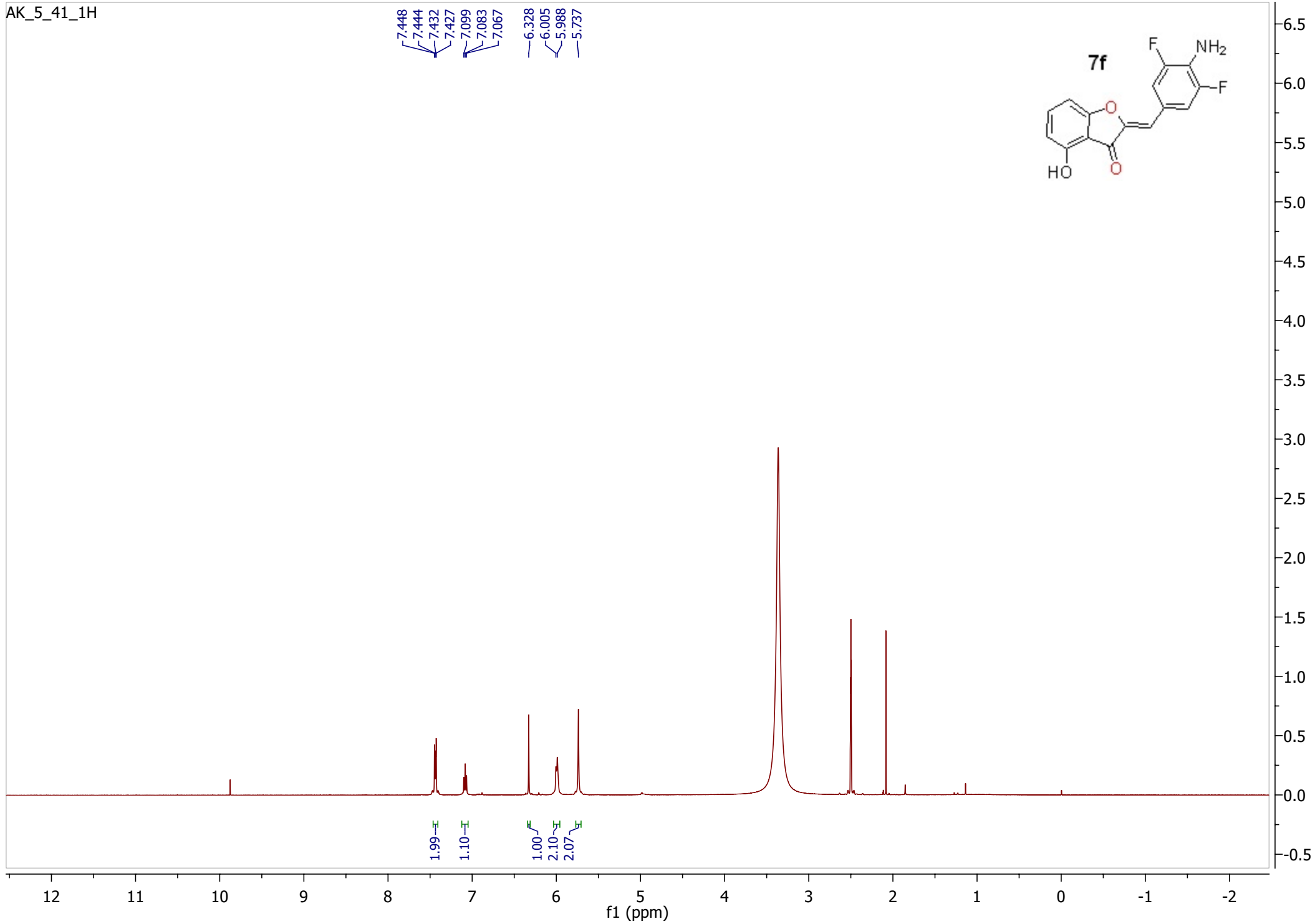
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1.08
1.00
1.10



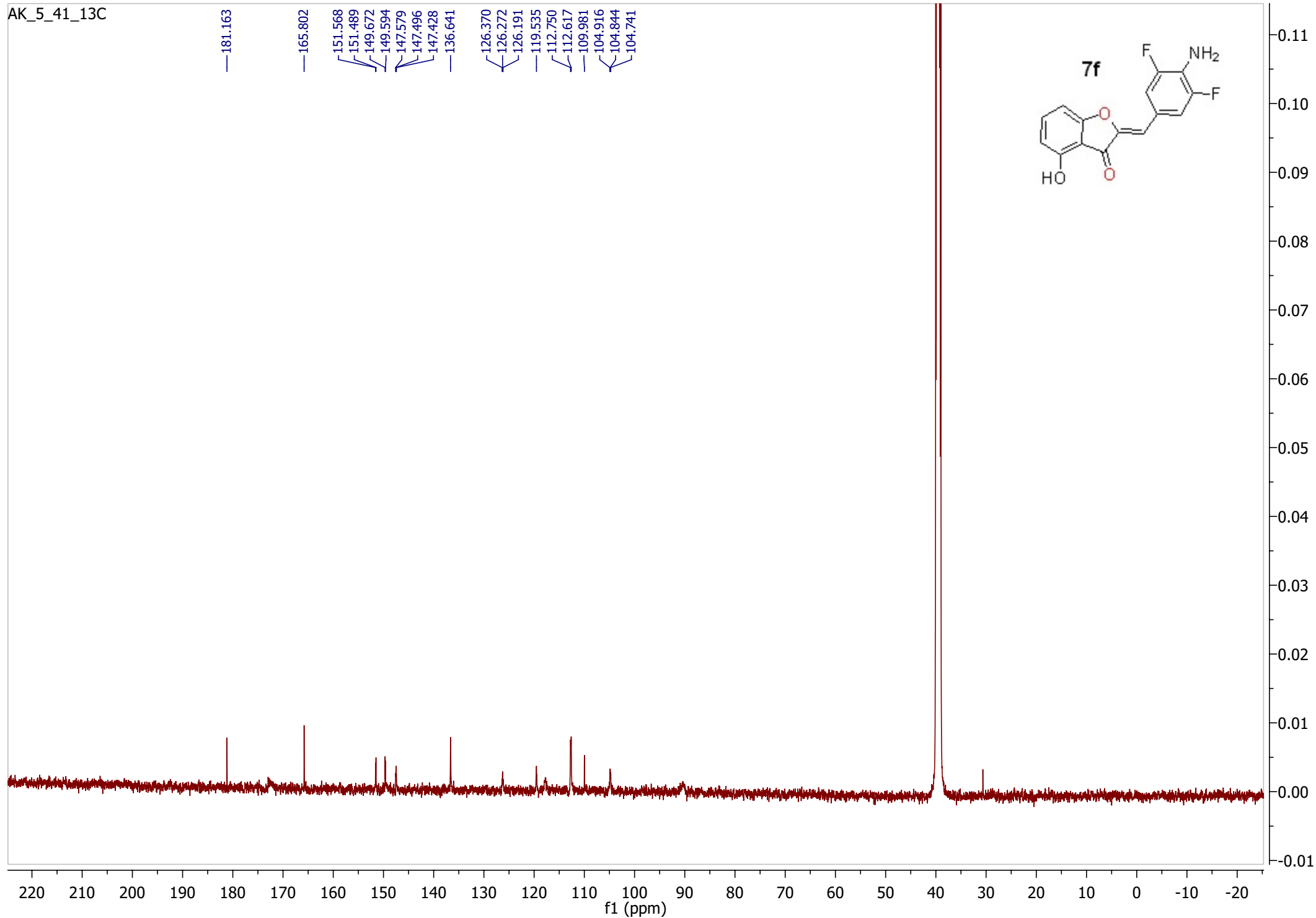
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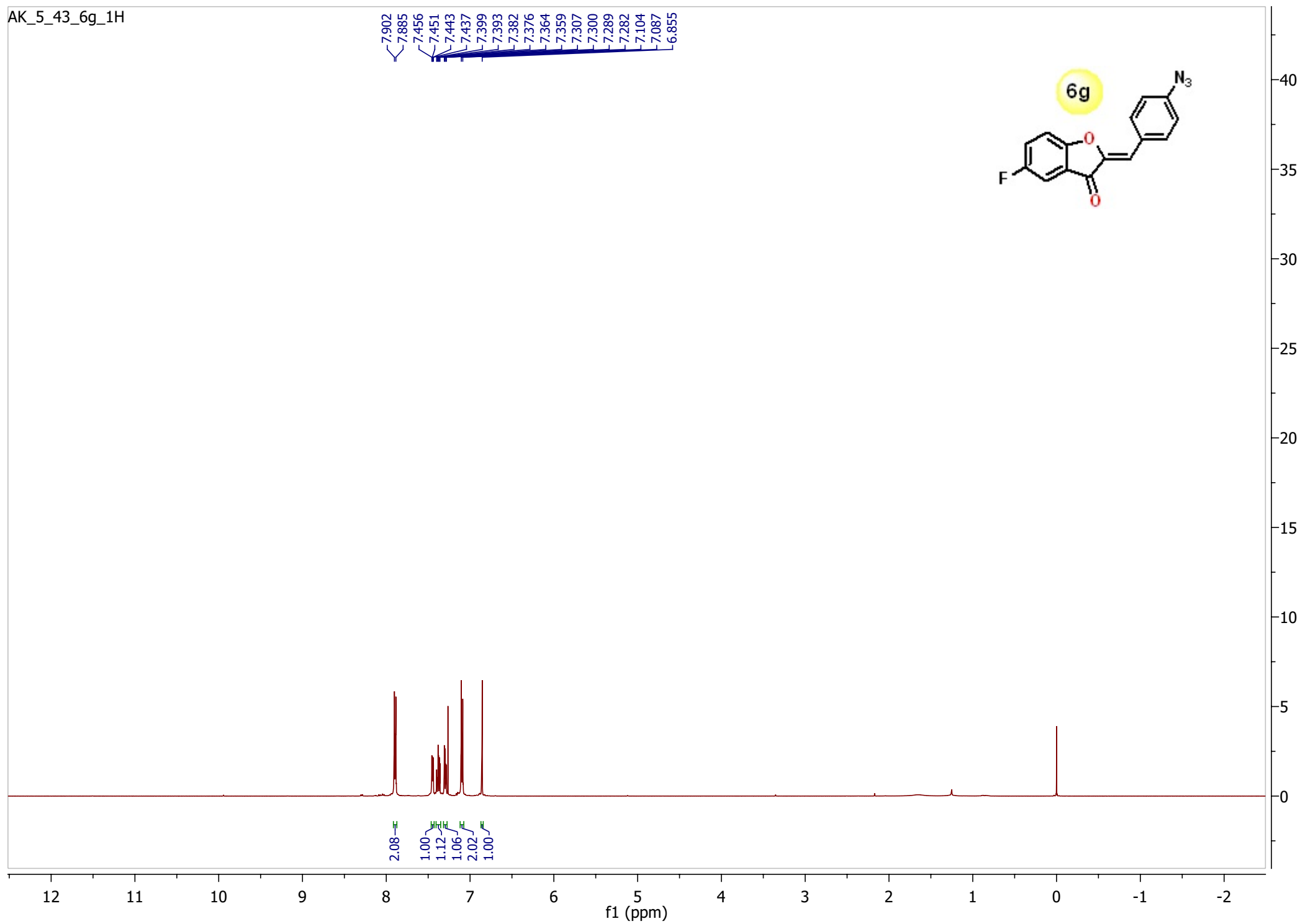
AK_5_41_1H



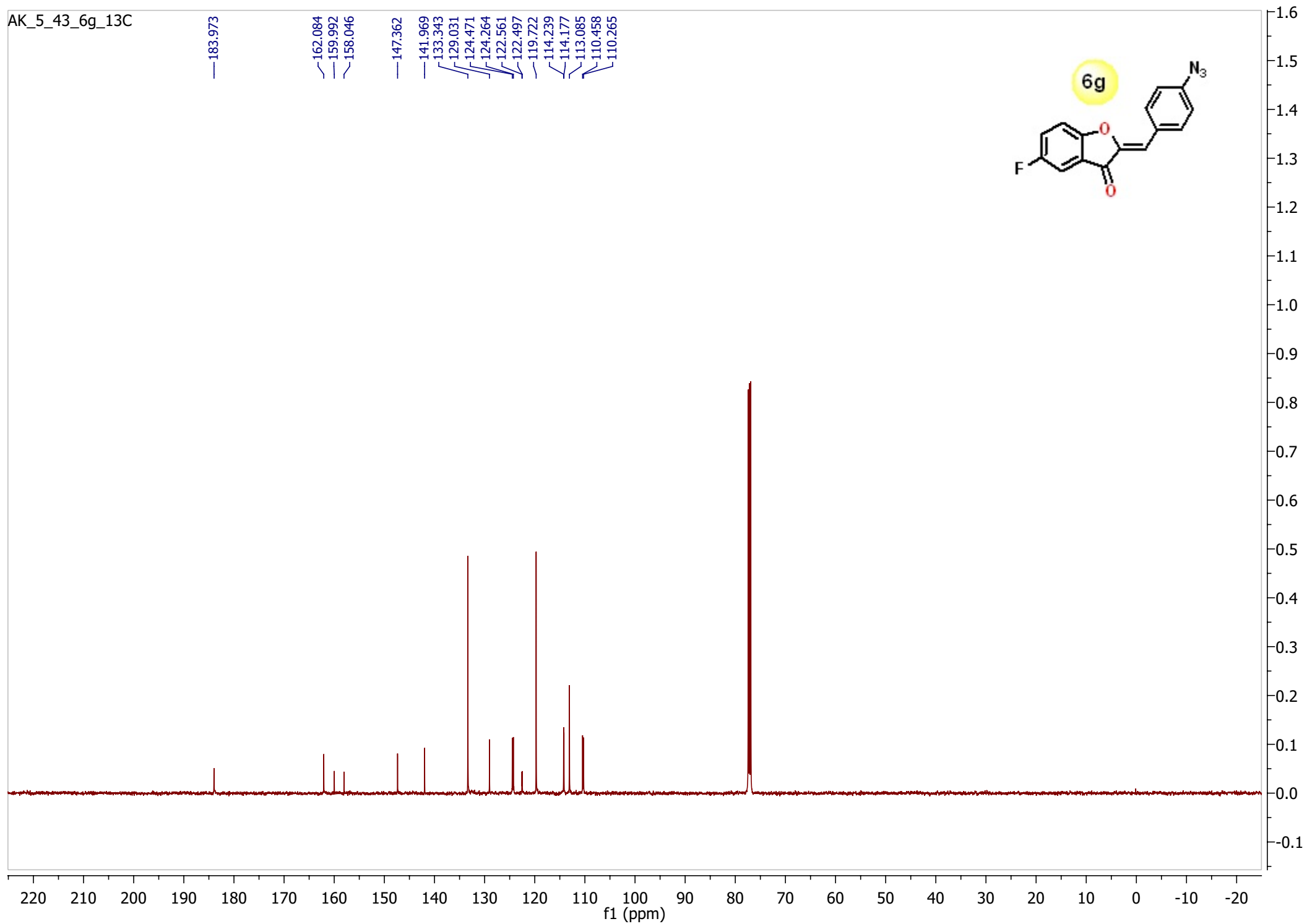
AK_5_41_13C



AK_5_43_6g_1H

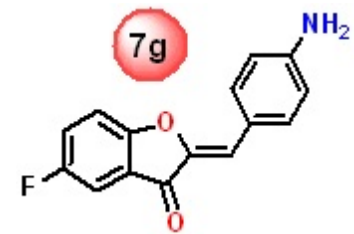


AK_5_43_6g_13C



AK_4_76

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7.607
7.600
7.591
7.582
7.577
7.573
7.563
7.558
6.877
6.671
6.654
6.156



2.03
1.98
1.11
1.00
2.02
1.80

12 11 10 9 8 7 6 5 4 3 2 1 0 -1 -2

f1 (ppm)

AK_4_76_amine

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159.771
156.583
152.048
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122.624
122.517
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116.236
114.773
114.666
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109.482
109.162

