

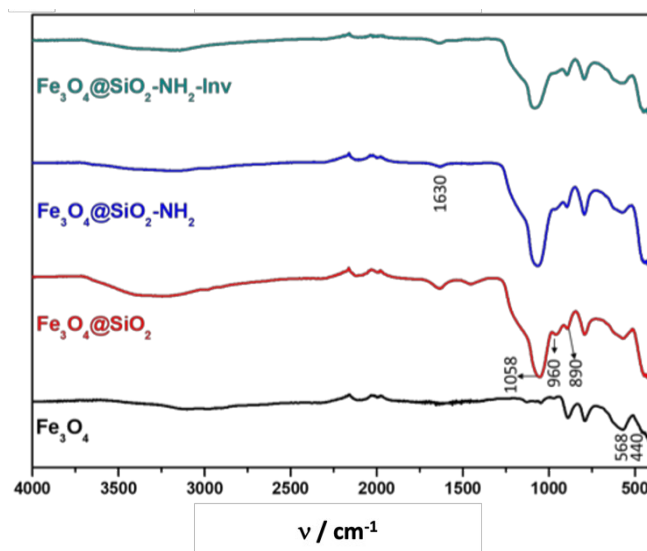
## Supporting Information

### Enzyme-functionalised, core/shell magnetic nanoparticles for selective pH-triggered sucrose capture

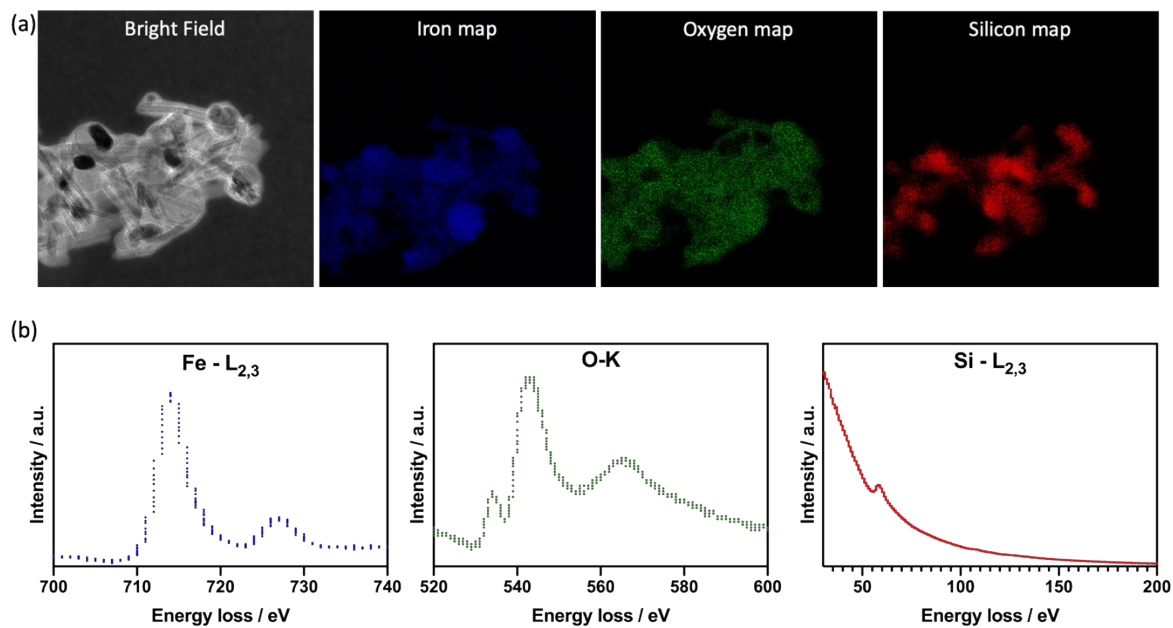
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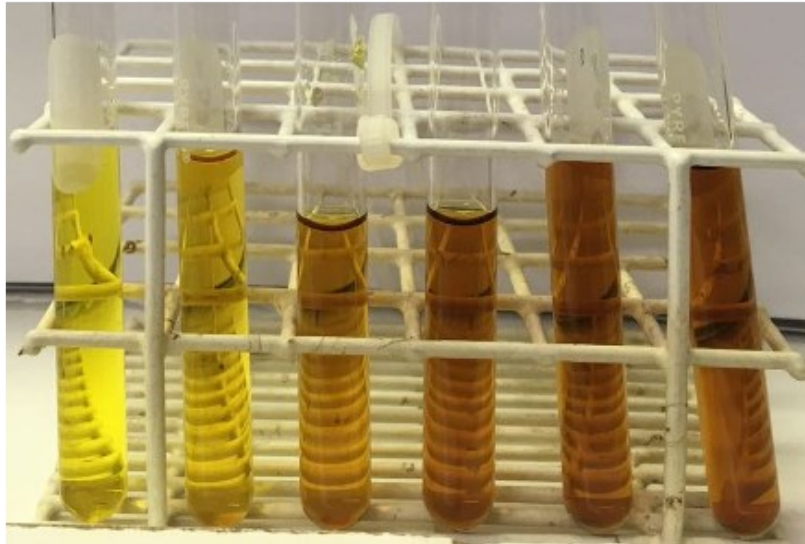
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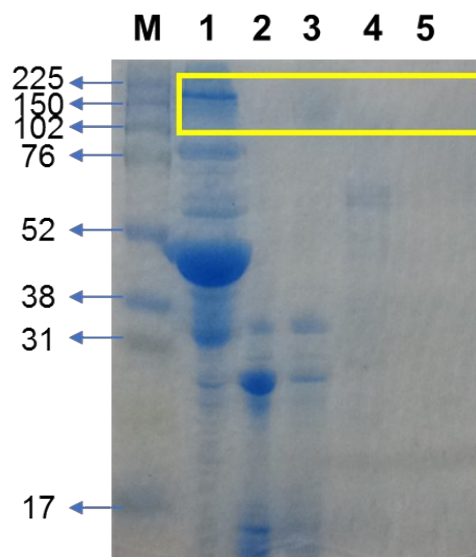
**Figure S1.** FT-IR spectra of the synthesised nanomaterials.



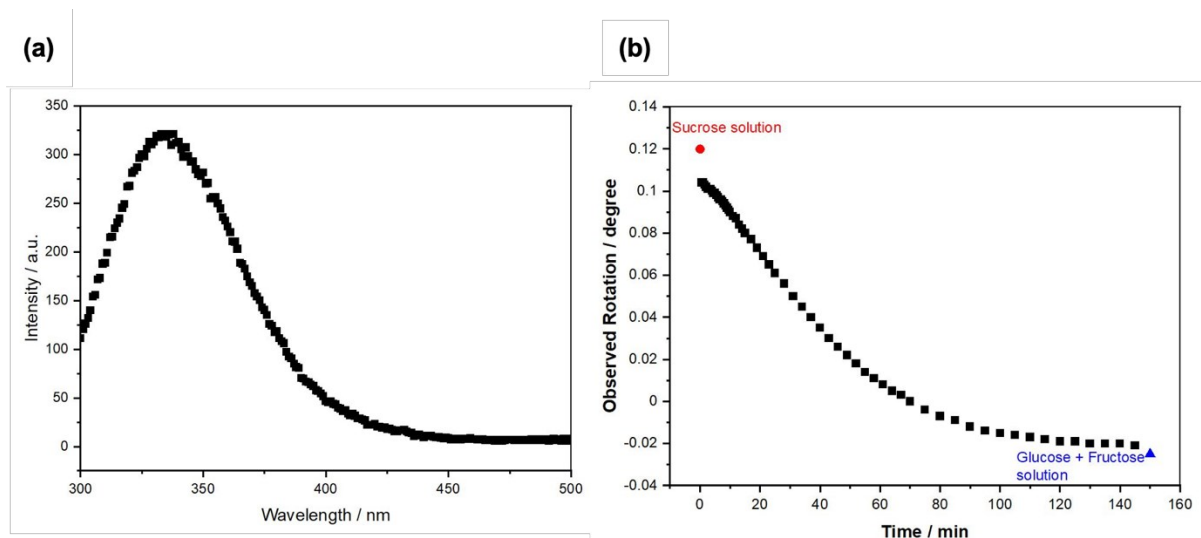
**Figure S2. (a)** ESI-TEM analysis on  $\text{Fe}_3\text{O}_4@SiO_2$  NPs: bright field (top left) and elemental mapping of iron (blue), oxygen (green) and silicon (red) atoms. **(b)** EEL spectra corresponding to the selected image, indicating iron (blue), oxygen (green) and silicon (red) atoms.



**Figure S3.** Photo images of the target enzyme activity test when increased sucrose concentration, from left to right, no invertase (blank, yellow) and to the highest sucrose concentration (the most intense brown).



**Figure S4.** SDS-PAGE gel image illustrating different stages of invertase purification. M - full-range rainbow MW marker (Amersham, GE Healthcare), 1 - extracted invertase, 2 - Superdex 200 eluate, 3 - DEAE-cellulose eluate, 4 and 5 - CM-sepharose eluates. Invertase is observed as a band between 100-200 kDa (yellow rectangle).



**Figure S5. (a)** Fluorescence spectrum of invertase at  $0.15 \text{ mg mL}^{-1}$  in sodium acetate buffer  $40 \text{ mmol L}^{-1}$ . **(b)** Polarimetry of sucrose solution ( $20 \text{ mg mL}^{-1}$ ) incubated with invertase ( $0.15 \text{ mg mL}^{-1}$ ), in sodium acetate buffer  $40 \text{ mmol L}^{-1}$  (pH 4.8).

**Table S1.** Comparison between the enzymatic activity of invertase immobilized on different nanoparticles

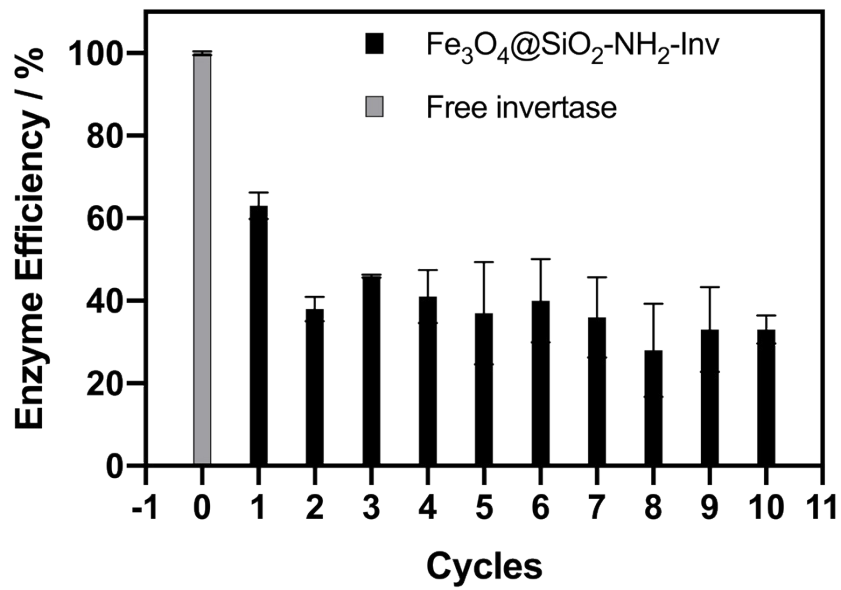
| Nanoparticle                                     | $K_M / \text{mM}$ | $V_{MAX} / \mu\text{M min}^{-1} \text{ mg enzyme}^{-1}$ | pH  | Temperature / °C | Reference  |
|--|-------------------|---|-----|------------------|------------|
| Free Invertase                                   | 67.7              | 50.4  | 4.8 | 50               | This study |
| $\text{Fe}_3\text{O}_4@\text{SiO}_2\text{-NH}_2$ | 98.3              | 97  |     |                  |            |
| Free Invertase                                   | 65.7              | 153.2   | 5   | 25               | 1          |
| PVlgMNP*   | 97.1              | 55.07   |     |                  |            |
| Free Invertase                                   | 65.7              | 153.2   | 5   | 25               | 2          |
| PAMAM-SPION**                                    | 92                | 40.8  |     |                  |            |
| Free Invertase                                   | 61.3              | 170.7   | 5   | 35               | 3          |
| Chitosan-MNPs***                                 | 81                | 60.8  |     |                  |            |
| Free Invertase                                   | 65.7              | 1670  | 4.5 | 55               | 4          |
| Chitosan NPs                                     | 205.7             | 1830 <sup>a</sup>                                       |     |                  |            |

\*Polyvinylimidazole grafted iron oxide nanoparticles

\*\*Polyamidoamine dendrimer grafted superparamagnetic iron oxide nanoparticles

\*\*\*Chitosan coated  $\gamma\text{-Fe}_2\text{O}_3$  magnetic nanoparticles

<sup>a</sup>U mL<sup>-1</sup>



**Figure S6.** Enzyme efficiency evaluation before and after immobilization onto Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub>-NH<sub>2</sub>-Inv using the enzyme activity test.

## References

1. K. Uzun, E. Çevik, M. Şenel, A. Baykal, *Bioprocess and Biosystems Engineering*, 2013, 36, 1807–1816.
2. K. Uzun, E. Çevik, M. Şenel, M. H. Sözeri, A. Baykal, M. F. Abasıyanık, M. S. Toprak, *Journal of Nanoparticle Research*, 2010, 12, 3057–3067.
3. P. P. Waifalkar, S. B. Parit, A. D. Chougale, S. C. Sahoo, P. S. Patil, P. B. Patil, *Journal of Colloid and Interface Science*, 2016, 482, 159-164.
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