## Enhanced fluorescence detection of nitroaromatic compound using bacteria embedded in porous poly lactic-co-glycolic acid microbeads

Tian Qiao<sup>a</sup>, Soohyun Kim<sup>a</sup>, Wonmok Lee<sup>b\*</sup>, and Hyunjung Lee<sup>a,\*</sup>

<sup>a</sup> Department of Materials Science and Engineering, Kookmin Univ.77 Jeongneung-ro,

Seongbuk-gu, Seoul, 02707, Republic of Korea

<sup>b</sup> Department of Chemistry, Sejong Univ., Neungdong-ro 209, Gwangjin-gu, Seoul,

143747, Republic of Korea

\* Corresponding authors.

Email address: <u>hyunjung@kookmin.ac.kr(H.Lee)</u>, <u>wonmoklee@sejong.ac.kr(W.Lee)</u>



**Fig. S1.** The structure of genetically engineered plasmid and mutated sequence information ( a) Structure of the plasmid of nitroaromatic sensitive bacteria (E. coli MG1655 with pPROBE  $-P_{yqiFmut}-gfp+$ )(b) Sequence of PyqjF – egfp+ (including mutated sequence) (c) Sensing mech anism of nitro aromatic compound inducible promoter-based biosensor



Fig. S2. SEM images to show morphology of fluorescent bacteria (a) and SEM images to sh ow existence of fluorescent bacteria on PLGA skeleton (b). Scale bars represent 500 nm



**Fig. S3.** The degradation behavior of open porous PLGA microbeads in PBS. (a) The FTIR spectra of porous PLGA microbeads before and after 14 day degradation. (b) T he derivative of TGA curves of open porous PLGA microbeads before and after 14 day degr adation.



Fig. S4. The different morphology and bacteria adhesion property of PLGA micrbeads as a f unction of the 1<sup>st</sup> stirring rpm.

The morphology of PLGA microbeads with 6000 1<sup>st</sup> rpm (a) ,12000 1<sup>st</sup> rpm (b) and 18000 1 st rpm (c), respectively. And (d),(e)and (f) are the relatively high magnification image of (a),( b) and (c), respectively.(g) is the average pore size of porous PLGA microbeads as a function of stirring speed during 1<sup>st</sup> emulsion. (h) The OD <sub>570nm</sub> value of PLGA microbeads after ba cteria incubation as a function of stirring speed during 1<sup>st</sup> emulsion.



**Fig. S5.** The CLSM morphology of the cross-section of PLGA microbeads with W/O ratio= 1:3.5 (a) and W/O ratio= 1:2 (b). Rhodamine B was used as the dry for CLSM and the excitation filter and emission filter was set up as 540 nm and 580 nm.

	10µg/mL DNT				100µg/mL DNT			
	M13 (0)	M13 (5 µg/mL)	M13 (20 µg/mL)	M	113 (0)	M13 (5 μg/mL)	M13 (20 µg/mL)	
3h		500µm	<u>500µm</u>	500µm	500µm	500µm	500µm	
	M13 (0)	M13 (5 µg/mL)	M13 (20 μg/mL)	M	113 (0)	M13 (5 µg/mL)	M13 (20 µg/mL)	
6h							643	
		500µm	500µm	500µm	<u>500µm</u>	500µm	500µm	
12h	M13 (0)	M13 (5 μg/mL)	М13 (20 µg/mL)	M	113 (0)	M13 (5 µg/mL)	M13 (20 µg/mL)	
		500µm	500µm	500µm	500µm	<u>500µm</u>	<u>500µm</u>	

Fig. S6. Fluorescence mode OM images of open porous PLGA microbeads attached with nitr oaromatic sensitive bacteria with 5  $\mu$ g/mL, 20  $\mu$ g/mL and without M13 bacteriophage in DNT solution.