

## Supporting Information

### **A thiocarbonate-caged fluorescent probe for specific visualization of peroxynitrite in living cells and zebrafish**

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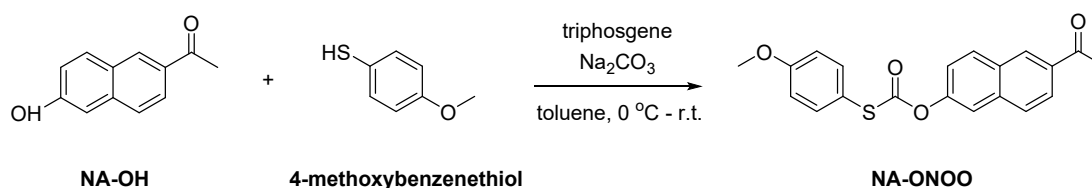
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## 1. Synthetic route of NA-ONOO



**Scheme S1.** Synthetic route of NA-ONOO

## 2. Preparation of analytes

ONOO<sup>-</sup>

0.6 M NaNO<sub>2</sub> and 0.7 M H<sub>2</sub>O<sub>2</sub> (acidified by 0.6 M HCl) were simultaneously and rapidly added into 1.2 M NaOH solution at 0 °C for stirring. After the solution is diluted 10 times, the concentration is calibrated by the absorbance at 301 nm.

(Extinction coefficient is 1670 cm<sup>-1</sup> M<sup>-1</sup>).

H<sub>2</sub>O<sub>2</sub>, TBHP, NaClO, NaNO<sub>2</sub>, KO<sub>2</sub>, Hcy, Cys, GSH

The above analytes with a concentration of 10 mM was prepared from the commercial available chemicals and solutions by ultra-pure water.

•OH

The hydroxyl radical was made by Fenton reaction, putting an equivalent amount of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) into the Iron dichloride solution (FeCl<sub>2</sub>).

NO

Nitric oxide (NO) was generated by sodium nitroprusside.

O<sup>2-</sup>

Superoxide (O<sup>2-</sup>) was generated from KO<sub>2</sub> in DMSO.

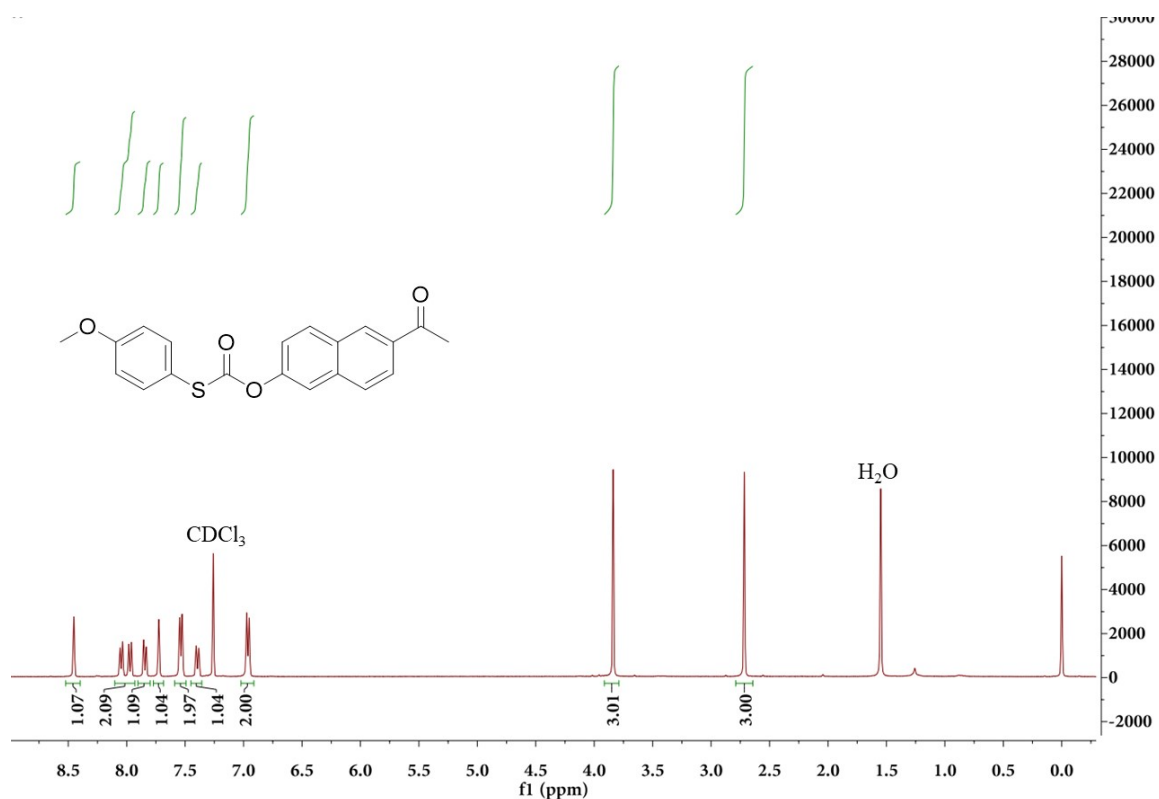
<sup>1</sup>O<sub>2</sub>

Singlet oxygen (<sup>1</sup>O<sub>2</sub>) was generated from NaOCl and H<sub>2</sub>O<sub>2</sub>.

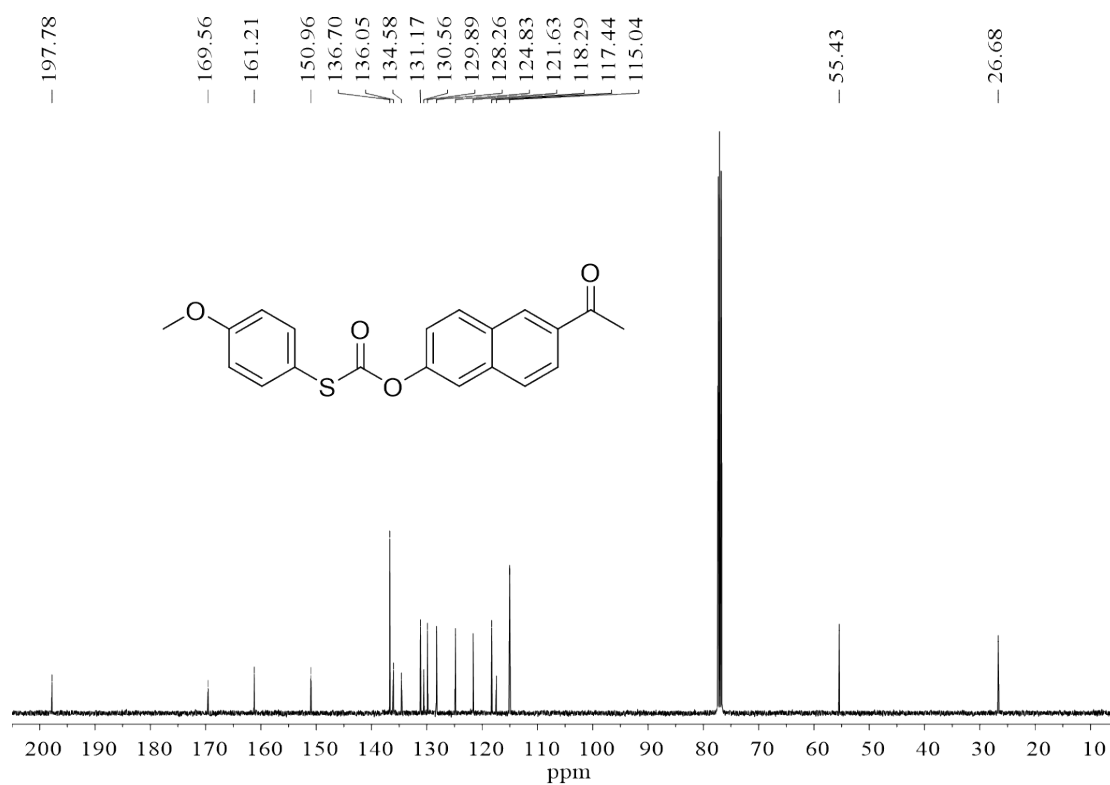
### 3. Cytotoxicity assays

HeLa cells were seeded into 96-well plates at a density of  $5 \times 10^3$  cells per well in culture media (DMEM) at 37 °C, 5% CO<sub>2</sub> and 95% air. Then, we cultured 0, 1, 5, 10, 20 and 50  $\mu\text{M}$  (final concentration) **NA-ONOO** with HeLa cells for 24 h. Finally, 10  $\mu\text{L}$  3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT, 5 mg mL<sup>-1</sup>) was added and cells were cultured for another 4 h. When the purple precipitate was clearly visible under the microscope, 100  $\mu\text{L}$  of DMSO was added to all the wells and swirled gently. Then, the absorbance in each well was measured, including that of the blanks, at 570 nm in a microtiter plate reader (Bio-Rad 680).

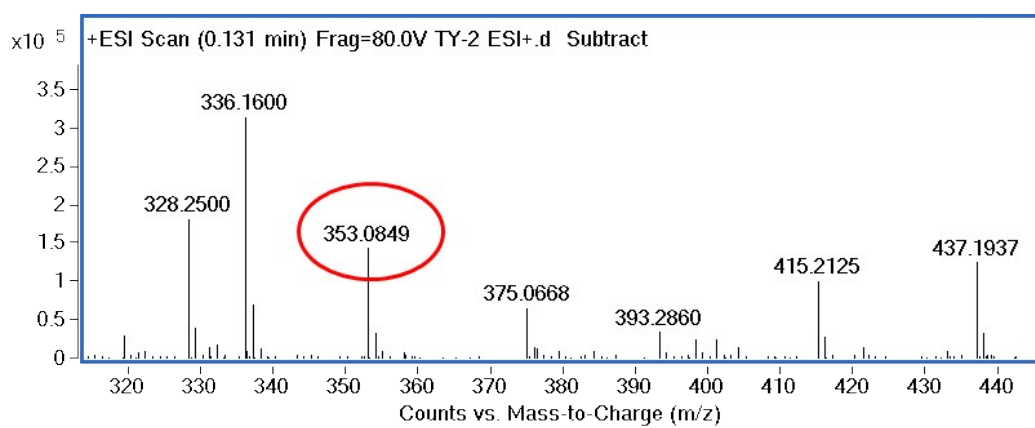
### 4. Characterization of NA-ONOO



**Fig. S1.** <sup>1</sup>H NMR spectrum of NA-ONOO



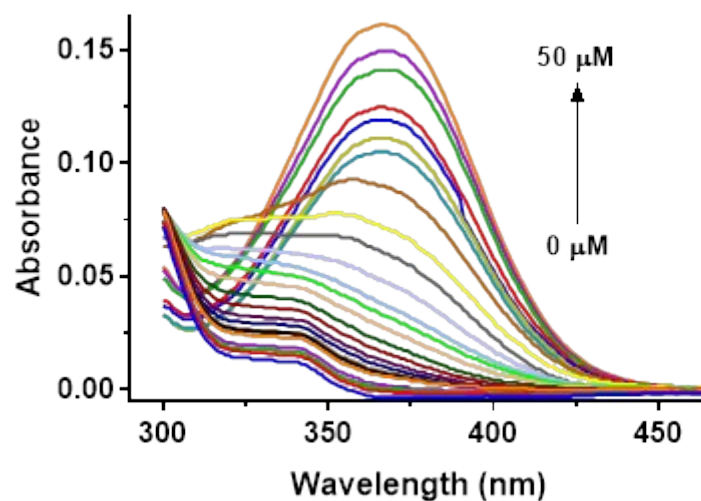
**Fig. S2.**  $^{13}\text{C}$  NMR spectrum of NA-ONOO



**g. S3.** HR-MS spectrum of NA-ONOO

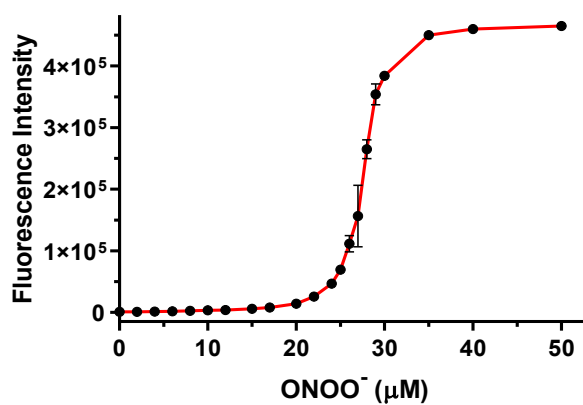
**Fi**

## 5. The dose-dependent absorption responses of probe NA-ONOO to ONOO<sup>-</sup>



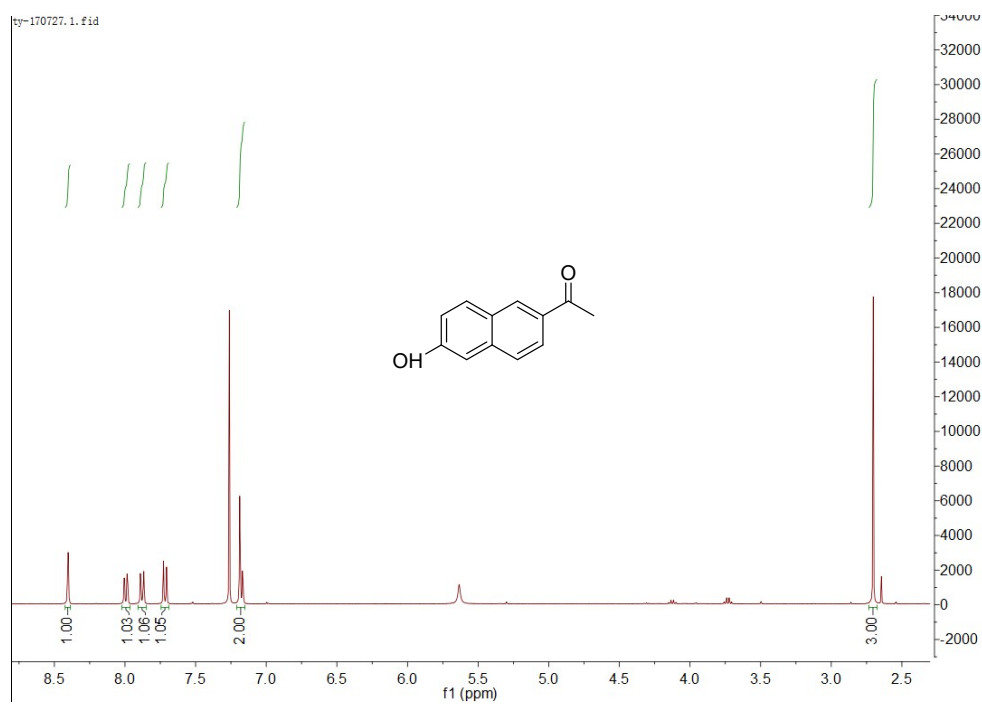
**Fig. S4.** Absorption spectra of probe NA-ONOO (10 μM) in the presence ONOO<sup>-</sup> (0–50 μM) in EtOH/PBS buffer (10 mM, pH 7.4, 1/1, V/V).

## 6. The dose-dependent fluorescence responses of probe NA-ONOO to ONOO<sup>-</sup>

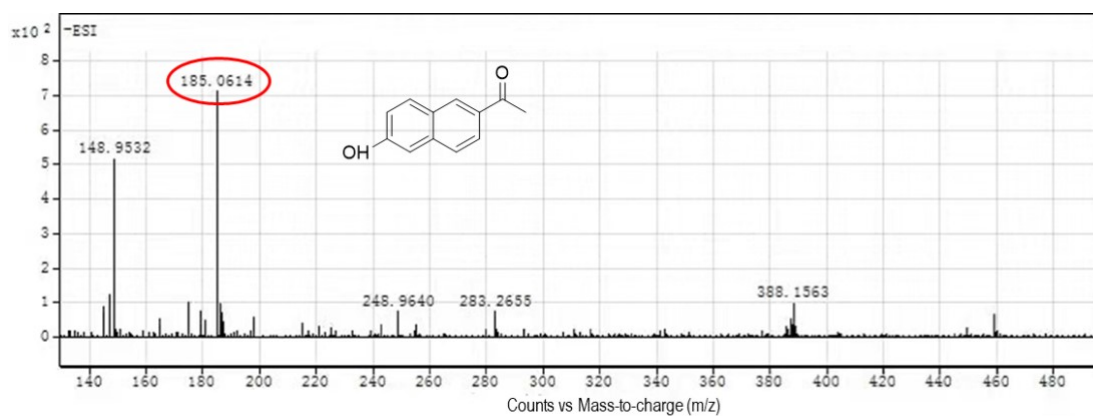


**Fig. S5.** (a) Fluorescence intensity profiles of NA-ONOO with 0–50 μM ONOO<sup>-</sup>.

## 7. Characterization of NA-ONOO reacting with ONOO<sup>-</sup>

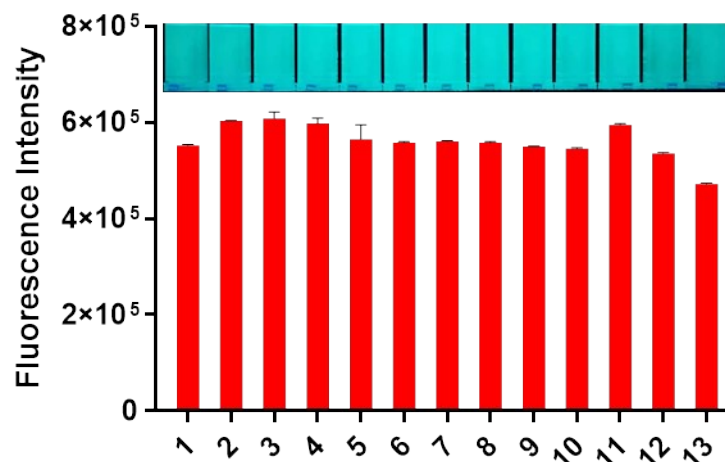


**Fig. S6.** <sup>1</sup>H NMR spectrum of NA-OH



**Fig. S7.** HR-MS spectrum of NA-OH

## 8. The interference experiments



**Fig. S8.** Fluorescence response of **NA-ONOO** ( $10 \mu\text{M}$ ) toward  $\text{ONOO}^-$  ( $50 \mu\text{M}$ ) in the presence of different analytes ( $200 \mu\text{M}$ ). (1) Blank; (2)  $\text{H}_2\text{O}_2$ ; (3)  $\text{OCl}^-$ ; (4) TBHP; (5)  $\text{NO}$ ; (6) Hcy; (7) Cys; (8) GSH; (9)  $\text{NO}_3^-$ ; (10)  $\text{NO}_2^-$ ; (11)  $^1\text{O}_2$ ; (12)  $\cdot\text{OH}$ ; (13)  $\text{O}_2^-$ . Inset: visual photographs of **NA-ONOO** in EtOH/PBS buffer ( $10 \text{ mM}$ , pH 7.4, V/V, 1/1) in the presence of  $\text{ONOO}^-$  with a series of interference biological reagents under handheld UV lamp.