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Electronic Supporting Information

2 **A ratiometric fluorescence strategy based on polyethyleneimine**
3 **surface-modified carbon dots and Eosin Y for ultrasensitive**
4 **determination of protamine and trypsin.**

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1 **Reagents**

2 The chemical reagents used in the experiments are of analytical grade without
3 further purification. The deionized water used in this experiment has a resistivity
4 greater than $18 \text{ M}\Omega \text{ cm}^{-1}$. PEI was provided by Aladdin Industrial Company. Tris-
5 HCl was purchased from Sinopharm Chemical Reagent Co. Ltd. Eosin Y, Protamine,
6 Arg, AA, Fru, Cys, GSH, Gly, Asp, ATP, Urease, G-ox, LZM and Pepsinand reduced
7 L-glutathione (GSH) were purchased from Sangon Biotech (Shanghai). Potassium
8 Chloride, Magnesium Chloride, Zinc Sulfate Sodium Chloride and Urea were
9 purchased from Beijing Chemical Works. Glucose and trypsin were purchased from
10 Aladdin. This experiment used a 10 mM Tris-HCl buffer solution to adjust the pH of
11 the reaction. And the configured solutions were all stored at 4°C .

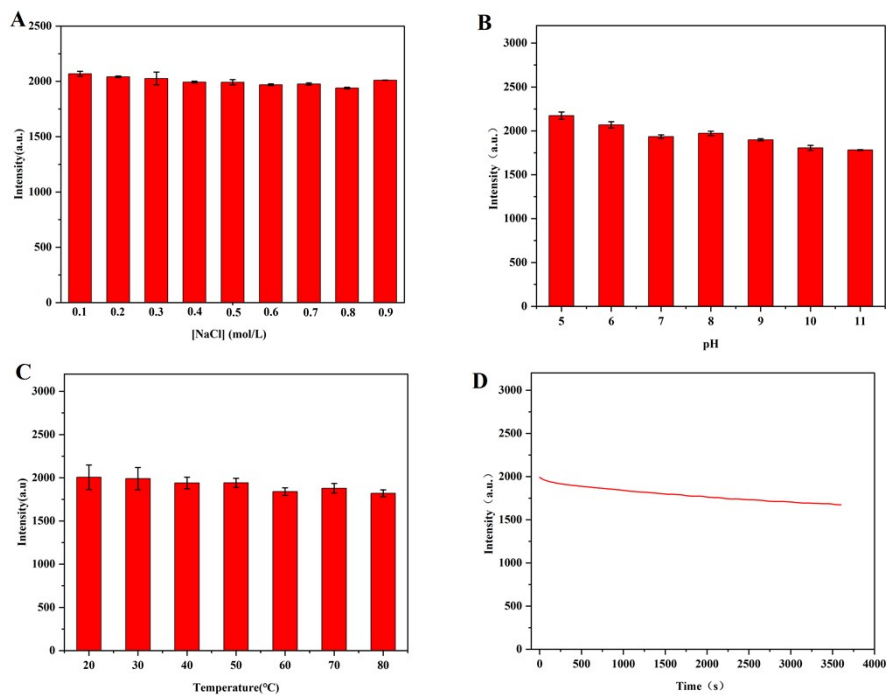
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13 **Instrumentation**

14 Fluorescence (FL) spectra were recorded by a RF-5301 PC
15 spectrofluorophotometer(Shimadzu, Japan). Ultraviolet-visible (UV-vis) absorption
16 spectra were acquired with the U-5100 UV-vis spectrophotometer. Powder X-ray
17 diffraction (XRD) patterns were conducted on a Rigaku D-Max 2550 diffractometer
18 (Rigaku, Japan). Fourier transform infrared (FT-IR) spectra were recorded on a
19 Nicolet 400 Fourier transform infrared spectrometer. Transmission electron
20 microscope (TEM) was collected on a JEM-2100F Transmission Electron Microscope
21 (JEOL, Japan). All temperature measurements were accomplished by using a water

1 bath. All pH measurements and the configured buffer solutions were performed using
2 a PHS-3C pH meter (Tuopu Co., Hangzhou, China).

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3 **Fig. S1.** Influences of (A) Ionic strength, (B) pH values, (C) Temperature, (D) Xenon
4 lamp irradiation time on the fluorescence of CDs-PEI at 350 nm excitation wavelength.

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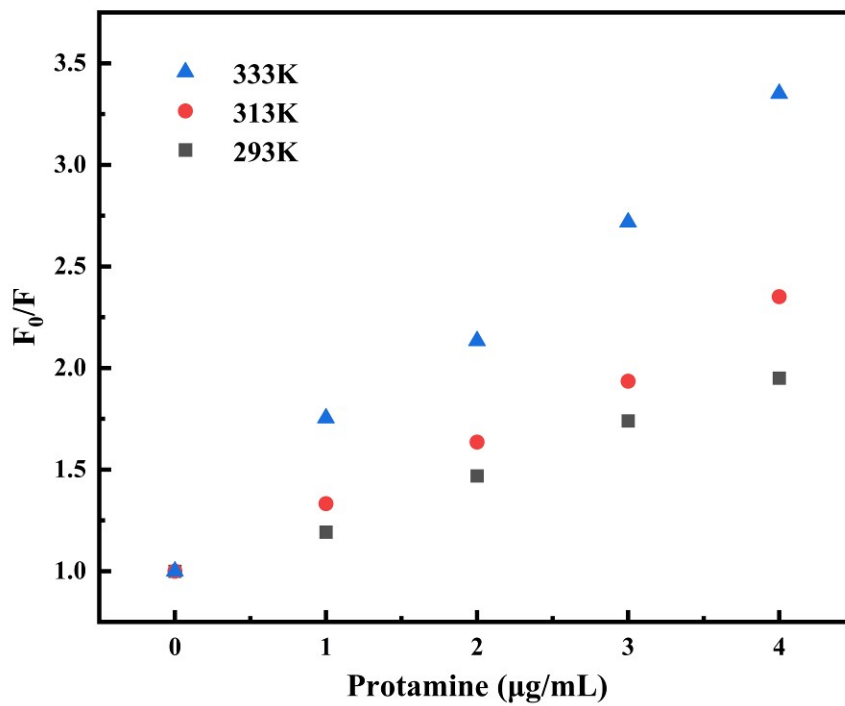
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3 **Fig. S2.** Stern–Volmer curves for the CDs-PEI/Eosin Y/Protamine system at three
 4 different temperatures. (F_0 and F were the fluorescence intensity of Eosin Y in the
 5 absence and presence of protamine, respectively.)

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1 **Table S1 Comparison with previous methods for the detection of protamine.**

Methods	Materials	Linear range ($\mu\text{g/mL}$)	LOD ($\mu\text{g/mL}$)	Reference
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Fluorescence	water soluble Perylene diimides	0.1-13 µg /mL	0.09 µg /mL	1
Methods	Materials	Linear range	LOD	Reference
Electrochemical	poly(thionine) modified glassy	4-22 µg /mL	0.28 µg /mL	2
Chemiluminescence	gold nanoclusters carbon electrode (Au NCs)	2.4-48 µg/mL	0.19 µg/mL	7
Colorimetric	Anionic poly(2-(2-(4-methylthiophen-3-ylloxy)ethyl)malonate acid)	0.1-30 µg /mL	0.1 µg /mL	3
Fluorescence	a pyrene derivative	0.5–8 µg/mL	0.5 µg/mL	4
Fluorescence	BZA-BOD@ZIF-90	0.25-0.4 µg/mL	0.07 µg /mL	5
Robust reverse phase-HPLC	Zorbax-SB C8 Column	0.5–5 mg/ml	0.06 mg/mL	6
Fluorescence	PEI-CDs and Eosin Y	0.1-5.2 µg/mL	0.03 µg/mL	This work

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Chemiluminescence	bovine serum albumin (BSA)-stabilized gold nanoclusters (Au NCs)	0.01-50 $\mu\text{g}/\text{mL}$	9 ng/mL	8
Electrochemical	luminol and black phosphorus nanosheets	0.1 – 5 $\mu\text{g}/\text{mL}$	63.3 ng/mL	9
Colorimetric	silver nanoparticles (AgNPs)	2.5-200 ng/mL	2 ng/mL	10
Fluorescence	silicon quantum dots (SiQDs) and triangular silver nanoprisms (TSNPRs)	0-40 ng/mL	8 ng/mL	11
Fluorescence	AgInZnS QD	0.1-4 $\mu\text{g}/\text{mL}$	0.04 $\mu\text{g}/\text{mL}$	12
Fluorescence	PEI-CDs and Eosin Y	0.4-56 ng/mL	0.21 ng/mL	This work

1 **Table S2** Comparison with previous methods for the detection of trypsin

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