

Palladium-Catalyzed One-Pot Phosphorylation of Phenols Mediated by Sulfuryl Fluoride

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I. General Information.

All commercially available organic compounds and DNA headpiece HP-NH₂ (5'- / 5phos / GAGTCA / iSp9 / iUniAmM / iSp9 / TGACTCCC-3', Figure 1) were purchased from commercial sources unless otherwise noted and used as received. The substrates **1** were prepared according to our previous paper.¹ ¹H NMR, ¹³C NMR, ³¹P NMR and ¹⁹F NMR spectra were recorded on Bruker AM-500 instruments. High-resolution mass spectra (HRMS-ESI) were obtained on an Agilent Technologies 6230 Accurate Mass TOF LC/MS instrument or an AB SCIEX 4600 QTOF MS instrument. All on-DNA reactions were performed in 1.5 mL or 5 mL Eppendorf tubes. On-DNA reaction were analyzed by LC-MS. Typically, samples were dissolved in an appropriate amount of distilled and deionized water (ddH₂O) and injected into a reverse-phase chromatography column (Xbridge Oligonucleotide BEH C18 column, 1.7 μm, 2.1×50 mm). The elution was carried out as followings: 5–95% solvent B over 4.5 min, 0.4 mL/min, λ = 260 nm; solvent A: 0.75% v/v hexafluoroisopropanol/ 0.038% v/v triethylamine in methanol/water = 5/95; solvent B: 0.75% v/v hexafluoroisopropanol/ 0.038% v/v triethylamine in methanol/water = 90/10. The effluents were analyzed by a Waters G2 TOF electrospray mass spectrometer in negative ion mode. Ignoring UV coefficient difference for all DNA products and assuming 100% of DNA total recovery, the yield of DNA products was determined from UV absorbance trace (260 nm) peak area using the equation below:

$$Yield (product, \%) = \frac{\% \text{ UV (product)}}{\% \text{ UV (DNA starting material before reaction)}} \times 100\%$$

Caution: Sulfuryl fluoride (SO₂F₂, MW = 102 g/mol, ρ = 4.17 g/cm³, bp = -55.20 °C) is a toxic gas that can resistant to hydrolysis up to 150 °C. The use of SO₂F₂ must be executed in a well ventilated fumehood. The excess SO₂F₂ can be quenched by basic aqueous medium.

II. Experimental Procedures and Characterizations:

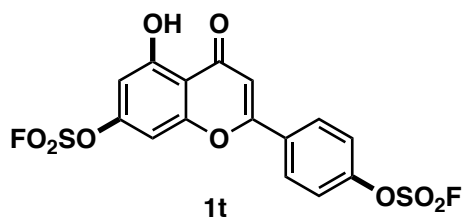
Table S1. Screening of the reaction conditions

entry	Derivation from the standard conditions	Yield (%) ^[b]
1 ^[a]	none	93 (87) ^[c]
2	without Pd(OAc) ₂ and dppf, 80 °C	0
3	without dppf, 80 °C	5
4	dppp (instead of dppf), 80 °C	48
5	BINAP (instead of dppf), 80 °C	53
6	Xphos (instead of dppf), 80 °C	58
7	Xantophos (instead of dppf), 80 °C	75
8	Pd(PPh ₃) ₂ (instead of Pd(OAc) ₂ and dppf), 80 °C	55
9	Pd(OAc) ₂ (10 mol%) and dppf (20 mol%), 80 °C	95
10	Pd(OAc) ₂ (2 mol%) and dppf (4 mol%), 80 °C	90
11	in the absence of argon, 80 °C	45
12	60 °C	92
13	40 °C	11
14	2a (2 equiv), CH ₃ CN-H ₂ O (4:1) as solvent, 80 °C	93
15	DMF as solvent, 80 °C	59
16	DMA as solvent, 80 °C	62
17	1,4-dioxane as solvent, 80 °C	47
18	Et ₃ N as base, 80 °C	90
19	Na ₂ CO ₃ as base, 80 °C	34

^[a]Standard conditions: **1a** (0.2 mmol), **2a** (0.25 mmol), DIPEA (0.4 mmol), Pd(OAc)₂ (5 mol %)/dppf (10 mol %) in CH₃CN (1 mL) were stirred at 80 °C for 5h under argon. ^[b]yield determined by LC-MS. ^[c]Isolated yield.

General Procedure for the Synthesis of Aryl Fluorosulfates

A 100 mL flask equipped with a magnetic stir bar was charged with the relative phenol (5 mmol, 1 equiv), Et₃N or DIPEA (10 mmol, 2 equiv) and DCM (25 mL). Sealed with a rubber septum, the flask was evacuated to low vacuum and backfilled with sulfuryl fluoride gas by a balloon. The resulting mixture was stirred at room temperature for 3~5 h. Upon completion, volatiles were removed in vacuo. The crude product was purified by flash chromatography on silica gel to give the desired aryl fluorosulfate.



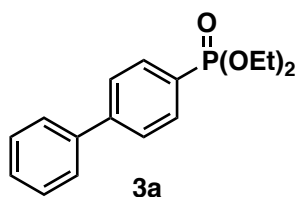
Yellow solid, 95% yield. ^1H NMR (500 MHz, DMSO- d_6) δ 12.96 (s, 1H), 8.40 – 8.34 (m, 2H), 7.90 – 7.85 (m, 2H), 7.65 (d, $J = 2.3$ Hz, 1H), 7.35 (s, 1H), 7.18 (d, $J = 2.3$ Hz, 1H). ^{13}C NMR (126 MHz, DMSO) δ 183.08, 163.32, 162.01, 156.91, 153.57, 152.30, 131.57, 130.05, 122.60, 110.88, 107.86, 105.15, 101.92. ^{19}F NMR (376 MHz, DMSO) δ 40.56, 39.74. HRMS (ESI): Calcd for $\text{C}_{15}\text{H}_9\text{F}_2\text{O}_9\text{S}_2$: $[\text{M}+\text{H}]^+$ 434.9656, found: m/z 434.9650.

Standard Conditions: Procedure for the Phosphorylation of Fluorosulfates

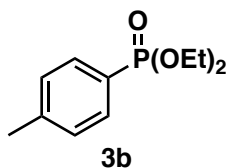
A 25 mL flask equipped with a magnetic stir bar was charged with the relative fluorosulfate **1** (0.2 mmol), P(O)H compound **2** (0.25 mmol), Pd(OAc) $_2$ (5 mol %), dppf (10 mol %), DIPEA (0.4 mmol) and CH $_3$ CN (1 mL). Sealed with a rubber septum, the flask was evacuated to low vacuum and backfilled with argon at room temperature. Then the resulting mixture was stirred at 80 °C for 5h. Upon completion, volatiles were removed in vacuo. The crude product was purified by flash chromatography on silica gel to give the desired product **3**.

One-Pot Synthesis: Procedure for the Phosphorylation of Phenols

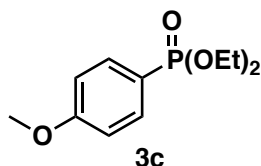
A 25 mL flask equipped with a magnetic stir bar was charged with the relative phenol (0.2 mmol), DIPEA (0.4 mmol) and CH $_3$ CN (1 mL). Sealed with a rubber septum, the flask was evacuated to low vacuum and backfilled with sulfuryl fluoride gas by a balloon. The resulting mixture was stirred at room temperature for 3 h. Upon completion, the sulfuryl fluoride gas balloon was removed and the flask was purged with argon for 3 times. After that, TMSOH (0.6 mmol) was added and stirred for 30 min to remove the fluoride ion, followed by addition of P(O)H compound **2** (0.25 mmol), Pd(OAc) $_2$ (5 mol %), dppf (10 mol %), DIPEA (0.4 mmol). Sealed with a rubber septum, the flask was evacuated to low vacuum and backfilled with argon at room temperature. Then resulting mixture was stirred at 80 °C for 5h. Upon completion, volatiles were removed in vacuo. The crude product was purified by flash chromatography on silica gel to give the desired product **3**.



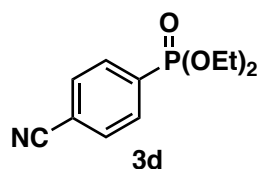
Colorless oil, 87% yield, (90% for one-pot). ^1H NMR (500 MHz, CDCl $_3$) δ 7.92 – 7.85 (m, 2H), 7.70 – 7.67 (m, 2H), 7.62 – 7.59 (m, 2H), 7.49 – 7.44 (m, 2H), 7.42 – 7.38 (m, 1H), 4.23 – 4.05 (m, 4H), 1.35 (t, $J = 7.1$ Hz, 6H). These data are in agreement with literature data.²



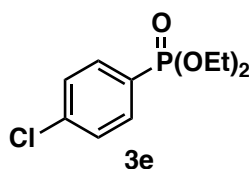
Colorless oil, 92% yield, (91% for one-pot). ^1H NMR (500 MHz, CDCl $_3$) δ 7.70 (dd, $J = 12.9, 7.8$ Hz, 2H), 7.29 – 7.26 (m, 2H), 4.22 – 3.99 (m, 4H), 2.40 (s, 3H), 1.31 (t, $J = 7.0$ Hz, 6H). These data are in agreement with literature data.³



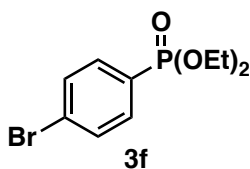
Colorless oil, 90% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.78 – 7.70 (m, 2H), 7.01 – 6.91 (m, 2H), 4.17 – 3.97 (m, 4H), 3.85 (s, 3H), 1.31 (t, $J = 7.0$ Hz, 6H). These data are in agreement with literature data.⁴



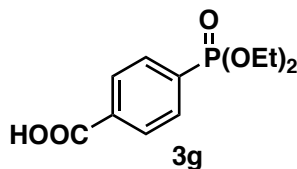
Colorless oil, 84% yield, (90% for one-pot). $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.92 (dd, $J = 12.9, 7.8$ Hz, 2H), 7.76 (dd, $J = 8.1, 2.8$ Hz, 2H), 4.24 – 4.06 (m, 4H), 1.34 (t, $J = 7.0$ Hz, 6H). These data are in agreement with literature data.⁵



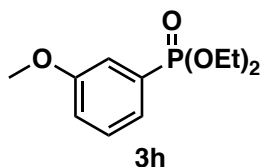
Colorless oil, 93% yield, (88% for one-pot). $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.75 (dd, $J = 12.5, 8.0$ Hz, 2H), 7.45 (dd, $J = 8.0, 2.1$ Hz, 2H), 4.23 – 4.00 (m, 4H), 1.32 (t, $J = 7.0$ Hz, 6H). These data are in agreement with literature data.⁵



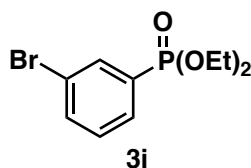
Colorless oil, 90% yield, (87% for one-pot). $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.67 (dd, $J = 12.7, 8.2$ Hz, 2H), 7.61 (dd, $J = 8.3, 3.4$ Hz, 2H), 4.20 – 4.02 (m, 4H), 1.32 (t, $J = 7.0$ Hz, 6H). These data are in agreement with literature data.⁶



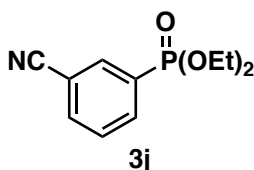
White solid, 93% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 11.23 – 11.03 (m, 1H), 8.19 (dd, $J = 8.1, 3.5$ Hz, 2H), 7.93 (dd, $J = 13.0, 7.9$ Hz, 2H), 4.27 – 4.09 (m, 4H), 1.34 (t, $J = 7.0$ Hz, 6H). These data are in agreement with literature data.⁷



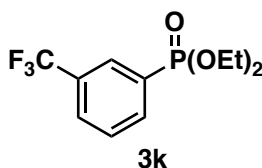
Colorless oil, 82% yield, (80% for one-pot). $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.40 – 7.31 (m, 3H), 7.11 – 7.05 (m, 1H), 4.19 – 4.03 (m, 4H), 3.84 (s, 3H), 1.32 (t, $J = 7.1$ Hz, 6H). These data are in agreement with literature data.⁸



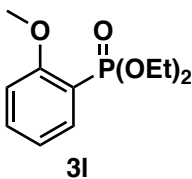
Colorless oil, 93% yield, $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.94 (d, $J = 12.5$ Hz, 1H), 7.74 (dd, $J = 12.4, 7.5$ Hz, 1H), 7.68 (dd, $J = 8.1, 1.7$ Hz, 1H), 7.35 (dt, $J = 9.1, 4.5$ Hz, 1H), 4.19 – 4.07 (m, 4H), 1.34 (t, $J = 6.9$ Hz, 6H). These data are in agreement with literature data.⁹



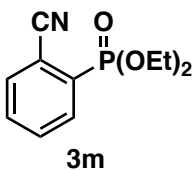
Colorless oil, 85% yield, (82% for one-pot). $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.08 (dd, $J = 13.3, 1.7$ Hz, 1H), 8.06 – 8.01 (m, 1H), 7.82 (dq, $J = 7.8, 1.2$ Hz, 1H), 7.60 (td, $J = 7.7, 3.5$ Hz, 1H), 4.25 – 4.06 (m, 4H), 1.34 (t, $J = 7.0$ Hz, 6H). These data are in agreement with literature data.¹⁰



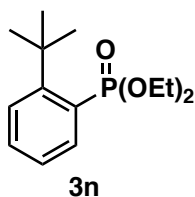
Colorless oil, 92% yield, (89% for one-pot). $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.07 (d, $J = 13.6$ Hz, 1H), 8.00 (dd, $J = 13.0, 7.6$ Hz, 1H), 7.81 (d, $J = 7.9$ Hz, 1H), 7.61 (td, $J = 7.7, 3.7$ Hz, 1H), 4.24 – 4.06 (m, 4H), 1.34 (t, $J = 7.1$ Hz, 6H). These data are in agreement with literature data.⁵



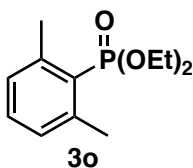
Colorless oil, 80% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.82 (ddd, $J = 14.9, 7.6, 1.8$ Hz, 1H), 7.54 – 7.47 (m, 1H), 7.05 – 6.98 (m, 1H), 6.98 – 6.91 (m, 1H), 4.24 – 4.07 (m, 4H), 3.90 (s, 3H), 1.33 (t, $J = 7.1$ Hz, 6H). These data are in agreement with literature data. These data are in agreement with literature data.¹¹



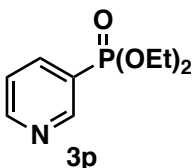
Colorless oil, 67% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.12 (dd, $J = 14.1, 7.4$ Hz, 1H), 7.84 – 7.77 (m, 1H), 7.73 – 7.62 (m, 2H), 4.32 – 4.13 (m, 4H), 1.38 (t, $J = 7.0$ Hz, 6H). These data are in agreement with literature data.¹²



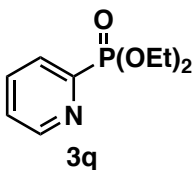
Colorless oil, 53% yield. ¹H NMR (500 MHz, DMSO-*d*₆) δ 7.89 (ddd, *J* = 14.9, 7.8, 1.6 Hz, 1H), 7.60 (td, *J* = 7.8, 7.4, 1.3 Hz, 1H), 7.53 (tt, *J* = 8.4, 1.7 Hz, 1H), 7.35 (tdd, *J* = 7.4, 2.6, 1.3 Hz, 1H), 4.09 – 3.95 (m, 4H), 1.51 (s, 9H), 1.24 (t, *J* = 7.1 Hz, 6H). ¹³C NMR (126 MHz, DMSO) δ 155.02, 154.92, 135.76, 135.70, 132.80, 132.78, 127.89, 127.77, 127.63, 126.21, 126.14, 126.04, 61.97, 61.93, 37.14, 37.12, 32.05, 16.53, 16.48. ³¹P NMR (202 MHz, DMSO) δ 19.88. HRMS (ESI): Calcd for C₁₄H₂₃O₃P: [M+H]⁺ 271.1463, found: *m/z* 271.1458.



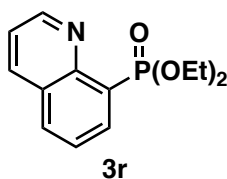
Colorless oil, 58% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.26 (t, 1H), 7.08 (dd, *J* = 7.6, 4.7 Hz, 2H), 4.16 (dp, *J* = 10.0, 7.2 Hz, 2H), 4.05 (dp, *J* = 10.0, 7.2 Hz, 2H), 2.64 (d, *J* = 1.6 Hz, 6H), 1.32 (t, *J* = 7.1 Hz, 6H). These data are in agreement with literature data.¹³



Colorless oil, 90% yield, (82% for one-pot). ¹H NMR (500 MHz, CDCl₃) δ 8.97 (dd, *J* = 6.6, 2.0 Hz, 1H), 8.77 (dt, *J* = 4.5, 2.1 Hz, 1H), 8.13 (ddt, *J* = 13.3, 7.8, 1.9 Hz, 1H), 7.43 (dt, *J* = 8.1, 4.1 Hz, 1H), 4.24 – 4.07 (m, 4H), 1.34 (t, *J* = 7.1 Hz, 6H). These data are in agreement with literature data.¹⁴

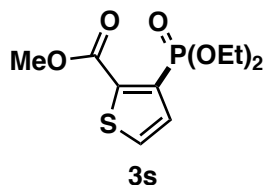


Colorless oil, 90% yield, (93% for one-pot). ¹H NMR (500 MHz, CDCl₃) δ 8.78 (d, *J* = 4.6 Hz, 1H), 7.95 (t, *J* = 7.2 Hz, 1H), 7.79 (tdd, *J* = 7.6, 5.5, 1.5 Hz, 1H), 7.41 (dd, *J* = 8.5, 4.3 Hz, 1H), 4.29 – 4.14 (m, 4H), 1.32 (t, *J* = 7.1 Hz, 6H). These data are in agreement with literature data.¹⁵

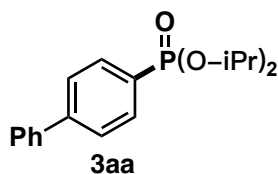


Colorless oil, 91% yield, (83% for one-pot). ¹H NMR (500 MHz, DMSO) δ 9.02 (dd, *J* = 4.2, 1.8 Hz, 1H), 8.47 (dt, *J* = 8.3, 2.1 Hz, 1H), 8.29 – 8.19 (m, 2H), 7.71 (ddd, *J* = 8.1, 7.1, 3.5 Hz, 1H),

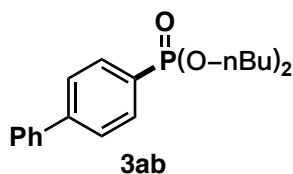
7.63 (dd, $J = 8.3, 4.2$ Hz, 1H), 4.17 (dq, $J = 8.7, 7.1, 3.8$ Hz, 4H), 1.24 (t, $J = 7.1$ Hz, 6H). ^{13}C NMR (126 MHz, DMSO) δ 151.36, 147.86, 147.81, 137.18, 136.26, 136.20, 133.55, 133.52, 129.46, 128.43, 128.35, 127.98, 126.46, 126.33, 122.54, 62.33, 62.28, 16.79, 16.74. ^{31}P NMR (202 MHz, DMSO) δ 16.00. HRMS (ESI): Calcd for $\text{C}_{13}\text{H}_{17}\text{NO}_3\text{P}$: $[\text{M}+\text{H}]^+$ 266.0946, found: m/z 266.0934.



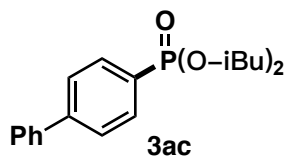
Colorless oil, 83% yield, (70% for one-pot). ^1H NMR (500 MHz, $\text{DMSO}-d_6$) δ 8.00 (dd, $J = 5.1, 3.0$ Hz, 1H), 7.42 (t, $J = 4.8$ Hz, 1H), 4.10 (dq, $J = 8.6, 7.0, 1.3$ Hz, 4H), 3.85 (s, 3H), 1.26 (t, $J = 7.0$ Hz, 6H). ^{13}C NMR (126 MHz, DMSO) δ 161.03, 161.01, 138.47, 138.35, 135.64, 134.11, 133.91, 133.80, 133.08, 132.93, 62.65, 62.60, 53.16, 16.67, 16.62. ^{31}P NMR (202 MHz, DMSO) δ 9.20. HRMS (ESI): Calcd for $\text{C}_{10}\text{H}_{16}\text{O}_5\text{PS}$: $[\text{M}+\text{H}]^+$ 279.0456, found: m/z 279.0445.



Colorless oil, 82% yield, (75% for one-pot). ^1H NMR (500 MHz, CDCl_3) δ 7.92 – 7.85 (m, 2H), 7.69 – 7.65 (m, 2H), 7.63 – 7.58 (m, 2H), 7.50 – 7.44 (m, 2H), 7.43 – 7.35 (m, 1H), 4.72 (dp, $J = 7.8, 6.2$ Hz, 2H), 1.39 (d, $J = 6.2$ Hz, 6H), 1.26 (d, $J = 6.2$ Hz, 6H). These data are in agreement with literature data.¹⁶

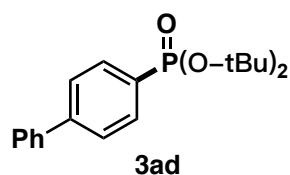


Colorless oil, 90% yield, (84% for one-pot). ^1H NMR (500 MHz, $\text{DMSO}-d_6$) δ 7.88 – 7.80 (m, 2H), 7.82 – 7.76 (m, 2H), 7.76 – 7.71 (m, 2H), 7.51 (dd, $J = 8.4, 6.9$ Hz, 2H), 7.47 – 7.40 (m, 1H), 4.06 – 3.92 (m, 4H), 1.60 (dq, $J = 8.5, 6.5$ Hz, 4H), 1.41 – 1.30 (m, 4H), 0.87 (t, $J = 7.4$ Hz, 6H). ^{13}C NMR (126 MHz, DMSO) δ 144.46, 144.44, 139.41, 132.44, 132.36, 129.56, 128.80, 128.32, 127.47, 127.44, 127.35, 126.82, 65.68, 65.63, 32.40, 32.36, 18.71, 13.84. ^{31}P NMR (202 MHz, DMSO) δ 17.96. HRMS (ESI): Calcd for $\text{C}_{20}\text{H}_{28}\text{O}_3\text{P}$: $[\text{M}+\text{H}]^+$ 347.1776, found: m/z 347.1763.

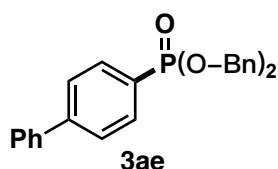


Colorless oil, 87% yield, (82% for one-pot). ^1H NMR (500 MHz, $\text{DMSO}-d_6$) δ 7.88 – 7.77 (m, 4H), 7.76 – 7.71 (m, 2H), 7.51 (dd, $J = 8.4, 6.9$ Hz, 2H), 7.47 – 7.40 (m, 1H), 3.77 (qt, $J = 9.8, 6.6$ Hz, 4H), 1.89 (dh, $J = 13.2, 6.6$ Hz, 2H), 0.90 (d, $J = 6.7$ Hz, 12H). ^{13}C NMR (126 MHz, DMSO) δ

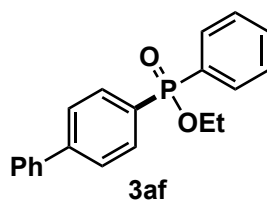
144.46, 144.44, 139.40, 132.45, 132.37, 129.58, 128.83, 128.24, 127.50, 127.45, 127.38, 126.73, 71.80, 71.75, 29.17, 29.12, 18.99. ^{31}P NMR (202 MHz, DMSO) δ 17.75. HRMS (ESI): Calcd for $\text{C}_{20}\text{H}_{28}\text{O}_3\text{P}$: $[\text{M}+\text{H}]^+$ 347.1776, found: m/z 347.1770.



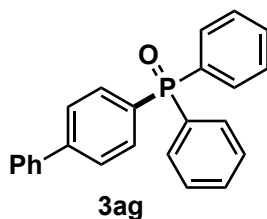
Colorless oil, 80% yield, (73% for one-pot). ^1H NMR (500 MHz, $\text{DMSO}-d_6$) δ 7.80 – 7.74 (m, 4H), 7.73 – 7.68 (m, 2H), 7.49 (dd, $J = 8.4, 6.9$ Hz, 2H), 7.44 – 7.38 (m, 1H), 1.42 (s, 18H). ^{13}C NMR (126 MHz, DMSO) δ 143.49, 143.47, 139.64, 133.70, 132.16, 132.07, 131.99, 129.50, 128.60, 127.38, 127.09, 126.97, 82.22, 82.16, 30.56, 30.53. ^{31}P NMR (202 MHz, DMSO) δ 9.76. HRMS (ESI): Calcd for $\text{C}_{20}\text{H}_{28}\text{O}_3\text{P}$: $[\text{M}+\text{H}]^+$ 347.1776, found: m/z 347.1768.



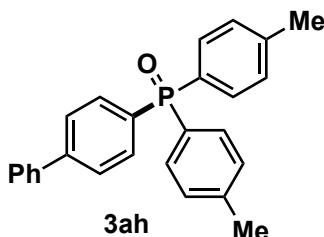
Colorless oil, 52% yield, (47% for one-pot). ^1H NMR (500 MHz, $\text{DMSO}-d_6$) δ 7.89 – 7.82 (m, 4H), 7.75 – 7.70 (m, 2H), 7.51 (dd, $J = 8.5, 6.9$ Hz, 2H), 7.45 – 7.41 (m, 1H), 7.40 – 7.31 (m, 10H), 5.10 (d, $J = 8.0$ Hz, 4H). ^{13}C NMR (126 MHz, DMSO) δ 144.71, 144.68, 139.36, 136.86, 136.81, 132.55, 132.47, 129.59, 128.95, 128.88, 128.71, 128.25, 127.79, 127.57, 127.47, 127.45, 126.28, 67.50, 67.46. ^{31}P NMR (202 MHz, DMSO) δ 18.96. HRMS (ESI): Calcd for $\text{C}_{26}\text{H}_{24}\text{O}_3\text{P}$: $[\text{M}+\text{H}]^+$ 415.1463, found: m/z 415.1456.



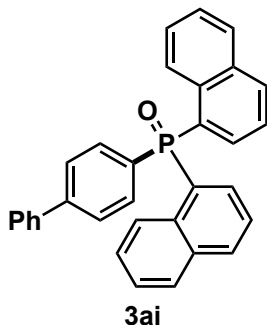
Colorless oil, 93% yield, (97% for one-pot). ^1H NMR (500 MHz, $\text{DMSO}-d_6$) δ 7.88 – 7.78 (m, 6H), 7.72 – 7.66 (m, 2H), 7.62 – 7.57 (m, 1H), 7.54 (ddd, $J = 8.5, 6.5, 3.4$ Hz, 2H), 7.48 (dd, $J = 8.4, 6.9$ Hz, 2H), 7.43 – 7.37 (m, 1H), 4.06 – 3.97 (m, 2H), 1.29 (t, $J = 7.0$ Hz, 3H). ^{13}C NMR (126 MHz, DMSO) δ 144.28, 144.26, 139.45, 132.88, 132.74, 132.72, 132.43, 132.35, 131.80, 131.75, 131.67, 131.57, 130.48, 129.54, 129.34, 129.23, 128.78, 127.58, 127.48, 127.46, 61.31, 61.26, 40.51, 40.43, 40.34, 40.26, 40.17, 40.09, 40.00, 39.93, 39.84, 39.67, 39.51, 16.85, 16.80. ^{31}P NMR (202 MHz, DMSO) δ 29.52. HRMS (ESI): Calcd for $\text{C}_{20}\text{H}_{20}\text{O}_2\text{P}$: $[\text{M}+\text{H}]^+$ 323.1201, found: m/z 323.1197.



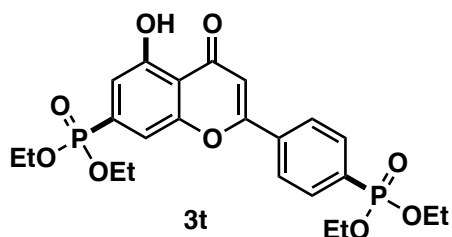
White solid, 90% yield, (91% for one-pot). ^1H NMR (500 MHz, Chloroform-*d*) δ 7.75 (tt, $J = 17.1$, 8.1 Hz, 8H), 7.65 – 7.56 (m, 4H), 7.53 – 7.45 (m, 6H), 7.42 (t, $J = 7.3$ Hz, 1H). These data are in agreement with literature data.¹⁷



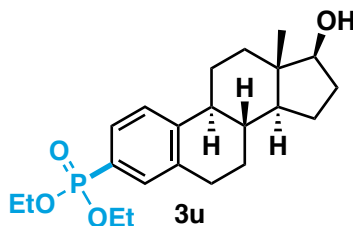
White solid, 91% yield, (88% for one-pot). ^1H NMR (500 MHz, DMSO-*d*₆) δ 7.81 (dd, $J = 8.2$, 2.4 Hz, 2H), 7.72 – 7.63 (m, 4H), 7.57 – 7.50 (m, 4H), 7.47 (dd, $J = 8.4$, 6.9 Hz, 2H), 7.42 – 7.37 (m, 1H), 7.34 (dd, $J = 8.2$, 2.6 Hz, 4H), 2.34 (s, 6H). ^{13}C NMR (126 MHz, DMSO) δ 143.86, 143.84, 142.53, 142.51, 139.45, 132.98, 132.62, 132.54, 132.16, 132.04, 131.96, 130.64, 129.85, 129.80, 129.75, 129.54, 128.75, 127.45, 127.42, 127.32, 21.55. ^{31}P NMR (202 MHz, DMSO) δ 25.58. HRMS (ESI): Calcd for C₂₆H₂₄OP: [M+H]⁺ 383.1563, found: m/z 383.1558



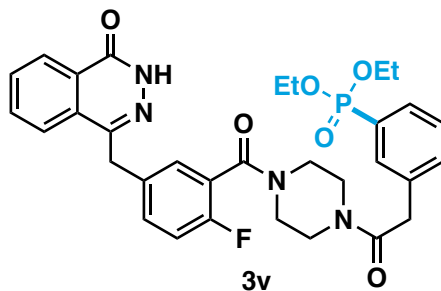
White solid, 90% yield. ^1H NMR (500 MHz, CDCl₃) δ 8.83 (d, $J = 8.2$ Hz, 2H), 8.06 – 7.97 (m, 2H), 7.93 (d, $J = 7.9$ Hz, 2H), 7.77 (dd, $J = 11.8$, 7.7 Hz, 2H), 7.69 (d, $J = 7.4$ Hz, 2H), 7.62 (d, $J = 7.6$ Hz, 2H), 7.53 (t, $J = 7.3$ Hz, 2H), 7.51 – 7.36 (m, 4H), 7.41 – 7.28 (m, 5H). ^{13}C NMR (126 MHz, CDCl₃) δ 139.89, 134.07, 133.32, 132.98, 128.97, 128.83, 128.18, 127.99, 127.52, 127.30, 126.62, 124.30. ^{31}P NMR (202 MHz, CDCl₃) δ 36.35. HRMS (ESI): Calcd for C₃₂H₂₄OP: [M+H]⁺ 455.1565, found: m/z 455.1559.



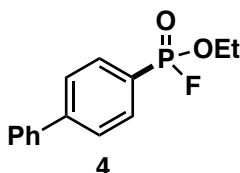
Yellow solid, 90% yield, (93% for one-pot). ^1H NMR (500 MHz, $\text{DMSO-}d_6$) δ 12.57 (s, 1H), 8.27 (dd, $J = 8.1, 3.5$ Hz, 2H), 7.83 (dd, $J = 12.7, 7.9$ Hz, 2H), 7.49 (d, $J = 14.5$ Hz, 1H), 7.25 (s, 1H), 6.97 (d, $J = 14.0$ Hz, 1H), 4.03 (dddt, $J = 14.8, 9.3, 7.1, 2.7$ Hz, 8H), 1.21 (dt, $J = 10.0, 7.0$ Hz, 12H). ^{13}C NMR (126 MHz, DMSO) δ 183.48, 163.90, 160.37, 160.20, 156.21, 156.01, 137.48, 136.02, 134.36, 134.33, 133.63, 132.37, 132.29, 132.16, 127.56, 127.44, 113.23, 113.15, 112.87, 112.85, 111.11, 111.02, 107.96, 62.94, 62.90, 62.57, 62.53, 40.57, 40.47, 40.40, 40.31, 40.23, 40.14, 39.97, 39.90, 39.81, 39.64, 39.47, 16.66, 16.61. ^{31}P NMR (202 MHz, DMSO) δ 16.13, 14.46. HRMS (ESI): Calcd for $\text{C}_{23}\text{H}_{29}\text{O}_9\text{P}_2$: $[\text{M}+\text{H}]^+$ 511.1287, found: m/z 511.1280.



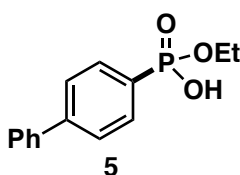
White solid, 92% yield, (83% for one-pot). ^1H NMR (500 MHz, $\text{DMSO-}d_6$) δ 7.46 – 7.35 (m, 3H), 4.52 (s, 1H), 4.05 – 3.90 (m, 4H), 3.53 (t, $J = 8.5$ Hz, 1H), 2.85 (dt, $J = 9.3, 4.6$ Hz, 2H), 2.32 (dd, $J = 14.0, 4.0$ Hz, 1H), 2.23 (ddd, $J = 13.8, 10.6, 3.9$ Hz, 1H), 1.86 (dddd, $J = 23.8, 12.1, 8.9, 4.1$ Hz, 3H), 1.60 (dddd, $J = 12.2, 9.8, 6.9, 3.2$ Hz, 1H), 1.45 – 1.35 (m, 3H), 1.35 – 1.24 (m, 2H), 1.22 (td, $J = 7.0, 1.1$ Hz, 8H), 0.67 (s, 3H). ^{13}C NMR (126 MHz, DMSO) δ 145.45, 137.47, 137.35, 132.19, 132.10, 128.88, 128.81, 126.38, 126.19, 126.07, 124.89, 80.44, 61.93, 61.89, 50.09, 44.53, 43.20, 38.34, 36.97, 30.32, 29.22, 26.93, 26.03, 23.22, 16.69, 16.64, 11.64. ^{31}P NMR (202 MHz, DMSO) δ 18.68. HRMS (ESI): Calcd for $\text{C}_{22}\text{H}_{34}\text{O}_4\text{P}$: $[\text{M}+\text{H}]^+$ 393.2195, found: m/z 393.2188.



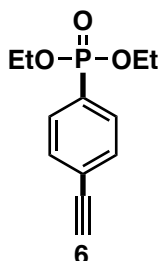
Brown solid, 85% yield, (81% for one-pot). ^1H NMR (500 MHz, $\text{DMSO-}d_6$) δ 12.60 (s, 1H), 8.27 (d, $J = 7.7$ Hz, 1H), 7.96 (d, $J = 8.0$ Hz, 1H), 7.92 – 7.80 (m, 2H), 7.59 (dd, $J = 14.3, 10.7$ Hz, 2H), 7.46 (ddt, $J = 15.3, 7.4, 3.8$ Hz, 3H), 7.37 (ddd, $J = 11.3, 6.4, 2.3$ Hz, 1H), 7.24 (t, $J = 9.0$ Hz, 1H), 4.33 (s, 2H), 4.00 (dt, $J = 14.8, 7.3$ Hz, 4H), 3.87 (s, 1H), 3.80 (s, 1H), 3.58 (d, $J = 16.4$ Hz, 4H), 3.43 (dt, $J = 19.8, 5.1$ Hz, 2H), 3.18 – 3.13 (m, 2H), 1.21 (dt, $J = 10.6, 7.0$ Hz, 6H). ^{13}C NMR (126 MHz, DMSO) δ 169.32, 164.58, 159.87, 155.90, 145.32, 136.96, 135.35, 134.03, 133.96, 132.58, 132.29, 132.05, 129.83, 129.75, 129.55, 129.43, 129.19, 129.06, 128.38, 128.05, 126.56, 125.92, 124.09, 116.50, 116.33, 62.14, 62.09, 46.99, 46.69, 45.83, 45.36, 41.98, 41.69, 41.39, 36.92, 16.65, 16.60. ^{19}F NMR (471 MHz, DMSO) δ -119.77. ^{31}P NMR (202 MHz, DMSO) δ 18.04, 18.01. HRMS (ESI): Calcd for $\text{C}_{32}\text{H}_{35}\text{FN}_4\text{O}_6\text{P}$: $[\text{M}+\text{H}]^+$ 621.2278, found: m/z 621.2272.



Colorless oil, 35% yield. ^1H NMR (500 MHz, DMSO) δ 7.92 (d, J = 11.5 Hz, 4H), 7.76 (d, J = 7.6 Hz, 2H), 7.54 (t, J = 7.5 Hz, 2H), 7.47 (t, J = 7.3 Hz, 1H), 4.36 – 4.27 (m, 2H), 1.35 (t, J = 7.0 Hz, 3H). ^{13}C NMR (126 MHz, DMSO) δ 146.02, 145.99, 139.01, 132.65, 132.56, 129.66, 129.19, 127.96, 127.83, 127.60, 124.14, 123.89, 122.54, 122.30, 64.70, 64.65, 16.62, 16.57. ^{31}P NMR (202 MHz, DMSO) δ 17.06 (d, J = 1031.4 Hz). ^{19}F NMR (376 MHz, DMSO) δ -63.25 (d, J = 1031.4 Hz). HRMS (ESI): Calcd for $\text{C}_{14}\text{H}_{15}\text{FO}_2\text{P}$: $[\text{M}+\text{H}]^+$ 265.0794, found: m/z 265.0787.



Colorless oil, 15% yield. ^1H NMR (500 MHz, DMSO- d_6) δ 7.81 – 7.74 (m, 4H), 7.73 – 7.68 (m, 2H), 7.49 (dd, J = 8.4, 6.9 Hz, 2H), 7.44 – 7.37 (m, 1H), 3.96 – 3.87 (m, 2H), 1.19 (t, J = 7.1 Hz, 3H). ^{13}C NMR (126 MHz, DMSO) δ 143.67, 143.64, 139.66, 132.18, 132.10, 131.22, 129.76, 129.54, 128.64, 127.39, 127.18, 127.07, 61.20, 61.16, 40.55, 40.46, 40.38, 40.29, 40.21, 40.12, 40.05, 39.95, 39.88, 39.79, 39.62, 39.45, 16.75, 16.70. ^{31}P NMR (202 MHz, DMSO) δ 14.99, 14.98. HRMS (ESI): Calcd for $\text{C}_{14}\text{H}_{16}\text{O}_3\text{P}$: $[\text{M}+\text{H}]^+$ 263.0837, found: m/z 263.0823.



To a mixture of **3f** (87 mg, 0.3 mmol), $\text{Pd}(\text{OAc})_2$ (6.7 mg, 0.03 mmol) and CuI (11 mg, 0.06 mmol) in triethylamine (Et_3N) (1.5 mL) was added (trimethylsilyl)acetylene (85 μL , 0.6 mmol). The reaction mixture was stirred under argon atmosphere at 60 $^\circ\text{C}$ for 6 h. After filtration, the filtrate was evaporated under reduced pressure, and dissolved in MeOH (1.5 mL). After that K_2CO_3 (124 mg, 0.9 mmol) was added. The mixture was stirred at room temperature for 3 h. After filtration, the filtrate was evaporated under reduced pressure. The crude product was diluted with dichloromethane, and was washed with water, and the dichloromethane layer was dried over MgSO_4 . After the solvent was removed, the residue was purified by flash chromatography to give 46 mg of **6** as colorless oil in 65% yield. ^1H NMR (500 MHz, CDCl_3) δ 7.83 – 7.75 (m, 2H), 7.62 – 7.54 (m, 2H), 4.25 – 4.03 (m, 4H), 3.26 (s, 1H), 1.33 (t, J = 7.0 Hz, 6H). These data are in agreement with literature data.¹⁸

III. The stability of **1a** in the presence of TMSOH.

Procedure: To a solution of **1a** (0.2 mmol) in CH₃CN (1 mL) was added TMSOH (0.6 mmol) and DIPEA (0.4 mmol). The mixture was stirred at room temperature and analysis by LC-MS at 0.5h, 1h and 3h.

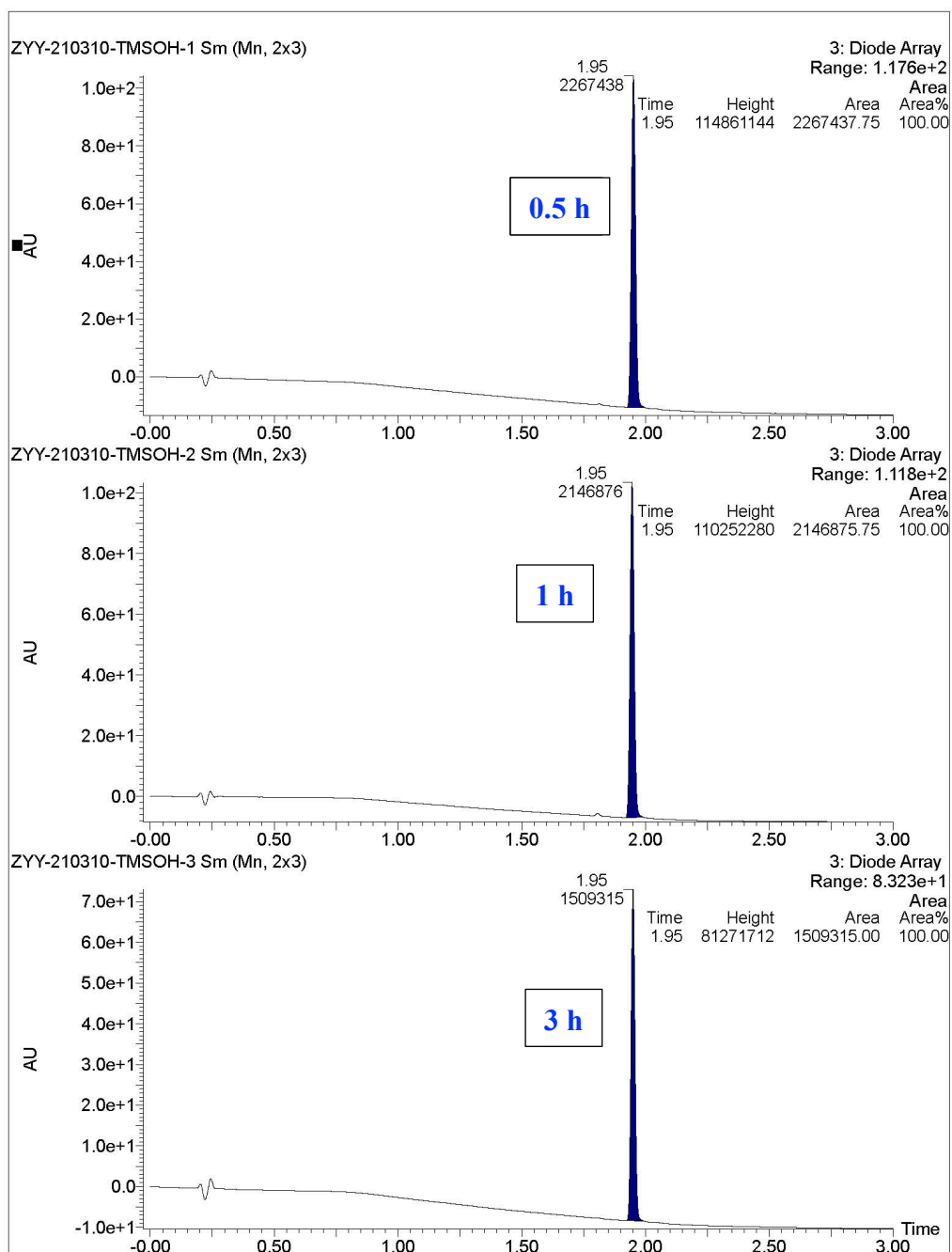
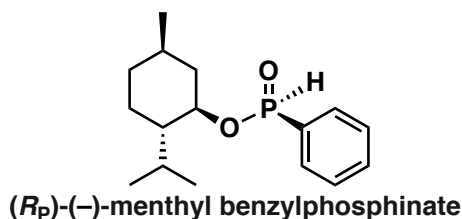
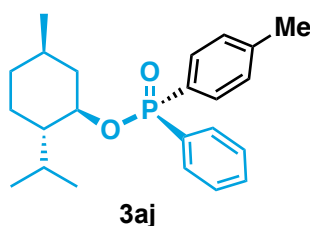


Figure 1. LC-MS analysis of the stability of **1a** in the presence of TMSOH and DIPEA.

IV. Synthesis of enantiomerically pure P-chiral 3aj.



Synthesis of (Rp)-(-)-menthyl benzylphosphinate. (the synthesis was according to the literature procedures with slightly modification¹⁹⁻²¹) A solution of (-)-menthol (50 g, 0.32 mol) and pyridine (25 mL, 0.32 mol) in hexane (150 mL) was added dropwise to dichlorophenylphosphine (44 mL g, 0.32 mol) in hexane (150 mL) at 0°C. After stirring at room temperature for 12 hours, the resulting pyridine hydrochloride was removed by filtration, and water (100 mL) was added slowly at 0°C. The two layers were separated, and the organic phase was washed with aqueous sodium bicarbonate solution (50 mL), dried over anhydrous Na₂SO₄, filtrated, and concentrated under reduced pressure to give 79 g of menthyl hydrogenophenylphosphinate with about 38% diastereomeric excess. The crude product was then diluted in hexane (10 mL) and stored at -40°C for 48h. The first batch was collected. After a second crystallization in hexane, 12.3 g (14% yield) of (Rp)-(-)-menthyl benzylphosphinate was obtained. ¹H NMR (500 MHz, CDCl₃) δ 7.66 (d, *J* = 552.5 Hz, 1H), 7.81 – 7.72 (m, 2H), 7.56 – 7.49 (m, 1H), 7.44 (td, *J* = 7.5, 3.2 Hz, 2H), 4.22 (qd, *J* = 10.5, 4.5 Hz, 1H), 2.20 – 2.07 (m, 2H), 1.69 (brs, 1H), 1.66 – 1.56 (m, 1H), 1.49 – 1.43 (m, 2H), 1.17 (td, *J* = 12.2, 10.9 Hz, 1H), 0.98 (qd, *J* = 13.6, 3.8 Hz, 1H), 0.89 (d, *J* = 7.0 Hz, 3H), 0.83 (d, *J* = 6.6 Hz, 3H), 0.79 (d, *J* = 6.9 Hz, 3H). ³¹P NMR (202 MHz, CDCl₃) δ 24.70. These data are in agreement with literature data.²¹



3aj (white solid, 85 % yield) was synthesized by using the one-pot procedure. ¹H NMR (500 MHz, CDCl₃) δ 7.80 – 7.79 (m, 2H), 7.68 – 7.62 (m, 2H), 7.52 – 7.46 (m, 1H), 7.45 – 7.38 (m, 2H), 7.22 (dd, *J* = 8.0, 3.2 Hz, 2H), 4.27 – 4.17 (m, 1H), 2.37 (s, 3H), 2.18 – 2.06 (m, 2H), 1.66 – 1.59 (m, 2H), 1.47 – 1.30 (m, 2H), 1.20 (td, *J* = 12.2, 10.8 Hz, 1H), 1.02 – 0.91 (m, 1H), 0.88 (d, *J* = 7.1 Hz, 2H), 0.83 (d, *J* = 6.4 Hz, 3H), 0.54 (d, *J* = 6.9 Hz, 3H). ³¹P NMR (202 MHz, DMSO) δ 16.13, 14.46. ³¹P NMR (202 MHz, CDCl₃) δ 29.77. These data are in agreement with literature data.²²

V. Synthesis of DNA conjugated phosphine oxide D2.

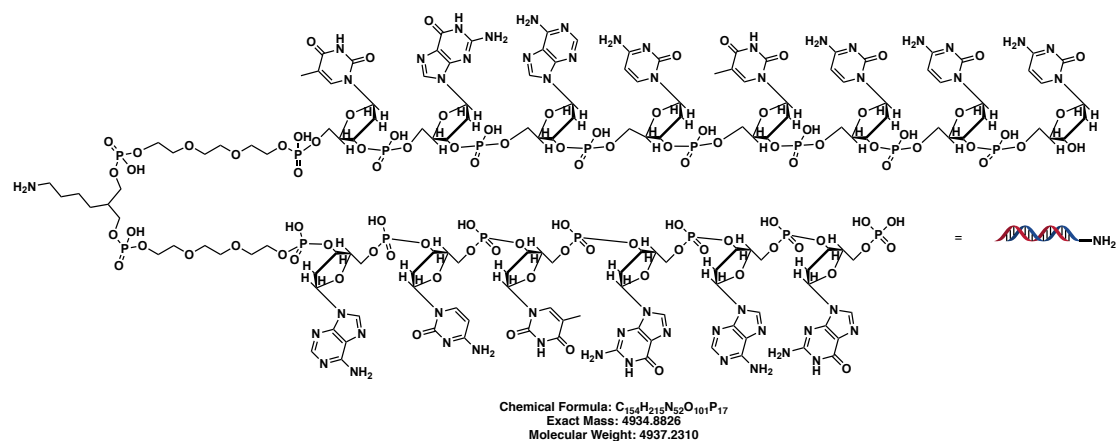


Figure 2. Structure of headpiece DNA (HP-DNA)

Synthesis of DNA conjugated aryl fluorosulfate D1:

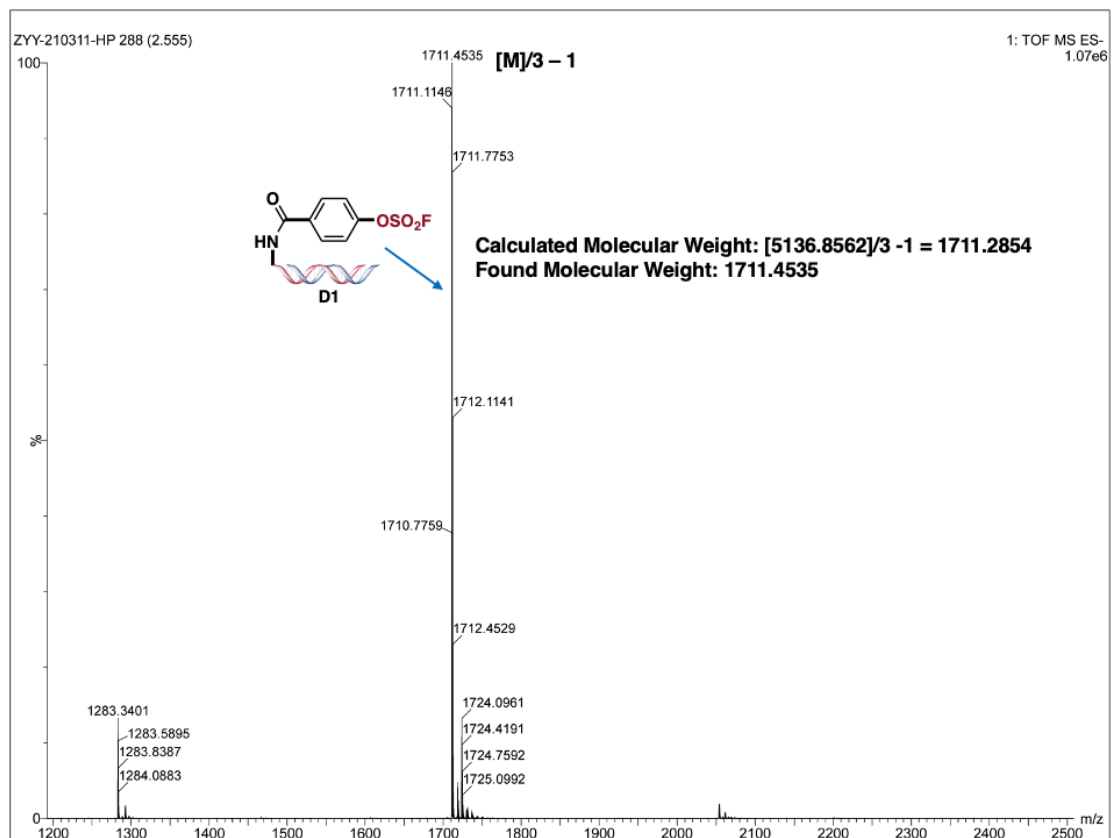
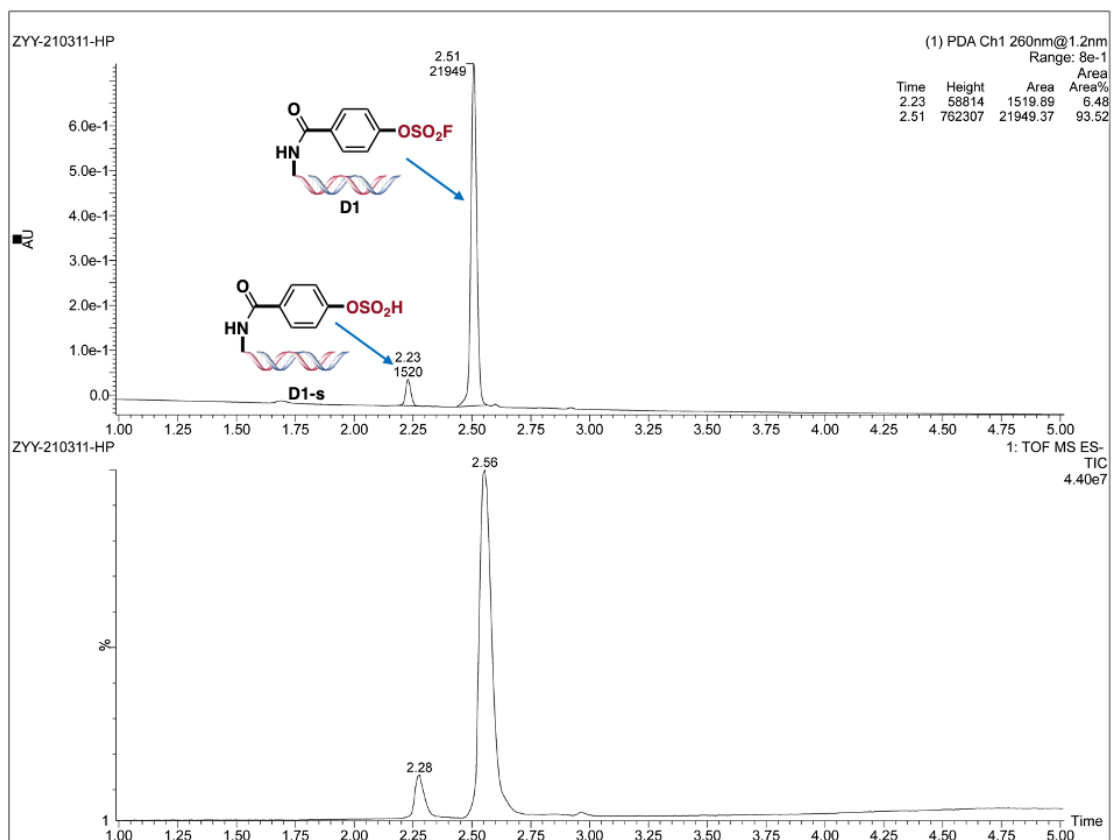
To a solution of DNA headpiece (100 nmol, 1 mM in water) in borate buffer 300 μ L (250 mM, pH = 9.4), was added a mixture of DMA solution of HATU (100 μ L, 200 mM in DMA), DIPEA (100 μ L, 400 mM in DMA) and acid (100 μ L, 200 mM in DMA). The resultant mixture was vortexed and incubated at 25 $^{\circ}$ C for 8 hours. Add 5 M NaCl solution (10% by volume) and cold ethanol (2.5 times by volume, ethanol stored at -20 $^{\circ}$ C) to the resultant supernatant. The mixture was vortexed and incubated at -80 $^{\circ}$ C for at least 30 minutes. The sample was centrifuged for 30 minutes at 4 $^{\circ}$ C in a microcentrifuge at 12,000 rpm to remove the supernatant. The resulting pellet (precipitate) was re-dissolved in ddH₂O. Yield of **D1** 93.5%.

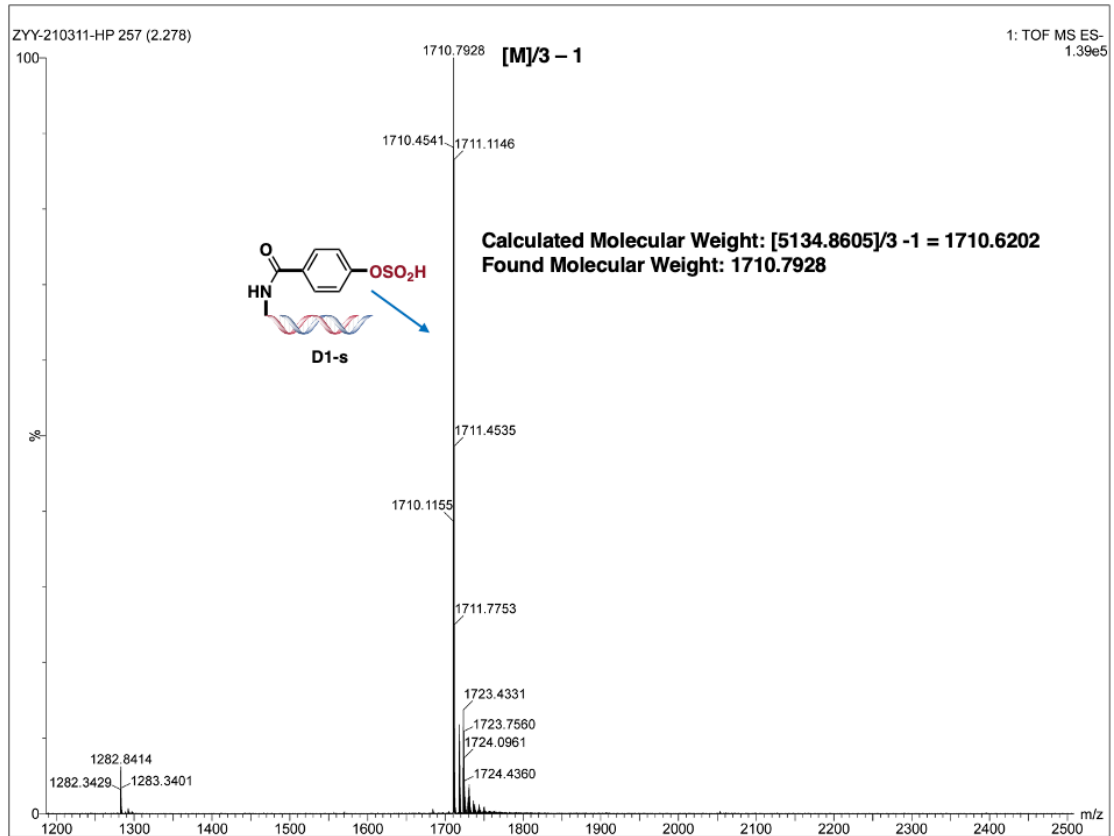
Synthesis of DNA conjugated phosphine oxide D2:

To the DNA conjugated aryl fluorosulfate **D1** (5 nmol, 5 μ L, 1 mM in water), was added 500 equiv. of phosphines oxide (5 μ L, 500 mM in DMF) followed by 500 equiv. of DIPEA (5 μ L, 500 M in DMF), 20 equiv. of Pd(OAc)₂ (5 μ L, 20 mM in DMF), 40 equiv. of dppf (5 μ L, 40 mM in DMF) and 75 μ L ddH₂O. The mixture was vortexed and incubated at 80 $^{\circ}$ C for 2 hours. After reaction, 30 equiv. of scavenger sodium diethyldithiocarbamic acid (compared with Pd(OAc)₂, 3 μ L, 1 M in ddH₂O) were added to the mixture, and the reaction mixture was heated at 60 $^{\circ}$ C for 30 minutes. The mixture was centrifuged at 4 $^{\circ}$ C for 10 min at 12,000 rpm, and the resultant supernatant was collected. Add 5 M NaCl solution (10% by volume) and cold ethanol (2.5 times by volume, ethanol stored at -20 $^{\circ}$ C) to the resultant supernatant. The mixture was vortexed and incubated at -80 $^{\circ}$ C for at least 30 minutes. The sample was centrifuged for 30 minutes at 4 $^{\circ}$ C in a microcentrifuge at 12,000 rpm to remove the supernatant. The resulting pellet (precipitate) was re-dissolved in ddH₂O (300 μ L) for LC-MS detection.

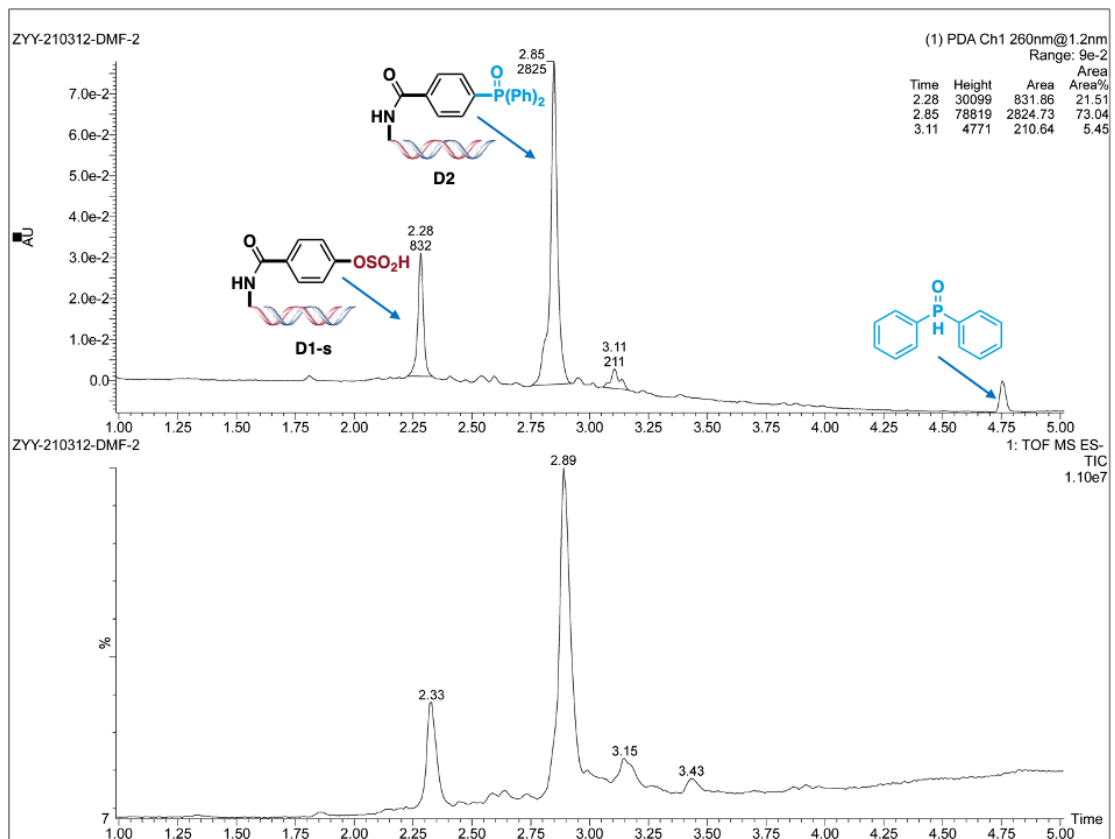
$$\text{Yield of D2: } 78.1\% = \frac{73.04\% \text{ UV (product)}}{93.52\% \text{ UV (DNA starting material before reaction)}} \times 100\%$$

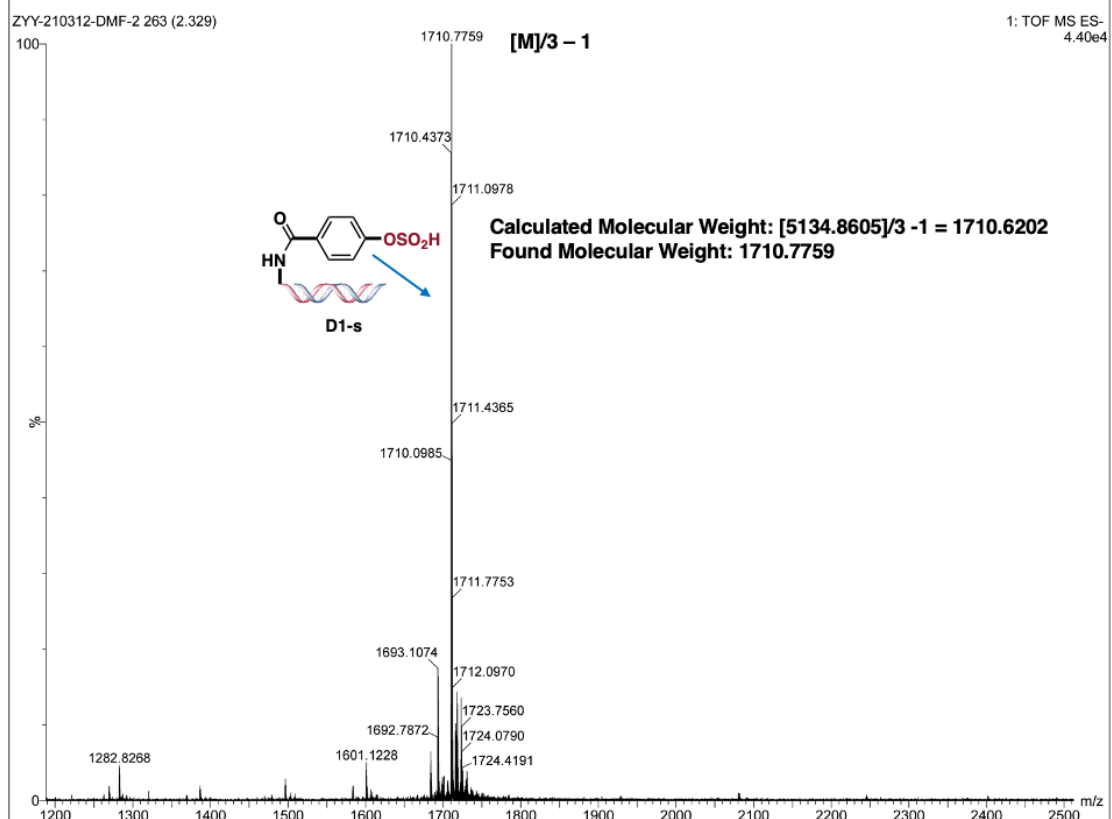
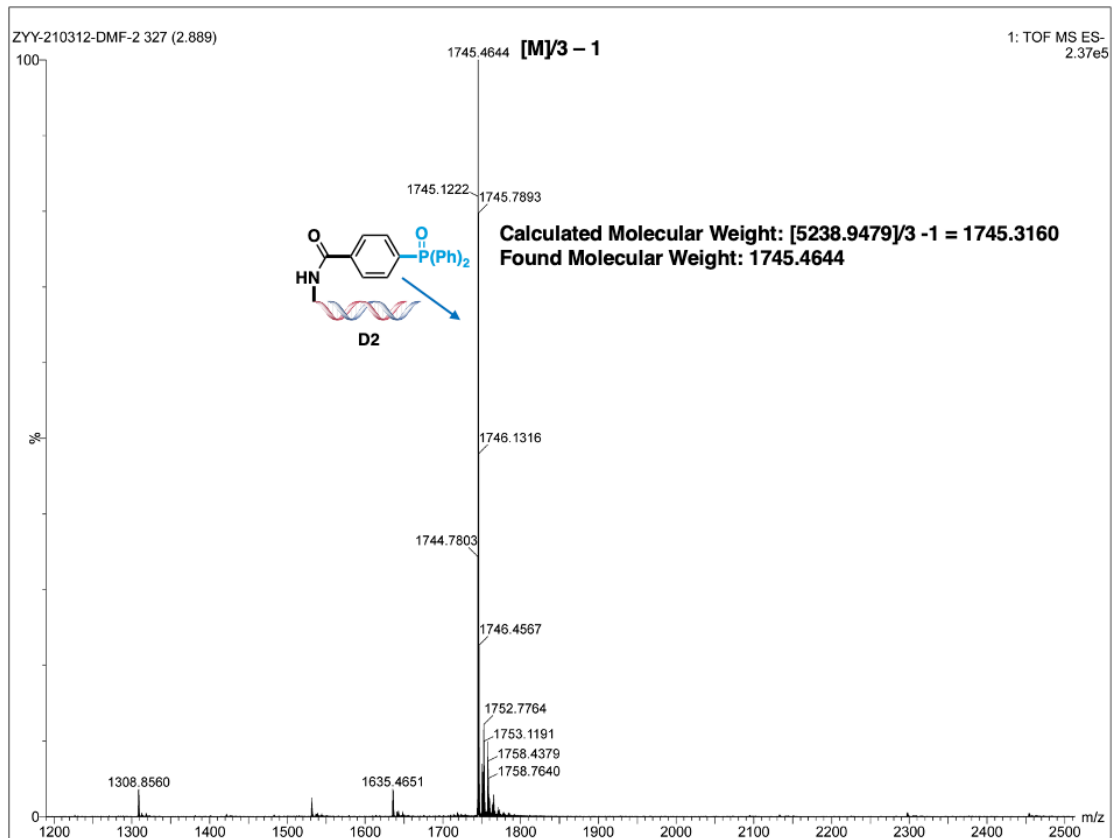
LC-MS analysis of D1



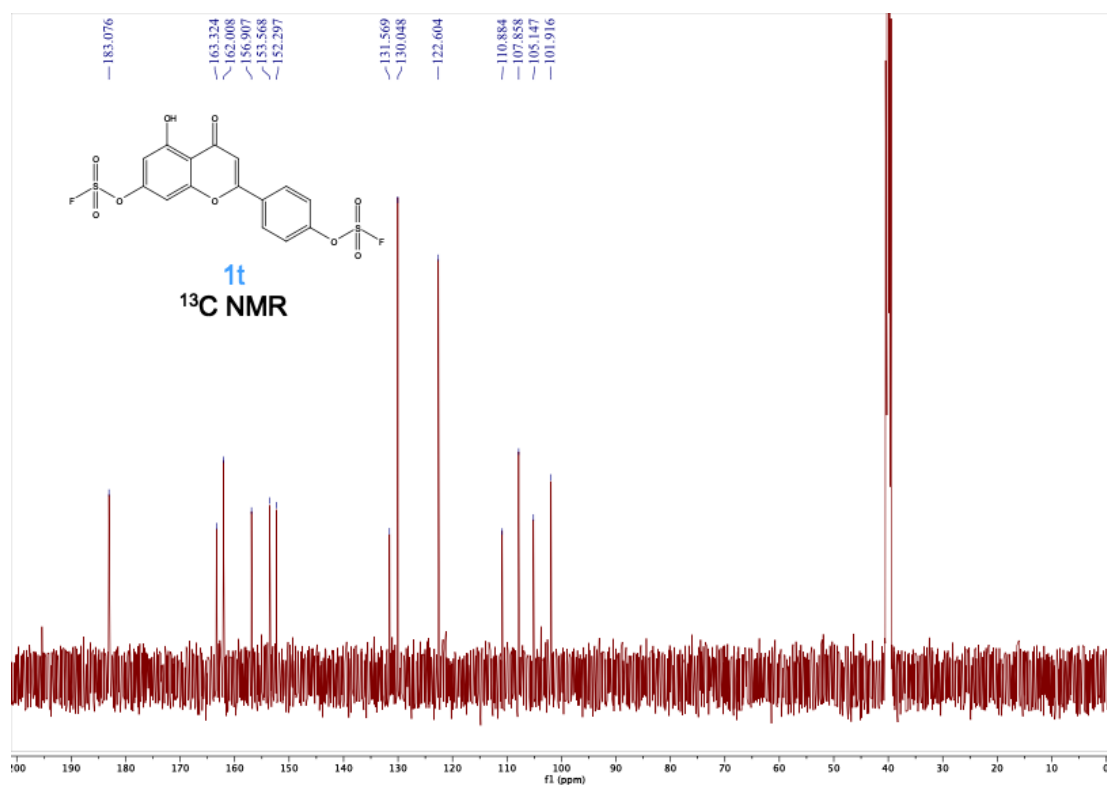
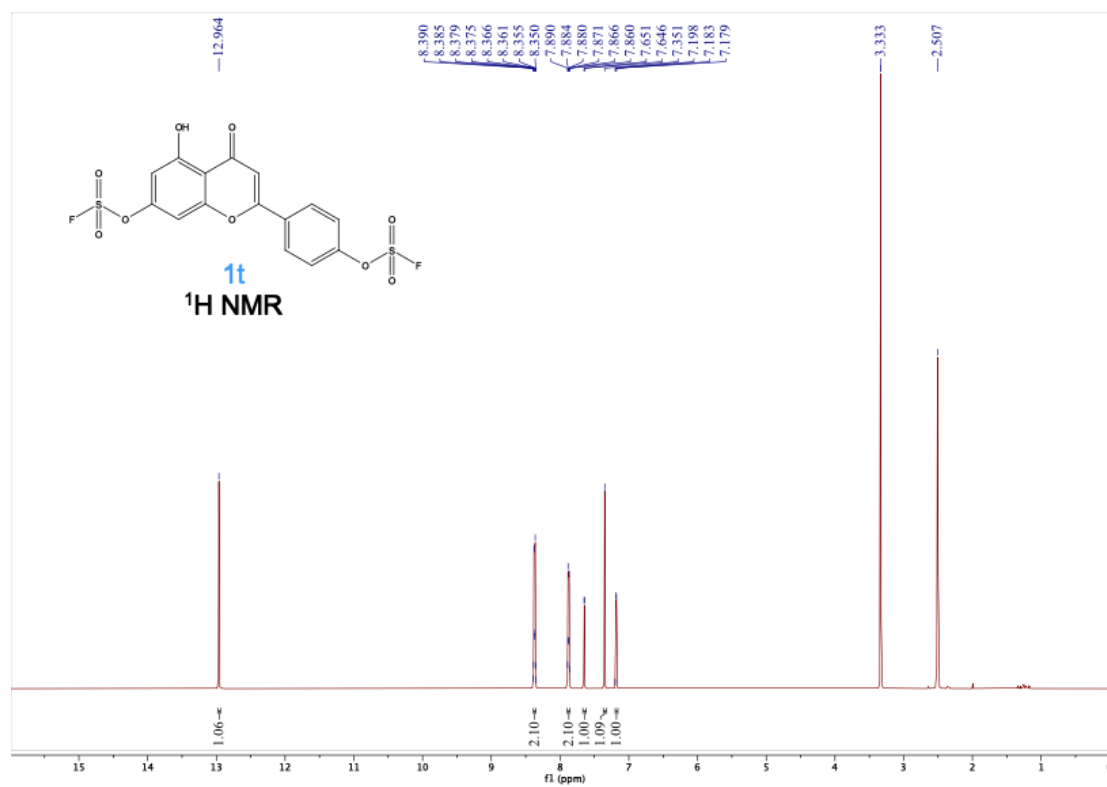


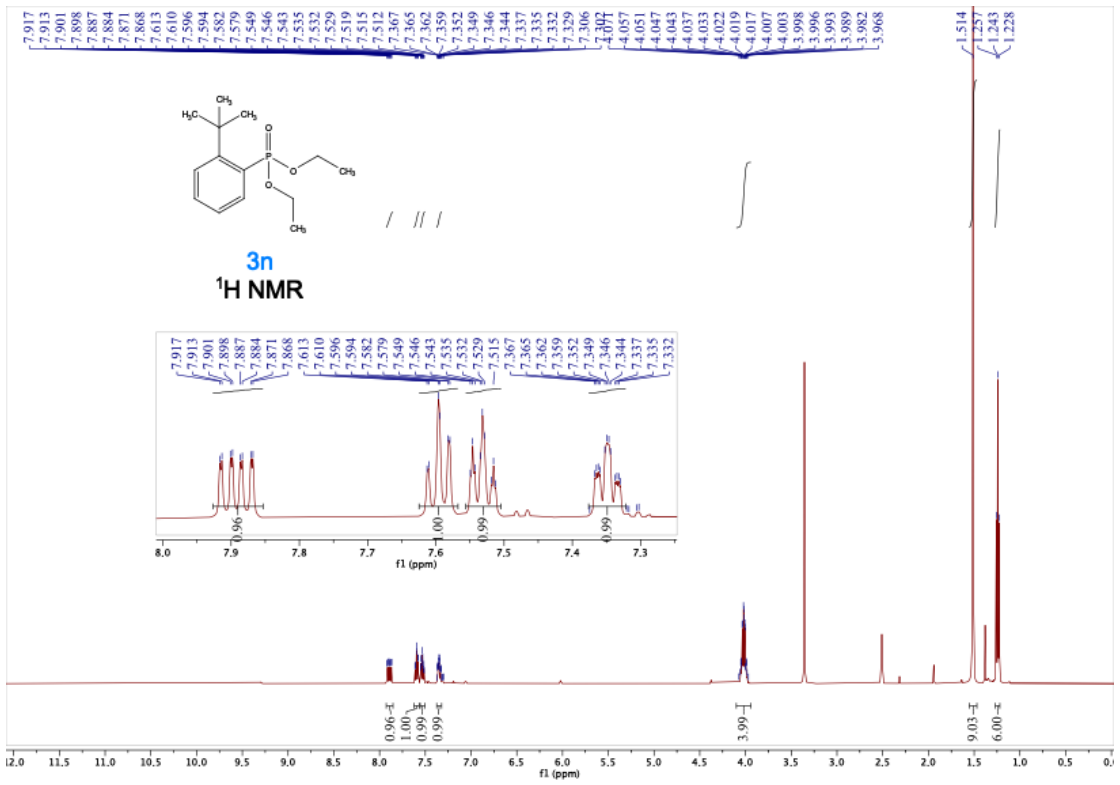
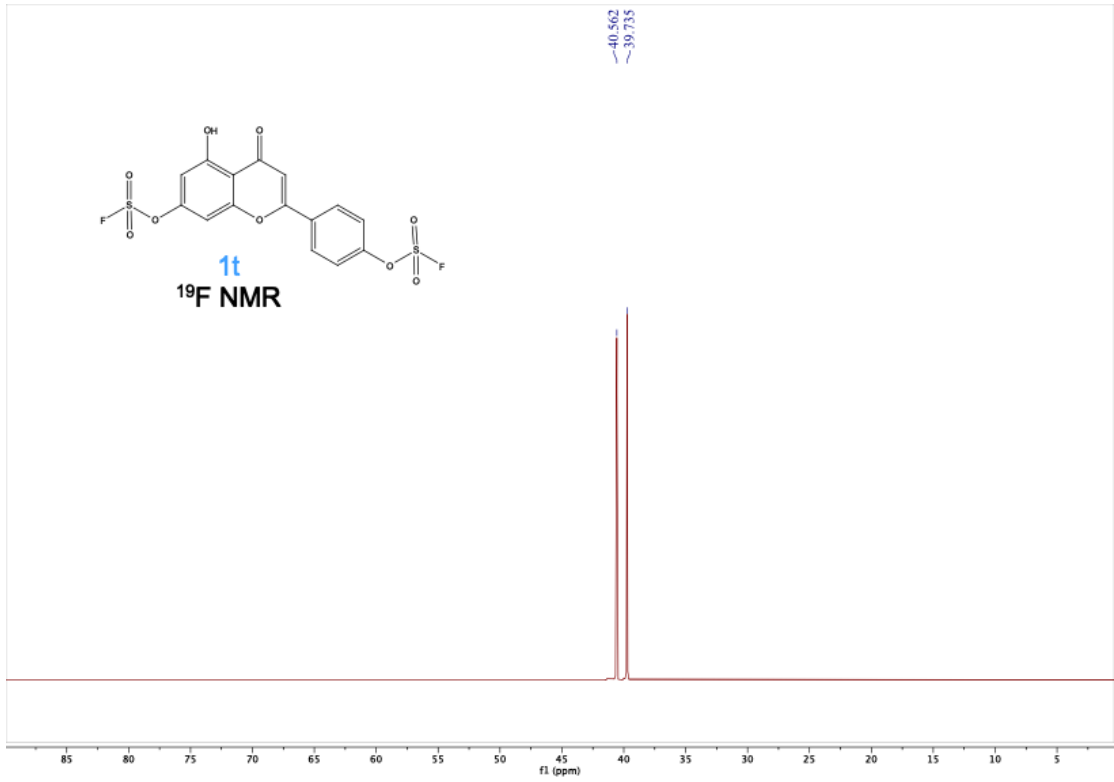
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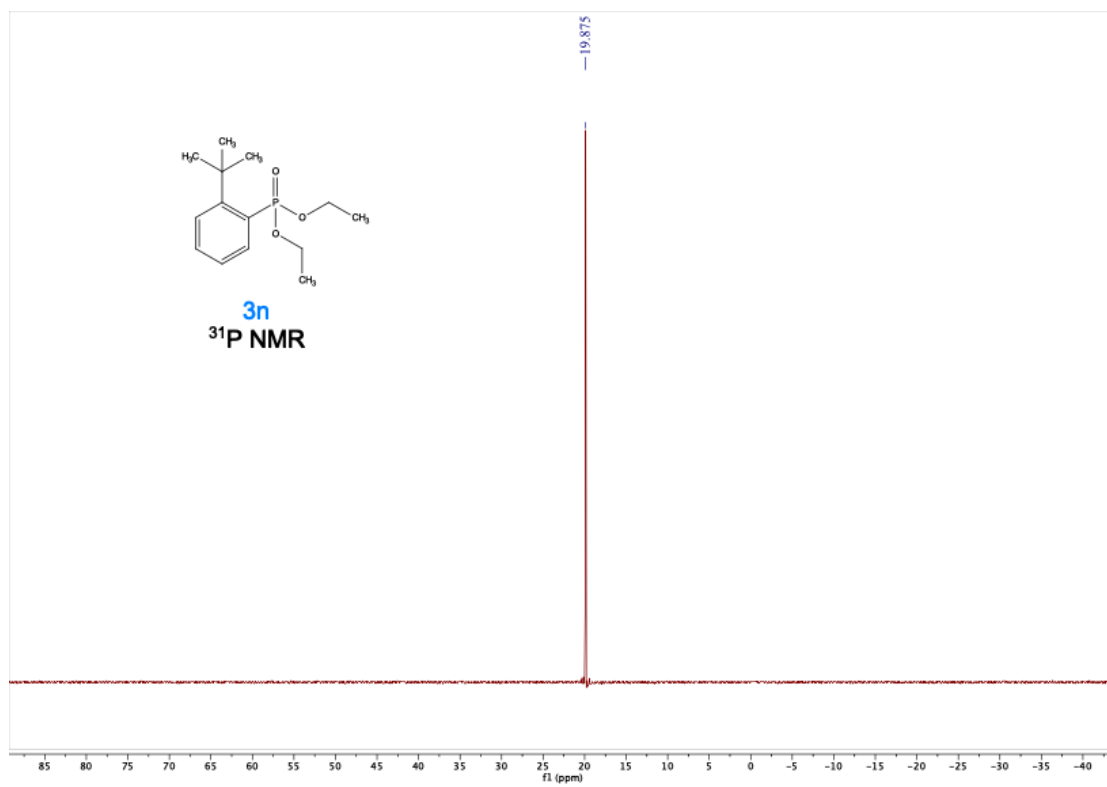
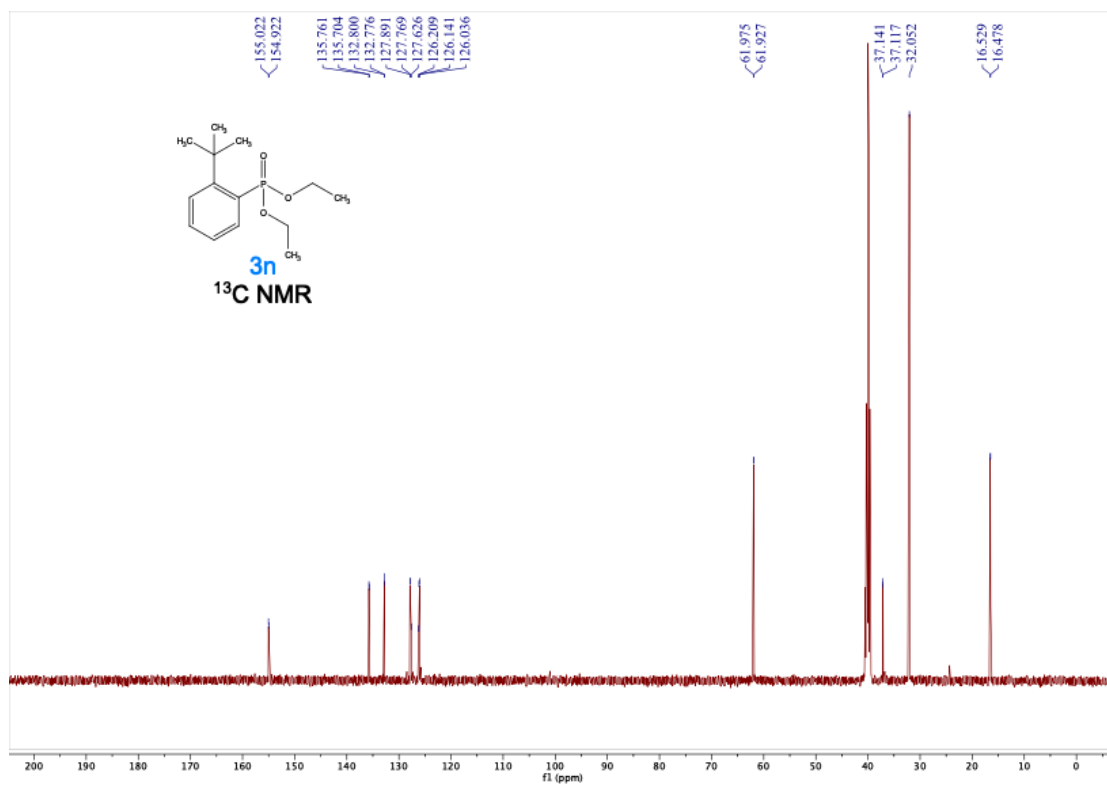


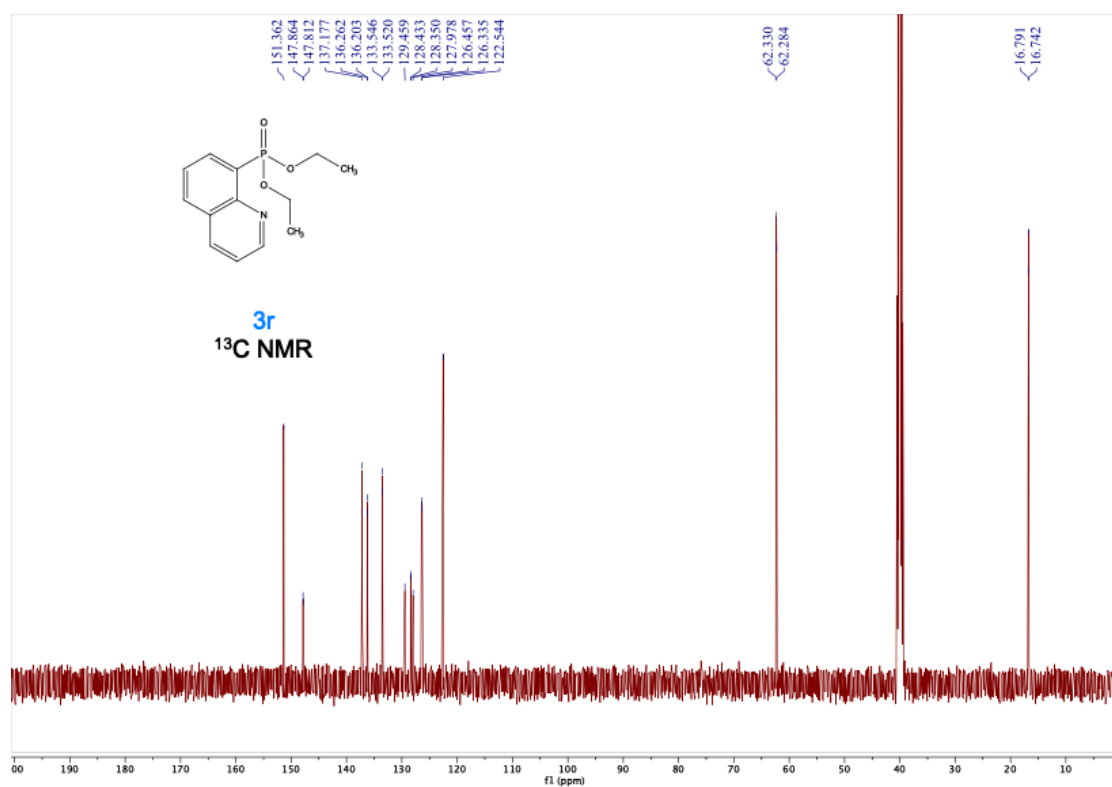
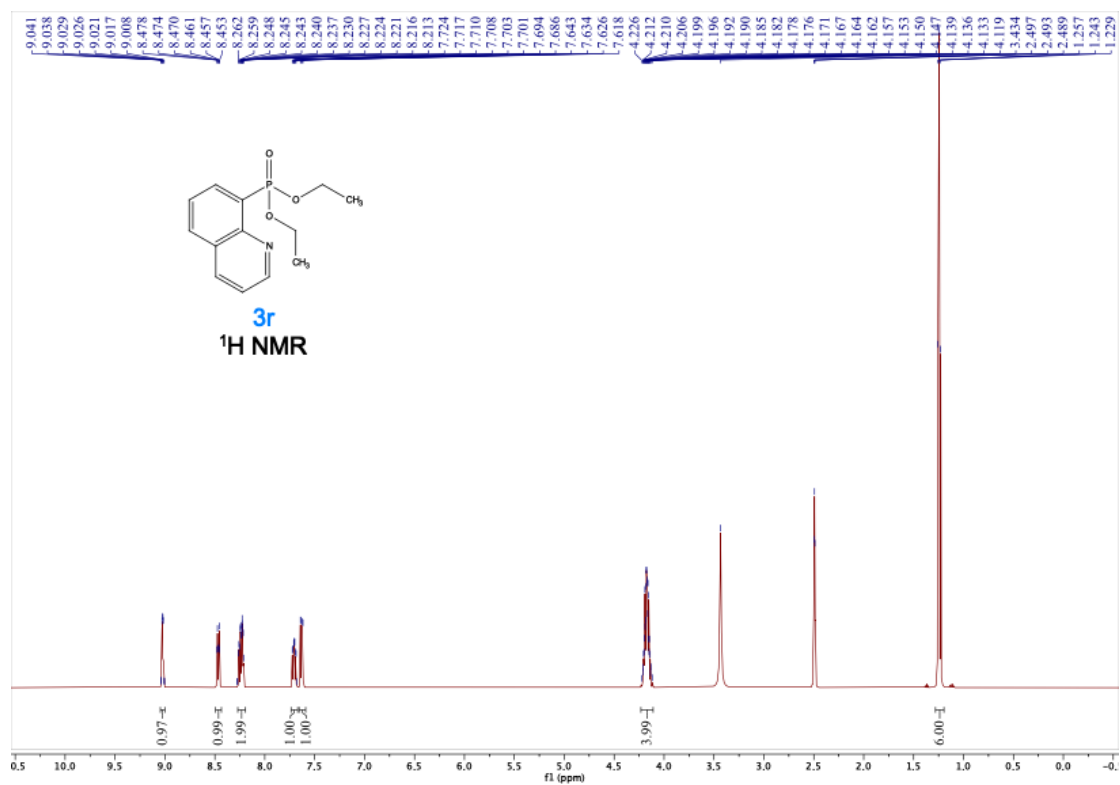


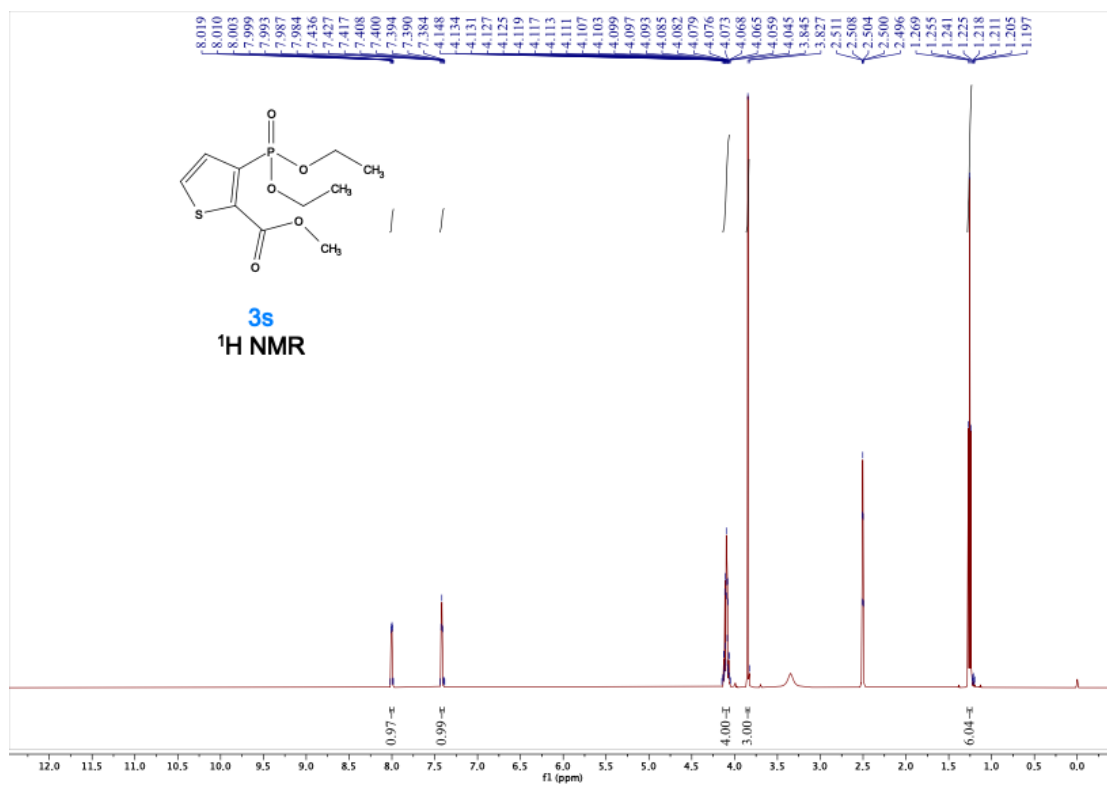
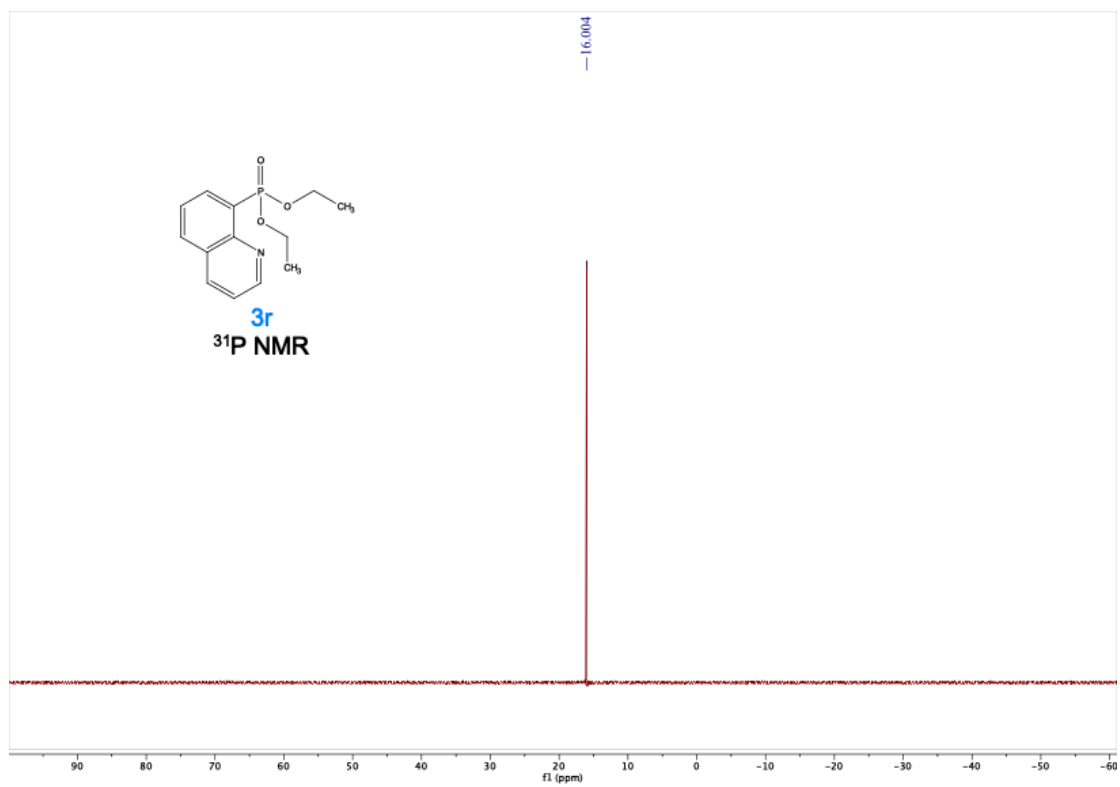
VI. Copies of ^1H NMR, ^{13}C NMR, ^{31}P NMR ^{19}F NMR Spectra

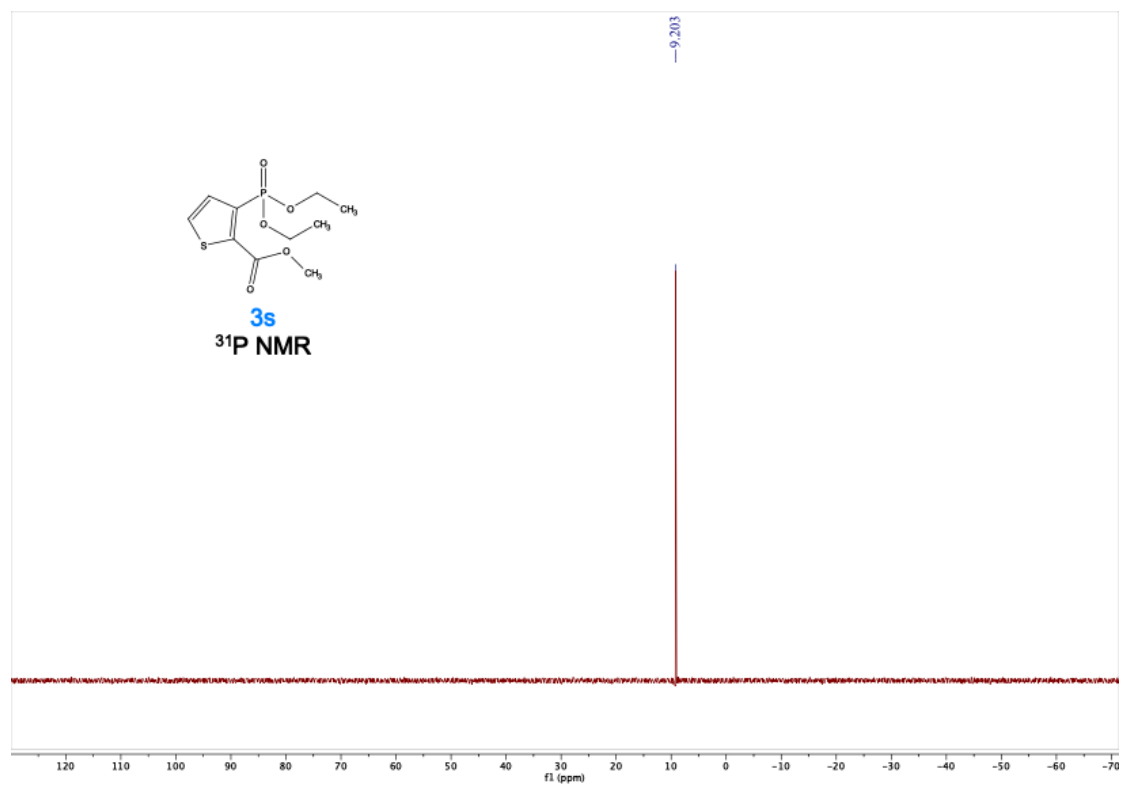
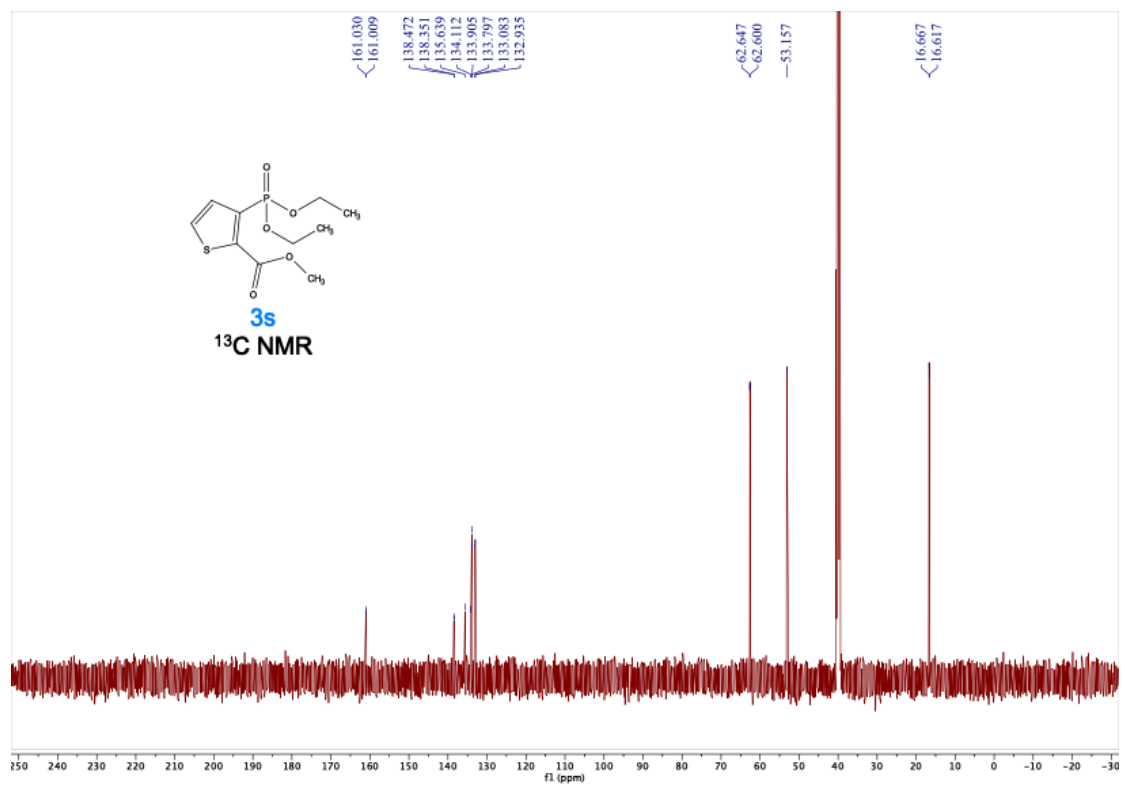


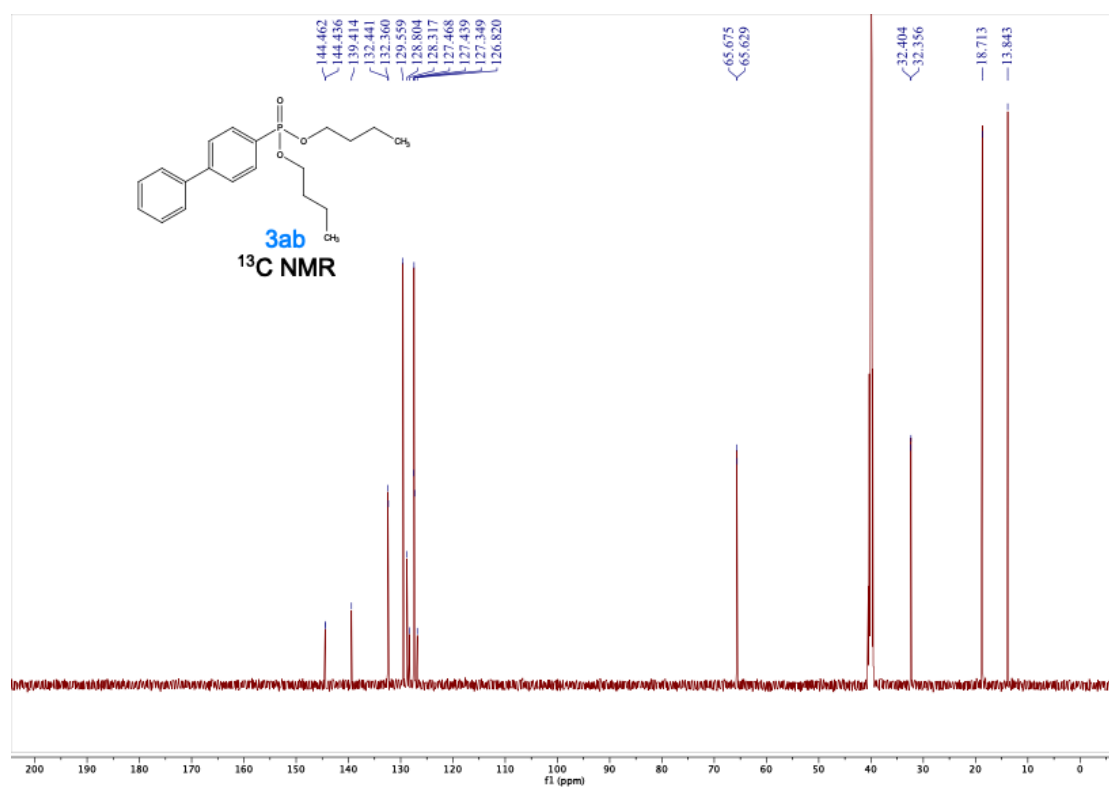
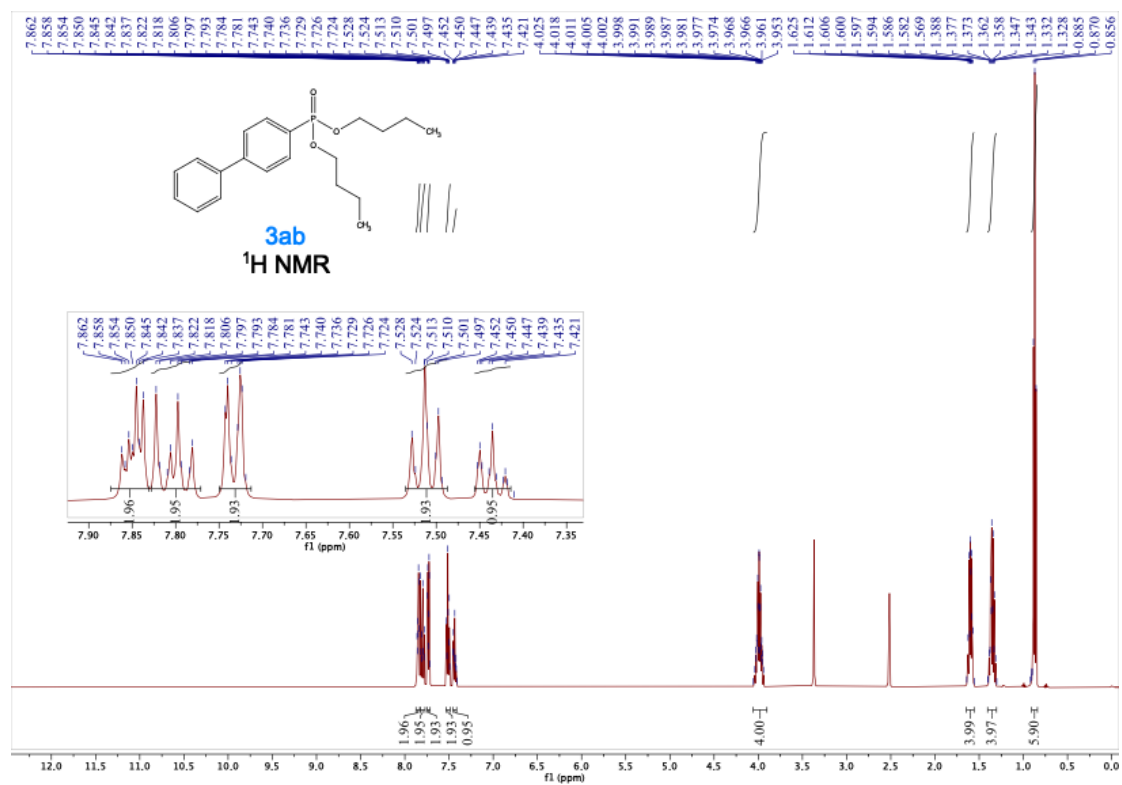


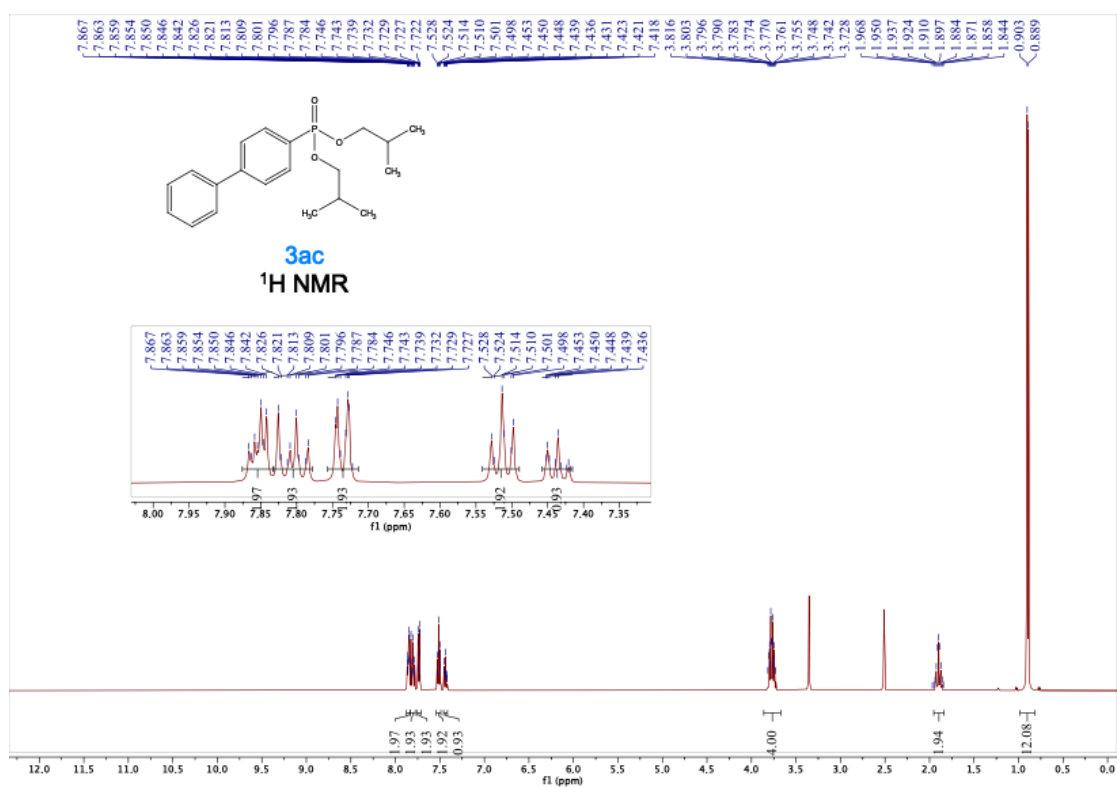
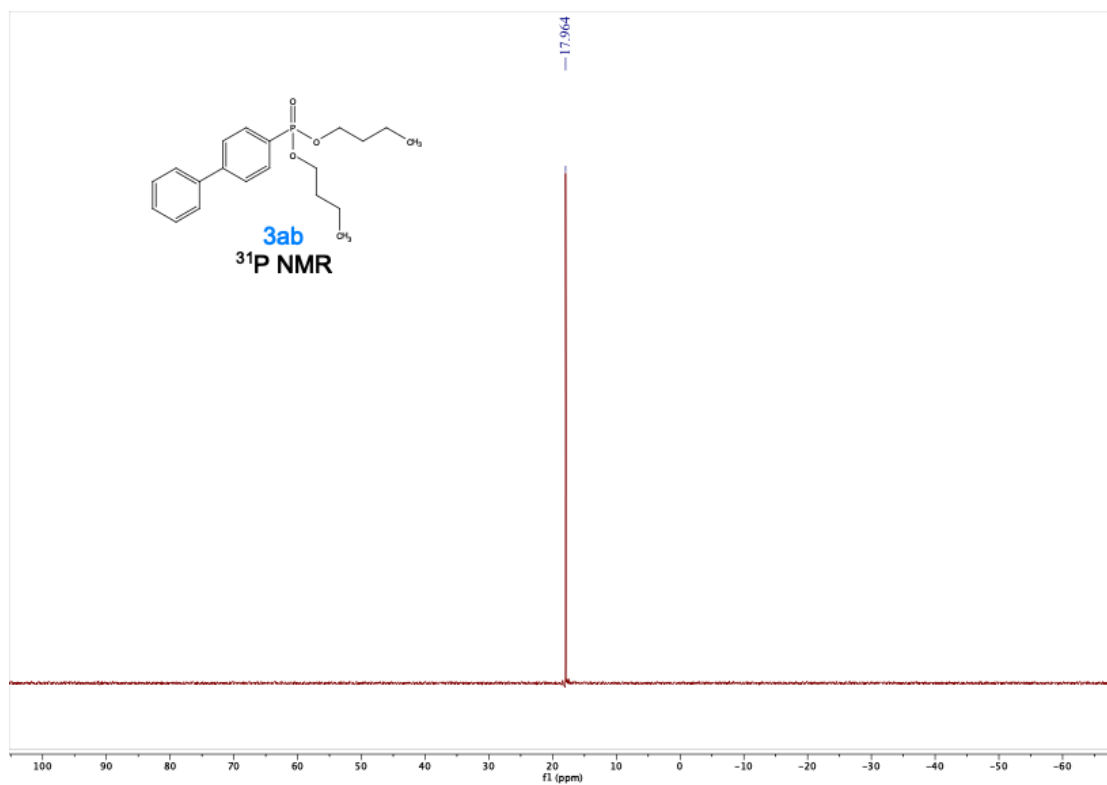


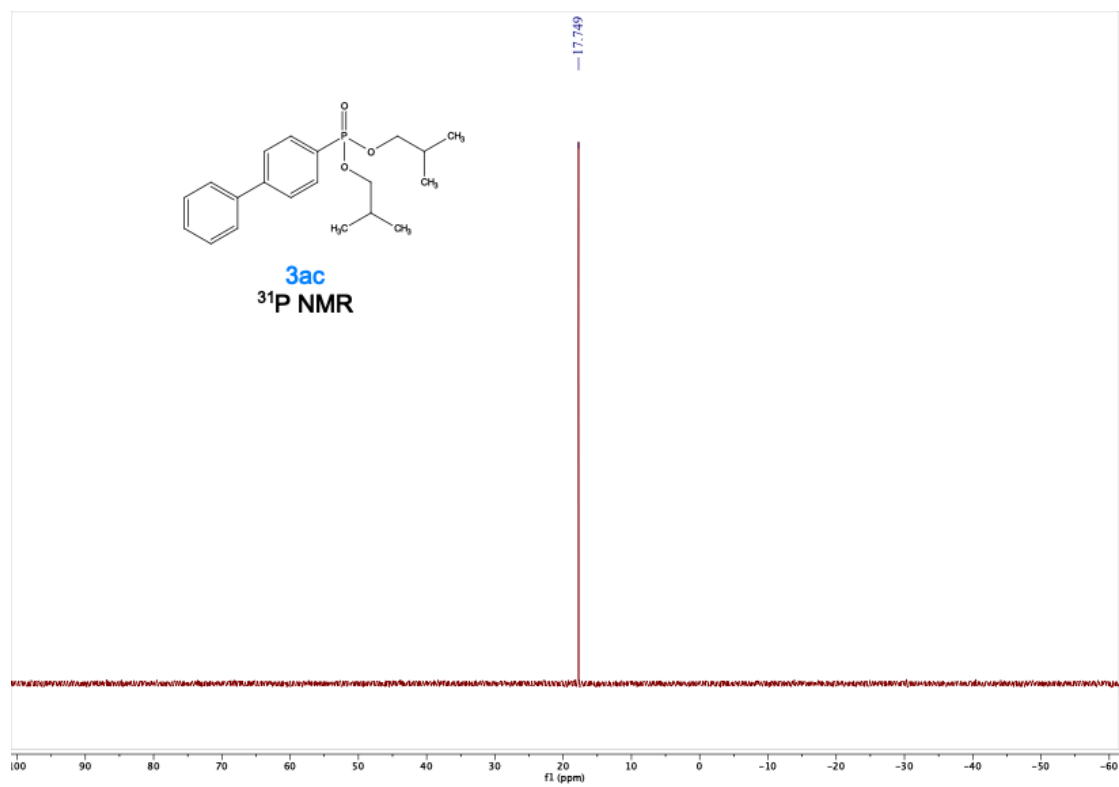
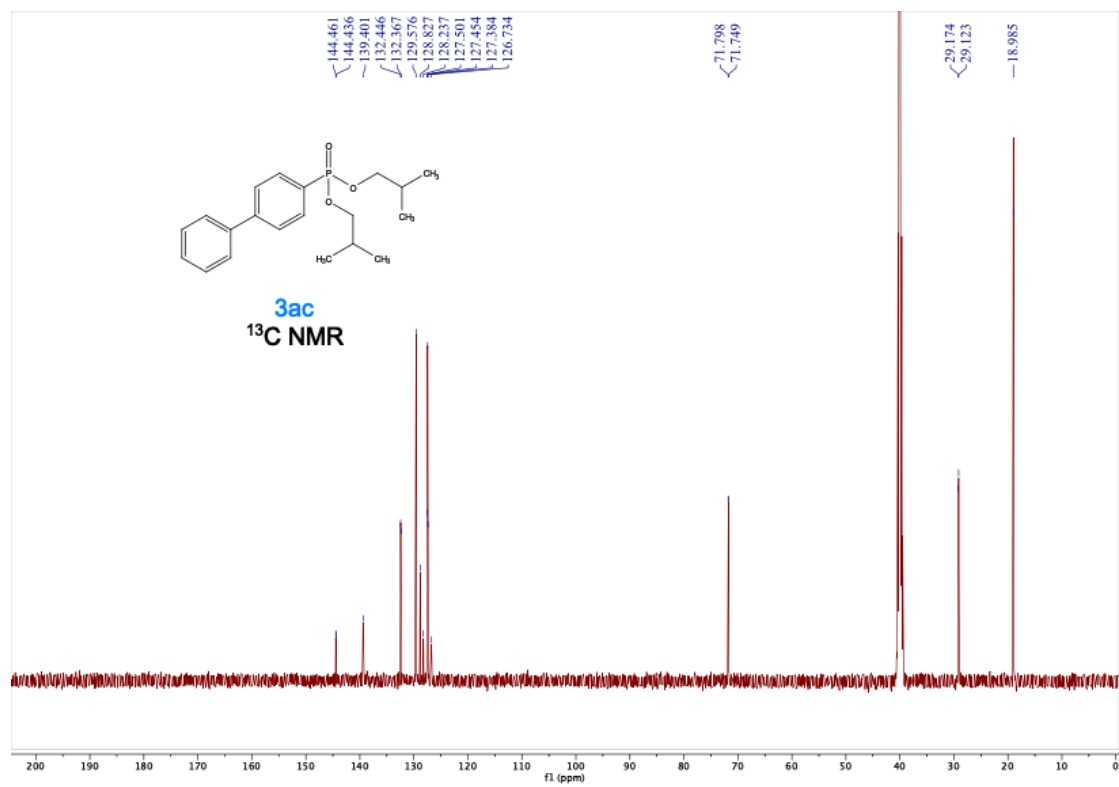


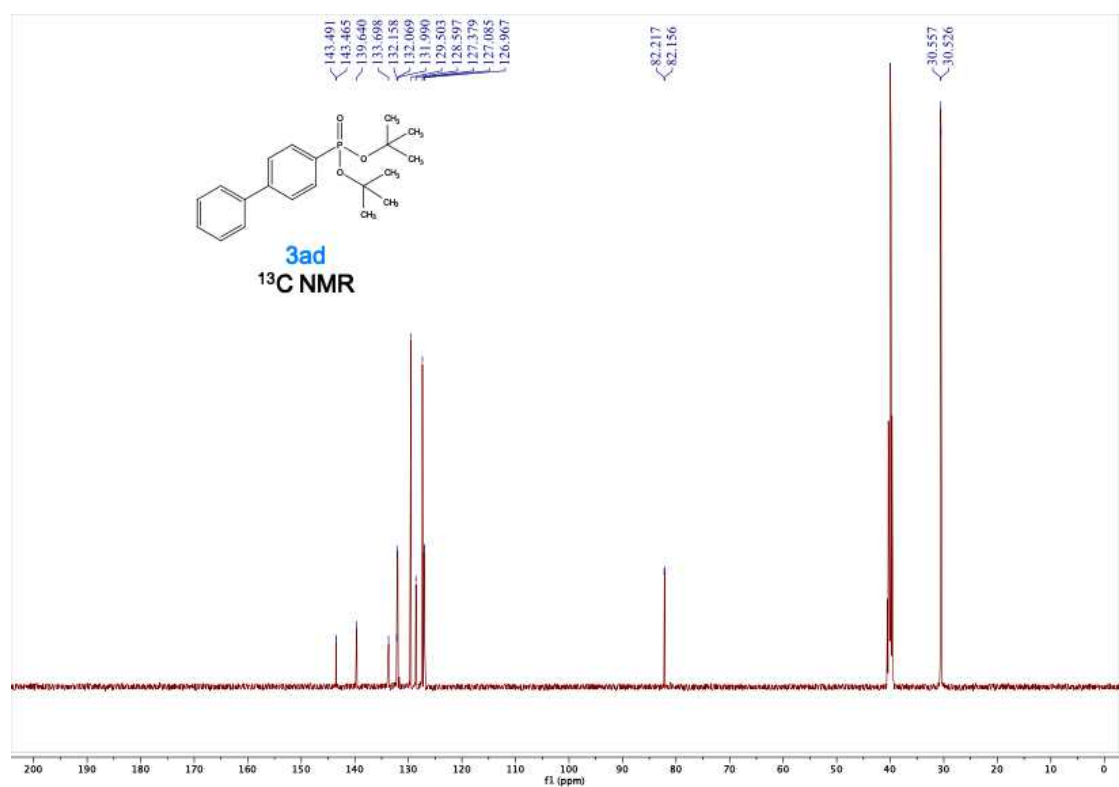
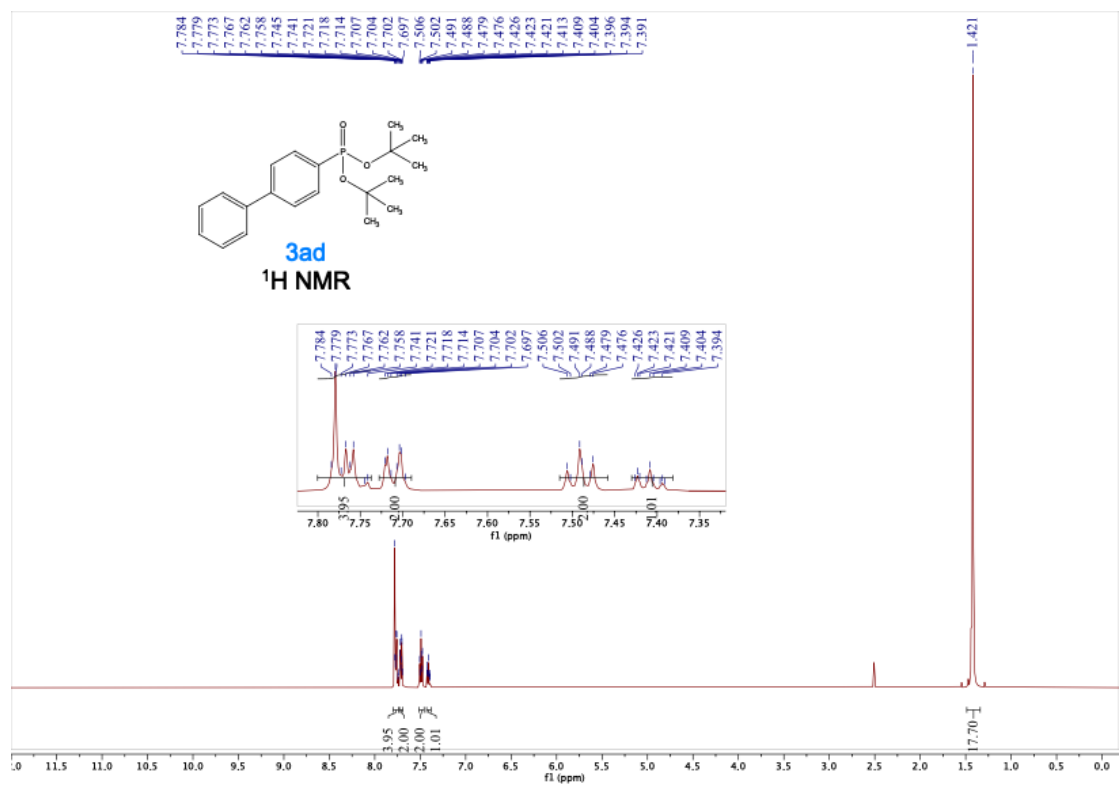


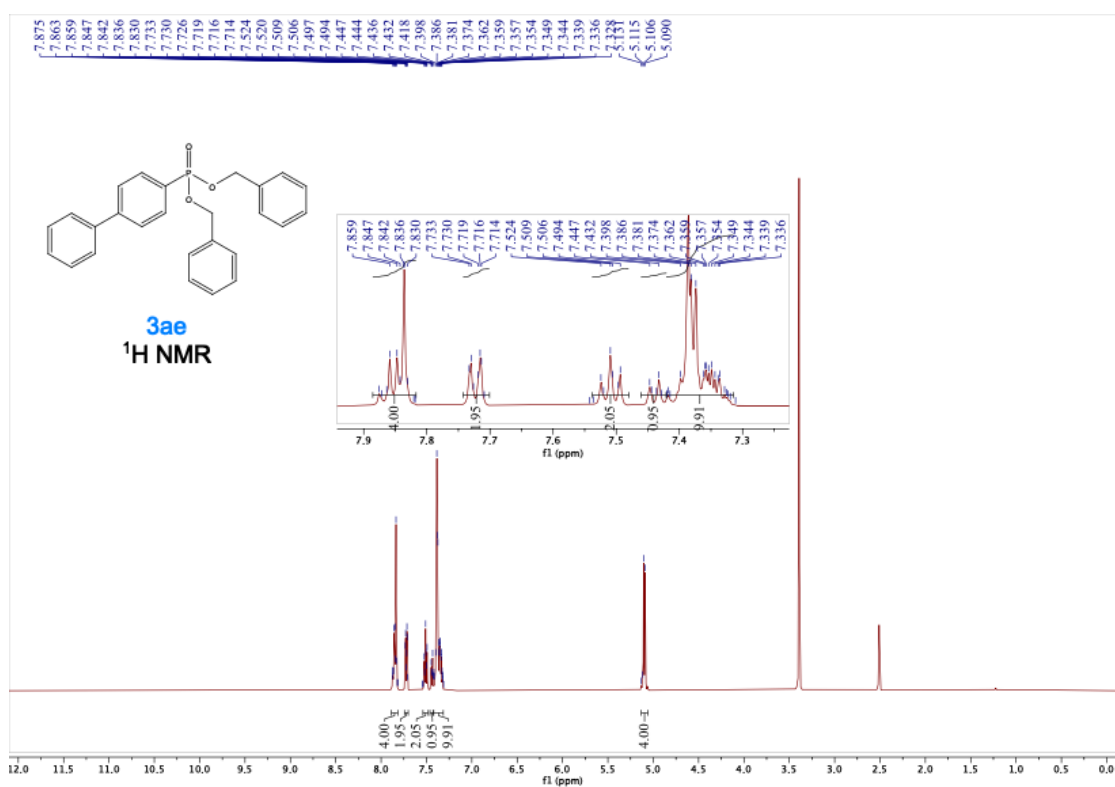
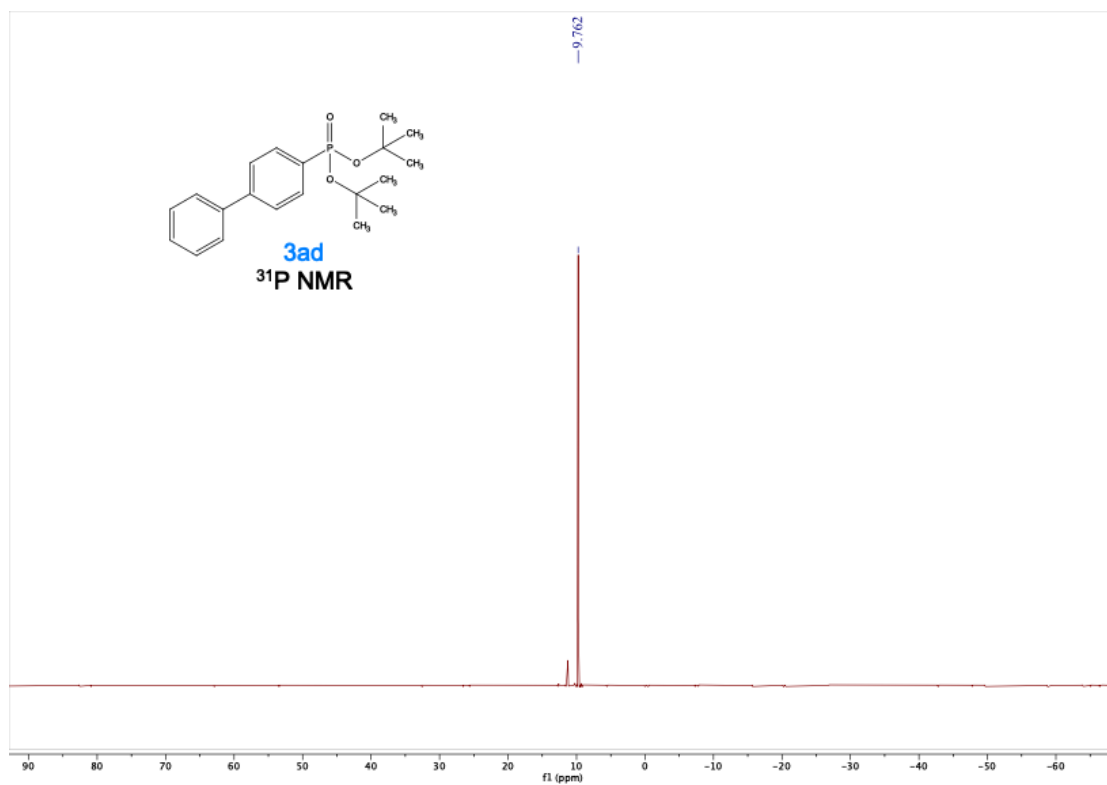


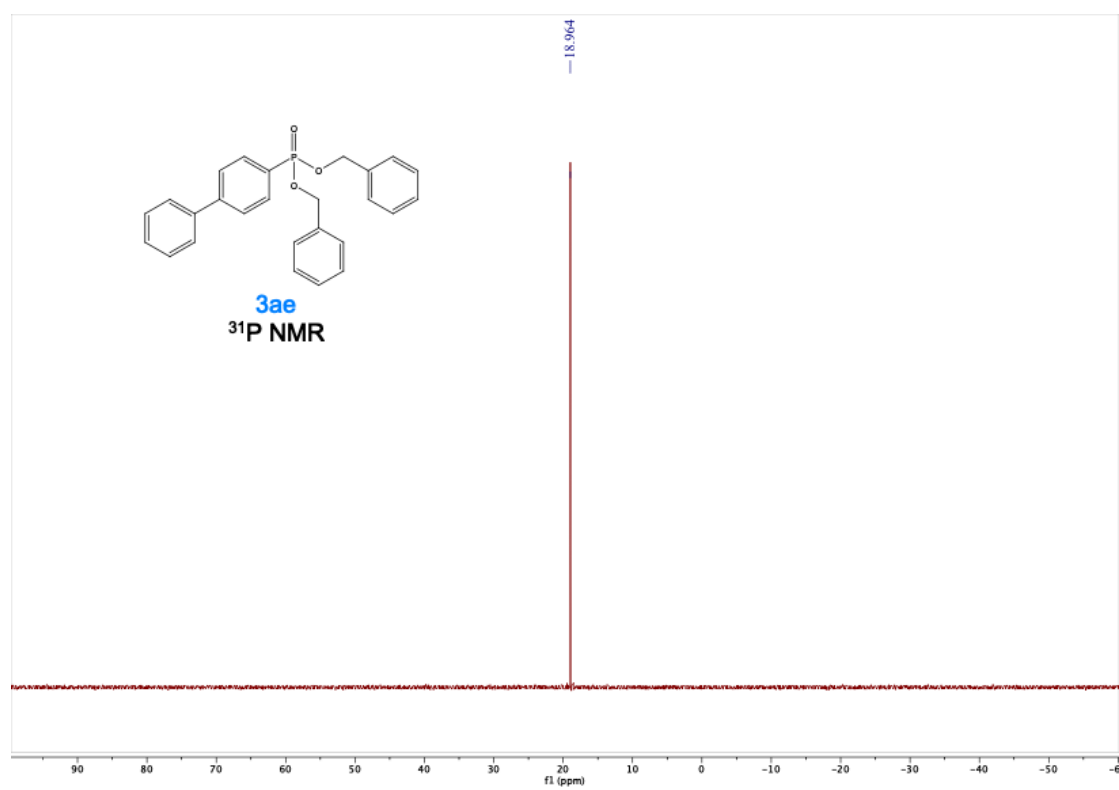
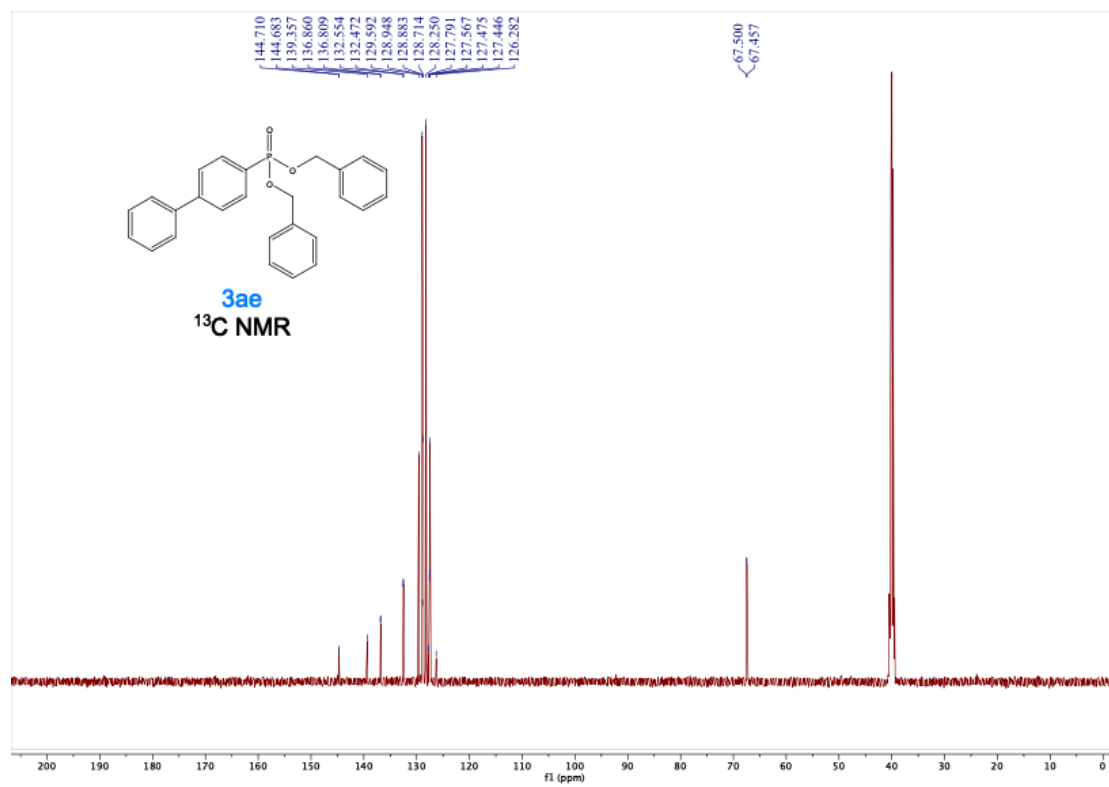


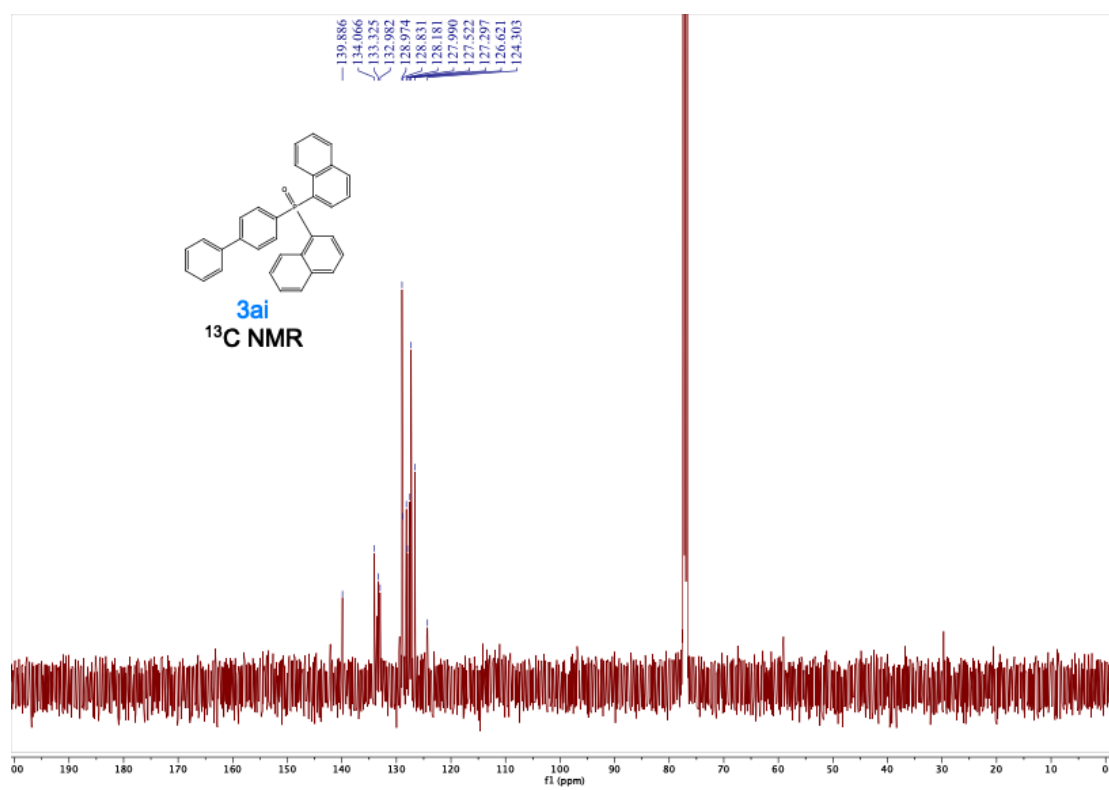
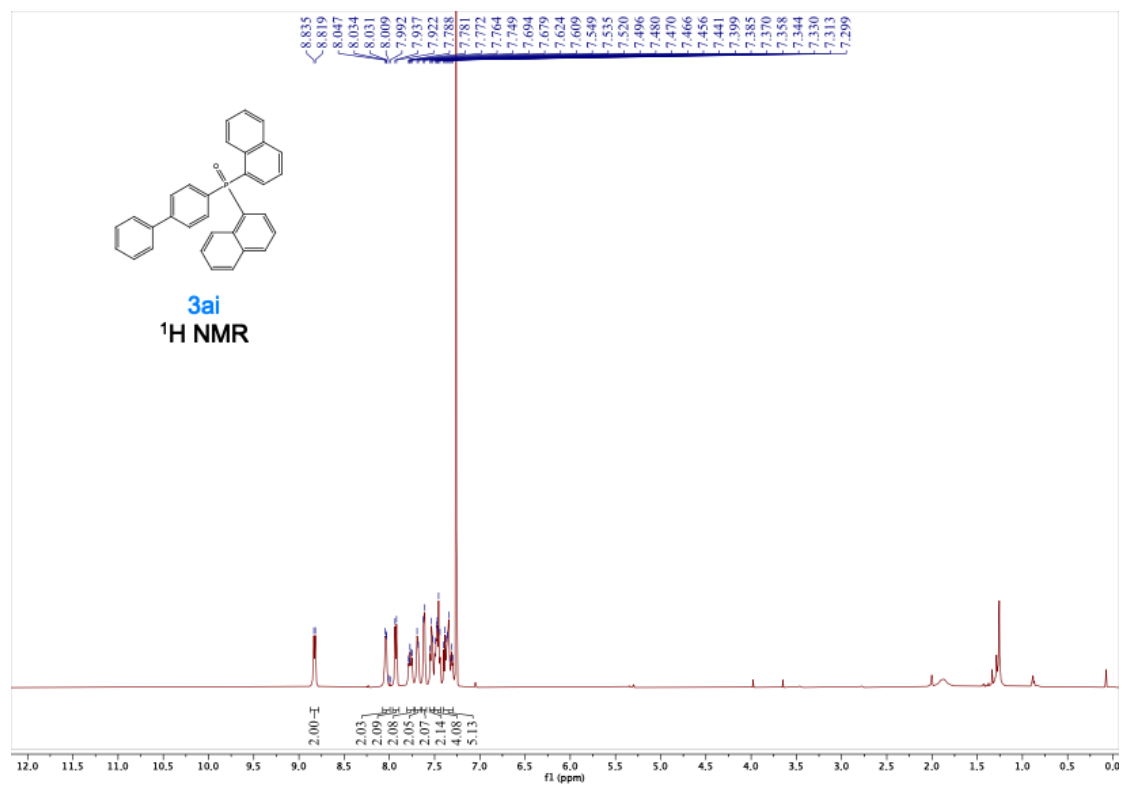


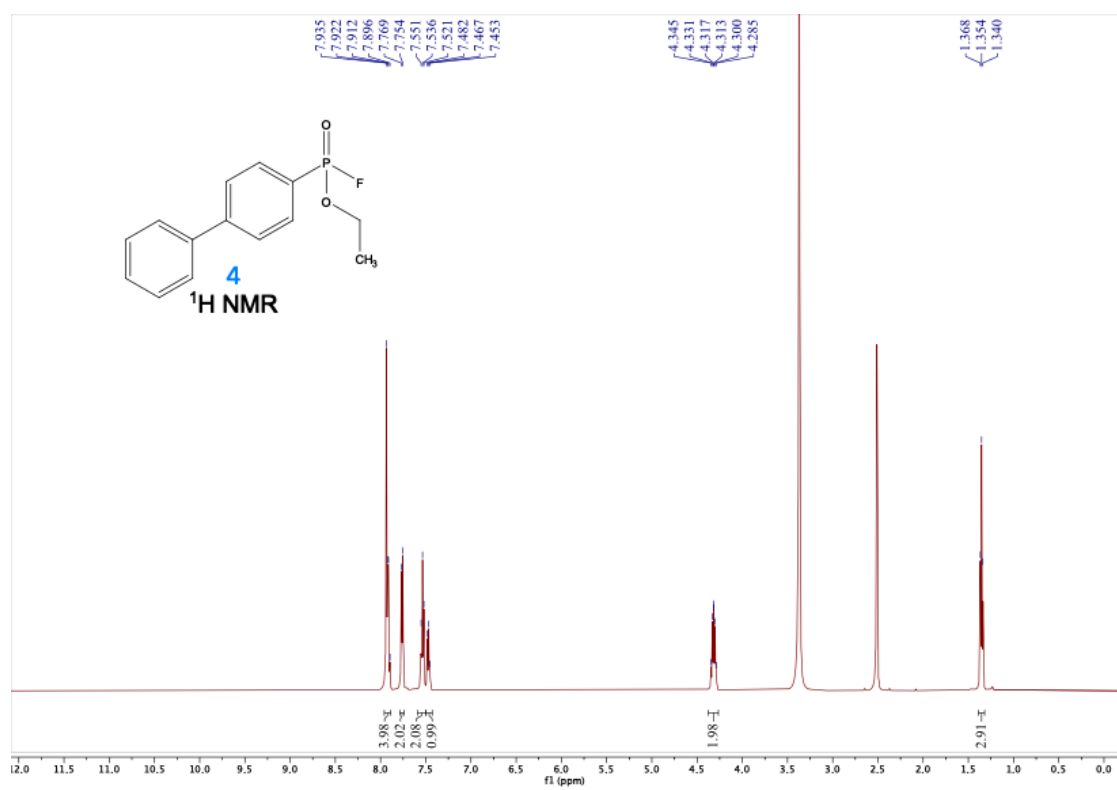
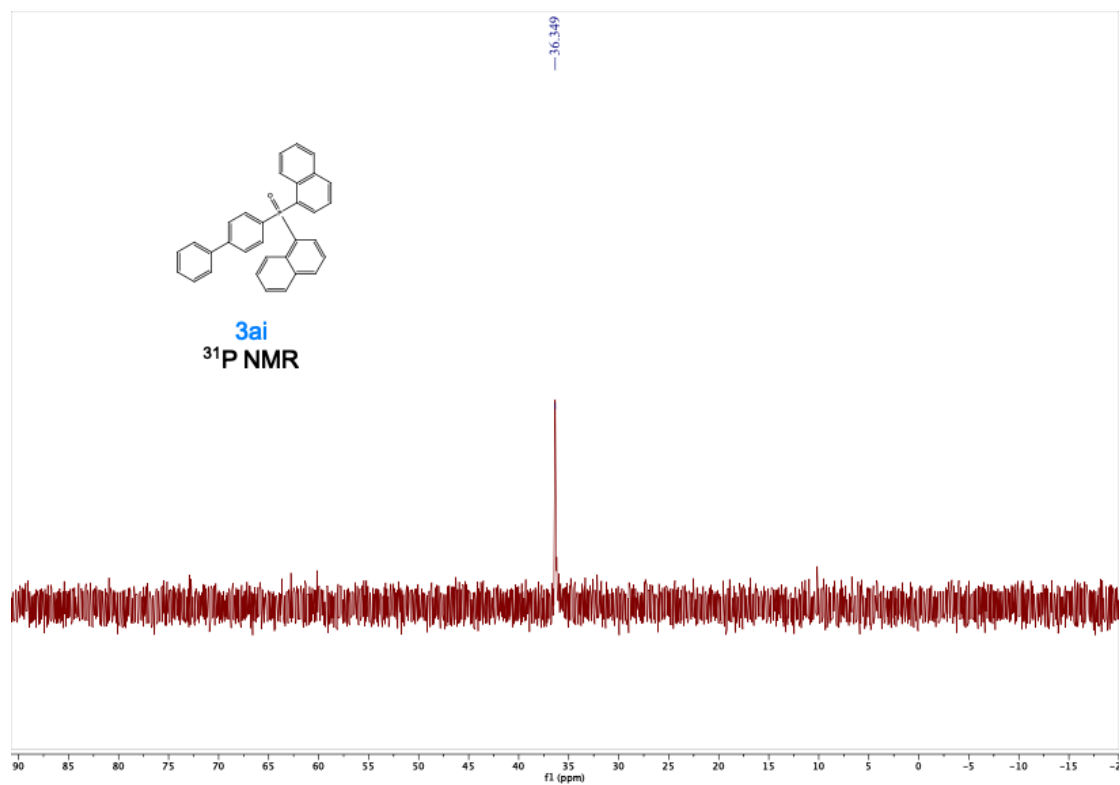


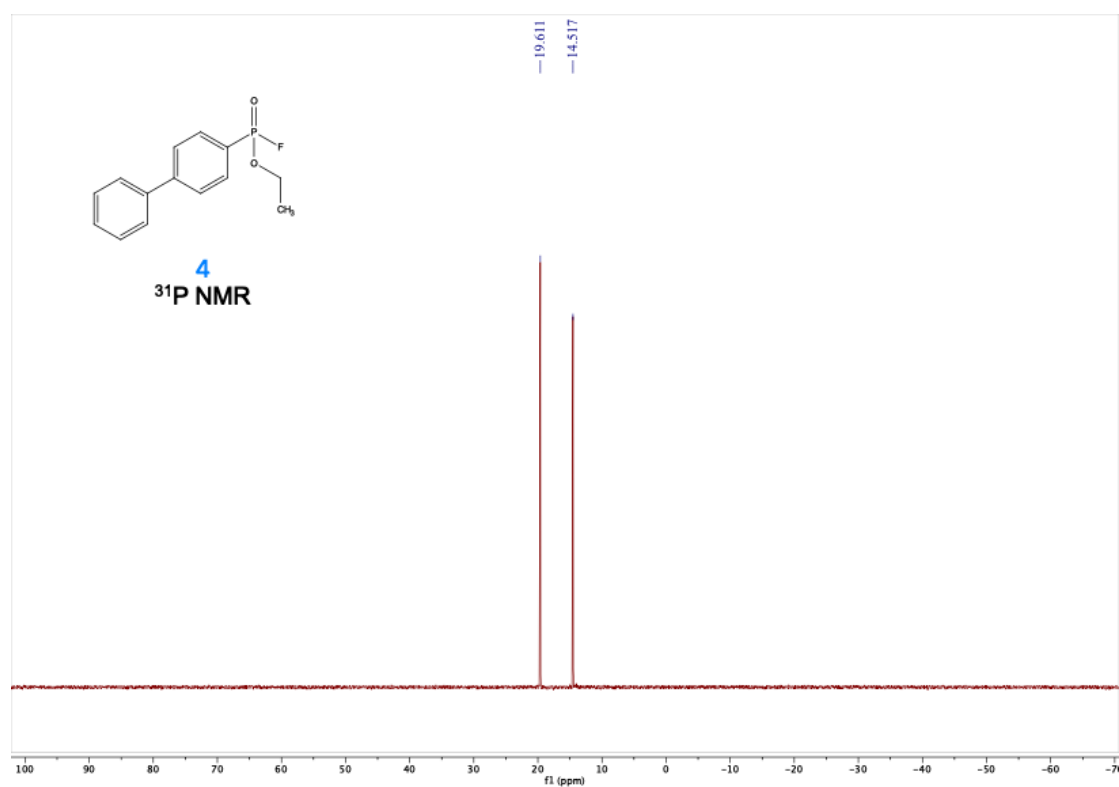
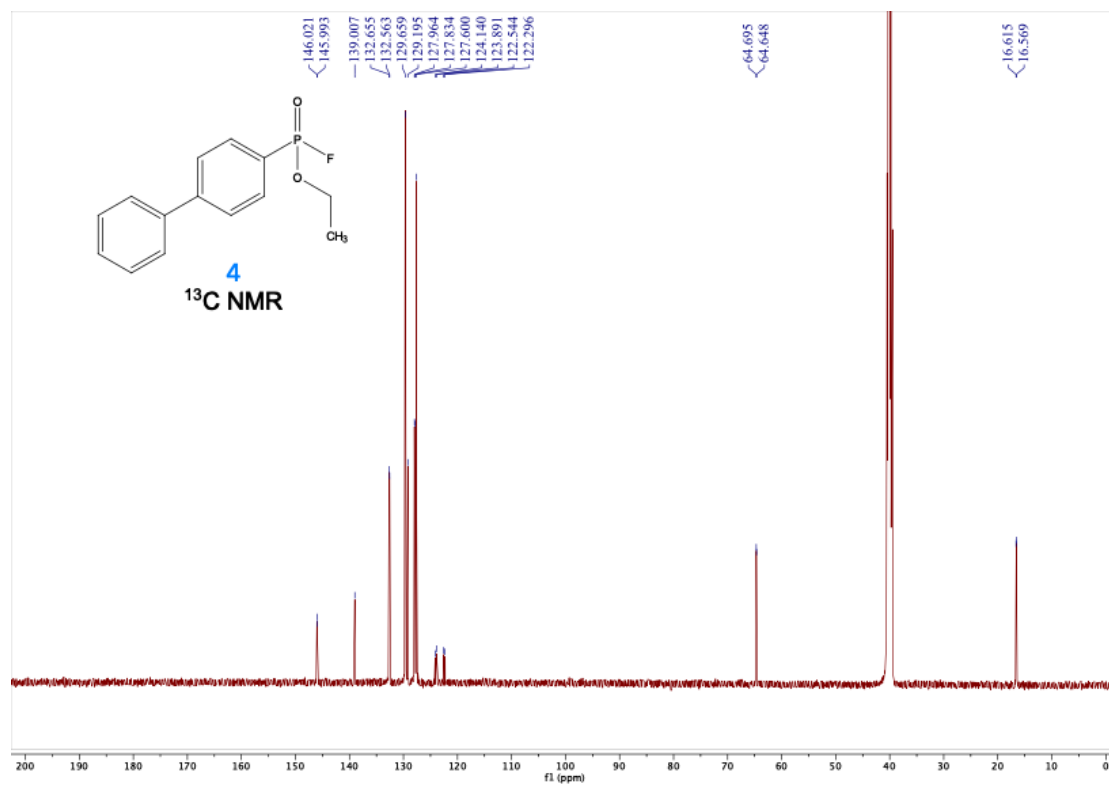


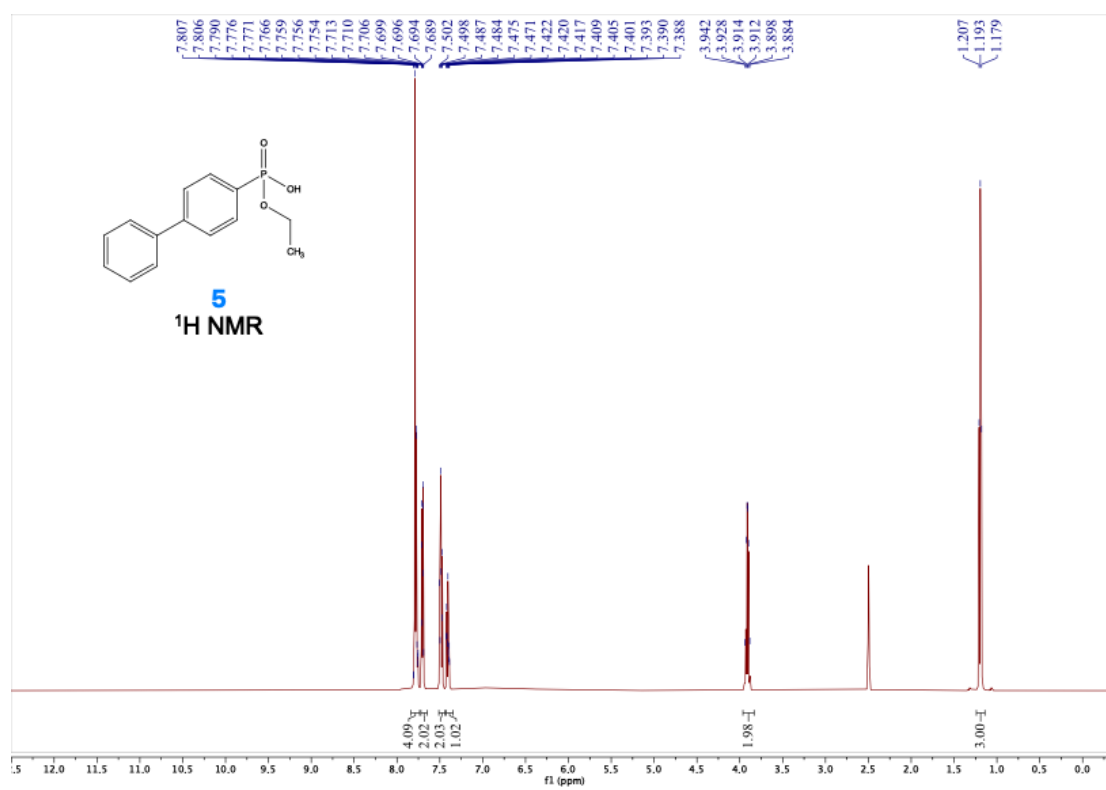
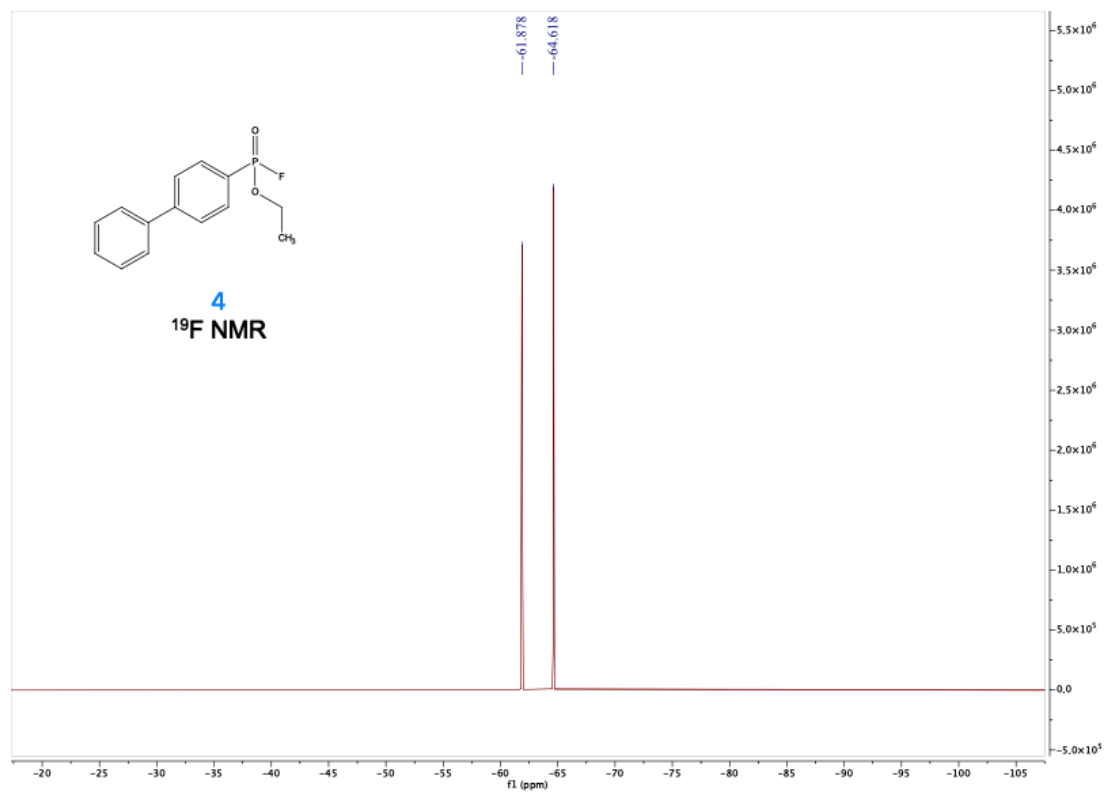


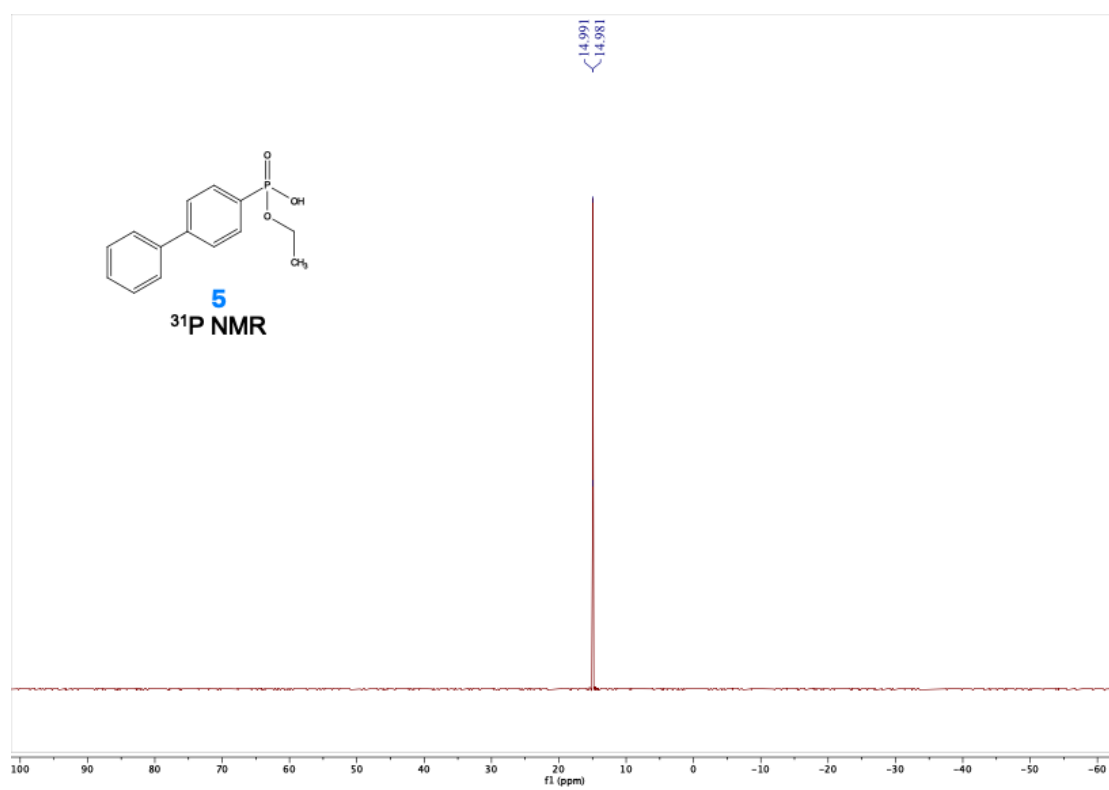
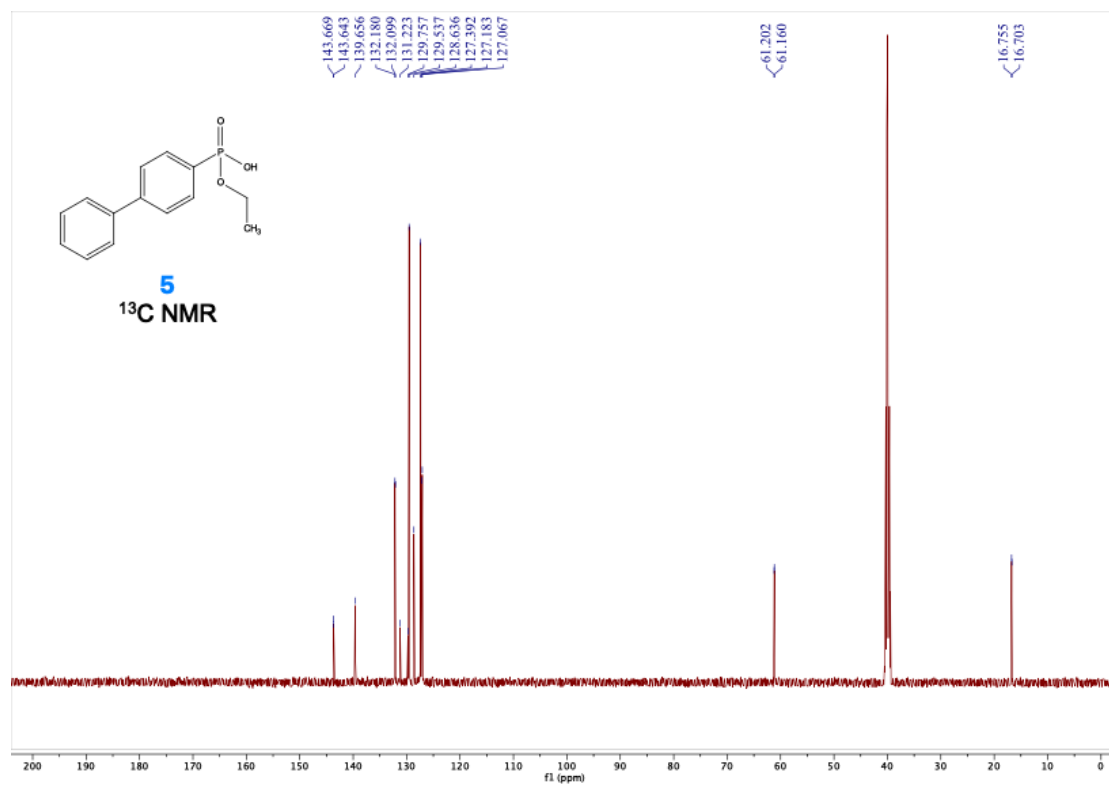


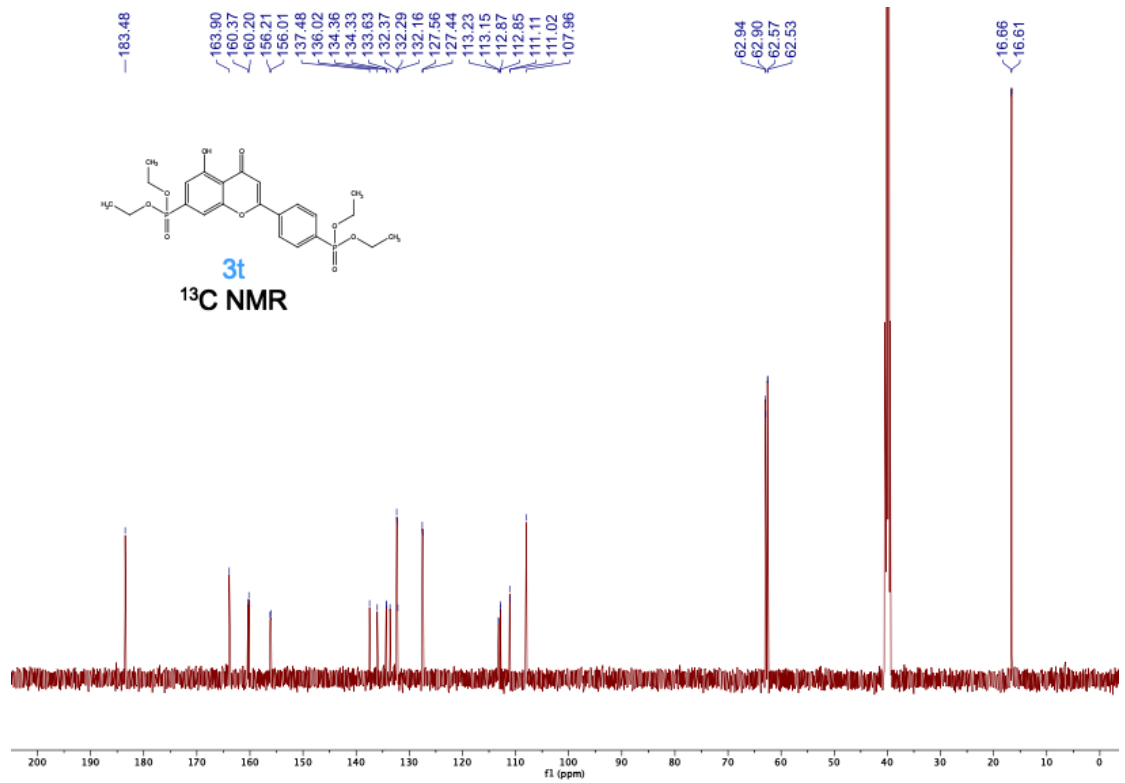
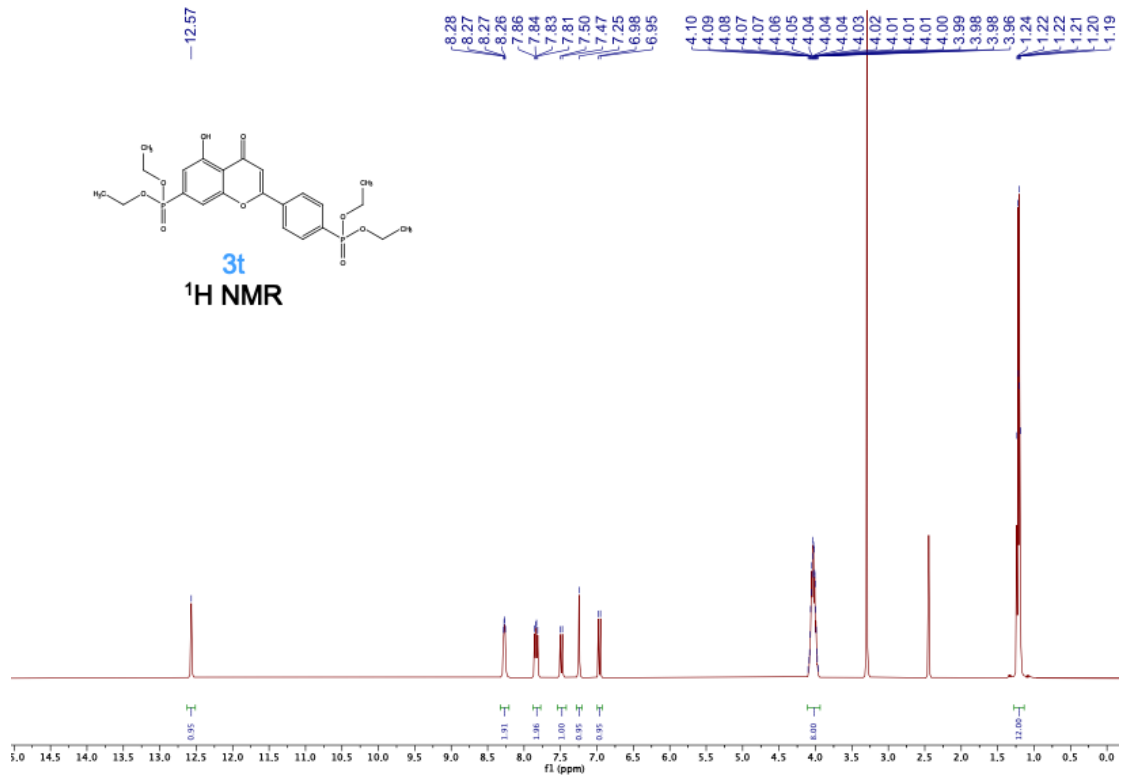


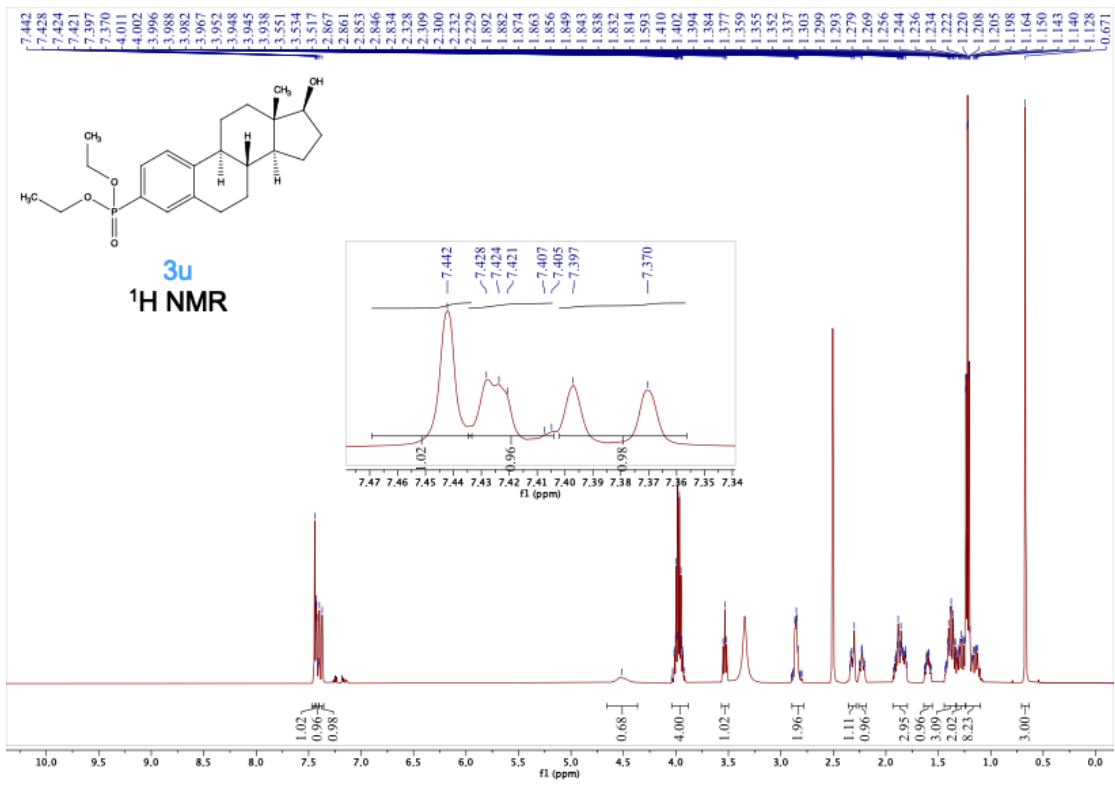
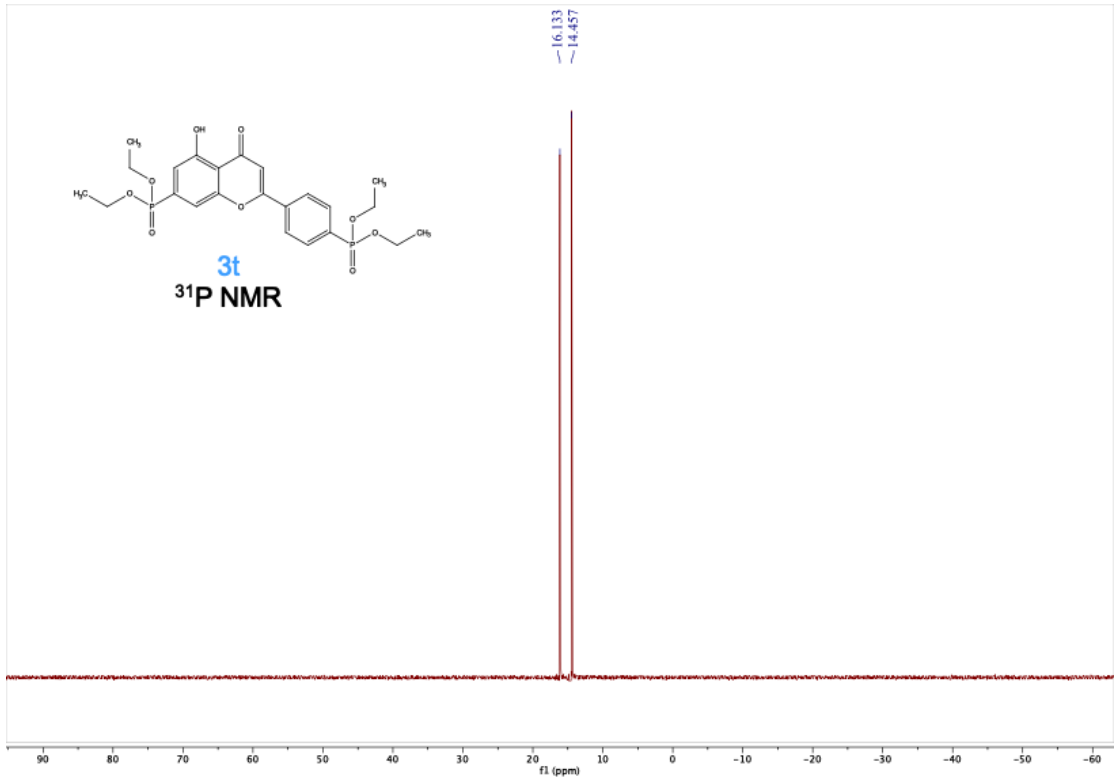


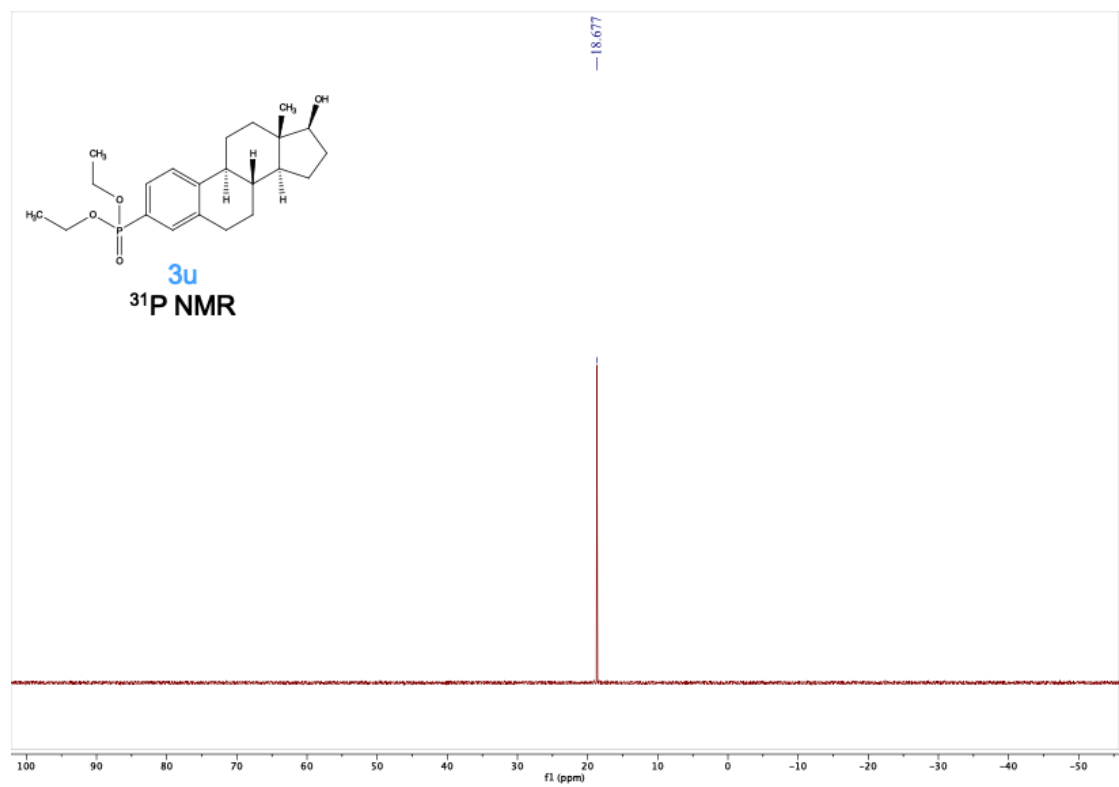
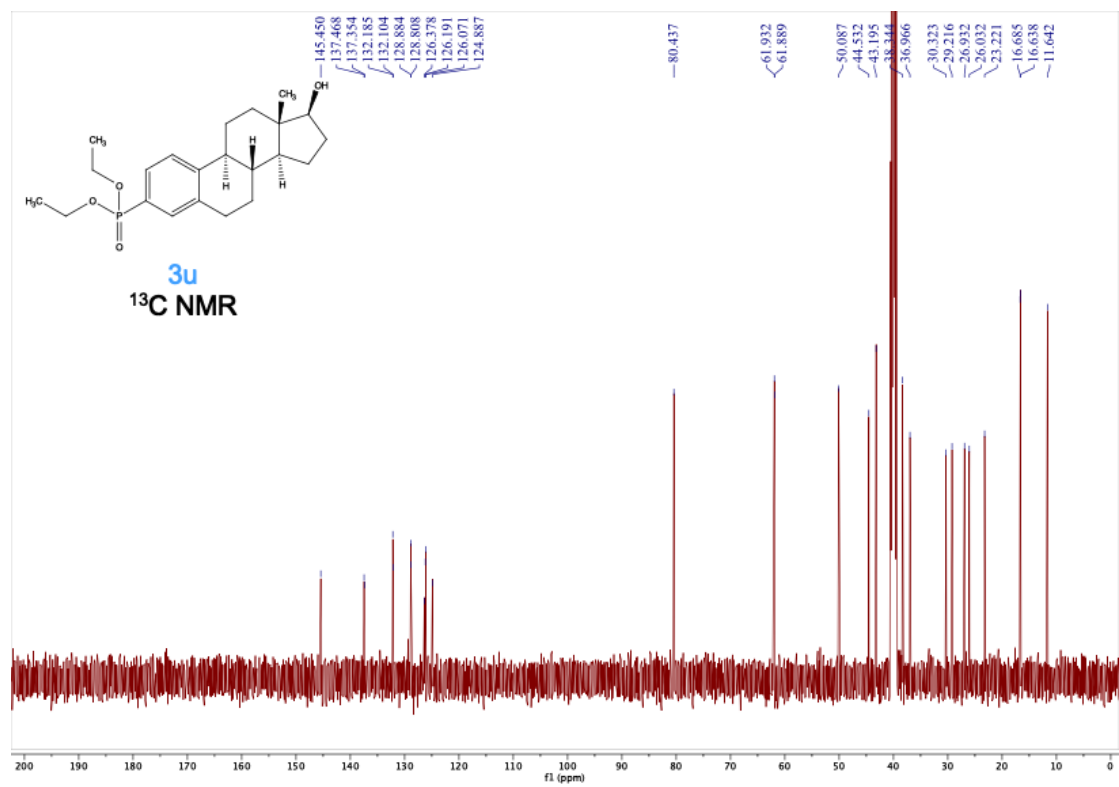


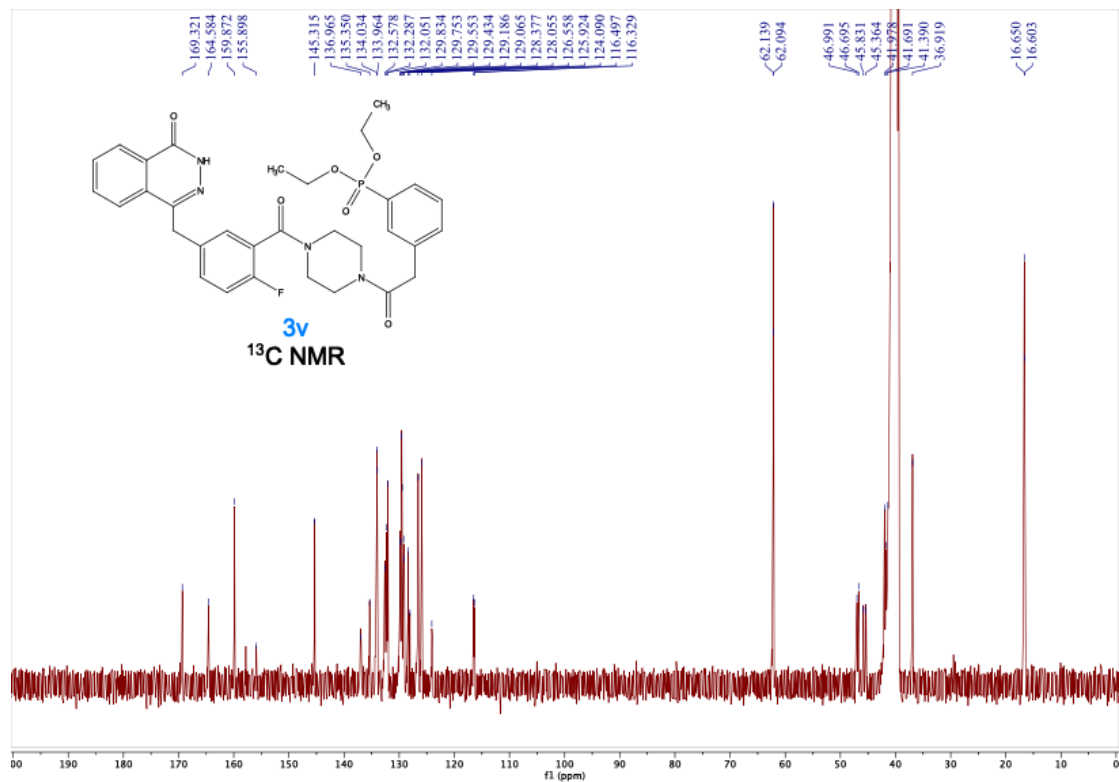
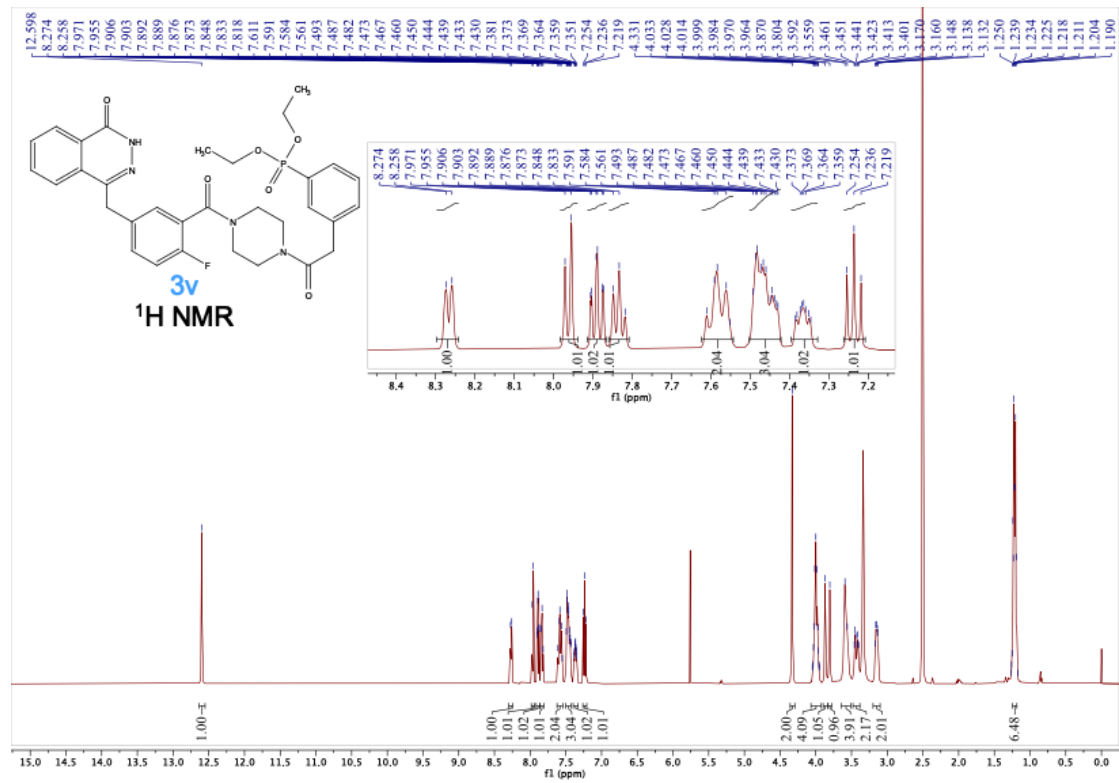


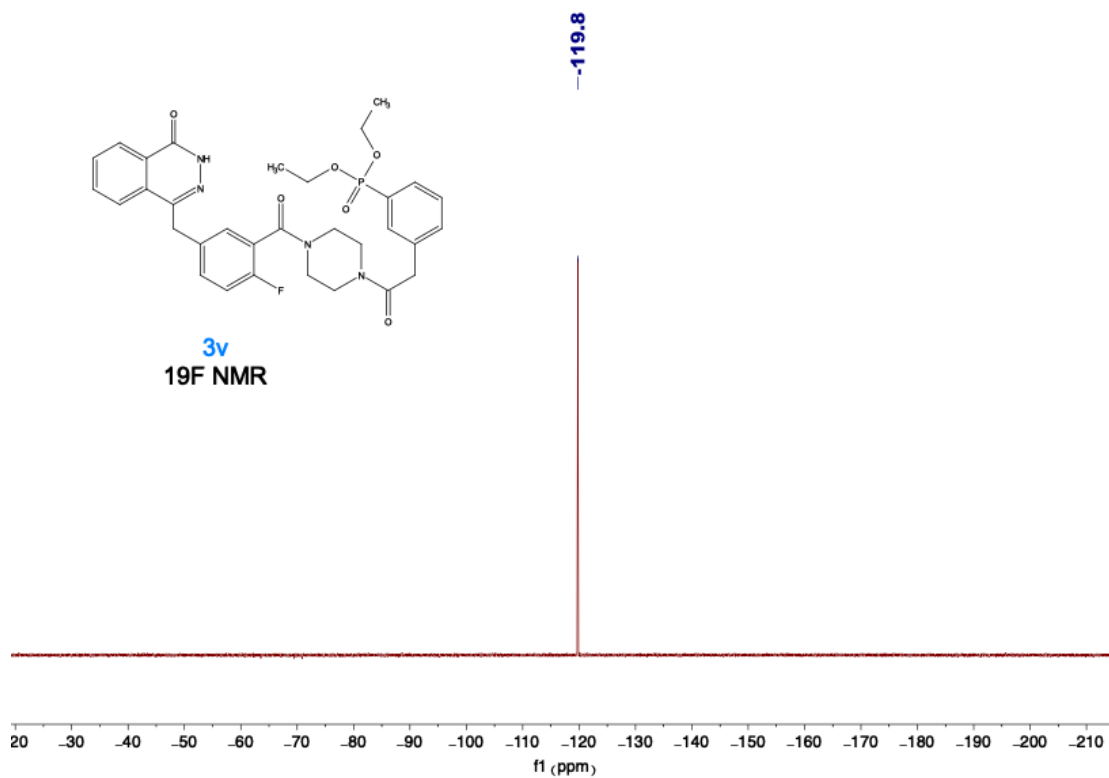
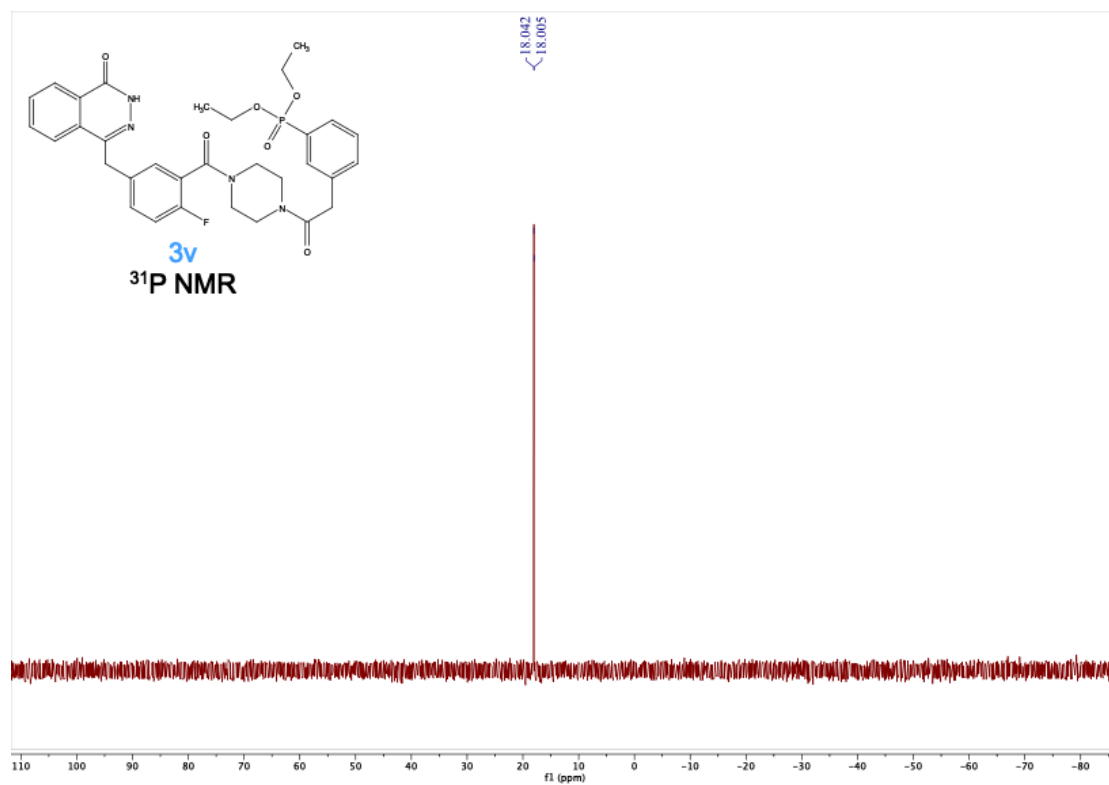


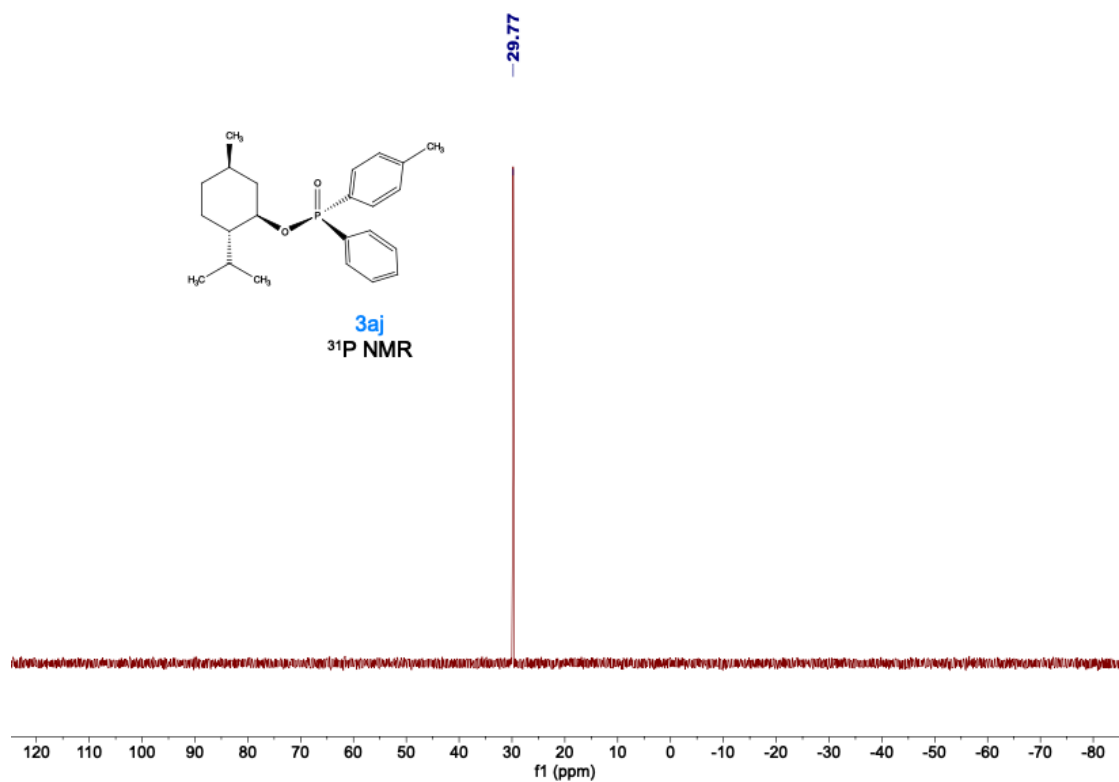
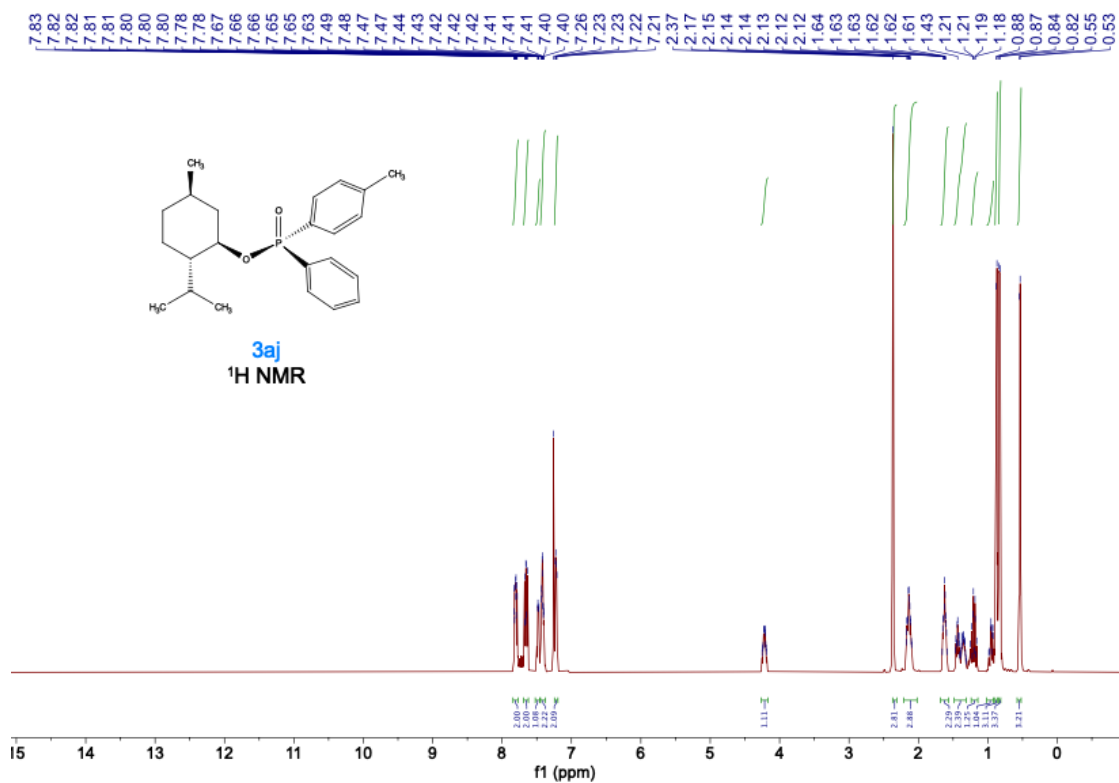


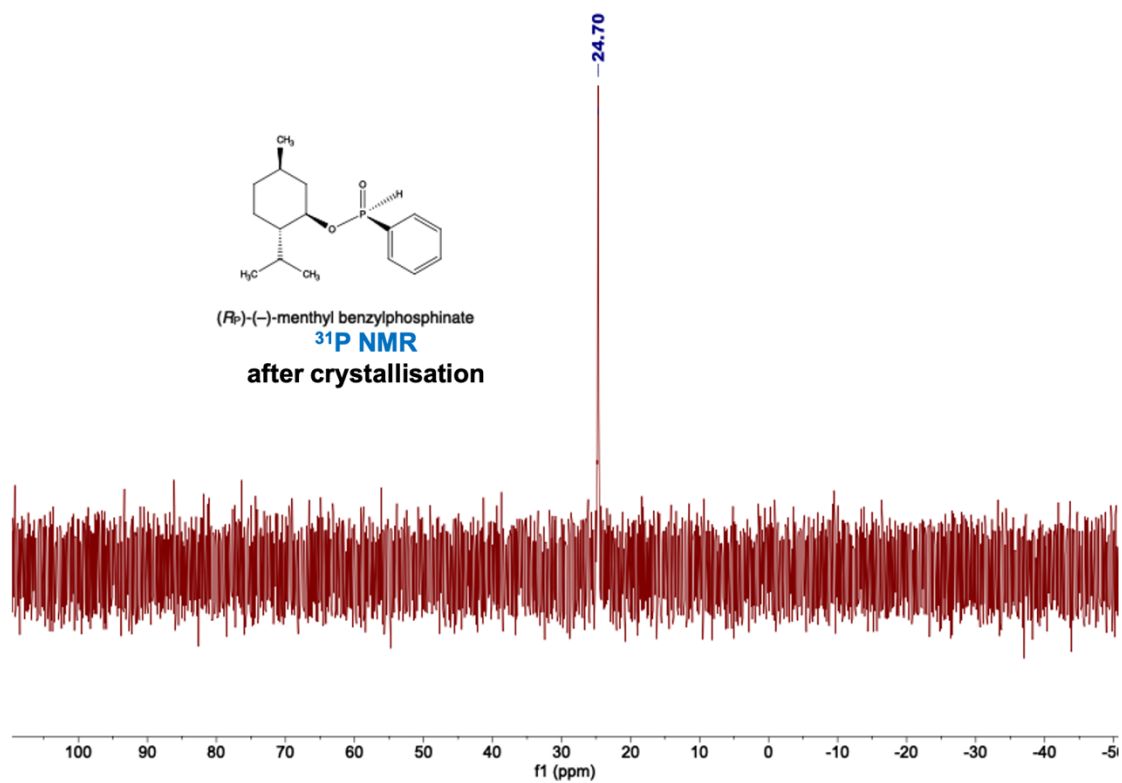
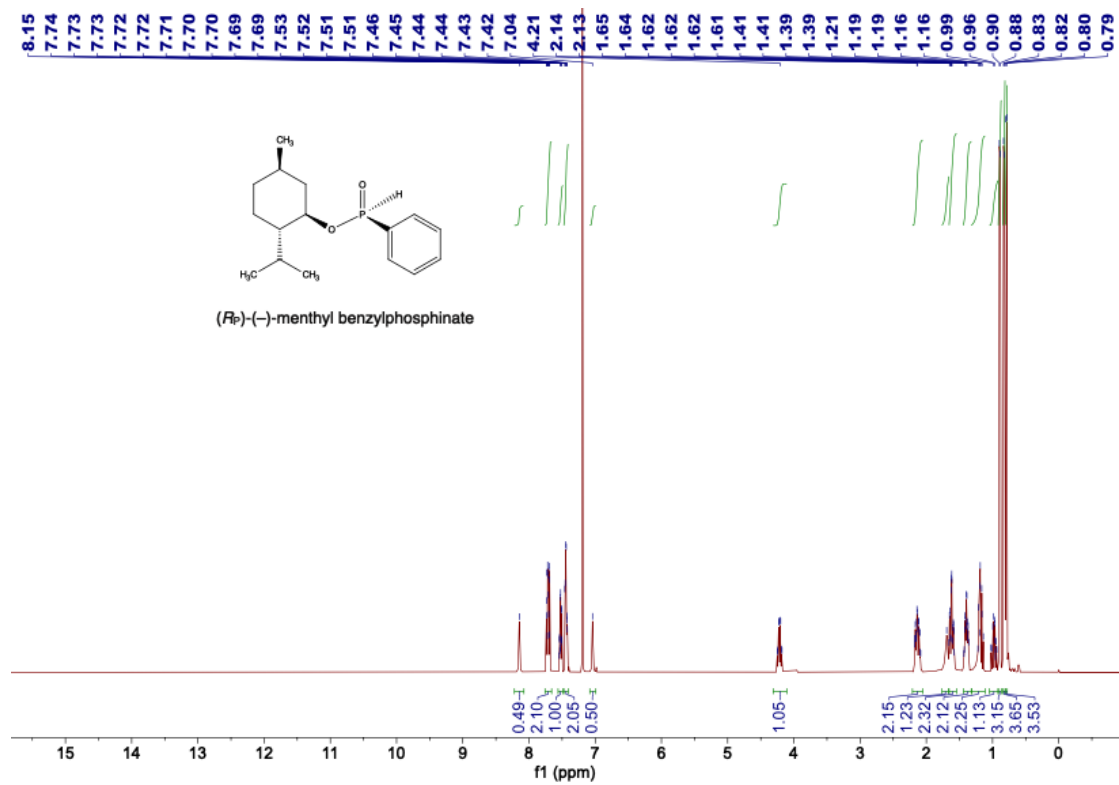


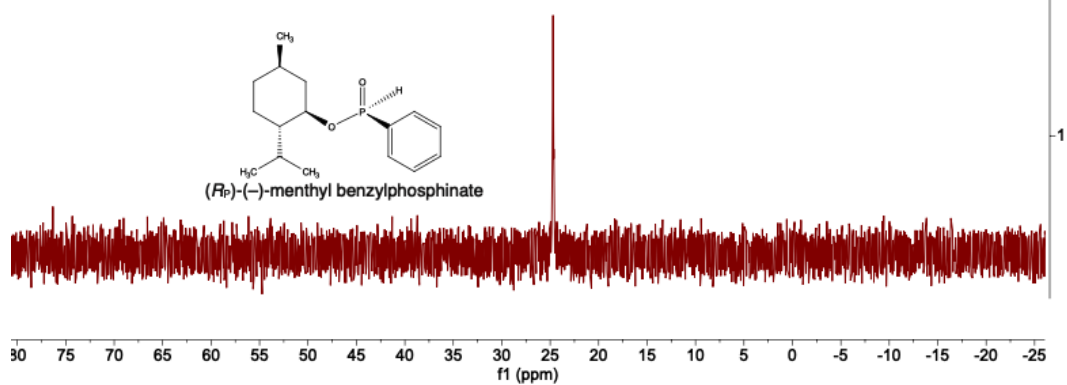
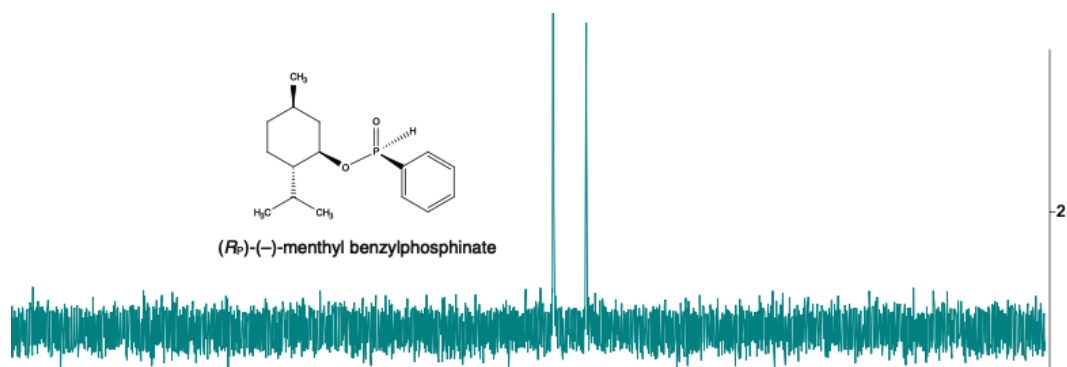
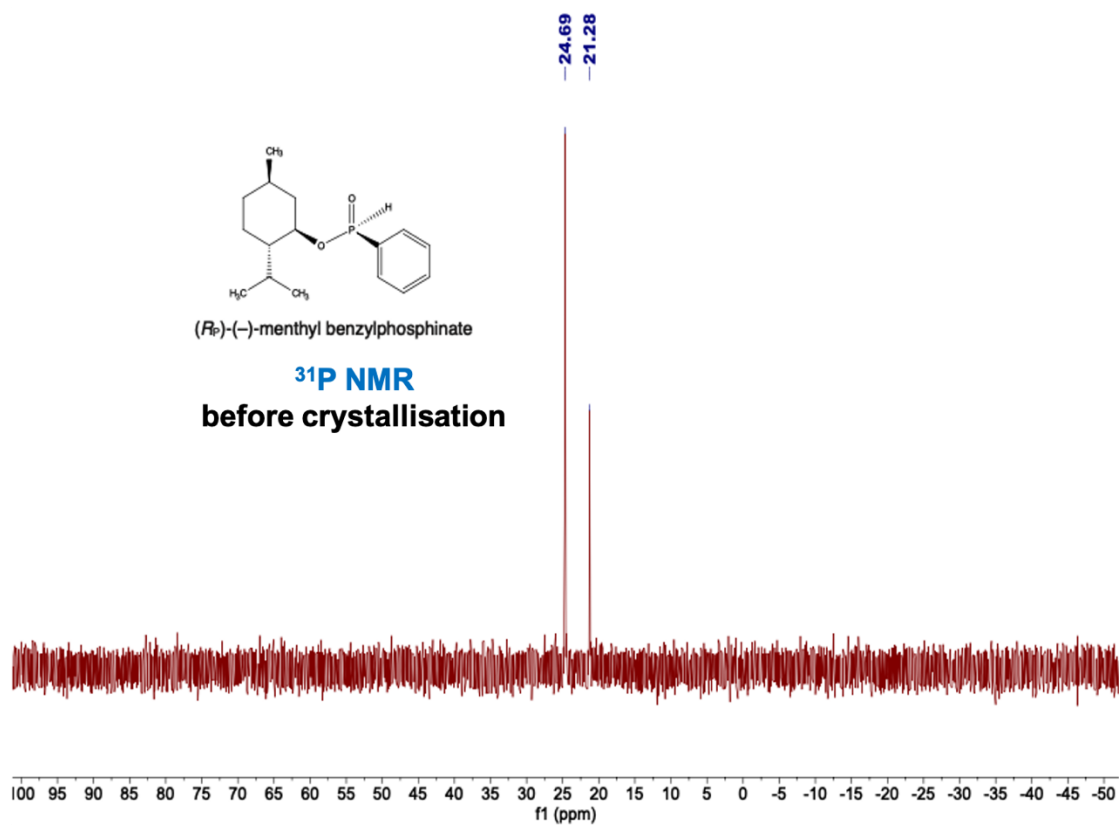












IV. Reference:

- (1) Zhang, S.; Xiong, H.; Lu, F.; Ma, F.; Gu, Y.; Ma, P.; Xu, H.; Yang, G. Synthesis of N -Acyl Sulfamates from Fluorosulfonates and Potassium Trimethylsilyloxy Imidates. *J. Org. Chem.* **2019**, *84* (23), 15380–15388. <https://doi.org/10.1021/acs.joc.9b02394>.
- (2) Zhao, Y. L.; Wu, G. J.; Han, F. S. Ni-Catalyzed Construction of C-P Bonds from Electron-Deficient Phenols via the in Situ Aryl C-O Activation by PyBroP. *Chem. Commun.* **2012**, *48* (47), 5868–5870. <https://doi.org/10.1039/c2cc31718d>.
- (3) Huang, C.; Tang, X.; Fu, H.; Jiang, Y.; Zhao, Y. Proline/Pipecolinic Acid-Promoted Copper-Catalyzed P -Arylation. *J. Org. Chem.* **2006**, *71* (13), 5020–5022. <https://doi.org/10.1021/jo060492j>.
- (4) Ackermann, L.; Born, R.; Spatz, J. H.; Meyer, D. Efficient Aryl-(Hetero)Aryl Coupling by Activation of C-Cl and C-F Bonds Using Nickel Complexes of Air-Stable Phosphine Oxides. *Angew. Chemie - Int. Ed.* **2005**, *44* (44), 7216–7219. <https://doi.org/10.1002/anie.200501860>.
- (5) Berrino, R.; Cacchi, S.; Fabrizi, G.; Goggiamani, A.; Stabile, P. Arenediazonium Tetrafluoroborates in Palladium-Catalyzed C-P Bond-Forming Reactions. Synthesis of Arylphosphonates, -Phosphine Oxides, and -Phosphines. *Org. Biomol. Chem.* **2010**, *8* (20), 4518–4520. <https://doi.org/10.1039/c0ob00243g>.
- (6) Edder, C.; Fréchet, J. M. J. Synthesis of Bridged Oligothiophenes: Toward a New Class of Thiophene-Based Electroactive Surfactants. *Org. Lett.* **2003**, *5* (11), 1879–1882. <https://doi.org/10.1021/ol034398q>.
- (7) Kabachnik, M. M.; Solntseva, M. D.; Izmer, V. V.; Novibora, Z. S.; Beletsbaya, I. P. Palladium-Catalyzed. Phase-Transfer Arylation of Dialkyl Phosphonates. *Russ. J. Org. Chem.* **1998**, *34*, 93–97.
- (8) Suto, Y.; Tsuji, R.; Kanai, M.; Shibasaki, M. Cu(I)-Catalyzed Direct Enantioselective Cross Aldol-Type Reaction of Acetonitrile. *Org. Lett.* **2005**, *7* (17), 3757–3760. <https://doi.org/10.1021/ol051423e>.
- (9) Andaloussi, M.; Lindh, J.; Sävmärker, J.; Sjöberg, P. J. R.; Larhed, M. Microwave-Promoted Palladium(II)-Catalyzed C-P Bond Formation by Using Arylboronic Acids or Aryltrifluoroborates. *Chem. - A Eur. J.* **2009**, *15* (47), 13069–13074. <https://doi.org/10.1002/chem.200901473>.
- (10) Liu, C.; Ji, C. L.; Zhou, T.; Hong, X.; Szostak, M. Decarbonylative Phosphorylation of Carboxylic Acids via Redox-Neutral Palladium Catalysis. *Org. Lett.* **2019**, *21* (22), 9256–9261. <https://doi.org/10.1021/acs.orglett.9b03678>.
- (11) Bonnaventure, I.; Charette, A. B. Probing the Importance of the Hemilabile Site of Bis(Phosphine) Monoxide Ligands in the Copper-Catalyzed Addition of Diethylzinc to N-Phosphinoylimines: Discovery of New Effective Chiral Ligands. *J. Org. Chem.* **2008**, *73* (16), 6330–6340.

- <https://doi.org/10.1021/jo800969x>.
- (12) Ghosh, I.; Shaikh, R. S.; König, B. Sensitization-Initiated Electron Transfer for Photoredox Catalysis. *Angew. Chemie - Int. Ed.* **2017**, *56* (29), 8544–8549. <https://doi.org/10.1002/anie.201703004>.
- (13) van der Knaap, T. A.; Klebach, T. C.; Lourens, R.; Vos, M.; Bickelhaupt, F. Oxidation Reactions of Phosphaalkenes. *J. Am. Chem. Soc.* **1983**, *105* (12), 4026–4032. <https://doi.org/10.1021/ja00350a047>.
- (14) Fu, W. C.; So, C. M.; Kwong, F. Y. Palladium-Catalyzed Phosphorylation of Aryl Mesylates and Tosylates. *Org. Lett.* **2015**, *17* (23), 5906–5909. <https://doi.org/10.1021/acs.orglett.5b03104>.
- (15) Xiang, C. B.; Bian, Y. J.; Mao, X. R.; Huang, Z. Z. Coupling Reactions of Heteroarenes with Phosphites under Silver Catalysis. *J. Org. Chem.* **2012**, *77* (17), 7706–7710. <https://doi.org/10.1021/jo301108g>.
- (16) Yang, J.; Xiao, J.; Chen, T.; Han, L. B. Nickel-Catalyzed Phosphorylation of Phenol Derivatives via C-O/P-H Cross-Coupling. *J. Org. Chem.* **2016**, *81* (9), 3911–3916. <https://doi.org/10.1021/acs.joc.6b00289>.
- (17) Yang, J.; Xiao, J.; Chen, T.; Han, L. B. Nickel-Catalyzed Phosphorylation of Aryl Triflates with P([Formula Presented]) Compounds. *J. Organomet. Chem.* **2016**, *820*, 120–124. <https://doi.org/10.1016/j.jorganchem.2016.07.026>.
- (18) Onouchi, H.; Maeda, K.; Yashima, E. A Helical Polyelectrolyte Induced by Specific Interactions with Biomolecules in Water [10]. *J. Am. Chem. Soc.* **2001**, *123* (30), 7441–7442. <https://doi.org/10.1021/ja0160647>.
- (19) W.B. Farnham, R.K. Murray, K. Mislow, Stereospecific alkylation of menthyl phenylphosphinate, *J. Am. Chem. Soc.* **92** (1970) 5809–5810. doi:10.1021/ja00722a083.
- (20) D. Moraleda, D. Gatineau, D. Martin, L. Giordano, G. Buono, A simple route to chiral phosphinous acid-boranes, *Chem. Commun.* (2008) 3031–3033. doi:10.1039/b802817f.
- (21) W.M. Wang, L.J. Liu, C.Q. Zhao, L.B. Han, Diastereoselective hydrolysis of asymmetric P-Cl species and synthesis of optically pure (RP)-(-)-menthyl H-phenylphosphinate, *European J. Org. Chem.* **2015** (2015) 2342–2345. doi:10.1002/ejoc.201500126.
- (22) X. Zhang, H. Liu, X. Hu, G. Tang, J. Zhu, Y. Zhao, Ni(II)/Zn Catalyzed Reductive Coupling of Aryl Halides with Diphenylphosphine Oxide in Water, *Org. Lett.* **13** (2011) 3478–3481. doi:10.1021/ol201141m.