

# Supporting Information

## **Catalytic asymmetric multicomponent reaction of isocyanide, isothiocyanate and alkylidene malonates**

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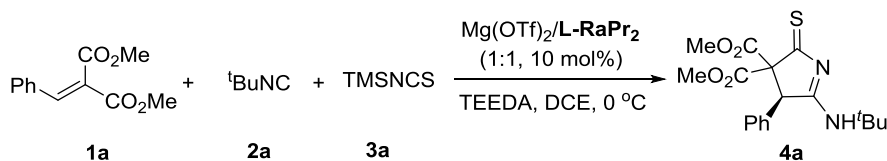
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## 1. General remarks

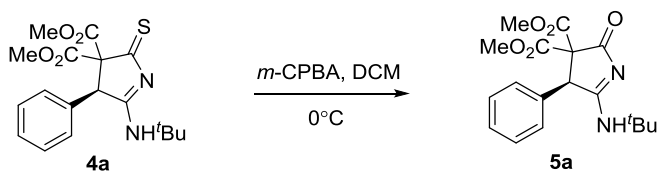
$^1\text{H}$  NMR spectra were recorded on Bruker ASCEND 400 MHz. Chemical shifts were recorded in ppm relative to tetramethylsilane and with the solvent resonance as the internal standard ( $\text{CDCl}_3$ ,  $\delta = 7.26$ ). Data were reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constants (Hz), integration.  $^{13}\text{C}\{^1\text{H}\}$  NMR data were collected on Bruker ASCEND 101 MHz with complete proton decoupling. Chemical shifts were reported in ppm from the tetramethylsilane with the solvent resonance as internal standard ( $\text{CDCl}_3$ ,  $\delta = 77.16$ ). Enantiomeric excesses were determined by chiral HPLC analysis on Daicel Chiralcel ADH, IA, IB, IC, IE, and Phenomenex Chiralcel Lux 5u Cellulose-2 at 23 °C with UV detector in comparison with the authentic racemates. Optical rotations were determined after flash column chromatography purification and reported as follows:  $[\alpha]_D^{25}$  (c: g/100 mL, in  $\text{CH}_2\text{Cl}_2$ ,  $\lambda = 589$  nm). HRMS were recorded on a commercial apparatus (ESI source). IR was recorded on Bruker Tensor II spectrometer with Plantium ATR accessory. All the reactions were carried out under an atmosphere of nitrogen in over-dried apparatus. All the solvents were purified by usual methods before use. Chromatography: Qingdao Haiyang silica gel, HG/T2354-92, H CP. Reagents purchased from commercial suppliers were used: *tert*-butyl isocyanide (Aladdin), TMSNCS (Alfa), TMSNCO (TCI), tetraethyl ethylenediamine (3A), magnesium trifluoromethanesulfonate (Alfa). The *N,N'*-dioxide ligands<sup>1</sup>, alkylidene malonates<sup>2</sup> and  $\text{Et}_3\text{HNCS}$ <sup>3</sup> were synthesized according to known procedures.

## 2. Experimental procedures

**General procedure 1** for preparation of racemic products **4a**: To an oven-dried tube were added  $\text{Mg}(\text{OTf})_2$  (0.01 mmol, 10 mol%),  $\text{Et}_3\text{N}$  (0.05 mmol, 50 mol%), dimethyl 2-benzylidenemalonate **1a** (0.10 mmol), *tert*-butyl isocyanide **2a** (0.15 mmol), TMSNCS **3a** (0.15 mmol), and  $\text{CH}_2\text{ClCH}_2\text{Cl}$  (1.0 mL). The mixture was stirred in  $\text{CH}_2\text{ClCH}_2\text{Cl}$  at 35 °C for 24 h and directly subjected to flash column chromatography on silica gel (eluent: petroleum ether/ethyl acetate = 2/1, v/v) to afford the racemic product **4a** as a yellow solid.

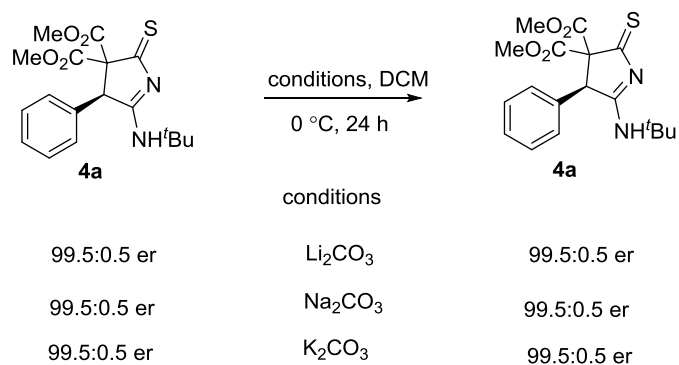


**General procedure 2** for preparation of enantioenriched products **4a**: To an oven-dried tube were added  $\text{Mg}(\text{OTf})_2$  (0.01 mmol, 10 mol%), **L-RaPr**<sub>2</sub> (0.01 mmol, 10 mol%), tetraethylenediamine (TEEDA) dimethyl 2-benzylidenemalonate (**1a**, 0.10 mmol) and  $\text{CH}_2\text{ClCH}_2\text{Cl}$  (1.0 mL). The mixture was stirred in  $\text{CH}_2\text{ClCH}_2\text{Cl}$  at 35 °C for 30 min. Then, TMSNCS **3a** (0.15 mmol) and *tert*-butyl isocyanide (**2a**, 0.15 mmol) were added to the mixture at 0 °C. The mixture was stirred in  $\text{CH}_2\text{ClCH}_2\text{Cl}$  at 0 °C for 96 h and directly subjected to flash column chromatography on silica gel (eluent: petroleum ether/ethyl acetate = 2/1, v/v) to afford the product **4a** as a yellow solid (29.1 mg, 80% yield, 95:5 er).



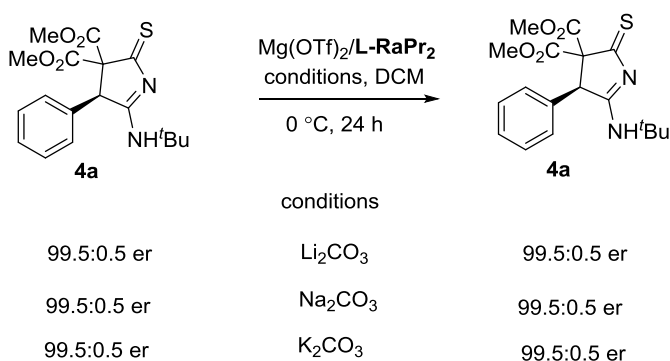
**General procedure 3** for synthesis of the oxo-product **5a**: To an oven-dried tube were added **4a** and  $\text{CH}_2\text{Cl}_2$ , then, *m*-CPBA (2 equiv) was added to the mixture at 0 °C. The mixture was stirred in  $\text{CH}_2\text{Cl}_2$  at 0 °C and directly subjected to flash column chromatography on basic  $\text{Al}_2\text{O}_3$  (upper layer) and silica gel (lower layer) (eluent: petroleum ether/ethyl acetate = 1/1, v/v) to afford the product **5a**.

## 3. Control experiments



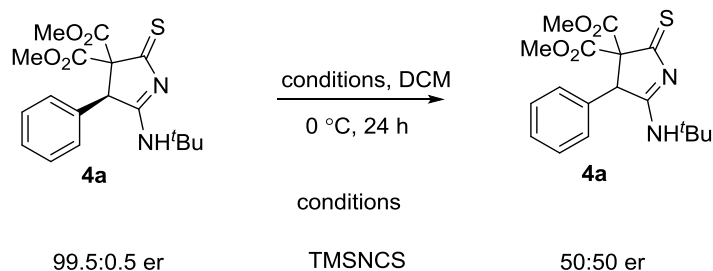
**Control experiment 1:** To an oven-dried tube were added **4a** (**4a** with 99.5:0.5 er could be easily obtained by recrystallization of petroleum ether and ethyl acetate) and CH<sub>2</sub>Cl<sub>2</sub>, then, base (0.5 equiv) was added to the mixture at 0 °C. The mixture was stirred in CH<sub>2</sub>Cl<sub>2</sub> at 0 °C for 24 h and directly subjected to flash column chromatography afforded the product **4a**.

These results indicated the addition of base has no effect on the optical purity of the product.



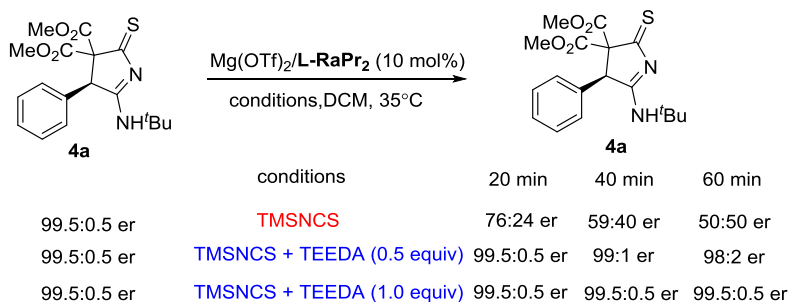
**Control experiment 2:** To an oven-dried tube were added Mg(OTf)<sub>2</sub> (10 mol%), L-RaPr<sub>2</sub> (10 mol%), base (0.5 equiv) and CH<sub>2</sub>Cl<sub>2</sub>, the mixture was stirred in CH<sub>2</sub>Cl<sub>2</sub> at 35 °C for 30 min. Then, **4a** was added to the mixture at 0 °C. The mixture was stirred in CH<sub>2</sub>Cl<sub>2</sub> at 0 °C for 24 h and directly subjected to flash column chromatography afforded the product **4a**.

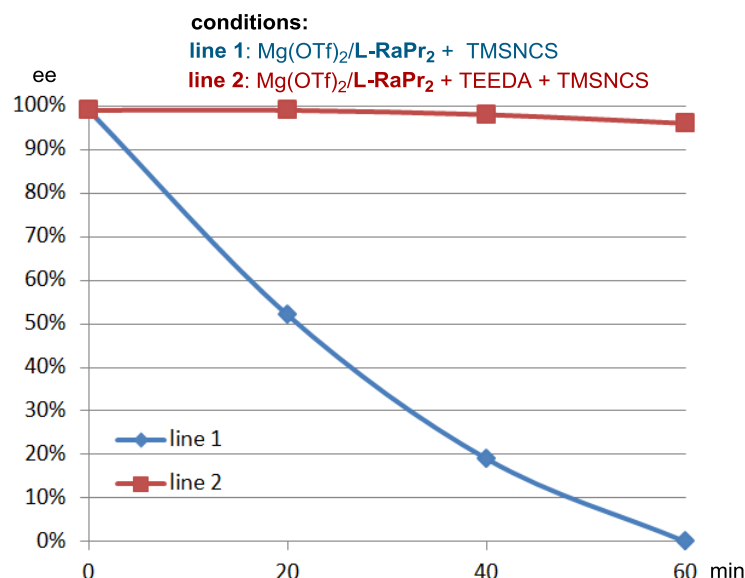
These results indicated that neither base nor chiral *N,N'*-dioxide/Mg<sup>II</sup> can promote the racemization of the product.



**Control experiment 3:** To an oven-dried tube were added **4a** and CH<sub>2</sub>Cl<sub>2</sub>, then, TMSNCS (1 equiv) was added to the mixture at 0 °C. The mixture was stirred in CH<sub>2</sub>Cl<sub>2</sub> at 0 °C for 24 h and directly subjected to flash column chromatography afforded the product **4a**.

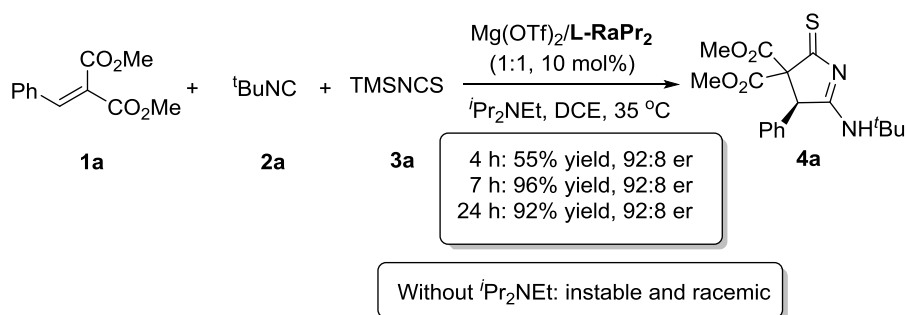
These results indicated that TMSNCS led to the racemization of the product efficiently, which was also investigated by H-D exchange experiment.





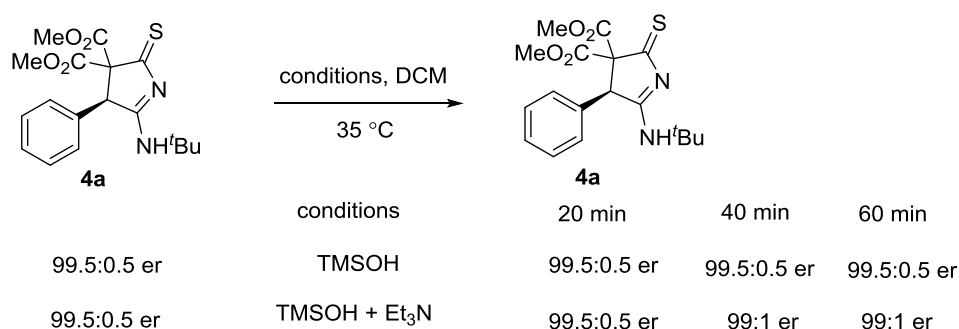
**Control experiment 4:** To an oven-dried tube were added Mg(OTf)<sub>2</sub> (10 mol%), L-RaPr<sub>2</sub> (10 mol%), **4a** (0.5 equiv), TEEDA and CH<sub>2</sub>Cl<sub>2</sub>, the mixture was stirred in CH<sub>2</sub>Cl<sub>2</sub> at 35 °C for 30 min. Then, TMSNCS (1.0 equiv) was added to the mixture at 35 °C and the stock solution was analysed by HPLC every 20 minutes.

It was found that the product underwent racemization quickly. When base additive (0.5 equiv) was added together, the racemization of the product was inhibited efficiently in the presence of chiral *N,N'*-dioxide/Mg<sup>II</sup> (10 mol%).



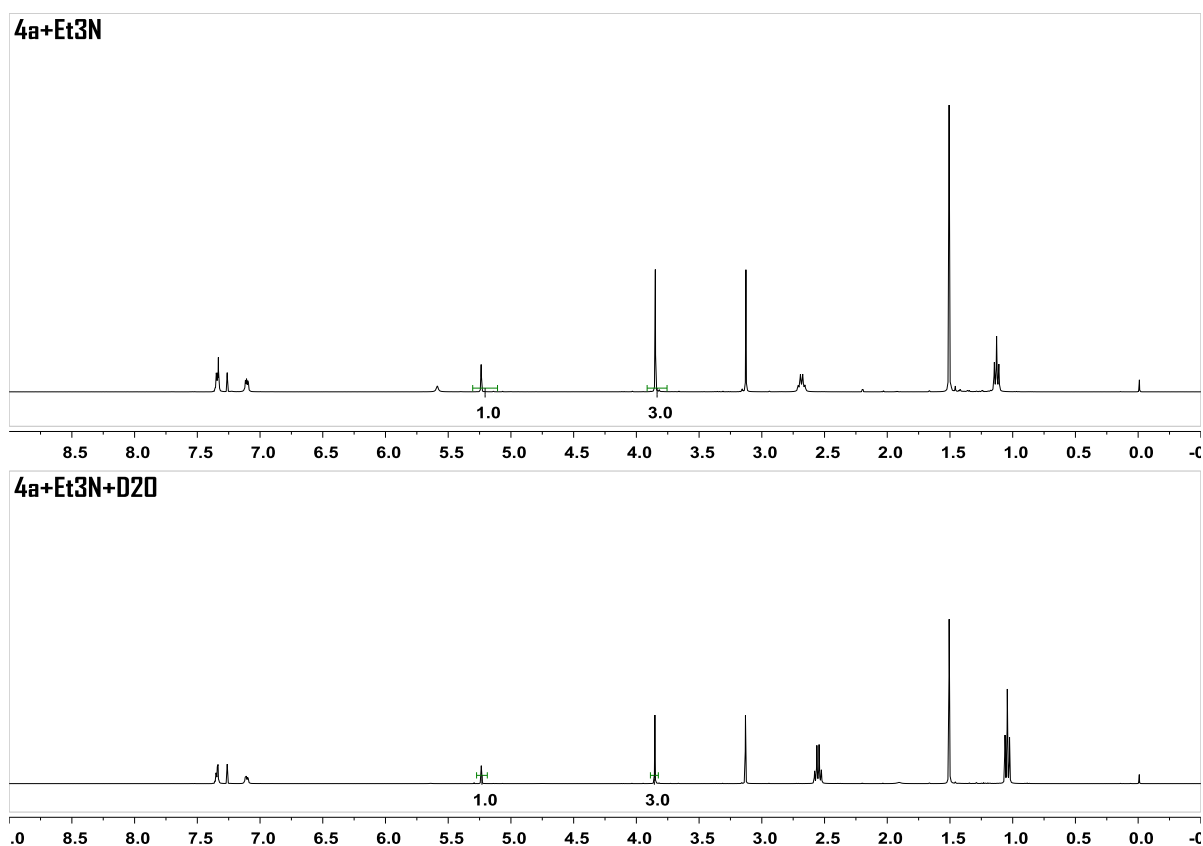
**Control experiment 5:** To an oven-dried tube were added Mg(OTf)<sub>2</sub> (0.01 mmol, 10 mol%), L-RaPr<sub>2</sub> (0.01 mmol, 10 mol%), *iPr*<sub>3</sub>NEt (0.05 mmol, 50 mol%), dimethyl 2-benzylidenemalonate **1a** (0.10 mmol), *tert*-butyl isocyanide **2a** (0.15 mmol), TMSNCS **3a** (0.15 mmol), and CH<sub>2</sub>ClCH<sub>2</sub>Cl (1.0 mL). The mixture was stirred in CH<sub>2</sub>ClCH<sub>2</sub>Cl at 35 °C.

These results indicated that the optical purity of the product was stable during the reaction process. The presence of base additive was crucile for high enantioselectivity. Otherwise, racemic product was afforded.

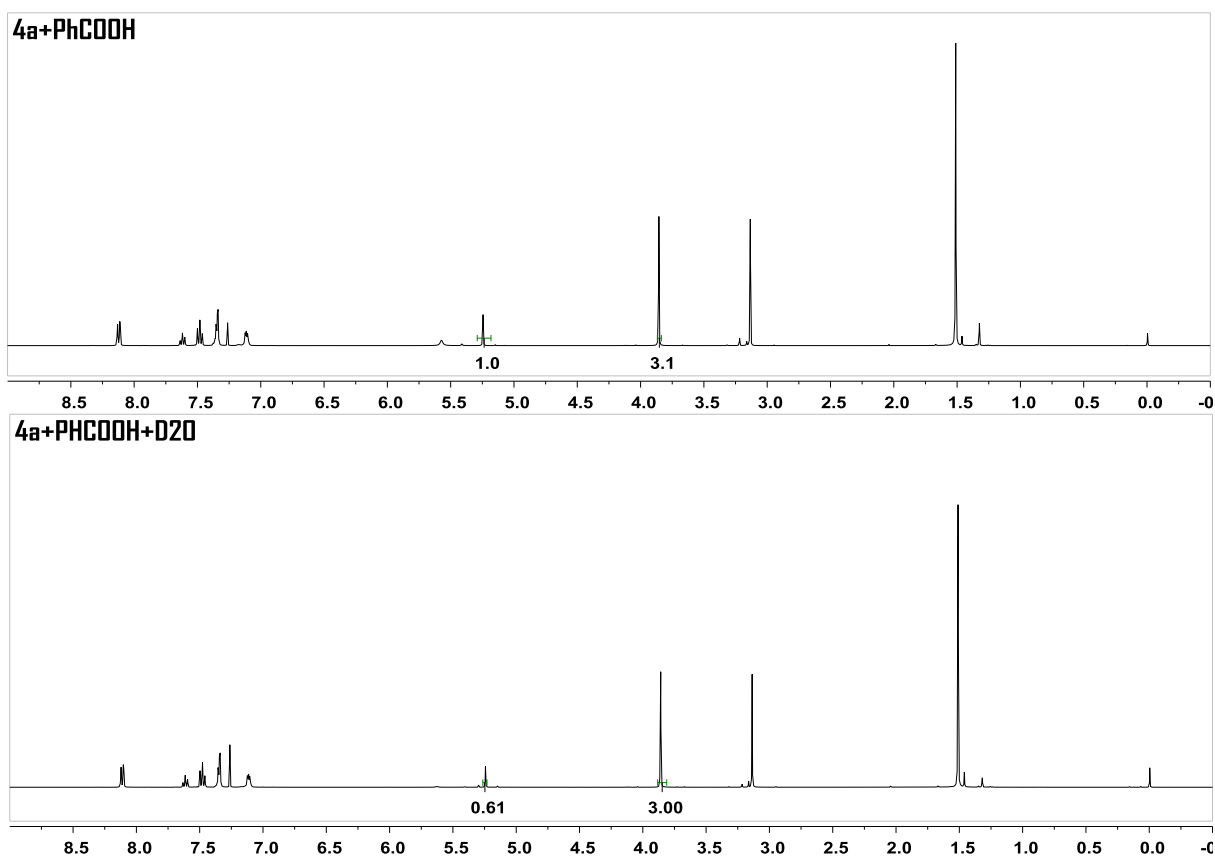


**Control experiment 6:** To an oven-dried tube were added **4a** and CH<sub>2</sub>Cl<sub>2</sub>, the mixture was stirred in CH<sub>2</sub>Cl<sub>2</sub> at 35 °C for 30 min. Then, TMSOH (1.0 equiv) was added to the mixture at 35 °C. The mixture was stirred in CH<sub>2</sub>Cl<sub>2</sub> at 35 °C and the stock solution was analysed by HPLC every 20 minutes.

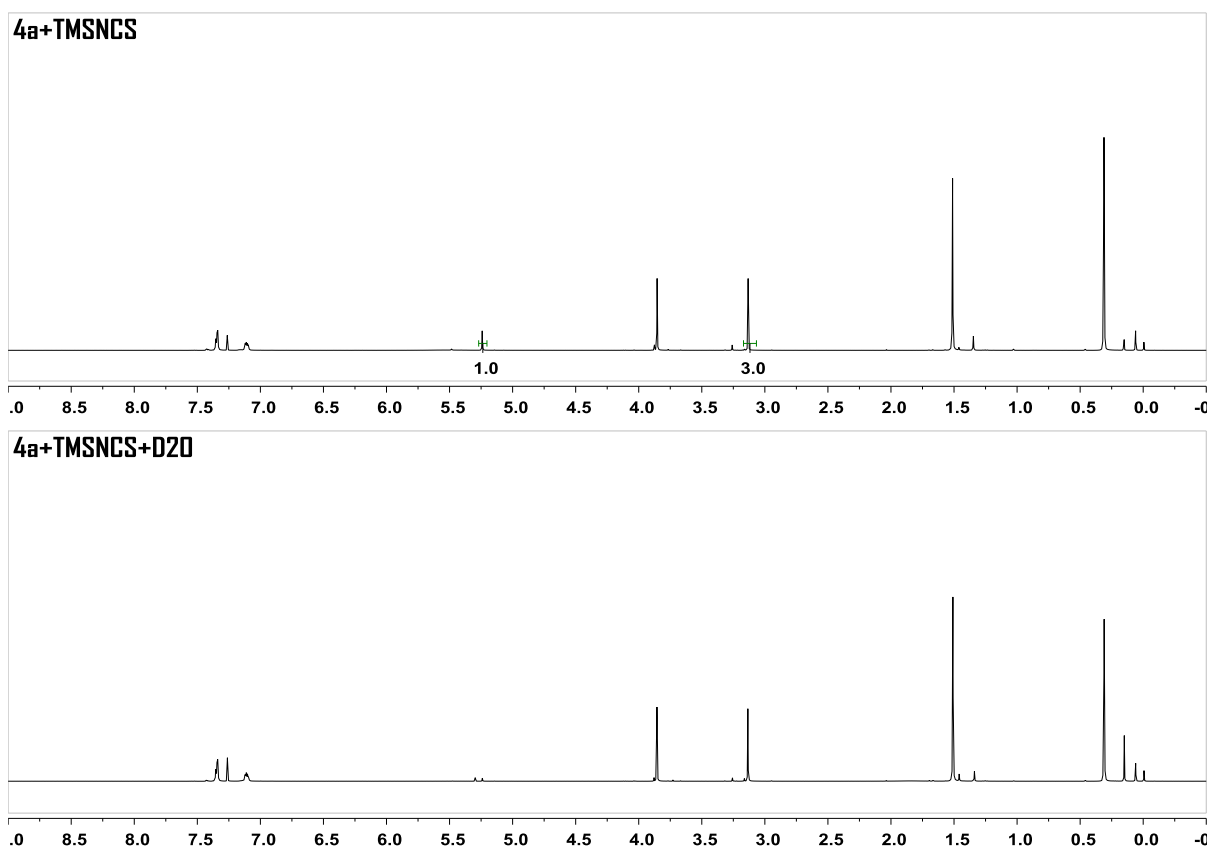
These results indicated that TMSOH has a negligible effect on the optical purity of the product. This control experiment implied that the HNCS generated in-situ was probably responsible for the racemization process.



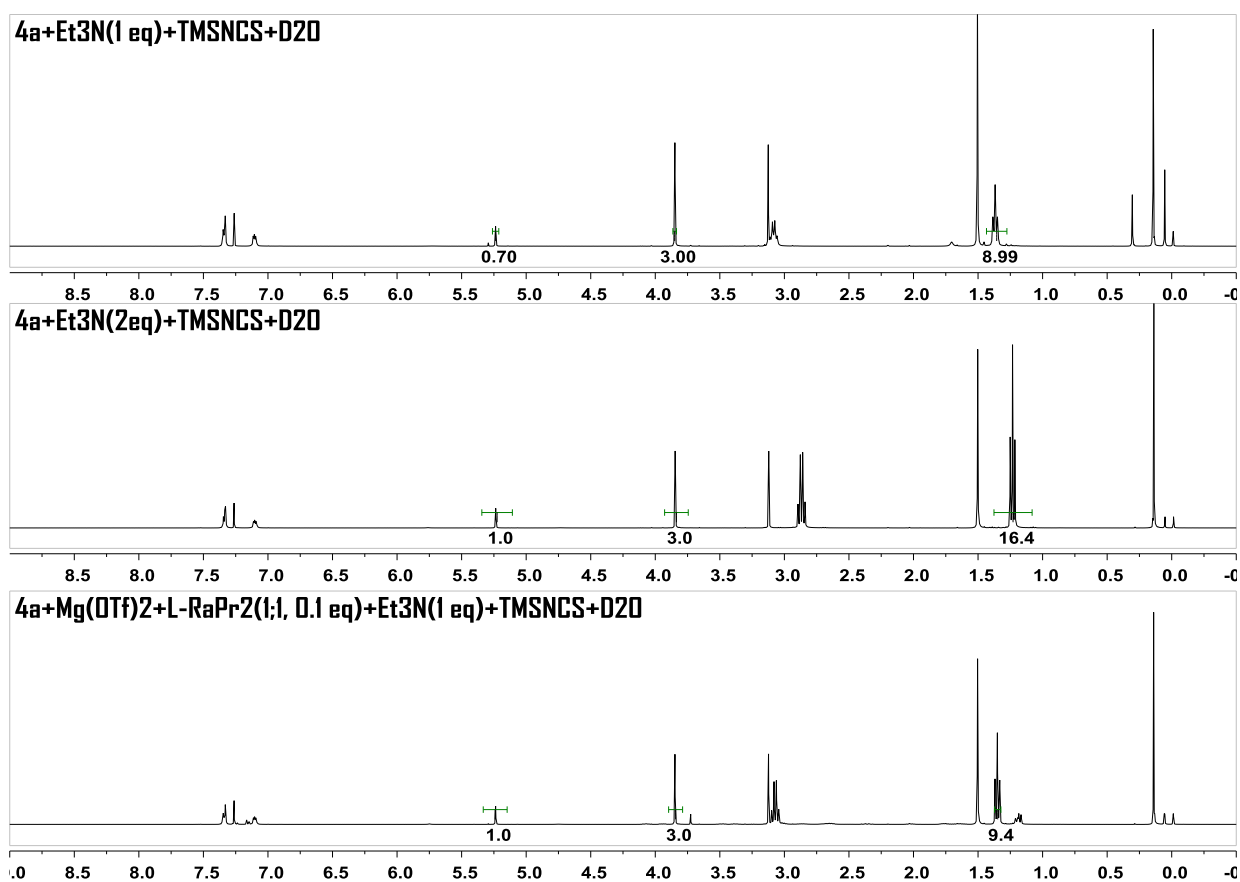
**Deuterization experiment 1:** To an oven-dried tube were added **4a**, Et<sub>3</sub>N(1 equiv) and CDCl<sub>3</sub>. The mixture was stirred in CDCl<sub>3</sub> at 35 °C for 30 min, then, D<sub>2</sub>O (5 μL) was added to the mixture at 35 °C. The mixture was stirred for 30 min. The <sup>1</sup>H NMR spectra indicated that Et<sub>3</sub>N is not able to accelerate H-D exchange at α-position of the product.



**Deuterization experiment 2:** To an oven-dried tube were added **4a**, PhCOOH (1 equiv) and CDCl<sub>3</sub>. The mixture was stirred in CDCl<sub>3</sub> at 35 °C for 30 min, then, D<sub>2</sub>O (5 μL) was added to the mixture at 35 °C. The mixture was stirred for 30 min. The <sup>1</sup>H NMR spectra indicated that PhCOOH is able to promote H-D exchange (ca. 40% after 0.5 h) at α-position of the product.

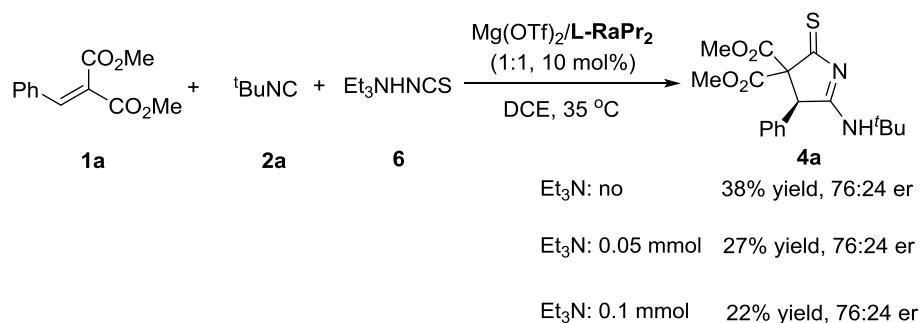


**Deuterization experiment 2:** To an oven-dried tube were added **4a**, TMSNCS (1 equiv) and CDCl<sub>3</sub>. The mixture was stirred in CDCl<sub>3</sub> at 35 °C for 30 min, then, D<sub>2</sub>O (5 μL) was added to the mixture at 35 °C. The mixture was stirred for 30 min. The <sup>1</sup>H NMR spectra indicated that TMSNCS enables to efficiently promote H-D exchange (ca. 100% after 0.5 h) at α-position of the product.

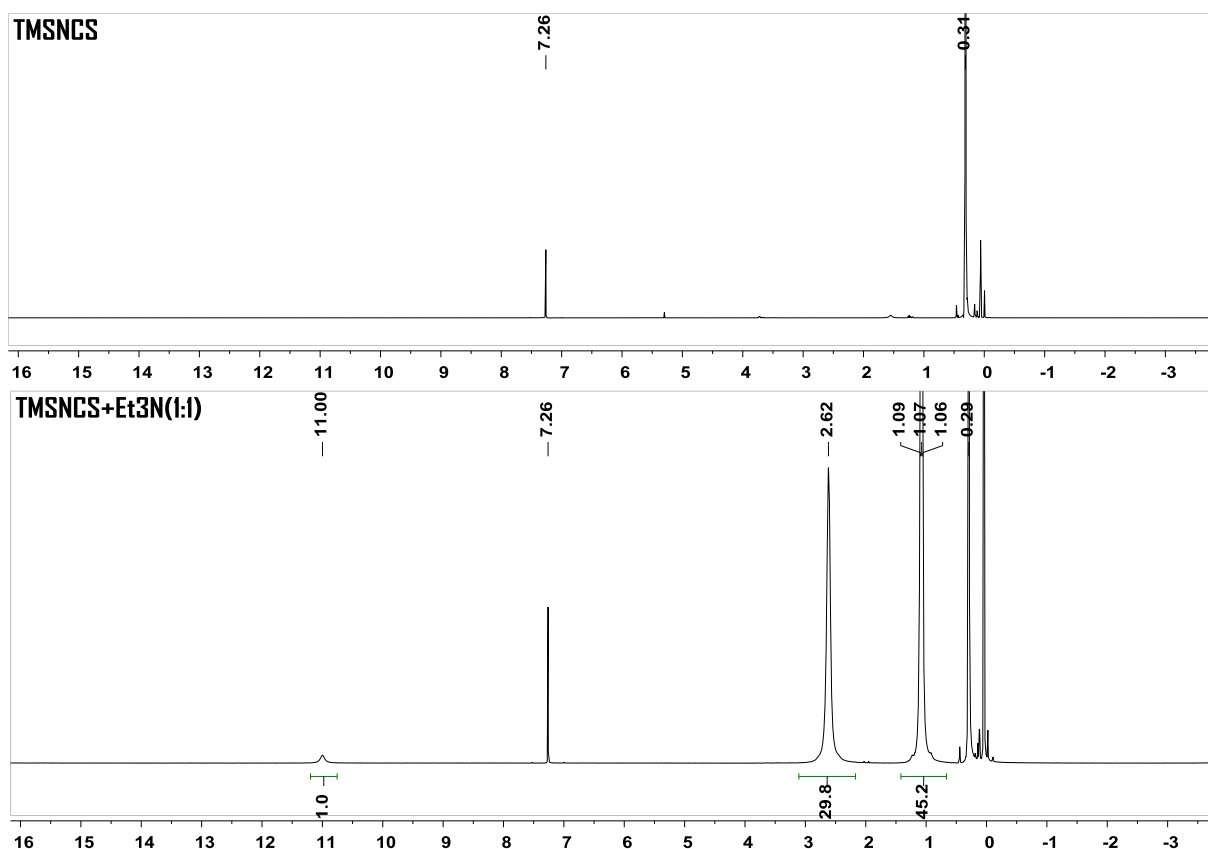


**Deuterization experiment 3:** To an oven-dried tube were added Mg(OTf)<sub>2</sub> (10 mol%), L-RaPr<sub>2</sub> (10 mol%), **4a**, Et<sub>3</sub>N (1 equiv) and CDCl<sub>3</sub>. The mixture was stirred in CDCl<sub>3</sub> at 35 °C for 30 min, then, TMSNCS and D<sub>2</sub>O (5 μL) was added to the mixture at 35 °C. Collectively, the <sup>1</sup>H

NMR spectra indicated that Et<sub>3</sub>N can inhibit the TMSNCS-mediated racemization process. Moreover, chiral *N,N'*-dioxide/Mg<sup>II</sup> (10 mol%) could enhance this performance of Et<sub>3</sub>N.

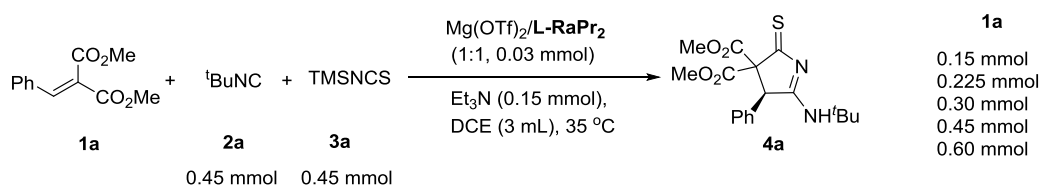


**Control experiment 7:** To an oven-dried tube were added Mg(OTf)<sub>2</sub> (0.01 mmol, 10 mol%), *L*-RaPr<sub>2</sub> (0.01 mmol, 10 mol%), dimethyl 2-benzylidenemalonate **1a** (0.10 mmol), *tert*-butyl isocyanide **2a** (0.15 mmol), Et<sub>3</sub>N•HNCs (0.10 mmol), Et<sub>3</sub>N and CH<sub>2</sub>ClCH<sub>2</sub>Cl (1.0 mL). The mixture was stirred in CH<sub>2</sub>ClCH<sub>2</sub>Cl at 35 °C for 24h.

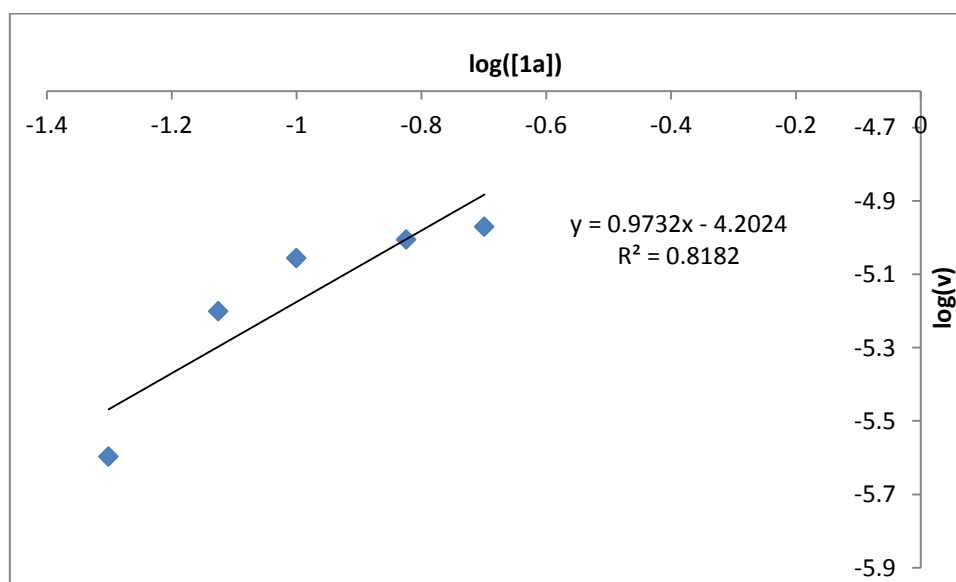
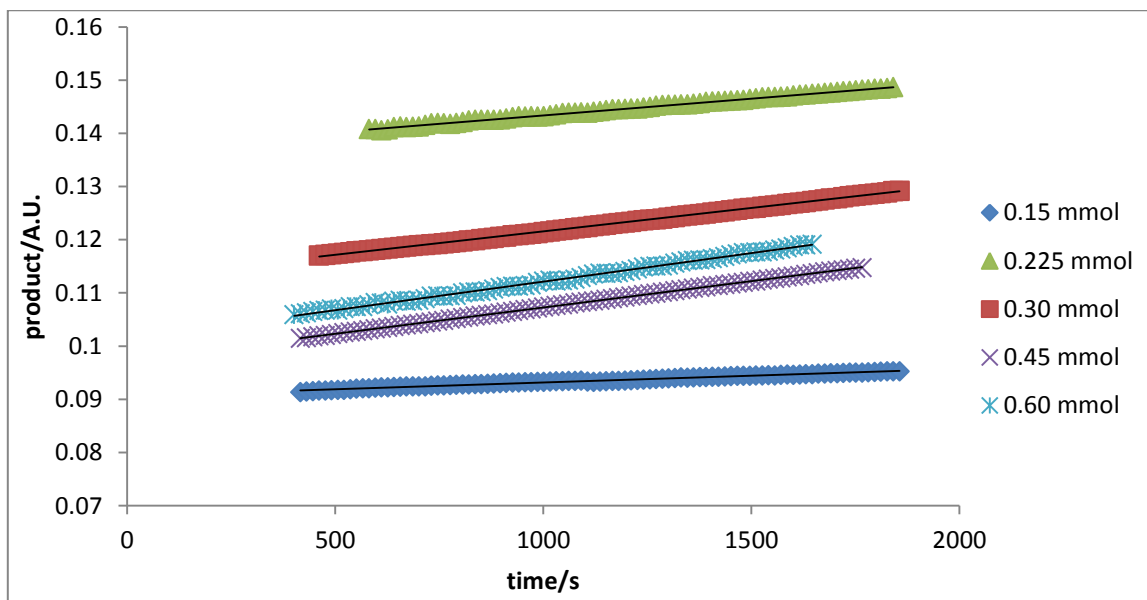


**Control experiment 8: Kinetic studies**

Kinetic analyses were performed using in situ attenuated total reflectance Fouriertransform infrared (ATR FTIR) spectroscopy to track the formation of product **4a** under synthetically relevant conditions. A Mettler Toledo SW License iC IR 701L instrument was treated as main experiment equipment. All of the kinetic experiments on each plot were performed using a single batch of reagents. Peak at 1335 cm<sup>-1</sup> was identified as the characteristic absorption of product **4a**.

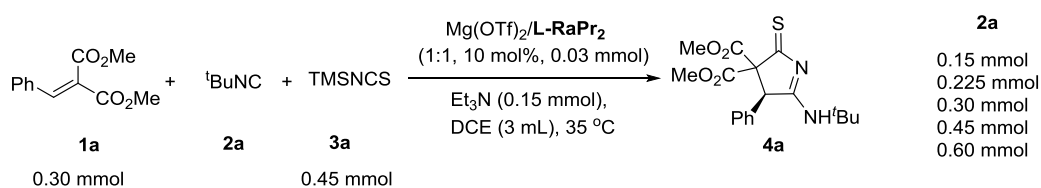


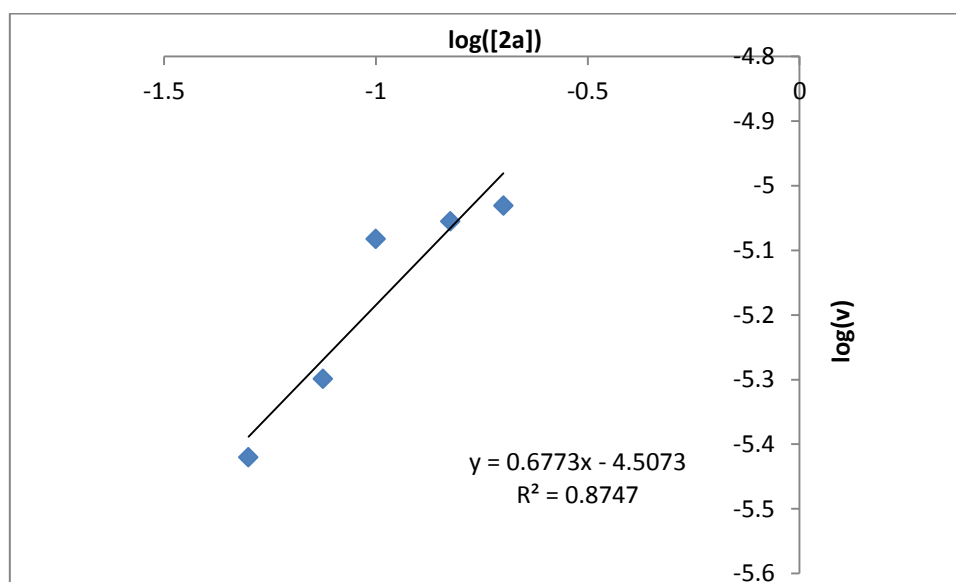
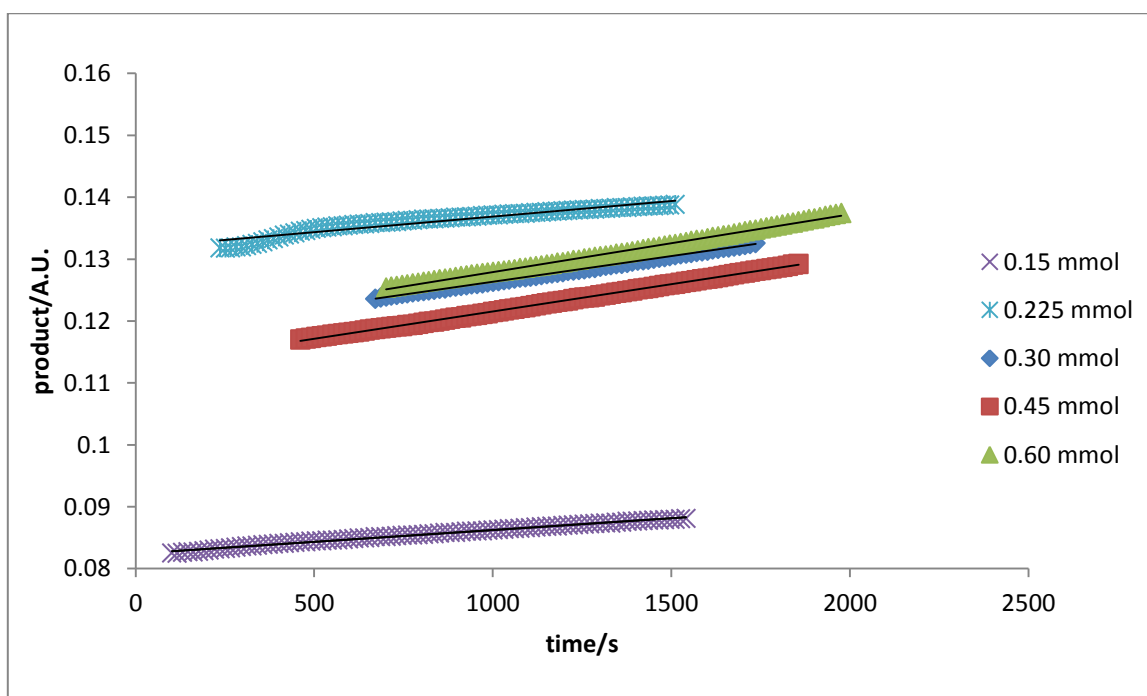




| [1a]/(M) | reaction rate/(A.U./s) | Log([1a])    | Log(v)       |
|----------|------------------------|--------------|--------------|
| 0.05     | 0.0000025315           | -1.301029996 | -5.596622068 |
| 0.075    | 0.0000062953           | -1.124938737 | -5.200983569 |
| 0.10     | 0.0000088052           | -1           | -5.004671991 |
| 0.15     | 0.0000098930           | -0.823908741 | -5.004671991 |
| 0.20     | 0.0000107028           | -0.698970004 | -4.97050259  |

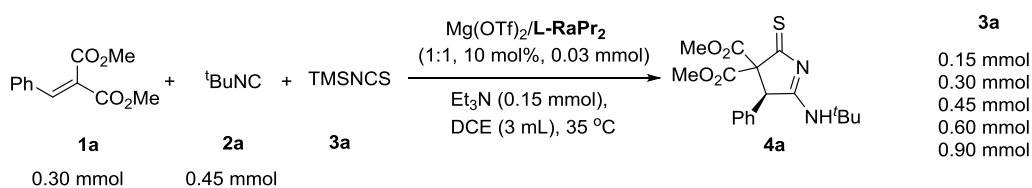
According to the formula  $v$  (Reaction rate) =  $k[1a]^{\alpha} [2a]^{\beta} [3a]^{\gamma} [L-RaPr_2/Mg(OTf)_2]^{\lambda} [Et_3N]^{\delta}$ , we can get the inference that  $\log(v) = -\alpha \log([1a]) + A$ . Calculating the data by means of excel,  $\alpha = 0.973$  was obtained, which indicates the reaction rate obeys a first order dependence on [1a].

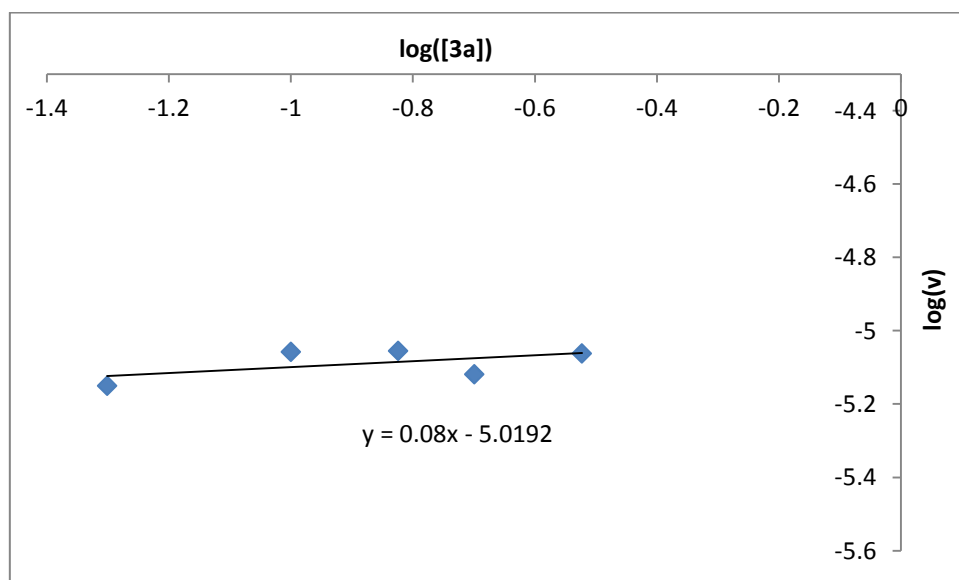
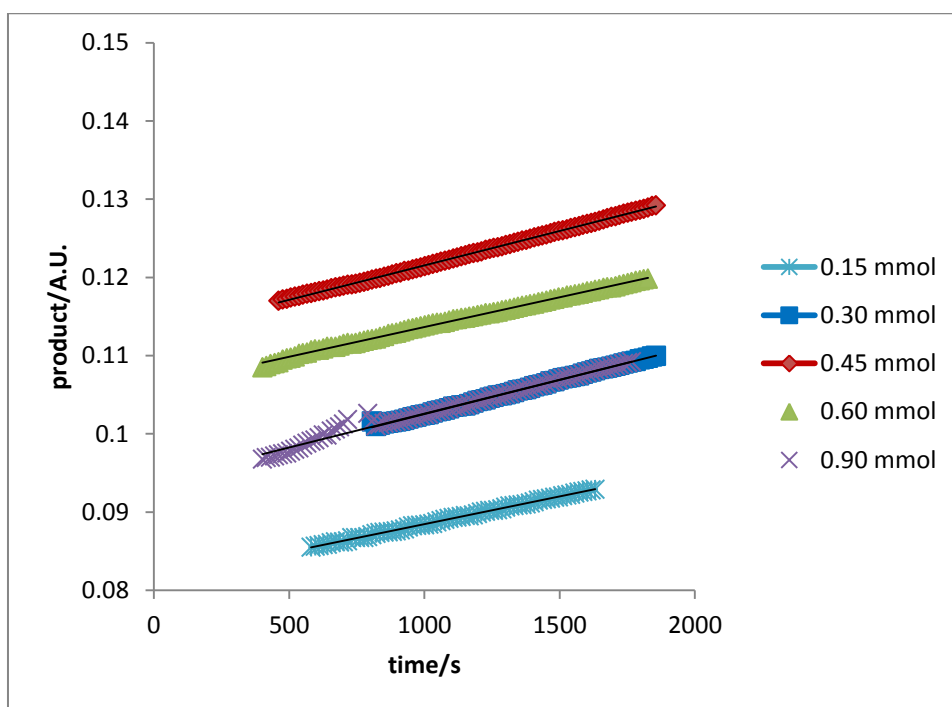




| [2a]/(M) | reaction rate/(A.U./s) | Log([2a])    | Log(v)       |
|----------|------------------------|--------------|--------------|
| 0.05     | 0.0000037986           | -1.301029996 | -5.420376436 |
| 0.075    | 0.0000050238           | -1.124938737 | -5.298967658 |
| 0.10     | 0.0000083253           | -1           | -5.082541756 |
| 0.15     | 0.0000088052           | -0.823908741 | -5.055260775 |
| 0.20     | 0.0000093144           | -0.698970004 | -5.030845116 |

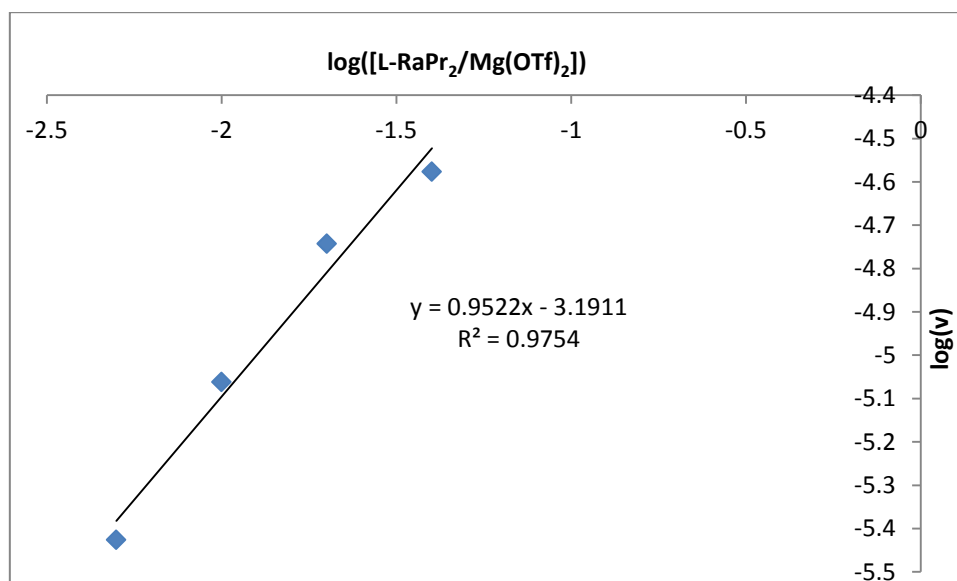
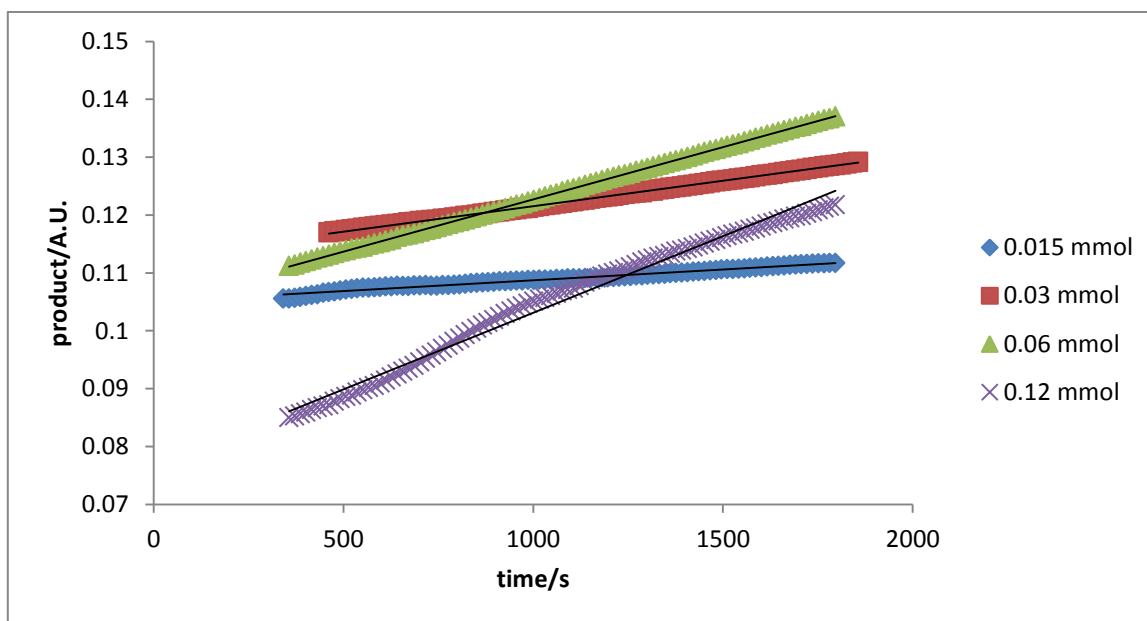
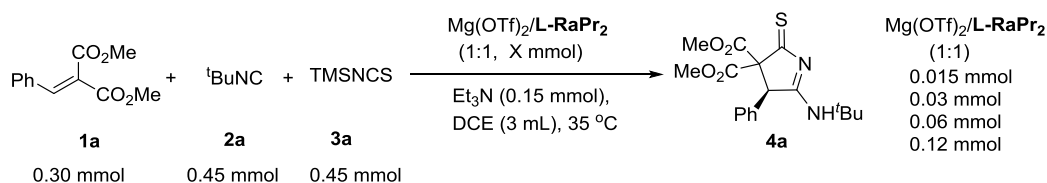
According to the formula  $v$  (Reaction rate) =  $k[1a]^{\alpha} [2a]^{\beta} [3a]^{\gamma} [L-RaPr_2/Mg(OTf)_2]^{\lambda} [Et_3N]^{\delta}$ , we can get the inference that  $\log(v) = \beta \log([2a]) + B$ . Calculating the data by means of excel,  $\beta = 0.677$  was obtained, which indicates the reaction rate obeys a fractional partial order dependence on [2a].





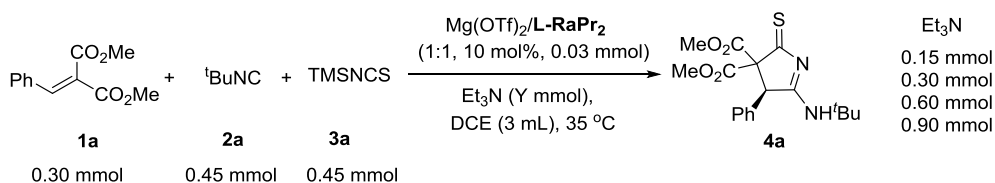
| [3a]/(M) | reaction rate/(A.U./s) | Log([3a])    | Log(v)       |
|----------|------------------------|--------------|--------------|
| 0.05     | 0.0000070921           | -1.301029996 | -5.150488454 |
| 0.10     | 0.0000087572           | -1           | -5.057634732 |
| 0.15     | 0.0000088052           | -0.823908741 | -5.055260775 |
| 0.20     | 0.0000076095           | -0.698970004 | -5.118643879 |
| 0.30     | 0.0000086749           | -0.522878745 | -5.061735523 |

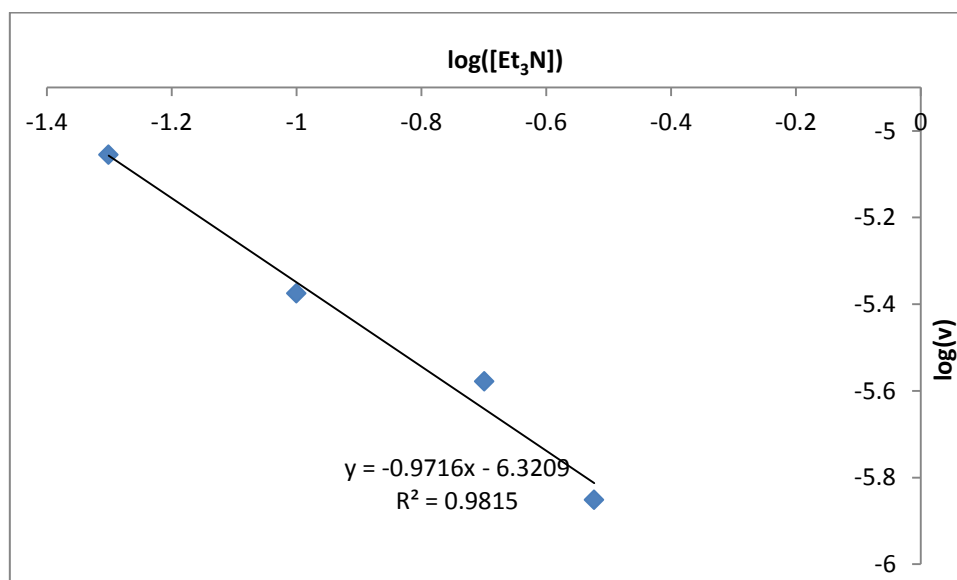
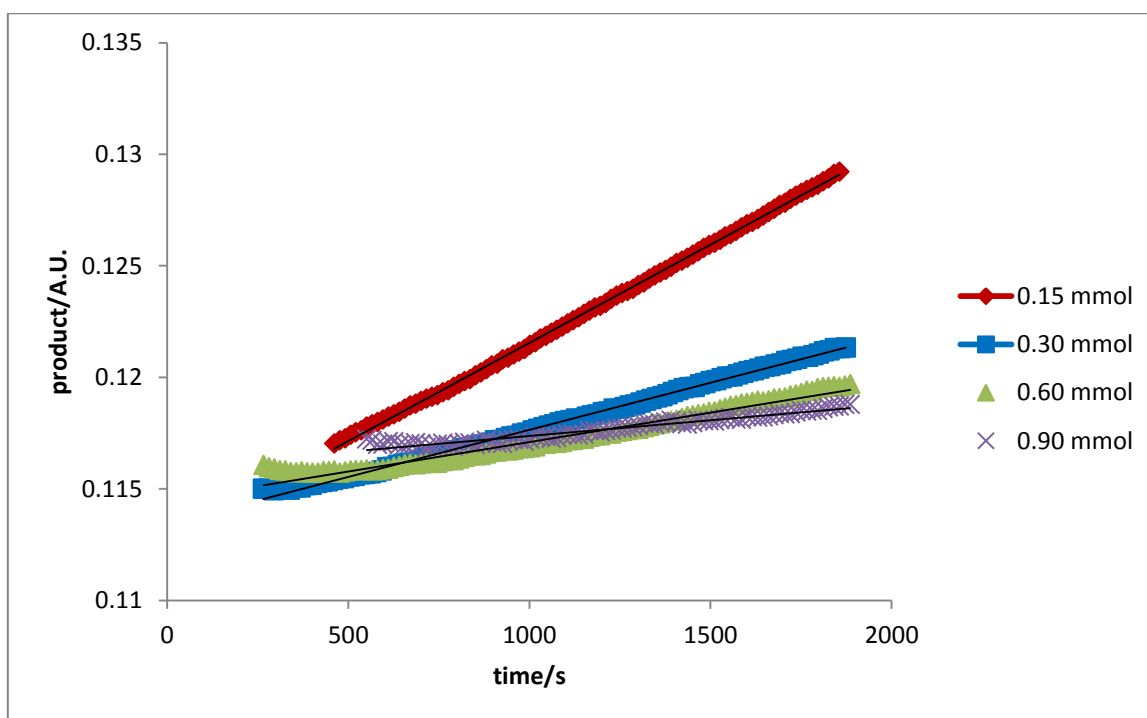
According to the formula  $v$  (Reaction rate) =  $k[1a]^{\alpha} [2a]^{\beta} [3a]^{\gamma} [L-RaPr_2/Mg(OTf)_2]^{\lambda} [Et_3N]^{\delta}$ , we can get the inference that  $\log(v) = \gamma \log([3a]) + C$ . Calculating the data by means of excel,  $\gamma = 0.08$  was obtained, which indicates the reaction rate obeys a zero order dependence on [3a].



| [catalyst]/(M) | reaction rate/(A.U./s) | Log([catalyst]) | Log(v)       |
|----------------|------------------------|-----------------|--------------|
| 0.005          | 0.0000037490           | -2.301029996    | -5.42608456  |
| 0.01           | 0.0000088052           | -2              | -5.062326672 |
| 0.02           | 0.0000180636           | -1.698970004    | -4.743195692 |
| 0.04           | 0.0000264860           | -1.397940009    | -4.576983625 |

According to the formula  $v$  (Reaction rate) =  $k[1a]^\alpha [2a]^\beta [3a]^\gamma [\text{L-RaPr}_2/\text{Mg}(\text{OTf})_2]^\lambda [\text{Et}_3\text{N}]^\delta$ , we can get the inference that  $\log(v) = \lambda \cdot \log([\text{L-RaPr}_2/\text{Mg}(\text{OTf})_2]) + D$ . Calculating the data by means of excel,  $\lambda = 0.952$  was obtained, which indicates the reaction rate obeys a first order dependence on  $[\text{L-RaPr}_2/\text{Mg}(\text{OTf})_2]$ .



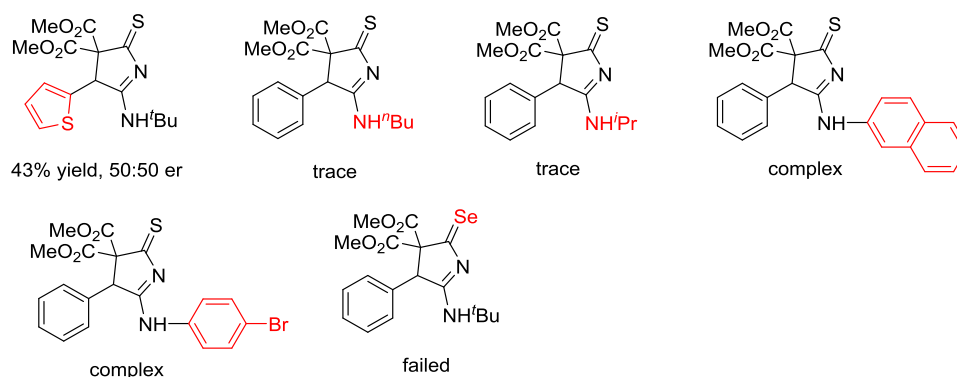


| [Et <sub>3</sub> N]/(M) | reaction rate/(A.U./s) | Log([Et <sub>3</sub> N]) | Log(v)       |
|-------------------------|------------------------|--------------------------|--------------|
| 0.05                    | 0.0000088052           | -1.301029996             | -5.055260775 |
| 0.10                    | 0.0000042119           | -1                       | -5.375521948 |
| 0.20                    | 0.0000026410           | -0.698970004             | -5.578231599 |
| 0.30                    | 0.0000014060           | -0.522878745             | -5.852014679 |

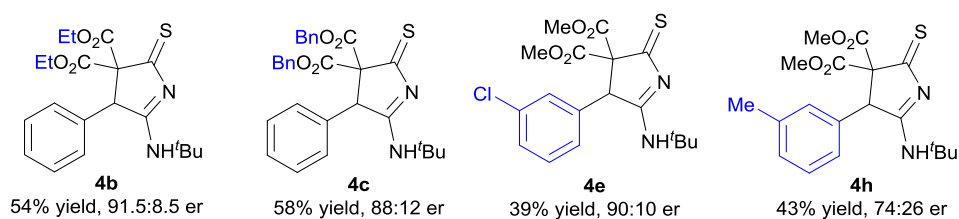
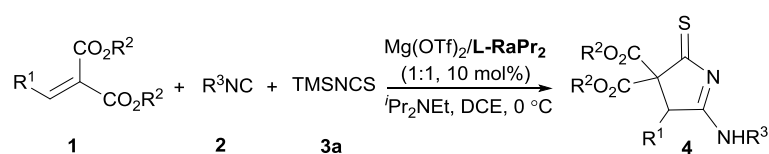
According to the formula  $v$  (Reaction rate) =  $k[1a]^{\alpha} [2a]^{\beta} [3a]^{\gamma} [L-RaPr_2/Mg(OTf)_2]^{\lambda} [Et_3N]^{\delta}$ , we can get the inference that  $\log(v) = \delta \log([Et_3N]) + E$ . Calculating the data by means of excel,  $\delta = -0.972$  was obtained, which indicates the reaction rate obeys a negative first-order dependence on  $[Et_3N]$ .

To get more insight into the reaction mechanism, kinetic studies via operando IR experiments were performed (for more details, see ESI). Peak at  $1335\text{ cm}^{-1}$  was identified as the characteristic absorption of the product **4a**. Kinetic studies exhibited that the initial rate of the reaction was approximate first-order kinetic dependence on the catalyst and alkyldiene malonate **1a** and fractional order kinetic dependence on isocyanide **2a**. In contrast, zero-order was observed for TMSNCS. These results imply that catalyst, the initial addition reaction between **1a** and **2a** in the presence of chiral catalyst were involved in the rate-determining step, TMSNCS was not the real reactive species and the addition of NCS group occurred after the rate-determining step. Interestingly, the reaction rate roughly showed negative first-order on base additive, and the presence of base probably generated ammonium thiocyanate to undergo the sequential addition reaction, balancing the initial addition of isocyanide and avoiding the generation of competitive byproducts and racemization process.

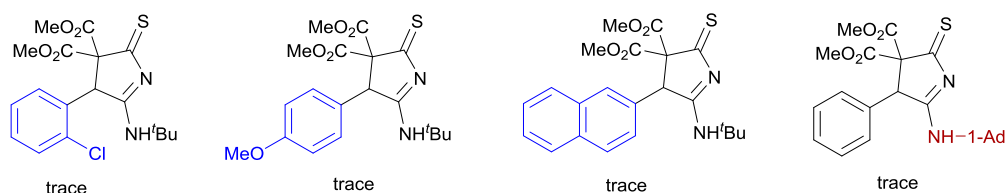
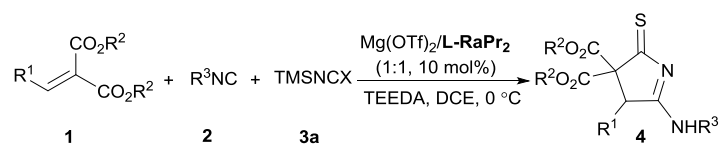
#### 4. Unsuccessful substrate scope



#### 5. Substrate scope with $i\text{Pr}_2\text{NEt}$



#### 6. Substrate scope with TEEDA at 0 °C



#### 7. Optimization of reaction conditions

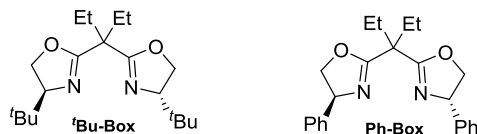
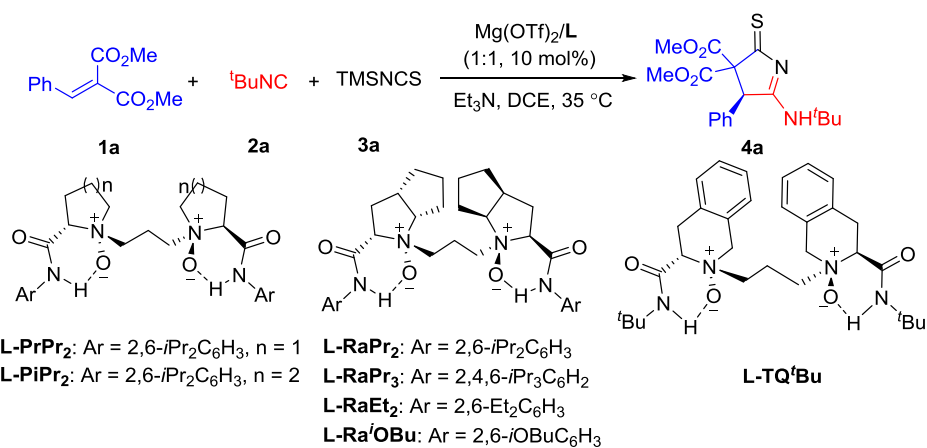
Table S1. Screening of the metal salts <sup>[a]</sup>



| entry | metal salts                        | yield (%) | er        |
|-------|------------------------------------|-----------|-----------|
| 1     | Mg(OTf) <sub>2</sub>               | 86        | 85.5:14.5 |
| 2     | Mg(ClO <sub>4</sub> ) <sub>2</sub> | 89        | 85:15     |
| 3     | Mg(NTf) <sub>2</sub>               | 91        | 83:17     |
| 4     | Zn(OTf) <sub>2</sub>               | messy     | -         |
| 5     | Cu(OTf) <sub>2</sub>               | messy     | -         |
| 6     | Ni(OTf) <sub>2</sub>               | messy     | -         |
| 7     | Sc(OTf) <sub>3</sub>               | messy     | -         |
| 8     | Yb(OTf) <sub>3</sub>               | messy     | -         |

[a] All reactions were carried out **1a** (0.10 mmol), **2a** (0.15 mmol), **3a** (0.15 mmol), M/L-RaPr<sub>2</sub> (1.0:1.0, 10.0 mol%) and Et<sub>3</sub>N (0.5 mmol) in CH<sub>2</sub>ClCH<sub>2</sub>Cl (1.0 mL) at 35 °C for 24 h. Isolated yield and er was determined by CSP-HPLC analysis.

**Table S2.** Screening of the ligands<sup>[a]</sup>

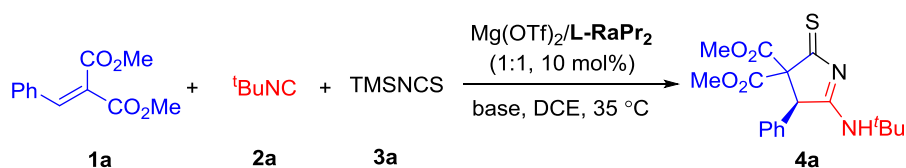


| entry | ligands                   | yield (%) | er        |
|-------|---------------------------|-----------|-----------|
| 1     | <b>L-PrPr<sub>2</sub></b> | 64        | 81.5:18.5 |
| 2     | <b>L-PiPr<sub>2</sub></b> | 96        | 75:25     |
| 3     | <b>L-RaPr<sub>2</sub></b> | 86        | 85.5:14.5 |
| 4     | <b>L-RaPr<sub>3</sub></b> | 90        | 77:23     |
| 5     | <b>L-RaEt<sub>2</sub></b> | 87        | 80:20     |

|    |  |    |       |
|----|--|----|-------|
| 6  | <b>L-Ra<sup>i</sup>OBu<sub>2</sub></b> | 35 | 65:35 |
| 7  | <b>L-PrCPh<sub>3</sub></b>             | 33 | 63:37 |
| 8  | <b>L-TQ<sup>t</sup>Bu</b>              | 81 | 47:53 |
| 9  | <b><sup>t</sup>Bu-Box</b>              | 93 | 50:50 |
| 10 | <b>Ph-Box</b>                          | 94 | 50:50 |

[a] All reactions were carried out **1a** (0.10 mmol), **2a** (0.15 mmol), **3a** (0.15 mmol), Mg(OTf)<sub>2</sub>/L (1.0:1.0, 10.0 mol%) and Et<sub>3</sub>N in CH<sub>2</sub>ClCH<sub>2</sub>Cl (1.0 mL) at 35 °C for 24 h. Isolated yield and er was determined by CSP-HPLC analysis.

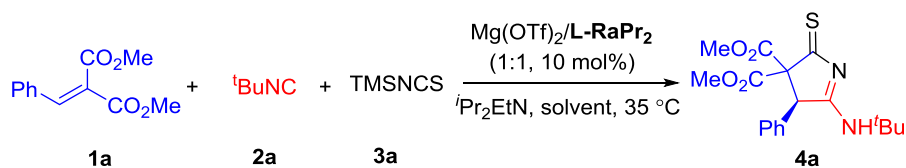
**Table S3.** Preliminary screening of the base <sup>[a]</sup>



| entry | base                             | yield (%) | er        |
|-------|----------------------------------|-----------|-----------|
| 1     | Li <sub>2</sub> CO <sub>3</sub>  | 87        | 50:50     |
| 2     | Na <sub>2</sub> CO <sub>3</sub>  | 70        | 76:24     |
| 3     | k <sub>2</sub> CO <sub>3</sub>   | 91        | 87:13     |
| 4     | Cs <sub>2</sub> CO <sub>3</sub>  | 58        | 66:34     |
| 5     | Et <sub>2</sub> NH               | 50        | 82:18     |
| 6     | Et <sub>3</sub> N                | 86        | 85.5:14.5 |
| 7     | <sup>i</sup> Pr <sub>2</sub> EtN | 88        | 92:8      |
| 8     | DMAP                             | 82        | 53.5:46.5 |
| 9     | DABCO                            | 95        | 53:47     |

[a] All reactions were carried out **1a** (0.10 mmol), **2a** (0.15 mmol), **3a** (0.15 mmol), Mg(OTf)<sub>2</sub>/L-RaPr<sub>2</sub> (1:1, 10.0 mol%) and base (0.05 mmol) in CH<sub>2</sub>ClCH<sub>2</sub>Cl (1.0 mL) at 35 °C for 24 h. Isolated yield and er was determined by CSP-HPLC analysis.

**Table S4.** Screening of the solvents <sup>[a]</sup>



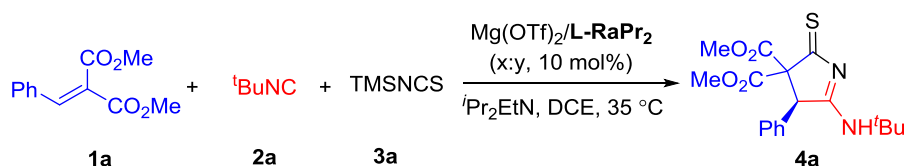
| Entry | solvent                              | yield (%) | er    |
|-------|--------------------------------------|-----------|-------|
| 1     | CH <sub>2</sub> ClCH <sub>2</sub> Cl | 88        | 92:8  |
| 2     | CH <sub>2</sub> Cl <sub>2</sub>      | 56        | 82:18 |



|   |                                     |       |       |
|---|-------------------------------------|-------|-------|
| 3 | CHCl <sub>3</sub>                   | 88    | 88:12 |
| 4 | CHCl <sub>2</sub> CHCl <sub>2</sub> | 72    | 85:15 |
| 5 | Toluene                             | trace | -     |
| 6 | Et <sub>2</sub> O                   | 69    | 85:15 |
| 7 | THF                                 | nr    | -     |
| 9 | PhCl                                | 50    | 79:21 |

[a] All reactions were carried out **1a** (0.10 mmol), **2a** (0.15 mmol), **3a** (0.15 mmol), Mg(OTf)<sub>2</sub>/L-RaPr<sub>2</sub> (1.0:1.0, 10.0 mol%) and <sup>i</sup>Pr<sub>2</sub>EtN (0.05 mmol) in solvent (1.0 mL) at 35 °C for 24 h. Isolated yield and er was determined by CSP-HPLC analysis

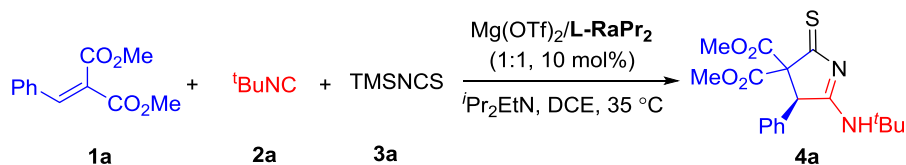
**Table S5.** Screening of the ratio of metal salt and ligand <sup>[a]</sup>



| entry            | M/L (x:y) | yield (%) | er       |
|------------------|-----------|-----------|----------|
| 1                | 1.5:1.0   | 80        | 90:10    |
| 2                | 1.2:1.0   | 93        | 91:9     |
| 3                | 1.0:1.0   | 88        | 92:8     |
| 4 <sup>[b]</sup> | 1.0:1.5   | 89        | 91.5:8.5 |
| 5 <sup>[c]</sup> | 1.0:2.0   | 95        | 91.5:8.5 |

[a] All reactions were carried out **1a** (0.10 mmol), **2a** (0.15 mmol), **3a** (0.15 mmol), Mg(OTf)<sub>2</sub>/L-RaPr<sub>2</sub> (x:y, 10.0 mol%) and <sup>i</sup>Pr<sub>2</sub>EtN (0.05 mmol) in CH<sub>2</sub>ClCH<sub>2</sub>Cl (1.0 mL) at 35 °C for 24 h. Isolated yield and er was determined by CSP-HPLC analysis

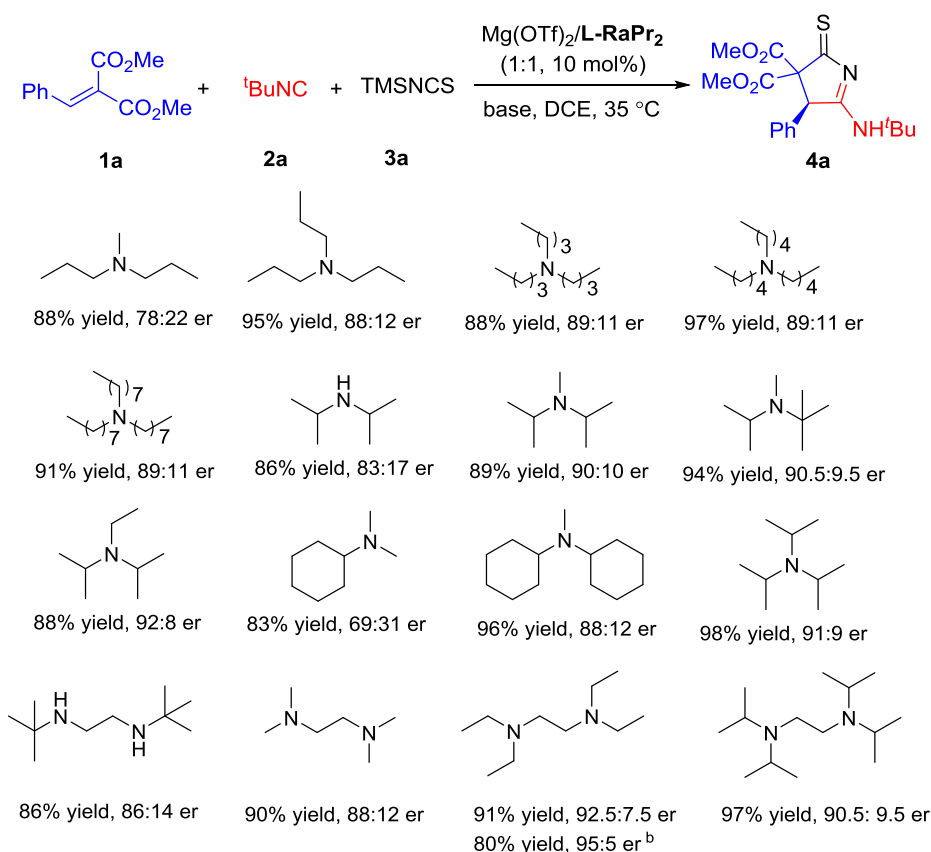
**Table S6.** Screening of the amount of base <sup>[a]</sup>



| entry | the amount of <sup>i</sup> Pr <sub>2</sub> EtN (x) | yield (%) | er   |
|-------|--|-----------|------|
| 1     | 0.25 equiv   | 83        | 91:9 |
| 2     | 0.5 equiv  | 88        | 92:8 |
| 3     | 1.0 equiv  | 72        | 91:9 |
| 4     | 1.5 equiv  | 51        | 91:9 |

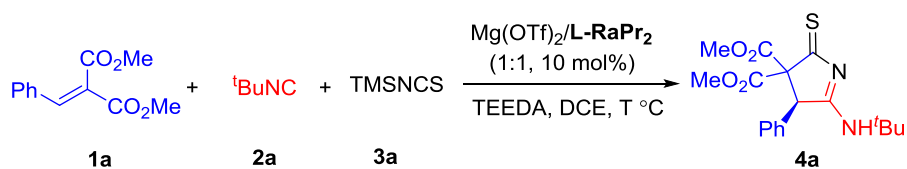
[a] All reactions were carried out **1a** (0.10 mmol), **2a** (0.15 mmol), **3a** (0.15 mmol), Mg(OTf)<sub>2</sub>/L-RaPr<sub>2</sub> (1.0:1.0, 10.0 mol%) and <sup>i</sup>Pr<sub>2</sub>EtN (x mmol) in DCE (1.0 mL) at 35 °C for 24 h. Isolated yield and er was determined by CSP-HPLC analysis.

**Table S7.** Screening of the base <sup>[a]</sup>



[a] All reactions were carried out **1a** (0.10 mmol), **2a** (0.15 mmol), **3a** (0.15 mmol), Mg(OTf)<sub>2</sub>/L-RaPr<sub>2</sub> (1.0:1.0, 10.0 mol%) and base (0.05 mmol; diamines was 0.025 mmol) in CH<sub>2</sub>ClCH<sub>2</sub>Cl (1.0 mL) at 35 °C for 24 h. Isolated yield and er was determined by CSP-HPLC analysis. [b] at 0 °C for 96 h.

**Table S8.** Screening of the temperature <sup>[a]</sup>

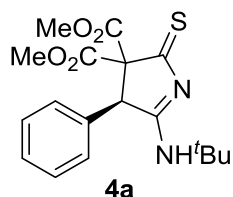


| entry            | temperature (°C) | yield (%) | er       |
|------------------|------------------|-----------|----------|
| 1 <sup>[b]</sup> | 35               | 91        | 92.5:7.5 |
| 2                | 0                | 80        | 95:5     |
| 3                | -5               | 70        | 95:5     |
| 4                | -20              | trace     | -        |
| 5 <sup>[c]</sup> | 0                | 48        | 94.5:5.5 |
| 6 <sup>[d]</sup> | 0                | 51        | 94:6     |

[a] All reactions were carried out **1a** (0.10 mmol), **2a** (0.15 mmol), **3a** (0.15 mmol), Mg(OTf)<sub>2</sub>/L-RaPr<sub>2</sub> (1.0:1.0, 10.0 mol%) and TEEDA (0.025 mmol) in CH<sub>2</sub>ClCH<sub>2</sub>Cl (1.0 mL) at T °C for 96 h. Isolated yield and er was determined by CSP-HPLC analysis. [b] 35°C for 24 h. [c] 4 Å MS was added as additive. [d] Conducting the reaction in glove box.

## 8. The analytical and spectral characterization data of products

### Dimethyl (S)-5-(*tert*-butylamino)-4-phenyl-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (**4a**)



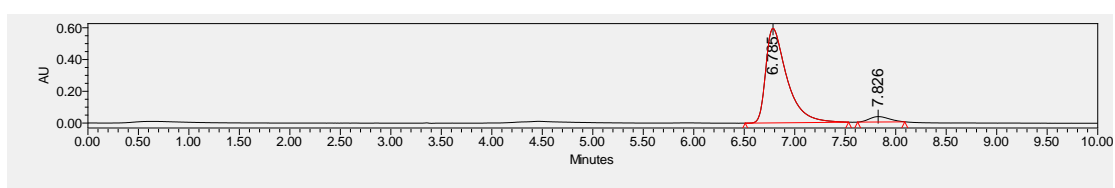
Yellow solid; m.p. 175–179 °C; 29.1 mg, 80% yield, 95:5 er;  $[\alpha]_D^{22} = -15.32$  ( $c = 0.44$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IF, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 6.76 min,  $t_R$  (minor) = 7.83 min.

IR (neat): 3309, 2952, 2361, 1728, 1599, 1531, 1455, 1434, 1322, 1288, 1207, 1124, 1099, 1049, 762 and 701  $\text{cm}^{-1}$ .

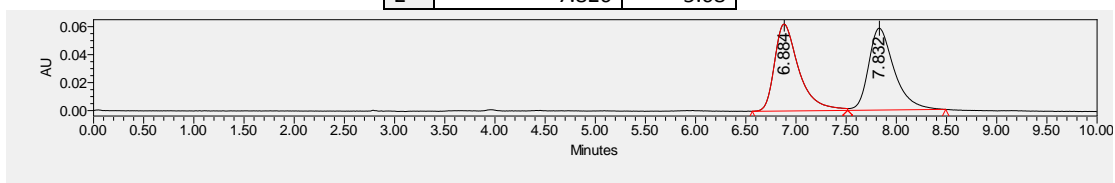
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.36 - 7.31$  (m, 3H), 7.12 – 7.06 (m, 2H), 5.68 (s, 1H), 5.23 (s, 1H), 3.83 (s, 3H), 3.11 (s, 3H), 1.50 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 217.7, 181.7, 166.9, 165.9, 133.7, 129.6, 129.1, 129.1, 79.4, 59.1, 56.0, 54.0, 52.3, 28.9$ .

HRMS (ESI-FT) calcd for  $\text{C}_{18}\text{H}_{23}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 363.1373, Found 363.1365.

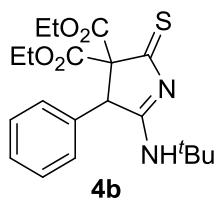


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 6.785          | 94.92  |
| 2 | 7.826          | 5.08   |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 6.884          | 50.28  |
| 2 | 7.832          | 49.72  |

### Diethyl 5-(*tert*-butylamino)-4-phenyl-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (**4b**)



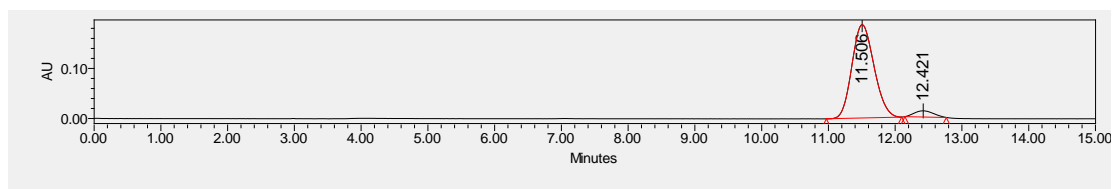
Yellow solid; m.p. 164–168 °C; 25.3 mg, 65% yield, 94.5:5.5 er;  $[\alpha]_D^{22} = -12.16$  ( $c = 0.36$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IE, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 11.51 min,  $t_R$  (minor) = 12.42 min.

IR (neat): 3307, 2978, 2930, 2361, 1724, 1598, 1534, 1456, 1367, 1332, 1228, 1125, 1198, 1048, 762 and 701  $\text{cm}^{-1}$ .

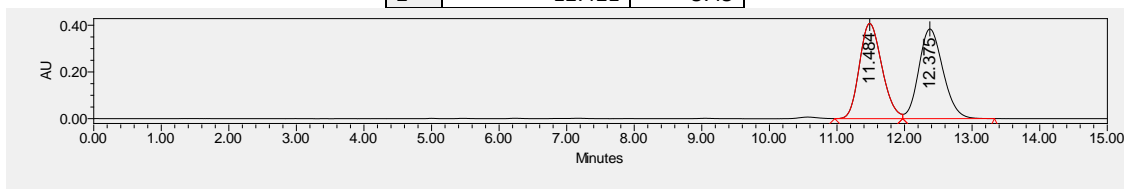
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.36 - 7.31$  (m, 3H), 7.15 – 7.09 (m, 2H), 5.52 (s, 1H), 5.21 (s, 1H), 4.35 – 4.26 (m, 2H), 3.74 – 3.63 (m, 1H), 3.51 – 3.40 (m, 1H), 1.49 (s, 9H), 1.30 (t,  $J = 7.2$  Hz, 3H), 0.80 (t,  $J = 7.2$  Hz, 3H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 218.2, 181.8, 166.5, 165.6, 134.0, 129.7, 129.1, 129.0, 79.1, 63.2, 61.7, 59.1, 55.9, 28.9, 14.0, 13.5$ .

HRMS (ESI-FT) calcd for  $\text{C}_{20}\text{H}_{27}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 391.1686, Found 391.1678.

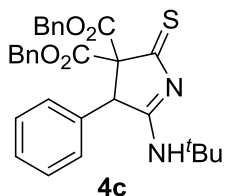


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 11.506         | 94.55  |
| 2 | 12.421         | 5.45   |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 11.484         | 49.70  |
| 2 | 12.375         | 50.30  |

**Dibenzyl 5-(*tert*-butylamino)-4-phenyl-2-thioxo-2,4-dihydro-3*H*-pyrrole-3,3-dicarboxylate (4c)**



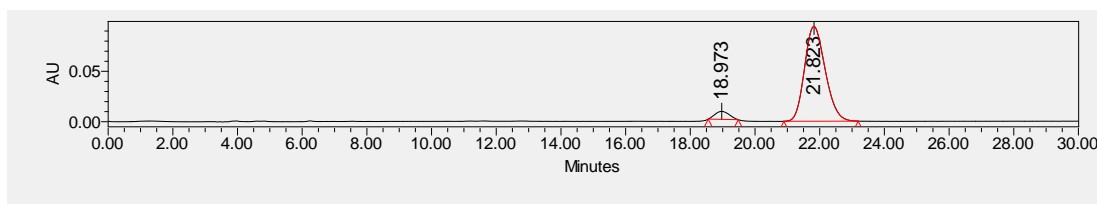
Yellow solid; m.p. 72–76 °C; 34.8 mg, 68% yield, 94.5:5.5 er;  $[\alpha]_D^{22} = -9.85$  ( $c = 0.68$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IE, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 21.82 min,  $t_R$  (minor) = 18.97 min.

IR (neat): 3313, 2926, 2360, 1723, 1597, 1531, 1455, 1369, 1331, 1288, 1207, 1121, 1096, 749 and 697  $\text{cm}^{-1}$ .

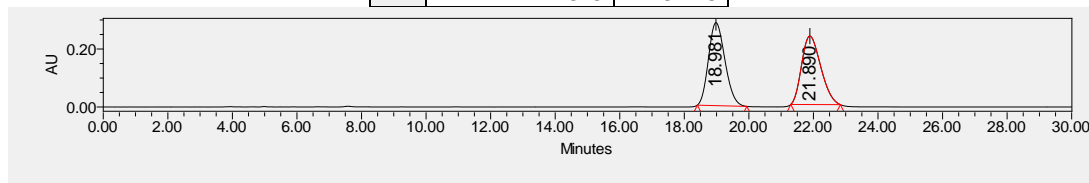
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.36 - 7.21$  (m, 11H), 7.11 – 7.05 (m, 2H), 6.99 – 6.77 (m, 2H), 5.53 (s, 1H), 5.31 – 5.07 (m, 3H), 4.66 (d,  $J = 12.4$  Hz, 1H), 4.24 (d,  $J = 12.4$  Hz, 1H), 1.48 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 217.5, 181.7, 166.3, 165.3, 135.2, 134.7, 133.7, 129.7, 129.2, 129.1, 128.5, 128.4, 128.3, 128.2, 79.3, 68.9, 67.5, 59.2, 56.0, 28.9$ .

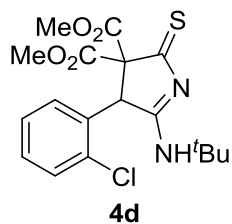
HRMS (ESI-FT) calcd for  $\text{C}_{30}\text{H}_{31}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 515.1999, Found 515.2001.



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 18.973         | 5.52   |
| 2 | 21.823         | 94.48  |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 18.981         | 51.13  |
| 2 | 21.890         | 48.87  |

**Dimethyl 5-(*tert*-butylamino)-4-(2-chlorophenyl)-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4d)**

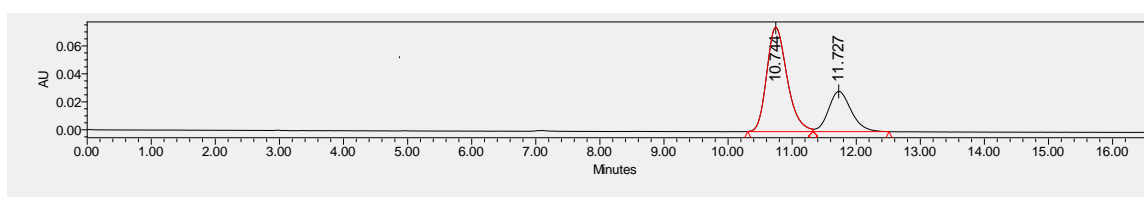
Yellow solid; m.p. 198–202 °C; 22.5 mg, 57% yield, 70:30 er;  $[\alpha]_D^{22} = +53.47$  ( $c = 0.40$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IE, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 10.75 min,  $t_R$  (minor) = 11.73 min.

IR (neat): 3310, 2953, 1728, 1600, 1532, 1476, 1435, 1338, 1293, 1207, 1124, 1099, 1050 and 760  $\text{cm}^{-1}$ .

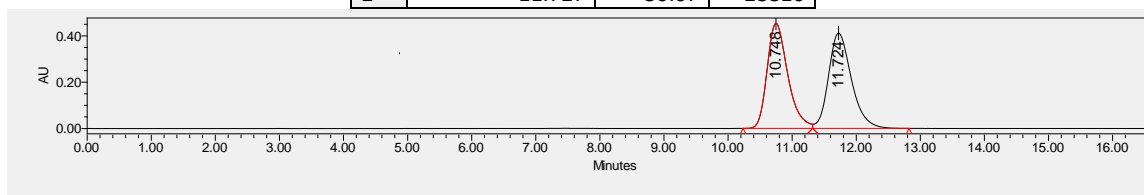
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.47 - 7.40$  (m, 1H), 7.31 – 7.22 (m, 2H), 7.00 – 6.89 (d,  $J = 7.5$  Hz, 1H), 5.75 (d,  $J = 2.4$  Hz, 1H), 5.57 (s, 1H), 3.85 (s, 3H), 3.12 (s, 3H), 1.52 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 218.1, 181.7, 166.5, 165.7, 136.2, 132.6, 130.3, 130.0, 129.8, 127.4, 78.4, 56.3, 55.7, 54.2, 52.3, 28.9$ .

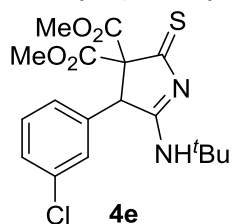
HRMS (ESI-FT) calcd for  $\text{C}_{18}\text{H}_{22}^{35}\text{ClN}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 397.0983, Found 397.0983;  $\text{C}_{18}\text{H}_{22}^{37}\text{ClN}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 399.0954, Found 399.0949.



|   | Retention Time | % Area | Height |
|---|----------------|--------|--------|
| 1 | 10.744         | 69.93  | 74609  |
| 2 | 11.727         | 30.07  | 28816  |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 10.748         | 49.80  |
| 2 | 11.724         | 50.20  |

**Dimethyl 5-(*tert*-butylamino)-4-(3-chlorophenyl)-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4e)**

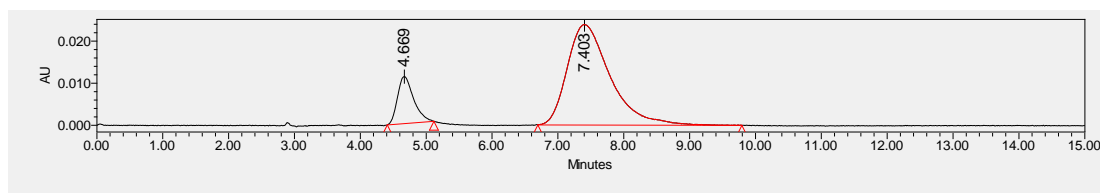
Yellow solid; m.p. 97–101 °C; 20.5 mg, 52% yield, 85:15 er;  $[\alpha]_D^{22} = -4.05$  ( $c = 0.37$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IG, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 7.40 min,  $t_R$  (minor) = 4.67 min.

IR (neat): 3309, 2971, 2360, 1729, 1602, 1533, 1476, 1433, 1336, 1288, 1206, 1125, 1099, 1050, 750 and 694  $\text{cm}^{-1}$ .

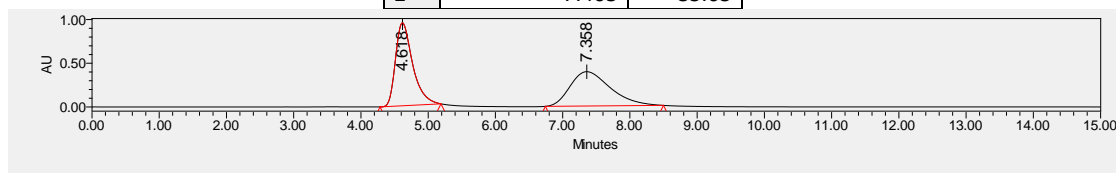
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.36 - 7.26$  (m, 2H), 7.17 – 7.11 (m, 1H), 7.01 – 6.95 (m, 1H), 5.55 (s, 1H), 5.20 (s, 1H), 3.86 (s, 3H), 3.22 (s, 3H), 1.52 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 217.3, 180.9, 166.8, 165.8, 135.7, 135.0, 130.4, 129.4, 79.3, 58.7, 56.3, 54.2, 52.5, 28.9$ .

HRMS (ESI-FT) calcd for  $\text{C}_{18}\text{H}_{22}^{35}\text{ClN}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 397.0983, Found 397.0980;  $\text{C}_{18}\text{H}_{22}^{37}\text{ClN}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 399.0954, Found 399.0956.

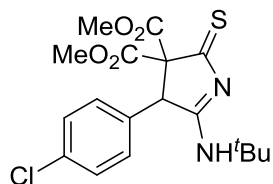


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 4.669          | 14.95  |
| 2 | 7.403          | 85.05  |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 4.618          | 50.67  |
| 2 | 7.358          | 49.33  |

**Dimethyl 5-(*tert*-butylamino)-4-(4-chlorophenyl)-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4f)**



**4f**

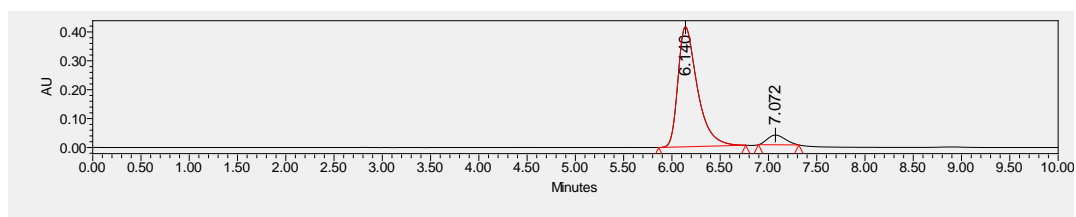
Yellow solid; m.p. 136–140 °C; 33.8 mg, 85% yield, 93:7 er;  $[\alpha]_D^{22} = -27.24$  ( $c = 0.62$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IF, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 6.14 min,  $t_R$  (minor) = 7.07 min.

IR (neat): 3312, 2953, 2361, 1729, 1601, 1534, 1492, 1434, 1288, 1207, 1125, 1094, 1050, 1017, 838, 799 and 749  $\text{cm}^{-1}$ .

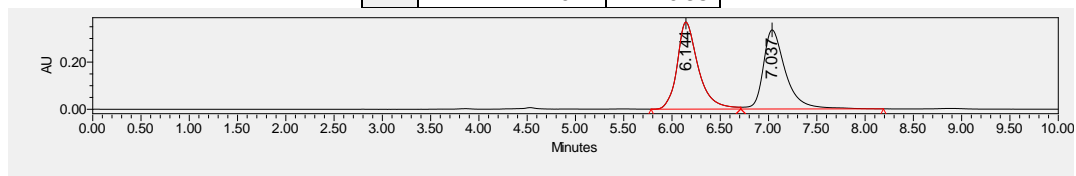
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.39 - 7.26$  (m, 2H), 7.08 – 7.00 (m, 2H), 5.67 (s, 1H), 5.20 (s, 1H), 3.84 (s, 3H), 3.19 (s, 3H), 1.49 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 217.3, 181.2, 166.7, 165.8, 135.2, 132.3, 130.9, 129.3, 79.2, 58.5, 56.2, 54.1, 52.5, 28.9$ .

HRMS (ESI-FT) calcd for  $\text{C}_{18}\text{H}_{22}^{35}\text{ClN}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 397.0983, Found 397.0992;  $\text{C}_{18}\text{H}_{22}^{37}\text{ClN}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 399.0954, Found 399.0961.

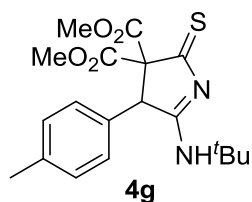


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 6.140          | 93.07  |
| 2 | 7.072          | 6.93   |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 6.144          | 50.99  |
| 2 | 7.037          | 49.01  |

#### Dimethyl 5-(*tert*-butylamino)-2-thioxo-4-(*p*-tolyl)-2,4-dihydro-3*H*-pyrrole-3,3-dicarboxylate (**4g**)



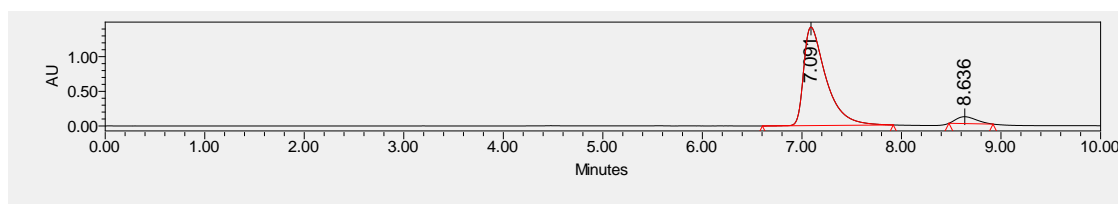
Yellow solid; m.p. 85–89 °C; 25.6 mg, 68% yield, 94:6 er;  $[\alpha]_D^{22} = -31.06$  ( $c = 0.40$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IF, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 7.09 min,  $t_R$  (minor) = 8.64 min.

IR (neat): 3311, 2952, 2360, 2341, 1729, 1600, 1533, 1433, 1337, 1288, 1208, 1125, 1099, 940 and 749  $\text{cm}^{-1}$ .

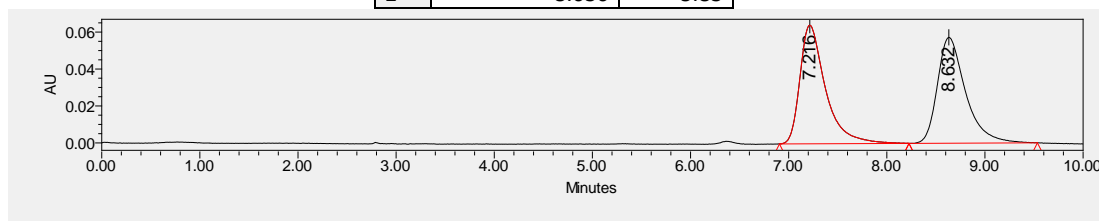
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.21 - 7.12$  (m, 2H), 7.03 – 6.92 (m, 2H), 5.58 (s, 1H), 5.20 (s, 1H), 3.84 (s, 3H), 3.17 (s, 3H), 2.33 (s, 3H), 1.50 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 218.0, 182.0, 167.0, 166.0, 139.1, 130.5, 129.8, 129.5, 79.4, 58.9, 56.0, 54.0, 52.4, 28.9, 21.3$ .

HRMS (ESI-FT) calcd for  $\text{C}_{19}\text{H}_{25}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 377.1530, Found 377.1528.

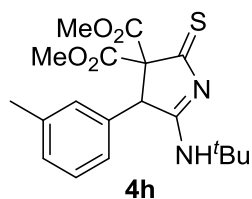


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 7.091          | 94.17  |
| 2 | 8.636          | 5.83   |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 7.216          | 50.44  |
| 2 | 8.632          | 49.56  |

#### Dimethyl 5-(*tert*-butylamino)-2-thioxo-4-(*m*-tolyl)-2,4-dihydro-3*H*-pyrrole-3,3-dicarboxylate (**4h**)



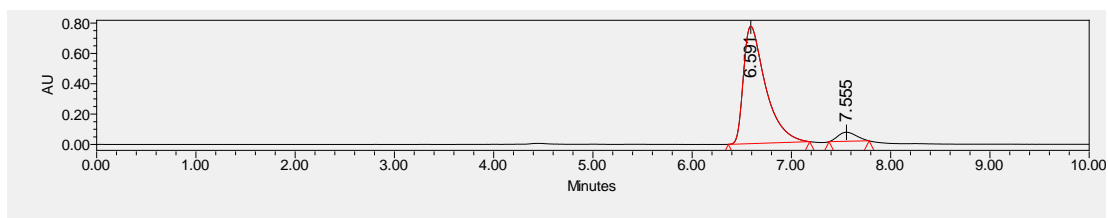
Yellow solid; m.p. 116–120 °C; 21.2 mg, 56% yield, 94:6 er;  $[\alpha]_D^{22} = -16.48$  ( $c = 0.35$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IF, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 6.59 min,  $t_R$  (minor) = 7.55 min.

IR (neat): 3310, 2953, 2361, 1729, 1560, 1534, 1433, 1336, 1289, 1207, 1124, 1098, 1050, 748, 750 and 703  $\text{cm}^{-1}$ .

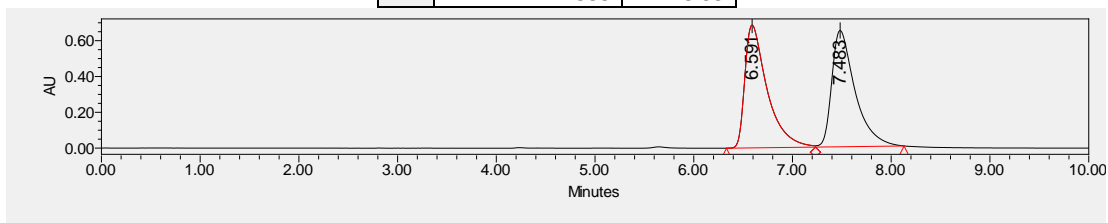
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.25 - 7.19$  (m, 1H), 7.16 – 7.12 (m, 1H), 6.95 – 6.91 (m, 1H), 6.90 – 6.85 (m, 1H), 5.55 (s, 1H), 5.20 (s, 1H), 3.85 (s, 3H), 3.16 (s, 3H), 2.33 (s, 3H), 1.51 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 218.0, 181.9, 167.0, 165.9, 139.0, 133.6, 130.5, 129.8, 129.0, 126.5, 79.4, 59.2, 56.0, 54.1, 54.0, 52.3, 29.0, 21.4$ .

HRMS (ESI-FT) calcd for  $\text{C}_{19}\text{H}_{25}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 377.1530, Found 377.1526.

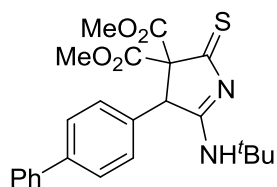


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 6.591          | 94.01  |
| 2 | 7.555          | 5.99   |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 6.591          | 50.27  |
| 2 | 7.483          | 49.73  |

**Dimethyl 4-([1,1'-biphenyl]-4-yl)-5-(*tert*-butylamino)-2-thioxo-2,4-dihydro-3*H*-pyrrole-3,3-dicarboxylate (4i)**



**4i**

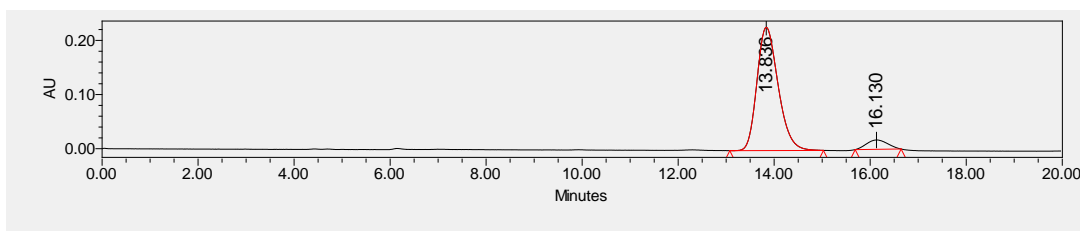
Yellow solid; m.p. 157–161 °C; 20.6 mg, 47% yield, 93:7 er;  $[\alpha]_D^{22} = -63.55$  ( $c = 0.31$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IE, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 13.84 min,  $t_R$  (minor) = 16.13 min.

IR (neat): 3309, 2971, 2360, 2341, 1729, 1560, 1533, 1487, 1434, 1337, 1288, 1207, 1124, 1098, 765, 750 and 699  $\text{cm}^{-1}$ .

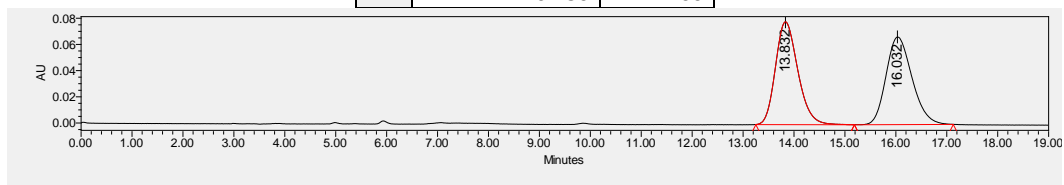
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.61 - 7.55$  (m, 4H), 7.48 – 7.42 (m, 2H), 7.40 – 7.35 (m, 1H), 7.21 – 7.14 (m, 2H), 5.63 (s, 1H), 5.29 (s, 1H), 3.87 (s, 3H), 3.17 (s, 3H), 1.53 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 217.8, 181.7, 167.0, 166.0, 141.9, 139.9, 132.6, 130.1, 129.1, 128.1, 127.7, 127.2, 127.1, 79.5, 58.9, 56.1, 54.1, 52.5, 29.0$ .

HRMS (ESI-FT) calcd for  $\text{C}_{24}\text{H}_{27}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 439.1686, Found 439.1687.



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 13.836         | 93.00  |
| 2 | 16.130         | 7.00   |

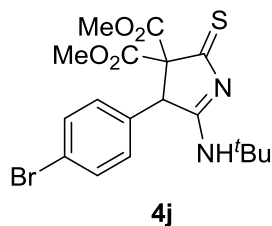


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 13.832         |        |
| 2 | 16.032         |        |



|   |        |       |
|---|--------|-------|
| 1 | 13.832 | 50.29 |
| 2 | 16.032 | 49.71 |

**Dimethyl 4-(4-bromophenyl)-5-(*tert*-butylamino)-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4j)**



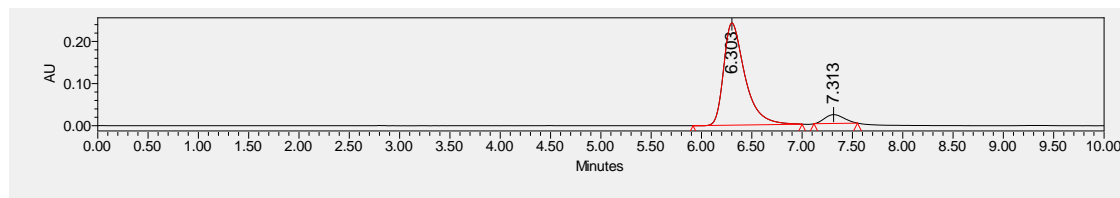
Yellow solid; m.p. 177–181 °C; 22.4 mg, 51% yield, 93:7 er;  $[\alpha]_D^{22} = -33.58$  ( $c = 0.55$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IF, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 6.30 min,  $t_R$  (minor) = 7.31 min.

IR (neat): 3311, 2952, 2360, 1728, 1600, 1532, 1488, 1434, 1407, 1335, 1286, 1207, 1125, 1099, 1049, 1012, 836, 798 and 748  $\text{cm}^{-1}$ .

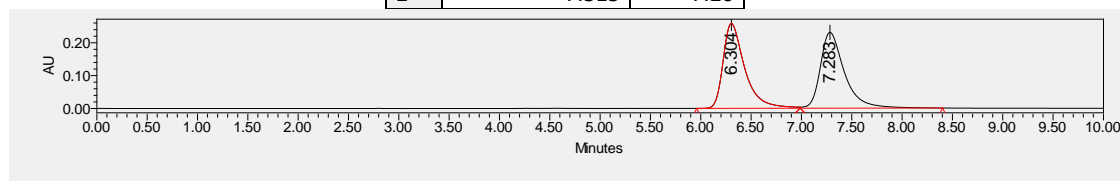
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.51 - 7.42$  (m, 2H), 7.06 – 6.93 (m, 2H), 5.64 (s, 1H), 5.18 (s, 1H), 3.84 (s, 3H), 3.19 (s, 3H), 1.50 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 217.3, 181.1, 166.7, 165.8, 132.8, 132.3, 131.2, 123.4, 79.1, 58.5, 56.2, 54.1, 52.5, 28.9$ .

HRMS (ESI-FT) calcd for  $\text{C}_{18}\text{H}_{22}^{79}\text{BrN}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 441.0478, Found 441.0485;  $\text{C}_{18}\text{H}_{22}^{81}\text{BrN}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 443.0458, Found 443.0462.

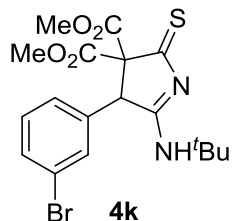


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 6.303          | 92.74  |
| 2 | 7.313          | 7.26   |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 6.304          | 50.23  |
| 2 | 7.283          | 49.77  |

**Dimethyl 4-(3-bromophenyl)-5-(*tert*-butylamino)-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4k)**



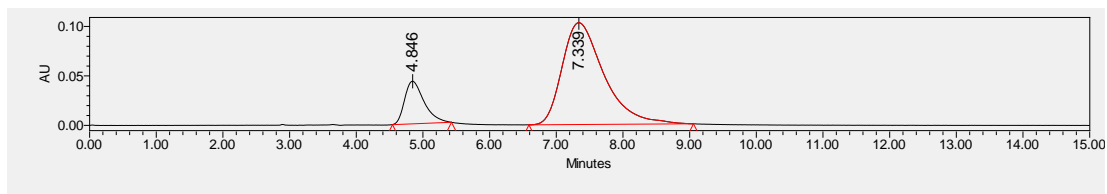
Yellow solid; m.p. 84–88 °C; 30.2 mg, 68% yield, 84:16 er;  $[\alpha]_D^{22} = -5.16$  ( $c = 0.50$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IG, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 7.34 min,  $t_R$  (minor) = 4.85 min.

IR (neat): 3309, 2952, 2360, 1729, 1601, 1532, 1475, 1433, 1335, 1287, 1206, 1125, 1099, 1049, 479 and 694  $\text{cm}^{-1}$ .

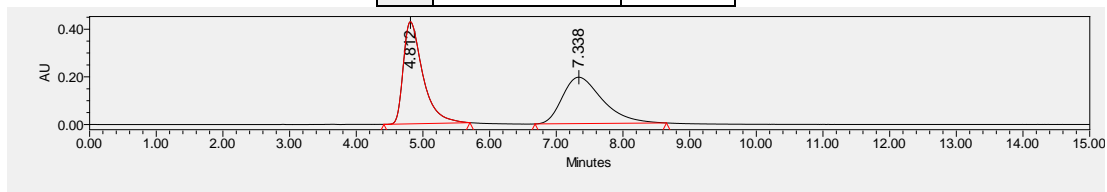
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.51 - 7.42$  (m, 2H), 7.06 – 6.93 (m, 2H), 5.64 (s, 1H), 5.18 (s, 1H), 3.84 (s, 3H), 3.19 (s, 3H), 1.50 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 217.2, 181.0, 166.7, 165.7, 136.0, 132.3, 130.7, 123.0, 121.1, 79.3, 58.6, 56.2, 54.2, 52.5, 28.9$ .

HRMS (ESI-FT) calcd for  $\text{C}_{18}\text{H}_{22}^{79}\text{BrN}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 441.0478, Found 441.0479;  $\text{C}_{18}\text{H}_{22}^{81}\text{BrN}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 443.0458, Found 443.0458.

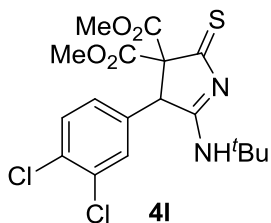


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 4.846          | 16.34  |
| 2 | 7.339          | 83.66  |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 4.812          | 51.62  |
| 2 | 7.338          | 48.38  |

**Dimethyl 5-(*tert*-butylamino)-4-(3,4-dichlorophenyl)-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4I)**



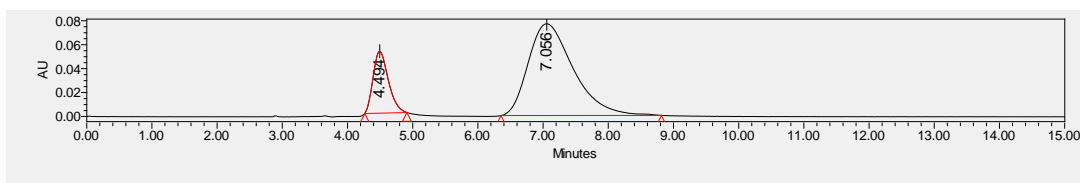
Yellow solid; m.p. 66–70 °C; 30.3 mg, 70% yield, 81.5:18.5 er;  $[\alpha]_D^{22} = -19.11$  ( $c = 0.65$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL **IG**, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 7.06 min,  $t_R$  (minor) = 4.49 min.

**IR** (neat): 3310, 2953, 1728, 1599, 1531, 1470, 1434, 1398, 1331, 1283, 1203, 1130, 1099, 1033, 946, 896, 737 and 677  $\text{cm}^{-1}$ .

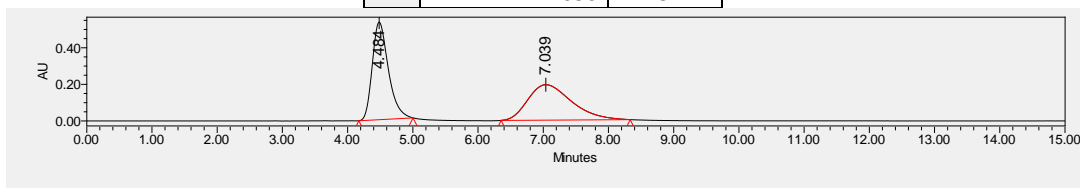
**$^1\text{H}$  NMR** (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.46 - 7.40$  (m, 1H), 7.26 – 7.22 (m, 1H), 6.98 – 6.87 (m, 1H), 5.66 (s, 1H), 5.18 (s, 1H), 3.85 (s, 3H), 3.26 (s, 3H), 1.51 (s, 9H).

**$^{13}\text{C}\{^1\text{H}\}$  NMR** (101 MHz,  $\text{CDCl}_3$ )  $\delta = 216.9, 180.5, 166.6, 165.7, 133.9, 133.6, 133.3, 131.7, 131.1, 128.6, 79.1, 58.1, 56.3, 54.3, 52.6, 28.9$ .

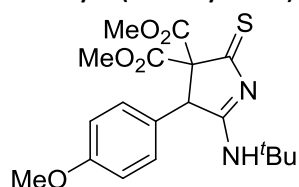
**HRMS** (ESI-FT) calcd for  $\text{C}_{18}\text{H}_{21}^{35}\text{Cl}_2\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 431.0594, Found 431.0599;  $\text{C}_{18}\text{H}_{21}^{35}\text{Cl}^{37}\text{ClN}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 433.0564, Found 433.0567;  $\text{C}_{18}\text{H}_{21}^{37}\text{Cl}_2\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 435.0535, Found 435.0534.



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 4.494          | 18.59  |
| 2 | 7.066          | 81.41  |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 4.484          | 50.46  |
| 2 | 7.039          | 49.54  |

**Dimethyl 5-(*tert*-butylamino)-4-(4-methoxyphenyl)-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4m)****4m**

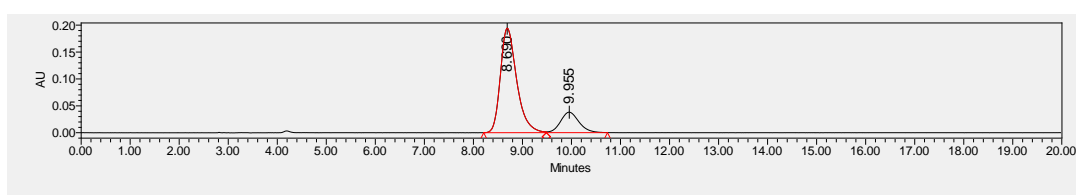
Yellow solid; m.p. 105–109 °C; 30.9 mg, 79% yield, 82:18 er;  $[\alpha]_D^{22} = -33.74$  ( $c = 0.49$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IF, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 8.69 min,  $t_R$  (minor) = 9.96 min.

IR (neat): 3310, 2957, 1728, 1603, 1514, 1435, 1333, 1287, 1207, 1098, 1033, 839 and 753  $\text{cm}^{-1}$ .

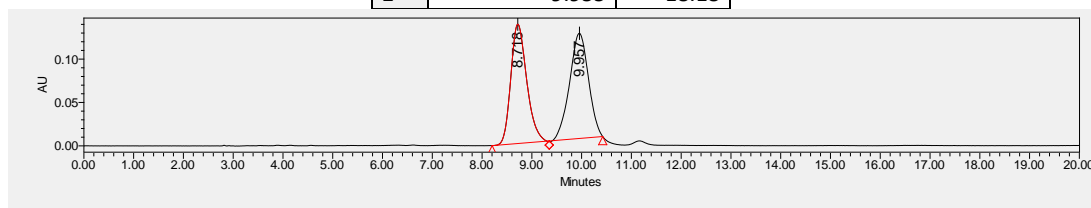
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.05 - 6.98$  (m, 2H), 6.90 – 6.80 (m, 2H), 5.63 (s, 1H), 5.19 (s, 1H), 3.84 (s, 3H), 3.79 (s, 3H), 3.19 (s, 3H), 1.50 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 217.9, 182.0, 167.0, 166.0, 160.0, 130.8, 125.3, 114.5, 79.3, 58.5, 56.0, 55.4, 54.0, 52.5, 28.9$ .

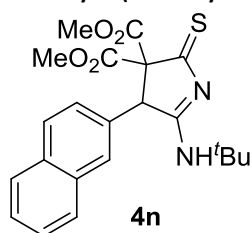
HRMS (ESI-FT) calcd for  $\text{C}_{19}\text{H}_{25}\text{N}_2\text{O}_5\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 393.1479, Found 393.1479.



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 8.690          | 81.82  |
| 2 | 9.955          | 18.18  |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 8.718          | 48.80  |
| 2 | 9.957          | 51.20  |

**Dimethyl 5-(*tert*-butylamino)-4-(naphthalen-2-yl)-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4n)****4n**

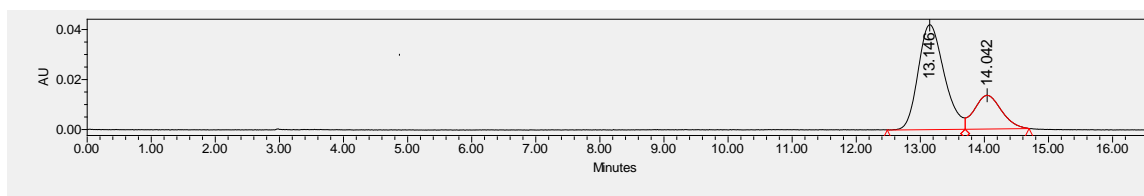
Yellow solid; m.p. 214–218 °C; 34.4 mg, 83% yield, 75:25 er;  $[\alpha]_D^{22} = -22.63$  ( $c = 0.55$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IE, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 13.15 min,  $t_R$  (minor) = 14.04 min.

IR (neat): 3310, 2971, 1729, 1601, 1531, 1435, 1335, 1287, 1207, 1125, 1098, 1051 and 752  $\text{cm}^{-1}$ .

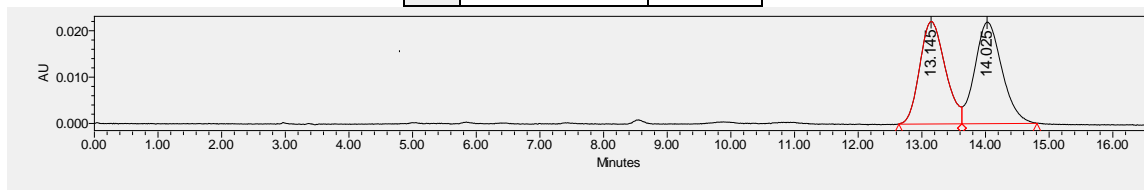
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.88 - 7.60$  (m, 3H), 7.68 – 7.59 (m, 1H), 7.56 – 7.49 (m, 2H), 7.18 – 7.09 (m, 1H), 5.62 (s, 1H), 5.41 (s, 1H), 3.87 (s, 3H), 3.02 (s, 3H), 1.51 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 217.9, 181.8, 167.0, 166.0, 133.2, 131.0, 129.0, 128.0, 127.9, 127.2, 127.0, 79.4, 59.4, 56.1, 54.1, 52.4, 28.9$ .

HRMS (ESI-FT) calcd for  $\text{C}_{22}\text{H}_{25}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 413.1530, Found 413.1531.

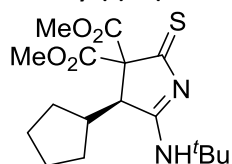


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 13.146         | 74.87  |
| 2 | 14.042         | 25.13  |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 13.145         | 48.74  |
| 2 | 14.025         | 51.26  |

**Dimethyl (S)-5-(*tert*-butylamino)-4-cyclopentyl-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4o)**



**4o**

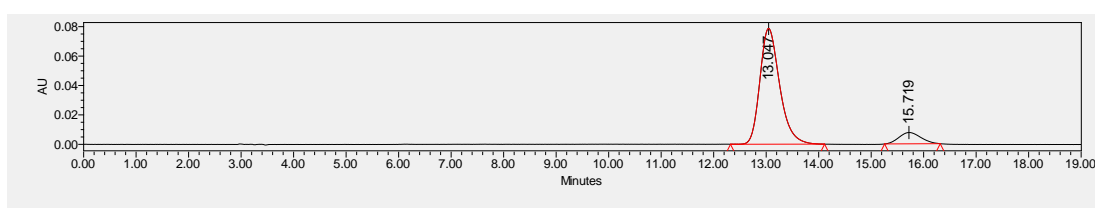
Yellow solid; m.p. 158–162 °C; 16.0 mg, 45% yield, 90:10 er;  $[\alpha]_D^{22} = -81.94$  ( $c = 0.31$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IE, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 13.05 min,  $t_R$  (minor) = 15.72 min.

IR (neat): 3342, 2957, 2872, 1730, 1597, 1526, 1435, 1332, 1294, 1207, 1103, 934 and 753  $\text{cm}^{-1}$ .

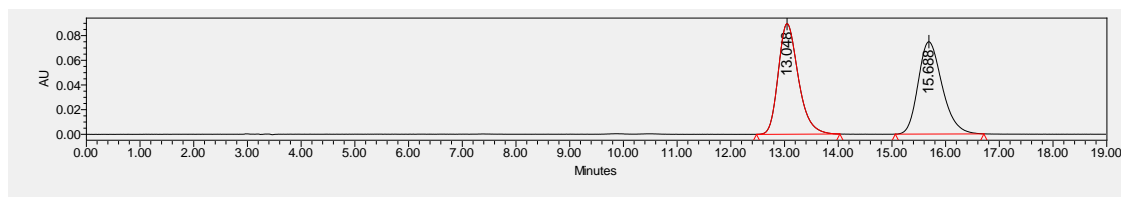
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta = 6.03$  (s, 1H), 3.85 – 3.79 (m, 4H), 3.75 (s, 3H), 2.18 – 2.07 (m, 1H), 1.91 – 1.79 (m, 2H), 1.71 – 1.66 (m, 2H), 1.63 – 1.56 (m, 2H), 1.53 (s, 9H), 1.29 – 1.20 (m, 2H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 218.0, 182.6, 167.5, 166.5, 77.9, 58.7, 56.0, 53.8, 53.0, 39.0, 32.3, 31.0, 29.0, 25.7, 24.8$ .

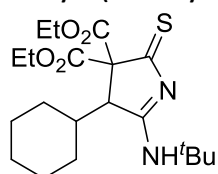
HRMS (ESI-FT) calcd for  $\text{C}_{17}\text{H}_{27}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 355.1686, Found 355.1685.



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 13.047         | 90.21  |
| 2 | 15.719         | 9.79   |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 13.048         | 50.09  |
| 2 | 15.688         | 49.91  |

**Diethyl 5-(*tert*-butylamino)-4-cyclohexyl-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4p)****4p**

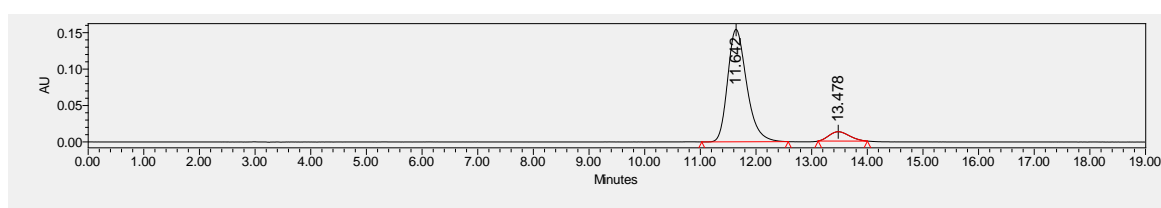
Yellow solid; m.p. 72–76 °C; 27.2 mg, 67% yield, 92:8 er;  $[\alpha]_D^{22} = +101.9$  ( $c = 0.55$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IE, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 11.64 min,  $t_R$  (minor) = 13.48 min.

IR (neat): 3345, 2929, 2855, 1727, 1594, 1529, 1451, 1340, 1284, 1208, 1123, 1098, 1032, 860 and 756  $\text{cm}^{-1}$ .

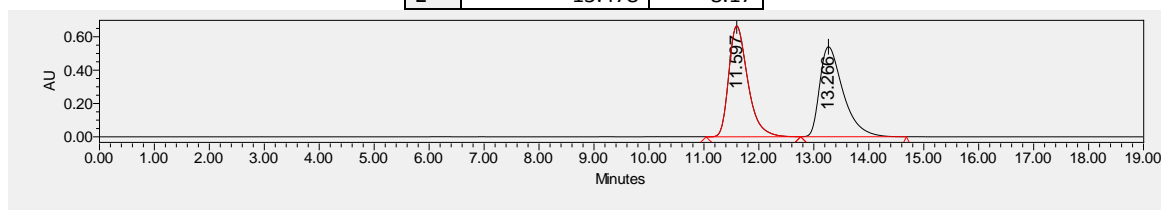
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 5.90$  (s, 1H), 4.32 – 4.09 (m, 4H), 3.49 (d,  $J = 2.4$  Hz, 1H), 1.82 – 1.74 (m, 2H), 1.71 – 1.61 (m, 4H), 1.53 (s, 9H), 1.31 – 1.21 (m, 8H), 1.11 – 0.98 (m, 2H), 0.80 – 0.69 (m, 1H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 217.7, 181.8, 166.7, 166.0, 63.1, 61.8, 59.3, 55.9, 39.0, 33.0, 29.1, 29.0, 27.2, 26.3, 26.2, 14.2, 13.8$ .

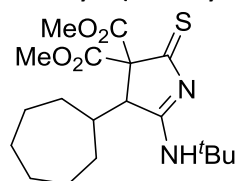
HRMS (ESI-FT) calcd for  $\text{C}_{20}\text{H}_{33}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 397.2156, Found 397.2158.



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 11.642         | 91.83  |
| 2 | 13.478         | 8.17   |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 11.597         | 50.02  |
| 2 | 13.266         | 49.98  |

**Dimethyl 5-(*tert*-butylamino)-4-cycloheptyl-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4q)****4q**

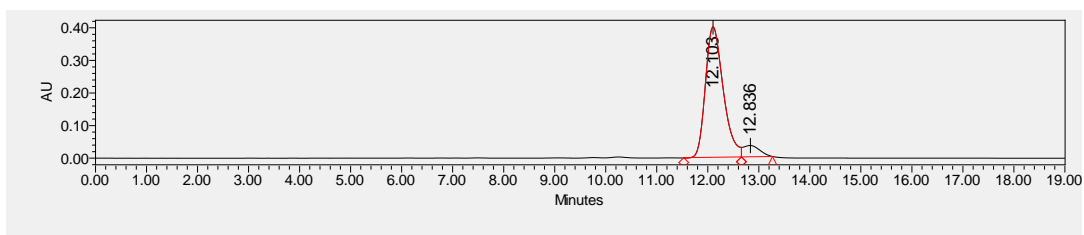
Yellow solid; m.p. 156–160 °C; 20.0 mg, 52% yield, 92.5:7.5 er;  $[\alpha]_D^{22} = +37.13$  ( $c = 0.40$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IE, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 12.10 min,  $t_R$  (minor) = 12.84 min.

IR (neat): 3344, 2925, 2857, 1730, 1593, 1526, 1436, 1340, 1286, 1207, 1122, 1066 and 751  $\text{cm}^{-1}$ .

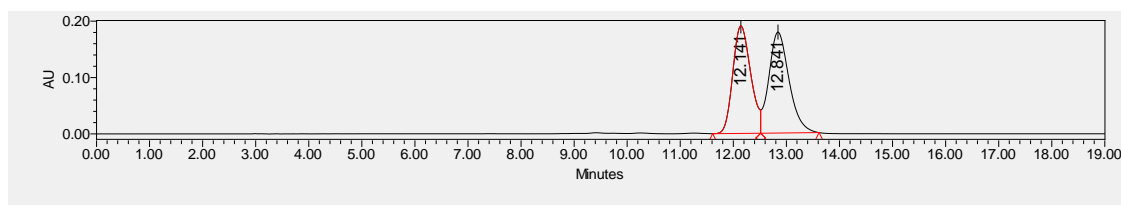
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 5.80$  (s, 1H), 3.79 – 3.74 (m, 7H), 1.84 – 1.61 (m, 6H), 1.53 (s, 9H), 1.49 – 1.37 (m, 5H), 1.35 – 1.33 (m, 2H), 1.11 – 0.99 (m, 1H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 218.1, 181.9, 167.3, 166.7, 60.3, 55.9, 54.1, 52.8, 52.8, 39.6, 35.6, 30.2, 29.1, 27.8, 27.2, 27.0$ .

HRMS (ESI-FT) calcd for  $\text{C}_{19}\text{H}_{31}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 383.1999, Found 383.1996.

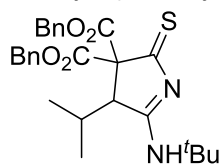


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 12.103         | 92.54  |
| 2 | 12.836         | 7.46   |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 12.141         | 48.46  |
| 2 | 12.841         | 51.54  |

**Dibenzyl 5-(*tert*-butylamino)-4-isopropyl-2-thioxo-2,4-dihydro-3*H*-pyrrole-3,3-dicarboxylate (4r)**



**4r**

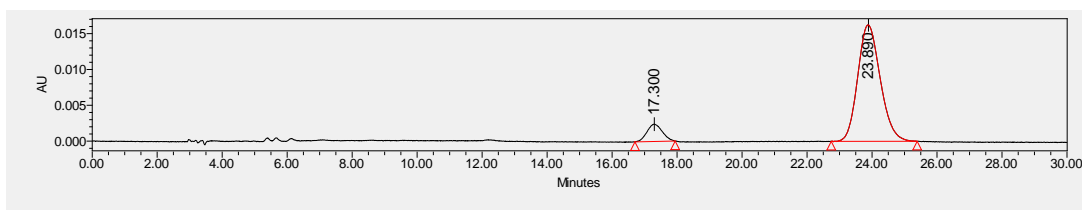
Yellow solid; m.p. 98–102 °C; 20.1 mg, 42% yield, 91:9 er;  $[\alpha]_D^{22} = +97.78$  ( $c = 0.40$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IE, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 23.89 min,  $t_R$  (minor) = 17.30 min.

IR (neat): 3345, 2968, 2326, 1728, 1596, 1525, 1459, 1365, 1312, 1287, 1207, 1131, 1046, 751 and 698  $\text{cm}^{-1}$ .

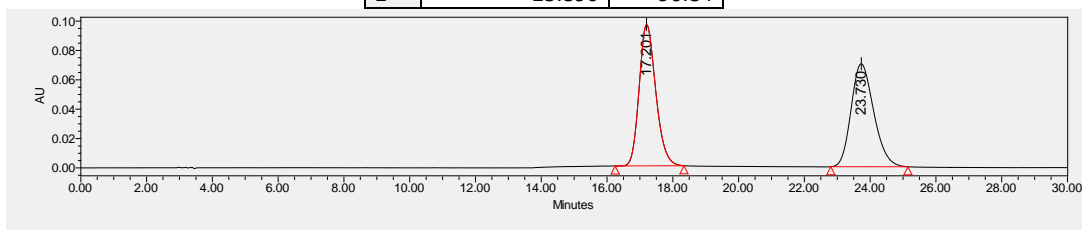
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.31 - 7.24$  (m, 10H), 5.74 (s, 1H), 5.27 – 5.01 (m, 4H), 3.56 (d,  $J = 2.6$  Hz, 1H), 1.86 – 1.77 (m, 1H), 1.50 (s, 9H), 1.01 (d,  $J = 6.8$  Hz, 3H), 0.73 (d,  $J = 6.8$  Hz, 3H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 217.1, 181.5, 166.4, 165.7, 135.1, 135.0, 128.7, 128.6, 128.6, 128.5, 128.3, 128.1, 68.7, 67.6, 59.5, 56.0, 29.1, 28.3, 22.4, 17.9$ .

HRMS (ESI-FT) calcd for  $\text{C}_{27}\text{H}_{33}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 481.2156, Found 481.2155.

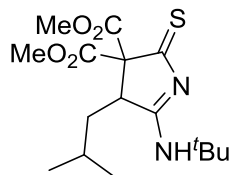


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 17.300         | 9.16   |
| 2 | 23.890         | 90.84  |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 17.201         | 50.14  |
| 2 | 23.730         | 49.86  |

**Dimethyl 5-(*tert*-butylamino)-4-isobutyl-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4s)**



**4s**

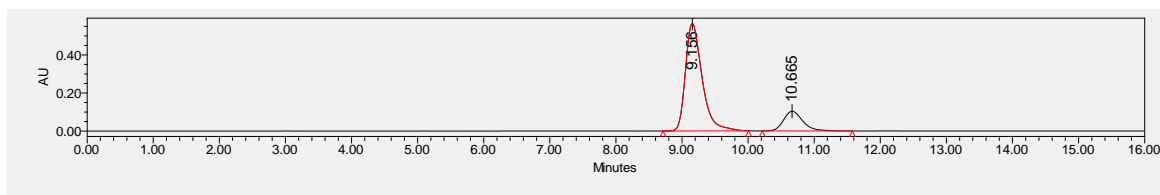
Yellow solid; m.p. 200–204 °C; 25.1 mg, 70% yield, 82:18 er;  $[\alpha]_D^{22} = -109.64$  ( $c = 0.28$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IE, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 9.16 min,  $t_R$  (minor) = 10.67 min.

IR (neat): 3329, 2958, 1729, 1597, 1520, 1435, 1335, 1298, 1204, 1119, 1074, 1045, 981 and 754  $\text{cm}^{-1}$ .

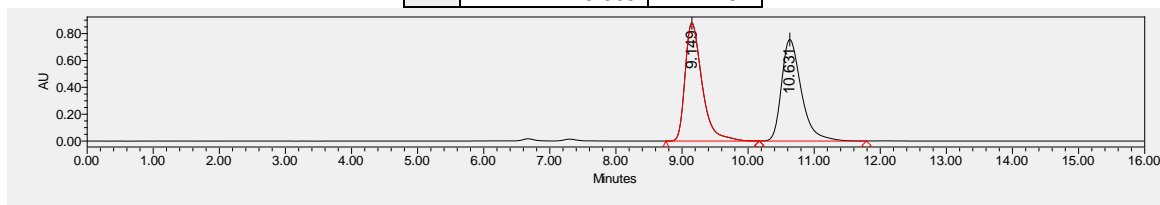
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 5.73$  (s, 1H), 4.00 (dd,  $J = 10.0, 5.2$  Hz, 1H), 3.84 (s, 3H), 3.74 (s, 3H), 1.54 (m, 10H), 1.35 – 1.23 (m, 2H), 0.93 (dd,  $J = 13.2, 6.4$  Hz, 6H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 218.5, 183.4, 167.4, 166.4, 77.3, 56.1, 53.7, 53.1, 51.2, 38.0, 29.0, 26.4, 23.4, 21.8$ .

HRMS (ESI-FT) calcd for  $\text{C}_{16}\text{H}_{27}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 343.1686, Found 343.1686.

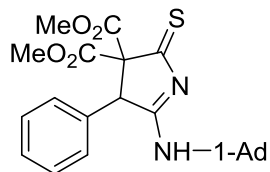


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 9.156          | 82.46  |
| 2 | 10.665         | 17.54  |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 9.149          | 49.95  |
| 2 | 10.631         | 50.05  |

**Dimethyl 5-(adamantan-1-ylamino)-4-phenyl-2-thioxo-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (4t)**



**4t**

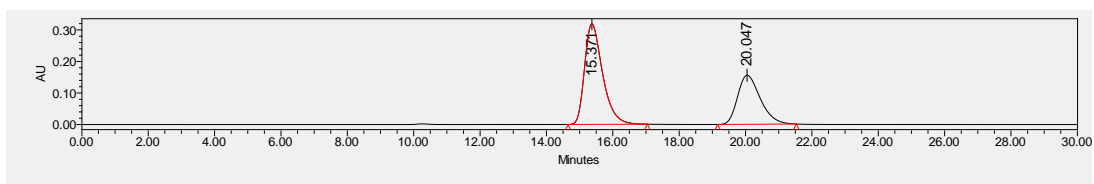
Yellow solid; m.p. 226–230 °C; 28.1 mg, 64% yield, 62:38 er;  $[\alpha]_D^{22} = -6.44$  ( $c = 0.40$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IE, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 320$  nm,  $t_R$  (major) = 15.37 min,  $t_R$  (minor) = 20.05 min.

IR (neat): 2911, 2853, 1728, 1600, 1529, 1453, 1333, 1289, 1084, 755 and 701  $\text{cm}^{-1}$ .

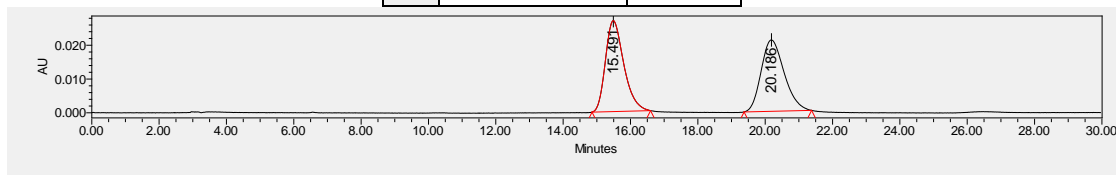
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.38 - 7.30$  (m 3H), 7.17 – 7.05 (m, 2H), 5.48 (s, 1H), 5.25 (s, 1H), 3.85 (s, 3H), 3.13 (s, 3H), 2.15 – 2.10 (m, 8H), 1.77 – 1.60 (m, 7H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 217.8, 181.6, 167.0, 165.9, 133.8, 129.7, 129.1, 129.1, 79.3, 59.2, 59.2, 56.7, 54.1, 54.0, 52.4, 41.5, 35.9, 29.5$ .

HRMS (ESI-FT) calcd for  $\text{C}_{24}\text{H}_{29}\text{N}_2\text{O}_4\text{S}^+$  ( $[\text{M}+\text{H}^+]$ ) = 441.1843, Found 441.1847.

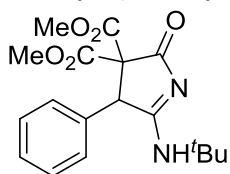


|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 15.371         | 61.93  |
| 2 | 20.047         | 38.07  |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 15.491         | 50.35  |
| 2 | 20.186         | 49.65  |

**Dimethyl 5-(*tert*-butylamino)-2-oxo-4-phenyl-2,4-dihydro-3H-pyrrole-3,3-dicarboxylate (5a)**



**5a**

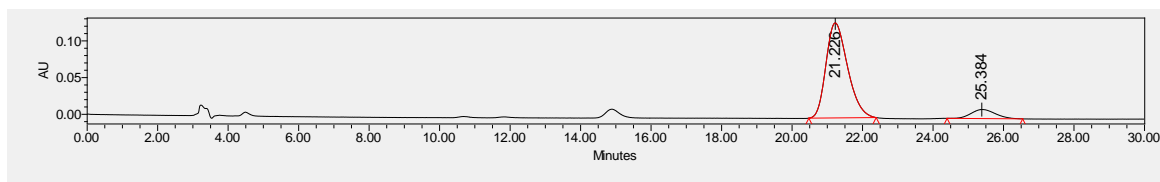
White solid; m.p. 210–214 °C; 3.9 mg, 11% yield, 89:11 er;  $[\alpha]_D^{22} = -102.14$  ( $c = 0.47$  in  $\text{CH}_2\text{Cl}_2$ ). HPLC DAICEL CHIRALCEL IE, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min,  $\lambda = 254$  nm,  $t_R$  (major) = 21.23 min,  $t_R$  (minor) = 25.38 min.

IR (neat): 3264, 3072, 2959, 1731, 1584, 1543, 1455, 1367, 1333, 1263, 1200, 1111, 1061, 759 and 701  $\text{cm}^{-1}$ .

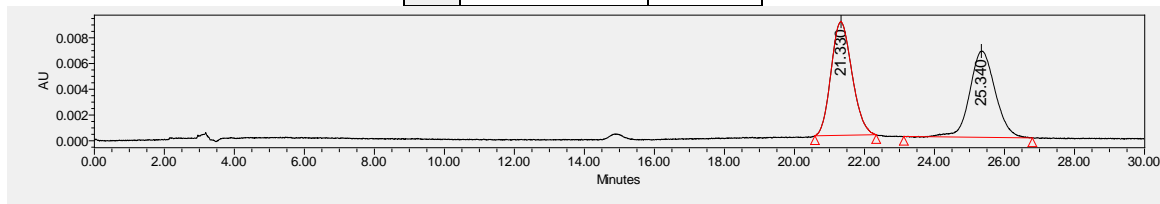
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.35 - 7.30$  (m, 3H), 7.16 – 7.09 (m, 2H), 5.53 (s, 1H), 5.13 (s, 1H), 3.84 (s, 3H), 3.14 (s, 3H), 1.45 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta = 181.3, 166.8, 165.8, 133.8, 129.8, 129.6, 129.0, 129.0, 71.0, 56.2, 55.3, 54.0, 54.0, 52.4, 28.5, 28.5$ .

HRMS (ESI-FT) calcd for  $\text{C}_{18}\text{H}_{23}\text{N}_2\text{O}_5^+$  ( $[\text{M}+\text{H}^+]$ ) = 347.1601, Found 347.1600.



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 21.226         | 89.72  |
| 2 | 25.384         | 10.28  |



|   | Retention Time | % Area |
|---|----------------|--------|
| 1 | 21.330         | 51.90  |
| 2 | 25.340         | 48.10  |



## 9. Copies of NMR spectra

Figure S1.  $^1\text{H}$  NMR of 4a

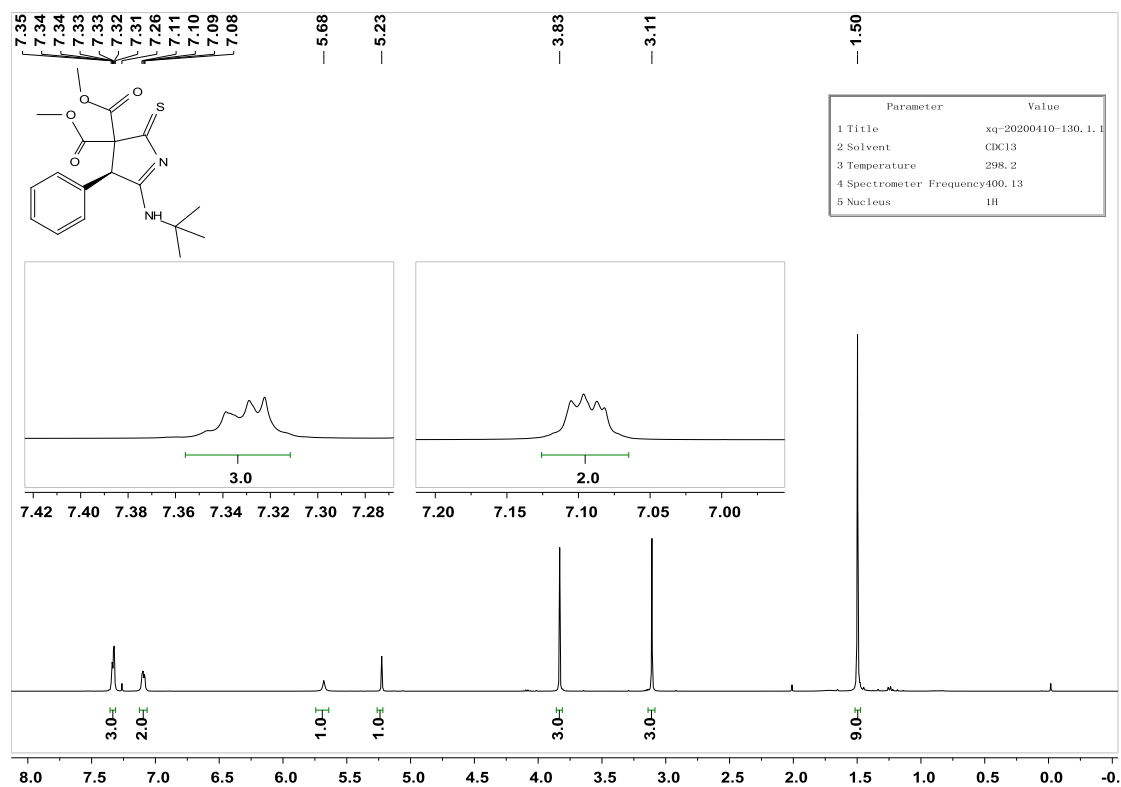


Figure S2.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4a

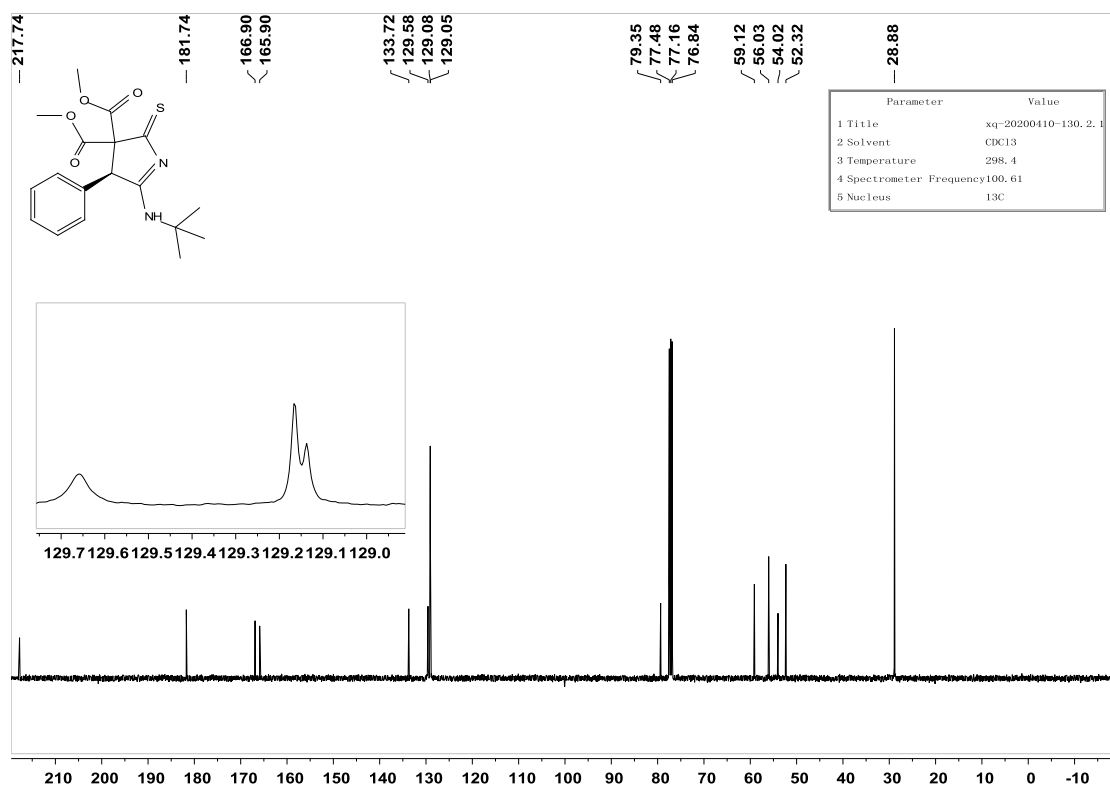


Figure S3.  $^1\text{H}$  NMR of 4b

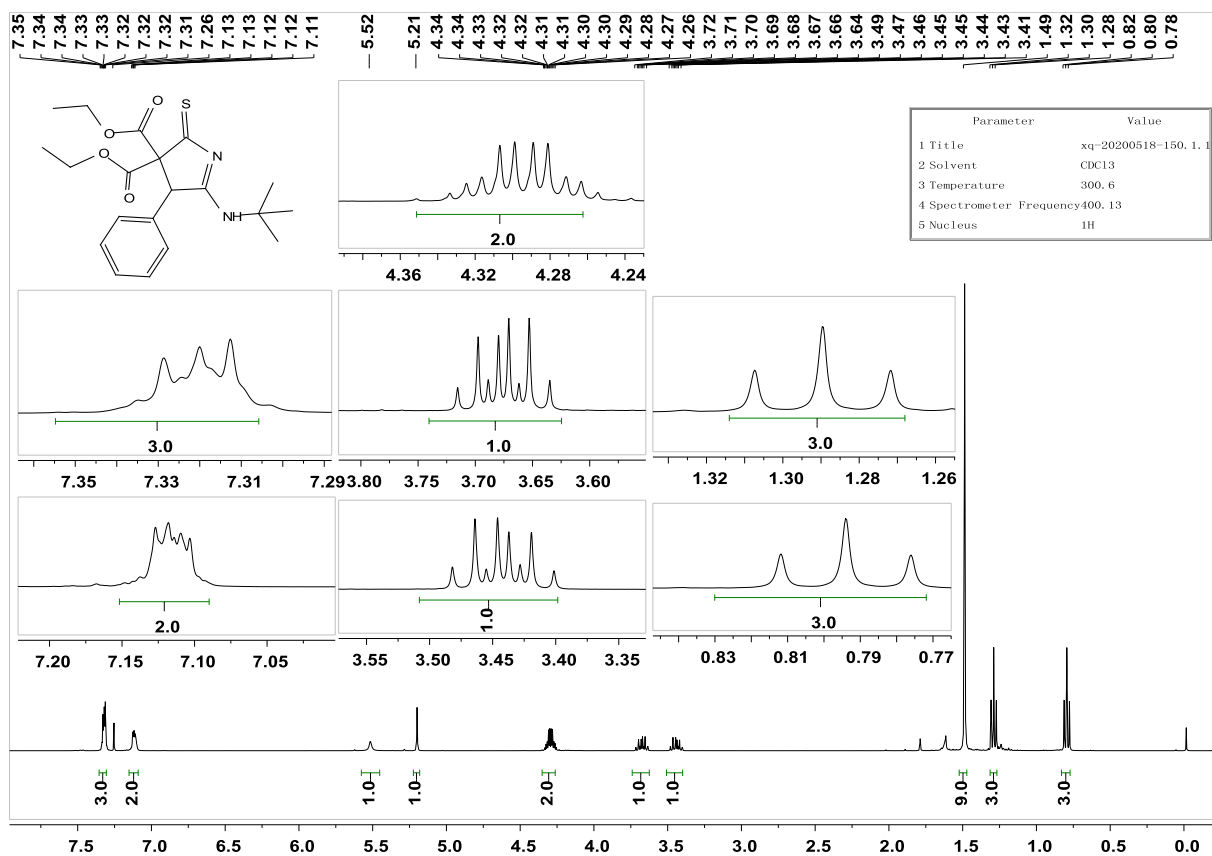


Figure S4.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4b

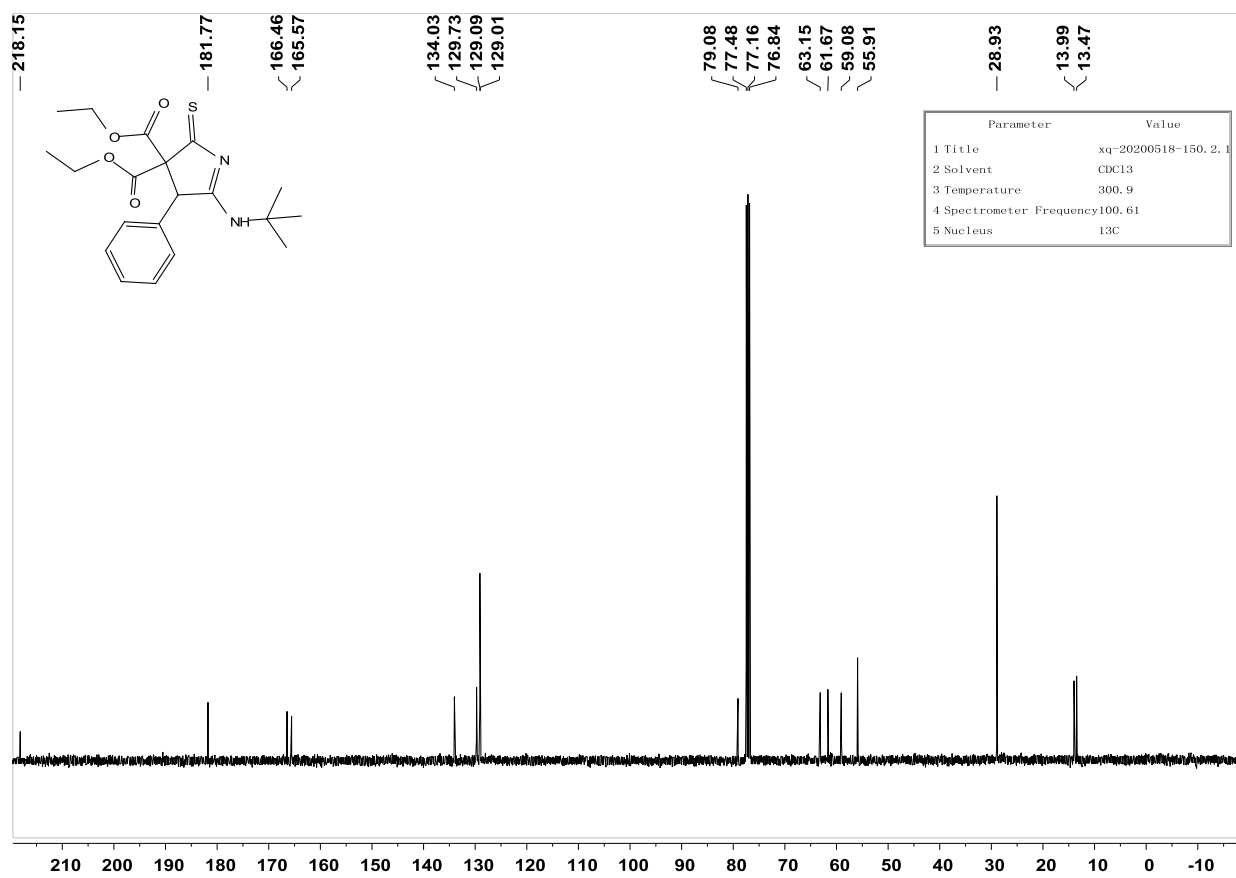


Figure S5.  $^1\text{H}$  NMR of 4c

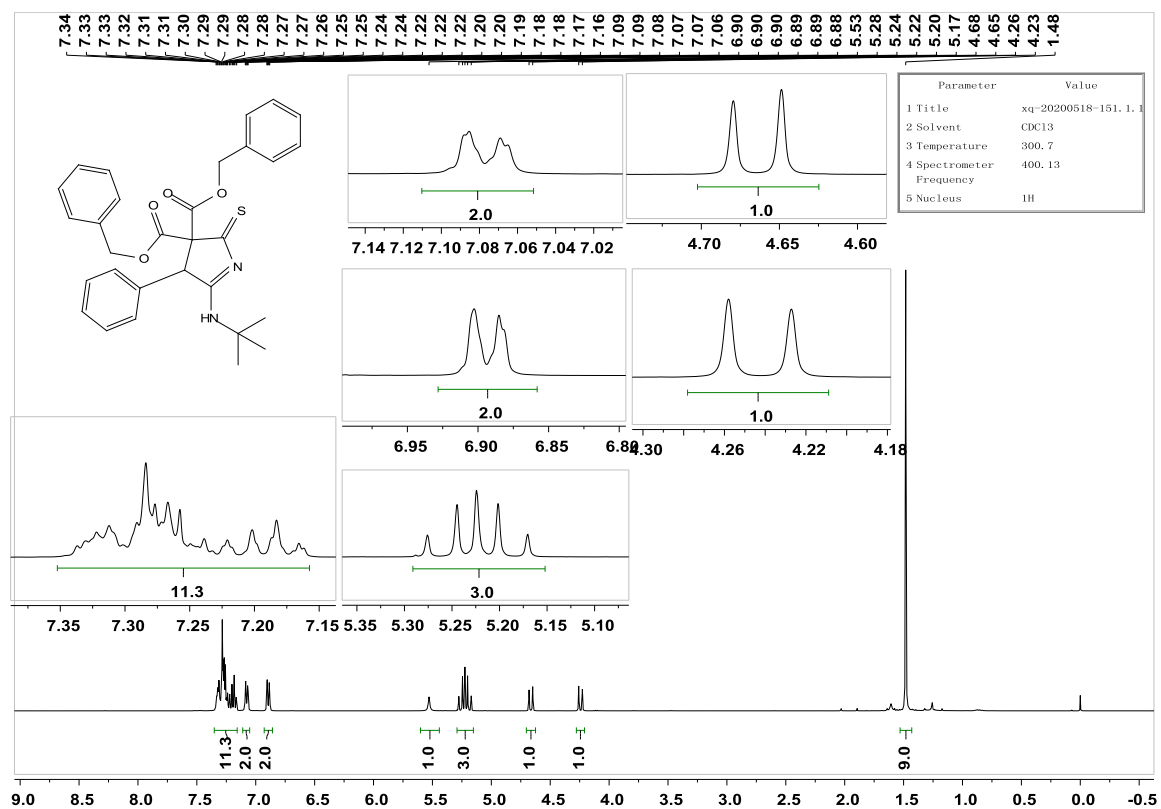


Figure S6.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4c

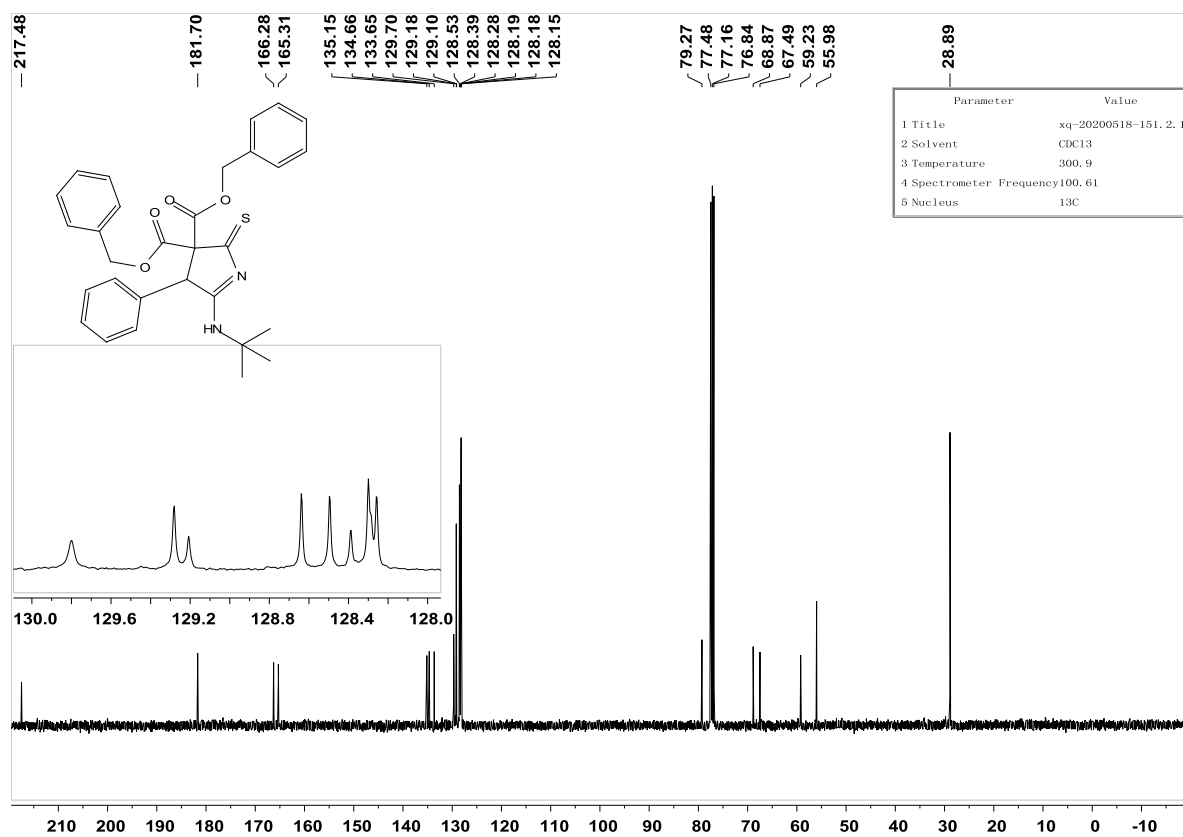


Figure S7.  $^1\text{H}$  NMR of 4d

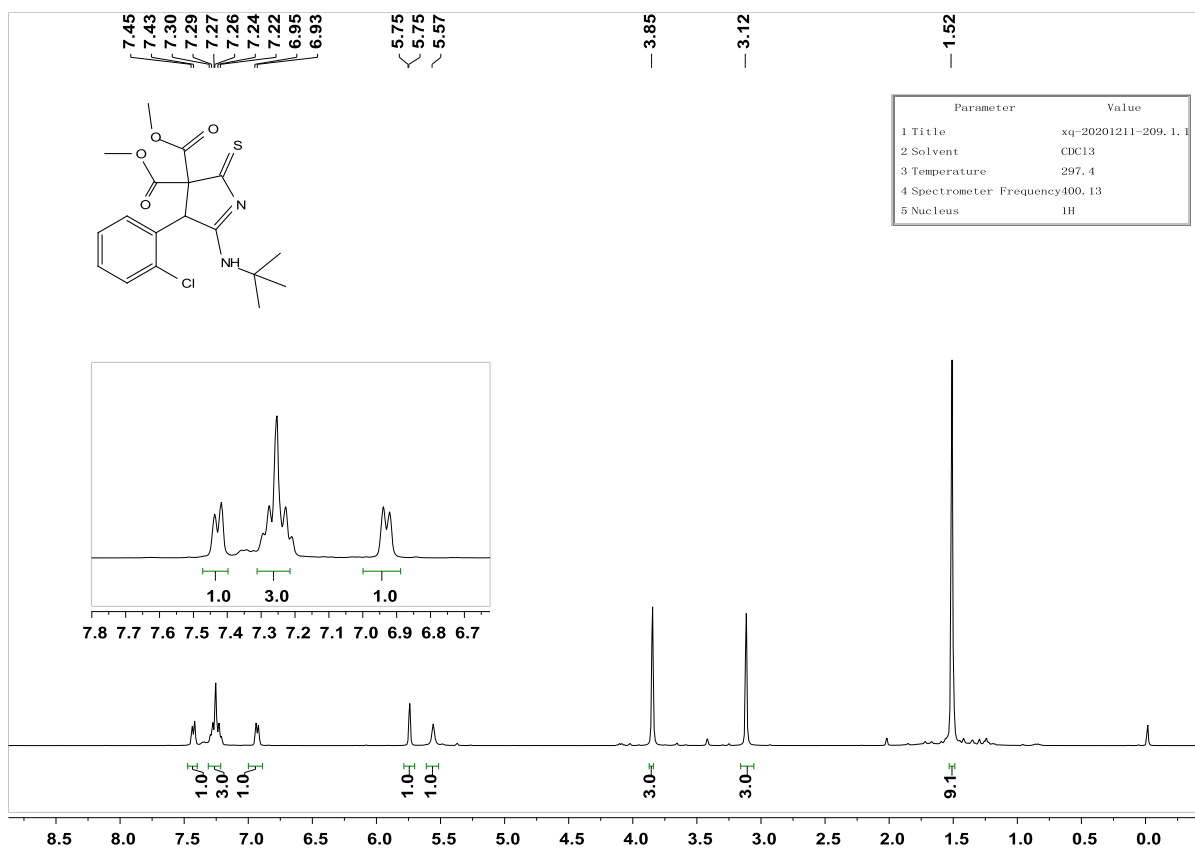


Figure S8.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4d

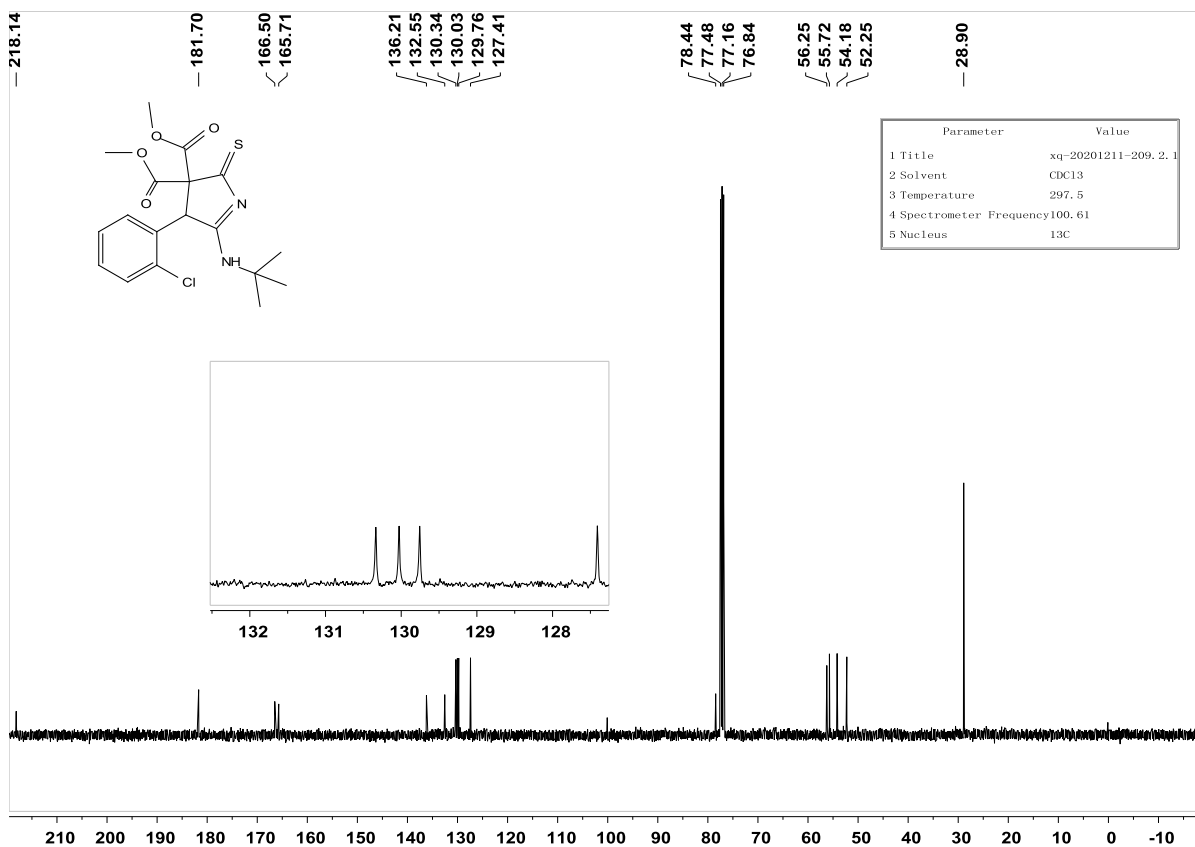


Figure S9.  $^1\text{H}$  NMR of 4e

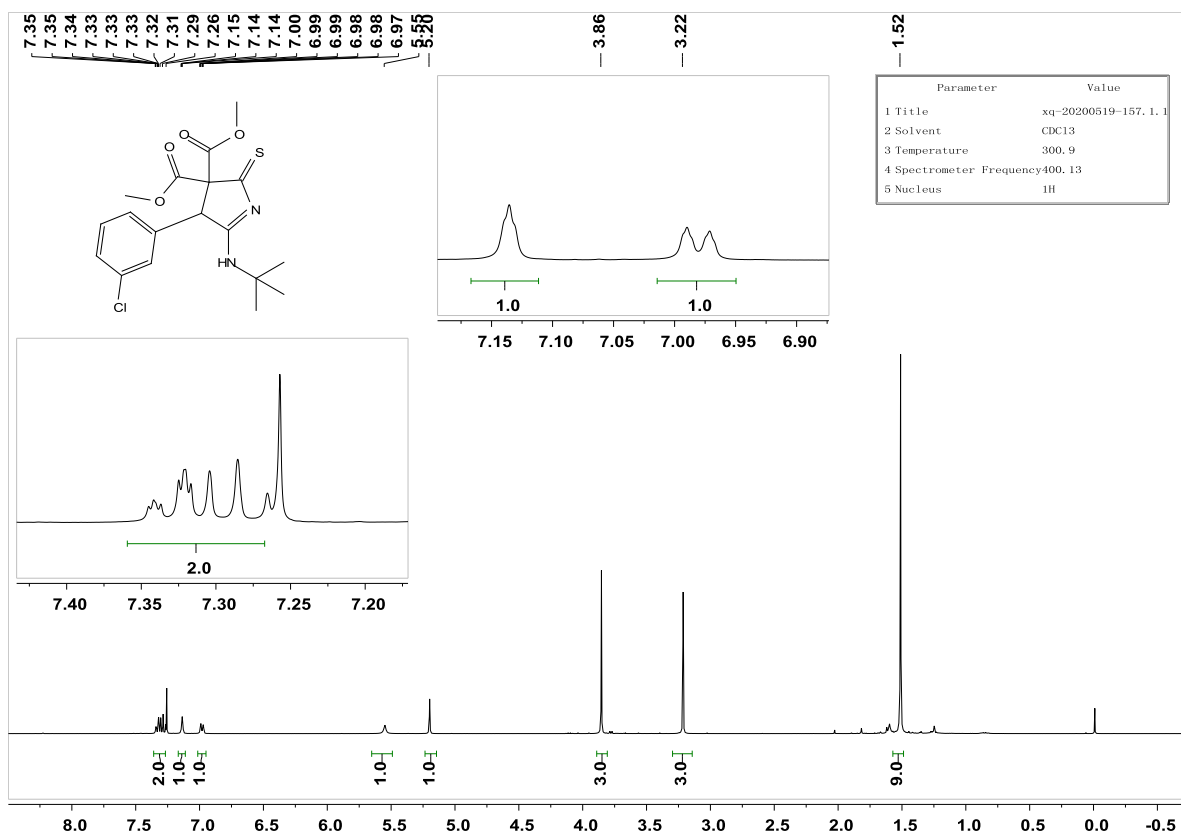


Figure S10.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4e

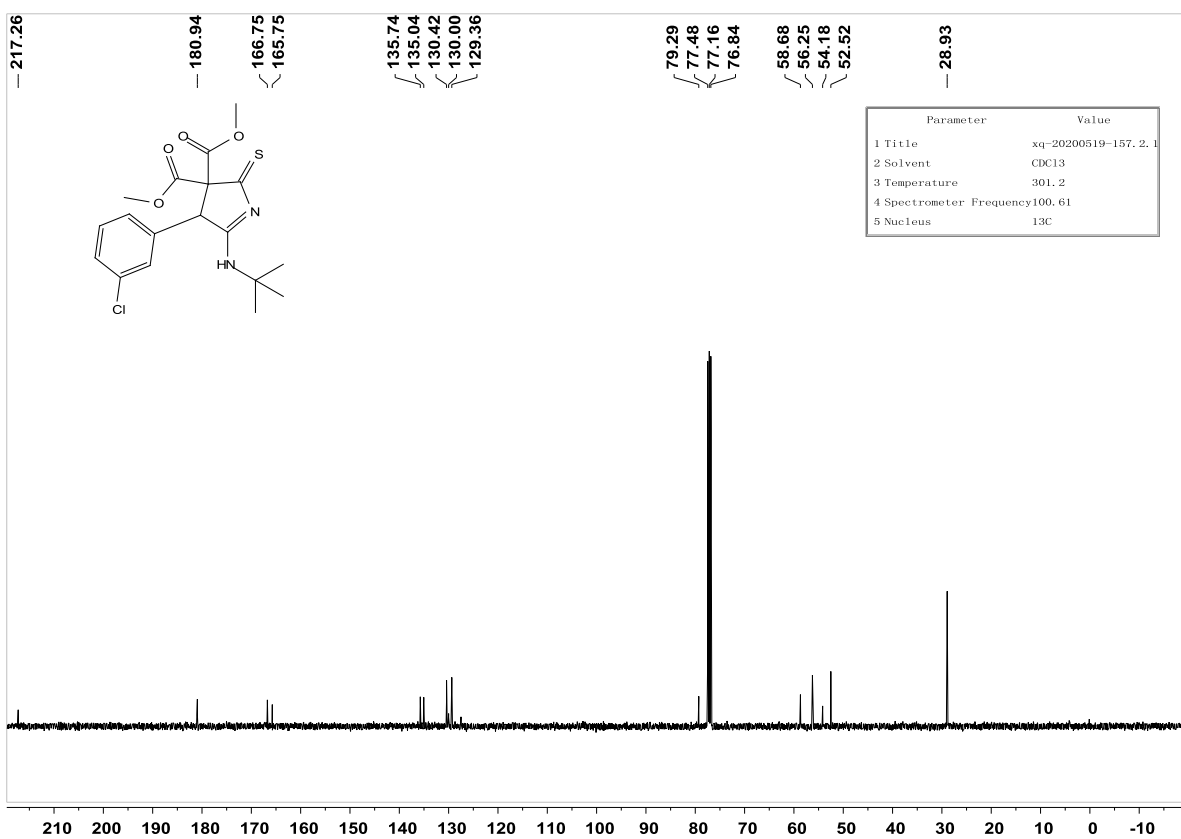


Figure S11.  $^1\text{H}$  NMR of 4f

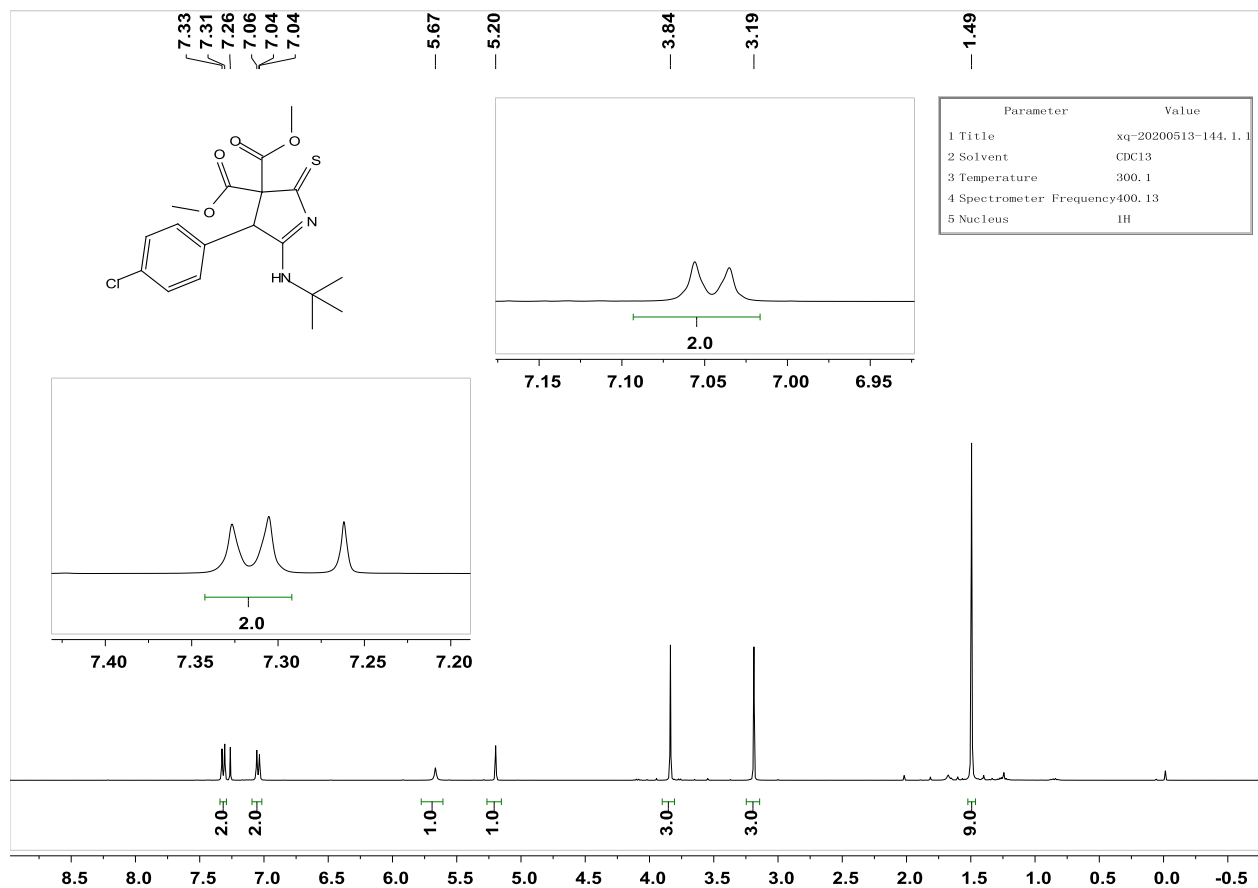


Figure S12.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4f

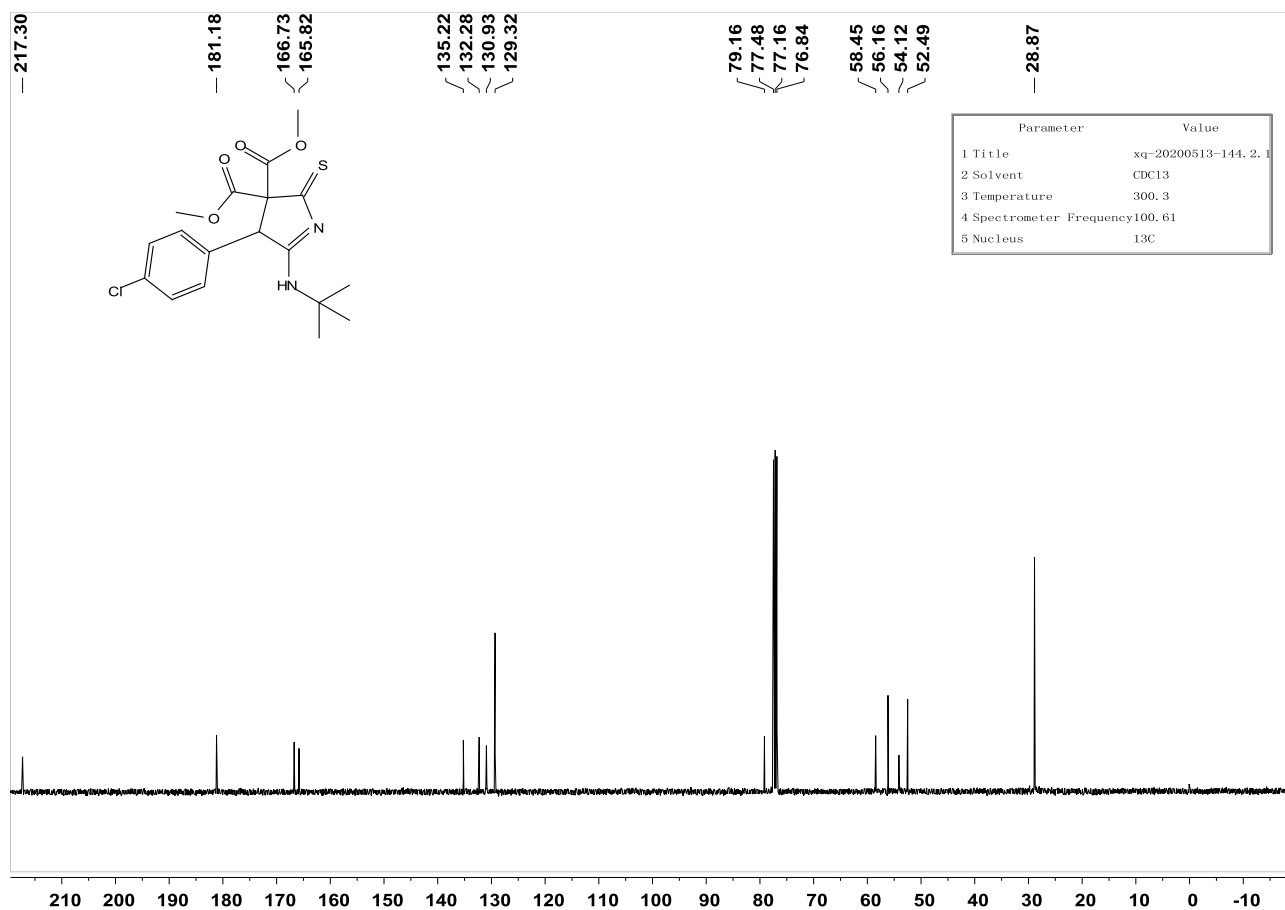


Figure S13.  $^1\text{H}$  NMR of 4g

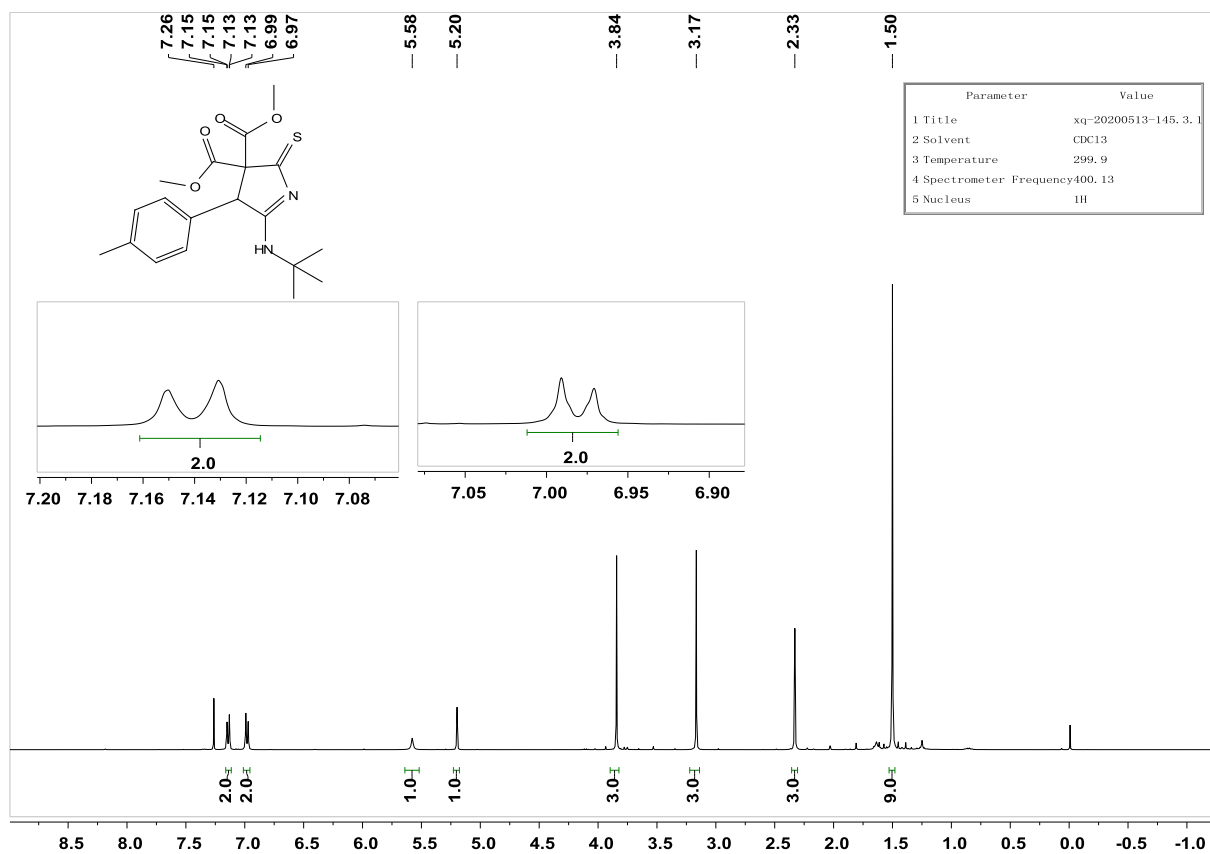


Figure S14.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4g

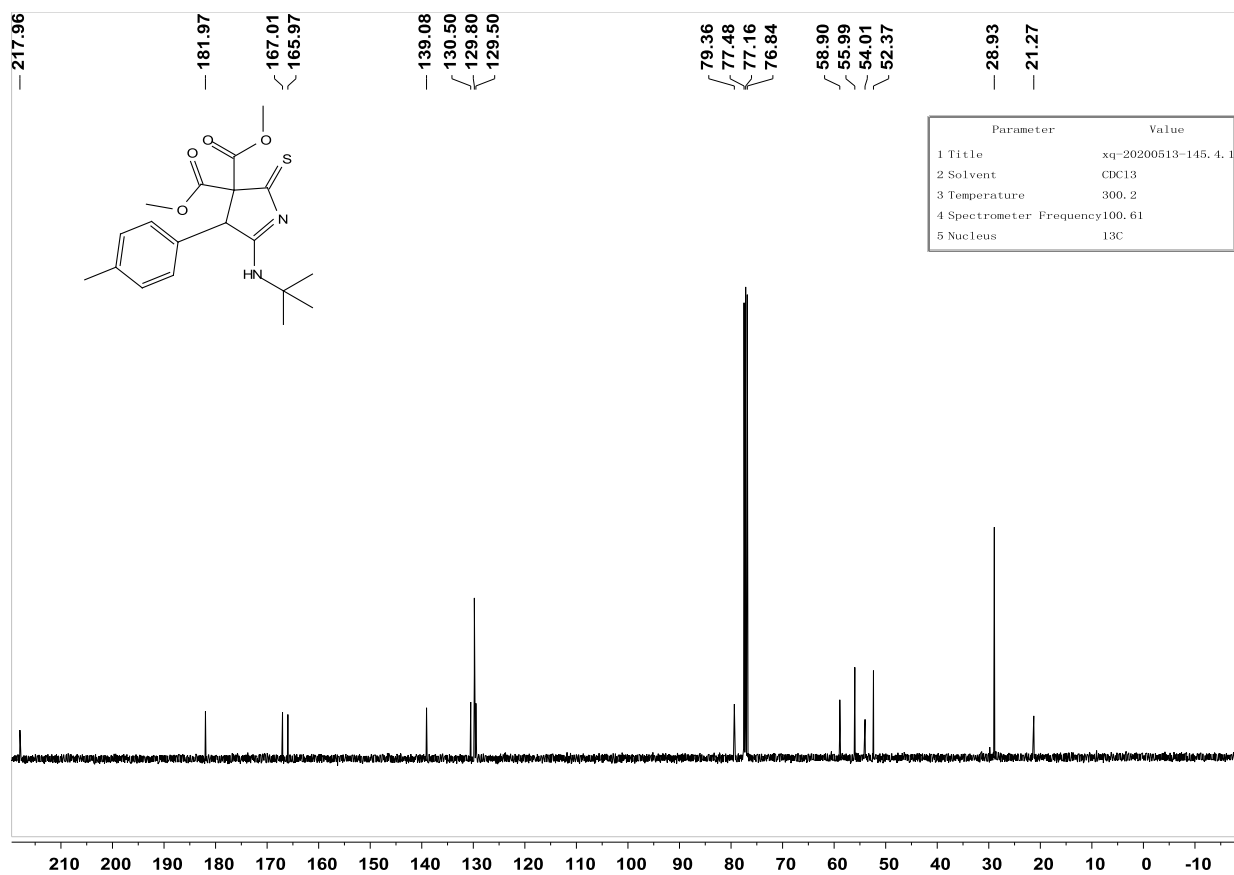


Figure S15.  $^1\text{H}$  NMR of 4h

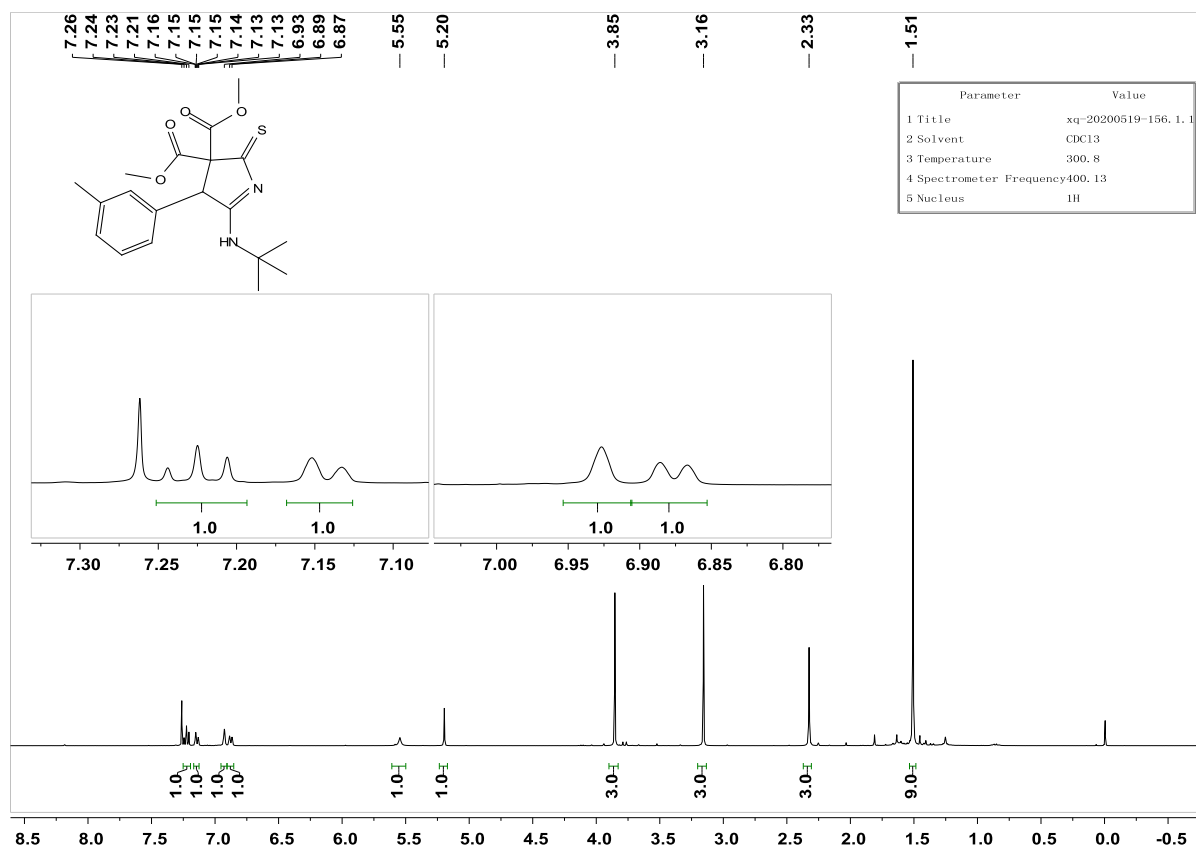


Figure S16.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4h

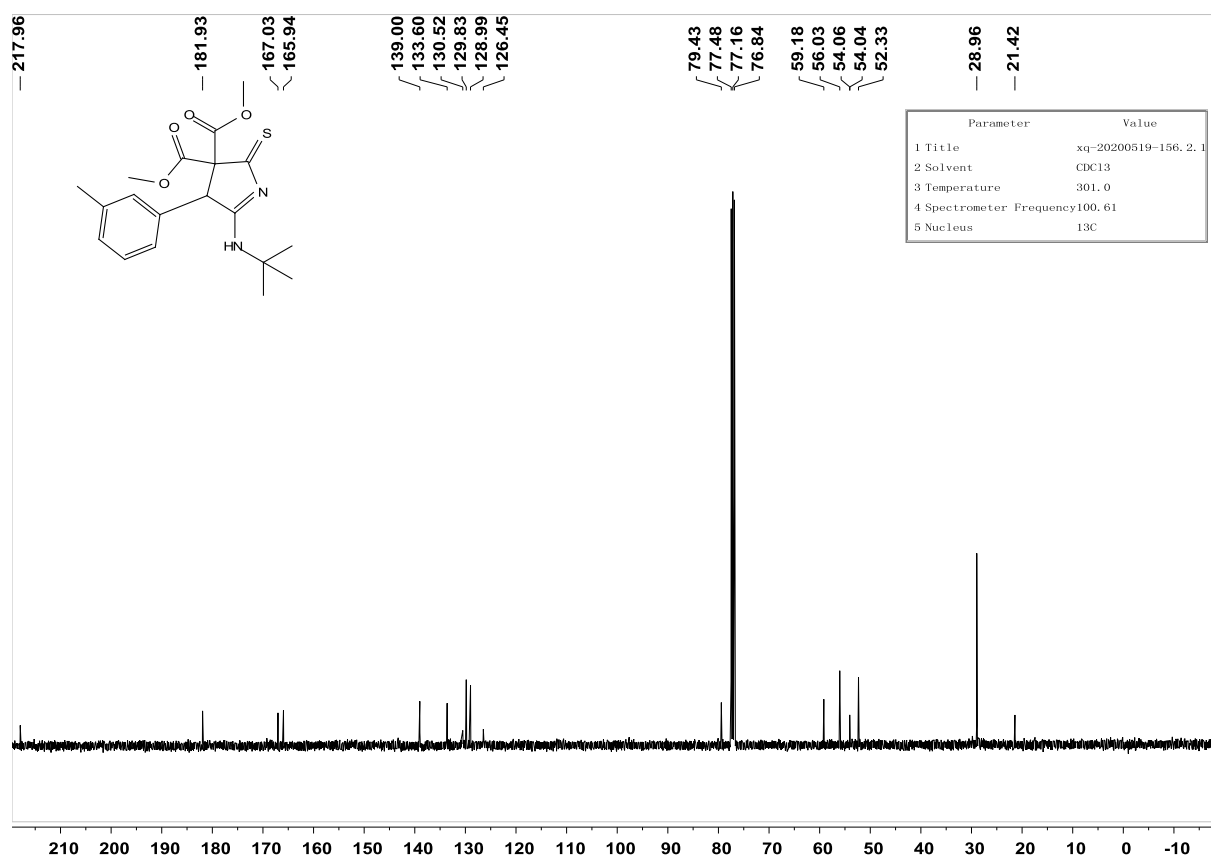




Figure S17.  $^1\text{H}$  NMR of 4i

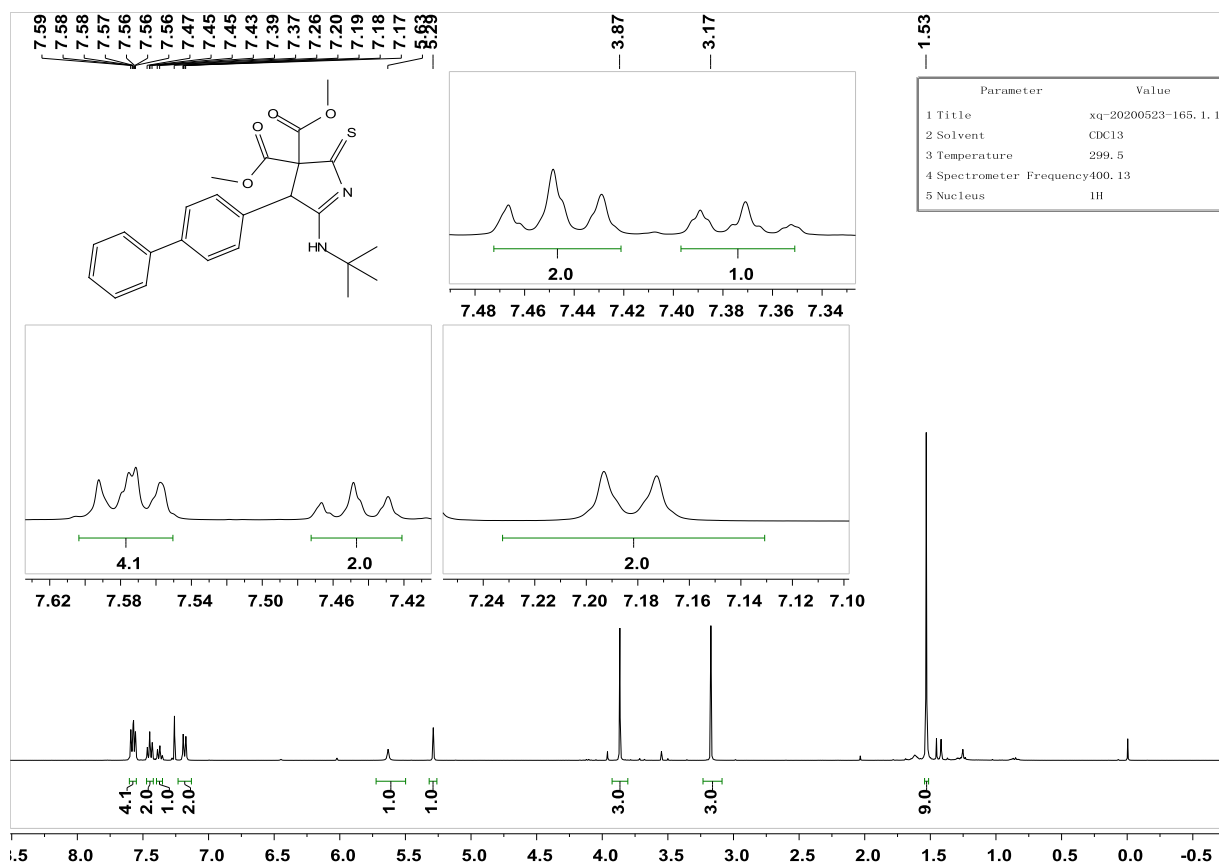


Figure S18.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4i

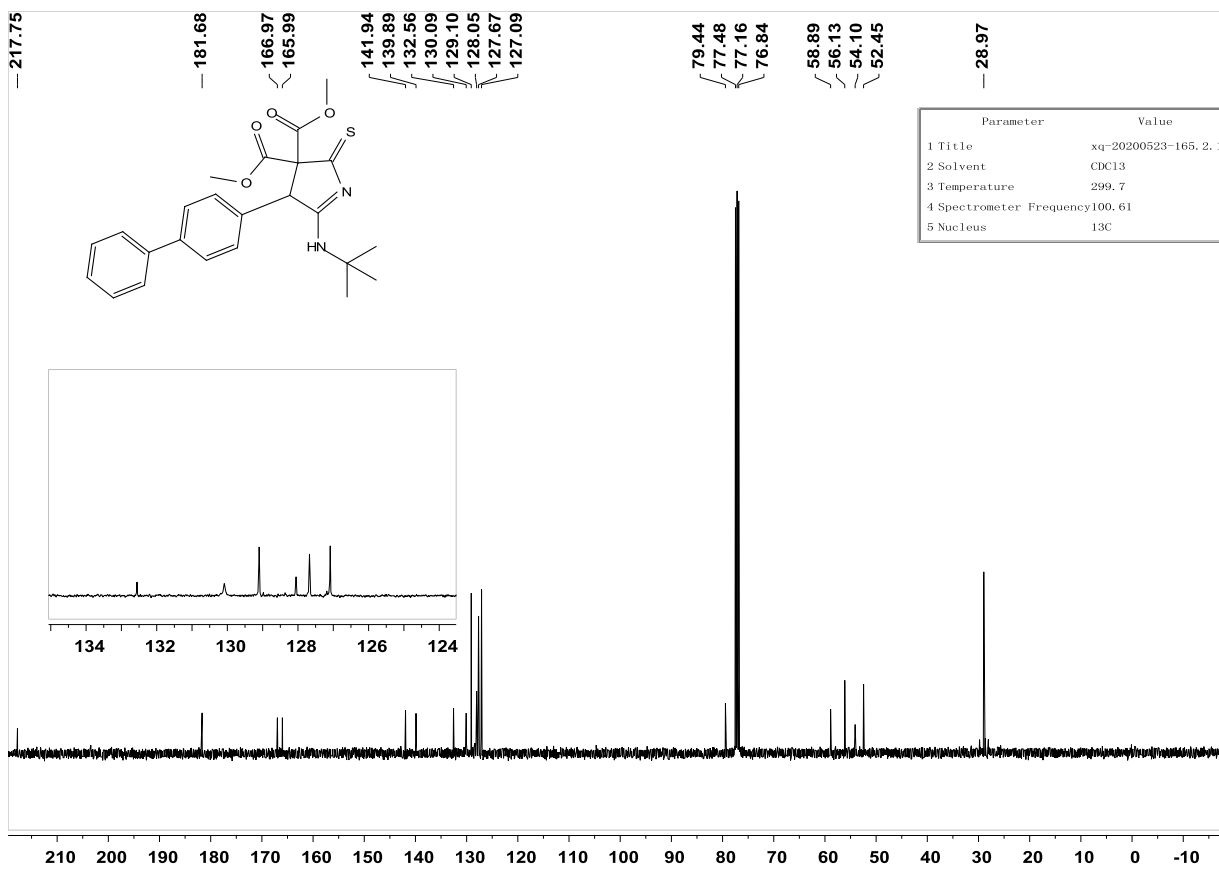


Figure S19.  $^1\text{H}$  NMR of 4j

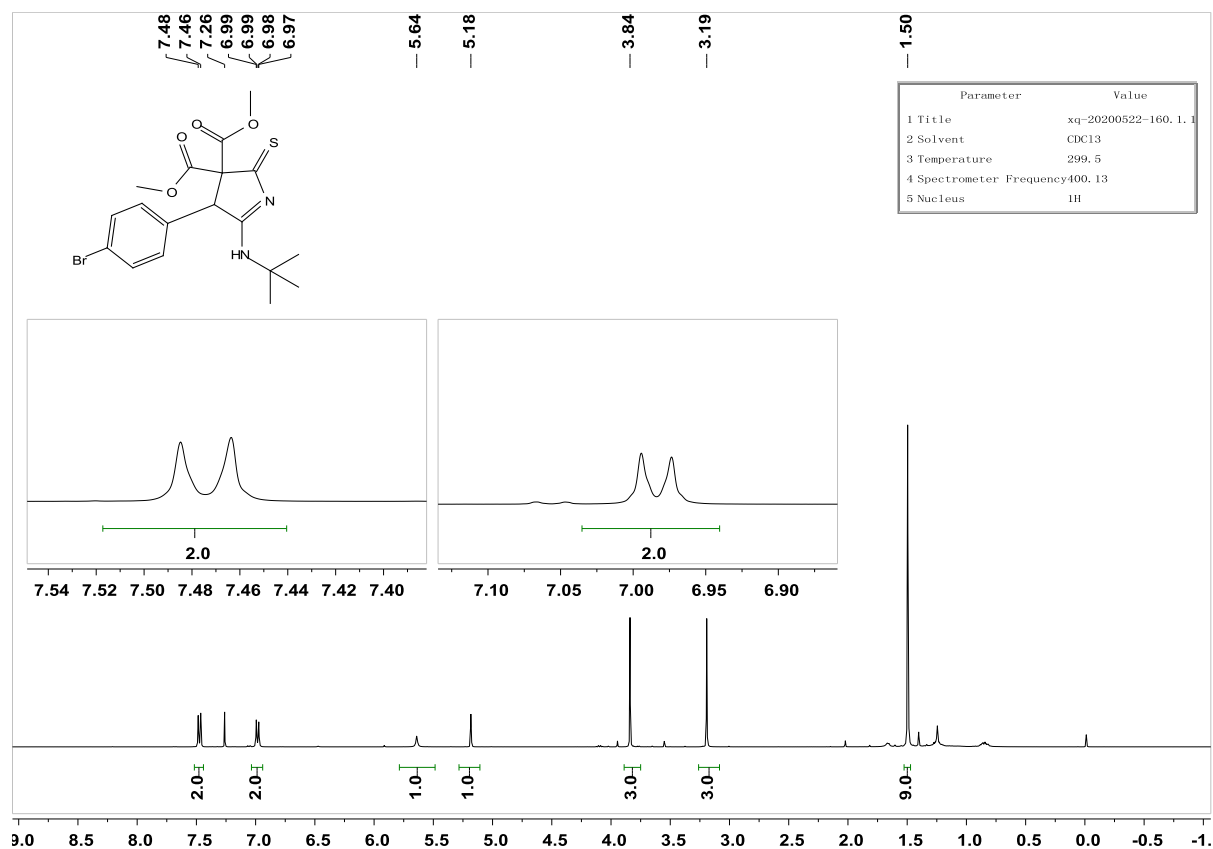


Figure S20.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4j

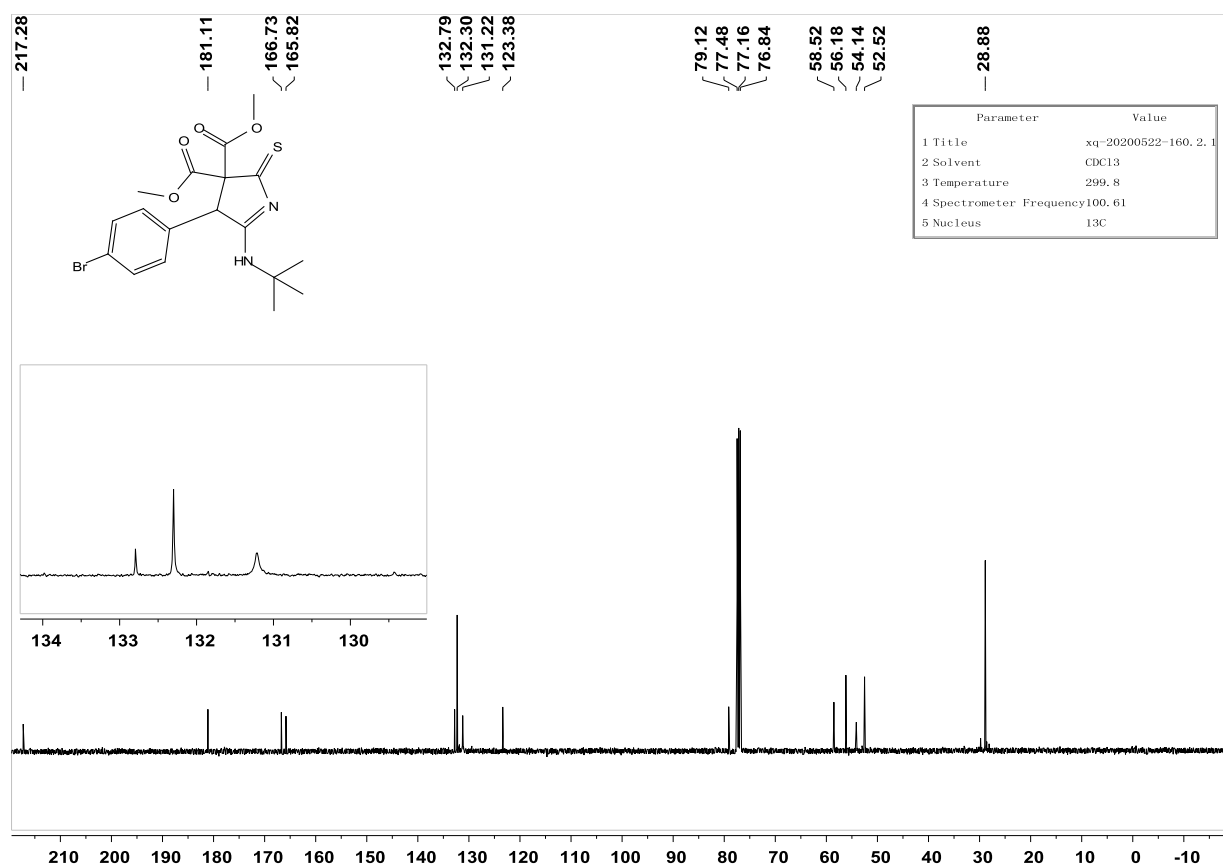


Figure S21.  $^1\text{H}$  NMR of 4k

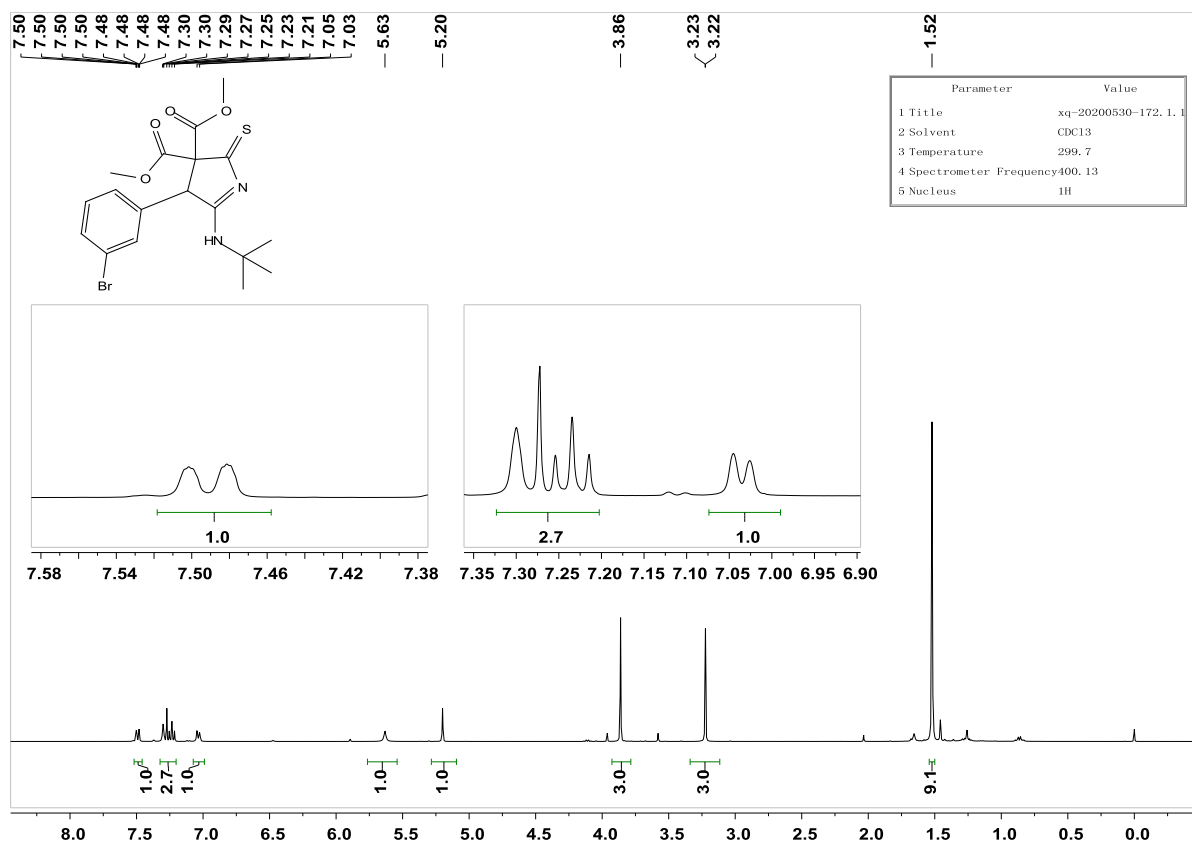


Figure S22.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4k

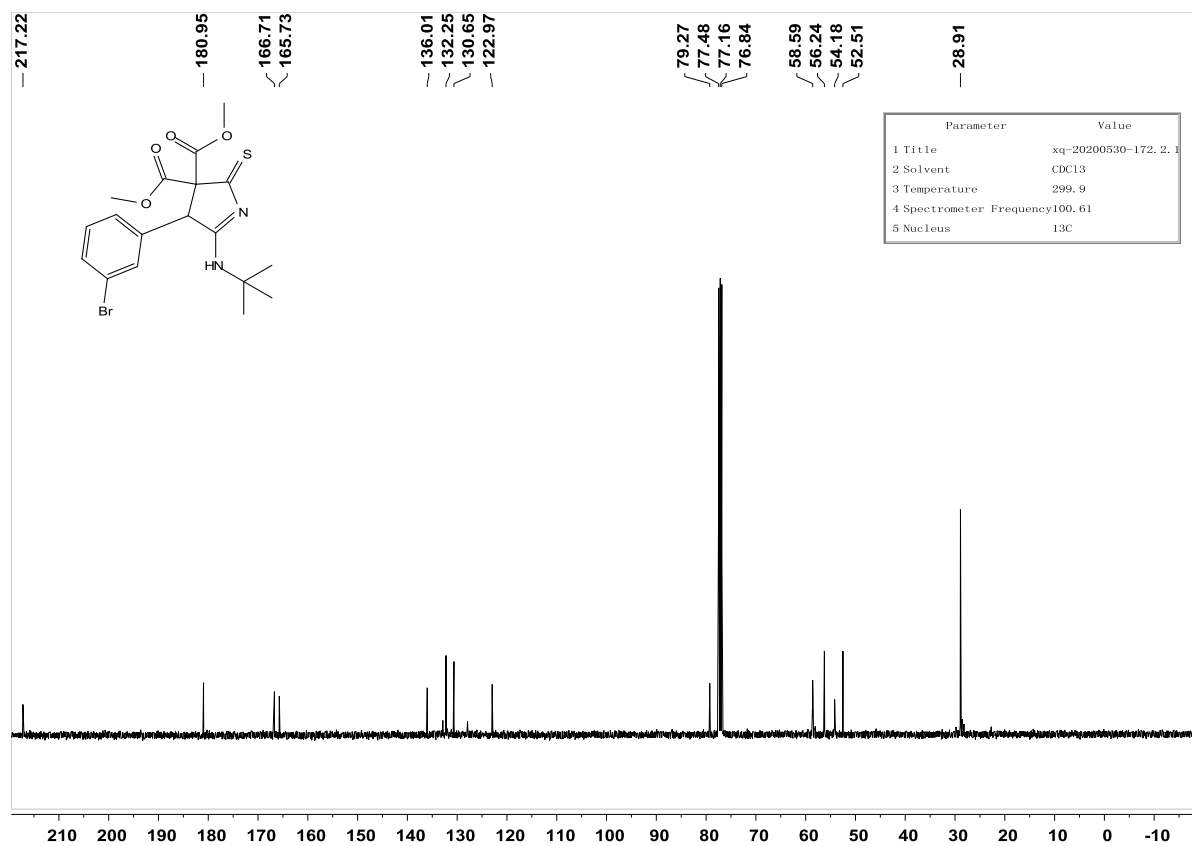


Figure S23.  $^1\text{H}$  NMR of 4l

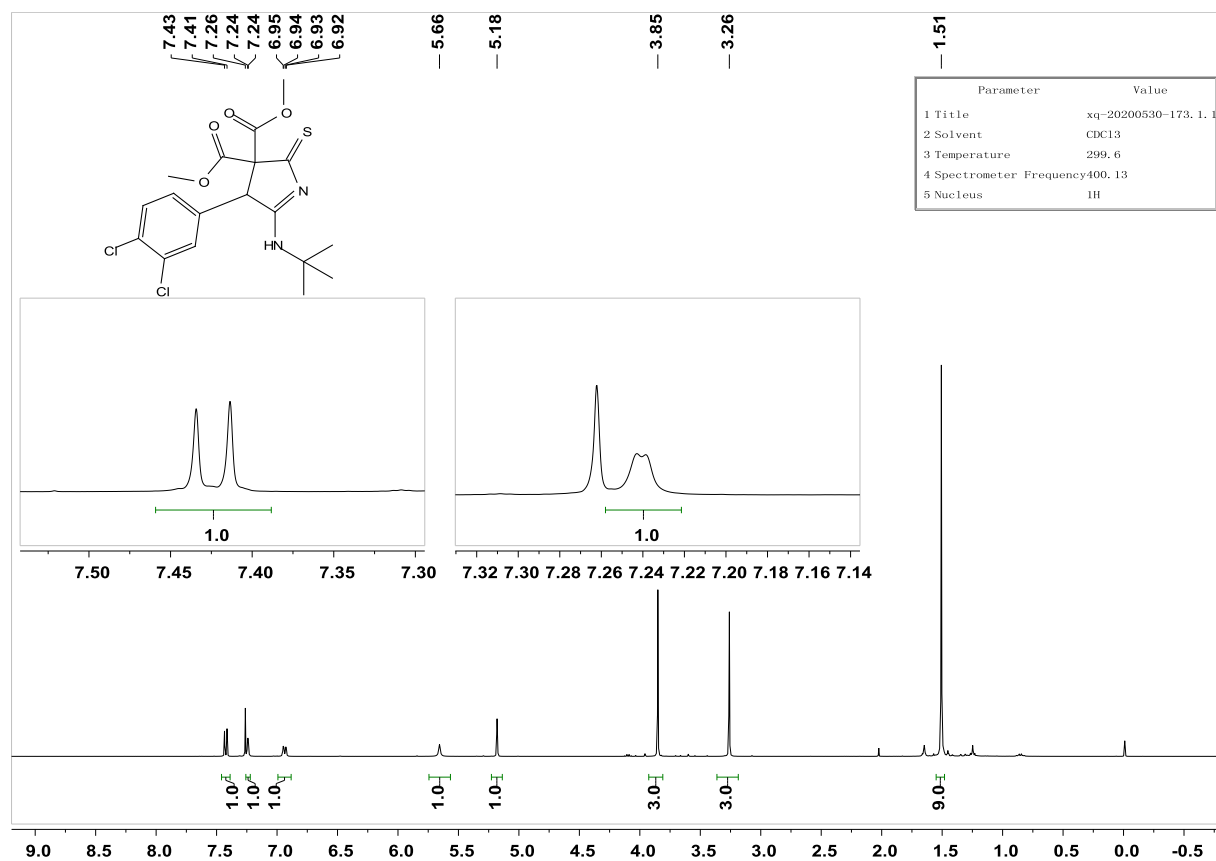


Figure S24.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4l

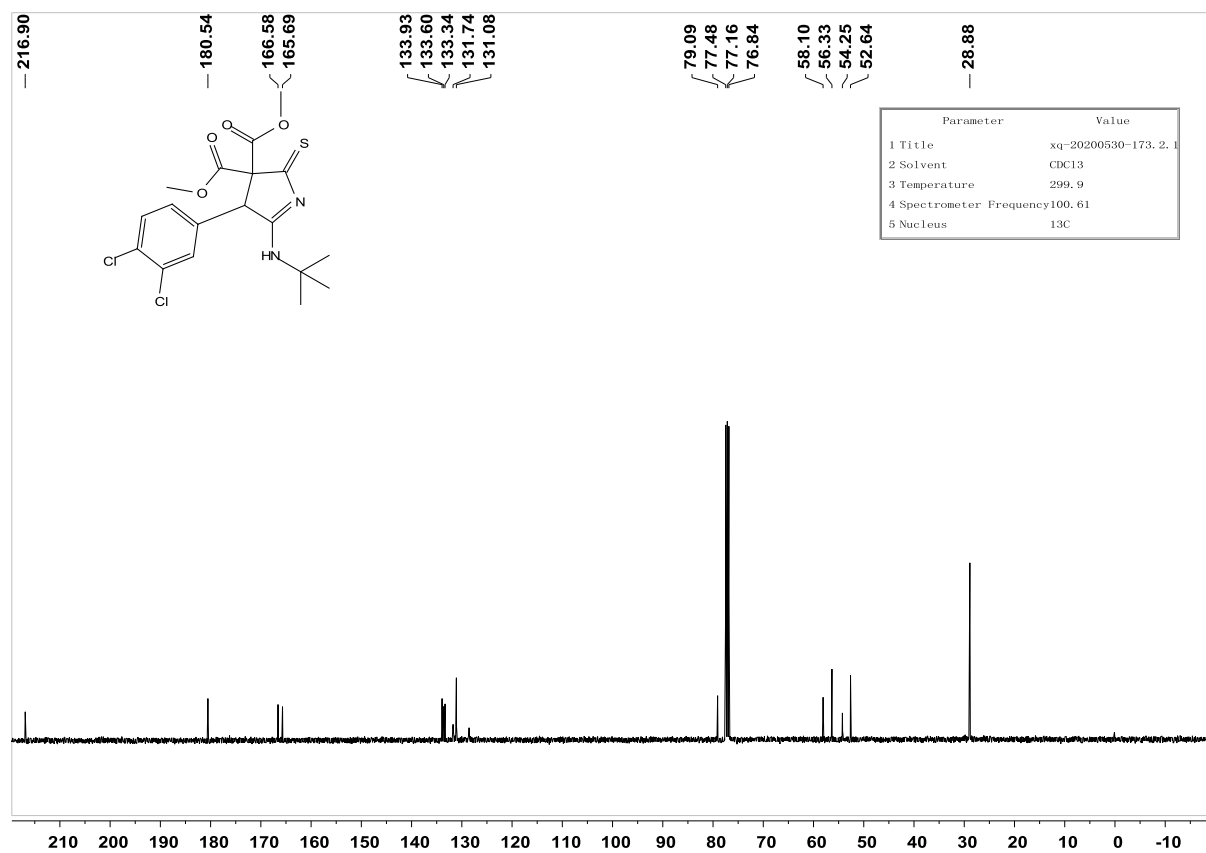


Figure S25.  $^1\text{H}$  NMR of 4m

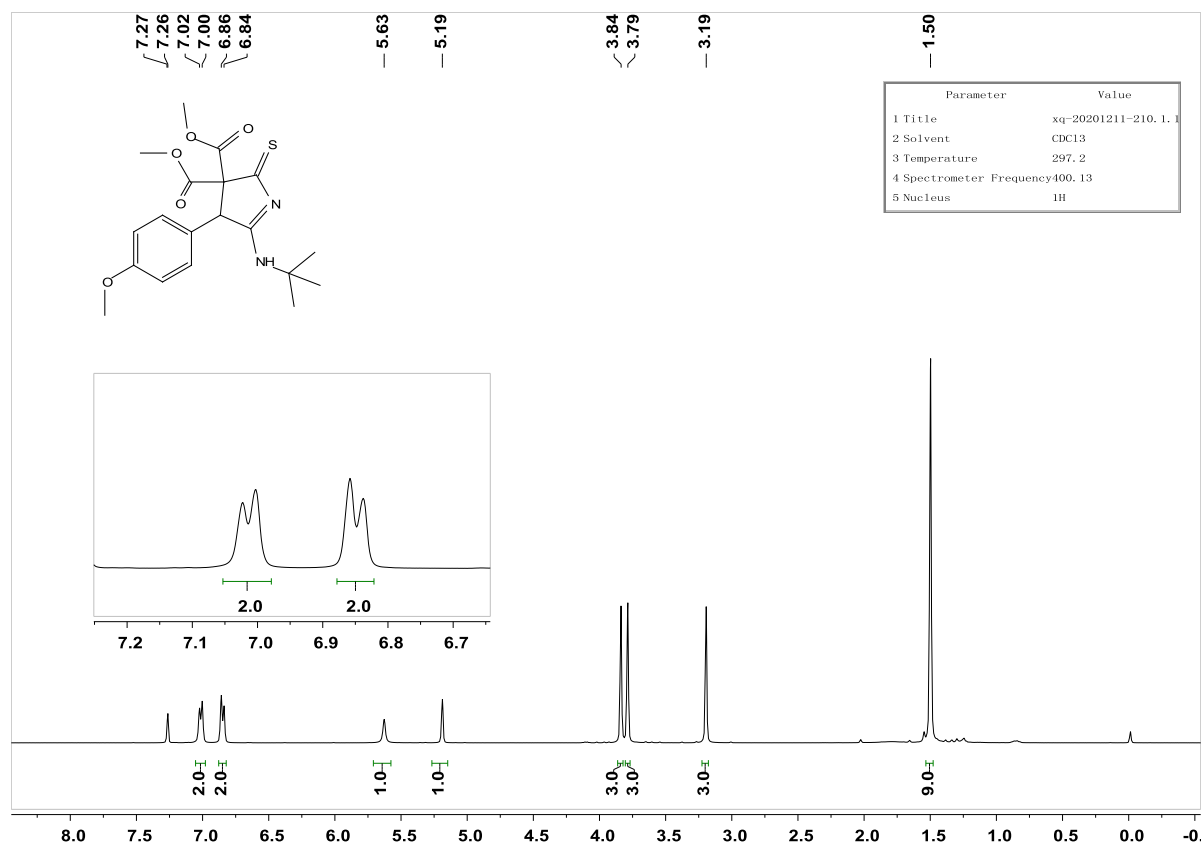


Figure S26.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4m

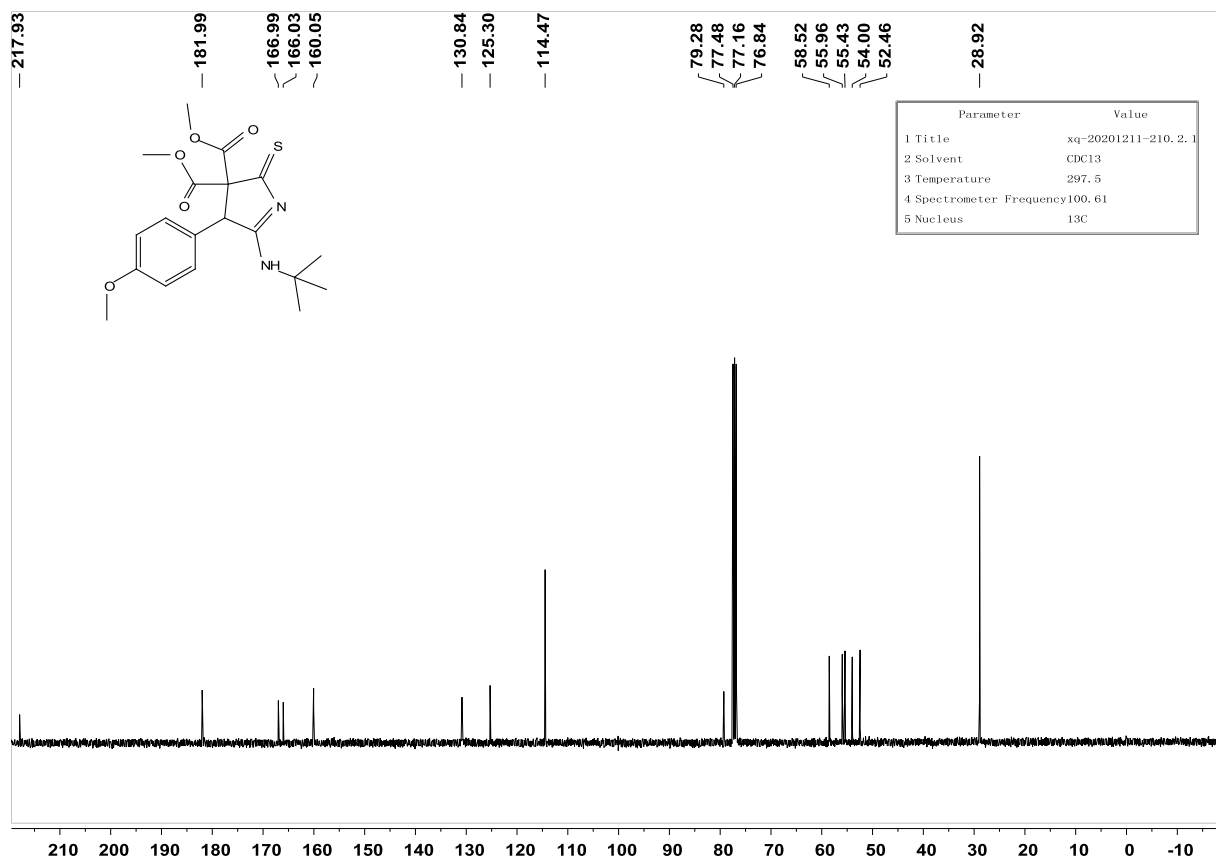


Figure S27.  $^1\text{H}$  NMR of 4n

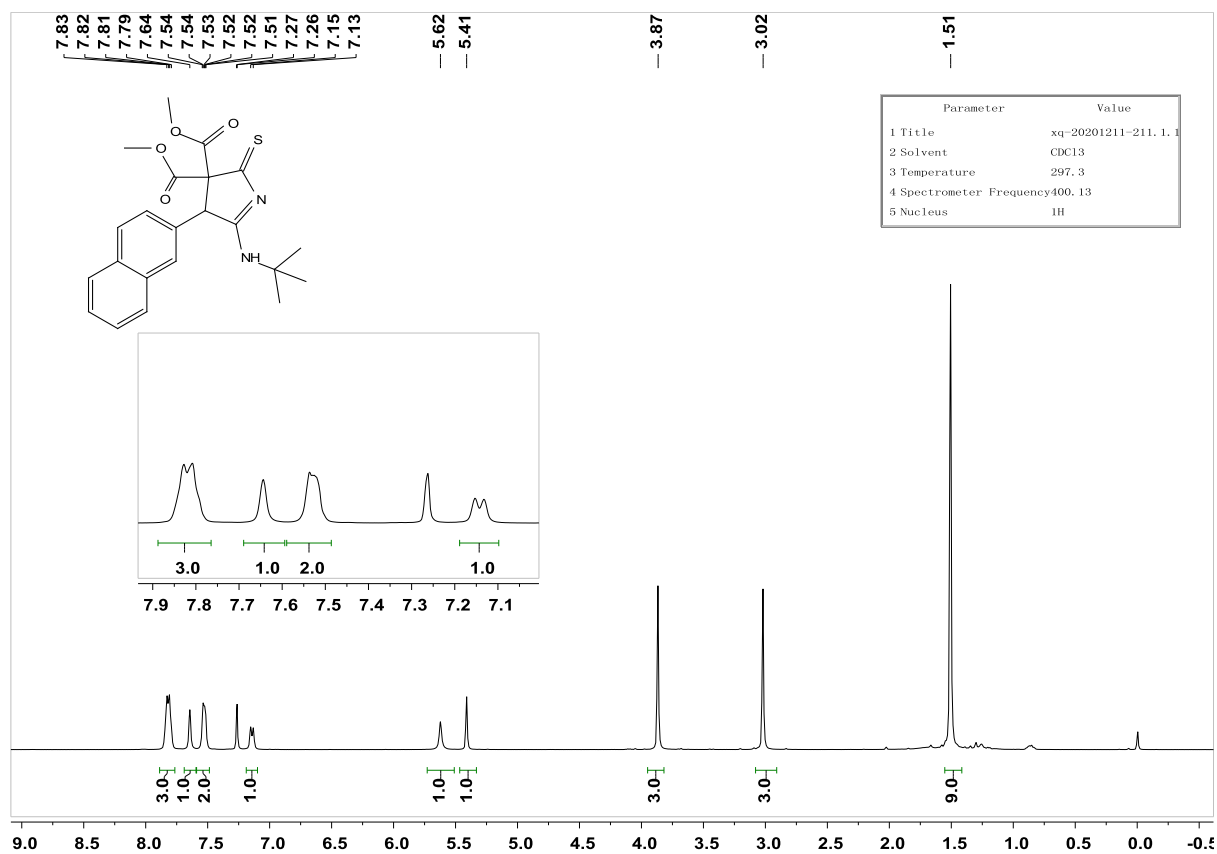


Figure S28.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4n

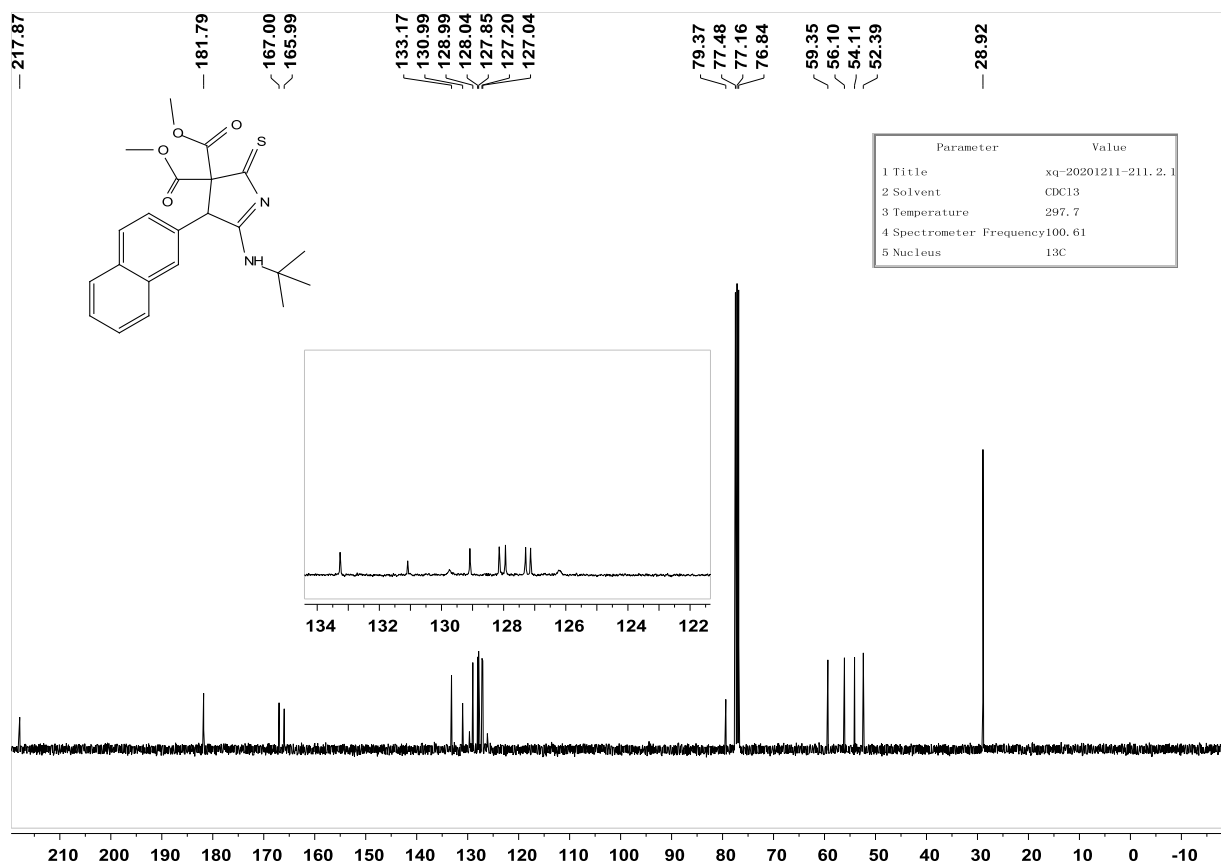


Figure S29.  $^1\text{H}$  NMR of 4o

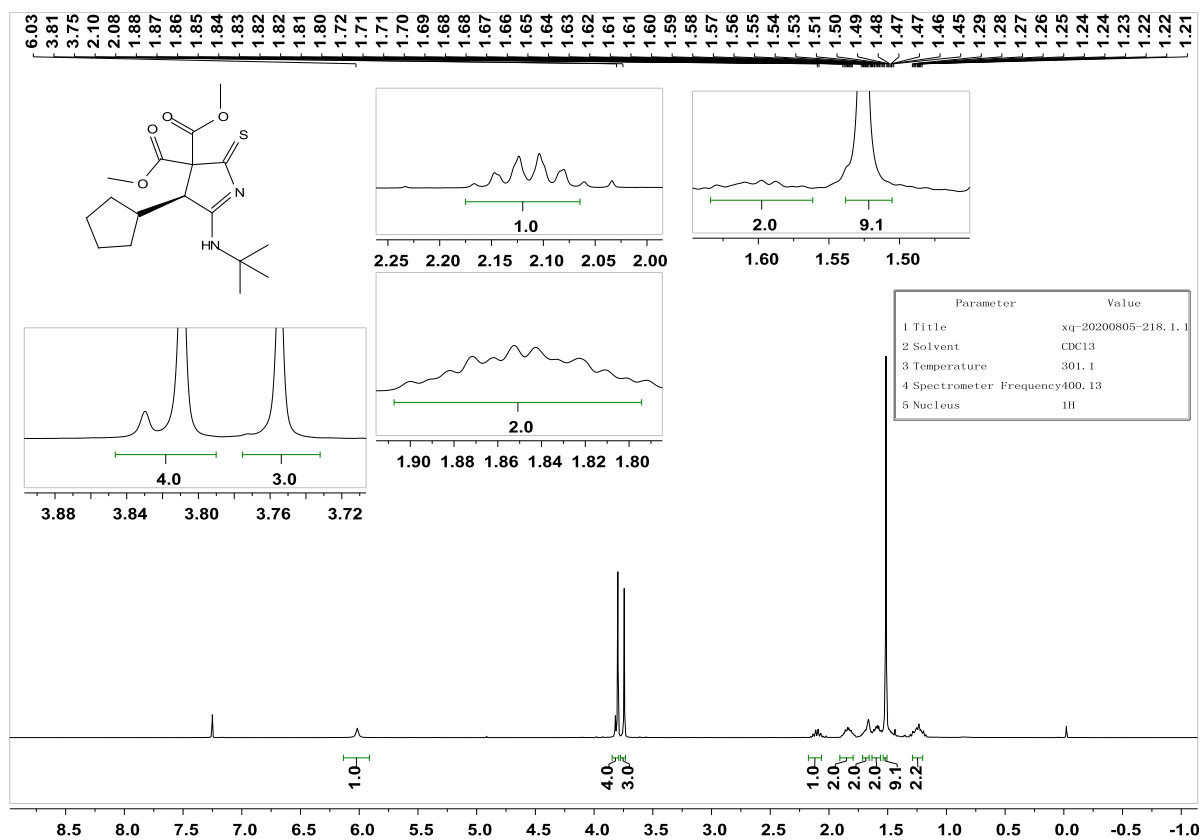


Figure S30.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4o

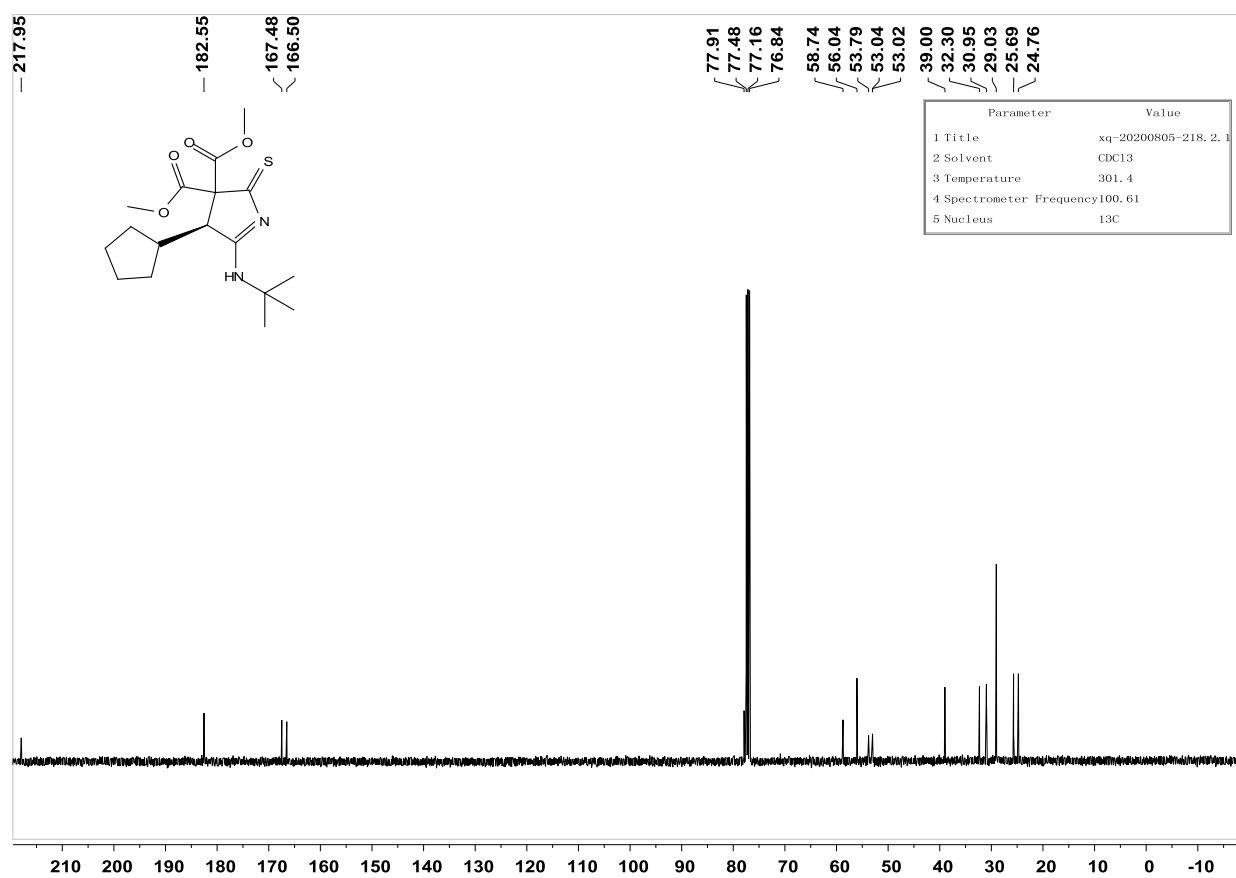


Figure S31.  $^1\text{H}$  NMR of 4p

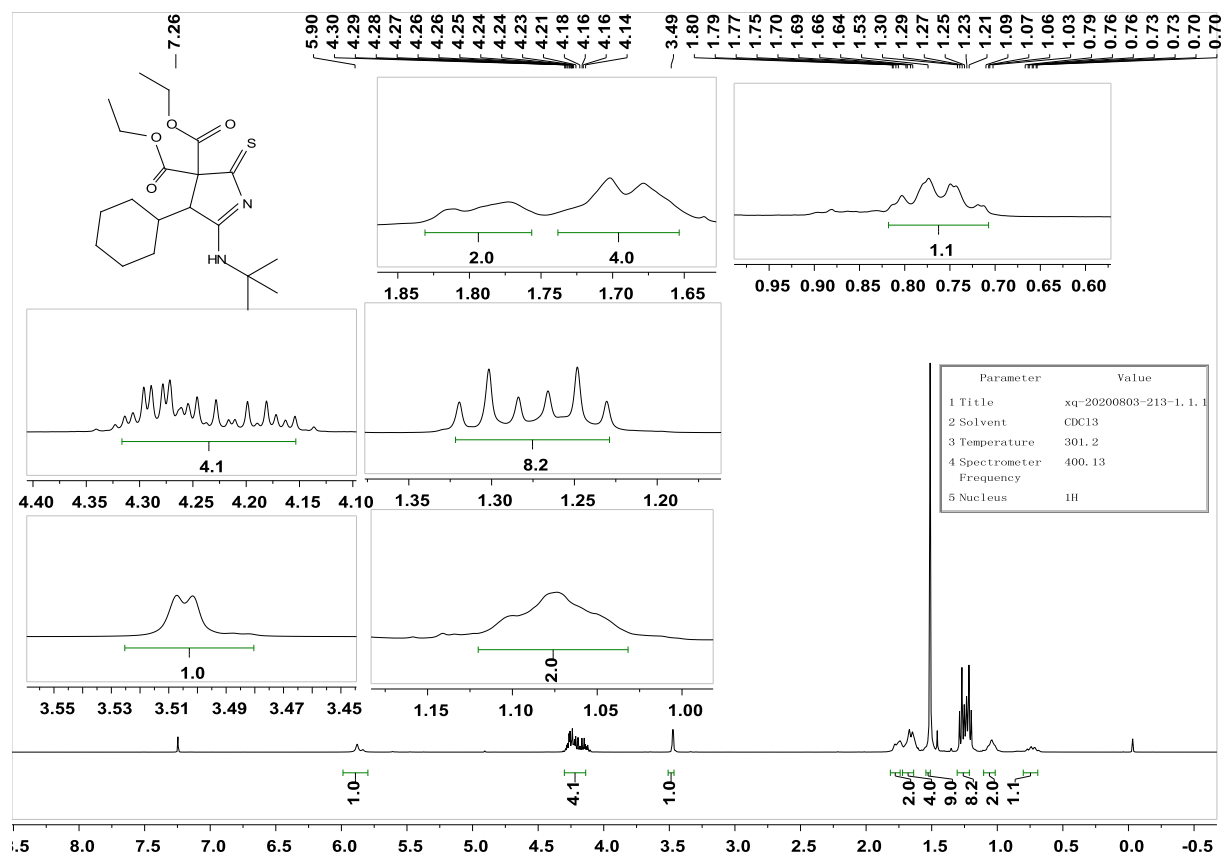


Figure S32.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4p

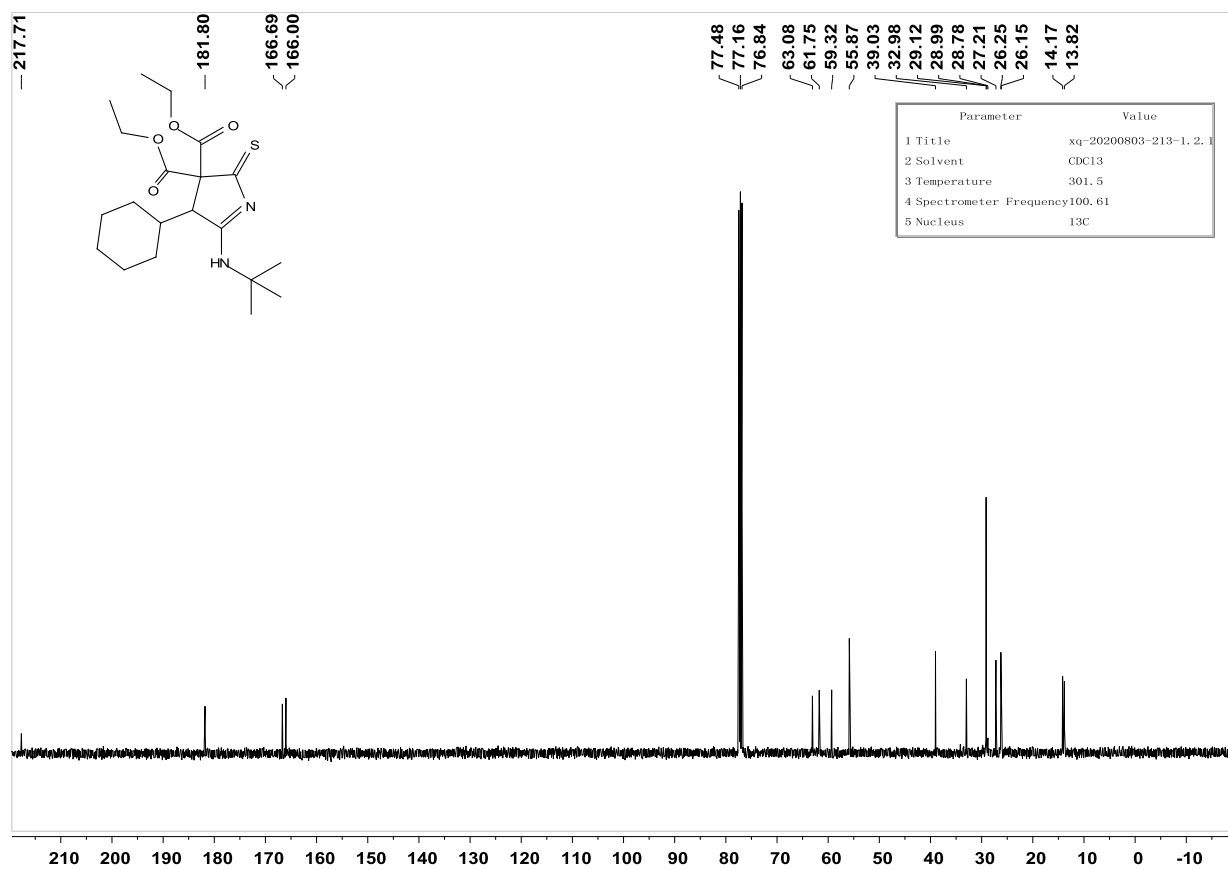




Figure S33.  $^1\text{H}$  NMR of 4q

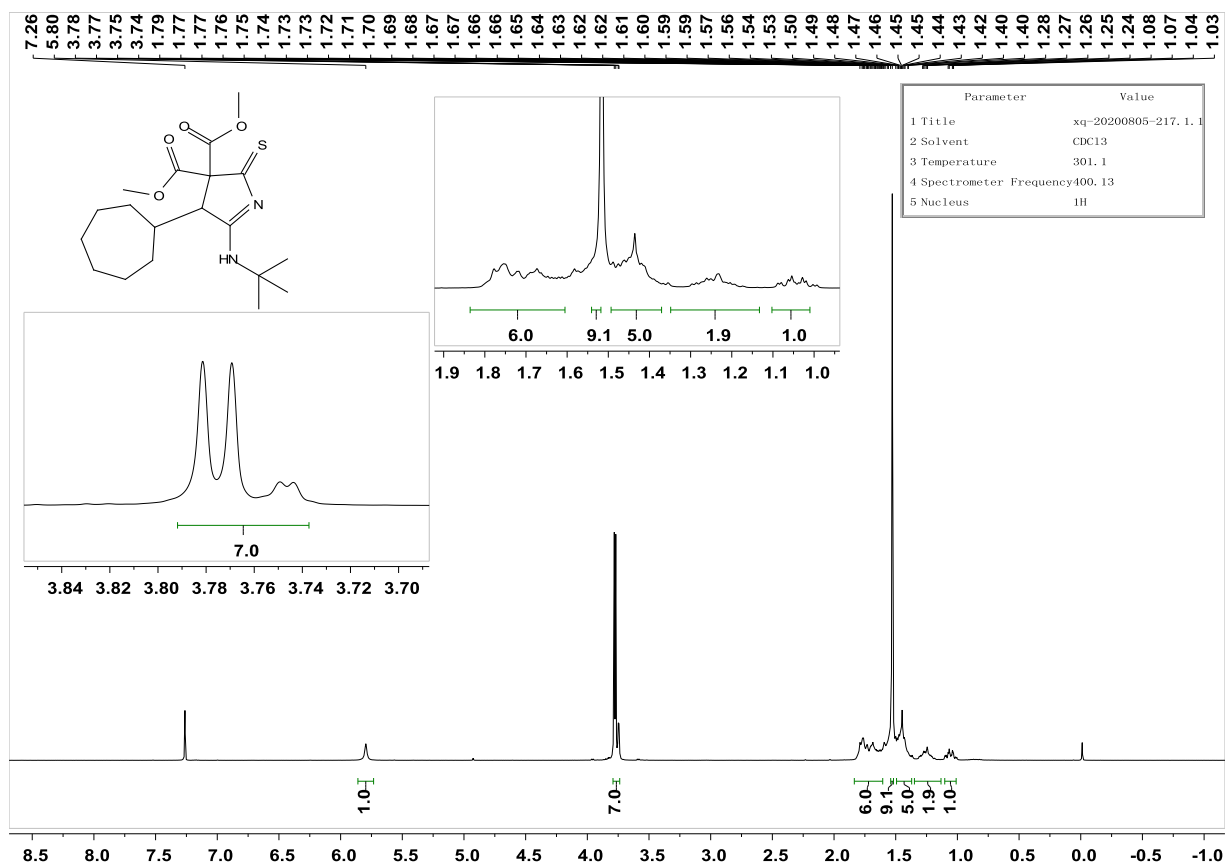


Figure S34.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4q

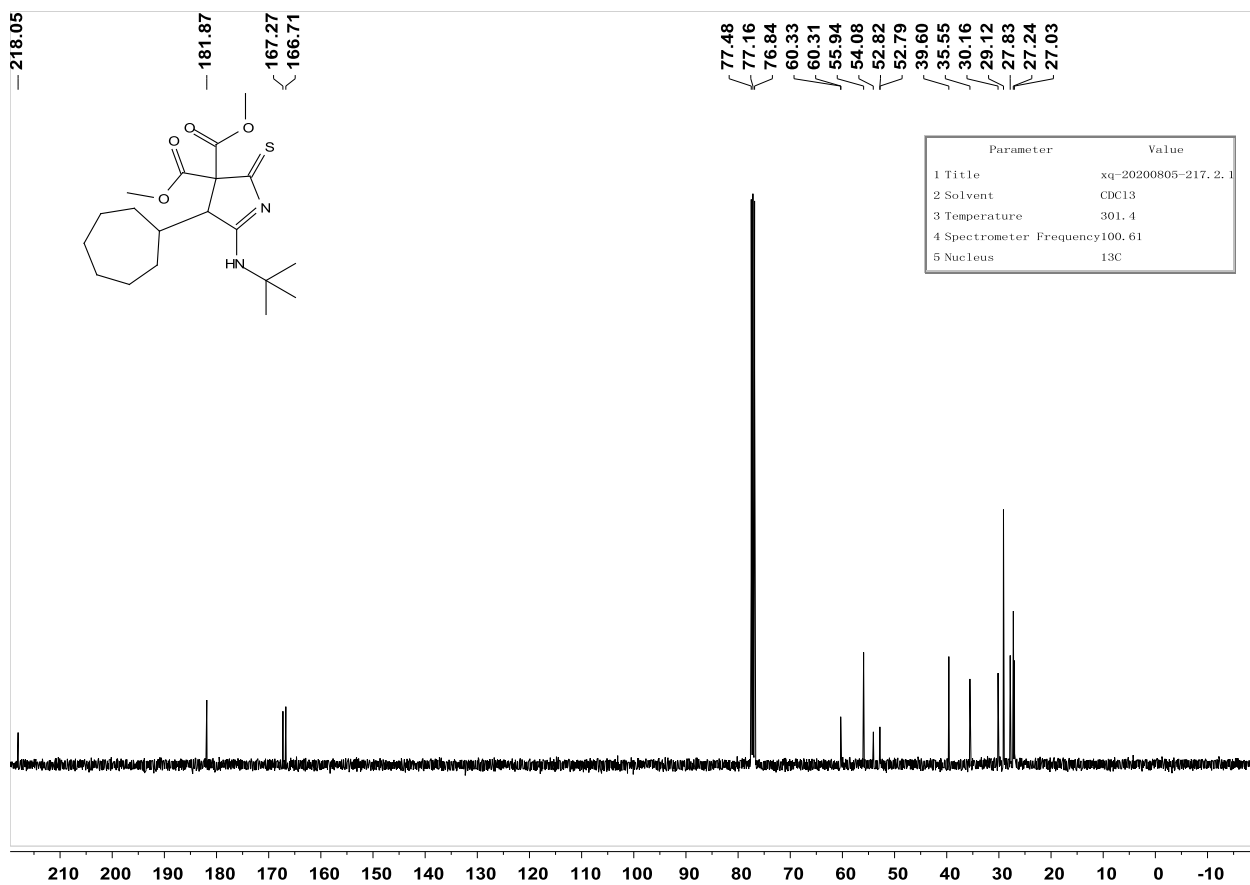


Figure S35.  $^1\text{H}$  NMR of 4r

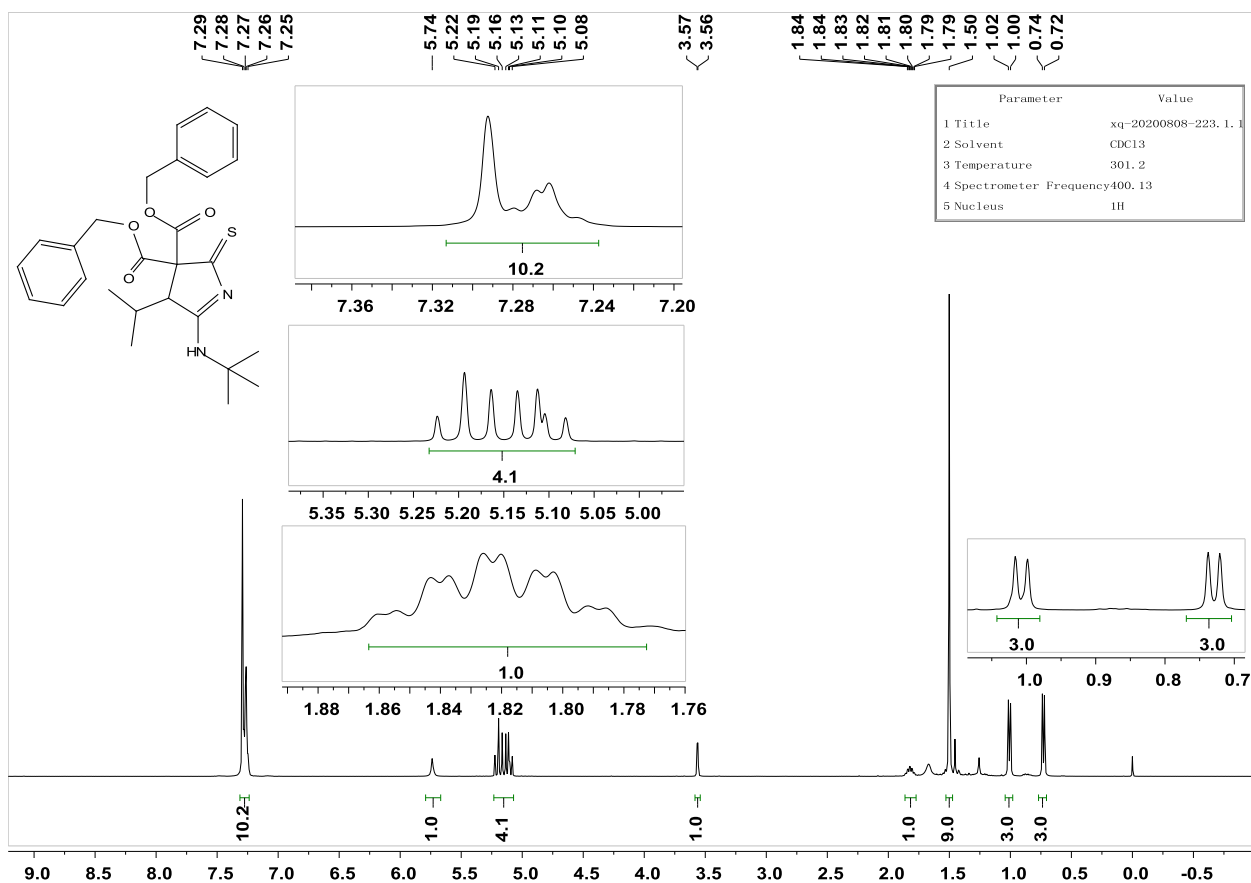


Figure S36.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4r

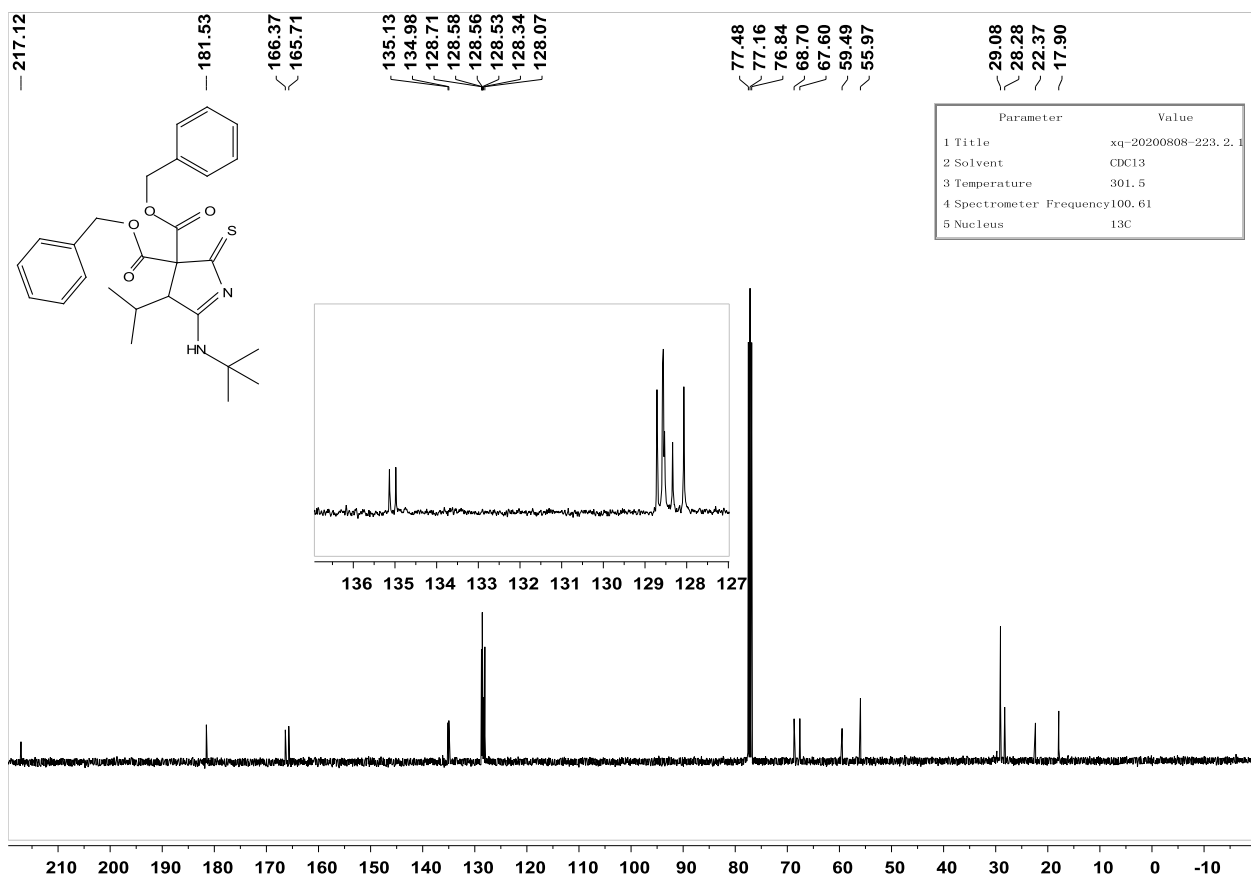


Figure S37.  $^1\text{H}$  NMR of 4s

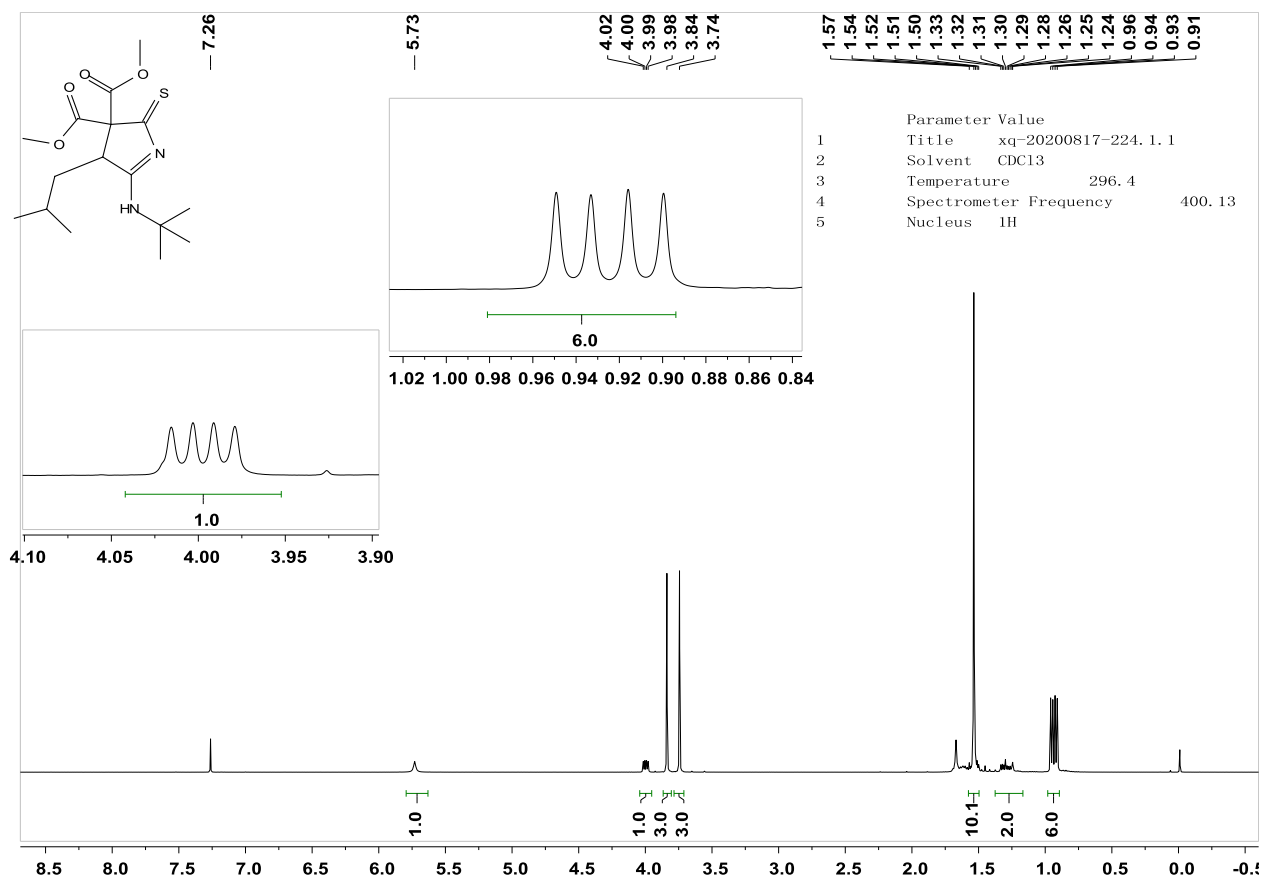


Figure S38.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4s

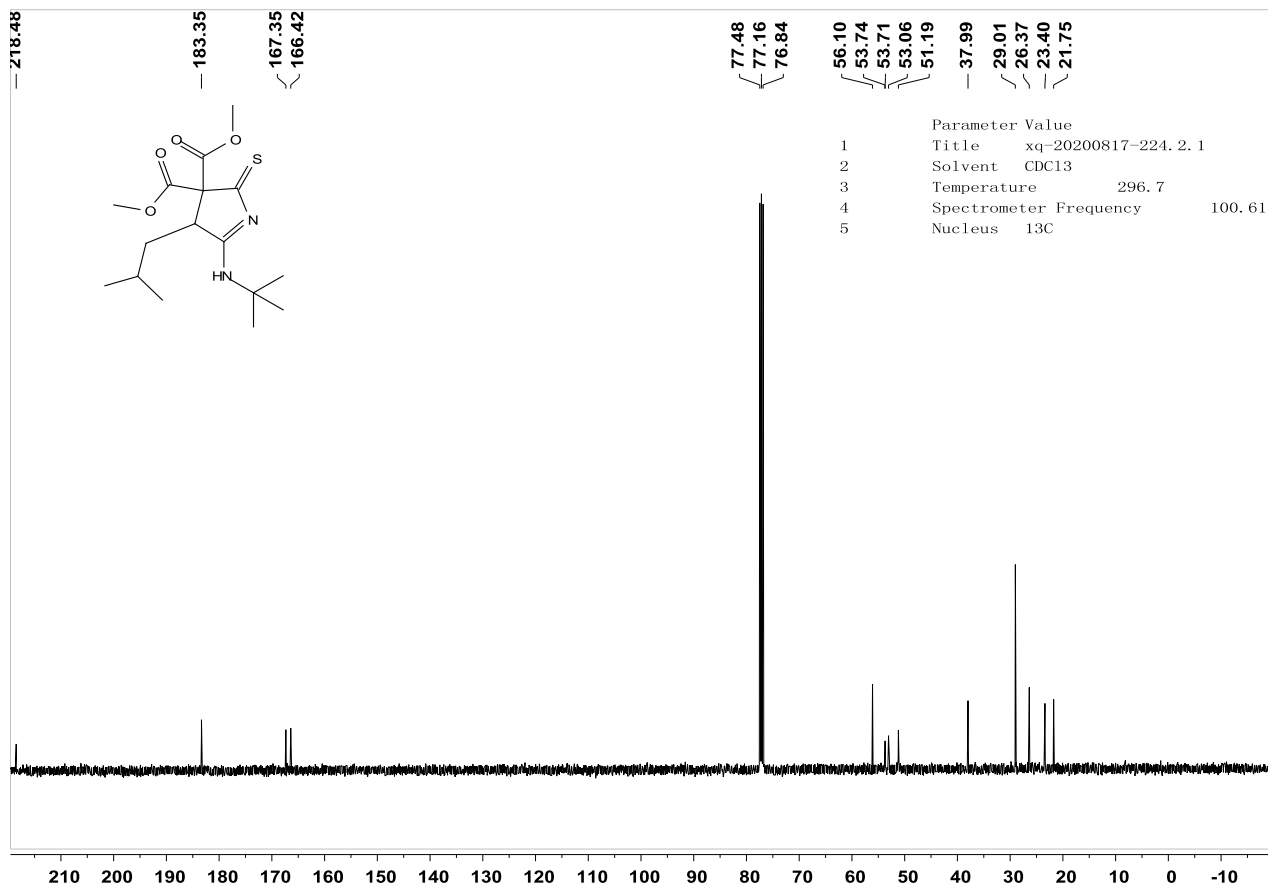


Figure S39.  $^1\text{H}$  NMR of 4t

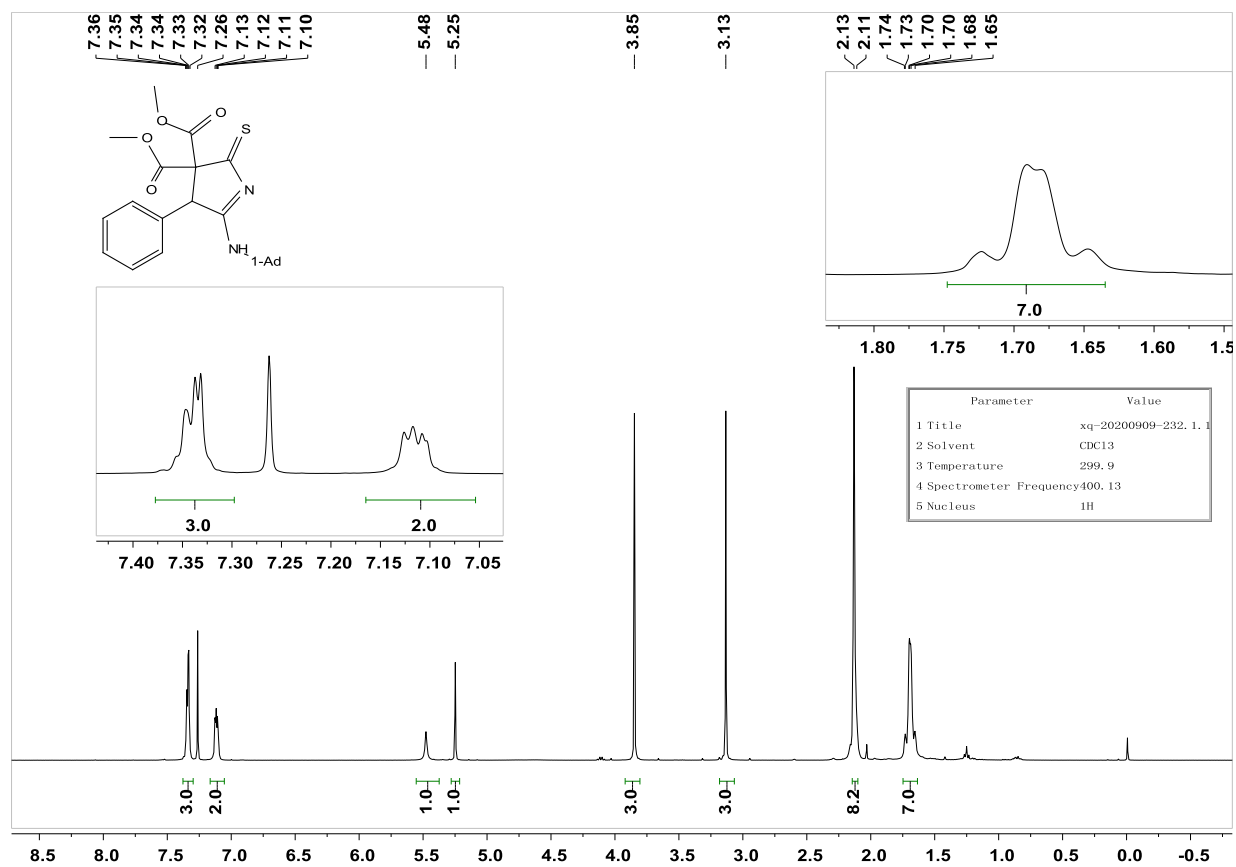


Figure S40.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 4t

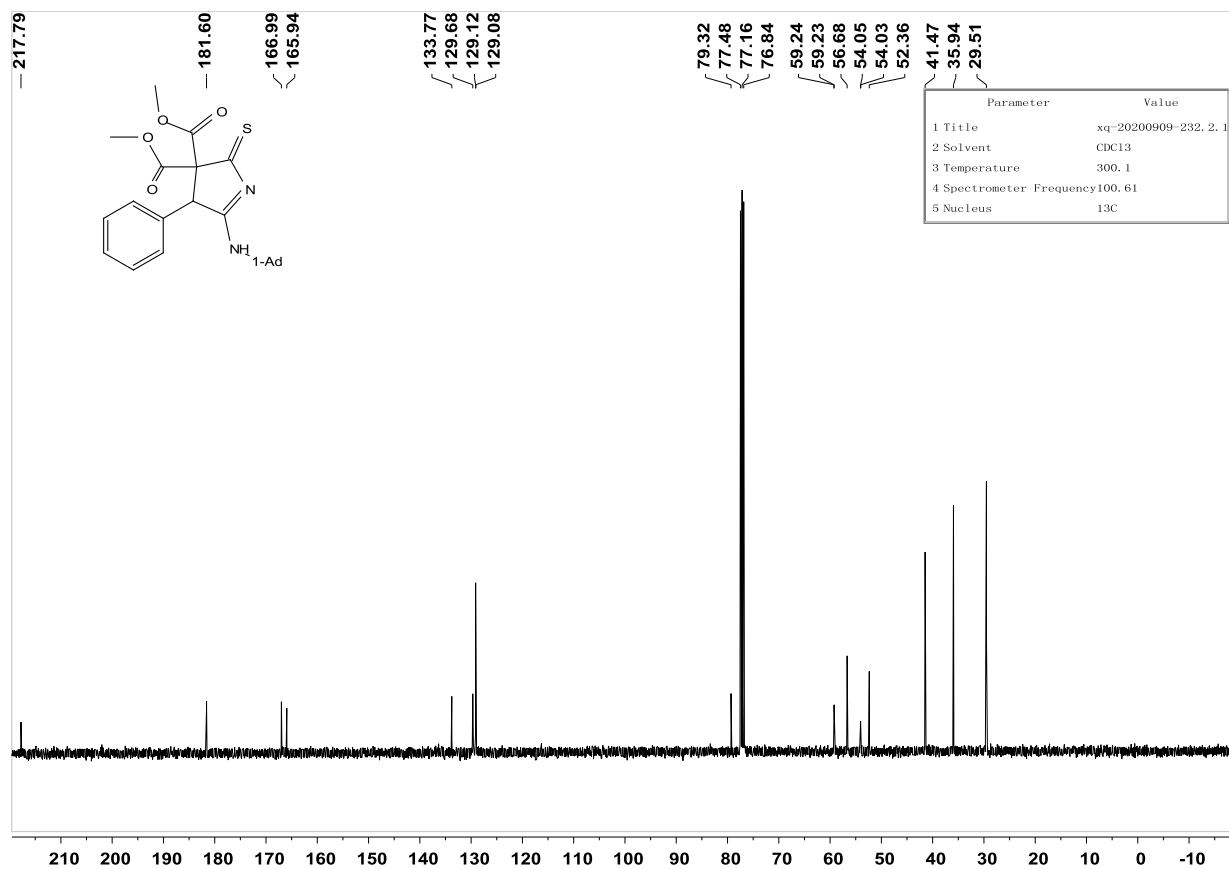


Figure S41.  $^1\text{H}$  NMR of 5a

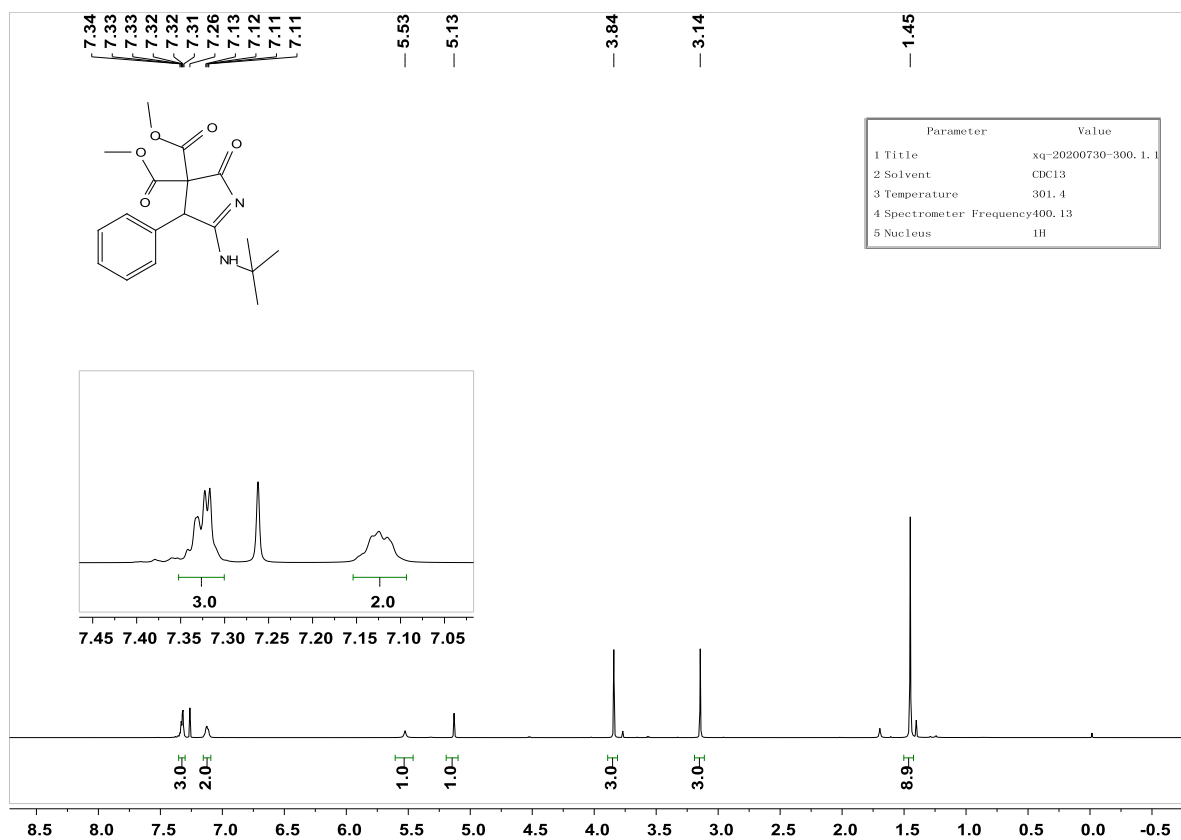


Figure S42.  $^{13}\text{C}\{^1\text{H}\}$  NMR of 5a

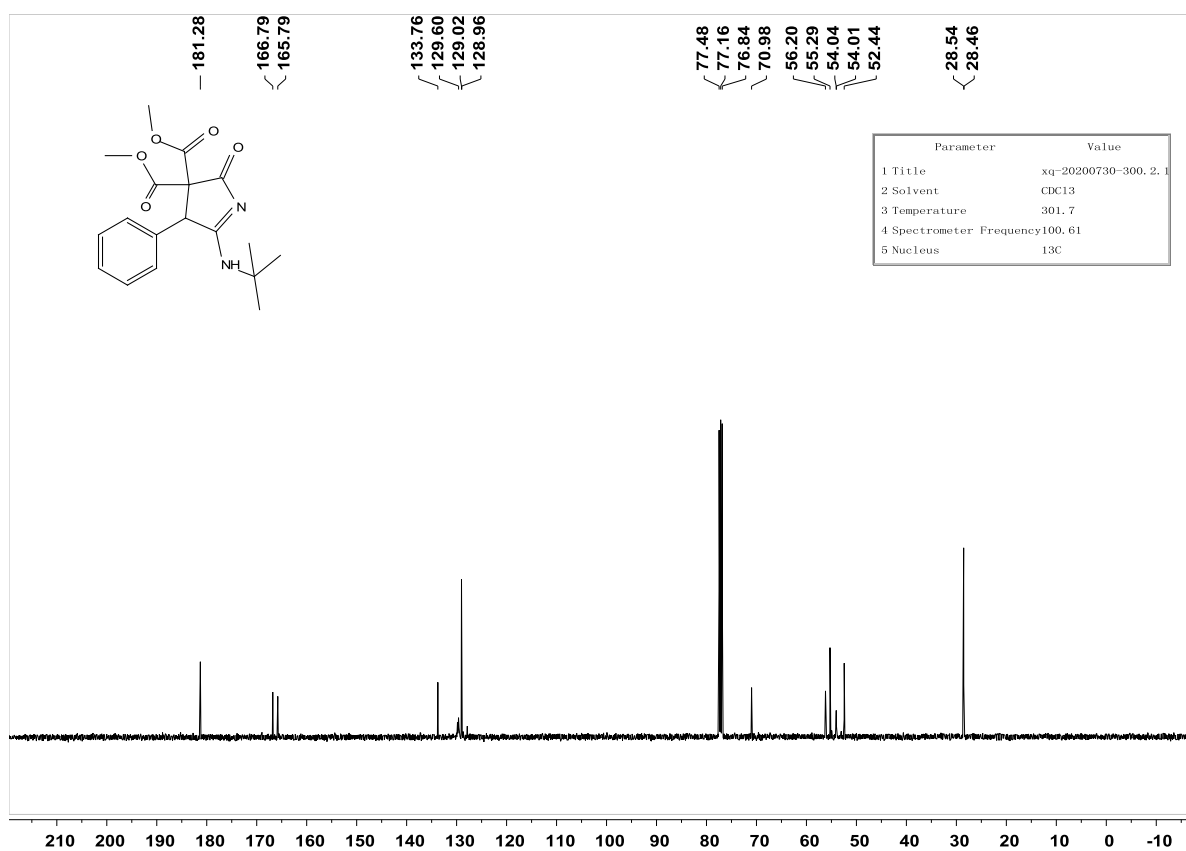


Figure S44. Cosy of 4a

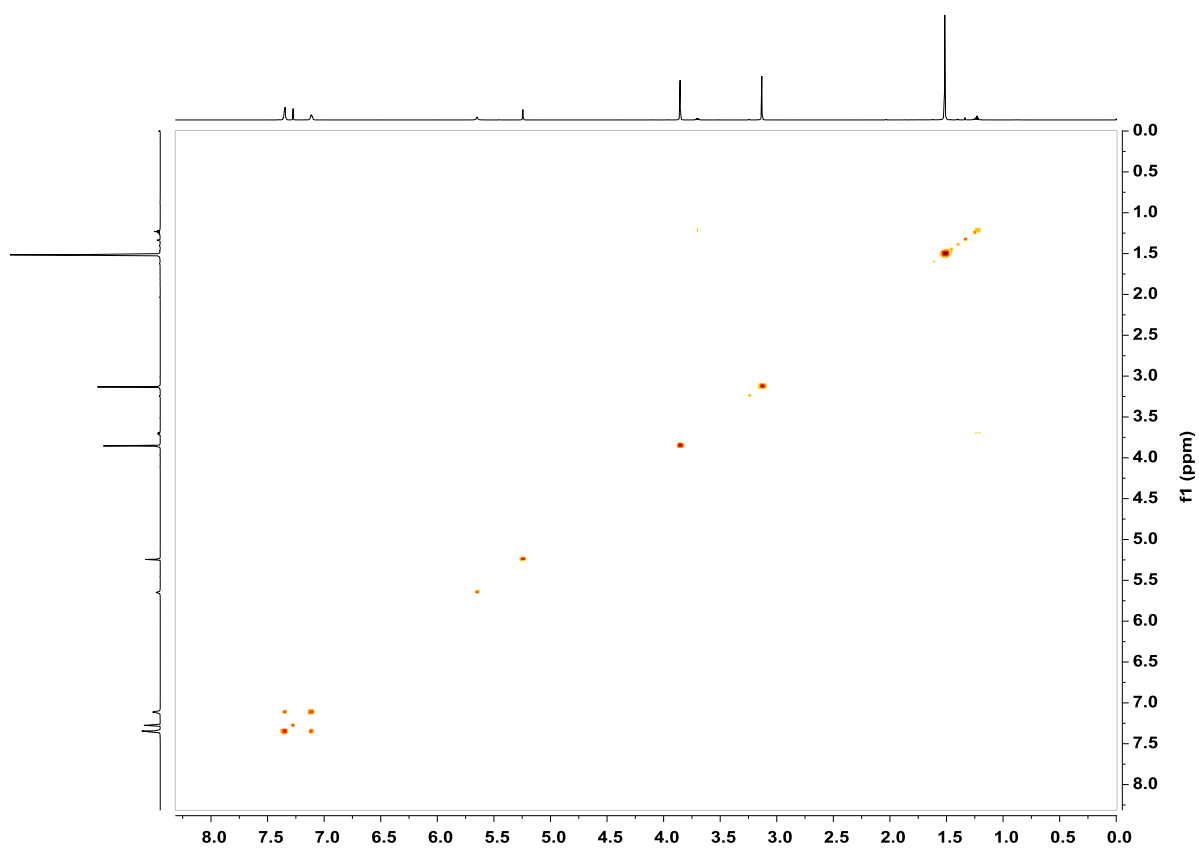


Figure S45. HSQC of 4a

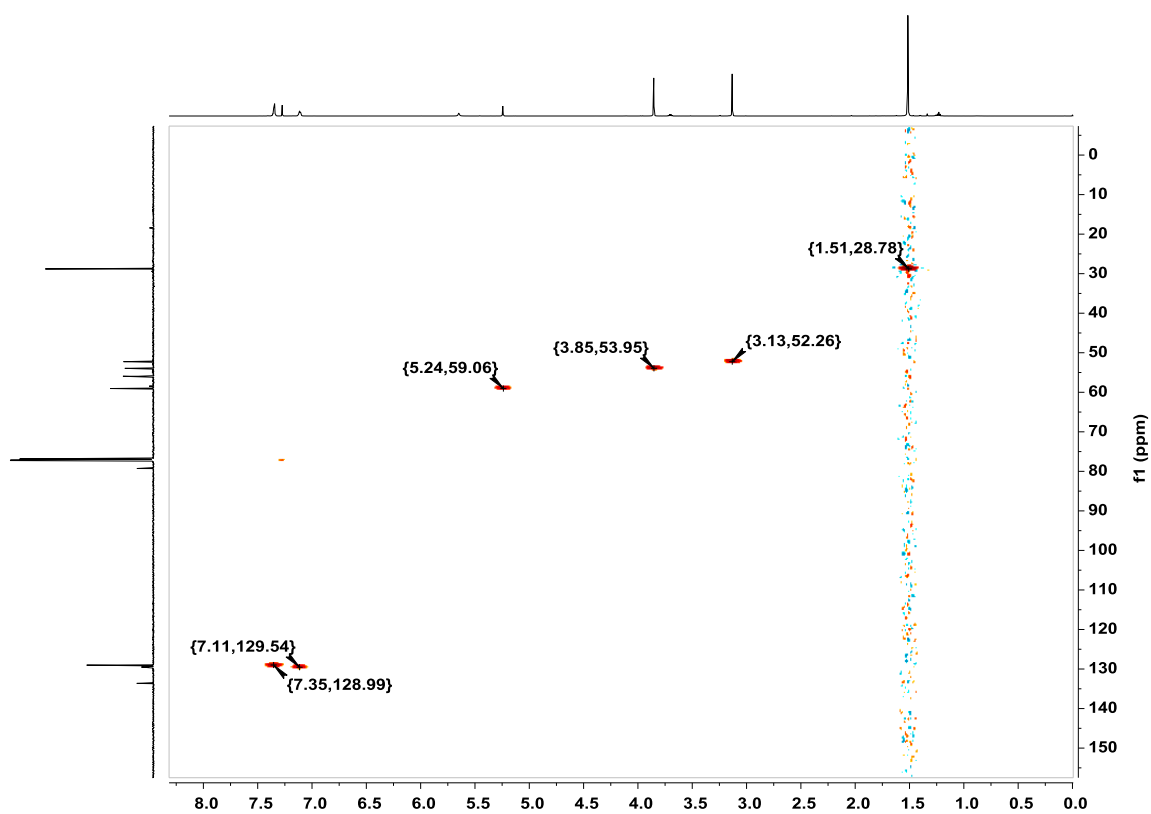


Figure S46. HSQC of 4a

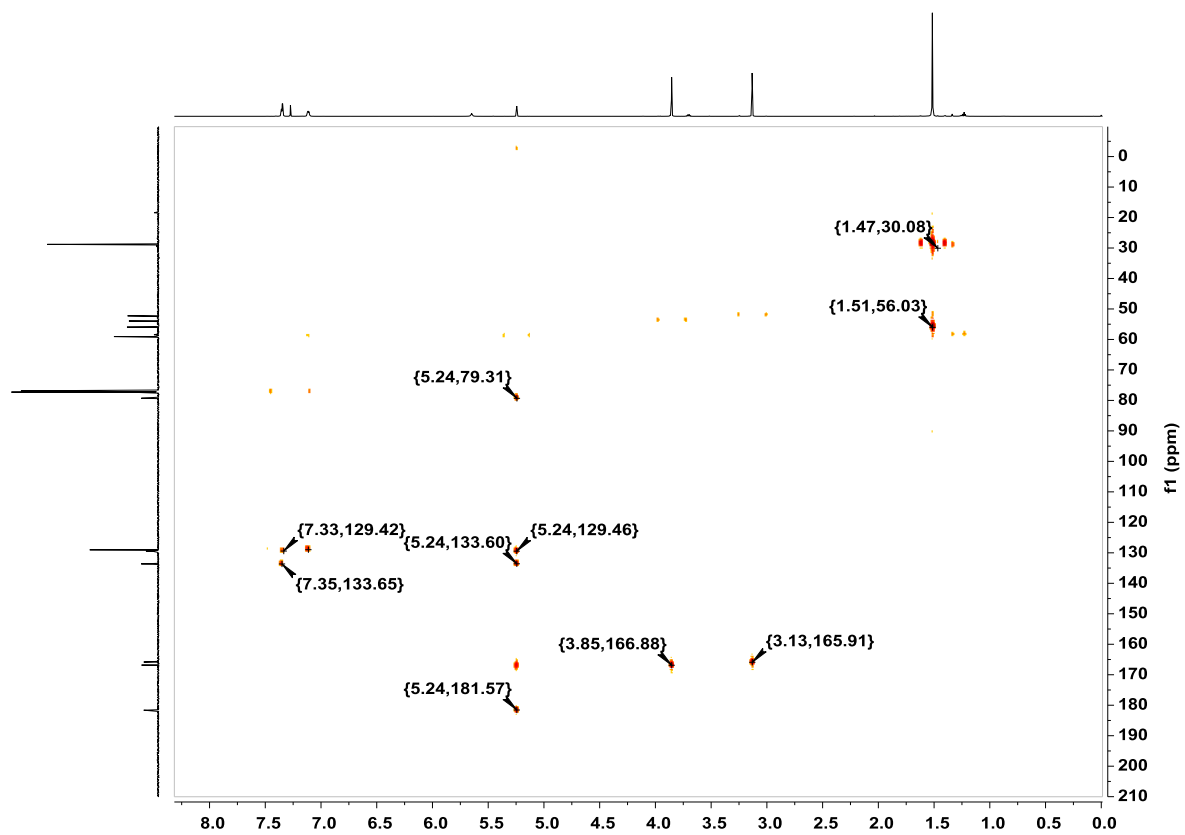
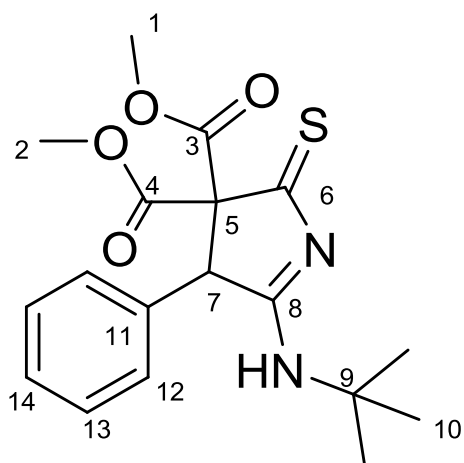
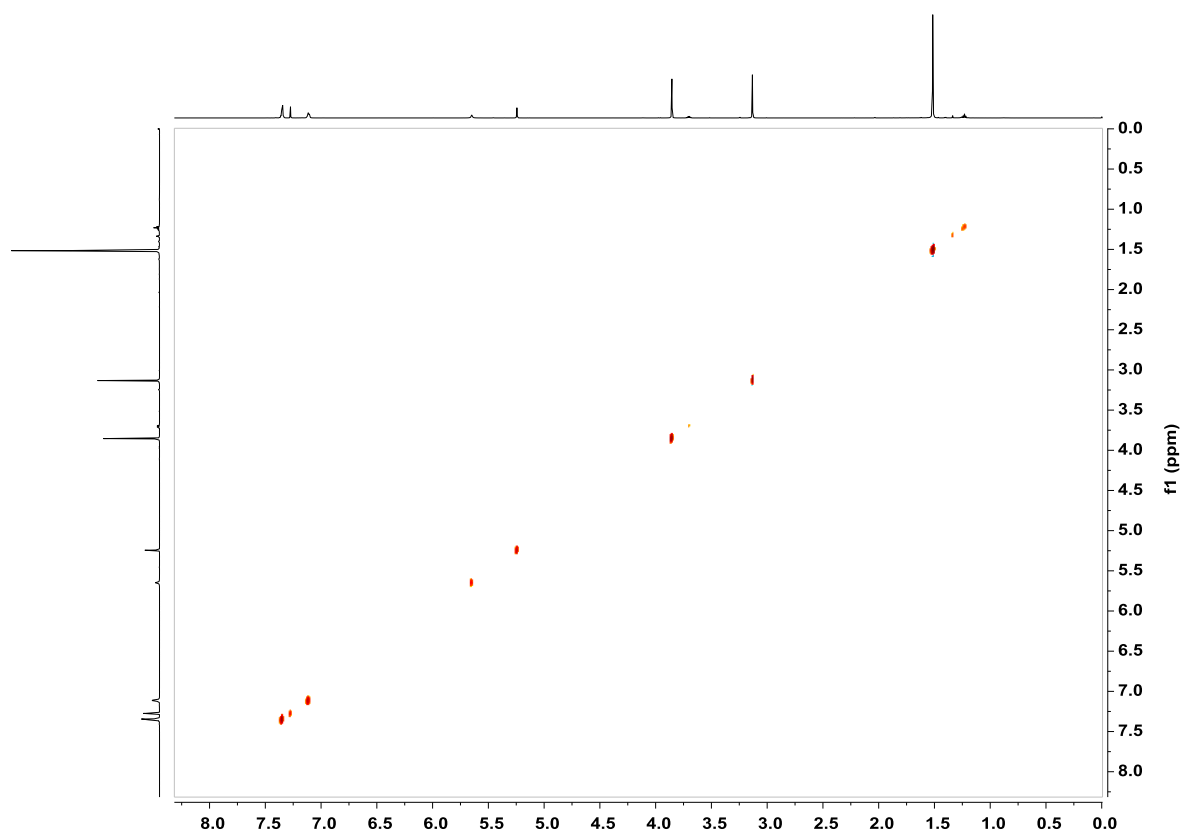


Figure S47. NOE of 4a



1, 3H (3.85, 53.95)    2, 3H (3.13, 52.26)    3, (166.88)    4, (165.91)    5, (79.31)    6, (181.57)    7, 1H (5.24, 59.06)    8, (129.46)    9, (56.03)    10, 9H (1.51, 28.78)    11, 1H (133.60)    12, 1H (7.32, 128.99)  
 13, 1H (7.11, 129.54)    14, 1H (7.35, 128.99)



Figure S48.  $^1\text{H}$  NMR of  $\text{Et}_3\text{NHNCs}$

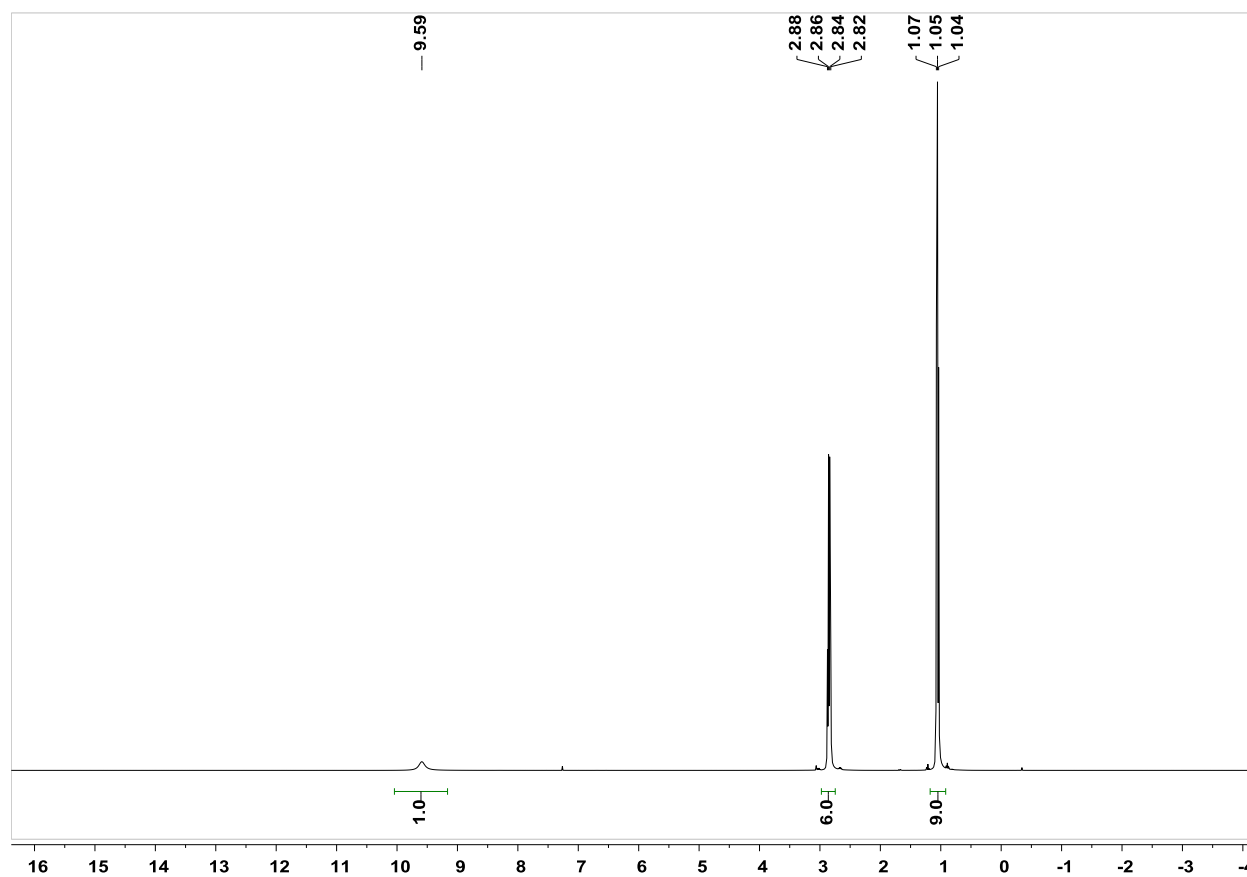
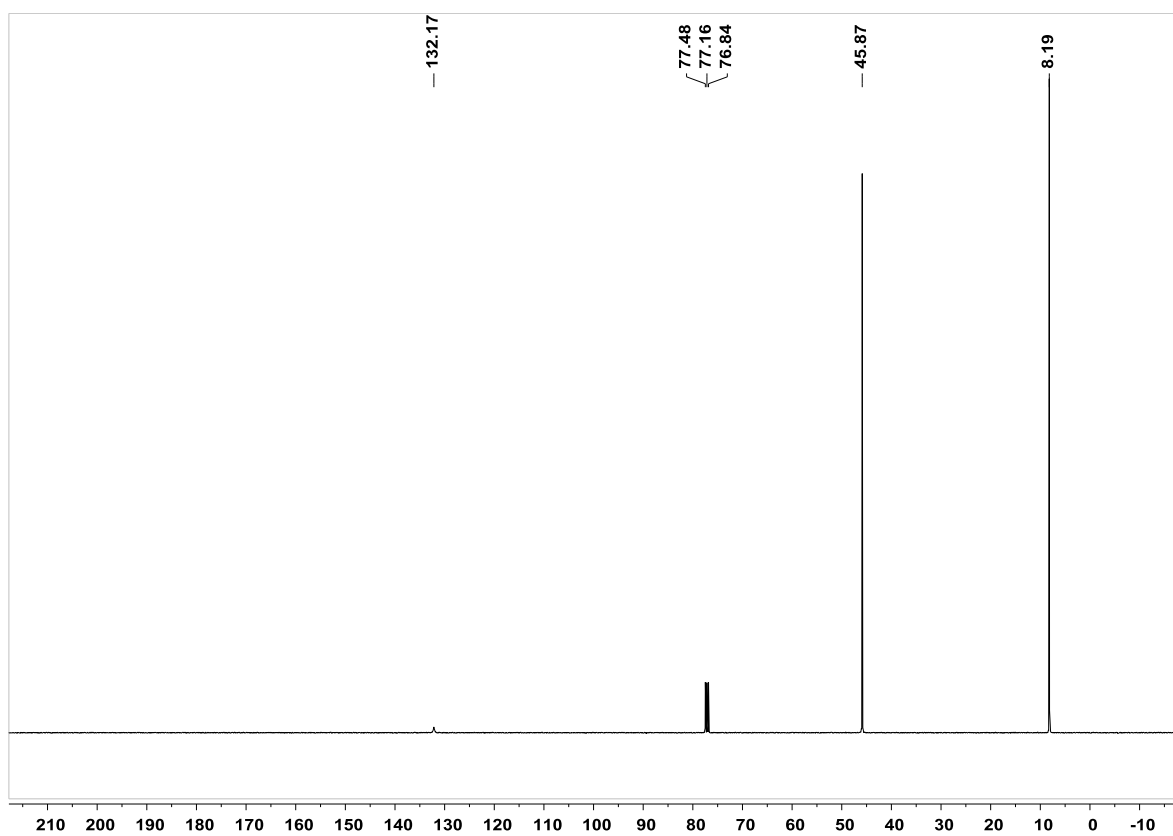


Figure S49.  $^{13}\text{C}$  NMR of  $\text{Et}_3\text{NHNCs}$



## 10. Absolute configuration of 4a and 4o

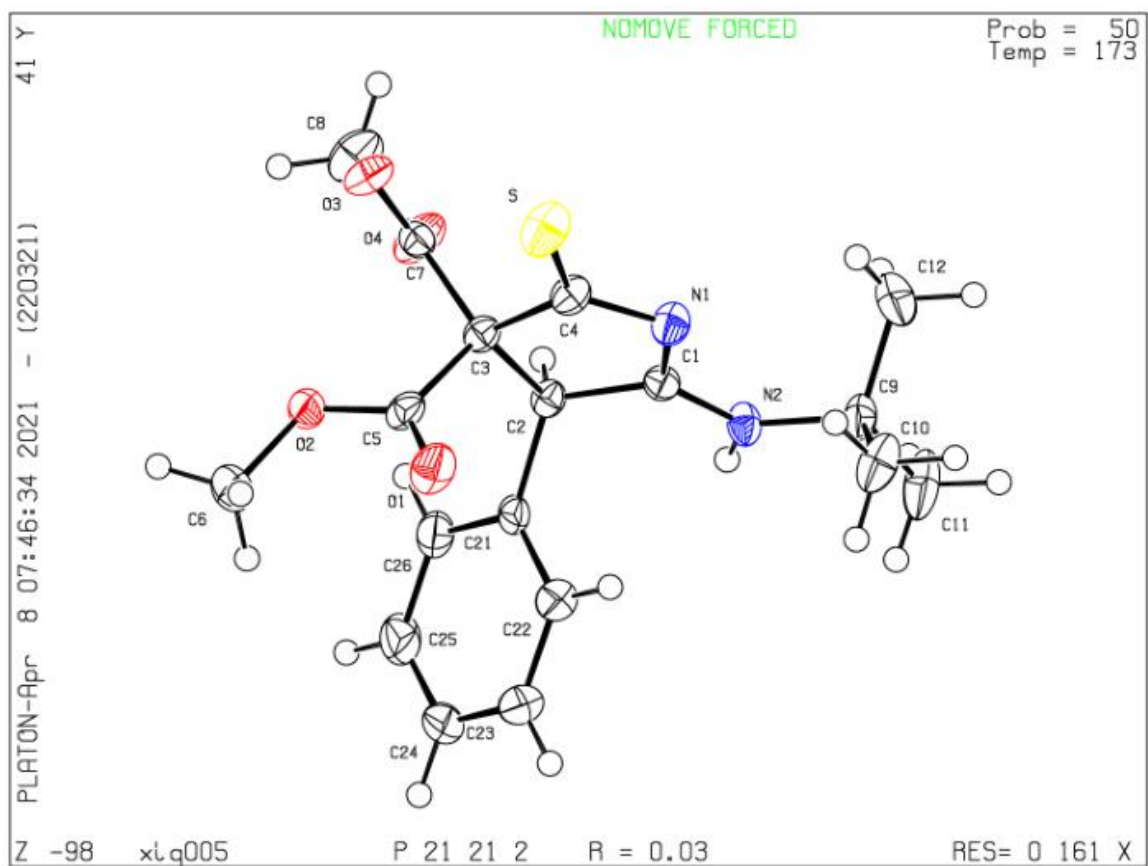


Figure S50. X-ray structure of 4a

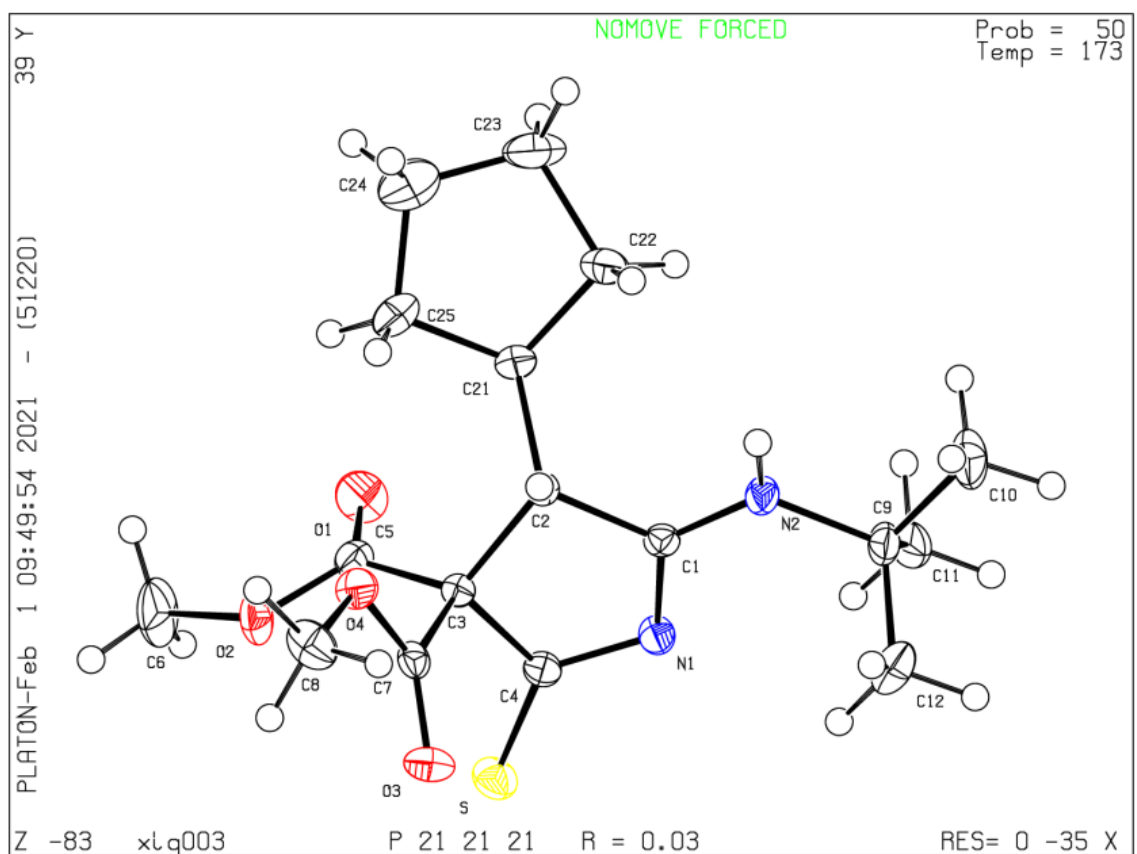


Figure S51. X-ray structure of **4o**Crystallographic Data for **4a** and **4o**.

|   | <b>4a</b>       | <b>4o</b>       |
|---|-----------------|-----------------|
| Formula   | C18 H22 N2 O4 S | C17 H26 N2 O4 S |
| Formula mass (amu)                                | 362.43          | 354.46          |
| Space group                                       | P 21 21 2       | P 21 21 21      |
| <i>a</i> (Å)                                      | 12.4981(5)      | 9.7300(2)       |
| <i>b</i> (Å)                                      | 17.3226(8)      | 11.7702(3)      |
| <i>c</i> (Å)                                      | 8.4593(4)       | 16.4807(4)      |
| $\alpha$ (deg)                                    | 90              | 90              |
| $\beta$ (deg)                                     | 90              | 90              |
| $\gamma$ (deg)                                    | 90              | 90              |
| <i>V</i> (Å <sup>3</sup> )                        | 1831.44(14)     | 1887.44(8)      |
| <i>Z</i>  | 4               | 4               |
| $\lambda$ (Å)                                     | 0.71073         | 1.54178         |
| <i>T</i> (K)                                      | 173 K           | 173 K           |
| $\rho_{\text{calcd}}$ (g cm <sup>-3</sup> )       | 1.314           | 1.247           |
| $\mu$ (mm <sup>-1</sup> )                         | 0.201           | 1.712           |
| Transmission factors                              | 0.898–0.989     | 0.637–0.781     |
| $\theta_{\text{max}}$ (deg)                       | 25.430          | 68.307          |
| No. of unique data, including $F_o^2 < 0$         | 3366            | 3442            |
| No. of unique data, with $F_o^2 > 2\sigma(F_o^2)$ | 3139            | 3401            |
| No. of variables                                  | 235             | 226             |
| $R(F)$ for $F_o^2 > 2\sigma(F_o^2)$ <sup>a</sup>  | 0.0308          | 0.0252          |
| $R_w(F_o^2)$ <sup>b</sup>                         | 0.0678          | 0.0704          |
| Goodness of fit                                   | 1.066           | 1.101           |

$$^a R(F) = \sum ||F_o| - |F_c|| / \sum |F_o|.$$

$$^b R_w(F_o^2) = [\sum [w(F_o^2 - F_c^2)^2] / \sum wF_o^4]^{1/2}; w^{-1} = [\sigma^2(F_o^2) + (Ap)^2 + Bp], \text{ where } p = [\max(F_o^2, 0) + 2F_c^2] / 3.$$

## 11. References

- (1) (a) Y. H. Wen, X. Huang, J. L. Huang, Y. Xiong, B. Qin, X. M. Feng, *Synlett* 2005, 2445. (b) Z. P. Yu, X. H. Liu, Z. H. Dong, M. S. Xie, X. M. Feng, *Angew. Chem. Int. Ed.* 2008, **47**, 1308.
- (2) (a) J. Wang, Y. Zhou, L. Zhang, Z. Li, X. Chen, H. Liu, *Org. Lett.* 2013, **15**, 1508. (b) Y. Guan, J. W. Attard, Visco, M. D. Fisher, T. J. Mattson, *Chem. Eur. J.* 2018, **24**, 7123.
- (3) I. A. Andreev, N. K. Ratmanova, A. U. Augustin, O. A. Ivanova, I. I. Levina, V. N. Khrustalev, D. B. Werz, I. V. Trushkov, *Angew. Chem. Int. Ed.* 2021, **60**, 7927.