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Supplementary Information

Efficient asymmetric syntheses of α-quaternary lactones and esters through chiral bifunctional sulfide-catalyzed desymmetrizing bromolactonization of α,α-diallyl carboxylic acids

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General Information

¹H and ¹³C NMR spectra were measured on JEOL JNM-AL 400 and JEOL JNM-ECZ 400R NMR instruments (400 MHz for ¹H NMR, 100 MHz for ¹³C NMR). Tetramethylsilane (TMS) served as the internal standard (0 ppm) for ¹H NMR, and CDCl₃ served as the internal standard (77.0 ppm) for ¹³C NMR. The following abbreviations were used to express the multiplicities: s = singlet; d = doublet; t = triplet; m = multiplet; br = broad. High-resolution mass spectra (HRMS) were measured on a JEOL JMS-700N. Infrared spectra (IR) were measured on a JASCO FT/IR-4200 spectrometer. Optical rotations were measured on a JASCO P-2100 polarimeter. High performance liquid chromatography (HPLC) was performed on Shimadzu LC-20AT and SPD-20A instruments using Daicel Chiralpak IA-3, IB-3, IC-3, ID-3, IF-3, IG-3, or Chiralcel OD-3 columns (4.6 mm × 250 mm). All reactions were monitored by thin-layer chromatography using Merck precoated TLC plates (silica gel 60GF-254, 0.25 mm), with visualization by the use of UV lamp (254 nm) or dyes. The products were purified by flash column chromatography on silica gel. Dehydrated solvents were purchased from Kanto Chemical.

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Table S1 Optimization of reaction solvents^a

Н	Br (10 mol %) (10 mol %) (10 mol %) (10 mol %) (10 mol %) (10 mol %) (12 equiv)	Ph Ph 2a	(S)-4b	SBu OH Ph
Entry	Solvent	Yield ^b (%)	dr ^c	er ^d
1	CH ₂ Cl ₂ (2.0 mL)	99	>20: 1	88:12
2	toluene (2.0 mL)	60	14:1	82:18
3	toluene (1.0 mL)-CH ₂ Cl ₂ (1.0 mL)	98	>20: 1	93: 7
4	hexane (1.0 mL)-CH ₂ Cl ₂ (1.0 mL)	99	>20: 1	91: 9
5	toluene (1.5 mL)-CH ₂ Cl ₂ (0.5 mL)	99	>20: 1	94: 6

^{*a*} Reaction conditions: **1a** (0.10 mmol), NBP (0.12 mmol), (*S*)-**4b** (10 mol %, 0.010 mmol), solvent (2.0 mL), –78 °C, 24 h. ^{*b*} Yield of isolated product **2a**. ^{*c*} The diastereomeric ratio (dr) was determined via ¹H NMR analysis. ^{*d*} The enantiomeric ratio (er) was determined via HPLC analysis on a chiral stationary phase.

Table S2 Effect of brominating reagents^a



^{*a*} Reaction conditions: **1a** (0.10 mmol), brominating reagent (0.12 mmol), (*S*)-**4b** (10 mol %, 0.010 mmol), toluene (1.5 mL)-CH₂Cl₂ (0.5 mL), –78 °C, 24 h. ^{*b*} Yield of isolated product **2a**. ^{*c*} The diastereomeric ratio (dr) was determined via ¹H NMR analysis. ^{*d*} The enantiomeric ratio (er) was determined via HPLC analysis on a chiral stationary phase.



Scheme S1 Effect of chiral catalysts.





When a reaction of **1n** was performed in the absence of a catalyst, product **2n** was obtained in a low yield with poor diastereoselectivity (1.1: 1 dr). Unfortunately, the reactions using chiral bifunctional sulfide catalysts did not improve the low diastereoselectivity of **2n**, although moderate levels of enantioselectivity were observed.



Scheme S3 Optimization of reaction conditions for substrate 1n.

Experimental Section

1. Synthesis of catalysts

Chiral sulfide catalysts (S)-3, (S)-4a–4e, (S)-5, and (S)-14 were prepared according to the literature.¹

2. Synthesis of substrates

Substrates 1 were prepared according to the literature method.² Substrate 7 was also prepared according to the literature method.³



1b: ¹H NMR (400 MHz, CDCl₃) δ 7.33 (d, J = 8.8 Hz, 2H), 7.25 (d, J = 8.8 Hz, 2H), 5.59–5.48 (m, 2H), 5.11–5.07 (m, 4H), 2.80 (dd, J = 7.6, 14.0 Hz, 2H), 2.73 (dd, J = 6.8, 13.6 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 181.1, 139.4, 133.1, 132.6, 128.6, 128.0, 119.2, 53.1, 38.5; IR

(neat): 3078, 3007, 2980, 2921, 1700, 1494, 1282, 1234, 1096, 1013, 995, 918, 826, 754, 725 cm⁻¹; HRMS (FAB) calcd for C₁₄H₁₅ClO₂: 250.0761 ([M]⁺), found 250.0761.



1c: ¹H NMR (400 MHz, CDCl₃) δ 7.28 (dd, J = 5.4, 9.0 Hz, 2H), 7.04 (dd, J = 8.8, 8.8 Hz, 2H), 5.59–5.49 (m, 2H), 5.11–5.07 (m, 4H), 2.81 (dd, J = 7.8, 13.8 Hz, 2H), 2.73 (dd, J = 6.8, 13.6 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 181.2, 161.7 (d, J = 245 Hz), 136.6 (d, J = 3.3 Hz), 132.7, 128.3

(d, J = 8.3 Hz), 119.1, 115.3 (d, J = 21.4 Hz), 52.9, 38.7; IR (neat): 3079, 2981, 2922, 1699, 1509, 1282, 1234, 1165, 995, 917, 832, 815, 734, 679, 650 cm⁻¹; HRMS (FAB) calcd for C₁₄H₁₅FO₂: 234.1056 ([M]⁺), found 234.1056.

1d: ¹H NMR (400 MHz, CDCl₃) δ 7.20 (d, J = 8.4 Hz, 2H), 7.16 (d, J = 8.8 Hz, 2H), 5.61–5.50 (m, 2H), 5.11–5.05 (m, 4H), 2.81 (dd, J = 7.6, 13.6 Hz, 2H), 2.75 (dd, J = 6.8, 14.0 Hz, 2H), 2.34 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 181.1, 137.9, 136.8, 133.2, 129.2, 126.4, 118.8, 52.9,

38.5, 21.0; IR (neat): 3077, 3007, 2980, 2947, 2923, 1698, 1515, 1282, 1234, 1132, 995, 915, 819, 730 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₈O₂: 230.1307 ([M]⁺), found 230.1307.



1e: ¹H NMR (400 MHz, CDCl₃) δ 7.24 (d, J = 8.8 Hz, 2H), 6.88 (d, J = 8.8 Hz, 2H), 5.60–5.50 (m, 2H), 5.11–5.05 (m, 4H), 3.80 (s, 3H), 2.81 (dd, J = 7.6, 14.0 Hz, 2H), 2.73 (dd, J = 6.8, 14.0 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 180.9, 158.5, 133.2, 132.9, 127.6, 118.8, 113.8,

55.2, 52.5, 38.5; IR (neat): 3077, 3004, 2979, 2954, 2936, 1699, 1514, 1292, 1252, 1185, 1035, 918, 830 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₈O₃: 246.1256 ([M]⁺), found 246.1256.



1f: ¹H NMR (400 MHz, CDCl₃) δ 7.61 (d, J = 8.4 Hz, 2H), 7.43 (d, J = 8.0 Hz, 2H), 5.58–5.48 (m, 2H), 5.12–5.08 (m, 4H), 2.84 (dd, J = 8.0, 14.0 Hz, 2H), 2.77 (dd, J = 6.8, 13.6 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 180.7, 144.9, 132.3, 129.5 (q, J = 32.1 Hz), 127.1, 125.4 (q, J

= 3.3 Hz), 124.0 (q, J = 271 Hz), 119.5, 53.7, 38.7; IR (neat): 3081, 2983, 1703, 1326, 1167, 1122, 1071, 1017, 920, 838 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₅F₃O₂: 284.1024 ([M]⁺), found 284.1024.



1g: ¹H NMR (400 MHz, CDCl₃) δ 7.34 (d, *J* = 8.8 Hz, 2H), 7.20 (d, *J* = 8.8 Hz, 2H), 5.59–5.48 (m, 2H), 5.12–5.07 (m, 4H), 2.82 (dd, *J* = 8.0, 14.0 Hz, 2H), 2.74 (dd, *J* = 6.8, 13.6 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 180.7, 148.2 (q, *J* = 1.7 Hz), 139.5, 132.5, 128.1, 120.7, 120.4

(q, J = 256 Hz), 119.3, 53.1, 38.7; IR (neat): 3080, 2983, 2925, 1703, 1510, 1254, 1212, 1162, 1116, 1019, 995, 920, 849, 686 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₅F₃O₃: 300.0973 ([M]⁺), found 300.0974.



1h: ¹H NMR (400 MHz, CDCl₃) δ 7.31–7.25 (m, 3H), 7.20 (td, *J* = 2.0, 7.2 Hz, 1H), 5.59–5.49 (m, 2H), 5.13–5.08 (m, 4H), 2.81 (dd, *J* = 7.6, 14.0 Hz, 2H), 2.74 (dd, *J* = 6.8, 13.6 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 180.7, 143.0, 134.5, 132.5, 129.7, 127.4, 126.9, 124.9, 119.4,

53.4, 38.5; IR (neat): 3078, 2980, 1699, 1594, 1573, 1282, 1234, 996, 918, 848, 771, 712, 687, 649 cm⁻¹; HRMS (FAB) calcd for C₁₄H₁₆ClO₂: 251.0839 ([M+H]⁺), found 251.0839.



1i: ¹H NMR (400 MHz, CDCl₃) δ 7.23 (d, J = 8.0 Hz, 1H), 7.12–7.08 (m, 3H), 5.60–5.50 (m, 2H), 5.12–5.06 (m, 4H), 2.82 (dd, J = 7.6, 13.6 Hz, 2H), 2.75 (dd, J = 6.8, 14.0 Hz, 2H), 2.36 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 181.3, 140.8, 138.0, 133.2, 128.3, 127.9, 127.2, 123.5,

118.8, 53.2, 38.5, 21.6; IR (neat): 3077, 3007, 2979, 2947, 2923, 1698, 1640, 1606, 1442, 1284, 1234, 995, 915, 776, 721, 705, 650 cm⁻¹; HRMS (FAB) calcd for $C_{15}H_{19}O_2$: 231.1385 ([M+H]⁺), found 231.1377.

i j: ¹H NMR (400 MHz, CDCl₃) δ 7.30–7.23 (m, 2H), 7.14 (dd, J = 6.8, 7.6 Hz, 1H), 7.05 (dd, J = 8.4, 10.8 Hz, 1H), 5.61–5.51 (m, 2H), 5.10–5.06 (m, 4H), 2.81 (dd, J = 6.8, 14.0 Hz, 2H), 2.73 (dd, J = 8.0, 14.0 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 180.5, 160.5 (d, J = 245 Hz), 132.6, 129.0 (d, J = 13.2 Hz), 128.9 (d, J = 9.1 Hz), 127.8 (d, J = 4.1 Hz), 123.8 (d, J = 3.3 Hz), 119.1, 115.7 (d, J = 22.2 Hz), 50.9, 38.1; IR (neat): 3082, 3009, 2982, 2926, 2876, 1698, 1488, 1446, 1300, 1286, 1241, 1217, 996, 920, 811, 755, 745, 686 cm⁻¹; HRMS (FAB) calcd for C₁₄H₁₅FO₂: 234.1056 ([M]⁺), found 234.1056.



1k: ¹H NMR (400 MHz, CDCl₃) δ 7.84–7.75 (m, 4H), 7.51–7.42 (m, 3H), 5.63–5.52 (m, 2H), 5.14–5.06 (m, 4H), 2.94 (dd, *J* = 7.6, 14.0 Hz, 2H), 2.89 (dd, *J* = 6.8, 13.6 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 180.6, 138.3, 133.1, 133.0, 132.4, 128.21, 128.15, 127.5, 126.2, 126.1,

125.4, 124.8, 119.0, 53.5, 38.5; IR (neat): 3075, 3061, 3007, 2979, 2923, 1698, 1639, 1282, 1233, 1136, 995, 917, 855, 817, 747 cm⁻¹; HRMS (FAB) calcd for C₁₈H₁₈O₂: 266.1307 ([M]⁺), found 266.1306.

11:⁴ ¹H NMR (400 MHz, CDCl₃) δ 7.25 (dd, J = 0.8, 4.8 Hz, 1H), 7.01– 6.97 (m, 2H), 5.69–5.59 (m, 2H), 5.15–5.09 (m, 4H), 2.89 (dd, J = 7.2, 13.6 Hz, 2H), 2.80 (dd, J = 7.2, 14.0 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 180.0, 145.0, 132.6, 126.6, 125.1, 124.6, 119.2, 51.7, 40.2; IR (neat): 3074, 3010, 2979, 2964, 2911, 1697, 1641, 1447, 1405, 1294, 1280, 1243, 1230, 996, 926, 889, 852, 703 cm⁻¹. **1m**:^{5 1}H NMR (400 MHz, CDCl₃) δ 7.31 (dd, J = 2.8, 4.8 Hz, 1H), 7.16 (dd, J = 1.2, 2.8 Hz, 1H), 7.06 (dd, J = 1.2, 4.8 Hz, 1H), 5.64-5.54 (m, 2H), 5.13–5.07 (m, 4H), 2.82 (dd, J = 7.6, 14.0 Hz, 2H), 2.75 (dd, J = 7.2, 14.0 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 180.9, 141.7, 132.9, 126.6, 125.6, 121.6, 118.9, 51.3, 39.0; IR (neat): 3080, 2981, 2954, 2925, 1699, 1281, 1235, 904, 863, 771, 727, 693, 649, 623 cm⁻¹.

1n: ¹H NMR (400 MHz, CDCl₃) δ 7.27 (t, J = 7.4 Hz, 2H), 7.20–7.16 (m, 3H), 5.84–5.74 (m, 2H), 5.20–5.13 (m, 4H), 2.61–2.57 (m, 2H), 2.44 (d, J = 7.2 Hz, 4H), 1.90–1.86 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 182.4, 141.9, 133.1, 128.4, 128.3, 125.9, 118.7, 49.2, 38.6, 37.1, 30.5; IR (neat): 3078, 3027, 2979, 2929, 1697, 1455, 1231, 994, 907, 730, 698 cm⁻¹; HRMS (FAB) calcd for C₁₆H₂₀O₂: 244.1463 ([M]⁺), found 244.1462.

10: ¹H NMR (400 MHz, CDCl₃) δ 7.29–7.18 (m, 5H), 5.90–5.80 (m, 2H), 5.19–5.14 (m, 4H), 2.94 (s, 2H), 2.42 (dd, J = 7.2, 14.4 Hz, 2H), 2.31 (dd, J = 7.6, 14.4 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 182.0, 137.0, 133.3, 130.1, 128.2, 126.7, 118.9, 50.7, 41.1, 38.1; IR (neat): 3077, 3031, 2979, 2924, 1698, 1639, 1454, 1446, 1282, 1248, 1232, 992, 916, 737, 701 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₈O₂: 230.1307 ([M]⁺), found 230.1305.

3. General procedure for stereoselective desymmetrizing bromolactonizations of 1

A solution of substrate 1 (0.10 mmol) and catalyst (*S*)-4c (10 mol %, 0.010 mmol) in toluene (1.5 mL)-CH₂Cl₂ (0.5 mL) was cooled to -78 °C. After stirring for 10 min at -78 °C, *N*-bromophthalimide (NBP) (0.12 mmol) was added to the cooled reaction solution. The reaction mixture was stirred for 24 h at -78 °C. After 24 h, the reaction mixture was quenched with saturated aqueous Na₂SO₃ (4.0 mL) at -78 °C and stirred for 10 min at -78 °C. The quenched reaction mixture was diluted with CH₂Cl₂ (2 mL) and H₂O (2 mL), and warmed to room temperature. The organic materials were extracted with CH₂Cl₂ for three times (5 mL × 3). The combined extracts were dried over Na₂SO₄ and concentrated. [The ¹H NMR analysis of the crude reaction mixture was performed at this stage to confirm diastereoselectivity.] The residue was purified by flash column

chromatography on silica gel (hexane/ethyl acetate as eluent) to give product 2. The enantioselectivity of the product 2 was determined by HPLC analysis on a chiral stationary phase.

2a:^{2a} $[\alpha]^{21}_{D}$ +52.6 (*c* = 0.84, CHCl₃, 95: 5 er); HPLC analysis: Daicel Chiralpak IB-3, hexane/2-propanol = 50:1, flow rate = 0.5 mL/min, 214 nm; retention time: 23.1 min (minor) and 39.9 min (major). ¹H NMR (400 MHz, CDCl₃) δ 7.44–7.37 (m, 4H), 7.34–7.29 (m, 1H), 5.71–5.60 (m, 1H), 5.15–5.10 (m, 2H), 4.48–4.41 (m, 1H), 3.59 (dd, *J* = 5.2, 10.8 Hz, 1H), 3.49 (dd, *J* = 6.8, 10.8 Hz, 1H), 2.82 (dd, *J* = 5.2, 13.2 Hz, 1H), 2.75–2.64 (m, 2H), 2.29 (dd, *J* = 10.8, 13.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 177.1, 138.3, 132.7, 129.0, 127.9, 126.1, 119.8, 75.1, 53.4, 43.5, 38.2, 32.8; HRMS (FAB) calcd for C₁₄H₁₅BrO₂: 294.0255 ([M]⁺), found 294.0252.



2b: $[\alpha]^{27}_{D}$ +44.8 (*c* = 1.3, CHCl₃, 92: 8 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 18.1 min (major) and 20.7 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.41–7.32 (m, 4H), 5.68–5.57 (m, 1H),

5.16–5.09 (m, 2H), 4.46–4.39 (m, 1H), 3.59 (dd, J = 5.2, 10.8 Hz, 1H), 3.48 (dd, J = 6.8, 10.8 Hz, 1H), 2.78 (dd, J = 5.2, 13.6 Hz, 1H), 2.71–2.61 (m, 2H), 2.30 (dd, J = 10.8, 13.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 176.7, 136.8, 134.0, 132.3, 129.2, 127.6, 120.1, 75.1, 53.0, 43.6, 38.1, 32.6; IR (neat): 1768, 1494, 1173, 1012, 926, 827 cm⁻¹; HRMS (FAB) calcd for C₁₄H₁₄BrClO₂: 327.9866 ([M]⁺), found 327.9864.



2c: $[\alpha]^{28}_{D}$ +48.1 (*c* = 1.3, CHCl₃, 94: 6 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 16.6 min (major) and 18.9 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.40 (dd, *J* = 5.2, 8.8 Hz, 2H), 7.08 (dd, *J*

= 8.4, 8.8 Hz, 2H), 5.68–5.57 (m, 1H), 5.15–5.09 (m, 2H), 4.47–4.40 (m, 1H), 3.59 (dd, J = 5.2, 10.8 Hz, 1H), 3.49 (dd, J = 6.4, 10.8 Hz, 1H), 2.79 (dd, J = 5.2, 13.6 Hz, 1H), 2.71–2.62 (m, 2H), 2.30 (dd, J = 10.8, 13.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 177.0, 162.2 (d, J = 246 Hz), 133.9 (d, J = 3.3 Hz), 132.4, 127.9 (d, J = 8.2 Hz), 120.0, 115.9 (d,

J = 21.4 Hz), 75.1, 52.8, 43.8, 38.2, 32.7; IR (neat): 1769, 1509, 1235, 1174, 1166, 1013, 927, 834 cm⁻¹; HRMS (FAB) calcd for C₁₄H₁₅BrFO₂: 313.0239 ([M+H]⁺), found 313.0238.



2d: $[\alpha]^{29}_D$ +37.2 (*c* = 1.2, CHCl₃, 94: 6 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 16.0 min (major) and 18.4 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.30 (d, *J* = 8.0 Hz, 2H), 7.19 (d, *J* = 8.0

Hz, 2H), 5.70–5.59 (m, 1H), 5.14–5.09 (m, 2H), 4.47–4.40 (m, 1H), 3.58 (dd, J = 4.8, 10.8 Hz, 1H), 3.48 (dd, J = 6.8, 10.8 Hz, 1H), 2.79 (dd, J = 5.6, 13.2 Hz, 1H), 2.72–2.62 (m, 2H), 2.34 (s, 3H), 2.26 (dd, J = 10.8, 13.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 177.3, 137.7, 135.2, 132.9, 129.7, 126.0, 119.6, 75.1, 53.0, 43.5, 38.2, 32.8, 20.9; IR (neat): 2922, 1771, 1514, 1233, 1173, 1014, 924, 816 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₇BrO₂: 308.0412 ([M]⁺), found 308.0411.



2e: $[\alpha]^{26}_{D}$ +46.3 (*c* = 1.0, CHCl₃, 94: 6 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 23.1 min (major) and 31.4 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.33 (d, *J* = 8.8 Hz, 2H), 6.90 (d, *J* = 8.8

Hz, 2H), 5.69–5.58 (m, 1H), 5.13–5.08 (m, 2H), 4.48–4.41 (m, 1H), 3.81 (s, 3H), 3.58 (dd, J = 4.8, 10.8 Hz, 1H), 3.48 (dd, J = 6.4, 10.8 Hz, 1H), 2.78 (dd, J = 4.8, 13.2 Hz, 1H), 2.66 (d, J = 8.0 Hz, 2H), 2.25 (dd, J = 10.4, 13.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 177.3, 159.1, 132.8, 130.0, 127.3, 119.6, 114.3, 75.1, 55.3, 52.6, 43.7, 38.2, 32.8; IR (neat): 1769, 1512, 1253, 1172, 1031, 1016, 1009, 925, 829 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₈BrO₃: 325.0439 ([M+H]⁺), found 325.0429.

2f: $[\alpha]^{26}_{D}$ +38.9 (*c* = 0.78, CHCl₃, 91: 9 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 15.6 min (major) and 17.4 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.66 (d, *J* = 8.0 Hz, 2H), 7.56 (d, *J* = 8.4

Hz, 2H), 5.69–5.58 (m, 1H), 5.18–5.11 (m, 2H), 4.46–4.39 (m, 1H), 3.60 (dd, *J* = 4.8, 10.8 Hz, 1H), 3.50 (dd, *J* = 6.4, 10.8 Hz, 1H), 2.83 (dd, *J* = 5.2, 13.2 Hz, 1H), 2.75–2.65

(m, 2H), 2.36 (dd, J = 10.4, 13.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 176.5, 142.4, 132.0, 130.3 (q, J = 32.1 Hz), 126.7, 126.0 (q, J = 4.1 Hz), 123.8 (q, J = 271 Hz), 120.4, 75.1, 53.4, 43.5, 38.0, 32.5; IR (neat): 1771, 1328, 1170, 1119, 1071, 1015 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₅BrF₃O₂: 363.0208 ([M+H]⁺), found 363.0208.



2g: $[\alpha]^{27}_{D}$ +39.2 (*c* = 0.94, CHCl₃, 91: 9 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 13.9 min (major) and 15.6 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.46 (d, *J* = 9.2 Hz, 2H), 7.24 (d, *J* =

8.8 Hz, 2H), 5.68–5.58 (m, 1H), 5.17–5.10 (m, 2H), 4.48–4.41 (m, 1H), 3.60 (dd, J = 4.8, 10.8 Hz, 1H), 3.49 (dd, J = 6.4, 10.8 Hz, 1H), 2.80 (dd, J = 5.2, 13.6 Hz, 1H), 2.73–2.62 (m, 2H), 2.32 (dd, J = 10.8, 13.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 176.7, 148.8, 136.9, 132.2, 127.8, 121.3, 120.4 (q, J = 257 Hz), 120.2, 75.1, 53.0, 43.7, 38.0, 32.6; IR (neat): 1776, 1509, 1257, 1213, 1169, 1015, 926 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₅BrF₃O₃: 379.0157 ([M+H]⁺), found 379.0156.

2h: $[\alpha]^{22}_{D} +53.4$ (*c* = 1.3, CHCl₃, 96: 4 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 14.5 min (major) and 15.6 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.41 (s, 1H), 7.35–7.28 (m, 3H), 5.68–5.58 (m, 1H), 5.17– 5.11 (m, 2H), 4.48–4.41 (m, 1H), 3.59 (dd, *J* = 5.2, 10.8 Hz, 1H), 3.50 (dd, *J* = 6.4, 10.8 Hz, 1H), 2.78 (dd, *J* = 5.2, 13.6 Hz, 1H), 2.73–2.62 (m, 2H), 2.32 (dd, *J* = 10.8, 13.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 176.5, 140.4, 135.0, 132.2, 130.3, 128.2, 126.4, 124.4, 120.2, 75.1, 53.3, 43.5, 38.0, 32.6; IR (neat): 1777, 1173, 1016, 926 cm⁻¹; HRMS (FAB) calcd for C₁₄H₁₅BrClO₂: 328.9944 ([M+H]⁺), found 328.9939.



2i: $[\alpha]^{20}_{D}$ +51.5 (c = 1.1, CHCl₃, 93: 7 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 12.1 min (major) and 13.4 min (minor). ¹H

NMR (400 MHz, CDCl₃) δ 7.32–7.11 (m, 4H), 5.71–5.60 (m, 1H), 5.18–5.10 (m, 2H), 4.47–4.40 (m, 1H), 3.58 (dd, J = 5.2, 10.4 Hz, 1H), 3.48 (dd, J = 6.4, 10.4 Hz, 1H), 2.79 (dd, J = 5.6, 13.2 Hz, 1H), 2.73–2.63 (m, 2H), 2.36 (s, 3H), 2.28 (dd, J = 10.4, 13.2 Hz,

1H); ¹³C NMR (100 MHz, CDCl₃) δ 177.2, 138.8, 138.3, 132.9, 128.8, 128.6, 126.8, 123.0, 119.7, 75.1, 53.4, 43.5, 38.2, 32.8, 21.6; IR (neat): 1775, 1171, 1015, 925 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₈BrO₂: 309.0490 ([M+H]⁺), found 309.0491.

2j: (Major diastereomer): $[\alpha]^{21}_D - 8.3$ (c = 1.2, CHCl₃, 86:14 er); HPLC analysis: Daicel Chiralpak IC-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 28.0 min (minor) and 31.5 min (major). ¹H NMR (400 MHz, CDCl₃) δ 7.57 (dt, J = 1.6, 8.0 Hz, 1H), 7.33–7.28 (m, 1H), 7.17-7.07 (m, 2H), 5.88-5.78 (m, 1H), 5.31-5.23 (m, 2H), 4.73-4.66 (m, 1H), 3.55 (dd, J = 5.6, 10.4 Hz, 1H, 3.36 (dd, J = 7.6, 10.8 Hz, 1H), 2.86–2.75 (m, 3H), 2.40 (ddd, J =2.0, 8.0, 14.0 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 177.1, 160.4 (d, J = 245 Hz), 131.7, 129.5 (d, J = 9.6 Hz), 128.9 (d, J = 11.5 Hz), 127.6 (d, J = 3.8 Hz), 124.3 (d, J = 2.9 Hz), 120.9, 116.4 (d, J = 23.0 Hz), 75.7, 49.6, 41.4, 40.0 (d, J = 2.8 Hz), 33.2; IR (neat): 1772, 1489, 1179, 757 cm⁻¹; HRMS (FAB) calcd for C₁₄H₁₅BrFO₂: 313.0239 ([M+H]⁺), found 313.0239. (Minor diastereomer): $[\alpha]^{22}_{D}$ +32.2 (c = 0.17, CHCl₃, 74:26 er); HPLC analysis: Daicel Chiralpak IC-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 18.7 min (major) and 19.6 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.34-7.25 (m, 2H), 7.15-7.09 (m, 2H), 5.83-5.72 (m, 1H), 5.22-5.18 (m, 2H), 4.49-4.42 (m, 1H), 3.57 (dd, J = 5.2, 10.8 Hz, 1H), 3.48 (dd, J = 6.4, 10.8 Hz, 1H), 2.86–2.76 (m, 3H), 2.39 (dd, J = 10.4, 13.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 176.6, 160.7 (d, J = 246 Hz), 132.8, 129.8 (d, J = 8.6 Hz), 128.2 (d, J = 3.8 Hz), 127.1 (d, J = 11.5 Hz),124.6 (d, J = 2.8 Hz), 120.4, 116.9 (d, J = 23.0 Hz), 75.7, 52.3 (d, J = 1.9 Hz), 40.2 (d, J= 3.9 Hz, 38.3 (d, J = 4.8 Hz), 32.7; IR (neat): 1781, 1491, 1178, 1018, 758 cm⁻¹; HRMS (FAB) calcd for C₁₄H₁₅BrFO₂: 313.0239 ([M+H]⁺), found 313.0239.



2k: $[\alpha]^{27}_{D}$ +61.0 (*c* = 1.8, CHCl₃, 95: 5 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 22.3 min (major) and 26.8 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.90–7.79 (m, 4H), 7.56–7.48 (m, 3H),

5.74–5.64 (m, 1H), 5.16–5.13 (m, 2H), 4.50–4.44 (m, 1H), 3.60 (dd, J = 5.2, 10.8 Hz, 1H), 3.51 (dd, J = 6.8, 10.8 Hz, 1H), 2.93 (dd, J = 5.2, 13.2 Hz, 1H), 2.78 (d, J = 6.8 Hz, 2H), 2.38 (dd, J = 10.8, 13.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 177.1, 135.6, 133.1,

132.7, 132.6, 129.1, 128.1, 127.5, 126.6, 126.5, 125.0, 123.9, 119.8, 75.2, 53.7, 43.4, 38.3, 32.8; IR (neat): 1769, 1173, 1014, 1005, 923, 817, 748 cm⁻¹; HRMS (FAB) calcd for C₁₈H₁₈BrO₂: 345.0490 ([M+H]⁺), found 345.0490.

21: $[\alpha]^{20}_{D}$ +27.9 (c = 1.3, CHCl₃, 95: 5 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 17.7 min (major) and 20.3 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.27 (dd, J = 1.2, 5.2 Hz, 1H), 7.05 (dd, J = 1.2, 3.6 Hz, 1H), 6.99 (dd, J = 3.6, 5.2 Hz, 1H), 5.73–5.62 (m, 1H), 5.18–5.13 (m, 2H), 4.70–4.63 (m, 1H), 3.60 (dd, J = 4.8, 10.8 Hz, 1H), 3.52 (dd, J = 6.4, 10.8 Hz, 1H), 2.80–2.69 (m, 3H), 2.35 (dd, J = 10.4, 13.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 176.1, 142.5, 132.2, 127.3, 125.2, 125.1, 120.1, 75.5, 50.8, 44.4, 39.5, 32.8; IR (neat): 1768, 1173, 1010, 926, 703 cm⁻¹; HRMS (FAB) calcd for C₁₂H₁₃BrO₂S: 299.9820 ([M]⁺), found 299.9820.

2m: $[\alpha]^{20}D + 41.6$ (*c* = 0.92, CHCl₃, 96: 4 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 20.4 min (major) and 24.1 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.37 (dd, *J* = 2.8, 5.2 Hz, 1H), 7.21 (dd, *J* = 1.2, 2.8 Hz, 1H), 7.14 (dd, *J* = 1.2, 5.2 Hz, 1H), 5.69–5.59 (m, 1H), 5.15–5.10 (m, 2H), 4.58–4.51 (m, 1H), 3.60 (dd, *J* = 4.8, 10.8 Hz, 1H), 3.50 (dd, *J* = 6.4, 10.4 Hz, 1H), 2.76–2.63 (m, 3H), 2.27 (dd, *J* = 10.4, 13.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 177.0, 139.3, 132.5, 127.1, 125.9, 121.5, 119.8, 75.4, 50.8, 43.1, 38.5, 32.9; IR (neat): 1774, 1173, 1016, 927, 789 cm⁻¹; HRMS (FAB) calcd for C₁₂H₁₄BrO₂S: 300.9898 ([M+H]⁺), found 300.9899.

Ph **2n**: (**Major diastereomer**): $[\alpha]^{27}_{D}$ +3.6 (c = 0.40, CHCl₃, 85:15 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 17.5 min (major) and 18.4 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.29 (tt, J = 1.6, 7.2 Hz, 2H), 7.22–7.18 (m, 3H), 5.85–5.75 (m, 1H), 5.25–5.19 (m, 2H), 4.64–4.57 (m, 1H), 3.57 (dd, J = 4.4, 10.8 Hz, 1H), 3.50 (dd, J = 6.4, 10.8 Hz, 1H), 2.81–2.73 (m, 1H), 2.59–2.52 (m, 1H), 2.46– 2.30 (m, 3H), 2.13 (dd, J = 9.2, 13.2 Hz, 1H), 2.00–1.88 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 179.2, 141.1, 132.0, 128.5, 128.3, 126.2, 120.2, 74.8, 48.3, 41.7, 38.4, 36.2, 34.0, 30.8; IR (neat): 3027, 2925, 1768, 1176, 1020, 925, 700 cm⁻¹; HRMS (FAB) calcd for C₁₆H₁₉BrO₂: 322.0568 ([M]⁺), found 322.0568. (**Minor diastereomer**): $[\alpha]^{27}_{D}$ –9.2 (*c* = 0.56, CHCl₃, 86:14 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 19.2 min (major) and 21.9 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.29 (tt, *J* = 1.6, 7.4 Hz, 2H), 7.21 (tt, *J* = 1.6, 7.4 Hz, 1H), 7.16 (dd, *J* = 1.2, 8.0 Hz, 2H), 5.80–5.70 (m, 1H), 5.23–5.18 (m, 2H), 4.68–4.61 (m, 1H), 3.57 (dd, *J* = 4.8, 10.8 Hz, 1H), 3.45 (dd, *J* = 6.4, 10.8 Hz, 1H), 2.74–2.61 (m, 2H), 2.51 (tdd, *J* = 1.2, 6.4, 14.0 Hz, 1H), 2.42 (dd, *J* = 8.4, 14.0 Hz, 1H), 2.28 (dd, *J* = 7.2, 13.6 Hz, 1H), 2.14 (dd, *J* = 8.8, 13.6 Hz, 1H), 1.97–1.85 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 179.0, 140.8, 132.6, 128.6, 128.2, 126.3, 120.1, 75.2, 47.9, 40.4, 39.1, 36.9, 33.8, 30.6; IR (neat): 2922, 1769, 1456, 1165, 1017, 924, 746, 700 cm⁻¹; HRMS (FAB) calcd for C₁₆H₁₉BrO₂: 322.0568 ([M]⁺), found 322.0568.

20: (1.3: 1 mixture of diastereomers): $[\alpha]^{26}_{D}$ –9.2 (c = 2.3, CHCl₃, 78:22 er for major diastereomer, 76:24 er for minor diastereomer); HPLC analysis: Daicel Chiralpak IG-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time for major diastereomer: 24.8 min (major) and 29.6 min (minor); retention time for minor diastereomer: 24.0 min (minor) and 30.6 min (major). ¹H NMR (400 MHz, CDCl₃) δ 7.34–7.25 (m, 3H), 7.22–7.18 (m, 2H), 5.86–5.72 (m, 1H), 5.24–5.17 (m, 2H), 4.48–4.41 (m, 0.44H), 3.47–3.40 (m, 0.56H), 3.33 (dd, J = 4.4, 10.4 Hz, 0.56H), 3.26 (dd, J = 6.4, 10.4 Hz, 0.56H), 3.19 (d, 13.2 Hz, 0.44H), 3.08– 3.04 (m, 1H), 2.75–2.69 (m, 1H), 2.63–2.51 (m, 1.44H), 2.39–2.18 (m, 2H), 2.09–2.03 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 179.64, 179.55, 136.5, 136.0, 132.6, 132.1, 130.3, 129.7, 128.74, 128.65, 127.4, 127.2, 120.4, 120.3, 75.1, 74.9, 50.1, 50.0, 43.8, 43.1, 42.5, 42.3, 34.8, 33.8, 33.1; IR (neat): 3030, 2978, 2921, 1769, 1170, 1013, 924, 703 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₇BrO₂: 308.0412 ([M]⁺), found 308.0413.

8:⁶ $[\alpha]^{27}_{D}$ +70.2 (*c* = 1.3, CHCl₃, 94: 6 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 50:1, flow rate = 0.5 mL/min, 214 nm; retention time: 37.5 min (minor) and 41.0 min (major). ¹H NMR (400 MHz, CDCl₃) δ 7.40–7.24 (m, 10H), 4.59–4.53 (m, 1H), 3.62 (dd, *J* = 4.8, 10.8 Hz, 1H), 3.52 (dd, *J* = 6.4, 10.8 Hz, 1H), 3.17 (dd, *J* = 5.2, 13.2 Hz, 1H), 2.83 (dd, *J* = 9.6, 13.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 176.2, 141.4, 139.3, 129.0, 128.5, 127.9, 127.6, 127.4, 127.2, 74.9, 58.2, 42.2, 32.6; IR (neat): 1778, 1168, 1026, 965, 697 cm⁻¹; HRMS (FAB) calcd for C₁₇H₁₅BrO₂: 330.0255 ([M]⁺), found 330.0255.

4. Transformations of products 2a and 8

To a solution of **2a** (0.20 mmol) in THF (5 mL) was added 10wt% Pd/C (0.030 mmol). The reaction mixture was warmed to 60 °C and stirred for 4 h under H₂ atmosphere. After 4 h, the reaction mixture was diluted with ethyl acetate and filtered over celite. The filtrate was concentrated and purified by flash column chromatography on silica gel (hexane/ethyl acetate = 50:1-5:1 as eluent) to give product **9**.

9: $[\alpha]^{21}D + 38.5$ (c = 1.5, CHCl₃, 95: 5 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 15.8 min (major) and 18.0 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.43–7.28 (m, 5H), 4.46–4.40 (m, 1H), 3.60 (dd, J = 4.8, 10.8 Hz, 1H), 3.51 (dd, J = 6.4, 10.8 Hz, 1H), 2.91 (dd, J = 5.2, 13.2 Hz, 1H), 2.22 (dd, J = 10.4, 13.2 Hz, 1H), 2.01 (ddd, J = 4.8, 12.4, 14.0 Hz, 1H), 1.83 (ddd, J = 4.8, 12.4, 14.0 Hz, 1H), 1.37–1.11 (m, 2H), 0.88 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 177.7, 138.3, 128.9, 127.7, 126.1, 74.9, 53.5, 41.6, 38.7, 33.0, 18.1, 14.2; IR (neat): 2960, 2933, 2872, 1773, 1178, 1002, 699 cm⁻¹; HRMS (FAB) calcd for C₁₄H₁₇BrO₂: 296.0412 ([M]⁺), found 296.0412.

To a solution of **9** (0.16 mmol) in toluene (8 mL) was added 2,2'azobis(isobutyronitrile) (AIBN) (0.048 mmol) and tributyltin hydride (0.48 mmol). The reaction mixture was warmed to 100 °C and stirred for 22 h. After 22 h, the reaction mixture was cooled to room temperature and quenched with saturated aqueous NaHCO₃ (15 mL). The organic materials were extracted with ethyl acetate for three times (5 mL × 3). The combined extracts were dried over Na₂SO₄ and concentrated. The residue was purified by flash column chromatography on silica gel (hexane/ethyl acetate = 70:1–10:1 as eluent) to give product **10**. **10**:³ $[\alpha]^{18}{}_{D}$ +57.8 (*c* = 1.4, CHCl₃, 95: 5 er); HPLC analysis: Daicel Chiralpak ID-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 20.7 min (major) and 22.3 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, *J* = 7.6 Hz, 2H), 7.36 (t, *J* = 7.6 Hz, 2H), 7.28 (t, *J* = 7.2 Hz, 1H), 4.40–4.31 (m, 1H), 2.81 (dd, *J* = 4.8, 13.2 Hz, 1H), 2.02–1.94 (m, 2H), 1.79 (ddd, *J* = 4.8, 12.4, 14.0 Hz, 1H), 1.42 (d, *J* = 6.0 Hz, 3H), 1.36–1.08 (m, 2H), 0.87 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 178.9, 138.8, 128.8, 127.4, 126.3, 73.7, 53.9, 41.9, 41.8, 20.5, 18.1, 14.2; IR (neat): 2960, 2933, 2872, 1761, 1184 cm⁻¹; HRMS (FAB) calcd for C₁₄H₁₈O₂: 218.1307 ([M]⁺), found 218.1307.

To a solution of 2a (0.10 mmol) in CH₃CN (1 mL) was added benzylamine (0.50 mmol). The reaction mixture was warmed to 70 °C and stirred for 48 h. After 48 h, the reaction mixture was concentrated. The residue was purified by flash column chromatography on silica gel (hexane/ethyl acetate = 30:1–1:1 as eluent) to give product **11**.

11: $[\alpha]^{20}_{D}$ +29.1 (*c* = 1.3, CHCl₃, 95: 5 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 3:1, flow rate = 0.5 mL/min, 214 nm; retention time: 12.6 min (minor) and 13.9 min (major). ¹H NMR (400 MHz, CDCl₃) δ 7.42–7.22 (m, 10H), 5.69–5.58 (m, 1H), 5.11–5.06 (m, 2H), 4.45–4.38 (m, 1H), 3.81 (s, 2H), 2.90 (dd, *J* = 3.6, 12.8 Hz, 1H), 2.82 (dd, *J* = 6.8, 13.2 Hz, 1H), 2.67 (d, *J* = 7.2 Hz, 2H), 2.60 (dd, *J* = 5.2, 13.2 Hz, 1H), 2.28 (dd, *J* = 10.8, 13.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 178.0, 139.8, 138.6, 133.1, 128.9, 128.4, 128.1, 127.7, 127.1, 126.2, 119.4, 77.1, 53.8, 53.0, 52.5, 43.5, 36.6; IR (neat): 2920, 2833, 1764, 1181, 1014, 923, 740, 699 cm⁻¹; HRMS (FAB) calcd for C₂₁H₂₃NO₂: 321.1729 ([M]⁺), found 321.1729.

To a solution of **2a** (0.10 mmol) in CH₃CN (1 mL) was added K₂CO₃ (0.30 mmol) and thiophenol (0.50 mmol). The reaction mixture was warmed to 75 °C and stirred for 3 h. After 3 h, the reaction mixture was cooled to room temperature and quenched with saturated aqueous NH₄Cl (5 mL). The organic materials were extracted with ethyl acetate for three times (5 mL \times 3). The combined extracts were dried over

 Na_2SO_4 and concentrated. The residue was purified by flash column chromatography on silica gel (hexane/ethyl acetate = 80:1-5:1 as eluent) to give product **12**.

12: $[\alpha]^{19}_{D}$ +73.1 (*c* = 1.7, CHCl₃, 95: 5 er); HPLC analysis: Daicel Chiralpak IA-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 19.5 min (major) and 21.0 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.39–7.20 (m, 10H), 5.67–5.57 (m, 1H), 5.12–5.04 (m, 2H), 4.41–4.34 (m, 1H), 3.37 (dd, *J* = 4.8, 13.2 Hz, 1H), 3.07 (dd, *J* = 7.6, 13.6 Hz, 1H), 2.80 (dd, *J* = 5.2, 13.2 Hz, 1H), 2.71–2.61 (m, 2H), 2.20 (dd, *J* = 10.8, 13.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 177.5, 138.4, 134.8, 132.9, 130.2, 129.1, 128.9, 127.7, 127.0, 126.1, 119.6, 75.6, 53.1, 43.7, 38.6, 38.4; IR (neat): 3060, 2929, 1771, 1175, 1005, 921, 740, 698 cm⁻¹; HRMS (FAB) calcd for C₂₀H₂₀O₂S: 324.1184 ([M]⁺), found 324.1183.

16 was synthesized from 8 in a similar manner for the synthesis of 10.

16:³ $[\alpha]^{26}_{D}$ +102.9 (c = 0.25, CHCl₃, 94: 6 er); HPLC analysis: Daicel Chiralcel OD-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 16.7 min (minor) and 17.5 min (major). ¹H NMR (400 MHz, CDCl₃) δ 7.39–7.22 (m, 10H), 4.53–4.45 (m, 1H), 3.06 (dd, J = 4.8, 12.8 Hz, 1H), 2.60 (dd, J = 10.4, 12.8 Hz, 1H), 1.47 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 177.2, 142.0, 139.9, 128.9, 128.3, 127.7, 127.6, 127.3, 127.2, 73.6, 58.6, 45.3, 20.4; IR (neat): 1762, 1180, 699 cm⁻¹; HRMS (FAB) calcd for C₁₇H₁₆O₂: 252.1150 ([M]⁺), found 252.1149.

5. General procedure for the synthesis of α-quaternary esters 13

To a solution of **2** (0.10 mmol) in MeOH (1.0 mL) was added K_2CO_3 (0.20 mmol) at room temperature. The reaction mixture was stirred for 48 h at 25 °C. After 48 h, the reaction mixture was quenched with H₂O (10 mL). The organic materials were extracted with ethyl acetate for three times (5 mL × 3). The combined extracts were dried over Na₂SO₄ and concentrated. The residue was purified by flash column chromatography on silica gel (hexane/ethyl acetate as eluent) to give product **13**.



13a: $[\alpha]^{21}_{D}$ +15.5 (*c* = 0.76, CHCl₃, 95: 5 er); HPLC analysis: Daicel Chiralpak IC-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 25.8 min (major) and 38.1 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.37–7.32 (m, 2H), 7.29–7.23 (m, 3H), 5.61–5.51

(m, 1H), 5.13–5.06 (m, 2H), 3.69 (s, 3H), 2.97 (dd, J = 7.2, 14.0 Hz, 1H), 2.90–2.84 (m, 2H), 2.66 (dd, J = 4.4, 4.8 Hz, 1H), 2.40 (dd, J = 5.2, 14.4 Hz, 1H), 2.35 (dd, J = 2.8, 5.2 Hz, 1H), 2.06 (dd, J = 6.0, 14.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 175.2, 141.5, 133.1, 128.5, 127.1, 126.2, 119.1, 53.2, 52.2, 49.0, 47.1, 40.3, 38.1; IR (neat): 1731, 1215, 923, 701 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₈O₃: 246.1256 ([M]⁺), found 246.1256.



13h: $[\alpha]^{21}_{D}$ +18.7 (*c* = 2.2, CHCl₃, 96: 4 er); HPLC analysis: Daicel Chiralpak IC-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 19.7 min (major) and 29.4 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.30–7.23 (m, 3H), 7.13 (td, *J* = 1.6, 7.2 Hz, 1H),

5.58–5.48 (m, 1H), 5.13–5.07 (m, 2H), 3.70 (s, 3H), 2.96 (dd, J=7.6, 14.0 Hz, 1H), 2.87–2.82 (m, 2H), 2.68 (dd, J=4.0, 4.8 Hz, 1H), 2.37 (dd, J=2.4, 5.2 Hz, 1H), 2.32 (dd, J=5.6, 14.4 Hz, 1H), 2.11 (dd, J=6.0, 14.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 174.6, 143.5, 134.5, 132.6, 129.7, 127.4, 126.6, 124.6, 119.5, 53.1, 52.4, 48.8, 46.9, 40.1, 37.9; IR (neat): 2952, 2925, 2853, 1731, 1215, 923 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₇ClO₃: 280.0866 ([M]⁺), found 280.0866.

131: $[\alpha]^{23}_{D}$ +9.7 (c = 2.0, CHCl₃, 95: 5 er); HPLC analysis: Daicel Chiralpak IF-3, hexane/2-propanol = 20:1, flow rate = 0.5 mL/min, 214 nm; retention time: 19.1 min (minor) and 21.9 min (major). ¹H NMR

(400 MHz, CDCl₃) δ 7.24 (dd, J = 1.6, 4.8 Hz, 1H), 6.99–6.95 (m, 2H), 5.68–5.57 (m, 1H), 5.16–5.08 (m, 2H), 3.73 (s, 3H), 3.00 (dd, J = 7.2, 14.0 Hz, 1H), 2.93–2.87 (m, 2H), 2.69 (dd, J = 4.4, 4.8 Hz, 1H), 2.42 (dd, J = 2.8, 4.8 Hz, 1H), 2.38 (dd, J = 5.6, 14.4 Hz, 1H), 2.20 (dd, J = 6.0, 14.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 173.9, 145.4, 132.8, 126.6, 124.8, 124.5, 119.3, 52.5, 51.4, 48.9, 47.0, 42.1, 39.7; IR (neat): 2924, 1732, 1216, 700 cm⁻¹; HRMS (FAB) calcd for C₁₃H₁₆O₃S: 252.0820 ([M]⁺), found 252.0820.

6. Determination of the absolute configuration of products

The absolute stereochemistry of products 10 and 16 were confirmed by comparison with reported data.³

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NMR Charts


























































































































































HPLC Charts
















































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