

Supporting Information

Enantioselective Total Synthesis of (+)-Rubrobramide, (+)-Talaramide A, and (-)-Berkeleyamide D by a Skeletal Diversification Strategy

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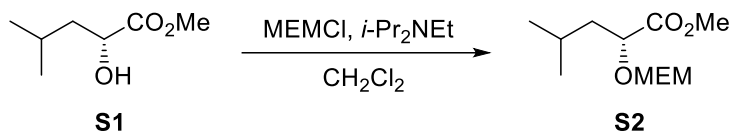
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Materials and Methods: ¹H NMR spectra were measured at 300 MHz (JNM-AL300, JEOL) or 400 MHz (JNM-AL400, ECZ400S, JEOL). Chemical shifts are expressed in ppm relative to tetramethylsilane ($\delta = 0$) as an internal standard (CDCl₃ or DMSO-*d*₆). Splitting patterns are indicated as follows: s, singlet; d, doublet; t, triplet; q, quartet; sept, septet; m, multiplet; br, broad peak. ¹³C NMR spectra were measured at 100 MHz (JNM-AL400, JEOL). The chemical shifts are reported in ppm, relative to the central line of the triplet at 77.0 ppm for CDCl₃ or the septet at 39.6 ppm for DMSO-*d*₆. Infrared spectra (IR) were measured on an IR spectrometer (VALOR-III, JASCO) and are reported in wavenumbers (cm⁻¹). High-resolution mass spectra (HRMS) were obtained using a mass spectrometer (JMS 700, JEOL) with a direct inlet system. Optical rotations were measured on a polarimeter (P-2200, JASCO) using a 100 mm pathlength cell. Melting points (m.p.) were measured on a Micro Melting Point system (Yanaco). Column chromatography was carried out on silica gel (40–100 mesh). Analytical thin-layer chromatography (TLC) was performed using 0.25 mm silica gel 60-F plates.

Experimental Procedures and Characterization Data

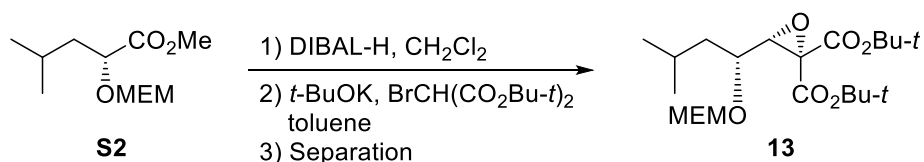
Methyl (*R*)-2-((2-methoxyethoxy)methoxy)-4-methylpentanoate (**S2**)



MEMCl (14.6 mL, 123 mmol) was slowly added to a stirred solution of secondary alcohol **S1** (7.02 g, 48.0 mmol) and *i*-Pr₂NEt (50 mL, 287 mmol) in CH₂Cl₂ (400 mL) over 5 min at 0 °C. The reaction mixture was heated to 40 °C (oil bath). After stirring for 48 h, the reaction was cooled to room temperature and quenched by addition of 1 N HCl. The organic layers were washed with saturated aqueous NaHCO₃ solution and dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (5:1 hexane/EtOAc) to afford MEM-protected ester **S2** (7.40 g, 66% yield) as a colorless liquid.

$[\alpha]_D^{26} +87.3$ (*c* 1.05, CHCl₃); IR (CHCl₃) 2955, 2873, 1753, 1176, 1114, 1028 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 4.76 (d, *J* = 1.0 Hz, 2H), 4.19 (dd, *J* = 8.8, 4.4 Hz, 1H), 3.74-3.71 (m, 2H), 3.54-3.51 (m, 2H), 3.38 (s, 3H), 1.84-1.74 (m, 1H), 1.70 (ddd, *J* = 14.2, 8.8, 5.4 Hz, 1H), 1.51 (ddd, *J* = 13.6, 8.8, 4.4 Hz, 1H), 0.93 (d, *J* = 6.8 Hz, 3H), 0.93 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.6, 95.3, 74.4, 71.6, 67.6, 59.0, 51.8, 41.8, 24.4, 23.1, 21.6; HRMS (FAB-DFMS) *m/z*: [M+H]⁺ Calcd for C₁₁H₂₃O₅ 235.1545; Found 235.1538.

Di-*tert*-butyl (S)-3-((*R*)-1-((2-methoxyethoxy)methoxy)-3-methylbutyl)oxirane-2,2-dicarboxylate (**13**)



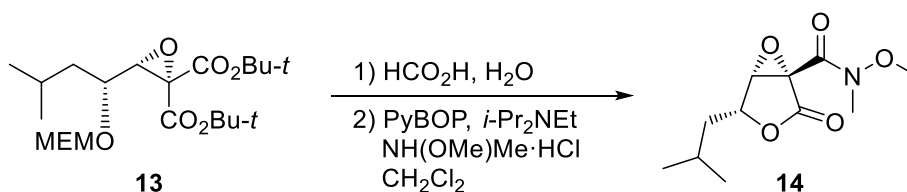
DIBAL-H (1.02 M solution in 31 mL hexane, 31 mmol) was slowly added to a stirred solution of ester **S2** (4.80 g, 20.5 mmol) in CH₂Cl₂ (200 mL) at -78 °C via dropping funnel over 20 min. After stirring for 1 h, the reaction was quenched by addition of MeOH and saturated aqueous Rochelle salt, and vigorously stirred for 1 h. The mixture was extracted with EtOAc three times. The combined organic layers were washed with water

and brine, dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude product was used in the next reaction without further purification.

A suspension of *t*-BuOK (2.40 g, 21.1 mmol) in toluene (50 mL) was added dropwise to di-*tert*-butyl bromomalonate (6.70 g, 22.7 mmol) in toluene (110 mL) at -78 °C. After stirring for 30 min, the crude aldehyde in toluene (50 mL) was added dropwise. After complete addition, the reaction mixture was allowed to warm to -45 °C gradually and was stirred for 12 h. The reaction was quenched by adding saturated aqueous NH₄Cl, and the mixture was extracted three times with EtOAc. The combined organic layers were washed with water and brine, dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (5:1 hexane/EtOAc) to afford epoxide **13** (5.23 g, 61% yield) as a colorless oil.

[α]_D²² +33.6 (*c* 1.23, CHCl₃); IR (CHCl₃) 2983, 2960, 1740, 1371, 1255, 1165, 1124, 1046, 1028 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 4.96 (d, *J* = 6.8 Hz, 1H), 4.75 (d, *J* = 6.8 Hz, 1H), 3.79-3.74 (m, 1H), 3.71-3.65 (m, 1H), 3.55-3.53 (m, 2H), 3.45 (dt, *J* = 8.6, 2.4 Hz, 1H), 3.41 (d, *J* = 8.6 Hz, 1H), 3.37 (s, 3H), 1.89-1.79 (m, 1H), 1.62 (ddd, *J* = 14.0, 10.0, 4.0 Hz, 1H), 1.50 (s, 9H), 1.48 (s, 9H), 1.26 (m, 1H), 0.93 (d, *J* = 6.3 Hz, 3H), 0.86 (d, *J* = 6.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.7, 163.5, 94.8, 83.7, 83.5, 73.0, 71.7, 67.4, 64.3, 59.8, 59.0, 40.5, 27.9 (3C), 27.8 (3C), 23.8, 23.6, 21.2; HRMS (FAB-DFMS) *m/z*: [M+H]⁺ Calcd for C₂₁H₃₉O₈ 419.2645; Found 419.2638.

(1*R*,4*R*,5*S*)-4-Isobutyl-*N*-methoxy-*N*-methyl-2-oxo-3,6-dioxabicyclo[3.1.0]hexane-1-carboxamide (**14**)



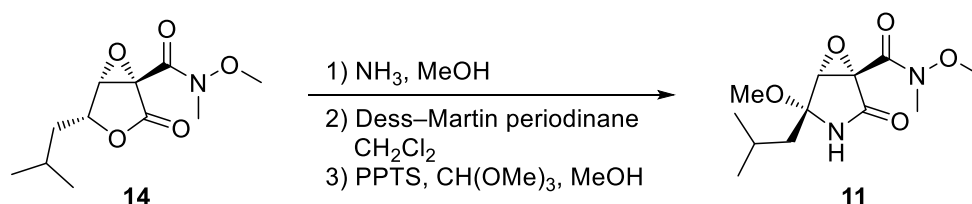
Epoxide **13** (2.00 g, 4.78 mmol) was dissolved in HCO₂H (30 mL) and H₂O (10 mL), and the solution was stirred at 35 °C for 12 h. The reaction mixture was concentrated *in vacuo*. The crude product was used in the next reaction without further purification.

N,*O*-dimethylhydroxylamine hydrochloride (1.40 g, 14.4 mmol), *i*-Pr₂NEt (5.0 mL, 34.4 mmol), and PyBOP (5.0 g, 9.60 mmol) were added successively to a stirred solution of the crude mono carboxylic acid **12** in CH₂Cl₂ (100 mL) at 0 °C. After stirring at room temperature for 2 h, the reaction was quenched by addition of 1 N HCl. The mixture was extracted with CH₂Cl₂ three times and washed with saturated aqueous NaHCO₃. The

combined organic layers were dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (1:1 hexane/EtOAc) to afford Weinreb amide **14** (1.00 g, 87% yield over two steps) as a colorless solid.

m.p. 67.5-69.5 °C; [α]_D²⁴ +61.4 (*c* 0.61, CHCl₃); IR (CHCl₃) 3019, 2963, 1787, 1687, 1682, 1468, 1223, 1216, 1072, 938 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 4.61 (dd, *J* = 6.8, 5.9 Hz, 1H), 4.21 (s, 1H), 3.74 (s, 3H), 3.28 (s, 3H), 1.89-1.75 (m, 2H), 1.63-1.56 (m, 1H), 0.97 (d, *J* = 6.8 Hz, 3H), 0.99 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 167.2, 161.5, 77.2, 62.0, 61.3, 58.9, 38.2, 32.2, 24.8, 22.9, 22.1; HRMS (FAB-DFMS) *m/z*: [M+H]⁺ Calcd for C₁₁H₁₈NO₅ 244.1185; Found 244.1184.

(1*S*,4*R*,5*R*)-4-Isobutyl-*N*,4-dimethoxy-*N*-methyl-2-oxo-6-oxa-3-azabicyclo[3.1.0]hexane-1-carboxamide (**11**)



NH₃ (21 mmol, 7 M solution in 3 mL MeOH) was added to a stirred solution of Weinreb amide **14** (770 mg, 3.16 mmol) in MeOH (20 mL) at 0 °C. After stirring at 0 °C for 30 min, additional NH₃ (14 mmol, 7 M solution in 2 mL MeOH) was added. After 30 min, the reaction was concentrated *in vacuo*. The crude product was used in the next reaction without further purification.

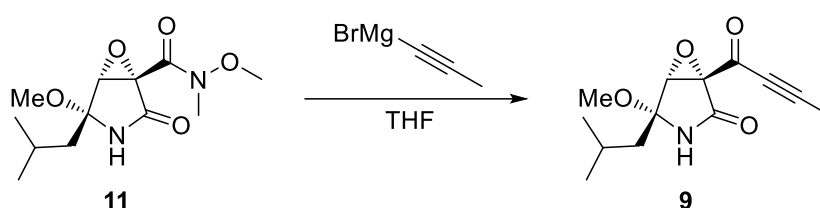
Dess–Martin periodinane (1.95 g, 4.60 mmol) was added to a solution of the crude product in CH₂Cl₂ (30 mL) and the mixture was stirred at room temperature for 1 h. The reaction was quenched by addition of saturated aqueous NaHCO₃ and Na₂S₂O₃ solution and stirred vigorously for 10 min. The mixture was extracted with CHCl₃ three times. The combined organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude product was used in the next reaction without further purification.

CH(OMe)₃ (3 mL) and PPTS (800 mg, 3.18 mmol) were added to a stirred solution of the crude product in MeOH (40 mL). The reaction was heated to reflux (oil bath) and stirred for 48 h. After cooling to room temperature, the reaction was quenched by addition of Et₃N (4.4 mL) and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (3:1 to 1:1 hexane/EtOAc) to afford lactam **11** (620 mg, 72% yield in three steps) as a colorless oil.

[α]_D²⁵ -45.6 (*c* 8.2, CHCl₃); IR (neat) 3503, 3272, 2957, 1725, 1673, 1464, 1062, 763 cm⁻¹;

^1H NMR (500 MHz, CDCl_3) δ 7.03 (br s, 1H), 3.99 (d, $J = 2.6$ Hz, 1H), 3.70 (s, 3H), 3.27 (s, 3H), 3.23 (s, 3H), 1.87 (sept, $J = 6.5$ Hz, 1H), 1.73 (dd, $J = 14.5, 5.1$ Hz, 1H), 1.58 (dd, $J = 14.5, 7.9$ Hz, 1H), 0.97 (d, $J = 6.5$ Hz, 3H), 0.94 (d, $J = 6.5$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 169.3, 162.4, 89.6, 61.9, 61.4, 60.3, 49.8, 41.5, 32.2, 24.1, 23.9, 23.4; HRMS (FAB-DFMS) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{12}\text{H}_{21}\text{N}_2\text{O}_5$ 273.1450; Found 273.1446.

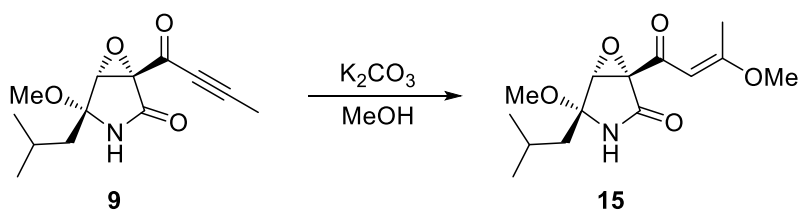
(1*R*,4*R*,5*R*)-1-(But-2-ynoyl)-4-isobutyl-4-methoxy-6-oxa-3-azabicyclo[3.1.0]hexan-2-one (**9**)



1-Propynyl magnesium bromide (4.5 mmol, 0.5 M in 9 mL THF) was added to a stirred solution of lactam **11** (200 mg, 0.734 mmol) in THF (7 mL) at -55 °C. The reaction was heated to -40 °C, stirred for 1 h, and quenched by addition of saturated aqueous NH_4Cl solution. The reaction mixture was extracted with EtOAc three times. The combined organic layers were dried over Na_2SO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (3:1 hexane/EtOAc) to afford alkyne ketone **9** (182 mg, 99% yield) as a yellow oil.

$[\alpha]_{\text{D}}^{25}$ -166.1 (c 4.5, CHCl_3); IR (neat) 3280, 2959, 2215, 1732, 1666, 1399, 763 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 6.88 (br s, 1H), 4.06 (d, $J = 2.9$ Hz, 1H), 3.25 (s, 3H), 2.07 (s, 3H), 1.93 (sept, $J = 6.3$ Hz, 1H), 1.75 (dd, $J = 14.6, 5.4$ Hz, 1H), 1.64 (dd, $J = 14.6, 7.8$ Hz, 1H), 1.01 (d, $J = 6.3$ Hz, 3H), 0.97 (d, $J = 6.3$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 175.5, 167.7, 96.3, 88.8, 77.6, 63.7, 61.9, 49.6, 42.3, 24.0, 23.8, 23.5, 4.5; HRMS (FAB-DFMS) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{13}\text{H}_{18}\text{NO}_4$ 252.1236; Found 252.1245.

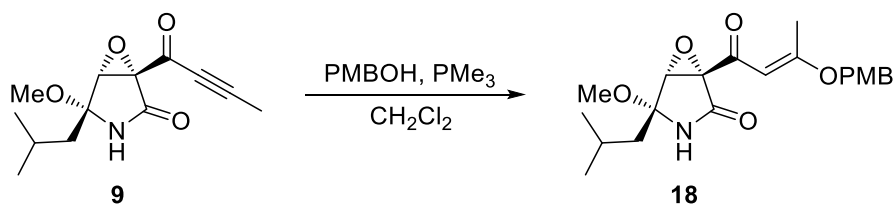
(1*R*,4*R*,5*R*)-4-Isobutyl-4-methoxy-1-(3-methoxybut-2-enoyl)-6-oxa-3-azabicyclo[3.1.0]hexan-2-one (**15**)



A solution of alkynyl ketone **9** (32.5 mg, 0.129 mmol) in MeOH (1.5 mL) was added to a suspension of K₂CO₃ (18 mg, 0.13 mmol) in MeOH (12 mL) over 5 min at 0 °C. After stirring for 10 min, the reaction was concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (3:1 hexane/EtOAc) to afford methoxy vinylogous ester **15** (34 mg, 93% yield) as a yellow oil.

[α]_D²⁶ -60.9 (*c* 1.5, CHCl₃); IR (neat) 3284, 2957, 1728, 1577, 1060, 810 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.41 (br s, 1H), 5.80 (s, 1H), 3.91 (d, *J* = 2.4 Hz, 1H), 3.71 (s, 3H), 3.27 (s, 3H), 2.37 (s, 3H), 1.94 (sept, *J* = 6.8 Hz, 1H), 1.72 (dd, *J* = 14.6, 5.9 Hz, 1H), 1.67 (dd, *J* = 14.6, 7.8 Hz, 1H), 1.02 (d, *J* = 6.8 Hz, 3H), 0.97 (d, *J* = 6.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 186.5, 178.0, 169.6, 94.5, 88.7, 63.4, 61.7, 56.1, 49.7, 42.8, 24.1, 23.9, 23.5, 20.7; HRMS (FAB-DFMS) *m/z*: [M+H]⁺ Calcd for C₁₄H₂₁NO₅ 284.1498; Found 284.1506.

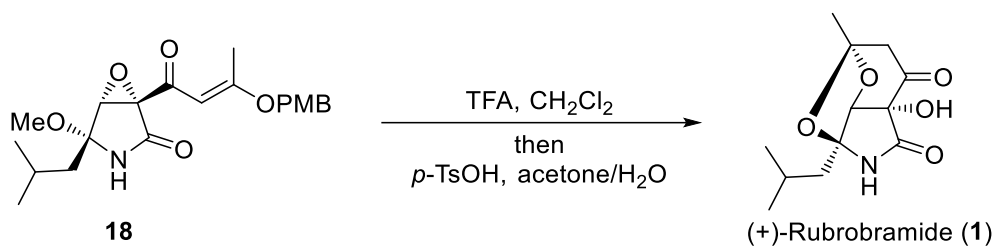
(1*R*,4*R*,5*R*)-4-Isobutyl-4-methoxy-1-(3-((4-methoxybenzyl)oxy)but-2-enoyl)-6-oxa-3-azabicyclo[3.1.0]hexan-2-one (**18**)



A solution of **9** (73 mg, 0.29 mmol) in CH₂Cl₂ (9 mL) was slowly added to a stirred solution of PMBOH (120 mg, 0.868 mmol) and PMe₃ (1.0 M in THF 0.06 mL, 0.06 mmol) in CH₂Cl₂ (20 mL) over 5 min at -20 °C. After stirring for 1 h, the reaction mixture was diluted with *n*-hexane and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (3:1 hexane/EtOAc) to afford **18** (101 mg, 89% yield) as a yellow oil.

[α]_D²³ -66.4 (*c* 2.1, CHCl₃); IR (neat) 3284, 2957, 1726, 1573, 1252, 1033, 824, 758 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.29 (d, *J* = 8.8 Hz, 2H), 6.92 (d, *J* = 8.8 Hz, 2H), 5.96 (s, 1H), 5.92 (br s, 1H), 4.86 (s, 2H), 3.91 (d, *J* = 2.7 Hz, 1H), 3.82 (s, 3H), 3.27 (s, 3H), 2.42 (s, 3H), 1.95 (sept, *J* = 6.8 Hz, 1H), 1.71 (dd, *J* = 6.8, 1.6 Hz, 2H), 1.04 (d, *J* = 6.8 Hz, 3H), 0.99 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 186.4, 177.0, 169.4, 159.9, 129.7 (2C), 126.8, 114.1 (2C), 95.6, 88.6, 70.9, 63.4, 61.6, 55.3, 49.7, 43.0, 24.1, 23.9, 23.6, 21.0; HRMS (FAB-DFMS) *m/z*: [M+H]⁺ Calcd for C₂₁H₂₇NO₆ 390.1917; Found 390.1910.

(+)-Rubrobramide (**1**)



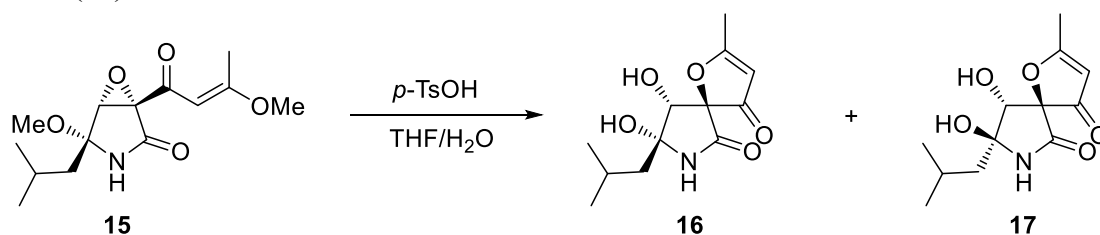
TFA (0.1 mL) was added to a stirred solution of *p*-methoxy benzyloxy vinyllogous ester **18** (44 mg, 0.11 mmol) in CH₂Cl₂ (10 mL). The reaction mixture was stirred at room temperature for 18 h and concentrated with toluene *in vacuo*. The crude product was used in the next reaction without further purification.

p-TsOH (33 mg, 0.17 mmol) was added to a stirred solution of the crude product in acetone (4.5 mL) and H₂O (1.5 mL). The reaction mixture was heated to reflux (oil bath) and stirred for 6 days. The reaction was quenched by addition of saturated aqueous NaHCO₃. The mixture was extracted with CHCl₃ three times, and the organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (3:1 hexane/EtOAc) to afford **1** (14.6 mg, 51% yield) as a colorless solid.

m.p. 131-132.5 °C; [α]_D²⁴ +224.1 (*c* 0.63, CHCl₃); IR (CHCl₃) 3275, 2959, 1748, 1717, 1389, 1230, 1191, 757 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.58 (br s, 1H), 4.88 (s, 1H), 4.23 (s, 1H), 2.97 (d, *J* = 18.5 Hz, 1H), 2.85 (d, *J* = 18.5 Hz, 1H), 1.92-1.87 (m, 1H), 1.86-1.81 (m, 2H), 1.63 (s, 3H), 1.03 (d, *J* = 6.5 Hz, 3H), 1.02 (d, *J* = 6.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 199.2, 168.4, 109.1, 92.8, 86.5, 82.4, 48.6, 45.5, 25.2, 24.3, 23.7, 23.5; HRMS (FAB-DFMS) *m/z*: [M+H]⁺ Calcd for C₁₂H₁₈NO₅ 256.1185; Found 256.1184.

(5*S*,8*R*,9*R*)-8,9-Dihydroxy-8-isobutyl-2-methyl-1-oxa-7-azaspiro[4.4]non-2-ene-4,6-dione (**16**)

(5*S*,8*S*,9*R*)-8,9-Dihydroxy-8-isobutyl-2-methyl-1-oxa-7-azaspiro[4.4]non-2-ene-4,6-dione (**17**)

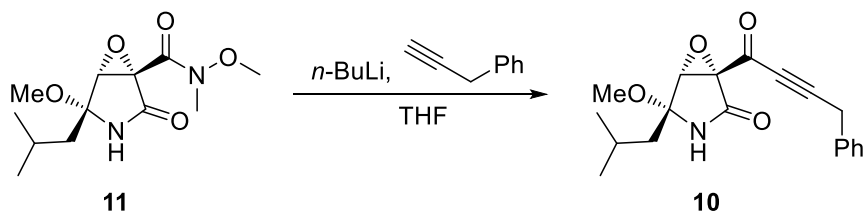


p-TsOH (19.5 mg, 0.103 mmol) was added to a stirred solution of methoxy vinylogous ester **15** (29.1 mg, 0.103 mmol) in THF (9 mL) and H₂O (1 mL). The reaction mixture was stirred at 40 °C for 5 days. The reaction was diluted with CHCl₃ and H₂O and extracted with CHCl₃ three times. The organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (1:1 to 1:3 hexane/EtOAc) to afford **16** (19.0 mg, 72% yield) and **17** (7.1 mg, 27% yield) as colorless solids.

Compound **16**: m.p. 136-139.5 °C; [α]_D²⁵ -27.8 (*c* 0.17, CHCl₃); IR (CHCl₃) 3281, 2958, 1731, 1686, 1596, 1336, 1162, 1124, 756 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.33 (br s, 1H), 6.05 (d, *J* = 5.9 Hz, 1H), 5.70 (s, 1H), 5.35 (s, 1H), 4.35 (d, *J* = 5.9 Hz, 1H), 2.37 (s, 3H), 1.85 (sept, *J* = 6.8 Hz, 1H), 1.62 (dd, *J* = 14.4, 6.3 Hz, 1H), 1.58 (dd, *J* = 14.4, 6.3 Hz, 1H), 0.92 (d, *J* = 6.8 Hz, 3H), 0.92 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 194.8, 164.1, 103.8, 95.9, 85.3, 73.9, 45.4, 24.3, 23.9, 23.4, 16.6; HRMS (FAB-DFMS) *m/z*: [M+Na]⁺ Calcd for C₁₂H₁₇NO₅Na 278.1004; Found 278.1004.

Compound **17**: m.p. 146.5 °C (decomp.); [α]_D²⁵ +25.3 (*c* 0.17, MeOH); IR (CHCl₃) 3410, 3243, 2952, 2959, 2867, 1732, 1680, 1584, 1338, 1157, 1127 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.95 (br s, 1H), 5.97 (d, *J* = 5.9 Hz, 1H), 5.78 (s, 1H), 5.44 (s, 1H), 4.38 (d, *J* = 5.9 Hz, 1H), 2.27 (s, 3H), 2.02 (dd, *J* = 14.1, 4.9 Hz, 1H), 1.94 (sept, *J* = 6.8 Hz, 1H), 1.59 (dd, *J* = 14.1, 6.8 Hz, 1H), 0.95 (d, *J* = 6.8 Hz, 3H), 0.92 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 197.1, 190.9, 163.7, 103.8, 94.8, 87.6, 81.8, 43.8, 25.0, 24.8, 22.5, 16.3; HRMS (FAB-DFMS) *m/z*: [M+Na]⁺ Calcd for C₁₂H₁₇NO₅Na 278.1004; Found 278.1000.

(1*R*,4*R*,5*R*)-4-Isobutyl-4-methoxy-1-(4-phenylbut-2-ynoyl)-6-oxa-3-azabicyclo[3.1.0]hexan-2-one (**10**)

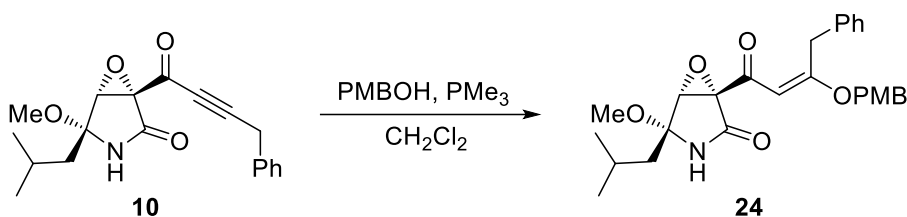


n-BuLi (0.93 mmol, 2.65 M in 0.35 mL *n*-hexane) was added to a stirred solution of 3-phenyl-1-propyne (74 mg, 0.27 mmol) in THF (5 mL) at -78 °C. After stirring for 30 min, lactam **11** (74 mg, 0.27 mmol) in THF (2 mL) was added. The reaction mixture was heated to -45 °C and stirred for 2 h. The reaction was quenched by addition of AcOH (0.05 mL, 0.29 mmol) in THF (1 mL) and H₂O, and the mixture was extracted with EtOAc three

times. The combined organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (3:1 hexane/EtOAc) to afford alkynyl ketone **10** (87 mg, 98% yield) as a yellow oil.

$[\alpha]_{\text{D}}^{23} +6.5$ (*c* 5.6, CHCl₃); IR (neat) 3259, 2958, 2213, 1728, 1668, 1398, 761, 698 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.37-7.28 (m, 5H), 5.68 (br s, 1H), 4.07 (d, *J* = 2.7 Hz, 1H), 3.84 (s, 2H), 3.22 (s, 3H), 1.94 (sept, *J* = 6.6 Hz, 1H), 1.71 (d, *J* = 6.6 Hz, 2H), 1.03 (d, *J* = 6.6 Hz, 3H), 0.99 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 175.4, 167.7, 133.4, 128.8 (2C), 127.9 (2C), 127.3, 97.2, 88.8, 79.6, 63.8, 62.1, 49.6, 42.3, 25.5, 24.0, 23.8, 23.4; HRMS (FAB-DFMS) *m/z*: [M+H]⁺ Calcd for C₁₉H₂₂NO₄ 328.1549; Found 328.1555.

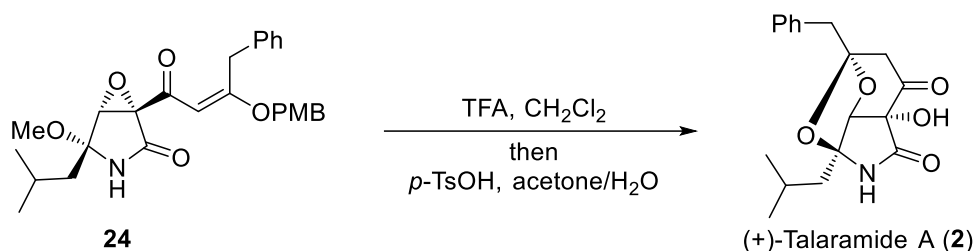
(1*R*,4*R*,5*R*)-4-Isobutyl-4-methoxy-1-(3-((4-methoxybenzyl)oxy)-4-phenylbut-2-enoyl)-6-oxa-3-azabicyclo[3.1.0]hexan-2-one (**24**)



A solution of alkynyl ketone **10** (44.0 mg, 0.134 mmol) in CH₂Cl₂ (3 mL) was slowly added to a stirred solution of PMBOH (56 mg, 0.41 mmol) and PMe₃ (1.0 M in THF, 0.05 mL, 0.05 mmol) in CH₂Cl₂ (10 mL) over 8 min at -20 °C. After stirring for 2 h, the reaction mixture was diluted with *n*-hexane and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (3:1 hexane/EtOAc) to afford *p*-methoxy benzyloxy vinylogous ester **24** (20.1 mg, 32% yield) as a yellow oil.

$[\alpha]_{\text{D}}^{22} -52.5$ (*c* 1.2, CHCl₃); IR (neat) 3275, 2956, 2937, 1726, 1570, 1251, 822 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.38-7.20 (m, 5H), 7.19 (d, *J* = 8.8 Hz, 2H), 6.88 (d, *J* = 8.8 Hz, 2H), 6.05 (br s, 1H), 5.96 (s, 1H), 4.86 (s, 2H), 4.25 (d, *J* = 13.7 Hz, 1H), 4.12 (d, *J* = 13.7 Hz, 1H), 3.90 (d, *J* = 2.4 Hz, 1H), 3.81 (s, 3H), 3.24 (s, 3H), 1.94 (sept, *J* = 6.8 Hz, 1H), 1.71 (s, 1H), 1.70 (d, *J* = 2.4 Hz, 1H), 1.03 (d, *J* = 6.8 Hz, 3H), 0.99 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 186.1, 177.1, 169.2, 159.8, 136.9, 129.4 (2C), 129.3 (2C), 128.3 (2C), 126.8, 126.6, 114.0 (2C), 95.8, 88.6, 70.9, 63.5, 61.6, 55.3, 49.6, 42.9, 38.7, 24.1, 23.9, 23.5; HRMS (FAB-DFMS) *m/z*: [M+H]⁺ Calcd for C₂₇H₃₂NO₆ 466.2230; Found 466.2235.

(-)-Talamamide (**2**)

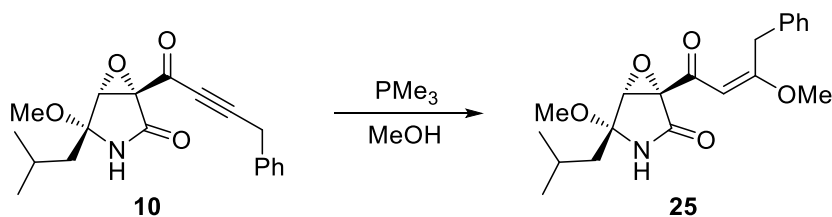


TFA (0.05 mL) was added to a stirred solution of **24** (28 mg, 0.060 mmol) in CH₂Cl₂ (5 mL). The reaction mixture was stirred at room temperature for 19 h and concentrated with toluene *in vacuo*. The crude product was used in the next reaction without further purification.

p-TsOH (11 mg, 0.058 mmol) was added to a stirred solution of the crude product in acetone (3 mL) and H₂O (1 mL). The reaction mixture was heated to reflux (oil bath) and stirred for 5 days. The reaction was quenched by addition of saturated aqueous NaHCO₃. The mixture was extracted with CHCl₃ three times, the organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (5:1 to 3:1 hexane/EtOAc) to afford talaramide (**2**) (12 mg, 62% yield) as a colorless solid.

m.p. 168-173 °C; [α]_D²⁴ +163.1 (*c* 0.82, CHCl₃); IR (CHCl₃) 3312, 3216, 2964, 2930, 1744, 1720, 760 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.37 (s, 1H), 7.33-7.23 (m, 5H), 6.58 (s, 1H), 4.76 (s, 1H), 3.11 (s, 2H), 2.79 (d, *J* = 18.5 Hz, 1H), 2.68 (d, *J* = 18.5 Hz, 1H), 1.71-1.62 (m, 1H), 1.58-1.51 (m, 2H), 0.84 (d, *J* = 7.3 Hz, 3H), 0.83 (d, *J* = 7.3 Hz, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 200.3, 168.7, 134.4, 130.7 (2C), 127.9 (2C), 126.7, 108.7, 93.0, 86.3, 83.2, 47.5, 44.2, 43.1, 23.8, 23.4, 23.0; HRMS (FAB-DFMS) *m/z*: [M+H]⁺ Calcd for C₁₈H₂₂NO₅ 332.1498; Found 332.1500.

(1*R*,4*R*,5*R*)-4-Isobutyl-4-methoxy-1-(3-methoxy-4-phenylbut-2-enoyl)-6-oxa-3-azabicyclo[3.1.0]hexan-2-one (**25**)

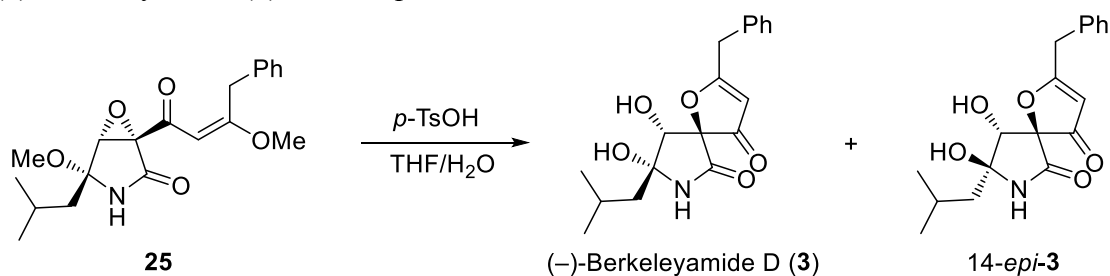


A solution of alkynyl ketone **10** (38.1 mg, 0.122 mmol) in CH₂Cl₂ (2 mL) was slowly added to a stirred solution of MeOH (0.05 mL, 1.2 mmol) and PMe₃ (1.0 M in THF, 0.02

mL, 0.02 mmol) in CH₂Cl₂ (10 mL) over 8 min at –20 °C. Additional PMe₃ (1.0 M in THF, 0.02 mL, 0.02 mmol) was added after stirring for 50 min and for 1.5 h. After stirring for 30 min, the reaction mixture was diluted with *n*-hexane and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (3:1 hexane/EtOAc) to afford vinylogous ester **25** (25.2 mg, 60% yield) as a red oil.

$[\alpha]_D^{26}$ –94.1 (*c* 1.35, CHCl₃); IR (neat) 3273, 2957, 1727, 1574, 1150, 812, 759, 701 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.35-7.18 (m, 5H), 6.39 (br s, 1H), 5.85 (s, 1H), 4.24 (d, *J* = 13.7 Hz, 1H), 4.09 (d, *J* = 13.7 Hz, 1H), 3.94 (d, *J* = 2.4 Hz, 1H), 3.71 (s, 3H), 3.27 (s, 3H), 1.94 (sept, *J* = 6.3 Hz, 1H), 1.73 (dd, *J* = 14.6, 5.9 Hz, 1H), 1.68 (dd, *J* = 14.6, 7.3 Hz, 1H), 1.03 (d, *J* = 6.3 Hz, 3H), 0.99 (d, *J* = 6.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 186.5, 178.2, 169.5, 136.8, 129.2 (2C), 128.3 (2C), 126.6, 94.7, 88.7, 63.4, 61.7, 56.4, 49.7, 42.7, 38.5, 24.1, 23.9, 23.5; HRMS (FAB-DFMS) *m/z*: [M+H]⁺ Calcd for C₂₀H₂₅NO₅ 360.1811; Found 360.1809.

(–)-Berkeleyamide (**3**) and 14-*epi*-**3**



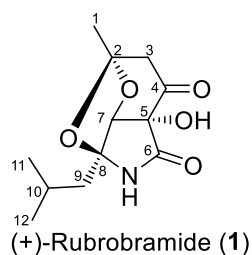
p-TsOH (7.4 mg, 0.039 mmol) was added to a stirred solution of **25** (14 mg, 0.039 mmol) in THF (2 mL) and H₂O (0.2 mL). The reaction mixture was stirred at 40 °C for 3 days. The reaction was diluted with CHCl₃ and H₂O and extracted with CHCl₃ three times. The organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (3:1 to 1:1 hexane/EtOAc) to afford berkeleyamide D (**3**) (6.3 mg, 49% yield) and 14-*epi*-**3** (5.4 mg, 42% yield) as colorless solids.

Berkeleyamide D (**3**): m.p. 114-121 °C; $[\alpha]_D^{26}$ –61.1 (*c* 0.54, MeOH); IR (CHCl₃) 3286, 2959, 1732, 1685, 1583, 758 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.39-7.29 (m, 5H), 6.72 (br s, 1H), 5.49 (s, 1H), 5.37 (s, 1H), 4.43 (d, *J* = 9.3 Hz, 1H), 4.02 (d, *J* = 17.6 Hz, 1H), 3.96 (d, *J* = 17.6 Hz, 1H), 3.03 (d, *J* = 10.2 Hz, 1H), 1.94 (sept, *J* = 6.3 Hz, 1H), 1.88 (dd, *J* = 14.6, 6.3 Hz, 1H), 1.62 (dd, *J* = 7.3, 6.3 Hz, 1H), 1.02 (d, *J* = 6.3 Hz, 3H), 1.01 (d, *J* = 6.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 199.4, 197.8, 163.9, 133.2, 129.2 (2C), 129.0 (2C), 127.8, 104.4, 95.3, 84.9, 75.2, 45.6, 37.4, 24.0, 23.9, 23.8; HRMS (FAB-

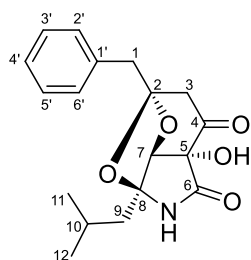
DFMS) m/z : $[M+H]^+$ Calcd for $C_{18}H_{22}NO_5$ 332.1498; Found 332.1497.

Compound 14-*epi*-**3**: m.p. 126.5-130.5 °C; $[\alpha]_D^{23}$ -18.1 (c 0.67, $CHCl_3$); IR ($CHCl_3$) 3474, 3363, 3019, 1744, 1710, 1675, 1572, 669 cm^{-1} ; 1H NMR (400 MHz, $DMSO-d_6$) δ 8.96 (br s, 1H), 7.41-7.22 (m, 5H), 5.99 (d, $J = 5.9$ Hz, 1H), 5.75 (s, 1H), 5.33 (s, 1H), 4.39 (d, $J = 5.9$ Hz, 1H), 3.96 (s, 2H), 1.99 (d, $J = 14.1$ Hz, 1H), 1.94 (sept, $J = 6.8$ Hz, 1H), 1.59 (dd, $J = 14.1, 6.8$ Hz, 1H), 0.94 (d, $J = 6.8$ Hz, 3H), 0.92 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, $DMSO-d_6$) δ 196.9, 192.5, 163.6, 134.8, 129.0 (2C), 128.6 (2C), 127.0, 104.0, 95.0, 87.6, 81.7, 43.8, 35.9, 25.0, 24.7, 22.5; HRMS (FAB-DFMS) m/z : $[M+H]^+$ Calcd for $C_{18}H_{22}NO_5$ 332.1498; Found 332.1520.

NMR chemical shifts of natural products and synthetic compounds

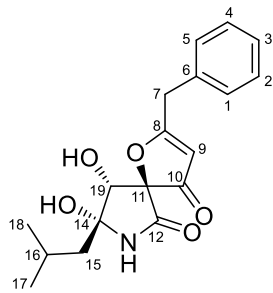


	Natural (CDCl ₃)		Synthetic (CDCl ₃)	
	¹ H (500 MHz)	¹³ C (125 MHz)	¹ H (400 MHz)	¹³ C (100 MHz)
1	1.61 (s)	25.2	1.63 (s)	25.2
2		109.0		109.1
3a	2.98 (d, <i>J</i> = 18.4 Hz)	48.6	2.97 (d, <i>J</i> = 18.5 Hz)	48.6
3b	2.83 (d, <i>J</i> = 18.4 Hz)		2.85 (d, <i>J</i> = 18.5 Hz)	
4		199.4		199.2
5		82.6		82.4
6		169.2		168.4
7	4.87 (s)	86.4	4.88 (s)	86.5
8		93.1		92.8
9	1.84 (m)	45.3	1.86-1.81 (m)	45.5
10	1.87 (m)	24.2	1.92-1.87 (m)	24.3
11	1.01 (d, <i>J</i> = 6.3 Hz)	23.4	1.02 (d, <i>J</i> = 6.5 Hz)	23.5
12	1.01 (d, <i>J</i> = 6.3 Hz)	23.8	1.03 (d, <i>J</i> = 6.5 Hz)	23.7
5-OH	4.42 (br s)		4.23 (s)	
6-NH	7.38 (br s)		6.58 (br s)	



(+)-Talaramide A (**2**)

	Natural (DMSO- <i>d</i> ₆)		Synthetic (DMSO- <i>d</i> ₆)	
	¹ H (500 MHz)	¹³ C (125 MHz)	¹ H (400 MHz)	¹³ C (100 MHz)
1	3.10 (s)	43.1	3.11 (s)	43.1
2		108.7		108.7
3a	2.68 (d, <i>J</i> = 18.6 Hz)	47.5	2.68 (d, <i>J</i> = 18.5 Hz)	47.5
3b	2.78 (d, <i>J</i> = 18.6 Hz)		2.79 (d, <i>J</i> = 18.5 Hz)	
4		200.4		200.3
5		83.3		83.2
6		168.8		168.7
7	4.76 (s)	86.3	4.76 (s)	86.3
8		93.0		93.0
9	1.55 (m)	44.3	1.58-1.51 (m)	44.2
10	1.86 (m)	23.0	1.71-1.62 (m)	23.0
11	0.83 (d, <i>J</i> = 6.6 Hz)	23.4	0.84 (d, <i>J</i> = 7.3 Hz)	23.4
12	0.82 (d, <i>J</i> = 6.6 Hz)	23.8	0.83 (d, <i>J</i> = 7.3 Hz)	23.8
1'		134.4		134.4
2'/6'	7.29 (m)	130.7	7.33-7.23 (m)	130.7
3'/5'	7.29 (m)	127.9	7.33-7.23 (m)	127.9
4'	7.25 (m)	126.8	7.33-7.23 (m)	126.7
5-OH	6.59 (br s)		6.58 (s)	
6-NH	9.37 (s)		9.37 (s)	



(-)-Berkeleyamide D (**3**)

	Natural (CDCl ₃)		Synthetic (CDCl ₃)	
	¹ H (300 MHz)	¹³ C (75 MHz)	¹ H (400 MHz)	¹³ C (100 MHz)
1, 5	7.33 (m)	129.2	7.39-7.29 (m)	129.2
2, 4	7.33 (m)	129.0	7.39-7.29 (m)	129.0
3	7.33 (m)	127.0	7.39-7.29 (m)	127.8
6		133.2		133.2
7	3.98 (d, <i>J</i> = 17.4 Hz), 3.96 (d, <i>J</i> = 17.4 Hz)	37.4	4.02 (d, <i>J</i> = 17.6 Hz), 3.96 (d, <i>J</i> = 17.6 Hz)	37.4
8		197.8		197.8
9	5.35 (br s)	104.4	5.37 (s)	104.4
10		199.4		199.4
11		95.3		95.3
12		164.1		163.9
13	6.78 (br s)		6.72 (br s)	
14		84.9		84.9
15	1.88 (m, 2H)	45.5	1.88 (dd, <i>J</i> = 14.6, 6.3 Hz), 1.62 (dd, <i>J</i> = 7.3, 6.3 Hz)	45.6
16	1.92 (m)	24.0	1.94 (sept, <i>J</i> = 6.3 Hz)	24.0
17	1.00 (d, <i>J</i> = 5.2 Hz, 3H)	23.9	1.02 (d, <i>J</i> = 6.3 Hz)	23.9
18	0.98 (d, <i>J</i> = 5.2 Hz, 3H)	23.8	1.01 (d, <i>J</i> = 6.3 Hz)	23.8
19	4.41 (d, <i>J</i> = 10.0 Hz)	75.1	4.43 (d, <i>J</i> = 9.3 Hz)	75.2
OH, C-19	3.04 (d, <i>J</i> = 10.0 Hz)		3.03 (d, <i>J</i> = 10.2 Hz)	
OH, C-14	5.48 (br s)		5.49 (s)	

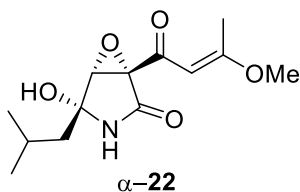
B3LYP 6-311G+ (d, p) Calculated Cartesian Coordinates

In the previous studies by Tsubaki's group, **3** was obtained as a single diastereomer at the hemiaminal position (C-14), which was attributed to the large energy difference between **3** and 14-*epi*-**3** (7.2 kcal/mol, see ref 9a). However, we obtained both **3** and 14-*epi*-**3** in a 1.2:1 ratio. In addition, structurally related molecules **16** and **17** (14-*epi*-**16**) were formed in a 2.7:1 ratio.

As our results cannot be explained by the relative thermodynamic stability of the spirocyclic products, we calculated the energy difference between the plausible intermediates α -**22** and β -**22** with a methyl substituent, which would lead to **16** and **17**, respectively (see Scheme 4 in the manuscript). The calculations revealed that the conformation of α -**22** with the lowest energy was more stable than that of β -**22** by 0.67 kcal/mol (see S17-S20), meaning that they would be present in a ratio of 2:1~3:1. We speculate that the mixture of α -**22** and β -**22** initially gave **16** and **17** in an almost 1:1 ratio, and over the long reaction time (3 days) for conversion of **15** to **16** and **17**, there was gradual isomerization from **17** to the more stable **16**, resulting in the 2.7:1 ratio. On the other hand, the similar reaction of **25** generated **3** and 14-*epi*-**3** in a 1.2:1 ratio, which can be rationalized by the shorter reaction time (2 days) for conversion of **25** to **3** and 14-*epi*-**3**.

Comparison of stability between α -22 and β -22 by DFT Calculations. Conformational analyses were performed using conformational search with MMFF and energy calculation with B3LYP/6-31G* level implemented in version 1.4.8 of the Spartan 18 software (Wavefunction, Inc., Irvine CA, America). The lower energy conformers of each compound, which differed from the most stable conformer by less than 2 kcal/mol, were optimized using DFT calculations at the B3LYP/6-311+G(d,p) level, that were implemented in the Gaussian 09 program package (Gaussian, Inc., Wallingford, CT, USA). The lowest energy conformations were determined by comparing the energies of each conformer.

α -22



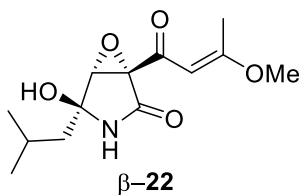
-937.665733 Hartree

Gibbs Free Energy: -588394.6241 kcal/mol

N	-1.450426	-0.214785	1.209393
C	-1.960267	-0.623023	-0.067659
C	-0.788129	-1.295605	-0.736744
C	0.376747	-1.255416	0.227204
C	-0.157658	-0.611407	1.452888
O	0.453216	-0.475795	2.498356
O	-2.968759	-1.572157	0.180400
O	-0.298737	-2.491846	-0.120165
C	1.743160	-1.083136	-0.329310
O	2.175701	-1.860847	-1.172159
C	2.529327	0.088231	0.159371
C	3.609907	0.598404	-0.465881
O	4.362960	1.661052	0.000894
C	4.131358	0.093213	-1.785221
C	4.073225	2.201743	1.285023
H	-0.663347	-1.159877	-1.797960

C	-2.485553	0.555222	-0.899739
C	-3.517692	1.483654	-0.219264
C	-4.810886	0.773132	0.178486
C	-3.846387	2.650342	-1.157887
H	-2.050267	0.076770	1.968504
H	-2.519309	-2.360868	0.533523
H	2.187550	0.518751	1.090259
H	4.801751	0.829165	-2.243722
H	4.700872	-0.830596	-1.644257
H	3.323520	-0.076696	-2.503581
H	4.803824	2.989663	1.490614
H	3.076701	2.654374	1.301327
H	4.171777	1.438366	2.063539
H	-2.927719	0.160932	-1.825136
H	-1.629999	1.173934	-1.205615
H	-3.069923	1.911438	0.686152
H	-5.239100	0.222894	-0.665889
H	-4.643324	0.072115	1.000927
H	-5.560072	1.493219	0.525763
H	-4.537074	3.353006	-0.679542
H	-2.940083	3.205457	-1.422305
H	-4.310762	2.296076	-2.084565

β -22



-937.664661Hartree

Gibbs Free Energy: -588393.9514 kcal/mol

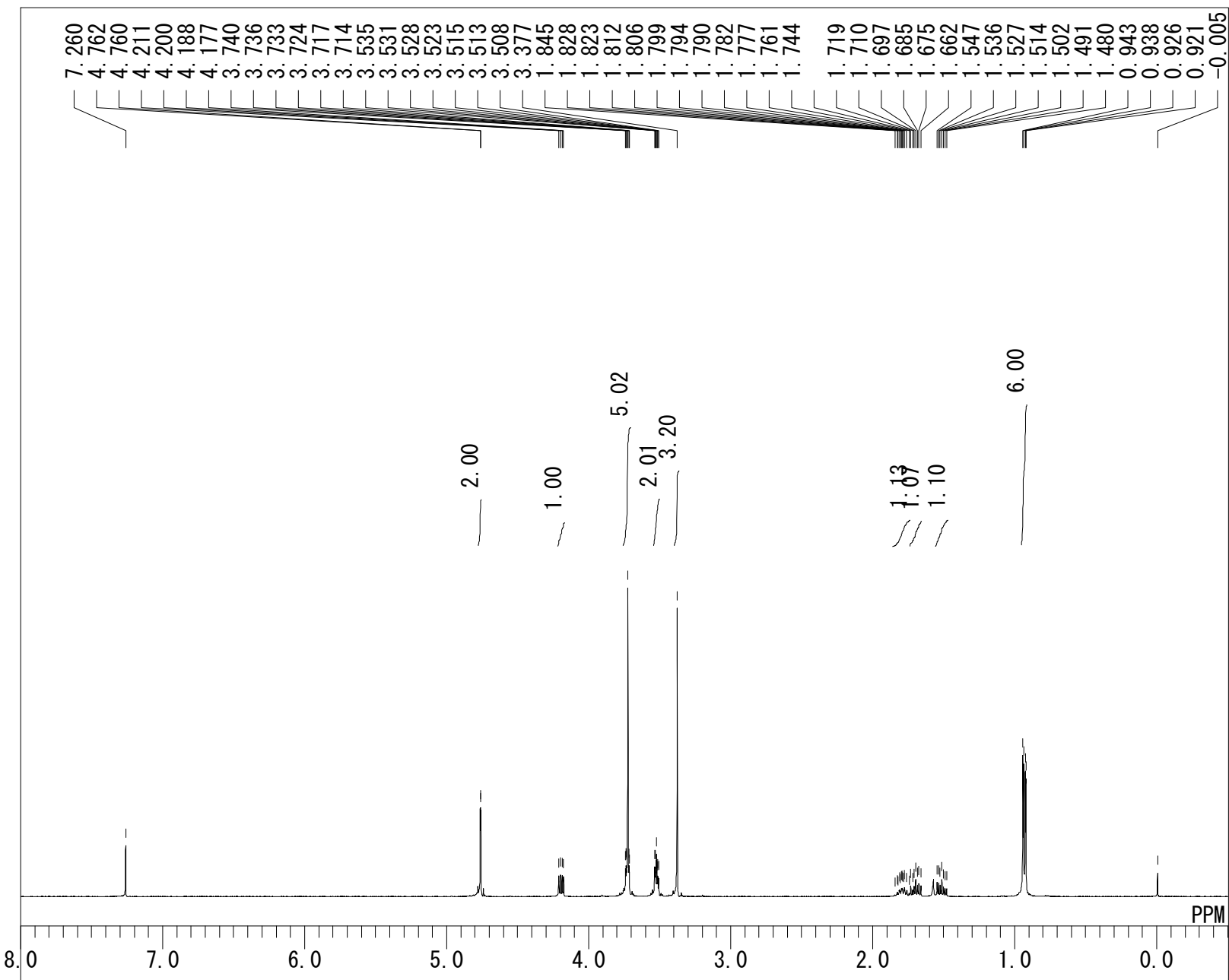
C	0.274632	-0.001288	-0.207067
C	0.113742	-1.515835	-0.115715
N	-1.174398	-1.733111	0.297686
C	-2.001742	-0.552665	0.545448
C	-1.041267	0.571640	0.131988
O	0.936619	-2.377745	-0.357424
O	-0.627506	0.517786	-1.221604
C	-3.320393	-0.646272	-0.235107
C	-4.411174	0.399052	0.070822
C	-5.738077	-0.065869	-0.550293
C	-4.062499	1.810540	-0.423426
O	-2.328655	-0.435067	1.923739
C	1.565005	0.793991	-0.023115
O	1.457956	2.000210	0.164762
C	2.800745	0.043156	-0.101552
C	4.032100	0.608825	0.058305
O	5.173335	-0.092984	-0.034321
C	4.309188	2.050753	0.347162
C	5.125221	-1.495686	-0.325232
H	-1.564670	-2.665486	0.269434
H	-1.128666	1.554885	0.576440
H	-3.731283	-1.638733	-0.015634
H	-3.071293	-0.633130	-1.301000
H	-4.540360	0.435253	1.157007
H	-5.658830	-0.140361	-1.640549
H	-6.038380	-1.046191	-0.168667
H	-6.541767	0.640496	-0.325000

H	-3.849222	1.812088	-1.497561
H	-4.901087	2.490996	-0.250222
H	-3.196606	2.232230	0.089715
H	-1.520220	-0.519143	2.443439
H	2.703321	-1.012251	-0.308500
H	3.746930	2.384711	1.220393
H	3.971055	2.675253	-0.483055
H	5.378097	2.191867	0.505493
H	6.162866	-1.821159	-0.350066
H	4.581442	-2.035049	0.453991
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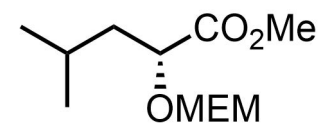
Computations were carried out with the SPARTAN' 18 series of programs: Y. Shao, L. Y. Molnar, Y. Jung, J. Kussmann, C. Ochsenfeld, S. T. Brown, A. T. B. Gilbert, L. V. Slipchenko, S. V. Levchnko, D. P. O'Neill, R. A DiStasio Jr., R. C. Lochan, T. Wang, G. J. O. Beran, N. A. Besley, J. M. Herbert, C. Y. Lin, T. Van Voorhis, S. H. Chien, A. Sodt, R. P. Steele, V. A. Rassolov, P. E. Maslen, P. P. Korambath, R. D. Adamson, B. Austin, J. Baker, E. F. C. Byrd, H. Dachsel, R. J. Doerksen, A. Dreuw, B. D. Dunietz, A. D. Dutoi, T. R. Furlani, S. R. Gwaltney, A. Heyden, S. Hirata, C-P. Hsu, G. Kedziora, R. Z. Khalliulin, P. Klunzinger, A. M. Lee, M. S. Lee, W. Z. Liang, I. Lotan, N. Nair, B. Peters, E. I. Proynov, P. A. Pieniazek, Y. M. Rhee, J. Ritchie, E. Rosta, C. D. Sherrill, A. C. Simmonett, J. E. Subotnik, H. L. Woodcock III, W. Zhang, A. T. Bell, A. K. Chakraborty, D. M. Chipman, F. J. Keil, A. Warshel, W. J. Hehre, H. F. Schaefer, J. Kong, A. I. Krylov, P. M. W. Gill and M. Head-Gordon, *Phys. Chem. Chem. Phys.* **8**, 3172 (2006).

Computations were carried out with the GAUSSIAN 09 series of programs: Frisch, M. J., Trucks, G. W., Schlegel, H. B., Scuseria, G. E., Robb, M. A., Cheeseman, J. R., Scalmani, G., Barone, V., Mennucci, B., Petersson, G. A., Nakatsuji, H., Caricato, M., Li, X., Hratchian, H. P., Izmaylov, A. F., Bloino, J., Zheng, G., Sonnenberg, J. L., Hada, M., Ehara, M., Toyota, K., Fukuda, R., Hasegawa, J., Ishida, M., Nakajima, T., Honda, Y., Kitao, O., Nakai, H., Vreven, T., Montgomery, J. A., Jr., Peralta, J. E., Ogliaro, F., Bearpark, M., Heyd, J. J., Brothers, E., Kudin, K. N., Staroverov, V. N., Kobayashi, R., Normand, J., Raghavachari, K., Rendell, A., Burant, J. C., Iyengar, S. S., Tomasi, J., Cossi, M., Rega, N., Millam, J. M., Klene, M., Knox, J. E., Cross, J. B., Bakken, V., Adamo, C.,

Jaramillo, J., Gomperts, R., Stratmann, R. E., Yazyev, O., Austin, A. J., Cammi, R., Pomelli, C., Ochterski, J. W., Martin, R. L., Morokuma, K., Zakrzewski, V. G., Voth, G. A., Salvador, P., Dannenberg, J. J., Dapprich, S., Daniels, A. D., Farkas, Ö., Foresman, J. B., Ortiz, J. V., Cioslowski, J., Fox, D. J. Gaussian 09 Revision D.01, Gaussian, Inc.: Wallingford, CT, 2009.



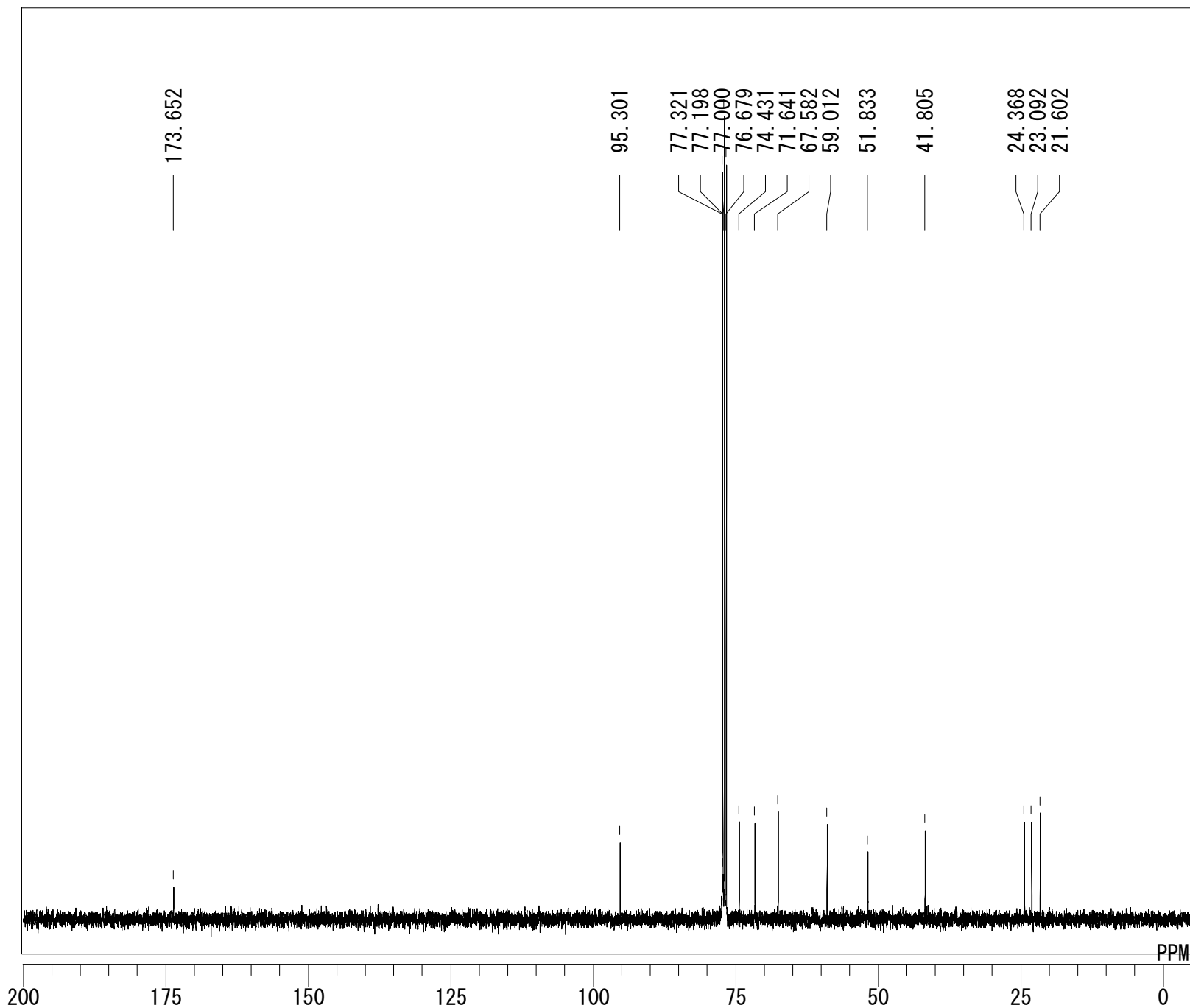
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EXMOD NON
OBFRQ 399.65 MHz
OBSET 124.00 KHz
OBFIN 10500.00 Hz
POINT 16384
FREQU 7992.01 Hz
SCANS 8
ACQTM 2.0500 sec
PD 4.9500 sec
PW1 5.80 usec
IRNUC 1H
CTEMP 24.0 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.12 Hz
RGAIN 19



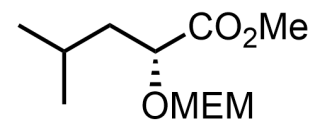
S2

¹H NMR

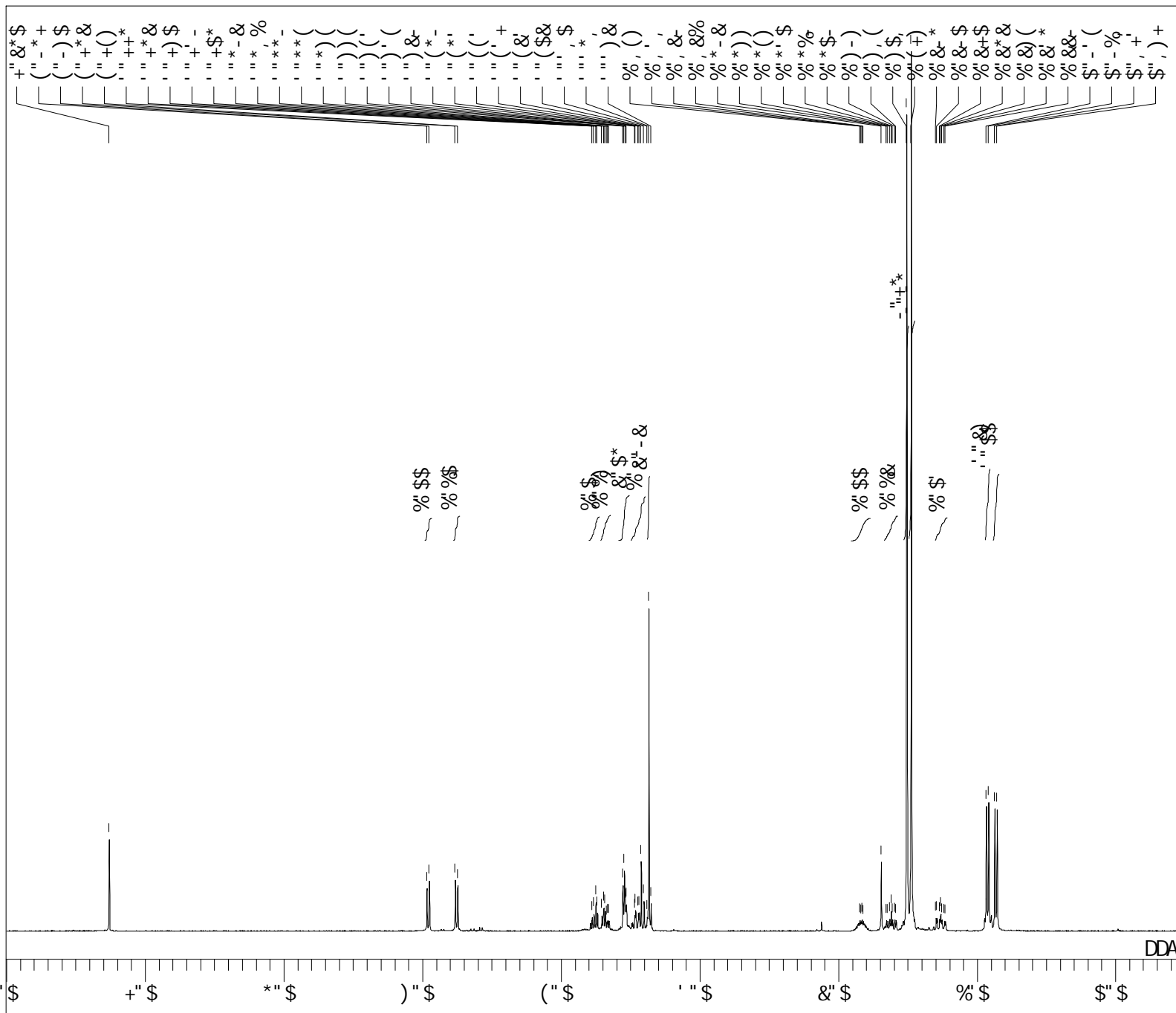
CDCl₃ (400 MHz)



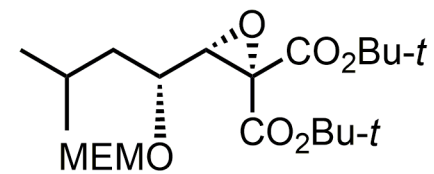
OBNUC 13C
 EXMOD BCM
 OBFRQ 100.40 MHz
 OBSET 125.00 KHz
 OBFIN 10500.00 Hz
 POINT 32768
 FREQU 27118.64 Hz
 SCANS 316
 ACQTM 1.2083 sec
 PD 1.7920 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 24.4 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 25



S2
¹³C NMR
 CDCl₃ (100 MHz)



C6: FE "*) A<n
 C6G9H %&(" \$\$?<n
 C6: =B %&)" \$\$ \$<n
 DC=BH %* (<n
 : F9EI +-&" \$% <n
 G75BG
 57EHA &"\$) \$\$ gYW
 D8 ("-\$) \$\$ gYW
 DK%)" *\$ i gYW
 =FBI 7 %<
 7H9AD &" - W
 G@JBH 787@
 9LF9: +" &* dda
 6: "\$ %& <n
 F; 5=B %



13
¹H NMR
 CDCl₃ (400 MHz)

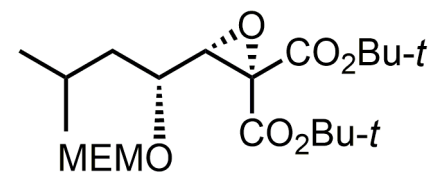
164.744
163.542

94.758
83.726
83.594
77.321
77.206
77.000
76.687
72.982
71.715
67.425
64.272
59.769
58.979

40.480

27.933
27.785
23.792
23.603
21.182

OBFRQ 100.40 MHz
OBSET 125.00 KHz
OBFIN 10500.00 Hz
POINT 32768
FREQ 27118.64 Hz
SCANS 1000
ACQTM 1.2083 sec
PD 1.7920 sec
PW1 5.80 usec
IRNUC 1H
CTEMP 24.7 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 1.20 Hz
RGAIN 25

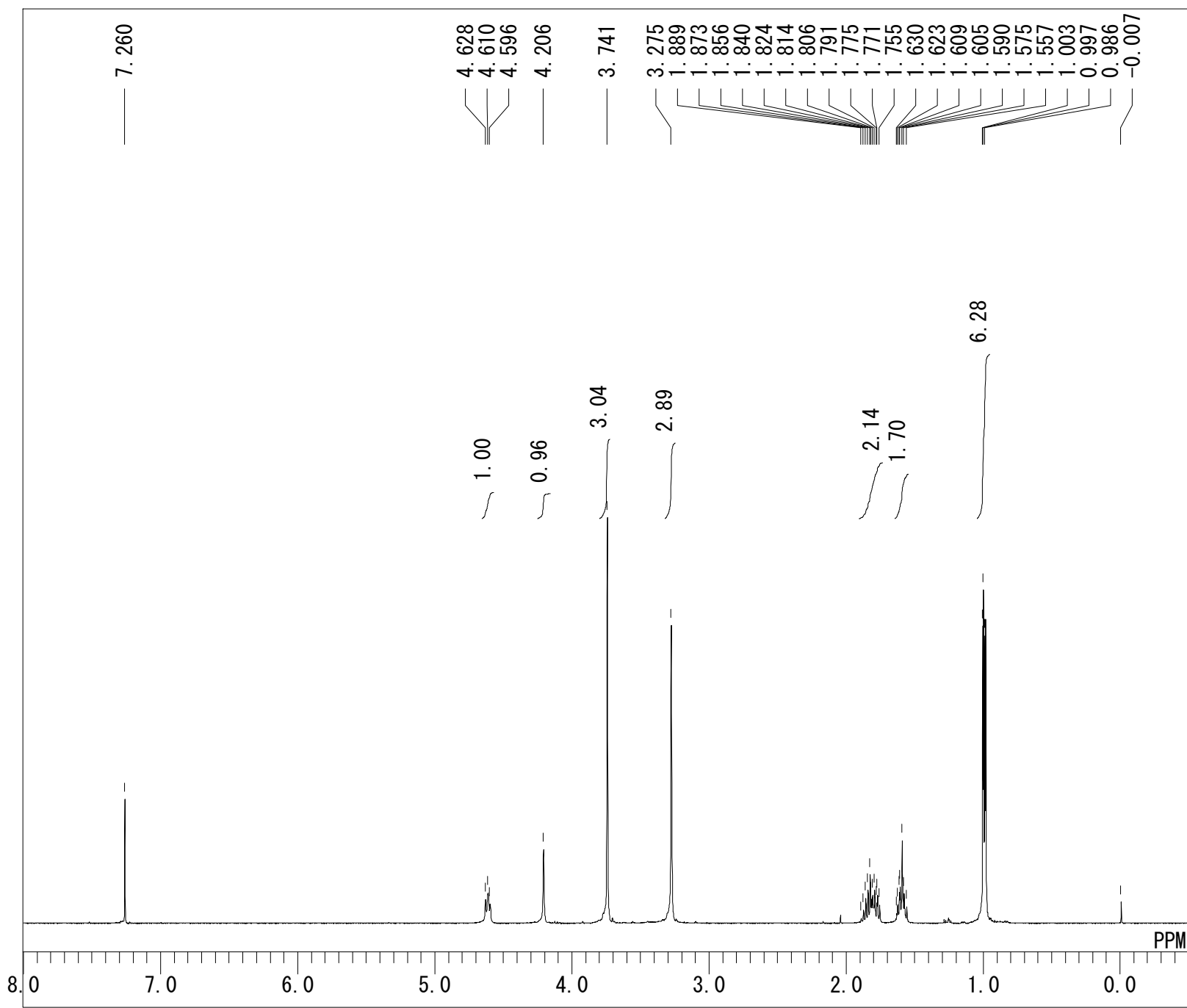


13

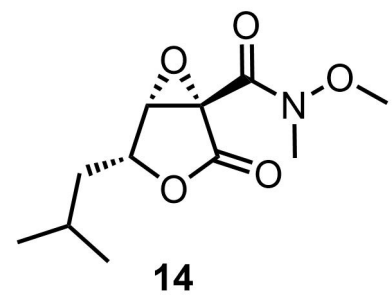
¹³C NMR

CDCl₃ (100 MHz)

PPM



OBFRQ 399.65 MHz
 OBSET 124.00 KHz
 OBFIN 10500.00 Hz
 POINT 16384
 FREQU 7992.01 Hz
 SCANS 8
 ACQTM 2.0500 sec
 PD 4.9500 sec
 PW1 5.60 usec
 IRNUC 1H
 CTEMP 21.8 c
 SLVNT CDCL3
 EXREF 7.26 ppm
 BF 0.09 Hz
 RGAIN 18



¹H NMR
CDCl₃ (400 MHz)

167.206
161.467

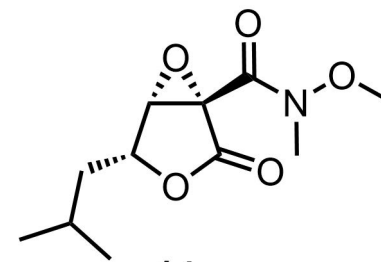
77.321
77.247
77.000
76.687

62.033
61.276
58.880

38.175
32.173

24.805
22.854
22.137

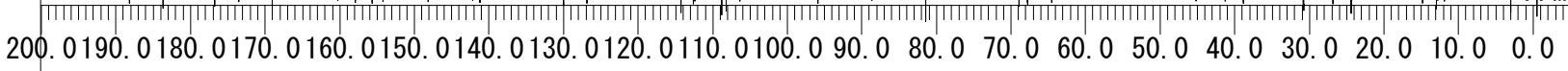
OBFRQ 100.40 MHz
OBSET 125.00 KHz
OBFIN 10500.00 Hz
POINT 32768
FREQ 27118.64 Hz
SCANS 600
ACQTM 1.2083 sec
PD 1.7920 sec
PW1 5.80 usec
IRNUC 1H
CTEMP 22.6 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 2.00 Hz
RGAIN 25

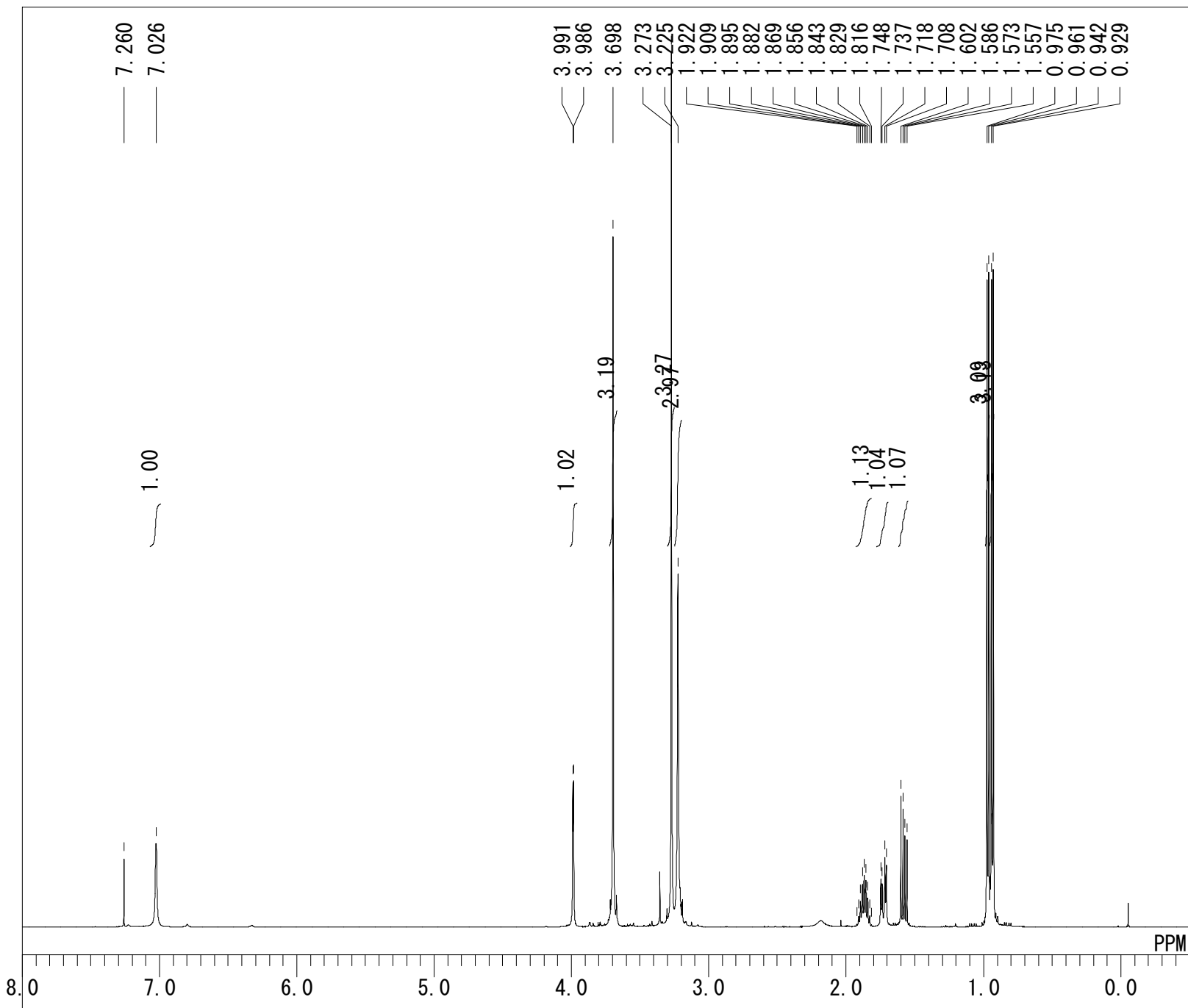


14

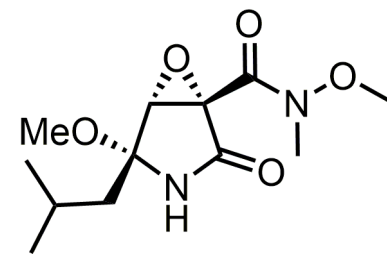
¹³C NMR

CDCl₃ (100 MHz)





OBFRO 495.13 MHz
 OBSET 4.38 KHz
 OBFIN 9.64 Hz
 POINT 26214
 FREQU 7429.42 Hz
 SCANS 16
 ACQTM 3.5285 sec
 PD 5.0000 sec
 PW1 6.25 usec
 IRNUC 1H
 CTEMP 23.8 c
 SLVNT CDCL3
 EXREF 7.26 ppm
 BF 0.09 Hz
 RGAIN 24



11

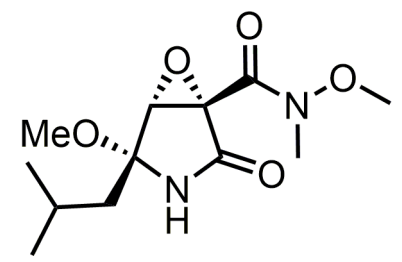
¹H NMR

CDCl₃ (500 MHz)

169.319
162.351

89.570
77.385
77.127
76.868
61.937
61.363
60.262
49.839
41.511
32.237
24.053
23.862
23.431
0.000

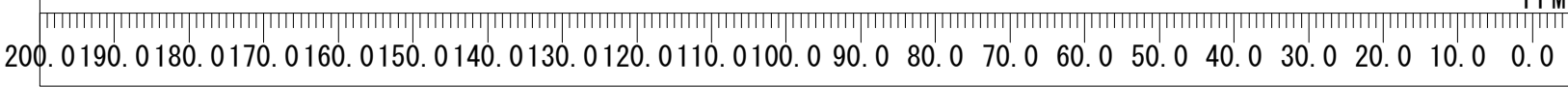
OBFRQ 124.51 MHz
OBSET 3.45 KHz
OBFIN 6.00 Hz
POINT 26224
FREQ 31250.00 Hz
SCANS 10000
ACQTM 0.8389 sec
PD 2.0000 sec
PW1 3.42 usec
IRNUC 1H
CTEMP 23.9 c
SLVNT CDCL3
EXREF 0.00 ppm
BF 0.09 Hz
RGAIN 60

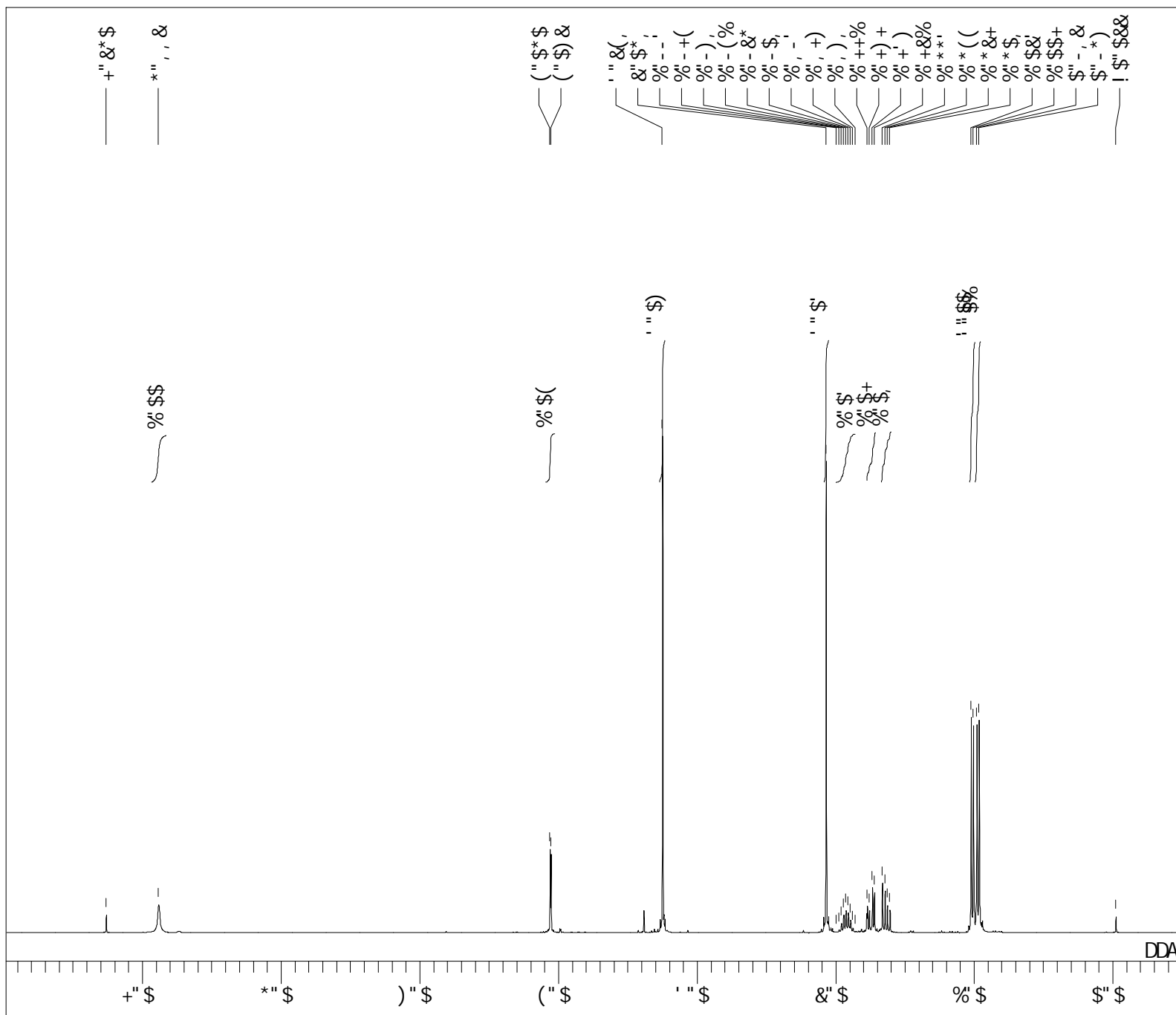


11

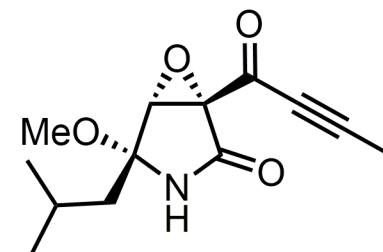
¹³C NMR

CDCl₃ (125 MHz)





C6: FE "*) A<n
 C6G9H %(" \$\$?<n
 C6: =B %) \$\$ \$\$ <n
 DC=BH %' (<n
 : F9EI +-&" \$% <n
 G75BG
 57EHA &"\$) \$\$: gYW
 D8 ("-\$) \$\$: gYW
 DK%)" *\$ \$: i gYW
 =FBI 7 %<
 7H9AD &(" , W
 G@|BH 787@
 9LF9: +"&* dda
 6: \$"%& <n
 F; 5=B %<



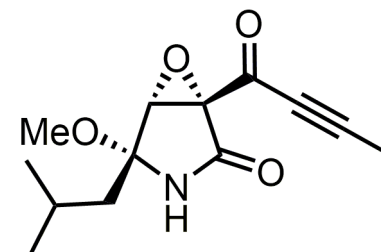
9

¹H NMR
CDCl₃ (400 MHz)

175.521
167.724

96.289
88.756
77.552
77.321
77.000
76.687
63.745
61.918
49.602
42.299
24.031
23.767
23.487
4.453

OBFRQ 100.40 MHz
OBSET 125.00 KHz
OBFIN 10500.00 Hz
POINT 32768
FREQ 27118.64 Hz
SCANS 1000
ACQTM 1.2083 sec
PD 1.7920 sec
PW1 5.80 usec
IRNUC 1H
CTEMP 25.3 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 1.20 Hz
RGAIN 25

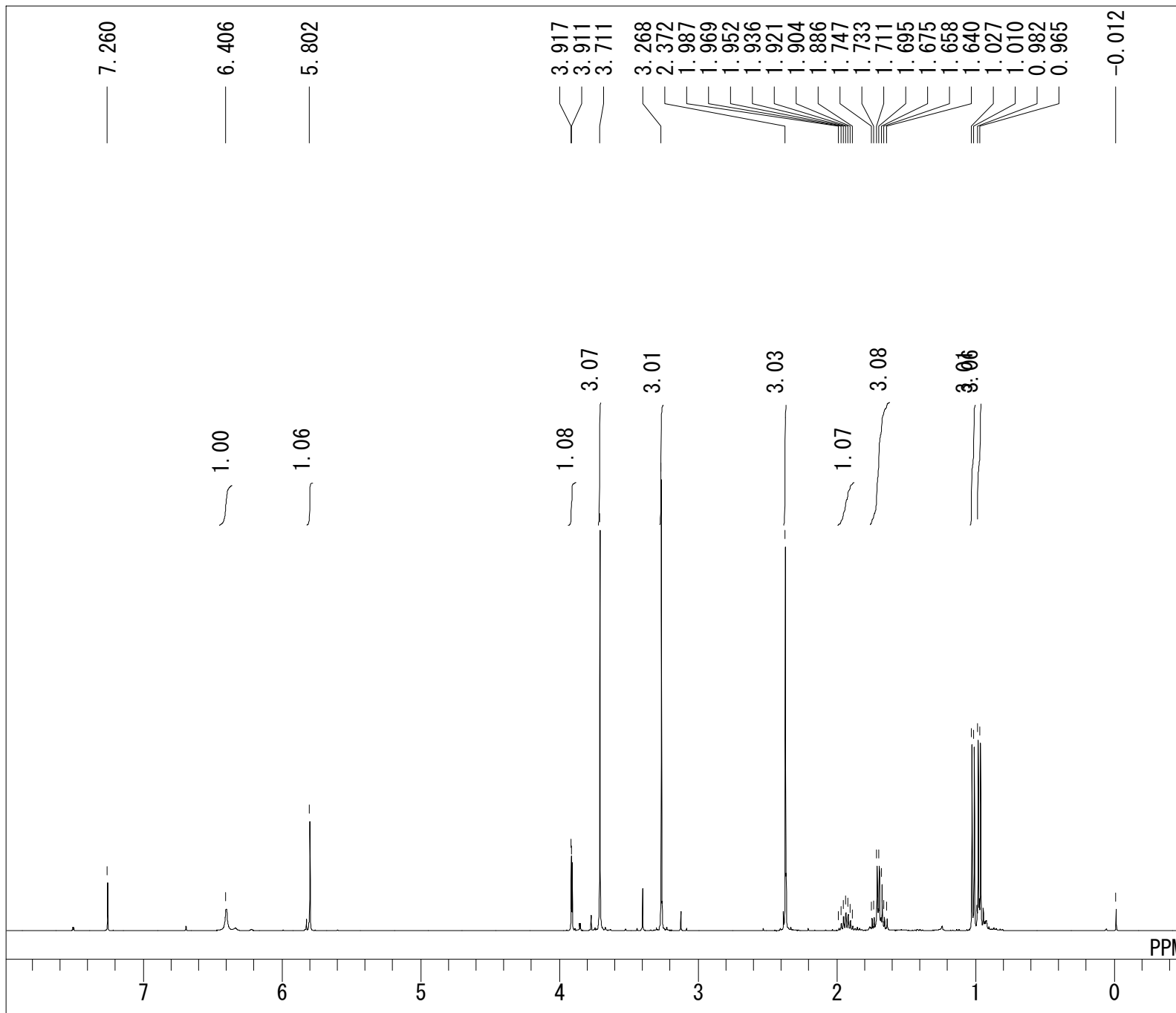


9

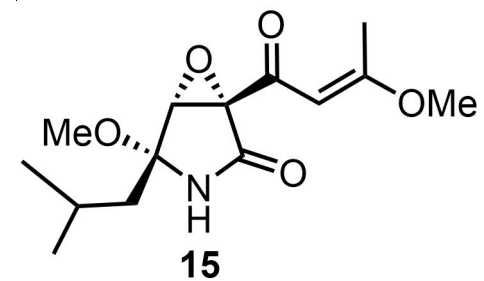
¹³C NMR

CDCl₃ (100 MHz)

PPM



OBFRQ 399.65 MHz
 OBSET 124.00 KHz
 OBFIN 10500.00 Hz
 POINT 16384
 FREQU 7992.01 Hz
 SCANS 8
 ACQTM 2.0500 sec
 PD 4.9500 sec
 PW1 5.60 usec
 IRNUC 1H
 CTEMP 23.7 c
 SLVNT CDCL3
 EXREF 7.26 ppm
 BF 0.12 Hz
 RGAIN 16



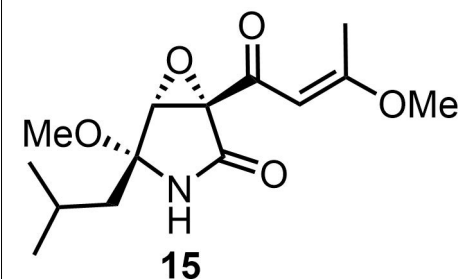
¹H NMR
 CDCl₃ (400 MHz)

186.536
178.032
169.626

94.478
88.707
77.321
77.000
76.679
63.367
61.728
56.105
49.651
42.768

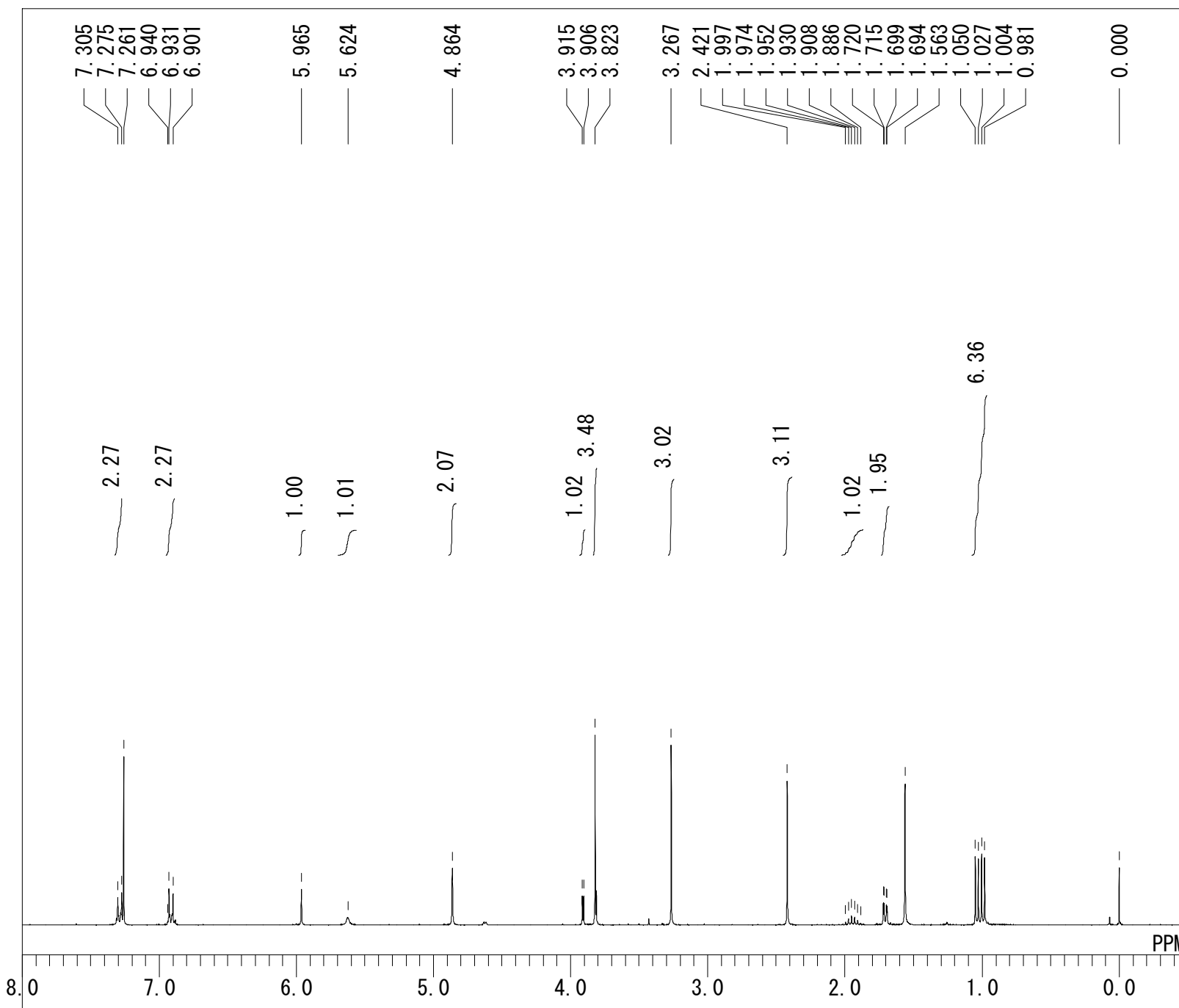
24.064
23.866
23.545
20.664

OBFRQ 100.40 MHz
OBSET 125.00 KHz
OBFIN 10500.00 Hz
POINT 32768
FREQ 27118.64 Hz
SCANS 2000
ACQTM 1.2083 sec
PD 1.7920 sec
PW1 5.80 usec
IRNUC 1H
CTEMP 24.4 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 25

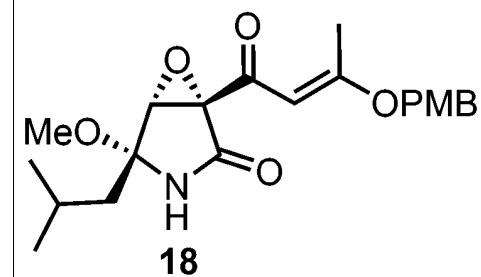


¹³C NMR
CDCl₃ (100 MHz)

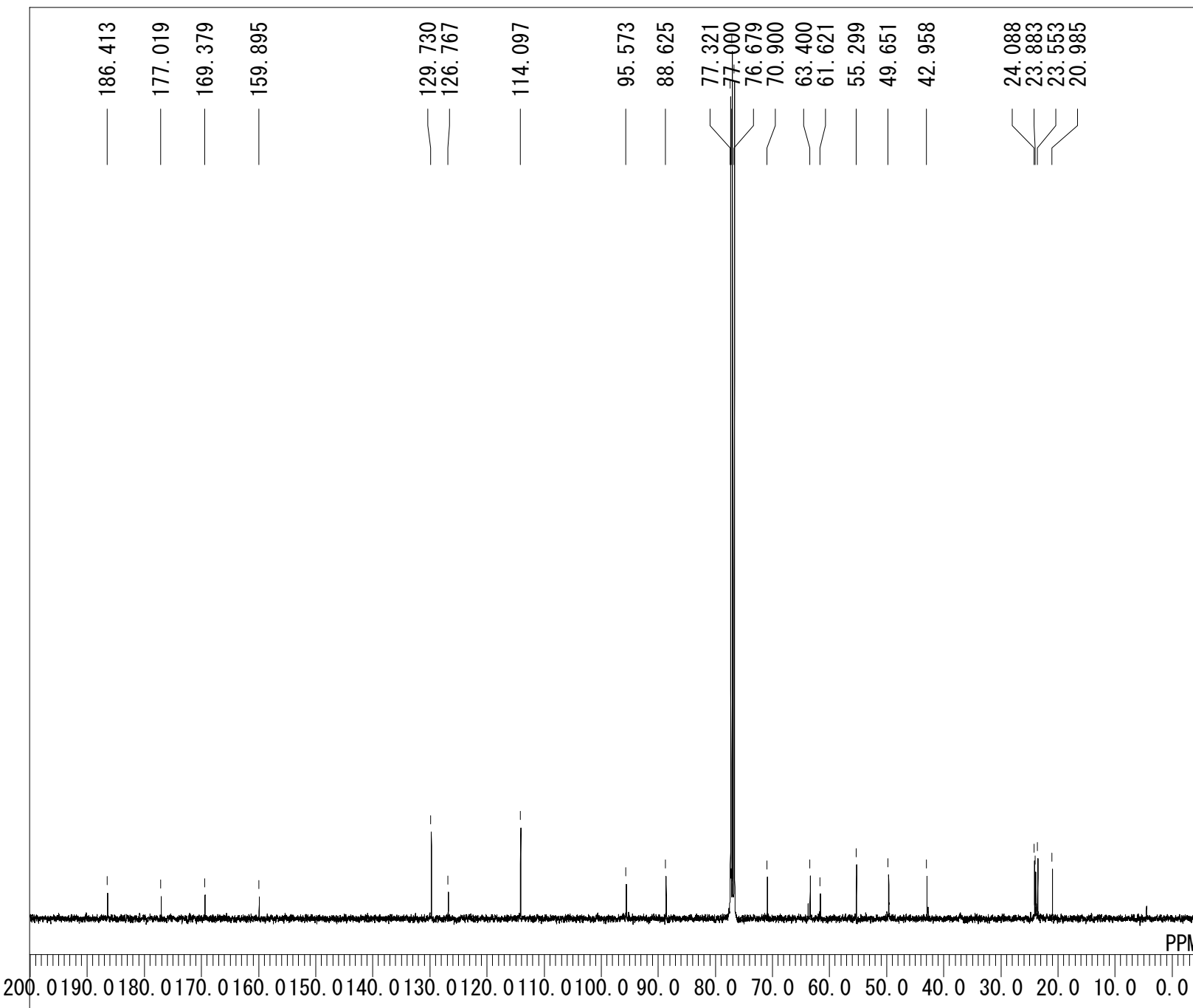
200.0 190.0 180.0 170.0 160.0 150.0 140.0 130.0 120.0 110.0 100.0 90.0 80.0 70.0 60.0 50.0 40.0 30.0 20.0 10.0 0.0



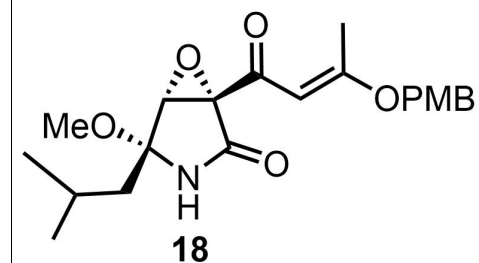
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 OBSET 130.00 KHz
 OBFIN 1150.00 Hz
 POINT 32768
 FREQU 6006.01 Hz
 SCANS 16
 ACQTM 5.4559 sec
 PD 1.5440 sec
 PW1 5.30 usec
 IRNUC 1H
 CTEMP 26.2 c
 SLVNT CDCL3
 EXREF 0.00 ppm
 BF 0.09 Hz
 RGAIN 21



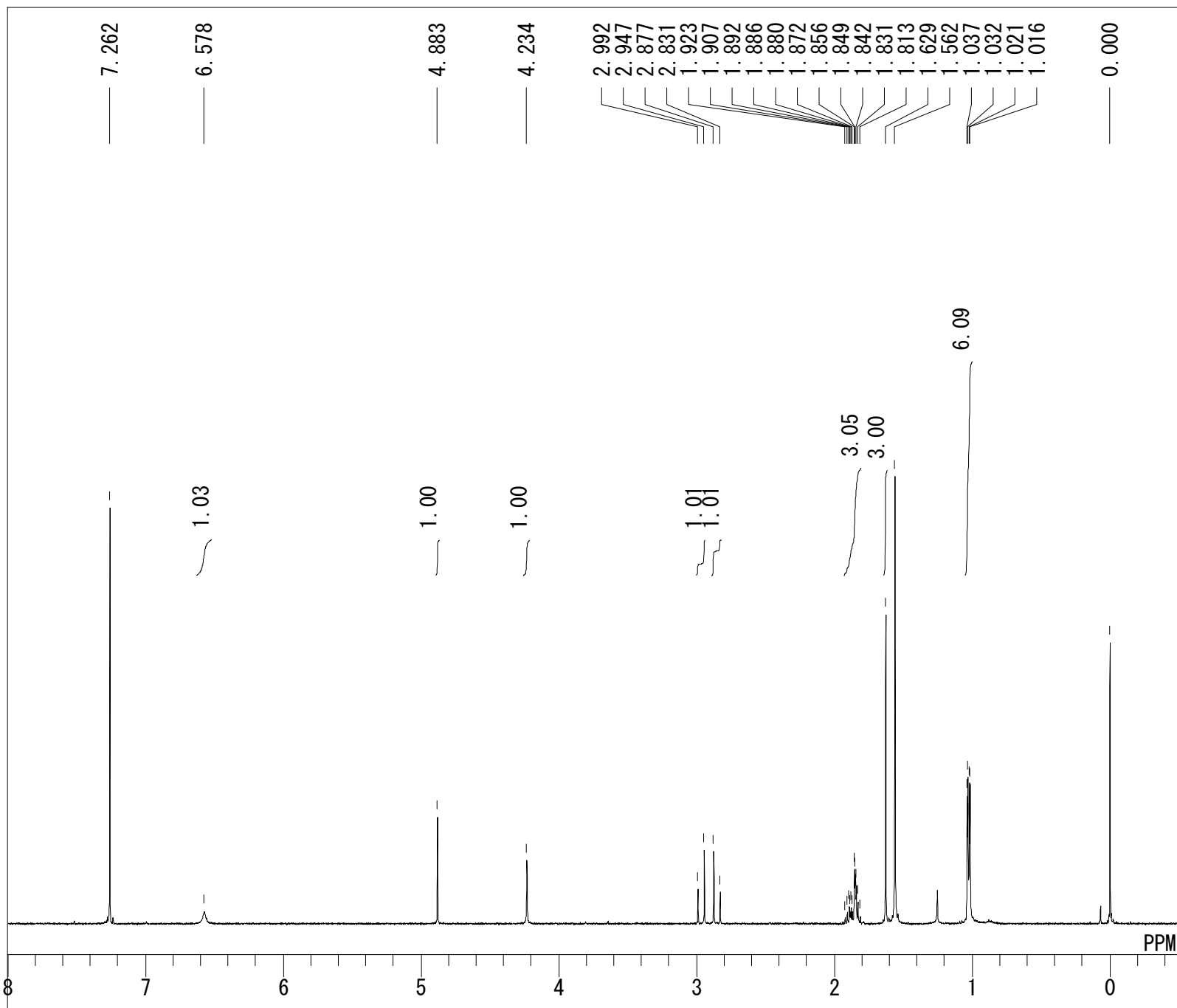
¹H NMR
 CDCl₃ (300 MHz)



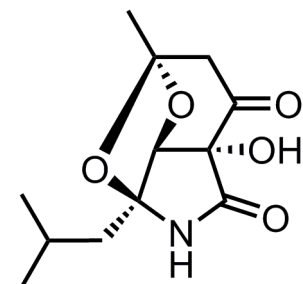
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 OBSET 125.00 KHz
 OBFIN 10500.00 Hz
 POINT 32768
 FREQU 27118.64 Hz
 SCANS 1500
 ACQTM 1.2083 sec
 PD 1.7920 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 23.5 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 2.00 Hz
 RGAIN 25



¹³C NMR
 CDCl₃ (100 MHz)



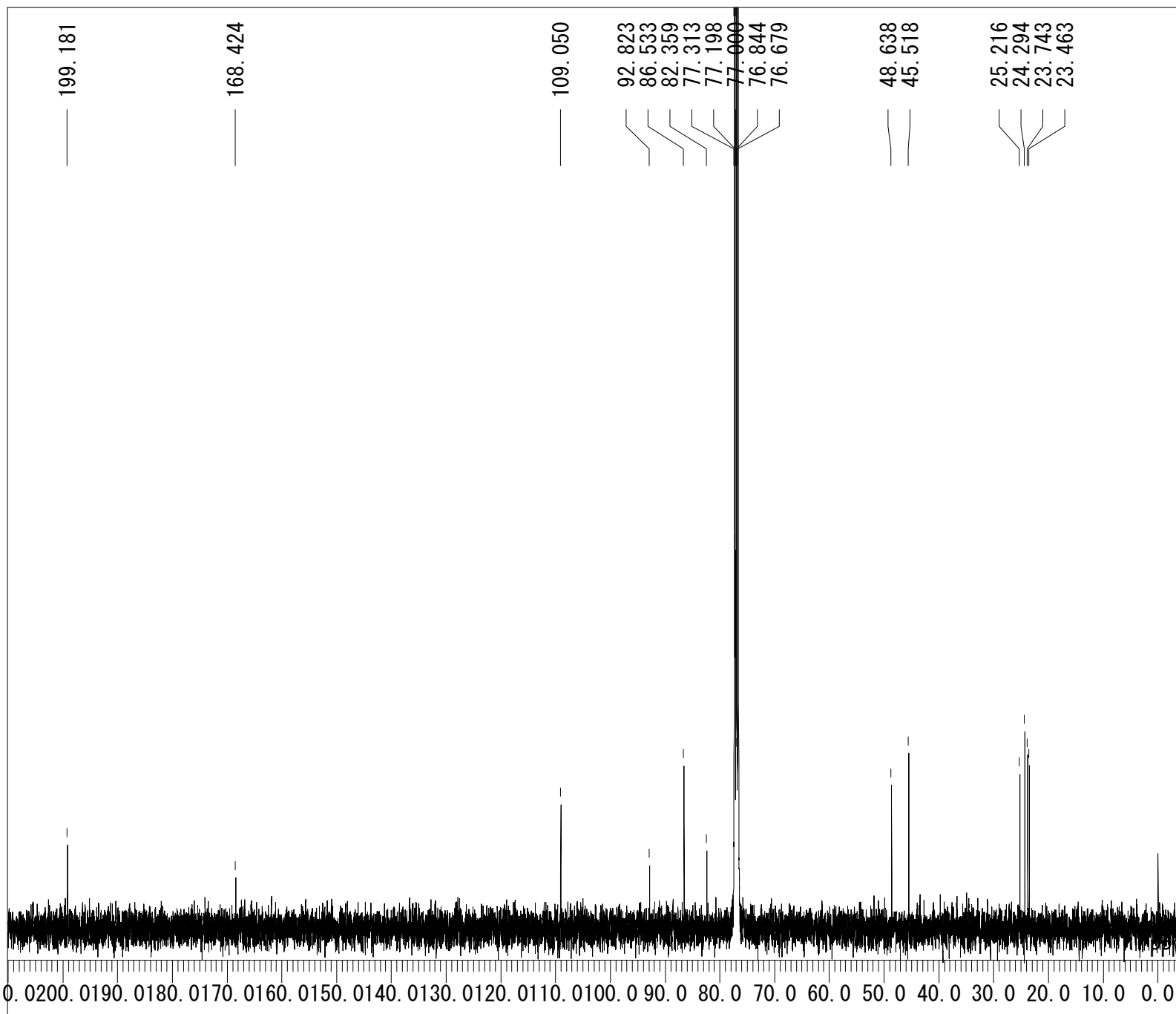
OBFRQ 399.65 MHz
 OBSET 124.00 KHz
 OBFIN 10500.00 Hz
 POINT 16384
 FREQU 7992.01 Hz
 SCANS 8
 ACQTM 2.0500 sec
 PD 4.9500 sec
 PW1 5.60 usec
 IRNUC 1H
 CTEMP 23.6 c
 SLVNT CDCL3
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 23



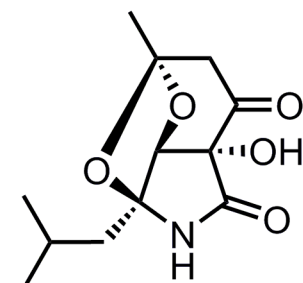
Rubrobramide (1)

¹H NMR

CDCl₃ (400 MHz)



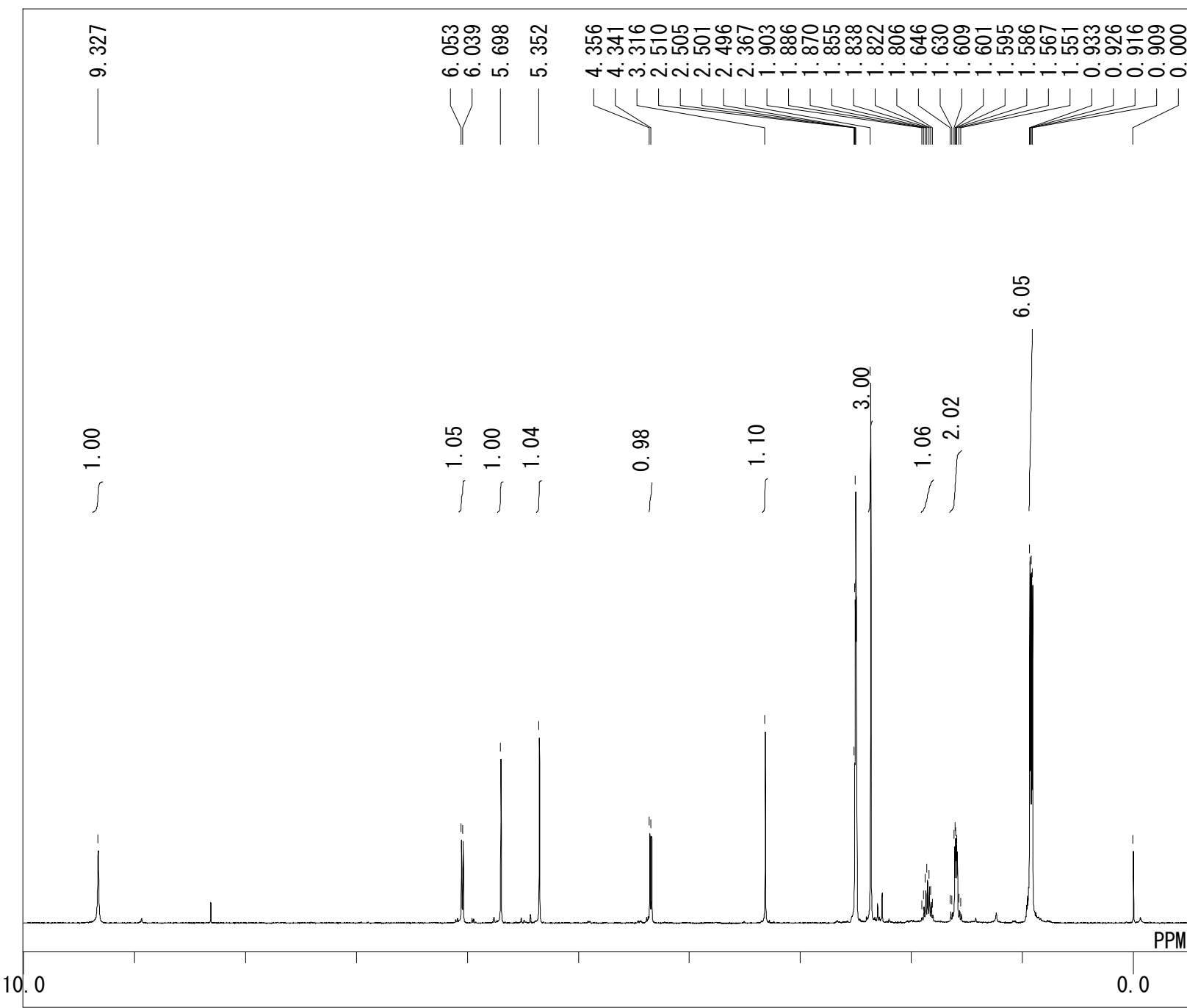
OBFRQ 100.40 MHz
 OBSET 125.00 KHz
 OBFIN 10500.00 Hz
 POINT 32768
 FREQU 27118.64 Hz
 SCANS 5000
 ACQTM 1.2083 sec
 PD 1.7920 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 24.1 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 25



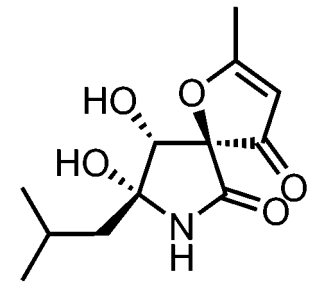
Rubrobramide (1)

¹³C NMR

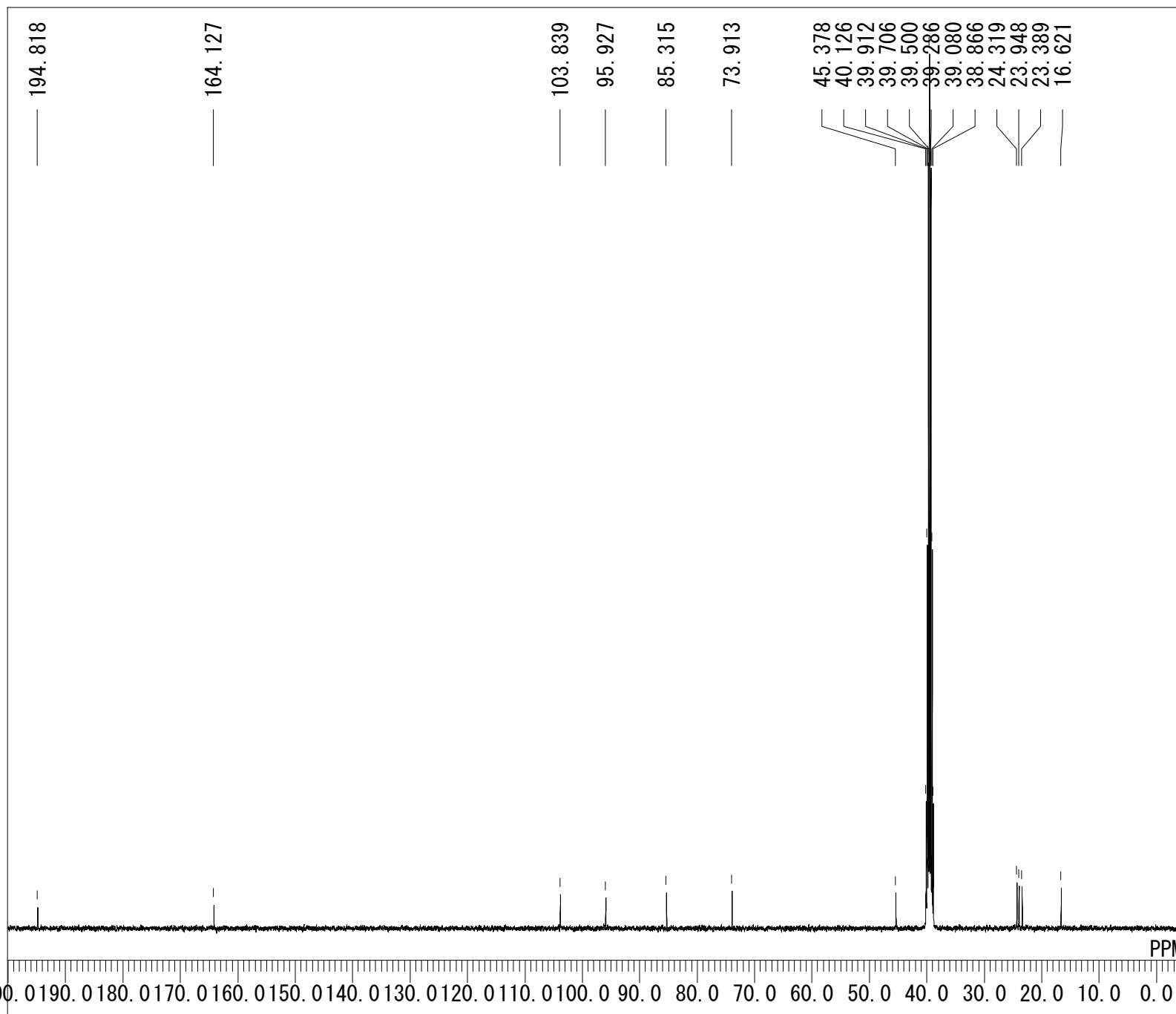
CDCl₃ (100 MHz)



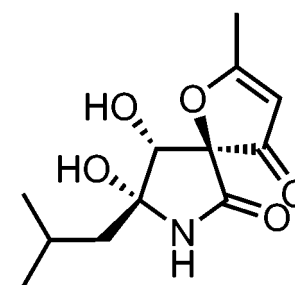
OBFREQ 399.65 MHz
 OBSET 124.00 KHz
 OBFIN 10500.00 Hz
 POINT 16384
 FREQU 7992.01 Hz
 SCANS 8
 ACQTM 2.0500 sec
 PD 4.9500 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 24.2 c
 SLVNT DMSO
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 19



16
¹H NMR
 DMSO-*d*₆ (400 MHz)



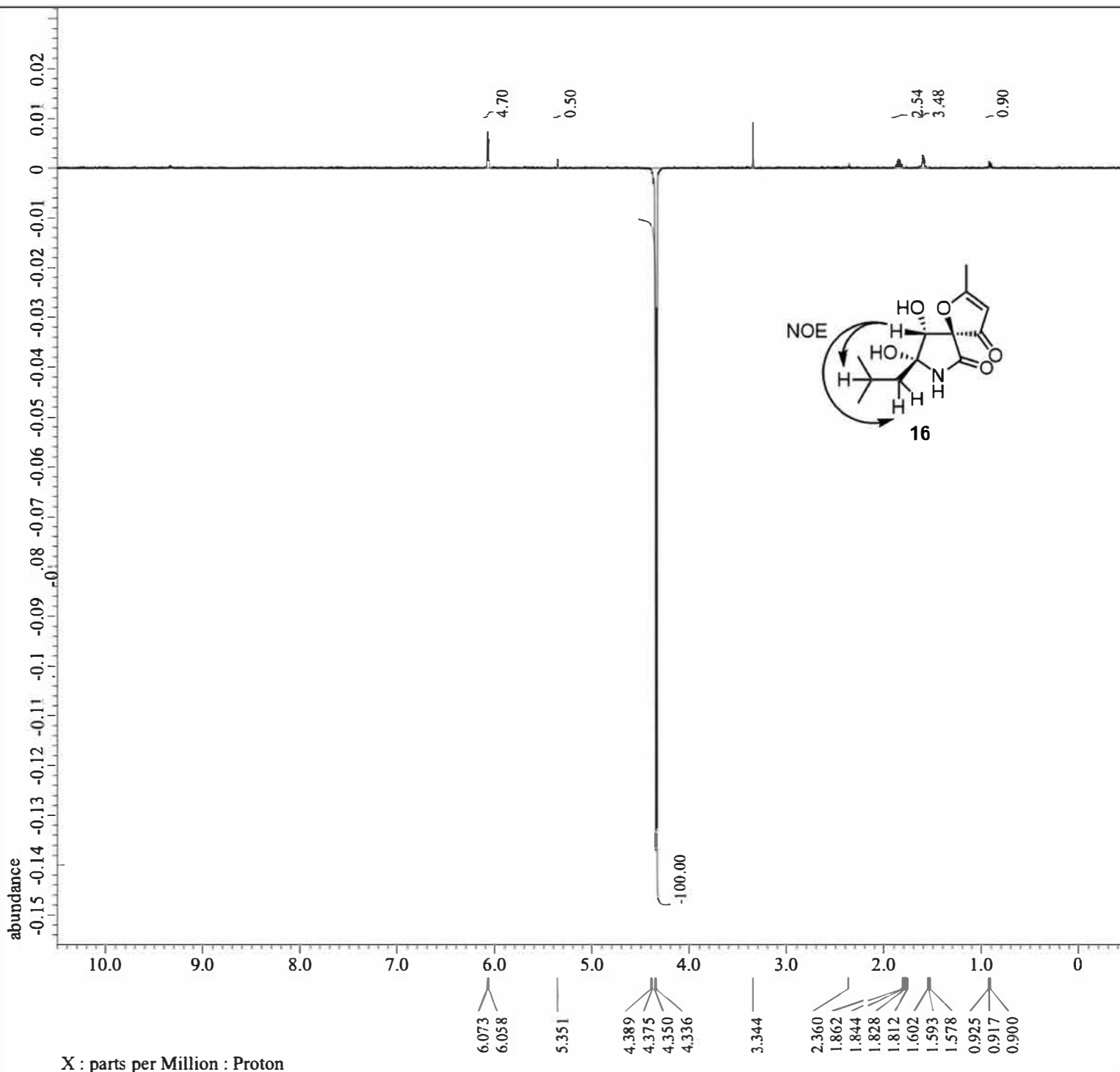
OBFREQ 100.40 MHz
 OBSET 125.00 KHz
 OBFIN 10500.00 Hz
 POINT 32768
 FREQU 27118.64 Hz
 SCANS 995
 ACQTM 1.2083 sec
 PD 1.7920 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 24.2 c
 SLVNT DMSO
 EXREF 39.50 ppm
 BF 2.00 Hz
 RGAIN 24



16

¹³C NMR

DMSO-*d*₆ (100 MHz)

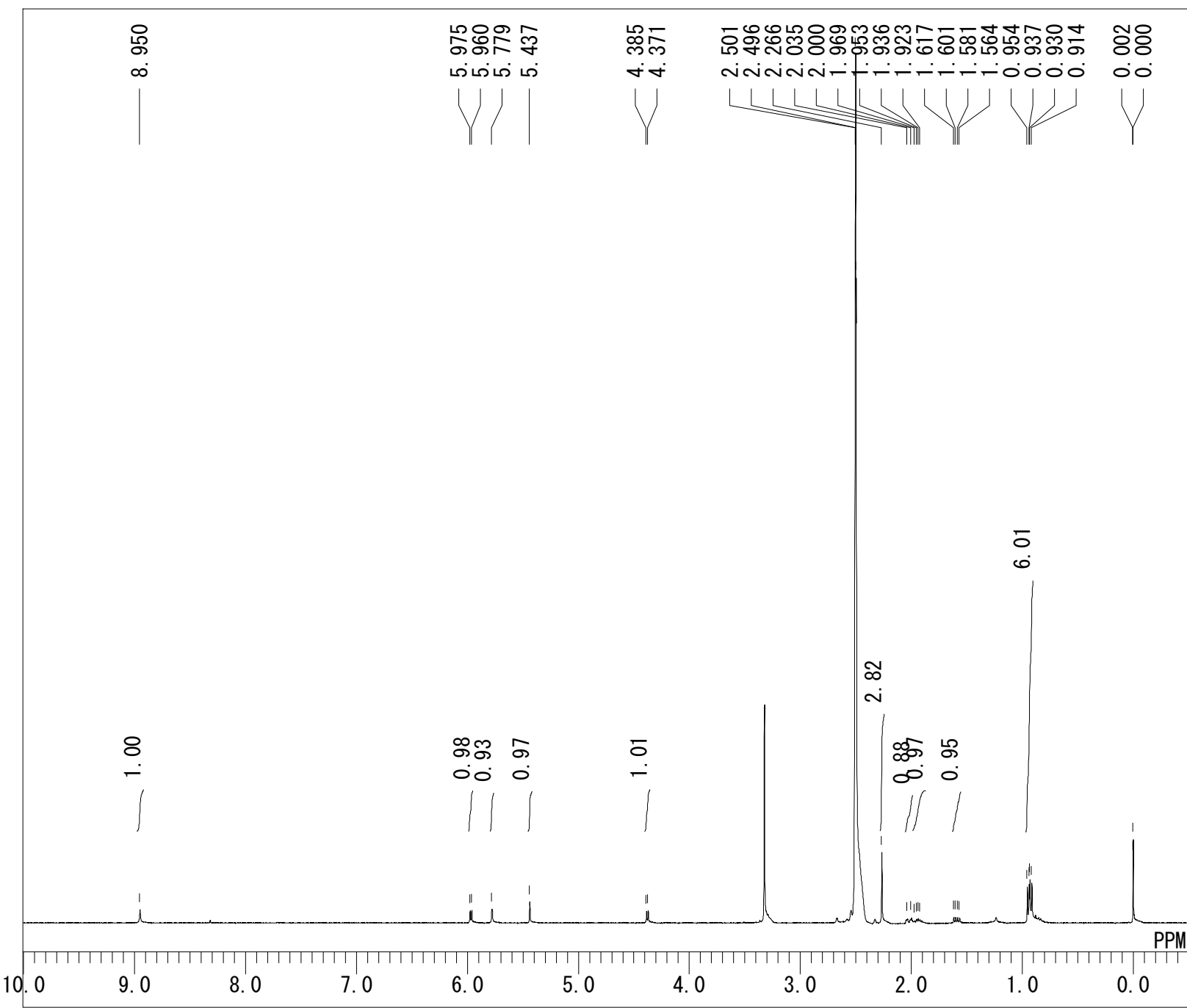


Filename = KST-Me-berkeleyamideD
 Author = delta
 Experiment = noe_id_dpfgse.jxp
 Sample Id = KST-Me-berkeleyamideD
 Solvent = DMSO-D6
 Creation_Time = 24-AUG-2021 19:55:42
 Revision_Time = 27-AUG-2021 10:30:55
 Current_Time = 27-AUG-2021 10:31:47

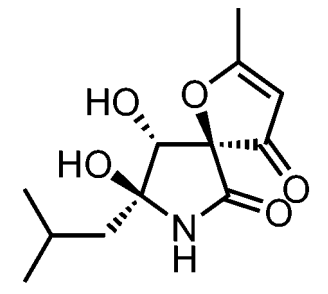
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 Data_Format = 1D COMPLEX
 Dim_Size = 13107
 Dim_Title = Proton
 Dim_Units = [ppm]
 Dimensions = X
 Spectrometer = JNM-ECZ400S/L1

Field_Strength = 9.389766 [T] (400 [MHz])
 X_Acq_Duration = 2.18628096 [s]
 X_Domain = 1H
 X_Freq = 399.78219838 [MHz]
 X_Offset = 5 [ppm]
 X_Points = 16384
 X_Prescans = 2
 X_Resolution = 0.45739775 [Hz]
 X_Sweep = 7.4940048 [kHz]
 X_Sweep_Clippped = 5.99520384 [kHz]
 Irr_Domain = Proton
 Irr_Freq = 399.78219838 [MHz]
 Irr_Offset = 5 [ppm]
 Tri_Domain = Proton
 Tri_Freq = 399.78219838 [MHz]
 Tri_Offset = 5 [ppm]
 Blanking = 2 [us]
 Clipped = FALSE
 Decimation_Reg = r: 834 (833), g: 49
 Scans = 16
 Total_Scans = 16

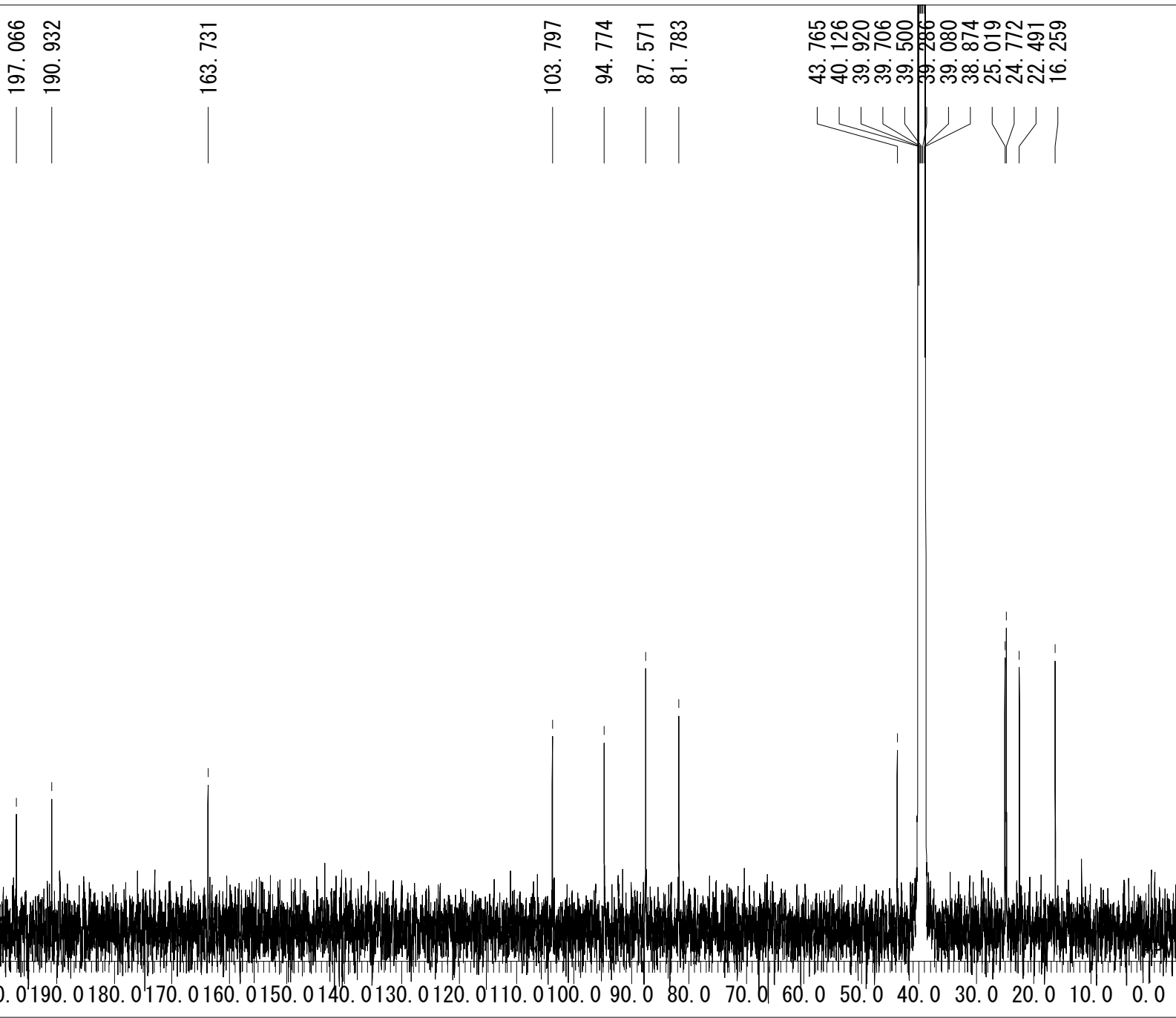
Relaxation_Delay = 7 [s]
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 Temp_Get = 21.4 [dC]
 Mix_Time = 0.5 [s]
 X_Acq_Time = 2.18628096 [s]
 X_Atn = 2.4 [dB]
 X_Pulse = 6.15 [us]
 Irr_Mode = Off
 Obs_Sel_180 = 40 [ms]
 Obs_Sel_Atn = 64.932 [dB]
 Obs_Sel_Offset = 4.3373704 [ppm]
 Obs_Sel_Shape = GAUSS
 Obs_Sel_Slp = 4.3373704 [ppm]
 Tri_Mode = Off
 Comment_1 = *** Pulse ***
 Comment_11 = *** NOESY mixing time ***
 Comment_111 = *** presat_time ***
 Comment_201 = *** obs_dante_presatu ***
 Comment_202 = *** irr_preaturation ***
 Comment_203 = *** tri_preaturation ***
 Comment_32 = *** Selective 180deg ***
 Comment_7 = *** Pulse Delay ***
 Comment_8 = *** Pulse Field Gradi ***
 Comment_900 = *** lock hold ***
 Dante_Loop = 699



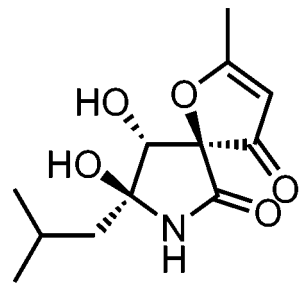
OBFRQ 399.65 MHz
 OBSET 124.00 KHz
 OBFIN 10500.00 Hz
 POINT 16384
 FREQU 7992.01 Hz
 SCANS 32
 ACQTM 2.0500 sec
 PD 4.9500 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 22.7 c
 SLVNT DMSO
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 21



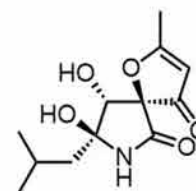
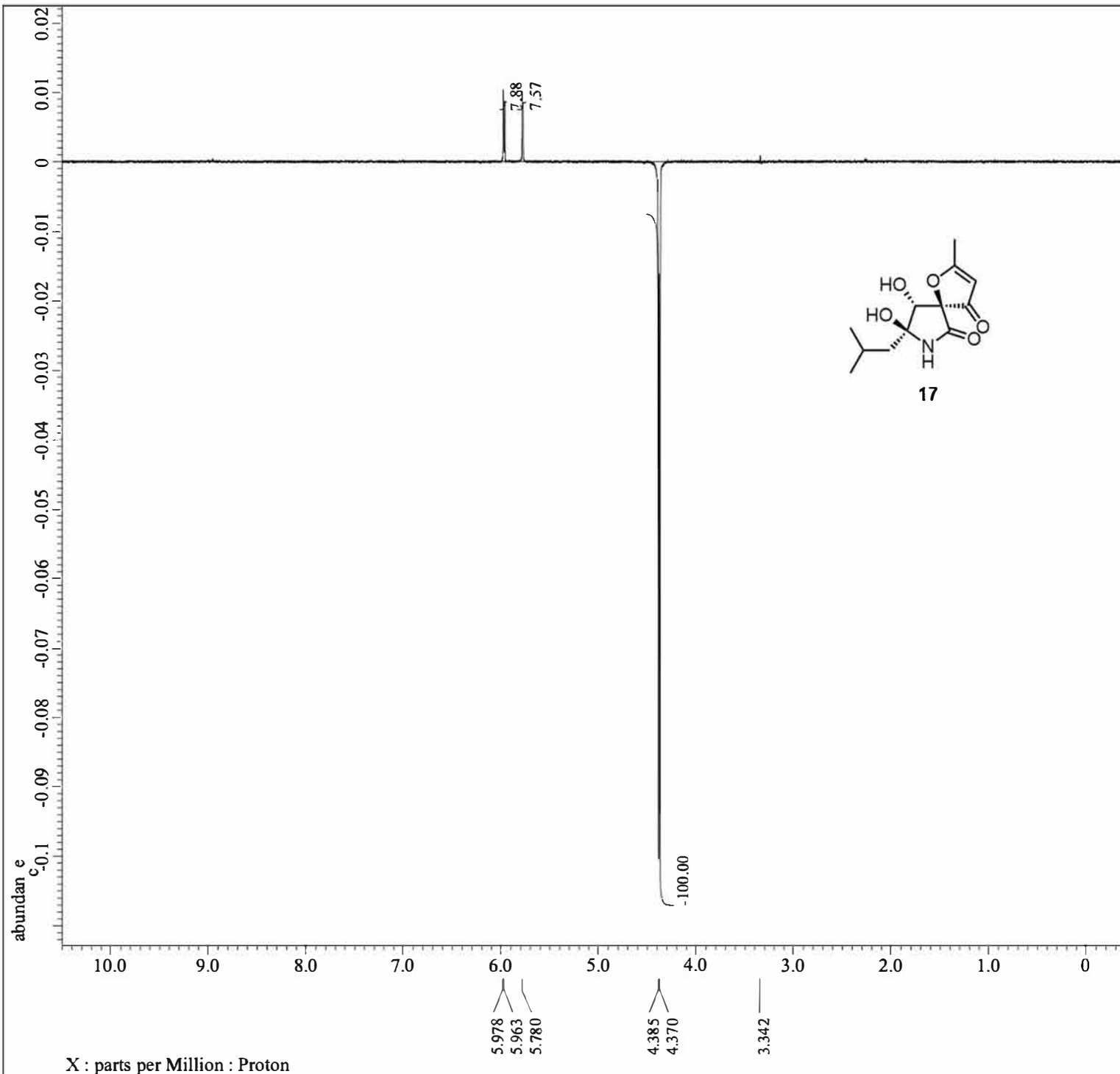
17
¹H NMR
 DMSO-d₆ (400 MHz)



OBFRQ 100.40 MHz
 OBSET 125.00 KHz
 OBFIN 10500.00 Hz
 POINT 32768
 FREQU 27118.64 Hz
 SCANS 2000
 ACQTM 1.2083 sec
 PD 1.7920 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 23.4 c
 SLVNT DMSO
 EXREF 39.50 ppm
 BF 2.00 Hz
 RGAIN 24



17
¹³C NMR
 DMSO-d₆ (100 MHz)



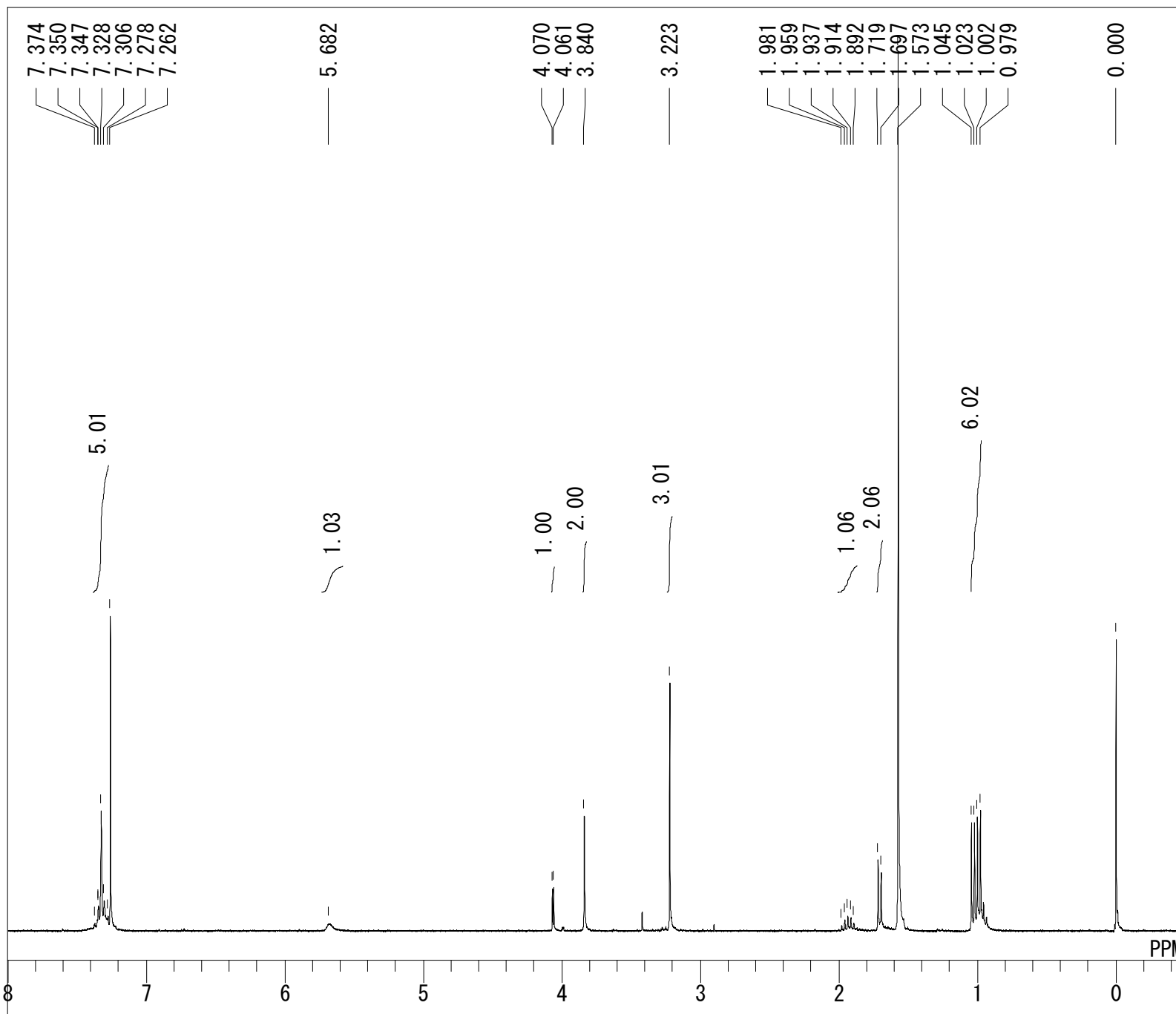
17

Filename = KST-Me-berkeleyamideD
 Author = delta
 Experiment = noe_ld_dpfge.jxp
 Sample Id = KST-Me-berkeleyamideD
 Solvent = DMSO-D6
 Creation Time = 25-AUG-2021 19:03:15
 Revision Time = 27-AUG-2021 10:46:56
 Current Time = 27-AUG-2021 10:47:19

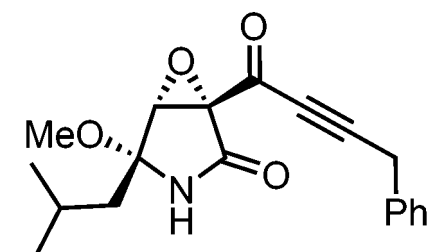
Comment = DPFGE NOE 1D
 Data Format = 1D COMPLEX
 Dim Size = 13107
 Dim Title = Proton
 Dim Units = [ppm]
 Dimensions = X
 Spectrometer = JNM-ECZ400S/L1

Field Strength = 9.389766 [T] (400 [MHz])
 X Acq Duration = 2.18628096 [s]
 X Domain = 1H
 X Freq = 399.78219838 [MHz]
 X Offset = 5 [ppm]
 X Points = 16384
 X Prescans = 2
 X Resolution = 0.45739775 [Hz]
 X Sweep = 7.4940048 [kHz]
 X Sweep Clipped = 5.99520384 [kHz]
 Irr Domain = Proton
 Irr Freq = 399.78219838 [MHz]
 Irr Offset = 5 [ppm]
 Tri Domain = Proton
 Tri Freq = 399.78219838 [MHz]
 Tri Offset = 5 [ppm]
 Blanking = 2 [us]
 Clipped = FALSE
 Decimation Req = r: 834 (833), g: 49
 Scans = 16
 Total Scans = 16

Relaxation Delay = 7 [s]
 Recvr Gain = 50
 Temp Get = 21.5 [dC]
 Mix Time = 0.5 [s]
 X Acq Time = 2.18628096 [s]
 X Atn = 2.4 [dB]
 X Pulse = 6.15 [us]
 Irr Mode = Off
 Obs Sel 180 = 40 [ms]
 Obs Sel Atn = 64.932 [dB]
 Obs Sel Offset = 4.36573172 [ppm]
 Obs Sel Shape = GAUSS
 Obs Sel Slp = 4.36573172 [ppm]
 Tri Mode = Off
 Comment 1 = *** Pulse ***
 Comment 11 = *** NOESY mixing time ***
 Comment 111 = *** presat time ***
 Comment 201 = *** obs_dante_presatu ***
 Comment 202 = *** irr_preaturation ***
 Comment 203 = *** tri_preaturation ***
 Comment 32 = *** Selective 180deg ***
 Comment 7 = *** Pulse Delay ***
 Comment 8 = *** Pulse Field Gradi ***
 Comment 900 = *** lock hold ***
 Dante Loop = 699



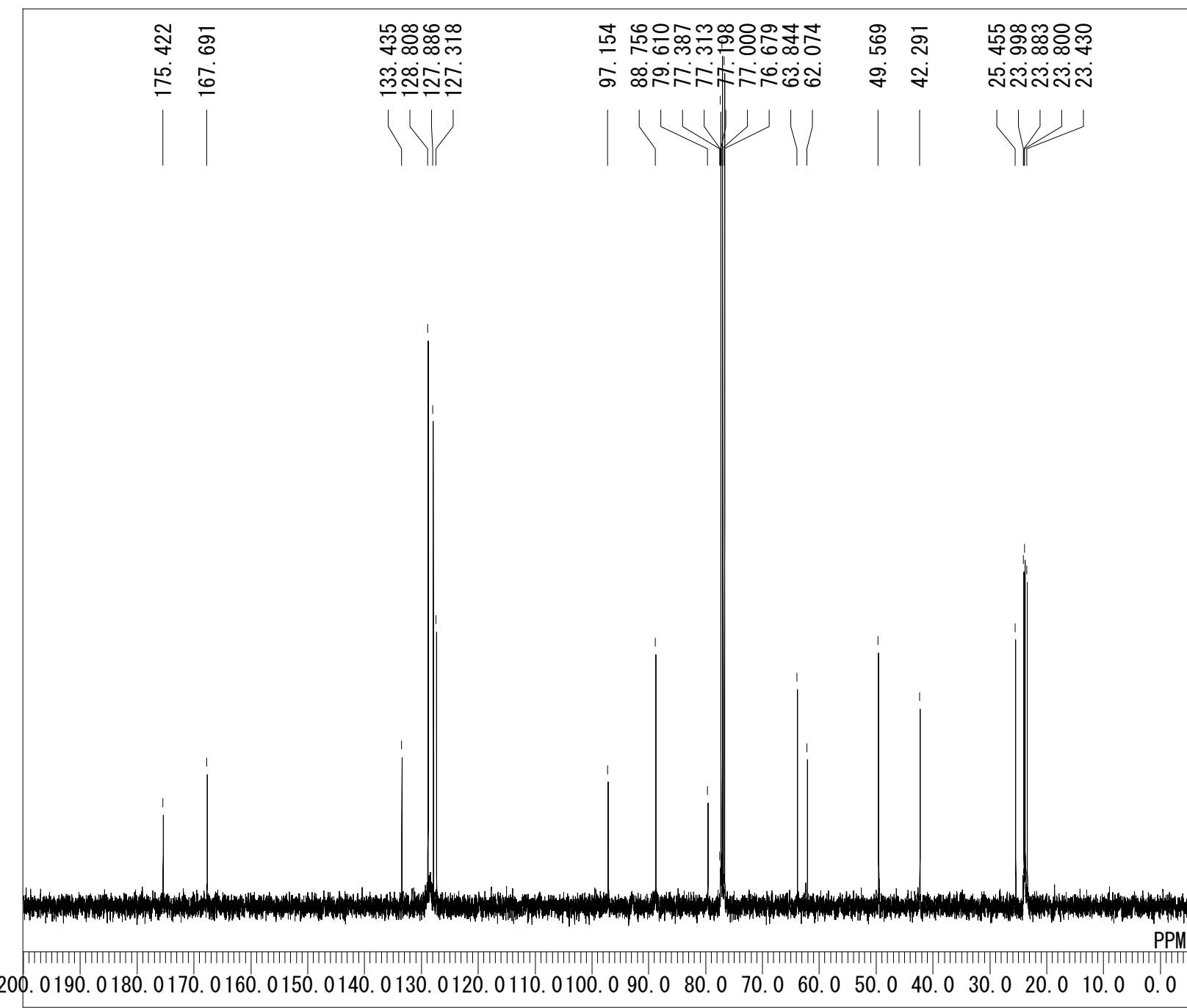
OBFRQ 300.40 MHz
 OBSET 130.00 KHz
 OBFIN 1150.00 Hz
 POINT 32768
 FREQU 6006.01 Hz
 SCANS 16
 ACQTM 5.4559 sec
 PD 1.5440 sec
 PW1 5.20 usec
 IRNUC 1H
 CTEMP 25.3 c
 SLVNT CDCL3
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 22



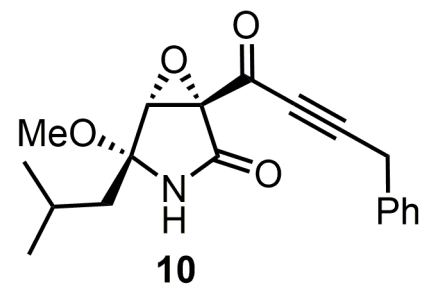
10

¹H NMR

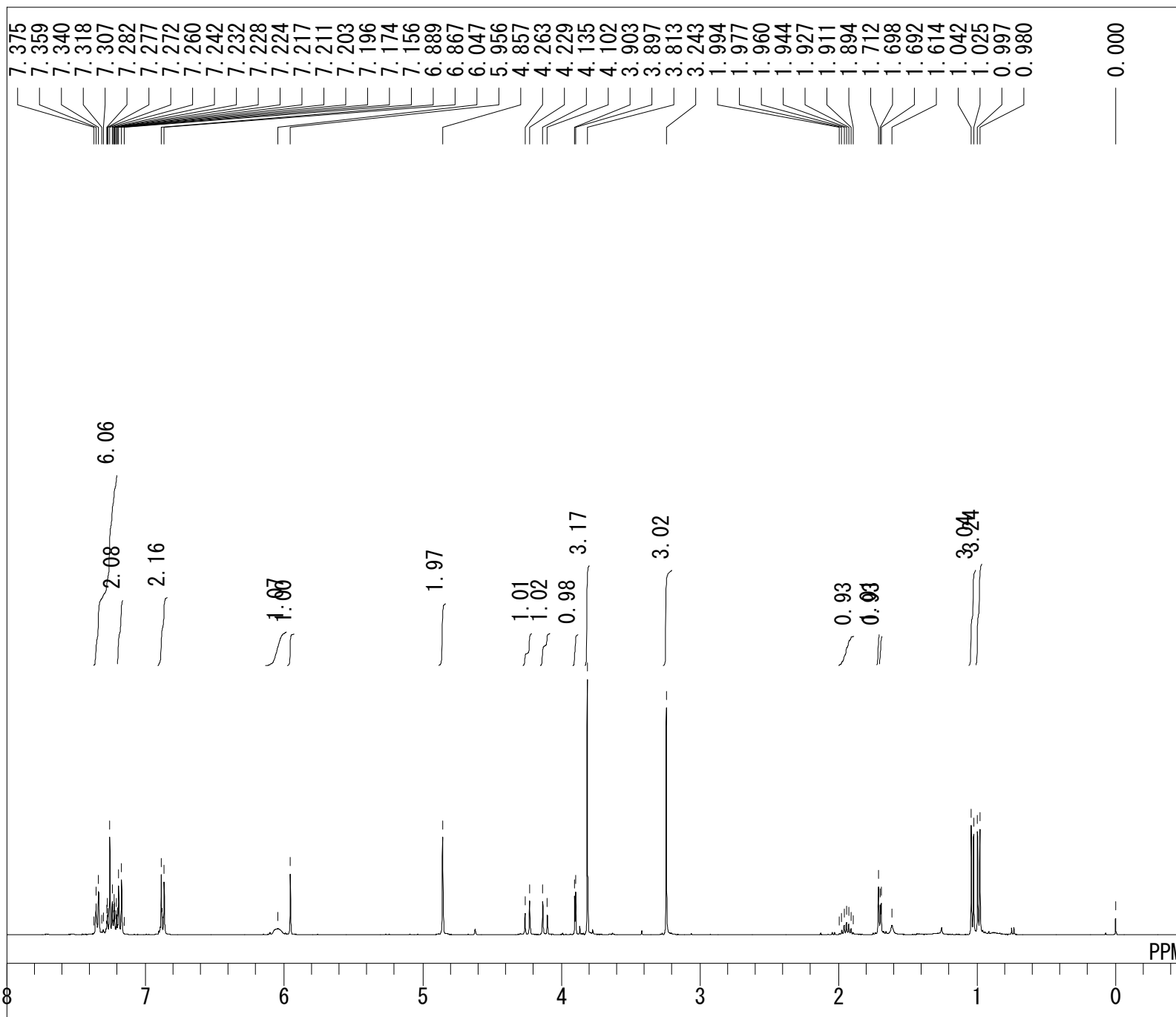
CDCl₃ (300 MHz)



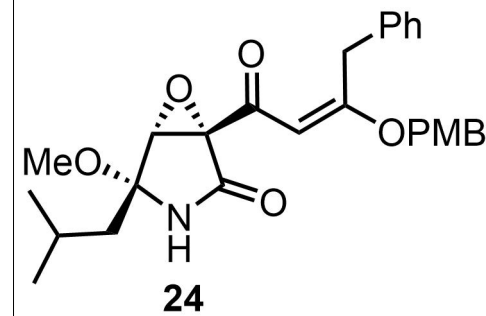
OBFRQ 100.40 MHz
 OBSET 125.00 KHz
 OBFIN 10500.00 Hz
 POINT 32768
 FREQU 27118.64 Hz
 SCANS 150
 ACQTM 1.2083 sec
 PD 1.7920 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 26.6 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 25



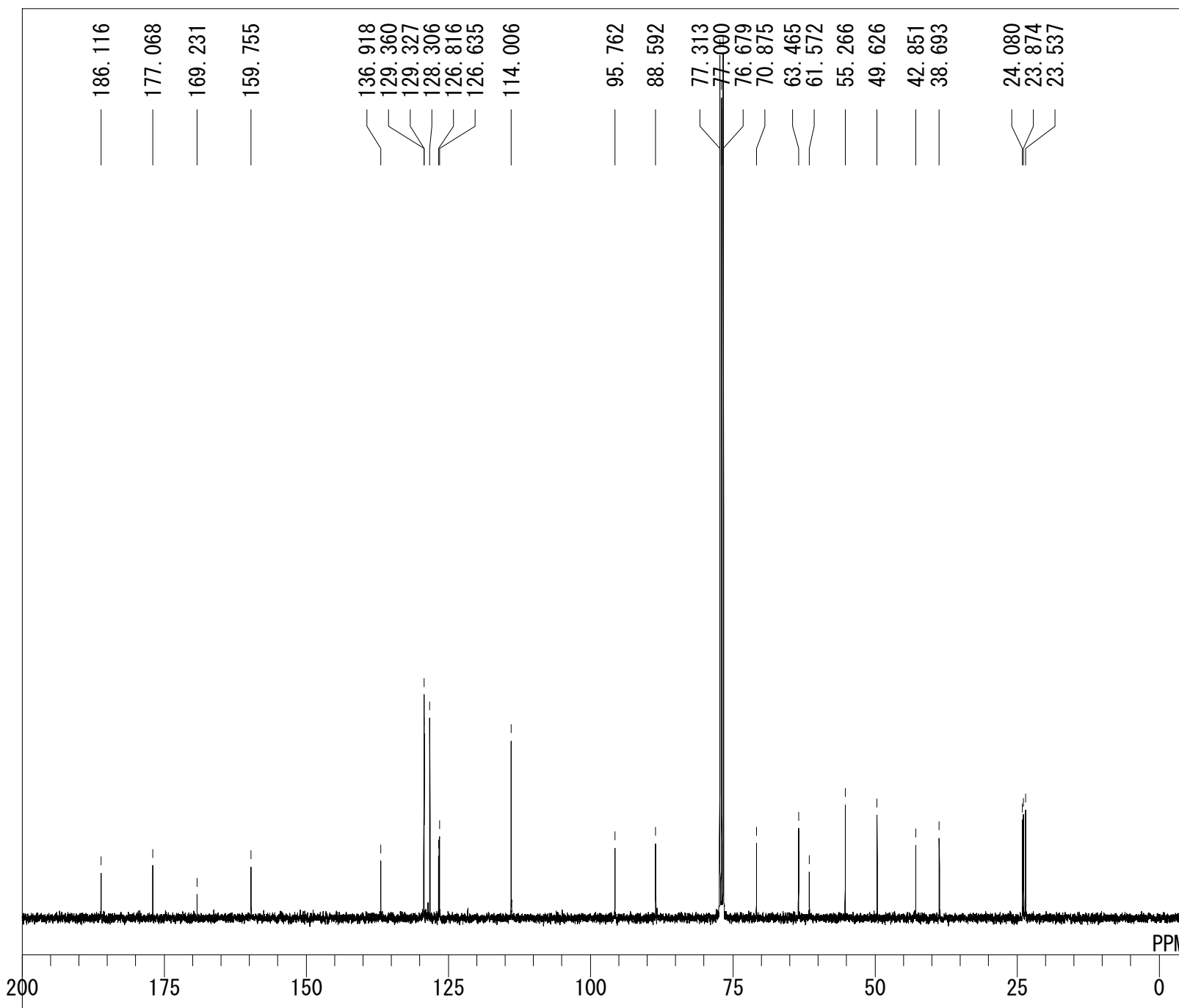
¹³C NMR
 CDCl₃ (100 MHz)



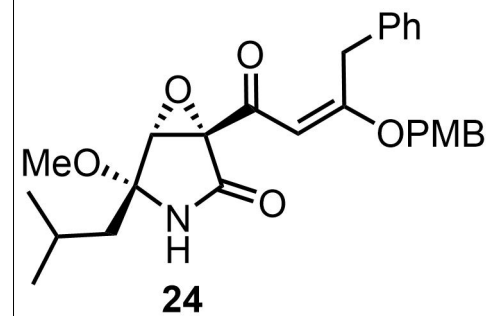
OBFRQ 399.65 MHz
 OBSET 124.00 KHz
 OBFIN 10500.00 Hz
 POINT 16384
 FREQU 7992.01 Hz
 SCANS 8
 ACQTM 2.0500 sec
 PD 4.9500 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 24.0 c
 SLVNT CDCL3
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 18



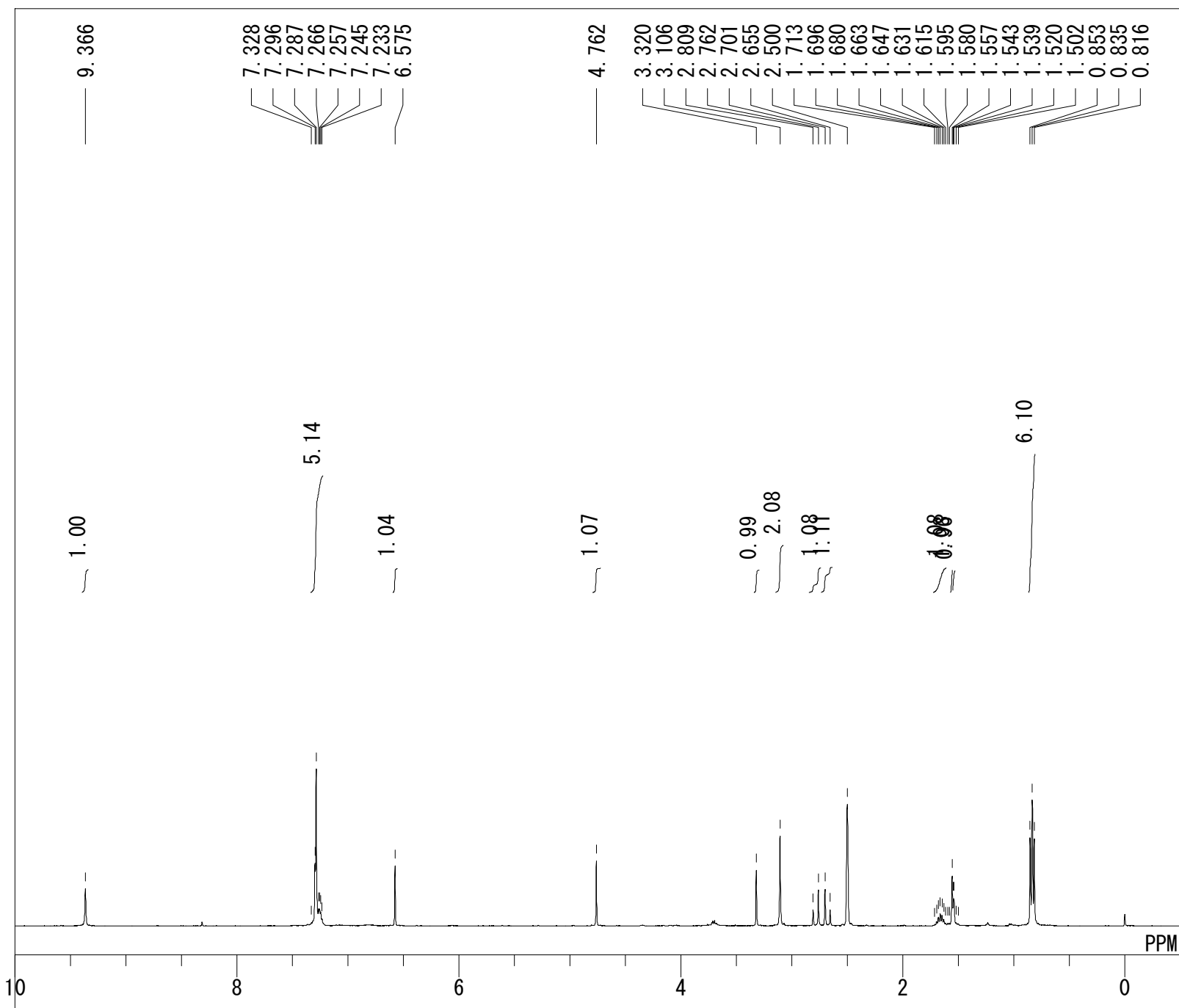
¹H NMR
 CDCl₃ (400 MHz)



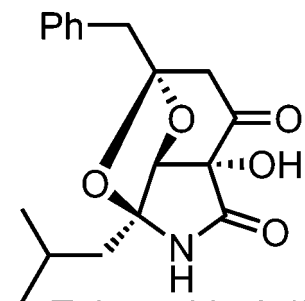
OBFRQ 100.40 MHz
 OBSET 125.00 KHz
 OBFIN 10500.00 Hz
 POINT 32768
 FREQU 27118.64 Hz
 SCANS 1000
 ACQTM 1.2083 sec
 PD 1.7920 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 24.3 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 25



¹³C NMR
 CDCl₃ (100 MHz)



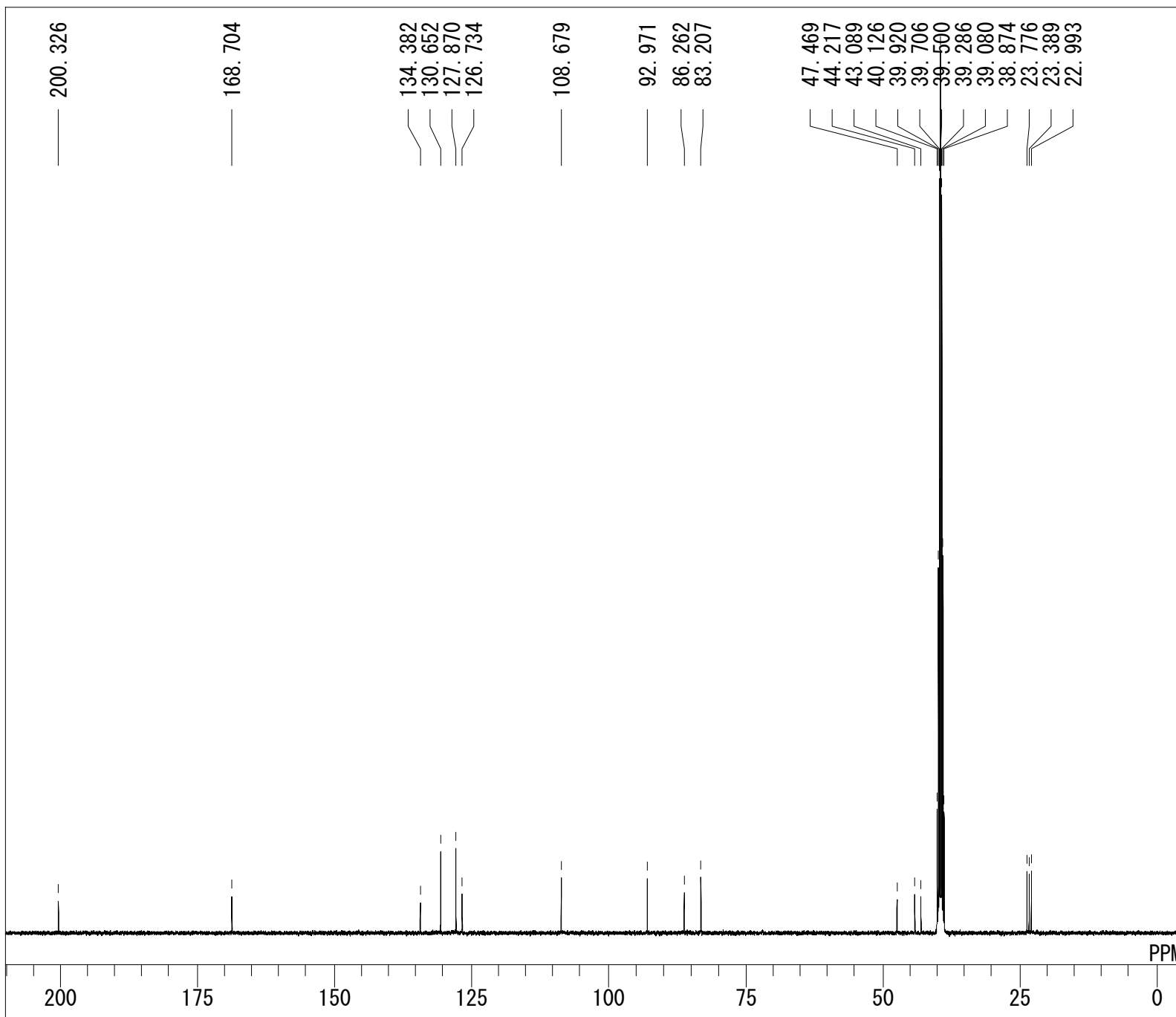
OBFRQ 399.65 MHz
 OBSET 124.00 KHz
 OBFIN 10500.00 Hz
 POINT 16384
 FREQU 7992.01 Hz
 SCANS 8
 ACQTM 2.0500 sec
 PD 4.9500 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 24.2 c
 SLVNT DMSO
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 18



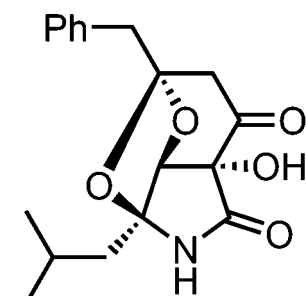
Taramide A (2)

¹H NMR

DMSO-*d*₆ (400 MHz)



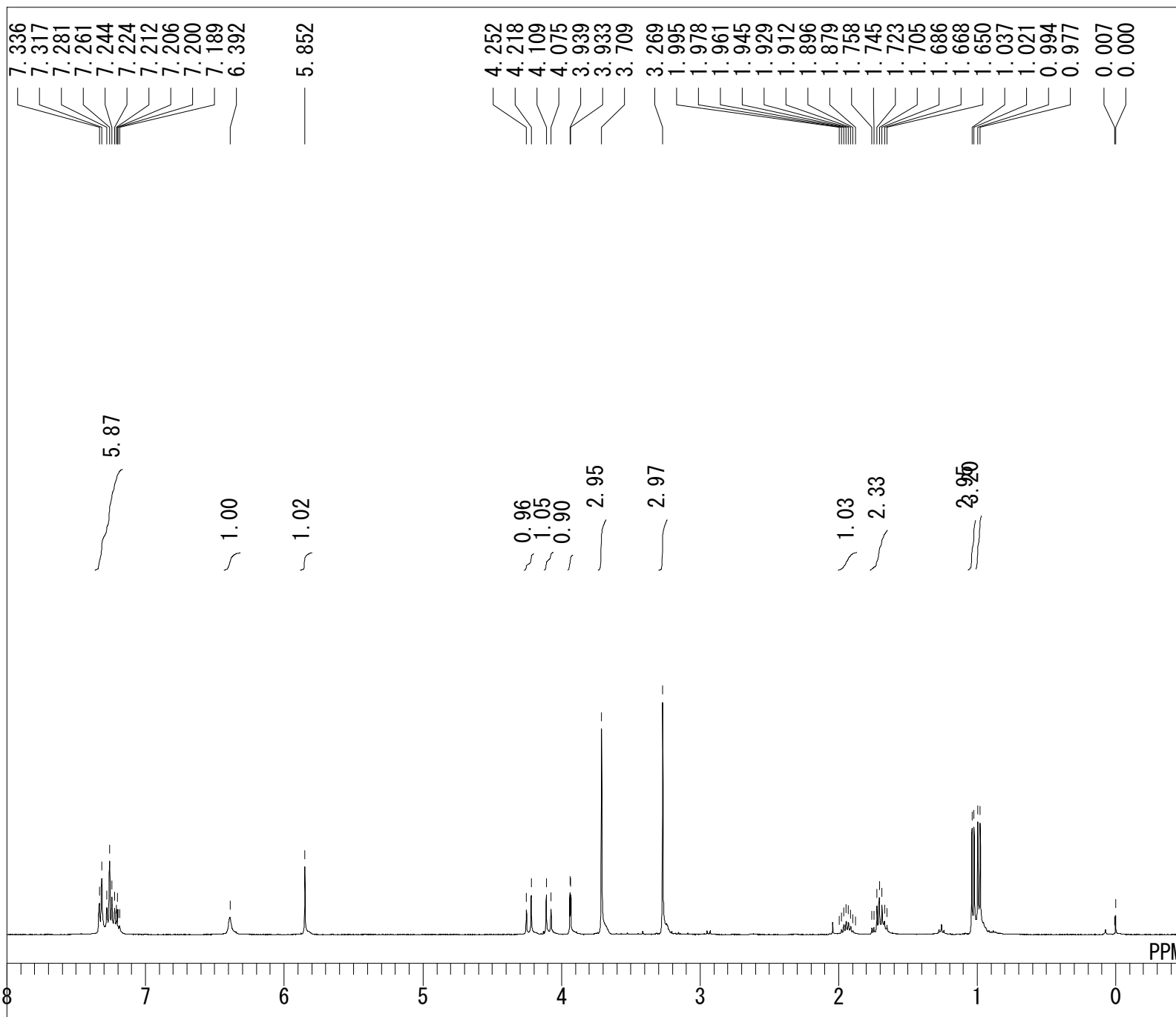
OBFRQ 100.40 MHz
 OBSET 125.00 KHz
 OBFIN 10500.00 Hz
 POINT 32768
 FREQU 27118.64 Hz
 SCANS 1972
 ACQTM 1.2083 sec
 PD 1.7920 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 24.3 c
 SLVNT DMSO
 EXREF 39.50 ppm
 BF 1.20 Hz
 RGAIN 24



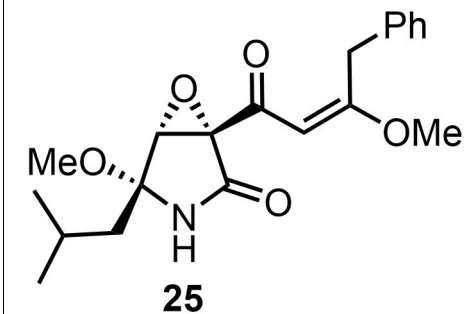
Taramide A (2)

¹³C NMR

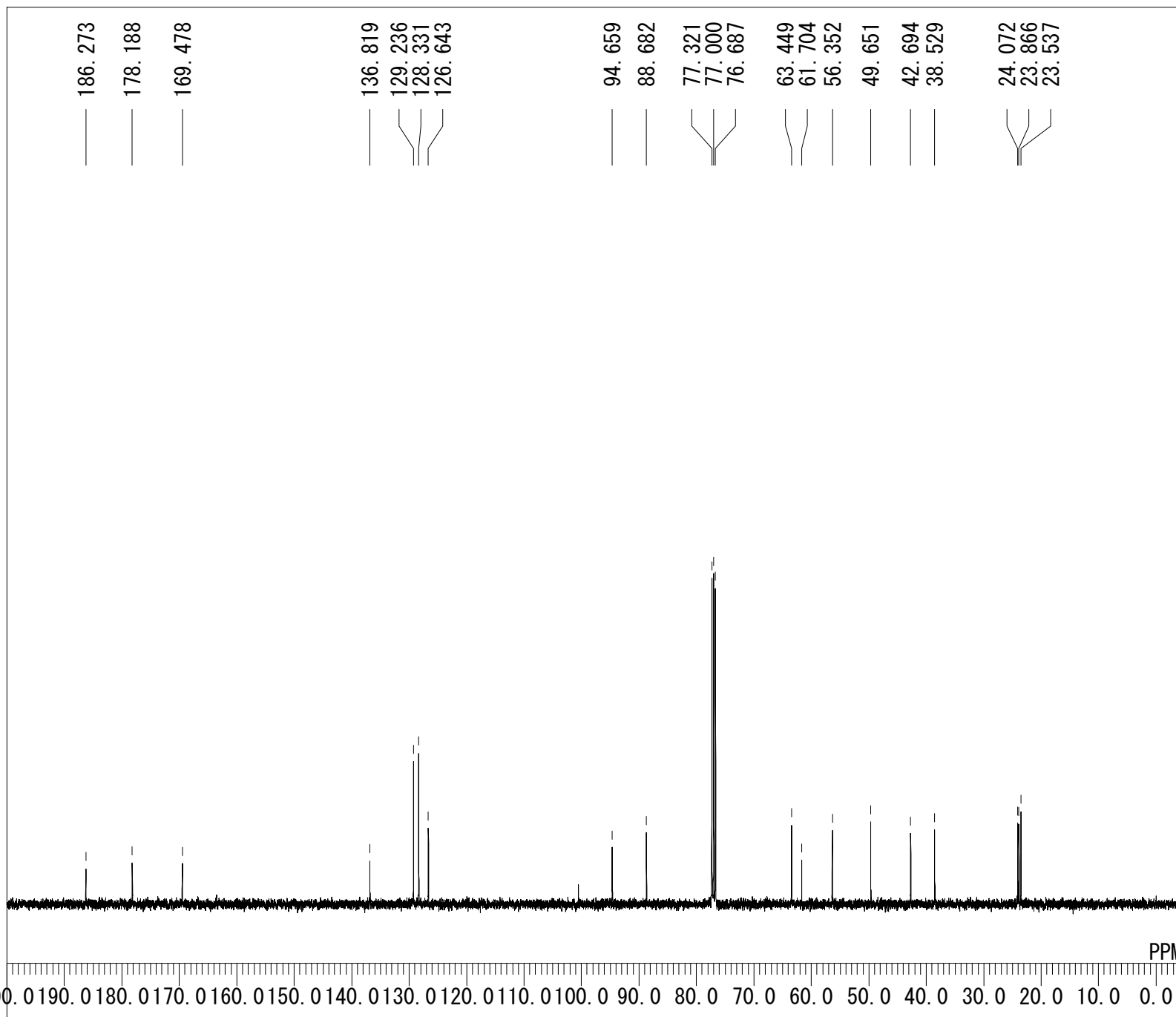
DMSO-*d*₆ (100 MHz)



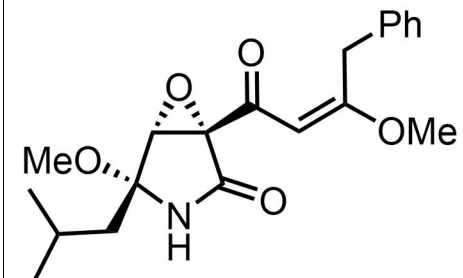
OBFRQ 399.65 MHz
 OBSET 124.00 KHz
 OBFIN 10500.00 Hz
 POINT 16384
 FREQU 7992.01 Hz
 SCANS 8
 ACQTM 2.0500 sec
 PD 4.9500 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 24.0 c
 SLVNT CDCL3
 EXREF 0.00 ppm
 BF 0.10 Hz
 RGAIN 16



¹H NMR
 CDCl₃ (400 MHz)



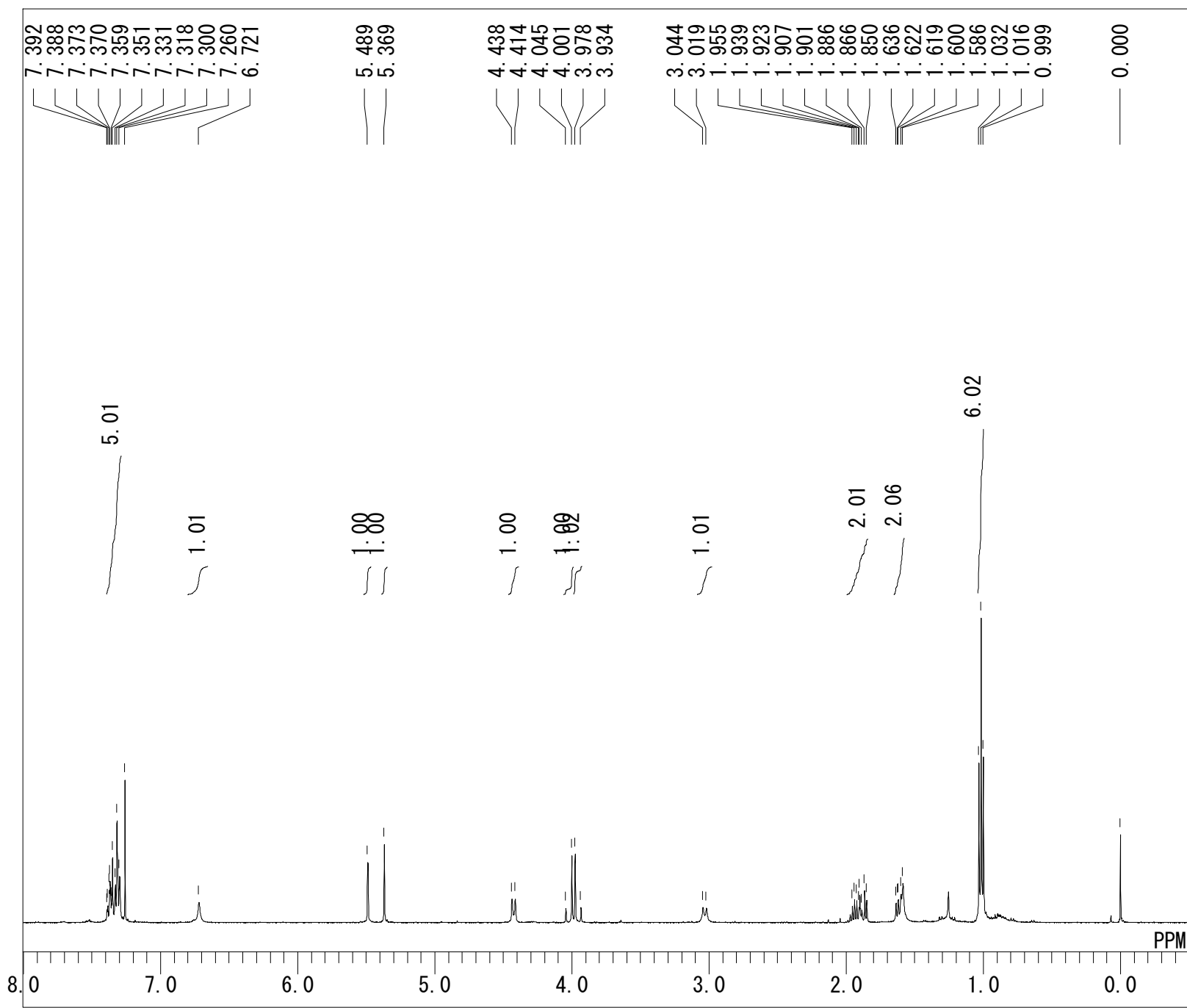
OBFRQ 100.40 MHz
 OBSET 125.00 KHz
 OBFIN 10500.00 Hz
 POINT 32768
 FREQU 27118.64 Hz
 SCANS 200
 ACQTM 1.2083 sec
 PD 1.7920 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 24.3 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 25



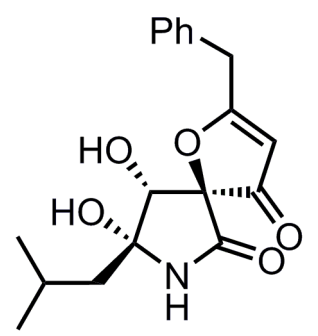
25

¹³C NMR

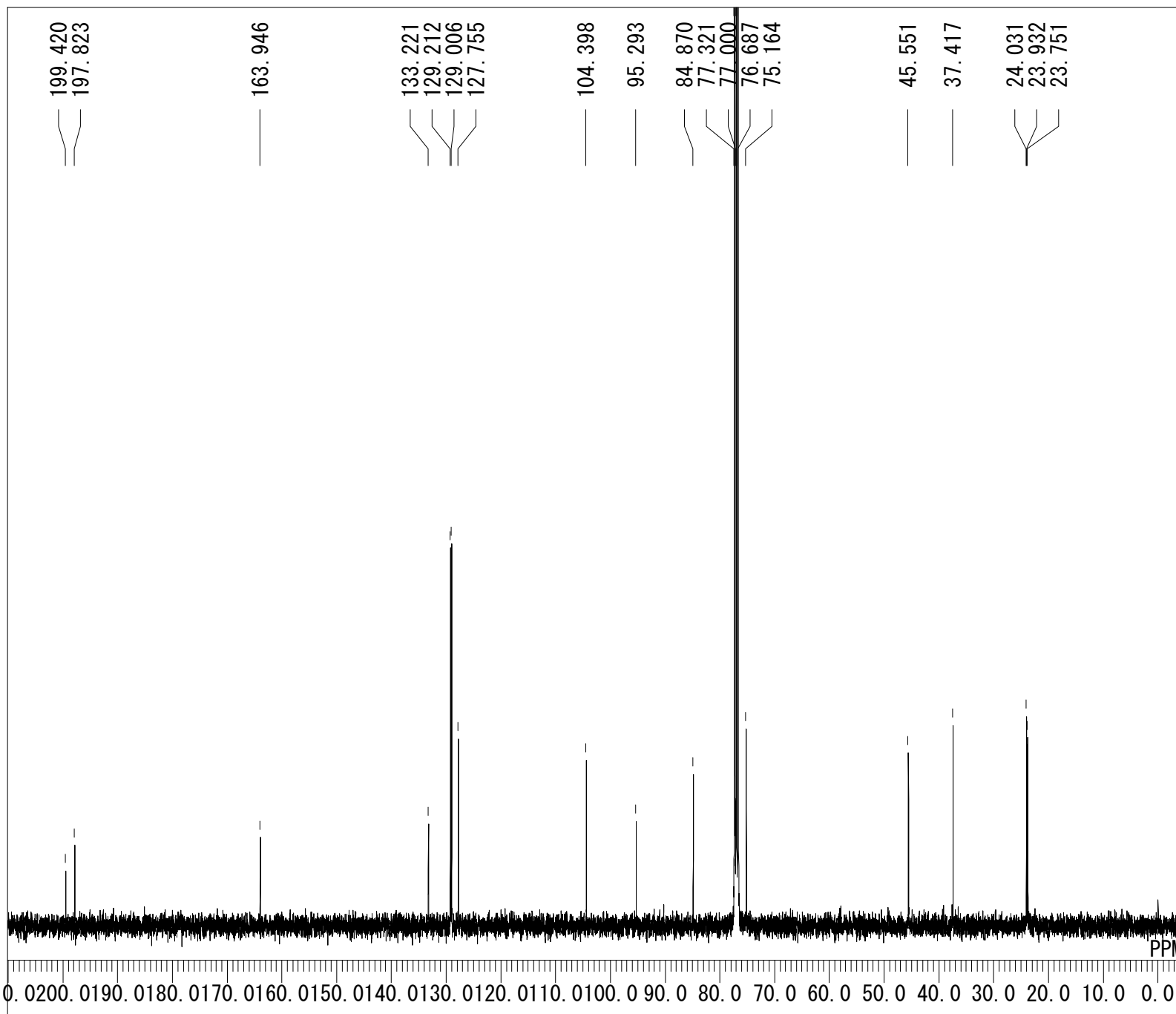
CDCl₃ (100 MHz)



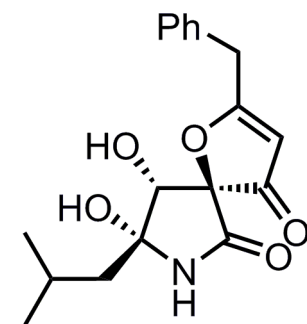
OBFREQ 399.65 MHz
 OBSET 124.00 KHz
 OBFIN 10500.00 Hz
 POINT 16384
 FREQU 7992.01 Hz
 SCANS 8
 ACQTM 2.0500 sec
 PD 4.9500 sec
 PW1 5.60 usec
 IRNUC 1H
 CTEMP 24.0 c
 SLVNT CDCL3
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 20



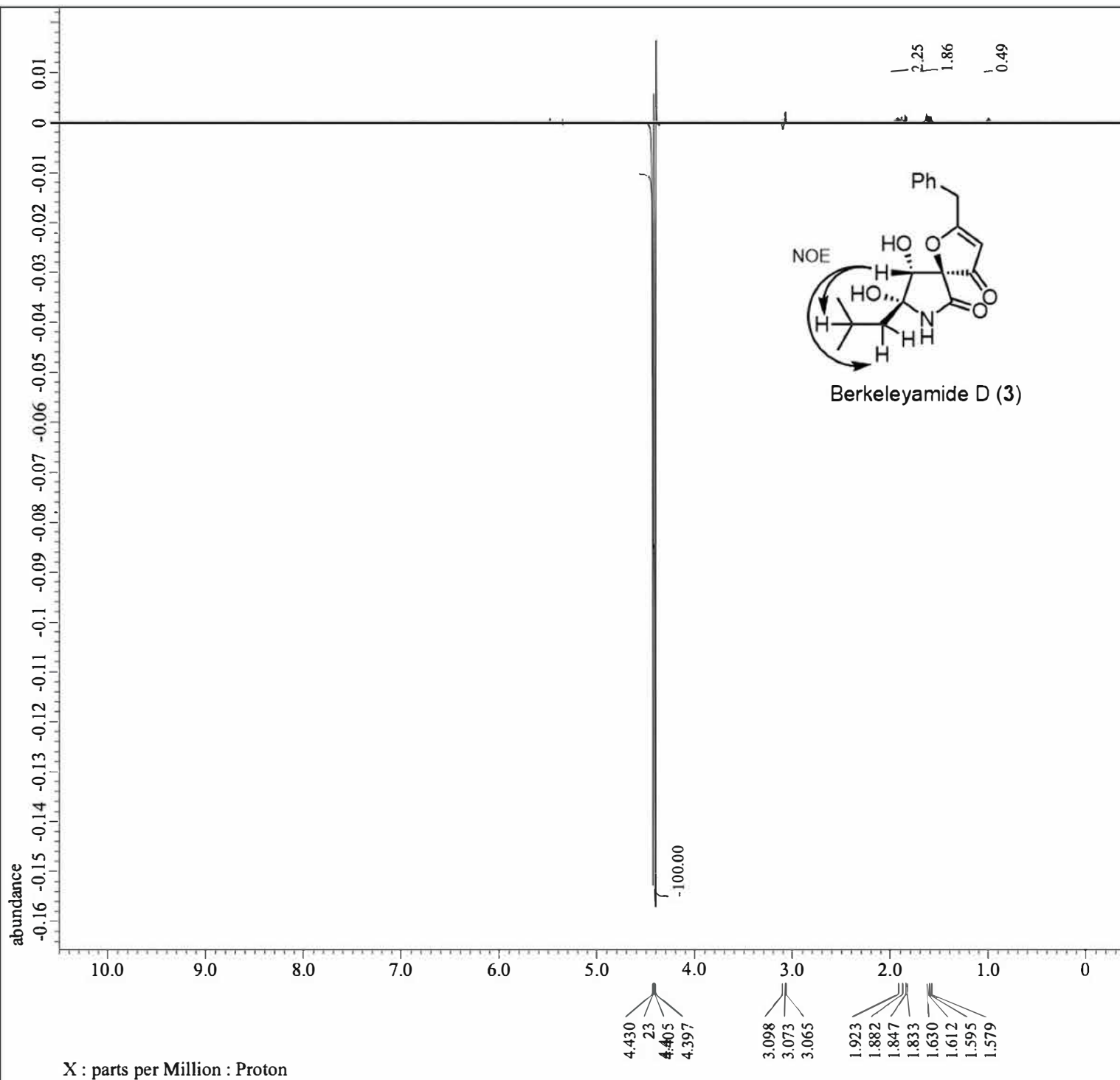
Berkeleyamide D (3)
¹H NMR
 CDCl₃ (400 MHz)



OBFREQ 100.40 MHz
 OBSET 125.00 KHz
 OBFIN 10500.00 Hz
 POINT 32768
 FREQU 27118.64 Hz
 SCANS 2000
 ACQTM 1.2083 sec
 PD 1.7920 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 24.6 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 1.20 Hz
 RGAIN 25



Berkeleyamide D (3)
¹³C NMR
 CDCl₃ (100 MHz)

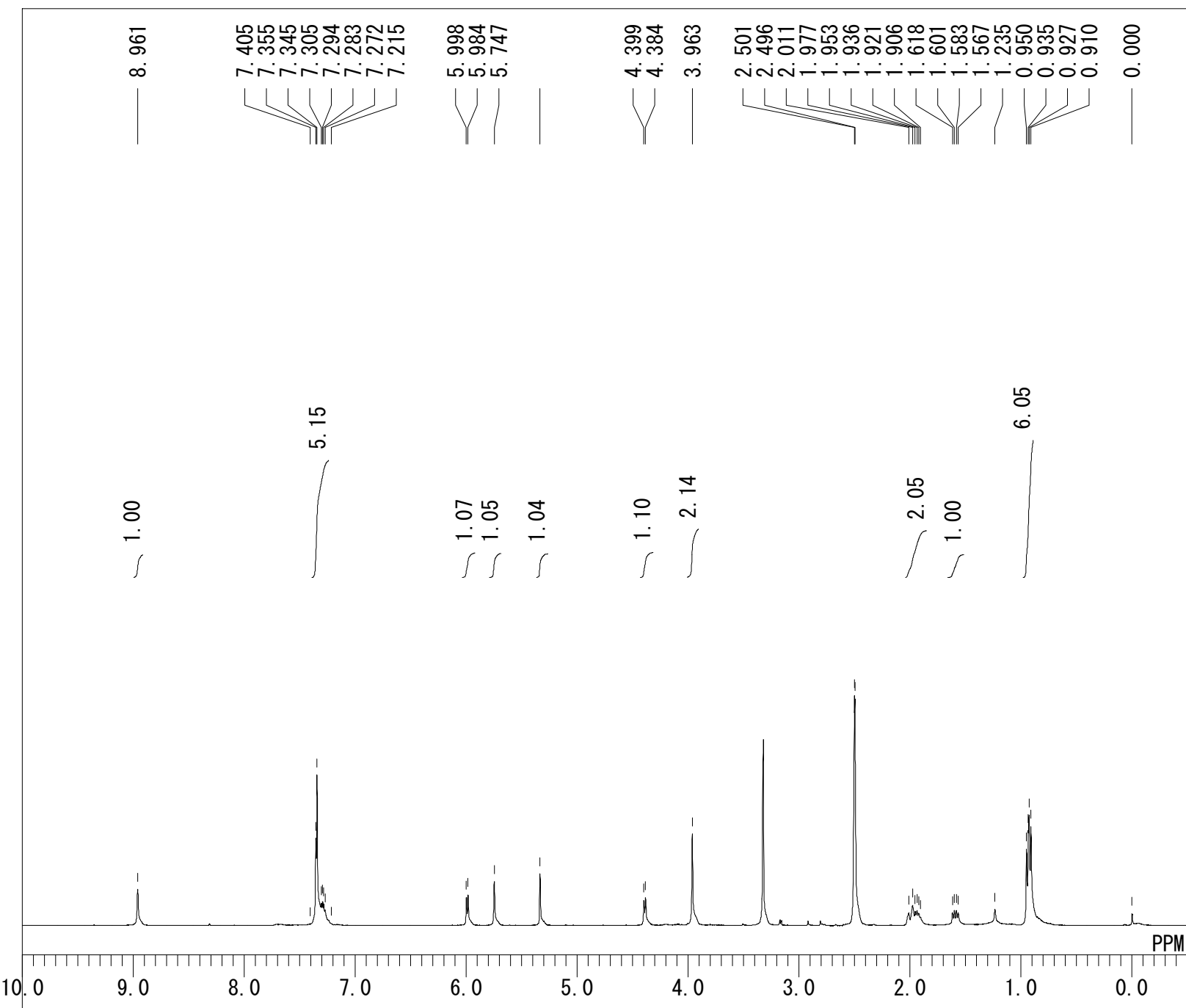


Filename = KST-berkeleyamideD_no
 Author = delta
 Experiment = noe_1d_dpfge.jxp
 Sample Id = KST-berkeleyamideD
 Solvent = CHLOROFORM-D
 Creation_Time = 23-AUG-2021 17:10:32
 Revision_Time = 27-AUG-2021 10:40:25
 Current_Time = 27-AUG-2021 10:40:57

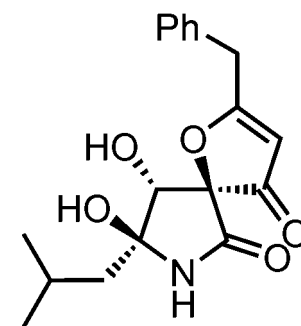
Comment = DPFGE NOE 1D
 Data_Format = 1D COMPLEX
 Dim_Size = 13107
 Dim_Title = Proton
 Dim_Units = [ppm]
 Dimensions = X
 Spectrometer = JNM-ECZ400S/L1

Field_Strength = 9.389766[T] (400 [MHz])
 X_Acq_Duration = 2.18628096[s]
 X_Domain = 1H
 X_Freq = 399.78219838 [MHz]
 X_Offset = 5 [ppm]
 X_Points = 16384
 X_Prescans = 2
 X_Resolution = 0.45739775 [Hz]
 X_Sweep = 7.4940048 [kHz]
 X_Sweep_Clippped = 5.99520384 [kHz]
 Irr_Domain = Proton
 Irr_Freq = 399.78219838 [MHz]
 Irr_Offset = 5 [ppm]
 Tri_Domain = Proton
 Tri_Freq = 399.78219838 [MHz]
 Tri_Offset = 5 [ppm]
 Blanking = 2 [us]
 Clipped = FALSE
 Decimation_Reg = r: 834 (833), g: 49
 Scans = 16
 Total_Scans = 16

Relaxation_Delay = 7 [s]
 Recvr_Gain = 50
 Temp_Get = 21.6 [dC]
 Mix_Time = 0.5 [s]
 X_Acq_Time = 2.18628096 [s]
 X_Atn = 2.4 [dB]
 X_Pulse = 6.15 [us]
 Irr_Mode = Off
 Obs_Sel_180 = 40 [ms]
 Obs_Sel_Atn = 64.932 [dB]
 Obs_Sel_Offset = 4.42566776 [ppm]
 Obs_Sel_Shape = GAUSS
 Obs_Sel_Slp = 4.42566776 [ppm]
 Tri_Mode = Off
 Comment_1 = *** Pulse ***
 Comment_11 = *** NOESY mixing time ***
 Comment_111 = *** presat_time ***
 Comment_201 = *** obs_dante_presatu ***
 Comment_202 = *** irr_preaturation ***
 Comment_203 = *** tri_preaturation ***
 Comment_32 = *** Selective 180deg ***
 Comment_7 = *** Pulse Delay ***
 Comment_8 = *** Pulse Field Gradi ***
 Comment_900 = *** lock hold ***
 Dante_Loop = 699



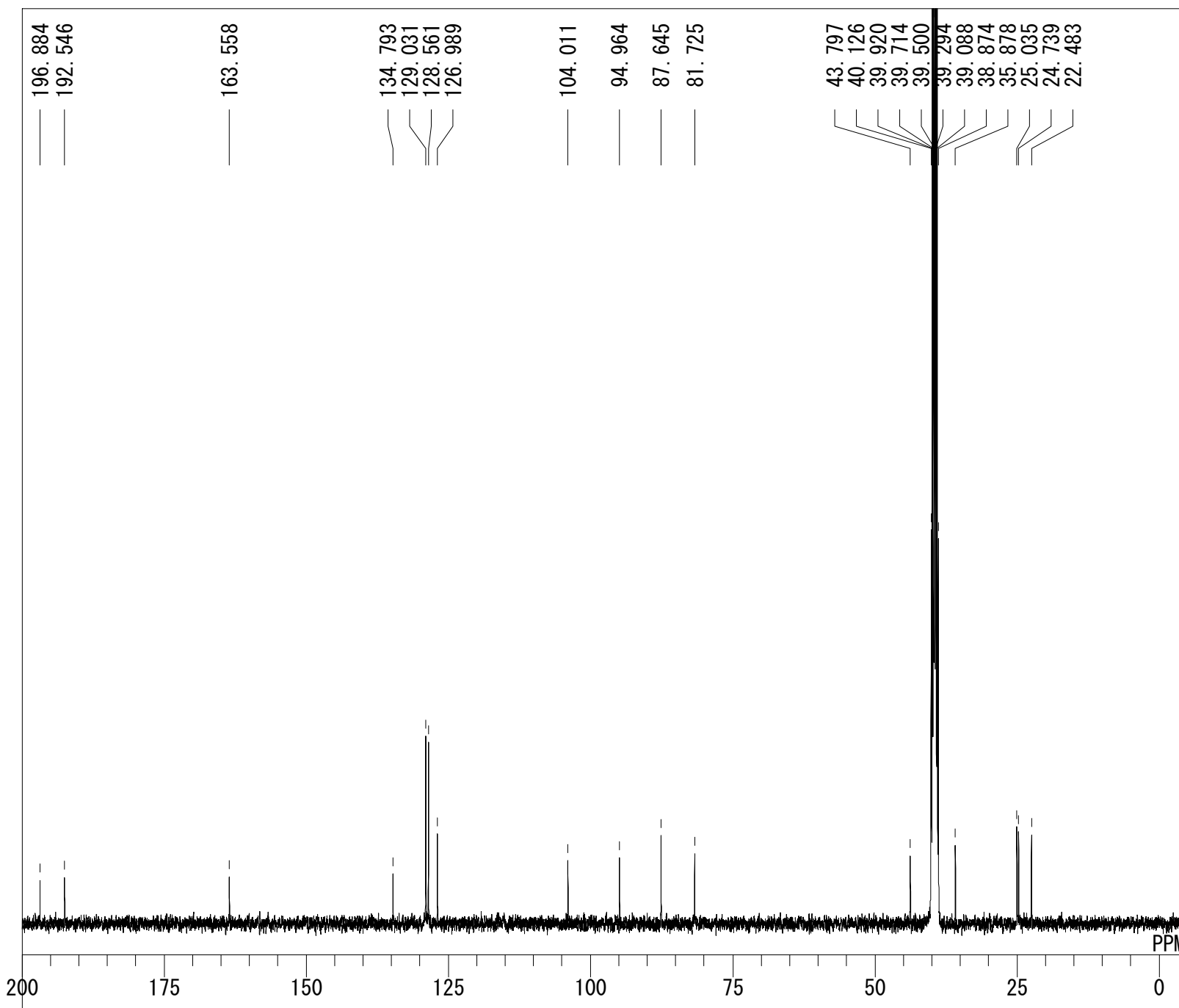
OBFRQ 399.65 MHz
 OBSET 124.00 KHz
 OBFIN 10500.00 Hz
 POINT 16384
 FREQU 7992.01 Hz
 SCANS 32
 ACQTM 2.0500 sec
 PD 4.9500 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 23.6 c
 SLVNT DMSO
 EXREF 0.00 ppm
 BF 0.12 Hz
 RGAIN 19



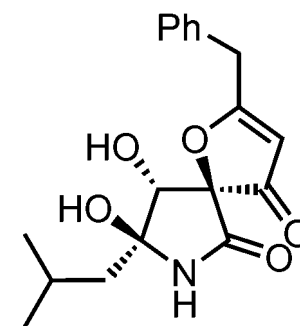
14-*epi*-3

¹H NMR

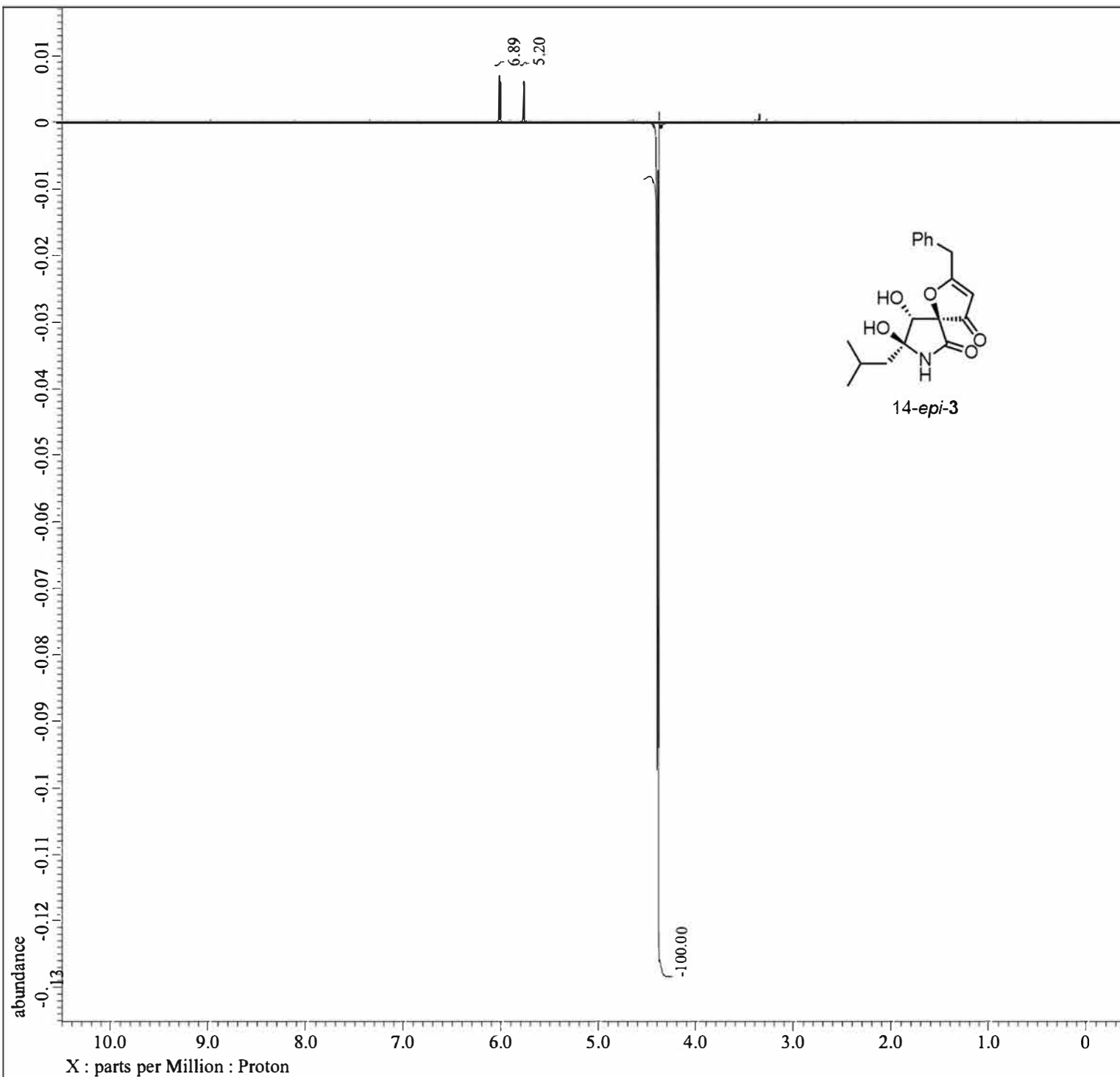
DMSO-*d*₆ (400 MHz)



OBFREQ 100.40 MHz
 OBSET 125.00 KHz
 OBFIN 10500.00 Hz
 POINT 32768
 FREQU 27118.64 Hz
 SCANS 1500
 ACQTM 1.2083 sec
 PD 1.7920 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 24.2 c
 SLVNT DMSO
 EXREF 39.50 ppm
 BF 2.00 Hz
 RGAIN 25



14-*epi*-3
¹³C NMR
 DMSO-*d*₆ (100 MHz)



Filename = KST-berkeleyamideD-is
 Author = delta
 Experiment = noe_ld_dpfge.jxp
 Sample Id = KST-berkeleyamideD-is
 Solvent = DMSO-D6
 Creation Time = 23-AUG-2021 18:13:17
 Revision Time = 27-AUG-2021 10:27:26
 Current Time = 27-AUG-2021 10:28:49

Comment = DPFGE NOE 1D
 Data Format = 1D COMPLEX
 Dim Size = 13107
 Dim Title = Proton
 Dim Units = [ppm]
 Dimensions = X
 Spectrometer = JNM-ECZ400S/L1

Field Strength = 9.389766[T] (400 [MHz])
 X Acq Duration = 2.18628096[s]
 X Domain = 1H
 X Freq = 399.78219838 [MHz]
 X Offset = 5 [ppm]
 X Points = 16384
 X Prescans = 2
 X Resolution = 0.45739775 [Hz]
 X Sweep = 7.4940048 [kHz]
 X Sweep Clipped = 5.99520384 [kHz]
 Irr Domain = Proton
 Irr Freq = 399.78219838 [MHz]
 Irr Offset = 5 [ppm]
 Tri Domain = Proton
 Tri Freq = 399.78219838 [MHz]
 Tri Offset = 5 [ppm]
 Blanking = 2 [us]
 Clipped = FALSE
 Decimation Req = r: 834 (833), g: 49
 Scans = 16
 Total Scans = 16

Relaxation Delay = 7[s]
 Recvr Gain = 50
 Temp Get = 21.4 [dC]
 Mix Time = 0.5 [s]
 X Acq Time = 2.18628096 [s]
 X Atn = 2.4 [dB]
 X Pulse = 6.15 [us]
 Irr Mode = Off
 Obs Sel 180 = 40 [ms]
 Obs Sel Atn = 64.932 [dB]
 Obs Sel Offset = 4.37970352 [ppm]
 Obs Sel Shape = GAUSS
 Obs Sel Slp = 4.37970352 [ppm]
 Tri Mode = Off
 Comment 1 = *** Pulse ***
 Comment 11 = *** NOESY mixing time ***
 Comment 111 = *** presat time ***
 Comment 201 = *** obs_dante_presatu ***
 Comment 202 = *** irr_preaturation ***
 Comment 203 = *** tri_preaturation ***
 Comment 32 = *** Selective 180deg ***
 Comment 7 = *** Pulse Delay ***
 Comment 8 = *** Pulse Field Gradi ***
 Comment 900 = *** lock hold ***
 Dante Loop = 699