

Electronic Supplementary Information for

Cytotoxic evaluation and DNA interaction of Ru^{II}-bipy complexes containing coumarin-based ligands

Patrícia S. V. B. de Almeida^a, Henrique Jefferson de Arruda^a, Gleyton Leonel S. Sousa^a, Felipe Vitório Ribeiro^a, José Aleixo de Azevedo-França^b, Larissa A. Ferreira^a, Guilherme P. Guedes^c, Heveline Silva^d, Arthur E. Kummerle^a and Amanda P. Neves^{a*}

^aInstituto de Química, Universidade Federal Rural do Rio de Janeiro, BR 465 Km 7, 23890-000, Seropédica, RJ, Brazil. E-mail: amandanevess@ufrj.br

^bInstituto de Ciências Exatas, Departamento de Química, Universidade Federal de Juiz de Fora, MG, Brazil.

^cInstituto de Química, Universidade Federal Fluminense, Niterói, RJ, Brazil.

^dInstituto de Ciências Exatas, Departamento de Química, Universidade Federal de Minas Gerais, MG, Brazil.

SUMMARY

I. Ethyl 3-(6-methyl-2-oxo-2 <i>H</i> -chromen-3-yl)-3-oxopropanoate (HL1).....	3
II. Ethyl 3-(7-(diethylamino)-2-oxo-2 <i>H</i> -chromen-3-yl)-3-oxopropanoate (HL2)	6
III. Ethyl 3-(8-methoxy-2-oxo-2 <i>H</i> -chromen-3-yl)-3-oxopropanoate (HL3).....	7
IV. <i>cis</i> -bis(2,2'-bipyridyl)-[3-(6-(methyl)-2-oxo-2 <i>H</i> -chromen-3-yl)-3-oxoethylpropanoate]-ruthenium (II) hexafluorophosphate (1).....	11
V. <i>cis</i> -bis(2,2'-bipyridyl)-[3-(7-(diethylamino)-2-oxo-2 <i>H</i> -chromen-3-yl)-3-oxoethylpropanoate] ruthenium (II) hexafluorophosphate (2)	14
VI. <i>cis</i> -bis(2,2'-bipyridyl)-[3-(8-(methoxy)-2-oxo-2 <i>H</i> -chromen-3-yl)-3-oxoethylpropanoate] ruthenium (II) hexafluorophosphate (3)	16
VII. X-Ray diffraction analysis.....	20
VIII. Biological studies	21

I. Ethyl 3-(6-methyl-2-oxo-2H-chromen-3-yl)-3-oxopropanoate (HL1)

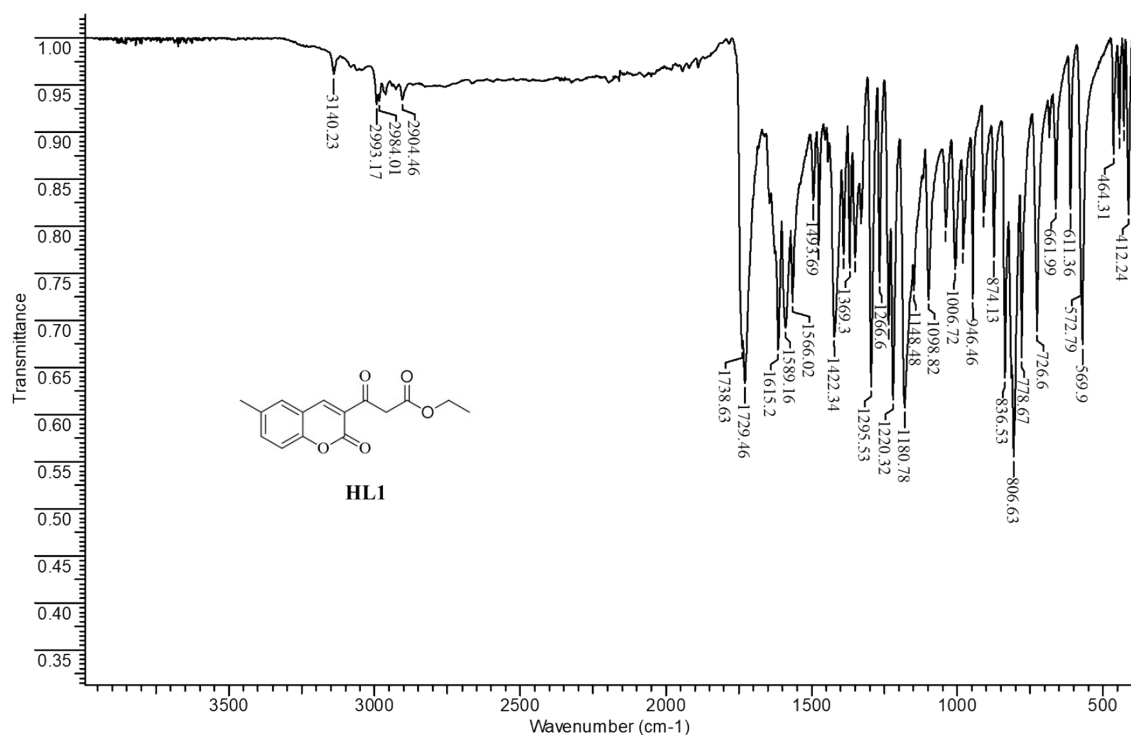


Figure S1: IR spectrum of HL1.

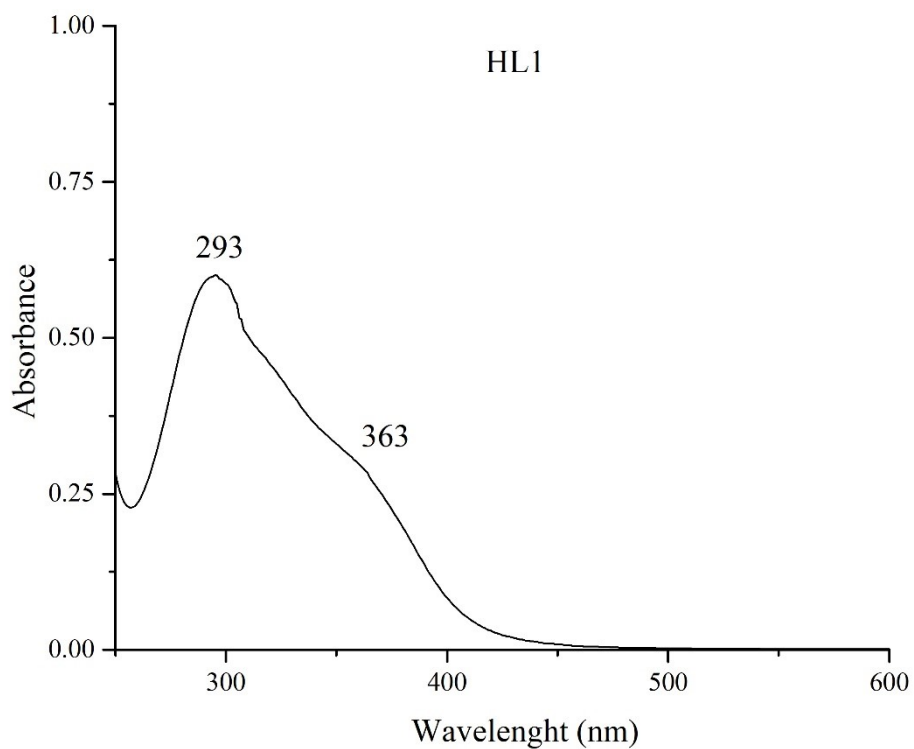


Figure S2: UV-Vis spectrum of HL1 in phosphate buffer at 5×10^{-5} M.

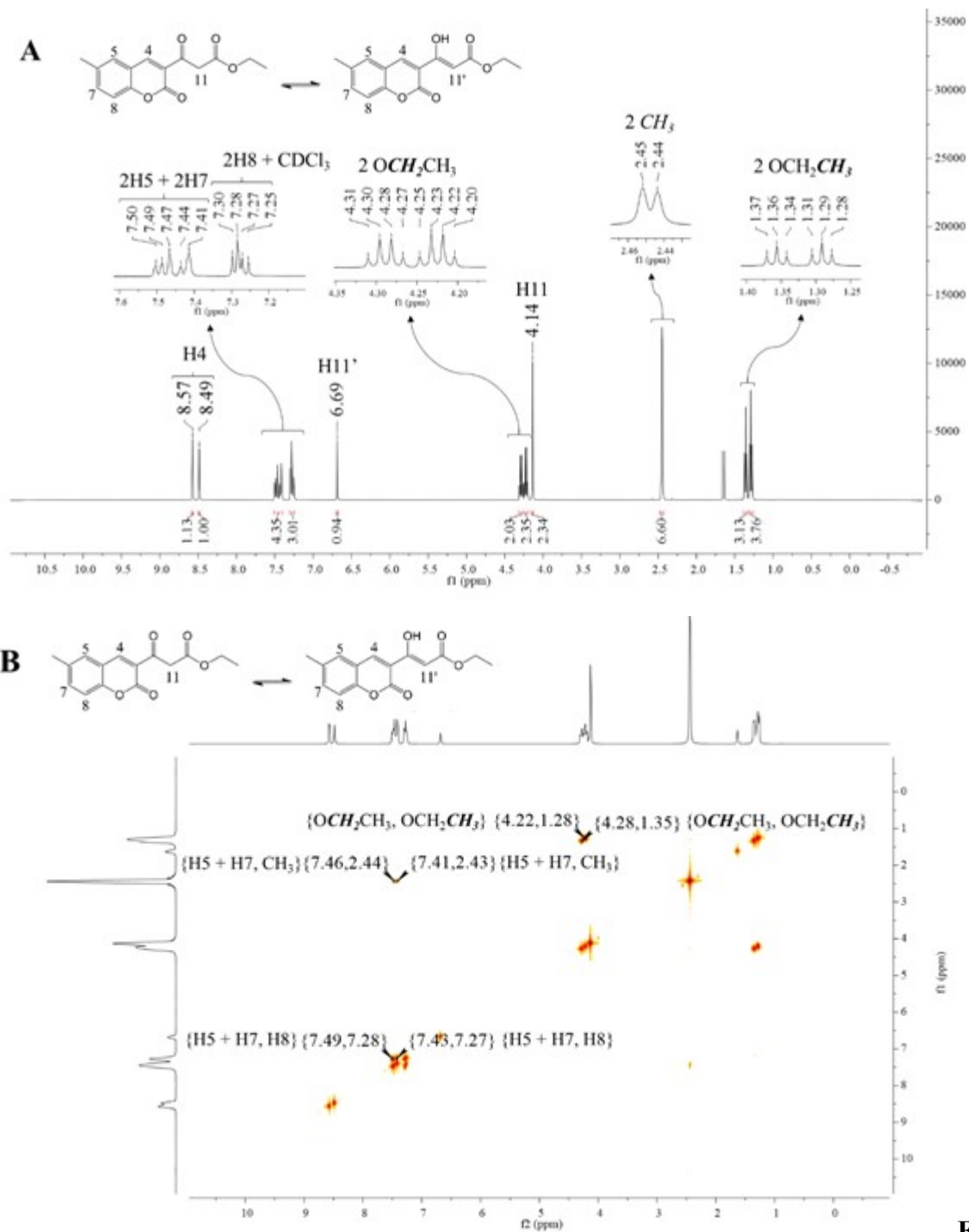


Figure S3: ^1H NMR spectrum of HL1 (A) and COSY spectrum (B). Solvent: CDCl₃.

20 #25-45 RT: 0.18-0.31 AV: 11 SM: 7G NL: 1.55E8
T: FTMS + p ESI Full ms [60.0000-800.0000]

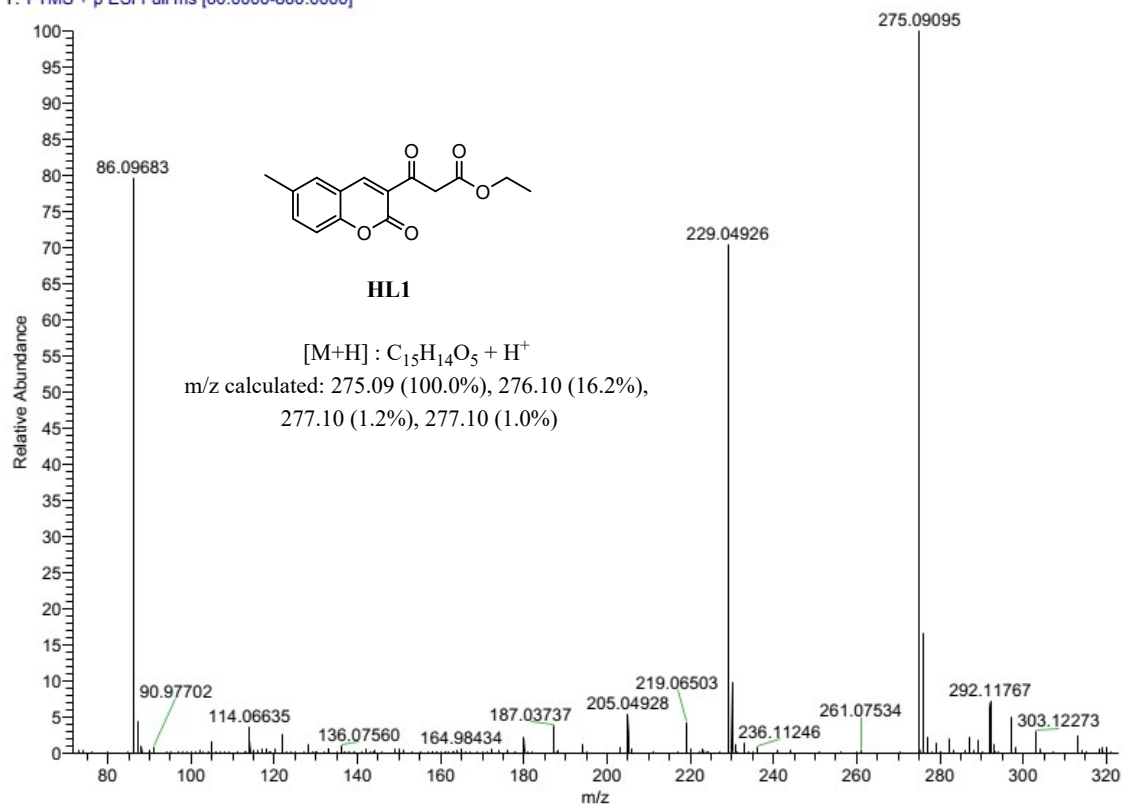


Figure S4: MS/ESI mass spectrum of **HL1**.

II. Ethyl 3-(7-(diethylamino)-2-oxo-2H-chromen-3-yl)-3-oxopropanoate (**HL2**)

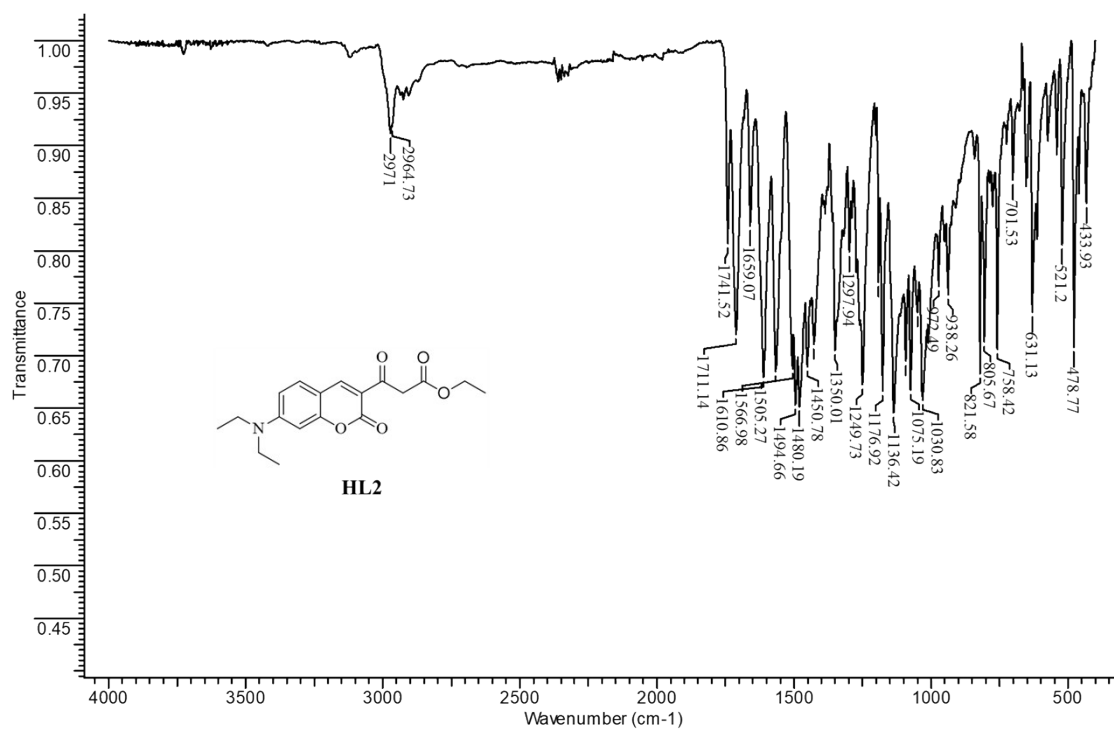


Figure S5: IR spectrum of **HL2**.

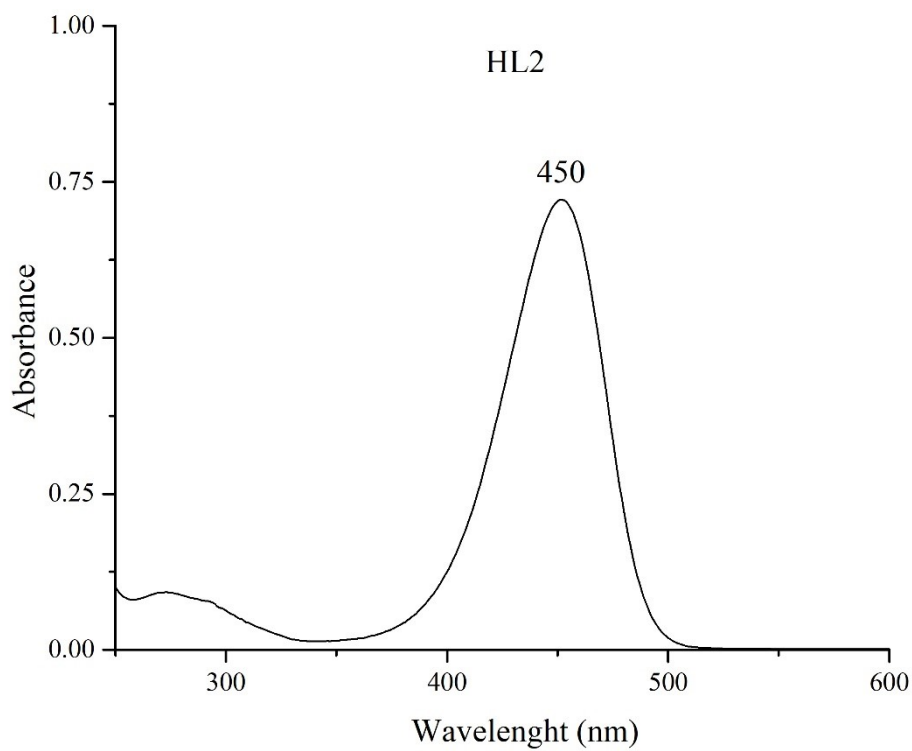
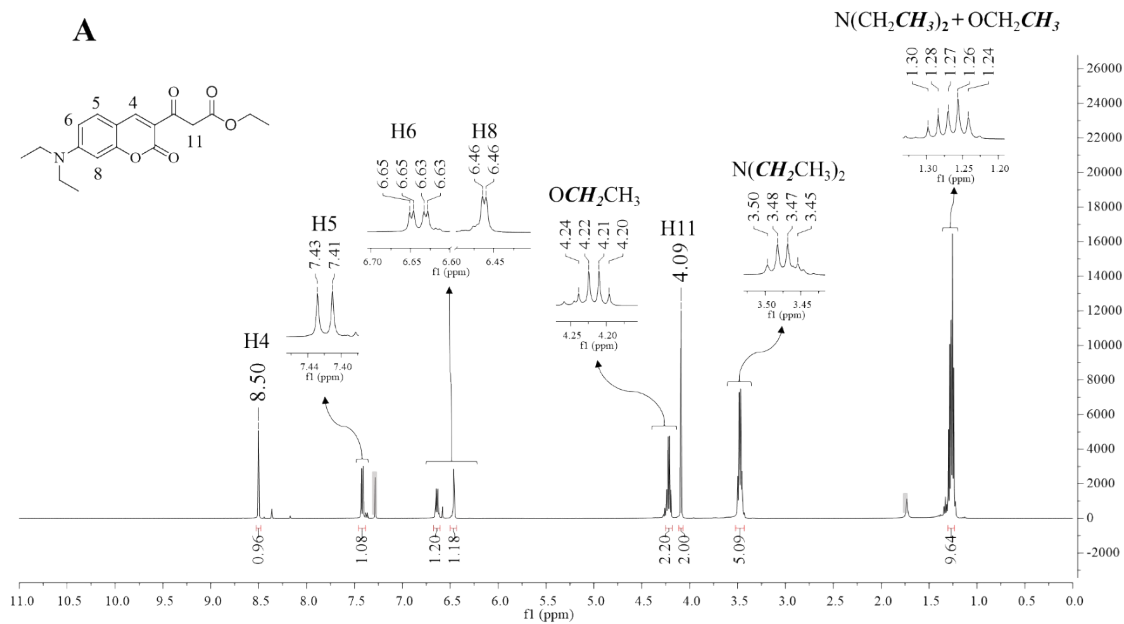
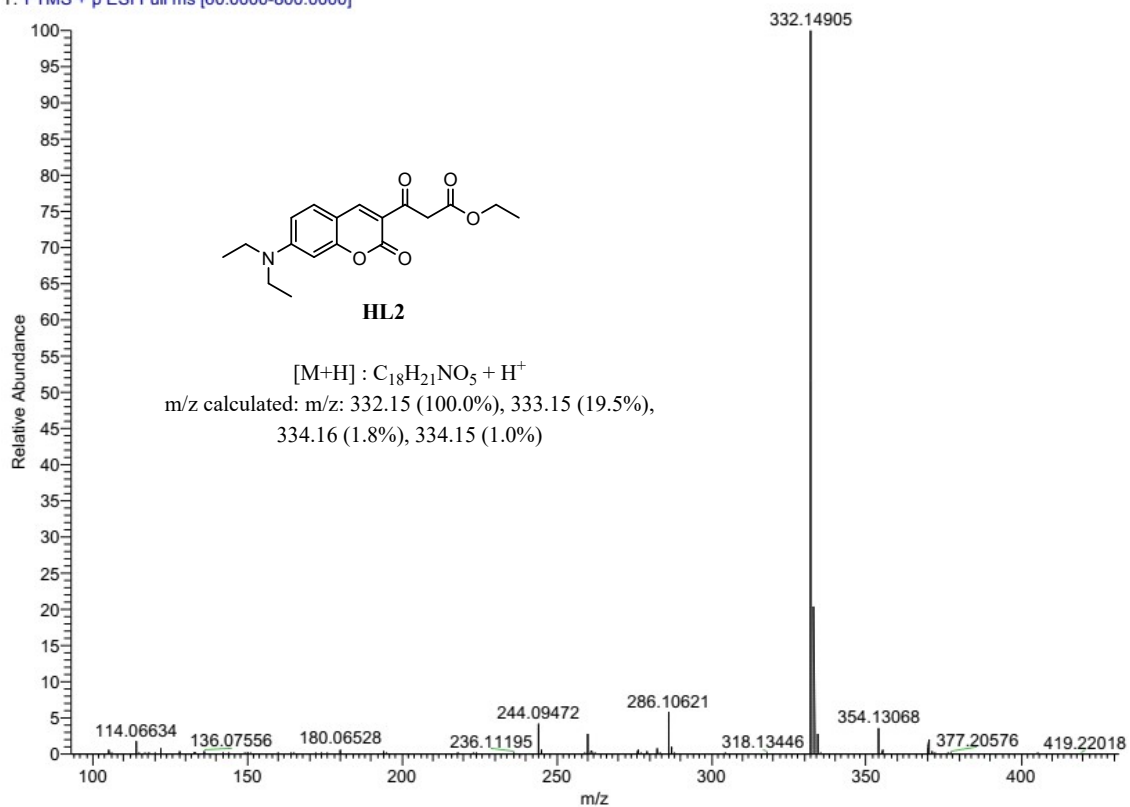


Figure S6: UV-Vis spectrum of **HL2** in phosphate buffer at 2×10^{-5} M.



19 #26-41 RT: 0.19-0.29 AV: 8 SM: 7G NL: 4.75E8
T: FTMS + p ESI Full ms [60.0000-800.0000]



III. Ethyl 3-(8-methoxy-2-oxo-2*H*-chromen-3-yl)-3-oxopropanoate (**HL3**)

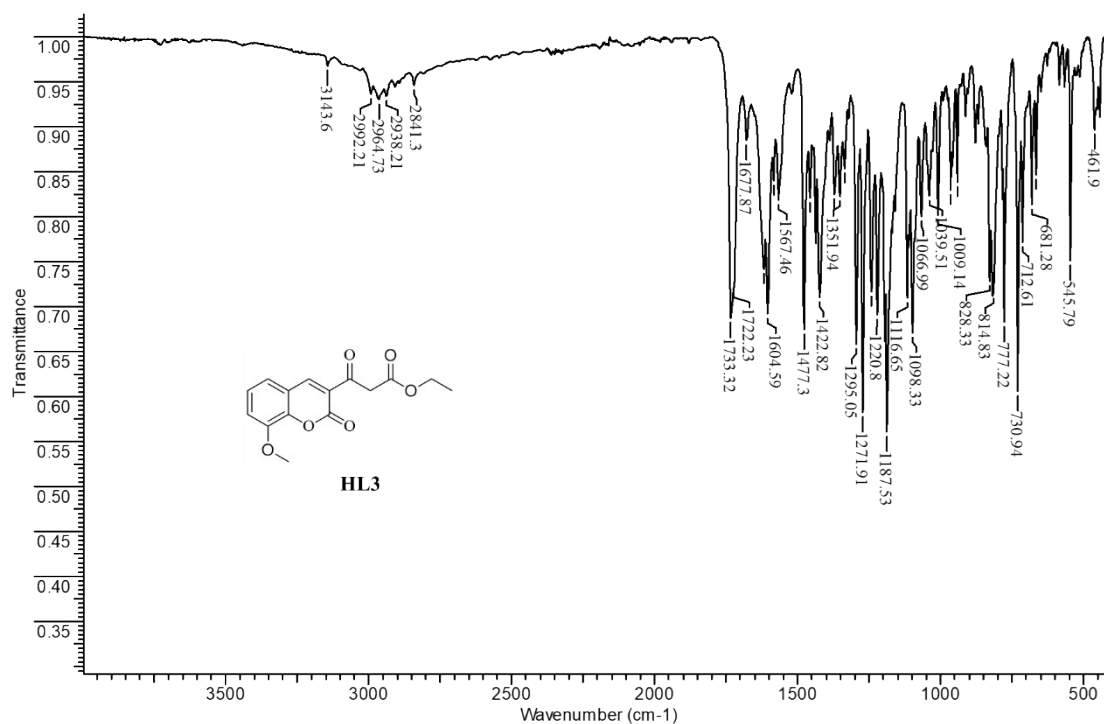


Figure S9: IR spectrum of **HL3**.

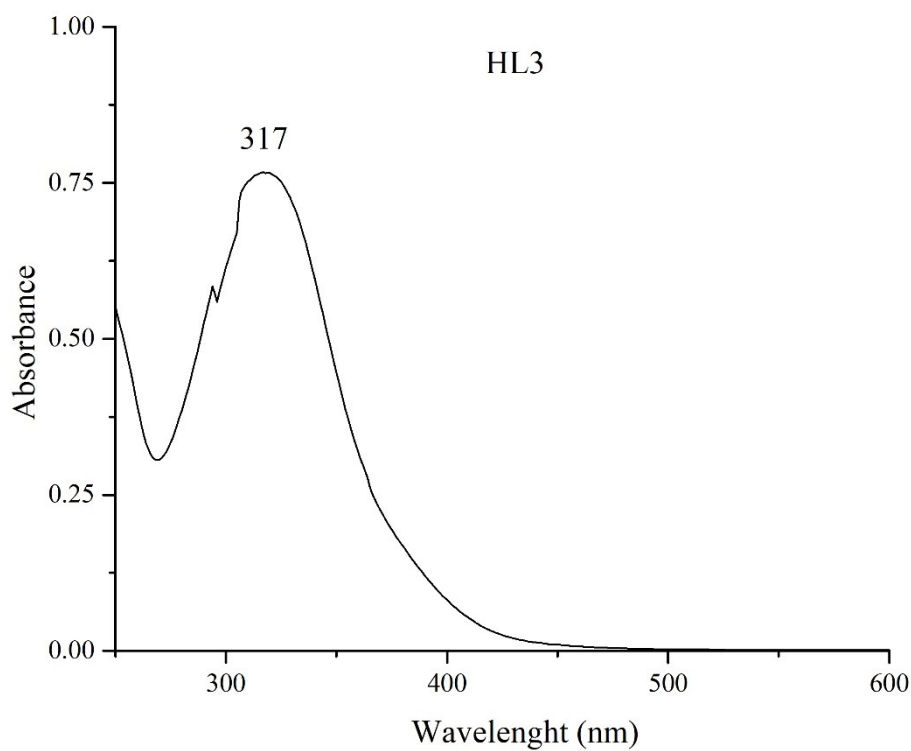


Figure S10: UV-Vis spectrum of **HL3** in phosphate buffer at 6×10^{-5} M.

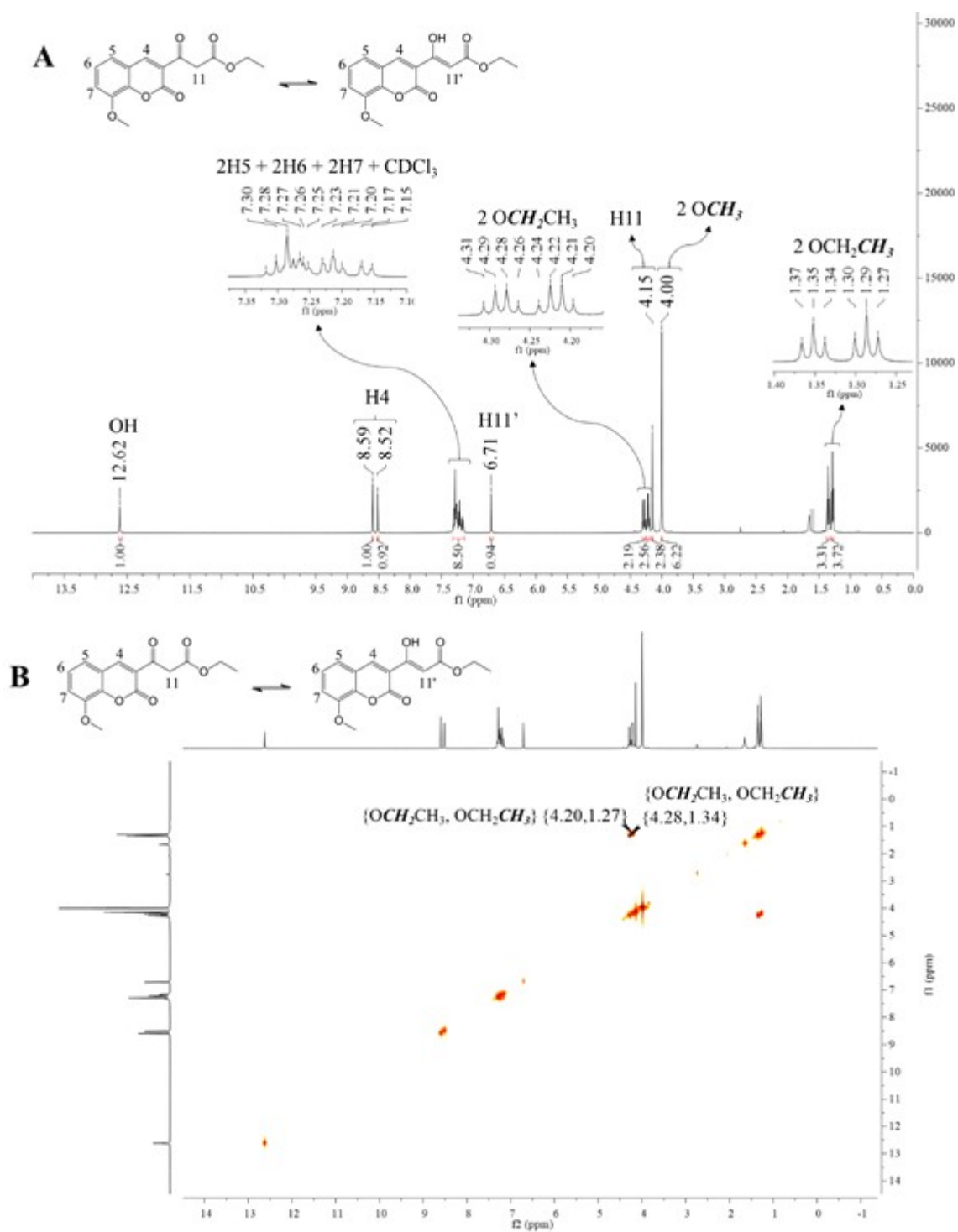


Figure S11: ^1H NMR spectrum of **HL3** (A) and COSY spectrum (B). Solvent: CDCl_3 .

21 #26-39 RT: 0.19-0.27 AV: 7 SM: 7G NL: 3.44E8
T: FTMS + p ESI Full ms [60.0000-800.0000]

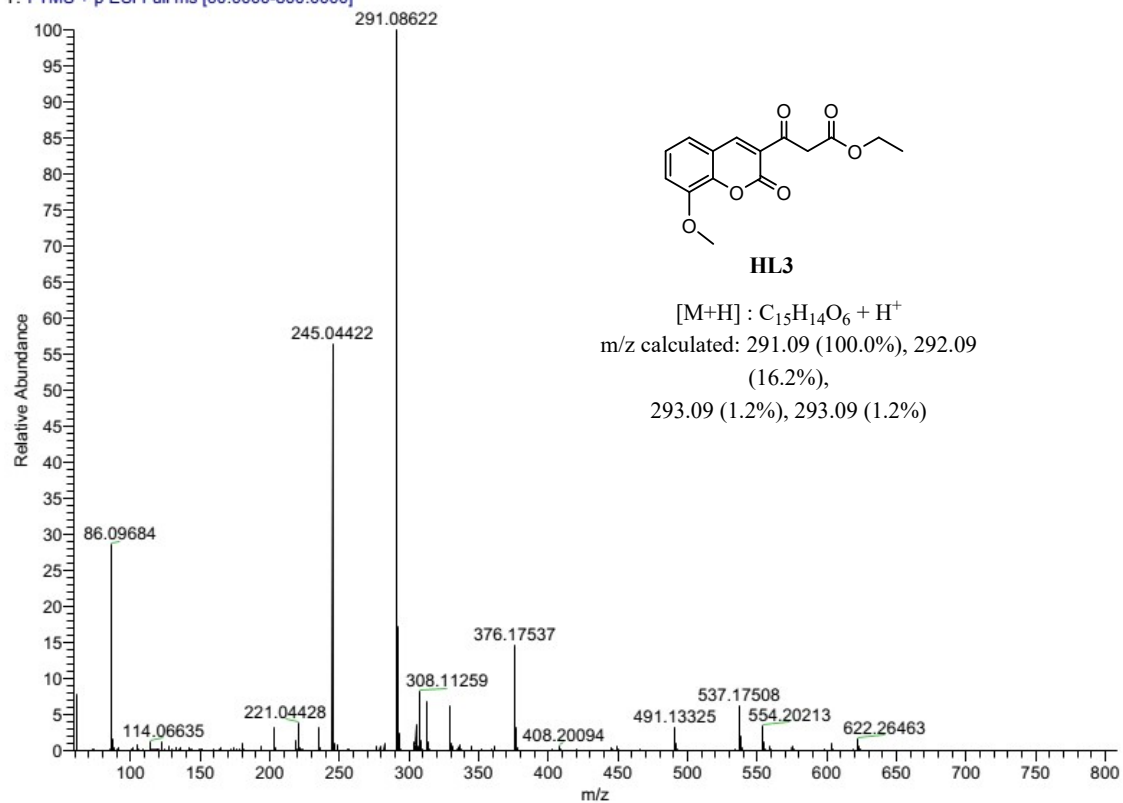


Figure S12: MS/ESI mass spectrum of **HL3**.

IV. *cis*-bis(2,2'-bipyridyl)-[3-(6-(methyl)-2-oxo-2*H*-chromen-3-yl)-3-oxoethylpropanoate]-ruthenium (II) hexafluorophosphate (1**)**

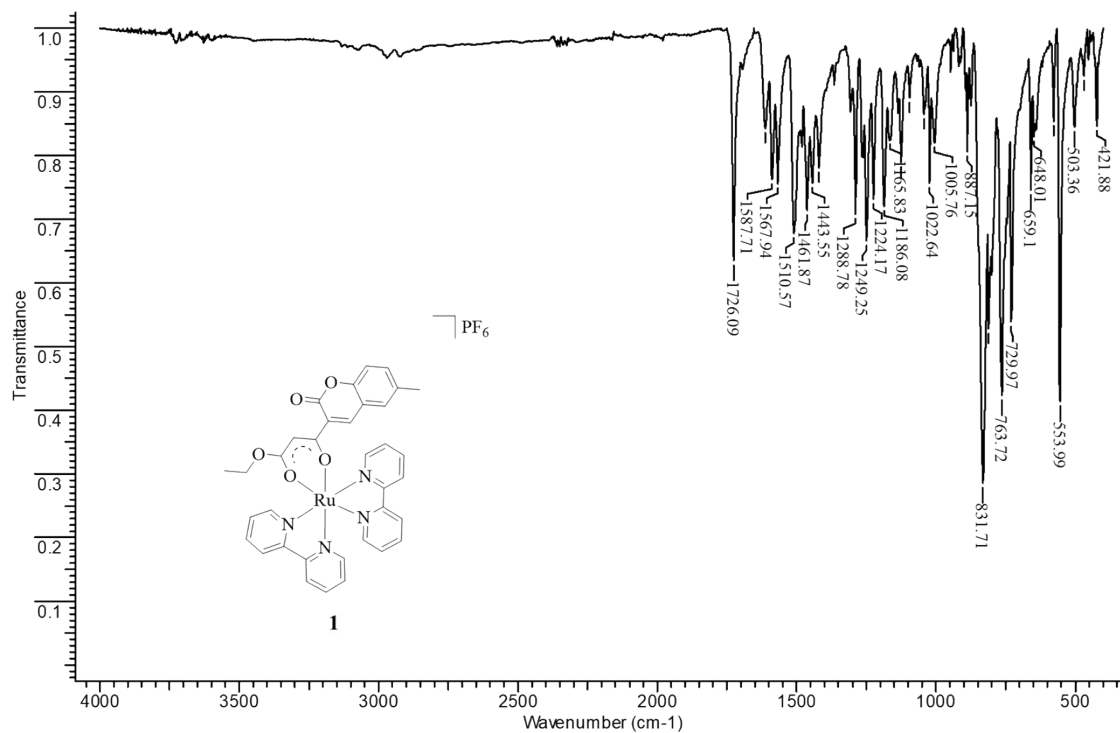


Figure S13: IR spectrum of **1**.

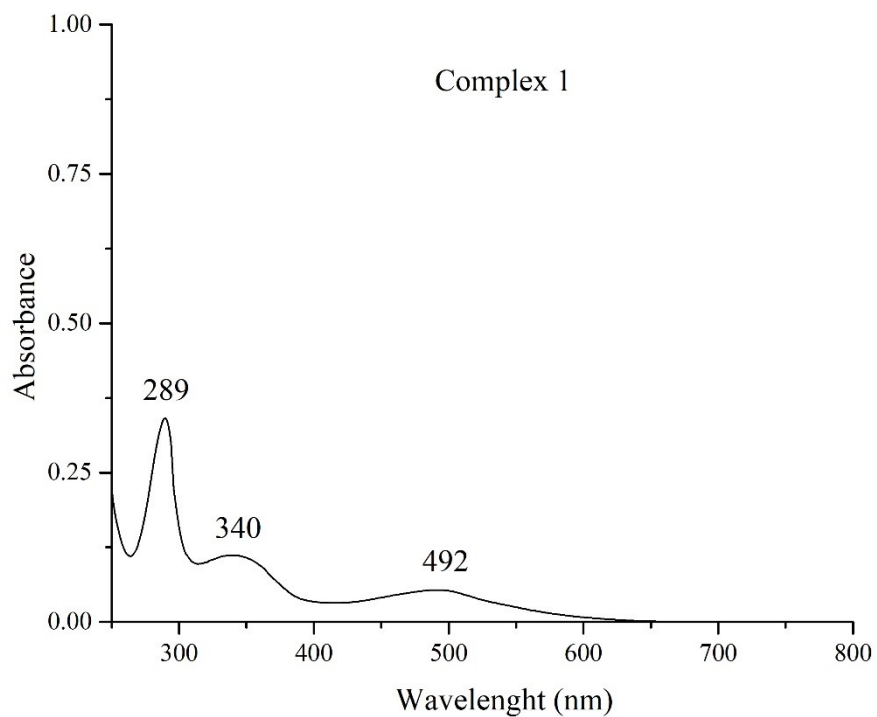


Figure S14: UV-Vis spectrum of **1** in phosphate buffer at 9×10^{-6} M.

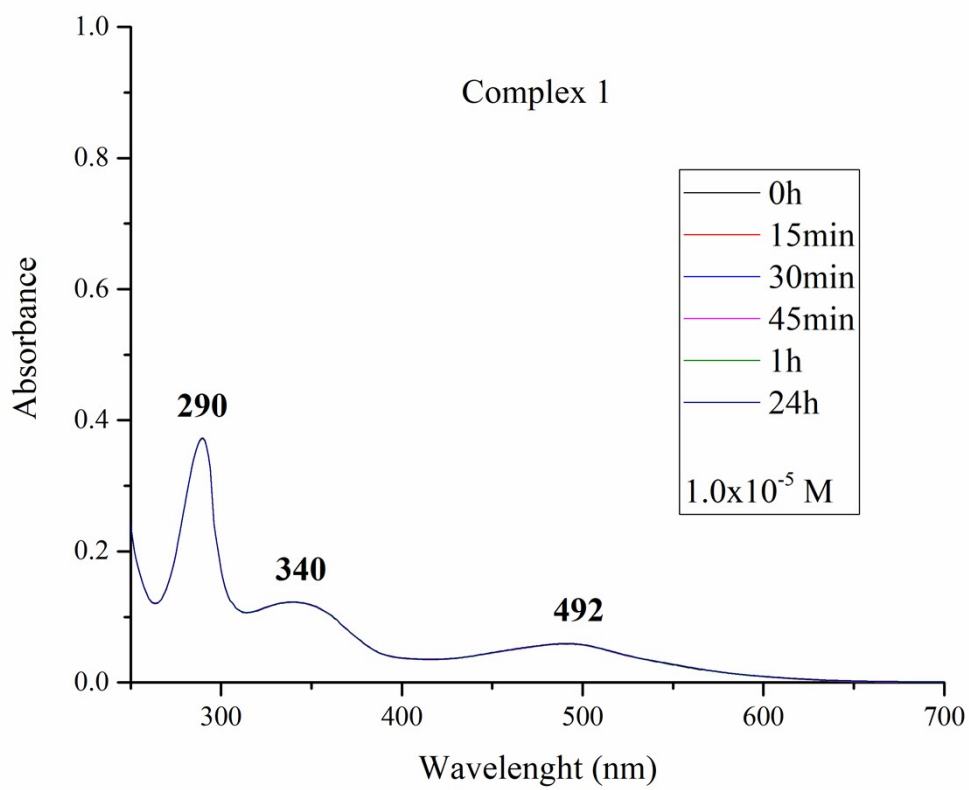


Figure S15: UV-Vis spectrum of **1** recorded over time within 24h, in phosphate buffer at 1.0×10^{-5} M.

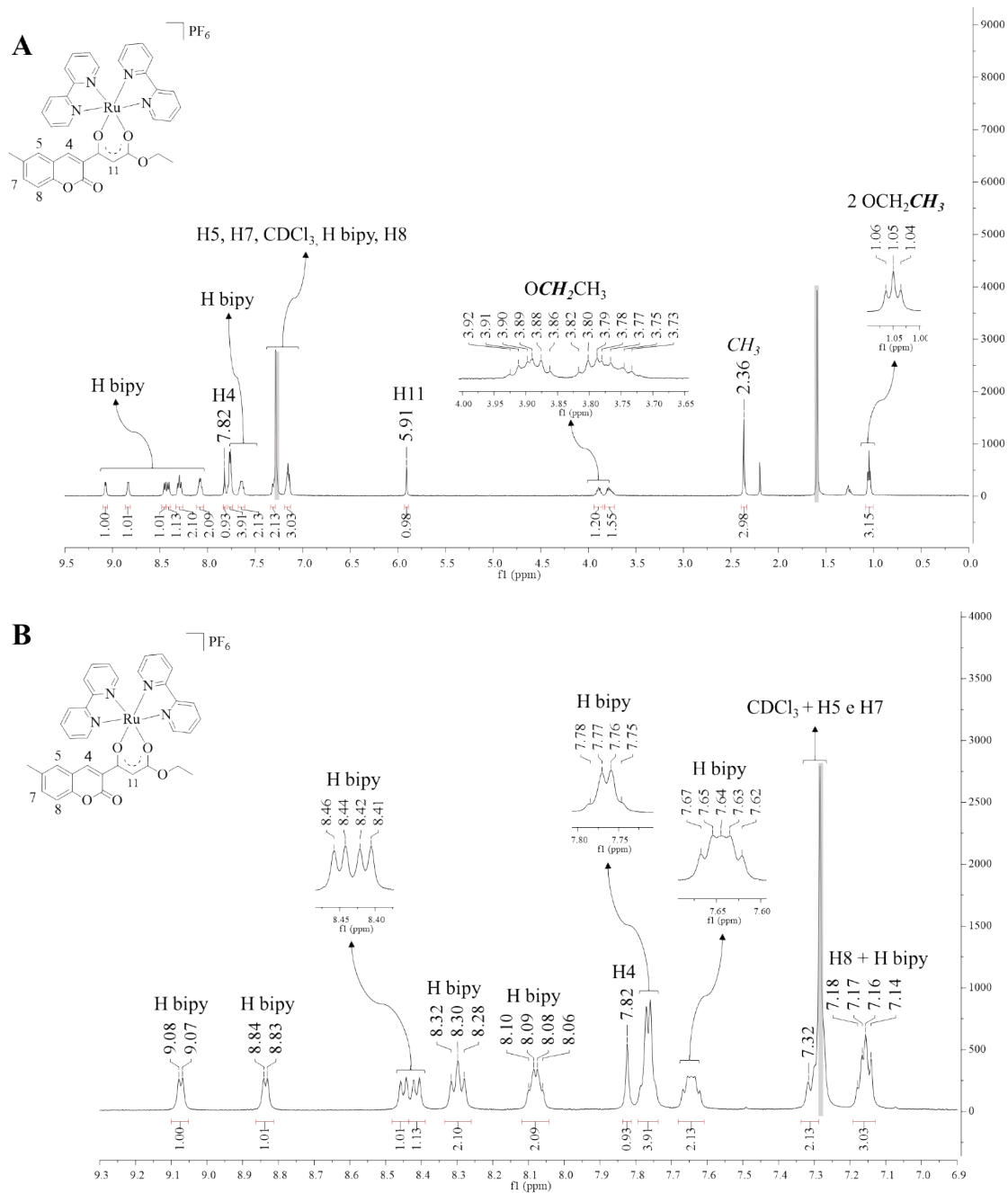


Figure S16: ^1H NMR spectra of **1** (A). Expanded region from 6.9 to 9.3 ppm (B). Solvent: CDCl_3 .

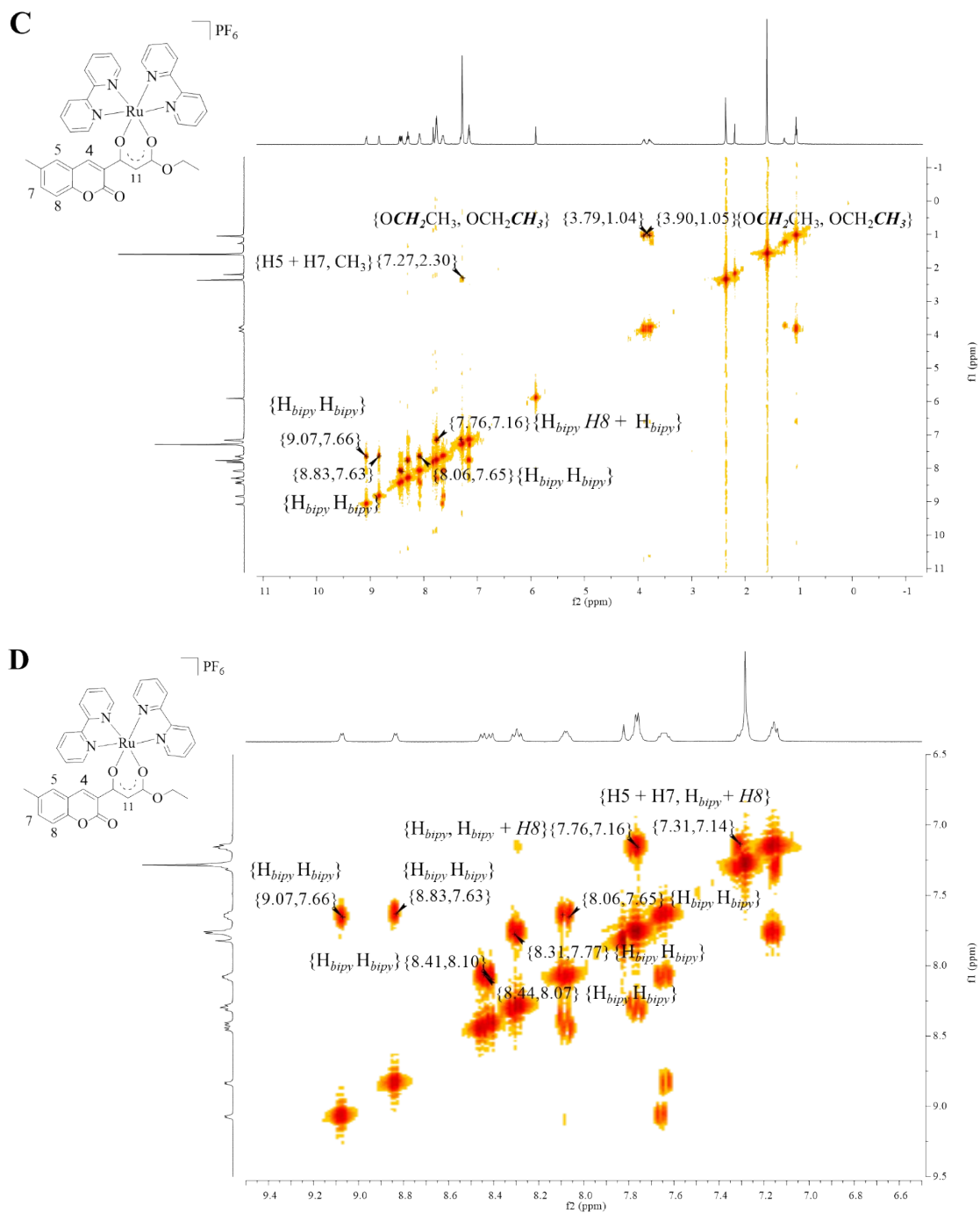


Figure S17: COSY spectra of **1** (C). Expanded region from 6.5 to 9.5 ppm (D). Solvent: CDCl_3 .

V. *cis*-bis(2,2'-bipyridyl)-[3-(7-(diethylamino)-2-oxo-2*H*-chromen-3-yl)-3-oxoethylpropanoate] ruthenium (II) hexafluorophosphate (**2**)

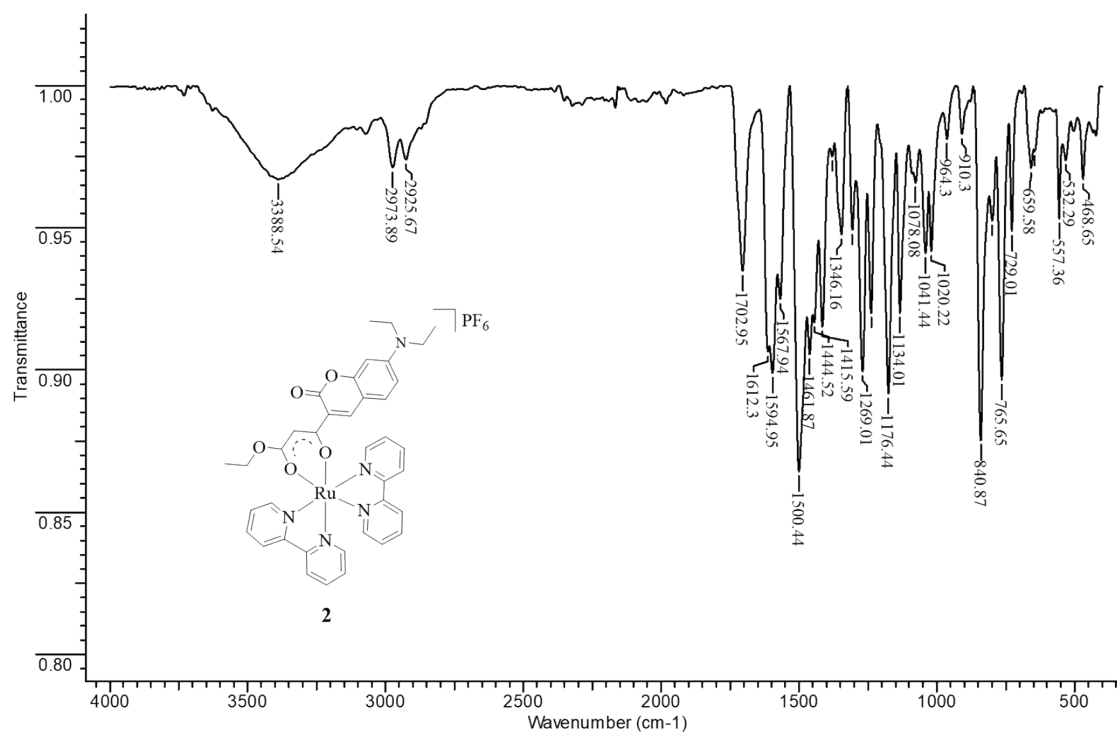


Figure S18: IR spectrum of **2**.

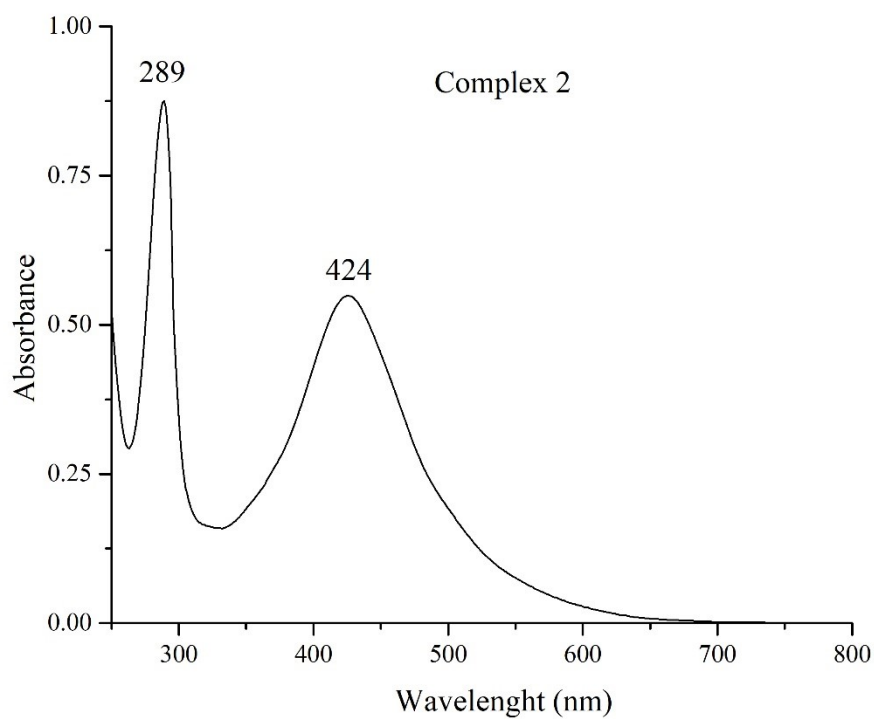


Figure S19: UV-Vis spectrum of the complex **2** in phosphate buffer at 2×10^{-5} M.

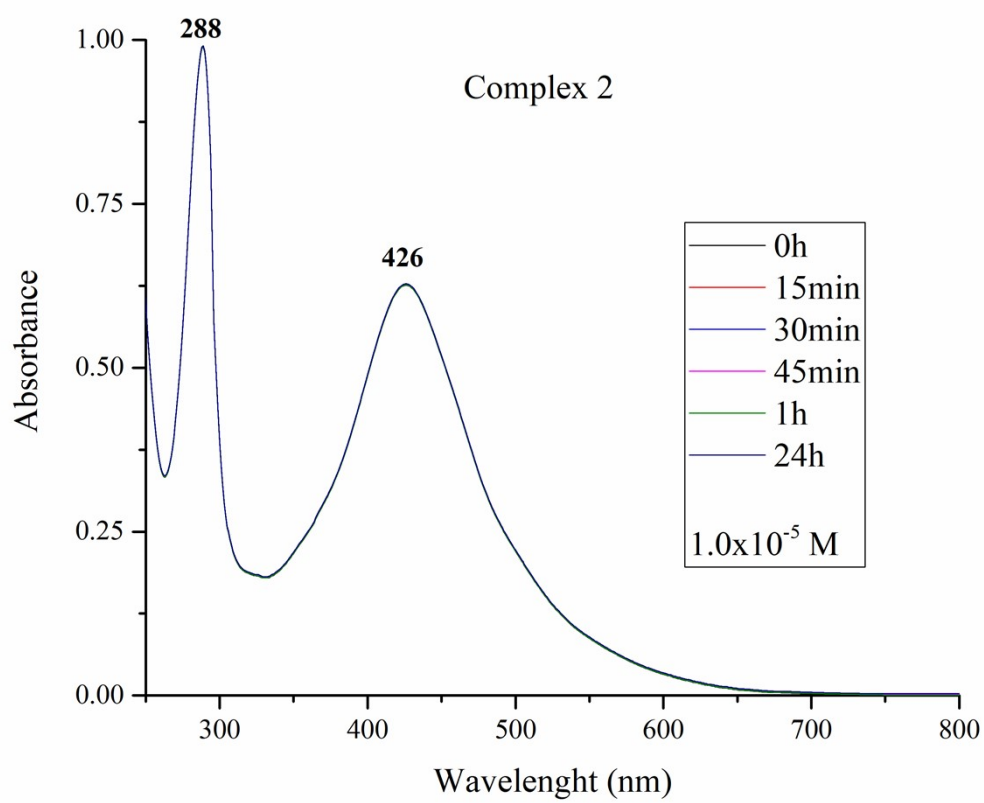


Figure S20: UV-Vis spectrum of **2** recorded over time within 24h, in phosphate buffer at $1.0 \times 10^{-5} \text{ M}$.

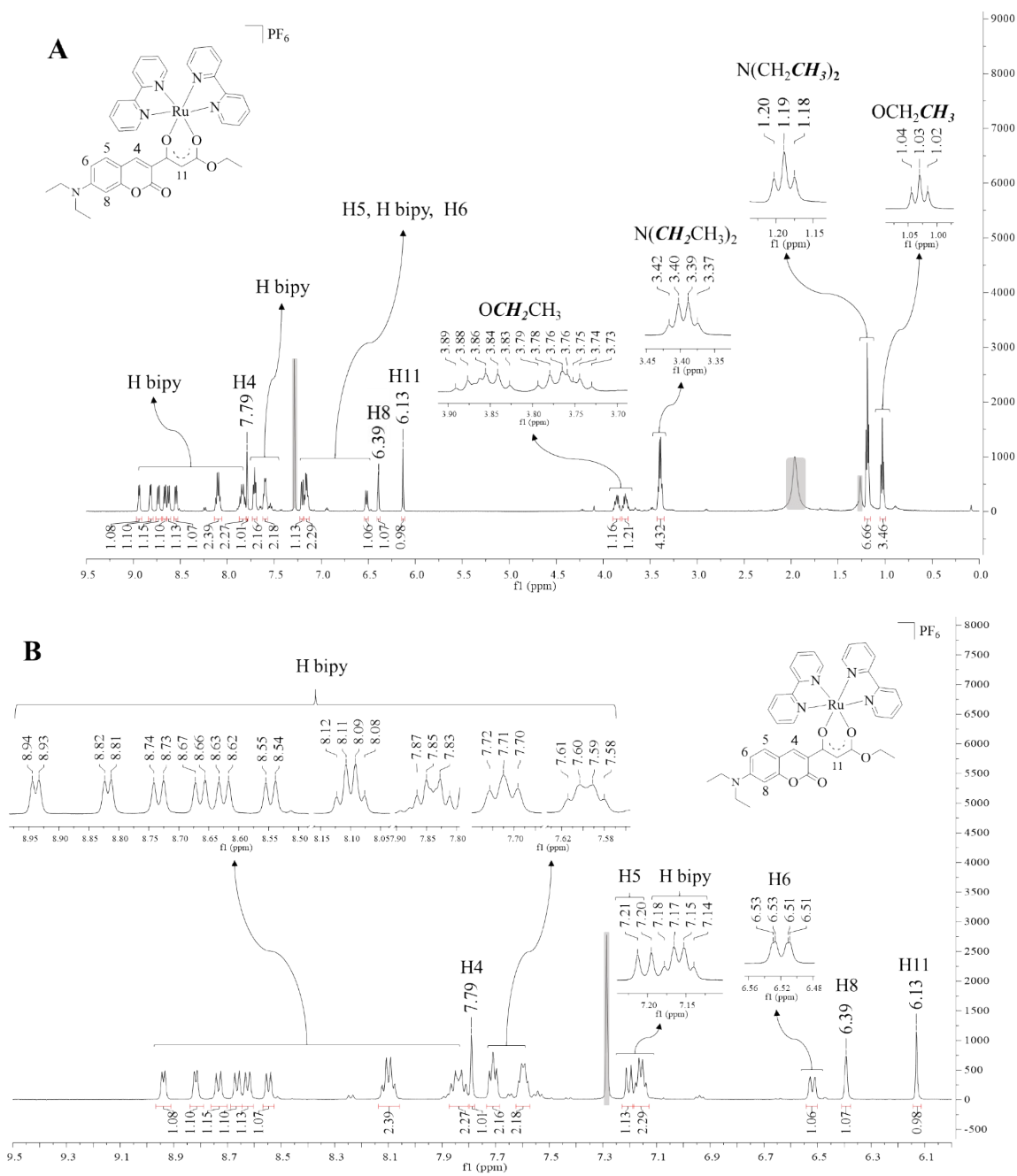


Figure S21: ^1H NMR spectra of **1** (A). Expanded region from 6.0 to 9.5 ppm (B). Solvent: CDCl_3

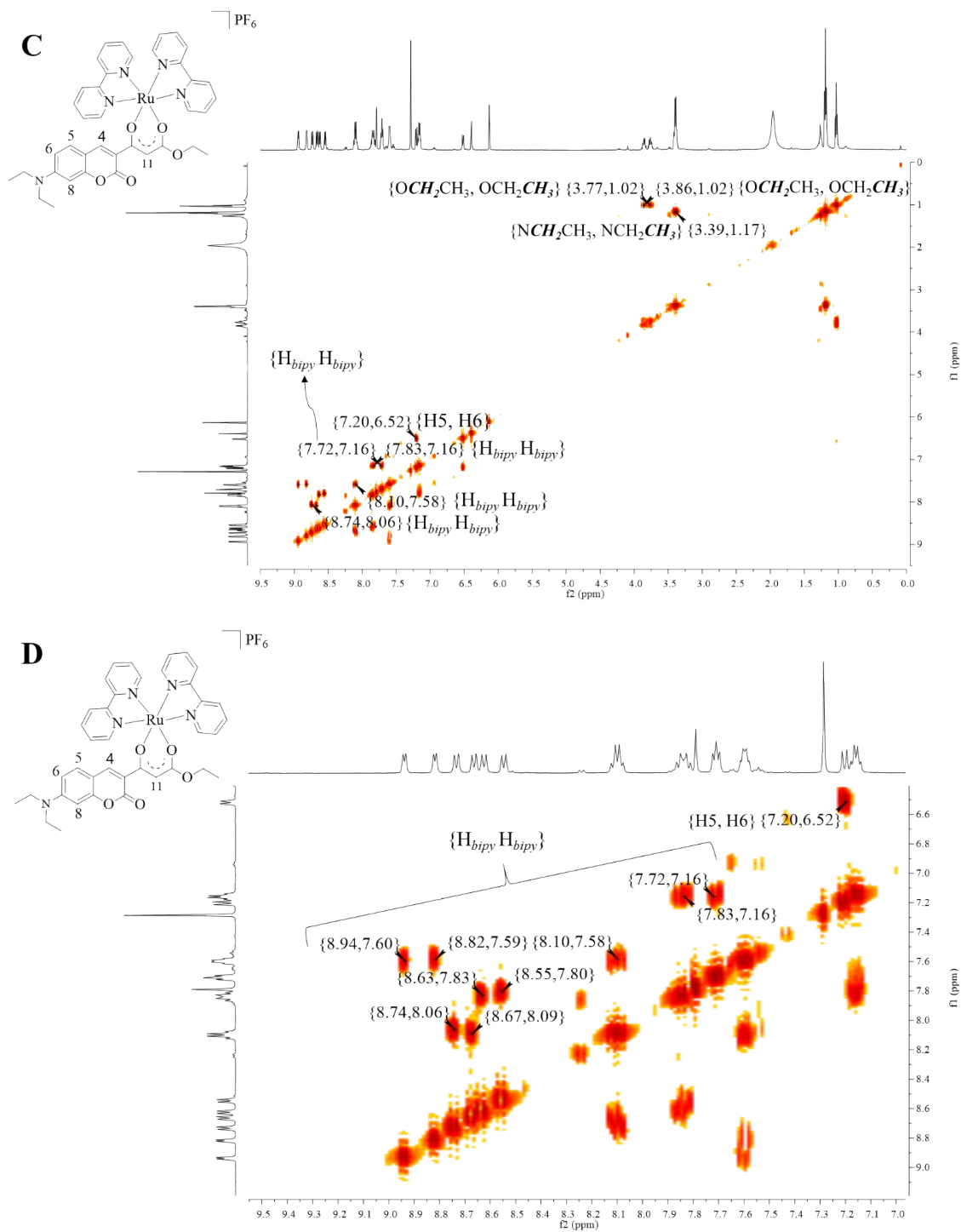


Figure S22: COSY spectra of **2** (C). Expanded region from 6.5 to 9.5 ppm (D). Solvent: $CDCl_3$.

VI. *cis*-bis(2,2'-bipyridyl)-[3-(8-(methoxy)-2-oxo-2*H*-chromen-3-yl)-3-oxoethylpropanoate] ruthenium (II) hexafluorophosphate (**3**)

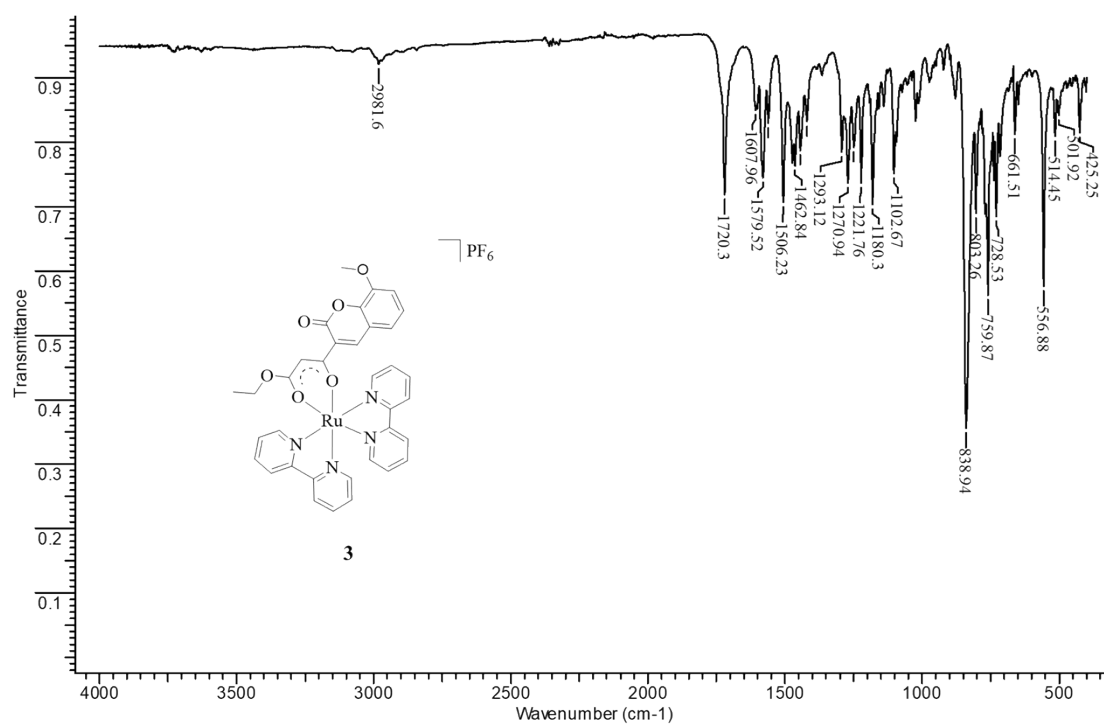


Figure S23: IR spectrum of **3**.

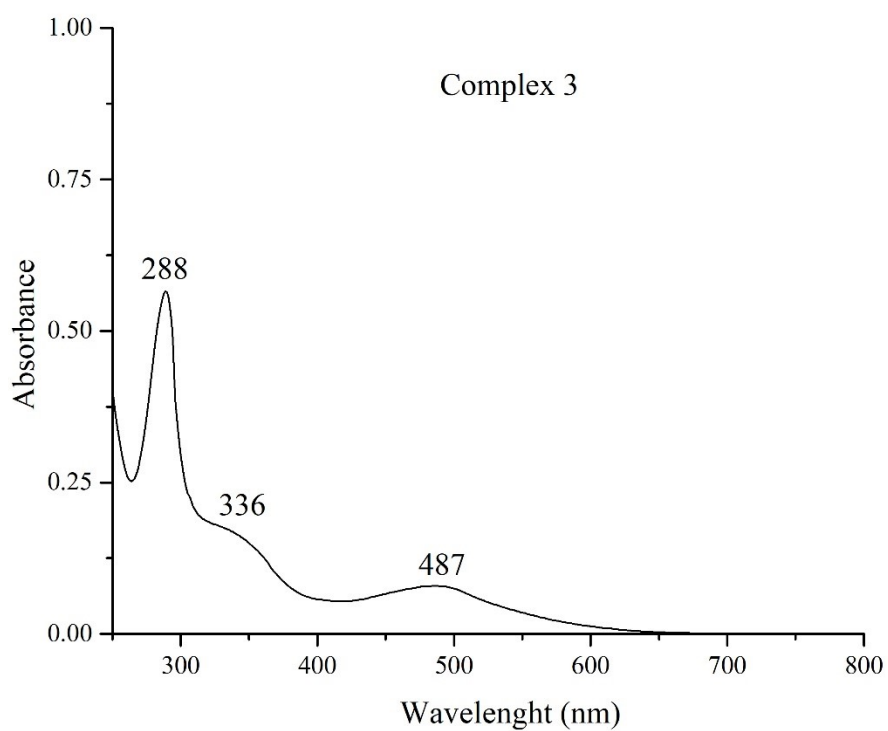


Figure S24: UV-Vis spectrum of **3** in phosphate buffer at 1.5×10^{-5} M.

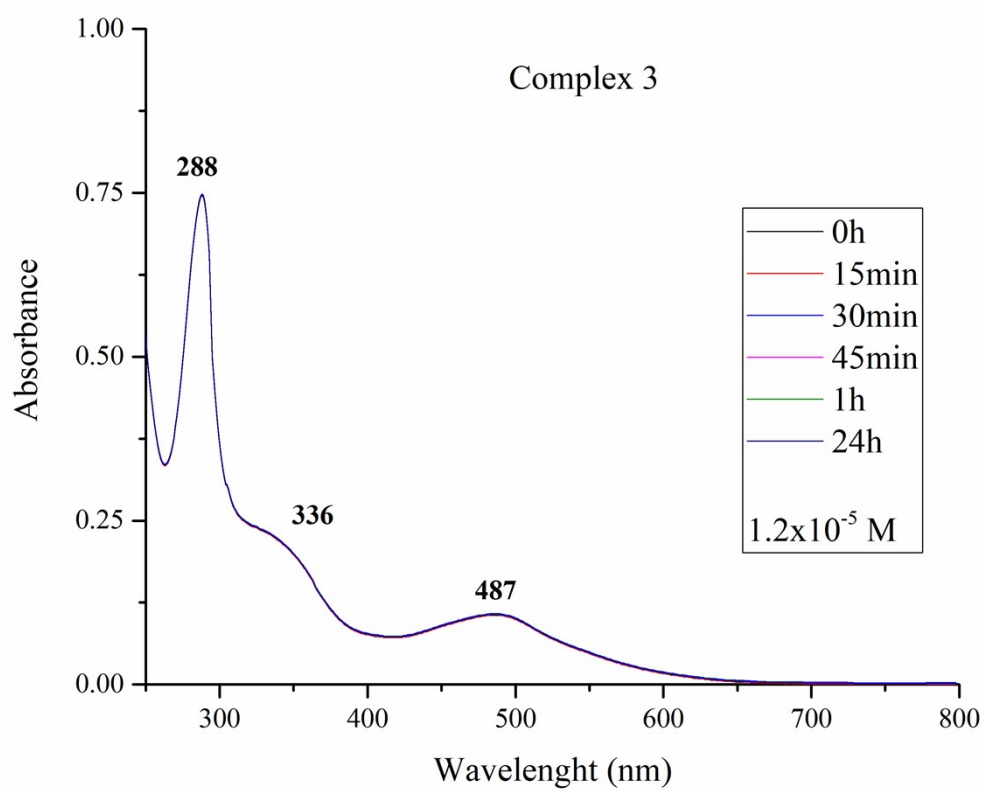


Figure S25: UV-Vis spectrum of **3** recorded over time within 24h, in phosphate buffer at 1.2×10^{-5} M.

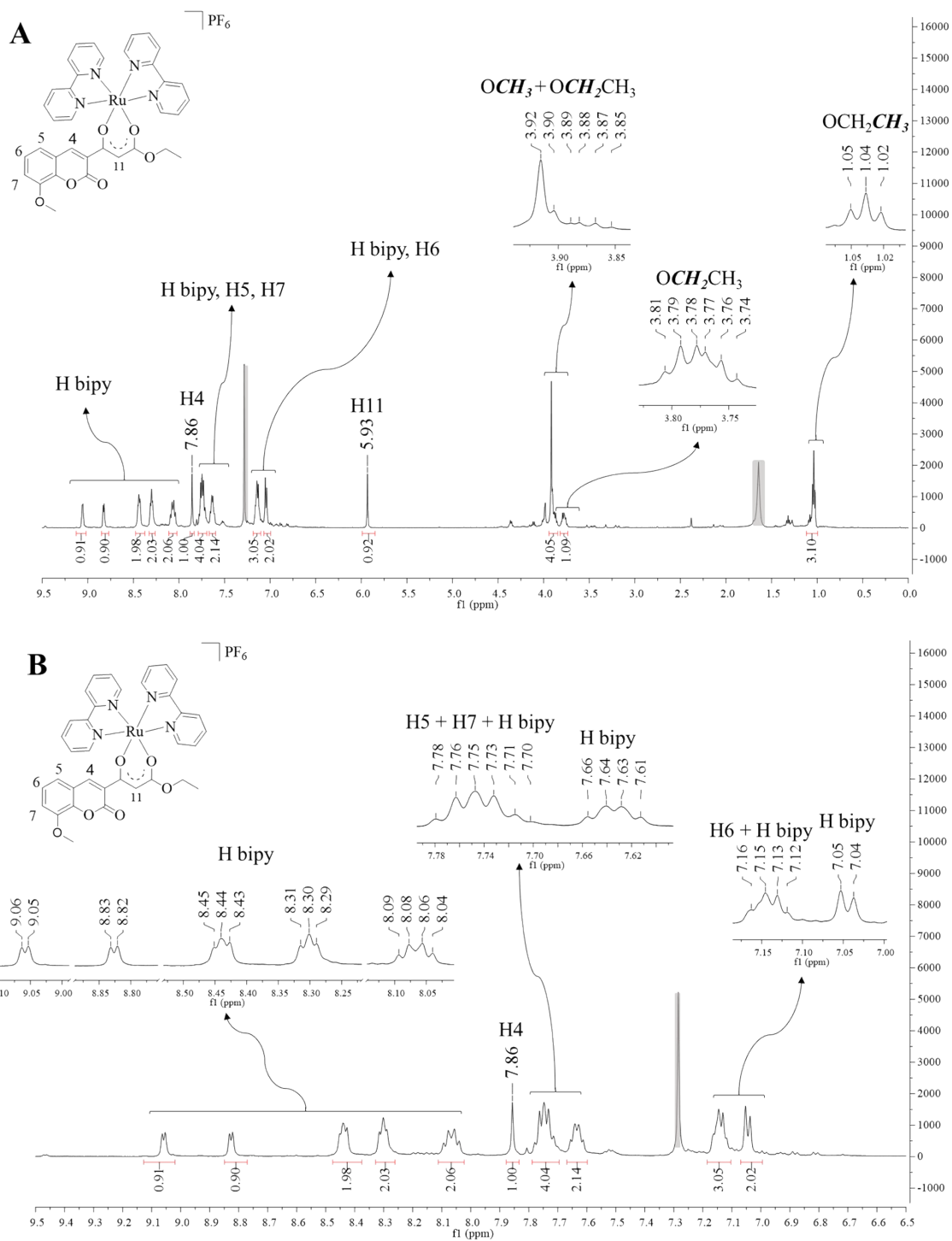


Figure S26: ^1H NMR spectra of **3** (A). Expanded region from 6.5 to 9.5 ppm (B). Solvent: CDCl_3

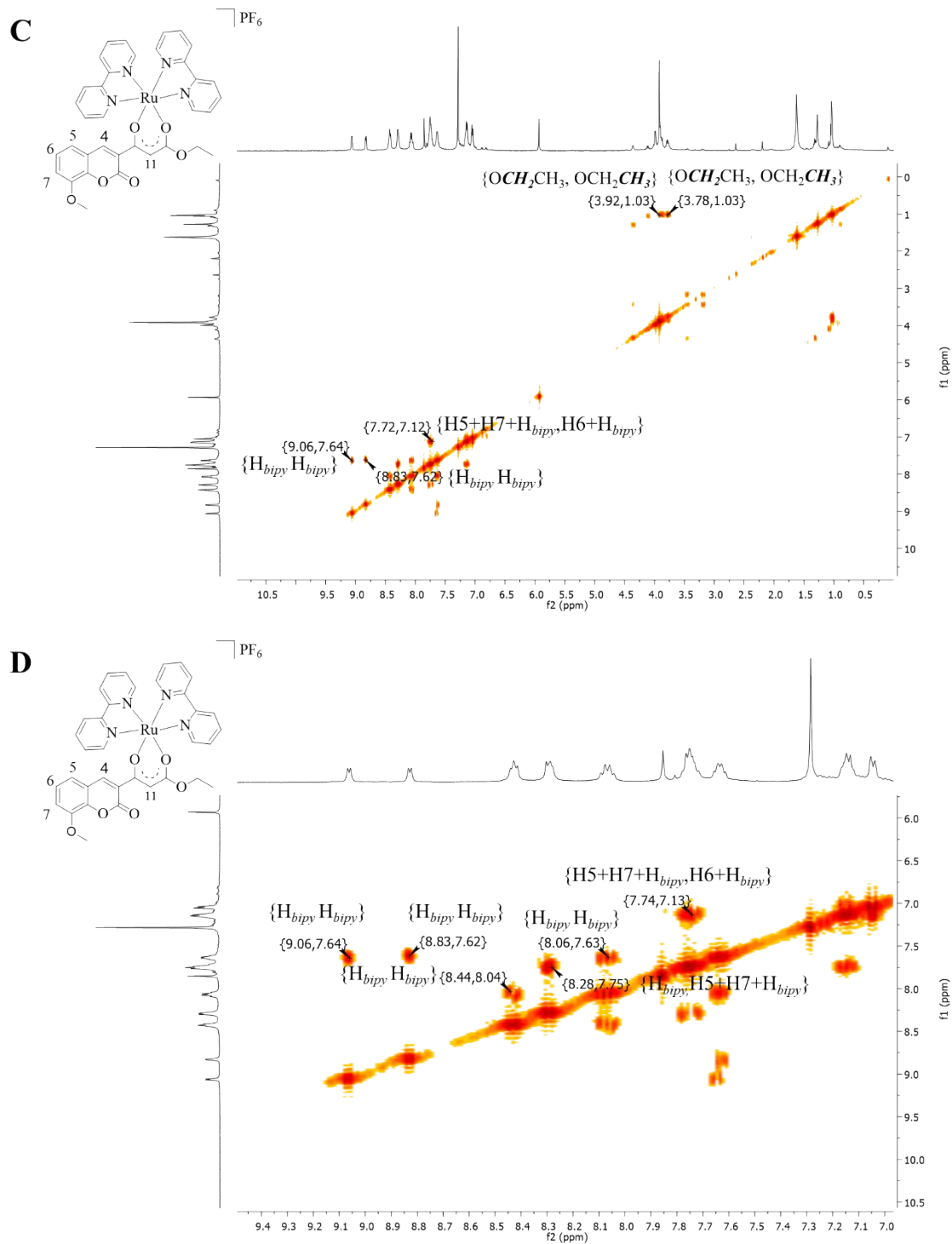


Figure S27: COSY spectra of **3** (**C**). Expanded region from 7.0 to 9.5 ppm (**D**). Solvent: CDCl₃.

VII. X-Ray diffraction analysis

Table S1: Summary of crystal structure, data collection and refinement for complex **2**.

Chemical formula	C ₃₈ H ₃₆ F ₆ N ₅ O ₅ PRu
Formula weigh	888.76
Crystal system	monoclinic
a/Å	28.3360(12)
b/Å	10.5670(3)
c/Å	26.8770(9)
α/°	90
β/°	110.5490(10)
γ/°	90
Unit cell volume (Å³)	7535.6(5)
T (K)	273(2)
Space group	C 2/c
Formula units per cell, Z	8
Radiation type	MoKα
Absorption coefficient (mm⁻¹)	0.539
Reflections measured	41132
Independents reflections	6637
<i>R</i>_{int}	0.0521
Final <i>R</i>_I values (<i>I</i>>2σ(<i>I</i>))	0.0361
Final <i>wR</i>(<i>F</i>²) values (<i>I</i>>2σ(<i>I</i>))	0.0773
Final <i>R</i>_I values (all data)	0.0522
Final <i>wR</i>(<i>F</i>²) values (all data)	0.0849
Goodness of (GOF) fit F²	1.046
CCDC deposition	2076893

Table S2: Hydrogen-bond geometry (Å, °) for **2**.

<i>D</i> —H··· <i>A</i>	<i>D</i> —H	<i>H</i> ··· <i>A</i>	<i>D</i> ··· <i>A</i>	<i>D</i> —H··· <i>A</i>
C35—H35...F5i	0.93	2.99	3.306(4)	101.4
C38—H38...O4	0.93	2.61	3.149(4)	117.8
C28—H28...F4	0.93	2.51	3.140(4)	125.5

Symmetry code: (i) -x+1, -y+2, -z+1.

VIII. Biological studies

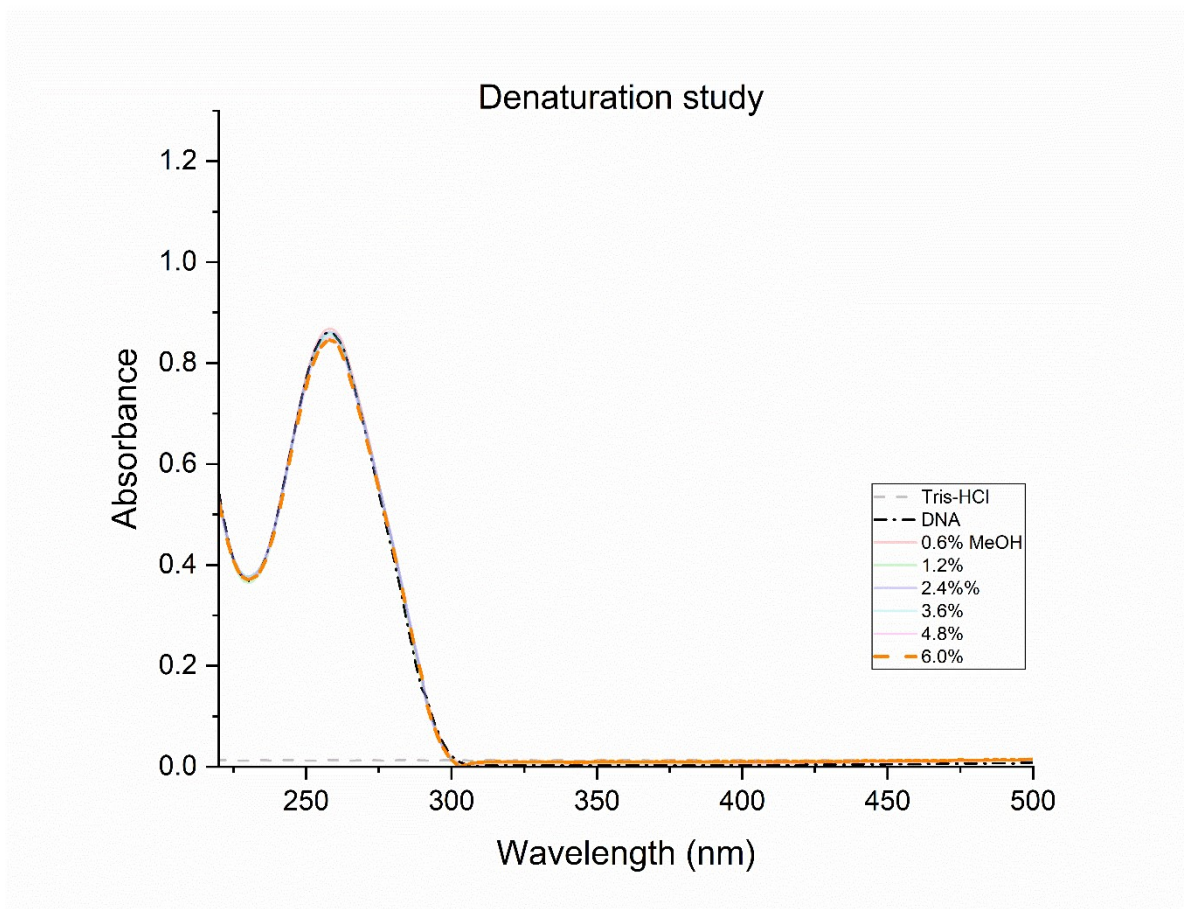


Figure S28. Absorption spectra of the ct-DNA in the presence of methanol in different proportions. Methanol = 0, 0.6, 1.2, 2.4, 3.6, 4.8 and 6.0%. ct-DNA at a concentration of 120 μ M in Tris-HCl buffer (pH 7.4).

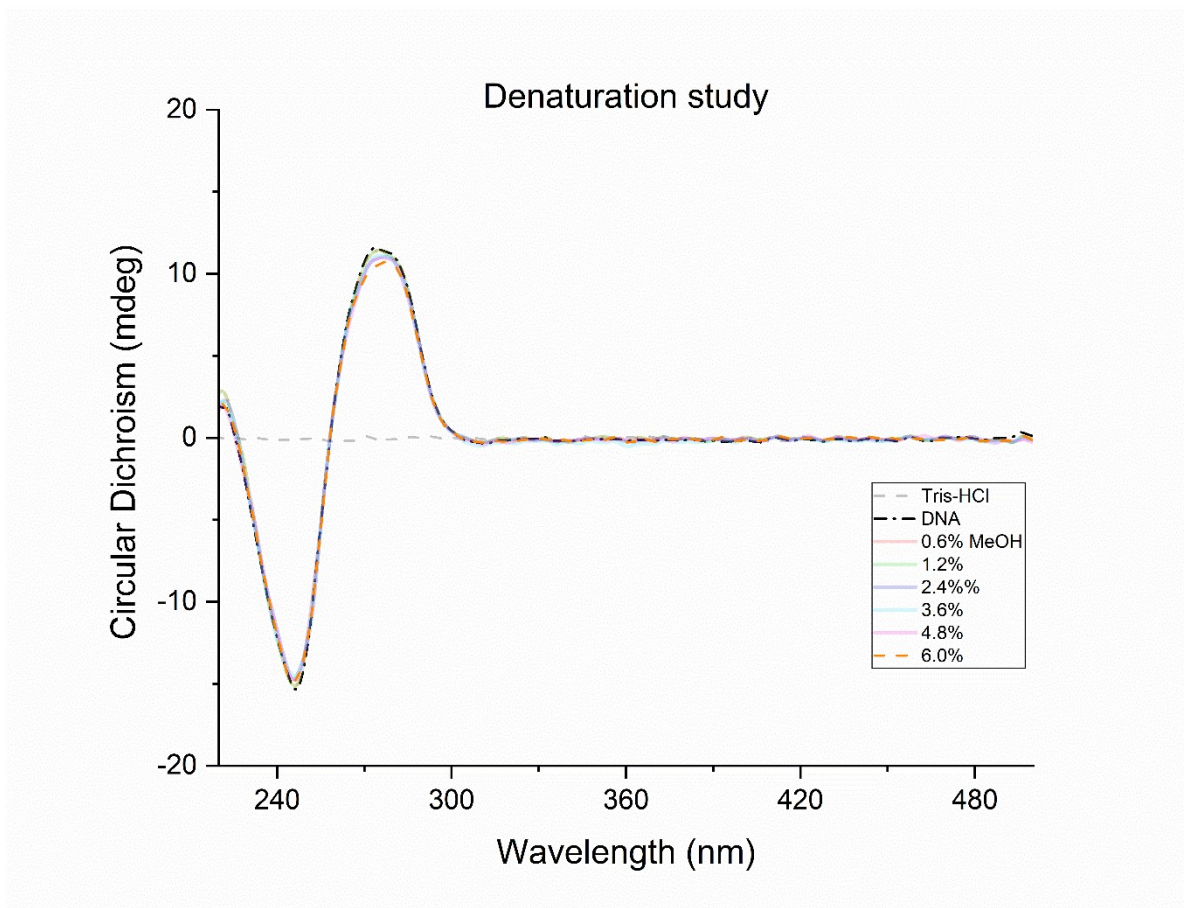


Figure S29. Circular dichroism of DNA at a concentration of 120 μ M in Tris-HCl buffer (pH 7.4), in the presence of increasing methanol proportions (0 to 6 %).

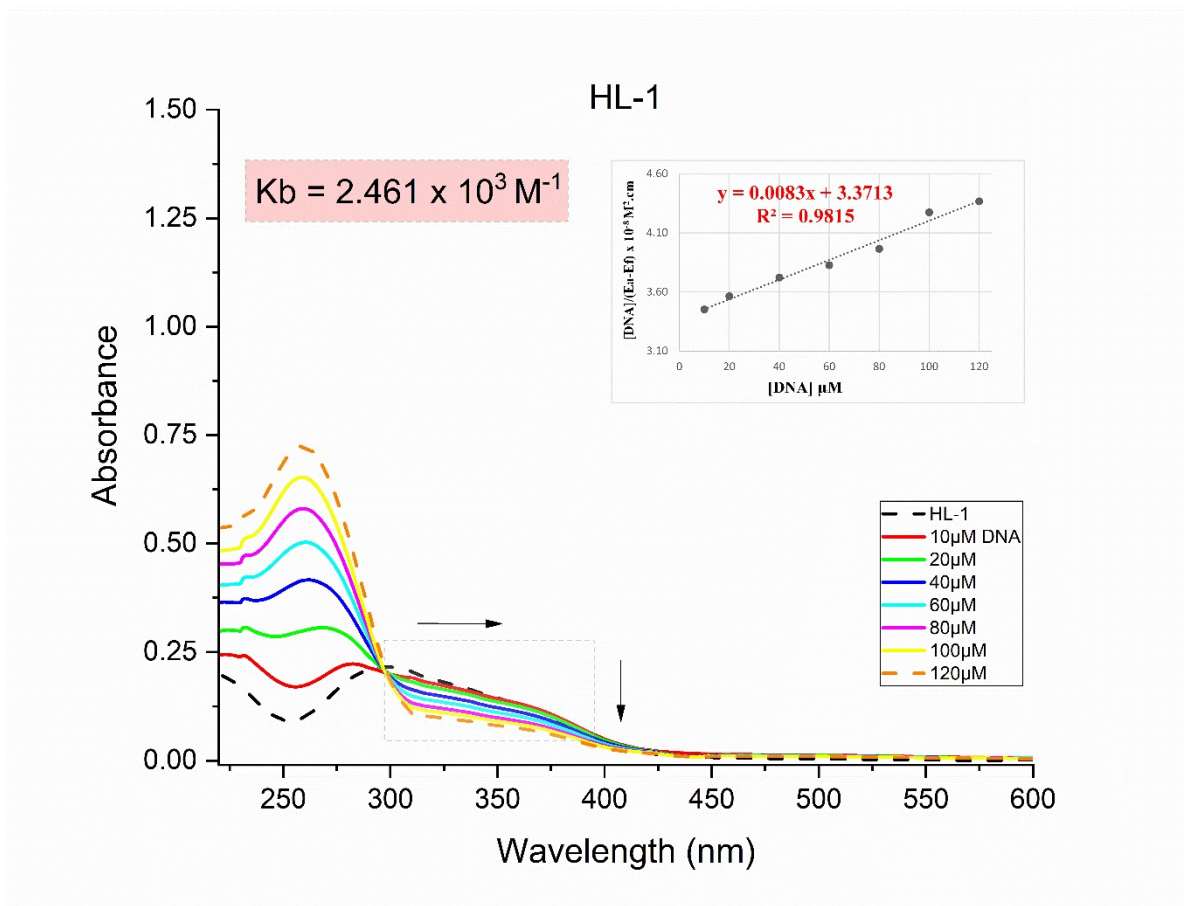


Figure S30. Absorption spectra of **HL1** at 30 μM in Tris-HCl buffer (pH 7.4) in the presence of ct-DNA in different concentrations. ct-DNA = 0, 10, 20, 40, 60, 80, 100 and 120 μM. λ (Kb) = 327 nm.

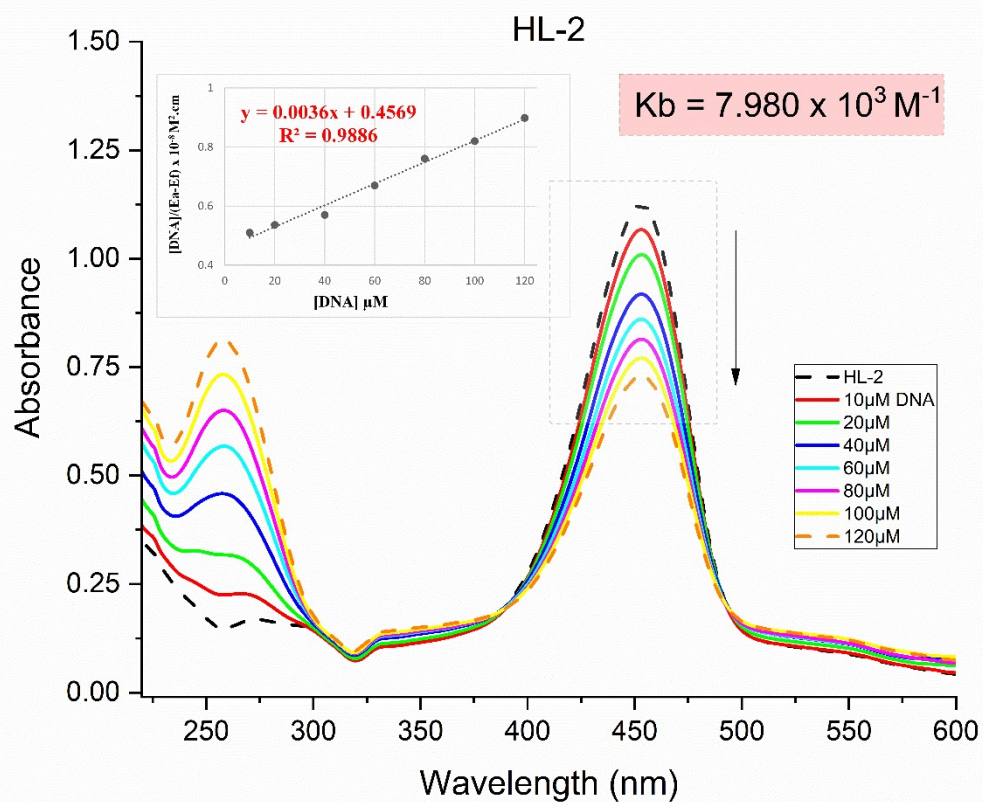


Figure S31. Absorption spectra of **HL2** at 30 μM in Tris-HCl buffer (pH 7.4) in the presence of ct-DNA in different concentrations. ct-DNA = 0, 10, 20, 40, 60, 80, 100 and 120 μM . λ (K_b) = 453 nm.

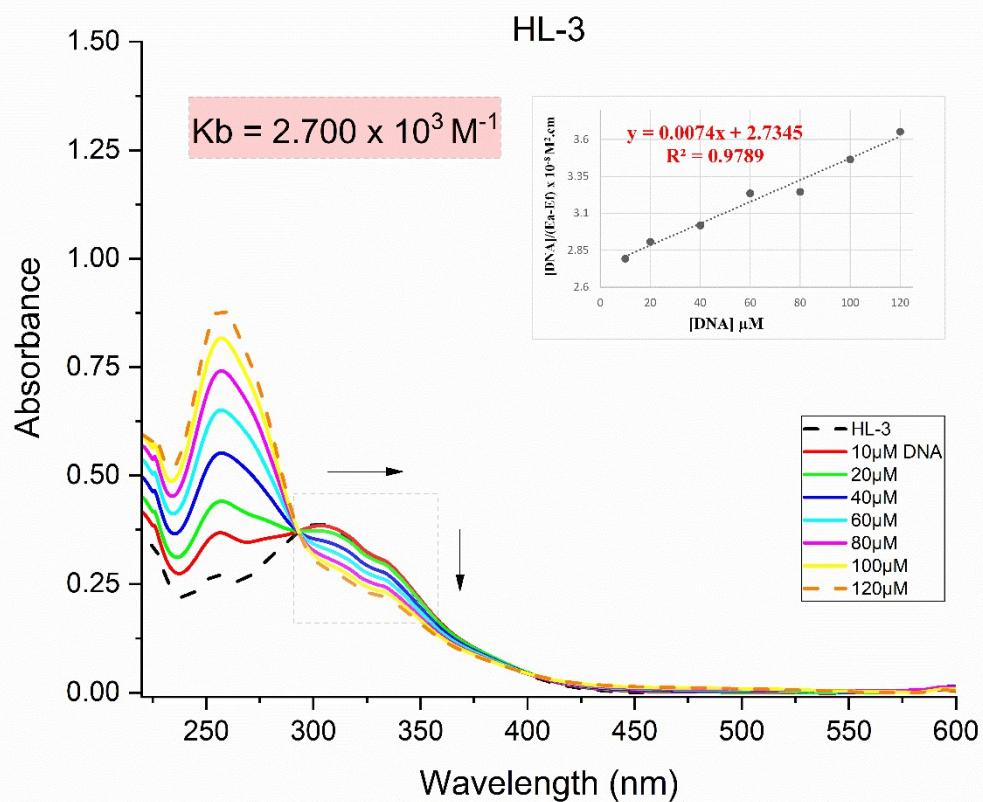


Figure S32. Absorption spectra of **HL3** at 30 μM in Tris-HCl buffer (pH 7.4) in the presence of ct-DNA in different concentrations. ct-DNA = 0, 10, 20, 40, 60, 80, 100 and 120 μM . λ (Kb) = 303 nm.

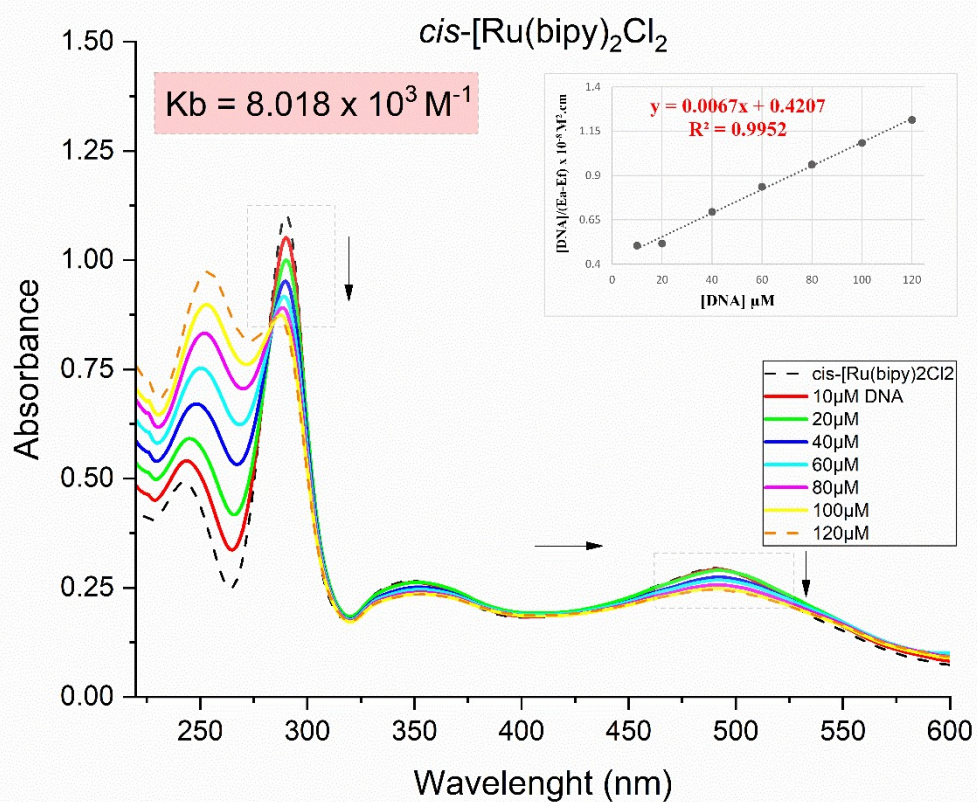


Figure S33. Absorption spectra of *cis*-[Ru(bipy)₂Cl₂] at 30 μM in Tris-HCl buffer (pH 7.4) in the presence of ct-DNA in different concentrations. ct-DNA = 0, 10, 20, 40, 60, 80, 100 and 120 μM. λ (Kb) = 490 nm.

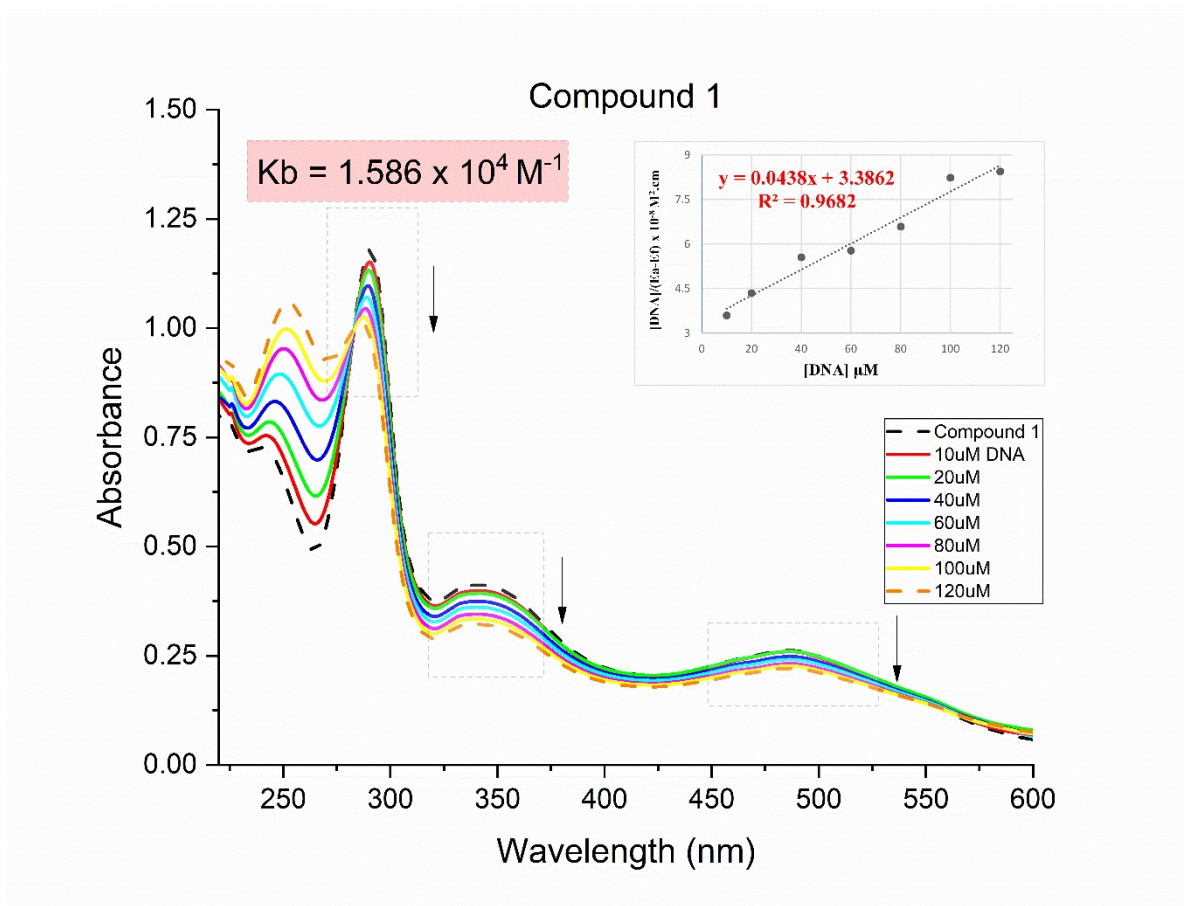


Figure S34. Absorption spectra of complex **1** at 30 μM in Tris-HCl buffer (pH 7.4) in the presence of ct-DNA in different concentrations. CtDNA = 0, 10, 20, 40, 60, 80, 100 and 120 μM . λ (Kb) = 487 nm.

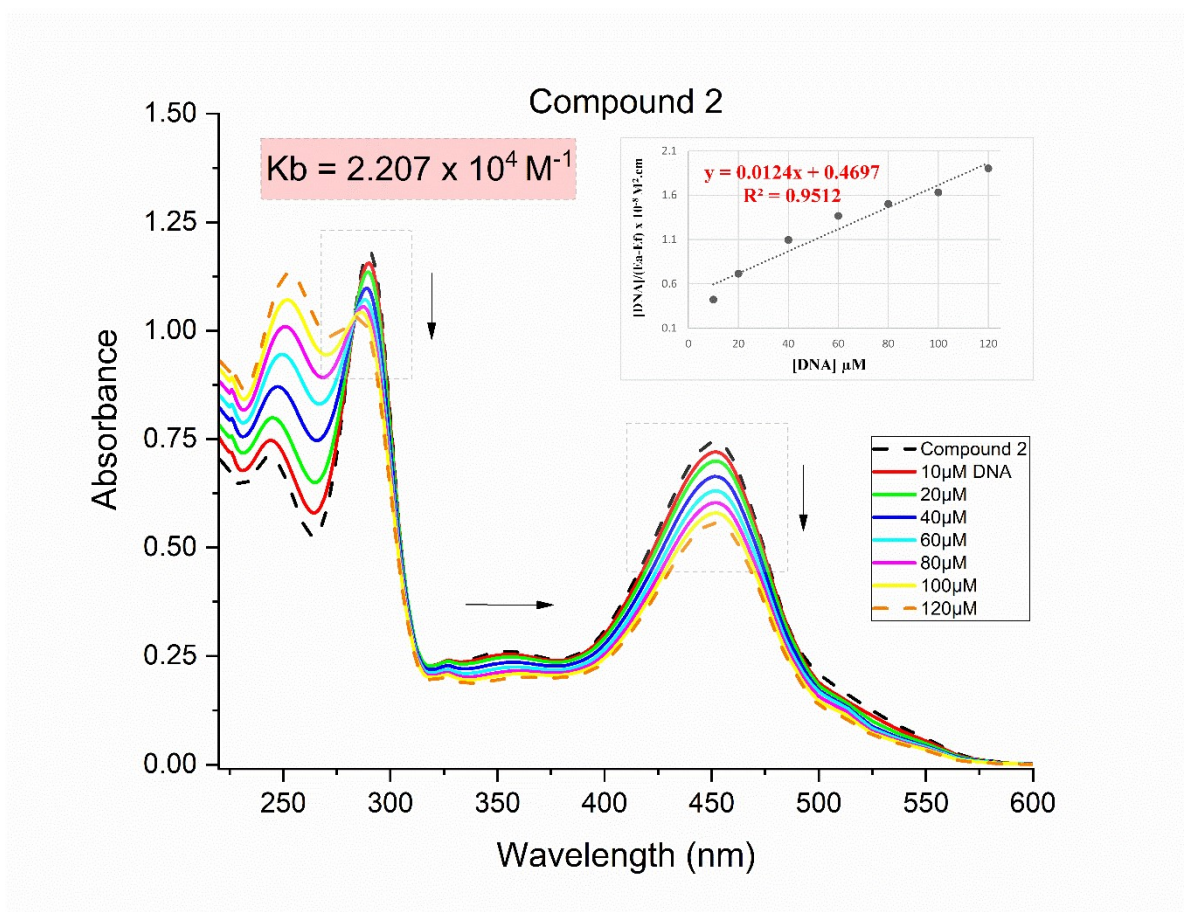


Figure S35. Absorption spectra of complex **2** at 30 μM in Tris-HCl buffer (pH 7.4) in the presence of ct-DNA in different concentrations. CtDNA = 0, 10, 20, 40, 60, 80, 100 and 120 μM . λ (Kb) = 450 nm.

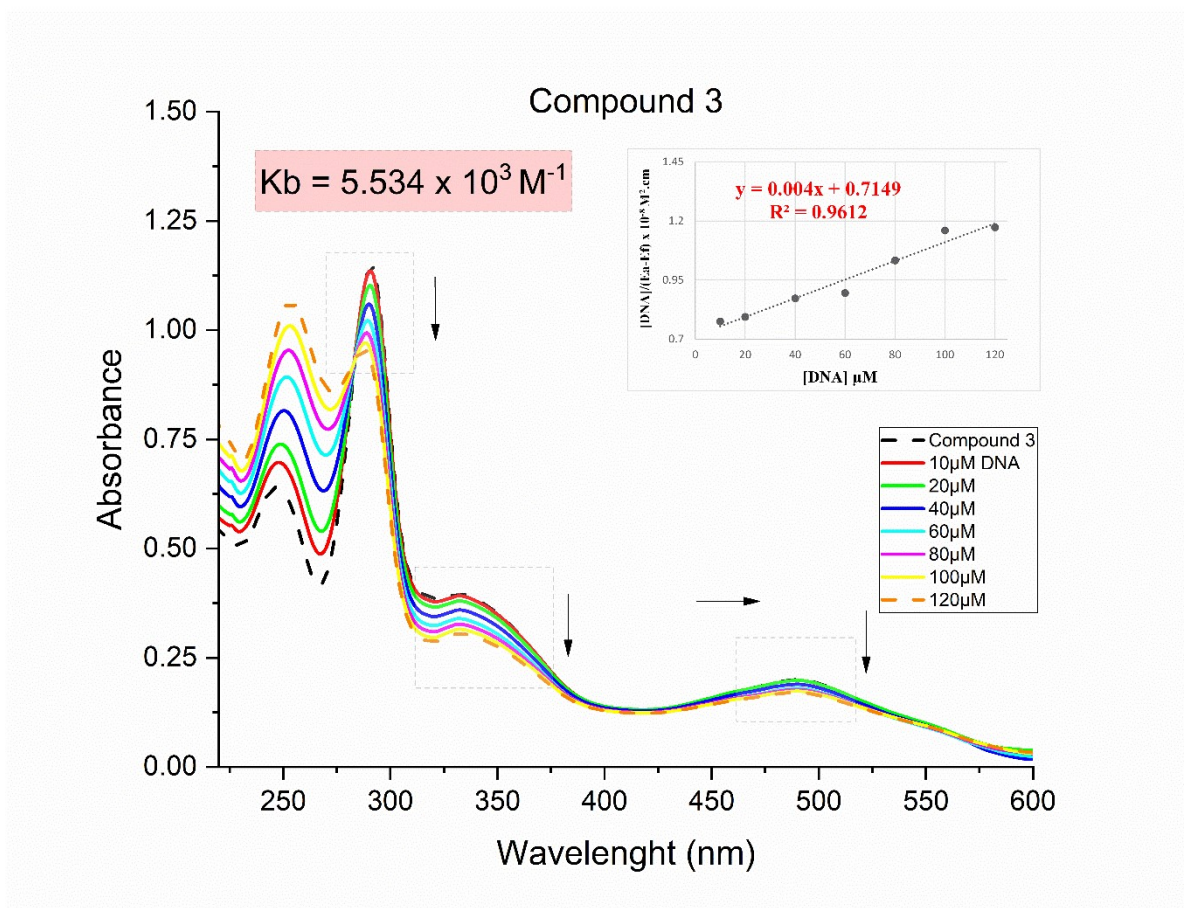


Figure S36. Absorption spectra of complex **3** at 30 μM in Tris-HCl buffer (pH 7.4) in the presence of ct-DNA in different concentrations. CtDNA = 0, 10, 20, 40, 60, 80, 100 and 120 μM . λ (Kb) = 490 nm.

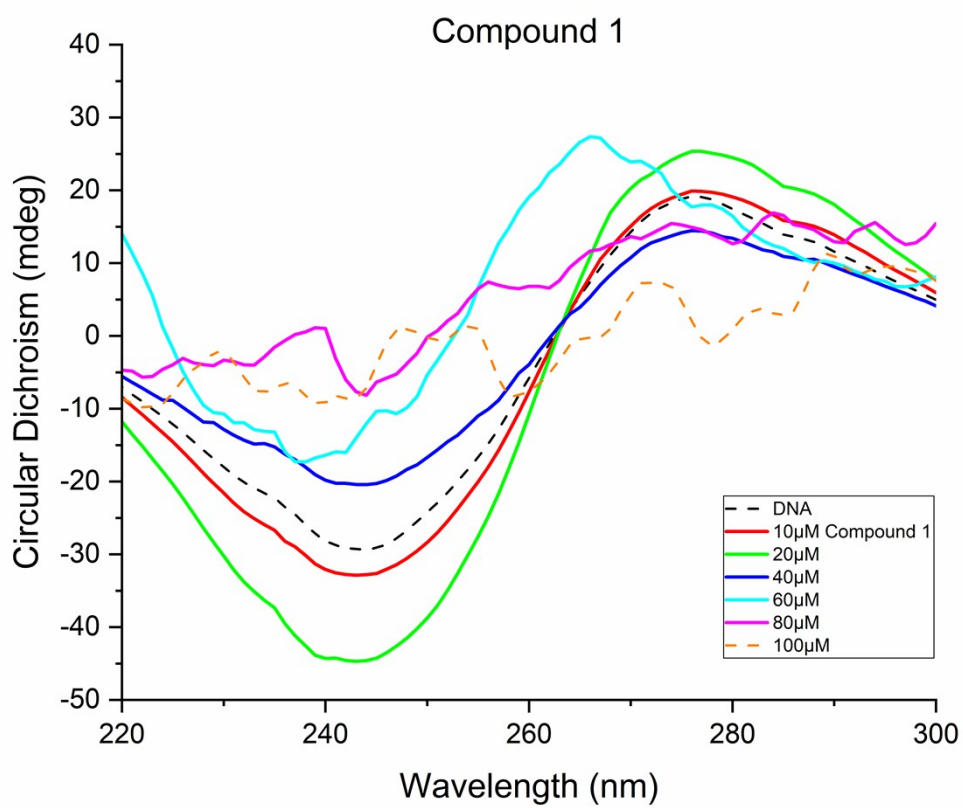


Figure S37. Circular dichroism of DNA at a concentration of 300 μM in Tris-HCl buffer (pH 7.4), in the presence of increasing concentrations of complex **1** (10, 20, 40, 60, 80 and 100 μM).

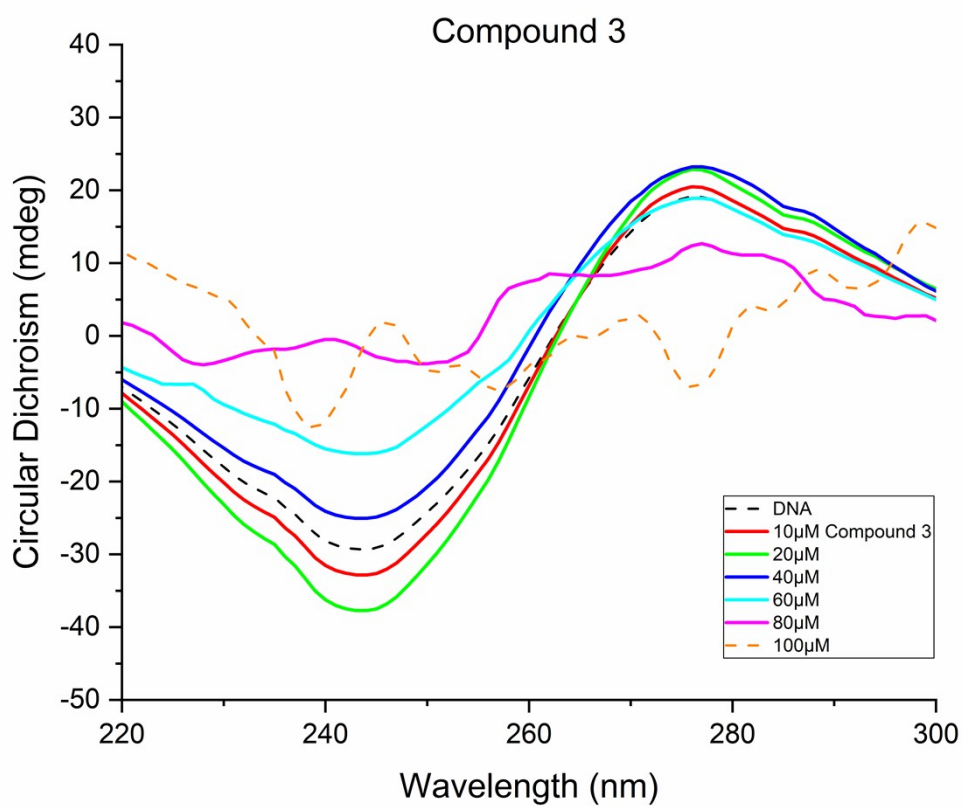


Figure S38. Circular dichroism of DNA at a concentration of 300 μM in Tris-HCl buffer (pH 7.4), in the presence of increasing concentrations of complex **3** (10, 20, 40, 60, 80 and 100 μM).

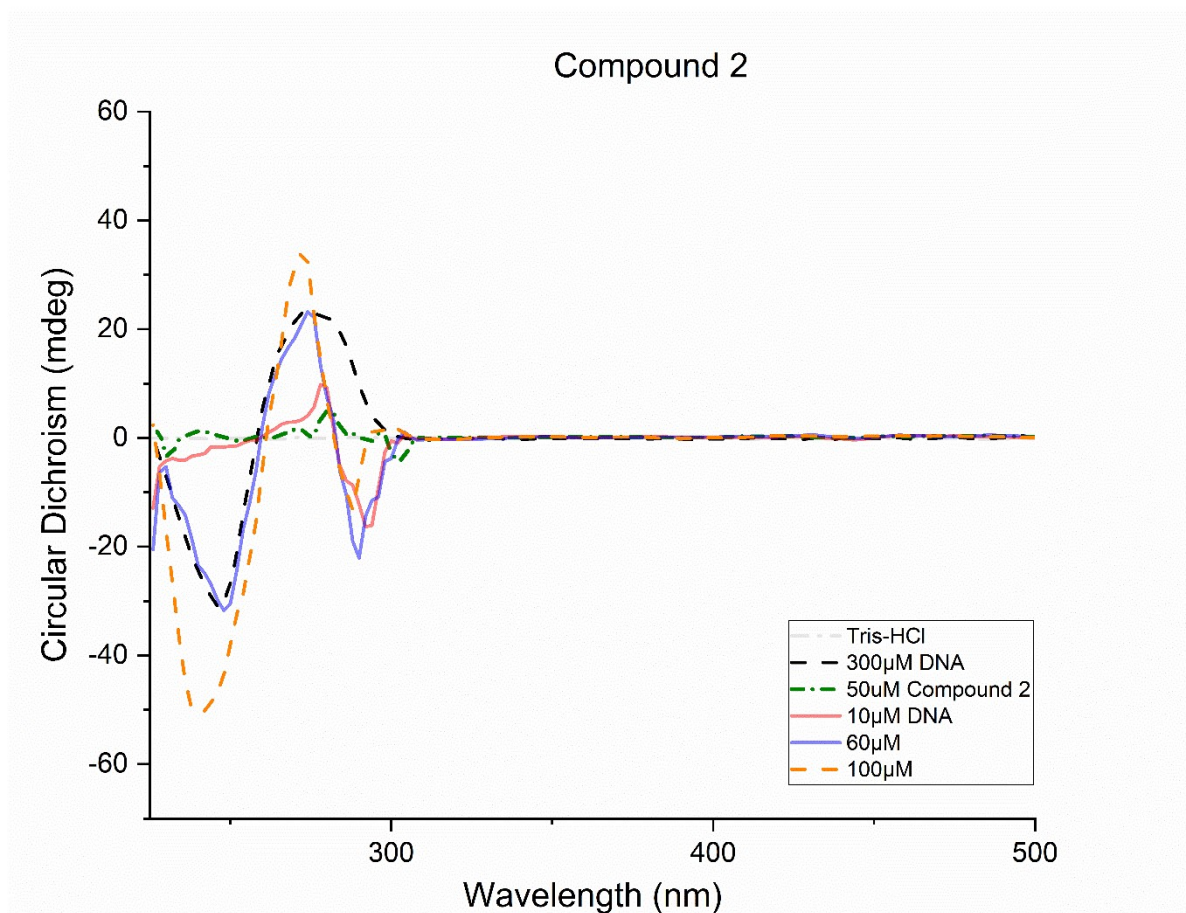


Figure S39. Circular dichroism spectra of the ct-DNA at different concentrations ($[DNA] = 10, 60$ and $100 \mu M$) in the presence of compound **2** at $50 \mu M$ in Tris-HCl buffer (pH 7.4).

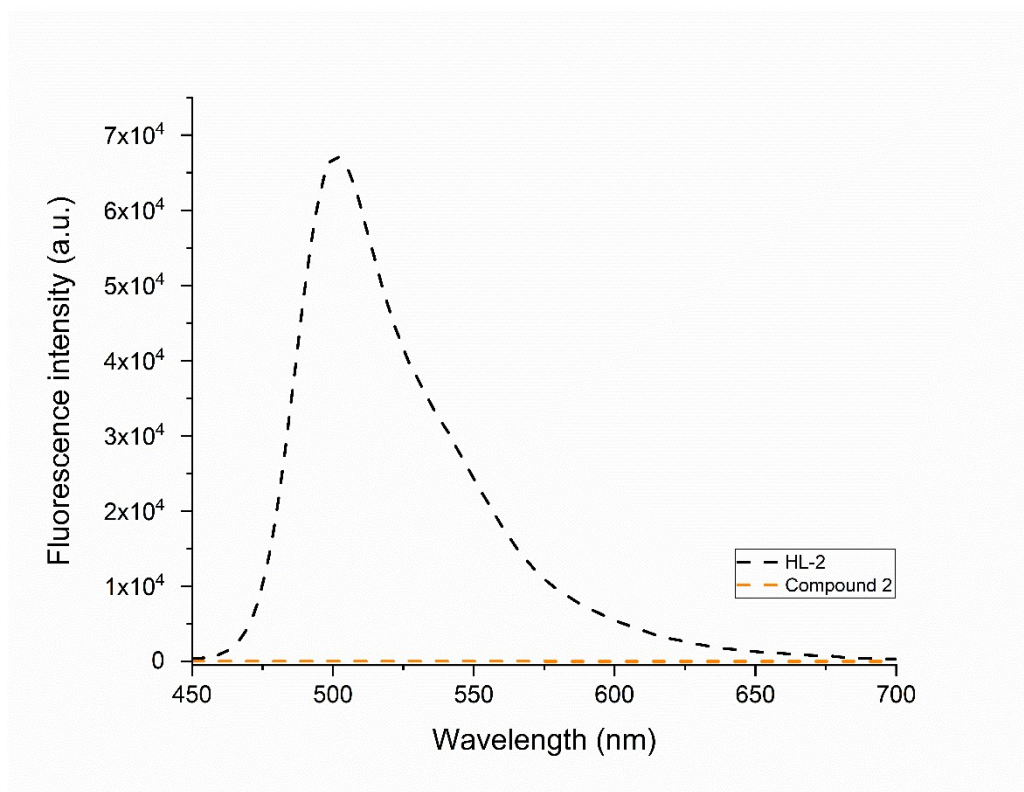


Figure S40. Fluorescence emission spectra of **HL2** and **2**, obtained at 50 μ M (λ_{ex} = 440 nm) in Tris-HCl buffer (pH 7.4) and recorded in an Edinburgh FLS980 spectrofluorometer.

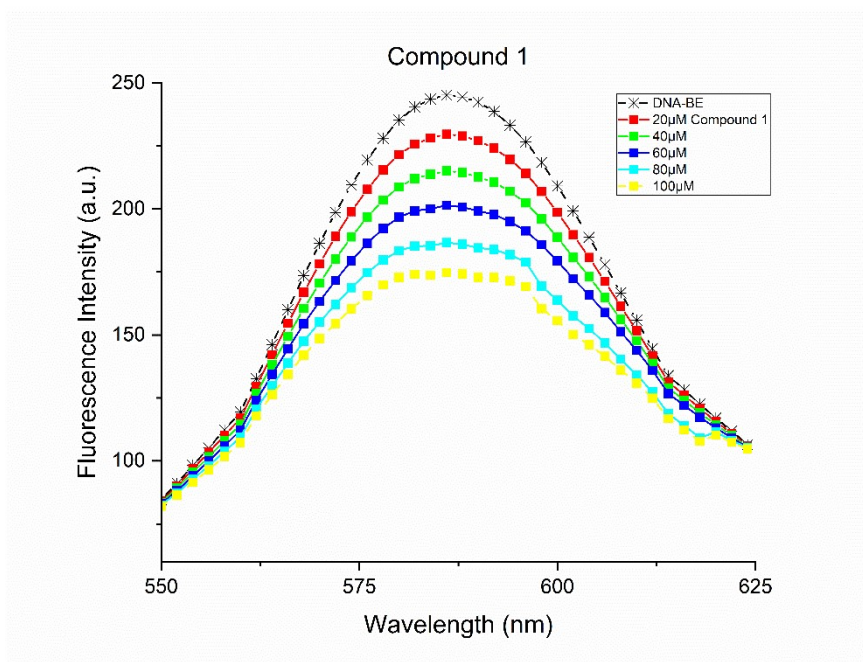


Figure S41. Emission quenching curves of EtdBr-DNA by complex 1 ($[\text{EtBr}] = 10 \mu\text{M}$, $[\text{DNA}] = 100 \mu\text{M}$, $[\text{Complex}] = 0\text{--}100 \mu\text{M}$).

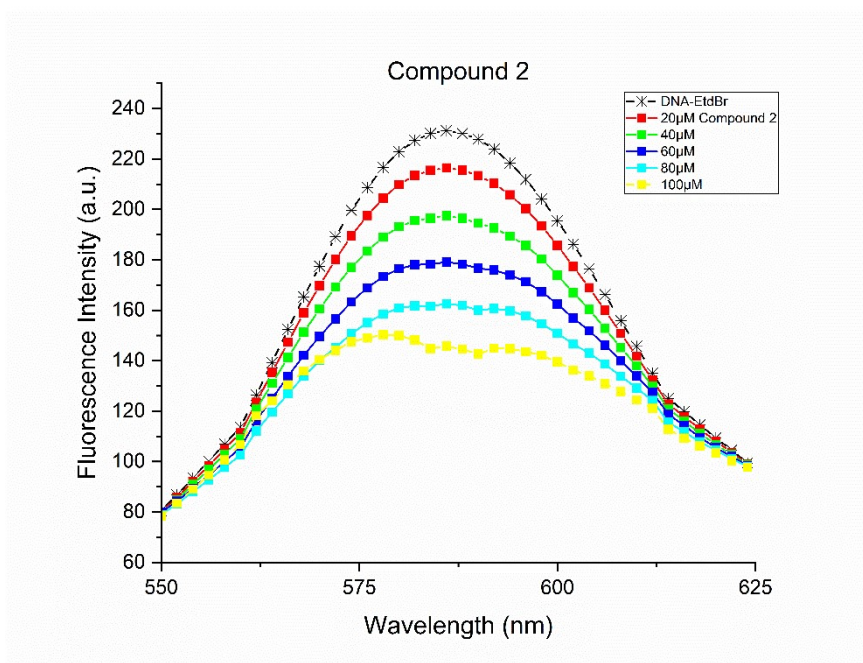


Figure S42. Emission quenching curves of EtdBr-DNA by complex 2 ($[\text{EtBr}] = 10 \mu\text{M}$, $[\text{DNA}] = 100 \mu\text{M}$, $[\text{Complex}] = 0\text{--}100 \mu\text{M}$).

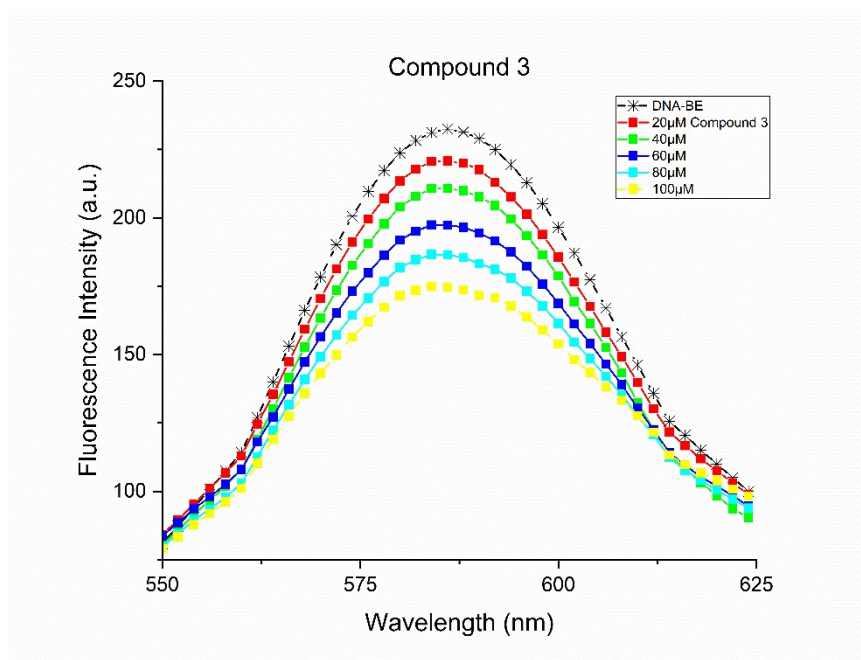


Figure S43. Emission quenching curves of EtdBr-DNA by complex **3** ([EtBr] = 10 μM , [DNA] = 100 μM , [Complex] = 0–100 μM).

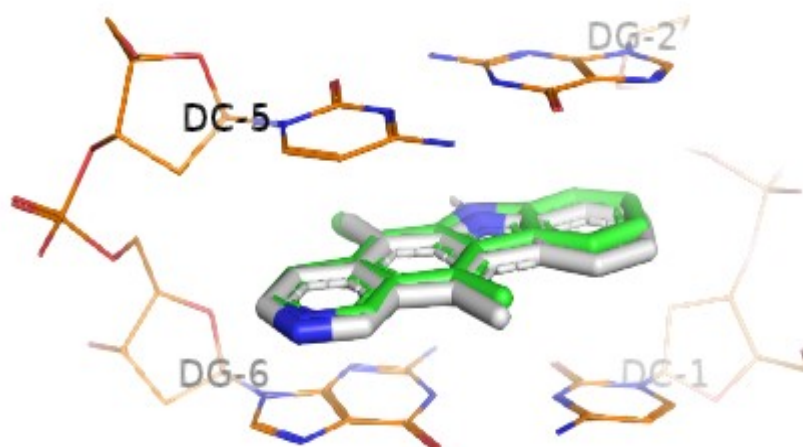


Figure S44. Redocking for target 1Z3F, available in PDB, Structure of ellipticine in complex with a 6-bp DNA, RMSD = 0.45.

Table S3: Molecular docking results for 1Z3F (PDB).

COMPOUNDS	SCORE
1	67.28
2	68.62
3	64.88
Ellipticine	75.45 (RMSD 0.45)