

**Bioavailability and toxicity of nanoscale/bulk rare earth oxides in soil: Physiological and ultrastructural alterations in *Eisenia fetida***

Muhammad Adeel<sup>a</sup>, Noman Shakoor<sup>a</sup>, Muhammad Arslan Ahmad<sup>b</sup>, Jason C. White<sup>c</sup>, Ghulam Jilani<sup>d</sup>, Yukui Rui<sup>a\*</sup>

<sup>a</sup>Beijing Key Laboratory of Farmland Soil Pollution Prevention and Remediation and College of Resources and Environmental Sciences, China Agricultural University, Beijing 100193, P.R. China

<sup>b</sup>Key Lab of Eco-restoration of Regional Contaminated Environment (Shenyang University), Ministry of Education, Shenyang 11044, PR China

<sup>c</sup>Connecticut Agricultural Experiment Station, New Haven, CT 06504, United States

<sup>d</sup>Institute of Soil Science, PMAS Arid Agriculture University, Rawalpindi 46300, Pakistan

\*Corresponding author:

Yukui Rui: [ruiyukui@163.com](mailto:ruiyukui@163.com)

Pages: 7

Figures: 5

Tables: 3

**Table of Contents**

1.1 Cast enzyme activities .....	- 2 -
2.1 NMs characterizations .....	- 2 -
References .....	- 7 -

## Supplementary information

### 1.1 Cast enzyme activities

Three enzymes were evaluated: acid phosphatase,  $\beta$ -glucosidase, and alkaline phosphatase<sup>1</sup>. The substrates used for the phosphatase and  $\beta$ -glucosidase protocols were p- Nitrophenyl phosphate (Sigma-Aldrich, St. Louis, MO) and 4-Nitro- phenyl  $\beta$ -D-glucopyranoside ( $\geq 98\%$ , Sigma-Aldrich, St. Louis, MO), respectively. The phosphatase samples were incubated for 45 min and  $\beta$ -glucosidase samples for 1 h. The reactions in the supernatants were then terminated by adding 1 N of NaOH. After adding ultrapure DI water for dilution, a Microplate Reader was used to read the developed colors at 410 nm. The phosphatase and  $\beta$ -glucosidase activity were both calculated as:

$$\text{Activity } (\mu\text{mol h}^{-1}\text{g}^{-1}) = \frac{\text{Final OD} - 1}{(\text{EC} = \mu\text{mol} = \text{mL}) / (1.5 \text{ mL/assay})} * (\text{incubation time, h}) * (\text{g sample suspension}) / (0.75 \text{ ml sample suspension})$$

where g DOM was the oven dried mass from 2 g of wet soil sample. The final OD value was calculated by subtracting the OD values of the sample controls and the substrate controls from the sample assays. The EC value for phosphatase and  $\beta$ -glucosidase was 2.2369 mL/ $\mu$ mol.

### 2.1 NMs characterizations

TEM results revealed the size of of nanoscale  $\text{La}_2\text{O}_3$ , bulk-  $\text{La}_2\text{O}_3$ , nanoscale  $\text{Yb}_2\text{O}_3$  and bulk-  $\text{Yb}_2\text{O}_3$  to be  $39 \pm 9$  nm,  $756 \pm 200$  nm,  $30 \pm 7$  nm and  $580 \pm 100$  nm, respectively. TEM observation showing that both nanoparticles are aggregated, but that the bulk particles were not aggregated (Figure S1 A). Zeta potential values of nanoscale  $\text{La}_2\text{O}_3$ , bulk  $\text{La}_2\text{O}_3$ , nanoscale  $\text{Yb}_2\text{O}_3$  and bulk  $\text{Yb}_2\text{O}_3$  were  $15 \pm 2$ ,  $19 \pm 1.5$ ,  $14 \pm 1.4$  and  $17 \pm 0.2$  mV (Table S1). These results suggest

moderate stability of both nanoscale and bulk materials. All bulk and nanoparticles readily dispersed into DI water. The dissolution percentage was significantly greater in acidic conditions (pH 4.5). However, both rare earth based NMs were only slightly dissolved in DI water at pH 7 (Figure S1 B).

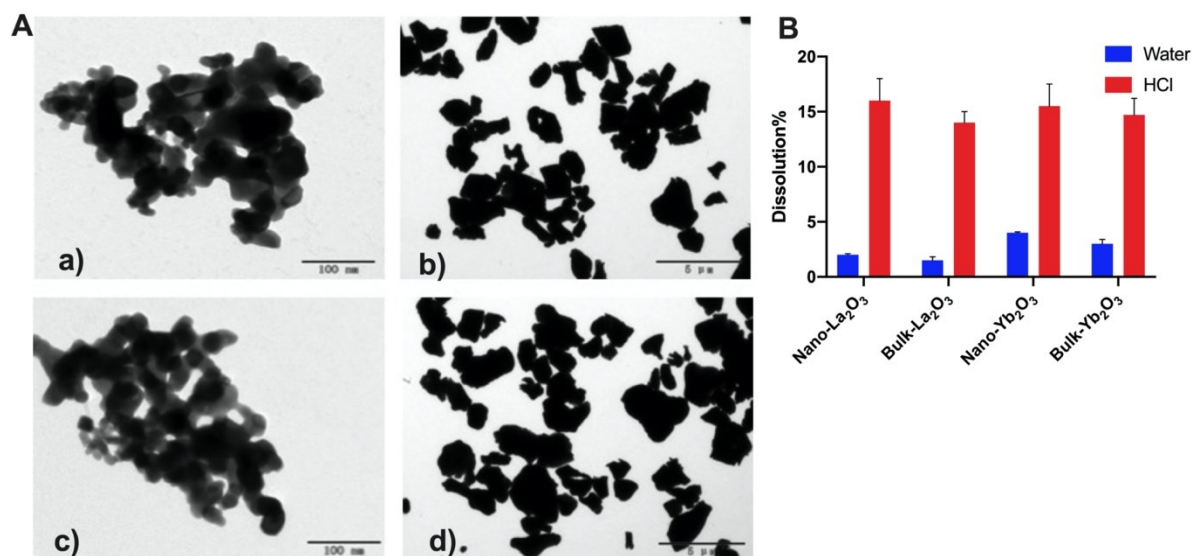


Figure S1 Morphological characterization. (A) TEM images of (a)nanoscale La<sub>2</sub>O<sub>3</sub> NPs, (b) bulk La<sub>2</sub>O<sub>3</sub>, (c) nanoscale Yb<sub>2</sub>O<sub>3</sub>, and (d)bulk Yb<sub>2</sub>O<sub>3</sub>. (B) Dissolution of nanoscale and bulk particles in DI water (pH=7) and acidic solution (pH=4.5)

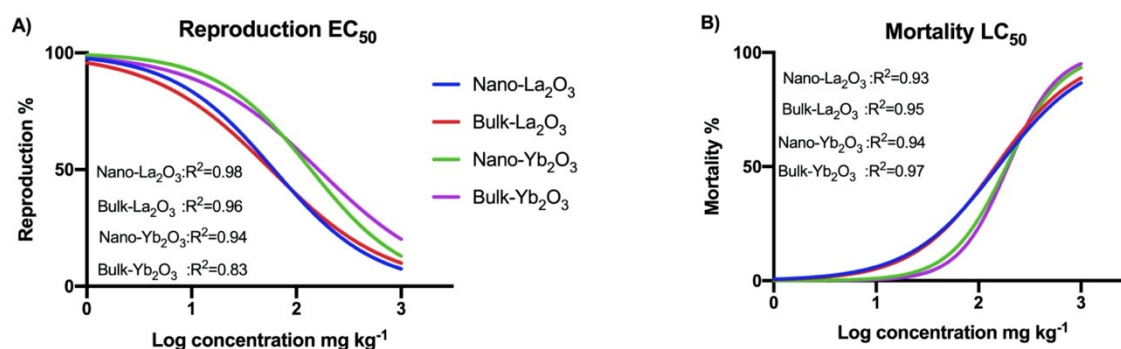


Figure S2 Dose–response curves for *Eisenia fetida* exposed to nanoscale and bulk REO for 28 days. (a) Reproduction (28-day median effective concentration (EC<sub>50</sub>) of nano La<sub>2</sub>O<sub>3</sub>, bulk La<sub>2</sub>O<sub>3</sub>, nano Yb<sub>2</sub>O<sub>3</sub> and bulk Yb<sub>2</sub>O<sub>3</sub> was. 60, 57, 138 and 162 mg kg<sup>-1</sup>, respectively (b) Mortality (28-day median lethal concentration (LC<sub>50</sub>) of nano La<sub>2</sub>O<sub>3</sub>, bulk La<sub>2</sub>O<sub>3</sub>, nano Yb<sub>2</sub>O<sub>3</sub>

and bulk  $\text{Yb}_2\text{O}_3$  was 155, 146, 186 and 193  $\text{mg kg}^{-1}$ . Results are presenting as average values and curves represent the models fit to data.  $p < 0.05$

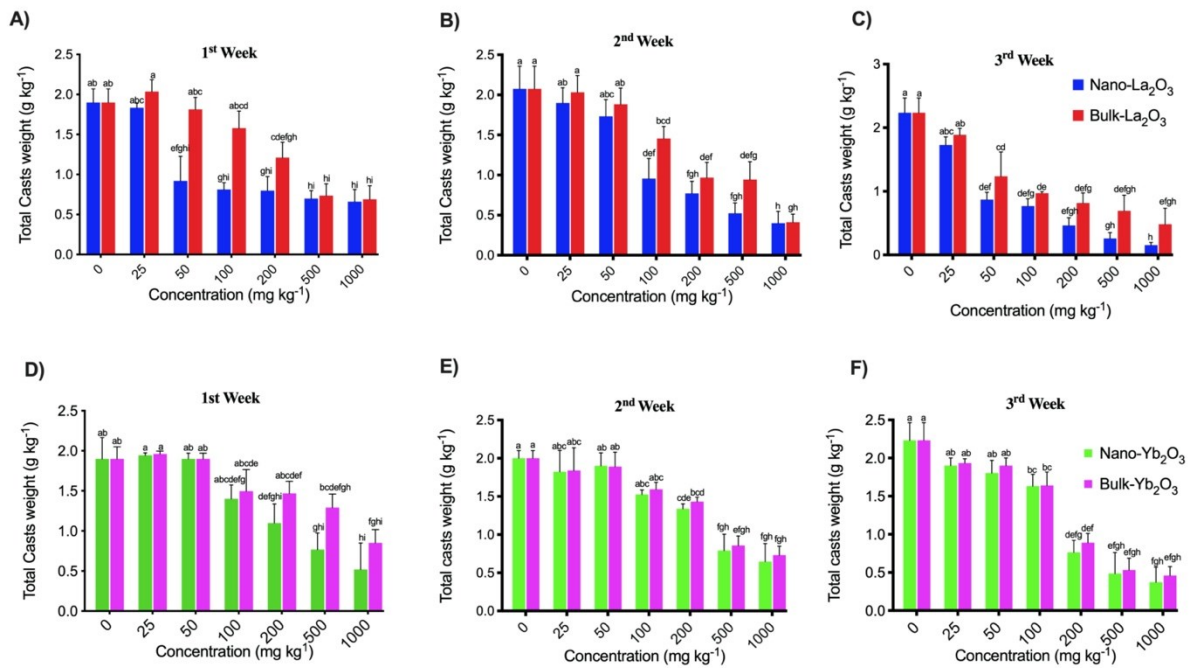


Figure S3. Weekly earthworm cast production under exposure to nanoscale and bulk REO based material (n=4,  $p < 0.05$ )

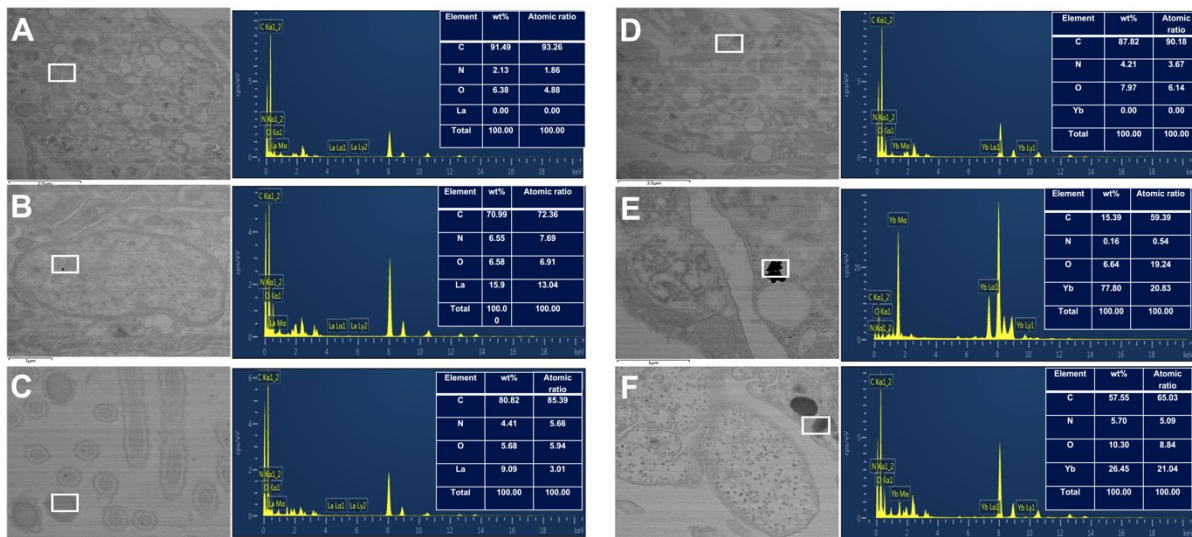


Figure S4. EDS analysis of earthworm tissue (A) Control (B) Nanoscale  $\text{La}_2\text{O}_3$  (C) Bulk  $\text{La}_2\text{O}_3$  (D) Control (E) Nanoscale  $\text{Yb}_2\text{O}_3$  (F) Bulk  $\text{Yb}_2\text{O}_3$

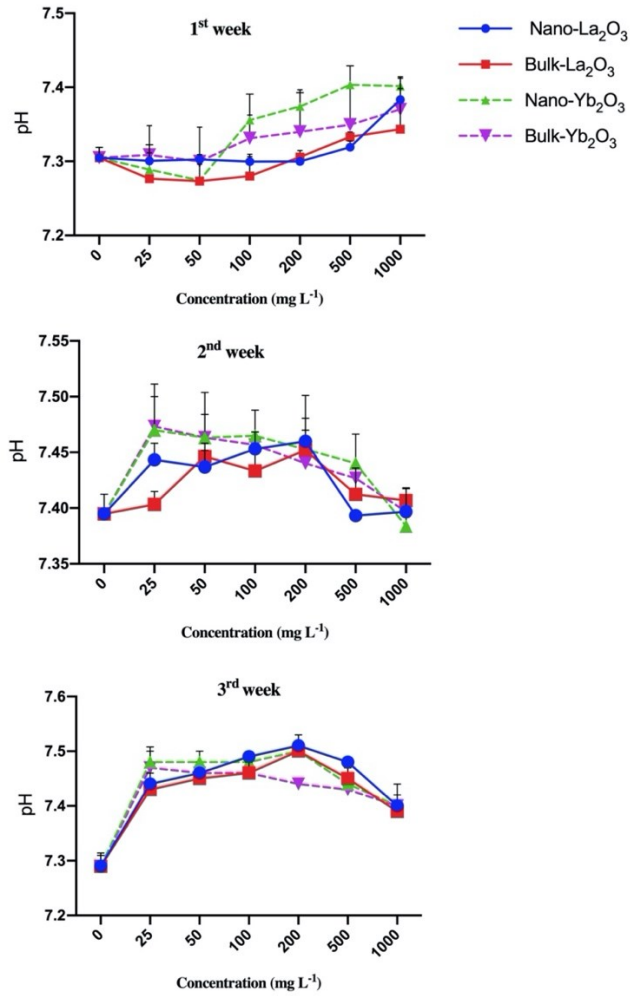


Figure S5. Weekly pH measurements in exposed soil

**Table S1. Soil characteristics**

Index	Mean Value
Soil property	Silty loam
Dry unit weight (g/cm <sup>3</sup> )	1.29
pH	6.9
Electrical conductivity (dS/m)	0.16
Rapidly available N (mg/kg)	20.37
Rapidly available P (mg/kg)	11.21
Rapidly available K (mg/kg)	73.64
Organic matter (mg/kg)	11.31
CEC (cmol/kg)	14.27
CaCO <sub>3</sub> (g/kg)	43.19
Rapidly available Fe (mg/kg)	22.91
Rapidly available Mn (mg/kg)	12.77
Rapidly available Cu (mg/kg)	3.17
Rapidly available Ni (mg/kg)	20.22

Table S2. Measurements of REO-NMs

REO	Purity(%)	TEM (nm)	$\zeta$ mv	DLS (nm)
Nano-La <sub>2</sub> O <sub>3</sub>	98.8	39±9	15±2	216±20
Bulk-La <sub>2</sub> O <sub>3</sub>	98.9	756±200	19±1.5	624±14
Nano-Yb <sub>2</sub> O <sub>3</sub>	99.9	30±7	14±1.4	207±13
Bulk-Yb <sub>2</sub> O <sub>3</sub>	99.9	580±100	17±0.2	557±15

Table S3 : Bioaccumulation factor (BAF) of nanoscale and bulk REO in earthworm over 28 days Mean ± SD (n = 3) with different letters in the column indicate significant ( $p < 0.005$ ) difference between the treatments.

mg kg <sup>-1</sup>	Nao-La <sub>2</sub> O <sub>3</sub>	Bulk-La <sub>2</sub> O <sub>3</sub>	Nano-Yb <sub>2</sub> O <sub>3</sub>	Bulk- Yb <sub>2</sub> O <sub>3</sub>
<b>25</b>	0.08±0.00096 <sup>fg</sup>	0.07±0.00020 <sup>fg</sup>	0.13±0.0033 <sup>a</sup>	0.11±0.00035 <sup>b</sup>
<b>50</b>	0.11±0.0057 <sup>cd</sup>	0.10±0.00030 <sup>de</sup>	0.13±0.0070 <sup>a</sup>	0.12±0.00035 <sup>a</sup>
<b>100</b>	0.10±0.00020 <sup>bc</sup>	0.06±0.00061 <sup>h</sup>	0.09±0.0002 <sup>e</sup>	0.0096±0.47 <sup>e</sup>
<b>200</b>	0.06±0.00027 <sup>h</sup>	0.04±0.00042 <sup>i</sup>	0.08±0.00035 <sup>f</sup>	0.07±0.00071 <sup>g</sup>
<b>500</b>	0.03±0.00022 <sup>i</sup>	0.03±0.00082 <sup>i</sup>	0.07±0.0002 <sup>fg</sup>	0.06±0.0012 <sup>h</sup>
<b>1000</b>	0.03±00045 <sup>i</sup>	0.03±0.00021 <sup>i</sup>	0.06±0.0004 <sup>h</sup>	0.03±0.00287 <sup>i</sup>

## References

1. Li, B.; Chen, Y.; Liang, W.-z.; Mu, L.; Bridges, W. C.; Jacobson, A. R.; Darnault, C. J. G., Influence of cerium oxide nanoparticles on the soil enzyme activities in a soil-grass microcosm system. *Geoderma* **2017**, *299*, 54-62.