

Supporting Information

Visible light-driven Metal-free Synthesis of Highly Substituted Pyrroles through C–H Functionalisation

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1. General Information

All reagents were purchased from commercial suppliers (Sigma-Aldrich, Oakwood and Combi-Blocks) and used without further purification, all solvents were analytical grade. Thin layer chromatography (TLC) was performed using silica gel GF254, 0.25 mm thickness, visualization was accomplished with short wave UV light or KMnO_4 staining solution followed by heating. Hydrogen nuclear magnetic resonance spectra (^1H NMR) were obtained at 500 MHz in CDCl_3 solutions, at ambient temperature. Carbon-13 nuclear magnetic resonance spectra (^{13}C NMR) were obtained at 125 MHz in CDCl_3 solutions, at ambient temperature. Chemical shifts (δ) are given in ppm and the residual solvent signals were used as references for ^1H and ^{13}C NMR spectra (CDCl_3 : $\delta\text{H} = 7.27$ ppm, $\delta\text{C} = 77.00$ ppm). High resolution mass spectra were recorded on Thermo Scientific LTQ FT Ultra and Q Exactive Orbitrap spectrometers working with an electrospray ionization (ESI). The Gas Chromatography coupled to Mass Spectrometry (GC-MS) analyses were performed using a Network GC system 6890N (Agilent Technologies Inc., Palo Alto, CA, USA), equipped with a HP-5MS 5% Phenyl Methyl Silox (25.0 m \times 250 μm \times 0.25 μ nominal) capillary column. The GC analyses were carried out in split mode (ratio 150:1) using helium as carrier gas at a flow rate of 504 mL/min (7.65 psi). The injection port temperature was 250 $^\circ\text{C}$; the oven was maintained at an initial temperature of 50 $^\circ\text{C}$ for 3 minutes, then programmed at 40 $^\circ\text{C}/\text{min}$ to a temperature of 280 $^\circ\text{C}$, where it was held, post-run, for 2 minutes. The MS detector was at 250 $^\circ\text{C}$, using H_2 flow at 40.00 mL/min, air at 400 mL/min and He makeup flow at 45.0 mL/min.

2. General Procedure

Benzophenone (0.5 mmol, 50 mol%), amines (1.0 mmol), methyl acetoacetate (2.0 mmol) and acid (1.0 mmol) were added to a test tube and capped with a rubber septum. The reaction mixture was stirred and irradiated using a compact fluorescent lamp (CFL, 50 W) at room temperature (kept at around 35 $^\circ\text{C}$ using a fan) for 72 h. After completion of the reaction (followed by TLC), the whole mixture was added to the top of a silica column and eluted using ethyl acetate/n-hexane (20:80) to afford the targeted product. For reactions employing levulinic acid the eluent was changed to ethyl acetate/n-hexane (30:70).

A description of the apparatus for the photoreaction is shown in Figure S1.

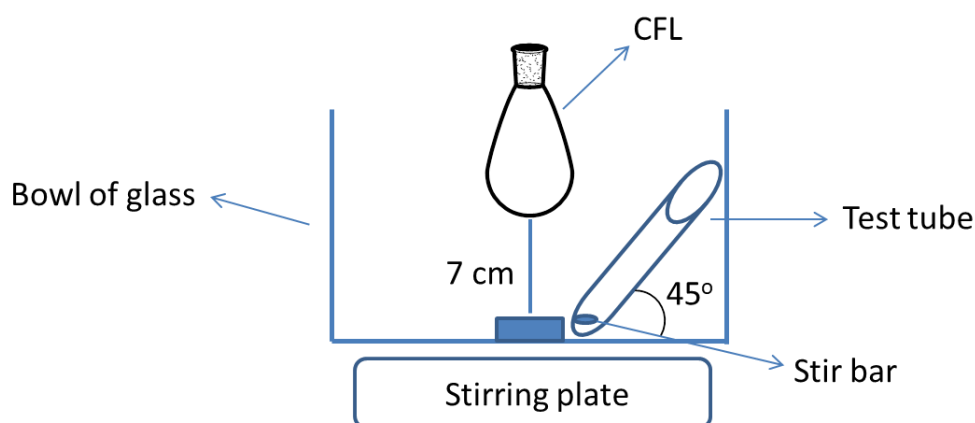


Figure S1. Homemade apparatus for the photochemical reaction.

Figure S2 presents the emission profile of the CFL lamp employed in this method. The overlap between benzophenone absorption band ($< 380 \text{ nm}$)¹ and CFL emission bands can be considered small. Under such conditions, access to the triplet state of benzophenone may be impaired and longer reaction times can be observed (e.g. 72 h reaction time). Alternatively, the triplet state of benzophenone might decay with different lifetimes depending on the solvent and media.² Under the neat condition described in our method, benzophenone triplet state lifetime may be reduced in such a way that productive quenching processes would be more difficult, thus having an effect

¹ <https://webbook.nist.gov/cgi/cbook.cgi?ID=C119619&Mask=400>. Accessed on July, 2021.

² G. Dormán, H. Nakamura, A. Pulsipher and G. D. Prestwich, *Chem. Rev.*, 2016, **116**, 15284–15398.

on the reaction time. For a brief discussion about even longer reaction times observed in our study, see item 6 of this Supporting Information.

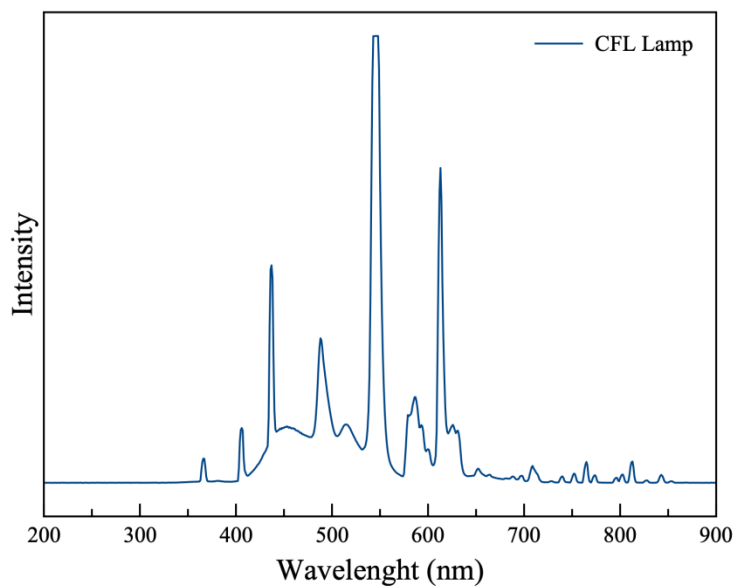
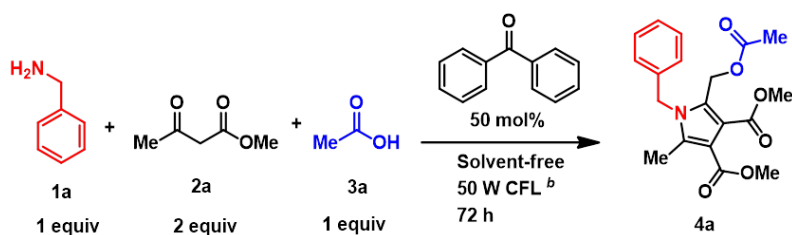
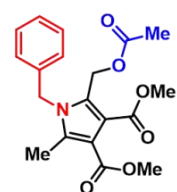
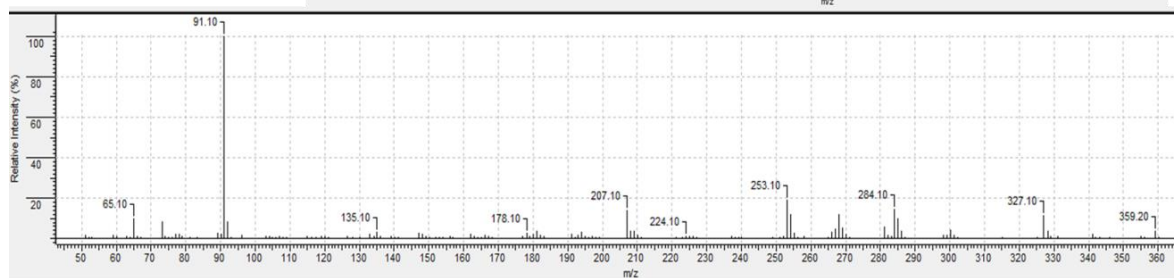
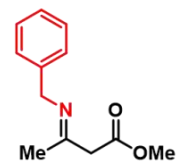
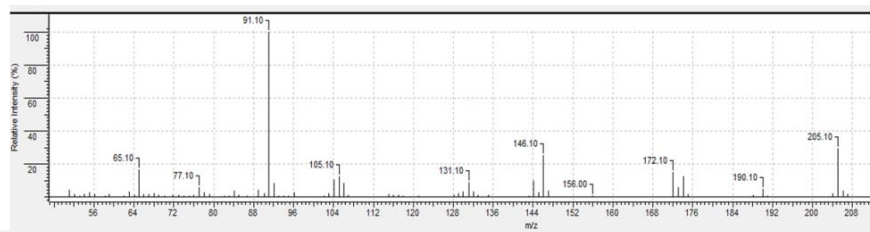


Figure S2. Emission profile for CFL lamp.

3. Kinetic profile



Benzophenone (0.5 mmol, 50 mol%), benzylamine (1.0 mmol), methyl acetoacetate (2.0 mmol) and acetic acid (1.0 mmol) were employed following the **General Procedure**. Aliquots of the reaction mixture were taken (10 min, 24 h, 48 h, 72 h and 96 h), diluted with ethyl acetate and analyzed by GC-MS (Figure S3). The experiment was done in triplicate.



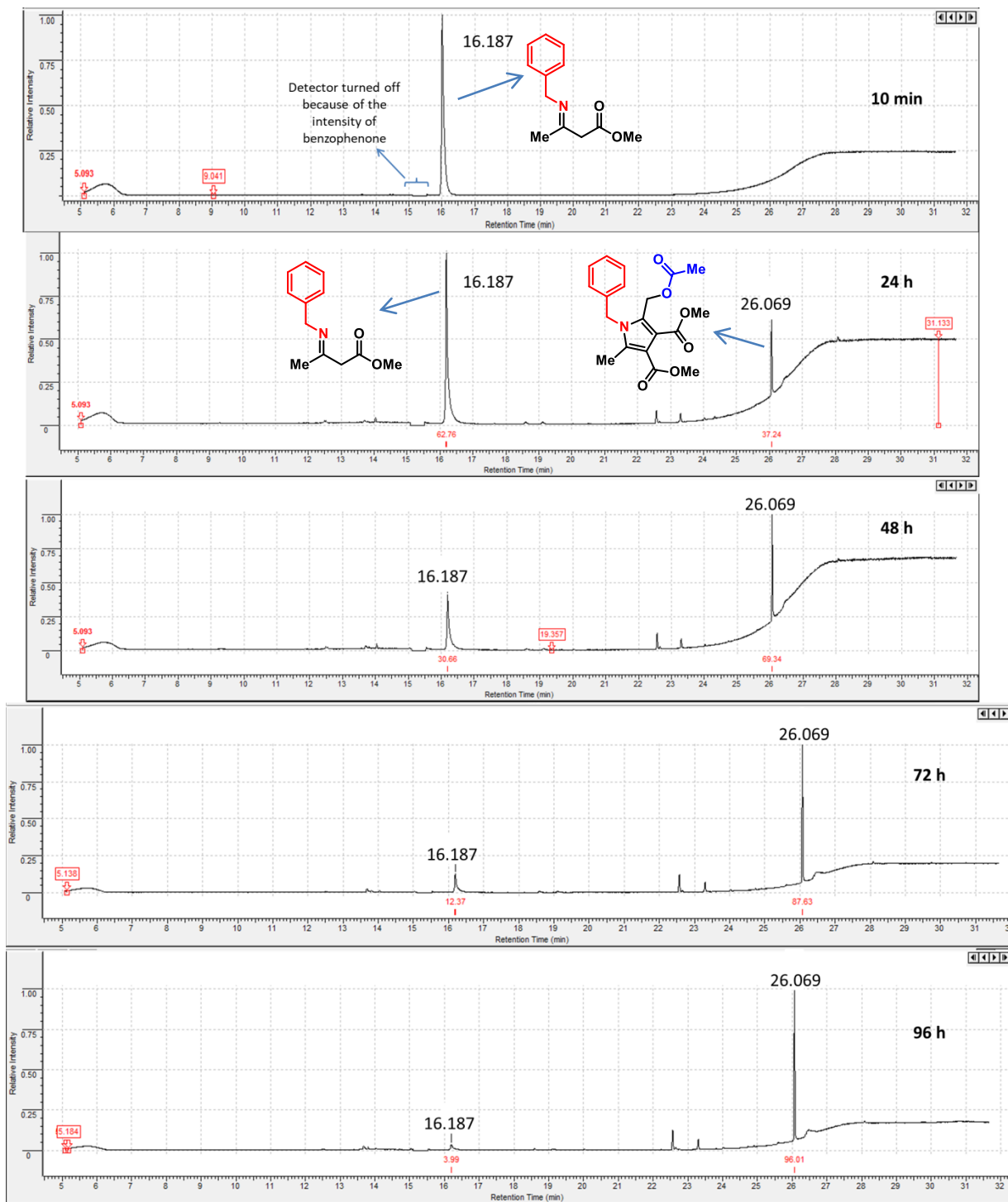


Figure S3. Kinetic profile.

4. Intermediate pyrroles

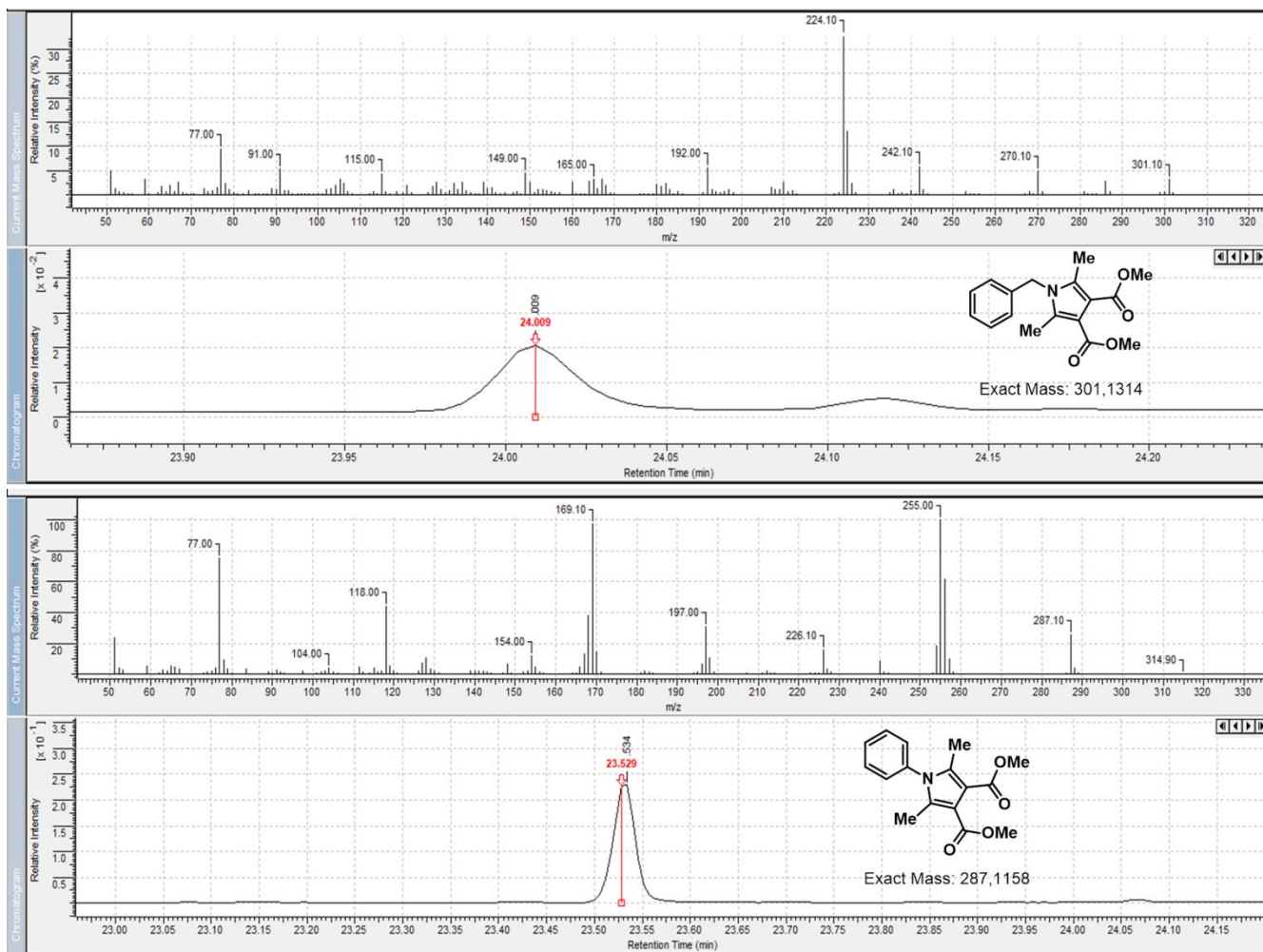
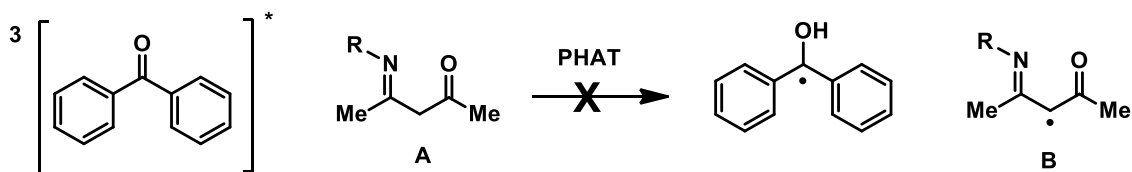


Figure S4. Intermediate pyrroles observed when employing benzylamine or aniline in the reaction.

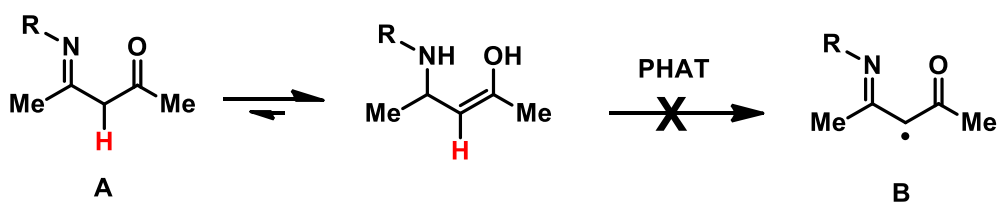
5. Possible reason for reaction failure when using either acetylacetone or dimethyl malonate.

Following the **General Procedure** with acetylacetone, benzylamine and acetic acid, we could observe by GC-MS the formation of the corresponding imine **A** in accordance to the mechanistic studies described in the manuscript. However, no product was obtained after 72 h irradiation and the imine remained unreacted.

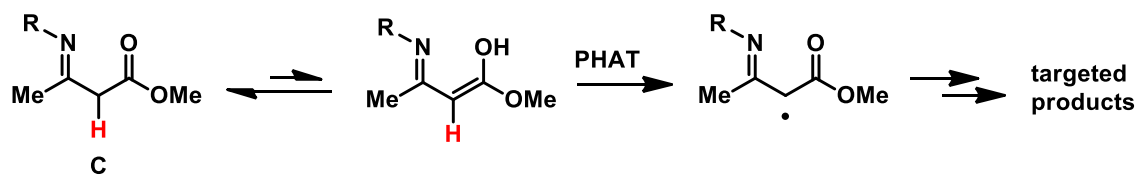
It suggests that photoinduced HAT transfer from imine **A** to the triplet state of benzophenone to furnish radical **B** is strongly unfavourable:



We hypothesize that, under our conditions, the enol form of imine **A** is highly favoured and considering that HAT parallels bond dissociation energies³, hydrogen transfer from sp^2 carbon becomes more difficult compared to the same transfer from sp^3 carbons:



On the other hand, the keto form of imine **C**, coming from methyl acetoacetate, might be favoured due to higher pK_a of α -hydrogens next to an ester group. In this scenario, imine **C** undergoes HAT more easily (from sp^3 carbon) allowing the transformation reach completion:



from methyl acetoacetate

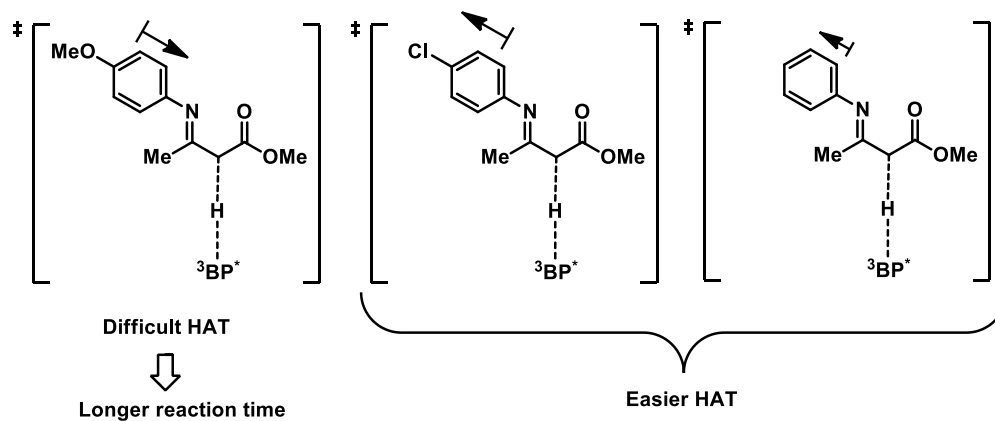
When employing dimethyl malonate, it is recovered unreacted and the process is shut down. This result emphasizes the necessity of imine formation to carry the transformation on.

6. Possible reason for longer reaction time when employing 4-methoxyaniline.

As reported in the manuscript, electron-rich 4-methoxyaniline has taken 6 days to reach 75% yield of the targeted product whereas other anilines have taken 72 h to furnish products in even higher yields. This outcome suggests an influence of electronic nature of anilines in the transformation. Here, we hypothesize that HAT step is the determining step in the reaction and possibly the electronic nature of anilines determines how fast the hydrogen transfer may occurs. The conjugation of the imines coming from anilines extends from the C–H bond susceptible to HAT towards the ring; hence, the influence of the electronics of the ring might be more substantial in this scenario.

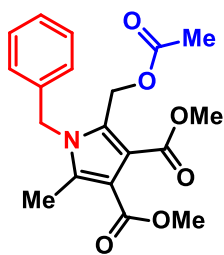
³ J. M. Mayer, *Acc. Chem. Res.*, 2011, **44**, 36–46.

Lanzalunga and collaborators⁴ reported polar effects in hydrogen atom transfer reactions from activated phenols and we believe we have the same kind of effect in the reaction employing 4-methoxyaniline. The polarization of the hydrogen atom transfer transition state because of the partial charge transfer may determine stabilization or destabilization of the transition state:



⁴ M. Bietti, E. Cucinotta, G. A. Dilabio, O. Lanzalunga, A. Lapi, M. Mazzonna, E. Romero-Montalvo and M. Salamone, *J. Org. Chem.*, 2019, **84**, 1778–1786.

7. Characterization of the products



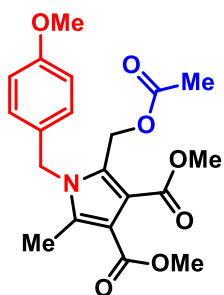
4a

Prepared from benzylamine, methyl acetoacetate and acetic acid following the general procedure to give the product as pale yellow oil (82% yield).

$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 7.32–7.26 (m, 3H), 6.88 (d, 2H, $J = 7.55$ Hz), 5.22 (s, 2H), 5.15 (s, 2H), 3.86 (s, 3H), 3.83 (s, 3H), 2.34 (s, 3H), 1.77 (s, 3H).

$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 170.35, 165.32, 165.24, 136.10, 135.85, 129.44, 128.93, 127.68, 125.43, 116.93, 112.88, 55.12, 51.86, 51.47, 47.40, 20.41, 10.92.

HRMS m/z (ESI): calcd. for $\text{C}_{19}\text{H}_{22}\text{NO}_6$ $[\text{M}+\text{H}]^+$ 360.1447, found 360.1444.



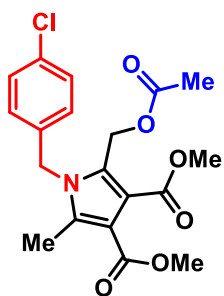
4b

Prepared from 4-methoxybenzylamine, methyl acetoacetate and acetic acid following the general procedure to give the product as pale orange oil (80% yield).

$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 6.83 (m, 4H), 5.21 (s, 2H), 5.08 (s, 2H), 3.86 (s, 3H), 3.82 (s, 3H), 3.78 (s, 3H), 2.34 (s, 3H), 1.83 (s, 3H).

$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 170.60, 165.57, 165.49, 159.34, 136.29, 129.62, 128.00, 126.98, 117.11, 114.54, 113.05, 55.53, 55.40, 52.08, 51.70, 47.19, 20.76, 11.19.

HRMS m/z (ESI): calcd. for $\text{C}_{20}\text{H}_{27}\text{NO}_7$ $[\text{M}+\text{H}]^+$ 390.1553, found 390.1552.



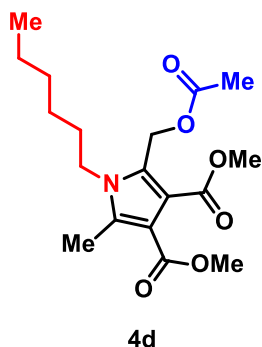
4c

Prepared from 4-chlorobenzylamine, methyl acetoacetate and acetic acid following the general procedure to give the product as pale yellow oil (85% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.28 (d, 2H, $J = 8.17$ Hz), 6.81 (d, 2H, $J = 8.17$ Hz), 5.20 (s, 2H), 5.11 (s, 2H), 3.86 (s, 3H), 3.83 (s, 3H), 2.32 (s, 3H), 1.80 (s, 3H).

¹³C NMR (125 MHz, Chloroform-*d*): δ 170.27, 165.17, 165.11, 135.85, 134.42, 133.64, 129.30, 129.09, 126.85, 117.11, 113.08, 55.02, 51.89, 51.51, 46.83, 20.44, 10.89.

HRMS m/z (ESI): calcd. for C₁₉H₂₁ClNO₆ [M+H]⁺ 394.1057, found 394.1055.

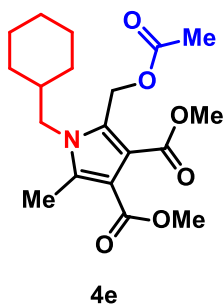


Prepared from hexylamine, methyl acetoacetate and acetic acid following the general procedure to give the product as pale yellow oil (94% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 5.23 (s, 2H), 3.84–3.79 (m+s+s, 8H), 2.42 (s, 3H), 2.05 (s, 3H), 1.66–1.60 (m, 2H), 1.38–1.27 (m, 6H), 0.89 (t, 3H, $J = 6.92$ Hz)

¹³C NMR (125 MHz, Chloroform-*d*): δ 170.40, 165.36, 165.33, 135.32, 128.42, 116.65, 112.46, 55.31, 51.76, 51.36, 44.37, 31.32, 30.80, 26.43, 22.42, 20.81, 13.89, 10.92.

HRMS m/z (ESI): calcd. for C₁₈H₂₈NO₆ [M+H]⁺ 354.1917, found 354.1915.

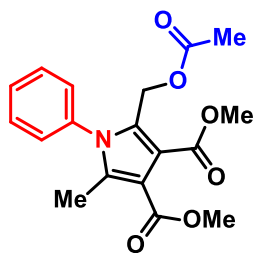


Prepared from cyclohexanemethylamine, methyl acetoacetate and acetic acid following the general procedure to give the product as pale yellow oil (95% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 5.23 (s, 2H), 3.83 (s, 3H), 3.79 (s, 3H), 3.69 (d, 2H, $J = 6.92$ Hz), 2.41 (s, 3H), 2.05 (s, 3H), 1.75–1.59 (m, 6H), 1.16 (m, 3H), 0.98 (m, 2H).

¹³C NMR (125 MHz, Chloroform-*d*): δ 170.51, 165.43, 165.36, 135.80, 128.98, 116.76, 112.51, 55.48, 51.75, 51.36, 50.28, 39.35, 30.74, 26.04, 25.79, 20.82, 11.49.

HRMS m/z (ESI): calcd. for C₁₉H₂₈NO₆ [M+H]⁺ 366.1917, found 366.1914.



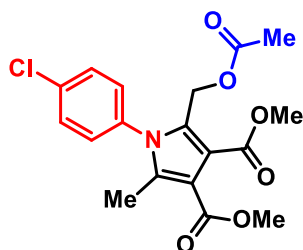
4f

Prepared from aniline, methyl acetoacetate and acetic acid following the general procedure to give the product as pale yellow oil (81% yield).

$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 7.50–7.54 (m, 3H), 7.20–7.24 (m, 2H), 4.96 (s, 2H), 3.87 (s, 3H), 3.85 (s, 3H), 2.19 (s, 3H), 1.91 (s, 3H).

$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 170.03, 165.32, 165.21, 136.79, 135.81, 129.97, 129.62, 129.57, 128.16, 116.85, 112.65, 55.64, 51.94, 51.55, 20.61, 11.86.

HRMS m/z (ESI): calcd. for $\text{C}_{18}\text{H}_{20}\text{NO}_6$ $[\text{M}+\text{H}]^+$ 346.1291, found 346.1293.



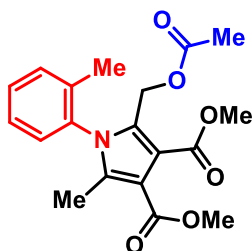
4g

Prepared from 4-chloro aniline, methyl acetoacetate and acetic acid following the general procedure to give the product as pale orange oil (84% yield).

$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 7.48 (d, 2H, $J = 8.80$ Hz), 7.16 (d, 2H, $J = 8.80$ Hz), 4.95 (s, 2H), 3.87 (s, 3H), 3.85 (s, 3H), 2.19 (s, 3H), 1.93 (s, 3H).

$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 169.97, 165.13, 165.04, 136.64, 135.84, 134.30, 129.88, 129.51, 117.21, 112.96, 55.54, 51.99, 51.61, 20.63, 11.85.

HRMS m/z (ESI): calcd. for $\text{C}_{18}\text{H}_{19}\text{ClNO}_6$ $[\text{M}+\text{H}]^+$ 380.0901, found 380.0903.



4h

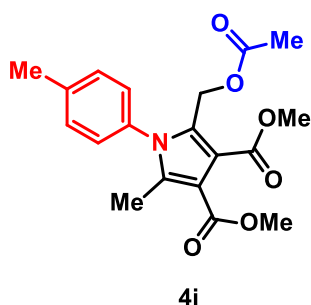
Prepared from *o*-toluidine (2-methyl aniline), methyl acetoacetate and acetic acid following the general procedure to give the product as pale yellow oil (83% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.41 (dd, 1H, $J = 6.92, 7.55$ Hz), 7.36 (d, 1H, $J = 6.92$ Hz), 7.32 (ap t, 1H, $J = 7.55$ Hz), 7.12 (d, 1H, $J = 7.55$ Hz), 4.93 (d, 2H, $J = 13.20$ Hz), 4.85 (d, 2H, $J = 13.20$ Hz), 3.87 (s, 3H), 3.85 (s, 3H), 2.11 (s, 3H), 1.98 (s, 3H), 1.87 (s, 3H).

¹³C NMR (125 MHz, Chloroform-*d*): δ 169.93, 165.29, 165.17, 136.58, 134.82, 131.13, 129.98, 129.39, 128.50, 127.08, 116.84, 112.60, 55.60, 51.87, 51.49, 20.49, 17.10, 11.41.

HSQC, HMBC, COSY, DEPT135, NOESY: See Spectral Data Section.

HRMS m/z (ESI): calcd. for C₁₉H₂₂NO₆ [M+H]⁺ 360.1447, found 360.1445.

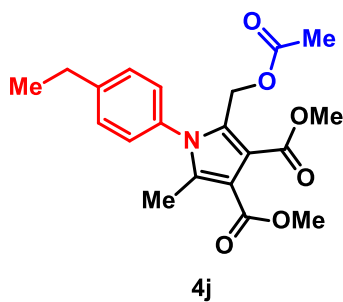


Prepared from 4-methyl aniline, methyl acetoacetate and acetic acid following the general procedure to give the product as pale brown oil (82% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.28 (d, 2H, $J = 8.17$ Hz), 7.08 (d, 2H, $J = 8.17$ Hz), 4.94 (s, 2H), 3.86 (s, 3H), 3.84 (s, 3H), 2.43 (s, 3H), 2.18 (s, 3H), 1.92 (s, 3H).

¹³C NMR (125 MHz, Chloroform-*d*): δ 170.09, 165.33, 165.22, 139.73, 136.90, 133.10, 130.13, 130.03, 127.84, 116.69, 112.51, 55.72, 51.87, 51.49, 21.19, 20.65, 11.84.

HRMS m/z (ESI): calcd. for C₁₉H₂₂NO₆ [M+H]⁺ 360.1447, found 360.1446.

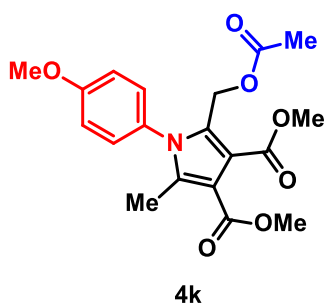


Prepared from 4-ethyl aniline, methyl acetoacetate and acetic acid following the general procedure to give the product as pale orange oil (89% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.31 (d, 2H, $J = 8.17$ Hz), 7.11 (d, 2H, $J = 8.17$ Hz), 4.94 (s, 2H), 3.86 (s, 3H), 3.84 (s, 3H), 2.73 (q, 2H, $J = 7.80$ Hz), 2.19 (s, 3H), 1.92 (s, 3H), 1.30 (t, 3H, $J = 7.55$ Hz).

¹³C NMR (125 MHz, Chloroform-*d*): δ 170.08, 165.35, 165.22, 145.88, 136.92, 133.24, 130.05, 128.90, 127.91, 116.66, 112.49, 55.76, 51.87, 51.51, 28.49, 20.63, 15.20, 11.88.

HRMS m/z (ESI): calcd. for C₂₀H₂₄NO₆ [M+H]⁺ 374.1604, found 374.1607.

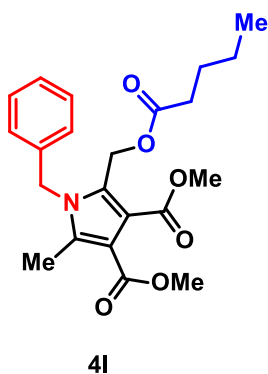


Prepared from 4-methoxy aniline, methyl acetoacetate and acetic acid following the general procedure to give the product as pale brown oil (71% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.12 (d, 2H, $J = 8.80$ Hz), 6.98 (d, 2H, $J = 8.80$ Hz), 4.95 (s, 2H), 3.88 (s, 3H), 3.87 (s, 3H), 3.85 (s, 3H), 2.18 (s, 3H), 1.94 (s, 3H).

¹³C NMR (125 MHz, Chloroform-*d*): δ 170.09, 165.33, 165.24, 160.16, 137.14, 130.21, 129.22, 128.24, 116.61, 114.62, 112.41, 55.72, 55.53, 51.89, 51.51, 20.66, 11.83.

HRMS m/z (ESI): calcd. for $C_{19}H_{22}NO_7$ $[M+H]^+$ 376.1396, found 376.1397.

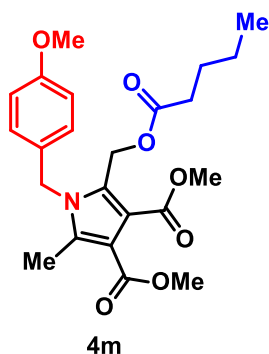


Prepared from benzylamine, methyl acetoacetate and valeric acid following the general procedure to give the product as pale yellow oil (82% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.30–7.26 (m, 3H), 6.88 (d, 2H, $J = 6.95$ Hz), 5.22 (s, 2H), 5.16 (s, 2H), 3.87 (s, 3H), 3.84 (s, 3H), 2.35 (s, 3H), 2.02 (t, 2H, $J = 7.55$ Hz), 1.43 (quint, 2H, $J = 7.55$ Hz), 1.24 (sext, 2H, $J = 7.55$ Hz), 0.85 (t, 3H, $J = 7.55$ Hz).

¹³C NMR (125 MHz, Chloroform-*d*): δ 173.18, 165.33, 165.22, 136.04, 135.89, 129.60, 128.91, 127.68, 125.43, 116.82, 112.89, 55.00, 51.84, 51.47, 47.39, 33.50, 26.65, 22.10, 13.59, 10.93.

HRMS m/z (ESI): calcd. for $C_{22}H_{28}NO_6$ $[M+H]^+$ 402.1917, found 402.1915.

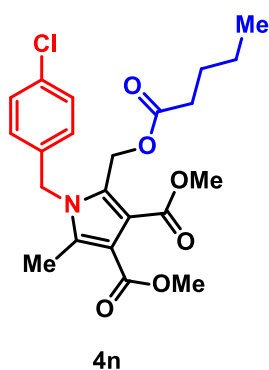


Prepared from 4-methoxybenzylamine, methyl acetoacetate and valeric acid following the general procedure to give the product as pale yellow oil (87% yield)

¹H NMR (500 MHz, Chloroform-*d*): δ 6.83 (m, 4H), 5.21 (s, 2H), 5.08 (s, 2H), 3.85 (s, 3H), 3.82 (s, 3H), 3.78 (s, 3H), 2.33 (s, 3H), 2.07 (t, 2H, $J = 7.55$ Hz), 1.44 (quint, 2H, $J = 7.55$ Hz), 1.22 (sext, 2H, $J = 7.55$ Hz), 0.85 (t, 3H, $J = 7.55$ Hz).

¹³C NMR (125 MHz, Chloroform-*d*): δ 173.17, 165.35, 165.23, 159.10, 136.00, 129.54, 127.80, 126.73, 116.77, 114.28, 112.84, 55.26, 55.04, 51.82, 51.45, 46.93, 33.59, 26.70, 22.12, 13.59, 10.96.

HRMS m/z (ESI): calcd. for C₂₃H₃₀NO₇ [M+H]⁺ 432.2022, found 432.2019.

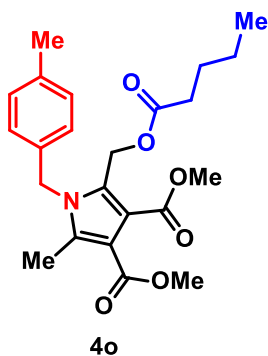


Prepared from 4-chlorobenzylamine, methyl acetoacetate and valeric acid following the general procedure to give the product as pale yellow oil (88% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.28 (d, 2H, $J = 8.17$ Hz), 6.82 (d, 2H, $J = 8.17$ Hz), 5.20 (s, 2H), 5.11 (s, 2H), 3.86 (s, 3H), 3.83 (s, 3H), 2.32 (s, 3H), 2.02 (t, 2H, $J = 7.65$ Hz), 1.41 (quint, 2H, $J = 7.55$ Hz), 1.21 (sext, 2H, $J = 7.55$ Hz), 0.85 (t, 3H, $J = 7.55$ Hz).

¹³C NMR (125 MHz, Chloroform-*d*): δ 173.14, 165.24, 165.12, 135.81, 134.48, 133.67, 129.50, 129.11, 126.87, 117.05, 113.14, 54.90, 51.90, 51.53, 46.83, 33.55, 26.68, 22.12, 13.59, 10.92.

HRMS m/z (ESI): calcd. for C₂₂H₂₇ClNO₆ [M+H]⁺ 436.1527, found 436.1524.

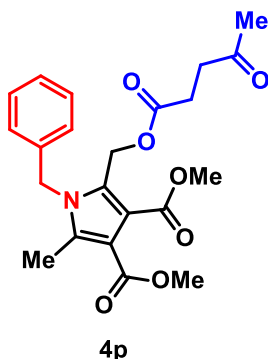


Prepared from 4-methylbenzylamine, methyl acetoacetate and valeric acid following the general procedure to give the product as pale yellow oil (80% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.10 (d, 2H, $J = 7.55$ Hz), 6.77 (d, 2H, $J = 7.55$ Hz), 5.20 (s, 2H), 5.10 (s, 2H), 3.86 (s, 3H), 3.83 (s, 3H), 2.33 (s, 3H), 2.32 (s, 3H), 2.04 (t, 2H, $J = 7.55$ Hz), 1.43 (quint, 2H, $J = 7.55$ Hz), 1.22 (sext, 2H, $J = 7.55$ Hz), 0.84 (t, 3H, $J = 7.55$ Hz).

¹³C NMR (125 MHz, Chloroform-*d*): δ 173.18, 165.38, 165.25, 137.43, 136.07, 132.84, 129.56, 125.41, 116.77, 112.81, 55.04, 51.83, 51.46, 47.23, 33.57, 26.68, 22.14, 20.97, 13.59, 10.96.

HRMS m/z (ESI): calcd. for C₂₃H₃₀NO₆ [M+H]⁺ 416.2073, found 416.2076.

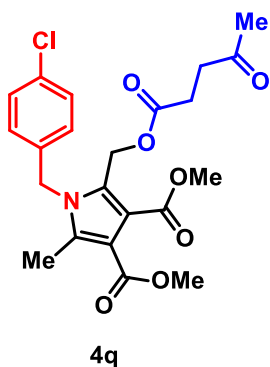


Prepared from benzylamine, methyl acetoacetate and levulinic acid following the general procedure to give the product as pale yellow oil (81% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.32–7.24 (m, 3H), 6.88 (d, 2H, $J = 6.92$ Hz), 5.20 (s, 2H), 5.16 (s, 2H), 3.85 (s, 3H), 3.82 (s, 3H), 2.58 (t, 2H, $J = 6.92$ Hz), 2.34 (s, 3H), 2.29 (t, 2H, $J = 6.92$ Hz), 2.12 (s, 3H).

¹³C NMR (125 MHz, Chloroform-*d*): δ 206.37, 172.09, 165.30, 165.17, 136.13, 135.93, 129.28, 128.91, 127.67, 125.46, 116.92, 112.84, 55.42, 51.84, 51.46, 47.35, 37.72, 29.72, 27.56, 10.93.

HRMS m/z (ESI): calcd. for C₂₂H₂₆NO₇ [M+H]⁺ 416.1709, found 416.1710.

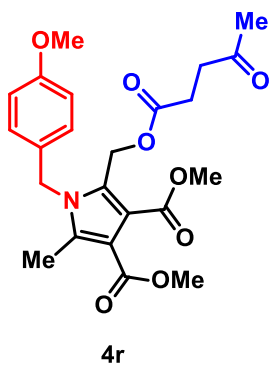


Prepared from 4-chlorobenzylamine, methyl acetoacetate and levulinic acid following the general procedure to give the product as pale yellow oil (79% yield).

$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 7.26 (d, 2H, $J = 8.17$ Hz), 6.82 (d, 2H, $J = 8.80$ Hz), 5.17 (s, 2H), 5.12 (s, 2H), 3.83 (s, 3H), 3.81 (s, 3H), 2.60 (t, 2H, $J = 6.92$ Hz), 2.31–2.29 (s+m, 5H), 2.12 (s, 3H).

$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 206.40, 172.06, 165.17, 165.05, 135.91, 134.50, 133.55, 129.11, 129.04, 126.92, 117.03, 113.00, 55.34, 51.84, 51.47, 46.75, 37.66, 29.65, 27.53, 10.87.

HRMS m/z (ESI): calcd. for $\text{C}_{22}\text{H}_{25}\text{ClNO}_7$ $[\text{M}+\text{H}]^+$ 450.1320, found 450.1317.



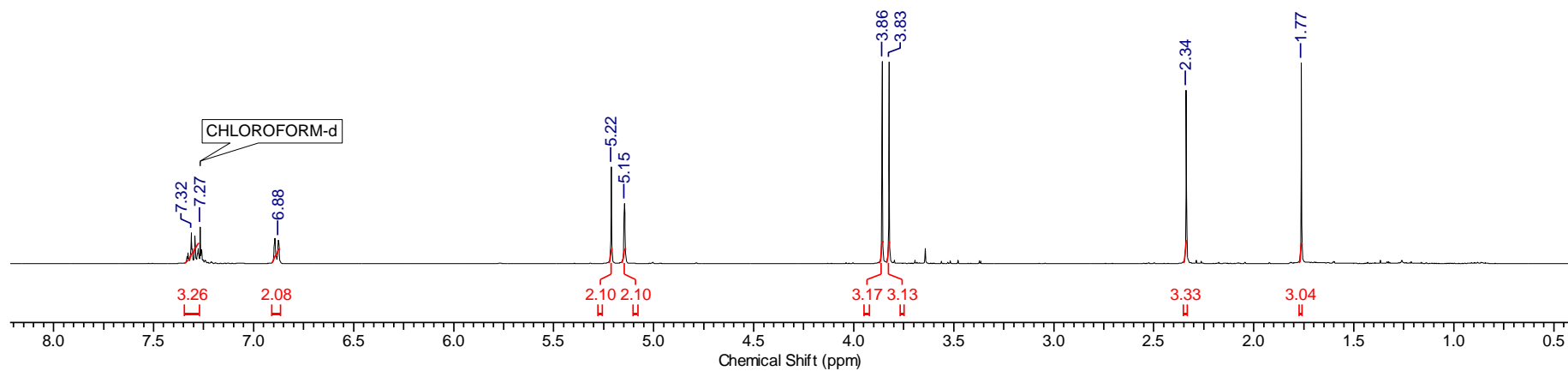
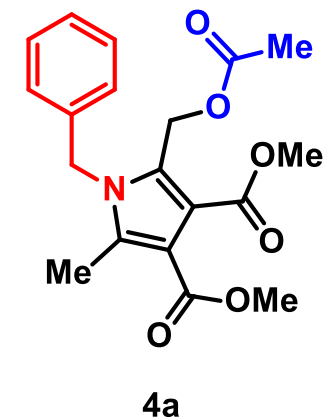
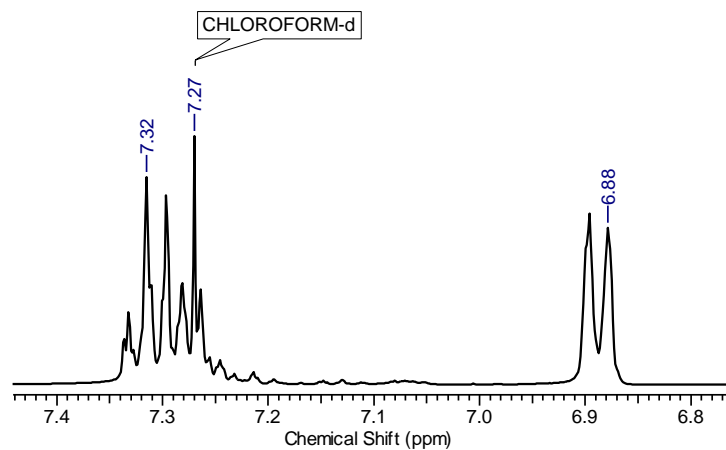
Prepared from 4-methoxybenzylamine, methyl acetoacetate and levulinic acid following the general procedure to give the product as pale yellow oil (78% yield).

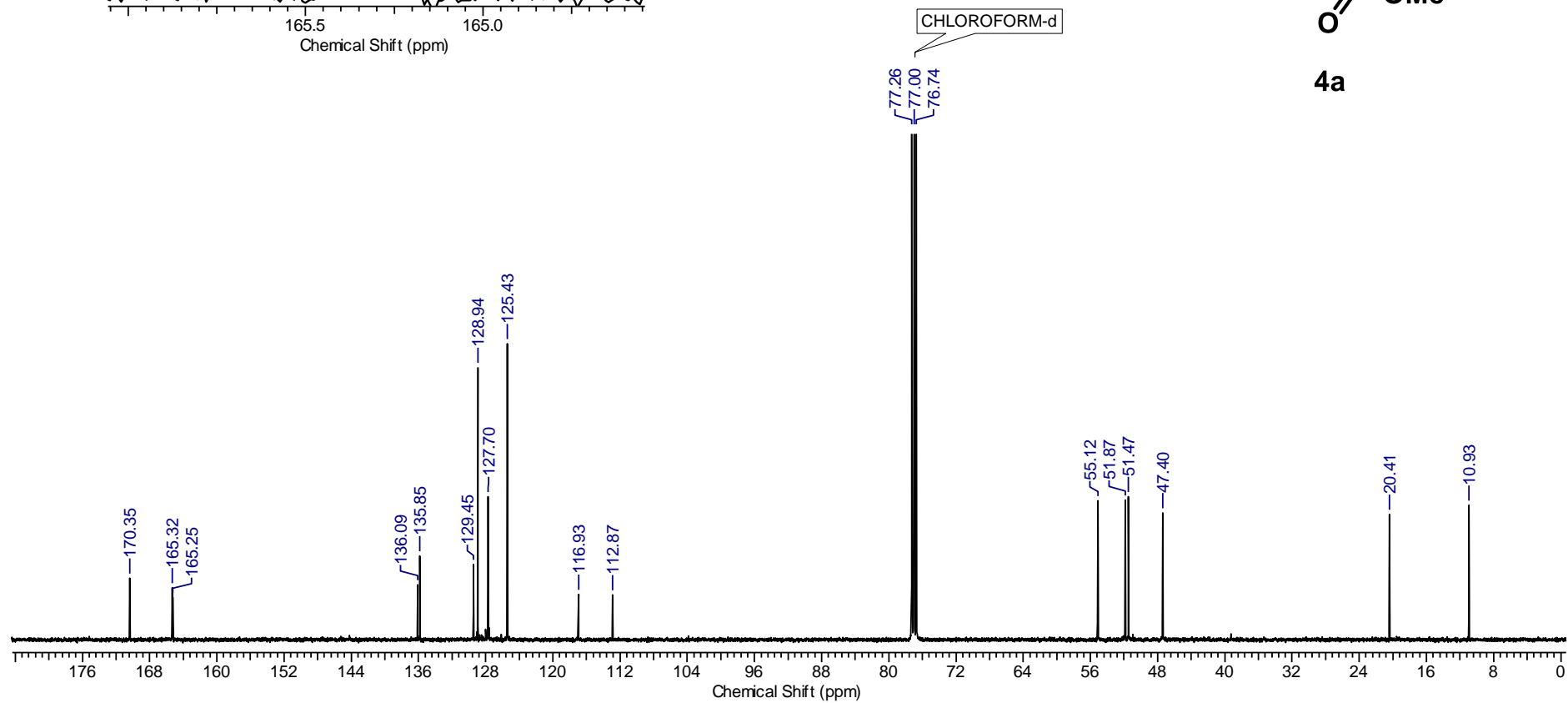
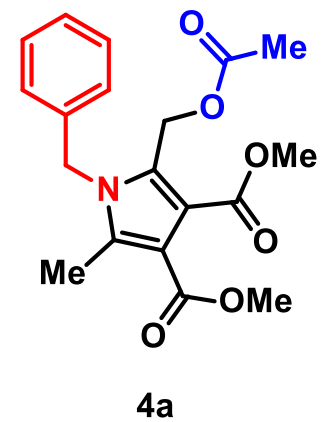
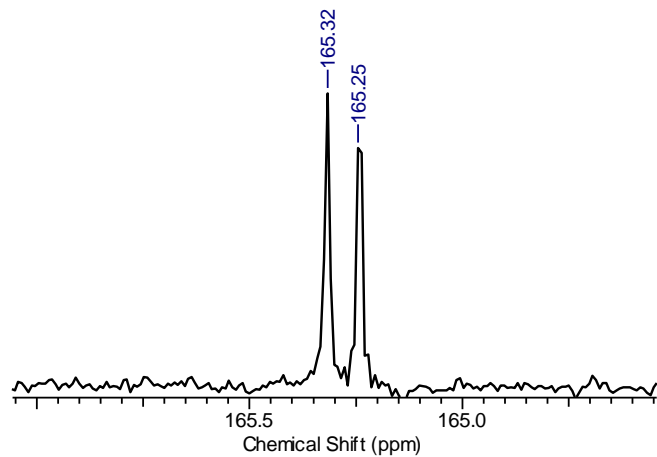
$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 6.83 (s, 4H), 5.20 (s, 2H), 5.09 (s, 2H), 3.85 (s, 3H), 3.82 (s, 3H), 3.77 (s, 3H), 2.61 (t, 2H, $J = 6.29$ Hz), 2.36–2.33 (s+m, 5H), 2.13 (s, 3H).

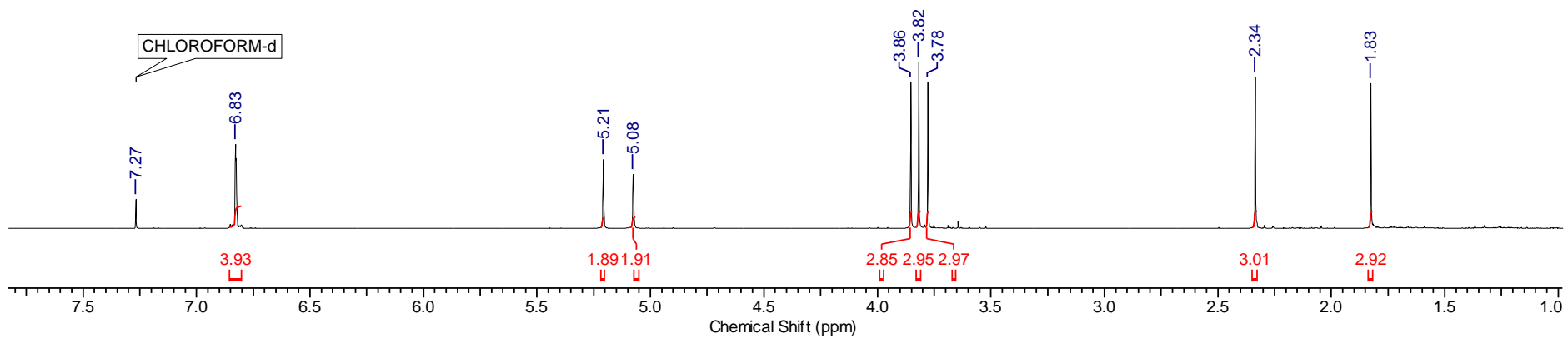
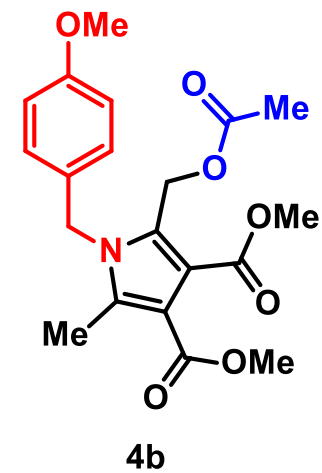
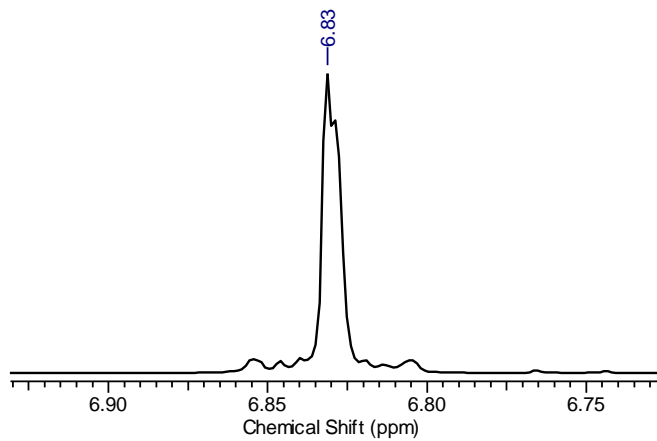
$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 206.42, 172.12, 165.35, 165.22, 159.09, 136.12, 129.23, 127.84, 126.79, 116.87, 114.30, 112.80, 55.48, 55.28, 51.86, 51.46, 46.91, 37.77, 29.72, 27.64, 10.98.

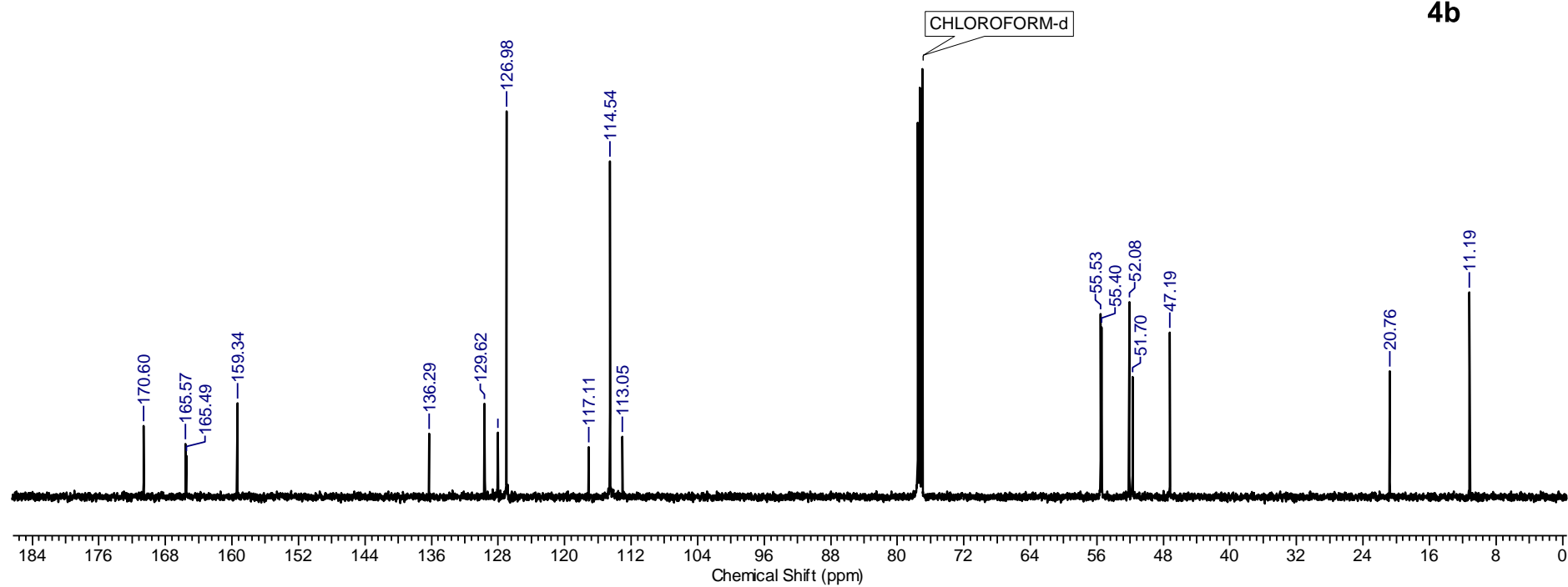
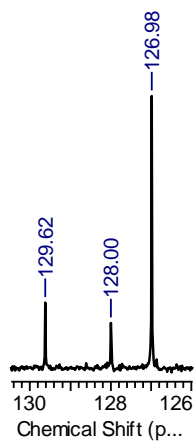
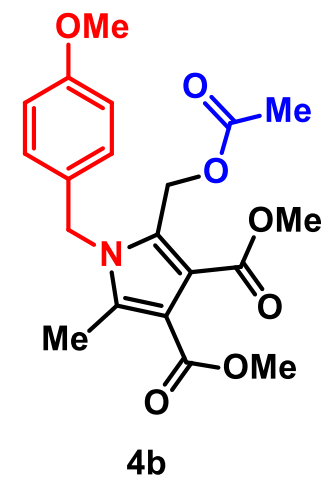
HRMS m/z (ESI): calcd. for $\text{C}_{23}\text{H}_{28}\text{NO}_8$ $[\text{M}+\text{H}]^+$ 446.1815, found 446.1817.

8. Spectral Data



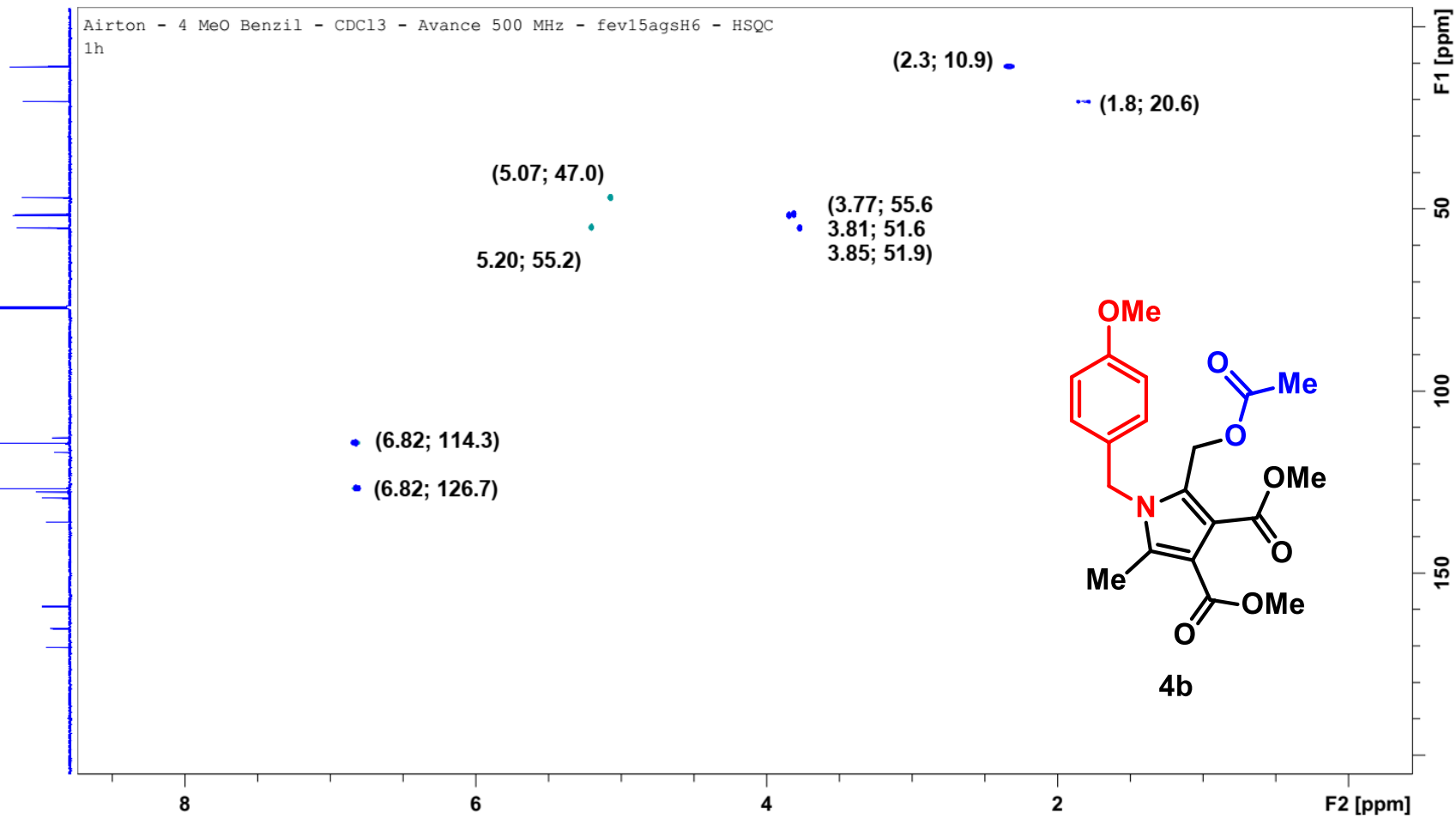






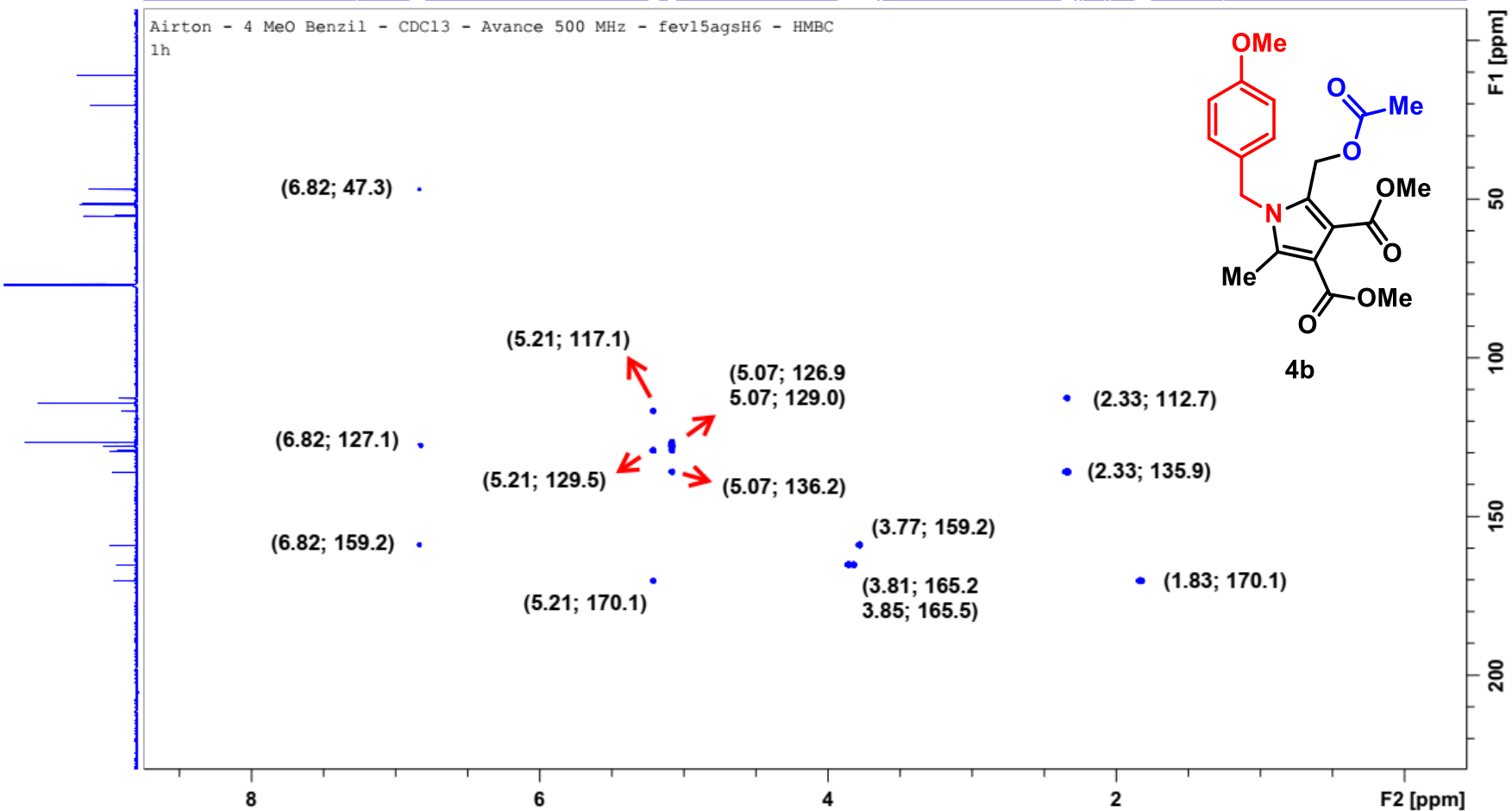
"fev15agsH6_4MeO benzilamina com MAA e acido acetico" 4 1 C:\Bruker\TopSpin4.0.7\examdata

Airton - 4 MeO Benzil - CDCl3 - Avance 500 MHz - fev15agsH6 - HSQC
1h

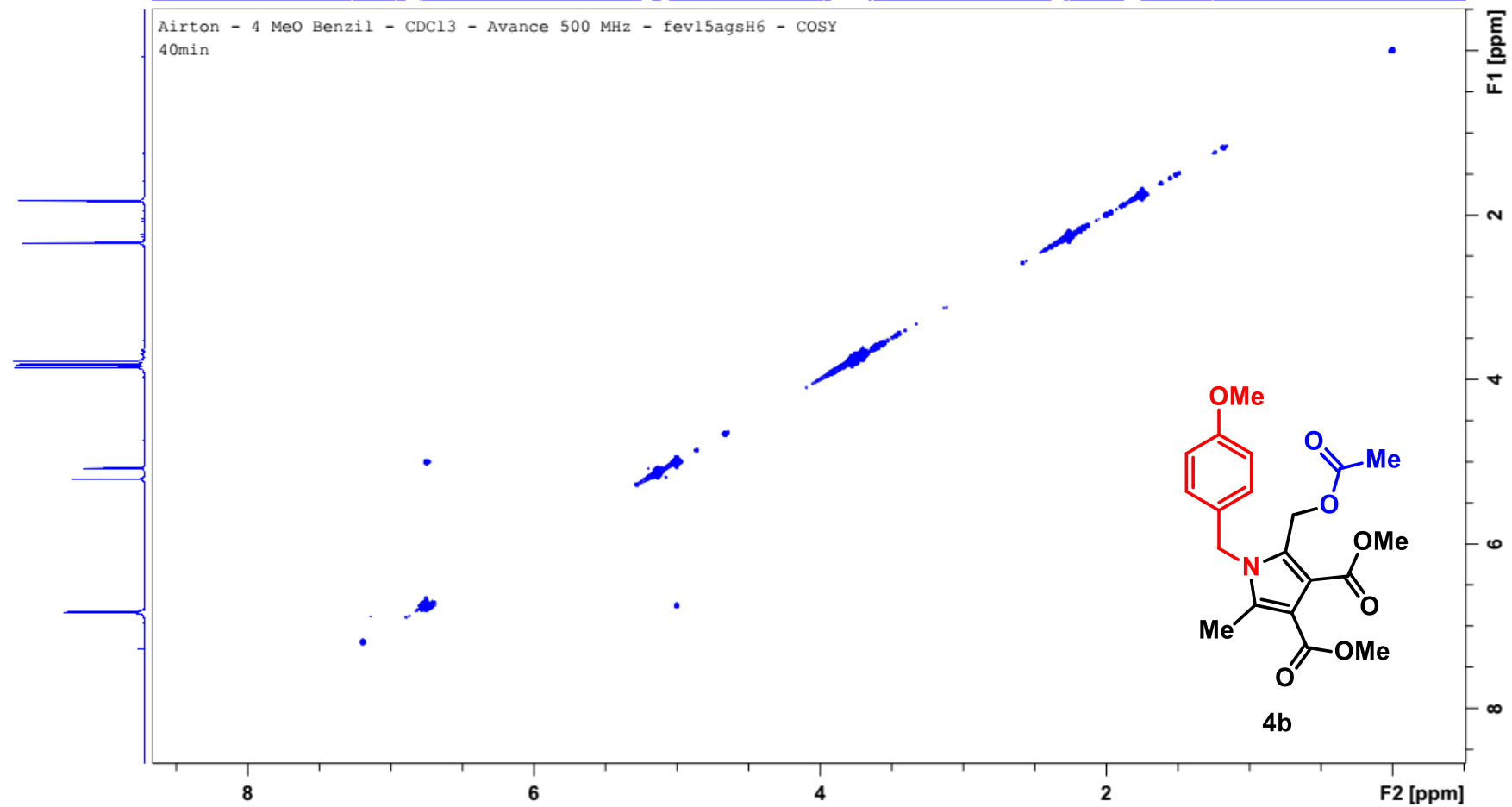


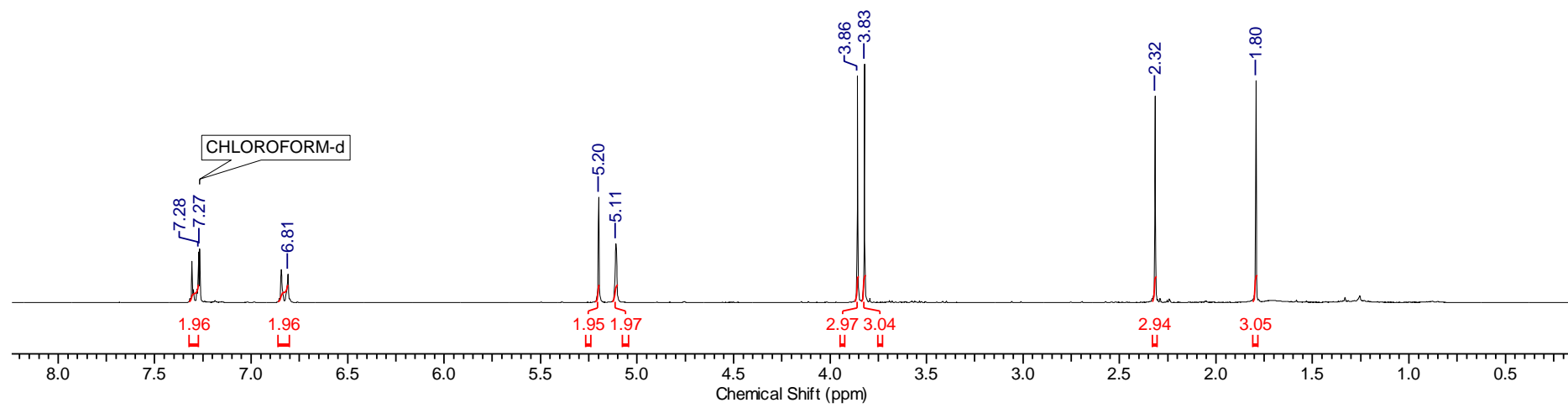
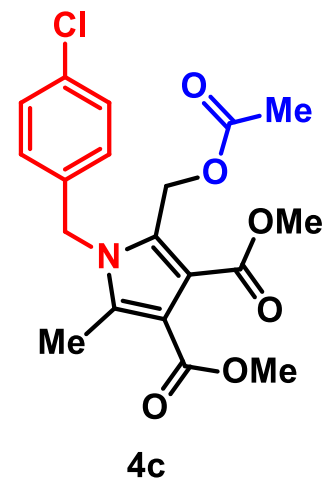
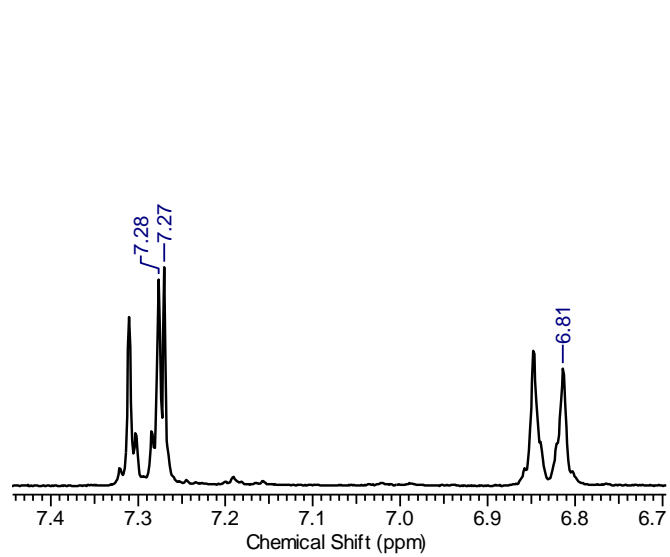
"fev15agsH6_4MeO benzilamina com MAA e acido acetico" 5 1 C:\Bruker\TopSpin4.0.7\examdata

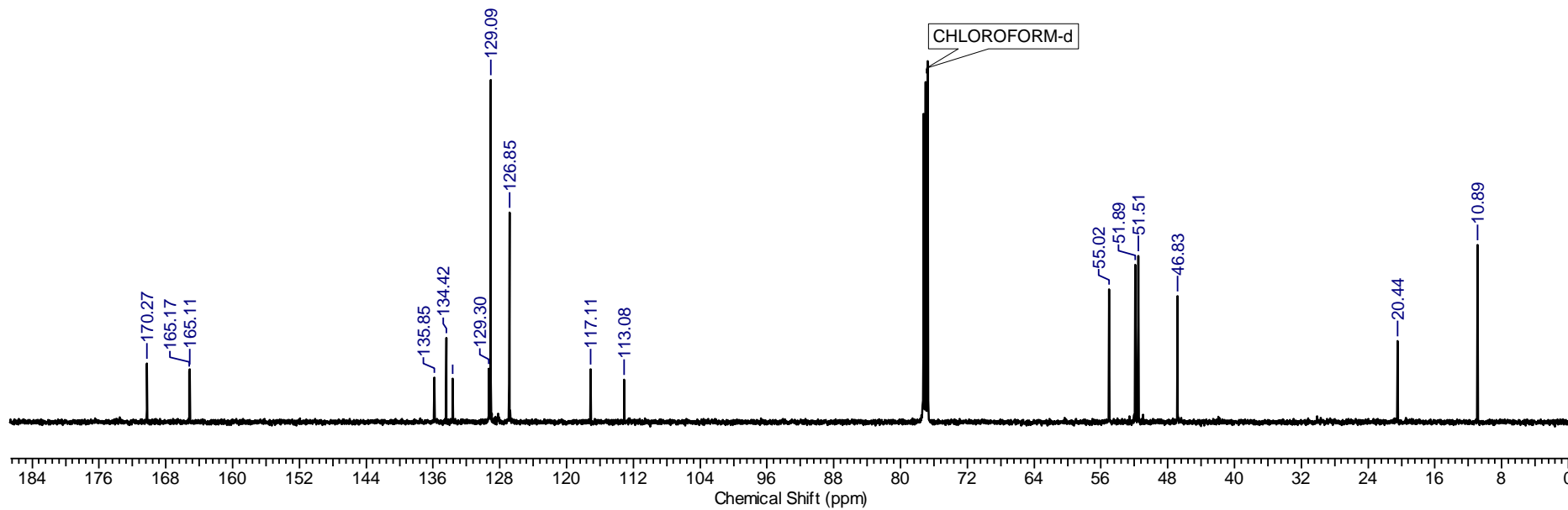
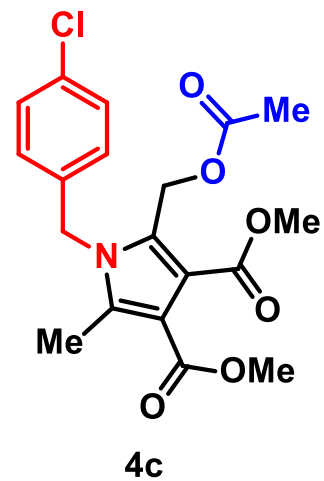
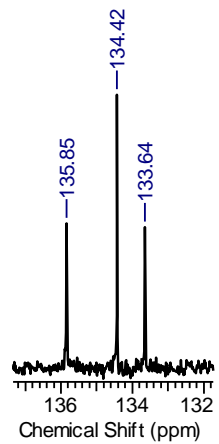
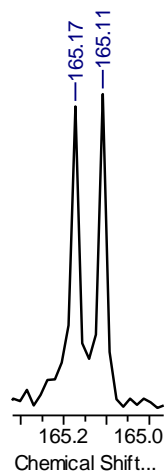
Airton - 4 MeO Benzil - CDC13 - Avance 500 MHz - fev15agsH6 - HMBC
1h

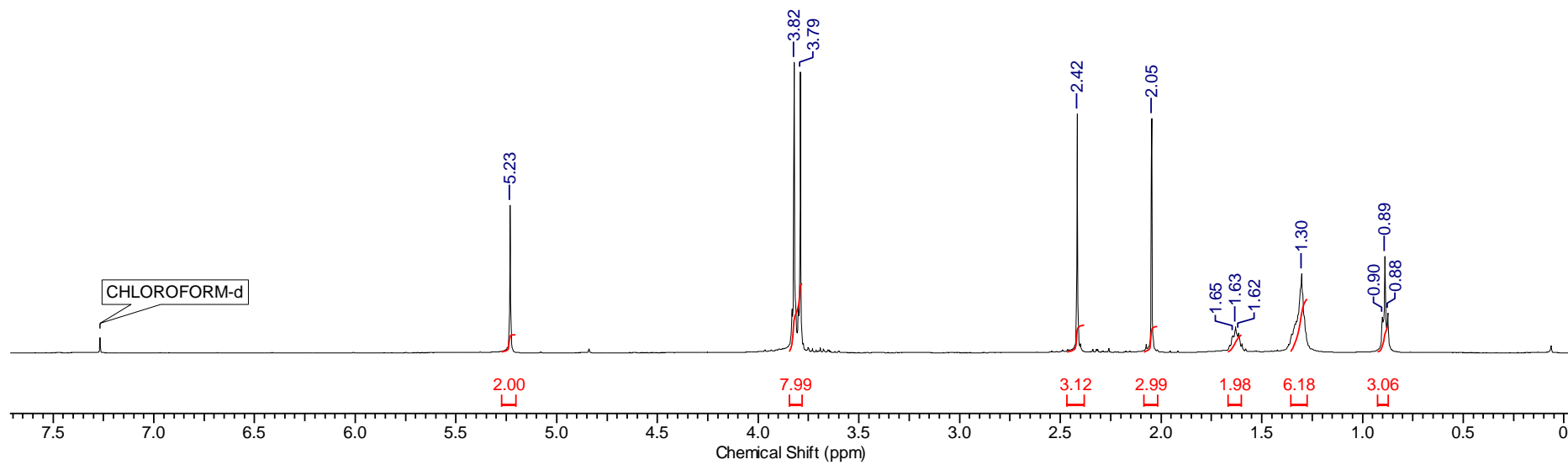
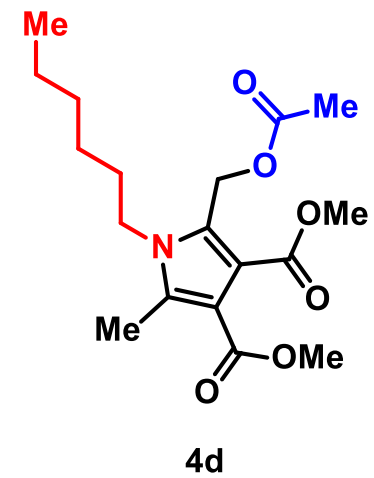
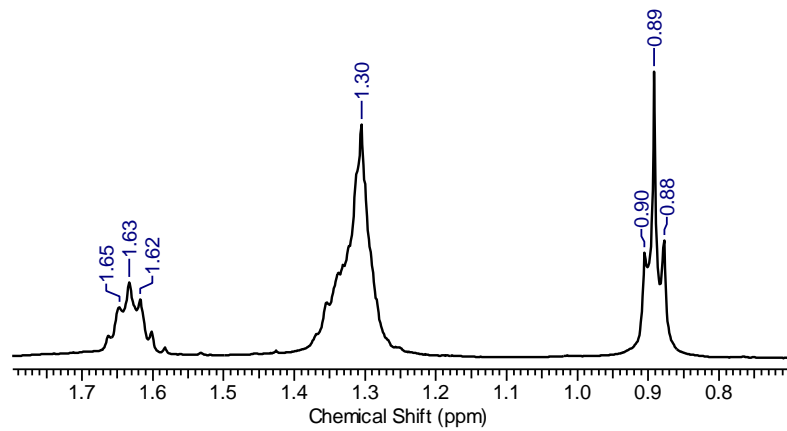


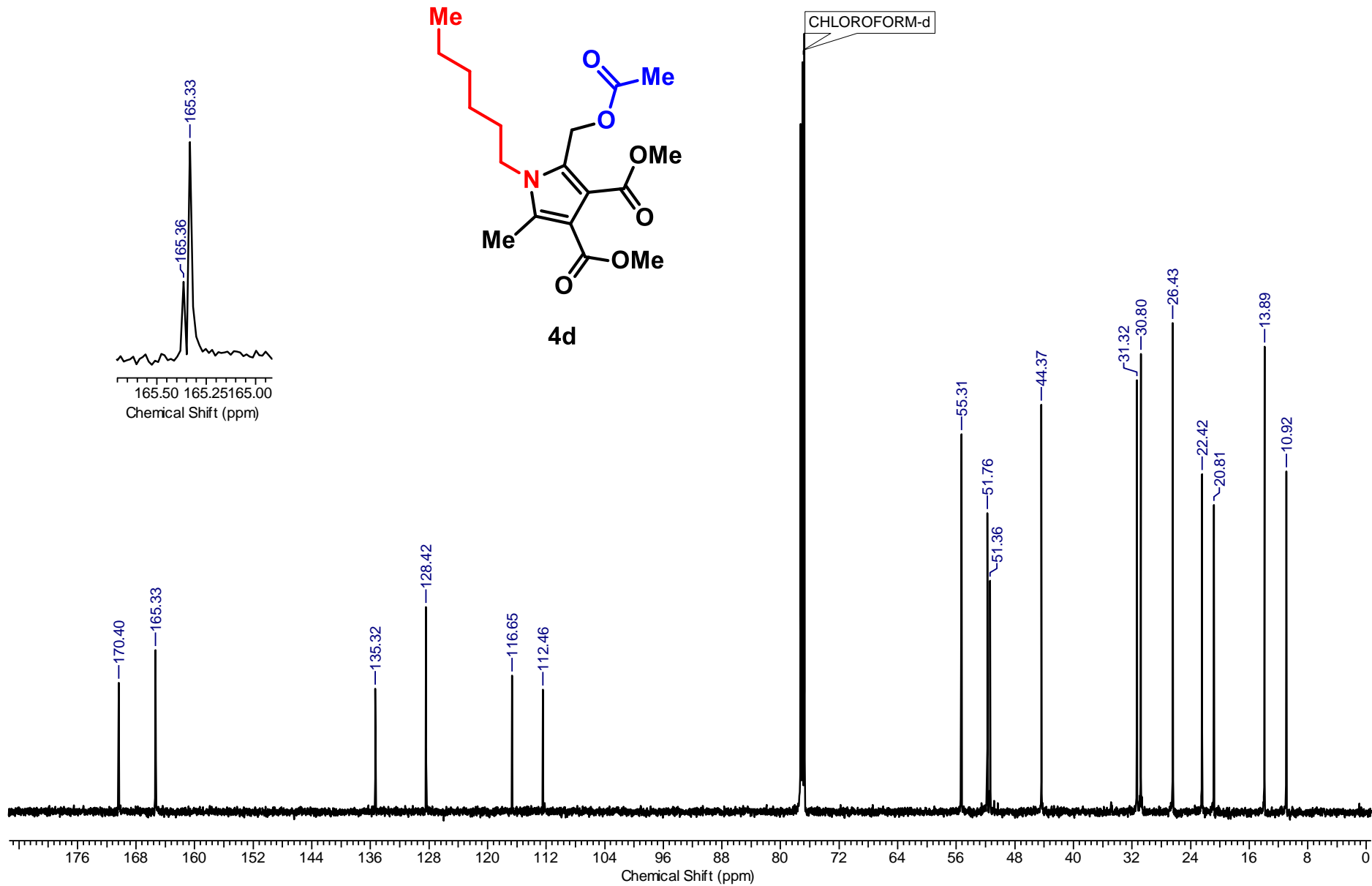
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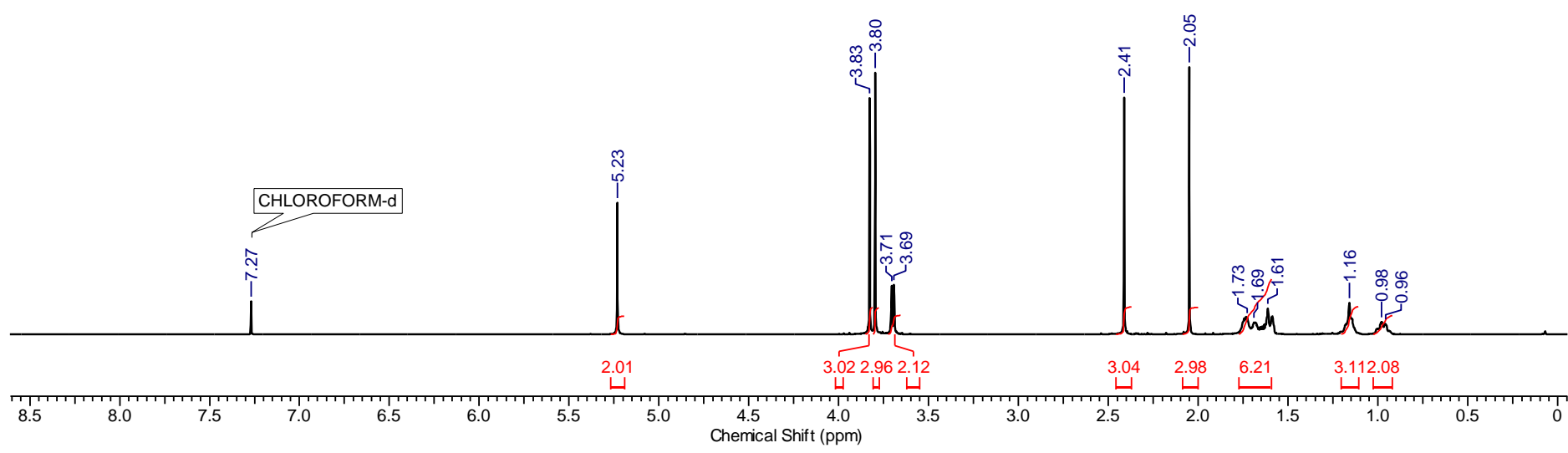
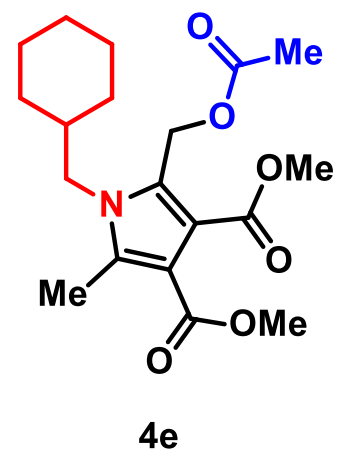
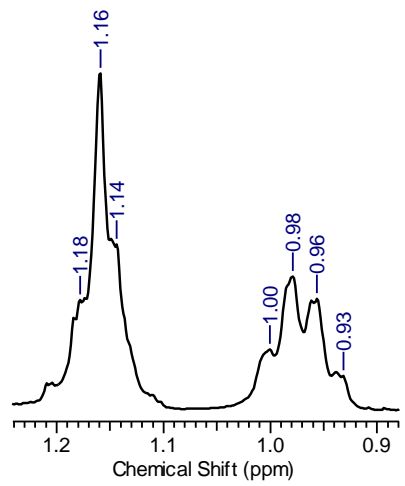
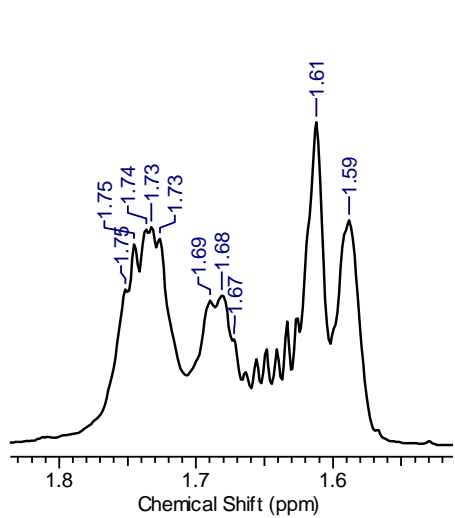


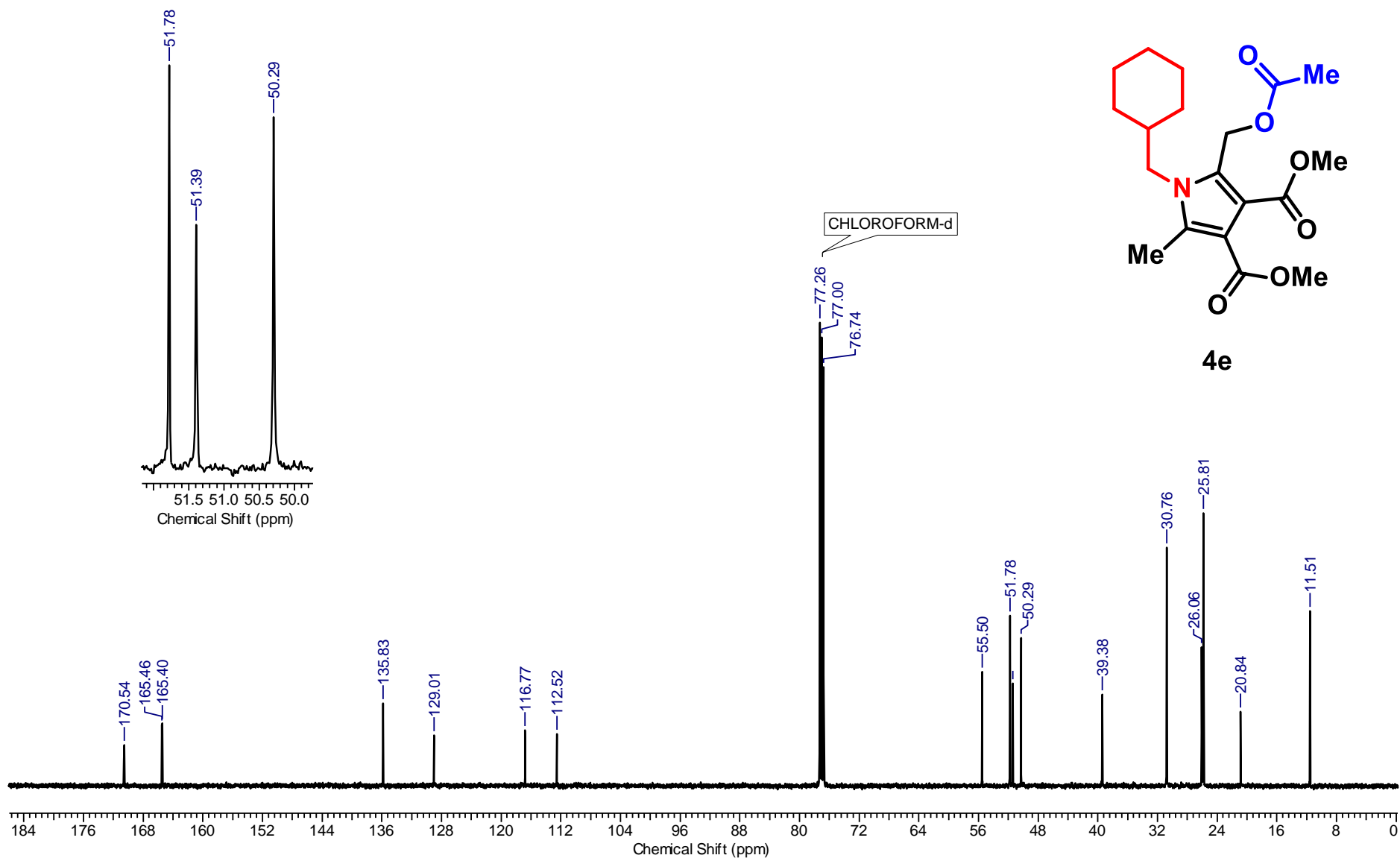


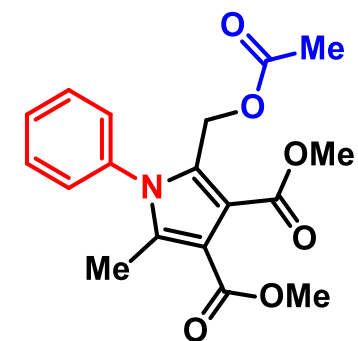
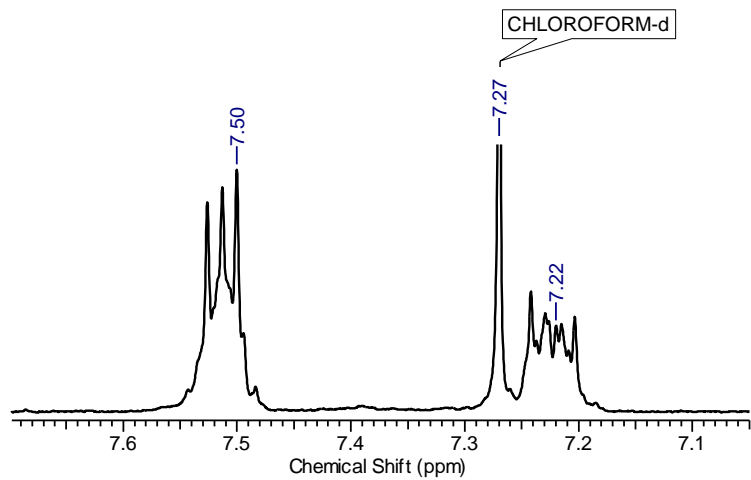




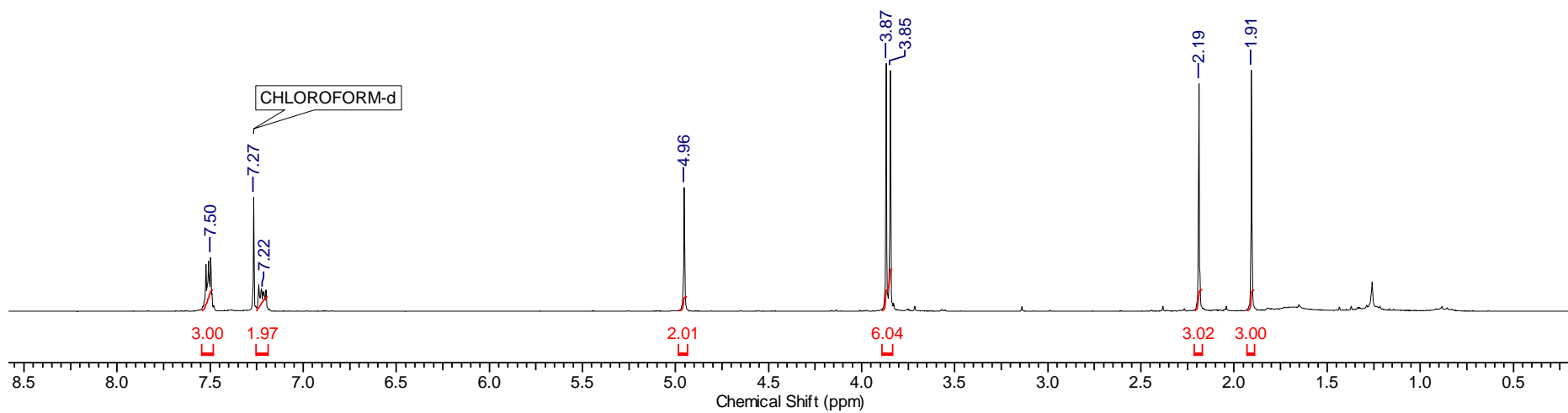


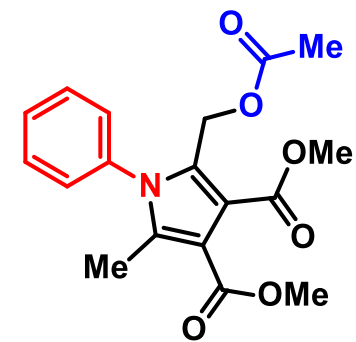




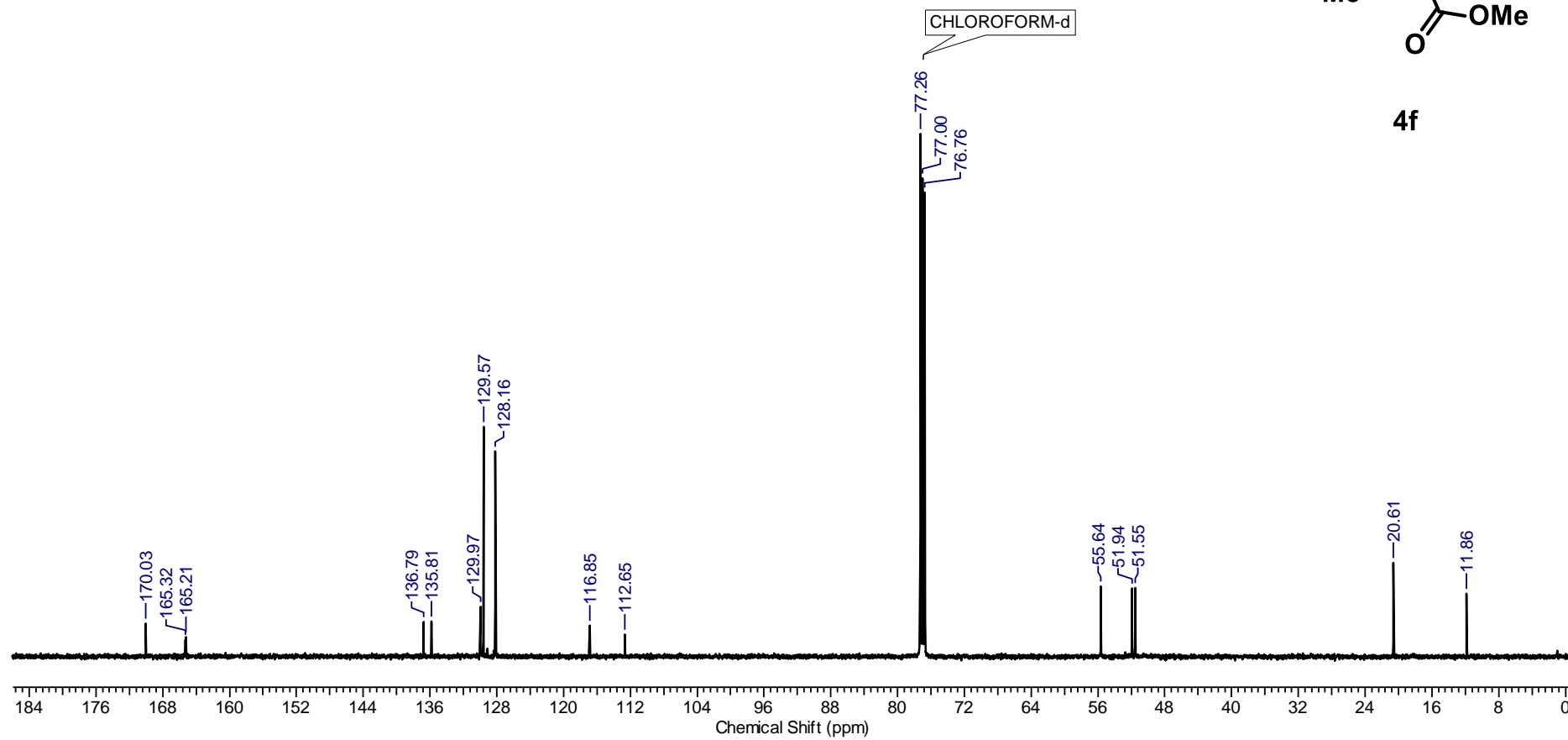


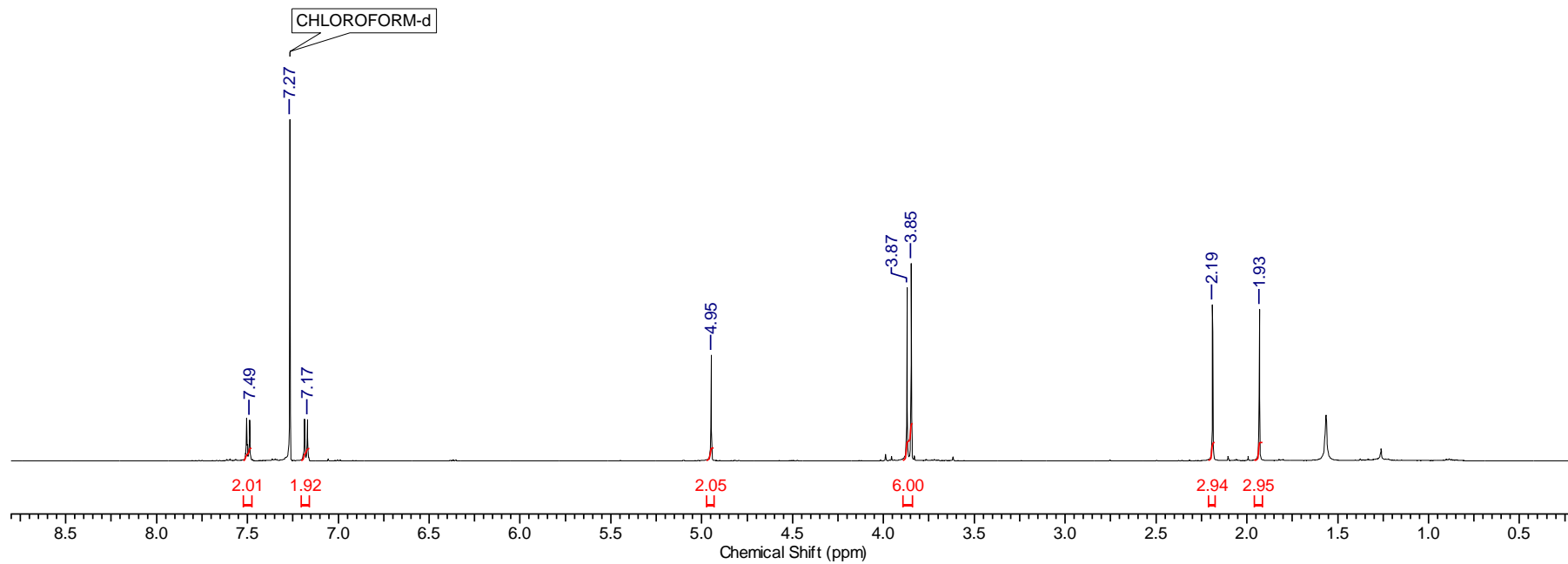
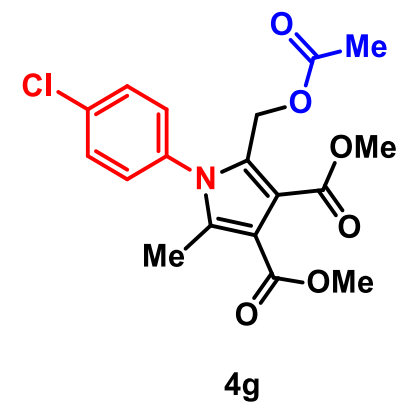
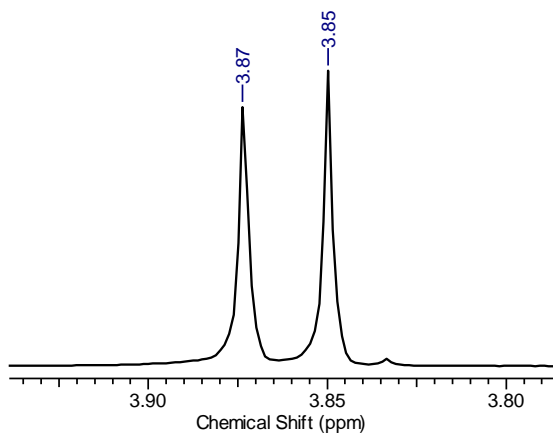
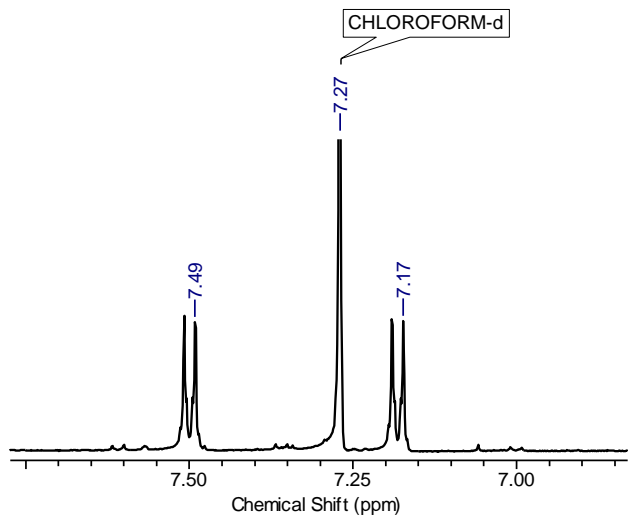
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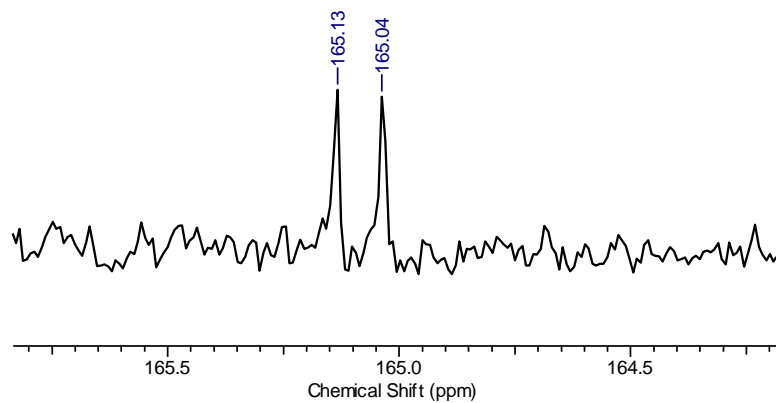




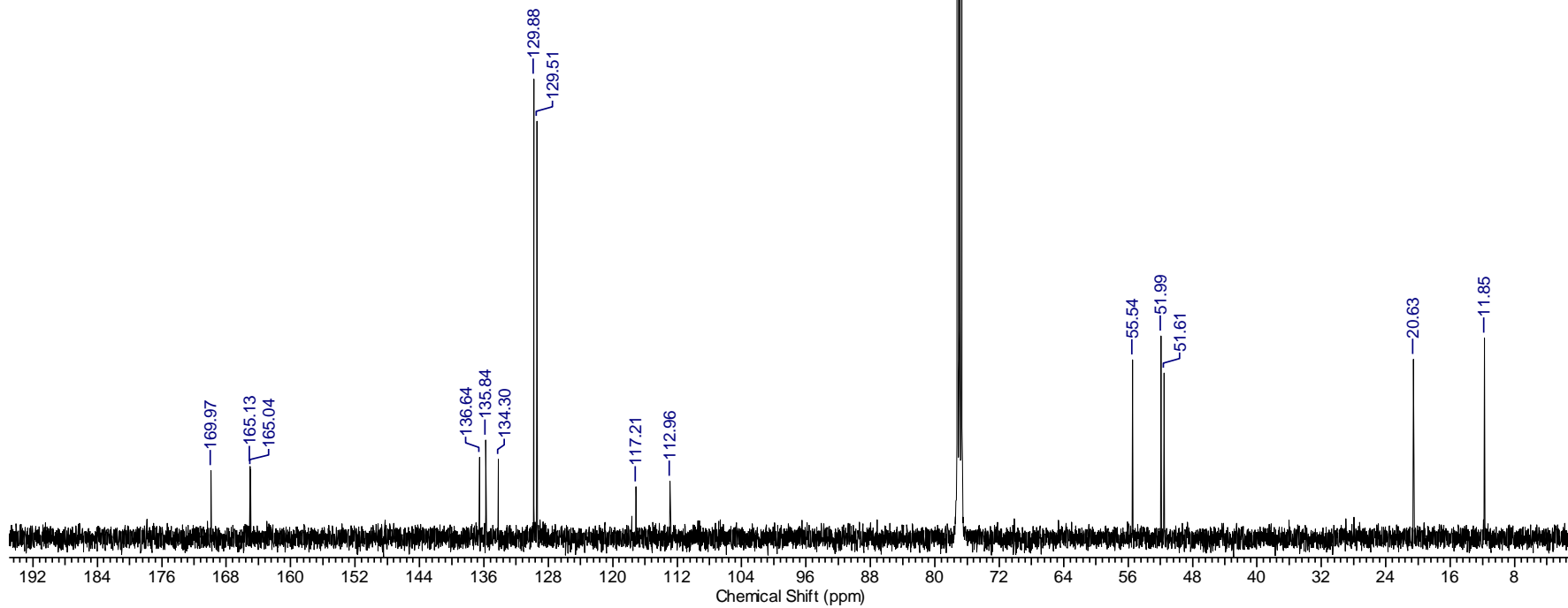
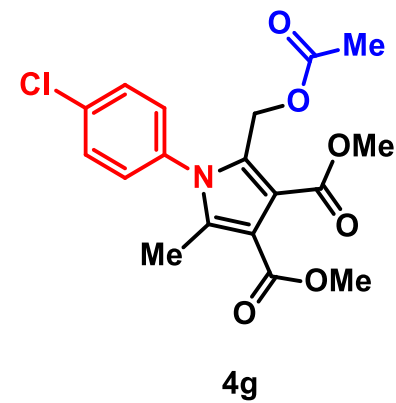
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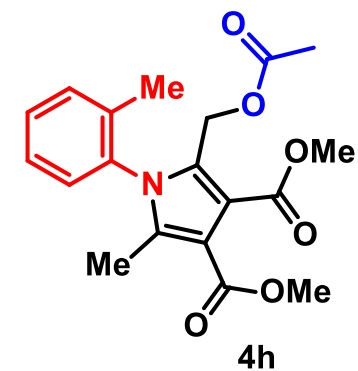
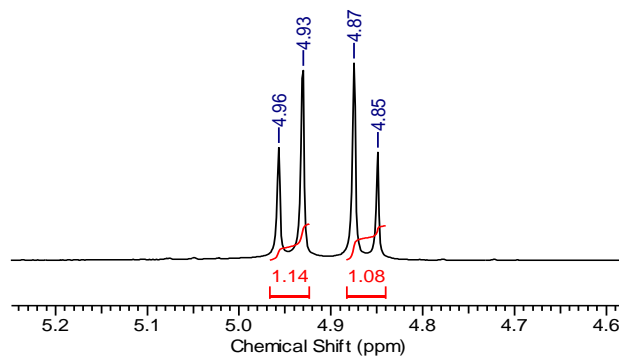
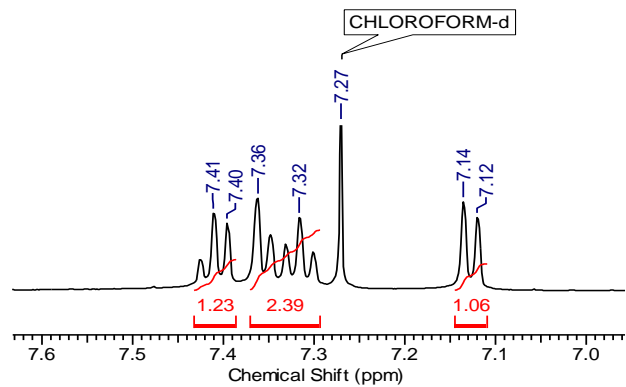




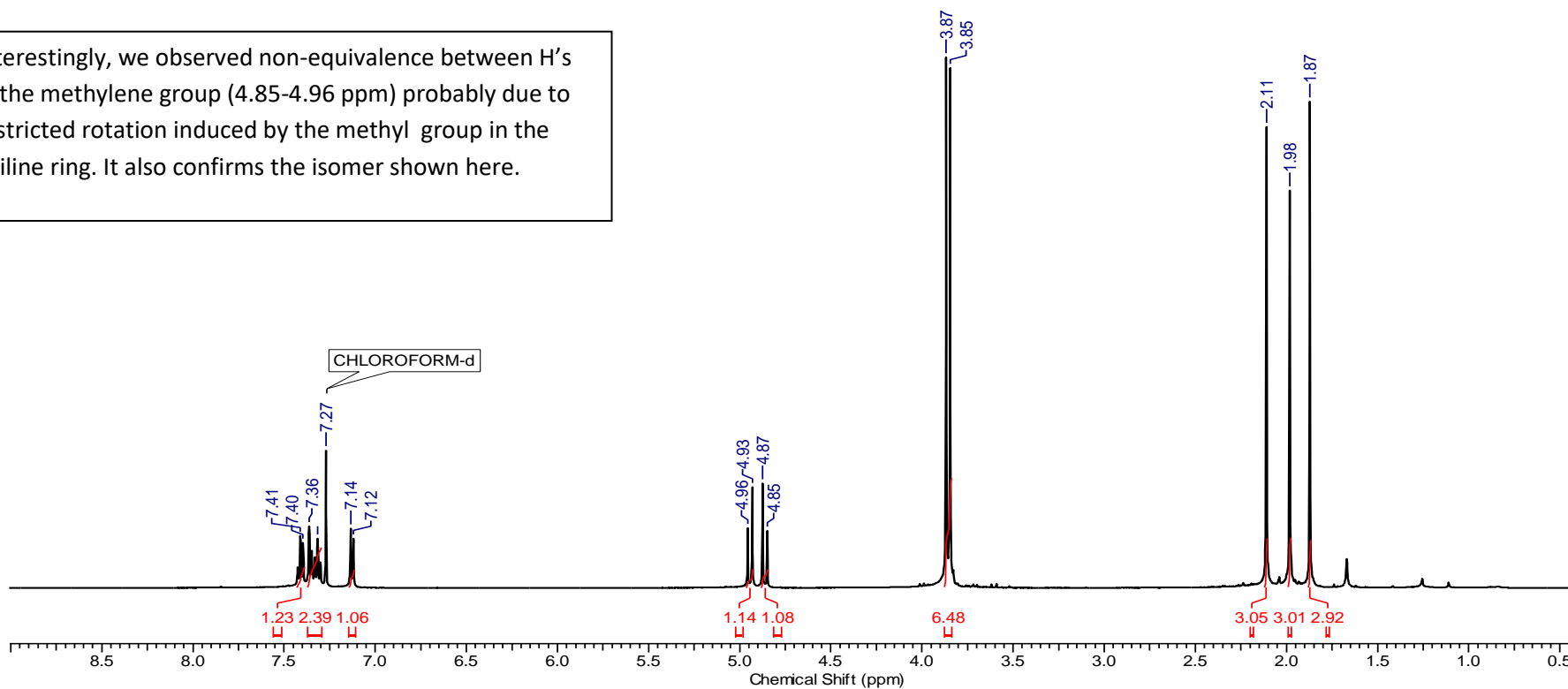


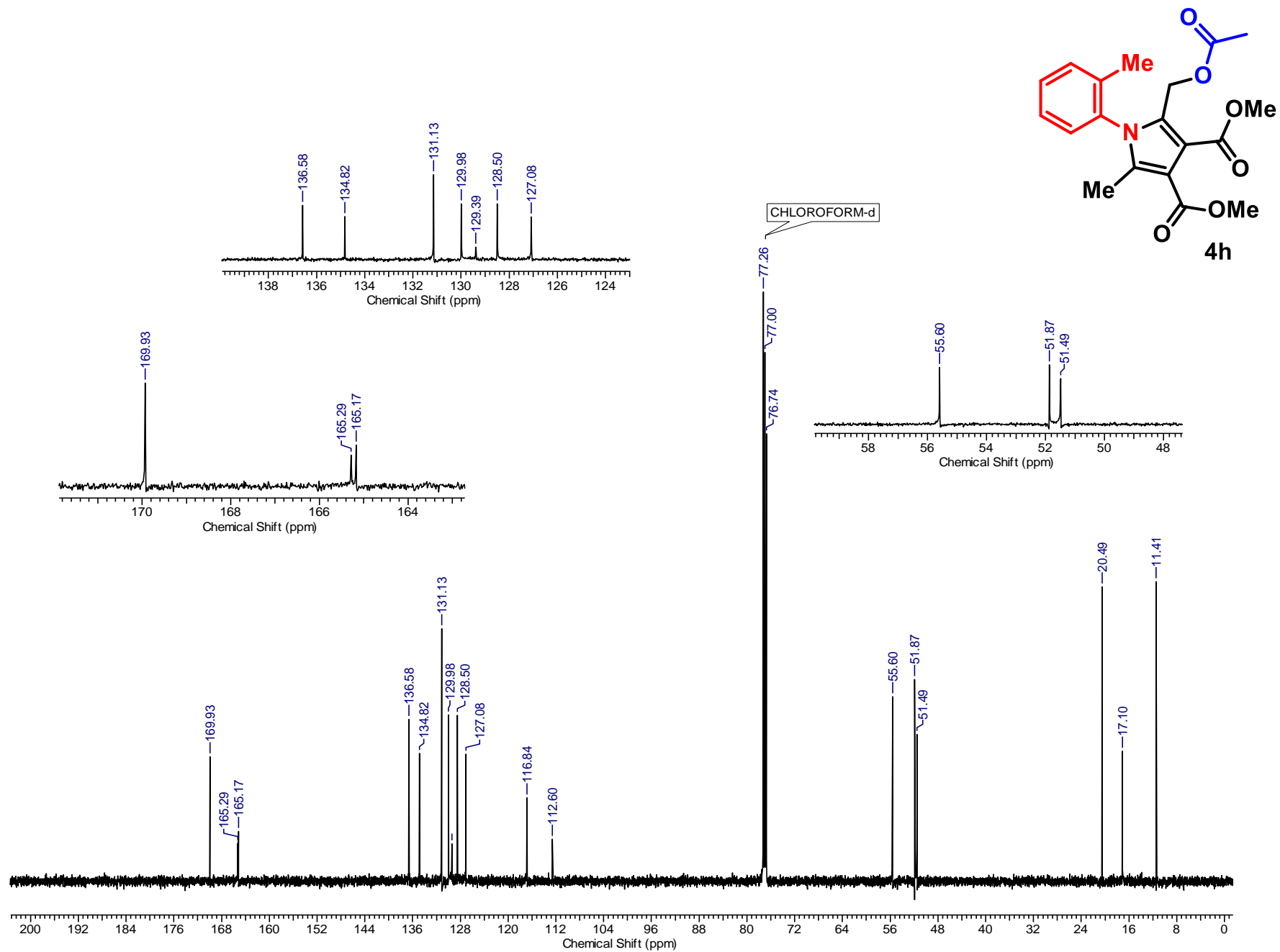
CHLOROFORM-d
77.26
77.00
76.74





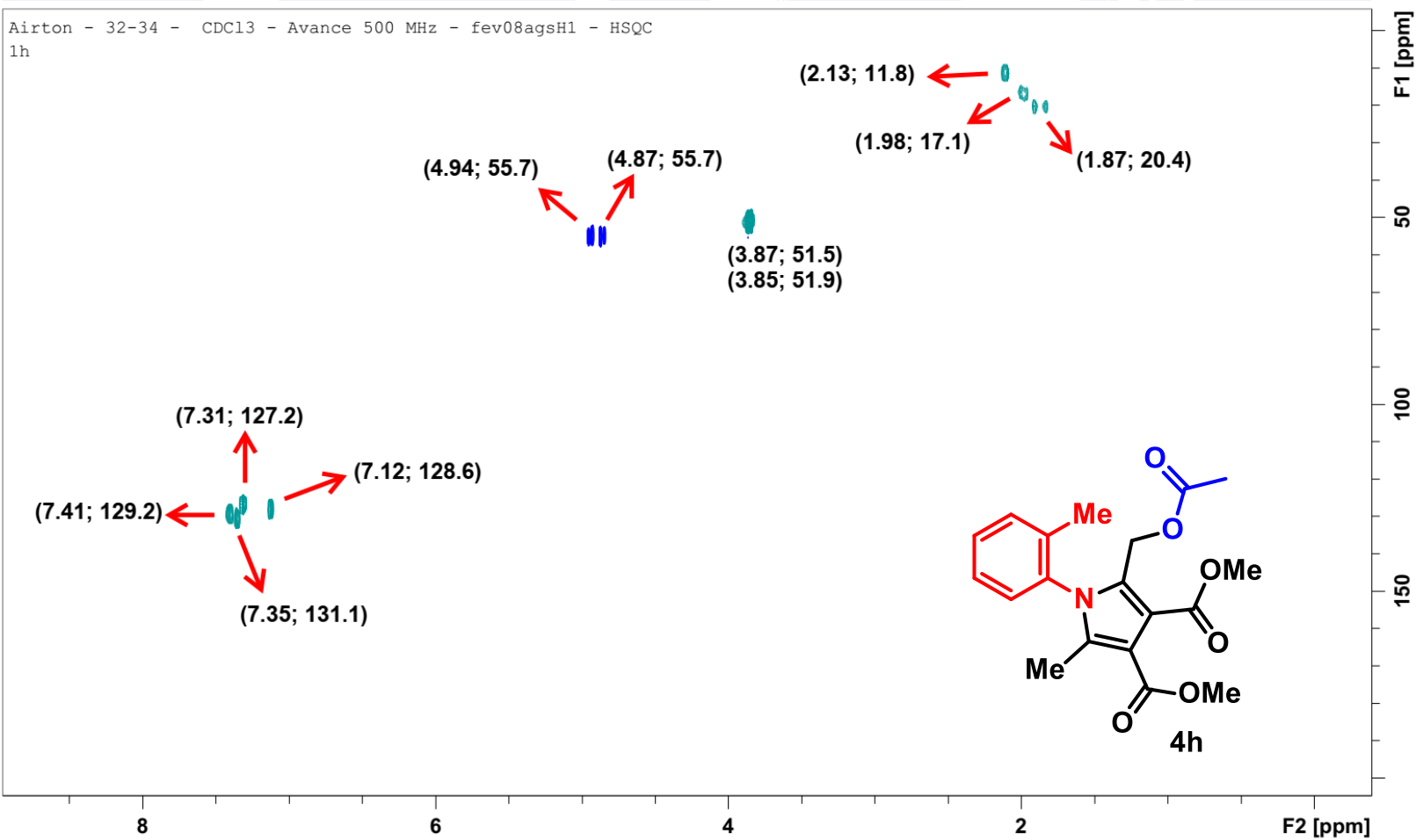
Interestingly, we observed non-equivalence between H's in the methylene group (4.85-4.96 ppm) probably due to restricted rotation induced by the methyl group in the aniline ring. It also confirms the isomer shown here.





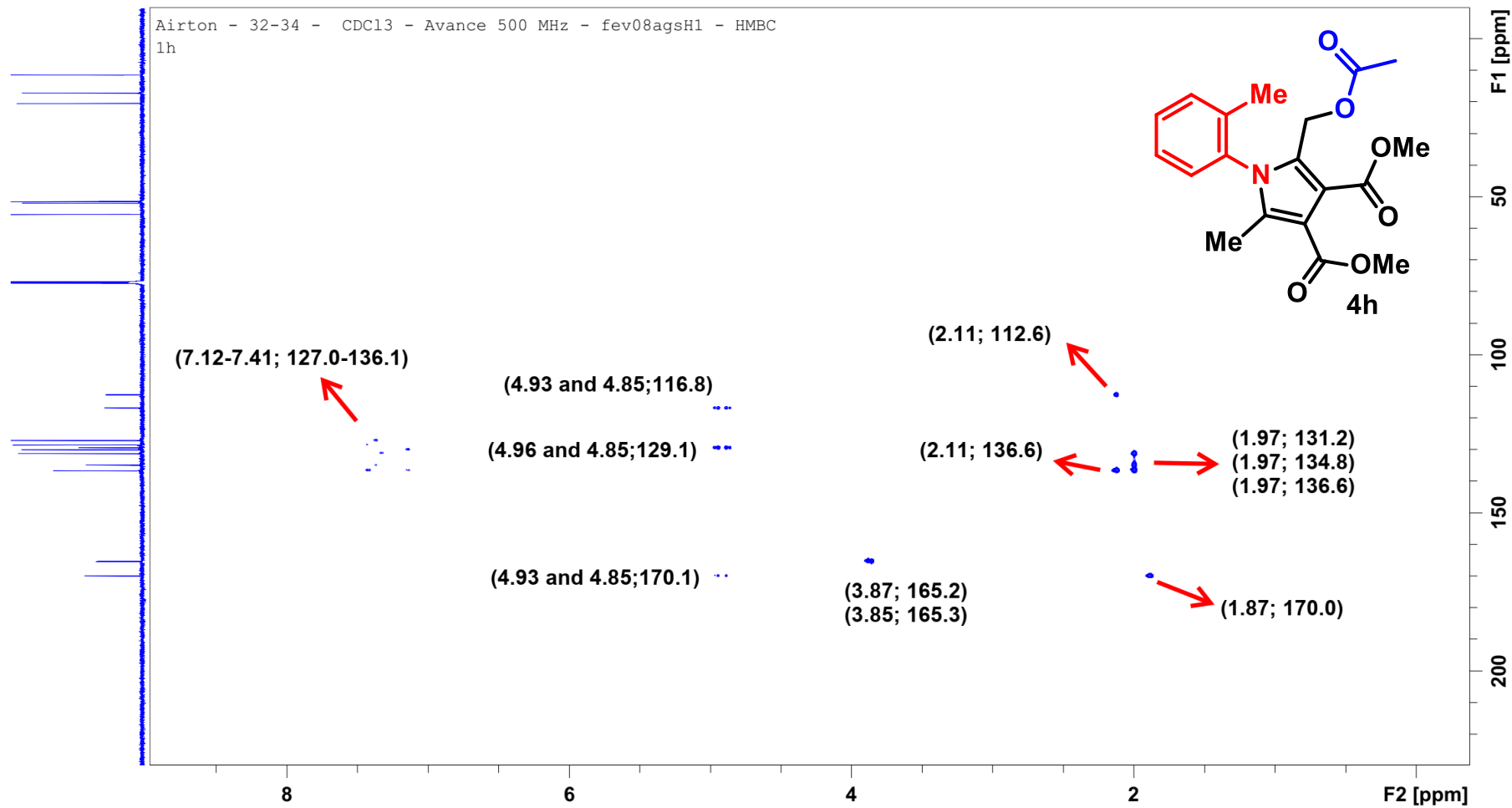
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Airton - 32-34 - CDC13 - Avance 500 MHz - fev08agsH1 - HSQC
1h

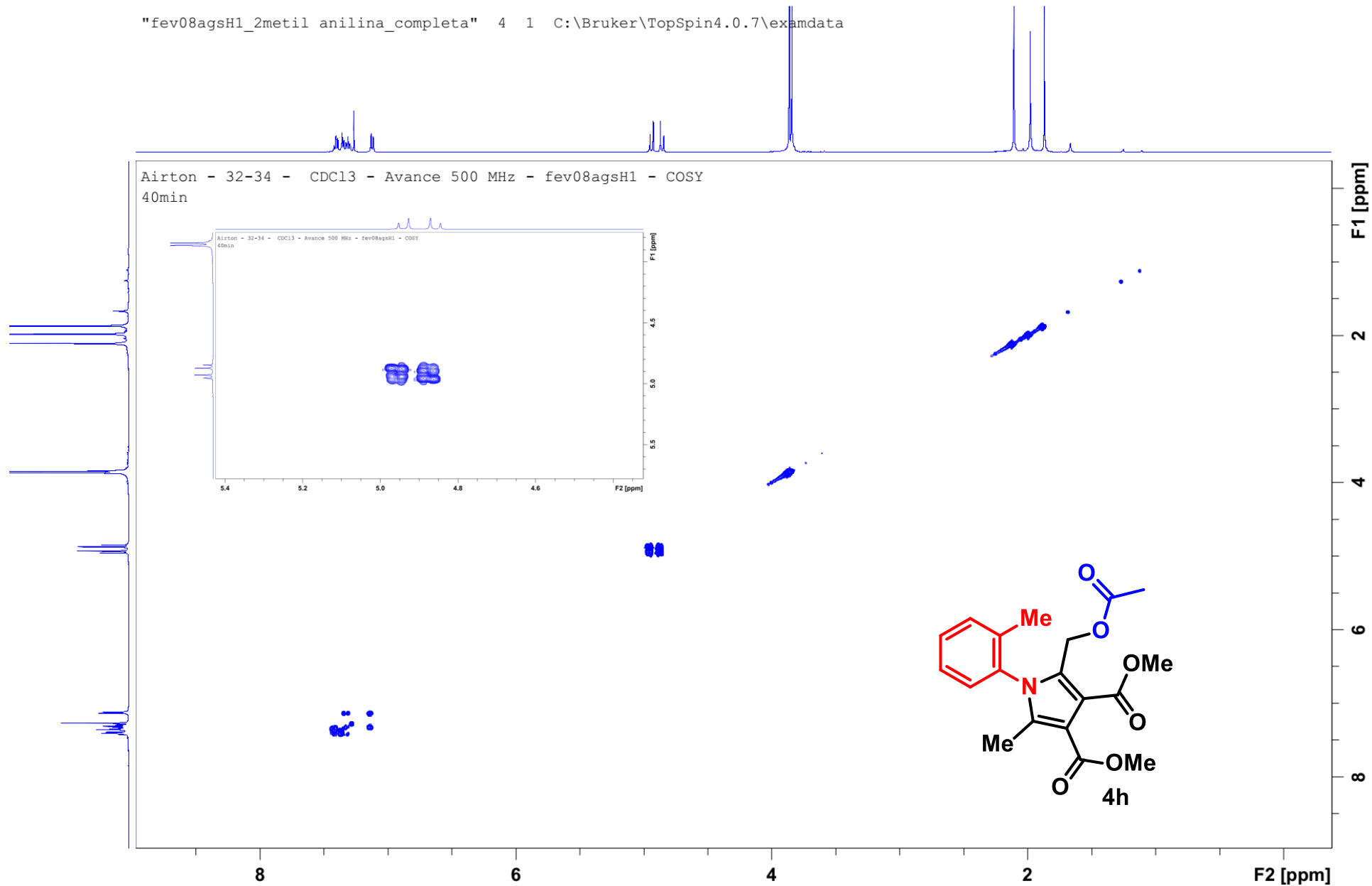


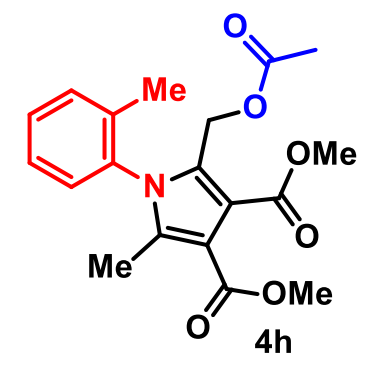
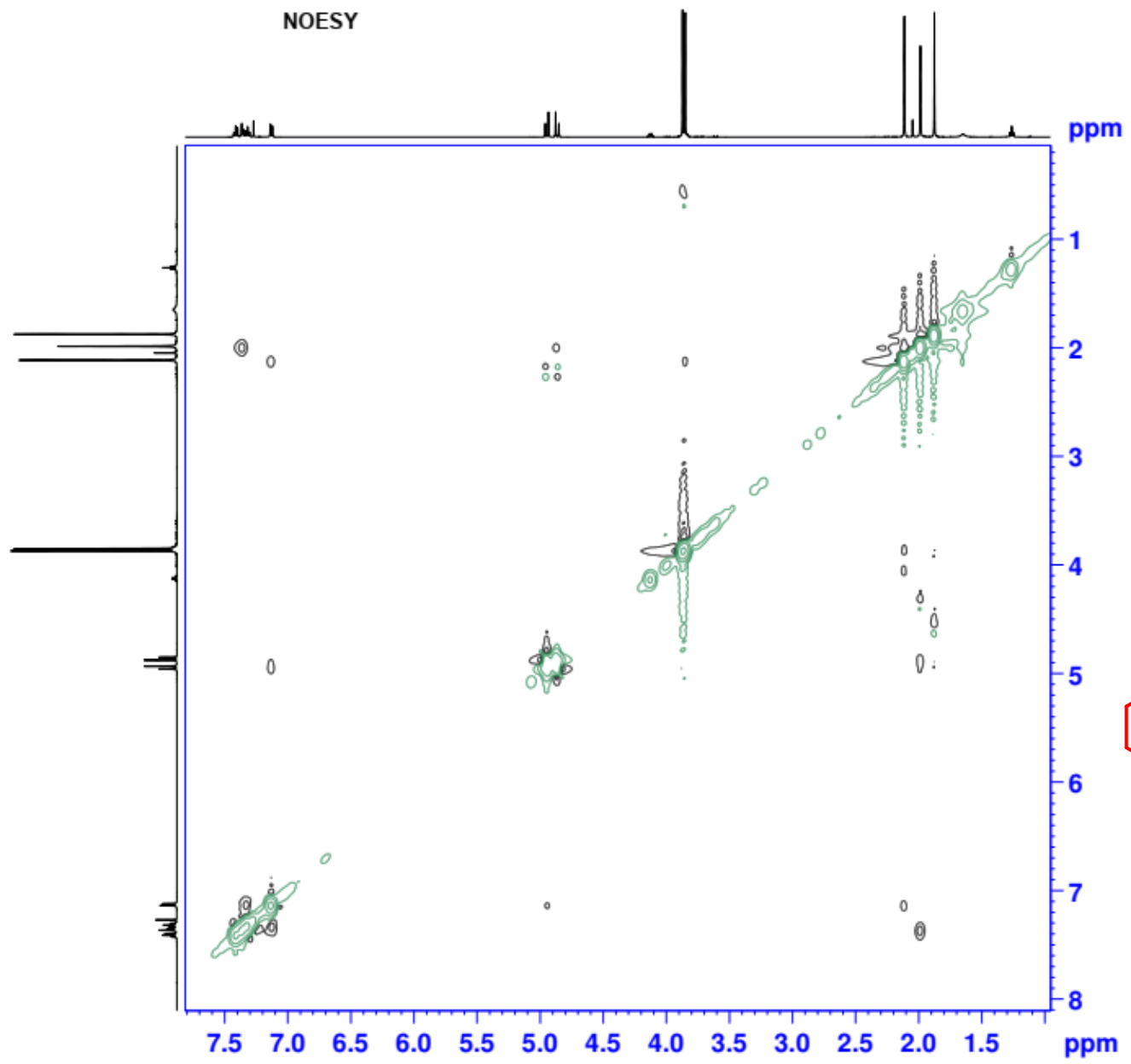
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Airton - 32-34 - CDCl3 - Avance 500 MHz - fev08agsH1 - HMBC
1h



"fev08agsH1_2metil anilina_completa" 4 1 C:\Bruker\TopSpin4.0.7\examdata

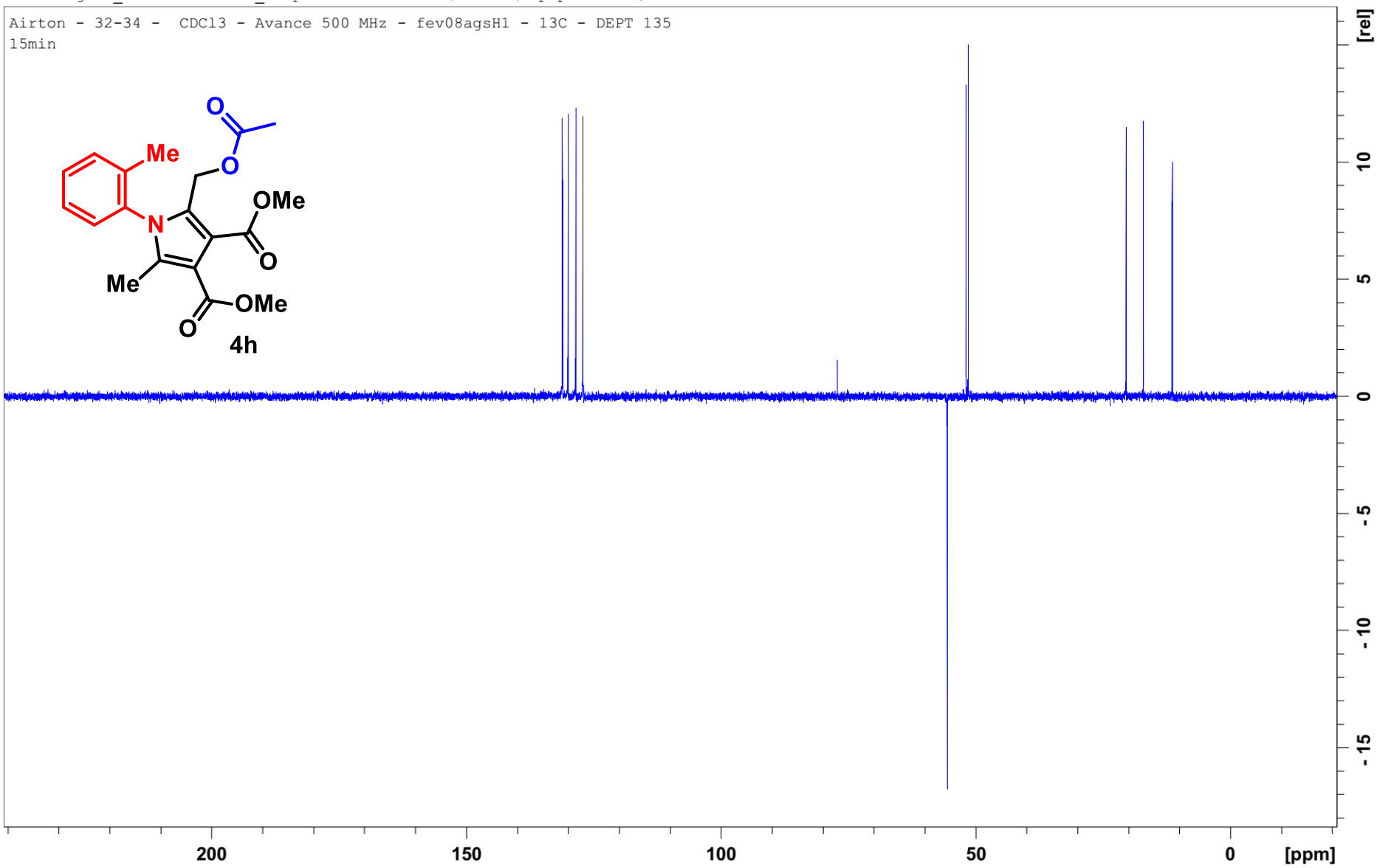


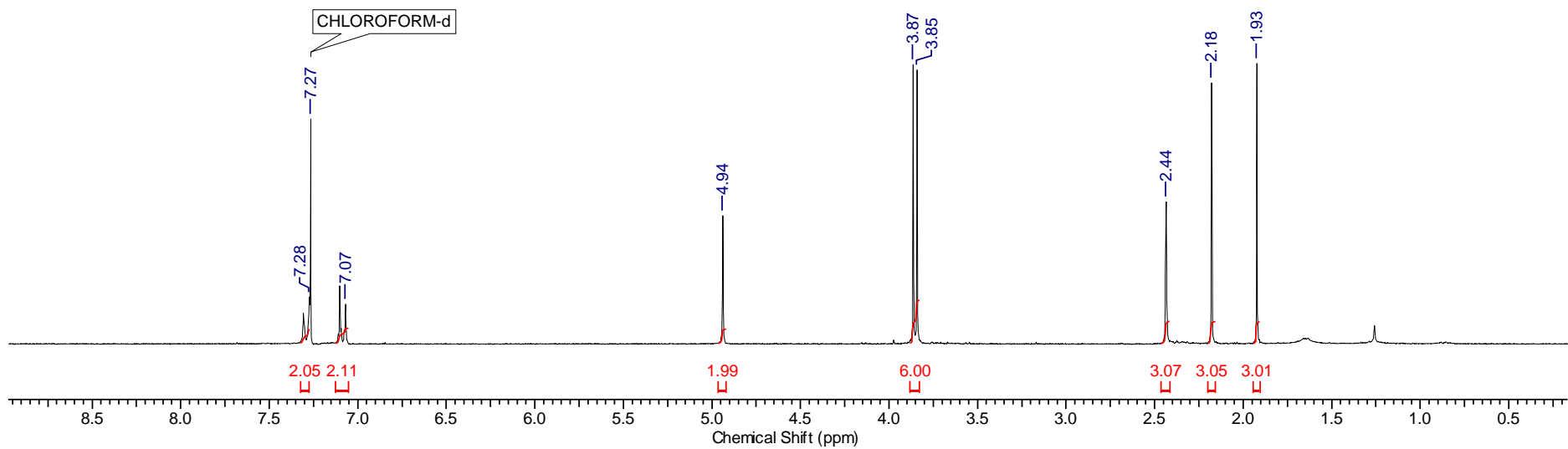
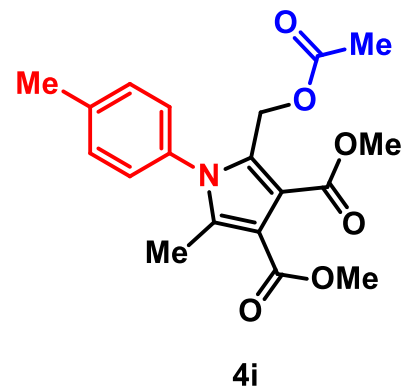
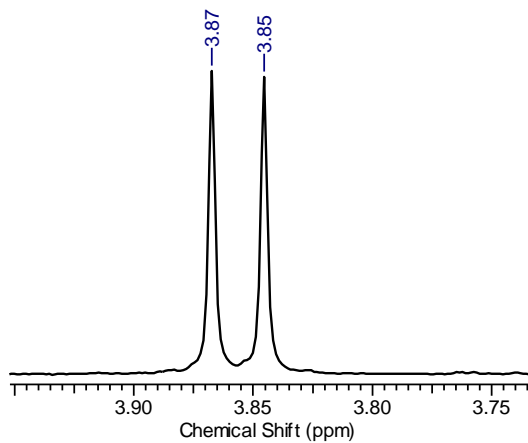
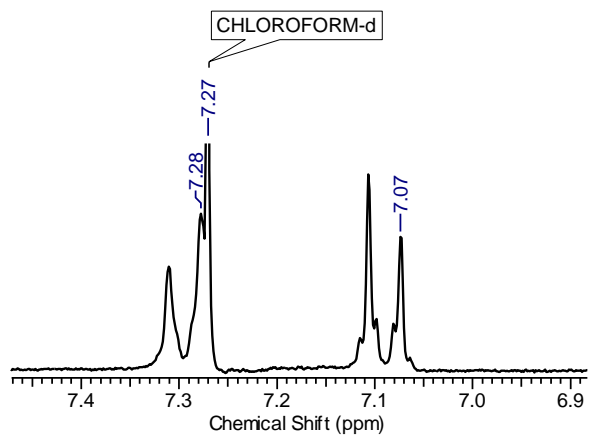


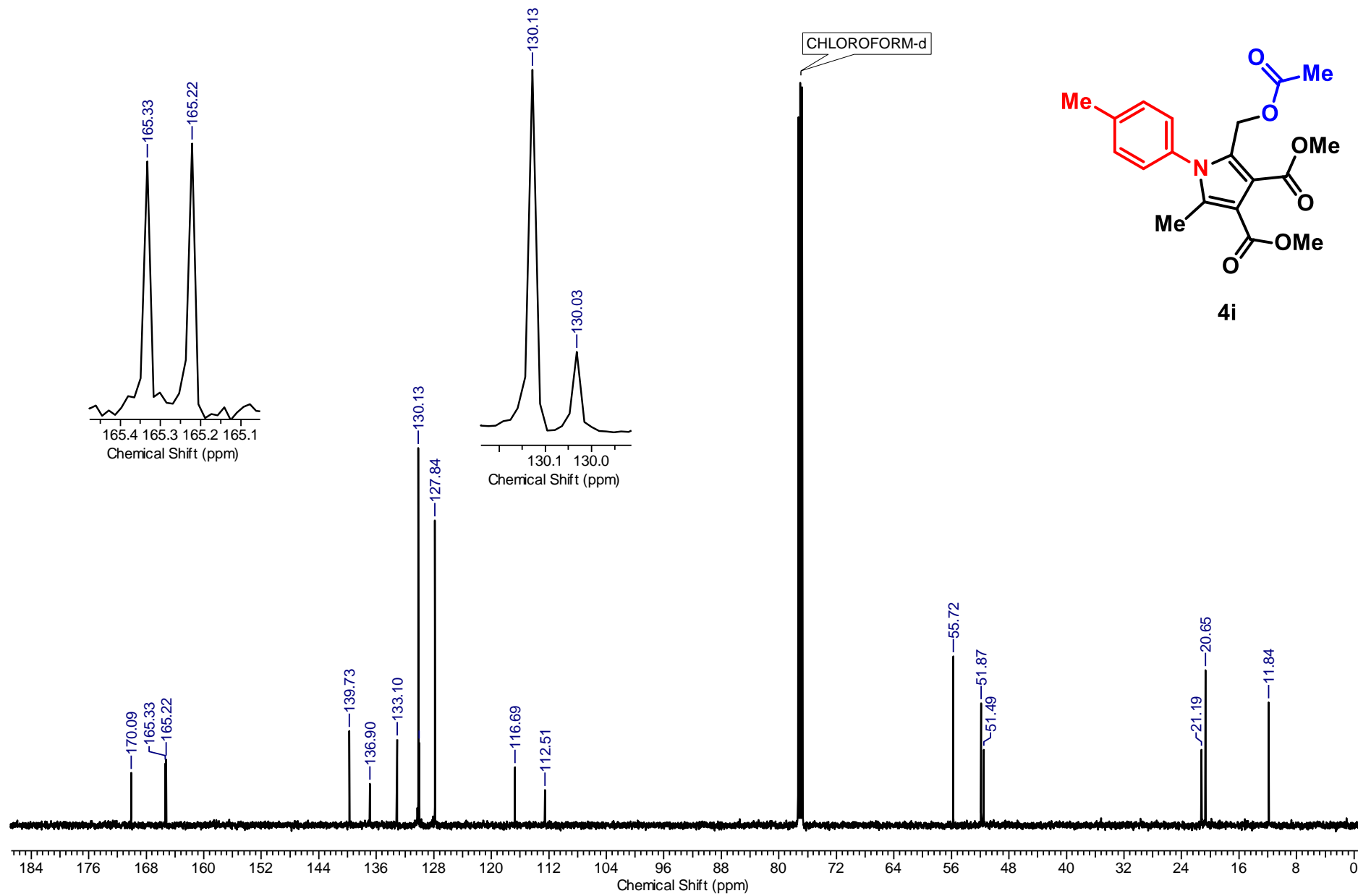
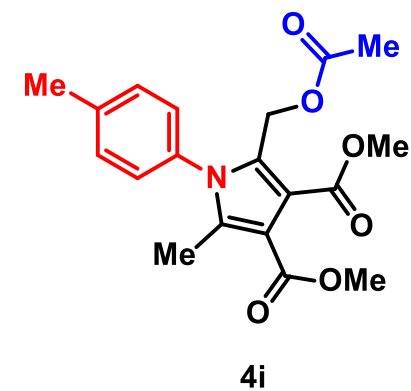
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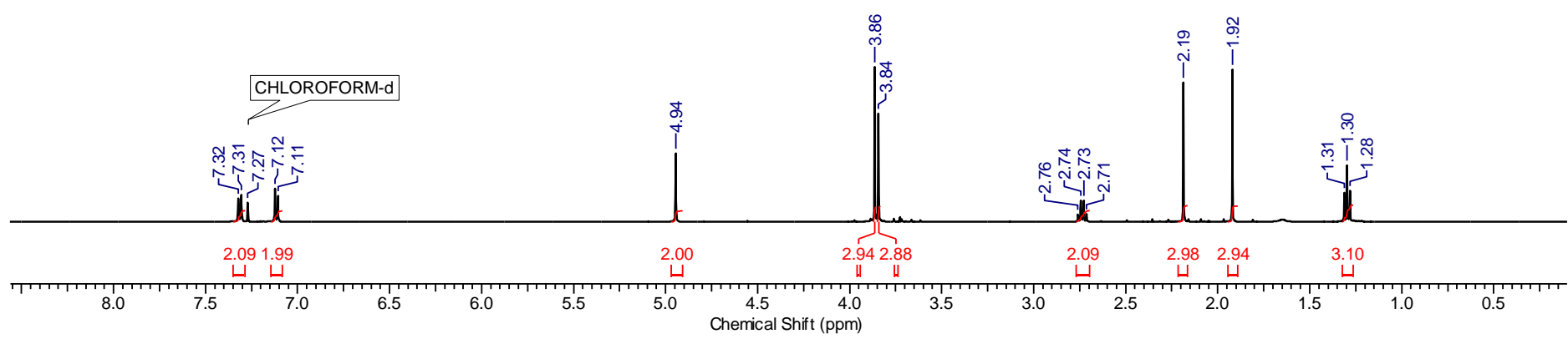
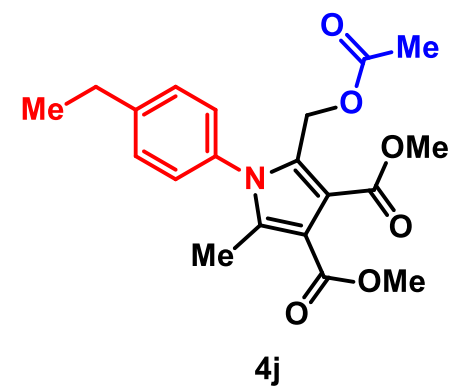
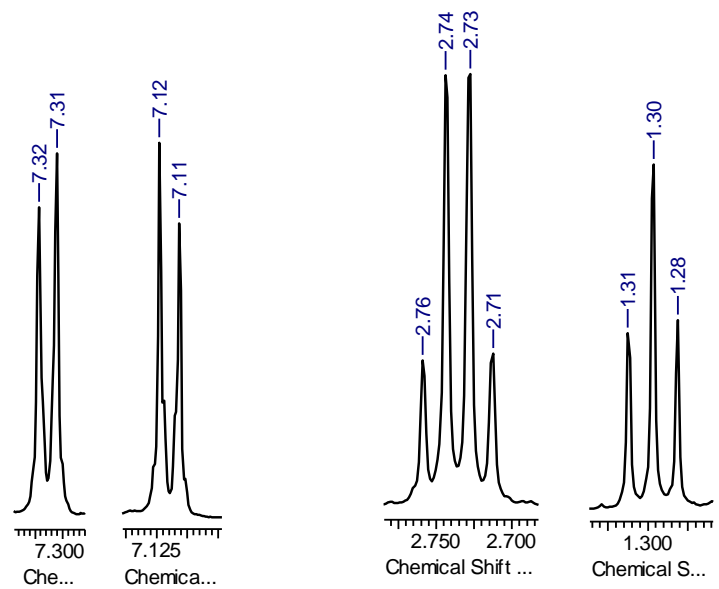
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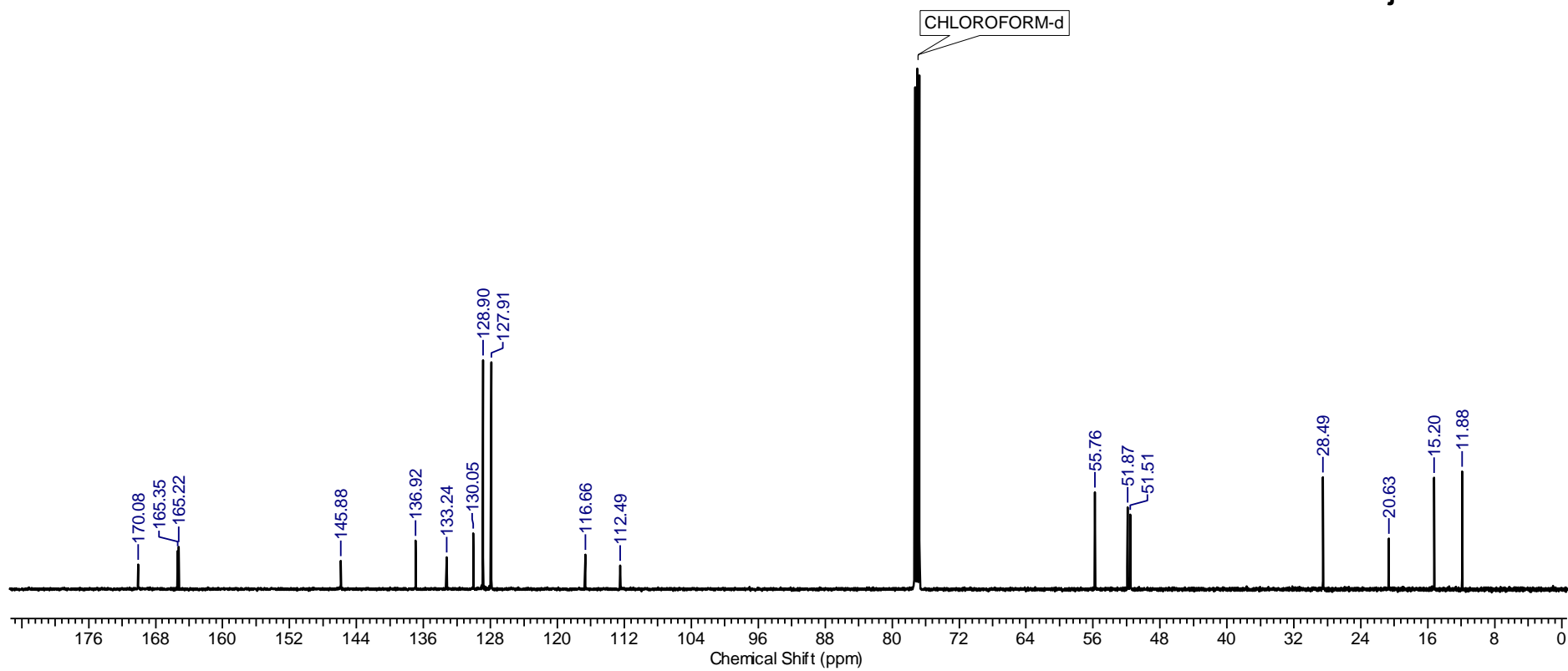
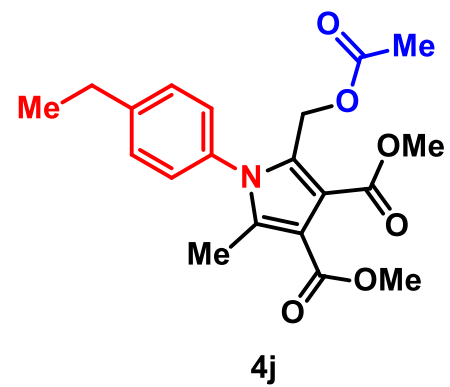
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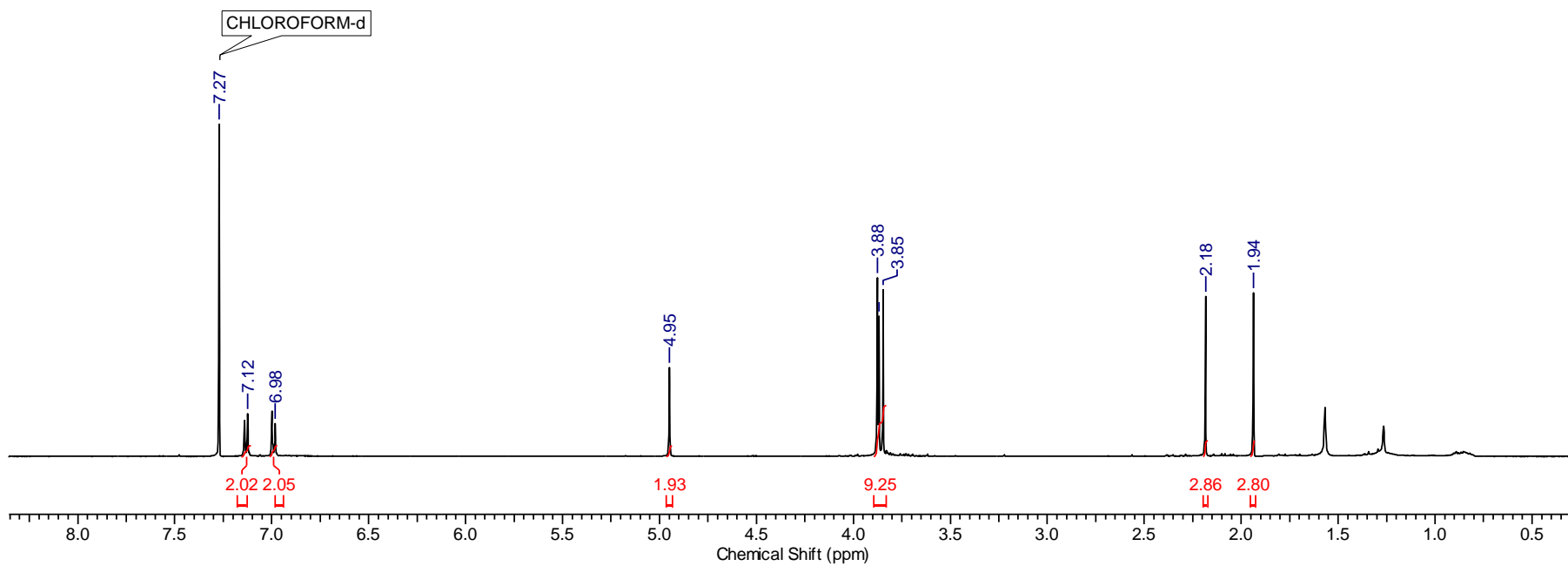
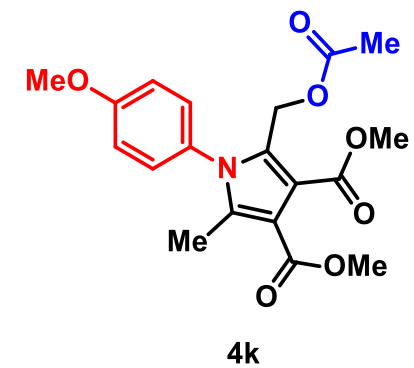
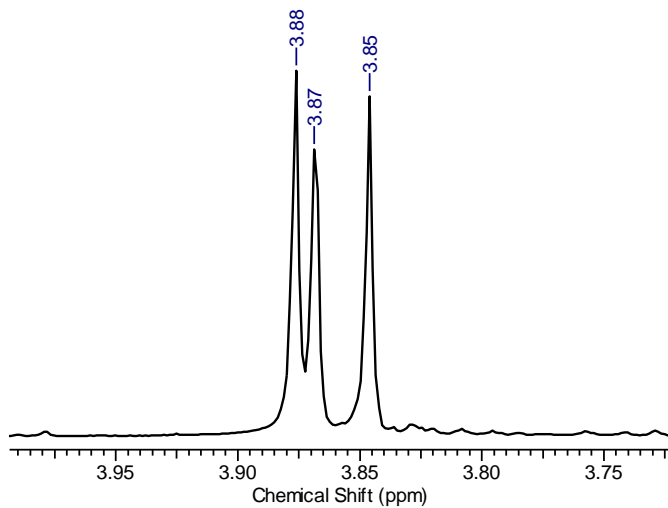
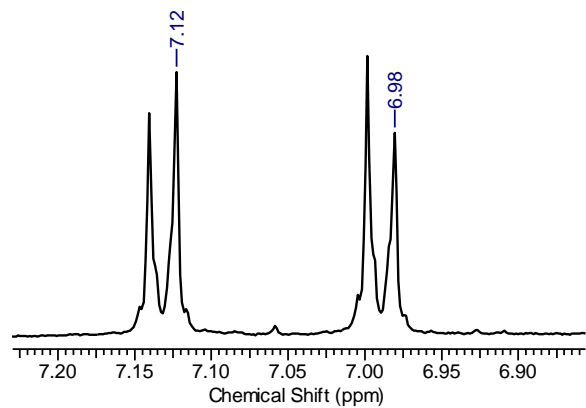


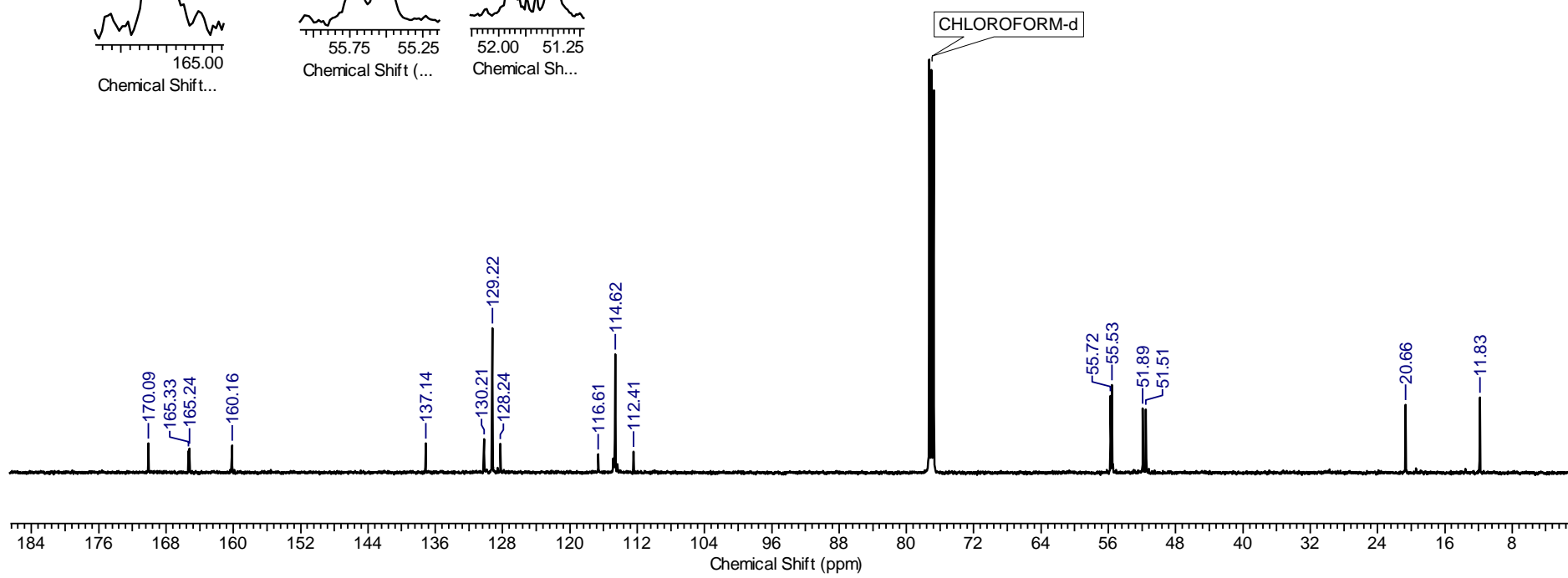
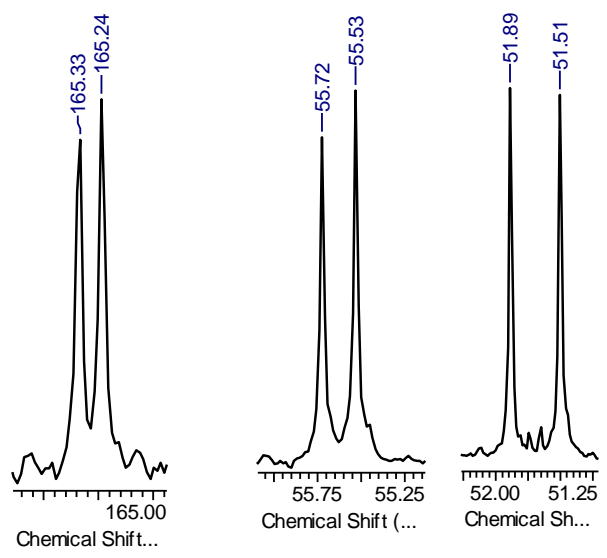
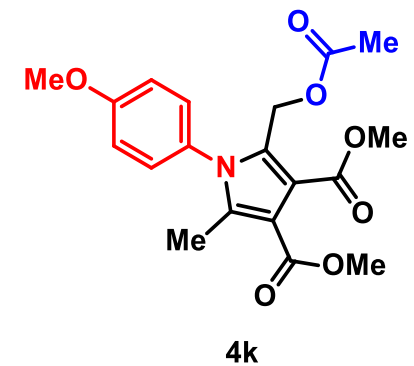


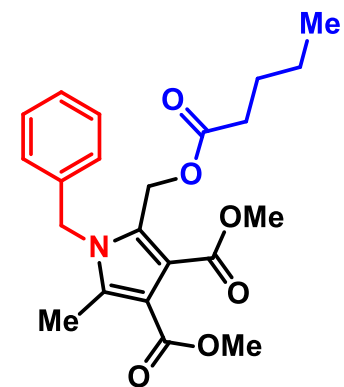
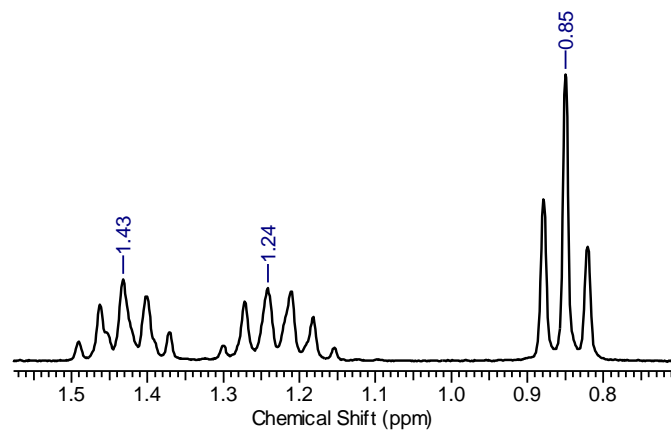
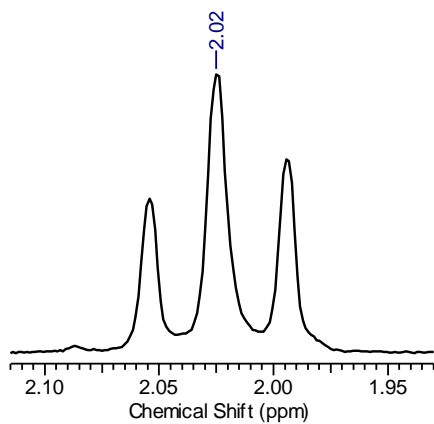




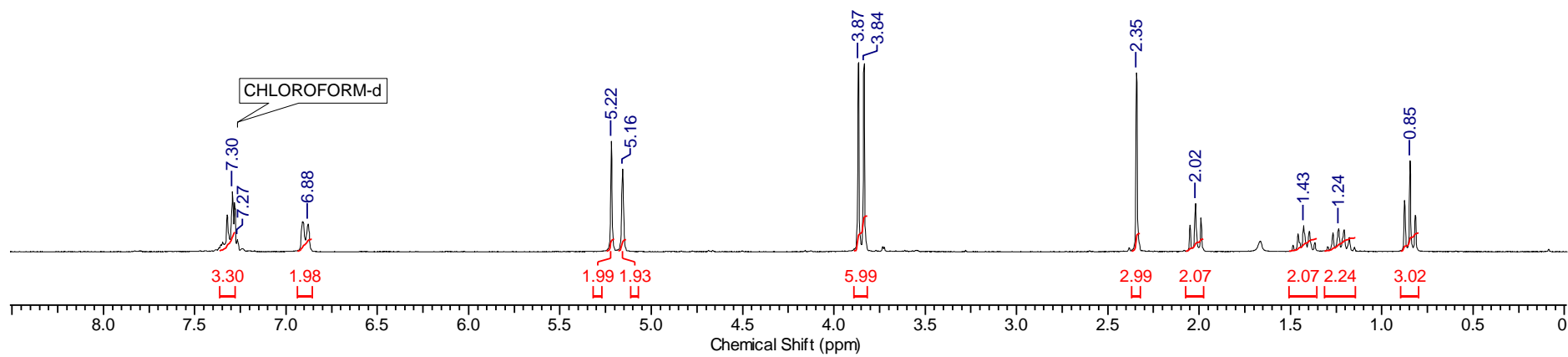


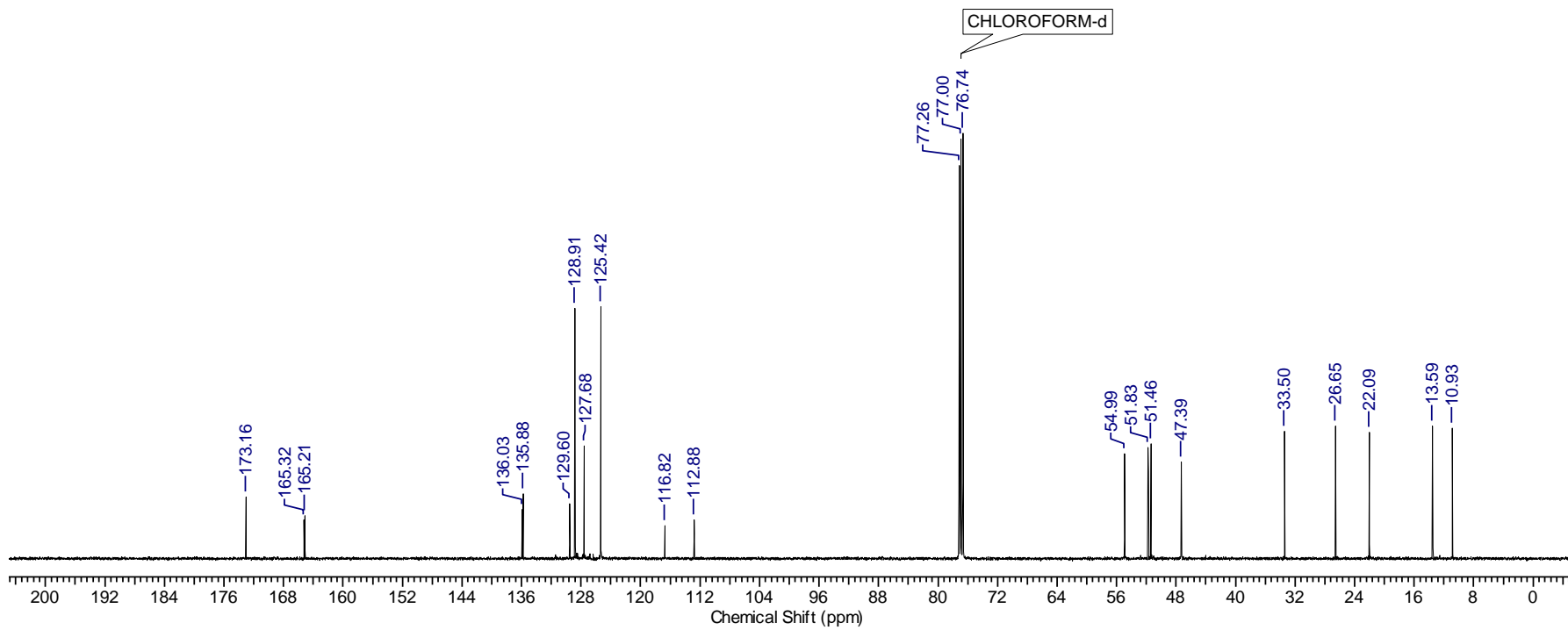
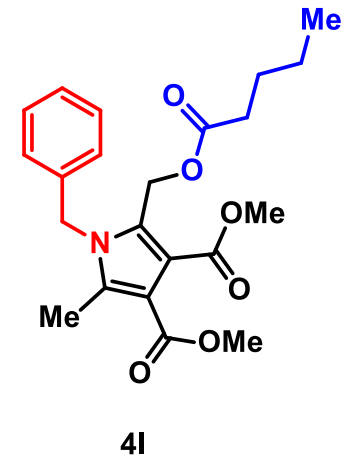
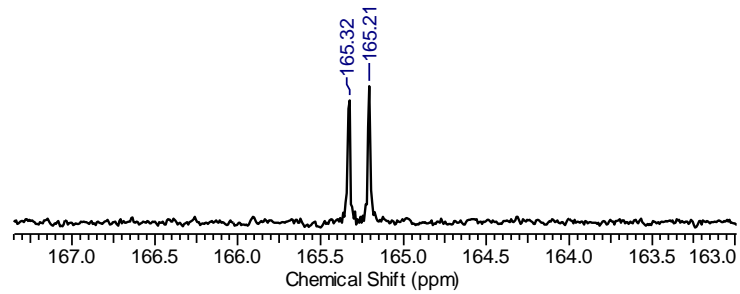


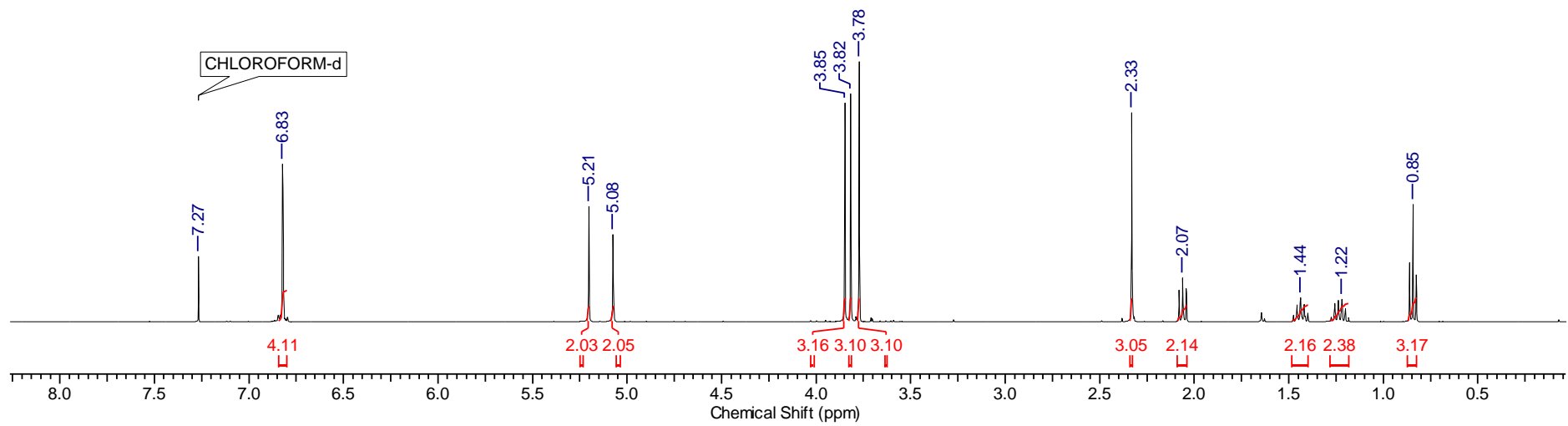
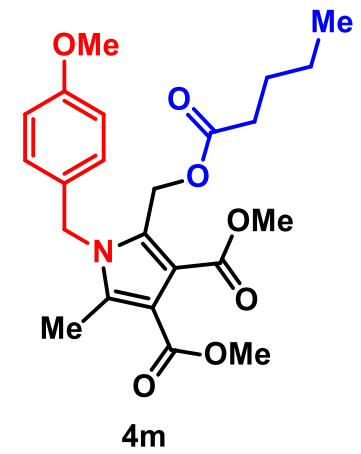
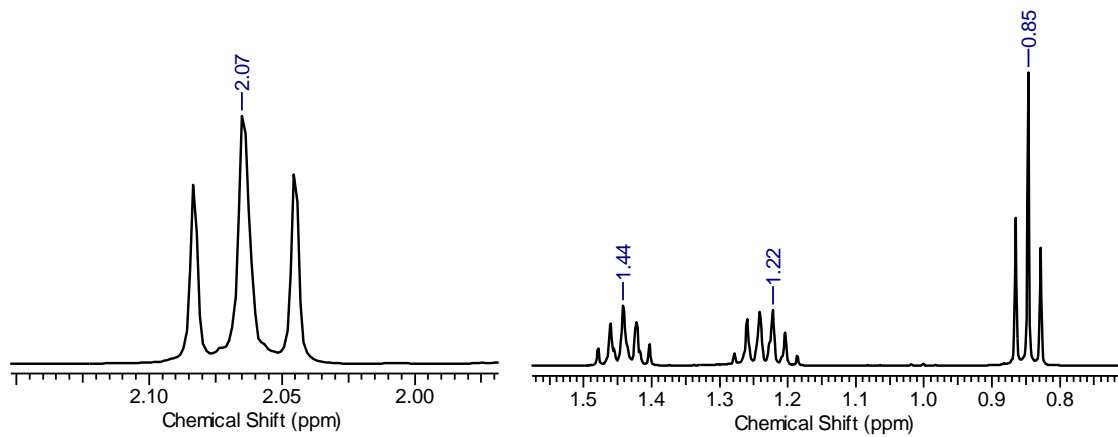


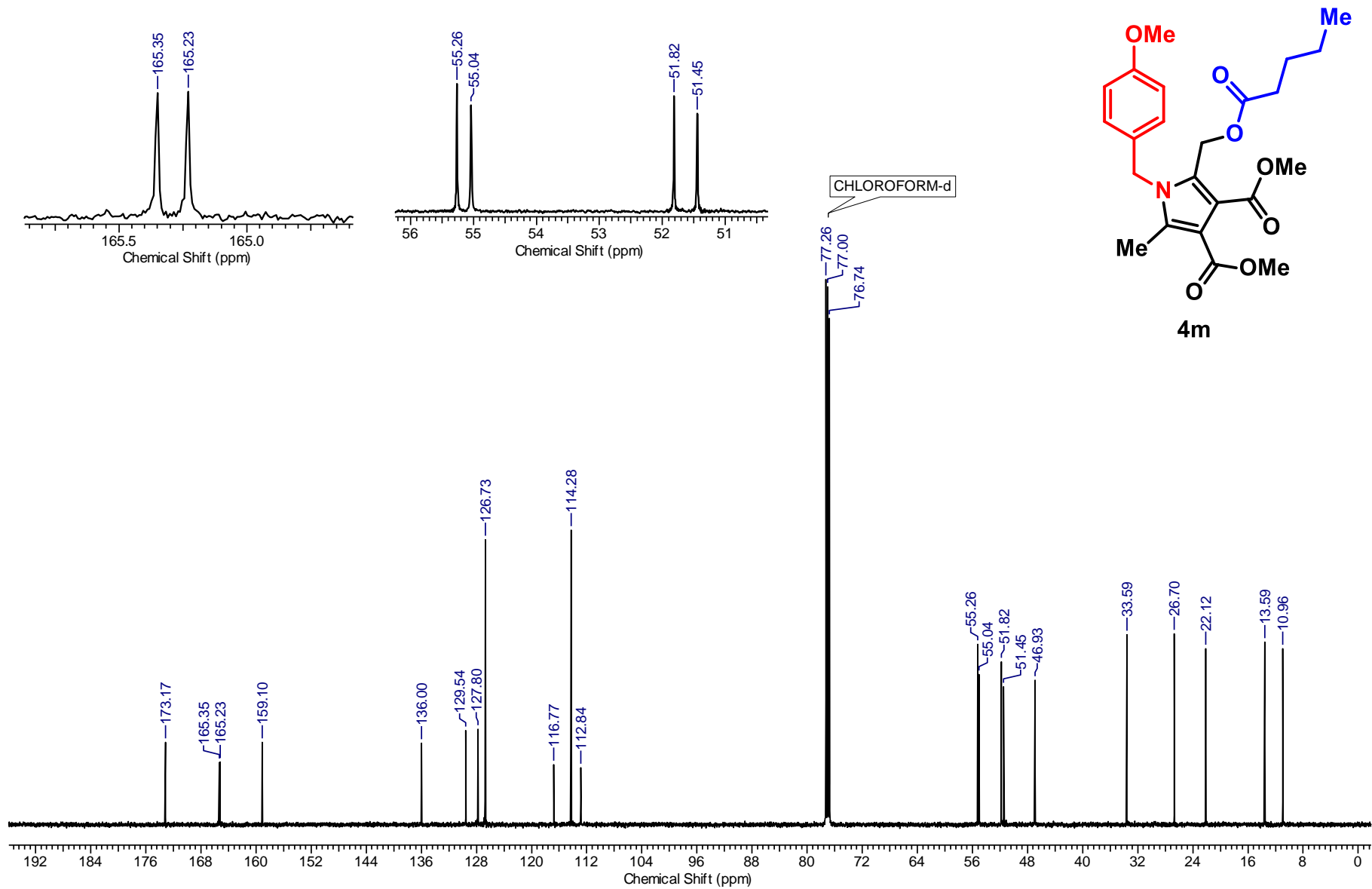


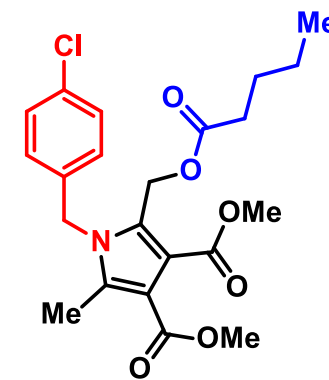
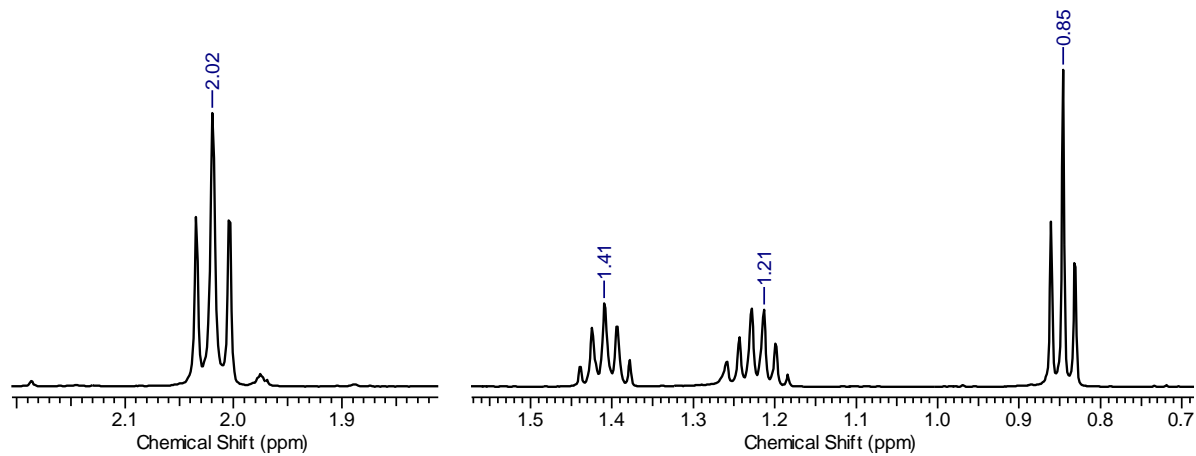
4I











4n

