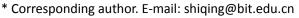
Bio-inspired engineering of a perfusable culture platform for guided three-dimensional nerve cell growth and

differentiation[†]

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D = 250µm

- **‡** These authors contributed equally to this work.
- ТСР D = 50µm D = 150µm (a) 600 8740 7655 2.564e+04 436.5 6570 2.198e+04 200 380.8 1.831e+04 200 5485 1.465e+04 4400 325.0 1.099 -200 3315 269.3



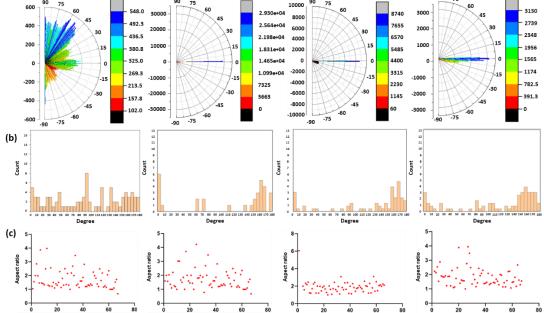


Fig.S1 Evaluation of the extent of orientation about cells 3D cultured in the microtube with different diameters (50µm, 150µm, 250µm), compared with cells 2D cultured on the tissue culture plates (TCP). (a) Polar histogram of the neurites' orientation angles; (b) Histogram of the nucleus orientation angle; (c) Scatter diagram of the nuclear aspect ratio.

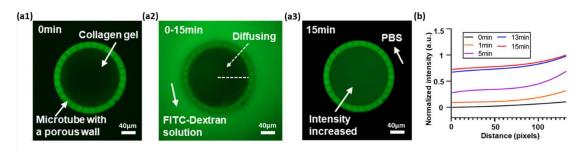


Fig.S2 (a1), (a2), (a3) The process of diffusion experiment; (b) The evolution of the intensity profiles along the dot line (marked in Fig.S2 (a2)).

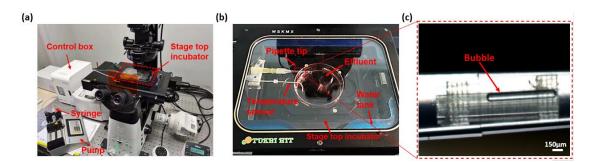


Figure S3. (a) Overview of the perfusion culture platform; (b) The perfusion culture apparatus setup in the stage top incubator; (c) An image taken during perfusion.

Symbol	Parameter	Value
D_g	Glucose diffusivity	1.0x10 ⁻⁹ (m ² /s)
R	Glucose consumption rate	8x10 ⁻³ (kg/(m ³ ·s))
С	Glucose concentration in the culture medium	4.5 (kg/m ³)
ε_c	Porosity of the collagen gel	90%
Kc	Permeability of the collagen gel	10 ⁻¹⁴ (m ²)
ρ	Culture medium density	0.893 (g/cm ³)
μ	Culture medium viscosity	8.3x10 ⁻³ (g/(cm·s))
q	Volume flow rate of the perfusion medium	5x10 ⁻¹² (m ³ /s)
Y	Mobility	50 (m·s/kg)
$\boldsymbol{\varepsilon}_i$	Interface thickness	6.5x10 ⁻⁶ (m)
σ	Surface tension coefficient	72. 92 (mN/m)
θ	Contact angle	10 (°)
p_0	Atmospheric pressure	101.325 (kPa)
g	Gravity acceleration	9.8 (m/s²)
r	Radius of the microtube	75 (μm)
h	Height of the microtube	2 (mm)
D_d	Diffusion coefficient of the gas molecules	1.8X10 ⁻⁹ (m²/s)
kн	Henry constant	7.3x10 ⁷ (Pa·kg/mol)
М	Molar mass of the gas	2.9x10 ⁻² (kg/mol)
R	Universal gas constant	8.314 (J/(mol·K))
Т	Absolute temperature	298.15 (K)

Table S1. List of the parameters of simulation model

Table S2. The statistical results of the cell viability of PC12 cells in the static and perfusable culture condition.

Day1/static	Day1/perfusable	Day7/static	Day7/perfusable
Live/Dead=1808/287	Live/Dead=4104/275	Live/Dead=3415/612	Live/Dead=3484/871
Cell viability=86.3%	Cell viability=94%	Cell viability=85%	Cell viability=81.6%
Live/Dead=2264/249	Live/Dead=1581/251	Live/Dead=1438/604	Live/Dead=5190/846
Cell viability=90%	Cell viability=86.2%	Cell viability=70.4%	Cell viability=85.2%
Live/Dead=3895/261	Live/Dead=2200/242	Live/Dead=1141/609	Live/Dead=9827/855
Cell viability=93.7%	Cell viability=91%	Cell viability=58.2%	Cell viability=91.7%

Mean±SD=90%±3%	Mean±SD=90%±3%	Mean±SD=71.2%±8%	Mean±SD=86.2%±6%
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