

## **Electronic Supplementary Information (ESI)**

### **Facile one-pot synthesis of Mn<sub>3</sub>O<sub>4</sub> nanorods and their analytical application**

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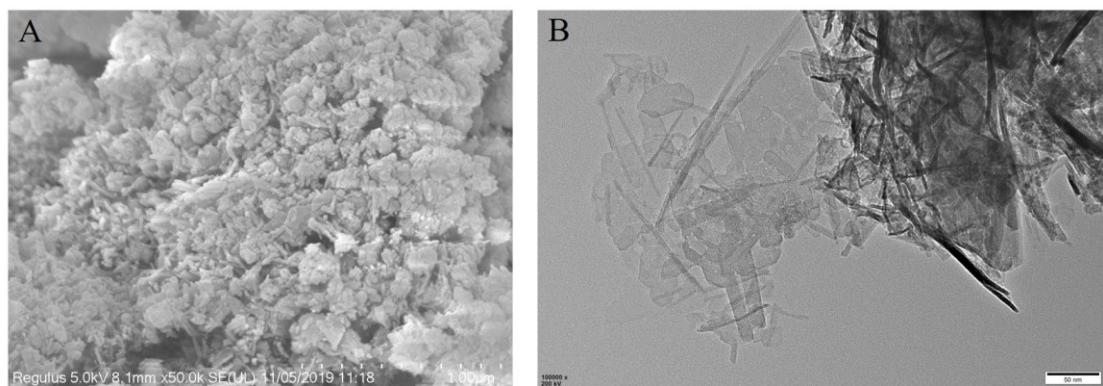
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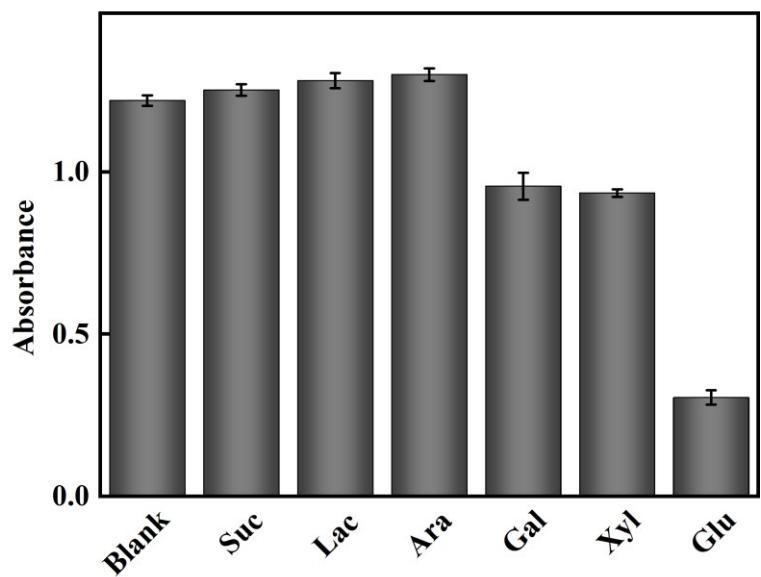
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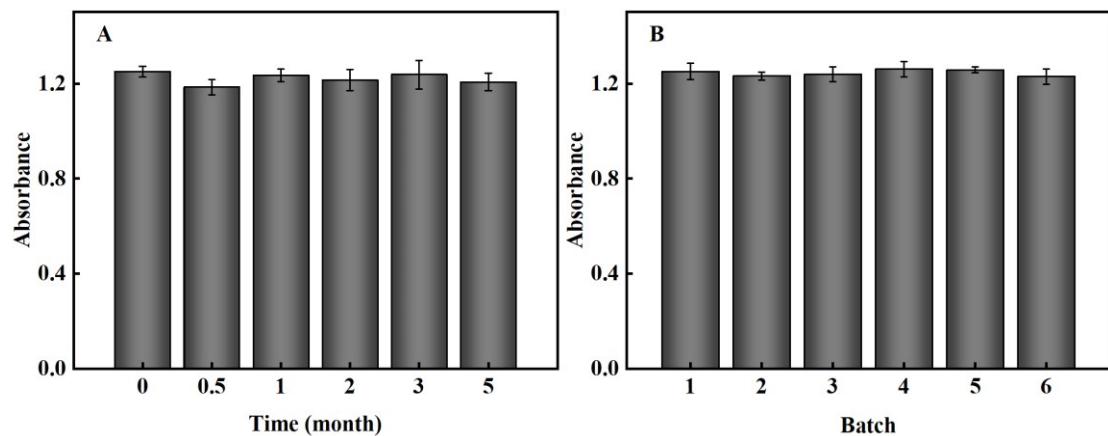
### **Additional Figures and Tables**



**Fig. S1** The large-scale (A) SEM and (B) TEM images of  $\text{Mn}_3\text{O}_4$  nanorods.



**Fig. S2** Selectivity of the proposed method for glucose detection. The concentration of Suc, Lac, Ara, Gal, Xyl and Glu is 2 mM, respectively. Error bars shown are standard deviations ( $n=3$ ).



**Fig. S3** (A) The stability of  $\text{Mn}_3\text{O}_4$  after storing different time. (B) The activity of  $\text{Mn}_3\text{O}_4$  prepared in different batches. Error bars shown are standard deviations ( $n=3$ ).

**Table S1** Comparison of this method with some others for H<sub>2</sub>O<sub>2</sub> detection.

Method	Materials and reagents	Linear range ( $\mu$ M)	LOD ( $\mu$ M)	Ref.
Colorimetry	Fluorescein	80-1200	30	1
Colorimetry	Acetate	6.0-200	3	2
Colorimetry	Au NPs <sup>a</sup>	ND <sup>b</sup>	2	3
Colorimetry	MoS <sub>2</sub>	5-100	1.5	4
Colorimetry	Co <sub>3</sub> O <sub>4</sub>	50-25000	10	5
Fluorometry	Cu nanoclusters	0.5-10	0.4	6
Fluorometry	CeO <sub>2</sub> -DNA	0-1	0.13	7
Colorimetry	Mn <sub>3</sub> O <sub>4</sub>	2-100	1.7	This work

<sup>a</sup>NPs: nanoparticles. <sup>b</sup>ND: not determined.

**Table S2** Comparison of this method with some others for glucose detection.

Method	Materials and reagents (Besides GOx)	Linear range ( $\mu\text{M}$ )	LOD ( $\mu\text{M}$ )	Ref.
Colorimetry	Acetate	6-150	4	2
Fluorometry	Cu nanoclusters	10-100	8	6
Fluorometry	CeO <sub>2</sub> -DNA	10-200	8.9	7
Colorimetry	Block copolymer	300-10000	200	8
Colorimetry	CoO-OMC <sup>a</sup>	100-5000	68	9
Colorimetry	HRP <sup>b</sup> -test paper	20-4000	14	10
Colorimetry	ATP-Fe <sub>3</sub> O <sub>4</sub>	0-4000	50	11
Fluorometry	Graphene QDs <sup>c</sup>	100-10000	30	12
Colorimetry	Au@p-SiO <sub>2</sub> <sup>d</sup>	20-500	20	13
Electrochemistry	GA-bacteria/GDH-bacteria/MWNTs/GCE <sup>e</sup>	100-2000	40	14
Electrochemistry	M13-E4@MnO <sub>2</sub> <sup>f</sup>	5-2000	1.8	15
GC-MS <sup>g</sup>	BA-MIPs <sup>h</sup>	2.8-168	0.7	16
Colorimetry	Mn <sub>3</sub> O <sub>4</sub>	5-200	4.4	This work

<sup>a</sup>OMC: ordered-mesoporous carbon. <sup>b</sup>HRP: horseradish peroxidase. <sup>c</sup>QDs: quantum dots. <sup>d</sup>Au@p-SiO<sub>2</sub>: Au nanoparticles encapsulated within porous silica; <sup>e</sup>GA-bacteria/GDH-bacteria/MWNTs/GCE: glucoamylase-displayed bacteria (GA-bacteria) and glucose dehydrogenase-displayed bacteria (GDH-bacteria) were co-immobilized on multi-walled carbon nanotubes (MWNTs) modified glassy carbon electrode (GCE).

<sup>f</sup>M13-E4@MnO<sub>2</sub>: genetically engineered M13 phage-templated MnO<sub>2</sub> nanowires.

<sup>g</sup>GC-MS: gas chromatograph equipped with a mass spectrometer. <sup>h</sup>BA-MIPs: boronate affinity-molecular imprinted polymers.

**Table S3** Performance of the proposed method for serum samples.

Sample	Known conc. (mM)	Spiked (mM)	Detected conc. (mM)	RSD (%)
# 1	4.65	2	6.78±0.24	3.53
# 2	6.96	2	9.12±0.19	2.08
# 3	13.13	2	15.27±0.61	3.99

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