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## Synthesis of novel hedgehog-shaped Bi<sub>2</sub>S<sub>3</sub> nanostructure for sensitive

## electrochemical glucose biosensor

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## **Experimental**

**Materials and Reagents:** The GOx molecules (EC 1.1.3.4, 108 U mg<sup>-1</sup>, from Aspergillus niger) were bought from Amresco. Glucose and Nafion were supplied by Sigma-Aldrich. Bismuth nitrate (Bi (NO<sub>3</sub>)<sub>3</sub>), cetyltrimethyl ammonium bromide (CTAB), trimellitic acid (C<sub>9</sub>H<sub>6</sub>O<sub>6</sub>), thiourea (CH<sub>4</sub>N<sub>2</sub>S), anhydrous ethanol, sodium phosphate (Na<sub>2</sub>HPO<sub>4</sub>), sodium dihydrogen phosphate (NaH<sub>2</sub>PO<sub>4</sub>), potassium chloride (KCl), potassium ferricyanide (K<sub>3</sub>Fe(CN)<sub>6</sub>), and potassium ferrocyanide (K<sub>4</sub>[Fe(CN)<sub>6</sub>]·3H<sub>2</sub>O) were purchased from Sinopharm Chemical Reagent Co., Ltd. Phosphate buffer solution (PBS, 0.1 mol·L<sup>-1</sup>) was the mixture of Na<sub>2</sub>HPO<sub>4</sub> and NaH<sub>2</sub>PO<sub>4</sub> solutions, and its pH was adjusted by H<sub>3</sub>PO<sub>4</sub> or NaOH solution. All reagents used in experiments were of analytical grade.

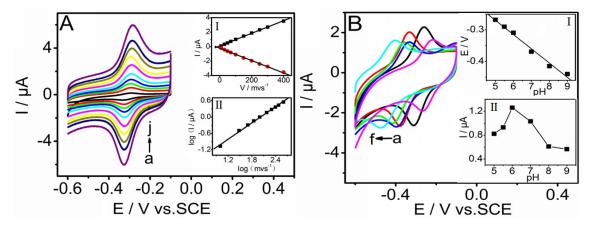
**Apparatus**: A CHI 852C electrochemical workstation was from Shanghai Chenhua Instrument Co. Ltd. (China), which was used to carry out electrochemical measurements. A three-electrode system, including a working electrode of glassy carbon electrode (GCE), a reference electrode of saturated calomel

electrode (SCE), and an auxiliary electrode of platinum wire were employed in the experiment. The cyclic voltametric tests were implemented at a scan rate of 100 mV·s<sup>-1</sup> in a cell with 10.0 mL PBS. All solution pH measurements were conducted on a S-25 digital pH-meter with glass combination electrode. A Hitachi S-4800 scanning electron microscope (Japan) was used to obtain scanning electron micrographs (SEM) at 15 kV of acceleration voltage. X-ray diffractometer spectra were measured by using polycrystalline X-ray diffractometer (XRD, D8 advance, Bruker AXS, Germany). Electrochemical impedance spectroscopy (EIS) was tested in a KNO<sub>3</sub> solution (0.1 mol·L<sup>-1</sup>) including 5 mM of  $K_3[Fe(CN)_6]/K_4[Fe(CN)_6]$  within a frequency range from 0.01 to 10 KHz. Fourier transform infrared (FT-IR) spectrum was recorded at a Tensor 27 spectrometer (Bruker Co., Germany).

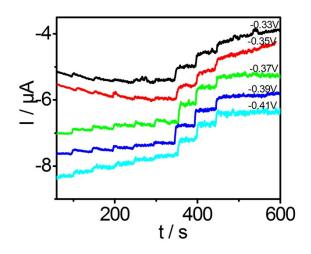
Synthesis of hedgehog-shaped  $Bi_2S_3$  nanostructure and electrochemical glucose biosensor: Hedgehog-shaped  $Bi_2S_3$  nanostructures were prepared by a composite soft template method by a simple hydrothermal route according to previous research<sup>S1</sup>. Firstly, 0.0210 g of trimellitic acid, 0.0365 g of CTAB, 0.3806 g of thiourea, 0.50 mL of bismuth nitrate solution (0.20 mol·L<sup>-1</sup>) and 9.50 mL of distilled water were transferred into a 50 mL beaker. The system was ultrasonically mixed to obtain a homogeneous light yellow solution. The obtained solution was added to a Teflon-lined reaction kettle, which was placed in a constant-temperature oven for 12 h at 120 °C. After that, the obtained black precipitates were centrifuged and washed using ethanol and distilled water several times in sequence, and finally dried at 60 °C under vacuum for 12 h to obtain black powder product.

The surface of GCEs were polished with 0.3- $\mu$ m and 0.05- $\mu$ m alumina slurry (Buhler) in turn, and cleaned carefully using distilled water. High purity N<sub>2</sub> stream was used to dry GCEs at room temperature. Then, 1.0 mg of HS-Bi<sub>2</sub>S<sub>3</sub> was weighted to distilled water (1.0 mL) and dispersed by ultrasonication. Next, GOx (1.0 mg) was added into 100  $\mu$ L of HS-Bi<sub>2</sub>S<sub>3</sub> suspension above, and kept shaking for at least 15 min. Next, 5.0  $\mu$ L of the mixed solution was coated on clean electrodes and dried naturally at room temperature. To avoid the leakage of the GOx from modified GCE, 5.0  $\mu$ L of Nafion (0.5%) was coated on surface of GOx/HS-Bi<sub>2</sub>S<sub>3</sub>/GCE. The prepared Nafion/GOx/HS-Bi<sub>2</sub>S<sub>3</sub>/GCE was stored in a refrigerator at 4 °C.

**Optimization of electrochemical detection conditions** 



**Fig.** *S1.* (A) cyclic voltammograms of the Nafion/GOx/HS-Bi<sub>2</sub>S<sub>3</sub>/GCE in 0.1 M pH 6.0 N<sub>2</sub>-saturated PBS at 10, 20, 50, 80, 100, 150, 200 and 300 mV s<sup>-1</sup> (from a to j), inset I: plots of anodic and cathodic peak currents vs. scan rate, inset II: plot of logarithm of  $i_{pc}$  vs. logarithm of v; and (B) Cyclic voltammograms of the Nafion/GOx/HS-Bi<sub>2</sub>S<sub>3</sub>/GCE in N<sub>2</sub>-saturated 0.1 M PBS with different pH values of (a-f) 5.0, 5.5 6.0, 7.0, 8.0 and 9.0 at a scan rate of 100 mV s<sup>-1</sup>, inset I: plot of formal potential vs. pH, and inset II: plot of peak current vs. pH.



*Fig. S2.* Amperometric response of Nafion/GOx/Bi<sub>2</sub>S<sub>3</sub>/GCE at different potential to successive additions of glucose in a stirred 0.1 M pH 6.0 PBS.

Sample	Reference (mM)	Proposed method (mM)	Relative errors (%)		
1	4.64	4.43	- 4.5		
2	4.88	5.03	3.1		
3	5.19	4.80	- 7.5		
4	6.39	5.92	- 7.4		
5	10.55	10.84	2.7		

Table S1	Detection	results of	glucose	in	human	serum	samples

## Reference

**S1.** M. L. Ye, F. Shi, M. Shen, W. F. Qin, C. L. Ren, Z. J. Yang, *Collioid. Surface. A* 2021, **613**, 126094.