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Supplementary information

One Step Synthesis of PY-NBD to Distinguish Cys/Hcy and GSH in

Aqueous Solution and Living Cells by Dual Channels

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1. Experimental reagents and instruments

All other chemicals were obtained from commercial suppliers and used without further purification. Silica gel (200-300 mesh, Qingdao Haiyang Chemical Co.) was used for column chromatography. ¹H NMR and ¹³C NMR spectra were recorded on a Bruker Advance at 400MHz or at 100 MHz, δ values are in parts per million relatives to TMS in DMSO-d₆. Mass spectra (MS) were measured with Bruker Apex IV FTMS using electrospray ionization (ESI). Absorption spectra were recorded on a Purkinje TU-1901 spectrophotometer. Fluorescence measurements were taken on a Hitachi F-7000 fluorescence spectrometer with a 10mm quartz cuvette. pH measurements were carried out with a pH acidometer (Mettler Toledo FE-30). Fluorescence imaging was observed under an Olympus IX81 confocal fluorescence microscope.



2. Characterization data

Fig. S1. ¹H NMR (400 MHz, DMSO-d₆) spectra of probe PY-NBD.



Fig. S2. ¹³C NMR (101 MHz, DMSO-d₆) spectra of probe PY-NBD.



Fig. S3. ESI-MS of probe PY-NBD.





Fig. S4. The stability of **PY-NBD** (5 μ M) in PBS (10 mM, pH 7.4, 50% CH₃CN, V/V) system and the fluorescence spectrum of response time to 50 μ M Hcy/Cys/GSH. (A) At 410nm, λ ex=345 nm. Slit: 2.5 nm/2.5 nm; (B) At 546nm, λ ex=473 nm. Slit: 5.0 nm/5.0 nm.

4. The capabilities of PY-NBD for detecting Hcy/Cys and GSH at different pH



Fig. S5. Fluorescence intensity change graph of PY-NBD (5 μ M) and Hcy/Cys/GSH (50 μ M) at different pH. (A) At 410nm, λ ex=345 nm. Slit: 2.5 nm/2.5 nm; (B) At 546nm, λ ex=473 nm. Slit: 5.0 nm/5.0 nm.



5. ESI-MS of PY-NBD after upon addition Cys, Hcy, and GSH

Fig. S7. ESI-MS of PY-NBD after upon addition Hcy



Fig. S8. ESI-MS of PY-NBD after upon addition GSH



6. Cytotoxicity assay

Fig. S9. Cell viability of HeLa cells treated with different concentrations of PY-NBD