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Supporting Information

Inclusion Complexation and Self-association of Cucurbit[n]uril (n = 6, 7)

and Diquat under Pseudo-physiological Conditions

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qNMR spectra were recorded over a spectral width of 12000 Hz using 8 scans without spinning. The pulse angle is 90°, and the repetition time is 64 s. DSS- d_6 (FUJIFILM Wako Chemicals Co) is used as an internal reference in qNMR. CB[6] or CB[7] were dissolved in NaCl-D₂O. Purities of CB[6] or CB[7] were calculated using integrated values of the methyl proton signal of the trimethylsilyl group of DSS- d_6 (Si-CH₃) and the H^a and H^c proton signals of CB[6] or CB[7]. In the case of CB[7], although DSS- d_6 forms the complex with CB[7], the integration value of Si-CH₃ did not affect on the quantitative (-0.744 ppm). Purities of CB[6] or CB[7] were 82.9 % and 87.2 %, respectively.



Fig. S1 qNMR spectra of CB[6].



Fig. S2 qNMR spectra of CB[7].

The stoichiometry for the inclusion complexation of CB[7] and DQ was investigated via Job's plot by using ¹H NMR measurement when the sum of the CB[7] and DQ concentrations ([CB[7]] and [DQ]) was maintained at 1 and 10 mM; changes in the observed chemical shift ($\Delta \delta_{obs}$) of the DQ H⁴ proton signal were used, and the stoichiometry was found from the plot of the $\Delta \delta_{obs}$ *[DQ] as function of the ratio [DQ]/([CB[7]] + [DQ]).



Fig. S3 Continuous variation plot (Job's plot) by using ¹H NMR measurement for the inclusion complexation of CB[7] and DQ: total concentration 1 mM in (a) pD1.2, (b) pD7.4, and (c) NaCl-D₂O; total concentration 10 mM in (d) pD1.2, (e) pD7.4, and (f) NaCl-D₂O.

The stoichiometry and association constant for the inclusion complexation between CB[7] and DQ were calculated via regression analysis at 30 - 65 °C in pD1.2, pD7.4, and NaCl-D₂O (Fig. S3, S5, and S7) while continuously increasing the CB[7] concentration ([CB[7]) from 0 to 5 mM in a constant concentration of DQ ([DQ]; 0.5 mM). The following Eqn (1) was used for regression analysis. *K* and *n* are, respectively, the association constant and stoichiometry of CB[7] and DQ, $\Delta \delta_{obs}$ means ($\delta_{DQ} - \delta_{obs}$), $\Delta \delta_{CX}$ indicates ($\delta_{DQ} - \delta_{CX}$), and δ_{DQ} , δ_{CX} , and δ_{obs} are the chemical shifts (in ppm) of the H⁴ protons of, correspondingly, DQ in a free state, the CB[7]-DQ inclusion complex, and the CB[7]-DQ mixture in the ¹H NMR spectra. In Fig. S3, S5, and S7, m1, m2, and m3 indicate that $\Delta \delta_{CX}$, *K*, and *n*, respectively. The thermodynamic parameters, that is, free energy change (ΔG), enthalpy change (ΔH), and entropy change (ΔS), for the inclusion complexation of CB[7] and DQ were derived from Van't Hoff plots by using association constants.

$$n \operatorname{CB}[7] + \operatorname{DQ} \underbrace{\overset{K}{\longleftarrow} \operatorname{CB}[7] - \operatorname{DQ}}_{[\operatorname{CB}[7]]} = \left[\frac{1}{\left(\frac{\Delta \ \delta \operatorname{cx}}{\Delta \ \delta \operatorname{obs}} - 1\right) K} \right]^{1/n} + n \frac{\Delta \ \delta \operatorname{obs}}{\Delta \ \delta \operatorname{cx}} [\operatorname{DQ}] \quad \cdots \quad \operatorname{Eqn}(1)$$





Fig. S4 Regression analysis by using Eqn (1) based on $\Delta \delta_{obs}$ of the H⁴ proton signal of DQ (0.5 mM DQ) at 30 - 65 °C in pD1.2: (a) 30 °C, (b) 35 °C, (c) 40 °C, (d) 45 °C, (e) 50 °C, (f) 55 °C, (g) 60 °C, and (h) 65 °C.



Fig. S5 Van't Hoff plots for the inclusion complexation between CB[7] and DQ (0.5 mM DQ) in pD1.2.





Fig. S6 Regression analysis by using Eqn (1) based on $\Delta \delta_{obs}$ of the H⁴ proton signal of DQ (0.5 mM DQ) at 30 - 65 °C in pD7.4: (a) 30 °C, (b) 35 °C, (c) 40 °C, (d) 45 °C, (e) 50 °C, (f) 55 °C, (g) 60 °C, and (h) 65 °C.



Fig. S7 Van't Hoff plots for the inclusion complexation between CB[7] and DQ (0.5 mM DQ) in pD7.4.





Fig. S8 Regression analysis by using Eqn (1) based on $\Delta \delta_{obs}$ of the H⁴ proton signal of DQ (0.5 mM DQ) at 30 – 65 °C in NaCl-D₂O: (a) 30 °C, (b) 35 °C, (c) 40 °C, (d) 45 °C, (e) 50 °C, (f) 55 °C, (g) 60 °C, and (h) 65 °C.



Fig. S9 Van't Hoff plots for the inclusion complexation between CB[7] and DQ (0.5 mM DQ) in NaCl-D₂O.

The association constant for the inclusion complexation of CB[7] and DQ was calculated via Benesi-Hildebrand method in pD1.2, pD7.4, and NaCl-D₂O while continuously increasing the CB[7] concentration ([CB[7]) from 0 to 5 mM in a constant concentration of DQ ([DQ]; 0.05 mM). The UV spectroscopy was conducted by using a UVmini-1240 (SHIMADZU, Japan); the absorbance was recorded at 330 nn and 35 °C. The following Eqn (2) was used for regression analysis. *K* is the association constant of CB[7] and DQ, ΔE means ($A_{obs}-A_{DQ}$), $\Delta \varepsilon$ indicates ($\varepsilon_{complex}-\varepsilon_{DQ}$), and A_{obs} and A_{DQ} , $\varepsilon_{complex}$ and ε_{DQ} are the absorbance of CB[7]-DQ mixture and DQ in a free state, and the molar absorption coefficient of the CB[7]-DQ inclusion complex and DQ in a free state.

$$CB[7] + DQ \xrightarrow{K} CB[7]-DQ$$

$$\frac{1}{\Delta E} = \frac{1}{K \Delta_{\mathcal{E}} [DQ][CB[7]]} + \frac{1}{\Delta_{\mathcal{E}} [DQ]} \cdots Eqn(2)$$



Fig. S10 Regression analysis by using Eqn (2) based on ΔE at 35 °C in (a) pD1.2, (b) pD7.4, and (c) NaCl-D₂O.

The *n* and *K* for the complexation between CB[7] and DQ were calculated via regression analysis by using Eqn (1) at 30 - 65 °C in NaCl-D₂O. The CB[7] concentration continuously was increased from 0 to 10 mM in a constant concentration (1 mM DQ). The ΔG , ΔH , and ΔS for the inclusion complexation of CB[7] and DQ were derived from Van't Hoff plots by using association constants. The association constant at 35 °C was 484 M⁻¹, and ΔG , ΔH , and $T\Delta S$ were -15.8 kJ/mol, -1.1 kJ/mol and -14.7 kJ/mol, respectively.





Fig. S11 Regression analysis by using Eqn (1) based on $\Delta \delta_{obs}$ of the H⁴ proton signal of DQ (1 mM DQ) at 30 - 65 °C in NaCl-D₂O: (a) 30 °C, (b) 35 °C, (c) 40 °C, (d) 45 °C, (e) 50 °C, (f) 55 °C, (g) 60 °C, and (h) 65 °C.



Fig. S12 Van't Hoff plots for the inclusion complexation between CB[7] and DQ (1 mM DQ) in NaCl- D_2O .



Fig. S13 Behavior of CB[7] or DQ, as revealed via ¹H NMR when increasing the CB[7] or DQ concentrations in pD1.2, pD7.4, and NaCl-D₂O: CB[7] in (a) pD1.2, (b) pD7.4, and (c) NaCl-D₂O; DQ in (d) pD1.2, (e) pD7.4, and (f) NaCl-D₂O. H^a, H^b, and H^c indicate the proton signals of CB[7]. H³, H⁴, H⁵, and H⁶ indicate the proton signals of DQ.



Fig. S14 Rotating-frame Overhauser effect (ROE) observed between CB[7] and DQ when forming the inclusion complex in pD7.4: (a) ¹H NMR spectrum of CB[7] and DQ; (b) one-dimensional (1D) ROESY spectrum irradiated to the H^a proton of CB[7]; (c) 1D ROESY spectrum irradiated to the H⁷ proton of DQ. The half-arrows in 1D ROESY spectra indicate positions of the irradiated proton signals.



Fig. S15 ROE observed between CB[7] and DQ when forming the inclusion complex in NaCl-D₂O: (a) ¹H NMR spectrum of CB[7] and DQ; (b) 1D ROESY spectrum irradiated to the H^a proton of CB[7]; (c) 1D ROESY spectrum irradiated to the H⁷ proton of DQ. The half-arrows in 1D ROESY spectra indicate positions of the irradiated proton signals.