Supporting Information

ROS-mediated liposomal dexamethasone: a new FA-targeted nanoformulation to combat rheumatoid arthritis *via* **inhibiting iRhom2/TNF-α/BAFF pathways** Yanqin Song^{a, b}, Muhammad Ismail^c, Qi Shan^a, Jianing Zhao^a, Yanping Zhu^a,

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Supplementary Figures

Scheme S1. Synthetic route of di-S-PC lipids.

Figure S1. (a) ¹H NMR and (b) ¹³C NMR (600 MHz, CDCl₃) spectra of C_{18} -S-COOH intermediate.

Figure S2. ¹H NMR (600 MHz, CDCl₃) spectrum of ROS-responsive di-S-PC lipids. Figure S3. ¹³C NMR (600 MHz, CH3OH- d_4) spectrum of ROS-responsive di-S-PC lipids.

Figure S4. The *in vitro* cellular association of Dex@FA-ROS-Lips. Confluent LPSactivated RAW264.7 cells exposed to free Dex solution or liposomes containg Dex $(50 \mu M)$ at various time intervals.

Figure S5. (a) Expression levels and (b) the quantified densitometric analysis of iRhom2, TNF- α and BAFF proteins detected by Western blotting (n = 3). β -actin antibody was used to ensure after incubation equal protein loading. Data are presented as Mean \pm S.D. and were statistically analyzed by one-way ANOVA. **P < 0.01 vs the control group; *P < 0.05, **P < 0.01 vs free Dex group.

Figure S6. (a) Schematic representation of AIA mice model protocol. 100 μ L of CFA with the heat-killed mycobacteria (10 mg/mL) was injected subcutaneously into SD males on day 0; (b) Arthritis first signs appeared 14 days after the first immunization step; (c) Hind paws and fore paws images according to the inflammation visual scores shown in table.

Figure S7. Hemolysis of Dex@FA-ROS-Lips: (a) Micrograph of RBCs exposed to 250 μg/mL ROS-Lips and Dex@FA-ROS-Lips after incubated at 37 °C for 2 h. 0.9%

saline (0% hemolysis) and distilled water (100% hemolysis) was set as negative and positive controls; (b) Images of tubes containing red blood cells (RBCs) incubated with different groups (after 3000 × centrifugation); (c) Hemolysis rate (%) of tested liposomes and the value less than 5% was considered safe for *in vivo* administration.

Scheme S1. Synthetic route of di-S-PC lipids.



As shown, ROS-responsive lipids (di-S-PC) were synthesized by a two-step reaction that C18-S-COOH intermediate was firstly prepared by conjugation between 6-Bromohexanoic acid and 1-Dodecanethiol in the presence of 25% KOH and further attached to GPC·TPBNa using DCC/DMAP condensation system.





Figure S1. (a) ¹H NMR and (b) ¹³C NMR (600 MHz, CDCl₃) spectra of C_{18} -S-COOH intermediate.



Figure S2. ¹H NMR (600 MHz, CDCl₃) spectrum of ROS-responsive di-S-PC lipids. ¹H NMR (600 MHz, CDCl₃): δ 5.40 (s, 1H, H-2"), 4.48 (d, J=6.4 Hz, 1H, H-1b"), 4.36

(m, 2H, H-4"), 4.14 (m, 1H, H-1a"), 3.53 (m, 4H, H-3", 5"), 3.46 (s, 4H, H-1, 1'), 3.25 (s, 9H, -N+(CH₃)₃), 2.61 (m, 4H, H-2, 2'), 1.46-1.25 (m, -CH₂-), 0.92 (m, 6H, H-17, 17') ppm.



Figure S3. ¹³C NMR (600 MHz, CH3OH-*d*₄) spectrum of ROS-responsive di-S-PC lipids. ¹³C NMR (500 MHz, CH₃OH-*d*₄): δ 77.82, 77.50, 77.17, 53.96, 49.18, 33.25, 32.65, 31.91, 29.77, 29.02, 22.64, 13.87 ppm.



Figure S4. The *in vitro* cellular association of Dex@FA-ROS-Lips. Confluent LPSactivated RAW264.7 cells exposed to free Dex solution or liposomes containg Dex (50 μ M) at various time intervals.



Figure S5. (a) Expression levels and (b) the quantified densitometric analysis of iRhom2, TNF- α and BAFF proteins detected by Western blotting (n = 3). β -actin antibody was used to ensure after incubation equal protein loading. Data are presented as Mean \pm S.D. and were statistically analyzed by one-way ANOVA. ^{**}P < 0.01 vs

а Arthritis assessment CFA **Drug injections** D 0 D 1 D 9 D 13 D 14 b Score 0 Score 1 Score 2 Score 3 Hind paw K С Fore paw Degree of inflammation Arthritis score 0 No inflammation, no swelling, no redness Hand/foot red 1 +/- slightly inflamed and/or swollen wrist/ankle +/- one inflamed finger Two inflamed fingers 2 +/- red hand/foot and high swollen wrist/ankle At least three inflamed fingers 3 + severe swollen wrist/ankle

the control group; *P < 0.05, **P < 0.01 vs free Dex group.

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