

Electronic Supplementary Information for:

One-Pot Synthesized Organosilica Nanospheres for Multiplexed Fluorescent Nanobarcoding and Subcellular Tracing

Xinfeng Du[†], Yifu Wang[†], Jingying Zhai[†], Chao Guo[†], Yupu Zhang[†], Wenyu Huang[†], Xueqing Ma[†],
Xiaojiang Xie^{†*}

[†]Department of Chemistry, Southern University of Science and Technology, Shenzhen, 518055, China

[†]Academy for Advanced Interdisciplinary Studies, Southern University of Science and Technology, Shenzhen, 518055, China

*Email: xiexj@sustech.edu.cn

Table of Contents:

Figure S1. Hydrodynamic size of the organosilica nanospheres utilizing F127 as template for different reaction time and different amount of F127.

Figure S2. XPS spectra of the organosilica nanospheres containing -C₁₁-N₃, -NH₂, -PEG-N₃, -SH, and -I groups.

Figure S3. FT-IR spectra the organosilica nanospheres containing amine, thiol, and azide groups.

Figure S4. Fluorescence intensity of the organosilica nanospheres loaded with different amount of BODIPY, and Cy5.

Figure S5. CLSM images of HeLa cells after incubated with organosilica nanospheres modified with Rhodamine for different time.

Figure S6. Flow cytometry of the fluorescent NBs in HeLa cells with different incubation time.

Figure S7. CLSM images for colocalization of nanospheres in HeLa cells.

Figure S8. 3D-stacking CLSM images of HeLa cells incubated with FITC-modified nanospheres and Hoechst-33342.

Figure S9. Fluorescence imaging of HeLa cells directly incubated with the dye FITC.

Figure S10. The absorbance and fluorescence spectra of BODIPY modified organosilica nanospheres.

Table S1. Size distribution and PDI of the organosilica nanoparticles prepared with different templates and organosilanes.

Table S2. Size distribution and PDI of organosilica nanospheres before and after 5 months storage.

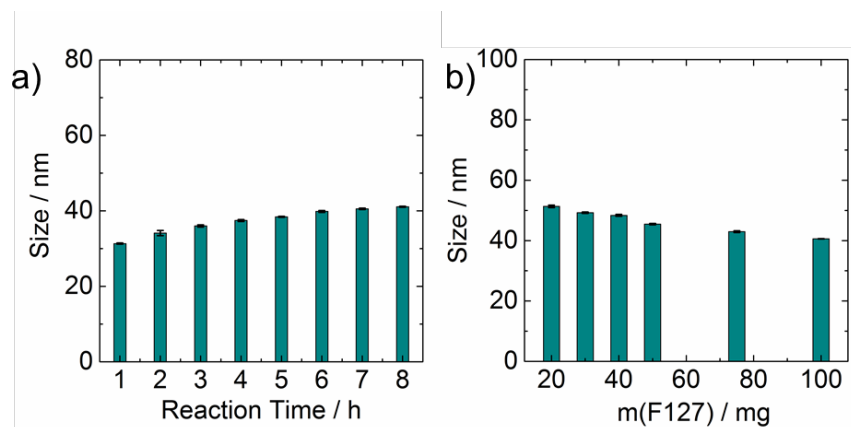


Figure S1. a) Hydrodynamic size of the organosilica nanospheres utilizing F127 as template for different reaction time. b) Hydrodynamic size of the organosilica nanospheres utilizing different amount of F127 as template.

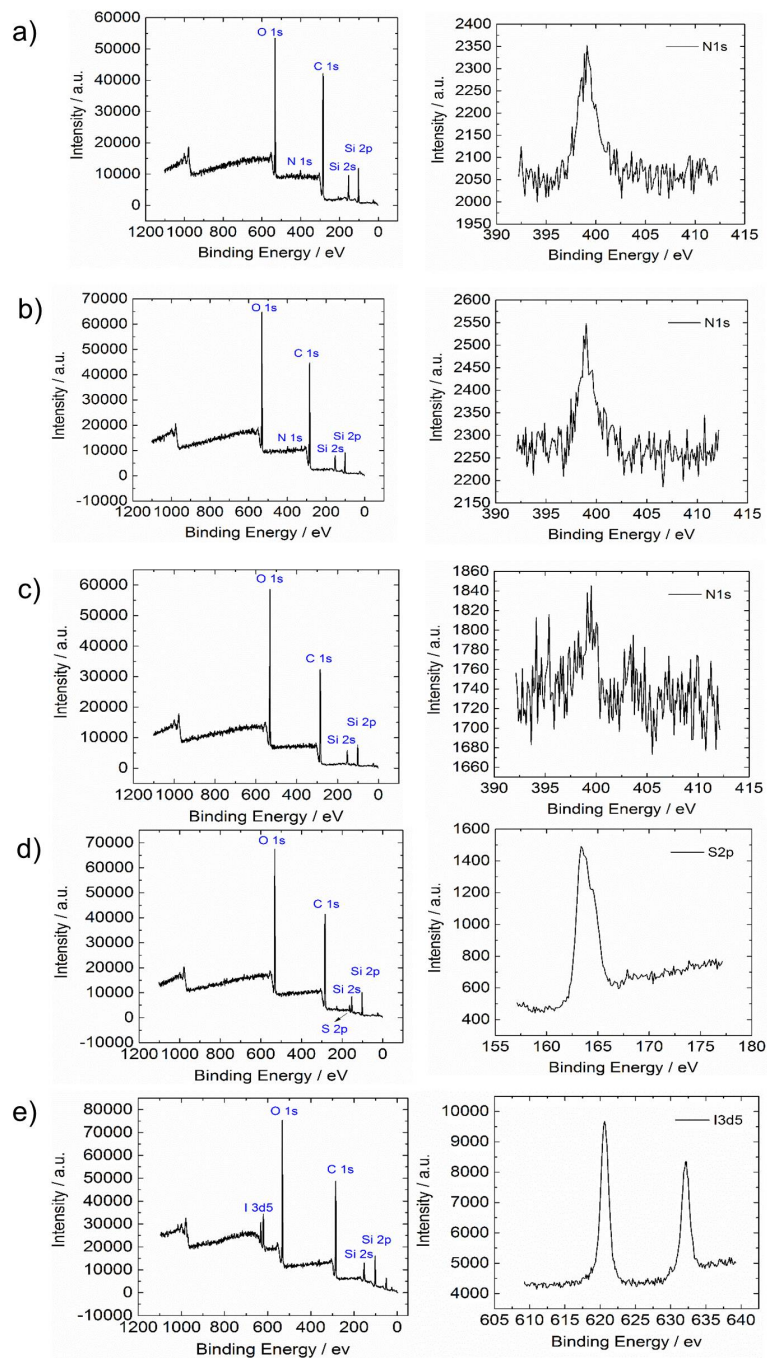


Figure S2. XPS spectra of the organosilica nanospheres containing a) -C₁₁-N₃, b) -NH₂, c) -PEG-N₃, d) -SH, and e) -I groups.

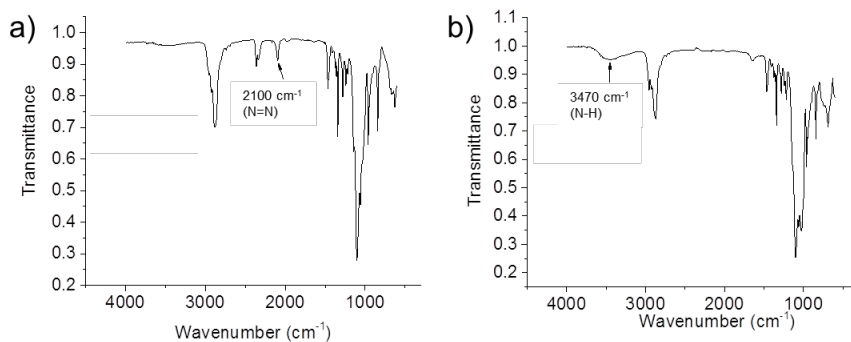


Figure S3. FT-IR spectra of the organosilica nanospheres containing amine, thiol, and azide groups.

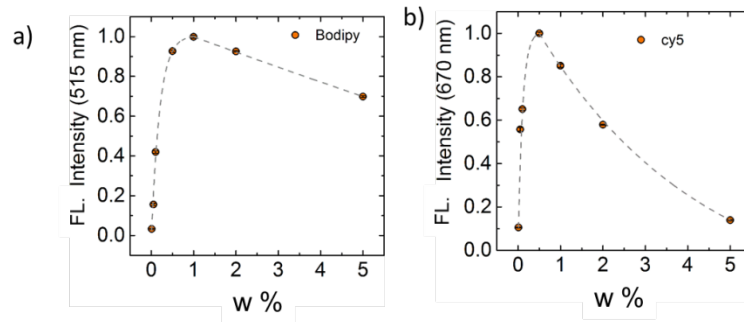


Figure S4. Fluorescence intensity of the organosilica nanospheres loaded with different amount of a) BODIPY, and b) Cy5.

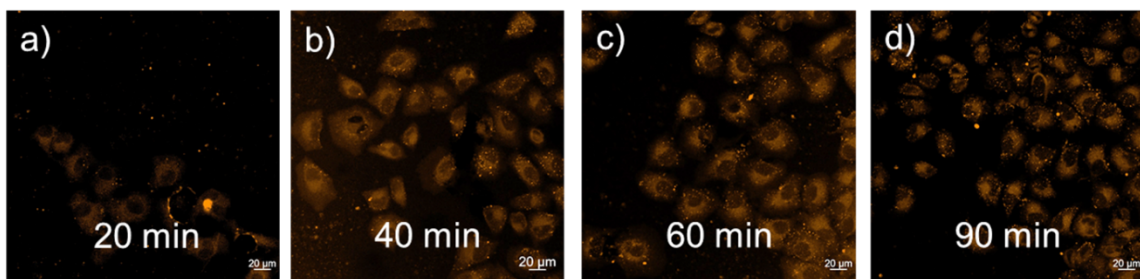


Figure S5. CLSM images of HeLa cells after incubated with organosilica nanospheres modified with Rhodamine for different time. Scale bar: 20 μm.

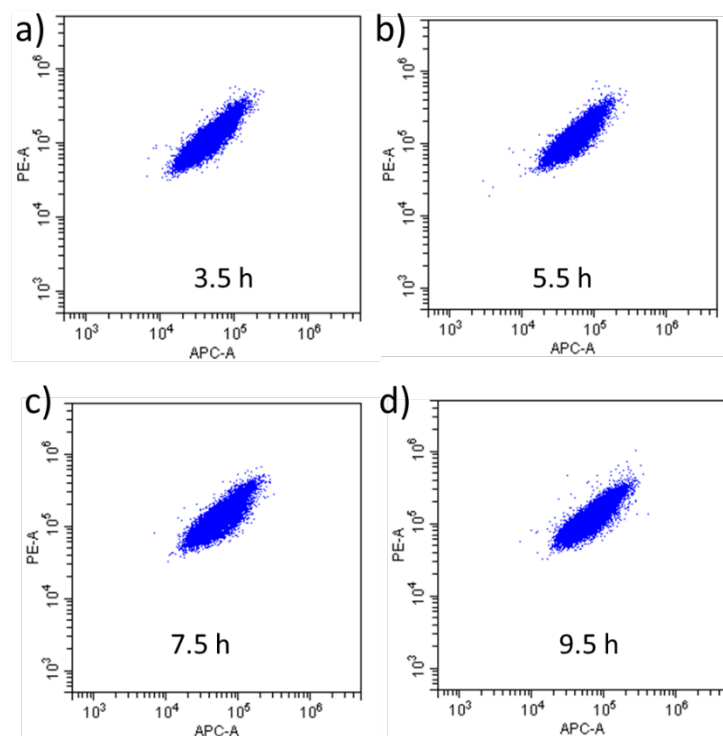


Figure S6. Flow cytometry of the fluorescent NBs in HeLa cells with different incubation time. The NBs were coded with Cy5 and RhITC.

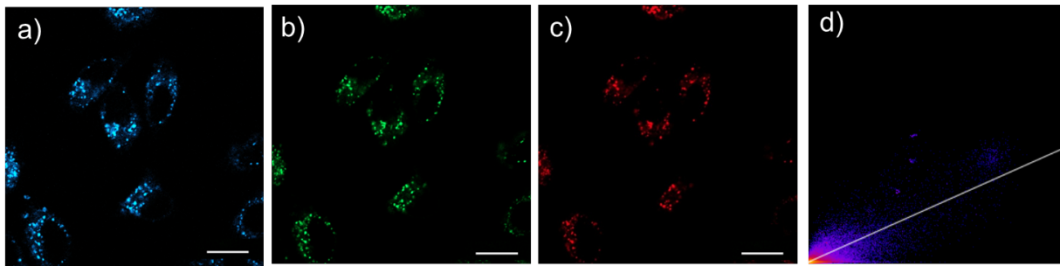


Figure S7. CLSM images of HeLa cells incubated with the organosilica nanospheres modified with MEQ and BODIPY as well as stained with LysoTracker Deep Red. a) Blue channel for MEQ. b) Green channel for BODIPY. c) Red channel for LysoTracker Deep Red. d) Co-localization of green channel and red channel. Scale bar: 20 μm .

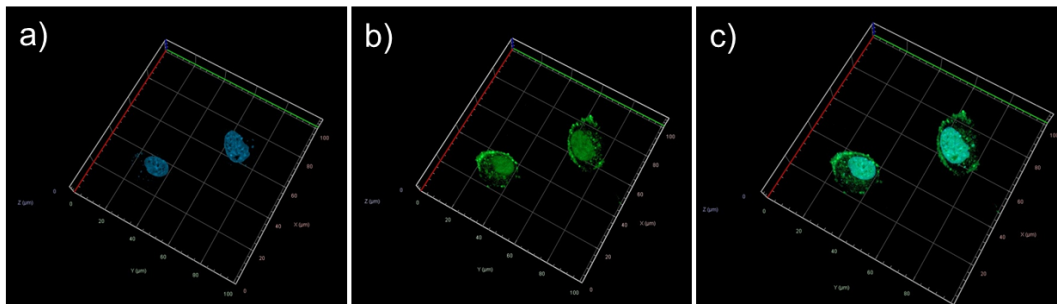


Figure S8. 3D-stacking CLSM images of HeLa cells incubated with FITC-modified nanospheres and Hoechst-33342. a) Blue channel for Hoechst-33342. b) Green channel for FITC. c) Overlay of a) and b)

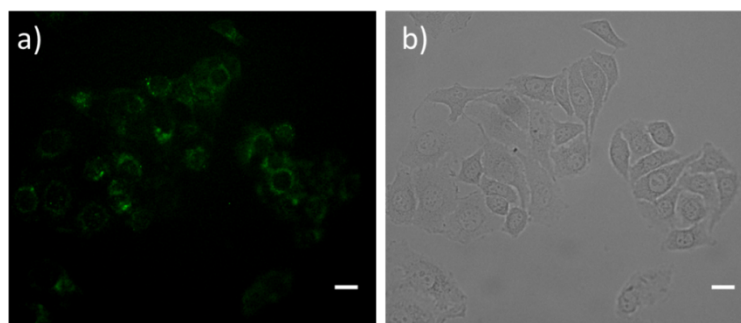


Figure S9. a) Fluorescence imaging of HeLa cells directly incubated with the dye FITC (1 μ M in the culture medium for 3 h). b) Bright field image. Scale bar: 20 μ m.

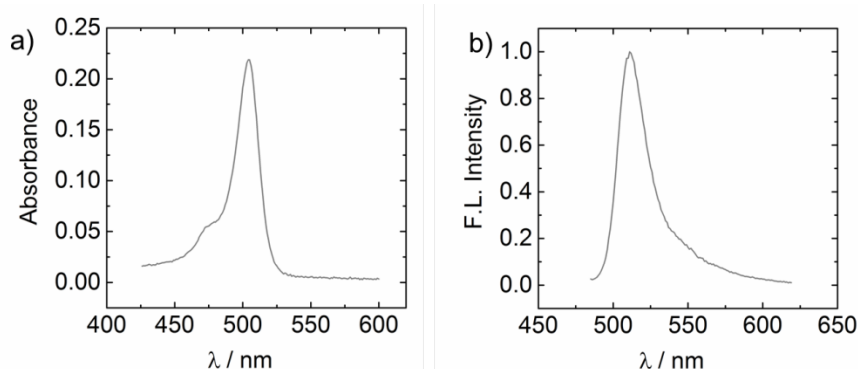


Figure S10. The absorbance and fluorescence spectra of BODIPY modified organosilica nanospheres.

Table S1. Size distribution and PDI of the organosilica nanoparticles prepared with different templates and organosilanes

| Template | Organosilanes | Size / nm | PDI |
|----------|---|-------------------|-------------------|
| F127 | TEOS | 382.7 \pm 22.91 | 0.69 \pm 0.063 |
| F127 | Si-propyl | 52.44 \pm 0.03 | 0.01 \pm 0.001 |
| F127 | Si-phenethyl | 38.53 \pm 0.10 | 0.005 \pm 0.004 |
| F127 | Si-propyl, Si-PEG-N ₃ | 51.68 \pm 0.06 | 0.03 \pm 0.005 |
| F127 | Si-propyl, Si-NH ₂ | 49.35 \pm 0.42 | 0.04 \pm 0.003 |
| F127 | Si-propyl, Si-SH | 52.38 \pm 0.19 | 0.04 \pm 0.014 |
| F127 | Si-propyl, Si-I | 38.61 \pm 0.25 | 0.08 \pm 0.009 |
| F127 | Si-propyl, Si-C ₁₁ -N ₃ | 41.08 \pm 0.62 | 0.15 \pm 0.012 |
| F127 | Si-propyl, Si-epoxy | 55.45 \pm 0.22 | 0.048 \pm 0.016 |
| F127 | Si-propyl, Si-PEG-N ₃ , Si-SH | 51.28 \pm 0.24 | 0.05 \pm 0.006 |
| Triton-X | Si-propyl | 15.61 \pm 0.32 | 0.19 \pm 0.0073 |
| Tween-20 | Si-propyl | 15.47 \pm 0.12 | 0.11 \pm 0.007 |
| PEG | Si-propyl | 202.8 \pm 2.62 | 0.16 \pm 0.008 |
| CTAB | Si-propyl | 22.83 \pm 10.17 | 0.38 \pm 0.151 |
| SDS | Si-propyl | 129.8 \pm 0.53 | 0.11 \pm 0.012 |

Table S2. Size distribution and PDI of organosilica nanospheres before and after 5 months storage.

| Nanospheres | Size / nm | PDI |
|------------------------|------------------|------------------|
| Newly prepared | 52.38 \pm 0.19 | 0.04 \pm 0.014 |
| After 5 months storage | 52.81 \pm 0.35 | 0.03 \pm 0.007 |