

Electronic Supplementary Information

Constrained beta-amino acid-containing miniproteins

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Peptide synthesis. All commercially available reagents and solvents were purchased from Sigma-Aldrich, Merck, Iris Biotech or Lipopharm and used without further purification. Fmoc (1*S*,2*S*)-*trans*-2-aminocyclopentanecarboxylic acid was purchased from the Synnovator and has >95% of purity. Peptide foldamers were obtained with an automated solid-phase peptide synthesizer (Biotage[®] Initiator+Alastra[™]) on H-Rink amide ChemMatrix[®] resin (loading: 0.59 mmol/g) in 100 mg scale. Fmoc deprotection was achieved using 20% piperidine in DMF for 3 + 10 min. A double-coupling procedure was performed with 0.5 M solution of DIC and 0.5 M solution of OXYMA in DMF, for α -amino acids 2 x 15 min, for β -amino acids 30 min, at 75 °C. Acetylation reaction was carried out using NMP/DIPEA/acetic anhydride (80:15:5) mixture. Cleavage of the peptides from the resin was accomplished with the mixture of TFA/TIS/H₂O (95:2.5:2.5) after 3 h of shaking or TFA/TIS/thioanisole/H₂O (85:5:5:5) after 4 h of shaking if in sequence was arginine residue. The crude peptide was precipitated with ice-cold diethyl ether and centrifuged (9000 rpm, 2 x 15 min, 4 °C). Obtained peptide foldamers were purified using the HPLC (Knauer Prep) with a preparative column 250 mm x 30 mm, Thermo Scientific[™] Hypersil GOLD[™] (C18, 12 μ m) and an analytical column 150 mm x 4.6 mm, Kinetex 100A (C18, 5 μ m). Solvents and gradients are given in Table S1. Yield of peptide synthesis was in 5-25% range. All peptides were at least 95% pure according to analytical HPLC.

Mass spectrometry. Peptides were studied by WATERS LCT Premier XE System consisting of high resolution mass spectrometer with electrospray ion source and a time of flight (TOF) analyzer (Table S2).

Circular dichroism. CD spectra were recorded on JASCO J-1500 with parameters: 0.2 nm resolution, 1.0 nm band width, 200 mdeg sensitivity, 4 s response time, 50 nm/min scanning speed. The CD spectra of the solvents alone were recorded and subtracted from the raw data. The CD intensity is given as mean residue molar ellipticity (θ [deg x cm² x dmol⁻¹]). Single CD spectra were measured at 20 °C. Peptides **1-15** were dissolved in water to 1 mg/mL concentration and diluted to 0.25 mg/mL with 100 mM potassium phosphate buffer pH 7.0. Peptides **16-29** were dissolved in 50 mM potassium phosphate buffer pH 7.0 at concentration of 1 mg/mL. Measurements were done in 0.02 cm cuvette path length and with 10 scans.

Measurement of T_m values. Solutions containing 0.25-1.00 mg/mL peptide in 100 mM potassium phosphate buffer pH 7.0 were used. Ellipticity was recorded at 222 nm using 1 mm

optical path length cuvette. The temperature was increased from 4 to 96 °C in increments of 2 °C. The melting point (T_m), was determined assuming:

$$Q_{obs} = \alpha Q_f + (1 - \alpha) Q_u$$

where: Q_{obs} – observed ellipticity, Q_f – ellipticity of folded miniprotein, Q_u – ellipticity of unfolded miniprotein, α – degree of folding;

Folded and unfolded ellipticities are linear function of temperature (T):

$$Q_f = m_f T + b_f$$

$$Q_u = m_u T + b_u$$

Where: m_f , b_f , m_u and b_u are coefficients of linear dependences.

Subsequently:

$$\alpha = \frac{1}{1 + K}$$

$$K = e^{-\frac{\Delta G^0}{RT}}$$

where: K – equilibrium constant, ΔG^0 - Gibbs free energy, R – gas constant.

Assuming heat capacity equal zero for mini-protein, Gibbs free energy can be evaluated as [S1]:

$$\Delta G^0 = \Delta H_m \left(1 - \frac{T}{T_m} \right)$$

where: ΔH_m – enthalpy; T_m – melting point temperature.

Combining all above equations, the dependence of observed ellipticity (Q_{obs}) on temperature could be obtained:

$$Q_{obs} = \frac{1}{1 + e^{-\frac{\Delta H_m \cdot \left(1 - \frac{T}{T_m} \right)}{RT}}} \cdot (b_f + m_f \cdot T - b_u - m_u \cdot T) + b_u + m_u \cdot T$$

This equation was used in non-linear fitting algorithm of the OriginPro9 program, to fit to experimental data and obtain T_m value. In most cases m_u value was assumed to be zero.

Temperature and guanidine-dependent denaturation. To examine the thermal unfolding of the peptide, stock solutions were prepared containing 0.25 mg/mL peptide in 100 mM potassium phosphate buffer pH 7.0, with 11 different concentrations of GuHCl. Each sample was incubated at room temperature for 1.5h. Ellipticity measurements were recorded at 222 nm with 1 mm path length cuvette and 3 independent scans. Others parameters were unchanged. The temperature was increased from 4 to 96 °C in increments of 2 °C.

The results for measurements with guanidine hydrochloride were analyzed using MatLab R2016a (The MathWorks, Inc.) and assuming following dependences [S2]. The observed ellipticity (Q_{obs}) depends in the equilibrium constant K and the ellipticity of folded (Q_f) and unfolded (Q_u) states.

$$Q_{obs} = \frac{1}{1 + K}(Q_u \cdot K + Q_f)$$

Parameters Q_f and Q_u depend on temperature and concentration of denaturing agent ($[GuHCl]$):

$$Q_u = a + bT + c[GuHCl]$$

$$Q_f = d + eT + f[GuHCl]$$

Where: a, b, c, d, e and f are coefficients in these linear dependences.

The equilibrium constant K is associated with the Gibbs free energy of peptide unfolding process (ΔG^0):

$$K = e^{\frac{-\Delta G^0}{RT}}$$

However, the value of ΔG^0 is determined by the equation:

$$\Delta G^0 = \Delta H^0 - T\Delta S^0 + \Delta C_p \left(T - T_0 + T \ln \left(\frac{T}{T_0} \right) \right) - m[GuHCl]$$

Nuclear magnetic resonance. The NMR experiments were performed on Bruker Avance III 600 MHz spectrometer using a 5 mm inverse detection QCI cryoprobe. The NMR spectra of 1mg of peptide **13** dissolved in 450 μ L of 50 mM phosphate buffer and 50 μ M D_2O of pH 7

were recorded at temperature 285 K. The temperature was controlled to ± 0.1 K. TOCSY and NOESY experiments were performed for chemical shift and structure assignment. All NMR spectra were acquired with suppression of solvent OH signal using the excitation sculpting pulse. Typical TOCSY - homonuclear Hartman (Hahn transfer using mlev17 sequence for mixing using two power levels for excitation and spinlock) and 2D NOESY were recorded in phase sensitive mode with the spectral width of 6009 Hz in both dimensions using 2048 data points and relaxation delay of 1.5s. These spectra were acquired with 1024 increments of 22 scans for TOCSY and 104 scans for 2D NOESY. Mixing times were set at 100 ms and 200 ms for TOCSY and 2D NOESY experiments, respectively. The data were acquired and processed using Topspin 3.1 (BrukerBioSpin, Rheinstetten, Germany). The processed spectra were assigned with the help of the SPARKY program (Goddard, T.D.; Kneller, D.G. *Sparky*, 3rd ed.; University of California; San Francisco, CA, USA 2001).

NMR structure calculation. All chemical shifts assigned to **13** can be seen in Table S3. Intensities of NOE signals were normalized to the distance between tryptophan's *beta* hydrogens and classified as follows: 1.8 – 2.5 Å: strong, 2.5 – 3.5 Å: medium, 3.5 – 4.5: weak (Table S4). Restraints to ambiguous CH₂ protons of trans-ACPC residues were omitted, while protons attached to the same carbon atoms were assumed to be a pseudo-atom. Structures were modelled using XPLOR-NIH software and images prepared in Discovery Studio Visualizer. Initially, 50 random conformations of a peptide were generated. Restraints derived from NOE contacts with upper distance limits: strong 2.5 Å (s), medium 3.5 Å (m) and weak 5 Å (w), and the lower distance limit set to 1.8 Å, were applied. Standard protocol for NMR structure calculations implemented in Xplor-NIH was used with the following steps: a) high temperature dynamics (3500 K, 800 ps or 8000 steps), b) simulated annealing performed from 3500 K to 25 K with 12.5 K step, at each temperature short dynamics was done (100 steps or 0.2 ps); c) gradient minimization of final structure. Finally, top lower energy structures were superimposed. Structure was deposited in RCSB Protein Data Bank.

Table S1. HPLC data for studied peptides

No.	Analytical HPLC		Preparative HPLC	
	Retention time [min]	gradient	Retention time [min]	Gradient
1	10.086	1	30.71	4
2	11.004	1	34.85	4
3	9.797	1	28.57	4
4	10.577	1	31.72	4
5	9.667	1	28.56	4
6	10.330	1	32.29	4
7	9.728	1	28.41	4
8	9.110	1	26.80	4
9	11.479	1	35.73	4
10	8.260	2	22.73	5
11	8.400	2	31.61	4
12	8.180	2	27.70	5
13	9.020	2	41.03	5
14	10.067	2	33.92	6
15	11.744	1	31.83	5
16	7.180	3	22.22	7
17	7.340	3	23.25	7
18	7.100	3	22.53	7
19	7.347	3	24.17	7
20	7.207	3	23.24	7
21	6.980	3	22.08	7
22	7.173	3	21.60	7
23	7.400	3	22.31	7
24	7.693	3	22.78	7
25	7.273	3	22.08.	7
26	7.420	3	22.87	7
27	7.475		23.27	7
28	7.460	3	23.26	7
29	7.547	3	23.80	7

Analytical HPLC. Solvent A: 99.95% H₂O, 0.05% TFA. Solvent B: 99.95% CH₃CN, 0.05% TFA.

(1) 1: Total flow: 0.9 mL/min, $t = 0$ min – 90% A, $t = 2$ min – 90% A, $t = 20$ min – 10% A, $t = 23$ min – 10% A, $t = 26$ min – 90 % A.

(2) Total flow: 1 mL/min, $t = 0$ min – 90% A, $t = 2$ min – 90% A, $t = 11$ min – 10% A, $t = 12$ min – 10% A, $t = 15$ min – 90 % A.

(3) Total flow: 1 mL/min, $t = 0$ min – 90% A, $t = 2$ min – 90% A, $t = 10$ min – 10% A, $t = 15$ min – 10% A, $t = 20$ min – 90 % A.

Preparative HPLC. Solvent A: 99.95% H₂O, 0.05% TFA. Solvent B: 99.95% CH₃CN, 0.05% TFA

(4): Total flow: 10 mL/min, $t = 0$ min – 90% A, $t = 6$ min – 90% A, $t = 14$ min – 80% A, $t = 20$ min – 70% A, $t = 38$ – 50% A, $t = 40$ min – 50% A, $t = 48$ min – 10% A, $t = 50$ min – 10% A, $t = 56$ min – 90% A, $t = 60$ – 90% A.

(5): Total flow: 10 mL/min, $t = 0 \text{ min} - 90\% \text{ A}$, $t = 6 \text{ min} - 90\% \text{ A}$, $t = 12 \text{ min} - 70\% \text{ A}$, $t = 20 \text{ min} - 70\% \text{ A}$, $t = 38 - 60\% \text{ A}$, $t = 40 \text{ min} - 60\% \text{ A}$, $t = 48 \text{ min} - 10\% \text{ A}$, $t = 50 \text{ min} - 10\% \text{ A}$, $t = 60 \text{ min} - 90\% \text{ A}$.

(6): Total flow: 10 mL/min, $t = 0 \text{ min} - 90\% \text{ A}$, $t = 6 \text{ min} - 90\% \text{ A}$, $t = 12 \text{ min} - 70\% \text{ A}$, $t = 20 \text{ min} - 60\% \text{ A}$, $t = 68 - 20\% \text{ A}$, $t = 70 \text{ min} - 20\% \text{ A}$, $t = 78 \text{ min} - 10\% \text{ A}$, $t = 80 \text{ min} - 10\% \text{ A}$, $t = 90 \text{ min} - 90\% \text{ A}$.

(7) Total flow: 10 mL/min, $t = 0 \text{ min} - 90\% \text{ A}$, $t = 6 \text{ min} - 90\% \text{ A}$, $t = 23 \text{ min} - 50\% \text{ A}$, $t = 25 \text{ min} - 10\% \text{ A}$, $t = 30 - 10\% \text{ A}$, $t = 33 \text{ min} - 90\% \text{ A}$.

Table S2. Description of MS spectra of peptides **1-29**

peptide	MS found [m/z]	MS theoretical [m/z]	Difference for major peak [ppm]
1	1106.0737 ([M+2H] ²⁺), 737.7387 ([M+3H] ³⁺),	1106.0732 ([M+2H] ²⁺), 737.7181 ([M+3H] ³⁺),	0.5
2	1104.5883 ([M+2H] ²⁺), 744.0534 ([M+2H+Na/K] ³⁺)	1104.5861 ([M+2H] ²⁺), 744.0534 ([M+2H+Na/K] ³⁺)	2.0
3	1105.0653 ([M+2H] ²⁺),	1105.0654 ([M+2H] ²⁺),	-0.1
4	1080.0745 ([M+2H] ²⁺),	1080.0758 ([M+2H] ²⁺),	-1.2
5	1105.1063 ([M+2H] ²⁺),	1105.0651 ([M+2H] ²⁺),	37.3
6	1097.5776 ([M+2H] ²⁺),	1097.5782 ([M+2H] ²⁺),	-0.5
7	1068.5680 ([M+2H] ²⁺),	1068.5675 ([M+2H] ²⁺),	0.5
8	1105.0659 ([M+2H] ²⁺),	1105.0654 ([M+2H] ²⁺),	0.5
9	1097.5586 ([M+2H] ²⁺), 744.7065 ([M+2H+Na/K] ³⁺)	1097.5601 ([M+2H] ²⁺), 744.694 ([M+2H+Na/K] ³⁺)	-1.4
10	1161.6075 ([M+2H] ²⁺), 774.7787 ([M+3H] ³⁺), 782.0988 ([M+2H+Na/K] ³⁺)	1161.6075 ([M+2H] ²⁺), 774.7409 ([M+3H] ³⁺), 782.0678 ([M+2H+Na/K] ³⁺)	0.0
11	1161.6062 ([M+2H] ²⁺), 782.0955 ([M+2H+Na/K] ³⁺)	1161.6075 ([M+2H] ²⁺), 782.0678 ([M+2H+Na/K] ³⁺)	-1.1
12	1161.6080 ([M+2H] ²⁺),	1161.6075 ([M+2H] ²⁺),	0.4
13	1103.5790 ([M+2H] ²⁺), 736.0533 ([M+3H] ³⁺),	1103.5782 ([M+2H] ²⁺), 736.0547 ([M+3H] ³⁺),	0.7
14	1096.5521 ([M+2H] ²⁺), 1107.5481 ([M+H+Na/K] ²⁺), 744.0598 ([M+2H+Na/K] ³⁺)	1096.5521 ([M+2H] ²⁺), 1107.5432 ([M+H+Na/K] ²⁺), 743.6863 ([M+2H+Na/K] ³⁺)	0.0
15	1095.0654 ([M+2H] ²⁺), 730.4116 ([M+3H] ³⁺), 743.0613 ([M+2H+Na/K] ³⁺)	1095.0649 ([M+2H] ²⁺), 730.3792 ([M+3H] ³⁺), 743.0306 ([M+2H+Na/K] ³⁺)	0.5
16	1177.3290 ([M+3H] ³⁺), 1209.9323 ([M+Na+2K] ³⁺), 883.2201 ([M+4H] ⁴⁺), 907.7009 ([M+Na+2K] ⁴⁺), 706.8076 ([M+5H] ⁵⁺)	1177.3188 ([M+3H] ³⁺), 1209.9493 ([M+Na+2K] ³⁺), 883.2411 ([M+4H] ⁴⁺), 907.4648 ([M+Na+2K] ⁴⁺), 706.7944 ([M+5H] ⁵⁺)	8.7
17	1171.3730 ([M+3H] ³⁺), 1204.014 ([M+Na+2K] ³⁺), 878.7485 ([M+4H] ⁴⁺), 903.0092 ([M+Na+2K] ⁴⁺), 703.2338 ([M+5H] ⁵⁺)	1171.3274 ([M+3H] ³⁺), 1203.9635 ([M+Na+2K] ³⁺), 878.7475 ([M+4H] ⁴⁺), 902.9656 ([M+Na+2K] ⁴⁺), 703.1996 ([M+5H] ⁵⁺)	38.9
18	1176.7262 ([M+3H] ³⁺), 1209.3905 ([M+Na+2K] ³⁺), 882.4794 ([M+4H] ⁴⁺), 906.9955 ([M+Na+2K] ⁴⁺), 706.3795 ([M+5H] ⁵⁺)	1176.6470 ([M+3H] ³⁺), 1209.282 ([M+Na+2K] ³⁺), 882.7372 ([M+4H] ⁴⁺), 906.9577 ([M+Na+2K] ⁴⁺), 706.3913 ([M+5H] ⁵⁺)	67.3
19	1162.2844 ([M+3H] ³⁺), 1194.6675 ([M+Na+2K] ³⁺), 871.7327 ([M+4H] ⁴⁺), 896.0294 ([M+Na+2K] ⁴⁺),	1162.3080 ([M+3H] ³⁺), 1194.9363 ([M+Na+2K] ³⁺), 871.9829 ([M+4H] ⁴⁺), 896.2068 ([M+Na+2K] ⁴⁺),	-20.3

	697.7919 ([M+5H] ⁵⁺)	697.7879 ([M+5H] ⁵⁺)	
20	1176.0098 ([M+3H] ³⁺), 1208.7823 ([M+Na+2K] ³⁺), 882.0013 ([M+4H] ⁴⁺), 906.7006 ([M+Na+2K] ⁴⁺), 706.0279 ([M+5H] ⁵⁺)	1175.9993 ([M+3H] ³⁺), 1208.6347 ([M+Na+2K] ³⁺), 882.2515 ([M+4H] ⁴⁺), 906.472 ([M+Na+2K] ⁴⁺), 706.0027 ([M+5H] ⁵⁺)	8.9
21	1176.6498 ([M+3H] ³⁺), 882.4855 ([M+4H] ⁴⁺), 706.1829 ([M+5H] ⁵⁺)	1176.6470 ([M+3H] ³⁺), 882.7372 ([M+4H] ⁴⁺), 706.3913 ([M+5H] ⁵⁺)	2.4
22	1171.3411 ([M+3H] ³⁺), 1203.964 ([M+Na+2K] ³⁺), 878.7408 ([M+4H] ⁴⁺), 903.0154 ([M+Na+2K] ⁴⁺), 703.2175 ([M+5H] ⁵⁺)	1171.3274 ([M+3H] ³⁺), 1203.9635 ([M+Na+2K] ³⁺), 878.7475 ([M+4H] ⁴⁺), 902.9656 ([M+Na+2K] ⁴⁺), 703.1996 ([M+5H] ⁵⁺)	11.7
23	1171.6315 ([M+3H] ³⁺), 878.9778 ([M+4H] ⁴⁺), 703.3821 ([M+5H] ⁵⁺)	1171.6433 ([M+3H] ³⁺), 878.9844 ([M+4H] ⁴⁺), 703.3891 ([M+5H] ⁵⁺)	-10.1
24	1171.6426 ([M+3H] ³⁺), 1203.3256 ([M+Na+2K] ³⁺), 878.7353 ([M+4H] ⁴⁺), 902.7455 ([M+Na+2K] ⁴⁺), 703.376 ([M+5H] ⁵⁺)	1171.6433 ([M+3H] ³⁺), 1204.2786 ([M+Na+2K] ³⁺), 878.9844 ([M+4H] ⁴⁺), 903.214 ([M+Na+2K] ⁴⁺), 703.3891 ([M+5H] ⁵⁺)	-0.6
25	1195.3237 ([M+3H] ³⁺), 1227.6448 ([M+Na+2K] ³⁺), 896.493 ([M+4H] ⁴⁺), 921.4927 ([M+Na+2K] ⁴⁺), 717.6011 ([M+5H] ⁵⁺)	1195.3345 ([M+3H] ³⁺), 1227.9695 ([M+Na+2K] ³⁺), 896.7528 ([M+4H] ⁴⁺), 920.9735 ([M+Na+2K] ⁴⁺), 717.6038 ([M+5H] ⁵⁺)	-9.0
26	1162.3097 ([M+3H] ³⁺), 1195.3408 ([M+Na+2K] ³⁺), 871.737 ([M+4H] ⁴⁺), 896.2319 ([M+Na+2K] ⁴⁺), 697.7875 ([M+5H] ⁵⁺)	1162.3080 ([M+3H] ³⁺), 1194.9387 ([M+Na+2K] ³⁺), 871.9829 ([M+4H] ⁴⁺), 896.2037 ([M+Na+2K] ⁴⁺), 697.7879 ([M+5H] ⁵⁺)	1.5
27	1165.6833 ([M+3H] ³⁺), 874.2466 ([M+4H] ⁴⁺), 699.6055 ([M+5H] ⁵⁺)	1165.6519 ([M+3H] ³⁺), 874.4909 ([M+4H] ⁴⁺), 699.7943 ([M+5H] ⁵⁺)	26.9
28	1155.6561 ([M+3H] ³⁺), 1188.6327 ([M+Na+2K] ³⁺), 866.7495 ([M+4H] ⁴⁺), 891.252 ([M+Na+2K] ⁴⁺), 693.5889 ([M+5H] ⁵⁺)	1155.6447 ([M+3H] ³⁺), 1188.2752 ([M+Na+2K] ³⁺), 866.9855 ([M+4H] ⁴⁺), 891.2064 ([M+Na+2K] ⁴⁺), 693.7899 ([M+5H] ⁵⁺)	9.9
29	1155.9686 ([M+3H] ³⁺), 1188.6478 ([M+Na+2K] ³⁺), 866.9707 ([M+4H] ⁴⁺), 891.548 ([M+Na+2K] ⁴⁺), 694.149 ([M+5H] ⁵⁺)	1155.9606 ([M+3H] ³⁺), 1188.7028 ([M+Na+2K] ³⁺), 867.2224 ([M+4H] ⁴⁺), 891.527 ([M+Na+2K] ⁴⁺), 693.9794 ([M+5H] ⁵⁺)	6.9

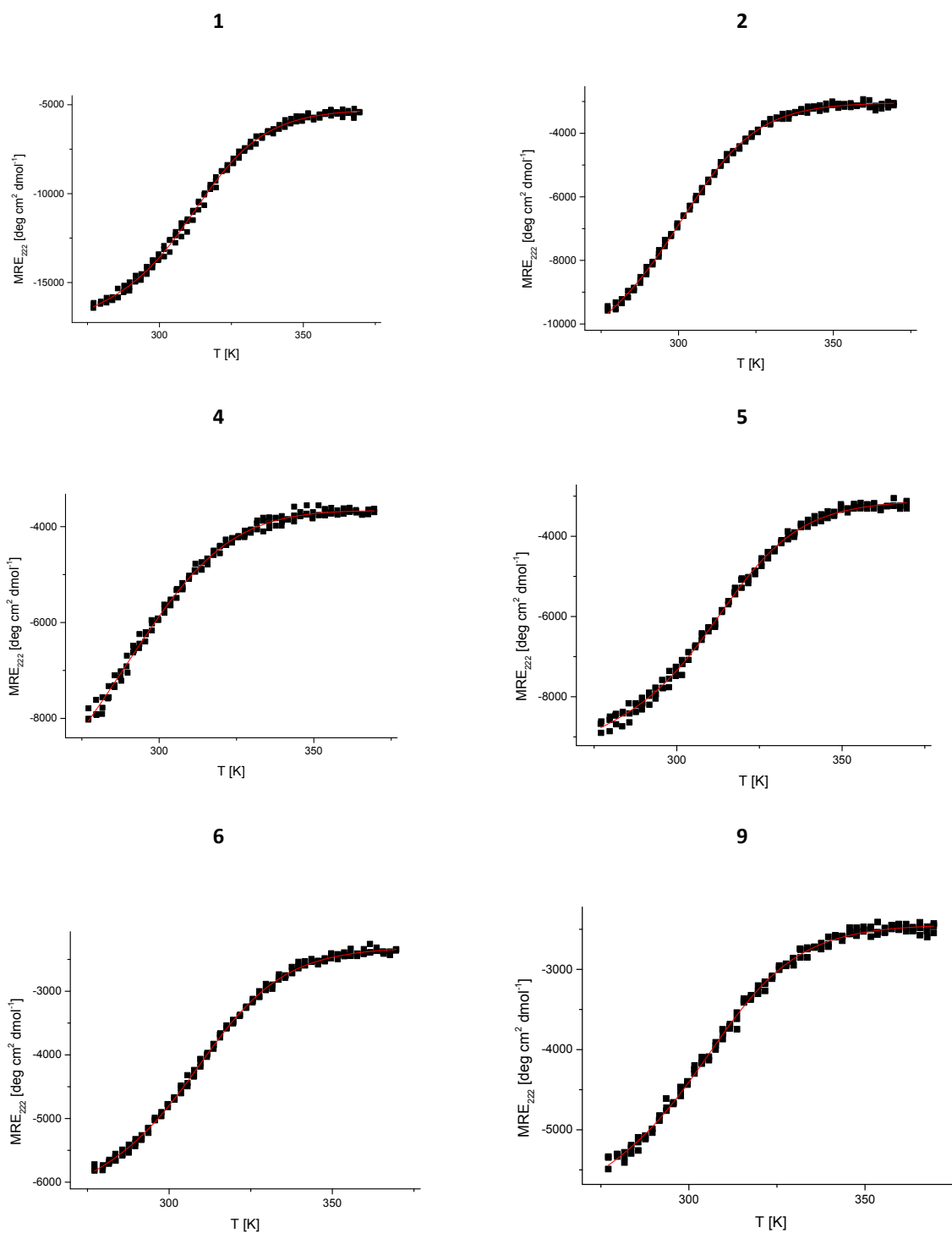


Figure S1. Plots of ellipticity (Θ_{222}) versus temperature (T) for peptides 1, 2, 4-6, 9, for which it was possible to determine T_m .

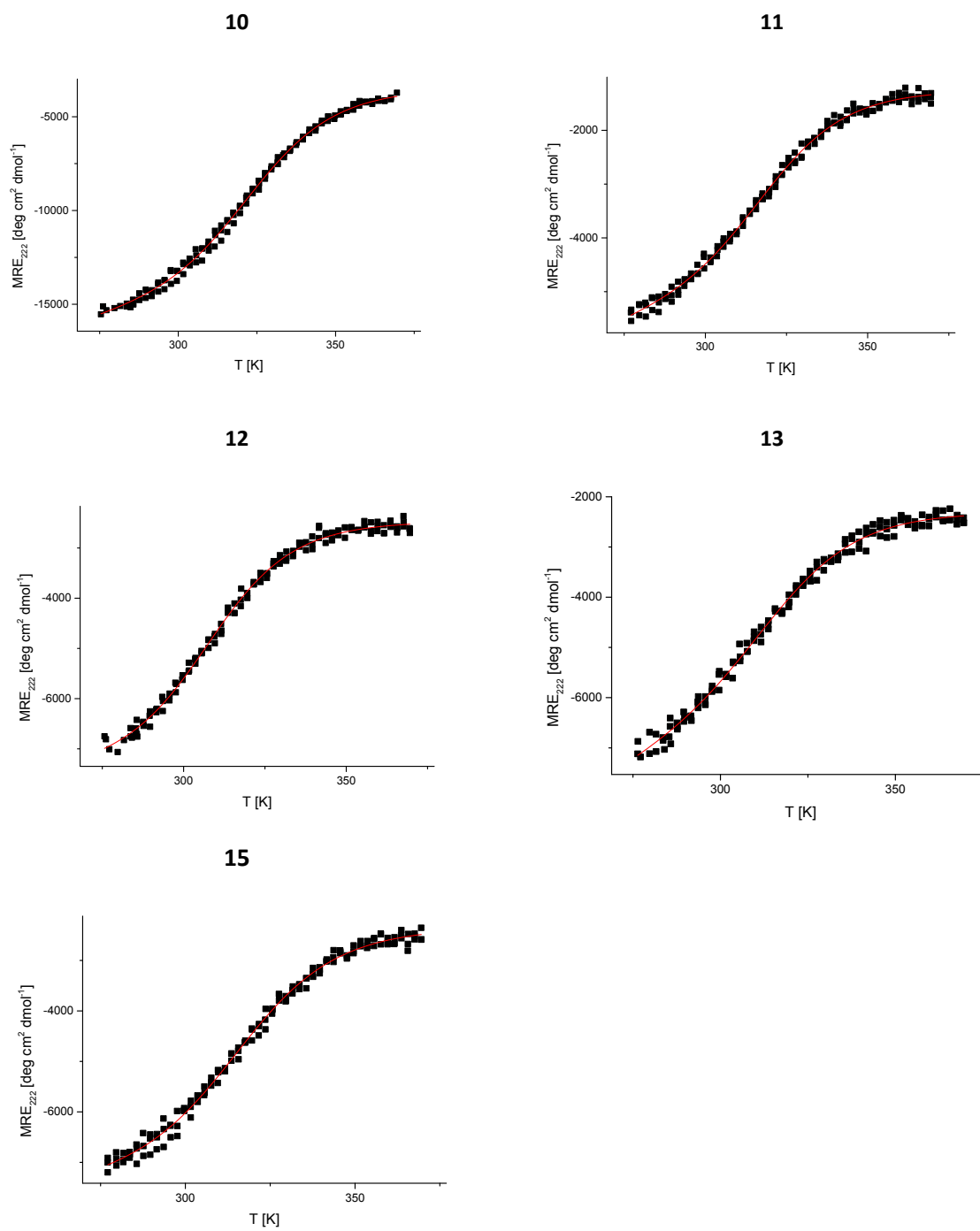


Figure S2. Plots of ellipticity (Θ_{222}) versus temperature (T) for peptides **10**, **11-13**, and **15**, for which it was possible to determine T_m .

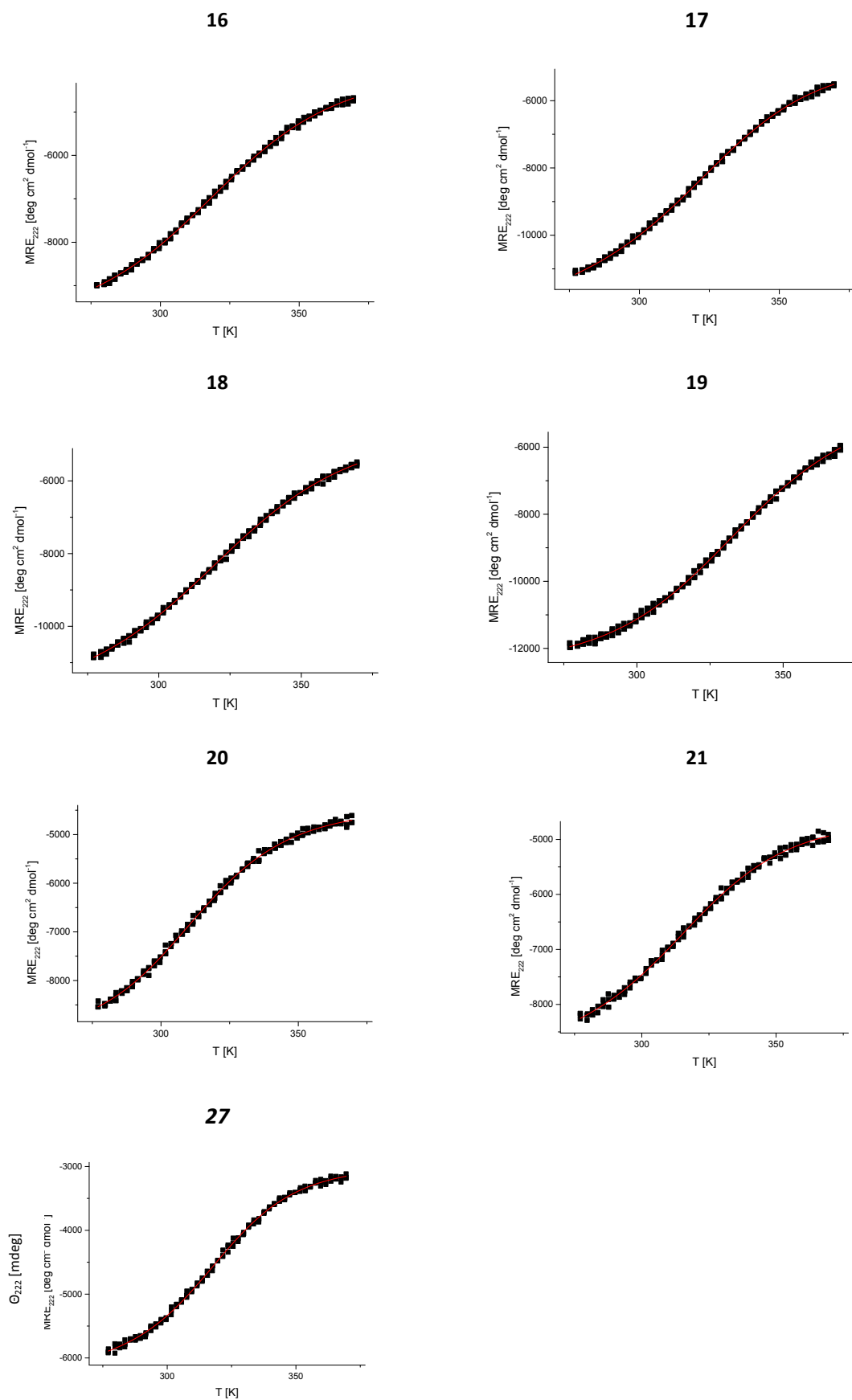
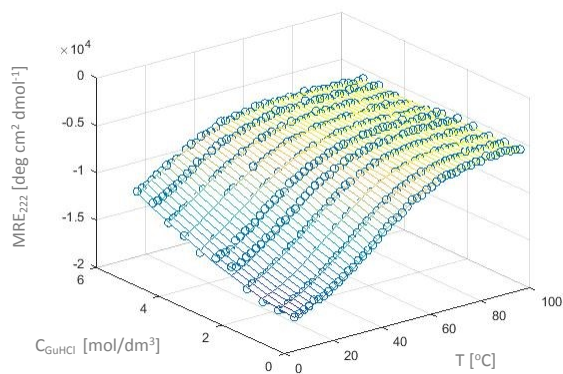
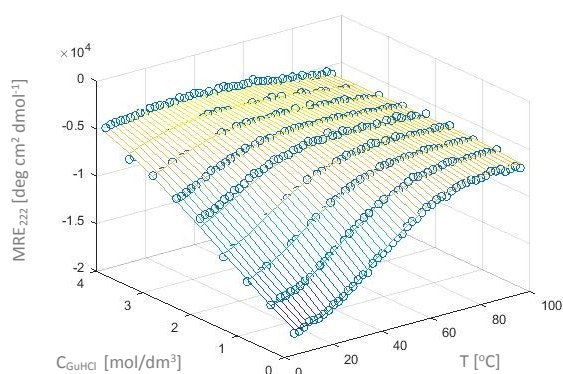


Figure S3. Plots of ellipticity (Θ_{222}) versus temperature (T) for peptides 16-21 and 27, for which it was possible to determine T_m .



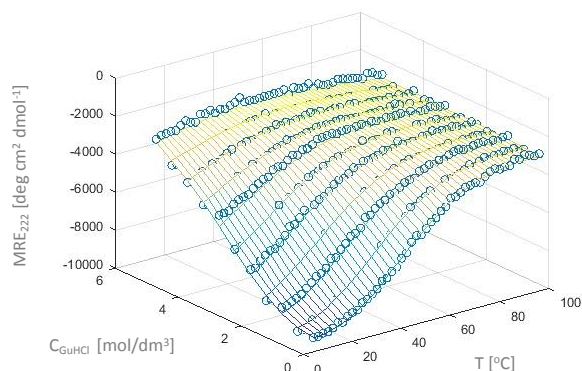
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ΔG°	0.30 ± 0.16 kcal/mol
ΔH°	10.6 ± 0.8 kcal/mol
$T\Delta S^\circ$	10.1 ± 0.6 kcal/mol
ΔC_p	0.16 ± 0.03 kcal/mol·K
m	0.23 ± 0.02 kcal/mol·M
K	0.60 ± 0.16



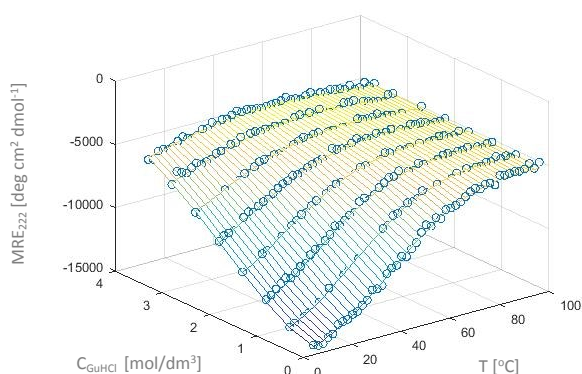
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ΔG°	0.48 ± 0.30 kcal/mol
ΔH°	11.7 ± 1.6 kcal/mol
$T\Delta S^\circ$	11.3 ± 1.3 kcal/mol
ΔC_p	-0.01 ± 0.05 kcal/mol·K
m	0.40 ± 0.04 kcal/mol·M
K	0.43 ± 0.18



11

ΔG°	0.28 ± 0.17 kcal/mol
ΔH°	9.6 ± 0.7 kcal/mol
$T\Delta S^\circ$	9.3 ± 0.6 kcal/mol
ΔC_p	0.12 ± 0.03 kcal/mol·K
m	0.38 ± 0.04 kcal/mol·M
K	0.61 ± 0.14



13

ΔG°	0.24 ± 0.07 kcal/mol
ΔH°	8.4 ± 0.6 kcal/mol
$T\Delta S^\circ$	8.2 ± 0.6 kcal/mol
ΔC_p	0.14 ± 0.03 kcal/mol·K
m	0.44 ± 0.05 kcal/mol·M
K	0.66 ± 0.10

Figure S4. CD signal at 222nm monitored in a function of guanidine hydrochloride concentration and temperature for peptide **1**, **5**, **11**, and **13** (experimental data are shown as dark green circles, while fitted thermodynamic parameters for the folding equilibrium are represented by the surface). Values of ΔG° , $T\Delta S^\circ$ and K are given for 293K.

Table S3. Chemical shifts of peptide **13** in 50mM phosphate buffer, pH 7. Cp stands for *trans*-(1*S*,2*S*)-2-amino-1-cyclopentanecarboxylic acid. Residue no 1 is acetyl group at *N*-terminus.

Residue	Proton	Chemical shift [ppm]
Ac 1	HA	1.81
Cp2	HN	8.10
	HA	2.48
	HB	3.98
	HG,HD,HE	1.45(4)
		1.52(3)
1.55(2)		
1.85(1)		
Leu3	HN	8.27
	HA	3.91
	HB	1.43
	HG	1.17
	HD	0.61(2) 0.69(1)
Tyr4	HN	8.10
	HA	4.06
	HB	2.80(2) 3.05(1)
	HD	6.79
	HE	6.53
Cp5	HN	8.08
	HA	2.38
	HB	4.05
	HG,HD,HE	1.40(5)
		1.53(4)
1.65(3)		
1.76(2) 1.88(1)		
Gln6	HN	7.93
	HA	3.61
	HB	1.86(2), 1.91(1)
	HG	2.12
	HN'	6.67(2) 7.37(1)
Trp7	HN	7.81
	HA	4.14
	HB	2.96(2) 3.59(1)
	HD1	6.83
	HE3	6.87
	HH2	6.92
	HZ2	7.00
	HZ3	6.82
HE1	9.48	
Leu8	HN	8.20
	HA	3.19
	HB	1.42(2) 1.51(1)
	HG	1.13
	HD	0.61(2) 0.69(1)
Cp9	HN	8.19
	HA	2.57
	HB	3.89
	HG,HD,HE	1.45(3)
1.65(2)		
1.71(1)		
Asp10	HN	7.96
	HA	4.23
	HB	2.57(2) 2.64(1)
Gly11	HN	7.05
	HA	3.26(2) 3.85(1)
Gly12	HN	8.50
	HA	0.80(1) 2.99(2)
Pro13	HA	4.34
	HB	2.23(1) 1.82(2)
	HG	1.90
	HD	3.22(2) 3.55(1)
Ser14	HN	7.59
	HA	4.17
	HB	3.66(2) 3.68(1)
Ser15	HN	8.02
	HA	3.93
	HB	3.38(2) 3.65(1)
Gly16	HN	7.71
	HA	3.61(2) 3.98(1)
Arg17	HN	8.00
	HA	4.70
	HB	1.58(2) 1.64(1)
	HG	1.42
	HD	3.02
	HN'	6.83(2)

		7.34(1)
Pro18	HA	4.49
	HB	2.09(1), 1.56(2)
	HG	1.76
	HD	3.42(2) 3.62(1)
Pro19	HA	3.26
	HB	0.21(2) 1.09(1)
	HG	1.40(2) 1.50(1)

	HD	2.53
Pro20	HA	4.12
	HB	2.00
	HG	1.66
	HD	2.78(2) 3.03(1)
Ser21	HN	8.05
	HA	4.07
	HB	3.56(2) 3.60(1)
NH ₂	HN1	6.91
	HN2	7.42

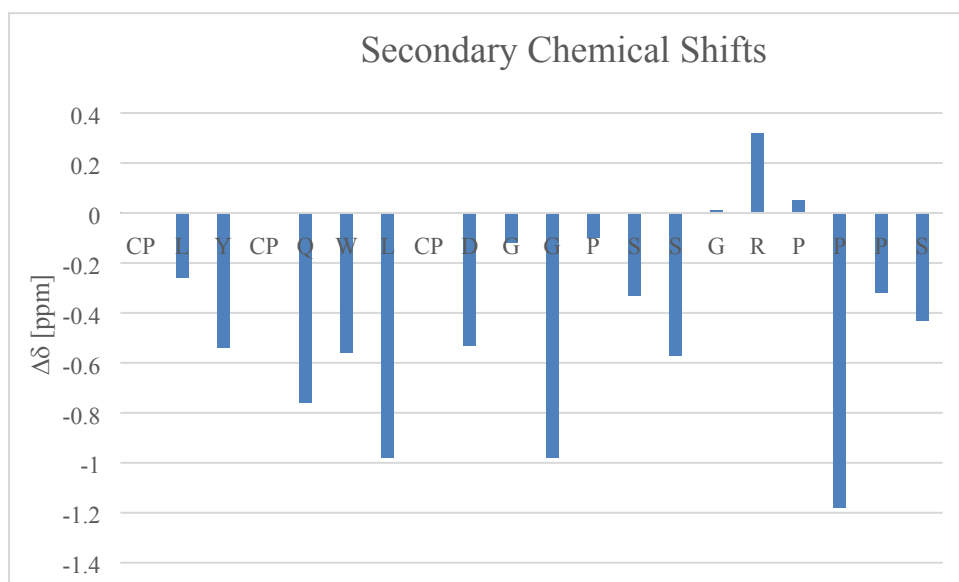


Figure S5. Differences between ¹H chemical shifts of HA protons of peptide **13** and random coil shifts for specific amino acid residues.

Table S4. Interproton contacts of peptide **13** in 50mM phosphate buffer, pH 7. Cp stands for *trans*-(1*S*,2*S*)-2-amino-1-cyclopentanecarboxylic acid. Residue no 1 is acetyl group at *N*-terminus.

Sequential (<i>i, i+1</i>)	Intensity
Ac1CH ₃ -Cp2HN	s
Cp2CH ₂ 1-L3HN	m
Cp2CH ₂ 3-L3HN	m
Cp2HA-L3HA	m
Cp2HA-L3HN	s
Cp2HB-L3HN	m
Cp2HN-L3HG	m
L3HA-Y4HN	s
L3HB-Y4HE	m
L3HD1-Y4HD	m
L3HD1-Y4HE	m
L3HD1-Y4HN	m
L3HD2-Y4HD	s
L3HD2-Y4HE	m
L3HD2-Y4HN	m
L3HN-Y4HA	w
L3HN-Y4HN	m
Y4HA-Cp5HN	s
Y4HB1-Cp5HN	s
Y4HB2-Cp5HN	s
Y4HD-Cp5HN	m
Y4HD-Cp5CH ₂ 1	m
Y4HD-Cp5CH ₂ 2	w
Y4HD-Cp5CH ₂ 3	s
Y4HE-Cp5CH ₂ 1	w
Y4HE-Cp5CH ₂ 2	s
Y4HN-Cp5CH ₂	s
Cp5CH ₂ 2-Q6HN	s
Cp5CH ₂ 3-Q6HN'1	m
Cp5HN-Q6HN	s
Cp5HA-Q6HN	s
Cp5HB-Q6HN	m
Q6HA-W7HN	s
Q6HB1-W7HN	s
Q6HB2-W7HN	s
Q6HG-W7HN	m
Q6HN-W7HN	m
W7HA-L8HN	m
W7HB1-L8HN	s
W7HB2-L8HN	m
W7HE3-L8HG	w
W7HE3-L8HA	m
W7HE3-L8HB1	m
W7HE3-L8HD1	m
W7HE3-L8HD2	m
W7HE3-L8HN	m
W7HH2-L8HD2	w
W7HH2-L8HD1	m
W7HN-L8HB2	m
W7HN-L8HN	m
W7HZ3-L8HD1	s
W7HZ3-L8HN	m
W7HZ3-L8HD2	m
W7HZ2-L8HD1	w
W7HZ2-L8HD2	w
L8HA-Cp9HN	s
L8HB1-Cp9HB	s
Cp9HN-D10HN	m
Cp9CH ₂ 2-D10HN	s
Cp9CH ₂ 3-D10HN	m
Cp9HA-D10HN	m
Cp9HA-D10HB2	w
Cp9HB-D10HN	s
D10HA-G11HN	m
D10HN-G11HN	m
G11HA1-G12HN	m
G11HA2-G12HN	m
G11HN-G12HN	s
G12HA1-P13HD2	s
G12HA2-P13HD2	m
G12HN-P13HD1	w
P13HA-S14HN	w
P13HD1-S14HN	w
S14HA-S15HN	m
S14HN-S15HN	m
S15HA-G16HN	s
G16HA1-R17HN	s
G16HA2-R17HN	s
G16HN-R17HN	s
G16HN-R17HA	m
R17HB1-P18HD1	s
R17HB1-P18HD2	s
P18HB1-P19HA	s
P18HG-P19HA	m
P18HB2-P19HA	s
P18HA-P19HA	w
P19HB2-P20HD2	w
P20HA-S21HN	s
P20HG-S21HN	s
S21HA-NH2HN2	m
Medium range (<i>i, i+2</i>)	Intensity
Cp2CH ₂ 3-Y4HE	s
L3HN-Cp5CH ₂ 1	w
Y4HD-Q6HB1	w
Cp5HB-W7HZ1	m
Cp5HB-W7HZ3	m
W7HA-Cp9HA	s
P13HA-S15HN	m
S14HA-G16HN	m
S15HN-R17HA	s
Medium range (<i>i, i+3</i>)	Intensity
Ac1HA-Y4HB1	m
Ac1HA-Y4HD	m
Ac1HA-Y4HE	w
Cp2HB-Cp5CH ₂ 2	m
Cp2HB-Cp5HA	s
L3HA-Q6HB1	s
L3HA-Q6HN	s
L3HB-Q6HA	m
L3HA-Q6HB2	s
L3HA-Q6HG	m
Y4HA-W7HE3	m
Y4HA-W7HN	m
Y4HA-W7HB2	s
Cp5HB-L8HN	s
Cp5HB-L8HD1	s
Cp5HB-L8HD2	m
Cp5HB-L8HG	m
Q6HA-Cp9HN	s
Q6HN'2-Cp9HA	m
Q6HA-Cp9HA	m
W7HA-D10HN	m
W7HA-D10HB1	w
L8HA-G11HN	s
G12HN-S15HB1	m
G12HN-S15HB2	m
S14HN-R17HA	m
R17HA-P20HD2	s
P19HA-NH ₂ HN1	s

Medium range ($i, i+4$)	Intensity
Cp2CH ₂ 3-Q6HN	m
Cp2HB-Q6HN	m
L3HA-W7HN	m
L8HA-G12HN	s
L8HD1-G12HN	w
L8HD2-G12HN	m
G11HA-S15HN	w

Long range ($i, i>4$)	Intensity
Y4HD-P19HB1	m
Y4HE-P19HB1	s
Y4HE-P19HB2	w
W7HA-G12HN	w
W7HH2-P13HA	m
W7HZ2-P13HA	s
W7HH2-P13HD1	w
W7HH2-P13HD2	m

W7HZ3-P13HD2	m
W7HZ2-P13HD2	s
W7HH2-P13HG	s
W7HZ2-P13HG	m
W7HH2-P13HB2	w
W7HZ2-P13HB2	w
W7HZ2-P19HA	m
W7HZ2-P19HG1	w
W7HN-P20HD1	w

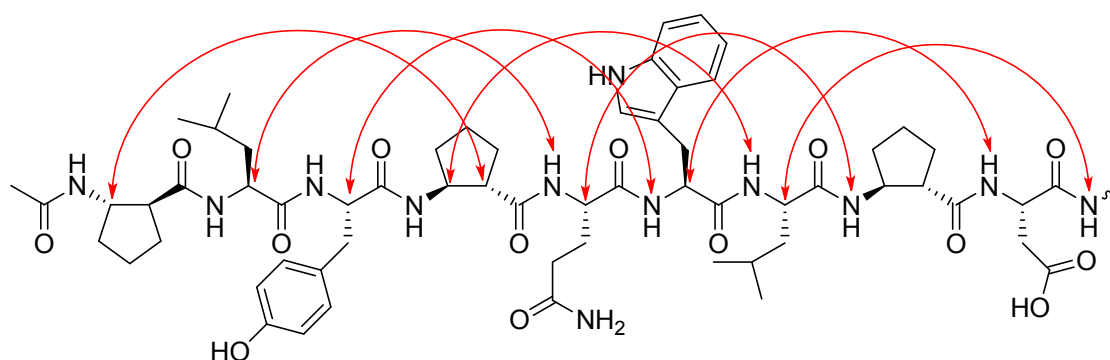


Figure S6. NOE $i-(i+3)$ contacts of N-terminal helix of peptide **13**.

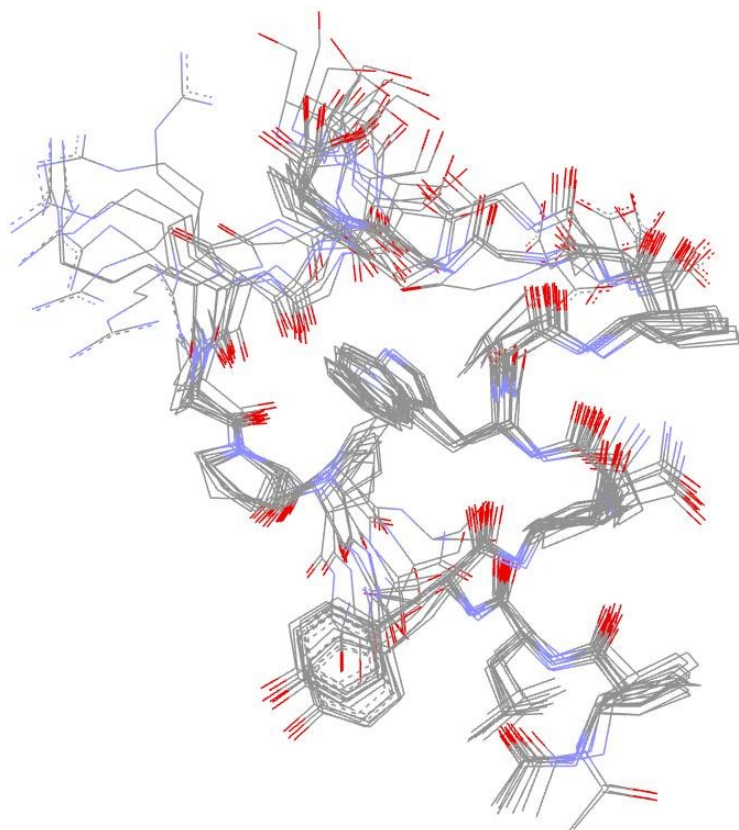


Figure S7. Superimposition of 10 lowest energy structures of peptide **13** obtained during molecular dynamics with NMR constraints.

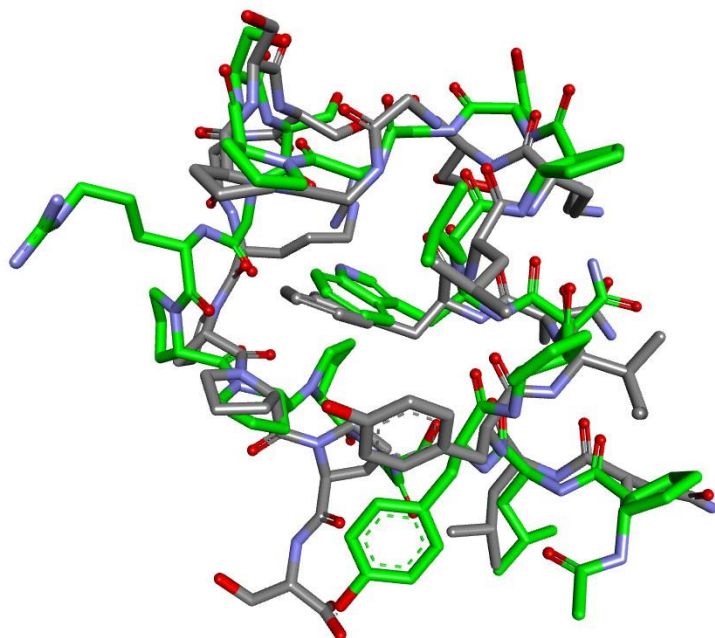


Figure S8. superimposition of average NMR (green carbon atoms) structure of peptide **13** and native Trp-cage (PDB id 1L2Y, grey carbon atoms).

Table S5. Statistics of NMR structure calculation

NOE restraints	
Total	135
Sequential ($ i-j = 1$)	77
Medium ($1 < i-j \leq 4$)	41
long range ($ i-j > 4$)	17
Average number of NOE violations	1.2
Average sum of upper limit violations	0.68 Å
Average sum of VdW violations	0.68 Å
Average RMDS (backbone)	0.75 Å

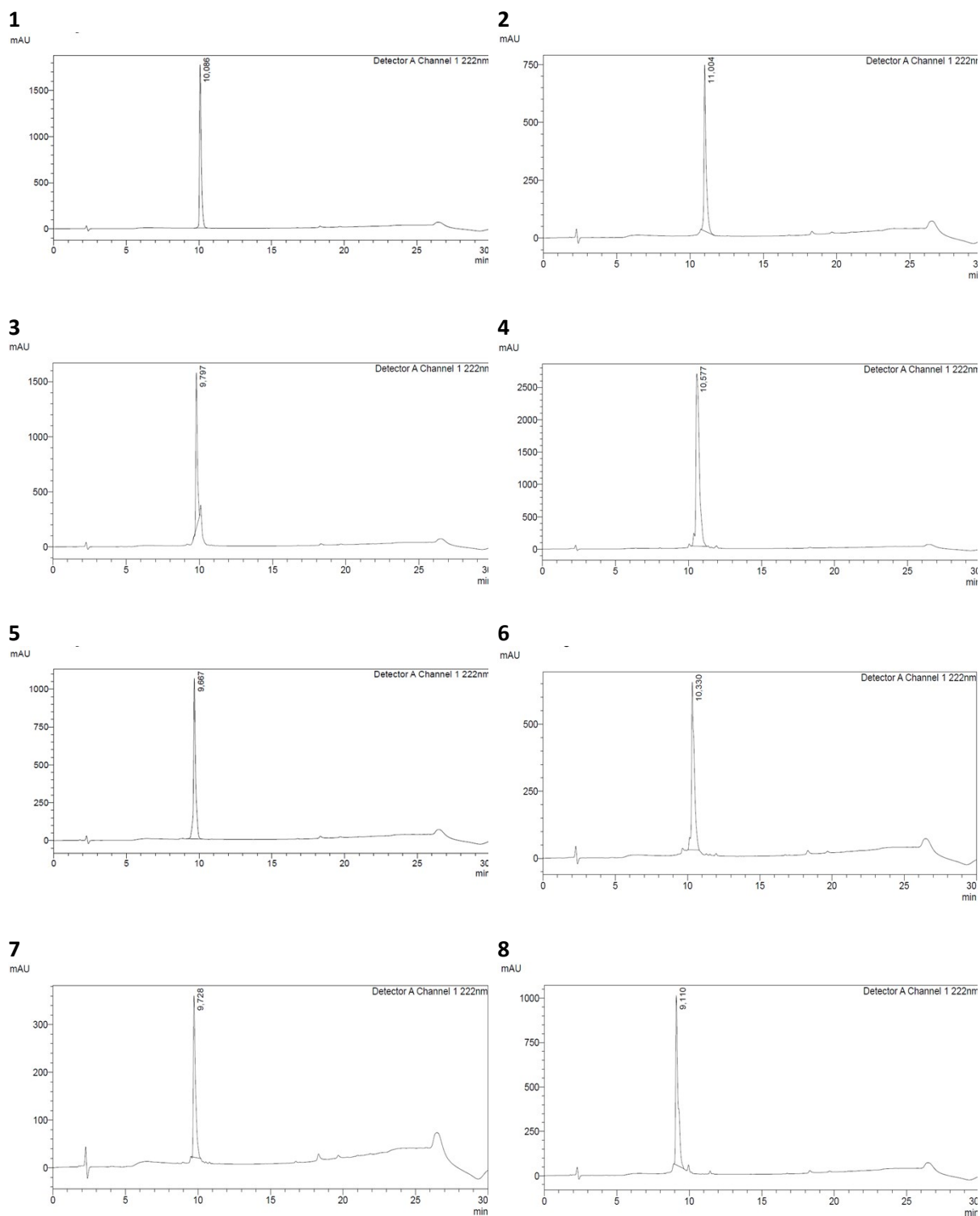


Figure S9. Analytical chromatograms for peptides 1-29.

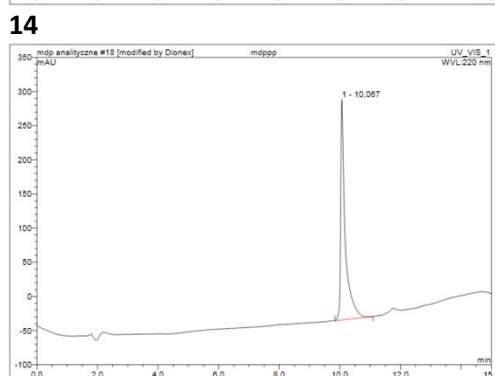
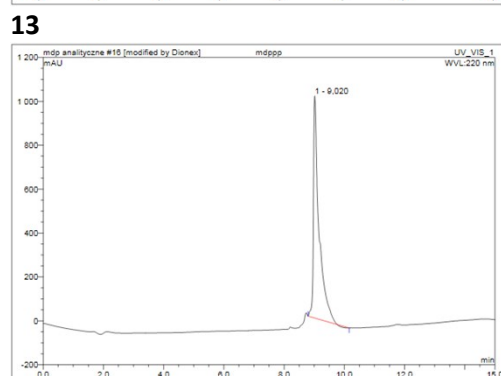
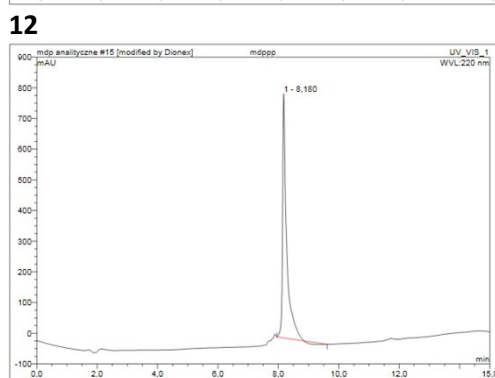
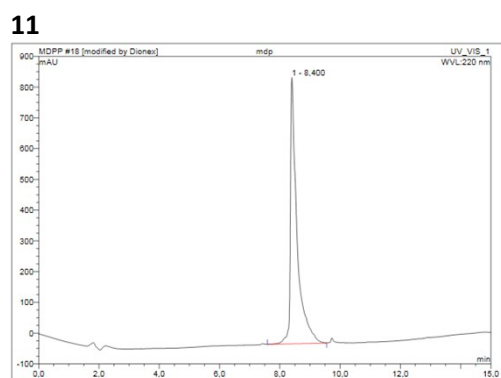
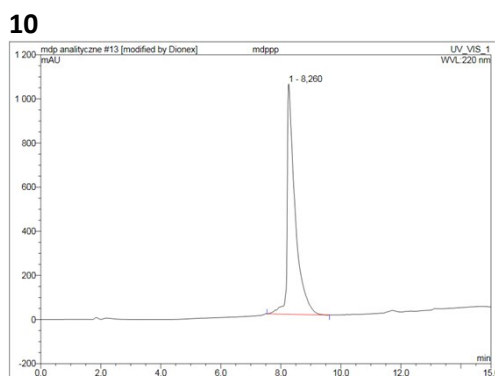
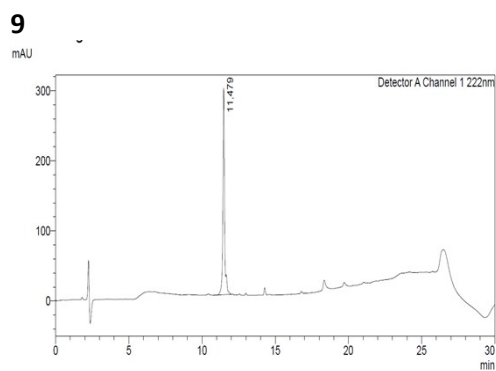
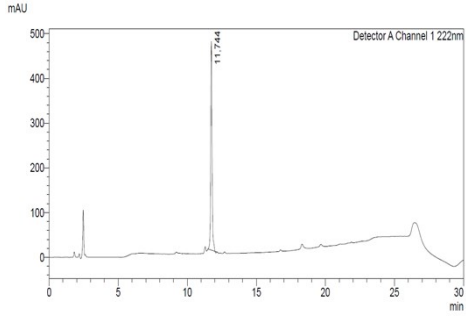
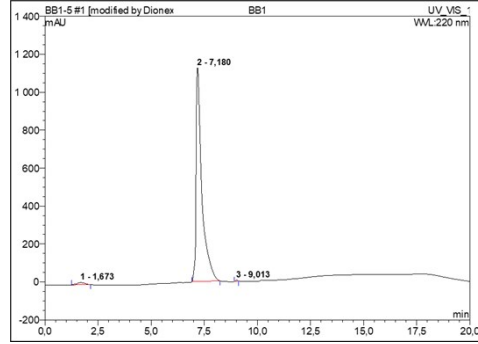


Figure S9 (continued). Analytical chromatograms for peptides 1-29.

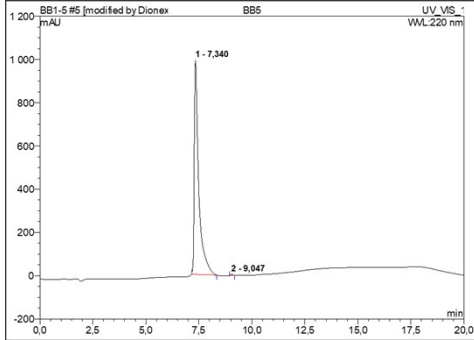
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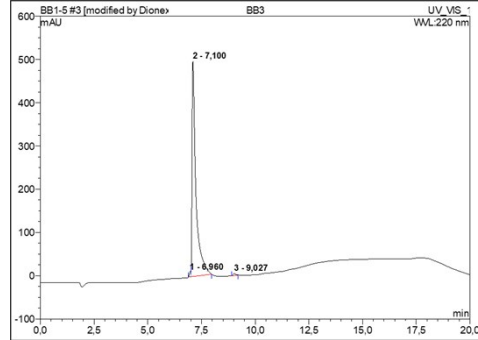
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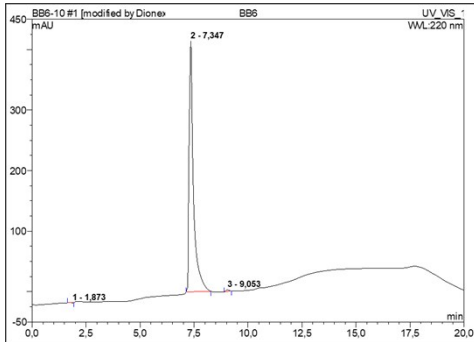
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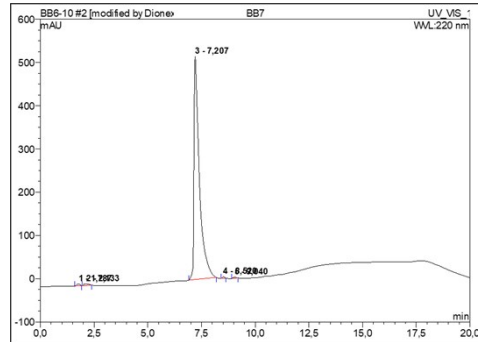
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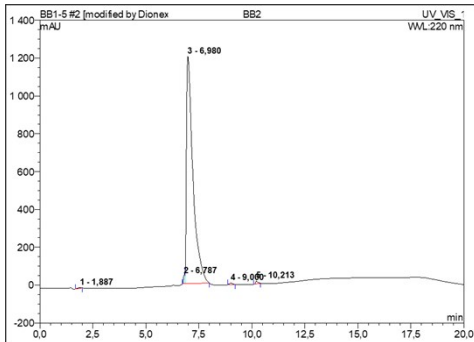
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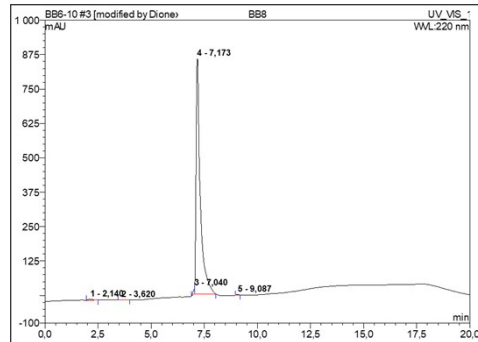
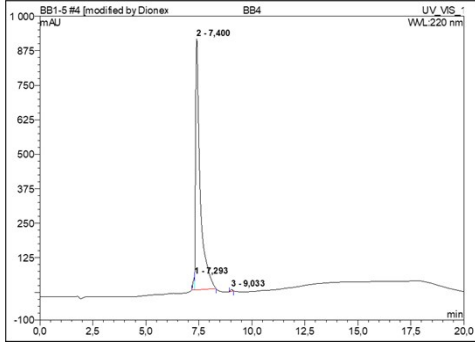
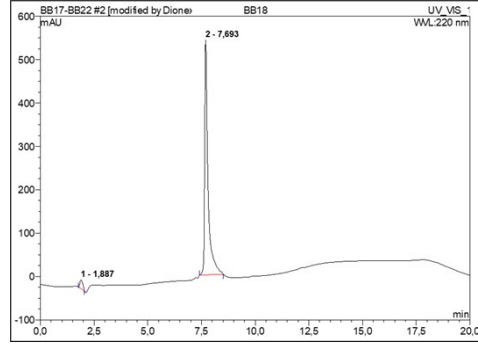


Figure S9 (continued). Analytical chromatograms for peptides 1-29.

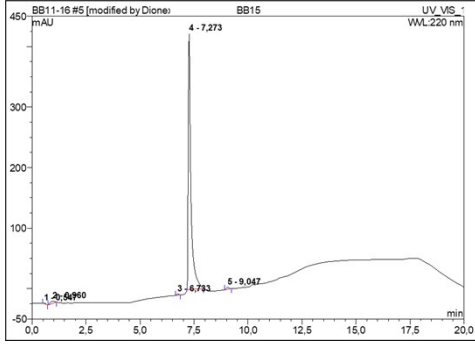
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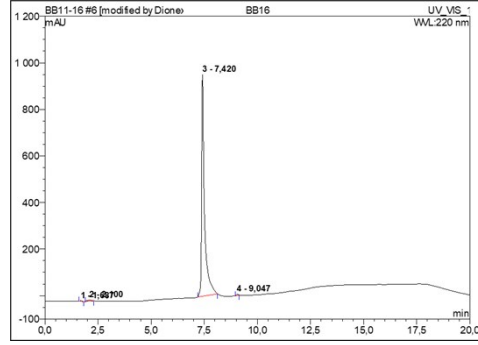
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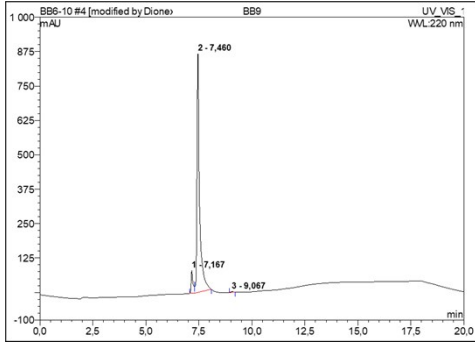
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27



29

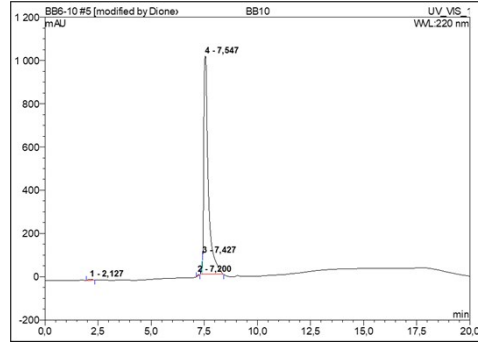


Figure S9 (continued). Analytical chromatograms for peptides 1-29.

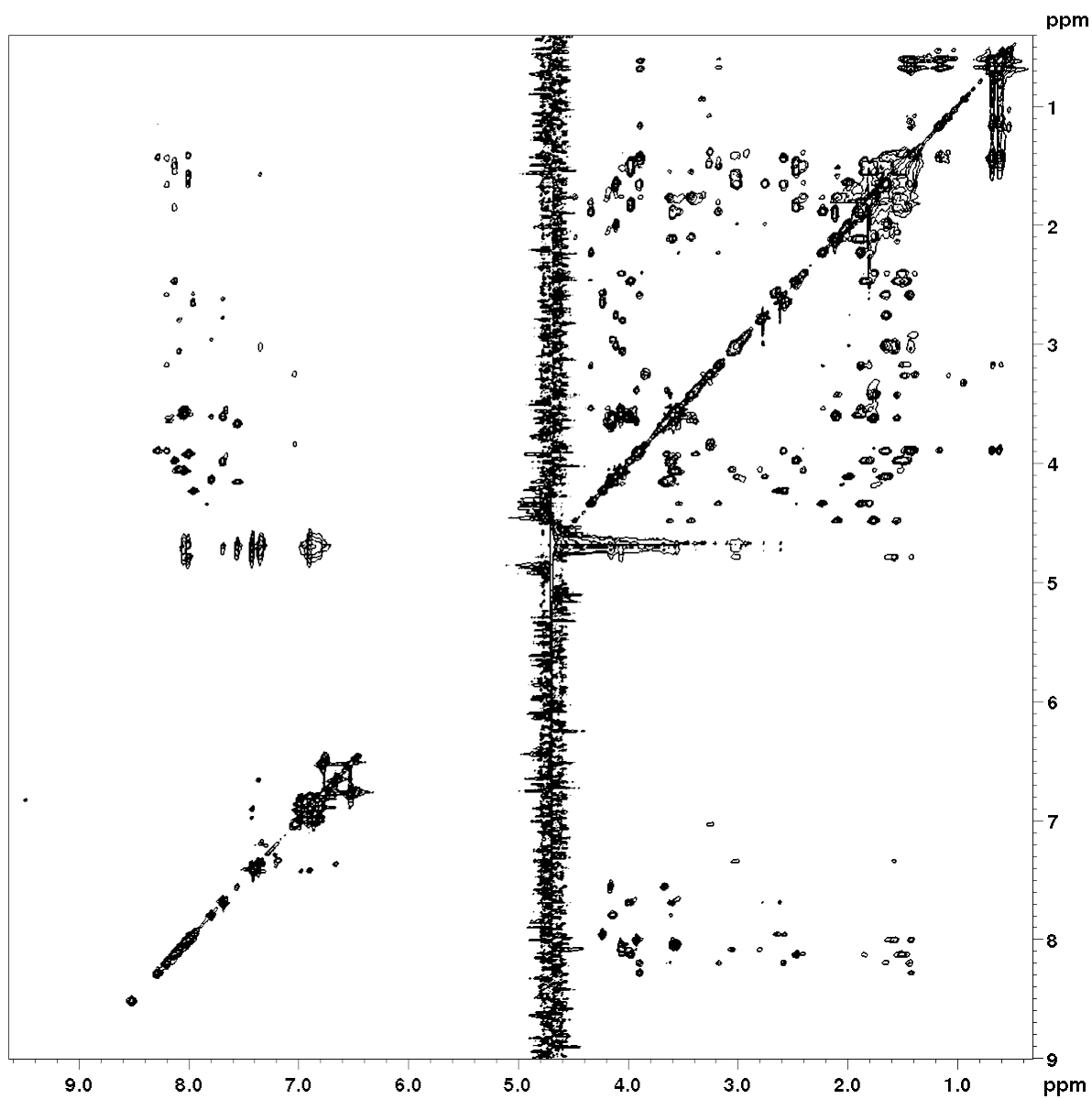


Figure S10. 2D TOCSY spectrum of peptide **13** dissolved in phosphate buffer 7.0 (10% D₂O) measured at 285K.

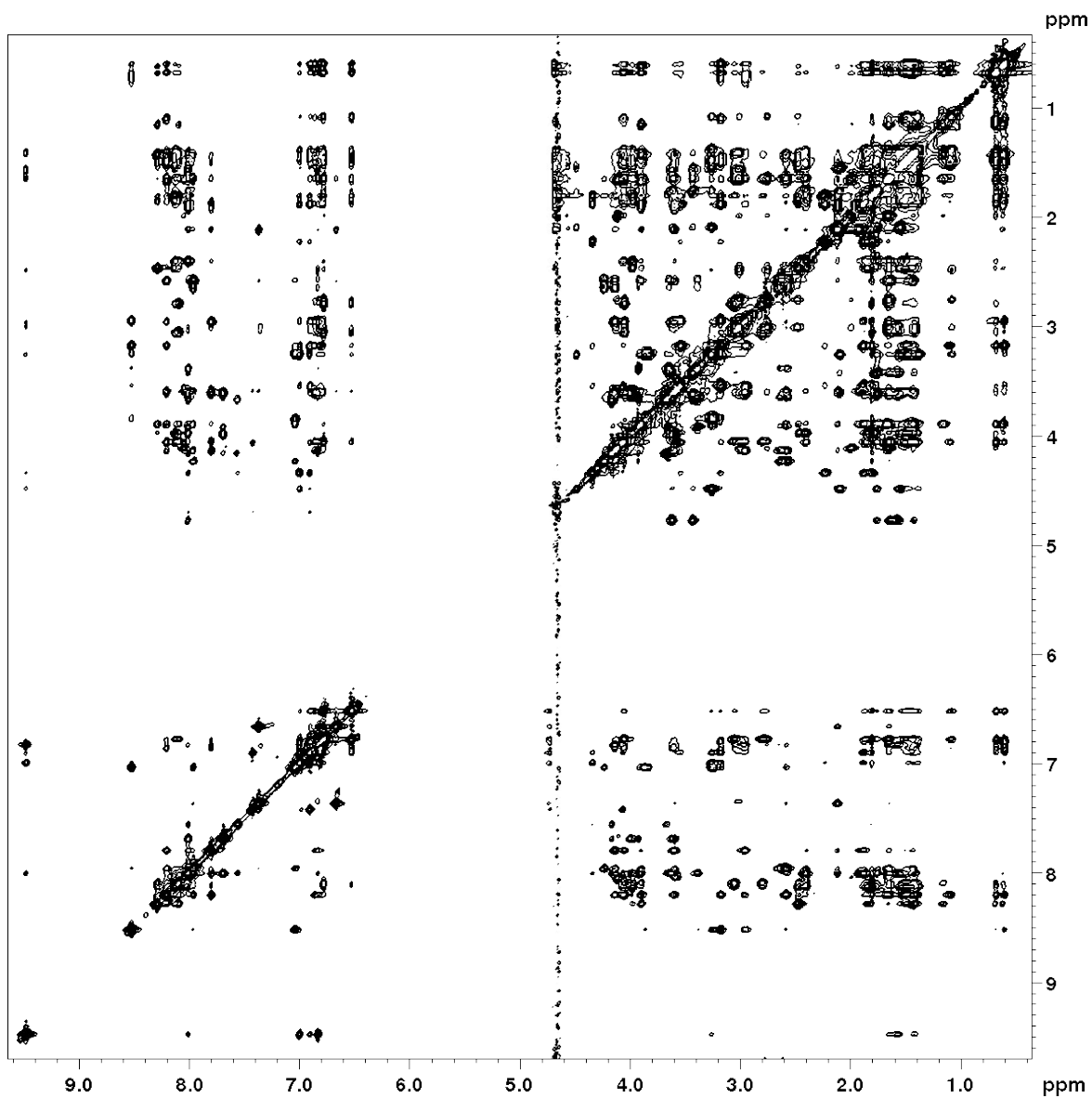


Figure S11. 2D NOESY spectrum of peptide **13** dissolved in phosphate buffer 7.0 (10% D₂O) measured at 285K.

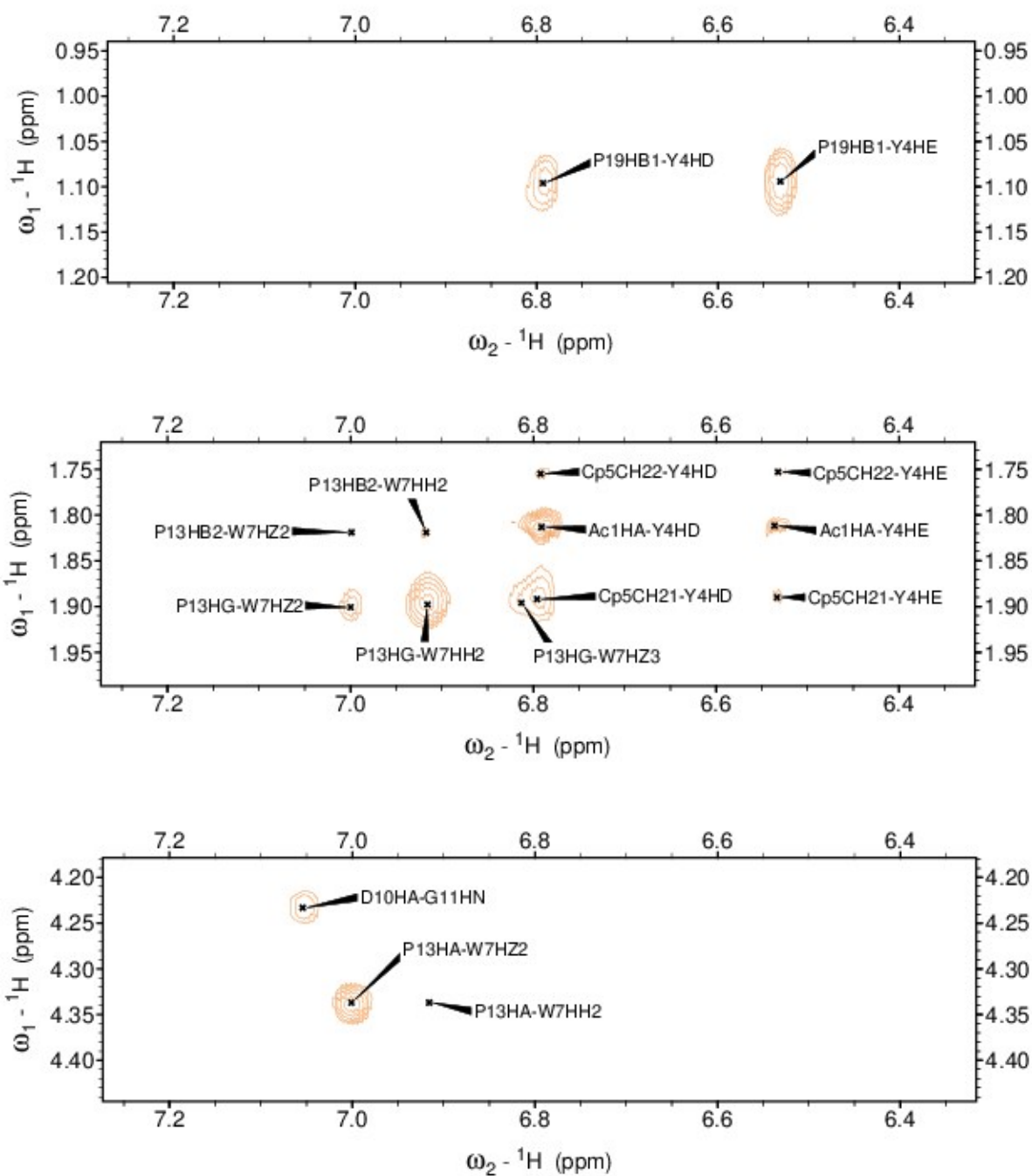


Figure S12. Annotated fragments of NOESY spectrum (phosphate buffer pH 7, 10% D₂O, 285K) of peptide **13** including long range inter-proton contacts.

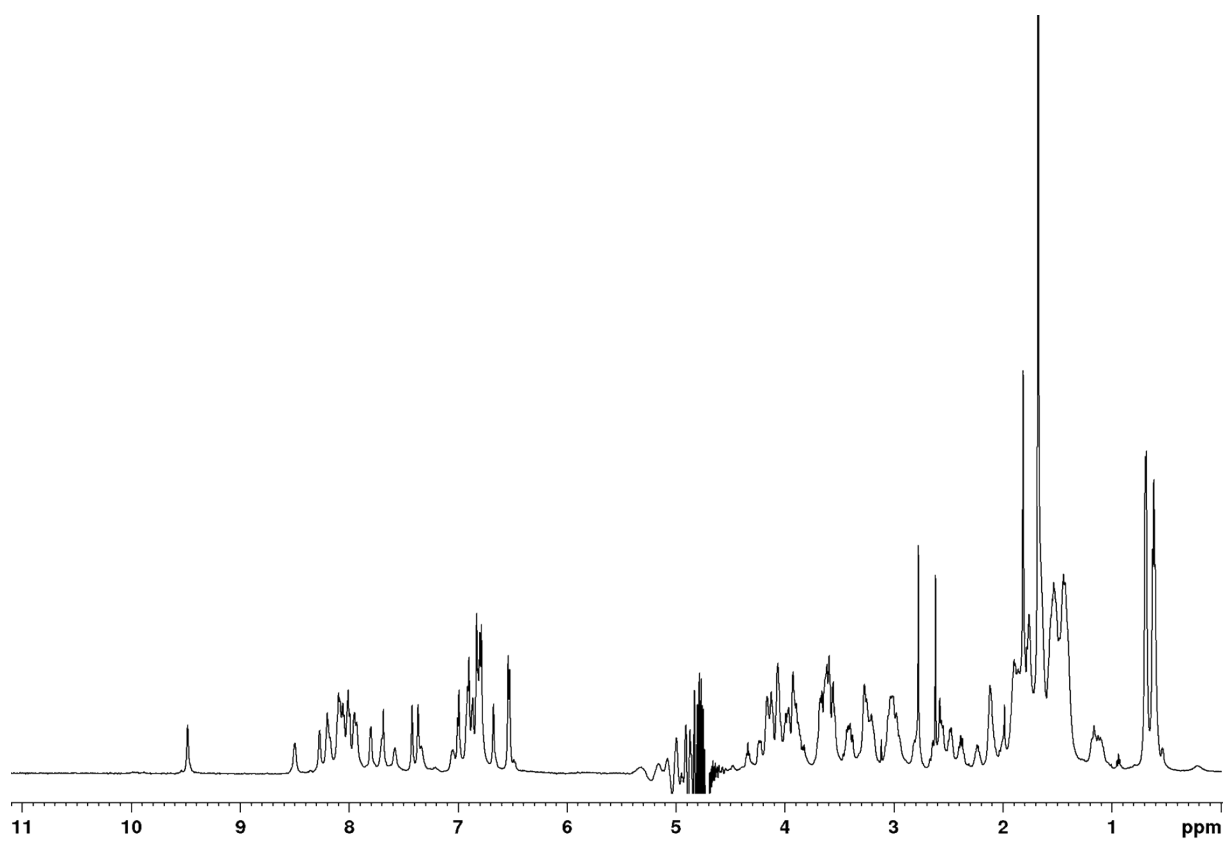
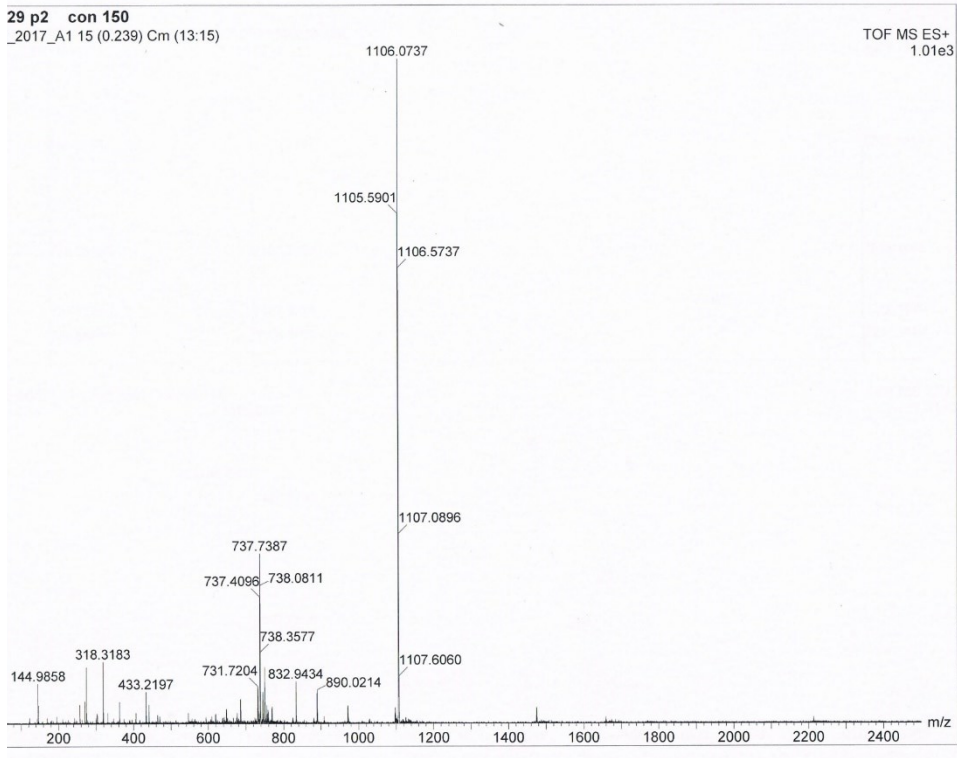


Figure S13. ^1H NMR spectrum of peptide **13** dissolved in phosphate buffer 7.0 (10% D_2O) measured at 285K.

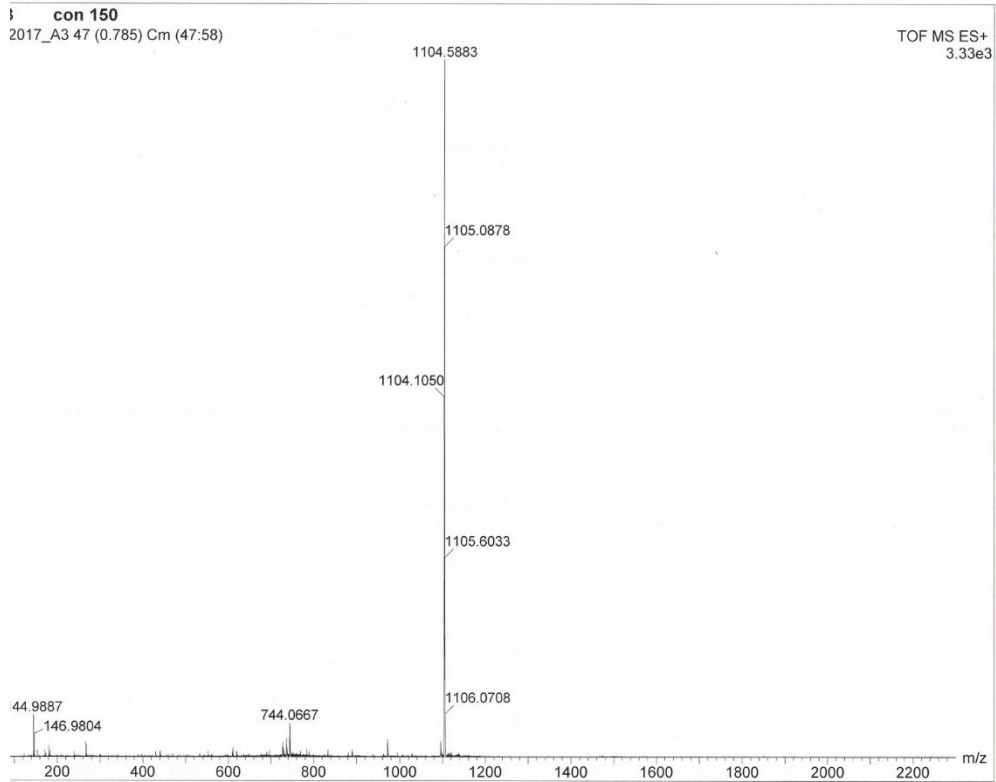
References

- S1. D. F. Kreidler, D. E. Mortenson, K. T. Forest, and S. H. Gellman, *J. Am. Chem. Soc.* **2016**, *138*, 6498-6505.
- S2. Z. E. Reinert Z. E., and W. S. Horne, *Chem. Sci.* **2014**, *5*, 3325-3330.

1



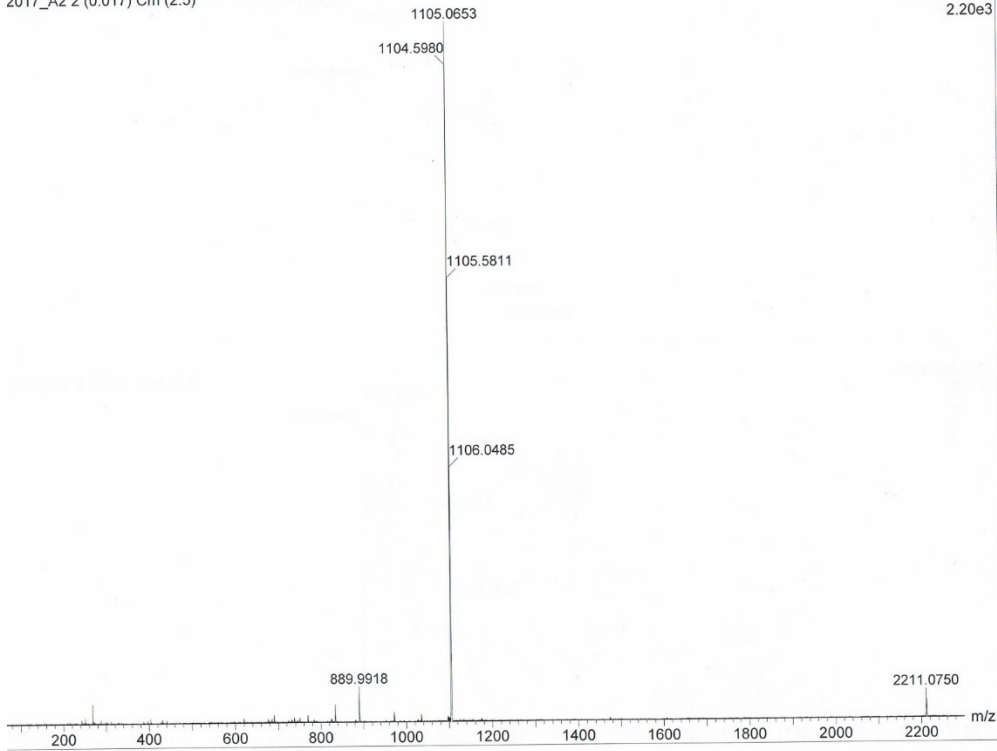
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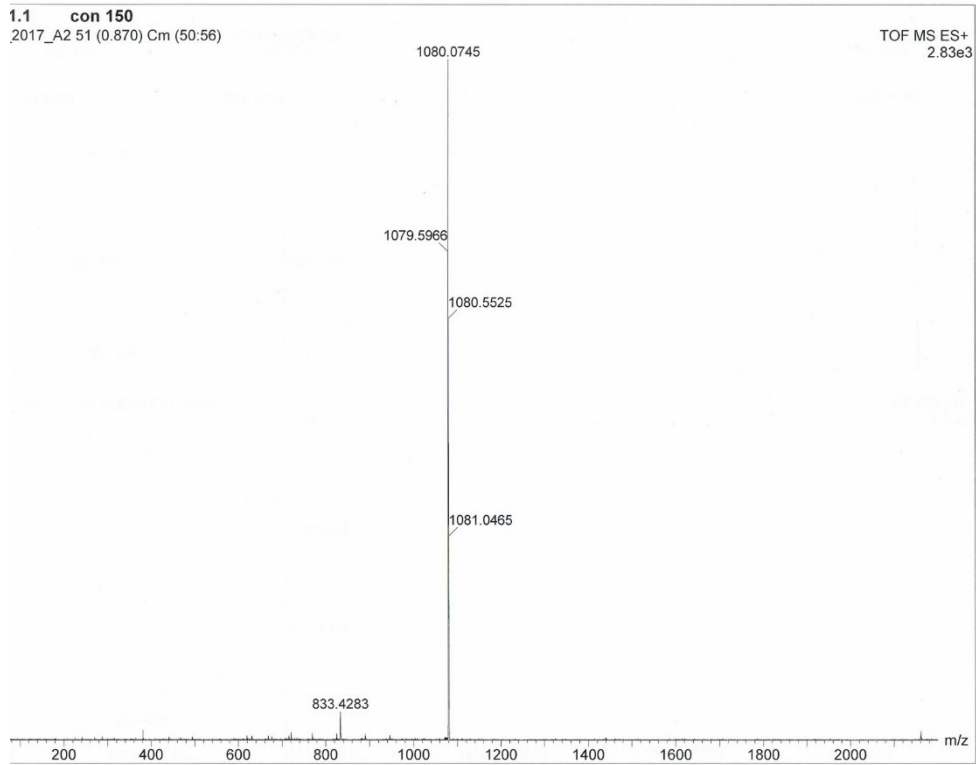
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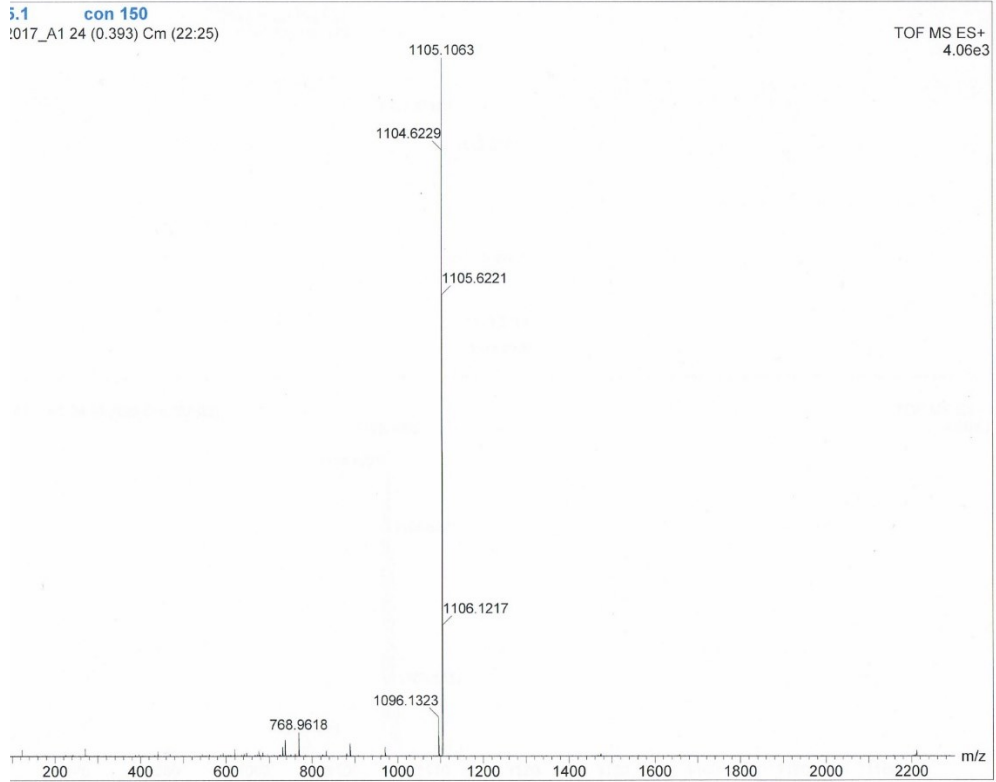
TOF MS ES+
2.20e3



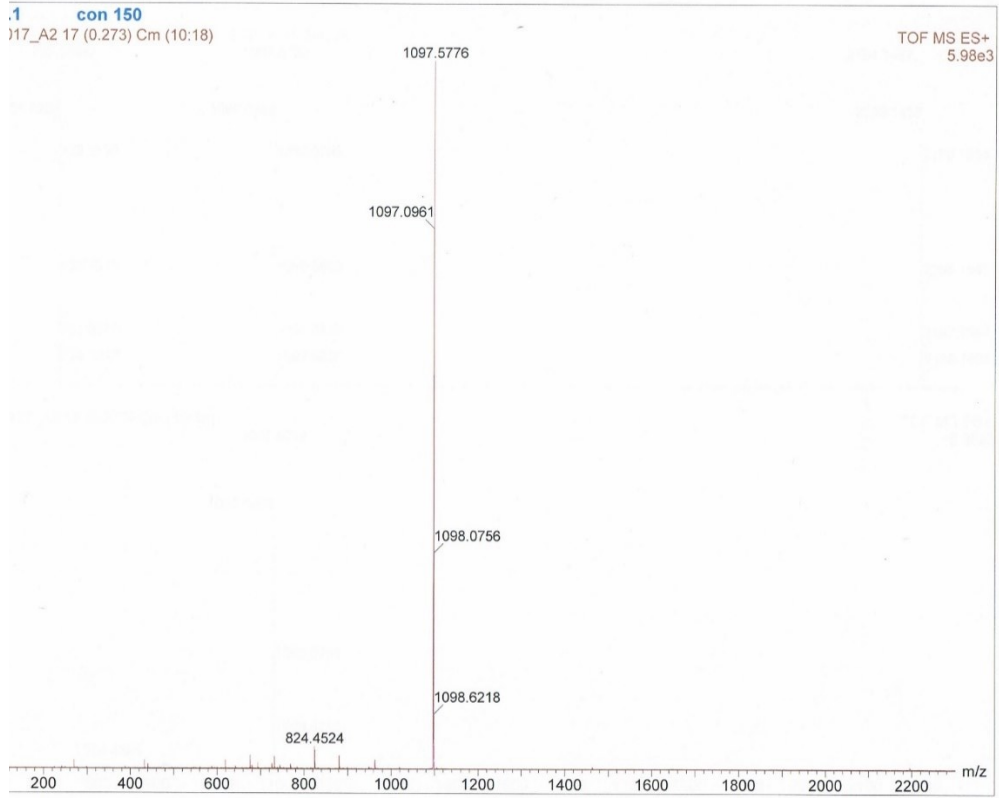
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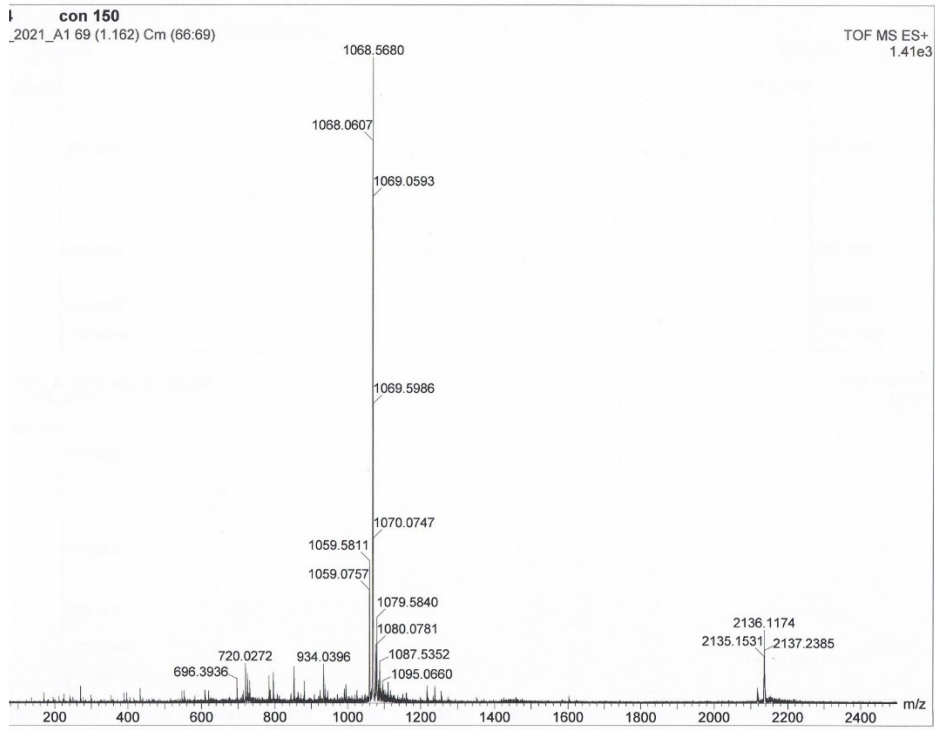
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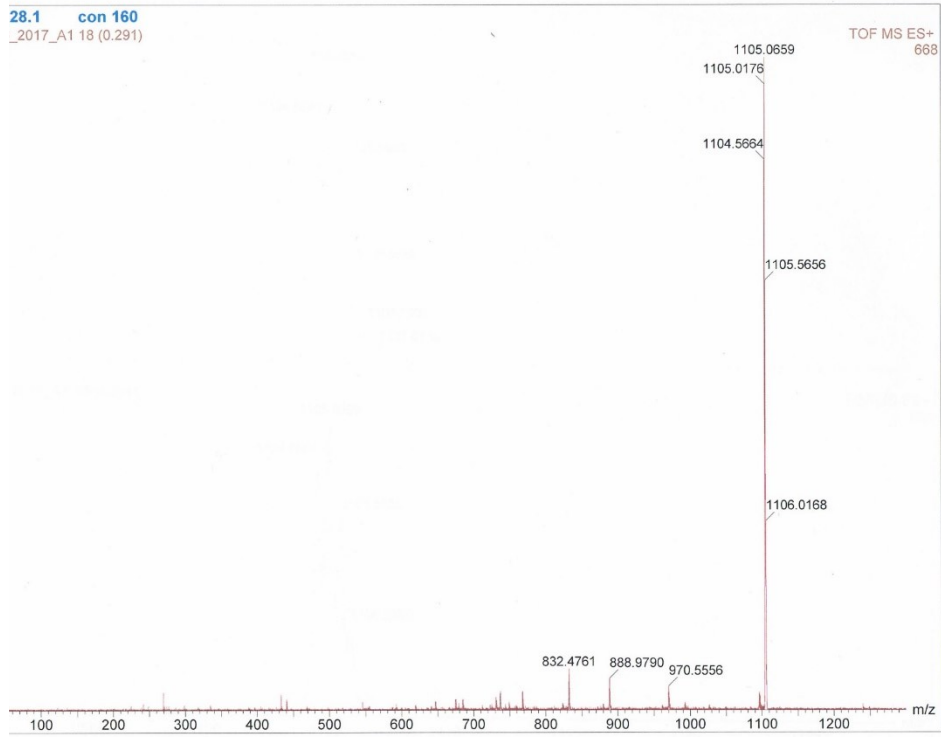
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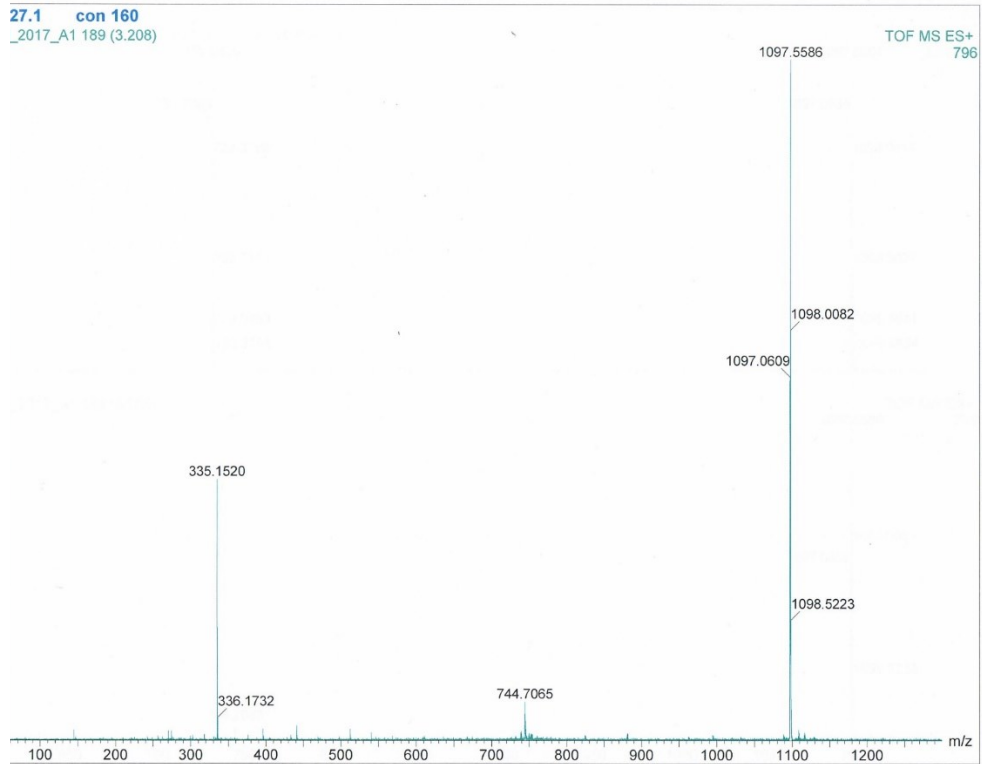
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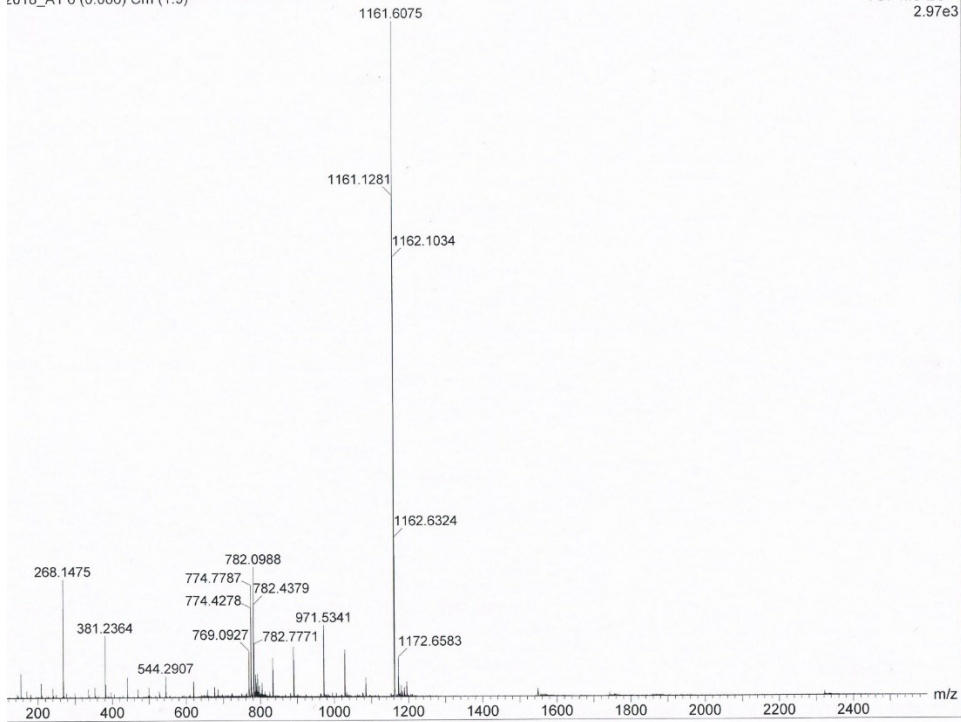
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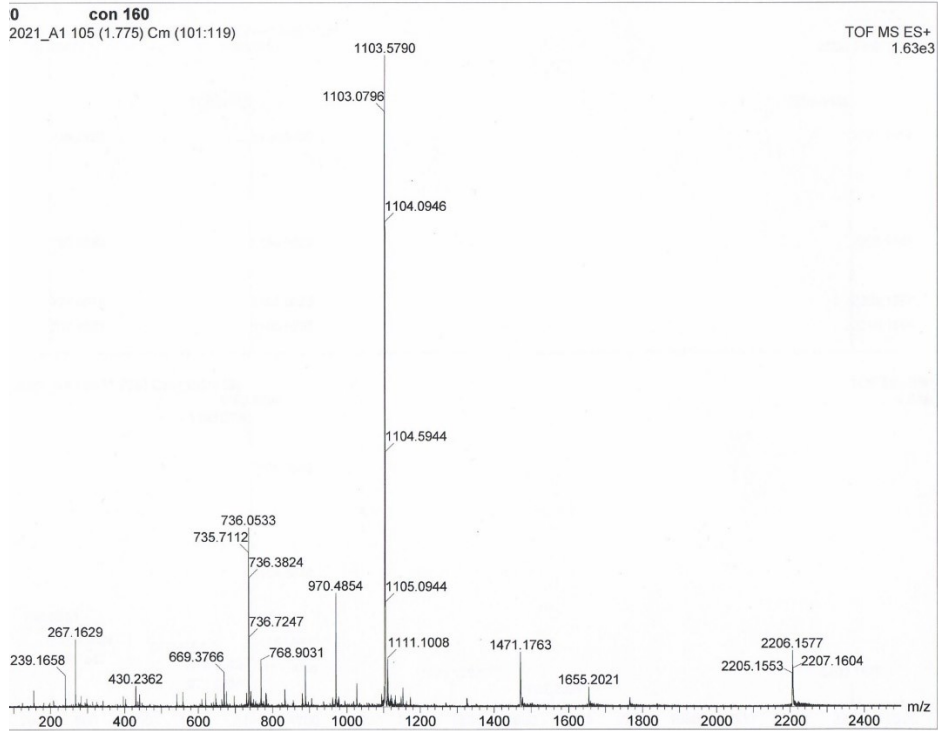
10

9 CpT.1 con 160
2018_A1 6 (0.086) Cm (1:9)

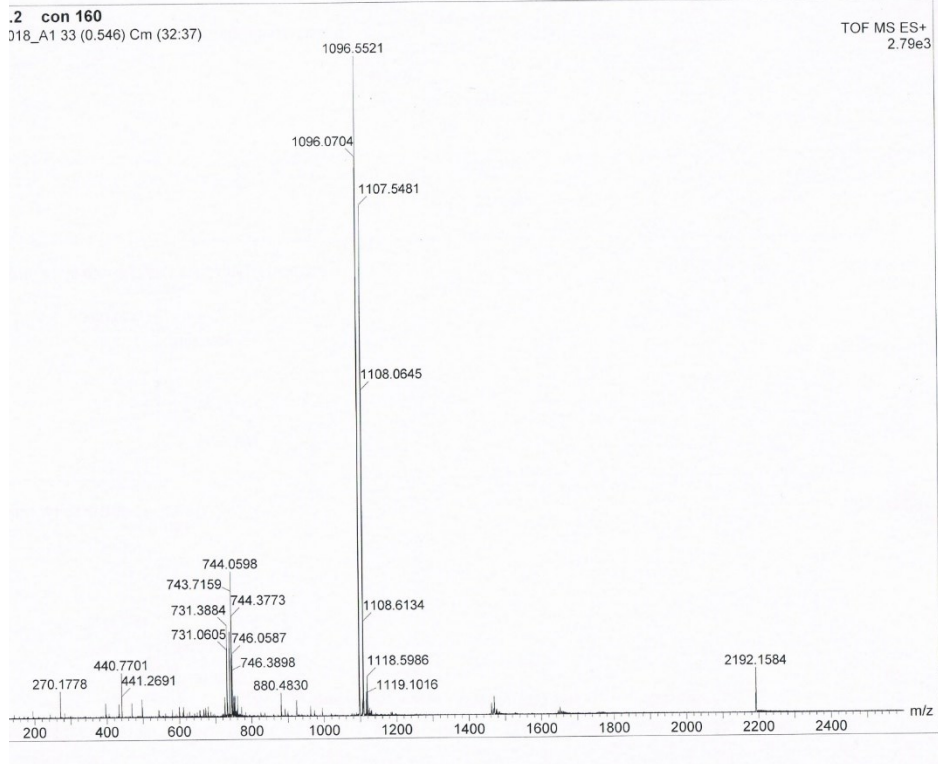
TOF MS ES+
2.97e3



11



12

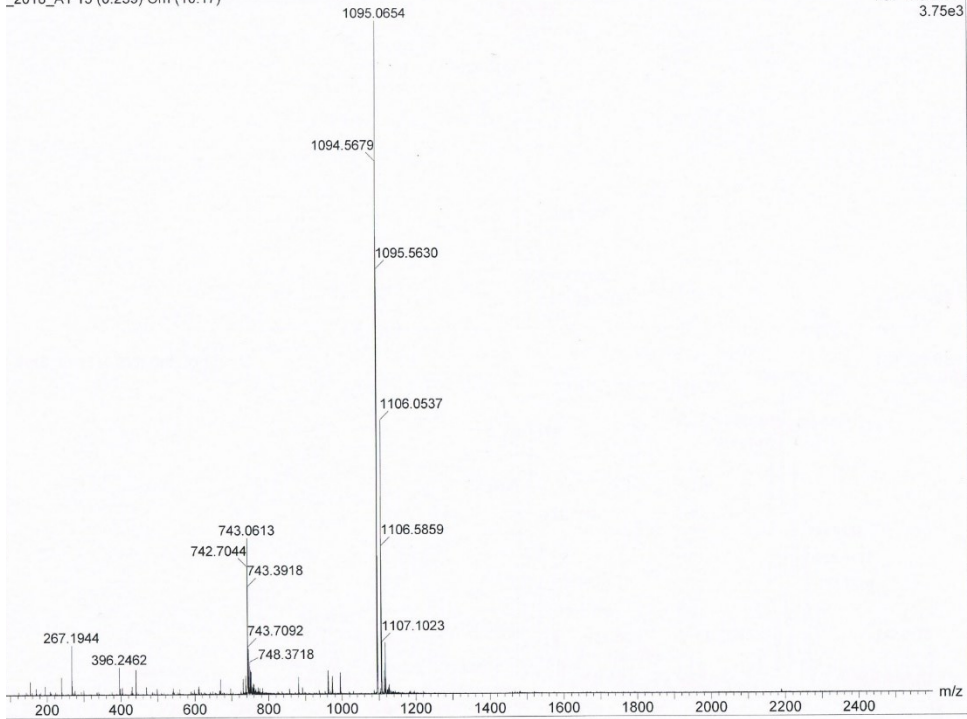


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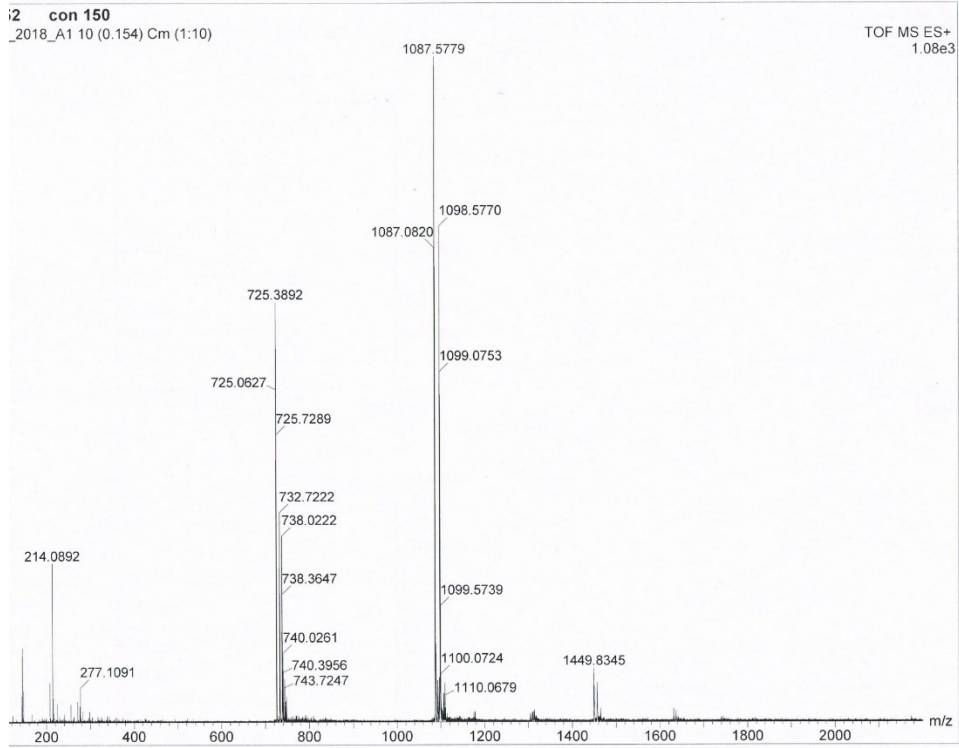
51 con 160

_2018_A1 15 (0.239) Cm (10:17)

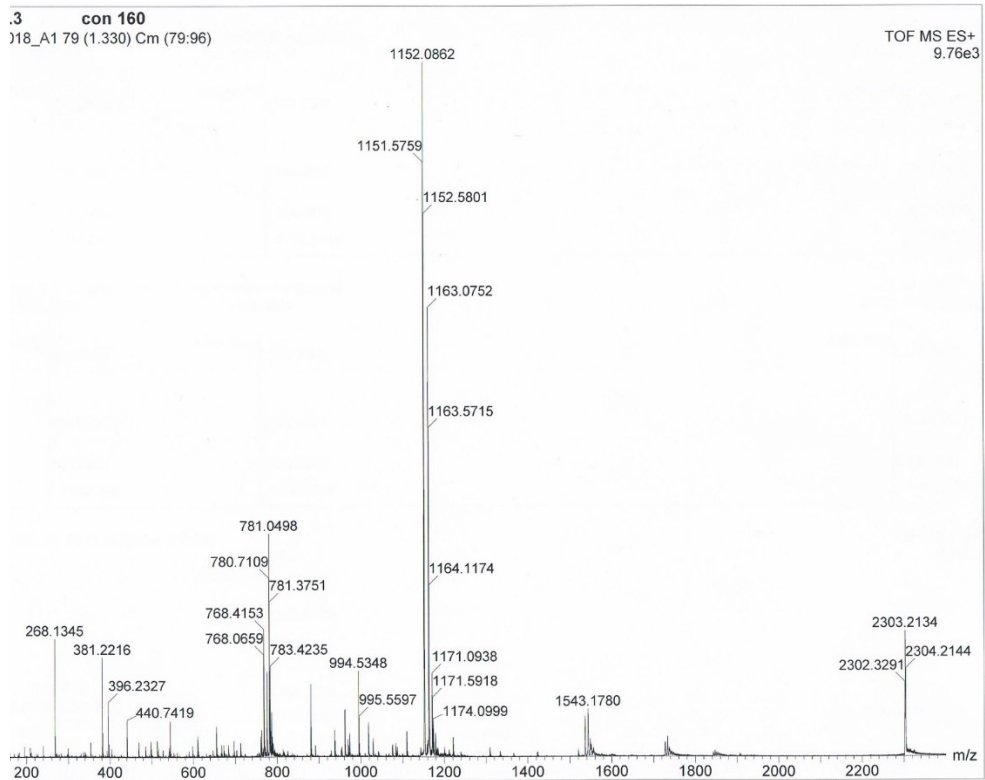
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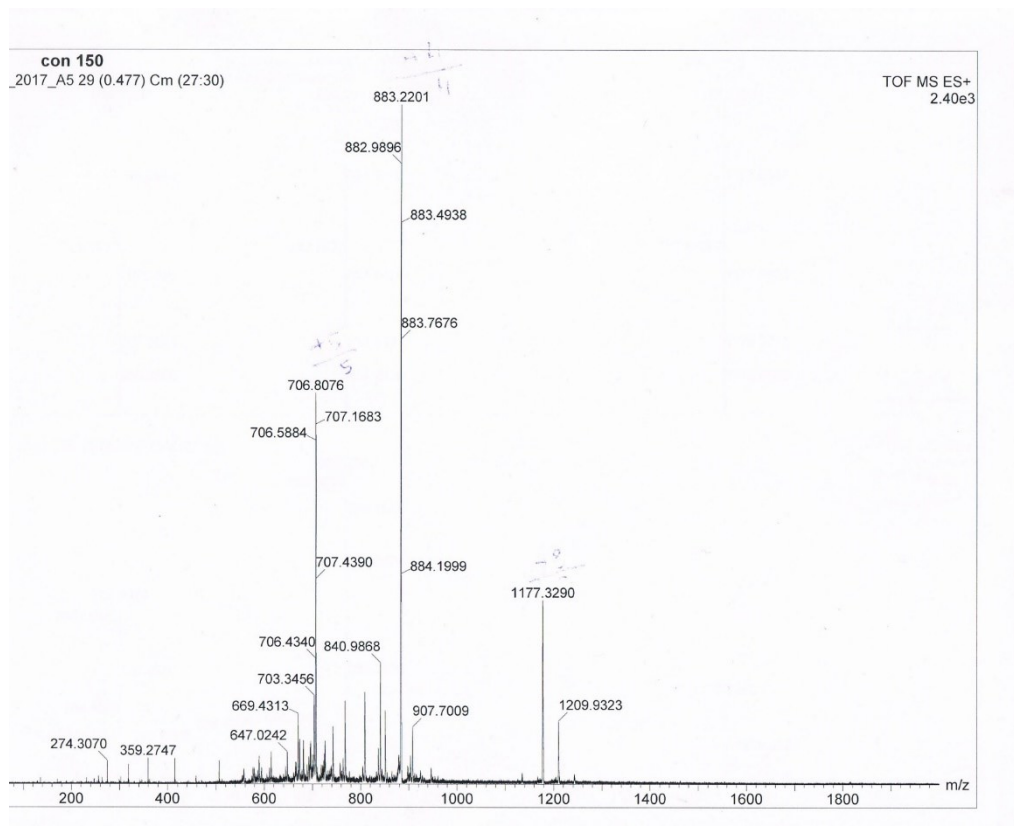
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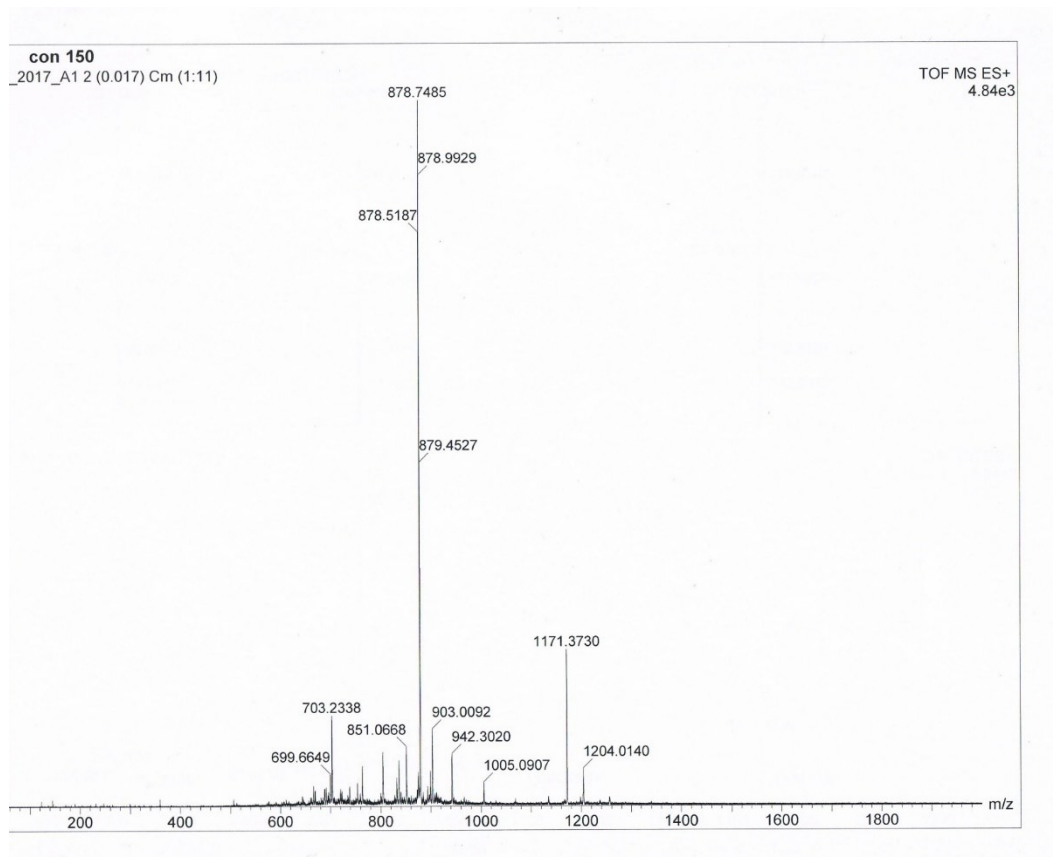
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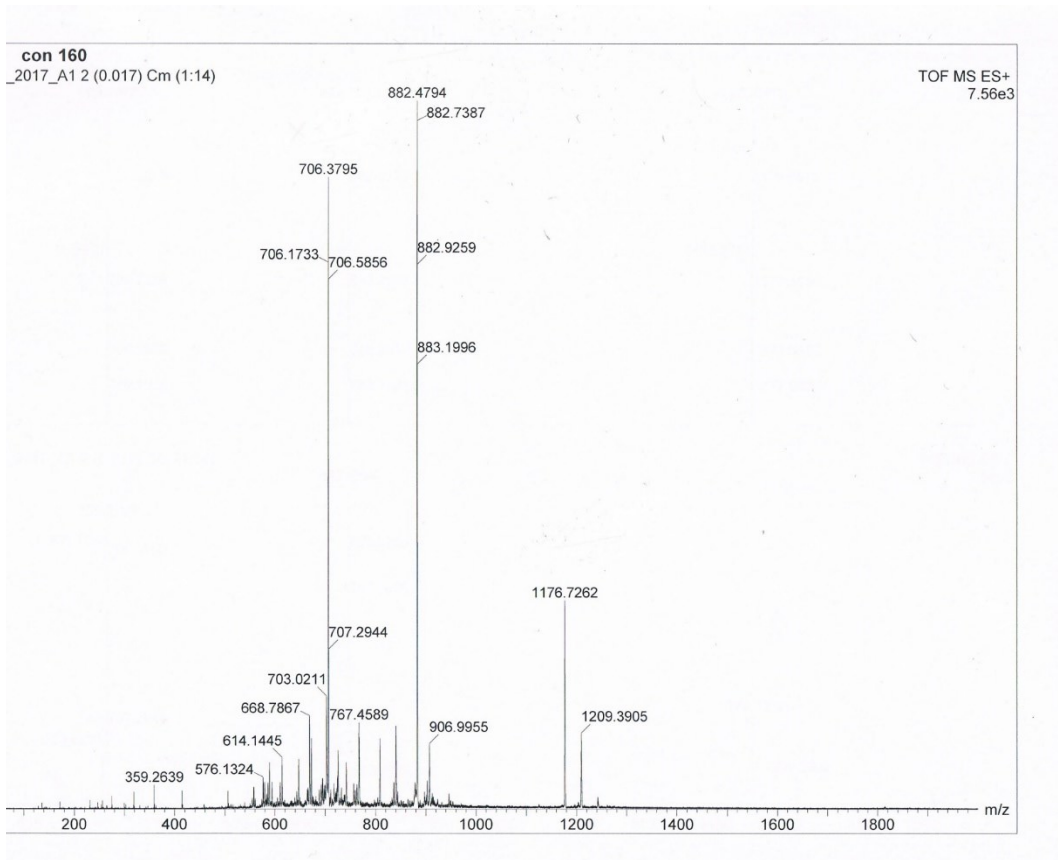
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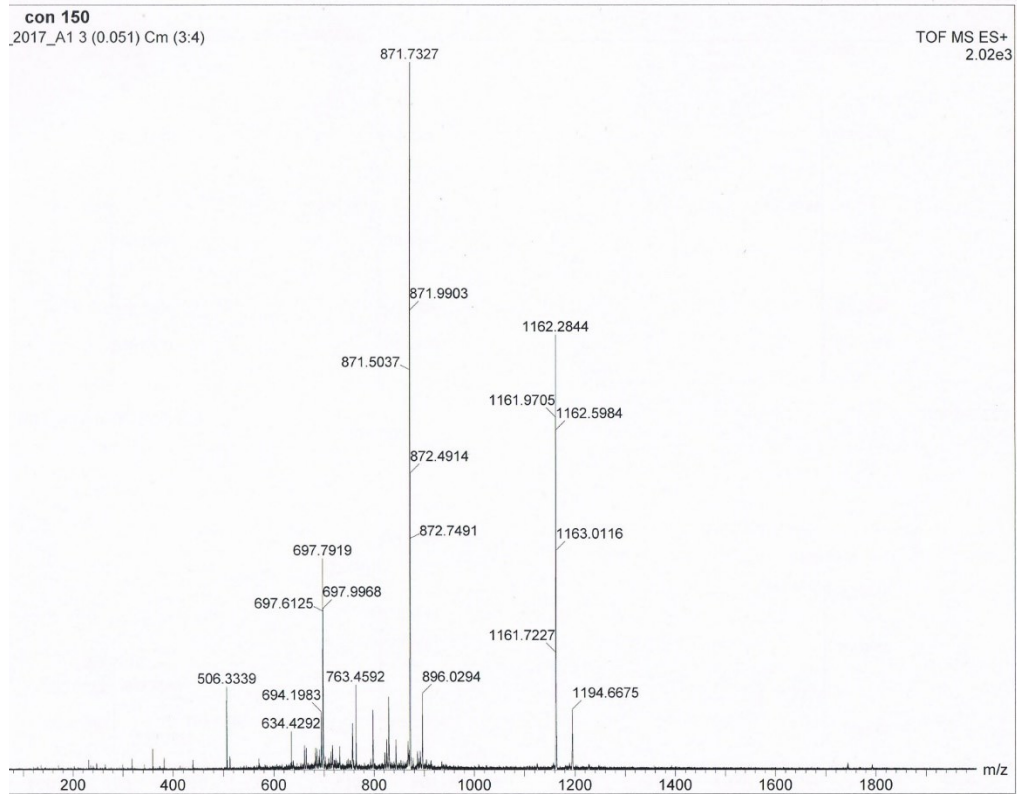
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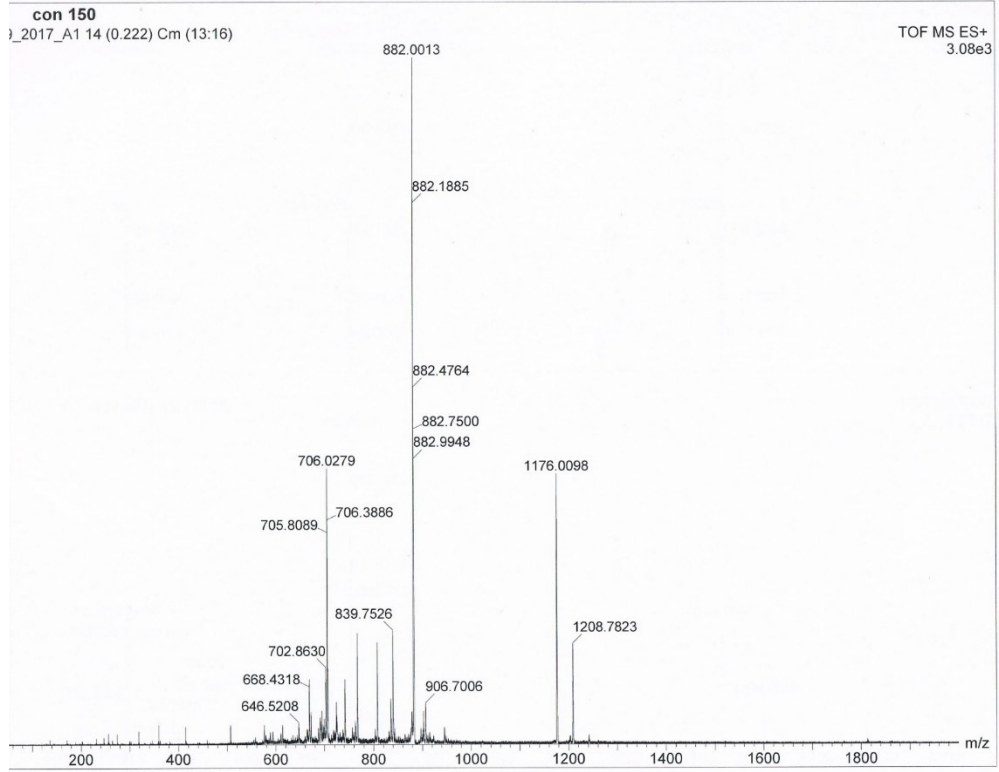
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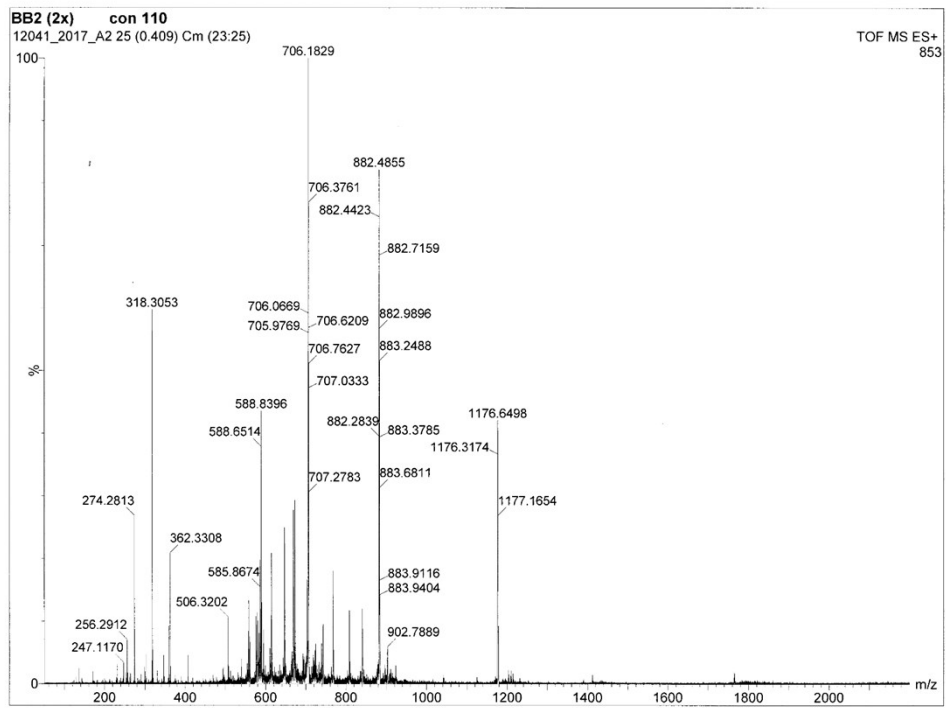
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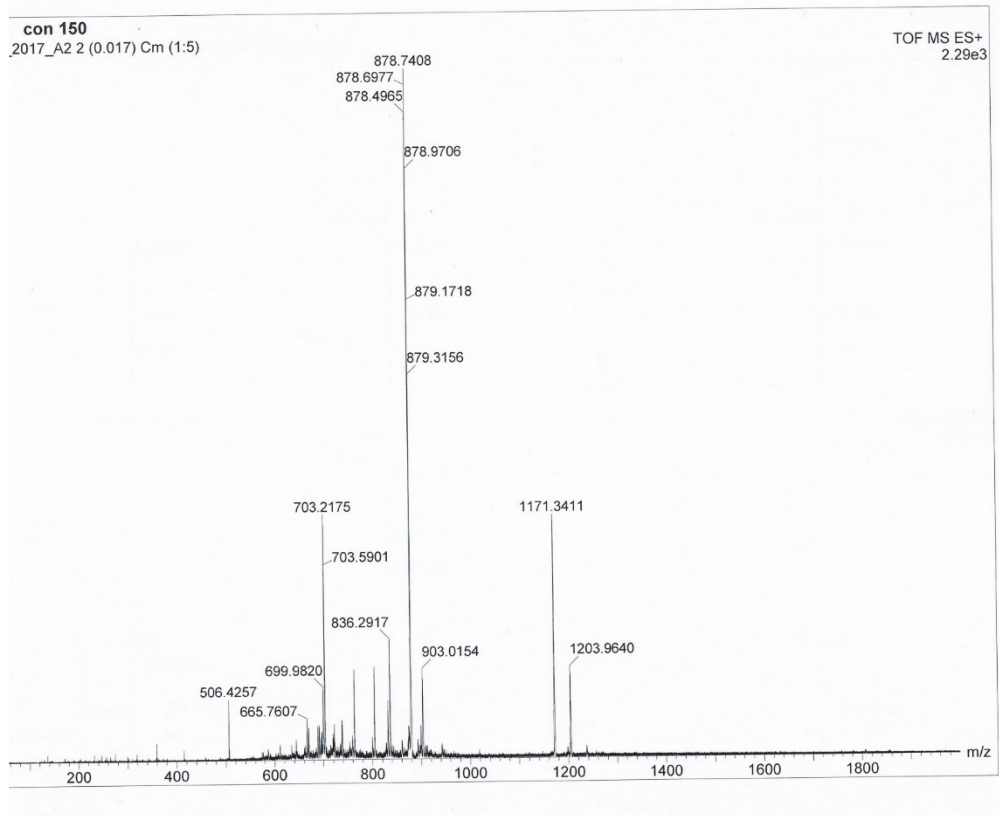
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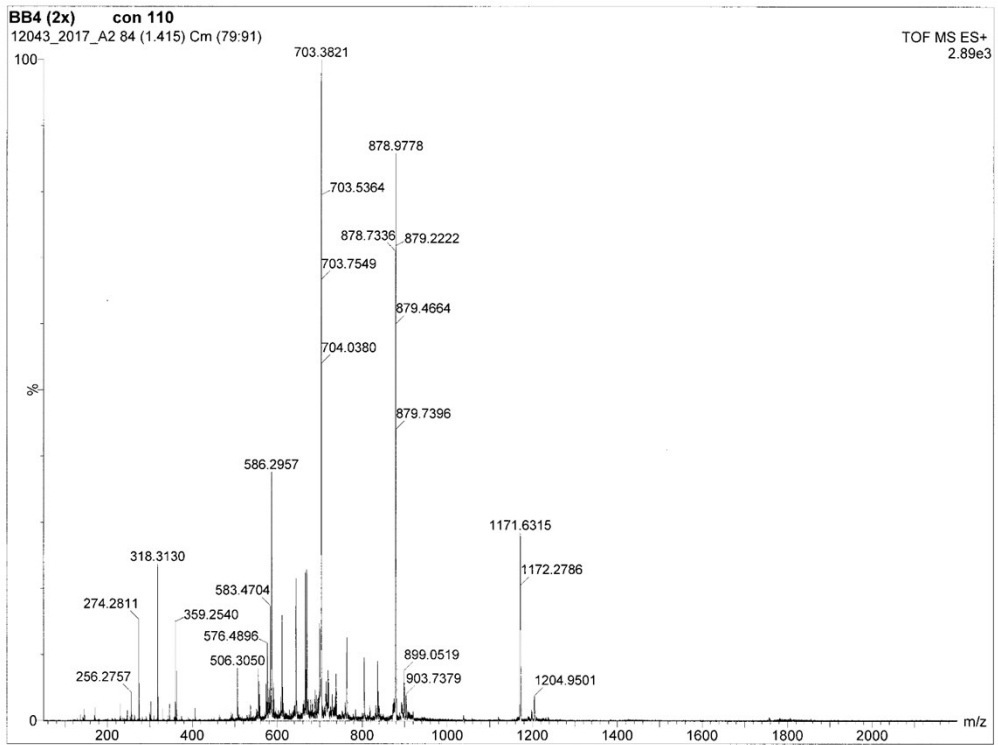
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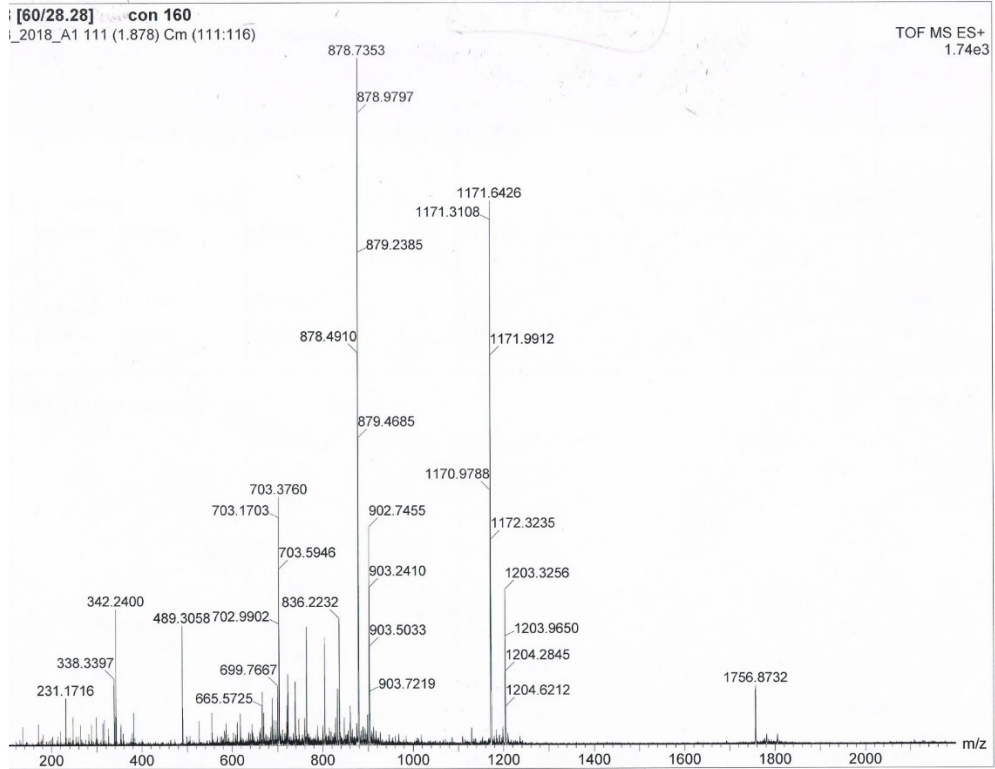
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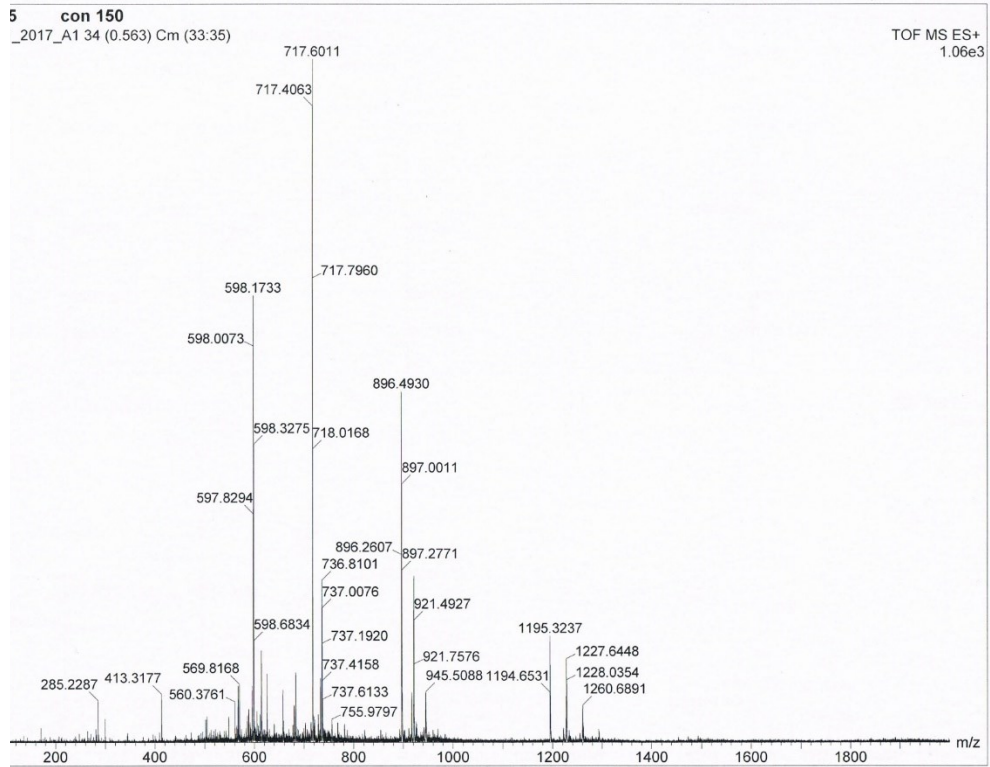
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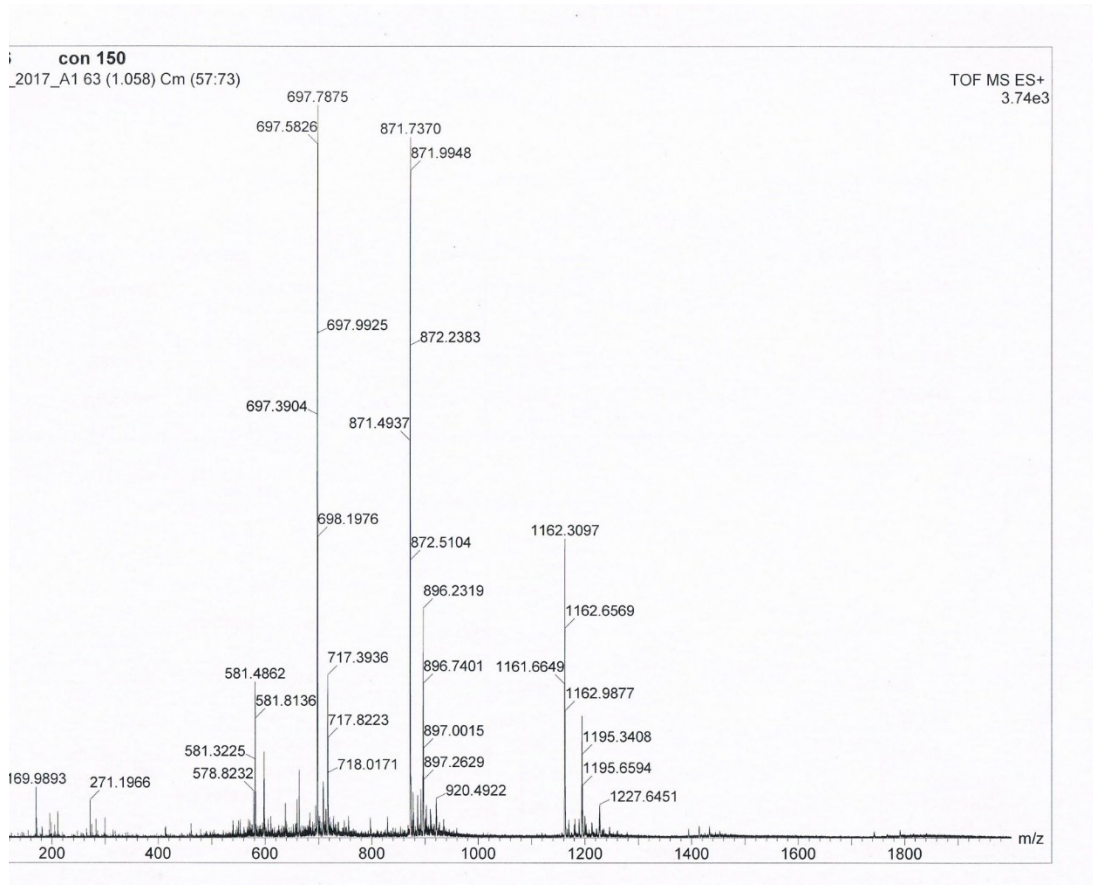
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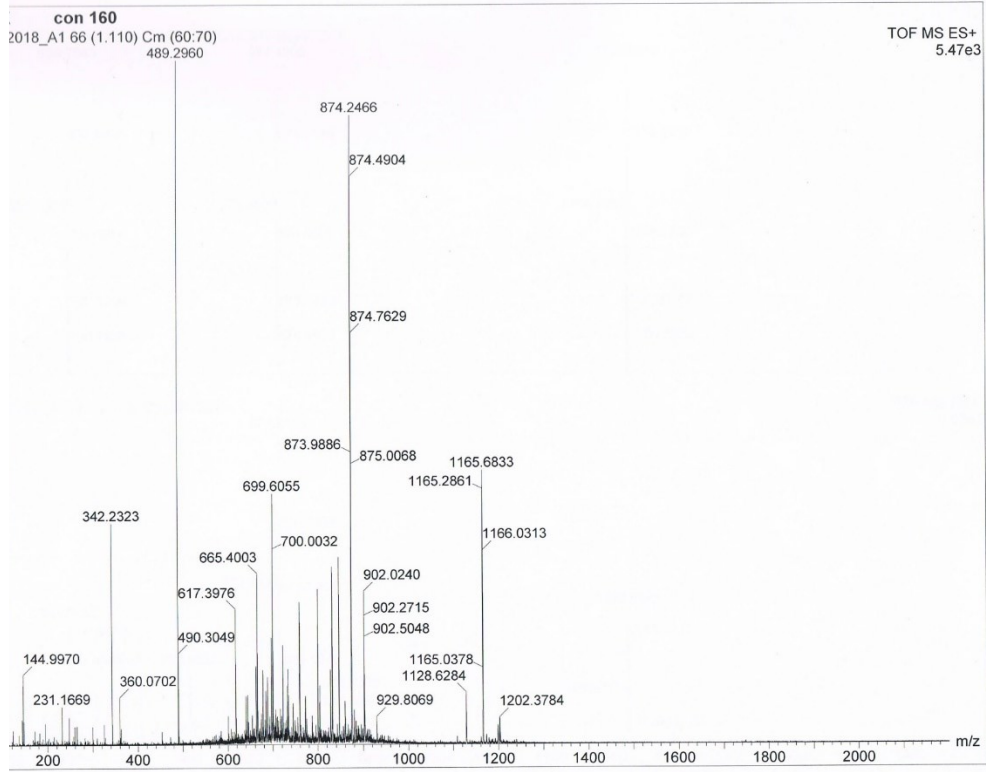
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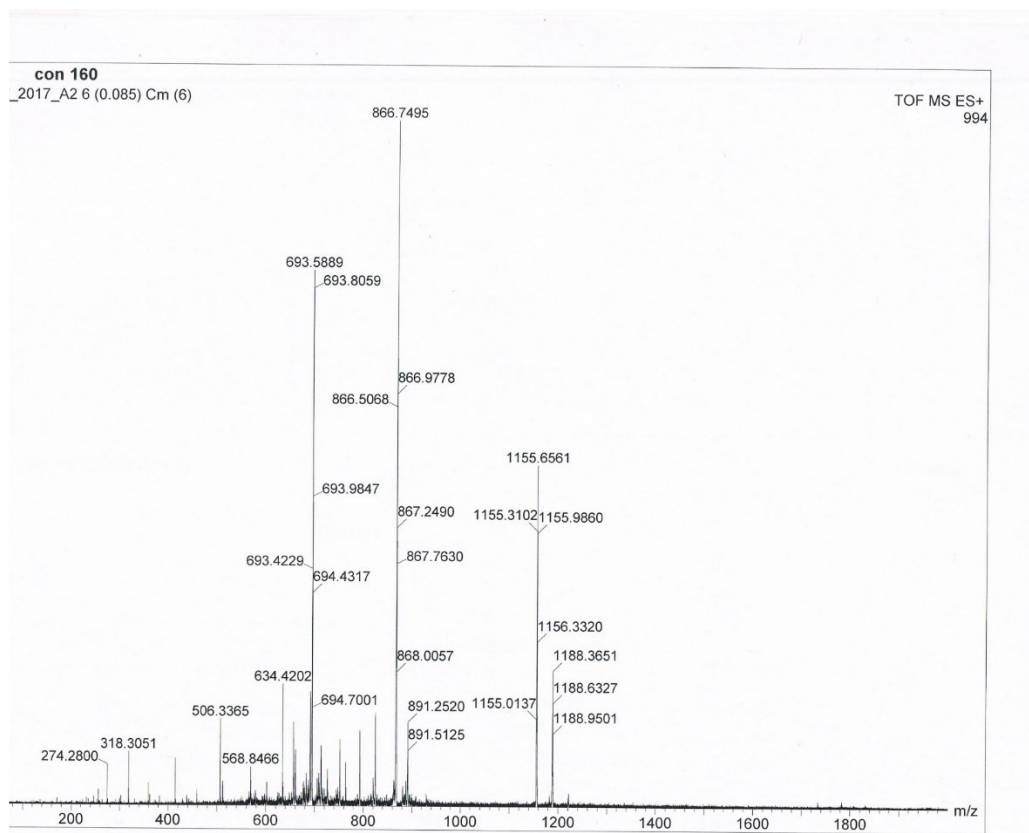
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